



UNIVERSITI PUTRA MALAYSIA

**IN VITRO PROPAGATION OF CITRUS HYSTRIX AND ASSESSMENT OF
GENETIC UNIFORMITY USING RAPD MARKERS**

ENG WEE HIANG

FP 2013 31



***IN VITRO PROPAGATION OF CITRUS HYSTRIX
AND ASSESSMENT OF GENETIC UNIFORMITY
USING RAPD MARKERS***

ENG WEE HIANG



**MASTER OF SCIENCE
UNIVERSITI PUTRA MALAYSIA**

2013



***IN VITRO PROPAGATION OF CITRUS HYSTRIX AND ASSESSMENT OF
GENETIC UNIFORMITY USING RAPD MARKERS***

By

ENG WEE HIANG



**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

July 2013

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia





Dedicated to:

My Parents
George Ngu

Abstract of thesis presented to the Senate of Universiti Putra Malaysia
in fulfilment of the requirement for the degree of Master of Science

IN VITRO PROPAGATION OF CITRUS HYSTRIX AND ASSESSMENT OF GENETIC UNIFORMITY USING RAPD MARKERS

By

ENG WEE HIANG

July 2013

Chairperson: Associate Professor Maheran Abdul Aziz, PhD

Faculty: Agriculture

Recent studies in *Citrus hystrix* largely focused on its usage in citriculture, pharmaceutical and nutritional values. This newly emerged plant urgently needs biotechnological studies especially on the establishment of its micropropagation protocol. The objectives of this study are (a) to develop an efficient micropropagation system using various juvenile explants of *C. hystrix* seedlings, (b) to elucidate genetic uniformity of plantlets derived from the juvenile explants using RAPD markers and (c) to develop an effective sterilization technique and induction of multiple shoot formation from nodal segments of mature field grown *C. hystrix*.

In vitro seedling and field-grown mature *C. hystrix* was used as source of explants to assess regeneration capability. In this study, various concentrations of BAP were assessed to determine the optimum concentration for regeneration. Leaves abscission occurred during regeneration stage. To overcome the problem, Ca-gluconate and

silver nitrate were amended into medium containing optimized concentration of BAP. In rooting stage, continuous auxin treatment and auxin pulse treatment were tested to determine efficient rooting method. Types of potting mixture were assessed to determine the best potting mixture for survival of deflasked *in vitro* plantlets. For sterilization study using field-grown mature plant, different types and combination of antibiotics were tested to reduce latent bacterial contamination. Medium with combination of BAP, GA₃ and AgNO₃ were tested to determine the best medium for shoot regeneration for field-grown mature plant. RAPD markers were used for screening uniformity of regenerants from *in vitro* seedling.

The optimum regeneration medium for shoot tip was MS medium + 2.22 µM BAP + 4 mM Ca-glu + 20 µM AgNO₃ + 30 g/L sucrose, inducing four shoots per explant. For epicotyls, hypocotyls, primary roots and cotyledons, the optimum regeneration medium was MS medium + 2.22 µM BAP + 10 µM AgNO₃ + 30 g/L sucrose which induced five, four, three and two shoots, respectively. For *in vitro* rooting, auxin pulse treatment with 9840 µM IBA in MS medium for 16 h prior to transfer to MS medium + 20 µM AgNO₃ produced significantly the highest number of roots (2 roots) and highest rooting percentage (87.50%). In the acclimatization study, medium consisting of soil : Peatgro : sand (1 : 1 : 1) was the best attaining a survival percentage of plantlets at 83.33%. Genetic stability of the plantlets was assessed using RAPD markers. Most of the plantlets were identical to the mother plant based on RAPD banding pattern. Eighteen out of twenty samples had Jaccard's similarity coefficient of 1.0000 indicating 90% of the regenerants were likely to be clones.

For effective sterilization of nodal segments derived from mature field grown *C. hystrix* the use of 500 mg/L cefotaxime as part of the surface sterilization procedure successfully reduced percentage of latent bacterial contamination and resulted in significantly the highest survival percentage (61.25%) after eight weeks of culture. The optimised regeneration medium for nodal segments of mature plants of *C. hystrix* was MS medium + 2.22 µM BAP + 20 µM AgNO₃ + 30 g/L sucrose inducing three shoots per explant. Through this study, an efficient micropropagation system has been developed that could provide a mean for mass propagation, and as a tool in genetic modification and *in vitro* germplasm conservation of *C. hystrix*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**PEMBIAKAN *IN VITRO* CITRUS HYSTRIX DAN PENILAIAN
KESAMAAN GENETIK MELALUI RAPD**

Oleh

ENG WEE HIANG

Julai 2013

Pengerusi: Profesor Madya Maheran Abdul Aziz, PhD

Fakulti: Pertanian

Kajian terkini dalam *Citrus hystrix* lebih tertumpu kepada kegunaannya dalam penanaman sitrus, perubatan dan nilai pemakanan. Tanaman ini amat memerlukan kajian bioteknologi terutamanya dalam pembangunan protokol pembiakan mikro. Objektif kajian ini adalah untuk (a) membangunkan sistem pembiakan mikro yang cekap dengan menggunakan pelbagai eksplan juvenil daripada anak benih *C. hystrix*, (b) menilai kesamaan genetik anak pokok daripada eksplan juvenil dengan menggunakan penanda RAPD dan (c) membangunkan teknik sterilisasi yang efektif dan mengaruh pembentukan pucuk berganda pada segmen nod *C. hystrix* yang diperoleh dari pokok matang di ladang.

