



UNIVERSITI PUTRA MALAYSIA

**FEMALE BROODSTOCK REPRODUCTIVE OUTPUT, LARVAL
REARING AND SPAT SURVIVAL OF BLACK LIPPED OYSTER**
(Pinctada margaritifera)

FARIBORZ EHTESHAMI
FP 2010 7

**FEMALE BROODSTOCK REPRODUCTIVE OUTPUT, LARVAL
REARING AND SPAT SURVIVAL OF BLACK LIPPED OYSTER**
(Pinctada margaritifera)

By

FARIBORZ EHTESHAMI

**Thesis submitted to the School of Graduates Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

July 2010



*I would like to dedicate this thesis
with love to the memory of my father
Heshmat Ehteshami and my mother
Jamileh yagmaeian to keep their
spirits alive*

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Doctor of Philosophy.

**FEMALE BROODSTOCK REPRODUCTIVE OUTPUT, LARVAL
REARING AND SPAT SURVIVAL OF BLACK LIPPED OYSTER
(*Pinctada margaritifera*)**

By

FARIBORZ EHTESHAMI

July 2010

Chairman: Dr Annie Christianus, PhD

Faculty: Agriculture

Overexploitation of *Pinctada margaritifera* as one of the natural resource was leading to a dramatic loss of its population in the north coast of Persian Gulf. Low abundance and density existing in the natural beds and poor larval recruitment prompted research on hatchery propagation of this species. Further research should be carried out to improve the survival and growth of larvae produced through artificial propagation.

This study addressed important issues in relation to the supplementation of polyunsaturated fatty acids (PUFA) in diet and their role in egg quality and biochemical composition, and larval growth and survival; microalgae biochemical

composition and its role in larvae culture; and spat settlement and transfer time to the sea farm.

In the first experiment, the effects of supplementary PUFA on oogenesis and hatching rate of *P. margaritifera* broodstock were compared with naturally fed oysters and those fed only microalgae. Supplementary food was effective ($P < 0.05$) on producing larger sized eggs (57.6 μm) and larvae (80.1 μm), and higher percentage of *P. margaritifera* D shape larvae (31.3%). Palmitic (16:0) and stearic (18:0) acid were the major saturated fatty acids in neutral and polar parts of gonad lipid. Oysters fed with supplementary PUFA had more docosahexaenoic acid (DHA) and less monounsaturated fatty acids (MUFA) in their gonad. The ratio of n-3/n-6 fatty acids in neutral lipid was the best representative of differences in conditioning of oysters for spawning and interpretation of the results of egg size and hatching performance.

In the second experiment, the effects of partial supplementation of the diet with PUFA on growth and survival of *P. margaritifera* D-shape and umbo larvae were investigated. PUFA supplemented in droplet form did not increase the growth and survival of D-shape and umbo *P. margaritifera* larvae compared to those fed fresh algae of *T. Iso* ($P > 0.05$). Considering results of size range of larvae, it can be concluded that through the grading process, a great number of larvae would be lost in treatments with supplementary diet compared to those fed with *T. Iso* only. D-shape and umbo larvae showed a similar performance in survival, with the highest related to larvae fed with fresh algae followed by 10, 30 and 100% of diet replaced with PUFA emulsions. While the lowest survival attributed to the unfed larvae.

The nutritional value of the three microalgal species used for the feeding *P. margaritifera* D-shape and umbo larvae: T. Iso, *Chaetoceros muelleri* and *C. calcitrans* in mono, binary and ternary species diets were evaluated. D-shape and umbo larval growth and survival were found to be the greatest with diet T. Iso mono and ternary species, respectively. *C. calcitrans* showed the lowest nutritional value for both larval stages. Growth of D-shape larvae was positively correlated with levels of MUFA, DHA, DHA/EPA and majority of the unsaturated fatty acids with 18 C including: 18:1n-9, 18:2n-6, 18:3n-3, 18:4n-3 and negatively correlated to 16:3n-4, 20:4n-6 (arachidonic acid) and EPA contents of microalgae.

