



UNIVERSITI PUTRA MALAYSIA

**A STUDY ON TWO MALAYSIAN ISOLATES OF
INFECTIOUS BRONCHITIS VIRUS**

SITI SURI BT. ARSHAD

FPV 1993 6

**A STUDY ON TWO MALAYSIAN ISOLATES OF
INFECTIOUS BRONCHITIS VIRUS**

By

SITI SURI BT. ARSHAD

Thesis Submitted in Fulfilment of the Requirement for
the Degree of Master of Science in the Faculty of
Veterinary Medicine and Animal Science
Universiti Pertanian Malaysia

April 1993



Dedicated to Hafiz, Sofie, Ani and Saiful



ACKNOWLEDGEMENTS

I would like to express my utmost appreciation and gratitude to my chairman Prof. Abdul Latif Ibrahim for his invaluable guidance, discussion and suggestion throughout the course of this study.

I would also like to express my thanks to Dr. Khatijah Mohd. Yusoff and Dr. Awang IPR for being my co-supervisors for this study.

I am grateful to the Majlis Penyelidikan dan Kemajuan Sains Negara (MPKSN) for awarding me a scholarship to pursue a post-graduate study.

I would like to offer my special thanks to the Institut Penyelidikan Haiwan, Ipoh and Makmal Diagnosa, Petaling Jaya for providing me with samples of the Avian Infectious Bronchitis Virus; Hajjah Rodiah Hussin, En. Kamaruddin Awang Isa, En. Rahim Osman and En. Raziman of the Virology Section for their technical assistance; Mr. Ho Oi Kuan and Puan Aminah Jusoh of Electron Microscope Unit for helping me in the field of electron microscopy and En. Fauzi Che Yusuf for the preparation of the photographic prints.

My thanks also go to Dr. An So Hwan of Veterinary Research Institute, Anyang, Korea and Prof. Syed Naqi of Cornell University, USA, for providing me with polyclonal and monoclonal antibodies against the Avian Infectious Bronchitis Virus.

A note of thanks to my roommates, Dr. Loretta Marie Cheow and Dr. Zubaidah Mahmood for their patience and understanding on sharing a small room during the duration of my study.

My thanks are also extended to Dr. Karim Sadun Ali, Dr. Harith Ibrahim, Dr.

Kumlung, Mdm. Esther Tan and Dr. Nihayah Mohamad for willingly sharing their knowledge, experiences and problems as post-graduate students.

My thanks also go to all members of the faculty who contributed in one way or another towards the completion of my study.

Last, but not least, I wish to express my deepest gratitude to my beloved husband for his continuous love and encouragement and to my three children for their love, support, understanding and sacrifice during the period of my study.

TABLE OF CONTENTS

ACKNOWLEDGEMENTS	iii
LIST OF TABLES	ix
LIST OF FIGURES	xi
LIST OF PLATES	xii
LIST OF ABBREVIATIONS	xv
ABSTRACT	xix
ABSTRAK	xxii
CHAPTER	
I INTRODUCTION	1
II LITERATURE REVIEW	6
Avian Infectious Bronchitis	6
General Properties of Avian IBV	8
Cytopathogenicity of Avian IBV	12
Antigenic Properties of Avian IBV	20
Proteins of Avian IBV	22
III CHARACTERISATION	34
Introduction	34
Materials and Methods	35
Viruses	35
Growth of Virus in Embryonated Egg	36
Titration of Virus	37

Purification of Virus	37
Negative Contrast Electron Microscopy	38
Preparation of Erythrocytes	38
Hemagglutination Test	39
Growth of IBV in Tissue Culture	39
Preparation of Cell Culture	39
Chicken Embryo Kidney (CEK) Cell Culture	39
Vero Cell Culture	40
Replication of IBV in CEK and Vero Cell Cultures	41
Cytopathogenicity Studies	42
Hematoxylin-Eosin (H & E) Staining	42
Indirect Immunoperoxidase Studies	43
Indirect Immunofluorescence Studies	44
Acridine Orange Staining	44
Results	46
Pathogenicity of IBV in Embryonated Egg	46
Titration of Virus	49
Purification of Virus	50
Morphology of Avian IBV	52
Hemagglutination Studies	55
Growth of Viruses in Cell Culture	56
Growth of Unpurified Virus in CEK and Vero Cell Cultures	56

