



**ISOLATION, CHARACTERISATION AND FUNCTIONAL ANALYSIS OF
ROOT-SPECIFIC PROMOTERS FROM MINING OF OIL PALM
TRANSCRIPTOME DATA**

By

SITI MASURA BINTI SUBHI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy**

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Chairman : Noor Azmi bin Shaharuddin, PhD

Faculty : Biotechnology and Biomolecular Sciences

Root-specific promoter is a valuable genetic engineering tool to improve oil palm resistance to diseases, malnutrition, or abiotic stresses that cause extensive losses in the oil palm industry. Since the availability of root-specific promoters from oil palm is still limited, this study was conducted to identify and characterise a strong promoter in a strict root-specific or preferential manner that may have great potential for root-system modification. Mining the transcriptome of various oil palm tissue-specific data generated from RNA Sequencing technology has resulted in discovering seven candidate genes for root-specific promoters. The expression pattern of these candidate genes was further validated through real-time quantitative polymerase chain reaction (RT-qPCR) analysis using various oil palm tissues, including root tissues at different developmental stages. Two transcripts, namely RSP-2 and RSP-3, that showed high expression in roots were selected for further research. RSP-2, annotated as an oil palm metallothionein gene (*EgMT*), was significantly

upregulated at 7 to 170-fold in roots. Likewise, RSP-3, which belongs to the proline-rich protein (*EgPRP1*) gene, was significantly upregulated at 7 to 55-fold. DNA vectors carrying a full-length promoter region and 5' promoter deletion fragments were constructed for the promoter functional study. Deletion analyses were performed according to the density of the cis-acting elements related to the root expression present in the promoter sequences. The promoter activity in transient expression was studied using the biolistic method in various oil palm tissues. The study indicated that *EgPRP1* and *EgMT* promoters could drive the expression of red fluorescence protein (DsRED) in oil palm root tissue. Since oil palm transformation was hampered by a long regeneration time, the stable promoter functional study was investigated further using tobacco as a model plant. The promoter activity was evaluated in seedlings, vegetative and reproductive tissues of T₁ generation using GUS histochemical assay. Promoter activity was evaluated based on the intensity of blue GUS staining. The RSP-3A construct (2000 bp upstream of *EgPRP1* start codon) showed root-preferential or dominant promoter activity in all the root developmental stages. Fine dissection revealed that the shortest promoter construct, designated as RSP-3D (665 bp upstream of *EgPRP1* start codon), conferred root-specific expression in mature plants. For *EgMT*, only the RSP-2D (1107 bp upstream of *EgMT* start codon) deletion construct directed strong GUS expression from early to mature root developmental stages. GUS expression directed by RSP-2A and RSP-2C constructs were detected in the roots of mature plants, with intensities lower than RSP-2D. However, the activity of *EgMT* promoters was also detectable in seed pods and immature seeds, albeit at lower levels than the *Cauliflower Mosaic Virus*

35S promoter (*CaMV35S*). Interestingly, the GUS expression driven by RSP-3A, RSP-2C and RSP-2D was detected at the flower and leaf-cutting sites, suggesting the promoters' activity was inducible by wounding. Results also showed that the promoters consist of cis-acting elements that act as negative regulators, which might be responsible for root specificity. The results further indicated that the 5' UTR and ATATT sequences defined as ROOTMOTIFAPOX1 cis-acting elements are essential for strong promoter activity. Overall, the functional study conclusively demonstrated that the genes identified through the mining of transcriptome data and RT-qPCR analysis that were upregulated during root developmental stages have a high potential to regulate a strong expression of transgenes in a root-specific or preferential manner. The results suggest that promoter regions of RSP-3A and RSP-3D from *EgPRP1* and RSP-2D from *EgMT* are potentially helpful in facilitating oil palm genetic engineering, notably improving crop yield through root-traits modification.

Keywords: Root-specific promoter, root-preferential promoter, expression study, oil palm, tobacco

SDG: GOAL 2: Zero hunger, GOAL 12 : Responsible consumption and production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMENCILAN, PENCIRIAN DAN ANALISIS FUNGSIONAL PROMOTER
SPESIFIK-AKAR DARIPADA PERLOMBONGAN DATA TRANSKRIPTOM
SAWIT**

Oleh

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Promoter spesifik-akar merupakan alat kejuruteraan genetik yang penting bagi meningkatkan ketahanan sawit terhadap penyakit, kekurangan nutrien atau tekanan abiotik yang mana menyebabkan kerugian besar dalam industri sawit. Memandangkan ketersediaan promoter spesifik akar daripada sawit masih terhad, kajian ini dijalankan dengan objektif untuk memencilkan kawasan promoter yang kuat dengan ciri spesifik atau dominan dalam tisu akar serta berpotensi untuk digunakan dalam pengubahsuaian sistem-akar. Perlombongan data transkriptom pelbagai tisu sawit yang dihasilkan daripada teknologi penjujukan asid ribonukleik (RNA-Seq) telah menghasilkan penemuan tujuh calon-calon gen untuk promoter spesifik-akar. Corak pengekspresan calon-calon gen ini telah dianalisis menggunakan kaedah tindak balas berantai polimerase kuantitatif masa-sebenar (RT-qPCR) dengan menggunakan pelbagai tisu sawit termasuk tisu akar pada peringkat perkembangan yang berbeza. Dua transkrip, iaitu RSP-2 dan RSP-3 yang

menunjukkan kadar pengekspresan gen yang tinggi dalam akar telah dipilih untuk kajian lanjut. RSP-2 yang dianotasi sebagai gen metallothionein (*EgMT*) diekspreskan dengan kadar yang tinggi dalam akar iaitu sebanyak lebih kurang 7 hingga 170 kali ganda. Begitu juga, RSP-3 yang merupakan gen proline rich protein (*EgPRP1*) diekspreskan dengan kadar yang tinggi iaitu lebih kurang 7 hingga 55 kali ganda. Vektor yang membawa jujukan penuh promoter dan serpihan 5' delesi promoter telah dibina untuk kajian fungsional. Analisis delesi dilakukan berdasarkan kepadatan elemen pengawalatur-cis yang berkaitan dengan pengekspresan spesifik-akar yang terdapat dalam jujukan promoter. Aktiviti promoter dalam pengekspresan sementara telah dikaji dalam pelbagai tisu sawit menggunakan kaedah biolistik. Kajian menunjukkan promoter *EgPRP1* dan *EgMT* mampu mengaruh pengekspresan protein berpendaflour merah (DsRED) dalam tisu akar sawit. Memandangkan tempoh regenerasi sawit tertransformasi adalah sangat panjang, kajian fungsi promoter yang stabil telah dijalankan seterusnya menggunakan tembakau sebagai tumbuhan model. Aktiviti promoter dinilai dalam sampel generasi T₁ iaitu anak benih (peringkat *in vitro*), tisu vegetatif dan tisu pembiakan secara ujian histokimia GUS. Aktiviti promoter dinilai berdasarkan intensiti biru pewarnaan GUS. Untuk promoter *EgPRP1*, rantau jujukan penuh iaitu RSP-3A (2000 bp ke hulu kodon pemula *EgPRP1*) menunjukkan keutamaan atau aktiviti yang dominan dalam semua peringkat pertumbuhan akar. Analisis delesi promoter menunjukkan rantau promoter terpendek iaitu RSP-3D (665 bp ke hulu kodon pemula *EgPRP1*) mengaruh pengekspresan gen secara spesifik-akar di peringkat pokok yang matang. Untuk promoter *EgMT*, didapati hanya delesi konstruk RSP-2D (1107 bp ke

hulu kodon pemula *EgMT*) mampu mengaruh pengekspresan GUS dalam akar dari mula peringkat awal perkembangan hingga pokok matang. Pengekspresan GUS yang diaruh oleh rantau jujukan penuh iaitu RSP-2A dan jujukan delesi RSP-2C dikesan dalam akar pokok matang, dengan intensiti yang lebih rendah berbanding promoter RSP-2D. Walau bagaimanapun, aktiviti promoter *EgMT* juga dapat dikesan dalam pod benih dan benih tidak matang, dengan tahap intensiti yang lebih rendah berbanding promoter promoter *Cauliflower Mosaic Virus 35S* (*CaMV35S*). Menariknya, pengekspresan GUS yang diaruh oleh promoter RSP-3A, RSP-2C dan RSP-2D juga dikesan di kawasan pemotongan bunga dan daun, menunjukkan aktiviti promoter boleh diaruh dengan rangsangan kecederaan. Hasil kajian menunjukkan kehadiran elemen-cis yang berfungsi sebagai pengawalatur negatif, yang mungkin bertanggungjawab untuk aktiviti spesifik akar. Keputusan juga mendapati kawasan 5' tidak diterjemahkan (5' UTR) dan jujukan ATATT yang ditakrifkan sebagai elemen-cis ROOTMOTIFAPOX1 adalah penting untuk aktiviti promoter yang kuat dalam akar. Secara keseluruhannya, kajian fungsional menunjukkan gen yang dikenalpasti melalui perlombongan data transkriptom dan (RT-qPCR), yang mempunyai kadar pengekspresan yang tinggi di setiap peringkat perkembangan akar berpotensi untuk mengaruh pengekspresan transgen yang kuat dalam tisu akar sama ada secara spesifik atau dominan. Keputusan menunjukkan bahawa promoter RSP-3A dan RSP-3D daripada *EgPRP1* dan RSP-2D daripada *EgMT* berpotensi membantu dalam kejuruteraan genetik sawit, terutamanya untuk meningkatkan hasil tanaman melalui pengubahsuaian trait-akar.

Kata kunci : Promoter spesifik-akar, Promoter dominan-akar, kajian pengekspresan, sawit, tembakau

SDG: MATLAMAT 2 : Kelaparan sifar, MATLAMAT 12 : Penggunaan dan penghasilan yang bertanggungjawab.



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LIST OF ABBREVIATIONS

5' RACE	5' Rapid amplification of cDNA ends
5'UTR	5'- untranslated region
A	Adenine
<i>AtNRT2.1</i>	<i>Arabidopsis thaliana</i> nitrate transporter 2
BA	6-Benzyladenine
<i>bar</i>	Bialaphos resistance gene
BLAST	Basic Local Alignment Search Tool
BLASTN	Basic Local Alignment Search Tool (nucleotide-nucleotide)
BLASTX	Basic Local Alignment Search Tool (translated nucleotide-protein)
bp	base pair
BSR	Basal stem rot
CaCl ₂	Calcium chloride
CAGE	Cap analysis gene expression
<i>CaMV35S</i>	Cauliflower Mosaic virus 35S promoter
<i>CaWRKY31</i>	WRKY protein
<i>CcHyPRP</i>	Hybrid-proline-rich protein
cDNA	complementary DNA
CIAP	Calf intestinal alkaline phosphatase
CPO	Crude palm oil
Cq	Cycle quantification
Ct	Cycle threshold
CTAB	Cetyl trimethylammonium bromide
DAG	Days after germination

DHS	DNas1 hypersensitive
DNA	Deoxyribonucleic acid
dNTP	Deoxynucleotide triphosphate
DsRED	Red fluorescent protein gene
dsRNA	Double strand RNA
DTT	Dithiothreitol
EDTA	Ethylenediaminetetraacetic acid
<i>EgMT</i>	<i>Elaeis guineensis</i> metallothionein
<i>EgPHT1</i>	High-affinity phosphate transporter
<i>EgPRP1</i>	<i>Elaeis guineensis</i> proline-rich protein
EST	Expressed sequence tag
ETBR	Ethidium bromide
FFB	Fresh fruit bunches
FPKM	per million mapped reads
GFP	Green fluorescent protein
<i>GmPRP2</i>	<i>Glycine max</i> proline rich protein
<i>GmTIP</i>	<i>Glycine max</i> tonoplast Intrinsic protein
GRAS	Gibberellin-responsive protein 2
GSPs	Gene-specific primers
GUS	β-glucuronidase gene
H ₂ O ₂	Hydrogen peroxidase
HTS	High-throughput sequencing
J1-1	Pepper defensin
Kb	Kilo base
KCl	Potassium chloride
LB	Luria-Bertani

LiCl	Lithium Chloride
<i>LjNRT2</i>	<i>Lotus japonica</i> nitrate transporter 2
MES	2-(N-morpholino)ethanesulfonic acid
MgCl ₂	Magnesium chloride
MgSO ₄	Magnesium sulfate
MPOB	Malaysian Palm Oil Board
mRNA	Messenger RNA
MS	Murashige and Skoog
<i>MsPRP2</i>	<i>Medicago sativa</i> proline rich protein
MT	Metallothionein
<i>MT3-B</i>	Oil palm metallothionein type 3
NAA	1-Naptheleneacetic acid
NaCl	Sodium chloride
NaPO ₄	Sodium Phosphate
<i>nptII</i>	Neomycin phosphotransferase II
NRRS	Nematode-responsive root-specific
<i>NtREL1</i>	Extensin-like protein
OD	Optical density
ORF	Open reading frame
<i>OsGRP7</i>	<i>Oryza sativa</i> glycine rich-protein 7
PCR	Polymerase chain reaction
PEG	Polyethylene-glycol
<i>PHT1</i>	Phosphate transferase
<i>Pht1;1</i>	<i>Hordeum vulgare</i> phosphate transferase
<i>pPRP3</i>	<i>Arabidopsis thaliana</i> proline rich protein
<i>PR10</i>	<i>Pinus monticola</i> Dougl. ex. D. Don pathogenesis-related protein

psi	Pounds per square inch
<i>PsPR10</i>	<i>Pinus strobus</i> pathogenesis-related protein
PVP	polyvinylpyrrolidone
<i>Pyk10</i>	<i>Arabidopsis thaliana</i> myrosinase
<i>RCc3</i>	HPS-like protein
RDA	Representational difference analysis
RFP	Red fluorescent protein
RNA	Ribonucleic acid
RNAi	RNA interference
RNA-Seq	Ribonucleic acid sequencing
RQN	RNA Quality Number
RT-qPCR	Real-time quantitative polymerase chain reaction
SAGE	Serial analysis of gene expression
SDS	Sodium dodecyl sulfate
<i>SLU7</i>	Pre-mRNA-splicing factor gene
SMRT	Single Molecule Sequencing Real Time
SSH	Subtractive hybridisation
TAE	1×Tris–Acetate EDTA
TE	Tris EDTA
TFBS	Transcription factor binding sites
<i>TIP2-2</i>	Tonoplast Intrinsic Protein
<i>TobRB7</i>	<i>Nicotiana tabacum</i> tonoplast intrinsic protein
Tris-HCl	TRIS Hydrochloride
TRs	Tandem repeats
TSS	Transcription start sites

UTR	Untranslated region
WAA	Week after anthesis
X-gal	5-Bromo-4-chloro-3-indolyl β -D-galactopyranoside
X-gluc	5-Bromo-4-Chloro-3-Indolyl- β -D-glucuronide
YEB	Yeast Extract Beef



CHAPTER 1

INTRODUCTION

Root is a below-ground part of a plant that provides support and firm soil anchorage. The roots absorb water and essential nutrients for plant growth. As they interact symbiotically with organisms, roots are also prone to attack by microbial pathogens that cause soil-borne diseases. The plant root system is also sensitive to environmental stimuli such as drought, salinity, and climate change. Soil-borne disease and the uncertainties of climate change could disrupt the root systems in absorbing water and nutrients (Longsaward *et al.*, 2023). The inefficient plant root system due to these factors could interfere with the plant's physiological processes and inhibit plant development, resulting in substantial losses in yield.

