



UNIVERSITI PUTRA MALAYSIA

**PRODUCTION OF FLAVONOIDS IN PEGAGA (CENTELLA ASIATICA
L. URBAN) CELL SUSPENSION CULTURE**

TAN SUAT HIAN

FBSB 2010 9



**PRODUCTION OF FLAVONOIDS IN PEGAGA (*CENTELLA ASIATICA* L.
URBAN) CELL SUSPENSION CULTURE**

TAN SUAT HIAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

September 2010



DEDICATED TO:

FATHER, MOTHER, BROTHERS & JASON CHEE

WHO ALWAYS HAVE FAITH IN ME

AND

THEIR SUPPORTS HAVE GUIDED ME TO GONE THROUGH

ALL THE OBSTACLES IN LIFE



Abstract of thesis submitted to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Doctor of Philosophy

**PRODUCTION OF FLAVONOIDS IN PEGAGA (*CENTELLA ASIATICA* L.
URBAN) CELL SUSPENSION CULTURE**

By

TAN SUAT HIAN

Septemeber 2010

Chairman: Professor Maziah Mahmood, PhD

Faculty: Biotechnology and Biomolecular Sciences

Centella asiatica L. Urban (Umbelliferea), locally known as ‘Pegaga’ was an herbal plant that had been used in traditional medicine in Asia for many centuries. Its medicinal values are attributed to the presence of flavonoid compounds. Since there was still no information available on the flavonoid production in cultured tissues, studied have been carried out in evaluating the distribution of the flavonoid content particularly kaempherol, quercertin, luteolin and apigenin in intact plants of the four accessions of *C. asiatica* collected several locations in Malaysia as well as in callus and cell suspension cultures. The total flavonoid content has been determined by using spectrophotometry methods. The flavonoid compounds present in the cell cultures have been analyzed with thin-layer chromatography (TLC) and high performance liquid chromatography (HPLC) techniques.

Results obtained from the studies revealed that the four accessions of *C. asiatica* which showed distinctive differentiation in morphological characteristic were



differed in the biochemical constituents especially in their flavonoid contents. The flavonoid constituents were detected in the range of 1.93 ± 0.094 to 8.99 ± 0.346 mg/g dry weight in the whole plant tissues. Flavonoids were also successfully detected in callus (0.98 ± 0.097 to 2.46 ± 0.021 mg/g dry weight) and cell suspension culture (0.67 ± 0.056 to 0.89 ± 0.044 mg/g dry weight). The flavonoid content found in cultured tissues were lower than that produced in the intact plant tissues. In the TLC analysis for the leaf tissues of intact plant, the possible flavonoid compounds might be presented were kaempferol, naringenin, luteolin, (+)-catechin and rutin. Similar results were also observed in callus, cell suspension and cell suspension media in the TLC analysis. Further investigation using HPLC showed that leaf tissues of intact plant accession UPM03 has rutin, luteolin, quercetin and kaempferol while other accessions only have rutin or/and kaempferol. An additional flavonoid compound called apigenin was detected in callus and cell suspension. As a result, culture conditions such as different cell size aggregations, pH values, light, inoculum sizes as well as interaction between plant growth regulators have been assessed to optimize and enhance the selected flavonoid production in the chosen cell suspension cultures. Cell suspension UPM03 in aggregation size of 250-500 μm with the highest cell growth rate (0.69 g fresh weight/day) was suitable to produce flavonoid in 100 mL shake flask of *C. asiatica* suspension cells. The cells grow better and produce more flavonoid (3.66 ± 0.13 mg/g dry weight) under the initial pH of 5.7 with the presence of light when supplemented with 3 mg/L 2,4-D and 1 mg/L kinetin. In addition, the responses of cells to selected elicitors and precursors have also been investigated. The biosynthesis of flavonoid was elicited by the addition of 1mg/L of both yeast extract (7.32 ± 0.17 mg/g dry weight) and salicylic

