



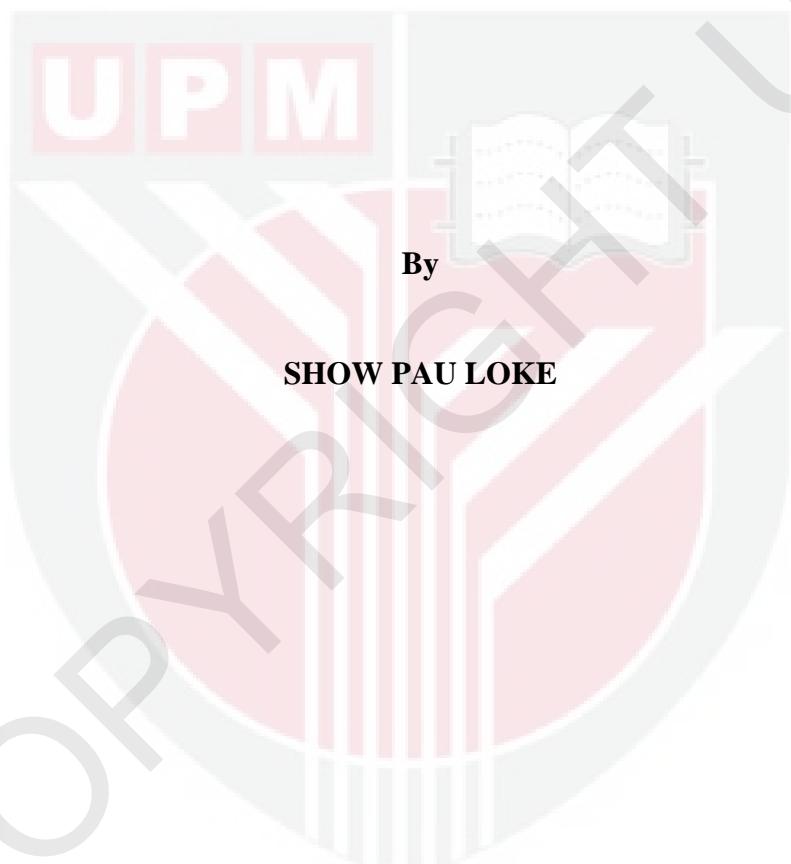
UNIVERSITI PUTRA MALAYSIA

***RECOVERY OF LIPASE FROM BURKHOLDERIA SP.
USING AQUEOUS TWO-PHASE SYSTEMS***

SHOW PAU LOKE

FK 2012 15

**RECOVERY OF LIPASE FROM *BURKHOLDERIA* SP. USING AQUEOUS
TWO-PHASE SYSTEMS**



**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in fulfilment of the requirements for the Degree of Doctor of Philosophy**

July 2012

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Doctor of Philosophy

**RECOVERY OF LIPASE FROM *BURKHOLDERIA* SP. USING AQUEOUS
TWO-PHASE SYSTEMS**

By

SHOW PAU LOKE

July 2012

Chairman : Mohd Shamsul Anuar, PhD

Faculty : Engineering

Conventional recovery methods for lipase are tedious and require several rounds of recovery steps. Therefore, the development of a cost-effective and recyclable protocol for the recovery of lipase is essential. Aqueous two-phase system (ATPS) can be used as an attractive alternative for the recovery of lipase from complex feedstock. This research is focused on the design of an ATPS protocol as a simplified and rapid recovery technique for the microbial lipase of *Burkholderia* sp. ST8. A multifactor experimental design based on a ‘change-one-factor-at-a-time’ approach was employed to study the effects of lipase production. A high lipase activity was achieved at a 250 rpm agitation speed for 36 hours of fermentation time in a cultivation medium containing 0.1 % (v/v) of Tween 80, 0.3 % (w/v) of nutrient broth and 0.1 % (w/v) of CaCl₂ at a pH of 9 with a volume ratio of 20:80 of medium volume to free volume of the flask which resulted in a high average production of 48.3 U/mL.