Oil palm (*Elaeis guineensis* Jacq) is a major Malaysian commodity crop threatened by soil-borne disease and climate change, resulting in substantial losses to the industry. *Ganoderma boninense*, which causes basal stem rot disease (BSR), develops from airborne spores and spreads in the soil through the roots (Naher *et al.*, 2013; Jazuli *et al.*, 2022). It is difficult to identify infected trees during the early stage of root disease since visible symptoms on aboveground parts appear only after the fungus has destroyed almost all of the root system (Siddiqui *et al.*, 2021; Longsaward *et al.*, 2023). BSR is also the main cause of the current oil palm tree crisis in many oil palm-producing countries, including Indonesia, Thailand, the Philippines, and Myanmar

(Zakaria, 2022). Besides, the oil palm industry also saw the effects of climate change, such as the El Nino and La Nina phenomena, that decreased crude palm oil production (Balu et al., 2018). Climate change is having an influence on agricultural crop yields as well as the frequency and severity of crop epidemic diseases all over the globe (Beddington, 2010; Stern and Stern, 2007; Ejaz et al., 2023). The drastic climate change will likely influence soil properties, which in turn may affect nutrient uptake by the palms (Rival, 2017). Adopting a genetic engineering approach to modify root architecture and function leads to developing stronger and healthier root systems that help maximise oil quality and yields. The strategy was successfully adopted for a wide variety of crops (Xue et al., 2016; Ramireddy et al., 2018; Luo et al., 2020). Genetic engineering is important for the improvement of oil palm yield since the global demand for food is estimated to increase by up to 70% by 2050 (van Dijk et al., 2021), and yield improvement should be focused on the available farmland (Masura et al., 2017; Rasid et al., 2020). To manipulate root systems, the availability of genetic engineering tools, particularly a regulatory element or promoter region, is fundamental to ensuring the genes of interest are expressed in a root-specific or preferential manner.

A promoter regulates the expression of a gene. By using the promoter specific to the root, the targeted recombinant proteins can be expressed in almost anything related to the root-soil interface. Protection against drought, increasing tolerance to salt and nutrient uptake, or increasing resistance to root pathogens are among the benefits of using root-specific promoters (Potenza et al., 2004). Several root-specific promoters have been isolated and

functionally characterised from plants, including rice (Li *et al.*, 2013), tobacco (Yamamoto *et al.*, 1991), *Arabidopsis* (Vijaybhaskar *et al.*, 2008), barley (Schünmann *et al.*, 2004), chickpea (Khandal *et al.*, 2020) and banana (James *et al.*, 2022). The efficiency of promoters is notably plant-species dependent, and the success of genetic modification also depends on its efficacious selection and use. The oil palm endogenous promoters useful for root manipulation were previously reported to be derived from Type-3 metallothionein-like (*MT3-B*) (Zubaidah and Siti Nor Akmar, 2005) and high-affinity phosphate transporter (*EgPHT1*) (Ahmadi *et al.*, 2018) genes. *MT3-B* promoter was inducible by the presence of Zn^{2+} and Fe^{2+} , while *EgPHT1* was triggered in response to Pi deficiency. Although strong inducible promoters can be a great benefit, sometimes that may limit their function in a broad application of genetic engineering. Since the published information on root-specific promoters from oil palm is still limited, discovering more root-specific promoters widens the range of promoters for genetic engineering. This is also necessary to fulfil the requirements of multiple transgenes stacking and cisgenesis technologies (Chen *et al.*, 2010a; Holme *et al.*, 2013; Masura *et al.*, 2019; Koul, 2022) to avoid gene silencing.

Numerous studies have reported that promoters beneficial for root modification are derived from root-specific or root-preferential genes (Yamamoto *et al.*, 1991; Vijaybhaskar *et al.*, 2008; Li *et al.*, 2013; Khandal *et al.*, 2020; James *et al.*, 2022). Therefore, it is hypothesised that the promoter sequences of genes that were up-regulated during oil palm root development (from *in vitro* tissue culture to field planting) could be used in genetic engineering as root-specific

promoters. Mining oil palm transcriptome data through RNA-sequencing (RNA-Seq) will accelerate gene discovery. The availability of oil genome data (Singh *et al.*, 2013) and abundant transcriptome data (Singh *et al.*, 2013; Ho *et al.*, 2016; Morris *et al.*, 2020) generated from various oil palm tissues facilitates the analysis using RNA-Seq. Among the promoters successfully identified using this approach were novel tissue-specific promoters from citrus and plum (Dasgupta *et al.*, 2020) and a root-specific promoter from peanut (Geng *et al.*, 2014).

The current study aimed to identify oil palm promoters suitable for expressing transgenes in a root-specific or preferential manner that may have great potential for oil palm's root system modification. Using specific promoters suitable for a specific target transgene is essential for fine-tuning the control of transgene expression and avoiding the disruption of normal plant growth. The root-specific promoter can be an important tool for precise gene modifications to protect and improve plants' root systems and enhance crop quality and yield. To accomplish the aim of the study, the work was carried out with four main objectives:

1. To screen and identify root-specific gene candidates from the existing oil palm transcriptome libraries.
2. To profile the expression levels of the selected genes using real-time quantitative polymerase chain reaction (RT-qPCR).
3. To isolate promoter regions and to carry out deletion promoter analyses.
4. To evaluate the activities of the promoters for stable and transient expressions in tobacco and oil palm tissues, respectively.

REFERENCES

- Abe, H., Urao, T., Ito, T., Seki, M., Shinozaki, K., & Yamaguchi-Shinozaki, K. (2003). *Arabidopsis AtMYC2 (bHLH) and AtMYB2 (MYB) function as transcriptional activators in abscisic acid signalling. Plant Cell*, 15(1), 63-78. <https://doi.org/10.1105/tpc.006130>.
- Adams, J. (2008). Transcriptome: connecting the genome to gene function. *Nature Education*, 1(1),195.
- Adams, M. D., Kelley, J. M., Gocayne, J. D., Dubnick, M., Polymeropoulos, M. H., Xiao, H., Merrill, C. R., Wu, A., Olde, B., & Moreno, R. F. (1991). Complementary DNA sequencing: expressed sequence tags and human genome project. *Science*, 252(5013), 1651–1656. <https://doi.org/10.1126/science.2047873>.
- Ahmadi, F., Siti Nor Akmar, A., Kadkhodaei, S., Ijab, S. M., Hamzah, L., Aziz, M. A., Rahman, Z. A., & Rabiah Syed Alwee, S. S. (2018). Functional characterization of the gene promoter for an *Elaeis guineensis* phosphate starvation-inducible, high-affinity phosphate transporter in both homologous and heterologous model systems. *Plant Physiology and Biochemistry*, 127, 320-335. <https://doi.org/10.1016/j.plaphy.2018.04.004>.
- Ahn, J. H., Choi, Y., Kwon, Y. M., Kim, S-G., Choild, Y. D., & Lee, J. S. (1996). A novel extensin gene encoding a hydroxyproline-rich glycoprotein requires sucrose for its wound-inducible expression in transgenic plants. *The Plant Cell*, 8, 1477-1490. <https://doi.org/doi: 10.1105/tpc.8.9.1477>.
- Ali, S., & Kim, W. C. (2019). A fruitful decade using synthetic promoters in the improvement of transgenic plants. *Frontiers in Plant Science*, 10, 1433. <https://doi.org/10.3389/fpls.2019.01433>.
- Alotaibi, S. S., Sparks, C. A., Parry, M. A. J., Simkin, A. J., & Raines, C. A. (2018). Identification of leaf promoters for use in transgenic wheat. *Plants (Basel)*, 7(2). <https://doi.org/10.3390/plants7020027>.
- Alpern, D., Gardeux, V., Russeil, J., Mangeat, B., Meireles-Filho, A. C. A., Breyse, R., Hacker, D. & Deplancke, B. (2019). BRB-seq: ultra-affordable high-throughput transcriptomics enabled by bulk RNA barcoding and sequencing. *Genome Biology*, 20(1), 71. <https://doi.org/10.1186/s13059-019-1671-x>.
- Altschul, S. F., Madden, T. L., Schäffer, A. A., Zhang, J., Zhang, Z., Miller, W., & Lipman, D. J. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research*, 25(17), 3389-3402. <https://doi:10.1093/nar/25.17.3389>.
- Apriyanto, A., Ernawan, R., Putra, M. K. A., Susilo, R., Ajambang, W., Pancoro, A., & Wibowo, C. S. (2023). Gene expression profiles of oil palm leaves

- from different oil yields and genetic population: Transcriptomic dataset. *Data Brief*, 48,109043. <https://doi.org/10.1016/j.dib.2023.109043>.
- Arai-Sanoh Y, T. T., Yoshinaga, S., Nakano, H., Kojima, M., Sakakibara, H., Kondo, M., & Uga, Y. (2014). Deep rooting conferred by DEEPER ROOTING 1 enhances rice yield in paddy fields. *Scientific Reports*, 4, 5563. <https://doi.org/10.1038/srep05563>.
- Asvini, B., & Jithesh. (2018). Impact of using artificial fertilizer in soil. *International Journal of Pure and Applied Mathematics*, 119, 47-55.
- Badai, S. S., Chan, K-L., Chan, P-K., Parveez, G. K. A., & Rasid, O. A. (2019). Identification of genes preferentially expressed in mesocarp tissue of oil palm using in silico analysis of transcripts. *J Oil Palm Res.*, 31, 540-549. <https://doi.org/10.21894/jopr.2019.0022>.
- Bahariah, B., Parveez, G.K.A., Masani, A.M.Y., & Khalid, N. (2014). The use of mannose selection system for gene transfer in tobacco plants (*Nicotiana Tabacum* L.), a model plant for oil palm transformation. *J Oil Palm Res.*, 26 (2), 154-162.
- Balu, N., Ismail, A., Hashim, N., Ismail, N., Shahari, D. N., Idris, N. A. N., Omar, N., Salleh, K. M., Hassan, N. A. M., & Kushairi, A. (2018). Malaysia : 100 years of resilient palm oil economic performance. *J Oil Palm Res.*, 30(1), 13-25. <https://doi.org/10.21894/jopr.2018.0014>.
- Baranowskij, N., Frohberg, C., Prat, S., & Willmitzer, L. (1994). A novel DNA binding protein with homology to Myb oncoproteins containing only one repeat can function as a transcriptional activator. *Embo J.*, 13(22), 5383-5392. <https://doi.org/10.1002/j.1460-2075.1994.tb06873.x>.
- Barrett, L. W., Fletcher, S., & Wilton, S. D. (2012). Regulation of eukaryotic gene expression by the untranslated gene regions and other noncoding elements. *Cell Mol. Life Sci.*, 69, 3613–3634. <https://doi.org/10.1007/s00018-012-0990-9>.
- Baumann, K., Paolis, A. D. P., & Gualberti, G. (1999). The DNA binding site of the Dof protein NtBBF1 is essential for tissue-specific and auxin-regulated expression of the *ro/B* oncogene in plants. *Plant Cell*, 11, <https://doi.org/10.1105/tpc.11.3.323>.
- Beddington, J. (2010). Food security: contributions from science to a new and greener revolution. *Philos. Trans. R. Soc. B.*, 365(1537), 61–71. <https://doi.org/10.1098/rstb.2009.0201>.
- Benfey, P. N., & Chua, N. H. (1989). Regulated genes in transgenic plants. *Science*, 244, 174–189. <https://doi.org/10.1126/science.244.4901.174>.
- Bernhardt, C., & Tierney, M. L. (2000). Expression of *AtPRP3*, a proline-rich structural cell wall protein from *Arabidopsis*, is regulated by cell-type-specific developmental pathways involved in root hair formation. *Plant Physiol.*, 122, 705–714. <https://doi.org/10.1104/pp.122.3.705>.

- Bevan, M. (1984). Binary *Agrobacterium* vectors for plant transformation. *Nucleic Acids Res.*, 12(22), 8711-8721. <https://doi.org/10.1093/nar/12.22.8711>.
- Bhattacharya, R., Koramutla, M. K., Negi, M., Pearce, G., & Ryan C. A. (2013). Hydroxyproline-rich glycopeptide signals in potato elicit signalling associated with defense against insects and pathogens. *Plant Science*, 207, 88–97. <https://doi.org/10.1016/j.plantsci.2013.03.002>.
- Boron, A. K., Van Orden, J., Nektarios Markakis, M., Mouille, G., Adriaensen, D., Verbelen, J. P., Höfte, H., & Vissenberg, K. (2014). Proline-rich protein-like PRPL1 controls elongation of root hairs in *Arabidopsis thaliana*. *J Exp. Bot.*, 65(18), 5485-5495. <https://doi.org/10.1093/jxb/eru308>.
- Boyle, B., & Brisson, N. (2001). Repression of the defense gene *PR-10a* by the single-stranded DNA binding protein SEBF. *Plant Cell*, 13(11), 2525-2537. <https://doi.org/10.1105/tpc.010231>.
- Brahmachary, M., Guilmatre, A., Quilez, J., Hasson, D., Borel, C., Warburton, P., & Sharp, A. J. (2014). Digital genotyping of macrosatellites and multicopy genes reveals novel biological functions associated with copy number variation of large tandem repeats. *PLoS Genet.*, 10(6), e1004418. <https://doi.org/10.1371/journal.pgen.1004418>.
- Bustin, S. A., Benes, V., Garson, J. A., Hellems, J., Huggett, J., Kubista, M., Kubista, M., Mueller, R., Nolan, T., Pfaffl, M. W., Shipley, G. L., Vandesompele, J., & Wittwer, C.T. (2009). The MIQE guidelines: Minimum information for publication of quantitative real-time PCR experiments. *Clinical Chemistry*, 55(4), 611-622. <https://doi.org/10.1373/clinchem.2008.112797>.
- Capdevila, M., & Atrian, S. (2011). Metallothionein protein evolution: a miniassay. *Journal of Biological Inorganic Chemistry*, 16(7), 977-989. <https://doi.org/10.1007/s00775-011-0798-3>.
- Chan, P-L., Rose, R. J., Abdul Murad, A. M., Zainal, Z., Low, E.T. L., Ooi, L. C-L., Yahya, S., & Singh, R. (2014). Evaluation of reference genes for quantitative real-time PCR in oil palm elite planting materials propagated by tissue culture. *PloS One*, 9(6), e99774-e99774. <https://doi.org/10.1371/journal.pone.0099774>.
- Chen, C., & Chen, Z. (2002). Potentiation of developmentally regulated plant defense response by AtWRKY18, a pathogen-induced *Arabidopsis* transcription factor. *Plant Physiol.*, 129(2), 706-716. <https://doi.org/10.1104/pp.001057>.
- Chen, L., Jiang, B., Wu, C., Sun, S., Hou, W., & Han, T. (2014). *GmPRP2* promoter drives root-preferential expression in transgenic *Arabidopsis* and soybean hairy roots. *BMC Plant Biology*, 14(1), 245. <https://doi.org/10.1186/s12870-014-0245-z>.

