## PRODUCTION OF MANNAN-DEGRADING ENZYMES BY *ASPERGILLUS NIGER* IN SHAKE FLASKS AND STIRRED TANK FERMENTER

By

### SITI NORITA BINTI MOHAMAD

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in the Fulfilment of the Requirements for the Degree of Master of Science

February 2005

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

### PRODUCTION OF MANNAN-DEGRADING ENZYMES BY ASPERGILLUS NIGER IN SHAKE FLASKS AND STIRRED TANK FERMENTER

By

#### SITI NORITA MOHAMAD

#### February 2005

#### Chairman: Professor Arbakariya Ariff, Ph.D.

Institute: Bioscience

Optimization of medium composition and culture conditions for mannan-degrading enzymes production by *Aspergillus niger* was carried out in shake flasks and 2 L stirred tank fermenter. Preliminary, three potential strains (*Sclerotium rolfsii, Rhizopus oryzae and A. niger*) were screened, and *A. niger* was used for subsequent study. The mannan-degrading enzymes were purified partially and characterized with regard to pH optima and stability, temperature optima and stability, and  $K_m$  and  $V_{max}$  values. The influence of agitation speed, aeration rate and incubation temperature on the production of mannan-degrading enzyme in batch fermentation using 2 L stirred tank fermenter were also investigated.

Highest level of  $\beta$ -mannanase activity was obtained when guar gum (1495 nkat mL<sup>-1</sup>) and bacteriological peptone (1744 nkat mL<sup>-1</sup>) were compared to other carbon (locust bean gum, cellulose, carboxymethylcellulose and glucose) and nitrogen (peptone from meat,

yeast extract, ammonium sulphate, nitrate and citrate) sources used. The conditions predicted for the maximum production of  $\beta$ -mannanase through the use of response surface methodology were at pH 5.47, 57 g L<sup>-1</sup> bacteriological peptone and 21.3 g L<sup>-1</sup> guar gum. The maximal  $\beta$ -mannanase, endoglucanase,  $\beta$ -mannosidase and galactosidase activity obtained from the predicted equation was of 2010.8, 34.8, 1.6 and 39.0 nkat mL<sup>-1</sup>, respectively.

The optimal temperatures for  $\beta$ -mannanase activity were 50°C and 60°C, with half-life  $(t_{1/2})$  of 6 h at 60°C and 4 h at 70°C. The optimal temperature for endoglucanase activity was 60°C, with  $t_{1/2}$  of 6 h at 60°C and 45 min at 70°C. The optimal temperature for  $\beta$ mannosidase was 70°C with  $t_{1/2}$  of 1.5 h at 70°C. While the optimal temperature for  $\alpha$ galactosidase activity was 50 to 60°C with  $t_{1/2}$  of 2.5 h at 60°C. The  $\beta$ -mannanase, endoglucanase and  $\alpha$ -galactosidase had a pH optima at 3.5 while  $\beta$ -mannosidase at pH 3.0. The enzymes characterized in this study were defined as acidic proteins. The  $\beta$ mannanase,  $\beta$ -mannosidase,  $\alpha$ -galactosidase and endoglucanase showed good stability at pH values of pH 3.5 - 7.0, pH 3.5 - 6.5, pH 3.5 - 5.0 and pH 4 - 7, respectively after a prolonged incubation (24 h at 50°C). High substrate specificity of crude culture filtrate, with low  $K_m$  value of  $\beta$ -mannanase (0.04 mg mL<sup>-1</sup>), endoglucanase (0.54 mg mL<sup>-1</sup>),  $\beta$ mannosidase (1.67 mM) and  $\alpha$ -galactosidase (1.34 mM) indicates the synergistic effect of the enzyme mixture had occurred. The value of  $V_{max}$  for  $\beta$ -mannanase, endoglucanase,  $\beta$ mannosidase and  $\alpha$ -galactosidase were 0.52, 0.12, 1.72 x 10<sup>-3</sup> and 4.68 x 10<sup>-3</sup> nmol mL<sup>-1</sup> min<sup>-1</sup>, respectively.

