



**INCORPORATION OF TRANSGENIC *Nannochloropsis* sp. INTO FISH FEED
AS VACCINE DELIVERY SYSTEM AGAINST VIBRIOSIS**

By

NOR IZZATI HUSNA BINTI NOOR HISHAM

**Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfilment of the
Requirements for the Degree of Master of Science**

December 2023

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of
the requirement for the degree of Master of Science

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December 2023

Chairman : Zetty Norhana Balia Yusof, Ph.D.
Faculty : Biotechnology and Biomolecular Sciences

Oral vaccination is one of the potential methods for preventing disease outbreaks, and this method is easier and causes less stress to the fish. However, there is a limitation for oral vaccination which the antigens were easily degraded due to the high acidity of the fish gut. To overcome this limitation, microalgae were used as a vaccine delivery system which it could provide antigen protection from being degraded. Thus, this study aims to evaluate the efficacy of oral vaccines on zebrafish, used as a host model against vibriosis, by incorporating transgenic *Nannochloropsis* sp. into fish feed. In this research, *Nannochloropsis* sp. was transformed with a designated vector (VCP_OmpK_ZA1) harbouring *OmpK* gene fragment, a gene coding for antigenic property protein from *Vibrio harveyi*. 10% of the verified transgenic *Nannochloropsis* sp. were mixed with commercial fish feed (TL+CP). Adult zebrafish were fed with the formulated fish feed for 5 consecutive days in week 1, and the booster given in week 3. For the control groups, zebrafish were fed with 10% wild-type *Nannochloropsis* sp.-formulated feed (WT+CP) and commercial feed (CP). The feeding trials were performed until week 6. The zebrafish then were evaluated through weekly analysis expression of genes via RT-qPCR, and the relative percentage of survival (RPS) and histopathology was analyzed upon bacterial challenge with *V. harveyi* in week 6. Based on the result obtained, the *IgZ* gene was significantly upregulated by 4-fold after 8 days post-first dose of vaccination while *TNF α* and *IL-1 β* genes were slightly upregulated after 1 day post-second dose of vaccination by 1.4-fold and 1.6-fold respectively. The RPS of the vaccinated fish (TL+CP) and unvaccinated fish (CP+WT) were 83% and 42%, respectively. For the histopathology analysis, the unvaccinated fish (CP and CP+WT) had severe histopathological changes and vaccinated fish (CP+TL) showed minor histopathological changes. This study has demonstrated that the transgenic microalgae-formulated feed has the potential to stimulate the immune response of the fish.

Keywords: Fish feed, *Nannochloropsis* sp., Oral vaccination, Transgenic, Vibriosis

SDG: GOAL 14: Life Below Water



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENAMBAHAN *Nannochloropsis* sp. TRANSGENIK KE DALAM MAKANAN IKAN SEBAGAI SISTEM PENYAMPAIAN VAKSIN UNTUK MELAWAN VIBRIOSIS

Oleh

NOR IZZATI HUSNA BINTI NOOR HISHAM

Disember 2023

Pengerusi : Zetty Norhana Balia Yusof, Ph.D.
Fakulti : Bioteknologi dan Sains Biomolekul

Pemvaksinan oral adalah salah satu kaedah yang berpotensi untuk mencegah wabak penyakit, dan kaedah ini lebih mudah dan mengurangkan tekanan kepada ikan. Walau bagaimanapun, terdapat had untuk vaksinasi oral yang mana antigennya mudah terdegradasi kerana keasidan yang tinggi di dalam usus ikan. Untuk mengatasi had ini, mikroalga digunakan sebagai sistem penyampaian vaksin yang boleh memberikan perlindungan kepada antigen daripada terdegradasi. Oleh itu, kajian ini bertujuan untuk menilai keberkesanan vaksin oral pada ikan zebra yang digunakan sebagai model haiwan untuk melawan penyakit vibriosis, dengan mencampurkan transgenik *Nannochloropsis* sp. ke dalam makanan ikan. Dalam penyelidikan ini, *Nannochloropsis* sp. telah diubah dengan vektor (VCP_OmpK_ZA1) yang membawa serpihan gen *OmpK*, gen pengekodan protein antigen daripada *Vibrio harveyi*. 10% daripada *Nannochloropsis* sp. transgenik yang disahkan dicampur dengan makanan ikan komersial (TL+CP). Ikan zebra dewasa diberi makan dengan makanan ikan yang diformulasikan ini selama 5 hari berturut-turut pada minggu pertama, dan vaksin penggalak diberikan pada minggu ketiga. Bagi kumpulan kawalan, ikan zebra diberi makan dengan 10% makanan rumusan *Nannochloropsis* sp.- jenis liar (WT+CP) dan makanan ikan komersial (CP). Eksperimen pemberian makanan ini berlangsung sehingga minggu ke-6. Ikan zebra kemudiannya dinilai menerusi ekspresi analisis gen melalui RT-qPCR pada setiap minggu, kemandirian peratus relatif (KPR) dan histopatologi pula dinilai setelah menjalani cabaran bakteria dengan *V. harveyi* pada minggu ke-6. Berdasarkan keputusan yang diperoleh, gen IgZ meningkat dengan ketara sebanyak 4 kali ganda selepas 8 hari selepas dos vaksinasi yang pertama. Bagi gen *TNF α* dan *IL-1 β* turut meningkat sebanyak 1.4 kali ganda dan 1.6 kali ganda masing-masing selepas 1 hari diberikan dos penggalak. KPR ikan yang divaksin (TL+CP) dan ikan yang tidak divaksinasi (CP+WT) masing-masing adalah 83% dan 42%. Untuk analisis histopatologi, ikan yang tidak divaksinasi (CP dan CP+WT) mengalami perubahan histopatologi yang teruk dan ikan yang divaksin (CP+TL) menunjukkan perubahan histopatologi yang masih terkawal. Kajian ini telah menunjukkan bahawa

makanan yang dirumuskan oleh mikroalga transgenik berpotensi untuk merangsang tindak balas imun ikan.

Kata Kunci: Makanan Ikan, *Nannochloropsis* sp., Transgenik, Vibriosis, Vaksinasi oral

SDG: GOL 14: Hidupan bawah air



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Zetty Norhana Binti Balia Yusof, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Chong Chou Min, PhD

Senior Lecturer

Faculty of Agriculture

Universiti Putra Malaysia

(Member)

Ina Salwany Binti Mohd Yasin, PhD

Associate Professor

Institute of Bioscience

Universiti Putra Malaysia

(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 16 May 2024

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LIST OF ABBREVIATIONS

%	Percentage
°C	Degree Celsius
A	Absorbance
Bp	Base pair
cDNA	Complementary DNA
Cells/ml	Cells per millilitre
Chl	Chlorophyll
DNA	Deoxyribonucleic acid
dNTP	Deoxyribonucleotide triphosphate
<i>et al.</i> ,	And friends
EtBr	Ethidium bromide
F, R	Forward, Reverse
FAO	Food and Agriculture Organization
g	Gram
h	Hours
L	Litre
min	Minutes
ml	Milliliter
NaCl	Sodium chloride
PCR	Polymerase chain reaction
qPCR	Quantitative PCR
Rpm	Revolutions per minute
rRNA	Ribosomal ribonucleic acid

RT-PCR	Reverse-transcriptase PCR
s	Seconds
sp.	Species
UPM	Universiti Putra Malaysia
UV-Vis	Ultraviolet-visible
V	Volt
µg	Microgram
µl	Microlitre



CHAPTER 1

INTRODUCTION

1.1 Background Study

In Malaysia, the total aquaculture production in 2021 was 1.75 million tonnes which comprises 1.33 million tonnes of fish captured and 417 187 million of farmed fish (Department of Fisheries Malaysia [DOF], 2022). Other than that, in 2021, 24 million production of aquatic plants and 242 million ornamental fish were recorded. The total sales recorded for all of the aquaculture productions were approximately RM 14.8 billion (DOF, 2022). This shows that aquaculture production in Malaysia is significant and profitable and this sector needs to be sustained throughout the year, especially for food production.