- Chen, L., Jiang, B., Wu, C., Sun, S., Hou, W., & Han, T. (2015). The characterization of *GmTIP*, a root-specific gene from soybean, and the expression analysis of its promoter. *Plant Cell, Tissue and Organ Culture*, 121(2), 259-274. <https://doi.org/10.1007/s11240-014-0682-2>.
- Chen, L., Tu, Z., Hussain, J., Cong, L., Yan, Y., Jin, L., Yang, G., & He, G. (2010b). Isolation and heterologous transformation analysis of a pollen-specific promoter from wheat (*Triticum aestivum* L.). *Mol. Biol. Rep.*, 37:737–744. <https://doi.org/10.1007/s11033-009-9582-7>.
- Chen, P. Y., Wang C. K., Soong, S.C., & To, K. Y. (2003). Complete sequence of the binary vector pBI121 and its application in cloning T-DNA insertion from transgenic plants. *Mol. Breed.*, 11, 287-293. <https://doi.org/10.1023/A:1023475710642>.
- Chen, Q-J., Xie, M., Ma, X-X., Dong, L., Chen, J., & Wang, X-C. (2010a). MISSA is a highly efficient *in vivo* DNA assembly method for plant multiple-gene transformation. *Plant Physiol.*, 153(1), 41–51. <https://doi.org/10.1104/pp.1109.152249>.
- Choi, D. W., Song, J. Y., Kwon, Y. M., & Kim, S. G. (1996). Characterization of cDNA encoding a proline-rich 14 kDa protein in developing cortical cells of the roots of bean (*Phaseolus vulgaris*) seedlings. *Plant Mol. Biol.*, 30, 973–982. <https://doi.org/10.1007/BF00020808>.
- Chow, C. N., Lee, T. Y., Hung, Y. C., Li, G. Z., Tseng, K. C., Liu, Y. H., Kuo, P. L., Zheng, H. Q., & Chang, W. C. (2019). PlantPAN 3.0: a new and updated resource for reconstructing transcriptional regulatory networks from ChIP-seq experiments in plants. *Nucleic Acids Res.*, 47(D1), D1155-d1163. <https://doi.org/10.1093/nar/gky1081>.
- Christensen, A. H., & Quail, P. H. (1996). Ubiquitin promoter-based vectors for high-level expression of selectable marker genes in monocotyledonous plant. *Transgenic Res.*, 5, 213-218. <https://doi.org/10.1007/BF01969712>.
- Chung, K. J., Hwang, S. K., Hahn, B. S., Kim, K. H., Kim, J. B., Kim, Y. H., Yang, J. S., & Ha, S. H. (2008). Authentic seed-specific activity of the *Perilla* oleosin 19 gene promoter in transgenic *Arabidopsis*. *Plant Cell Rep.*, 27, 29–37. <https://doi.org/10.1007/s00299-007-0440-6>.
- Claros, M. G., & Canovas, F. M. (1999). RNA isolation from plant tissues: A practical experience for biological undergraduate. *Biochemical Education*, 27, 110-113. [https://doi.org/10.1016/S0307-4412\(98\)00289-1](https://doi.org/10.1016/S0307-4412(98)00289-1).
- Cobbett, C., & Goldbrough, P. B. (2002). Phytochelatins and metallathioneins: roles in heavy metal detoxification and homeostasis. *Annu. Rev. Plant Biol.*, 53, 159–182. <https://doi.org/10.1146/annurev.arplant.53.100301.135154>.

- Cook, R. J. (2001). Management of wheat and barley root diseases in modern farming systems. *Australasian Plant Pathology*, 30(2), 119-126. <https://doi.org/10.1071/AP01010>.
- da Cunha Cruz, Y., Martins Scarpa, A. L., Pereira, M. P., Mauro de Castro, E., & Pereira, F. J. 2020. Root anatomy and nutrient uptake of the cattail *Typha domingensis* Pers. (*Typhaceae*) grown under drought condition. *Rhizosphere*, 16,100253. <https://doi.org/10.1016/j.rhisph.2020.100253>.
- Dasgupta, K., Hotton, S., Belknap, W., Syed, Y., Dardick, C., Thilmony, R., & Thomson, J. G. (2020). Isolation of novel citrus and plum fruit promoters and their functional characterization for fruit biotechnology. *BMC Biotechnol.*, 20(1), 43. <https://doi.org/10.1186/s12896-020-00635-w>.
- De Bauw, P., Vandamme, E., Lupembe, A., Mwakasege, L., Senthilkumar, K., Dramé, K. N., & Merckx, R. (2019). Anatomical root responses of rice to combined phosphorus and water stress – relations to tolerance and breeding opportunities. *Functional Plant Biology*, 46(11), 1009-1022. <https://doi.org/10.1071/FP19002>.
- Deikman J, X. R., Kneissl, M. L., Ciardi, J. A., Kim, K. N., & Pelah, D. (1998). Separation of cis elements responsive to ethylene, fruit development, and ripening in the 5'-flanking region of the ripening-related *E8 gene*. *Plant Mol. Biol.*, 37, 1001–1011. <https://doi.org/10.1023/A:1006091928367>.
- Díaz-Martín, J., Almoguera, C., Prieto-Dapena, P., Espinosa, J. M., & Jordano, J. (2005). Functional interaction between two transcription factors involved in the developmental regulation of a small heat stress protein gene promoter. *Plant Physiol.*, 139(3), 1483-1494. <https://doi.org/10.1104/pp.105.069963>.
- Dietz-Pfeilstetter, A. (2010). Stability of transgene expression as a challenge for genetic engineering. *Plant Sci.*, 179, 164–167. <https://doi.org/10.1016/j.plantsci.2010.1004.1015>.
- Dignam, B. E. A., Marshall, S. D. G., Wall, A. J., Mtandavari, Y. F., Gerard, E. M., Hicks, E., Cameron, C., Aalders, L. T., Shi, S., & Bell, N. L. (2022). Impacts of soil-borne disease on plant yield and farm profit in dairying soils. *Journal of Sustainable Agriculture and Environment*, 1(1), 16-29. <https://doi.org/10.1002/sae2.12009>.
- Dong, C. J., Wang, Y., Yu, S. S., & Liu, J. Y. (2010). Characterization of a novel rice metallothionein gene promoter: its tissue specificity and heavy metal responsiveness. *J Integr. Plant Biol.*, 52(10), 914-924. <https://doi.org/10.1111/j.1744-7909.2010.00966.x>.
- Doyle, J. J., & Doyle, J. L. (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue *Phytochem. Bull*, 19, 11-15.

- Duan, L., Yu, J., Xu, L., Tian, P., Hu, X., Song, X., & Pan, Y. (2019). Functional characterization of a type 4 metallothionein gene (*CsMT4*) in cucumber. *Horticultural Plant Journal*, 5(3), 120-128. <https://doi.org/10.1016/j.hpj.2019.04.002>.
- Dunn, M. A., White, A. J., Vural, S., & Hughes, M. A. (1998). Identification of promoter elements in a low-temperature-responsive gene (*blt4.9*) from barley (*Hordeum vulgare* L.). *Plant Mol. Biol.*, 38(4), 551-564. <https://doi.org/10.1023/a:1006098132352>.
- Đúranová, H., Šimora, V., Ďurišová, L., Olexiková, L., Kovár, M., & Požgajová, M. 2023. Modifications in ultrastructural characteristics and redox status of plants under environmental stress: A Review. *Plants*, 12(8), 1666. <https://doi.org/10.3390/plants12081666>.
- Ebener, W., Fowler, T. J., Suzuki, H., Shaver, J., & Tierney, M. L. (1993). Expression of *DcPRP1* is linked to carrot storage root formation and is induced by wounding and auxin treatment. *Plant Physiol*, 101(1), 259-65. <https://doi.org/10.1104/pp.101.1.259>.
- Edwards, K., Johnstone, C., & Thompson, C. (1991). A simple and rapid method for the preparation of plant genomic DNA for PCR analysis. *Nucleic Acids Res.*, 19, 1349. <https://doi.org/10.1093/nar/19.6.1349>.
- Ejaz, M. R., Jaoua, S., Ahmadi, M., & Shabani, F. (2023). An examination of how climate change could affect the future spread of *Fusarium spp.* around the world, using correlative models to model the changes. *Environmental Technology & Innovation*, 31, 103177. <https://doi.org/10.1016/j.eti.2023.103177>.
- Ellerström, M., Stålberg, K., Ezcurra, I., & Rask, L. (1996). Functional dissection of a napin gene promoter: Identification of promoter elements required for embryo and endosperm-specific transcription. *Plant Molecular Biology*, 32(6), 1019-1027. <https://doi.org/10.1007/BF00041385>.
- Elmayan, T., & Tepfer, M. (1995). Evaluation in tobacco of the organ specificity and strength of the *rolD* promoter, domain A of the 35S promoter and the 35S2 promoter. *Transgenic Res.*, 4(6), 388-396. <https://doi.org/10.1007/bf01973757>.
- Elmayan, T., & Vaucheret, H. (1996). Expression of single copies of a strongly expressed 35S transgene can be silenced post transcriptionally. *Plant J.*, 9, 787-797.
- Eulgem, T., Rushton, P. J., Schmelzer, E., Hahlbrock, K., & Somssich, I. E. (1999). Early nuclear events in plant defence signalling: rapid gene activation by WRKY transcription factors. *Embo J.*, 18(17), 4689-4699. <https://doi.org/10.1093/emboj/18.17.4689>.

- Evans, I. M., Swinhoe, R., Gatehouse, L. N., Gatehouse, J. A., Boulter, D. (1988). Distribution of root mRNA species in other vegetative organs of pea (*Pisum sativum* L.). *Molecular and General Genetics*, 214(1), 153-157. <https://doi.org/10.1007/BF00340194>.
- Fahmideh, L., Khodadadi, E., Khodadadi, E., Zeinalzadeh, E., Dao, S., Köse, Ş., & Kafil, H. S. (2023). Transcriptome analysis methods: From the serial analysis of gene expression and microarray to sequencing new generation methods. *Biointerface Research in Applied Chemistry*, 13(6), 543. <https://doi.org/510.33263/BRIAC33136.33543>.
- Fehlberg, V., Vieweg, M. F., Dohmann, E. M., Hohnjec, N., Pühler, A., Perlick, A. M., & Küster, H. (2005). The promoter of the leghaemoglobin gene *VfLb29*: functional analysis and identification of modules necessary for its activation in the infected cells of root nodules and in the arbuscule-containing cells of mycorrhizal roots. *J Exp. Bot.*, 56(413), 799-806. <https://doi.org/10.1093/jxb/eri074>.
- Fei, H. M., Chaillou, S., Hirel, B., Mahon, J. D., & Vessey, J. K. (2003). Overexpression of a soybean cytosolic glutamine synthetase gene linked to organ-specific promoters in pea plants grown in different concentrations of nitrate. *Planta*, 216, 467-474. <https://doi.org/10.1007/s00425-002-0873-7>.
- Ferreira, T. M. M., Filho, J. A. F., Leão, A. P., de Sousa, C. A. F., & Souza, M. T. J. (2022). Structural and functional analysis of stress-inducible genes and their promoters selected from young oil palm (*Elaeis guineensis*) under salt stress. *BMC Genomics*, 23(1), 735. <https://doi.org/10.1186/s12864-022-08926-6>.
- Fess, T. L., Kotcon, J. B., & Benedito, V. A. (2011). Crop breeding for low input agriculture: A sustainable response to feed a growing world population. *Sustainability*, 3, 1742-1772. <https://doi.org/10.3390/su3101742>
- Fizree, P. M. A. A., Shaharuddin, N. A., Ho, C. L., Masura, S. S., Manaf, M. A. A., Parveez, G. K. A., & Masani, M. Y. A. (2019). Evaluation of transient *DsRED* gene expression in oil palm embryogenic calli. *Scientia Horticulturae*, 257, 108679. <https://doi.10.1016/j.scienta.2019.108679>.
- Foster, S. J., Park, T. H., Pel, M., Brigneti, G., Sliwka, J., Jagger, L., van der Vossen, E., & Jones, J. D. G. (2009). *Rpi-vnt1.1*, a *Tm-2²* homolog from *Solanum venturii*, confers resistance to potato late blight. *Mol. Plant Microbe Interact.*, 22, 589-600. <https://doi.org/10.1094/MPMI-22-5-0589>.
- Fraisier, V., Gojon, A., Tillard, P., & Daniel-Vedele, F. (2000). Constitutive expression of a putative high-affinity nitrate transporter in *Nicotiana plumbaginifolia*: Evidence for post-transcriptional regulation by a reduced nitrogen source. *The Plant Journal*, 23, 489-496. <https://doi.org/10.1046/j.1365-313x.2000.00813.x>.

- Freisinger, E. (2011). Structural features specific to plant metallothioneins. *Journal of Biological Inorganic Chemistry*, 16(7), <https://doi.org/10.1007/s00775-011-0801-z>.
- Frith, M. C., Pheasant, M., & Mattick, J. S. (2005). The amazing complexity of the human transcriptome. *European Journal of Human Genetics*, 13, 894–897. <https://doi.org/10.1038/sj.ejhg.5201459>.
- Frohman, M. A., Dush, M. K., & Martin, G. R. (1988). Rapid production of full length cDNAs from rare transcripts: Amplification using a single gene-specific oligonucleotide primer. *Proc. Natl. Acad. Sci. USA*, 85, 8998–9002. <https://doi.org/10.1073/pnas.85.23.8998>.
- Fujino, K., Obara, M., & Sato, K. (2014). Diversification of the plant-specific hybrid glycine-rich protein (*HyGRP*) genes in cereals. *Frontiers in Plant Science*, 5, 489–489. <https://doi.org/10.3389/fpls.2014.00489>.
- Geng, L., Duan, X., Liang, C., Shu, C., Song, F., & Zhang, J. (2014). Mining tissue-specific contigs from peanut (*Arachis hypogaea* L.) for promoter cloning by deep transcriptome sequencing. *Plant and Cell Physiology*, 55(10), 1793–1801. <https://doi.org/10.1093/pcp/pcu111>.
- Gamuyao, R., Chin, J. H., Pariasca-Tanaka, J., Pesaresi, P., Catausan, S., Dalid, C., Slamet-Loedin, I., Tecson-Mendoza, E. M., Wissuwa, M., & Heuer, S. (2012). The protein kinase *Pstol1* from traditional rice confers tolerance of phosphorus deficiency. *Nature (London)*, 488, 535–539. <https://doi.org/10.1038/nature11346>. PMID: 22914168.
- Ghanem, M. E., Hichri, I., Smigocki, A. C., Albacete, A., Fauconnier, M. L., Diatloff, E., Martinez-Andujar, C. Lutts, S. Dodd, I. C., & Pérez-Alfocea, F. 2011. Root-targeted biotechnology to mediate hormonal signalling and improve crop stress tolerance. *Plant Cell Rep.*, 30(5), 807–823. <https://doi.org/10.1007/s00299-011-1005-2>.
- Gonsalves, D. (1998). Control of papaya ringspot virus in papaya: a case study. *Annu. Rev. Phytopathol.*, 36, 415–437. <https://doi.org/10.1146/annurev.phyto.36.1.415>.
- Gonsalves, D. (2006). Transgenic papaya: Development, release, impact, and challenges. *Advances in Virus Research*, 67, 317–354.
- Gothandam, K. M., Nalini, E., Karthikeyan, S., & Shin, J. S. (2010). OsPRP3, a flower specific proline-rich protein of rice, determines extracellular matrix structure of floral organs and its overexpression confers cold-tolerance. *Plant Mol. Biol.*, 72, 125–135. <https://doi.org/10.1007/s11103-009-9557-z>.
- Guignard, M. S., Leitch, A. R., Acquisti, C., Eizaguirre, C., Elser, J. J., Hessen., D. O., Jeyasingh, P. D., Neiman, M., Richardson, A. E., Soltis, P. S., Soltis, D. E., Stevens, C. J., Trimmer, M., Weider, L. J., Woodward, G., & Leitch, I. J. (2017). Impacts of nitrogen and phosphorus: From genomes