The third experiment was conducted to investigate the effect of collector materials and position on *P. margaritifera* spat attachment. Settlement on polyethylene pipes (54.6%) was significantly higher ($P < 0.05$) than on plastic baskets (25%). Significantly higher ($P < 0.05$) spat catch was recorded from collectors installed close to the bottom as compared to the top part of the settlement tank. Numbers of dead spat after settlement on pipe (1.4%) and basket (1.5%) were not significantly different ($P > 0.05$). Possible factors causing the pattern of settlement reveals that it could be related to the material used for collector, light sensitivity and gravity force. These results indicated that polyethylene pipes positioned horizontally in the lower part of the tank are suitable for the settlement of *P. margaritifera* pediveliger larvae in hatchery.

In the fourth experiment, *P. margaritifera* spat were transferred to the sea farms in Hendurabi and Lavan Islands at 25, 50, and 65 days post settlement, while one group was maintained in the hatchery. Retaining the spat in the hatchery for more than 25 days did not improve the growth and survival ($P > 0.05$). Spat grown in Hendurabi

were twice the size of those grown in hatchery and were at least 1 cm longer than those grown in Lavan. Spat from two propagation trials upon reaching 55 days old, were deployed to the Hendurabi on 5th September and 7th October 2008, respectively. Culture was carried out for five months. Spat from the first deployment were significantly larger in size than the second one ($P < 0.05$). Daily growth rate (DGR) was observed to be higher in September (warmer month) as compared to February (winter). Statistical analyses showed highly positive correlation between DGR and water temperature, whereas food abundance had a negligible effect.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Doktor Falsafah

**MEMPERBAIKI HASIL PEMBIAKAN INDUK BETINA,
TUMBESARAN DAN KEMANDIRIAN LARVA BENTUK-D,
UMBO DAN SPAT *Pinctada margaritifera***

Oleh

FARIBORZ EHTESHAMI

Julai 2010

Pengerusi: Dr. Annie Christianus, PhD

Fakulti: Pertanian

Eksploitasi terhadap sumber semulajadi *Pinctada margaritifera* dengan dramatiknya telah mengurangkan populasinya di utara pantai Teluk Farsi. Taburan dan densiti yang rendah di tapak semulajadi dan kekurangan sumber untuk larva telah menggalakkan kajian ke atas pengeluaran spesis ini di hatcheri. Kajian lanjutan adalah perlu untuk meningkatkan kadar kemandirian dan tumbesaran larva yang dihasilkan melalui pengeluaran artifisial.

Kajian ini menekankan isu penting berkaitan dengan penambahan asid lemak poli tidak tepu (PUFA) dalam gizi dan peranannya ke atas kualiti telur dan komposisi biokimia, dan tumbesaran dan kemandirian larva; komposisi biokimia alga mikro

dan peranannya dalam kultur larvi; dan pelekatan dan jangkamasa perpindahan spat ke kultur di laut.

Pada eksperimen pertama, kesan ke atas oogenesis dan kadar penetasan hasil dari induk *P. margaritifera* dibandingkan diantara induk yang diberi makanan dengan penambahan PUFA dengan makanan semulajadi dan alga mikro. Makanan tambahan berjaya ($P < 0.05$) menghasilkan telur (57.6 μm) dan larva yang bersaiz lebih besar (80.1 μm), dan peratusan larva-D yang lebih tinggi (31.3%). Asid palmitik (16:0) dan stearik (18:0) merupakan asid lemak tepu yang utama terdapat di bahagian neutral dan kutub pada lipid gonad. Tiram yang diberi PUFA tambahan mempunyai asid dokosahekaenoik (DHA) yang lebih tinggi dan asid lemak mono tak tepu (MUFA) yang rendah dalam komposisi gonadnya. Nisbah n-3/n-6 pada lipid neutral merupakan penunjuk yang sangat sesuai untuk pembezaan dalam penyesuaian tiram untuk pembiasaan dan menerangkan keputusan untuk saiz dan hasil penetasan telur.

