



UNIVERSITI PUTRA MALAYSIA

***BREAKING OF SEED DORMANCY, PLANTLET REGENERATION
AND ANTIOXIDANT ACTIVITIES OF BUNIAM PERSICUM***

YOUSEF EMAMIPOOR

FBSB 2012 20

**BREAKING OF SEED DORMANCY, PLANTLET
REGENERATION AND ANTIOXIDANT
ACTIVITIES OF *BUNIMUM PERSICUM***

YOUSEF EMAMIPOOR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirement for the Degree of Doctor of Philosophy**

August 2012

DEDICATION

I wish to dedicate this thesis to my father, for his great spirit, my mother for her merciful heart, my wife, Mahboubeh, for her love and patience, and my dear children, Ladan, Emad and Foad, for their faith in me - to them I am much indebted..



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy.

BREAKING OF SEED DORMANCY, PLANTLET REGENERATION AND ANTIOXIDANT ACTIVITIES OF *BUNIUM PERSICUM*

by

YOUSEF EMAIPOOR

August 2012

Chairman: Professor Maziah Mahmood, PhD

Faculty: Biotechnology and Biomolecular Sciences

Bunium persicum (Boiss) Fedtsch is a valuable medicinal plant that is facing extinction. A study was conducted to adopt various strategies and techniques to conserve and protect the biodiversity of *B. persicum*. Germination of dormant seeds and also maturation of seedlings under *in vitro* condition to overcome seed dormancy and reduce the juvenile period were studied. Then, the efficiency of direct and indirect shoot regeneration, the capacity of somatic embryogenesis and development of micropropagation methods were investigated. In addition, the antioxidant potential, total phenolic compounds and flavonoid contents of seeds were compared with explants and their related calli.

The seeds were treated at room (25 °C) and chilling (2-5 °C) temperatures with or without plant growth regulators (PGRs). The germination rate of dormant seed was 54.7, 46.7 and 53.3% under moist-chilling, moist-room with 35.2 mg/L gibberellic acid (GA₃)

and moist-chilling with 1.4 mg/L thidiazuron (TDZ) conditions respectively. In addition, results showed that treatment of seeds with a combination of 1.4 mg/L TDZ with 35.2 mg/L GA₃ under moist-chilling conditions caused maximum seed germination, which was 93.7%. Furthermore, transferring of germinated seeds to Murashige and Skoog (MS), Gamborg (B5), Driver and Kuniyuki (DKW) and MSB (MS minerals with B5 vitamins) for maturation demonstrated that ½ MSB media was the most suitable medium for seedling development.

Successful direct regeneration of *B. persicum* obtained 14.5 ± 1.5 shoots per root explant on MSB medium supplemented with 0.02 mg/L methyl jasmonate (MJ). Application of root, corm and leaf explants from six-month-old seedlings on MSB medium supplemented with various auxins showed that root-derived and corm-derived calli on MSB medium supplemented with 1.0 mg/L 2,4-dichlorophenoxyacetic acid (2, 4-D) by 77.1% and 74.9% respectively induced somatic embryogenesis calli. The somatic embryos transferred to medium supplemented with different concentrations of benzylaminopurine (BAP), kinetin, spermidine, forchlorfenuron (CPPU), chlormequat chloride (CCC), paclobutrazol (PBZ), casein hydrolysate (CH), poly ethylene glycol (PEG) and banana powder, led to maximum plantlet regeneration, which was 65.8 ± 2.6 obtained in ½ MSB medium supplemented with 20 g/L banana powder. Consequently, induction of somatic embryogenesis under 1.0 mg/L 2,4-D was found to be more suitable than other auxins and capacity of banana powder for plantlet development by having indol acetic acid (IAA), cytokinins and gibberellins (GAs) was more than other additives and PGRs.