- to natural ecosystems and agriculture. *Front. Ecol. Evol.*, 5, 70. <https://doi.org/10.3389/fevo.2017.00070>.
- Gujjar, R. S., Pathak, A. D., Karkute, S. G., & Supaibulwatana, K. (2019). Multifunctional proline rich proteins and their role in regulating cellular proline content in plants under stress. *Biologia Plantarum*, 63, 448-454. <https://doi.org/10.32615/bp.32019.32078>.
- Gunadi, A., Dean, E. A., & Finer, J. J. (2019). Transient transformation using particle bombardment for gene expression analysis. *Methods Mol. Biol.*, 1864, 67-79. https://doi.org/10.1007/978-1-4939-8778-8_5.
- Guo, W. J., Bundithya, W., & Goldsbrough, P. B. (2003). Characterization of the *Arabidopsis* metallothionein gene family: Tissue-specific expression and induction during senescence and in response to copper. *New Phytol.*, 159, 369–381. <https://doi.org/10.1046/j.1469-8137.2003.00813.x>.
- Hanin, A. N., Parveez, G. K. A., Rasid, O. A., & Masani, M. Y. A. (2020). Biolistic-mediated oil palm transformation with alfalfa glucanase (*AGLU1*) and rice chitinase (*RCH10*) genes for increasing oil palm resistance towards *Ganoderma boninense*. *Industrial Crops and Products*, 144, 112008. <https://doi.org/10.1016/j.indcrop.2019.112008>.
- Hashemipetroudi, S. H., Nematzadeh, G., Ahmadian, G., Yamchi, A., & Kuhlmann, M. (2018). Assessment of DNA contamination in RNA samples based on ribosomal DNA. *J Vis. Exp.*, 131, 55451. <https://doi.org/10.3791/55451>.
- He, X., Qu, B., Li, W., Zhao, X., Teng, W., Ma, W., Ren, Y., Li, B., Li, Z., & Tong, Y. (2015). The nitrate-inducible NAC transcription factor TaNAC2-5A controls nitrate response and increases wheat yield. *Plant Physiology*, 169, 1991–2005. <https://doi.org/10.1104/pp.15.00568>.
- Hernandez-Garcia, C. M., & Finer, J. J. (2014). Identification and validation of promoters and cis-acting regulatory elements. *Plant Science*, 217-218, 109-119. <https://doi.org/10.1016/j.plantsci.2013.12.007>.
- Hiei, Y., & Komari, T. (2008). Stable inheritance of transgenes in rice plants transformed by *Agrobacterium tumefaciens*. *Rice Genetics III*, 131-142. <https://doi.org/10.1038/3511>.
- Higo, K., Ugawa, Y., Iwamoto, M., & Korenaga, T. (1999). Plant cis-acting regulatory DNA elements (PLACE) database: 1999. *Nucleic Acids Research*, 27(1), 297-300. <https://doi.org/10.1093/nar/27.1.297>.
- Ho, C.-L., Tan, Y.-C., Yeoh, K.-A., Ghazali, A.-K., Yee, W.-Y., & Hoh, C.-C. (2016). *De novo* transcriptome analyses of host fungal interactions in oil palm (*Elaeis guineensis* Jacq.). *BMC Genomics*, 17, 66. <https://doi.org/10.1186/s12864-016-2368-0>
- Holbein, J., Franke, R. B., Marhavý, P., Fujita, S., Górecka, M., Sobczak, M., Geldner, N., Schreiber, L., W. Grudler, F. M., & Siddique, S. (2019). Root

- endodermal barrier system contributes to defence against plant-parasitic cyst and root-knot nematodes. *The Plant Journal*, 100(2), 221-236. <https://doi.org/210.1111/tpj.14459>.
- Holme, I. B., Wendt, T. & Holm, P. B. (2013). Intragenesis and cisgenesis as alternatives to transgenic crop development. *Plant Biotechnol. J.*, 11(4), 395–407. <https://doi.org/10.1111/pbi.12055>.
- Hsieh, H.-M., Liu, W.-K., & Huang, P. C. (1995). A novel stress-inducible metallothionein-like gene from rice. *Plant Mol Biol.*, 28, 381–389. <https://doi.org/10.1007/BF00020388>.
- Hsieh, H. M., Liu, W. K., Chang, A., & Huang, P. C. (1996). RNA expression patterns of a type 2 metallothionein-like gene from rice. *Plant Mol Biol.*, 32(3), 525-529. <https://doi:10.1007/bf00019104>.
- Hsieh, P.-H., Oyang, Y.-J., & Chen, C.-Y. (2019). Effect of de novo transcriptome assembly on transcript quantification. *Scientific Reports*, 9(1), 8304. <https://doi.org/10.1038/s41598-019-44499-3>.
- Huang, L. Y., Zhang, F., Qin, Q., Wang, W. S., & Fu, B. Y. (2015). Identification and validation of root-specific promoters in rice. *J Integr. Agr.*, 14(1), 1–10. [https://doi.org/10.1016/S2095-3119\(14\)60763-2](https://doi.org/10.1016/S2095-3119(14)60763-2).
- Hubank, M., & Schatz, D.G. (1994). Identifying differences in mRNA expression by representational difference analysis of cDNA. *Nucleic Acids Res.*, 22, 5640–5648. <https://doi.org/10.1093/nar/22.25.5640>.
- Hudson, O., Fulton, J. C., Dong, A. K., Dufault, N. S., & Ali, M. E. (2021). *Fusarium oxysporum f. sp. niveum* molecular diagnostics past, present and future. *Int. J. Mol. Sci.*, 22(18), 9735. <https://doi.org/10.3390/ijms22189735>.
- Hushiarian, R., Yusof, N. A., & Dutse, S. W. (2013). Detection and control of *Ganoderma boninense*: strategies and perspectives. *SpringerPlus*, 2, 555. <https://doi.org/10.1186/2193-1801-2-555>.
- Ihsan, M. Z., Ahmad, S. J., Shah, Z. H., Rehman, H. M., Aslam, Z., Ahuja, I., Bones, A. M., & Ahmad, J. N. (2017). Gene mining for proline based signaling proteins in cell wall of *Arabidopsis thaliana*. *Frontiers in Plant Science*, 8, 215995. <https://doi.org/215910.213389/fpls.212017.200233>.
- Ilardi, V., & Tazzava, M. (2015). Biotechnological strategies and tools for Plum pox virus resistance: Trans-, intra-, cis-genesis, and beyond. *Frontiers in Plant Science*, 6, 379. <https://doi.org/10.3389/fpls.2015.00379>.
- Izawati, A. M. D., Masani, A. M. Y., Parveez, G.K.A., & Ismanizan, I. (2017). 2-deoxyglucose selection system for *Agrobacterium*-mediated transformation of *Solanum lycopersion* and *Nicotiana tabacum* plants. *Australian J. Crop Sci.*, 7, 813-820. <https://doi.org/10.21475/ajcs.17.11.07.pne425>.

- James, A., Paul, J., Souvan, J., Cooper, T., Dale, J., Harding, R., & Deo, P. (2022). Assessment of root-specific promoters in banana and tobacco and identification of a banana *TIP2* promoter with strong root activity. *Frontiers in Plant Science*, 13. <https://doi.org/10.3389/fpls.2022.1009487>.
- Janssen, E. M., Liu, C., & Van der Fels-Klerx, H. J. (2018). *Fusarium* infection and trichothecenes in barley and its comparison with wheat. *World Mycotoxin Journal* 11(1): 33 - 46. <https://doi.org/10.3920/WMJ2017.2255>.
- Jazuli, N. A., Kamu, A., Chong, K. P., Gabda, D., Hassan, A., Abu Seman, I., & Ho, C. M. (2022). A review of factors affecting *Ganoderma* basal stem rot disease progress in oil palm. *Plants (Basel)*, 11(19), 2462. <https://doi.org/10.3390/plants11192462>.
- Jeong, J. S., Kim, Y. S., Baek, K. H., Jung, H., Ha, S. H., Do, C. Y., Kim, M., Reuzeau, C., & Kim, J. K. (2010). Root-specific expression of *OsNAC10* improves drought tolerance and grain yield in rice under field drought conditions. *Plant Physiology and Biochemistry*, 153, 185-197. <https://doi.org/10.1104/pp.110.154773>.
- Jones, M.O., Manning, K., Andrews, J., Wright, C., Taylor, I. B., & Andrew, J. T. (2008). The promoter from *SIREO*, a highly-expressed, root-specific *Solanum lycopersicum* gene, directs expression to cortex of mature roots. *Functional Plant Biology*, 35(12), 1224-1233. <https://doi.org/10.1071/FP08139>.
- Joshi, C. P. (1987). An inspection of the domain between putative tata box and translation start site in 79 plants genes. *Nucleic Acids Res.*, 15, 993-1001. <https://doi.org/10.1093/nar/15.16.6643>.
- Joshi, I., Kumar, A., Kohli, D., Bhattacharya, R., Sirohi, A., Chaudhury, A., & Jain, P. K. (2022). Gall-specific promoter, an alternative to the constitutive *CaMV35S* promoter, drives host-derived RNA interference targeting *Mimsp2* gene to confer effective nematode resistance. *Frontiers in Plant Science*, 13. <https://doi.org/10.3389/fpls.2022.1007322>.
- Joshi, R., Pareek, A., & Singla-Pareek, S.-L. (2016). Chapter 9 - Plant Metallothioneins: Classification, distribution, function, and regulation. In *Plant Metal Interaction; Emerging Remediation Techniques* pp. 239-261. <https://doi.org/10.1016/B978-0-12-803158-2.00009-6>.
- Jourdan, C., & Rey, H. (1997). Architecture and development of the oil-palm (*Elaeis guineensis* Jacq.) root system. *Plant and Soil*, 189, 33-48. <https://doi.org/10.1023/A:1004290024473>.
- Jourdan, C., Michaux-Ferrière, M., & Perbal, G. (2000). Root system architecture and gravitropism in the oil palm. *Annals of Botany*, 85(6), 861-868. <https://doi.org/10.1006/anbo.2000.1148>.

- Kagaya, Y., Ohmiya, K., & Hattori, T. (1999). RAV1, a novel DNA-binding protein, binds to bipartite recognition sequence through two distinct DNA-binding domains uniquely found in higher plants. *Nucleic Acids Res.*, 27(2), 470-478. <https://doi.org/10.1093/nar/27.2.470>.
- Kakrana, A., Kumar, A., Satheesh, V., Abdin, M. Z., Subramaniam, K., Bhattacharya, R. C., Srinivasan, R., Sirohi, A., & Jain, P. K. (2017). Identification, validation and utilization of novel nematode-responsive root-specific promoters in *Arabidopsis* for inducing host-delivered RNAi mediated root-knot nematode resistance. *Frontiers in Plant Science*, 8(2049). <https://doi.org/10.3389/fpls.2017.02049>.
- Kaniewski, W. K., & Thomas, P. (2004). The potato story. *AgBioForum*, 7, 41–46.
- Katan, J. (2017). Diseases caused by soilborne pathogens: Biology, management and challenges. *Journal of Plant Pathology*, 99(2), 305-315. <https://doi.org/10.4454/jpp.v99i2.3862>.
- Kavi Kishor, P. B., & Sreenivasulu, N. (2014). Is proline accumulation per se correlated with stress tolerance or is proline homeostasis a more critical issue?. *Plant Cell Environ.*, 37(2), 300-311. <https://doi.org/10.1111/pce.12157>.
- Kavi Kishor, P. B., Hima Kumari, P. H., Sunita, M. S., & Sreenivasulu, N. 2015. Role of proline in cell wall synthesis and plant development and its implications in plant ontogeny. *Front. Plant Sci.*, 6:544. <https://doi.org/10.3389/fpls.2015.00544>.
- Keller, B., & Lamb, C. J. (1989). Specific expression of a novel cell wall hydroxyproline-rich glycoprotein gene in lateral root initiation. *Genes Dev.*, 3, 1639-1646. <https://doi.org/10.1101/gad.3.10.1639>.
- Kersey, P. J. (2019). Plant genome sequences: past, present, future. *Current Opinion in Plant Biology*, 48, 1-8. <https://doi.org/10.1016/j.pbi.2018.11.001>.
- Khandal, H., Gupta, S. K., Dwivedi, V., Mandal, D., Sharma, N. K., Vishwakarma, N. K., Pal, L., Choudhary, M., Francis, A. Malakar, P., Singh, N. P., Sharma, K., Sinharoy, S., Singh, N. P., Sharma, R., & Chattopadhyay, D. (2020). Root-specific expression of chickpea cytokinin oxidase/dehydrogenase 6 leads to enhanced root growth, drought tolerance and yield without compromising nodulation. *Plant Biotechnol. J.*, 18(11), 2225-2240. <https://doi.org/10.1111/pbi.13378>.
- Kim, D. W., Lee, S. H., Choi, S. B., Won, S. K., Heo, Y. K., Cho, M., Park, Y. I., & Cho, H. T. (2006). Functional conservation of a root hair cell-specific cis-element in angiosperms with different root hair distribution patterns. *Plant Cell*, 18(11), 2958-2970. <https://doi.org/10.1105/tpc.106.045229>.

- Kim, H. J., Kim, Y. K., Park, J. Y., & Kim, J. (2002). Light signalling mediated by phytochrome plays an important role in cold-induced gene expression through the C-repeat/dehydration responsive element (C/DRE) in *Arabidopsis thaliana*. *Plant J.*, 29(6), 693-704. <https://doi.org/10.1046/j.1365-313x.2002.01249.x>.
- Kısa, D., Öztürk, L., Doker, S., & Gökçe, İ. (2017). Expression analysis of metallothioneins and mineral contents in tomato (*Lycopersicon esculentum*) under heavy metal stress. *Journal of the Science of Food and Agriculture*, 97(6), 1916-1923. <https://doi.org/10.1002/jsfa.7995>.
- Kong, K., Makabe, S., Ntui, V. O., Khan, R. S., & Nakamura, I. (2014). Synthetic chitinase gene driven by root-specific *LjNRT2* and *AtNRT2.1* promoters confers resistance to *Fusarium oxysporum* in transgenic tobacco and tomato. *Plant Biotechnology Reports*, 8(2), 151-159. <https://doi.org/10.1007/s11816-013-0303-2>.
- Kościelniak, P., Glazińska, P., Kęsy, J., & Zadworny, M. (2021). Formation and development of taproots in deciduous tree species. *Frontiers in Plant Science*, 12. <https://doi.org/10.3389/fpls.2021.772567>.
- Koul, B. (2022). Cisgenics and crop improvement. In B. Koul (Ed.), *Cisgenics and Transgenics*, (pp. 107–129). Springer; Singapore. https://doi.org/10.1007/978-1981-1019-2119-1003_1003.
- Koyama, T., Ono, T., Shimizu, M., Jinbo, T., Mizuno, R., Tomita, K., Mitsukawa, N., Kawazu, T., Kimura, T., Ohmiya, K., & Sakka, K. (2005). Promoter of *Arabidopsis thaliana* phosphate transporter gene drives root-specific expression of transgene in rice. *J Biosci. Bioeng.*, 9 (1), 38-42. <https://doi.org/10.1263/jbb.99.38>.
- Kumar, S., Kumar, S., & Mohapatra, T. (2021). Interaction between macro- and micro-nutrients in plants. *Frontiers in Plant Science*, 12, 665583. <https://doi.org/10.3389/fpls.2021.665583>.
- Lacombe, E., Van Doorselaere, J., Boerjan, W., Boudet, A. M. and Grima-Pettenati, J. (2000). Characterization of cis-elements required for vascular expression of the *cinnamoyl CoA reductase* gene and for protein-DNA complex formation. *Plant J.*, 23(5), 663-676. <https://doi.org/10.1046/j.1365-313x.2000.00838.x>.
- Lagrange, T., Franzetti, B., Axelos, M., Mache, R. and Lerbs-Mache, S. 1993. Structure and expression of the nuclear gene coding for the chloroplast ribosomal protein L21: developmental regulation of a housekeeping gene by alternative promoters. *Mol Cell Biol*, 13(4), 2614-2622. <https://doi.org/10.1128/mcb.13.4.2614-2622.1993>.
- Lang, Z., Zhou, P., Yu, J., Ao, G., & Zhao, Q. 2008. Functional characterization of the pollen-specific *SBgLR* promoter from potato (*Solanum tuberosum* L.). *Planta*, 227, 387–396. <https://doi.org/10.1007/s00425-007-0625-9>.