In the fish farm, there is a higher possibility for infectious diseases to occur as the fish are more susceptible to bacterial pathogens due to the higher density of the farmed fish and the limitation of using clean water (Watts *et al.*, 2017). The common bacterial pathogens that cause fish disease are *Vibrio*, *Edwardsiella*, *Aeromonas*, and *Streptococcus* (Sudheesh *et al.*, 2012; Wamala *et al.*, 2018). *Vibrio* spp. is a significant pathogenic microbe that can cause severe impacts on the health of both animals and humans. The outbreak of vibriosis disease raises concerns, especially when it affects a large number of fish, particularly in fish farms, as it can cause mass mortality, leading to huge losses in the aquatic industry.

Antibiotics have been widely introduced for controlling diseases; however, it is difficult to determine the suitable amount to completely diminish the pathogens. Furthermore, extensive use of antibiotics will result in the development of antibiotic-resistant bacteria which can cause harm to humans, as well as the environment. In Peninsular Malaysia, tetracycline is one of the common antibiotics used to treat and prevent disease outbreaks in aquatic animals. A study by Thiang *et al.* (2022), detected low concentrations of tetracycline residues in five farms in the region, suggesting its usage in the area. Additionally, the study also reported that out of 93 different bacteria types, 48 were found to carry the *tet* gene, including *Enterobacter*, *Vibrio*, *Photobacterium*, and *Pseudoalteromonas*. The presence of tetracycline residues and the emergence of bacteria carrying the *tet* gene pose risks to aquatic plants, such as algae. These findings suggest that the use of antibiotics may have contributed to the development of antibiotic-resistant pathogens. Therefore, the utilization of antibiotics for treating or preventing disease outbreaks in aquatic animals needs to be carefully regulated and monitored.

Vaccination is another strategy for treating bacterial and viral diseases. Vaccination works by stimulating the immune system of the fish to produce antibodies against the pathogen which will protect the fish from the pathogen as well as prevent the disease from recurring (Mondal and Thomas, 2022). Most commercial vaccines are made of inactivated bacterial pathogens, and they are introduced into the fish through injection (Adam *et al.* 2021, Du *et al.* 2022) However, this practice is costly and reduces the vaccine efficacy as it introduces stress to the fish.

Recently, an alternative was developed by introducing genetically modifying microalgae *Nannochloropsis* sp. into a vaccine carrier to vaccinate the fish orally (Abidin *et al.*, 2020b; Abidin *et al.*, 2021a). This invention is known to be a promising technology that could become one of the practical methods for vaccinating a larger amount of fish or small-sized fish, helping to reduce the use of antibiotics in aquatic animals. Other than that, the features of the microalgae itself are potential for vaccine candidates as they have rigid cell walls which can protect the antigen from easily degraded and improve the efficiency of the vaccine effects on fish (Kwon *et al.*, 2013; Specht & Mayfield, 2014). Also, *Nannochloropsis* sp. plays a significant role in nutrition enrichment for aquatic animals as they have high lipid content and high levels of eicosapentaenoic fatty acid (Guimarães *et al.*, 2021). Thus, by utilizing transgenic *Nannochloropsis* sp. as an oral vaccine for the fish, they will provide protection as well as nutrition to the fish. However, studies on using the microalgae such as *Nannochloropsis* sp. as a vaccine delivery system for vaccinating the fish via oral route are still lacking.

1.2 Problem Statements

The problem statements in this research study were:

1. Outbreak of the diseases will cause a huge loss of aquaculture production.
2. A long-term treatment with an antibiotic will lead to the development of antibiotic- resistance bacteria.
3. Oral vaccination has low efficiency as the oral vaccine tends to degrade in the fish gut due to high acidity.
4. Lack of studies related to the utilization of the microalgae as a vaccine delivery system for fish

1.3 Objective

The research on the development of transgenic *Nannochloropsis* sp. by Abidin *et al.* in 2021a had not been tested on fish. Therefore, this study aimed to continue exploring the effectiveness of the newly developed vaccine. The primary objective was to evaluate the efficacy of oral vaccines on zebrafish, used as a host model against vibriosis, by incorporating transgenic *Nannochloropsis* sp. into their feed.

The specific objectives for this research study were:

1. To transform *Nannochloropsis* sp. with vector VCP_OMP_K_ZA1 harbouring OmpK gene fragment.
2. To formulate a feed-based oral vaccine and evaluate the efficiency of the transgenic *Nannochloropsis* sp. – formulated feed in inducing an immune response in zebrafish through gene expression analysis (Rt-qPCR), relative percentage survival (RPS) evaluation and histopathological changes.