Growth of Purified Virus in CEK and Vero Cell Cultures	56
Growth of the IBV Adapted CEK in Vero Cell Cultures	56
Cytopathogenicity Studies	61
Hematoxylin-Eosin Staining	61
Indirect Immunoperoxidase Studies	66
Comparison of the Peroxidase Stain in the Culture Systems	66
Cross Reactivity of the Serotypes	66
Positive Peroxidase Revealed by Unadapted Virus	66
Indirect Immunofluorescence Studies	71
Acridine Orange Staining	74
Discussion	77
Summary	83
IV SEROTYPING AND PROTEIN ANALYSES	84
Introduction	84
Materials and Methods	85
Viruses	85
Antibodies	85
Serological Properties	86
Virus Neutralisation Test	86
Indirect Immunoperoxidase Studies	87
Analysis of Protein	87

Sample Preparation	87
Polyacrylamide Gel Electrophoresis (PAGE)	87
Western Blotting Analysis	89
Immunodetection Studies	91
Results	92
Neutralisation Indices	92
Cross Neutralisation Determined by IIP Studies	94
Protein Profile Studies	96
Immunodetection Studies	100
Discussion	110
Summary	115
V GENERAL DISCUSSION AND CONCLUSION	117
BIBLIOGRAPHY	123
APPENDICES	138
APPENDIX A: MEDIA	138
APPENDIX B: BUFFER	142
APPENDIX C: STAINING	148
APPENDIX D: ADDITIONAL TABLES	153
APPENDIX E: SUBSTRATE	161
APPENDIX F: MISCELLANEOUS	164
VITA	168

LIST OF TABLES

Table	Page
1 Virus Titre Determined by Egg Infective Dose 50% (EID ₅₀).	49
2 Hemagglutination of the Various IBV Strains.	55
3 Adaptability of the IBV Strains in Cell Culture.	57
4 Serum Neutralisation Test to Determine Neutralisation Indices.	93
5 Indirect Immunoperoxidase Assay of the Various IBV Strains.	95
6 Western Blot Analyses of Anti-IBV Antisera to the Nucleoprotein of IBV.	107
7 Western Blot Analyses of Anti-IBV Antisera to the Matrix Protein of IBV.	108
8 Western Blot Analyses of Anti-IBV Antisera to the Spike Protein of IBV.	109
9 Titration of M41 Virus Strain by Estimating 50% Endpoint by the Method of Reed and Muench.	154
10 Titration of H120 Virus Strain by Estimating 50% Endpoint by the Method of Reed and Muench.	154
11 Titration of H52 Virus Strain by Estimating 50% Endpoint by the Method of Reed and Muench.	155
12 Titration of PJ41 Virus Strain by Estimating 50% Endpoint by the Method of Reed and Muench.	155
13 Titration of PJ43 Virus Strain by Estimating 50% Endpoint by the Method of Reed and Muench.	156
14 Determination of NI between M41 Antigen and Isolates (PJ41 and PJ43) Antisera.	156
15 Determination of NI between H120 Antigen and Isolates (PJ41 and PJ43) and Reference (Connecticut and Arkansas) Antisera.	157
16 Determination of NI between H52 Antigen and Isolates (PJ41 and PJ43) Antisera.	157