Pada eksperimen kedua, kesan penambahan separa PUFA ke dalam gizi larva ke atas tumbesaran dan kemandirian larva bentuk-D dan umbo *P. margaritifera* telah dikaji. Penambahan PUFA dalam bentuk titisan tidak meningkatkan tumbesaran dan kemandirian larva bentuk-D dan umbo berbanding dengan larva yang diberi T. Iso ($P > 0.05$). Berdasarkan julat untuk saiz larva, kesimpulan dapat dibuat bahawa semasa proses pengredan, kehilangan sejumlah besar larva akan berlaku untuk larva yang diberi gizi dengan penambahan PUFA berbanding dengan hanya diberi T. Iso. Larva bentuk-D dan umbo menunjukkan kadar kemandirian yang sama, dengan kadar tertinggi pada larva yang diberi alga segar diikuti dengan gizi yang diberi penggantian emulsi PUFA sebanyak 10, 30 and 100%. Manakala kadar kemandirian yang paling rendah adalah pada larva yang tidak diberi sebarang makanan.

Nilai nutrient untuk tiga spesis alga mikro yang diberikan untuk larva bentuk-D dan umbo *P. margaritifera*: *T. Iso*, *Chaetoceros muelleri* dan *C. calcitrans* dalam bentuk gabungan spesis mono, binari dan ternari telah dinilai. Tumbesaran dan kadar kemandirian larva bentuk-D dan umbo yang paling tinggi adalah dengan gizi *T. Iso* gabungan spesis mono dan ternari, masing-masingnya. *C. calcitrans* memberikan nilai nutrient paling rendah untuk kedua-dua peringkat larva tersebut. Tumbesaran larva bentuk-D menunjukkan korelasi positif dengan tahap MUFA, DHA, DHA/EPA dan kebanyakan asid lemak tak tepu dengan 18 C termasuk: 18:1n-9, 18:2n-6, 18:3n-3, 18:4n-3, manakala korelasi negatif dengan 16:3n-4, 20:4n-6 (asid arakidonik) dan kandungan EPA alga mikro.

Eksperimen ketiga dijalankan untuk mengkaji kesan bahan dan kedudukan penggutip untuk penggutipan spat *P. margaritifera*. Pelekatan pada paip polietilene (54.6%) ketara lebih tinggi ($P < 0.05$) berbanding dengan bakul plastik (25%). Kutipan spat ketara lebih tinggi ($P < 0.05$) dicatatkan untuk penggutip yang diletakkan dekat dengan dasar berbanding dengan bahagian atas tangki pemendapan. Jumlah spat yang mati selepas pelekatan ke atas paip (1.4%) dan bakul (1.5%) tidak menunjukkan perbezaan yang ketara ($P > 0.05$). Faktor yang menyebabkan corak pelekatan kemungkinan mempunyai kaitan dengan bahan yang digunakan untuk penggutip, sensitiviti terhadap cahaya dan tarikan graviti. Keputusan ini menunjukkan bahawa paip polietilene yang diletak secara horizontal di bahagian bawah tangki adalah sesuai untuk pelekatan larva pediveliger *P. margaritifera* pediveliger di hatcheri.

Pada eksperimen keempat, spat *P. margaritifera* dipindahkan ke tapak di laut di Pulau Hendurabi dan Lavan pada 25, 50, dan 65 hari selepas pelekatan, manakala

satu kumpulan lagi diletakkan di hatcheri. Meletakkan spat di hatcheri lebih daripada 25 hari tidak menunjukkan kadar tumbesaran dan kemandirian yang ketara ($P > 0.05$). Spat yang dikultur di Hendurabi bersaiz dua kali ganda berbanding dengan kumpulan yang dikultur di hatcheri dan sekurang-kurangnya 1 sm lebih panjang berbanding dengan kumpulan di Lavan. Spat daripada dua pengeluaran, apabila mencapai 55 hari, dilepaskan di Hendurabi pada 5^{hb} September dan 7^{hb} Oktober 2008, tiap satunya. Kultur dijalankan selama lima bulan. Spat dari perlepasan pertama ketara lebih besar ($P < 0.05$) berbanding dengan kumpulan yang kedua. Kadar tumbesaran harian (DGR) didapati lebih tinggi pada bulan September (musim panas) berbanding dengan bulan Februari (musim sejuk). Analisis statistik menunjukkan korelasi positif yang tinggi di antara DGR dan suhu air, manakala taburan kehadiran makanan tidak memberikan sebarang kesan.