Successful indirect shoot regeneration of *B. persicum* was obtained from culture of leaf-derived callus on MSB medium supplemented with 0.4 mg/L 2,4-D. The callus was implanted on ½ MSB medium supplemented with 0.2, 0.4, 0.6, 0.8, and 1.0 mg/L various cytokinins including BAP, kinetin, isopentyl aminopurine (2iP) and zeatin. Results indicated that the most suitable cytokinins for shoot regeneration were 0.6 mg/L kinetin with 34.2 ± 0.6 plantlets per culture. In addition, CPPU, TDZ, spermidine and additives including banana powder, yeast extract and casein hydrolysate influenced callus proliferation more than regeneration. Subsequently, results indicated that efficiency of embryogenesis callus was more than indirect and direct shoot regenerated calli. Also, the effect of different concentrations of sucrose, BAP, PBZ and GA₃ on size of *B. persicum* corms was investigated. Results showed that 90 g/L sucrose with 164.9 ± 2.8 g corm fresh weight (FW) was the most suitable sucrose concentration for growth of corm and shoot numbers.

The antioxidant activity, total phenolic compounds and flavonoids of seed were compared with root, corm, leaf and their related calli. Results of FRAP assay (based on trolox equivalent) showed that leaf segments of six-month-old *B. persicum* with 16.14 mg TE/ g DW had maximum antioxidant activity. Also, result of DPPH assay (based on gallic acid equivalent) for derived calli indicated that corm-derived callus on medium supplemented with 1.0 mg/L NAA (CN) with 12.61 mg GE/g DW showed maximum scavenging of free radicals. However, CN with 18.72 mg GE/g DW had maximum phenolic compounds and root-derived callus under 1.0 mg/L picloram (RP) had maximum flavonoid, which was 7.50 mg RE/g DW.

The results showed that not only seed but also leaf, corm, root and their related derived callus could be used as natural plant antioxidants. However, *B. persicum* seeds had higher amounts of naringenin (170.3 $\mu\text{g/g DW}$), kaempferol (132.7 $\mu\text{g/g DW}$) and quercetin +(120.3 $\mu\text{g/g DW}$), but leaves had higher amount of naringin (240.6 $\mu\text{g/g DW}$), corms had higher amount of naringenin (146.7 $\mu\text{g/g DW}$) while roots had higher amounts of rutin (160.6 $\mu\text{g/g DW}$).

Results showed that a combination of GA_3 (35.2 mg/L) and TDZ (1.4 mg/L) with 93.7% breaking of seed dormancy and subsequent seedling maturation on $\frac{1}{2}$ MSB medium was the suitable method for cultivation via seed. The result also showed that direct regeneration from root explants with 14.5 ± 1.5 , indirect shoot regeneration from leaf-derived callus with 34.2 ± 0.6 and indirect somatic embryodensis from root-derived callus with 65.8 ± 2.3 plantlets/culture respectively could be reliable methods for *B. persicum* regeneration. Furthermore, leaf and corm-derived callus with 16.1 mg TE/g DW and $11.2 \pm 0.1\text{mg GE/g DW}$ antioxidant activity respectively, were found to be suitable as seed replacement. The presence of various flavonoids in seeds rather than leaf, corm and root could be related to more activity of flavonoid biosynthesis enzymes in seeds.

Tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah.

**PEMECAHAN DORMANSI BENIH, PERTUMBUHAN SEMULA DAN
AKTIVITI ANTIOKSIDAN BAGI *BUNIUM PERSICUM***

oleh

YOUSEF EMAIPOOR

Ogos 2012

Pengerusi: Profesor Maziah Mahmood, PhD

Fakulti: Bioteknologi dan Sains Biomolekul

Bunium persicum (Boiss.) B. Fedtsch adalah satu tumbuhan ubatan yang berharga dan diancam kepupusan. Suatu kajian telah dijalankan bagi mengamalkan beberapa strategi dan teknik untuk memulihara dan memelihara pertumbuhan *B. persicum*. Pendomestikan *B. persicum*, pertumbuhan semula secara langsung, jaringan pertumbuhan semula secara somatic embryogenesis telah dijalankan secara jaringan mikropropagas ke atas tumbuhan ini.

Selain daripada itu, potensi antioksidan, jumlah sebatian phenolic dan kanduangan flavonoid benih dibandingkan dengan eksplant yang berkaitan dengan mereka.

