

UNIVERSITI PUTRA MALAYSIA

ISOLATION AND CULTURE OF GREEN ALGAE HAEMATOCOCCUS PLUVIALIS AND ITS MOLECULAR DIVERSITY

MOSTAFA NOROOZI

FS 2011 67

ISOLATION AND CULTURE OF GREEN ALGAE HAEMATOCOCCUS PLUVIALIS AND ITS MOLECULAR DIVERSITY



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Doctor of Philosophy

November 2011

DEDICATED TO

To my inspiring mother

To my lovely wife, Saeidah

To my handsome sons, Hossein and Mohammad Hesan

To my supporting siblings



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

ISOLATION AND CULTURE OF GREEN ALGAE HAEMATOCOCCUS PLUVIALIS AND ITS MOLECULAR DIVERSITY

By

MOSTAFA NOROOZI

November 2011

Chairman: Hishamuddin Bin Omar, PhD

Faculty: Science

Haematococcus pluvialis (Flotow) is a unicellular green alga from volvocale algae living in temporary shallow freshwater ponds and has many applications for humans, poultry and fishery due to its ability to produce astaxanthin. The main objective of this study was isolation of new strains from the water bodies in Iran and to evaluate the growth of nine strains of *H. pluvialis* obtained from four countries (four strains from Iran, two strains from Finland, one strain from Switzerland and three strains from USA), in distinct growth media and also determine their genetic diversity based on ISSR and RAPD.

G

The traditional methods and our new found method were used to isolate the four Iranian species. In the new method the algal suspension were exposed to high light intensity to make the cells transform to big, red cyst form which is easy to see and easier to separate from other organisms. The isolated algae were cultured in Bold medium as general algal medium to obtain enough biomass to inoculums to other media. All the cultures were

done under sterile condition using pre-autoclaved chemicals and microbiological filter passed air for aeration. Two methods were used to detect the biomass of cultivated cells including 0.8 micron Milipore filter paper and DMSO method to extract and measure chlorophyll concentration in all media and strains. The astaxanthin concentration was measured by spectrophotometer and HPLC methods which DMSO was used to extract pigment from the cyst of *H. pluvialis*. In this research, the cultured media were centrifuged to precipitate the biomass and DNA was extracted from 10 different strains of *H. pluvialis* using the Dellaporta method with the liquid nitrogen and mortar. Genetic similarity study was carried out using 14 ISSR and 12 RAPD primers.

The biomass obtained from Bold culture were used for the purpose of inoculation to four other media namely: NIES, OHM, Mixotroph and COMBO (the COMBO medium was used for the first time to culture *H. pluvialis*). The results of this study depicted that Mixotroph growth medium gave the best biomass of 0.577 g dry wt/l followed by COMBO and OHM media respectively. Moreover, the strains showed geographical differentiations as a result the Iran4 and USA2 strains produced highest biomass and the Finland2 showed the slowest growth.

Molecular markers are suitable tools for the purpose of finding out genetic variations in organisms; however there is no study conducted on ISSR or RAPD molecular markers regarding this organism. The molecular weights of the bands produced ranged from 3.4 to 0.14 Kb. The PCA and dendrogram clustered the *H. pluvialis* strains into various groups according to their geographical origins. The lowest genetic similarity was between the Iran2 and USA1 strains and the highest genetic similarity was between Finland1 and Finland2. The results showed that ISSR and RAPD markers are useful for the genetic

diversity studies of *Haematococcus* as they showed clustering of strains from similar geographical origins. As a conclusion different strains of *H. pluvialis* shows various abilities in physiological, metabolites and biochemical traits. The various characteristics of strains explain the necessity of survey on new resources to discover more efficient strains.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PEMENCILAN, KULTUR ALGA HIJAU ALGA *HAEMATOCOCCUS PLUVIALIS* DAN KEPELBAGAIAN GENETIK MOLEKULARNYA

Oleh

MOSTAFA NOROOZI

November 2011

Pengerusi: Hishamuddin Bin Omar, PhD

Fakulti: Sains

Haematococcus pluvialis (Flotow) adalah alga hijau alga unisel dari kumpulan volvocale yang biasa ditemui di badan air tawar cetek sementara dan mempunyai banyak kegunaan bagi manusia, ternakan dan perikanan kerana kemampuannya untuk menghasilkan astaxanthin. Tujuan utama dari penelitian ini adalah untuk menilai pertumbuhan sepuluh strain *Haematococcus pluvialis* yang diperolehi daripada empat negara (empat strain dari Iran, dua strain dari Finland, satu strain dari Switzerland dan tiga strain dari Amerika Syarikat) yang dikultur dalam media pertumbuhan yang berbeza dan juga untuk menentukan kepelbagaian genetik berdasarkan kaedah ISSR dan RAPD.