- Laurell, H., Iacovoni, J. S., Abot, A., Svec, D., Maoret, J. J., Arnal, J. F., & Kubista, M. (2012). Correction of RT-qPCR data for genomic DNA-derived signals with ValidPrime. *Nucleic Acids Res.*, 40(7), e51. <https://doi.org/10.1093/nar/gkr1259>.
- Le, N. Q., Yapp, E. K., Nagasundaram, N., & Yeh, H. (2019). Classifying promoters by interpreting the hidden information of DNA sequences via deep learning and combination of continuous FastText N-Grams. *Frontiers in Bioengineering and Biotechnology*, 7. <https://doi.org/10.3389/fbioe.2019.00305>.
- Leach, F., & Aoyagi, K. (1991). Promoter analysis of the highly expressed *rolC* and *rolD* root-inducing genes of *Agrobacterium rhizogenes*: enhancer and tissue-specific DNA determinants are dissociated. *Plant Sci.*, 207, 37-44. <https://doi.org/10.1016/j.plantsci.2013.02.002>.
- Ledger, S. E., & Gardner, R. C. (1994). Cloning and characterization of five cDNAs for genes differentially expressed during fruit development of kiwifruit (*Actinidia deliciosa* var. *deliciosa*). *Plant Molecular Biology*, 25(5), 877-886. <https://doi.org/10.1007/BF00028882>.
- Lee, H-H., Kim, J-S., Hoang, Q.T.N., Kim, J-I., & Kim, Y.S. (2018). Root-specific expression of defensin in transgenic tobacco results in enhanced resistance against *Phytophthora parasitica* var. *nicotianae*. *European Journal of Plant Pathology*, 151(3), 811-823. <https://doi.org/10.1007/s10658-018-1419-6>.
- Lehmann, S., Funck, D., Szabados, L., & Rentsch, D. (2010). Proline metabolism and transport in plant development. *Amino Acids*, 39(4), 949-962. <https://doi.org/10.1007/s00726-010-0525-3>.
- Leifeld, J. (2013). Low-input arming: A way towards climate-friendly agriculture?. *Carbon Management*, 4(1), 31-41. <https://doi.org/10.4155/cmt.12.70>.
- Lescot, M., Déhais, P., Thijs, G., Marchal, K., Moreau, Y., Van de Peer, Y., Rouzé, P., & Rombauts, S. (2002). PlantCARE, a database of plant cis-acting regulatory elements and a portal to tools for in silico analysis of promoter sequences. *Nucleic Acids Research*, 30(1), 325-327. <https://doi.org/10.1093/nar/30.1.325>.
- Li X, Zhao, J., Tan, Z, Zeng, R., & Liao, H. (2015). *GmEXPB2*, a cell wall beta-expansin, affects soybean nodulation through modifying root architecture and promoting nodule formation and development. *Plant Physiology*, 169, 2640–2653. <https://doi.org/10.1104/pp.15.01029>.
- Li Y., Liu, S., Yu Z., Liu Y., & Wu, P. (2013). Isolation and characterization of two novel root-specific promoters in rice (*Oryza sativa* L.). *Plant Science*, 207, 37-44. <https://doi.org/10.1016/j.plantsci.2013.02.002>.

- Li, J., Brader, G., Kariola, T., & Palva, E.T. (2006). WRKY70 modulates the selection of signaling pathways in plant defense. *Plant J.*, 46(3), 477-491. <https://doi.org/10.1111/j.1365-313X.2006.02712.x>.
- Li, J., Xu, R-F., Qin, R-Y., Ma, H., Li, H., Zhang, Y-P., Li, L. Wei, P-C., & Yang J-B. (2014). Isolation and functional characterization of a novel rice constitutive promoter. *Plant Cell Rep.*, 33(10), 1651–1660. <https://doi.org/10.1007/s00299-014-1644-1>.
- Li, Y., Fu, X., Zhao, M., Zhang, W., Li, B., An, D., Li, J., Zhang, A. Liu, R., & Liu, X. (2018). A genome-wide view of transcriptome dynamics during early spike development in bread wheat. *Scientific Reports*, 8(1), 15338-15338. <https://doi.org/10.1038/s41598-018-33718-y>.
- Li, Y., Liu, X., Chen, R., Tian, J., Fan, Y., & Zhou, X. (2019). Genome-scale mining of root-preferential genes from maize and characterization of their promoter activity. *BMC Plant Biology*, 19, 584. <https://doi.org/10.1186/s12870-019-2198-8>.
- Li, Z., Yang, J., Peng, J., Cheng, Z., Liu, X., Zhang, Z., Bhadauria, V., Zhao, W., & Peng, Y. (2021). Transcriptional landscapes of long non-coding RNAs and alternative splicing in *Pyricularia oryzae* revealed by RNA-Seq. *Frontiers in Plant Science*, 12, 723636. <https://doi.org/10.3389/fpls.2021.723636>.
- Li, Z. T., Kim, K-H., Jasinski, J. R., Creech, M. R., & Gray, D. J. (2012). Large-scale characterization of promoters from grapevine (*Vitis spp.*) using quantitative anthocyanin and GUS assay systems. *Plant Science*, 196, 132-142. <https://doi.org/10.1016/j.plantsci.2012.08.009>.
- Lia, Y., Liua, S., Yuc, Z., Liua, Y., & Wua, P. (2013). Isolation and characterization of two novel root-specific promoters in rice (*Oryza sativa* L.). *Plant Science*, 207, 37-44. <https://doi.org/10.1016/j.plantsci.2013.02.002>.
- Liang, P., & Pardee, A.B. (1992). Differential display of eukaryotic messenger RNA by means of the polymerase chain reaction. *Science*, 257, 967–971. <https://doi.org/10.1126/science.1354393>.
- Liang, P., Averboux, L., & Pardee, A. B. (1993). Distribution and cloning of eukaryotic mRNAs by means of differential display: Refinements and optimization. *Nucleic Acids Res*, 21, 3269–3275. <https://doi.org/10.1093/nar/21.14.3269>.
- Liao, Y-L., Shen, Y-B., Chang, J., Zhang, W-W., Cheng, S.-Y., & Xu, F. (2015). Isolation, expression, and promoter analysis of *GbWRKY2*: A novel transcription factor gene from *Ginkgo biloba*. *International Journal of Genomics*, 607185. <https://doi.org/10.1155/2015/607185>.
- Lim, F.-H., Rasid, O. A., Idris, A. S., As'wad, A. W. M., Vadamalai, G., Parveez, G. K. A., & Wong, M.-Y. (2023). Induced expression of *Ganoderma*

- boninense* Lanosterol 14 α -Demethylase (*ERG11*) during interaction with oil palm. *Molecular Biology Reports*, 50(3), 2367-2379. <https://doi.org/10.1007/s11033-022-08131-4>.
- Lisitsyn, N., Lisitsyn, N., & Wigler, M. (1993). Cloning the differences between two complex genomes. *Science*, 259, 5640–5648. <https://doi.org/10.1126/science.8438152>.
- Liu, F., Zheng, K., Chen, H. C., & Liu, Z.F. (2018). Capping-RACE: a simple, accurate, and sensitive 5' RACE method for use in prokaryotes. *Nucleic Acids Res.*, 46(21), e129. <https://doi.org/10.1093/nar/gky739>.
- Liu, J.-J., & Ekramoddoullah, A. (2003). Root-specific expression of a western white pine PR10 gene is mediated by different promoter regions in transgenic tobacco. *Plant Molecular Biology*, 52, 103-120. <https://doi.org/10.1023/A:1023930326839>.
- Liu, M., Zhao, J., Wang, J., Liu, Z., & Liu, G. (2017). Phylogenetic analysis of 25 plant species representing 19 angiosperm families and one gymnosperm family based on 390 orthologous genes. *Plant Systematics and Evolution*, 303(3), 413-417. <https://doi.org/10.1007/s00606-016-1380-9>.
- Liu, Q., Ding, C., Lang, X., Guo, G., Chen, J., & Su, X. (2019). Small noncoding RNA discovery and profiling with sRNA tools based on high-throughput sequencing. *Briefings in Bioinformatics*, 22(1), 463-473. <https://doi.org/10.1093/bib/bbz151>.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2^{- $\Delta\Delta C_T$} Method. *Methods*, 25(4), 402-408. <https://doi.org/10.1006/meth.2001.1262>.
- Longsaward, R., Pengnoo, A., Kongsawadworakul, P., & Viboonjun, U. (2023). A novel rubber tree PR-10 protein involved in host-defense response against the white root rot fungus *Rigidoporus microporus*. *BMC Plant Biology*, 23(1), 157. <https://doi.org/10.1186/s12870-023-04149-3>.
- Loomis, W. D. (1974). Overcoming problems of phenolics and quinones in the isolation of plant enzymes and organelles. *Meth. Enzymol.*, 31, 528–545. [https://doi.org/10.1016/0076-6879\(74\)31057-9](https://doi.org/10.1016/0076-6879(74)31057-9).
- Low, E.T.L., Jayanthi, N. Chan, K.L., Sanusi, N.S.N.M., Halim.,M.A.A., Rosli, R., Azizi,N., Amiruddin, N., Angel, L.P.L., Ong-Abdullah, M., Singh, R., Mohamad Arif A.A.,I Sambanthamurthi, R., Parveez, G.K.A., & Kushairi, A. (2017). The oil palm genome revolution. *J Oil Palm Res.*, 29(4), 456-468. <https://doi.org/10.21894/jopr.2017.00018>.
- Low, E. T. L., Singh, R., Rajanaidu, N., Ong-Abdullah, M., Ooi, L. C. L., Lakey, N. D., Smith, S. W., Ordway, J. M., & Sambathamurthi, R. (2016). New frontiers for the oil palm industry through genome technology. *The*

- Planter*, Kuala Lumpur 92, (1087), 701-710. <https://doi.org/10.56333/tp.2016.009>.
- Lowe, R., Shirley, N., Bleackley, M., Dolan, S., & Shafee, T. (2017). Transcriptomics technologies. *PLOS Computational Biology*, 13(5), e1005457. <https://doi.org/10.1371/journal.pcbi.1005457>.
- Lu, G., & Moriyama, E. N. (2004). Vector NTI, a balanced all-in-one sequence analysis suite. *Briefings in Bioinformatics*, 5(4), 378-388. <https://doi.org/10.1093/bib/5.4.378>.
- Lü, S., Gu, H., Yuan, X., Wang, X., Wu, A., Qu, L., & Liu, J. (2007). The GUS reporter-aided analysis of the promoter activities of a rice metallothionein gene reveals different regulatory regions responsible for tissue-specific and inducible expression in transgenic *Arabidopsis*. *Transgenic Res.*, 16, 177–119. <https://doi.org/10.1007/s11248-006-9035-1>.
- Luo, B., Xu, M., Zhao, L., Xie, P., Chen, Y., Harwood, W., Xu, G., Fan, X., & Miller, A.J. (2020). Overexpression of the high-affinity nitrate transporter *OsNRT2.3b* driven by different promoters in barley improves yield and nutrient uptake balance. *International Journal of Molecular Sciences*, 21(4), 1320. <https://doi.org/10.3390/ijms21041320>.
- Ma, W. Y., Li, J. J., Qu, B. Y., He, X., Zhao, X. Q., Li, B., Fu, X. D., & Tong, Y. P. (2014). Auxin biosynthetic gene *TAR2* is involved in low nitrogen-mediated reprogramming of root architecture in *Arabidopsis*. *The Plant Journal*, 78, 70–79. <https://doi.org/10.1111/tpj.12448>.
- Ma, Y., Xue, M., Zhang, X., & Chen, S. (2023). Genome-wide analysis of the metallothionein gene family in cassava reveals its role in response to physiological stress through the regulation of reactive oxygen species. *BMC Plant Biol.*, 23(1), 227. <https://doi.org/10.1186/s12870-023-04174-2>.
- MacKenzie, M., & Argyropoulos, C. (2023). An Introduction to nanopore sequencing: past, present, and future considerations. *Micromachines*, 14, 459. <https://doi.org/10.3390/mi14020459>.
- Manaf, M. A. A., Izawati, A. M. D., Zubaidah, R., Masani, A. M. Y., Safiza, M., Lim, F. H., Nurniwalis, A. W., Omar, O. A., & Parveez, G. K. A. (2017). Biotechnology for diversification and improved resilience of the oil palm. *The Planter*, Kuala Lumpur, 93(1093), 237-249. <https://doi.org/10.56333/tp.2017.006>.
- Martorella, M., Kasela, S., Garcia-Flores R., Gokden, A., Castel, S.E., & Lappalainen, T. (2022). Non invasive, low-cost RNA-sequencing enhances discovery potential of transcriptome studies. *bioRxiv*. <https://doi.org/10.1101/2022.1109.1106.506813>.
- Maruyama-Nakashita, A., Nakamura, Y., Watanabe-Takahashi, A., Inoue, E., Yamaya, T., & Takahashi, H. (2005). Identification of a novel cis-acting

- element conferring sulfur deficiency response in *Arabidopsis* roots. *Plant J.* 42(3), 305-314. <https://doi.org/10.1111/j.1365-313X.2005.02363.x>.
- Masani, M. Y. A., Izawati, A. M. D., Rasid, O. A., & Parveez, G. K. A. (2018). Biotechnology of oil palm: Current status of oil palm genetic transformation. *Biocatal. Agric. Biotechnol.*, 15, 335-347. <https://doi.org/10.1016/j.bcab.2018.07.008>.
- Masura, S. S., Parveez, G. K. A., & Ismanizan, I. (2010). Isolation and characterization of oil palm constitutive promoter derived from ubiquitin extension protein (*uep1*) gene. *New Biotechnology*, 27, 289-299. <https://doi.org/10.1016/j.nbt.2010.01.337>.
- Masura, S. S., Rasid, O. A., Shaharuddin, N. A., Masani, A. M. Y., Chan, K. L., Low, E. T. L., Badai, S. S., Hanin, A. N., Rahmah, A. R. S., Abdullah, M. P., Azzeme, A. M., & Parveez, G. K. A. (2022). Identification of oil palm root-specific genes through mining of RNA-Seq data and RT-qPCR analysis. *J Oil Palm Res.*, 34 (2), 261-275. <https://doi.org/210.21894/jopr.22021.20036>.
- Masura, S. S., Parveez, G. K. A., & Low, E. T. L. (2011). Isolation and characterization of an oil palm constitutive promoter from a translationally control tumor protein (TCTP) gene. *Plant Physiology and Biochemistry*, 40, 701-708. <https://doi.org/10.1016/j.plaphy.2011.04.003>.
- Masura, S. S., Parveez, G. K. A., & Rasid, O. A. (2019). Isolation and characterisation of an oil palm constitutive promoter derived from ubiquitin extension protein (*uep2*) gene. *J Oil Palm Res.*, 31(1), 28-41. <https://doi.org/10.21894/jopr.2019.0001>.
- Masura, S. S., Tahir, N. I., Rasid, O. A., Ramli, U. S., Othman, A., Masani, M. Y. A., Parveez, G. K. A., & Kushairi, A. (2017). Post-genomic technologies for the advancement of oil palm research. *J Oil Palm Res.*, 29(4), 469-486. <https://doi.org/10.21894/jopr.2017.00013>.
- Matsumura, I. (2015). Why Johnny can't clone: Common pitfalls and not so common solutions. *Biotechniques*, 59(3), iv-xiii. <https://doi.org/10.2144/000114324>.
- Mattoo, A. K., Murata, T., Pantastico, E. B., Chachin, K., Ogata, K., & Phan, T. C. (1975). Chemicals changes during ripening and senescence. In E. B. Pantastico (Ed), *Post-harvest physiology, handling and utilization of tropical and subtropical fruit and vegetables* (pp. 103-127). AVI, Westport, Conn.
- McElrone, A. J., Choat, B., Gambetta, G. A., & Brodersen, C. R. (2013). Water uptake and transport in vascular plants. *Nature Education Knowledge*, 4(5), 6.