A fermentation in 2 L stirred tank fermenter using optimized medium yielded 678 nkat mL<sup>-1</sup>  $\beta$ -mannanase, associated with 1.25 nkat mL<sup>-1</sup>  $\beta$ -mannosidase, 18.46 nkat mL<sup>-1</sup>  $\alpha$ -galactosidase and 40.15 nkat mL<sup>-1</sup> endoglucanase at impeller tip speed of 0.82 m s<sup>-1</sup>, aeration rate of 0.1 vvm and incubation temperature of 35°C. Higher degree of agitation speed and aeration rate had an inhibitory effect on the production of mannan-degrading enzymes.

Abstrak tesis dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan Ijazah Master Sains

## PENGHASILAN ENZIM BAGI MENDEGRADASIKAN MANNAN OLEH ASPERGILLUS NIGER MENGGUNAKAN KELALANG PENGGONCANG DAN TANGKI FERMENTER PENGADUK

Oleh

## SITI NORITA MOHAMAD

#### Februari 2005

#### Pengerusi: Profesor Arbakariya Ariff, Ph.D.

Institut: Biosains

Pengoptimaan komposisi media dan keadaan kultur untuk penghasilan enzim bagi mendegradasikan mannan oleh *A. niger* dijalankan di dalam kelalang penggoncang dan tangki fermenter pengaduk 2 L. Sebagai permulaan, tiga strain (*Sclerotium rolfsii, Aspergillus niger* and *Rhizopus oryzae*) yang mempunyai potensi telah dipilih dan *A. niger* telah digunakan untuk kajian yang seterusnya. Enzim utk mendegradasikan mannan telah ditulinkan separa dan dicirikan untuk penentuan pH optima dan kestabilan, suhu optima dan kestabilan, dan nilai  $K_m$  and  $V_{max}$ . Kesan kelajuan putaran, kadar pengudaraan dan suhu inkubasi kepada penghasilan enzim bagi mendegradasikan mannan tangki fermenter suapan sesekelompok menggunakan tangki fermenter pengaduk, 2 L juga dikaji.

Tahap tertinggi bagi aktiviti  $\beta$ -mannanase didapati apabila gum guar (1495 nkat mL<sup>-1</sup>) dan pepton bakteriologikal (1744 nkat mL<sup>-1</sup>) dibanding dengan sumber karbon (gum kekacang locust, selulos, karboksimetilselulos and glukosa) and nitrogen (pepton daripada daging, yis ekstrak, ammonium sulfat, nitrat and sitrat) lain yang telah digunakan. Anggaran keadaan bagi penghasilan maksimum β-mannanase melalui penggunaan kaedah tindakbalas permukaan (response surface methodology) adalah pada pH 5.47, 57 g L<sup>-1</sup> pepton bakteriologikal dan 21.3 g L<sup>-1</sup> gum guar. Anggaran aktiviti maksimum bagi β-mannanase, endoglukanase, β-mannosidase and galaktosidase yang didapati dari persaman, masing-masing adalah 2010.8, 34.8, 1.6 and 39.0 nkat mL<sup>-1</sup>.

