



UNIVERSITI PUTRA MALAYSIA

**CLONING, EXPRESSION, AND PURIFICATION OF RIBOFLAVIN
SYNTHASE FROM PHOTOBACTERIUM SP. J15**

GOL MOHAMMAD DORRAZEHI

FBSB 2014 6



**CLONING, EXPRESSION, AND PURIFICATION OF RIBOFLAVIN
SYNTHASE FROM *PHOTOBACTERIUM* SP. J15**

By

GOL MOHAMMAD DORRAZEHI

**thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

February 2014

A Special Dedication to
My Father for his unconditional love and support; for his believing in me and to lig
to find the best way.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia
in fulfilment of the requirement for the degree of Master of Science

**CLONING, EXPRESSION, AND PURIFICATION OF RIBOFLAVIN
SYNTHASE FROM *PHOTOBACTERIUM* SP. J15**

By

GOL MOHAMMAD DORRAZEHI

February 2014

Chair: Mohd Shukuri Bin Mohamad Ali, PhD
Faculty: Biotechnology and Biomolecular Sciences

Riboflavin synthase (RiS) is a homotrimeric enzyme that catalyzes the final reaction of riboflavin (vitamin B₂) biosynthesis from two molecules of 6,7-dimethyl-8-ribityllumazine. Gram-negative bacteria and certain yeasts are unable to absorb riboflavin or riboflavin derivatives from the environment. The RiS is nonexistent in humans while it only presents in bacteria and yeasts, which are dependent on their own production of riboflavin and will not survive in lack of the RiS function; hence the RiS is a potential target for the development of antimicrobial agents to target Gram-negative pathogens. The aim of this research was to clone and express the RiS in heterologous system and investigate the oligomerization of the RiS monomers. The RiS in current study is from a Malaysian isolated strain, *Photobacterium* sp. strain J15. Protein sequence alignment of this RiS shows high similarity to the RiS of *E. coli*. Analysis of the DNA sequence of this riboflavin synthase shows 6 rare codons, in which common *E. coli* strains are not able to supply the corresponding tRNAs, thus the *E. coli* strain Rosetta-gami B (DE3) pLysS was selected as the expression host. The gene encoding RiS was cloned into pET-32b(+) vector, which carried ampicillin resistance gene and utilized T7 promoter for high-level expression of a Trx-His tagged protein. The heterologous expression of RiS was performed by transformation of recombinant pET-32b(+) vector into *E. coli* Rosetta-gami B(DE3)pLysS. High-level expression of soluble RiS was optimized for expression temperature at 20 °C, inducer (IPTG) concentration at 1 mM, and time of induction for 20 h. The purification of the His-tagged enzyme was performed by affinity chromatography and was subjected to western-blot for verification. The purified fusion enzyme was cleaved by thrombin to remove Trx-His tag and purified by anion exchange chromatography to get the mature enzyme. Specific activity of mature RiS was significantly higher than fusion RiS. The size of mature RiS was estimated as 26.5 kDa by gel filtration chromatography. Site directed mutagenesis of I190V and chimerization of RiS was experimented to improve the trimer formation of quaternary structure of RiS but it didn't affect the trimerization of monomers, therefore, formation of trimer of RiS monomers was confirmed by cross-linking experiments. A melting point of 42.3 °C and a secondary structure composition of 13.6 % helix, 25.1 % beta, 12.5 % turn and 38.5 % random was measured by Circular-dichroism (CD) analysis. Through this work the RiS of *Photobacterium* sp. J15 was successfully cloned and expressed into heterologous system. The RiS is

found to function only in dimeric or trimeric form. The availability of functional enzyme by heterologous expression enables further characterization and structural studies towards drug development at pharmaceutical level.