Three different novel techniques for the direct recovery of lipase based on sustainable ATPS have been developed. The recycling concept in the system is based on the principals of green chemistry, with good efficiency and economic viability. The first novel method is a recycling hydrophilic organic solvent/inorganic salt aqueous two-phase flotation (ATPF) system, which integrates solvent sublation (SS) and aqueous two-phase extraction (ATPE) in the recovery of lipase from fermentation broth. A purification factor of 14 and a lipase yield of 99 % were achieved in the optimized ATPF system with recovery of alcohol and salt are 70% and 61 %, respectively. The second and third lipase recovery techniques were explored by using a recyclable temperature-induced polymer, an ethylene oxide-propylene oxide (EOPO) polymer in an ATPF system as well as an ATPS. As for the second method, a recycling EOPO/ammonium sulphateATPF was developed for the recovery of lipase from fermentation broth. Under the optimal conditions, the average yield and purification factor were 98.5 % and 15, respectively. It was also demonstrated that EOPO and salt was recovered up to 91 % and 75 % in the ATPF system. Direct lipase recovery using recycling an EOPO/potassium phosphate ATPS was studied as the third protocol in this thesis. The average purification factor of lipase and the yield obtained from four successive purifications were 15.3 and 99.1 %, respectively. There was no significant difference in using fresh or recycled chemicals in these three novel methods based on ATPS. The choice of method in lipase recovery based on ATPS is subjected to the requirement of the purity and cost of the system. Hence, the simplicity and effectiveness of these three lipase recovery methods based on sustainable ATPS were proven in this study.

Lastly, an extractive fermentation technique was employed using a thermoseparating reagent to form a two-phase system for simultaneous cell cultivation and downstream processing. A 10 % (w/w) solution of EOPO with a molecular mass of 3900 g/mol and a pH of 8.5, a 200 rpm speed, and 30 °C were selected as the optimal conditions for lipase production (55 U/ml). Repetitive batch fermentation was performed by continuous replacement of the top phase every 24 hours, which resulted in an average cell growth mass of 4.8 g/L for 10 extractive batches over 240 hours. The dry cell mass in the bioreactor was 22 % higher than that in the flasks. *Burkholderia* sp. ST8 lipase was successfully produced and recovered by using thermoseparating reagents in a single step - extractive fermentation.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMULIHANLIPASE DARIPADA *BURKHOLDERIA SP.* BERDASARKAN
SISTEM DUA-FASA AKUEUS**

Oleh

SHOW PAU LOKE

Julai 2012

Pengerusi : Mohd Shamsul Anuar, PhD

Fakulti : Kejuruteraan

Kaedah-kaedah konvensional untuk pemulihan lipase adalah memerlukan dan memerlukan beberapa langkah pemulihan yang rumit. Oleh sebab itu, pembangunan protokol pemulihan lipase yang berkos efektif serta berupaya dikitar semula adalah amat mustahak. Sistem dua-fasa akueus (SDFA) boleh digunakan sebagai kaedah alternatif untuk pemulihan lipase daripada stok suapan yang kompleks. Dengan itu, kajian penyelidikan ini memfokuskan kepada reka-bentuk protokol SDFA sebagai teknik pemulihan lipase daripada mikroorganisma *Burkholderia sp.* ST8. Eksperimen yang berdasarkan pendekatan 'perubahan-satu-faktor-pada-satu-masa' telah diaplikasikan untuk mengoptimumkan penghasilan lipase. Pada keadaan optimum iaitu 250 putaran per minit pergolakan kelajuan selama 36 jam masa penapaian dalam medium yang mengandungi 1 % (i/i) Tween 80, 0.3 % (i/i) nutrien bubur dan 0.1 % (b/i) CaCl₂ pada pH 9 dengan nisbah kapasiti 20:80 medium kepada kelantangan kelalang, menghasilkan lipase dengan purata sebanyak 48.3 U/mL dalam kajian ini.