Pendomestikan berhadapan dengan dua cabaran penting. Malah benih *B. persicum* adalah sangat dorman dan benih hanya boleh di germenasikan pada kawasan bersuhu sejuk semulajadi. Benih yang kering dan lembap di rawat pada kadar suhu yang berbeza

termasuk pada suhu bilik (25°C) dan suhu sejuk (2-5 °C). Keputusan menunjukkan bahawa terdapat germinasi benih maksimum pada kadar 54.7, 46.7 dan 53.3% di bawah keadaan lembap dan sejuk selepas 60 hari. Keputusan juga menunjukkan perawatan benih dengan 35.2 mg/L GA₃ di bawah keadaan suhu lembap bilik adalah sebanyak 46.7% kadar germinasi dan dengan 1.4 mg/L TDZ di bawah keadaan penyejukan lembap adalah 53.3% dengan dipengaruhi oleh dormansi pematangan benih. Selain daripada itu, keputusan juga menunjukkan perawatan benih dengan kombinasi 1.4 mg/L TDZ dan 35.2 mg/L GA₃ di bawah keadaan sejuk lembap menghasilkan germinasi benih yang maksimum iaitu 93.7%. Tambahan pula, pemindahan benih yang telah digermenasikan kepada MS, B5, DKW dan MSB (mineral MS dengan vitamin B5) untuk proses pematangan menunjukkan bahawa ½ MSB media adalah yang paling sesuai untuk perkembangan anak benih.

Pertumbuhan semula secara langsung yang berjaya bagi *B. persicum* telah diperolehi daripada eksplant dan menunjukkan maksimum perincian 14.5 ± 1.5 bagi setiap eksplant. Medium MSB dengan 0.02 mg/L methyl jasmonate (MJ) digunakan untuk proses pertumbuhan semula. Selain itu, keputusan menunjukkan akar, umbi dan daun daripada eksplant dan juga benih adalah bahan yang tidak sesuai bagi proses pertumbuhan semula bagi MSB, MS, B5 dan DKW yang di tambah dengan 1, 2, 5 dan 6 mg/L BAP atau kinetin.

Pertumbuhan semula secara tidak langsung yang berjaya bagi *B. persicum* bermula dengan percambahan akar, umbi dan daun yang bermula dari anak benih berusia 6 bulan. Eksplant daun pada medium MSB dengan tambahan 0.4 mg/l 2, 4-D digunakan untuk

induksi kalus. Kalus daun dikulturkan di dalam $\frac{1}{2}$ medium MSB dengan tambahan 0.2, 0.4, 0.6, 0.8, dan 1.0 mg/L perbezaan cytokinins. Medium tersebut bersama dengan 0.6 mg/L kinetin dan 34.2 tumbuhan semula untuk setiap kultur adalah bahan yang paling sesuai untuk pertumbuhan semula bagi kalus daun. Selain daripada itu, kesan daripada perbezaan konsentrasis sukrosa, BAP, PBZ dan GA₃ pada saiz umbi *B. persicum* telah dikaji. Keputusan menunjukkan bahawa pada $\frac{1}{2}$ MSB, hanya konsentrasi sukrosa yang mempengaruhi saiz umbi dan bilangan jaringan untuk tumbuhan baru. Keputusan juga menunjukkan 90 g/L sukrosa dengan 164.9 \pm 2.8 g umbi FW adalah konsentrasi sukrosa paling sesuai untuk perkembangan umbi dan bilangan jaringan.

Embriogenesis somatik yang berjaya diperoleh daripada umbi atau kalus akar di dalam media MSB dengan tambahan 1.0 mg/L 2,4-D. Keputusan menunjukkan kalus akar dengan 77.1% pembentukan embryogenesis somatic mempengaruhi embryogenesis somatik. Dengan itu, embrio somatik di pindahkan ke medium $\frac{1}{2}$ MSB (dengan tambahan konsentrasi berbeza dari BAP, kinetin, spermidine, CPPU, CCC, PBZ, casein hydrolysate, PEG dan tepung pisang) membawa kepada pertumbuhan semula tanaman muda sebanyak 65.8 \pm 2.6 dengan perolehan medium $\frac{1}{2}$ MSB pada tambahan 20 g/L tepung pisang.