REFERENCES

- Abdul Hannan, M. D., Mahbubur Rahman, M. D., Nurunnabi Mondal, M. D., Suzan Chandra, D., Chowdhury, G., & Tofazzal Islam, M. D. (2019). Molecular Identification of *Vibrio alginolyticus* Causing Vibriosis in Shrimp and Its Herbal Remedy. *Polish Journal of Microbiology*, 68(4), 429–438. <https://doi.org/10.33073/pjm-2019-042>
- Abidin, A. A. Z., Suntarajh, M., & Yusof, Z. N. B. (2020a). Microalgae as a vaccine delivery system to aquatic organism. In M. A. Alam, J. Xu, & Z. Wang (Eds.), *Microalgae Biotechnology For Food, Health and High Value Products*, 353–372. Springer. <https://doi.org/10.1007/978-981-15-0169-2>
- Abidin, A. A. Z., Suntarajh, M., & Yusof, Z. N. B. (2020b). Transformation of a Malaysian species of *Nannochloropsis*: Gateway to construction of transgenic microalgae as vaccine delivery system to aquatic organisms. *Bioengineered*, 11(1), 1071–1079. <https://doi.org/10.1080/21655979.2020.1822106>
- Abidin, A. A. Z., Othman, N. A., Yusoff, F. M., & Yusof, Z. N. B. (2021a). Determination of transgene stability in *Nannochloropsis* sp. transformed with immunogenic peptide for oral vaccination against vibriosis. *Aquaculture International*, 29(2), 477–486. <http://dx.doi.org/10.1007/s10499-020-00634-w>
- Abidin, A. A. Z., Yokthongwattana, C., & Yusof, Z. N. B. (2021b). Carotenogenesis in *Nannochloropsis oculata* under salinity and oxidative stress. *Sains Malaysiana*, 50(2), 327–337. <http://dx.doi.org/10.17576/jsm-2021-5002-05>
- Adams, A., & Subasinghe, R. (2021). Fish vaccines. *Veterinary Vaccines*, 113–118. <https://doi.org/10.1002/9781119506287.ch9>
- Akbar, I., Radhakrishnan, D. K., Venkatachalam, R., Sathrajith, A. T., & S, S. (2014). Standartization of the bioencapsulation of probiotics and oil emulsion in *Artemia parthenogenetica*. *International Journal of Research in Fisheries and Aquaculture*, 4(3), 122–125.
- Amalina, N. Z., Santha, S., Zulperi, D., Amal, M. N., Yusof, M. T., Zamri-Saad, M., & Ina-Salwany, M. Y. (2019). Prevalence, antimicrobial susceptibility and plasmid profiling of *Vibrio* spp. isolated from cultured groupers in Peninsular Malaysia. *BMC Microbiology*, 19(1). <https://doi.org/10.1186/s12866-019-1624-2>
- Arredondo, P., & Arciniega, G. M. (2001). Strategies and Techniques for Counselor Training Based on the Multicultural Counseling Competencies. *Journal of Multicultural Counseling and Development*, 29(4), 263–273. <https://doi.org/10.1002/j.2161-1912.2001.tb00469.x>
- Assefa A., & Abunna, F. (2018). Maintenance of fish health in aquaculture: Review of epidemiological approaches for prevention and control of infectious disease of fish. *Veterinary Medicine International*, 2018, 5432497. <https://doi.org/10.1155/2018/5432497>

- Atujona, D., Huang, Y., Wang, Z., Jian, J., & Cai, S. (2019). *Vibrio harveyi* (VirB11) recombinant vaccine development against vibriosis in orange-spotted grouper (*Epinephelus coioides*). *Aquaculture Research*, 50(9), 2628-2634. <http://dx.doi.org/10.1111/are.14220>
- Baker-Austin, C., Oliver, J. D., Alam, M., Ali, A., Waldor, M. K., Qadri, F., & Martinez-Urtaza, J. (2018). *Vibrio* spp. infections. *Nature Reviews Disease Primers*, 4(1), 1–19. <https://doi.org/10.1038/s41572-018-0005-8>
- Behera, T., & Swain, P. (2014). Antigen encapsulated alginate-coated chitosan microspheres stimulate both innate and adaptive immune responses in fish through oral immunization. *Aquaculture International*, 22(2), 673-688. <https://doi.org/10.1007/s10499-013-9696-8>
- Bøgwald, J., & Dalmo, R. A. (2019). Review on Immersion Vaccines for Fish: An Update 2019. *Microorganisms*, 7(12), 627. <https://doi.org/10.3390/microorganisms7120627>
- Camacho, F., Macedo, A., & Malcata, F. (2019). Potential industrial applications and commercialization of microalgae in the functional food and feed industries: A short review. *Marine Drugs*, 17(6), 312. <https://doi.org/10.3390/md17060312>
- Charoonnart, P., Purton, S., & Saksmerprome, V. (2018). Applications of microalgae biotechnology for disease control in aquaculture. *Biology*, 7(2), 24. <https://doi.org/10.3390/biology7020024>
- Charoonnart, P., Worakajit, N., Zedler, J. A., Meetam, M., Robinson, C., & Saksmerprome, V. (2019). Generation of microalga *Chlamydomonas reinhardtii* expressing shrimp antiviral dsRNA without supplementation of antibiotics. *Scientific Reports*, 9(1), 3164. <https://doi.org/10.1038/s41598-019-39539-x>
- Chen, K., Wang, C. Q., Fan, Y. Q., Xie, Y. S., Yin, Z. F., Xu, Z. J., Zhang, H. L., Cao, J. T., Han, Z. H., Wang, Y., & Song, D. Q. (2014). The evaluation of rapid cooling as an anesthetic method for the zebrafish. *Zebrafish*, 11(1), 71–75. <https://doi.org/10.1089/zeb.2012.0858>
- Chen, J., Ghorai, M. K., Kenney, G., & Stubbe, J. (2008). Mechanistic studies on bleomycin-mediated DNA damage: multiple binding modes can result in double-stranded DNA cleavage. *Nucleic Acids Research*, 36(11), 3781–3790. <https://doi.org/10.1093/nar/gkn302>
- Chin, Y. K., Al-saari, N., Zulperi, Z., Mohd-Aris, A., Salleh, A., Silvaraj, S., Mohamad, A., Lee, J., Zamri-Saad, M., & Ina-Salwany, M. Y. (2019). Efficacy of bath vaccination with a live attenuated *Vibrio harveyi* against vibriosis in Asian seabass fingerling, *Lates calcarifer*. *Aquaculture Research*, 51(1), 389–399. <https://doi.org/10.1111/are.14386>
- Dadar, M., Dhama, K., Vakharia, V. N., Hoseinifar, S. H., Karthik, K., Tiwari, R., Khandia, R., Munjal, A., Salgado-Miranda, C., & Joshi, S. K. (2017). Advances

- in aquaculture vaccines against fish pathogens: Global status and current trends. *Reviews in Fisheries Science and Aquaculture*, 25(3), 184–217. <https://doi.org/10.1080/23308249.2016.1261277>
- Dallaire-Dufresne, S., Tanaka, K. H., Trudel, M. V., Lafaille, A., & Charette, S. J. (2014). Virulence, genomic features, and plasticity of *Aeromonas salmonicida* subsp. *salmonicida*, the causative agent of fish furunculosis. *Veterinary Microbiology*, 169(1–2), 1–7. <https://doi.org/10.1016/j.vetmic.2013.06.025>
- Defoirdt, T. (2013). Virulence mechanisms of bacterial aquaculture pathogens and antivirulence therapy for aquaculture. *Reviews in Aquaculture*, 6(2), 100–114. <https://doi.org/10.1111/raq.12030>
- Defoirdt, T., Sorgeloos, P., & Bossier, P. (2011). Alternatives to antibiotics for the control of bacterial disease in Aquaculture. *Current Opinion in Microbiology*, 14(3), 251–258. <https://doi.org/10.1016/j.mib.2011.03.004>
- De Marco, G., Cappello, T., & Maisano, M. (2023). Histomorphological changes in fish gut in response to prebiotics and probiotics treatment to improve their health status: A Review. *Animals*, 13(18), 2860. <https://doi.org/10.3390/ani13182860>
- Deng, Y., Zhang, Y., Chen, H., Xu, L., Wang, Q., & Feng, J. (2020). Gut–Liver immune response and gut microbiota profiling reveal the pathogenic mechanisms of *Vibrio harveyi* in Pearl Gentian Grouper (*Epinephelus lanceolatus*♂ × *E. fuscoguttatus*♀). *Frontiers in Immunology*, 11. <https://doi.org/10.3389/fimmu.2020.607754>
- Department of Fisheries. (2022). *Annual Reports 2021*. DOF. <https://www.dof.gov.my/en/resources/i-extension-en/annual-report/#>
- Dey, A., Ghosh, K., & Hazra, N. (2015). An overview on bioencapsulation of live food organisms with probiotics for better growth and survival of freshwater fish juveniles. *International Journal of Research in Fisheries and Aquacultures*, 5(2), 74–83.
- Dong, H. T., Taengphu, S., Sangsuriya, P., Charoensapsri, W., Phiwsaiya, K., Sornwatana, T., Khunrae, P., Rattanarojpong, T., & Senapin, S. (2017). Recovery of *Vibrio harveyi* from scale drop and muscle necrosis disease in farmed barramundi, *lates calcarifer* in Vietnam. *Aquaculture*, 473, 89–96. <https://doi.org/10.1016/j.aquaculture.2017.02.005>
- Du, Y., Hu, X., Miao, L., & Chen, J. (2022). Current status and development prospects of aquatic vaccines. *Frontiers in Immunology*, 13. <https://doi.org/10.3389/fimmu.2022.1040336>
- Fathi, S., Harun, A. N., Rambat, S., & Tukiran, N. A. (2018). Current issues in Aquaculture: Lessons from Malaysia. *Advanced Science Letters*, 24(1), 503–505. <https://doi.org/10.1166/asl.2018.12051>