Suhu optima bagi aktiviti β-mannanase ialah 50°C dan 60°C, dengan jangka separa hayat (t<sub>1/2</sub>) selama 6 jam pada 60°C dan 4 jam pada 70°C. Suhu optima bagi aktiviti endoglukanase ialah 60°C, dengan t<sub>1/2</sub> selama 6 jam pada 60°C dan 45 minit pada 70°C. Suhu optima bagi β-mannosidase adalah pada 70°C dengan t<sub>1/2</sub> selama 1.5 jam pada 70°C. Sementara itu, suhu optima bagi aktiviti α-galaktosidase ialah pada 50 ke 60°C dengan t<sub>1/2</sub> selama 2.5 jam pada 60°C. β-mannanase, endoglukanase dan α-galaktosidase mempunyai pH optima yang sama iaitu pada 3.5 sementara β-mannosidase pada pH 3.0. Enzim yang dicirikan di dalam kajian ini didefinasikan sebagai protein berasid. Kestabilan yang baik pada pH ditunjukkan oleh β-mannanase, β-mannosidase, α-galaktosidase and endoglukanase, masing-masing pada 50°C). Kespesifikan substrat yang tinggi bagi filtrasi kultur kasar, dengan nilai K<sub>m</sub> yang rendah bagi β-mannanase (0.04 mg mL<sup>-1</sup>), endoglukanase (0.54 mg mL<sup>-1</sup>), β-mannosidase (1.67 mM) and α-galaktosidase (1.34 mM) menggambarkan yang kesan sinergistik telah berlaku pada campuran enzim.

Nilai  $V_{max}$  untuk  $\beta$ -mannanase, endoglukanase,  $\beta$ -mannosidase and  $\alpha$ -galaktosidase masing-masing adalah 0.52, 0.12, 1.72 x 10<sup>-3</sup> and 4.68 x 10<sup>-3</sup> nmol mL<sup>-1</sup> min<sup>-1</sup>.

Fermentasi dalam tangki fermenter pengaduk 2 L menggunakan medium yang telah dioptimakan menghasilkan 678 nkat mL<sup>-1</sup>  $\beta$ -mannanase, berserta dengan 1.25 nkat mL<sup>-1</sup>  $\beta$ -mannosidase, 18.46 nkat mL<sup>-1</sup>  $\alpha$ -galaktosidase dan 40.15 nkat mL<sup>-1</sup> endoglukanase pada kelajuan hujung pemutar 0.82 m s<sup>-1</sup>, kadar pengudaran 0.1 vvm dan suhu inkubasi pada 35°C. Peningkatan darjah kelajuan putaran dan kadar pengudaraan mempunyai kesan perencatan pada penghasilan enzim untuk mendegradasikan mannan.

#### ACKNOWLEDGEMENTS

All praise to Allah S.W.T. who has guide my safely, through every mile, grant me wealth, give me health and most of all give me care and love well. I thank Allah S.W.T. for giving me the strength to finish my study.

I would like to express my sincere appreciation and deepest gratitude to my supervisor, Professor Dr. Arbakariya Ariff for his invaluable guidance, kind and suggestions during the course of this study. My deep appreciation is also extent to the members of my supervisory committee, Dr. Hirzun Mohd Yusof and Dr. Rosfarizan Mohamad for their constructive criticism, guidance and suggestions that have been a great help.

I would also like to express my gratitude to the Director General and Director of Research, of Fisheries Department to for their permission to pursue the study. Thanks also extended to the former and current Head of the Centre of Fisheries Department in Batu Berendam, Mr. Hambal Hj. Hanafi and Mr. Hj Rosly Hassan for supporting me to continue the study. Sincere appreciations also go to staff of Fermentation Unit, Mr Sobri, Ms. Lyana and Mr. Rizal for their assistance throughout my study.

Special thanks goes to my friends, Kak Noorull, Julia, Fidh, Musa, Bazli, Mai, Ina, Siew Ling, Yan Peng, Chin Ming, Xiao Cui, and Mr. Naza for their kind, patience and support during my study. I also wish to express my thanks to all my friends in Freshwater Fisheries Research Center, Batu Berendam especially to Ayong, Kahar, Reha, Zie and

Kak Nab for helping me during the period of my study. A great appreciation also dedicated to Khiriyiah, for spending her precious time commenting this thesis.