- Meister, R., Rajani, M. S., Ruzicka, D., & Schachtman, D. P. (2014). Challenges of modifying root traits in crops for agriculture. *Trends in Plant Science*, 19, 779–788. <https://doi.org/10.1016/j.tplants.2014.08.005>.
- Mena, M., Cejudo, F. J., Isabel-Lamonedá, I., & Carbonero, P. (2002). A role for the DOF transcription factor BPBF in the regulation of gibberellin-responsive genes in barley aleurone. *Plant Physiol.*, 130(1), 111-119. <https://doi.org/10.1104/pp.005561>.
- Mhimdi, M., & Manuel, J. (2020). Understanding of adventitious root formation: What can we learn from comparative genetics?. *Frontiers in Plant Science*, 11. <https://doi.org/10.3389/fpls.2020.582020>.
- Mohan, C., Jayanarayanan, A. N. & Narayanan, S. (2017). Construction of a novel synthetic root-specific promoter and its characterization in transgenic tobacco plants. *3 Biotech.*, 7(4), 234. <https://doi.org/10.1007/s13205-017-0872-9>.
- Mollier, P., Hoffmann, B., Orsel, M., & Pelletier, G. (2000). Tagging of a cryptic promoter that confers root-specific gus expression in *Arabidopsis thaliana*. *Plant Cell Reports*, 19(11), 1076-1083. <https://doi.org/10.1007/s002990000241>.
- Moon, H., & Callahan, A. M. (2004). Developmental regulation of peach ACC oxidase promoter-GUS fusions in transgenic tomato fruits. *J Exp. Bot.*, 55, 1519–1528. <https://doi.org/10.1093/jxb/erh162>.
- Morgan, J. B., & Connolly, E. L. (2013). Plant-soil interactions: nutrient uptake. *Nat Educ. Knowl.*, 4(8), 2.
- Morioka, M. S., Kawaji, H., Nishiyori-Sueki, H., Murata, M., Kojima-Ishiyama, M., Carninci, P., & Itoh, M. (2020). Cap Analysis of Gene Expression (CAGE): A quantitative and genome-wide assay of transcription start sites. *Methods, Mol. Biol.*, 2120, 277-301. https://doi.org/10.1007/978-1-0716-0327-7_20.
- Morita, A., Umemura, T., Kuroyanagi, M., Futsuhara, Y., Perata, P., & Yamaguchi, J. (1998). Functional dissection of a sugar-repressed alpha-amylase gene (*RAmy1 A*) promoter in rice embryos. *FEBS Lett.*, 423(1), 81-85. [https://doi.org/10.1016/s0014-5793\(98\)00067-2](https://doi.org/10.1016/s0014-5793(98)00067-2).
- Morris, P. E., Chan, P. L., Cheng-Li, L.O., Ong, P. W., Kamaruddin, K., Marjuni, M., Amiruddin, M. D., Chan, K. L., Rosli, R., Low, E. T. L., Singh, R., Shaharuddin, N. A., Murphy, M., Manaf, M. A. A., & Siti Nor Akmar, A. (2020). Transcriptome analysis across various mesocarp developmental stages of MPOB-Angola *dura*. *J Oil Palm Res.*, 32(3), 406-419. <https://doi.org/10.21894/jopr.2020.0034>.
- Mudge, S. R., Smith, F. W., & Richardson, A. E. (2003). Root-specific and phosphate-regulated expression of phytase under the control of a phosphate transporter promoter enables *Arabidopsis* to grow on phytate

- as a sole P source. *Plant Science*, 165(4), 871-878. [https://doi.org/10.1016/S0168-9452\(03\)00286-3](https://doi.org/10.1016/S0168-9452(03)00286-3).
- Naher, U. A., Othman, R., & Panhwar, Q. A. (2013). Beneficial effects of mycorrhizal association for crop production in the tropics: A review. *Journal of Agriculture and Biology International*, 15(5), 1021-1028.
- Nan, L., Lin, H., Guan, Y., & Chen F. (2002). Functional analysis of cis-acting sequences regulating root-specific expression in transgenic tobacco. *Chin. Sci. Bull.*, 47, 1441-1445. <https://doi.org/10.1360/02tb9318>.
- Ng, S. K., von Uexkull, H., & Rolf, H. (2003). Botanical aspects of the oil palm relevant to crop management. In T. Fairhurst and R. Hardter (Eds), *Oil Palm; Management for large and sustainable yields* (pp. 14-15). Switzerland: Potash and Phosphate Institute of Canada (PPIC) and International Potash Institute (IPI), Basel. ISBN: 9810484852.
- Nibau, C., Gibbs, D.J., & Coates, J. C. (2008). Branching out in new directions: the control of root architecture by lateral root formation. *New Phytologist*, 179(3), 595–614. <https://doi.org/10.1111/j.1469-8137.2008.02472.x>.
- Nishiuchi, T., Shinshi, H., & Suzuki, K. (2004). Rapid and transient activation of transcription of the *ERF3* gene by wounding in tobacco leaves: possible involvement of *NtWRKYs* and autorepression. *J Biol. Chem.*, 279(53), 55355-55361. <https://doi.org/10.1074/jbc.M409674200>.
- Nitz, I., Berkefeld, H., Puzio, P.S., & Grundler, F. M. V. (2001). *Pyk10*, a seedling and root specific gene and promoter from *Arabidopsis thaliana*. *Plant Sci.*, 161, 337-346. [https://doi.org/10.1016/s0168-9452\(01\)00412-5](https://doi.org/10.1016/s0168-9452(01)00412-5).
- Noh, S. A., Lee, H. S., Huh, G. H., Oh, M. J., Paek, K. H., Shin, J. S., & Bae, J. M. (2012). A sweet potato *SRD1* promoter confers strong root-, taproot-, and tuber-specific expression in *Arabidopsis*, carrot and potato. *Transgenic Res.*, 21, 265-278. <https://doi.org/10.1007/s11248-011-9528-4>.
- Notka, F., Liss, M., & Wagner, R. (2011). Industrial scale gene synthesis. *Methods in Enzymology*, 498, 247-275. <https://doi.org/10.1016/B978-0-12-385120-8.00011-5>.
- Novick, A., & Weiner, M. (1957). Enzyme Induction as an All-or-None Phenomenon. *Proc. Natl. Acad. Sci. USA*, 43, 553–566. <https://doi.org/10.1073/pnas.43.7.553>.
- Ogawa, M., Hanada, A., Yamauchi, Y., Kuwahara, A., Kamiya, Y., & Yamaguchi, S. (2003). Gibberellin biosynthesis and response during *Arabidopsis* seed germination. *Plant Cell*, 15(7), 1591-1604. <https://doi.org/10.1105/tpc.011650>.
- Ohgishi, M., Oka, A., Morelli, G., Ruberti, I., & Aoyama, T. (2001). Negative autoregulation of the *Arabidopsis* homeobox gene *ATHB-2*. *Plant J.*, 25(4), 389-398. <https://doi.org/10.1046/j.1365-313x.2001.00966.x>.

- Oikonomopoulos, S., Bayega, A., Fahiminiya, S., Djambazian, H., Berube, P., & Ragoussis, J. (2020). Methodologies for transcript profiling using long-read technologies. *Frontiers in Genetics*, 11, 525110. <https://doi.org/10.3389/fgene.2020.00606>.
- O'Leary, N.A., Wright, M.W., Brister, J.R., Ciufu, S., Haddad, D., McVeigh, R., Robbertse, B., Smith-White, B., Ako-Adjei, D., Astashyn, A., Badretdin, A., Bao, Y., Blinkova, O., Brover, V., Chetvernin, V., Choi, J., Cox, E., Ermolaeva, O., Farrell, C. M., Goldfarb, T., Gupta, T., Haft, D., Hatcher, E., Hlavina, W., Joardar, V. S., Kodali, V. K., Li, W., Maglott, D., Masterson, P., McGarvey, K.M., Murphy, M.R., O'Neill, K., Pujar, S., Rangwala, S.H., Rausch, D., Riddick, L.D., Schoch, C., Shkeda, A., Storz, S.S., Sun, H., Thibaud-Nissen, F., Tolstoy, I., Tully, R.E., Vatsan, A.R., Wallin, C., Webb, D., Wu, W., Landrum, M.J., Kimchi, A., Tatusova, T., DiCuccio, M., Kitts, P., Murphy, T.D., & Pruitt, K.D. (2016). Reference sequence (RefSeq) database at NCBI: current status, taxonomic expansion, and functional annotation. *Nucleic Acids Res.*, 44(D1), D733-745. <https://doi.org/10.1093/nar/gkv1189>.
- Onyango, S. O., Roderick, H., Tripathi, J. N., Collins, R., Atkinson, H. J., Richard O. Oduor, R.O., & Tripathi., L. (2016). The *ZmRCP-1* promoter of maize provides root tip specific expression of transgenes in plantain. *J Biol. Res-Thessaloniki*, 23, 4. <https://doi.org/10.1186/s40709-016-0041-z>.
- Pahalvi, H. N., Rafiya, L., Rashid, S., Nisar, B., & Kamili, A. N. (2021). Chemical fertilizers and their impact on soil health. In *Microbiota and Biofertilizers*. https://doi.org/10.1007/978-3-030-61010-4_1, Vol 2.
- Panhwar, Q. A., Ali, A., Naher, U. A., & Memon, M. Y. (2019). Chapter 2 - Fertilizer management strategies for enhancing nutrient use efficiency and sustainable wheat production. In S. Chandran, M.R. Unni, & S. Thomas (Eds.), *Organic Farming* (pp. 17-39). Woodhead Publishing. <https://doi.org/10.1016/B978-0-12-813272-2.00002-1>.
- Panth, M., Hassler, S. C., & Baysal-Gurel, F. (2020). Methods for management of soilborne diseases in crop production. *Agriculture*, 10(1), 16.
- Park, H. C., Kim, M. L., Kang, Y. H., Jeon, J. M., Yoo, J. H., Kim, M. C., Park, C. Y., Jeong, J. C., Moon, B. C., Lee, J. H., Yoon, H. W., Lee, S. H., Chung, W. S., Lim, C. O., Lee, S. Y., Hong, J. C., & Cho, M. J. (2004). Pathogen- and NaCl-induced expression of the S_{CaM}-4 promoter is mediated in part by a GT-1 box that interacts with a GT-1-like transcription factor. *Plant Physiol.*, 135(4), 2150-2161. <https://doi.org/10.1104/pp.104.041442>.
- Parveez, G. K. A. (1998). *Optimization of parameters involved in transformation of oil palm using the biolistic method*, PhD Thesis, Universiti Putra Malaysia.

- Parveez, G. K. A., Rasid, O. A., Masani, A. M. Y., & Sambanthamurthi, R. (2015). Biotechnology of oil palm: strategies towards manipulation of lipid content and composition. *Plant Cell Rep.*, 34, 533-543.
- Patankar, H. V., Al-Harrasi, I., Al Kharusi, L., Jana, G. A., Al-Yahyai, R., Sunkar, R., & Yaish, M. W. (2019). Overexpression of a *Metallothionein 2A* gene from date palm confers abiotic stress tolerance to yeast and *Arabidopsis thaliana*. *Int. J. Mol. Sci.*, 20(12), 2871. <https://doi.org/10.3390/ijms20122871>.
- Pauli, S., Rothnie, H. M., Chen, G., He, X., & Hohn, T. (2004). The cauliflower mosaic virus 35S promoter extends into the transcribed region. *J Virol.*, 78(22), 12120-12128. <https://doi.org/10.1128/jvi.78.22.12120-12128.2004>.
- Pearce, G., Siems, W. F., Bhattacharya, R., Chen, Y-C., & Ryan, C. A. (2007). Three hydroxyproline-rich glycopeptides derived from a single petunia polyprotein precursor activate *defensin I*, a pathogen defense response gene. *Journal of Biological Chemistry*, 282(24), 17777-17784. <https://doi.org/10.1074/jbc.M701543200>.
- Pervez, M. T., Hasnain, M. J. U., Abbas, S. H., Moustafa, M. F., Aslam, N., & Shah, S. S. M. (2022). A comprehensive review of performance of next-generation sequencing platforms. *BioMed Research International*, 3457806. <https://doi.org/10.1155/2022/3457806>.
- Plesch, G., Ehrhardt, T., & Mueller-Roeber, B. (2001). Involvement of TAAAG elements suggests a role for Dof transcription factors in guard cell-specific gene expression. *Plant J.*, 28(4), 455-464. <https://doi.org/10.1046/j.1365-313x.2001.01166.x>.
- Potenza, C., Aleman, L., & Sengupta-Gopalan, C. (2004). Targeting transgene expression in research, agricultural and environmental applications: Promoters used in plant transformation. *In Vitro Cell Dev. Biol. Plant*, 40, 1-22. <https://doi.org/10.1079/IVP2003477>.
- Pruitt, K. D., Tatusova, T., & Maglott, D. R. (2005). NCBI Reference Sequence (RefSeq): a curated non-redundant sequence database of genomes, transcripts and proteins. *Nucleic Acids Research* 33(Database issue), D501-D504. <https://doi.org/10.1093/nar/gki025>.
- Qaderi, M. M., Martel, A. B., & Dixon, S. L. (2019). Environmental factors influence plant vascular system and water regulation. *Plants (Basel)*, 8(3), <https://doi.org/10.3390/plants8030065>.
- Qu, B., He, X., Wang, J., Zhao, Y., Teng, W., Shao, A., Zhao, X., Ma, W., Wang, J., Li, B., Li, Z., & Tong, Y. (2015). A wheat CCAAT box binding transcription factor increases the grain yield of wheat with less fertilizer input. *Plant Physiology*, 167, 411-423. <https://doi.org/10.1104/pp.114.246959>.

- Quilez, J., Guilmatre, A., Garg, P., Highnam, G., Gymrek, M., Erlich, Y., Joshi, R. S., Mittelman, D., & Sharp, A. J. (2016). Polymorphic tandem repeats within gene promoters act as modifiers of gene expression and DNA methylation in humans. *Nucleic Acids Res.*, 44(8), 3750-3762. <https://doi.org/10.1093/nar/gkw219>.
- Quinn, J. M., & Merchant, S. (1995). Two copper-responsive elements associated with the *Chlamydomonas* *Cyc6* gene function as targets for transcriptional activators. *Plant Cell*, 7(5), 623-628. <https://doi.org/10.1105/tpc.7.5.623>.
- Rahman, A. K. A., Abdullah, R., Balu, N., & Shariff, F. M. (2013). The impact of La Niña and El Niño events on crude palm oil prices: An econometric analysis. *Oil Palm Ind. Econ. J.*, (13), 38–51.
- Raj, A., & van Oudenaarden, A. (2008). Nature, nurture, or chance: stochastic gene expression and its consequences. *Cell*, 135(2), 216-226. <https://doi.org/10.1016/j.cell.2008.09.050>.
- Rajasheker, G., Nagaraju, M., Varghese, R. P., Jalaja, N., Somanaboina, A. K., Singam, P., Ramakrishna, C., Penna, S., Sreenivasulu, N., & Kishor, P. B. (2022). Identification and analysis of proline-rich proteins and hybrid proline-rich proteins super family genes from *Sorghum bicolor* and their expression patterns to abiotic stress and zinc stimuli. *Frontiers in Plant Science*, 13, 952732. <https://doi.org/10.3389/fpls.2022.952732>.
- Ramireddy, E., Hosseini, S. A., Eggert, K., Gillandt, S., Gnad, H., von Wirén, N., & Schmölling, T. (2018). Root engineering in barley: Increasing cytokinin degradation produces a larger root system, mineral enrichment in the shoot and improved drought tolerance. *Plant Physiology*, 177(3), 1078-1095. <https://doi.org/10.1104/pp.18.00199>.
- Rani, B., & Sharma, V. K. (2017). Transcriptome profiling: Methods and applications- A review. *Agricultural Reviews*, 38(4), 271-281. <https://doi.org/10.18805/ag.R-1549>.
- Rao, X., Huang, X., Zhou, Z., & Lin, X. (2013). An improvement of the 2^{-ΔΔCT} method for quantitative real-time polymerase chain reaction data analysis. *Biostat. Bioinforma. Biomath.*, 3(3), 71-85.
- Rashidi, B., Mehrabi, S., Demchenko, K., & Pawlowski, K. (2011). The *Casuarina glauca* metallothionein I promoter in nodulated transgenic hairy roots of the actinorhizal plant *Datisca glomerata*. *Functional Plant Biology*, 38, 728–737. <https://doi.org/10.1071/FP10216>.
- Rasid, O. A., Masura, S. S., Hanin, A. N., Izawati, A. M. D., Bohari, B., Masani, M. Y. A. & Parveez, G. K. A. (2020). Oil palm transgenic research: Challenges, update and future outlook. In M. Ithnin, and A. Kushairi (Eds.) *The oil palm genome. Compendium of plant genome*, (pp. 69-81).

Springer, Cham. https://doi.org/10.1007/1978-1003-1030-22549-22540_22546.