- Feng, S., Feng, W., Zhao, L., Gu, H., Li, Q., Shi, K., Guo, S., & Zhang, N. (2014). Preparation of transgenic *Dunaliella salina* for immunization against white spot syndrome virus in crayfish. *Archives of Virology*, 159, 519–525. <https://doi.org/10.1007/s00705-013-1856-7>
- Ferdous, U. T., & Yusof, Z. N. B. (2021). Medicinal prospects of antioxidants from algal sources in cancer therapy. *Frontiers in Pharmacology*, 12, 593116. <https://doi.org/10.3389/fphar.2021.593116>
- Ferrer-Ledo, N., Stegemüller, L., Janssen, M., Wijffels, R. H., & Barbosa, M. J. (2023). Growth and fatty acid distribution over lipid classes in *Nannochloropsis Oceanica* acclimated to different temperatures. *Frontiers in Plant Science*, 14. <https://doi.org/10.3389/fpls.2023.1078998>
- Firdaus-Nawi, M., Yusoff, S.M., Yusof, H., Abdullah, S.-Z. & Zamri-Saad, M. (2013). Efficacy of feed-based adjuvant vaccine against *Streptococcus agalactiae* in *Oreochromis* spp. in Malaysia. *Aquaculture Research*, 45(1), 87-96. <https://doi.org/10.1111/j.1365-2109.2012.03207.x>
- Finney, D. J. (1952). Probit analysis: A statistical treatment of the sigmoid response curve (2nd ed.). Cambridge University Press.
- Fisheries Research Institute (FRI). 2017. The state of the aquatic resources. Department of Fisheries, Malaysia.
- Guimarães, A. M., Guertler, C., do Vale Pereira, G., da Rosa Coelho, J., Costa Rezende, P., Nóbrega, R. O., & do Nascimento Vieira, F. (2021). *Nannochloropsis* spp. as feed additive for the Pacific white shrimp: effect on midgut microbiology, thermal shock resistance and immunology. *Animals*, 11(1), 150. <https://doi.org/10.3390/ani11010150>
- Gong, H., Wang, Q., Lai, Y., Zhao, C., Sun, C., Chen, Z., Tao, J., & Huang, Z. (2021). Study on immune response of organs of *epinephelus coioides* and *carassius auratus* after immersion vaccination with inactivated *Vibrio harveyi* vaccine. *Frontiers in Immunology*, 11. <https://doi.org/10.3389/fimmu.2020.622387>
- He, Y., Peng, H., Liu, J., Chen, F., Zhou, Y., & Chen, H. W. (2017). *Chlorella* sp. transgenic with Scy-hepc enhancing the survival of *Sparus macrocephalus* and hybrid grouper challenged with *Aeromonas hydrophila*. *Fish & Shellfish Immunology*, 73, 22-29. <https://doi.org/10.1016/j.fsi.2017.11.051>
- Hegde, A., Kabra, S., Basawa, R.M. et al., (2023) Bacterial diseases in marine fish species: current trends and future prospects in disease management. *World J Microbiol Biotechnol* 39, 317. <https://doi.org/10.1007/s11274-023-03755-5>
- Hibberd, D. J. (1981). Notes on the taxonomy and nomenclature of the algal classes Eustigmatophyceae and Tribophyceae (synonym Xanthophyceae). *Botanical Journal of the Linnean Society*, 82(2), 93–119. <https://doi.org/10.1111/j.1095-8339.1981.tb00954.x>

- Holten-Andersen, L., Doherty, T. M., Korsholm, K. S., & Andersen, P. (2004). Combination of the cationic surfactant dimethyl dioctadecyl ammonium bromide and synthetic mycobacterial cord factor as an efficient adjuvant for tuberculosis subunit vaccines. *Infection and Immunity*, 72(3), 1608–1617. <https://doi.org/10.1128/iai.72.3.1608-1617.2004>
- Hong, Y., Hu, C., Bai, J., Fan, D., Lin, A., Xiang, L., & Shao, J. (2023). Essential role of an ERV-derived env38 protein in adaptive humoral immunity against an exogenous SVCV infection in a zebrafish model. *PLOS Pathogens*, 19(4). <https://doi.org/10.1371/journal.ppat.1011222>
- Ina-Salwany, M. Y., Al-saari, N., Mohamad, A., Mursidi, F. A., Mohd-Aris, A., Amal, M. N., Kasai, H., Mino, S., Sawabe, T., & Zamri-Saad, M. (2018). Vibriosis in fish: A review on disease development and revention. *Journal of Aquatic Animal Health*, 31(1), 3–22. <https://doi.org/10.1002/aah.10045>
- Izumiya, H., Furukawa, M., Ogata, K., Isobe, J., Watanabe, S., Sasaki, M., Ichinose, K., Arakawa, E., Morita, M., Kurane, I., & Ohnishi, M. (2017). A double-quadratic model for predicting *Vibrio* species in water environments of Japan. *Archives of microbiology*, 199(9), 1293–1302. <https://doi.org/10.1007/s00203-017-1402-1>
- Jansen, K. U., Gruber, W. C., Simon, R., Wassil, J., & Anderson, A. S. (2021). The impact of human vaccines on bacterial antimicrobial resistance. A review. *Environmental chemistry letters*, 19(6), 4031–4062. <https://doi.org/10.1007/s10311-021-01274-z>
- Jumatli, A., & Ismail, M. S. (2021). Promotion of sustainable aquaculture in Malaysia. In F. A. Aya, L. D. de la Peña, N. D. Salayo, & E. A. Tendencia (Eds.), *Proceedings of the international workshop on the promotion of sustainable aquaculture, aquatic animal health, and resource enhancement in Southeast Asia*, 31–40. Tigbauan, Iloilo, Philippines: Aquaculture Department, Southeast Asian Fisheries Development Center.
- Kiatarangul, A., Maneenin, S., Purton, S., Areechon, N., Hirono, I., Brocklehurst, T. W., & Unajak, S. (2020). An oral delivery system for controlling white spot syndrome virus infection in shrimp using transgenic microalgae. *Aquaculture*, 521, 1-8. <https://doi.org/10.1016/j.aquaculture.2020.735022>
- Kilian, O., Benemann, C. S., Niyogi, K. K., & Vick, B. (2011). High-efficiency homologous recombination in the oil-producing alga *Nannochloropsis* SP. *Proceedings of the National Academy of Sciences*, 108(52), 21265–21269. <https://doi.org/10.1073/pnas.1105861108>
- Kim, J. Y., & Lee, J.-L. (2017). Correlation of total bacterial and *Vibrio* spp. populations between fish and water in the aquaculture system. *Frontiers in Marine Science*, 4. <https://doi.org/10.3389/fmars.2017.00147>
- Kordon, A. O., Pinchuk, L., & Karsi, A. (2021). Adaptive immune system in fish. *Turkish Journal of Fisheries and Aquatic Sciences*, 22(4). <https://doi.org/10.4194/trjfas20235>