Abstrak tesis yang dikemukakan kepada Senat of Universiti Putra Malaysia
Sebagai memenuhi keperluan untuk ijazah Master Sains

**PENGLONAN, EKSPRESI, DAN PENULENAN RIBOFLAVIN SINTASE
DARI *PHOTOBACTERIUM* SP. J15**

Oleh

GOL MOHAMMAD DORRAZEHI

Februari 2014

Pengerusi: Mohd Shukuri Bin Mohamad Ali, PhD

Fakulti: Bioteknologi dan Sains Biomolekul

Riboflavin sintase (RiS) merupakan enzim homotrimeric yang memangkin tindak balas terakhir dalam biosintesis riboflavin (vitamin B2) daripada dua molekul 6,7-dimetil-8-ribityllumazine. Bakteria Gram-negatif dan yis tidak mampu menyerap riboflavin atau hasilan riboflavin dari alam sekitar. RiS ini juga tidak wujud pada manusia dan ia hanya ditemui dalam bakteria dan yis, yang bergantung kepada pengeluaran riboflavin mereka sendiri dan tidak akan dapat hidup jika kekurangan fungsi RiS; oleh itu RiS menjadi sasaran untuk pembangunan agen antimikrob terutamanya patogen Gram-negatif. Tujuan kajian ini adalah untuk mengklon dan mengekspresi RiS dalam sistem heterologous dan akhirnya untuk menyiasat oligomerisasi RiS dari monomernya. RiS dalam kajian semasa ini adalah berasal dari strain terpencil di Malaysia; *Photobacterium* sp. J15. Penjajaran jujukan protin RiS ini menunjukkan persamaan yang tinggi kepada RiS dari *E. coli*. Analisis jujukan DNA daripada riboflavin sintase ini menunjukkan 6 codon yang jarang ditemui, di mana strain *E. coli* biasa tidak mampu untuk membekalkan tRNAs yang diperlukan, dengan itu strain *E. coli* Rosetta-Gami B (DE3) pLysS dipilih untuk menjadi perumah ekspresi. Gen yang mengekod RiS telah diklon ke dalam vektor pET-32b (+), yang membawa gen rintangan terhadap ampisillin dan menggunakan penganjur T7 untuk mengekspresi protein yang telah ditanda dengan Trx-His pada tahap tinggi. Ekspresi RiS secara heterologous dilakukan dengan menjalankan transformasi rekombinan pET-32b (+) ke dalam vektor *E. coli* Rosetta-Gami B (DE3) pLysS. Ekspresi RiS yang larut pada tahap tinggi telah dioptimumkan suhu eskpresinya pada 20 ° C, kepekatan pencetus (IPTG) sebanyak 1 mM, dan masa induksi selama 20 jam. Penulenan enzim yang ditanda dengan His telah dilakukan dengan kromatografi kecenderungan GDQ ZHVWHQK' WHODK GLODNXNDQ XQWXN SHQJHVDKDQ (C gabungan yang tulen telah dileraikan oleh thrombin untuk membuang penanda Trx-His dan ditulenan pula dengan kromatografi pertukaran anion untuk mendapatkan enzim yang matang. Aktiviti spesifik untuk RiS matang adalah jauh lebih tinggi daripada RiS yang bergabung. Saiz RiS yang matang dianggarkan sebanyak 26.5 kDa oleh kromatografi penurasan gel. Mutagenesis tapak yang diarah bagi I190V dan chimerisasi RiS telah dieksperimenkan untuk meningkatkan pembentukan trimer bagi struktur kuaterner RiS tetapi ia tidak menjejaskan trimerisasi daripada monomer, oleh itu pembentukan trimer daripada monomer RiS telah disahkan oleh eksperimen silang-hubung. Takat lebur pada 42.3 °C dan komposisi struktur

sekunder sebanyak 13.6% heliks, 25.1% beta, 12.5% lengkung dan 38.5% rawak, diukur dengan analisis circular-dichroism (CD). RiS daripada *Photobacterium* sp. J15 telah berjaya diklon dan diekspres di dalam sistem heterologous. RiS daripada *Photobacterium* sp. J15 didapati hanya akan berfungsi dalam bentuk dimeric atau trimeric. Kehadiran enzim yang berfungsi daripada ekspresi heterologous membolehkan pencirian lanjut dan kajian struktur dijalankan.

ACKNOWLEDGEMENTS

Foremost, I would like to express my sincere gratitude to my advisor Dr. Adam Leow Thean Chor for the continuous support of my MSc study and research, for his patience, motivation, enthusiasm, and immense knowledge. His guidance helped me in all the time of research and writing of this thesis.

I also would like to thank my supervisory committee, Dr. Mohd Shukuri Mohamad Ali, Prof. Abu Bakar Salleh, Prof. Raja Noor Zaliha Raja Abd Rahman and Dr. Normi Mohd Yahaya for the useful comments, remarks and engagement through the learning process of this research.