ACKNOWLEDGEMENTS

First and foremost, I would like to thank my supervisor Dr. Annie Christianus for her invaluable guidance and assistance during this study. I felt very fortunate to have an opportunity to work under her patient supervision. As well, I sincerely express my thanks to Assoc. Prof. Sharr Azni Harmin and Assoc. Prof. Che Roos Saad as members of the supervisory committee for providing me with the advice and direction during this dissertation. Their helps and contribution makes this dissertation possible.

Thanks to the technical and administrative staff of Bandar-Lengeh Mollusk Research Station, Mr. Arganji for his generosity in providing the facilities, Messrs Mahijoo, Safari and Moradii for technical helps and scuba diving operations. A special gratitude is expressed to my friend and researcher of station Mr. Hossein Rameshi for his kind support from the beginning to the end of the study.

I am indebted to Dr. Naser Agh, Head of Artemia and Aquatic Animals Research Institute for his assistance in biochemical analysis.

The research for this work was carried out at the Bandar-Lengeh Mollusk Research Station and financially supported by Iranian Fisheries Research Organization. I would not have completed this study without their support.

I want to acknowledge all my friends in Iran and Malaysia for their friendship and support and I wish the best for them in work, education and life.

Finally I would like to thank my brothers and sisters for their enduring support, helping me in the choices I have made and for their dedication. Thanks also to my niece, Mahkameh, her husband Mohammad and my other niece, Niloofar for making my wife and I feel at home.

Above and beyond all, my heartfelt gratitude to my lovely wife, Mehrnoosh Jadda her endless love, priceless, perpetual, indispensable help, support and everything made all this possible.

I certify that an Examination Committee has met on (date of viva voce) to conduct the final examination of Fariborz Ehteshami on his PhD thesis entitled “Female broodstock reproductive output, larval rearing and spat survival of pearl oyster, *Pinctada margaritifera*” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommended that the student be awarded the Doctor of Philosophy degree.

Members of the Examination Committee were as follows:

Abdul Razak Alimon, PhD

Professor
Faculty of Animal Science
Universiti Putra Malaysia
(Chairman)

Aziz Arshad, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Tan Chin Ping, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Internal Examiner)

Mehdi Saveh Doroudi, PhD

Professor
Aquaculture Division
Primary Industries & Resources
Australia
(External Examiner)

BUJANG KIM HUAT, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia
Date:



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Annie Christianus, PhD

Senior lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Sharr Azni Harmin, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Che Roos Saad, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

HASANAH MOHD GHAZALI, PhD

Professor and Dean
School of Graduate studies
Universiti Putra Malaysia

Date: 12 August 2010



DECLARATION

I declare that the thesis is my original work expect for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

Fariborz Ehteshami

Date

TABLE OF CONTENTS

	Page
ABSTRACT	iii
ABSTRAK	vii
ACKNOWLEDGEMENTS.....	xi
APPROVAL	xiii
DECLARATION.....	xv
LIST OF TABLES	xx
LIST OF FIGURES	xxii
LIST OF ABBREVIATIONS	xxv
 CHAPTER	
1.	Introduction
1.1 Specific objectives	3
2.	Literature review
2.1 Taxonomy and distribution	4
2.2 Reproduction	6
2.3 Artificial propagation	8
2.4 Feeding and digestion	9
2.5 Nutritional value of microalgae	11
2.5.1 General	11
2.5.2 Biochemical composition	14
2.6 Nutritional requirements of bivalves	21
2.6.1 Particle size	21
2.6.2 Protein	23
2.6.3 Carbohydrates	24
2.6.4 Lipids	25
2.6.5 Minerals and vitamins	28
2.7 Nutritional requirements during the reproduction process	29
2.8 Nutritional requirements during the larval stage	33
2.9 Spat collectors	39
2.10 Spat transfer	42
3.	General methodology
	45