acid (7.11 ± 0.16 mg/g dry weight). Elicitation with 400 μ M methyl jasmonate have increased flavonoid content to 7.82 ± 0.2 mg/g dry weight. However, the addition of 1mg/L casein hydrolysate (4.91 ± 0.15 mg/g dry weight) and chitosan (4.29 ± 0.13 mg/g dry weight) have no significant effect if compared to the control culture (5.08 ± 0.22 mg/g dry weight). The usage of 60 mg/L phenylalanine and 40 mg/L tyrosine have resulted in 14-fold and 3-fold increase in production of flavonoid, respectively. Further study on the synergistic effect of combination of precursor (60 mg/L phenylalanine) and elicitor (400 μ M) showed that the cell suspension UPM03 not only produce the highest flavonoid content in cell (67.07 ± 6.47 mg/g DW) but also in the media (7.73 ± 0.12 mg/g). Thus, it was proven that the synergistic bioprocess was a very useful strategy to enhance the flavonoid production particularly luteolin in *in vitro* cultures of *C. asiatica*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGHASILAN FLAVONOID MELALUI AMPAIAN SEL PEGAGA
(*CENTELLA ASIATICA* L. URBAN)**

Oleh

TAN SUAT HIAN

September 2010

Pengerusi: Professor Maziah Mahmood, PhD

Fakulti: Bioteknologi dan Sains Biomolekul

Centella asiatica L. Urban (Umbelliferea) yang dikenali sebagai pegaga oleh masyarakat tempatan merupakan sejenis pokok herba yang telah digunakan selama beberapa abad di negara Asia sebagai ubat tradisional. Nilai perubatannya adalah disebabkan kehadiran kompoun flavonoid. Oleh kerana ketiadaan maklumat dalam penghasilan flavonoid dalam tisu yang telah dikulturkan, maka kajian telah dijalankan untuk mengenalpasti taburan flavonoid terutamanya kaempferol, kuersetin, luteolin dan apigenin dalam 4 aksesori pokok induk *C. asiatica* yang telah dikumpulkan dari beberapa kawasan di Malaysia serta dalam kultur kalus and sel ampaiian. Jumlah kandungan flavonoid telah ditentukan dengan penggunaan cara spektrofotometri. Komposisi flavonoid dalam kultur sel telah dianalisis dengan teknik TLC dan HPLC.

Keputusan telah menunjukkan bahawa 4 aksesori *C. asiatica* mempunyai pembezaan dari segi ciri-ciri morfologi yang menonjol dan berbeza dalam komposisi biokimia

terutamanya dalam kandungan flavonoid. Kandungan flavonoid dalam pokok induk dikesan dalam lingkungan 1.93 ± 0.094 ke 8.99 ± 0.346 mg/g berat kering. Kandungan flavonoid juga berjaya dikesan dalam kalus (0.98 ± 0.097 ke 2.46 ± 0.021 mg/g berat kering) dan sel ampaian (0.67 ± 0.056 ke 0.89 ± 0.044 mg/g berat kering). Kandungan flavonoid yang dikesan dalam kultur tisu adalah lebih rendah daripada pokok induk. Dalam analisi TLC untuk daun anak pokok, flavonoid kompond yang mungkin hadir ialah kaempferol, naringerin, luteolin, (+)-katekin dan rutin. Keputusan yang sama juga dikesan dalam kalus, sel ampaian dan media sel ampaian dalam TLC analisis. Kajian seterusnya dengan HPLC menunjukkan tisu daun anak pokok aksesori UPM03 mempunyai rutin, luteolin, kuersetin dan kaempferol manakala aksesori lain hanya mempunyai rutin atau/dan kaempferol. Oleh itu, keadaan kultur seperti saiz gumpalan yang berbeza, nilai pH, kehadiran cahaya, saiz inokula serta interaksi hormon telah ditaksir untuk mengoptimumkan dan menambah penghasilan flavonoid yang terpilih dalam kultur sel ampaian yang dipilih. Sel ampaian dengan saiz gumpalan 250-500 μm dengan kadar penumbuhan yang paling tinggi (0.69 g berat basah/hari) sesuai digunakan untuk penghasilan menggunakan 100 mL kalalang goncang. Sel-sel ampaian ini tumbuh dengan lebih baik dan menghasilkan lebih banyak flavonoid (3.66 ± 0.13 mg/g berat kering) dalam keadaan pH 5.7 diikuti dengan kehadiran cahaya apabila ditambahkan dengan 3 mg/L 2,4-D dan 1 mg/L kinetin. Tambahan pula, kajian telah dijalankan pada tindakbalas sel terhadap penggunaan elisitor dan pembekalan prekursor tertentu. Penghasilan flavonoid telah digalakkan dengan penambahan 1 mg/L kedua-duanya ekstrak ragi (7.32 ± 0.16 mg/g berat kering) dan asid salisailik (7.11 ± 0.16 mg/g berat kering). Penggunaan 400 μM metil jasmonat telah menambahkan kandungan