Finally, I would like to express my highest gratitude to Hjh Rokiah Md. Diah, Hj. Mohamad Hj. Abu, Hasan Ali, all my brothers, sisters in-law, niece and nephew; thank you for your understanding, caring and moral support given during the period of my study.

## **TABLE OF CONTENTS**

## Page

ABSTRACT	ii
ABSTRAK	V
ACKNOWLEDGEMENTS	viii
APPROVAL	Х
DECLARATION	xii
LIST OF TABLES	xvii
LIST OF FIGURES	xix
LIST OF ABBEVIATIONS	xxiii

# CHAPTER

1.	INTRODU	JCTION	1
2.	MANNAN	HYDROLYSIS AND THE FERMENTATION PROCESS	
	FOR MAN	NAN-DEGRADING ENZYMES PRODUCTION	6
	2.1 Mannar	1	6
	2.1.1	Glucomannan	8
	2.1.2	Galactomannan	9
	2.1.3	Galactoglucomannan	10
	2.2 Hydrol	ysis of Mannan	11
	2.3 Enzym	es Involves in Hydrolysis of Mannan	16
	2.3.1	$\beta$ –Mannanase	16
	2.3.2	$\beta$ –Mannosidase	17
	2.3.3	α-Galactosidase	17
	2.3.4	Endoglucanase	19
	2.4 Develo	pment of Fermentation Process for the Production of	
	Mannar	n-Degrading Enzymes	20
	2.4.1	Microorganisms	20
	2.4.2	Medium Composition	21
	2.4.	2.1 Effect of Carbon Source	22
	2.4.	2.2 Effect of Nitrogen Source	25
	2.4.	2.3 Effect of C : N Ratio	25
	2.4.3	Culture Condition	30
	2.4.	3.1 Effect of pH	30
	2.4.	3.2 Effect of Temperature	31
	2.4.4	Effect of Aeration and Agitation	32
	2.4.5	Enzymes Purification and Characterization	33
	2.5 Conclu	ding Remark	40

3	GENERAL MATERIALS AND METHODS	42
	3.1. Chemical Reagents	42
	3.2. Microorganisms and Maintenance	42
	3.3. Medium	43
	3.4. Analytical Procedures	45
	3.4.1. Determination of Cell Concentration	45
	3.4.2. Determination of Total Reducing Sugars	45
	3.4.3. Determination of Mannanase and Endoglucanase Activity	46
	3.4.4. Determination of $\alpha$ -Galactosidase ( <i>pNP</i> - $\alpha$ Gal) and	
	β-Mannosidase ( <i>p</i> NP- $β$ Man) Activity	47
	3.5 Statistical Analysis	48
	3.6 Experimental Work Plan	48
4.	PRODUCTION OF ENZYMES IMPORTANT IN MANNAN-	
	HYDROLYSIS - COMPARISON AMONG THREE DIFFERENT	
	FUNGAL STRAINS	50
	4.1 Introduction	50
	4.2 Materials and Methods	51
	4.2.1 Microorganisms	51
	4.2.2 Medium	51
	4.2.3 Fermentation	53
	4.2.4 Analytical Methods	53
	4.3 Results and Discussion	54
	4.3.1 Growth Morphology of the Potential Strain on Agar Plate	54
	4.3.2 Enzyme Production Using Different Carbon Sources	60
	4.4 Conclusions	65
5	EFFECT OF DIFFERENT CARBON AND NITROGEN	
	SOURCES AND INITIAL CULTURE pH ON GROWTH	
	OF ASPERGILLUS NIGER AND PRODUCTION OF	
	MANNAN-DEGRADING ENZYMES	66
	5.1. Introduction	66
	5.2. Materials and Methods	68
	5.2.1. Microorganisms	68
	5.2.2. Medium 5.2.2 Ecomonatorian	68
	5.2.5. Fermentation 5.2.4. Experimental Design for Optimum Nitrogen Concentration	09
	5.2.4. Experimental Design for Optimum Nitrogen Concentration	60
	allu pri Value	09
	5.2.5. Allalytical Methods	/1
	5.3.1 Effect of Different Carbon Sources	72 72
	5.3.2 Effect of Different Nitrogen Sources	/ Z Q 1
	5.3.2. Effect of $C \cdot N$ Ratio	01 90
	5.3.4 Ontimization of the Rectariological Pantone Concentration	09
	and nH Value	02
		14