Di samping itu, pengukuran daripada jumlah flavonoid, sebatian phenolic dan aktiviti antioksi dan umbi, daun, akar dan kalus menunjukkan bahawa kalus akar di bawah 1.0mg/L picloram mempunyai maksimum flavonoid iaitu 7.50 mg rutin bersamaan / g DW. Walau bagaimanapun, kalus umbi di bawah 1.0 mg NAA (C.C., NAA) dengan 18.72 mg asid gallic bersamaan / g DW mempunyai maksimum sebatian fenol.

Berdasarkan daripada kaedah DPPH, pemerangkapan maksimum radikal bebas dipersembahkan oleh C.C. NAA dengan 12.61 mg asid gallic bersamaan /g DW. Berdasarkan dari ujian FRAP, segmen daun dengan 16.14 mg trolox bersamaan /g DW mempunyai maksimum aktiviti antioksidan.

Kehadiran flavanoid seperti quercetin, kaempferol, rutin, naringenin di dalam ekstrak benih menunjukkan bahawa benih boleh digunakan sebagai tumbuhan antioksidan semula jadi. Walau bagaimanapun, ekstrak benih *B. persicum* dengan jumlah kompaun antioksidan semuladi yang berpatutan boleh digantikan dengan aktioksidan sintetik.

ACKNOWLEDGEMENT

First, I would like to record my gratitude to my highly respected supervisor, Professor Dr. Maziah Mahmood, for her supervision, advice, and guidance from early stage of this research as well as giving me extraordinary experiences throughout the work. My sincere thanks also go to members of my supervisory committee, Prof. Madya Dr. Rosfarizan Mohamad and Prof. Dr. Mawardi Rahmani from the Faculty of Science, Universiti Putra Malaysia.

I would not have finished this project without the support of my family who have always helped me whenever I needed them. My deepest appreciation goes to my wife and children whose patience, understanding and support have been my source of strength and inspiration. To my friends who helped me in researching on different fields concerning this project, I am also much indebted.

My appreciation also goes to all my lab mates from Lab 235, Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences, for sharing their knowledge and friendship with me throughout this research.

I thank Dr. Jahangir Porhemmat, the Deputy Minister and Head of Agricultural Research, Education and Extension Organization (AREEO) and Dr. Bahman Panahi, Dean of Kerman Agricultural and Natural Resources Research Center (KANRC) for their financial support.

I certify that a Thesis Examination Committee has met on 10 August 2012 to conduct the final examination of Yousef Emami Poor on his thesis entitled “Breaking of Seed Dormancy, Plantlet Regeneration, and Antioxidant Activities of *Bunium persicum* (Boiss.)” in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Norhani Abdullah, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Syahida Ahmad, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Internal Examiner)

Noor Azmi Shaharuddin, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Internal Examiner)

Praveen K. Saxena, PhD

Professor

University of Guelph

Canada

(External Examiner)

SEOW HENG FONG, PhD

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 22 October 2012

This thesis was submitted to the senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the supervisory committee are as follows:

Maziah Mahmood, PhD

Professor
Faculty of Biotechnology
Universiti Putra Malaysia
(Chairman)

Mawardi Rahmani, PhD

Professor
Faculty of Chemistry
Universiti Putra Malaysia
(Member)

Rosfarizan Mohamad, PhD

Associate Professor
Faculty of Biotechnology
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School Of Graduate Studies
Universiti Putra Malaysia

Date: 10 August 2012

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously and is not concurrently submitted for any other degree at Universiti Putra Malaysia or other institutions.