- Kwon, K.-C., Verma, D., Singh, N. D., Herzog, R., & Daniell, H. (2013). Oral delivery of human biopharmaceuticals, autoantigens and vaccine antigens bioencapsulated in plant cells. *Advanced Drug Delivery Reviews*, 65(6), 782–799. <https://doi.org/10.1016/j.addr.2012.10.005>
- Kwon, K., Lamb, A., Fox, D., & Jegathese, S. J. (2019). An evaluation of microalgae as a recombinant protein oral delivery platform for fish using green fluorescent protein (GFP). *Fish & Shellfish Immunology*, 87, 414-420. <https://doi.org/10.1016/j.fsi.2019.01.038>
- Laith, A., Abdullah, M., Nurhafizah, W., Hussein, H., Aya, J., Effendy, A., & Najiah, M. (2019). Efficacy of live attenuated vaccine derived from the *Streptococcus agalactiae* on the immune responses of *Oreochromis niloticus*. *Fish & Shellfish Immunology*, 90, 235-243. <https://doi.org/10.1016/j.fsi.2019.04.052>
- Laktuka, K., Kalnbalkite, A., Sniega, L., Logins, K., & Lauka, D. (2023). Towards the sustainable intensification of aquaculture: Exploring possible ways forward. *Sustainability*, 15(24), 16952. <https://doi.org/10.3390/su152416952>
- Lanh, P. T., Nguyen, H. M., Duong, B. T., Hoa, N. T., Thom, L. T., Tam, L. T., Thu, H. T., Nha, V. V., Hong, D. D., Mouradov, A., Koyande, A. K., Show, P.-L., & Van Quyen, D. (2021). Generation of microalga *Chlamydomonas reinhardtii* expressing VP28 protein as oral vaccine candidate for shrimps against White Spot Syndrome Virus (WSSV) infection. *Aquaculture*, 540, 736737. <https://doi.org/10.1016/j.aquaculture.2021.736737>
- Lee, Y. S. (2017). *Future of Vaccination in Warmwater Aquaculture. Strengthening capacities, policies and national action plans on prudent and responsible use of antimicrobials in fisheries Final Workshop in cooperation with AVA Singapore and INFOFISH*. Singapore; Singapore. Retrieved from <https://www.fao.org/fi/static-media/MeetingDocuments/WorkshopAMR17/presentations/21.pdf>.
- Lekang, O. (2015). Feeding equipment. In D. A. Davis (Ed.), *Feed and feeding practices in aquaculture: Series in food science, technology and nutrition*, 349-368. Woodhead Publishing. <https://doi.org/10.1016/b978-0-08-100506-4.00014-3>
- Levine, M., & Szein, M. (2004). Vaccine development strategies for improving immunization: The role of modern immunology. *Nature Immunology*, 5(5), 460–464. <https://doi.org/10.1038/ni0504-460>
- Levraud, J.-P., Rawls, J. F., & Clatworthy, A. E. (2022). Using zebrafish to understand reciprocal interactions between the nervous and immune systems and the microbial world. *Journal of Neuroinflammation*, 19(1). <https://doi.org/10.1186/s12974-022-02506-x>
- Lillehaug A. Vaccination strategies and procedures. In: Gudding R, Lillehaug A, Evensen O, editors (2014). *Fish Vaccination*. Oxford: Willey Blackwell;. p.

- Liu, L., Xiao, J., Zhang, M., Zhu, W., Xia, X., Dai, X., Pan, Y., Yan, S., & Wang, Y. (2018). A *Vibrio owensii* strain as the causative agent of AHPND in cultured shrimp, *Litopenaeus Vannamei*. *Journal of Invertebrate Pathology*, 153, 156–164. <https://doi.org/10.1016/j.jip.2018.02.005>
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2^{(-Delta Delta C(T))} Method. *Methods (San Diego, Calif.)*, 25(4), 402–408. <https://doi.org/10.1006/meth.2001.1262>
- Ma, J., Bruce, T. J., Jones, E. M., & Cain, K. D. (2019). A review of fish vaccine development strategies: Conventional methods and modern biotechnological approaches. *Microorganisms*, 7(11), 569. <https://doi.org/10.3390/microorganisms7110569>
- Ma, K., Bao, Q., Wu, Y., Chen, S., Zhao, S., Wu, H., & Fan, J. (2020). Evaluation of microalgae as immunostimulants and recombinant vaccines for diseases prevention and control in Aquaculture. *Frontiers in Bioengineering and Biotechnology*, 8. <https://doi.org/10.3389/fbioe.2020.590431>
- Manchanayake, T., Salleh, A., Amal, M. N., Yasin, I. S., & Zamri-Saad, M. (2023). Pathology and pathogenesis of *Vibrio* infection in fish: A Review. *Aquaculture Reports*, 28, 101459. <https://doi.org/10.1016/j.aqrep.2022.101459>
- Matsumoto, M., Araki, K., Hayashi, K., Takeuchi, Y., Shiozaki, K., Suetake, H., & Yamamoto, A. (2017). Adjuvant effect of recombinant interleukin-12 in the Nocardiosis formalin-killed vaccine of the amberjack *Seriola dumerili*. *Fish & Shellfish Immunology*, 67, 263–269. <https://doi.org/10.1016/j.fsi.2017.06.025>
- Maulu, S., Hasimuna, O. J., Haambiya, L. H., Monde, C., Musuka, C. G., Makorwa, T. H., Munganga, B. P., Phiri, K. J., & Nsekanabo, J. D. (2021). Climate change effects on aquaculture production: Sustainability Implications, mitigation, and adaptations. *Frontiers in Sustainable Food Systems*, 5. <https://doi.org/10.3389/fsufs.2021.609097>
- McClure, M. M., McIntyre, P. B., & McCune, A. R. (2006). Notes on the natural diet and habitat of eight danionin fishes, including the zebrafish *Danio rerio*. *Journal of Fish Biology*, 69(2), 553–570. <https://doi.org/10.1111/j.1095-8649.2006.01125.x>
- Mekasha, S. & Linke, D. (2021). Secretion Systems in Gram-Negative bacterial fish pathogens. *Frontiers in microbiology*, 12, 782673. <https://doi.org/10.3389/fmicb.2021.782673>
- Michelet, L., Lefebvre-Legendre, L., Burr, S. E., Rochaix, J. -D. & Goldschmidt-Clermont, M. (2011). Enhanced chloroplast transgene expression in a nuclear mutant of *Chlamydomonas*. *Plant Biotechnology Journal*, 9(5), 565–574.