 \bigcirc

Media Bold digunakan untuk mendapatkan pengeluaran biojisim utama bagi tujuan inokulasi bagi empat media lain iaitu: NIES, OHM, Mixotroph dan COMBO (media COMBO digunakan buat pertama kalinya untuk pengkulturan *H. pluvialis*). Biojisim homogeny yang tumbuh dalam media berbeza ditapis melalui kertas penapis Millipore

0.8µm untuk mendapatkan berat kering. Kepekatan klorofil ditentukan melalui kaedah dimetilsulfoksida. Keputusan kajian ini menunjukkan bahawa media pertumbuhan Miksotrof memberikan biojisim terbaik dengan berat kering 0,577 g / l diikuti oleh medium COMBO dan media OHM. Selain itu, strain menunjukkan perbezaan geografi antra hasil strain Iran4 dan USA2 yang menghasilkan biojisim tertinggi manakala Finland2 menunjukkan pertumbuhan paling lambat.

Penanda molekul adalah alat yang sesuai untuk tujuan mengetahui variasi genetik dalam organisma, namun tiada kajian yang telah dilakukan menggunakan penanda molecul ISSR atau RAPD terhadap organisma ini. Dalam kajian ini, sampel DNA diekstrak dari 10 strain *H. pluvialis* yang berbesa menggunakan kaedah Dellaporta. Kajian persamaan genetik dijalankan dengan menggunakan primer 14 ISSR dan 12 RAPD.

Kesamaan genetik terendah adalah antara strain Iran2 dan USA1 manakala kesamaan genetik tertinggi adalah antara Finland1 dan Finland2. Keputusan kajian menunjukkan bahawa penanda ISSR dan RAPD adalah berguna untuk kajian kepelbagaian genetik *Haematococcus* yang telah menunjukkan hasil kluster strain dari asal geografi yang serupa.

ACKNOWLEDGEMENTS

First of all, Alhamdullillah to Allah S.W.T for giving me this opportunity and strength to complete my study.

I wish to express my heartfelt thanks to Dr. Hishamuddin Bin Omar for his help and invaluable advice, encouragement and unfailing patience throughout the course of this study. His guidance is truly appreciated and his constant guidance towards the completion of this thesis. A thank you note also goes to Prof. Dr. Tan Soon Guan and Associated Prof. Dr. Suhaimi Napis for their advice and support.

I would like to thank Dr. Mohammad Amin Hejazi to initiate thinking of my theis subject and also his assistance in providing CCAP strains of *Haematococcus pluvialis*.

My heartfelt thanks, gratitude and appreciation goes to my family especially my wife and my son for their endless effort in persuading me to complete this thesis and not forgetting my mother and brothers for their support and encouragement throughout the finishing point of this thesis.

Finally, this thesis is dedicated to all those who believe in the virtue of learning.

I certify that a Thesis Examination Committee has met 30th November to conduct the final examination of Mostafa Noroozi on his thesis entitled "Isolation and culture of green algae *Haematococcus pluvialis* and its molecular diversity" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Rusea Go, PhD

Associate Professor Department of Biology, Faculty of Science Universiti Putra Malaysia (Chairman)

Muskhazli Mustafa PhD

Associate Professor Department of Biology, Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Faridah Qamar<mark>uz Zam</mark>an, Ph<mark>D</mark>

Associate Professor Department of Biology, Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Tasneem Fatma, PhD

Professor Department of Biosciences, Faculty of Natural Sciences Jamia Millia Islamia a Central University New Delhi, India (External Examiner)

SEOW HENG FONG, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Hishamuddin Bin Omar, PhD

Senior Lecturer Faculty of Science University Putra Malaysia (Chairman)

Tan Soon Guan, PhD

Professor Doctor Faculty of Biotechnology and Biomolecular Science University Putra Malaysia (Member)

Suhaimi Napis, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Science University Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institutions.



TABLE OF CONTENTS

			Page		
DE AB AC AP DE LIS LIS CH	EDICA STRA STRA CKNO PROV CLAI ST OF ST OF ST OF	ATED ACT AK WLEDGEMENTS VAL RATION 5 TABLES 5 FIGURES 5 FIGURES 5 ABBREVIATIONS	ii iii vi viii ix xi xvi xvi xvi xix		
1	INT	RODUCTION	1		
	1.1	Statement of problem	5		
	1.2	Objectives	6		
2	2 LITERATURE REVIEW				
	2.1	About Haematococcus	7		
		2.1.1 Taxonomic Description of <i>Haematococcus</i>	7		
		2.1.2 Contributors	9		
		2.1.3 Classification of <i>Haematococcus</i>	9		
		2.1.4 Habitat	10		
	2.2	Biology of <i>Haematococcus</i>	12		
		2.2.1 Cellular Morphology and Organells	13		
	2.3	Cell Wall	16		
	2.3.1 Ultrastructural Aspects of the Cell Boundary of Hapluvialis		ococcus 16		
	2.4	Microalgae Nutritional Properties	18		
	2.4.1 Astaxanthin and othe important Biochemical Compos <i>Haematococcus</i>				
	2.5 Carotenogenesis				
		2.5.1 Involvement of ROS in the Carotenogenesis Process	22		
	2.6	Astaxanthin	24		
		2.6.1 Astaxanthin structure	26		
		2.6.2 Radical trapping and Antioxidant Activity of Astaxanthi	n 27		