	Page
17 Determination of NI between PJ41 Antigen and Isolate (PJ43) and References (M41, H120, H52, Connecticut and Arkansas) Antisera.	158
18 Determination of NI between PJ43 Antigen and Isolate (PJ41) and References (M41, H120, H52, Connecticut and Arkansas) Antisera.	158

x



LIST OF FIGURES

Figure		Page
1	Assembly for Electrophoretic Blotting Procedure.	90
2	Molecular Weight Determination of Virus Polypeptides.	160

LIST OF PLATES

Plate		Page
1	Photograph Showing the 18-Day-Old Normal Embryo (Left) and the Constricted, Spherical Shaped Infected Embryo (Right).	48
2	Photograph Showing the Lesion on the 16-Day-Old Normal Embryo (Left) and the Dwarfed, Infected Embryo (Right) Caused by Avian IBV.	48
3	The Sedimentation Bands (Arrows) of IBV Particles in the 20-50% (w/v) Sucrose Gradient.	51
4	Photograph Showing the Typical Coronavirus With Spikes (Arrows) Surrounded the Virion.	54
5	The Virus Particles Devoid of Spikes (Arrows) in M41 Strain Preparation.	54
6	The Unstained Control CEK Cell Culture.	59
7	The Unstained IBV Infected CEK Cell Culture Displayed a Ballooning of the Cells and Spiderweb Appearance (Arrows).	59
8	The Unstained Control Vero Cell Culture.	60
9	The Unstained IBV Infected Vero Cell Culture Displayed a Ballooning of the Cells (Arrow).	60
10	H & E Staining of Control CEK Cell Culture.	63
11	H & E Staining of IBV Infected CEK Cells at 72 Hours Post Infection.	63
12	H & E Staining of the Control Vero Cells Culture.	65
13	H & E Staining of the IBV Infected Vero Cells at 72 Hours Post Infection. Note the Early Syncytial Cells that Consist of 3 - 4 Nuclei (Arrow a).	65
14	Photograph Showing Complete Absence of Peroxidase Activity of the Uninfected CEK Cells Except for the Non-Specific Background Staining.	68

	Page
15 IIP Staining of the IBV in CEK Cells Positive Reactions Appeared as Brownish Discoloration of Cell Clumps (Arrow).	68
16 The Control Vero Cells Showing the Complete Absence of Peroxidase Activity Except for Non-Specific Background Staining.	70
17 IIP Staining of the IBV Infected Vero Cell. Note the Brownish Coloration (Arrow) in the Cytoplasm of the Infected Cells.	70
18 IIF Staining of the IBV Infected Vero Cells Showing the Shiny Apple Green Discoloration of the Cytoplasm (Arrow a) and Darkly Stained Nuclei (Arrow b).	73
19 AO Staining of the Control Vero Cells.	76
20 AO Staining of the IBV Infected Vero Cells 36 Hours Post Infection Showing the Red-Flame Discoloration of the Cytoplasm (Arrow).	76
21 Comparison of Coomasie Blue Stained Protein Profiles of Three Reference Strains and Two Local Isolates of Avian IBV by SDS-PAGE. Lane 1 Molecular Markers; Lane 2 Reference M41; Lane 3 Reference H120; Lane 4 Reference H52; Lane 5 Isolate PJ41; Lane 6 Isolate PJ43; Lane 7 Unpurified PJ41; Lane 8 Unpurified PJ43. Note the Presence of Only One Band (Arrow) in All the Purified IBV Strains Compared to Unpurified IBV.	98
22 Comparison of Silver Stained Protein Profiles of Three Reference Strains and Two Local Isolates of Avian IBV by SDS-PAGE. Lane 1 Molecular Markers; Lane 2 Reference M41; Lane 3 Reference H120; Lane 4 Reference H52; Lane 5 Isolate PJ41; Lane 6 Isolate PJ43. Note the Similar Migration Pattern of All the IBV Strains Showing Eight Bands (Arrows) Sedimented at 91 kd, 87 kd, 74 kd, 64 kd, 58 kd, 53 kd 33 kd and 81 kd.	99