<https://doi.org/10.1111/j.1467-7652.2010.00564.x>

- Miccoli, A., Manni, M., Picchietti, S., & Scapigliati, G. (2021). State-of-the-art vaccine research for aquaculture use: The case of three economically relevant fish species. *Vaccines*, 9(2), 140. <https://doi.org/10.3390/vaccines9020140>
- Mohamad, N., Amal, M. N., Yasin, I. S., Zamri Saad, M., Nasruddin, N. S., Al-saari, N., Mino, S., & Sawabe, T. (2019). Vibriosis in cultured marine fishes: A Review. *Aquaculture*, 512, 734289. <https://doi.org/10.1016/j.aquaculture.2019.734289>
- Mohd-Aris, A., Muhamad-Sofie, M., Zamri-Saad, M., Daud, H. M., & Ina-Salwany, M. Y. (2019). Live vaccines against bacterial fish diseases: A review. *Veterinary World*, 12(11), 1806–1815. <https://doi.org/10.14202/vetworld.2019.1806-1815>
- Mohd Yazid, S. H., Mohd Daud, H., Azmai, M. N. A., Mohamad, N., & Mohd Nor, N. (2021). Estimating the Economic Loss Due to Vibriosis in Net-Cage Cultured Asian Seabass (*Lates calcarifer*): Evidence From the East Coast of Peninsular Malaysia. *Frontiers in Veterinary Science*, 8, 644009. <https://doi.org/10.3389/fvets.2021.644009>
- Mondal, H., & Thomas, J. (2022). A review on the recent advances and application of vaccines against fish pathogens in aquaculture. *Aquaculture International*, 30(4), 1971–2000. <https://doi.org/10.1007/s10499-022-00884-w>
- Munang'andu, H. M., Mutoloki, S., & Evensen, Ø. (2015). An overview of challenges limiting the design of protective mucosal vaccines for finfish. *Frontiers in Immunology*, 6. <https://doi.org/10.3389/fimmu.2015.00542>
- Muktar, Y., Tesfaye, S., & Tesfaye, B. (2016). Present status and future prospects of fish vaccination: A review. *Journal of Veterinary Science and Technology*, 7(2), 299. <https://doi.org/10.4172/2157-7579.1000299>
- Mutoloki, S., Munang'andu, H. M., & Evensen, Ø. (2015). Oral vaccination of fish – antigen preparations, uptake, and immune induction. *Frontiers in Immunology*, 6, 519. <https://doi.org/10.3389/fimmu.2015.00519>
- Nag, D., Farr, D. A., Walton, M. G., & Withey, J. H. (2020). Zebrafish models for pathogenic vibrios. *Journal of Bacteriology*, 202(24). <https://doi.org/10.1128/jb.00165-20>
- Nagasawa, K., & Cruz-Lacierda, E. R. (Eds.). (2004). Diseases of cultured groupers. Tigbauan, Iloilo, Philippines: Aquaculture Department, Southeast Asian Fisheries Development Center.
- Neely, M. N., Pfeifer, J. D., & Caparon, M. (2002). *Streptococcus*-zebrafish model of bacterial pathogenesis. *Infection and Immunity*, 70(7), 3904–3914. <https://doi.org/10.1128/iai.70.7.3904.2002>

- Ningqiu, L., Junjie, B., Shuqin, W., Xiaozhe, F., Haihua, L., Xing, Y., & Cunbin, S. (2008). An outer membrane protein, *OmpK*, is an effective vaccine candidate for *Vibrio harveyi* in Orange-spotted grouper (*Epinephelus coioides*). *Fish & Shellfish Immunology*, 25(6), 829–833. <https://doi.org/10.1016/j.fsi.2008.09.007>
- Okuthe, G. E., & Bhomela, B. (2020). Morphology, histology and histochemistry of the digestive tract of the banded tilapia, tilapia sparrmanii (*perciformes: Cichlidae*). *Zoologia*, 37, 1–14. <https://doi.org/10.3897/zoologia.37.e51043>
- Pridgeon, J. W., & Klesius, P. H. (2012). Major bacterial diseases in aquaculture and their vaccine development. *CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources*, 7(048), 1-16. <https://doi.org/10.1079/pavsnr.20127048>
- Pajdak-Czaus, J., Schulz, P., Terech-Majewska, E., Szweda, W., Siwicki, A. K., & Platt-Samoraj, A. (2021). Influence of Infectious Pancreatic Necrosis Virus and *Yersinia ruckeri* co-Infection on a non-Specific immune system in rainbow trout (*Oncorhynchus mykiss*). *Animals*, 11(7), 1974. <https://doi.org/10.3390/ani11071974>
- Qosimah, D., Santoso, S., Maftuch, M., Khotimah, H., Fitri, L. E., Aulanni'am, A., & Suwanti, L. T. (2023). *Aeromonas hydrophila* induction method in adult zebrafish (*Danio rerio*) as animal infection models. *Veterinary world*, 16(2), 250–257. <https://doi.org/10.14202/vetworld.2023.250-257>
- Quang, H. T. (2020). Genetic diversity and toxic genes analysis of *Vibrio* spp. isolated from white leg shrimp and marine fishes cultured in Tam G Lang Lagoon in thua thien Hue Province, Vietnam. *Indian Journal of Science and Technology*, 13(13), 1412–1422. <https://doi.org/10.17485/ijst/v13i13.161>
- Ramesh Kumar, P and Kalidas, C and Tamilmani, G and Sakthivel, M and Nazar, A K A and Maharshi, V Ashok and Rao, S K Srinivasa and Gopakumar, G (2014). Microbiological and histopathological investigations of *Vibrio alginolyticus* infection in cobia *Rachycentron canadum* (Linnaeus, 1766) cultured in sea cage. *Indian Journal of Fisheries*, 61 (1). pp. 124-127.
- Ridzuan, M. S. M., Abdullah, A., Ramly, R., Mansor, N. N., Ramli, N., & Firdaus-Nawi, M. (2022). Current status and advances of fish vaccines in Malaysia. *Veterinary world*, 15(2), 465–482. <https://doi.org/10.14202/vetworld.2022.465-482>
- Riera Romo, M., Pérez-Martínez, D., & Castillo Ferrer, C. (2016). Innate immunity in vertebrates: an overview. *Immunology*, 148(2), 125–139. <https://doi.org/10.1111/imm.12597>
- Rodriguez I., Novoa, B., & Figueras, A. (2008). Immune response of zebrafish (*Danio rerio*) against a newly isolated bacterial pathogen *Aeromonas hydrophila*. *Fish & Shellfish Immunology*, 25(3), 239–249. <https://doi.org/10.1016/j.fsi.2008.05.002>