- Raven, P. H., Evert, R.F., & Eichorn, S.E. (1992). *Biology of Plants*. New York: Worth.
- Redillas, M. C., Jeong, J. S., Kim, Y. S., Jung, H., Bang, S. W., Choi, Ha, S. H., Reuzeau, C., & Kim, J. K. (2012). The overexpression of OsNAC9 alters the root architecture of rice plants enhancing drought resistance and grain yield under field conditions. *Plant Biotechnol. J.*, 10, 792–805. <https://doi.org/10.1111/j.1467-7652.2012.00697.x>.
- Rieping, M., & Schöffl, F. (1992). Synergistic effect of upstream sequences, CCAAT box elements, and HSE sequences for enhanced expression of chimaeric heat shock genes in transgenic tobacco. *Mol. Gen. Genet.*, 231(2), 226-232. <https://doi.org/10.1007/bf00279795>.
- Rival, A. (2017). Breeding the oil palm (*Elaeis guineensis* Jacq.) for climate change. *Oilseeds & Fats Crops and Lipids*, 24, D107. <https://doi.org/10.1051/ocl/2017001>.
- Rushton, P. J., Reinstädler, A., Lipka, V., Lippok, B., & Somssich, I. E. (2002). Synthetic plant promoters containing defined regulatory elements provide novel insights into pathogen- and wound-induced signaling. *Plant Cell*, 14(4), 749-762. <https://doi.org/10.1105/tpc.010412>.
- Rushton, P. J., Torres, J. T., Parniske, M., Wernert, P., Hahlbrock, K., & Somssich, I. E. (1996). Interaction of elicitor-induced DNA-binding proteins with elicitor response elements in the promoters of parsley *PR1* genes. *Embo J.* 15(20), 5690-5700.
- Saba-Mayoral, A., Bassie, L., Christou, P., & Capell, T. (2022). Development of a facile genetic transformation system for the Spanish elite rice paella genotype Bomba. *Transgenic Research*, 31(3), 325-340. <https://doi.org/10.1007/s11248-022-00303-z>.
- Safitri, L., Suryanti, S., Kautsar, V., Kurniawan, A., & Santiabudi, F. (2018). Study of oil palm root architecture with variation of crop stage and soil type vulnerable to drought. *IOP Conf. Series: Earth and Environmental Science*, 141, 012031. <https://doi.org/10.1088/1755-1315/1141/1081/012031>.
- Safiza, M., Anita, J., Mohamad Arif, A. M., & Fazliza, M. A. (2015). Differentially expressed genes in *G. boninense* naturally-infected palms. *Proceedings of the PIPOC 2015 International Palm Oil Congress* (pp. 617-621).Kuala Lumpur.
- Salerno, M.-I., Gianinazzi, S., Arnould, C., & Gianinazzi-Pearson, V. (2004). Ultrastructural and cell wall modifications during infection of *Eucalyptus viminalis* roots by a pathogenic *Fusarium oxysporum* strain. *Journal of*

General Plant Pathology, 70(3), 145-152 <https://doi.org/10.1007/s10327-004-0107-x>.

Salzman, R. A., Fujita, T., Salzman, K. Z., Hasegawa, P. M., & Bressan, R.A. (1999). An improved RNA isolation method for plant tissues containing high levels of phenolic compounds of carbohydrates. *Plant Mol. Biol.*, 11, 11-17. <https://doi.org/10.1023/A:1007520314478>.

Sambanthamurthi, R., Singh, R., Parveez, G. K. A., Ong-Abdullah, M., & Kushairi, A. (2009). Opportunities for oil palm via breeding and biotechnology. In S. M. Jain & P. M. Priyadarshan (Eds.), *Breeding Plantation Tree Crops*, (pp. 377-422). New York: Springer. https://doi.org/10.1007/978-0-387-71201-7_11.

Sanders, R., Mason, D. J., Foy, C. A., & Huggett, J. F. (2014). Considerations for accurate gene expression measurement by reverse transcription quantitative PCR when analysing clinical samples. *Analytical and Bioanalytical Chemistry*, 406(26), 6471-6483. <https://doi.org/10.1007/s00216-014-7857-x>.

Savary, S., Teng, P. S., Willocquet, L., & Nutter, F. (2006). Quantification and modeling of crop losses: A review of purposes. *Annu. Rev. Phytopathol.*, 44, 89-112. <https://doi.org/10.1146/annurev.phyto.44.070505.143342>.

Sawant, S. S., Singh, P. K., Madalana, R., & Tuli, R. (2001). Designing of an artificial expression cassette for the high level expression of transgenes in plants. *Theor. Appl. Genet.*, 102, 653-644. <https://doi.org/10.1007/s001220051691>.

Schummer, M., Ng, W., Nelson, P. S., Bumgarner, R. E., & Hood, L. (1997). Inexpensive handheld device for the construction of high-density nucleic acid arrays. *Biotechniques*, 23, 1087-1092. <https://doi.org/10.2144/97236st06>.

Schünmann, P. H., Richardson, A. E., Vickers, C. E., & Delhaize, E. (2004). Promoter analysis of the barley *Pht1;1* phosphate transporter gene identifies regions controlling root expression and responsiveness to phosphate deprivation. *Plant Physiol.*, 136(4), 4205-4214. <https://doi.org/10.1104/pp.104.045823>.

Shabb, J. B., Muhonen, W. W., & Mehus, A. A. (2017). Chapter Twenty - Quantitation of human metallothionein isoforms in cells, tissues, and cerebrospinal fluid by mass spectrometry. *Methods in Enzymology*, 586, 413-431. <https://doi.org/10.1016/bs.mie.2016.11.004>.

Shang, X., Zhu, L., Duan, Y., He, Q., Zhao, M., Yu, Y., & Guo, W. (2022). An easy and rapid transformation protocol for transient expression in cotton fiber. *Frontiers in Plant Science*, 13, 837994. <https://doi.org/10.3389/fpls.2022.837994>.

- Shirsat, A., Wilford, N., Croy, R., & Boulter, D. (1989). Sequences responsible for the tissue specific promoter activity of a pea legumin gene in tobacco. *Mol. Genet. Genomics*, 215, 326-331. <https://doi.org/10.1007/BF00339737>.
- Shrawat, A. K., Carroll, R.T., DePauw, M., Taylor, G. J., & Good, A.G. (2008). Genetic engineering of improved nitrogen use efficiency in rice by the tissue-specific expression of alanine aminotransferase. *Plant Biotechnol. J.*, 6, 722–732. <https://doi.org/10.1111/j.1467-7652.2008.00351.x>.
- Siddiqui, Y., Surendran, A., Paterson, R. R. M., Ali, A., & Ahmad, K. (2021). Current strategies and perspectives in detection and control of basal stem rot of oil palm. *Saudi Journal of Biological Sciences*, 28(5), 2840-2849. <https://doi.org/10.1016/j.sjbs.2021.02.016>.
- Sierro, N., Battey, J. N. D., Ouadi, S., Bakaher, N., Bovet, L., Willig, A., Goepfert, S., Peitsch, M. C., & Ivanov, N. V. (2014). The tobacco genome sequence and its comparison with those of tomato and potato. *Nature Communications*, 5(1), 3833. <https://doi.org/10.1038/ncomms4833>.
- Singh, R., Meilina, O. A., Low, E. T. L., Arif, A. M., Rozana, R., Rajanaidu, N., Ooi, C. L. L., Ooi, S. E., Chan, K. L., Mohd Amin, H., Norazah, A., Jayanthi, N., Bacher, B., Lakey, N., Smith, S. W., He, D., Hogan, M., Budiman, M. A., Lee, E. K., Desalle, R., Kudrna, D., Goicoechea, J. L., Wing, R. A., Wilson, R. K., Fulton, R. S., Ordway, J. M., Martienssen, R. A., & Sambanthamurthi, R. (2013). Oil palm genome sequence reveals divergence of interfertile species in old and new worlds. *Nature*, 500(7462), 335–339. <https://doi.org/10.1038/nature12309>.
- Sirimat, S., & Sakulsathaporn, A. (2019). Optimization of explant surfacemsterilization conditions and multiple shoot induction in threatened plant *Phanera sirindhorniae*. *J Adv. Agric. Technol.*, 6(4), 263-266. <https://doi.org/10.18178/joaat.6.4.263-266>.
- Skrypina, N. A., Timofeeva, A. V., Khaspekov, G. L., Savochkina, L. P., & Beabealashvili, R. (2003). Total RNA suitable for molecular biology analysis. *J Biotechnol.*, 105(1-2), 1-9. [https://doi.org/10.1016/s0168-1656\(03\)00140-8](https://doi.org/10.1016/s0168-1656(03)00140-8).
- Srinath, T., Reddy, V. D., & Rao, K. V. (2017). Isolation and functional characterization of a novel stress inducible promoter from pigeonpea (*Cajanus cajan* L). *Plant Cell, Tissue and Organ Culture*, 53(3), 487-494. <https://doi.org/10.1007/s11240-016-1123-1>.
- Steffens, B., & Rasmussen, A. (2016). The physiology of adventitious roots. *Plant Physiol.*, 170(2), 603-617. <https://doi.org/10.1104/pp.15.01360>.
- Steinkellner, S., Mammarler, R., & Vierheilig, H. (2005). Microconidia germination of tomato pathogen *Fusarium oxysporum* in the presence of root exudate. *J Plant Interacts*, 1, 23–30. <https://doi.org/10.1080/17429140500134334>.

- Stern, N., & Stern, N. H. (2007). The Economics of Climate Change: The Stern Review. *Cambridge University Press*.
- Stougaard, J., Jørgensen, J. E., Christensen, T., Kühle, A., & Marcker, K. A. (1990). Interdependence and nodule specificity of cis-acting regulatory elements in the soybean leghemoglobin *lbc3* and *N23* gene promoters. *Mol. Gen. Genet.*, 220(3), 353-360. <https://doi.org/10.1007/bf00391738>.
- Strizhov, N., Ábrahám, E., Ökrész, L., Blickling, S., Zilberstein, A., Schell, J., Koncz, C., & Szabados, L. (1997). Differential expression of two *P5CS* genes controlling proline accumulation during salt-stress requires ABA and is regulated by ABA1, ABI1 and AXR2 in *Arabidopsis*. *The Plant Journal*, 12(3), 557-569. <https://doi.org/10.1111/j.0960-7412.1997.00557.x>.
- Sugimoto, K., Takeda, S., & Hirochika, H. (2003). Transcriptional activation mediated by binding of a plant GATA-type zinc finger protein AGP1 to the AG-motif (AGATCCAA) of the wound-inducible *Myb* gene *NtMyb2*. *Plant J.*, 36(4), 550-564. <https://doi.org/10.1046/j.1365-313x.2003.01899.x>.
- Sun, L., Wang, J., Song, K., Sun, Y., Qin, Q., & Xue, Y. (2019). Transcriptome analysis of rice (*Oryza sativa* L.) shoots responsive to cadmium stress. *Scientific Reports*, 9(1), 10177. <https://doi.org/10.1038/s41598-019-46684-w>.
- Suzuki, H., Wagner, T., & Tierney, M. L. (1993). Differential expression of two soybean (*Glycine max* L.) proline-rich protein genes after wounding. *Plant Physiol.*, 101(1283-1287), 1283-1287. <https://doi.org/10.1104/pp.101.4.1283>.
- Svensson, J. T., Crosatti, C., Campoli, C., Bassi, R., Stanca, A. M., Close, T. J., & Cattivelli, L. (2006). Transcriptome analysis of cold acclimation in barley albina and xantha mutants. *Plant Physiol.*, 141(1), 257-270. <https://doi.org/10.1104/pp.105.072645>.
- Swapna, L., Khurana, R., Vijaya, Kumar, S., Tyagi, A.K., & Rao, K. V. (2011). Pollen-specific expression of *Oryza sativa Indica* pollen allergen gene (*OSIPA*) promoter in rice and *Arabidopsis* transgenic systems. *Mol. Biotechnol.*, 48, 49–59. <https://doi.org/10.1007/s12033-010-9347-5>.
- Székely, G., Ábrahám, E., Cséplő, Á., Rigó, G., Zsigmond, L., Csiszár, J., Ayaydin, F., Strizhov, N., Jásik, J., Schmelzer, E., Koncz, C., & Szabados, L. (2008). Duplicated *P5CS* genes of *Arabidopsis* play distinct roles in stress regulation and developmental control of proline biosynthesis. *The Plant Journal*, 53(1), 11-28. <https://doi.org/10.1111/j.1365-1313X.2007.03318.x>.
- Tameshige, T., Hirakawa, Y., Torii, K. U., & Uchida, N. (2015). Cell walls as a stage for intercellular communication regulating shoot meristem development. *Front. Plant Sci.*, 6, 324. <https://doi.org/10.3389/fpls.2015.00324>.

- Tamilarasan, S., & Rajam, M. V. (2013). Engineering crop plants for nematode resistance through host-derived RNA interference. *Cell Dev. Biol.*, 2, 2. <https://doi.org/10.4172/2168-9296.1000114>.
- Tedder, T. F., Strueli, M., Schlossman, S. F., & Saito, H. (1988). Isolation and structure of a cDNA encoding the B1 (CD20) cell-surface antigen of human B lymphocytes. *Proc. Natl. Acad. Sci. USA*, 85, 208–212. <https://doi.org/10.1073/pnas.85.1.208>.
- Tizaoui, K., & Kchouk, M. E. (2012). Genetic approaches for studying transgene inheritance and genetic recombination in three successive generations of transformed tobacco. *Genetics and Molecular Biology*, 35(3), 640-649. <https://doi.org/10.1590/S1415-47572012000400015>.
- Tollefson, J. (2011). Brazil cooks up transgenic bean. *Nature*, 478(7368), 168. <https://doi.org/10.1038/478168a>.
- Toyofuku, K., Umemura, T., & Yamaguchi, J. (1998). Promoter elements required for sugar-repression of the *RAmy3D* gene for α -amylase in rice. *FEBS Lett.*, 428(3), 275-280. [https://doi.org/10.1016/s0014-5793\(98\)00518-3](https://doi.org/10.1016/s0014-5793(98)00518-3).
- Trapnell, C., Pachter, L., & Salzberg, S. L. (2009). Discovering splice junctions with RNA-Seq. *Bioinformatics* 25, 1105–1111. <https://doi.org/10.1093/bioinformatics/btp120>.
- Trapnell, C., Williams, B. A., Pertea, G., Mortazavi, A., Kwan, G., van Baren, M. J., Salzberg, S. L., Wold, B. J., Pachter, L. (2010). Transcript assembly and quantification by RNA-Seq reveals unannotated transcripts and isoform switching during cell differentiation. *Nature Biotechnology*, 28(5), 511-515. <https://doi.org/10.1038/nbt.1621>.
- Tremousaygue, D., Manevski, A., Bardet, C., Lescure, N., & Lescure, B. (1999). Plant interstitial telomere motifs participate in the control of gene expression in root meristems. *Plant J.* 20(5), 553-561. <https://doi.org/10.1046/j.1365-3113x.1999.00627.x>.
- Tricoli, D. M., Carney, K. J., Russell, P. F., McMaster, J. R., Groff, D. W., Hadden, K. C., Himmel, P. T., Hubbard, J. P., Boeshore, M. L., & Quemada, H.D. (1995). Field-evaluation of transgenic squash containing single or multiple virus coat protein gene constructs for resistance to cucumber mosaic virus, watermelon mosaic virus 2, and zucchini yellow mosaic virus. *Nature Biotechnology*, 13, 1458–1465. <https://doi.org/10.1038/nbt1295-1458>.
- Uchida, R. (2000). Essential nutrients for plant growth: nutrient functions and deficiency symptom. In J. A. Silva and R. Uchida (Eds.), *Plant nutrient management in Hawaii's soils, approaches for tropical and subtropical agriculture*, (pp. 32-55). College of Tropical Agriculture and Human Resources, University of Hawaii, Manoa.