	Page	
23	Western Blot of Three Reference Strains and Two Local Isolates of Avian IBV Detected by Polyclonal Antibodies (A) of Massachusetts, Arkansas and Connecticut serotypes, H52 and H120, PJ 41 Isolate. Lane 1 Molecular Markers; Lane 2 Reference M41; Lane 3 Reference H120; Lane 4 Reference H52; Lane 5 Isolate PJ41; Lane 6 Isolate PJ43. Note the Similar Migration Pattern of All the IBV Strains Showing Three Bands (Arrows) Sedimented at 87 kd, 58 kd and 33 kd.	102
24	Western Blot of Three Reference Strains and Two Local Isolates of Avian IBV Detected by Polyclonal Antibodies (B) of M 41 and PJ 43 Isolate. Lane 1 Molecular Markers; Lane 2 reference M41; Lane 3 Reference H120; Lane 4 Reference H52; Lane 5 Isolate PJ41; Lane 6 Isolate PJ43. Note the Presence of Only One Band (Arrow) in All the IBV Strains.	103
25	Western Blot of Three Reference Strains and Two Local Isolates of Avian IBV Detected by ND virus Antisera. Lane 1 Molecular Markers; Lane 2 Reference M41; Lane 3 Reference H120; Lane 4 Reference H52; Lane 5 Isolate PJ41; Lane 6 Isolate PJ43. Note the Absent of Band Indicating No Cross Reaction Between the IBV and NDV Virus Groups.	104
26	Western Blot of Three Reference Strains and Two Local Isolates of Avian IBV Detected by Group Specific Monoclonal Antibodies of Avian IBV. Lane 1 Molecular Markers; Lane 2 Reference M41; Lane 3 Reference H120; Lane 4 Reference H52; Lane 5 Isolate PJ41; Lane 6 Isolate PJ43. Note the Presence of Only One Band (Arrow) in All the IBV Strains.	105

LIST OF ABBREVIATIONS

AGPT	Agar Gel Precipitation Test
AO	Acridine Orange
Ark	Arkansas
ATV	Antibiotic-Trypsin-Versene
BSA	Bovine Serum Albumin
BHK	Cell line Derived from Baby Hamster Kidney
°C	Degree Celcius
CEK	Chicken Embryo Kidney
CELi	Chicken Embryo Liver
CELO	Chick Embryo Lethal Orphan
CELu	Chicken Embryo Lung
CK	Chicken Kidney
cm ³	Cubic Centimeter
Conn	Connecticut
cpe	Cytopathic Effect
CRD	Chronic Respiratory Disease
CsCl	Cesium Chloride
DME	Dulbecco's Modified Eagle Medium
DNA	Deoxyribonucleic Acid
Dr.	Doctor
D.W	Distilled water

EID₅₀	Egg Infective Dose 50 Percent
ELISA	Enzyme-Linked Immunosorbent Assay
g	Gravity
gm	Gram
H & E	Hematoxylin-Eosin
H120	Holland 120
H52	Holland 52
HA	Hemagglutination
HI	Hemagglutination Inhibition
HRP	Horse-Radish Peroxidase
HCV	Human Coronavirus
IB	Infectious Bronchitis
IBD	Infectious Bursal Disease
IBV	Infectious Bronchitis Virus
IIF	Indirect Immunofluorescent
IgG	Immunoglobulin G
IIP	Indirect Immunoperoxidase
IP	Immunoperoxidase
IPH	Institut Penyelidikan Haiwan
kd	Kilodalton
M	Matrix
M	Molar
MDPJ	Makmal Diagnosa Petaling Jaya