- Rønneseth, A., Castillo, D., D'Alvise, P., Tønnesen, Ø., Haugland, G., Grotkjaer, T., Engell-Sørensen, K., Nørremark, L., Bergh, Ø., Wergeland, H. I., & Gram, L. (2017). Comparative assessment of *Vibrio* virulence in marine fish larvae. *Journal of Fish Diseases*, 40(10), 1373–1385. <https://doi.org/10.1111/jfd.12612>
- Rudtanatip, T., Boonsri, B., Praiboon, J., & Wongprasert, K. (2019). Bioencapsulation efficacy of sulfated galactans in adult *Artemia salina* for enhancing immunity in shrimp *Litopenaeus vannamei*. *Fish & Shellfish Immunology*, 94, 90-98 <https://doi.org/10.1016/j.fsi.2019.08.065>
- Sarker, P. K., Kapuscinski, A. R., Lanois, A. J., Livesey, E. D., Bernhard, K. P., & Coley, M. L. (2016). Towards sustainable aquafeeds: Complete substitution of fish oil with marine microalga *Schizochytrium* sp. improves growth and fatty acid deposition in juvenile Nile tilapia (*Oreochromis niloticus*). *Plos One*, 11(6), e0156684. <https://doi.org/10.1371/journal.pone.0156684>
- Schmidt, J. G., Korbut, R., Ohtani, M., & Jørgensen, L. von. (2017). Zebrafish (*Danio rerio*) as a model to visualize infection dynamics of *Vibrio Anguillarum* following intraperitoneal injection and Bath Exposure. *Fish & Shellfish Immunology*, 67, 692–697. <https://doi.org/10.1016/j.fsi.2017.06.052>
- van Seventer, J. M., & Hochberg, N. S. (2017). Principles of infectious diseases: transmission, diagnosis, prevention, and control. *International Encyclopedia of Public Health*, 22–39. <https://doi.org/10.1016/B978-0-12-803678-5.00516-6>
- Shah, M.R., Lutz, G.A., Alam, A. et al.(2018). Microalgae in aquafeeds for a sustainable aquaculture industry. *Journal of Applied Phycology* 30, 197–213. <https://doi.org/10.1007/s10811-017-1234-z>
- Shahin, K., Pirezan, F., Rogge, M., Lafrentz, B. R., Shrestha, R. P., Hildebrand, M., Lu, F., HogenEsch, H., & Soto, E. (2020). Development of IglC and GroEL recombinant vaccines for francisellosis in Nile tilapia, *Oreochromis niloticus*. *Fish & Shellfish Immunology*, 105, 341-349. <https://doi.org/10.1016/j.fsi.2020.07.045>
- Shoemaker, C. A., Klesius, P. H., Evans, J. J. & Arias, C. R. (2009). Use of modified live vaccines in aquaculture. *Journal of the World Aquaculture Society*, 40(5),573-585.
- Sikic, B. I. (1986). Biochemical and cellular determinants of bleomycin cytotoxicity. *Cancer Surveys*, 5(1), 81–91. <https://europepmc.org/article/med/2439200>
- Soliman, W. S., Shaapan, R. M., Mohamed, L. A., & Gayed, S. S. (2019). Recent biocontrol measures for fish bacterial diseases, in particular to probiotics, bio-encapsulated vaccines, and phage therapy. *Open Veterinary Journal*, 9(3), 190-195. <https://doi.org/10.4314/ovj.v9i3.2>
- Sommerset, I., Krossøy, B., Biering, E., & Frost, P. (2005). Vaccines for fish aquaculture. *Expert Review of Vaccines*, 4(1), 89-101.

<https://doi.org/10.1586/14760584.4.1.89>

- Spence, R., Gerlach, G., Lawrence, C., & Smith, C. (2007). The behaviour and ecology of the zebrafish, *Danio rerio*. *Biological Reviews*, 83(1), 13–34. <https://doi.org/10.1111/j.1469-185x.2007.00030.x>
- Specht, E. A., & Mayfield, S. P. (2014). Algae-based oral recombinant vaccines. *Frontiers in Microbiology*, 5, 60. <https://doi.org/10.3389/fmicb.2014.00060>
- Sudheesh, P. S., Al-Ghabshi, A., Al-Mazrooei, N., & Al-Habsi, S. (2012). Comparative pathogenomics of bacteria causing infectious diseases in fish. *International Journal of Evolutionary Biology*, 2012, 1–16. <https://doi.org/10.1155/2012/457264>
- Sullivan, C., & Kim, C. H. (2008). Zebrafish as a model for infectious disease and immune function. *Fish & Shellfish Immunology*, 25(4), 341–350. <https://doi.org/10.1016/j.fsi.2008.05.005>
- Sultana, R., Khatoon, H., Rahman, M. R., Haque, M. E., Nayma, Z., & Mukta, F. A. (2022). Potentiality of nannochloropsis sp. as partial dietary replacement of fishmeal on growth, proximate composition, pigment and breeding performance in guppy (poecilia reticulata). *Bioresource Technology Reports*, 18, 101112. <https://doi.org/10.1016/j.biteb.2022.101112>
- Teame, T., Zhang, Z., Ran, C., Zhang, H., Yang, Y., Ding, Q., Xie, M., Gao, C., Ye, Y., Duan, M., & Zhou, Z. (2019). The use of zebrafish (*Danio rerio*) as biomedical models. *Animal Frontiers : The Review Magazine of Animal Agriculture*, 9(3), 68–77. <https://doi.org/10.1093/af/vfz020>
- Thiang, E. L., Chai, C. Y., Lee, C. W., Takada, H., Wang, A. J., Chai, L. C., & Bong, C. W. (2022). Tetracycline resistance and prevalence of tetracycline resistance genes in bacteria from marine aquaculture farms in Peninsular Malaysia. *Sains Malaysiana*, 51(2), 345–357. <https://doi.org/10.17576/jsm-2022-5102-02>
- Vartak, A., & Sucheck, S. J. (2016). Recent advances in subunit vaccine carriers. *Vaccines*, 4(2), 12. <https://doi.org/10.3390/vaccines4020012>
- Vieler, A., Wu, G., Tsai, C.-H., Bullard, B., Cornish, A. J., Harvey, C., Reca, I.-B., Thornburg, C., Achawanantakun, R., Buehl, C. J., Campbell, M. S., Cavalier, D., Childs, K. L., Clark, T. J., Deshpande, R., Erickson, E., Armenia Ferguson, A., Handee, W., Kong, Q., ... Benning, C. (2012). Genome, functional gene annotation, and nuclear transformation of the heterokont oleaginous alga *Nannochloropsis oceanica* CCMP1779. *PLoS Genetics*, 8(11). <https://doi.org/10.1371/journal.pgen.1003064>
- Villumsen, K. R., Neumann, L., Ohtani, M., Strøm, H. K., & Raida, M. K. (2014). Oral and anal vaccination confers full protection against enteric redmouth disease (ERM) in rainbow trout. *PloS one*, 9(4), e93845.