5.3.3.1 Mathematical Modelling for Mannan-Degrading Enzymes	92
Activity	98
5.3.3.2 Surface Plots for Mannan-degrading Enzymes Activity	103
5.3.3.3 Verifications	105
5.4 Conclusions	

# 6. PARTIAL PURIFICATION AND CHARACTERIZATION OF MANNAN- DEGRADING ENZYMES PRODUCED BY

ASPERGI	LLUS NIGER	106
6.1 Introd	uction	106
6.2 Materi	als and Methods	107
6.2.1	Microorganisms and Fermentation	107
6.2.2	Partial Purification of Enzymes	107
6.2	2.2.1 Separation and Ultrafiltration	107
6.2	2.2.2 Salting Out Using Ammonium Sulphate Precipitation	109
6.2.3	Determination of pH Optimum and Stability	109
6.2.4	Determination of Temperature Optimum and Stability	110
6.2.5	Determination of Kinetic Parameters	110
6.2.6	Hydrolysis of Guar Gum (GG) and Locust Bean Gum (LBG)	111
6.2.7	Analytical Methods	112
6.3 Result	s and Discussion	112
6.3.1	Enzymes Purification	112
6.3.2	pH Optimum and Stability	115
6.3.3	Temperature Optimum and Stability	121
6.3.4	Kinetic and Enzyme Hydrolysis	127
6.4 Conclu	usions	132

### 7 EFFECT OF AGITATION, AERATION AND INCUBATION TEMPERATURE ON THE PRODUCTION OF MANNAN-DEGRADING ENZYMES IN 2 L STIRRED TANK FERMENTER 7.1 Introduction

7.2 Materials and Methods 13	35
7.2.1 Microrganisms 13	35
7.2.2 Medium Composition and Preparation 13	35
7.2.3 Fermenter and Fermentation 13	36
7.2.4 Analytical Methods 13	37
7.3 Results and Discussion 14	11
7.3.1 Effect of Agitation Speed 14	11
7.3.1.1 Mannan-Degrading Enzymes Production 14	11
7.3.1.2 Growth of Aspergillus niger 14	14
7.3.1.3 Profile of Dissolved Oxygen Tension, pH and	
Reducing Sugar 14	16
7.3.1.4 The Productivity, Yield and Specific Activity of Enzymes 15	50
7.3.2 Effect of Aeration Rate 15	51
7.3.2.1 Mannan-Degrading Enzymes Production 15	51

134

7.3.2.2 Growth of Aspergillus niger	154
7.3.2.3 Profile of Dissolved Oxygen Tension, pH and	
Reducing Sugar	155
7.3.2.4 The Productivity, Yield and Specific Activity of Enzyr	nes 159
7.3.3 Effect of Incubation Temperature	161
7.3.3.1 Mannan-Degrading Enzymes Production	161
7.3.3.2 Growth of Aspergillus niger	164
7.3.3.3 Profile of Dissolved Oxygen Tension, pH and	
Reducing Sugar	165
7.3.3.4 The Productivity, Yield and Specific Activity of Enzyr	nes 168
7.3.4 Comparison of Maximum Productivity, Yield and Spee	cific
Enzyme Activity in Fermentation Using 2 L Stirred T	`ank
Fermenter and Fermentation in Shake Flask	169
7.4 Conclusions	171
8 GENERAL DISCUSSION, CONCLUSIONS AND SUGGESTION	NS 173
8.1 General Discussion	173
8.2 Conclusions	175
8.3 Suggestions For Further Work	177
REFERENCES	180
APPENDICES	
BIODATA OF THE AUTHOR	218

## DECLARATION

I hereby declare that the thesis is based on my original work expect for quotation and citation which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other at UPM or other institutions.