3.1 Microalgae Culture	45
3.2 Broodstock monitoring	48
3.3 Hatchery techniques for spat production	50
3.3.1 Selection and transportation of broodstock	50
3.3.2 Seawater management and aeration	51
3.3.3 Spawning	51
3.3.4 Determination of egg numbers	53
3.3.5 Measurements	54
3.4 Biochemical analyses.....	56
3.5 Statistical methods	59
4. The effect of polyunsaturated fatty acids (PUFAs) on reproductive output of female <i>Pinctada margaritifera</i>	61
4.1 Introduction	61
4.2 Methodology	63
4.2.1 Experimental design	63
4.2.2 Biochemical analyses	66
4.2.3 Statistical methods	66
4.3 Results	67
4.3.1 Effect of partial replacement of the algal diet with enrichment oil on fecundity of <i>P. margaritifera</i>	67
4.3.2 Effect of partial replacement of the algal diet with enrichment oil on biochemical composition of gonad	69
4.4 Discussion	75
4.5 Conclusion	79
5. Effect of dietary polyunsaturated fatty acids on <i>Pinctada margaritifera</i> larval growth	81
5.1 Introduction	81
5.2 Methodology	84
5.2.1 Spawning and larvae culture	84
5.2.2 Microalgae	84
5.2.3 Biochemical analyses	86
5.2.4 Experiment 1: Effect of partial replacement of the algal diet with PUFAs supplementation on growth and survival of <i>P. margaritifera</i> D-shape larvae	86
5.2.5 Experiment 2: Effect of partial replacement of the algal	

diet with PUFAs supplementation on growth and survival of <i>P. margaritifera</i> umbo-stage larvae	87
5.2.6 Experiment 3: Evaluation of mono, binary and ternary feeding of three tropical microalgae for <i>P. margaritifera</i> D-shape larvae	88
5.2.7 Experiment 4: Evaluation of mono, binary and ternary feeding of three tropical microalgae for <i>P. margaritifera</i> umbo-stage larvae	89
5.2.8 Statistical methods	89
5.3 Results	91
5.3.1 Microalgae	91
5.3.2 Experiment 1: Effect of partial replacement of the algal diet with PUFAs supplementation on growth and survival of <i>P. margaritifera</i> D-shape larvae	95
5.3.3 Experiment 2: Effect of partial replacement of the algal diet with PUFAs supplementation on growth and survival of <i>P. margaritifera</i> umbo larvae	97
5.3.4 Experiment 3: Evaluation of mono, binary and ternary feeding of three tropical microalgae for <i>P. margaritifera</i> D-shape larvae	99
5.3.5 Experiment 4: Evaluation of mono, binary and ternary feeding of three tropical microalgae for <i>P. margaritifera</i> umbo larvae	102
5.4 Discussion	105
5.5 Conclusion	112
6. Evaluation of selected substrates for collection of hatchery reared black-lip pearl oyster (<i>Pinctada margaritifera</i>) spat	114
6.1 Introduction	114
6.2 Methodology	116
6.2.1 Experimental design	116
6.2.2 Statistical methods	117
6.3 Results	118
6.4 Discussion	120
6.5 Conclusion	122
7. Effects of location and age of <i>Pinctada margaritifera</i> spat transfer from hatchery on its growth and survival in the sea	123
7.1 Introduction	123

7.2 Methodology	125
7.2.1 Hatchery aspects	125
7.2.2 Experimental design	125
7.2.3 Statistical methods	128
7.3 Results	129
7.4 Discussion	140
7.5 Conclusion	143
8. General discussions and recommendations	145
8.1 Female broodstock and egg	145
8.2 D-shape and umbo larvae	146
8.3 Spat	147
REFERENCES	151
BIODATA OF STUDENT	180
LIST OF PUBLICATIONS.....	181