Anak benih *in vitro* dan pokok matang dari ladang bagi *C. hystrix* telah digunakan sebagai bahan tanaman untuk menilai keupayaan pertumbuhan semulanya. Dalam

kajian ini, pelbagai kepekatan BAP telah dinilai untuk menentukan kepekatan BAP yang paling sesuai untuk pertumbuhan semula. Keguguran daun berlaku ketika peringkat pertumbuhan semula. Bagi mengatasi masalah ini, Ca-gluconate dan argentum nitrat telah ditambahkan ke dalam medium yang mengandungi kepekatan BAP yang terbaik. Di peringkat pengakaran, rawatan auksin berterusan dan rawatan auksin sementara dikaji bagi mengenalpasti kaedah merangsang pengakaran terbaik. Pelbagai jenis campuran medium memasu dinilai bagi menentukan campuran terbaik untuk anak pokok yang dikeluarkan dari berkas kultur. Bagi kajian pembasmian pencemar-pencemar pokok matang dari ladang, pelbagai jenis antibiotik dan campuran antibiotik-antibiotik telah digunakan untuk mengurangkan pencemaran bakteria lanjutan. Medium yang terdiri daripada kombinasi BAP, GA₃ dan AgNO₃ dinilai bagi menentukan medium yang paling sesuai untuk pertumbuhan semula pokok matang. Penanda-penanda RAPD digunakan untuk menilai keseragaman genetik anak-anak pokok hasil pertumbuhan semula *in vitro*.

Medium pertumbuhan semula yang optimum untuk pucuk adalah medium MS + 2.22 μM BAP + 4 mM Ca-glu + 20 μM AgNO₃ + 30 g/L sukrosa yang menghasilkan empat pucuk per eksplan. Bagi epikotil, hipokotil, akar utama dan kotiledon, medium regenerasi optimum adalah medium MS + 2.22 μM BAP + 10 μM AgNO₃ + 30 g/L sukrosa masing-masing menghasilkan empat, empat, tiga dan dua pucuk. Untuk pengakaran, rawatan auksin sementara dengan 9840 μM IBA dalam medium MS selama 16 jam sebelum dipindahkan ke medium MS + 20 μM AgNO₃ menghasilkan jumlah akar (2 akar) dan peratusan pengakaran (87.50%) tertinggi. Dalam kajian penyesuaian, medium yang terdiri daripada tanah : Peatgro : pasir (1 : 1 : 1)

merupakan campuran yang terbaik dengan mencapai peratusan hidup tertinggi (83.33%) untuk anak pokok berakar. Kesamaan genetik anak pokok dinilai dengan menggunakan penanda RAPD. Berdasarkan corak jalur RAPD, didapati kebanyakan anak pokok adalah seiras dengan pokok induk. Lapan belas daripada dua puluh sampel menpunyai nilai kesamaan koefficient Jacaard 1.0000 menunjukkan 90% anak pokok berkemungkinan adalah klon.

Untuk pensterilan segmen nod *C. hystrix* dari pokok matang di ladang penggunaan 500 mg/L cefotaxime sebagai sebahagian daripada prosidur pensterilan telah berjaya menurunkan peratusan pencemaran bakteria dan menghasilkan peratusan hidup tertinggi (61.25%) yang ketara selepas lapan minggu. Medium regenerasi optimum untuk keratan nod dari pokok matang adalah medium MS + 2.22 μM BAP + 20 μM AgNO₃ + 30 g/L sukrosa menghasilkan tiga pucuk per eksplan. Melalui kajian ini, sistem pembiakan mikro yang cekap telah dibangunkan sesuai untuk kegunaan pembiakan bahan tanaman secara besar-besaran, dan sebagai kaedah pengubahsuaian genetik dan pengekalan germplasma *C. hystrix* secara *in vitro*.

ACKNOWLEDGEMENTS

I would like to express my sincere gratitude to members of the Supervisory Committee, Associate Professor Dr. Maheran Abdul Aziz from Department of Agriculture Technology and Associate Professor Dr. Uma Rani A/P Sinniah from Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia for their valuable advice and encouragements during the course of my study and preparation of this manuscript.

I am indebted to Ministry of Education Malaysia for granting me study leave and financial support to pursue my Master Degree at Universiti Putra Malaysia. Deepest thanks to my beloved parents for their endless encouragement and support during the tenure of my study. My appreciation is done to Mr. George Ngu K.K. for sources of inspirations in achieving my dream in higher education.

Finally, I am blessed to be able to carry out my research among industrious laboratory staffs and intelligent research teams at *In vitro* Laboratory of Department of Agriculture Technology. Their assistances and friendliness are indispensable and unforgettable.