6

	2.6.3 Factors Responsible for Astaxanthin Formation	28	
2.7	Astaxanthin detection methods		
2.8	Algal Isolation		
2.9	Culture of Haematococcus		
2.10	Factors Affecting Growth of Algae and Haematococcus	35	
	2.10.1 Light	35	
	2.10.2 Nutrients	36	
	2.10.3 Temperature	38	
	2.10.4 pH	39	
	2.10.5 Salinity and Iron	39	
2.11	Culture Systems	40	
2.12	History of Growth Medium	44	
	2.12.1 Autotrophic Media	46	
	2.12.2 Mixotrophic Media	47	
	2.12.3 Fed-batch Production of Astaxanthin in Bubble Column	49	
	2.12.4 Heterotrophic Medium	50	
2.13	Multivariable Method Test	51	
2.14	One and Two Step Production of <i>Haematococcus</i>	52	
2.15	Molecular Approach to Haematococcus	53	
	2.15.1 Molecular Genetic Markers and Their Characteristics	55	
	2.15.2 Molecular Marker Criteria	56	
2.16	DNA quality Detection		
2.17	Fingerprinting Studies	59	
	2.17.1 RAPD	59	
	2.17.2 ISSR	60	
	2.17.3 Microsatellite	61	
	2.17.4 AFLP	62	
	2.17.5 Isozyme	62	
	2.17.6 Microsatellite primed-PCR	63	
	2.17.7 Unanchored Primers:	63	
	2.17.8 Anchored Primers	64	
2.18	Genetic Analysis	64	
2.19	Clustering	65	
	2.19.1 UPGMA Method	66	
	2.19.2 Neighbor Joining Method (NJ)	67	

 \mathbf{C}

3	ISOLATION AND MORPHOLOGY		
	3.1	Introduction	68
		3.1.1 Isolation	68
		3.1.2 Microscopic Images	69
	3.2	Material and Methods	70
		3.2.1 Algal Isolation and Purification	70
		3.2.2 Morphology	73
	3.3	Results and Discussion	74
		3.3.1 Haematococcus Isolation	74
		3.3.2 Morphology	76
	3.4	Conclussion	80
4	GROWTH STUDY		
	4.1	Introduction	82
	4.2	Material and Methods	84
		4.2.1 Algal Culture Conditions	84
		4.2.2 Growth Media Preparation	84
		4.2.3 Inoculation	86
		4.2.4 Biomass Measurement and Dry Biomass Method	89
	4.3	Results and Discussion	91
		4.3.1 Dry Weight Growth Assessment	92
	4.4	Conclussion	105
5	PIG	MENT ANALYSIS	106
	5.1	Introduction	106
	5.2	Material and Methods	108
		5.2.1 Chlorophyll Measurement	108
		5.2.2 Astaxanthin Extraction and Measurement	109
	5.3	Results and Discussion	112
		5.3.1 Astaxanthin Measurement	112
		5.3.2 Spectrophotometer Measurement of Astaxanthin	112
		5.3.3 HPLC of Astaxanthin	115
		5.3.4 Chlorophyll Measurement	115
	5.4	Conclussion	123
6	MO	LECULAR MARKER ANALYSIS	124

 \bigcirc

xiv

	6.1	Introduction	124
	6.2	Material and Methods	125
		6.2.1 DNA Isolation	125
		6.2.2 Modified Dellaporta Protocol	126
		6.2.3 DNA Quality	127
		6.2.4 Agarose Gel Electrophoresis	128
		6.2.5 Gel Documentation	129
		6.2.6 Polymerase Chain Reaction (PCR)	129
		6.2.7 Primer Optimization	131
		6.2.8 Temperature Optimization	131
		6.2.9 Mg ⁺² Optimization	131
	6.3	Analysis	132
		6.3.1 UPGMA and Neighbor Joining Methods	133
	6.4	NTSYSpc	134
	6.5	Clustering	134
	6.6	Band Matching Similarity Coefficients	135
	6.7	Shannon-Weaver Diversity Index	136
	6.8	Analysis of Molecular Variance (AMOVA)	137
	6.9	PopGene	137
	6.10	Results and Discussion	138
		6.10.1 DNA Quality	139
		6.10.2 Analysis of Molecular Variance	139
		6.10.3 Jacard's Similarity Index	141
		6.10.4 Primer and Band Characteristics	145
		6.10.5 Dendrogram for ISSR, RAPD and Pooled ISSR-RAPD Data	ı 151
	6.11	Conclussion	155
7	CON	ICLUSSION AND RECOMENDATIONS	158
	7.1	Recommendations	159
	REF	ERENCES	160
	APP	ENDIX	175
	LIST	F OF PUBLICATIONS	194
	BIO	DATA OF STUDENT	197