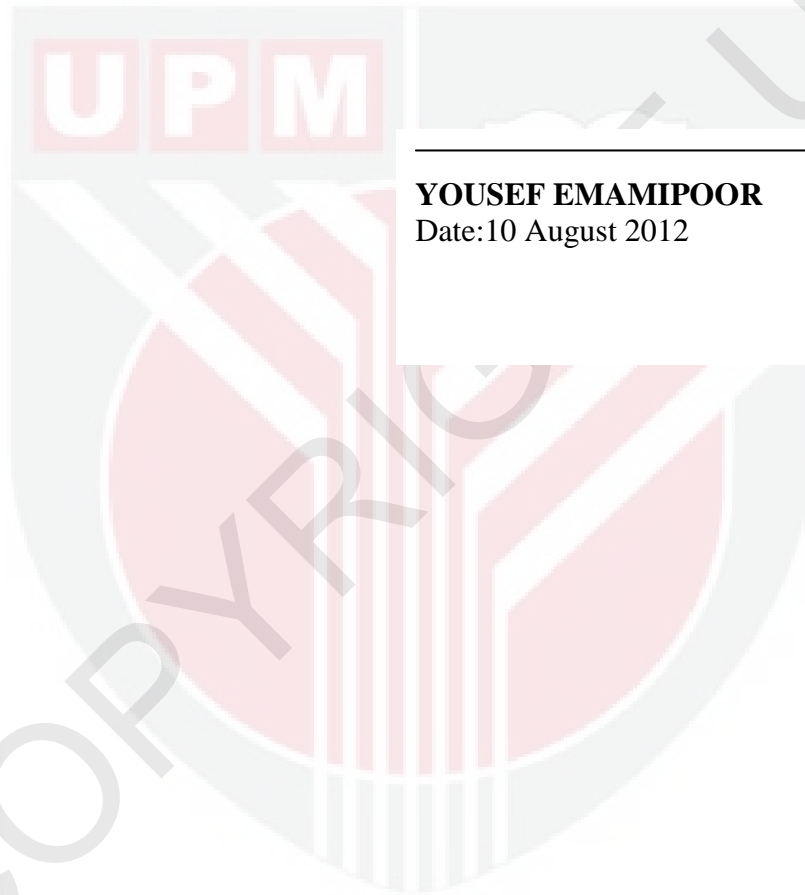


TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vii
ACKNOWLEDGEMENTS	xi
APPROVAL	xii
DECLARATION	xiv
LIST OF TABLES	xix
LIST OF FIGURES	xxii
LIST OF ABBREVIATIONS	xxvii
CHAPTER	
1 INTRODUCTION	1
1.1 Importance of Biotechnology in Medicinal Plants	1
1.2 Objectives	4
2 LITERATURE REVIEW	5
2.1 Bunium Persicum (Boiss.) B. Fedstch	5
2.1.1 Importance of B. persicum	10
2.2 Chemical Ingredients of B. persicum Seeds	11
2.2.1 Essential oil	11
2.2.2 Antioxidant activity	13
2.2.3 Phenolic compounds	15
2.2.4 Flavonoids	16
2.3 Propagation of B. persicum	24
2.3.1 Propagation by seed	24
2.3.2 Propagation by corm	28
2.4 Micropropagation of B. persicum	28
2.4.1 Direct regeneration	30
2.4.2 Indirect shoot regeneration	31
2.5 Factors influencing plant regeneration	34
2.5.1 Explants	34
2.5.2 Plant growth regulators (PGRs)	35
2.5.3 Additives	47
2.6 Tuberization	48
2.6.1 The influence of plant growth regulators	49
2.6.2 The influence of temperature	50
2.6.3 The influence of sucrose concentration	50
3 MATERIALS AND METHODS	51
3.1 Plant Materials	51
3.1.1 Seed collection	51
3.2 Germination of B. persicum Seeds	54