Tiga teknik pemulihan terus lipase yang berbeza berdasarkan lestari SDFA telah dikajikan. Konsep kitar semula adalah berprinsipkan kimia hijau, baik dari segi kecekapan dan ekonomi. Kaedah pertama pemulihan lipase yang novel adalah pengitaran semula pelarut organik hirofilik/garam bukan organik pengapungan dua-fasa akueus (PDFA) yang mengintegrasikan kaedah sublation pelarut (SP) dan pengekstrakan dua-fasa akueus dalam pemulihan lipase daripada media penapaian. PDFA telah dioptimumkan pada keadaan 40 mL isipadu 50 % (b/b) 2-propanol, 1.0 L isipadu 250 g/L kalium fosfat, pH 8.5, 100 % (i/i) stok suapan, 30 mL/min kadar aliran N₂ selama 30 minit dalam tiub kolorimeter berjejari 8 cm dengan keliangan G4 (5-15 µm) kaca cakera tersinter. Faktor penulenan pada 14 dan hasil lipase sebanyak 99 % telah dicapai dalam PDFA ini. Teknik pemulihan lipase yang kedua dan ketiga telah dieksplorasi dengan menggunakan induksi suhu polimer, etilena oksida-propilena oksida (EOPO) polimer dalam PDFA dan SDFA. Bagi kaedah kedua, EOPO/ammonium sulfat PDFA untuk pemulihan lipase daripada media penapaian telah dihasilkan. Pada keadaan optimum, purata kecekapan pemisahan dan faktor penulenan masing-masing adalah sebanyak 76 % dan 15. Keadaan ini juga menunjukkan bahawa EOPO dalam PDFA telah dipulihkan sebanyak 91 %. Pemulihan terus lipase dengan penggunaan kitaran semula EOPO/kalium fosfat SDFA telah dikaji sebagai protokol ketiga dalam tesis ini. Pada SDFA ini, lipase yang tulen telah dipindahkan kepada fasa polimer yang terdiri daripada EOPO 3900 dengan kepanjangan talian garis ikatan 48.5 % (b/b), nisbah isipadu sebanyak 2.3 dan stok suapan sebanyak 20 % (b/b) pada pH 7. Purata faktor penulenan lipase dan hasilan yang diperoleh dari keempat-empat pemulihan yang berturutan adalah masing-masing 15.3 dan 99.1 %. Di samping itu, didapati bahawa tiada perbezaan yang ketara dengan menggunakan bahan kimia yang segar atau dikitar semula dalam

tiga teknik pemulihan lipase berdasarkan SDFA. Pemilihan teknik adalah berdasar keperluan darjah ketelunan and cost yang mampu. Oleh yang demikian, kecekapan dan keberkesanan dalam tiga kaedah pemulihan lipase berdasarkan SDFA telah terbukti dalam kajian ini.

Akhir sekali, teknik penapaian ekstraktif telah diaplikasi dengan menggunakan reagen induksi suhu untuk membentuk satu sistem yang mempunyai kebaikan penghasilan sel dan pemulihan lipase serentak. Satu pelarut 10 % (b/b) EOPO dengan jisim molekul 3900 g/mol dan pH 8.5, kelajuan 200 putaran per minit dan 30 °C telah dipilih sebagai syarat-syarat optimum untuk penghasilan lipase (55 U/ml). Kelompok fermentasi yang berulangan telah dikaji dengan penggantian fasa pada setiap 24 jam, yang menyebabkan pertumbuhan sel purata jisim sebanyak 4.8 g/L untuk 10 kelompok ekstraktif iaitu, 240 jam. Berat kereing sel di dalam bioreaktor adalah 22 % lebih tinggi daripada dalam kelalang. *Burkholderia* sp. ST8 lipase telah berjaya dihasilkan dan ditulenan dengan menggunakan reagen induksi suhu polimer dengan hanya satu langkah - penapaian ekstraktif.

ACKNOWLEDGEMENTS

I wish to express my utmost gratitude to my main supervisor, Dr. Mohd Shamsul Anuar for his guidance and support during the course of this research. I would also like to thank my supervisory committee members, Prof. Dr. Ling Tau Chuan, Prof. Dr. Arbakariya Ariff, and Assoc. Prof. Dr. Yus Aniza Yusof for their encouragement and excellent advice.

Special thanks to all the laboratory officers of the Department of Process and Food Engineering (KPM) and the Laboratory of Immuno Therapeutics and Vaccines (LIVES) for their consistent and kind assistance. Further, I would like to acknowledge Dr. Hii Siew Ling from the Universiti Tunku Abdul Rahman for providing the bacterial strain used in this research.