flavonoid ke 7.82 ± 0.2 mg/g berat kering. Walau bagaimanapun, penambahan 1 mg/L kasin hidrolisat (4.91 ± 0.15 mg/g berat kering) dan kitosan (4.29 ± 0.13 mg/g berat kering) tidak mempunyai kesan yang ketara jika dibandingkan dengan sampel kawalan (5.08 ± 0.22 mg/g berat kering). Penggunaan 60 g/L fenilalanina dan 40 mg/L tirosina telah meningkatkan penghasilan flavonoid masing-masing sebanyak 14-kali ganda dan 3-kali ganda. Kajian tentang kesan sinergistik dalam kombinasi prekursor dan elisitor menunjukkan ia adalah strategi yang amat berguna dalam meningkatkan penghasilan flavonoid terutamanya luteolin dalam kultur *in vitro* *C. asiatica*. Kajian seterusnya dalam kesan kombinasi precursor (60 mg/L phenylalanina) dan elisitor (400 μ M tyrosina) menunjukkan sel ampaiian UPM03 bukan sahaja menghasilkan kandungan flavonoid yang terbanyak dalam sel (67.07 ± 6.47 mg/g berat kering) malahan dalam media (7.73 ± 0.12 mg/g). Kesimpulan, proses penggabungan penggunaan ialah satu strategi untuk meningkatkan penghasilan flavonoid terutamanya luteolin dalam kultur *C. asiatica*.

ACKNOWLEDGEMENTS

I am grateful to Buddha that gives me the physical and mental strength to carry on my study to completion. The philosophy and lectures of Buddha has guides me throughout my life.

First and foremost, I would like to take this opportunity to express my heartiest gratitude to the chairman of my supervisory committee, Prof. Dr. Maziah Mahmood, for her invaluable advice, excellence guidance, contribution, patient and criticism, not only in making the completion of this thesis a success, but also guided me to be a better person. Special thanks again to Prof. Dr. Maziah who gave me a golden opportunity to participate in the 3rd Global Summit on Medicinal and Aromatic Plants (GOSMAP'03) in Chiang Mai, November, 2007. I sincerely thank to my co-supervisors of my supervisory committee, Prof. Dr. Arbakariya B. Ariff and Dr. Syahida Ahmad for their support, assistance, friendliness and as well as the suggestion throughout this research.

My sincere appreciation is extended to assoc. Prof. Dr. Radzali Muse for his invaluable assistance and helpful suggestion. I hope he can fully recover from his illness soon. This research could not be accomplished without the help from all staff of the Department of Biochemistry and Microbiology. Their cooperation really assist me to finish this research.



With appreciations, to all my labmates from lab 235 for sharing their knowledge, friendship, ideas, as well as their life experiences during the course of this project. Acknowledgement also extended to Nisha and Vijadren for their assistance in reading the draft leading to an improvement of this thesis.

Finally yet importantly, I would like to express my heartiest appreciation and thanks to father, mother, brothers and Jason Chee. Thanks for all their care, understanding and patience throughout the course of this project. Their love and care in many ways really kept me going and contributed to the accomplishment of my study.

May God bless all of you.