## **SITI NORITA BINTI MOHAMAD**

Date:

## LIST OF TABLES

Table		Page
1	Hemicellulose content of typical plant materials.	6
2	Production of enzymes by various microorganisms on different carbon and nitrogen sources.	28-29
3	The properties of $\beta$ -mannanases from different sources of microorganisms with regard to optimum pH and stability, and temperature optimum and stability.	36
4	The properties of endoglucanase from different sources of microorganisms with regard to optimum pH and stability, and temperature optimum and stability.	37
5	The properties of $\beta$ -mannosidase from different sources of microorganisms with regard to optimum pH and stability, and temperature optimum and stability.	38
6	The properties of $\alpha$ -galactosidase from different sources of microorganisms with regard to optimum pH and stability, and temperature optimum and stability.	39
7	Composition of basal medium and trace elements.	44
8	Actual factor levels corresponding to coded factor levels.	70
9	Production of mannan-degrading enzymes by <i>A. niger</i> in batch fermentation using different carbon sources.	91
10	Production of mannan-degrading enzymes by <i>A. niger</i> in batch fermentation using different nitrogen sources.	91
11	Full factorial central composite design matrix of two variables in coded units along with the observed responses for $\beta$ -mannanase, endoglucanase, $\beta$ -mannosidase and $\alpha$ -galactosidase during growth of <i>A. niger</i> in batch fermentation.	95
12	Analysis of variance (ANOVA) for quadratic model for (A) $\beta$ - mannanase activity, (B) endoglucanase activity, (C) $\beta$ - mannosidase activity and (D) $\alpha$ -galactosidase activity from the data of central composite design experiments.	96

13	Composition of calculated optimized medium for $\beta$ -mannanase production by <i>A. niger</i> . pH value of the medium was adjusted to 5.47 with 1 M phosphoric acid prior to sterilization.	104
14	Comparison between experimental data using optimized medium and calculated data from model equations for enzyme production by <i>A. niger</i> .	104
15	Partial purification of (A) $\beta$ -mannanase, (B) endoglucanase, (C) $\beta$ -mannosidase and (D) $\alpha$ -galactosidase from <i>A. niger</i> fermentation broth.	114
16	Reducing sugar produced from the galactomannan hydrolysis.	131
17	Biomass production of A. niger at different agitation speeds	146
18	Fermentation kinetics of $\beta$ -mannanase, endoglucanase, $\beta$ -mannosidase and $\alpha$ -galactosidase by <i>A. niger</i> at different agitation speeds in 2 L stirred tank fermenter	151
19	Biomass production of A. niger at different aeration rates	155
20	Fermentation kinetics of $\beta$ -mannanase, endoglucanase, $\beta$ -mannosidase and $\alpha$ -galactosidase by <i>A. niger</i> at different aeration rates in 2 L stirred tank fermenter	160
21	Biomass production of <i>A. niger</i> at different incubation temperatures	165
22	Fermentation kinetics of $\beta$ -mannanase, endoglucanase $\beta$ -mannosidase and $\alpha$ -galactosidase production by <i>A. niger</i> at incubation temperatures in 2 L stirred tank fermenter	169
23	Comparison of fermentation kinetics of maximum mannan- degrading enzymes production by <i>A. niger</i> in fermentation using 2 L stirred tank fermenter and fermentation in shake flask	171