I wish to extend my sincere appreciation to my labmates and friends (Dr. Ooi Chien Wei, Dr. Ramanan, Dr. Tan Joo Shun, Tam, Lo, Fatemeh, Fadzlie, Yu Kiat, Teck Kim, Yin Hui and Grace Ng) for frequently helping me and sharing their invaluable knowledge with me. Last but not least, I would like to thank my family for their earnest love, tolerance and sacrifice. I could not have completed this thesis without all of them.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Shamsul Anuar, PhD

Senior Lecturer
Faculty of Engineering
Universiti Putra Malaysia
(Chairman)

Ling Tau Chuan, PhD

Professor
Faculty of Sciences
University of Malaya
(External Member)

Arbakariya Ariff, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Yus Aniza Yusof, PhD

Associate Professor
Faculty of Engineering
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

DECLARATION

I declare that this thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



SHOW PAU LOKE

Date: 19th July 2012

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	v
ACKNOWLEDGEMENTS	viii
APPROVAL	ix
DECLARATION	xi
LIST OF TABLES	xvi
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xx
 CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Research problems	5
1.3 Objectives	6
1.4 Summary of the research	6
2 LITERATURE REVIEW	9
2.1 Recovery strategies for biomolecules	9
2.1.1 Conventional methods	10
2.1.2 Alternative methods	11
2.2 Aqueous two-phase system (ATPS)	14
2.2.1 Design of an ATPS	14
2.2.2 Advantages of an ATPS	17
2.3 Types of ATPS	18
2.3.1 Polymer/polymer ATPS	18
2.3.2 Polymer/salt ATPS	19
2.3.3 Alcohol/salt ATPS	20
2.4 Temperature-induced phase partitioning for protein recovery	21
2.5 Applications of an ATPS	23
2.5.1 Recovery of bioproducts	23
2.6 Process integration with ATPS	24
2.6.1 Extractive bioconversion	25
2.6.2 Extractive fermentation	27
2.6.3 Cell disruption integrated with product recovery	30
2.7 Solvent sublation (SS)	30
2.7.1 Advantages of SS	31
2.7.1.1 High separation efficiency	31
2.7.1.2 High concentration coefficient	31
2.7.1.3 Low dosage of organic solvent	32
2.7.1.4 Soft separation process	32
2.7.1.5 Simple operation	32
2.8 Aqueous two-phase flotation (ATPF)	33
2.9 Lipase	34
2.9.1 Characteristic of lipase	38

2.9.2	Bacteria lipase	40
2.9.3	Applications of lipases	41
2.10	Critical review of the current state of knowledge in lipase recovery based on ATPS	42
3	ENHANCEMENTLIPASE PRODUCTION DERIVED FROM <i>BUKHODERIA</i> SP. ST8 BY STUDY THE EFFET OF FERMENTATION CONDITIONS	43
3.1	Introduction	43
3.2	Materials and Methods	43
3.2.1	Materials	44
3.2.2	Cultivation of <i>Burkholderia</i> sp. ST8 cells	44
3.2.3	Lipase activity assay	46
3.2.4	Bicinchoninic acid (BCA) assay	46
3.4	Results and Discussion	47
3.4.1	Effect of medium volume ratio (V_R) to free volume of the shake flask	47
3.4.2	Effect of nutrient broth concentration	48
3.4.3	Effect of metal ion concentration	49
3.4.4	Effect of carbon source concentration	50
3.4.5	Effect of pH	51
3.4.6	Effect of fermentation duration	53
3.4.7	Effect of fermentation temperature	54
3.4.8	Effect of agitation speed	55
3.5	Conclusion	56
4	RECOVERY OF LIPASE DERIVED FROM <i>BURKHOLDERIA</i> SP. ST8 USING SUSTAINABLE AQUEOUS TWO-PHASE FLOTATION COMPOSED OF RECYCLING HYDROPHILIC ORGANIC SOLVENT AND INORGANIC SALT	57
4.1	Introduction	57
4.2	Materials and Methods	60
4.2.1	Chemicals and apparatus	60
4.2.2	Media and culture condition	60
4.2.3	Optimization of the ATPF	61
4.2.4	Recycling of the ATPF	61
4.2.5	Analytical methods	63
4.2.6	Calculations	63
4.2.7	SDS-PAGE analysis	65
4.3	Results and Discussion	66
4.3.1	Selection of phase components – based ATPF	66
4.3.2	Optimization of a 2-propanol/potassium phosphate ATPF	69
4.3.3	Scaling-up of the 2-propanol/potassium phosphate ATPF	75
4.3.4	Recycling of the phase-forming components	77
4.4	Conclusion	79
5	DIRECT RECOVERY OF LIPASE DERIVED FROM	81