LIST OF TABLES

Table	Page
2.1 Time series growth data for three species of pearl oyster.	9
2.2 Major classes and genera of microalgae cultured for aquaculture use.....	13
2.3 The effect of different culture media on cellular density (10^6 cells ml $^{-1}$) and proximate composition (pg cell $^{-1}$) of three marine microalgae.	19
2.4 Glossary of fatty acid nomenclature.	27
3.1 Composition and preparation of Guillard's F ₂ medium.	47
4.1 Reproductive stages in the gonadal development of the genus <i>Pinctada</i> based on macroscopic observations.	66
4.2 Effect of diet on fecundity and embryonic development in <i>P. margaritifera</i>	69
4.3 Effect of diet on lipid, protein and carbohydrate (mg g $^{-1}$ dry weight), and neutral lipid (NL, in % of total lipid) of female gonad of <i>P. margaritifera</i>	70
4.4 Categories and ratios (% of total fatty acids) of fatty acids in the neutral lipids of female gonads of <i>P. margaritifera</i>	72
4.5 Categories and ratios (% of total fatty acids) of fatty acids in the polar lipids of female gonads of <i>P. margaritifera</i>	73
5.1 Diet of <i>Pinctada margaritifera</i> larvae in (1) D and (2) umbo stages.	88
5.2 Quantitative diet of <i>Pinctada margaritifera</i> (cell ml $^{-1}$) (3) D-shape and (4) umbo-stage larvae.	90
5.3 Growth rate (μ , in division day $^{-1}$) and mean value \pm standard error of dry weight (DW, in pg cell $^{-1}$), ash free dry weight (AFDW, in pg cell $^{-1}$) and the proximate composition (mg g $^{-1}$ DW) of microalgae used as a diet.	91
5.4 Quantitative categories of fatty acids (mg g $^{-1}$ dry weight) in three tropical microalgae investigated in this study.	93
5.5 Distribution of shell length (APM, in μm) of <i>P. margaritifera</i> D-shape larvae for different percentiles at the end of Experiment 1.	96
5.6 Distribution of shell length (APM, in μm) of <i>P. margaritifera</i> umbo larvae for different percentiles at the end of Experiment 2.	98
5.7 Pearson correlation between increase in APM of larvae during	

Experiments 3 and 4, and major nutrient components of microalgae used as a diet in this study.	104
7.1 Spat transfer to Lavan and Hendurabi Islands undertaken from October to November 2007 and from September to October 2008.	127
7.2 Summary on 150 days spat culture in the sea and hatchery in fall and winter. 128	
7.3 Mortality rate (%) of <i>P. margaritifera</i> spat deployed to Lavan and Hendurabi Islands 25 (T1L, T1H), 40 (T2L, T2H), and 55 (T3L, T3H) days post settlement.	134
7.4 Comparison of daily growth rate (DGR) of <i>P. margaritifera</i> spat in fall (from 5 Sep 2008 to 3 Dec 2008, bold numbers) and winter (from 6 Dec 2008 to 4 March 2009).	136

LIST OF FIGURES

Figure	Page
2.1 Schematic classification of <i>Pinctada margaritifera</i>	4
2.2 Distribution of black-lip pearl oyster, <i>P. margaritifera</i> (dark blue).	6
2.3 Anatomy of <i>Pinctada fucata</i>	11
2.4 Average percentage compositions of the PUFAs of microalgae commonly used in aquaculture.	16
2.5 Comparison of the percentages of essential amino acids in microalgae (grey bars) and Pacific oyster (<i>C. gigas</i>) larvae (black bars).....	20
3.1 Study areas inside the Persian Gulf. Lavan, Shidvar and Hendurabi Islands for broodstock and spat culture and Bandar-Lengeh for propagation of oysters and larvae culture.	49
3.2 On-bottom culture of broodstock in natural bed.	49
3.3 Spawning and larval culture room with individual spawning vessel in front.	53
3.4 Picture of egg (top right), D-shape larvae (top left), and adult oyster (below)....	55
4.1 Schematic distribution of adult oysters in treatments.	65
4.2 Reproductive output of <i>P. margaritifera</i> directly spawned (T1) and after 24 days (T2, T3 and T4).	68
4.3 Reproductive stages of female <i>P. margaritifera</i> at the end of conditioning period.	68
4.4 Neutral fatty acids compositions in female gonads of <i>P. margaritifera</i> . Expressed as the percentage of total fatty acids of the fraction.	74
4.5 Polar fatty acids compositions in female gonads of <i>P. margaritifera</i> . Expressed as the percentage of total fatty acids of the fraction.	74
5.1 Fatty acid composition (mg g ⁻¹) (A) Saturated, (B) Monounsaturated and (C) Polyunsaturated of three species of microalgae.	94
5.2 Shell length (APM, in µm ± SD) of <i>P. margaritifera</i> D-shape larvae at the end of Experiment 1.	96
5.3 Survival (mean ± SD) of <i>P. margaritifera</i> D-shape larvae at the end of Experiment 1.	97