I certify that a Thesis Examination Committee has met on 15 July 2013 to conduct the final examination of Eng Wee Hiang on his thesis entitled "*In Vitro* Propagation of *Citrus hystrix* and Assessment of Genetic Uniformity Using RAPD Markers" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Siti Nor Akmar Abdullah, PhD

Professor

Faculty of Agriculture
Universiti Putra Malaysia
(Chairperson)

Saleh Kadzimin, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Nur Ashikin Psyquay Abdullah, PhD

Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Normah Mohd Noor, PhD

Professor
Institute of Systems Biology (INBIOSIS)
Universiti Kebangsaan Malaysia
(External Examiner)

NORITAH OMAR, PhD
Associate Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 19 September 2013

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Maheran Abdul Aziz, PhD

Associate Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Chairperson)

Uma Rani A/P Sinniah, PhD

Associate Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: 15 July 2013

Name and Matric No.: ENG WEE HIANG GS27971

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	v
ACKNOWLEDGEMENTS	viii
APPROVAL	ix
DECLARATION	xi
TABLE OF CONTENTS	xii
LIST OF TABLES	xv
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xxi
CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Objectives	4
2 LITERATURE REVIEW	5
2.1 <i>Citrus hystrix</i>	5
2.1.1 Taxonomy and Characteristics	5
2.1.2 Potential of <i>Citrus hystrix</i>	7
2.2 Methods of Propagation in Citrus	8
2.2.1 Conventional Propagation	8
2.2.2 Micropropagation	9
2.3 Sterilization	10
2.3.1 Types of Sterilant	11
2.3.2 Antibiotics	12
2.4 Shoot Proliferation	14
2.4.1 Types of Explants	14
2.4.2 Plant Growth Regulators	16
2.5 Rooting	18
2.6 Acclimatization	20
2.7 Calcium in Micropropagation	22
2.8 Ethylene and Effects on Tissue Culture	24
2.8.1 Leaf Abscission	25
2.8.2 Leaf Expansion	25
2.8.3 Shoot Production	26
2.8.4 Root Production	26
2.8.5 Unorganized Growth	27
2.8.6 Overcoming Effects of Ethylene	27
2.9 Somaclonal Variation in Tissue Cultured Plants	28

2.9.1 RAPD	30
2.9.2 Advantages and disadvantages of RAPD	31
3 SHOOT REGENERATION FROM VARIOUS EXPLANTS OF <i>IN VITRO</i> SEEDLING	33
3.1 Introduction	33
3.2 Materials and Methods	35
3.2.1 Preparation of Explants	35
3.2.2 Medium Preparation and Treatments	36
3.2.3 Explant Culture Conditions and Incubation	37
3.2.4 Experimental Design, Parameter Recorded and Data Analysis	37
3.3 Results and Discussion	38
3.3.1 Shoot Regeneration from Shoot	38
3.3.2 Shoot Regeneration from Epicotyl	48
3.3.3 Shoot Regeneration from Cotyledon	55
3.3.4 Shoot Regeneration from Hypocotyl	62
3.3.5 Shoot Regeneration from Primary Root	69
3.4 General Discussion	76
3.5 Conclusion	81
4 IN VITRO ROOTING AND ACCLIMATIZATION OF REGENERANTS	82
4.1 Introduction	82
4.2 Material and Methods	84
4.2.1 <i>In Vitro</i> Rooting Study	84
4.2.2 Acclimatization Study	88
4.3 Results	90
4.3.1 <i>In Vitro</i> Rooting Study	90
4.3.2 Acclimatization Study	96
4.4 Discussion	98
4.4.1 <i>In Vitro</i> Rooting Study	98
4.4.2 Acclimatization Study	100
4.5 Conclusion	102
5 GENETIC UNIFORMITY OF REGENERANTS OF <i>C. hystrix</i> USING RAPD	103
5.1 Introduction	103
5.2 Materials and Methods	105
5.2.1 Plant materials	105
5.2.2 Total DNA Isolation	105
5.2.3 Total DNA Assessment	107
5.2.4 RAPD Assay	107
5.2.5 Gel Scoring	111
5.2.6 Data Analysis	111
5.3 Results	114

5.4 Discussion	123
5.5 Conclusion	127
6 SURFACE STERILIZATION AND SHOOT REGENERATION OF NODAL SEGMENTS FROM FIELD GROWN MATURE PLANTS	128
6.1 Introduction	128
6.2 Material and Methods	130
6.2.1 Surface Sterilization of Nodal Segments	130
6.2.2 Shoot Regeneration from Nodal Segments	134
6.3 Results	136
6.3.1 Surface Sterilization of Nodal Segments	136
6.3.2 Shoot Regeneration from Nodal Segments	141
6.4 Discussion	145
6.4.1 Surface Sterilization of Nodal Segments	145
6.4.2 Shoot Regeneration from Nodal Segments	148
6.5 Conclusion	151
7 GENERAL DISCUSSION, GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	152
7.1 General Discussion	152
7.2 General Conclusion	158
7.3 Recommendation for Future Research	160
REFERENCES	161
APENDICES	176
BIODATA OF STUDENT	201
LIST OF PUBLICATIONS	202