I certify that an Examination Committee met on 7th September 2010 to conduct the final examination of Tan Suat Hian on her Doctor of Philosophy thesis entitled “Production of Flavonoids in *Centella asiatica* L. urban (Pegaga) Cell Suspension Culture” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Norhani Abdullah, PhD

Professor
Faculty Biotechnology and Science Biomolecules
Universiti Putra Malaysia
(Chairman)

Mohd Arif Syed, PhD

Professor
Faculty Biotechnology and Science Biomolecules
Universiti Putra Malaysia
(Internal Examiner)

Nor’ Aini Abdul Rahman, PhD

Faculty Biotechnology and Science Biomolecules
Universiti Putra Malaysia
(Internal Examiner)

Praveen K. Saxena, PhD

Professor
Department of Plant Agriculture
University of Guelph
Guelph, Ontario, N1G 2W1
Canada
(External Examiner)

BUJANG KIM HUAT, Ph.D.

Professor/Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Maziah Mahmood, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Arbakariya b. Ariff, PhD

Professor

Faculty of Bioprocess Technology

Universiti Putra Malaysia

(Member)

Syahida Ahmad, PhD

Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

HASANAH MOHD GHAZALI, PhD

Professor and Dean

School of Graduate School

Universiti Putra Malaysia

Date:

DECLARATION

I declare that the thesis is my original work except for quotations and citations that have been duly acknowledged. I also declare that it has not been previously and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or other institutions.

TAN SUAT HIAN

Date: 6/9/10

TABLE OF CONTENTS

	Page
DEDICATION	II
ABSTRACT	III
ABSTRAK	VI
ACKNOWLEDGEMENTS	IX
APPROVAL	XI
DECLARATION	XIII
LIST OF TABLES	XVIII
LIST OF FIGURES	XIX
LIST OF ABBREVIATIONS	XXIII
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 <i>Centella asiatica</i> (L.) Urban	4
2.1.1 Taxonomy of <i>Centella asiatica</i>	5
2.1.2 Morphological Description	6
2.1.3 Medicinal properties of <i>Centella asiatica</i>	8
2.2 Secondary Metabolites	11
2.2.1 Flavonoid	12
2.2.2 Flavonoid Biosynthesis Pathway	15
2.2.3 Flavonoids in <i>C. asiatica</i>	17
2.3.4 Flavonoids in Nutrition and Health	18
2.3 Plant Cell culture	20
2.3.1 Callus Culture	24
2.3.2 Cell Suspension Culture	26
2.3.3 Valuable Secondary Product From <i>in vitro</i> Culture	29
2.4 Yield Improvement Strategies	31
2.4.1 Effects of Explants	32
2.4.2 Effects of the Culture Environment	33
2.4.2 Screening and Selection of Highly Productive Cell Lines	38
2.4.3 Plant Growth Regulators	39
2.4.4 Effects of Aggregate Cell Size	41
2.4.5 Inoculums Size	43
2.4.6 Precursors feeding	44
2.4.7 Elicitation	46
3 MORPHOLOGICAL CHARACTERISTICS AND BIOCHEMICAL PROFILES OF MALAYSIAN <i>CENTELLA ASIATICA</i>	55
3.1 Introduction	55
3.2 Materials and Methods	56