- Uga, Y., Sugimoto, K., Ogawa, S., Rane, J., Ishitani, M., Hara, N., Kitomi, Y., Inukai, Y., Ono, K., Kanno, N., Inoue, H., Takehisa, H., Motoyama, R., Nagamura, Y., Wu, J., Matsumoto, T., Takai, T., Okuno, K., & Yano, M. (2013). Control of root system architecture by *DEEPER ROOTING 1* increases rice yield under drought conditions. *Nature Genetics*, 45, 1097–1102. <https://doi.org/10.1038/ng.2725>.
- Urao, T., Yamaguchi-Shinozaki, K., Urao, S., & Shinozaki, K. (1993). An *Arabidopsis myb* homolog is induced by dehydration stress and its gene product binds to the conserved MYB recognition sequence. *Plant Cell*, 5(11), 1529-1539. <https://doi.org/10.1105/tpc.5.11.1529>.
- van der Does, H. C., Constantin, M. E., Houterman, P. M., Takken, F.L.W., Cornelissen, B. J. C., Haring, M. A., van den Burg, H. A., & Rep, M. (2019). *Fusarium oxysporum* colonizes the stem of resistant tomato plants, the extent varying with the R-gene present. *European Journal of Plant Pathology*, 154(1), 55-65. <https://doi.org/10.1007/s10658-018-1596-3>.
- van Dijk, M., Morley, T., Rau, M. L., & Saghai, Y. (2021). A meta-analysis of projected global food demand and population at risk of hunger for the period 2010–2050. *Nature Food*, 2(7), 494-501. <https://doi.org/10.1038/s43016-021-00322-9>.
- Vaughn, J. N., Ellingson, S. R., Mignone, F., & von Arnim, A. G. (2012). Known and novel post-transcriptional regulatory sequences are conserved across plant families. *RNA*, 18, 368–384. <https://doi.org/10.1261/rna.031179.111>.
- Velculescu, V. E., Zhang, L., Vogelstein, B., & Kinzler, K. W. (1995). Serial analysis of gene expression. *Science*, 270, 484–487. <https://doi.org/10.1126/science.270.5235.484>.
- Venter, M., & Botha, F. C. (2010). Synthetic promoter engineering. In E.C. Pua, and M.R. Davey (Eds.) In *Plant developmental biology—biotechnological perspectives*, (pp. 393–414). Berlin/Heidelberg: Springer. https://doi.org/10.1007/978-3-642-04670-4_20.
- Vieweg, M. F., Frühling, M., Quandt, H. J., Heim, U., Bäumlein, H., Pühler, A., Küster, H., & Andreas, M. P. (2004). The promoter of the *Vicia faba* L. leghemoglobin gene *VfLb29* is specifically activated in the infected cells of root nodules and in the arbuscule-containing cells of mycorrhizal roots from different legume and nonlegume plants. *Mol. Plant Microbe Interact.*, 17(1), 62-69. <https://doi.org/10.1094/mpmi.2004.17.1.62>.
- Vignols, F., Jose-Estanyol, M., Caparros-Ruiz, D., Rigau, J., & Puigdomenech P. (1999). Involvement of a maize proline-rich protein in secondary cell wall formation as deduced from its specific mRNA localization. *Plant Mol. Biol.*, 39(5), 945-52. <https://doi.org/10.1023/A:1006129703262>.

- Vijaybhaskar, V., Subbiah, V., Kaur, J., Vijayakumari, P., & Siddiqi, I. (2008). Identification of a root-specific glycosyltransferase from *Arabidopsis* and characterization of its promoter. *J. Biosci.*, 33, 185-193. <https://doi.org/10.1007/s12038-008-0036-5>.
- Werner, T., Nehnevajova, E., Köllmer, I., Novák, O., Strnad, M., Krämer, U., & Schmülling, T. (2010). Root-specific reduction of cytokinin causes enhanced root growth, wheat (*Triticum aestivum* L.). *Mol. Biol. Rep.*, 37, 737–744. <https://doi.org/10.1105/tpc.109.072694>.
- Wingender, E., Chen, X., Fricke, E., Geffers, R., Hehl, R., Liebich, I., Krull, M., Matys, V., Michael, H., Ohnhäuser, R., Prüss, M., Schacherer, F., Thiele, S., (2001). The TRANSFAC system on gene expression regulation. *Nucleic Acids Research*, 29(1), 281-283. <https://doi.org/10.1093/nar/29.1.281>.
- Winicov, I., Valliyodan, B., Xue, L., & Hooper, J. K. (2004). The *MsPRP2* promoter enables strong heterologous gene expression in a root-specific manner and is enhanced by overexpression of *Alfin 1*. *Planta*, 219, 925-935. <https://doi.org/10.1007/s00425-004-1296-4>.
- Wu, K. Malik., K., Tian, L., Hu, M., Froster, E., Brown, D., & Miki, B. (2001). Enhancers and core promoter elements are essential for the activity of a cryptic gene activation sequence from tobacco, tCUP. *Mol. Genet. Genomics*, 265, 763–770. <https://doi.org/10.1007/s004380100478>.
- Xiao, K., Liu, J., Dewbre, G., Harrison, M., & Wang, Z-Y. (2006). Isolation and characterization of root-specific phosphate transporter promoters from *Medicago truncatula*. *Plant Biology*, 8(4), 439-449. <https://doi.org/10.1055/s-2005-873053>.
- Xiong, L. L., Xue, L. L., Al-Hawwas, M., Huang, J., Niu, R.Z., Tan, Y. X., Xu, Y., Su, Y. Y., Liu, J., & Wang, T. H. (2020). Single-nucleotide polymorphism screening and RNA sequencing of key messenger RNAs associated with neonatal hypoxic-ischemia brain damage. *Neural Regen. Res.*, 15(1), 86-95. <https://doi.org/10.4103/1673-5374.264469>.
- Xu, N., Hagen, G., & Guilfoyle, T. (1997). Multiple auxin response modules in the soybean SAUR 15A promoter. *Plant Sci.*, 126, 193-201. [https://doi.org/10.1016/S0168-9452\(97\)00110-6](https://doi.org/10.1016/S0168-9452(97)00110-6).
- Xu, X., Chen, C., Fan, B., & Chen, Z. (2006). Physical and functional interactions between pathogen-induced *Arabidopsis* *WRKY18*, *WRKY40*, and *WRKY60* transcription factors. *Plant Cell*, 18, 1310-1326. <https://doi.org/10.1105/tpc.105.037523>.
- Xu, X., Guo, S., Chen, K., Song, H., Liu, J., Guo, L., Qian, Q., & Wang, H. (2010). A 796 bp *PsPR10* gene promoter fragment increased root-specific expression of the GUS reporter gene under the abiotic stresses and signal molecules in tobacco. *Biotechnol Lett.*, 32, 1533–1539. <https://doi.org/10.1007/s10529-010-0312-y>.

- Xu, Y., Buchholz, W. G., DeRose, R. T., & Hall, T.C. (1995). Characterization of a rice gene family encoding root-specific proteins. *Plant Mol. Biol.*, 27, 237-248. <https://doi.10.1007/BF00020180>.
- Xue, G. -P., Rae, A. L., White, R. G., Drenth, J., Richardson, T., & McIntyre, C. L. (2016). A strong root-specific expression system for stable transgene expression in bread wheat. *Plant Cell Reports*, 35(2), 469-481. <https://doi.10.1007/s00299-015-1897-3>.
- Xun, H., Zhang, X., Yu, J., Pang, J., Wang, S., Liu, B., Dong, Y., Jiang, L., & Guo, D. (2021). Analysis of expression characteristics of soybean leaf and root tissue-specific promoters in *Arabidopsis* and soybean. *Transgenic Research*, 30(6), 799-810. <https://doi.org/10.1007/s11248-021-00266-7>.
- Yahya, Z., Husin, A., Talib, J., Othman, J., Ahmed, O.H., & Jalloh, M. B. (2010). Oil Palm (*Elaeis guineensis*) roots response to mechanization in Bernam series soil. *American Journal of Applied Sciences*, 7(3), 343-348. <https://doi.org/10.3844/ajassp.2010.343.348>.
- Yamamoto, S., Nakano, T., Suzuki, K., & Shinshi, H. (2004). Elicitor-induced activation of transcription via W box-related cis-acting elements from a basic chitinase gene by WRKY transcription factors in tobacco. *Biochim. Biophys. Acta*, 1679(3), 279-287. <https://doi.org/10.1016/j.bbaexp.2004.07.005>.
- Yamamoto, Y. T., Taylor, C.G., Acedo, G.N., Cheng, C.L., & Conkling, M.A. (1991). Characterization of cis-acting sequences regulating root-specific gene expression in tobacco. *The Plant Cell*, 3, 371-382. <https://doi.org/10.1105/tpc.3.4.371>.
- Yan, K., Ablimit, M., Liu, S., Liu, Z., & Wang, Y. (2023). A novel metallothionein gene *HcMT* from halophyte shrub *Halostachys caspica* respond to cadmium and sodium stress. *Plant Physiology and Biochemistry*, 201, 107763. <https://doi.org/https://doi.org/10.1016/j.plaphy.2023.107763>.
- Yang, G. P., Ross D. T., Kuang, W. W., Brown, P. O., & Weigel, R. J. (1999). Combining SSH and cDNA microarrays for rapid identification of differentially expressed genes. *Nucleic Acids Res.*, 27(6), 1517–1523. <https://doi.10.1093/nar/27.6.1517>.
- Yang, M., Zhang, F., Wang, F., Dong, Z., Cao, Q., & Chen, M. (2015). Characterization of a type 1 metallothionein gene from the stresses-tolerant plant *Ziziphus jujuba*. *Int. J. Mol. Sci.* 16: 16750–16762. <https://doi.org/10.13390/ijms160816750>.
- Yang, R. C., Xu, H. L., Yu, W. G., Lu, C. G., Long, M. S., Liu, C.Q., Pan, N. S., & Chen, Z. L. (1995). Transgenic tomato plants expressing cucumber mosaic virus coat protein (CMV-cp) and their resistance to cucumber mosaic virus (CMV). *Acta Agric. Jiangsu*, 11, 42–46.

- Yang, Z., Gao, Z., Zhou, H., He, Y., Liu, Y., Lai, Y., Zheng, J., Li, X., & Liao, H., (2021). GmPTF1 modifies root architecture responses to phosphate starvation primarily through regulating *GmEXPB2* expression in soybean. *The Plant Journal*, 107(2), 525-543. <https://doi.org/510.1111/tpj.15307>.
- Yeap, W.-C., Loo, J. M., Wong, Y. C., & Kulaveerasingam, H. (2014). Evaluation of suitable reference genes for qRT-PCR gene expression normalization in reproductive, vegetative tissues and during fruit development in oil palm. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 116(1), 55-66. <https://doi.org/10.1007/s11240-013-0382-3>.
- Yu, C. Y., Liu, Y. H., Zhang, A.D., Su, S., Yan, A., Huang, L., Ali, I., Liu, Y., Forde, B.G., & Gan, Y. B. (2015). MADS-box transcription factor *OsMADS25* regulates root development through affection of nitrate accumulation in rice. *PloS One*. 10, e0135196. <https://doi.org/10.1371/journal.pone.0135196>.
- Zakaria, L. (2022). Basal stem rot of oil palm - The pathogen, disease incidence, and control methods. *Plant Dis*. <https://doi.org/10.1094/pdis-02-22-0358-fe>.
- Zavallo, D., Bilbao, M. L., Esteban Hopp, H., & Heinz, R. (2010). Isolation and functional characterization of two novel seed-specific promoters from sunflower (*Helianthus annuus* L.). *Plant Cell Reports*, 29, 239–248. <https://doi.org/10.1007/s00299-010-0816-x>.
- Zeng, Y., & Yang, T. (2002). RNA isolation from highly viscous samples rich in polyphenols and polysaccharides. *Plant. Mol. Biol. Rep.*, 20, 417a-417e. <https://doi.org/10.1007/BF02772130>.
- Zhang, C., Pan, S., Chen, H., Cai, T., Zhuang, C., Deng, Y., Zhuang, Y., Zeng, Y., Chen, S., & Zhuang, W. (2016). Characterization of *NtREL1*, a novel root-specific gene from tobacco, and upstream promoter activity analysis in homologous and heterologous hosts. *Plant Cell Reports*, 35(4), 757-769. <https://doi.10.1007/s00299-015-1918-2>.
- Zhang, L., Zhou, W., Velculescu, V. E., Kern, S. E., Hruban, R. H., Hamilton, S. R., Vogelstein, B., & Kinzler, K. W. (1997). Gene expression profiles in normal and cancer cells. *Science*, 276, 1268–1272. <https://doi.10.1126/science.276.5316.1268>.
- Zhang, W., Fang, D., Dong, K., Hu, F., Ye, Z., & Cao, J. (2023a). Insights into the environmental factors shaping lateral root development. *Physiologia Plantarum*, 175(2), e13878. <https://doi.org/13810.1111/ppl.13878>.
- Zhang, X., Gong, X., Yu, H., Su, X., Cheng, S., Huang, J., Lei, Z., Li, M. and Ma, F. (2023b). The proline-rich protein *MdPRP6* confers tolerance to salt stress in transgenic apple (*Malus domestica*). *Scientia Horticulturae*, 308, 111581. <https://doi.org/10.1016/j.scienta.2022.111581>.

- Zhang, Y., Tan, L., Zhu, Z., Yuan, L., Xie, D., & Sun, C. (2015). *TOND1* confers tolerance to nitrogen deficiency in rice. *The Plant Journal*, 81, 367–376. <https://doi.org/10.1111/tpj.12736>.
- Zhao, N., Sun, X., Hou, S., Chen, G., Zhang, H., Han, Y., Zhou, J., Wang, X., & Zhang, Z. (2022). N addition mitigates water stress via different photosynthesis and water traits for three native plant species in the Qinghai-Tibet plateau. *Agriculture*, 12(11), 1873. <https://doi.org/10.3390/agriculture12111873>.
- Zhou J, Xie, J., Liao, H., & Wang, X. (2014). Overexpression of β -expansin gene *GmEXPB2* improves phosphorus efficiency in soybean. *Physiologia Plantarum*, 150, 194–204. <https://doi.org/10.1111/ppl.12077>.
- Zhou, D. X., Li, Y. F., Rocipon, M., & Mache, R. (1992). Sequence-specific interaction between *S1F*, a spinach nuclear factor, and a negative cis-element conserved in plastid-related genes. *J Biol. Chem.*, 267(33), 23515-23519. [https://doi.org/10.1016/S0021-9258\(18\)35869-1](https://doi.org/10.1016/S0021-9258(18)35869-1).
- Zhou, J. M., & Goldsbrough, P. B. (1995). Structure, organization and expression of the metallothionein gene family in *Arabidopsis*. *Molecular and General Genetics* 248, 318–328. <https://doi.org/10.1007/BF02191599>.
- Zubaidah, R., & Siti Nor Akmar, A. (2005). The effects of metal ions on root-specific expression of the oil palm *MT3-B* gene promoter. *Proceedings of the PIPOC 2005 International Palm Oil Congress*, (pp. 1104–1110). Kuala Lumpur.
- Zubaidah, R., & Siti Nor Akmar, A. (2010). Functional characterisation of the oil palm type 3 metallothionein-like gene (*MT3-B*) promoter. *Plant Molecular Biology Reporter*, 28(3), 531-541. <https://doi.org/10.1007/s11105-009-0177-1>.
- Zubaidah, R., Nurniwalis, A. W., Siti Nor Akmar, A., & Parveez, G. K. A. (2017). The use of *Arabidopsis thaliana* model system for testing oil palm promoter: Case study on oil palm *MT3-A* promoter. *J Oil Palm Res.*, 29(2), 189-196. <https://doi.org/10.21894/jopr.2017.2902.04>.
- Zulkifli, Y., Norziha, A., Naqiuddin, M. H., Fadila, A. M., Nor Azwani, A. B., Suzana, M., Samsul, K. R., Ong-Abdullah, M., Singh, R., Parveez, G. K. A., & Kushairi, A. (2017). Designing the oil palm of the future. *J Oil Palm Res.*, 29, 440 – 455. <https://doi.org/10.21894/jopr.2017.00015>.