## LIST OF FIGURES

Figures	\$	Page
1	A typical molecular structure of 1,4-β-mannan.	7
2	Konjac glucomannan with acetyl group.	9
3	(A) D-galacto-D-mannan from guar ( <i>Oyamopsis tetragonolobus</i> ) and (B) D-galacto-D-mannan from carob ( <i>Ceratonia siliqua</i> )	10
4	A structure of 0-acetyl-galactoglucomannans .	11
5	The non reducing end of guar gum is shown schematically. The polymannose chain ( $\beta$ -linkage) is substituted every 2 residues by a galactosyl molecule ( $\alpha$ -linkage). The hydroxyl groups are shown as thick bars in their correct equatorial or axial positions. The arrows represent the glycosidic links recognized by $\beta$ -mannanase, $\beta$ -mannosidase and $\alpha$ -galactosidase.	14
6	Structure of monosaccharide, galactose, glucose and mannose.	15
7	Effect of different carbon sources to the production of mannanase by <i>A. niger</i> NCH-189.	23
8	Mannanolytic systems of <i>Aureobasidium pullulans</i> : cellular localization of enzyme components and regulation of their synthesis.	24
9	C : N ratio of $\beta$ -mannanase production by <i>Rhodothermus marinus</i> grown on locust bean gum (C) and yeast extract (N).	27
10	Effect of pH on mannanase activity and stability of <i>Aspergillus niger</i> .	35
11	Flow diagram of the experimental work.	49
12	Schematic diagram of the position of mycelium and/or spore on the glass microscope slide.	52
13	Schematic diagram of the position of glass microscope slide in the Petri dish from, A) the top view and B) the side view.	52
14	Photograph of A. niger cultured on potato dextrose agar (A) view	57

from the back plate and (B) view from top plate.

15	A. niger conidia and conidiophore	57
16	Photograph of <i>R. oryzae</i> grown on potato dextrose agar.	58
17	R. oryzae sporangium and sporangiophore	58
18	Photograph of, (A) <i>S. rolfsii</i> after 5 days grown on PDA (B) matured sclerotia production by <i>S. rolfsii</i> after 9 days of incubation.	59
19	Hyphae from sclerotial S. rolfsii germination	59
20	Effect of different carbon sources: locust bean gum (LBG), guar gum (GG), $\alpha$ -cellulose and glucose on the production of (A) $\beta$ -mannanase and (B) endoglucanase by <i>A. niger, S. rolfsii and R. oryzae.</i>	63
21	Effect of different carbon sources: locust bean gum (LBG), guar gum (GG), $\alpha$ -cellulose and glucose on the production of (A) $\beta$ -mannosidase and (B) $\alpha$ -galactosidase by <i>A. niger, S. rolfsii and R. oryzae.</i>	64
22	Production of (A) $\beta$ -mannanase and (B) endoglucanase during growth of <i>A. niger</i> in different carbon sources.	75
23	Production of (A) $\beta$ -mannosidase and (B) $\alpha$ -galactosidase during growth of <i>A</i> . <i>niger</i> on different carbon sources.	77
24	The profiles of (A) biomass production, (B) pH and (C) reducing sugar production and consumption during enzymes fermentation by <i>A. niger</i> using different carbon sources.	80
25	The production of (A) $\beta$ -mannanase and (B) endoglucanase during growth of <i>A</i> . <i>niger</i> in different nitrogen sources.	84
26	(A) $\beta$ -Mannosidase and (B) $\alpha$ -galactosidase production by <i>A. niger</i> in batch fermentation using different nitrogen sources.	85
27	(A) Biomass production, (B) reducing sugar, (C) nitrogen consumption and (D) pH profile during mannan-degrading enzymes fermentation by <i>A. niger</i> using different nitrogen sources.	88
28	Contour and surface plot of the model equation fitted the experimental data of the central composite design based on the	101

influence of variation in pH value and bacteriological peptone concentration to the production of (A)  $\beta$ -mannanase and (B) endoglucanase in *A. niger* fermentation.