BURKHOLDERIA SP. ST8 IN A RECYCLING AQUEOUS TWO-PHASE FLOTATION

5.1	Introduction	81
5.2	Materials and Methods	83
5.2.1	Chemicals and apparatus	83
5.2.2	Media and culture condition	83
5.2.3	Lipase activity assay	83
5.2.4	BCA assay	84
5.2.5	Calculations	84
5.2.6	Optimization of separation parameters	86
5.2.7	Recycling of phase components on the ATPF	86
5.2.8	SDS-PAGE analysis	88
5.3	Results and Discussion	88
5.3.1	Effect of ammonium sulphate concentration and M _w of EOPO	88
5.3.2	Effect of pH	90
5.3.3	Effect of initial volume and concentration of EOPO	91
5.3.4	Effect of crude feedstock concentration and total volume	92
5.3.5	Effect of nitrogen flow rate on the ATPF	93
5.3.6	Effect of flotation time on the ATPF	94
5.3.7	Effect of recycling the phase-forming components in the ATPF	95
5.4	Conclusion	98

6

PRIMARY RECOVERY OF LIPASE DERIVED FROM BURKHOLDERIA SP. ST8 AND RECYCLING OF PHASE COMPONENTS IN AN AQUEOUS TWO-PHASE SYSTEM

6.1	Introduction	99
6.2	Materials and Methods	101
6.2.1	Materials	101
6.2.2	Media and culture condition	102
6.2.3	Lipase activity assay	102
6.2.4	Protein assay	102
6.2.5	Phase diagram	103
6.2.6	Partition experiments	103
6.2.7	Recycling of phase components in the ATPS	104
6.2.8	Calculations	106
6.2.9	SDS-PAGE analysis	107
6.3	Results and Discussion	107
6.3.1	Effect of EOPO M _w	107
6.3.2	Effect of TLL on lipase partitioning	110
6.3.3	Effect of V _{Ron} lipase partitioning	113
6.3.4	Effect of crude feedstock concentration on lipase partitioning	114
6.3.5	Effect of pH on lipase partitioning	115
6.3.6	Recovery of lipase and recycling of the phase components	117
6.4	Conclusion	121

7	EXTRACTIVE FERMENTATION FOR IMPROVED PRODUCTION AND RECOVERY OF LIPASE DERIVED FROM <i>BURKHOLDERIA</i> SP. ST8 USING A THERMOSEPARATING POLYMER IN AQUEOUS TWO-PHASE SYSTEMS	122
7.1	Introduction	122
7.2	Materials and Methods	126
7.2.1	Materials	126
7.2.2	Media and culture condition	126
7.2.3	Recycling of polymer and cells in repetitive fermentation	127
7.2.4	Analytical methods	128
7.2.5	SDS-PAGE analysis	129
7.2.6	Calculations	129
7.3	Results and Discussion	130
7.3.1	Effect of thermoseparating reagent and its concentration	130
7.3.2	Effect of pH	133
7.3.3	Effect of orbital speed	134
7.3.4	Effect of temperature	136
7.3.5	Repetitive ATPS fermentation in flask and bioreactor	137
7.3.6	Cell growth for homogeneous and ATPS fermentation	139
7.3.7	SDS-PAGE analysis	141
7.4	Conclusion	142
8	GENERAL DISCUSSION, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	143
8.1	General discussion	143
8.2	Conclusions	149
8.3	Recommendations for future research	151
REFERENCES		153
APPENDICES		167
BIODATA OF STUDENT		169
LIST OF PUBLICATIONS		170