<https://doi.org/10.1371/journal.pone.0093845>

- Wahidin, S., Idris, A., & Shaleh, S. R. (2013). The influence of light intensity and photoperiod on the growth and lipid content of microalgae *Nannochloropsis* sp.. *Bioresource Technology*, 129, 7–11. <https://doi.org/10.1016/j.biortech.2012.11.032>
- Wamala, S. P., Mugimba, K. K., Mutoloki, S., Evensen, Ø., Mdegela, R., Byarugaba, D. K., & Sørum, H. (2018). Occurrence and antibiotic susceptibility of fish bacteria isolated from *Oreochromis niloticus* (Nile tilapia) and *Clarias gariepinus* (African catfish) in Uganda. *Fisheries and Aquatic Sciences*, 21, 6. <https://doi.org/10.1186/s41240-017-0080-x>
- Wang, D., Ning, K., Li, J., Hu, J., Han, D., Wang, H., Zeng, X., Jing, X., Zhou, Q., Su, X., Chang, X., Wang, A., Wang, W., Jia, J., Wei, L., Xin, Y., Qiao, Y., Huang, R., Chen, J., Han, B., ... Xu, J. (2014). *Nannochloropsis* genomes reveal evolution of microalgal oleaginous traits. *PLoS genetics*, 10(1), e1004094. <https://doi.org/10.1371/journal.pgen.1004094>
- Wang, J., Zou, L. L., & Li, A. X. (2014). Construction of a *Streptococcus iniae* sortase A mutant and evaluation of its potential as an attenuated modified live vaccine in Nile tilapia (*Oreochromis niloticus*). *Fish & Shellfish Immunology*, 40(2), 392–398. <https://doi.org/10.1016/j.fsi.2014.07.028>
- Wang, Q., Ji, W., & Xu, Z. (2020). Current use and development of fish vaccines in China. *Fish & Shellfish Immunology*, 96, 223–234. <https://doi.org/10.1016/j.fsi.2019.12.010>
- Watts, J., Schreier, H. J., Lanska, L., & Hale, M. S. (2017). The rising tide of antimicrobial resistance in aquaculture: Sources, sinks and solutions. *Marine Drugs*, 15(6), 158. <https://doi.org/10.3390/md15060158>
- Wilson, J. M., & Castro, L. F. C. (2010). Morphological diversity of the gastrointestinal tract in fishes. *Fish Physiology*, 1–55. [https://doi.org/10.1016/s1546-5098\(10\)03001-3](https://doi.org/10.1016/s1546-5098(10)03001-3)
- Wolf, J. C., & Maack, G. (2017). Evaluating the credibility of histopathology data in environmental endocrine toxicity studies. *Environmental Toxicology and Chemistry*, 36(3), 601–611. <https://doi.org/10.1002/etc.3695>
- Wolf, J. C., & Wheeler, J. R. (2018). A critical review of histopathological findings associated with endocrine and non-endocrine hepatic toxicity in fish models. *Aquatic Toxicology*, 197, 60–78. <https://doi.org/10.1016/j.aquatox.2018.01.013>
- Xu, H., Zeng, Y. H., Yin, W. L., Lu, H. B., Gong, X. X., Zhang, N., Zhang, X., Long, H., Ren, W., Cai, X. N., Huang, A. Y., & Xie, Z. Y. (2022). Prevalence of bacterial coinfections with *Vibrio harveyi* in the industrialized flow-through Aquaculture systems in Hainan Province: A neglected high-risk lethal causative agent to hybrid grouper. *International Journal of Molecular Sciences*, 23(19),

- Xu, X., Liu, K., Wang, S., Guo, W., Xie, Z., & Zhou, Y. (2017). Identification of pathogenicity, investigation of virulent gene distribution and development of a virulent strain-specific detection PCR method for *Vibrio harveyi* isolated from Hainan province and Guangdong Province, China. *Aquaculture*, 468, 226–234. <https://doi.org/10.1016/j.aquaculture.2016.10.015>
- Yaacob, E. N., Norouzitallab, P., De Geest, B. G., Bajek, A., Dierckens, K., Bossier, P., & Vanrompay, D. (2020). Recombinant DnaK orally administered protects axenic European sea bass against vibriosis. *Frontiers in Immunology*, 10, 3162. <https://doi.org/10.3389/fimmu.2019.03162>
- Ye, H., Xu, Z., Tao, Z., Li, W., Li, Y., Yang, A., Wang, W., Yin, X., & Yan, X. (2021). Efficacy and safety of *Pseudomonas plecoglossicida* mutant ΔtssD-1 as a live attenuated vaccine for the large yellow croaker (*Larimichthys crocea*). *Aquaculture*, 531, 735976. <https://doi.org/10.1016/j.aquaculture.2020.735976>
- Ye, Y., Liu, M., Yu, L., Sun, H., & Liu, J. (2024). *Nannochloropsis* as an emerging algal chassis for light-driven synthesis of lipids and high-value products. *Marine Drugs*, 22(2), 54. <https://doi.org/10.3390/md22020054>
- Yun, S., Giri, S. S., Kim, H. J., Kim, S. G., Kim, S. W., Kang, J. W., Han, S. J., Kwon, J., Oh, W. T., Chi, C., Jun, J. W., & Chang Park, S. (2019). Enhanced bath immersion vaccination through microbubble treatment in the cyprinid loach. *Fish & Shellfish Immunology*, 91, 12–18. <https://doi.org/10.1016/j.fsi.2019.05.021>
- Zahran, E., Elbahnaswy, S., Ibrahim, I., & Khaled, A. A. (2021). *Nannochloropsis oculata* enhances immune response, transcription of stress, and cytokine genes in Nile tilapia subjected to air exposure stress. *Aquaculture Reports*, 21, 100911. <https://doi.org/10.1016/j.aqrep.2021.100911>
- Zanella, L., & Vianello, F. (2020). Microalgae of the genus *Nannochloropsis*: Chemical composition and functional implications for human nutrition. *Journal of Functional Foods*, 68, 103919. <https://doi.org/10.1016/j.jff.2020.103919>
- Zhai, Y., Shi, D., He, P., Cai, C., Yin, R., and Jia, R. (2019). Effect of trans-vp28 gene *Synechocystis* sp. PCC6803 on growth and immunity of *Litopenaeus vannamei* and defense against white spot syndrome virus (WSSV). *Aquaculture* 512:734306. <https://doi.org/10.1016/j.aquaculture.2019.734306>
- Zhang, J.-M., & An, J. (2007). Cytokines, Inflammation, and Pain. *International Anesthesiology Clinics*, 45(2), 27–37. <https://doi.org/10.1097/aia.0b013e318034194e>
- Zhang, W., Liao, Z., Hu, F., Chen, X., & Huang, X. (2019). Protective immune responses of recombinant outer membrane proteins *OmpF* and *OmpK*

of *Aeromonas hydrophila* in European eel (*Anguilla anguilla*). *Aquaculture Research*, 50(12), 3559–3566. <https://doi.org/10.1111/arc.14311>

Zhang, Y. A., Salinas, I., Li, J., Parra, D., Bjork, S., Xu, Z., LaPatra, S. E., Bartholomew, J., & Sunyer, J. O. (2010). IgT, a primitive immunoglobulin class specialized in mucosal immunity. *Nature immunology*, 11(9), 827–835. <https://doi.org/10.1038/ni.1913>