29 102 Contour and surface plot of the model equation and fitted the experimental data of the central composite design based on the influence of variation in pH value and bacteriological peptone concentration to the production of (A)  $\beta$ -mannosidase and (B)  $\alpha$ galactosidase in A. niger fermentation. 30 A photograph of tangential flow membrane filtration unit. 108 31 Effect of pH, on the activity of (A)  $\beta$ -mannanase, (B) 117 endoglucanase, (C)  $\beta$ -mannosidase and (D)  $\alpha$ -galactosidase from A. niger. 32 Effect of the pH, on the stability of partially purified (A)  $\beta$ -120 galactosidase from A. niger. 33 Effect of temperature on the activity of (A)  $\beta$ -mannanase, (B) 125 endoglucanase, (C)  $\beta$ -mannosidase and (D)  $\alpha$ -galactosidase from A. niger. 34 Effect of temperature on the stability of (A)  $\beta$ -mannanase, (B) 126 endoglucanase, (C)  $\beta$ -mannosidase and (D)  $\alpha$ -galactosidase from A. niger. Langmuir plots for the determination of  $K_m$  and  $V_{max}$  values for 35 128 partially purified enzyme from A. niger towards its characterizing in hydrolysis of different substrates, (A) locust bean gum (LBG), (B) carboxymethylcellulose (CMC), (C) 4-nitrophenyl-β-Dmannopyranoside 4-nitrophenvl-α-D-(pNPMan) and (D) galactopyranoside (pNPGal). 36 A photograph of 2 L stirred tank fermenter. 139 37 Schematic diagram and dimensions of a 2 L stirred tank fermenter 140 38 143 Effect of different agitation speeds (impeller tip speed) on enzyme production by A. niger in 2 L stirred tank fermenter, (A) Bmannanase, (B) endoglucanase, (C)  $\beta$ -mannosidase and (D)  $\alpha$ galactosidase. 39 Effect of different agitation speeds (impeller tip speed) on the 149 profile of (A) dissolved oxygen, (B) pH and (C) reducing sugar

during growth of A. niger in 2 L stirred tank fermenter.

40	Effect of different aeration rates on enzyme production by <i>A. niger</i> in 2 L stirred tank fermenter, (A) $\beta$ -mannanase, (B) endoglucanase, (C) $\beta$ -mannosidase and (D) $\alpha$ -galactosidase.	153
41	Effect of different aeration rates on the profile of (A) dissolved oxygen, (B) pH and (C) reducing sugar during growth of <i>A. niger</i> in 2 L stirred tank fermenter.	158
42	Effect of different incubation temperatures on enzyme production by <i>A. niger</i> in 2 L stirred tank fermenter, (A) $\beta$ -mannanase, (B) endoglucanase, (C) $\beta$ -mannosidase and (D) $\alpha$ -galactosidase.	163
43	Effect of different incubation temperature on the profile of (A) dissolved oxygen, (B) pH value and (C) reducing sugar during growth of <i>A. niger</i> in 2 L stirred tank fermenter.	167

# LIST OF ABBREVIATIONS

nkat mL <sup>-1</sup>	•	nanokatal/ millilitre
$\mathrm{U}\mathrm{mL}^{-1}$	:	Unit/millilitre
mg mL <sup>-1</sup>	:	milligram/millilitre
PDA	:	Potato Dextrose Agar
<i>p</i> NP-αGal	:	<i>p</i> -nitrophenyl-α-D-galactoside
<i>p</i> NP-αMan	:	<i>p</i> -nitrophenyl-β-D-mannoside
h	:	hour
min	:	Minute
L	:	litre
mM	:	millimolar
μL	:	microlitre
GG	:	Guar gum
LBG	:	Locust bean gum
СМС	:	Carboxymethylcellulose
°C	:	Degree Celsius
DNS	:	Dinitrosalicylic acid
DOT	:	Dissolved oxygen tension
rpm	:	Rotation per minute
Re	:	Reynolds number based on diameter of rotation
vvm	:	Volume of air per min/ volume of fermentation media