3.2.1	Sterilization of seeds	54
3.2.2	Effect of temperature and moisture on breaking of seed dormancy	54
3.2.3	Effect of plant growth regulators on breaking of seed dormancy	56
3.2.4	Effect of combination of TDZ and GA3 on breaking of seed dormancy	57
3.2.5	Seedling acclimatization	58
3.2.6	Data analysis	59
3.3	Development of <i>B. persicum</i> seedlings	59
3.3.1	Selection of basal media for in vitro growth of <i>B. persicum</i> seedlings	59
3.4	Influence of media, explants and PGRs on direct shoot regeneration of <i>B. persicum</i>	60
3.4.1	Rooting of plantlets	62
3.4.2	Hardening and acclimatization of regenerated plantlets	62
3.5	Establishment of potential calli for indirect regeneration of <i>B. persicum</i>	63
3.5.1	Callus initiation media	63
3.5.2	Culture conditions	64
3.5.3	Potential medium in callus formation	64
3.5.4	Potential explants for callus induction	65
3.5.5	Callus growth	66
3.5.6	Callus proliferation medium	67
3.5.7	Callus maintenance medium	67
3.6	Indirect Regeneration of <i>B. persicum</i>	69
3.6.1	Plant regeneration through somatic embryogenesis	69
3.6.2	Influence of kinetin, BAP, spermidine, CPPU, PBZ, CCC, CH, PEG and banana powder on multiplication of somatic embryos from root-derived somatic embryogenesis callus	72
3.6.3	Shoot regeneration from potential leaf derived callus	73
3.7	Antioxidant activity of <i>B. persicum</i> seeds	77
3.7.1	Extraction method	77
3.7.2	Solvents	78
3.7.3	Antioxidant activity of explants and derived calli	81
3.7.4	Total flavonoid content	82
3.7.5	Total phenolic compounds	83
3.7.6	Quantitative analysis of flavonoids by high-performance Liquid Chromatography	84

4

RESULTS AND DISCUSSION	86	
4.1	Breaking of Seed Dormancy of <i>B. persicum</i>	86
4.1.1	Influence of chilling temperature and moisture	86
4.1.2	Influence of temperature and plant growth regulators on seed germination	91

4.1.3	Effect of combination of TDZ, GA3 and chilling temperature on seed germination	99
4.1.4	Acclimatization of one month old seedlings	103
4.1.5	In vitro micropropagation of <i>B. persicum</i> from seed	105
4.2	Direct Regeneration of <i>B. persicum</i>	108
4.2.1	Influence of medium and explant	108
4.2.2	Rooting of regenerated shoots	110
4.2.3	Hardening and acclimatization	112
4.3	Establishment of potential explants of <i>B. persicum</i> for callus induction	113
4.3.1	Selection of medium for callus induction	114
4.3.2	Influence of different explants on callus induction	116
4.3.3	Proliferation of leaf, corm and root-derived calli	124
4.3.4	Callus growth curve	127
4.3.5	Influence of 2,4-D and cytokinins on callus growth	131
4.4	Potential Embryogenic Callus of <i>B. persicum</i>	136
4.4.1	Influence of 2,4-D on induction of embryogenic callus	136
4.4.2	Regeneration of plantlets from corm-derived somatic embryos	139
4.4.3	Regeneration of plantlets from root-derived somatic embryos	144
4.4.4	Development of protocol for regeneration via somatic embryogenesis	150
4.5	Shoot Regeneration of <i>B. persicum</i> from Leaf-derived Callus	150
4.5.1	Effect of cytokinins on shoot regeneration of <i>B. persicum</i>	152
4.5.2	Maturation of <i>B. persicum</i> plantlets	166
4.5.3	Development of protocols for direct and indirect shoot regeneration of <i>B. persicum</i>	173
4.6	Total Antioxidant Activity of <i>B. persicum</i>	175
4.6.1	Effect of solvent on antioxidant activity of <i>B. persicum</i> seeds	175
4.6.2	Antioxidant activity of methanolic extracts of seed, leaf, corm and root from <i>B. persicum</i> in habitat	177
4.6.3	Antioxidant activity of leaf, corm and root explants of <i>B. persicum</i> and their derived calli	178
4.7	Total Phenolic Content	183
4.7.1	Seed and Explants of <i>B. persicum</i> from habitat	183
4.7.2	Leaf, corm, root explants from six-month-old plant and their derived calli	184
4.8	Total Flavonoid Content	185
4.8.1	Seed and explants of <i>B. persicum</i> from habitat	185
4.8.2	Leaf, corm, root explants and their derived calli from six-month-old plant	186
4.9	Quantification of Flavonoid Compounds by TLC and HPLC	188
4.9.1	TLC analysis of flavonoids	188
4.9.2	HPLC analysis of flavonoids	189

4.9.3	Flavonoid profiles in different parts of <i>B. persicum</i>	192
5	CONCLUSION	196
	REFERENCES	200
	APPENDICES	217
	BIODATA OF STUDENT	235