M41	Massachusetts 41
mA	Miliampere
Mass	Massachusetts
MHV	Murine Hepatitis Virus
ml	Mililitre
mm	Milimeter
mol. wt.	Molecular Weight
ND	Newcastle Disease
NDV	Newcastle Disease Virus
NI	Neutralisation Index
nm	Nanometer
N	Nucleoprotein
NTE	Sodium Tris EDTA buffer
OD	Optical Density
PAGE	Polyacrylamide Gel Electrophoresis
PBS	Phosphate Buffer Saline
PfU	Plaque Forming Unit
PJ	Petaling Jaya
PK 15	Cell line Derived from Pig Kidney
PTA	Phosphotungsten Acid
RNA	Ribonucleic Acid
S	Spike
SDS	Sodium dodecyl sulfate

TBS	Tween Buffer Saline
TCID ₅₀	Tissue Culture Infective Dose 50 Percent
TGEV	Transmissible Gastroenteritis Virus
<i>ug</i>	Microgram
UK	United Kingdom
<i>ul</i>	Microliter
USA	United States of America
<i>um</i>	Micrometer
Vero	Cell line Derived from Kidney Tissue of Green African Monkey
VN	Virus Neutralisation
v/v	Volume to volume
w/v	Weight to Volume

Abstract of thesis submitted to the Senate of Universiti Pertanian Malaysia
in fulfilment of the requirements for the degree of Master of Science.

**A STUDY ON TWO MALAYSIAN ISOLATES OF INFECTIOUS BRONCHITIS
VIRUS**

By

SITI SURI ARSHAD

April 1993

Chairman: Prof. Abdul Latif Ibrahim

Faculty : Veterinary Medicine and Animal Science

This research was carried out to investigate whether the outbreak of Infectious Bronchitis (IB) in Malaysia is due to the vaccine virus or the emergence of a new serotype which is different from the vaccine strains. The study involves the comparison between the two local isolates (PJ41 and PJ43) and the three reference strains (H120, H52 and M41) of Massachusetts derivative.

Initial characterisation of the two isolates involved a study on their basic properties such as morphologies, hemagglutinating activity, pathogenicity in embryonated chicken eggs and their adaptability in primary and secondary tissue cultures with the aid of Acridine Orange (AO), indirect immunoperoxidase (IIP) and indirect immunofluorescent (IIF) staining (to compare localisation of the RNA) and the Hematoxylin and Eosin staining (H & E) (to compare morphologic changes in cell culture).

Both isolates and reference strains revealed a corona-like appearance under electron microscopy and formed pathognomonic lesions of toe curling and dwarfing of the embryo. The virus strains were indistinguishable in their ability to produce cytopathic effects in both primary and secondary cell cultures. The AO staining indicated that the virus possesses a RNA. IIP and IIF tests indicated that they are located in the cytoplasm of the cell. The hemagglutinating activity however, showed that this activity is not always their properties when one of the isolates, (PJ43), and one of the reference strains, (H52), were unable to agglutinate erythrocytes.

The results showed that the local isolates have many of the properties of IBV reference strains. The isolates and the reference strains were then serotyped and their proteins were analysed. Neutralisation of the isolates and the reference strain were seen with the homologous antisera and not with the heterologous antisera in the embryonated chicken egg. In contrast, staining of the viral protein was seen with both homologous and heterologous antisera in both the IIP test as well as the Western Blot analyses. The latter test showed that IBV possesses three structural proteins whereby the matrix (M) being a conserved protein.

In conclusion, this is the first detailed study conducted on local isolates of IBV in Malaysia. The study shows that they are related to the Massachusetts strain and hence the hypothesis stated for this study is proven true. However, it cannot be differentiated either from vaccine strain or the field strain unless a detail study on the amino acid content on the spike protein could be carry out. The study has also provided more information on the characteristics of local isolates and the

relationship of these isolates with the Massachusetts field strain and the vaccine strain. Such an information is very important in the planning of any control program. The success of any vaccination lies on the ability to fully characterise the isolate and to select an appropriate vaccine strain.

Abstrak daripada tesis yang dikemukakan kepada Senat Universiti Pertanian Malaysia bagi memenuhi keperluan Ijazah Master Sains.