5.4 Shell length (APM, in $\mu\text{m} \pm \text{SD}$) of <i>P. margaritifera</i> umbo larvae at the end of Experiment 2.	98
5.5 Survival (mean \pm SD) of <i>P. margaritifera</i> umbo larvae at the end of Experiment 2.	99
5.6 Increase in antero-posterior shell length (APM, mean \pm SD) of <i>P. margaritifera</i> D-shape larvae at the end of Experiment 3.....	101
5.7 Survival (mean \pm SD) of <i>P. margaritifera</i> D-shape larvae at the end of Experiment 3.	101
5.8 Increase in antero-posterior shell length (APM, mean \pm SD) of <i>P. margaritifera</i> umbo larvae at the end of Experiment 4.	103
5.9 Survival rate (mean \pm SD) of <i>P. margaritifera</i> umbo larvae at the end of Experiment 4.	103
6.1 Substrates for spat settlement.	117
6.2 Schematic distribution of collectors in settlement tank.	117
6.3 Shell height, hinge length and thickness (mean \pm standard deviation, n=60 for each of 8 replicates) of 55 days old <i>Pinctada margaritifera</i> spat on two types of collectores..	119
6.4 Performance of substrate for the settlement (%), and mortality (%) of <i>Pinctada margaritifera</i> spat. Settlement was calculated 10 days after introducing of pediveliger to settlement tank and post settlement mortality, at the end of experiment.	119
6.5 Mean number (\pm standard deviation, n=4 replicates) of <i>P. margaritifera</i> spat on two types of collectors at two heights.	120
7.1 Changes in mean (\pm Standard deviation, SD, n=30) shell height of <i>P. margaritifera</i> spat deployed to Lavan and Hendurabi Islands 25 (T1L, T1H), 40 (T2L, T2H), and 55 (T3L, T3H) days post settlement.	130
7.2 A: Growth curve of <i>Pinctada margaritifera</i> in the hatchery (Control). B: Scatter plot of residuals by fit values for cubic model.	131
7.3 A: Growth curve of <i>Pinctada margaritifera</i> in Hendurabi (T1H). B: Scatter plot of residuals by fit values for cubic model.	132
7.4 A: Growth curve of <i>Pinctada margaritifera</i> in Lavan (T1L). B: Scatter plot of residuals by fit values for cubic model.	133
7.5 Daily growth rate (DGR, mean \pm SD) of <i>P. margaritifera</i> spat deployed to the sea farm on 5 th September (T4) and 7 th October 2008(T5).	135

7.6 Shell heights (mean ± SD) of <i>P. margaritifera</i> spat deployed to the sea farm, Hendurabi Island, on 5 th Sep (T4) and 7 th Oct 2008 (T5) sampled at 15-day intervals.	137
7.7 Mortality rate (%) of <i>P. margaritifera</i> spat (mean ± SD) after 150 days culture in sea farm (T4 and T5) and hatchery with fixed temperature (Control 1 and 2) and adjusted temperature (Control 3 and 4).	138
7.8 Values for environmental factors (mean ± Standard error, SE) in Hendurabi from September 2008 to March 2009. A: Temperature, B: Chlorophyll-a.	139
8.1 General life cycle of pearl oyster and important results made in this study	150