3.2.1	General Chemicals and Supplies	56
3.2.2	Plant Materials	56
3.2.3	Identification of Morphological Characteristics	58
3.2.4	Preparation of Crude Extract for Biochemical Assay	58
3.2.5	Total Soluble Protein Assay	59
3.2.6	Total Glucose Assay	60
3.2.7	Total Chlorophyll Assay	60
3.2.8	Beta-carotene Assay	61
3.2.9	Statistical Analysis	61
3.3	Results and Discussion	62
3.3.1	Identification of Morphological Characteristics	62
3.3.2	Total Soluble Protein Content	66
3.3.3	Total Glucose Content	69
3.3.4	Chlorophyll Content	71
3.3.5	Beta-carotene Content	73
3.4	Conclusions	75
4	CALLUS INDUCTION OF <i>CENTELLA ASIATICA</i>	77
4.1	Introduction	77
4.2	Material and Methods	78
4.2.1	Callus Induction on Leaf Explants	78
4.2.2	Callus Maintenance Medium	80
4.2.3	Callus Growth Curve	80
4.2.4	Analysis of Flavonoids Content	82
4.2.6	Statistical Analysis	82
4.3	Results and Discussion	82
4.3.1	Callus Induction from Leaf Explants	82
4.3.2	Callus Maintenance Medium	92
4.3.3	Callus Growth Curve	99
4.3.4	Growth and Flavonoid Content in the Leaf-derived Callus of the Four Accessions	106
4.4	Conclusions	111
5	ESTABLISHMENT OF CELL SUSPENSION CULTURES OF <i>C. ASIATICA</i> FOR THE PRODUCTION OF FLAVONOIDS	112
5.1	Introduction	112
5.2	Materials and Methods	113
5.2.1	Establishment of Cell Suspension Culture and Maintenance	113
5.2.2	Cell Suspension Growth Curve	114
5.2.3	Analysis of Flavonoid Content	114
5.2.4	Statistical Analysis	115
5.3	Results and Discussion	115
5.3.1	Establishment of Cell Suspension Cultures	115
5.3.2	Growth Curve of the Cell Suspension Cultures	122
5.4	Conclusions	125

6	ANALYSIS OF FLAVONOID COMPOUNDS USING TLC, SPECTROPHOTOMETRY AND HPLC METHODS	126
6.1	Introduction	126
6.2	Materials and Methods	126
	6.2.1 Plant Materials	126
	6.2.2 Reference Compounds	127
	6.2.3 Extraction and Hydrolysis Conditions	127
	6.2.4 Determination of Total Flavonoid Content with Spectrophotometer	128
	6.2.5 Thin Layer Chromatography Analysis	129
	6.2.6 Quantitative Analysis by High-performance Liquid Chromatography (HPLC)	129
6.3	Results and Discussion	130
	6.3.1 Determination of Flavonoids Content by TLC Analysis	130
	6.3.2 Optimization of the Peak	134
	6.3.3 Flavonoid Profiles in Different Accessions of <i>C. asiatica</i>	139
	6.3.4 Determination of Flavonoid Content by Spectrophotometry	143
6.4	Conclusions	147
7	EFFECTS OF DIFFERENT CULTURE CONDITIONS ON <i>C. ASIATICA</i> CELL SUSPENSION CULTURES	148
7.1	Introduction	148
7.2	Materials and Methods	149
	7.2.1 Cell Suspension Cultures	149
	7.2.2 Culture Conditions Studied	149
	7.2.3 Analysis of Flavonoid Content	151
	7.2.4 Statistical Analysis	152
7.3	Results and Discussion	152
	7.3.1 Effects of Aggregate Fraction	152
	7.3.2 Effects of Inoculum Size	166
	7.3.3 Effects of Initial pH Values	170
	7.3.4 Effects of Light Irradiation	174
	7.3.5 Effects of Plant Growth Regulators	177
7.4	Conclusions	180
8	EFFECTS OF DIFFERENT ELICITORS AND PRECURSORS ON <i>C. ASIATICA</i> CELL SUSPENSION CULTURES	181
8.1	Introduction	181
8.2	Materials and Methods	183
	8.2.1 <i>In Vitro</i> Cultures and Culture Conditions	183
	8.2.2 Elicitor Preparation	183
	8.2.3 Precursor Preparation	184
	8.2.4 Development of Synergistic Bioprocess for Productivity Enhancement	185
	8.2.3 Extraction and Analysis	185
	8.2.4 Statistical Analysis	186
8.3	Results and Discussion	186

8.3.1	Effect of the Elicitors	186
8.3.2	The Effect of Addition of Precursors	197
8.3.3	Development of Synergistic Bioprocess for Productivity Enhancement	206
8.4	Conclusions	212
9	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	213
9.1	Summary and General Conclusions	214
9.2	Future Research	216
	REFERENCES	217
	APPENDICES	240
	BIODATA OF STUDENT	245
	LIST OF PUBLICATIONS	246