KAJIAN KE ATAS DUA ISOLAT VIRUS BRONKITIS BERJANGKIT MALAYSIA

Oleh

SITI SURI BT ARSHAD

April 1993

Pengerusi: Professor Abdul Latif Ibrahim

Fakulti : Kedoktoran Veterinar dan Sains Peternakan

Penyelidikan ini dijalankan untuk menyiasat samada wabak Bronkitis Berjangkit (IB) di Malaysia disebabkan oleh vaksin virus atau kehadiran serotip baru yang berbeza dari strain vaksin. Kajian ini melibatkan perbandingan di antara dua isolat tempatan (PJ41 dan PJ43) dengan tiga strain rujukan (H120, H52 dan M41) yang berasal dari rentetan Massachusetts.

Pencirian terawal dua isolat ini melibatkan kajian ke atas sifat khas seperti morfologi, aktiviti pengaglutinatan, kepatogenan dalam telur ayam berembrio dan penyesuaianya dalam kultur tisu primer dan sekunder dengan bantuan pewarnaan akridina jingga, imunoperoksidase tak langsung (IIP) dan imunopendarfluor tak langsung (IIF) (untuk perbandingan kedudukan RNA) dan pewarnaan Hematoksilin-Eosin (H&E) (untuk perbandingan perubahan morfologi dalam kultur sel).

Kedua-dua isolat dan strain rujukan menunjukkan rupa bak korona di bawah mikroskopi elektron dan lesi yang patognomonik iaitu lentikan jari kaki dan kerencatan embrio. Virus-virus tersebut tidak dapat dibezakan melalui keupayaan

dalam menghasilkan kesan sitopati pada kedua-dua sel primer dan sekunder. Pewarnaan akridina jingga membuktikan bahawa virus tersebut adalah RNA dan ujian IIP dan IIF menunjukkan kedudukan virus dalam sitoplasma sel. Sifat pengaglutinatan bagaimanapun menunjukkan bahawa ianya tidak semestinya sifat khas apabila salah satu isolat (PJ43) dan strain rujukan (H52)-tidak berupaya untuk mengaglutinat eritrosit.

Keputusan kajian menunjukkan bahawa isolat tempatan mempunyai banyak sifat khas strain rujukan IBV. Isolat dan strain rujukan kemudiannya diserotip dan proteininya dianalisis. Peneutralan ke atas isolat dan strain rujukan dapat dilihat dengan antiserum homologus dan tidak dapat dilihat dengan antiserum heterologus dalam telur ayam berembrio. Sebaliknya, pewarnaan protein virus dapat dilihat dengan kedua-dua antiserum homologus dan heterologus dalam ujian IIP dan analisis Western Blot. Analisis Western Blot menunjukkan IBV mempunyai tiga protein struktur di mana matriknya (M) adalah protein kekal.

Kesimpulannya, kajian ini merupakan satu kajian terperinci yang pertama yang pernah dilaksanakan di Malaysia terhadap isolat IBV tempatan. Pengkajian menunjukkan bahawa isolat tersebut adalah berkaitan dengan strain Massachusetts dan dengan itu membuktikan bahawa hipotesis bagi kajian ini adalah benar. Walaubagaimanapun, ianya tidak dapat dibezakan samada dari strain vaksin atau strain lapangan sehingga kajian terperinci keatas kandungan acid amino dalam protein spikanya dapat dijalankan. Pengkajian ini juga memberi lebih maklumat tentang ciri isolat tempatan dan perkaitannya dengan strain lapangan Massachusetts dan strain vaksin. Maklumat seumpama ini sangat berguna dalam

perancangan sebarang program pengawalan. Kejayaan sesuatu pemvaksinan bergantung kepada kebolehan dalam mencirikan keseluruhan isolat dan dalam memilih strain vaksin yang bersesuaian.