The Enzyme and Microbial Technology Research group was a place where I found the way of research. The substantial contribution of the group to complete this project is admirable. Thanks to all the principal lecturers and group members for their strict and extensive comments and sharing their research experience during weekly meeting. There was a light up to bright a dark way for me to move forward.

I would like to thank my labmates, who were like my sisters and brothers and are an unforgettable part of my life; Zul, Adi, Lutfi, Rahim, Zarir, Farzad, Hisham, Rofandi, Randa, Ely, Mehrnoosh, Ferrol, Baya, Fiza, Maya, Ati, Dura, Ira, Hoda, Farah Hani, Hafidza, Zurith, Su Hi, Jonathan, Fu, Firdos and all those who although not individually mentioned here but had contributed directly and indirectly to my research and thesis.

My appreciation to all loved ones, who have supported me throughout entire process, both by keeping me harmonious and helping me putting pieces together; to all my friends who shared happiness and their great times with me in Malaysia.

Last but not least, my deepest thanks to my parents, sisters and brothers who have always been there for me even from a very far distance they were just beside me. Their encouragements, unconditional love and support made me who I am. I will be grateful forever for your love.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd Shukuri Bin Mohamad Ali, PhD

Senior lecturer

Faculty of Biotechnology and Biomolecular Science

Universiti Putra Malaysia

(Chairman)

Adam Leow Thean Chor, PhD

Senior lecturer

Faculty of Biotechnology and Biomolecular Science

Universiti Putra Malaysia

(Member)

Abu Bakar Salleh, PhD

Professor

Faculty of Biotechnology and Biomolecular Science

Universiti Putra Malaysia

(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

Declaration by Graduate Student

I hereby confirm that:

- x this thesis is my original work;
- x quotations, illustrations and citations have been duly referenced;
- x this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- x intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- x written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- x there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Gol Mohammad Dorrazehi, GS31726

TABLE OF CONTENTS

	Page
ABSTRACT	iii
ABSTRAK	v
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xviii
 CHAPTER	
1 INTRODUCTION	1
1.1 Problem statement	2
1.2 Objectives	2
 2 LITERATURE REVIEW	 3
2.1 Riboflavin	3
2.2 Riboflavin Synthase	3
2.2.1 Application of riboflavin synthase	5
2.2.2 Heterologous expression of riboflavin synthase	5
2.3 Multimeric enzymes	7
2.4 Mechanism and structure of riboflavin synthase	7
2.5 Site directed mutagenesis	13
2.6 Protein engineering and chimeragenesis	14
2.7 Cross-linking of protein-protein interactions	14
2.8 Sources of riboflavin synthase	14
2.8.1 <i>Photobacterium</i>	15
 3 MATERIALS AND METHODS	 16
3.1 Materials	16
3.2 General methods	16
3.3 Strains and plasmids	16
3.4 Cloning of RiS gene using pTrcHis TOPO TA Expression Kit	17
3.4.1 Extraction of genomic DNA of <i>Photobacterium</i> sp. strain J1517	17
3.4.2 Amplification of riboflavin synthase gene	17
3.4.3 Cloning of RiS gene into pTrcHis vector	18
3.4.4 Preparation of <i>E. coli</i> competent cells	18
3.4.5 Transformation of Recombinant pTrcHis into <i>E. coli</i> Top10	18
3.4.6 Direct PCR analysis of positive clones	18

3.4.7	Preparation of glycerol stock from positively transformed colony	19
3.5	Expression of RiS gene in <i>E. coli</i> Top10	19
3.6	SDS-PAGE analysis of riboflavin synthase expression in <i>E. coli</i> Top10	19
3.7	Cloning of RiS gene into pET-32b(+) Vector	20
3.7.1	Amplification of RS gene	20
3.7.2	Double Digestion of PCR product and Vector	20
3.7.3	Purification of digested RiS gene and pET-32b(+) vector	21
3.7.4	Ligation of RiS gene into pET32b(+) Vector	21
3.7.5	Heat-Shock transformation of <i>E. coli</i> Top10	21
3.7.6	Colony PCR	21
3.7.7	Sequencing of recombinant plasmid (pET/RiS)	21
3.8	Expression of RiS gene in <i>E. coli</i> Rosetta-gami B(DE3)pLysS	21
3.8.1	Extraction of recombinant PET-32b(+) plasmid	21
3.8.2	Transformation of recombinant plasmid pET/RiS into <i>E. coli</i> Rosetta-gami B(DE3)pLysS	22
3.9	Expression of RiS in <i>E. coli</i> Rosetta-gami B(DE3)pLysS	22
3.9.1	Optimization of inducer concentration effect on RiS expression	22
3.9.2	Optimization of Temperature effect on RiS Expression	22
3.9.3	Optimization of incubation time of RiS expression	22
3.10	Purification of recombinant riboflavin synthase	23
3.10.1	Preparation of crude lysate	23
3.10.2	Affinity purification of RiS	23
3.11	Western-Blot analysis of purified riboflavin synthase	23
3.12	Tag cleavage and purification of mature riboflavin synthase	23
3.12.1	Cleavage of fusion RiS by thrombin	23
3.12.2	Desalting of thrombin cleavage mixture	24
3.12.3	Anion-exchange chromatography purification of mature RiS	24
3.13	Assay of riboflavin synthase	24
3.14	Gel filtration and size determination of mature RiS	24
3.15	Circular-Dichroism (CD) analysis of mature RiS	25
3.16	Protein modeling and sequence analysis of RiS	25
3.16.1	Multiple sequence alignment of riboflavin synthase	25
3.16.2	Protein structure analysis	25
3.17	Protein engineering for assembly of quaternary structure	25
3.17.1	Site Directed Mutagenesis (SDM)	25
3.17.1.1	Transformation of mutant plasmid pET/I190V into <i>E. coli</i> Rosetta-gami B(DE3)pLysS	26
3.17.1.2	Expression of the mutant RiS	26
3.17.1.3	Analysis of trimeric formation of RiS	26
3.17.2	Chimerization of RiS by replacing the C-terminal helix	26
3.17.3	Cross-linking of RiS monomers	27
4	RESULTS AND DISCUSSION	28
4.1	Cloning of RiS Gene in pTrcHis vector	28
4.1.1	Genomic DNA extraction	28

4.1.2	PCR cloning of RiS gene	29
4.1.3	Analysis of positive transformants	30
4.2	Expression of RiS gene in <i>E. coli</i> Top10	32
4.2.1	SDS-PAGE analysis of RiS expression in <i>E. coli</i> Top10	32
4.3	Examining the codon usage of RiS Gene	33
4.4	Restriction site analysis of RiS gene and Primer design	34
4.5	Cloning of RiS gene in pET-32b(+) system	35
4.5.1	RiS Gene amplification by RiS primers	35
4.5.2	Double digestion and ligation of RiS gene and vector	35
4.5.3	Transformation of recombinant plasmid pET/RiS into <i>E. coli</i> Top10	36
4.5.4	Analysis of positive transformants	36
4.6	Transformation of recombinant pET/RiS into <i>E. coli</i> Rosetta-gami B(DE3)pLysS	38
4.6.1	Expression of RiS in <i>E. coli</i> Rosetta-gami B(DE3)pLysS	39
4.6.2	SDS-PAGE analysis of expressed proteins	40
4.7	Optimization of temperature effect on RiS expression	41
4.8	Optimization of IPTG concentration on RiS expression	41
4.9	Optimization of incubation time on RiS expression	45
4.10	Purification of His-tagged RiS	46
4.11	Western blot analysis of purified RiS	47
4.12	Activity assay evaluation of RS enzyme	48
4.13	Thrombin cleavage of fusion RiS	49
4.14	Size determination of mature RiS by gel filtration	51
4.15	Circular Dichroism analysis of mature RiS	52
4.16	Sequence analysis and Protein Modelling of RiS	53
4.17	Site directed mutation of RiS	57
4.18	Chimerization of RiS from <i>Photobacterium</i> sp. J15	60
4.19	Cross-linking of RiS from <i>Photobacterium</i> sp. J15	61
5	CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	
5.1	General discussion	63
5.2	Conclusion	63
5.2	Recommendation for future research	64
	REFERENCES	65
	APPENDICES	70
	BIODATA OF STUDENT	77
	LIST OF PUBLICATIONS	78