



**BIOACTIVITIES AND PHYTOCHEMICAL AND TOXICITY ANALYSES OF
ETHANOLIC KAYU MANIS HUTAN [*Cinnamomum iners* (Reinw. ex Nees
and T. Nees) Blume] LEAF EXTRACT AND ITS APPLICATION IN FOOD**

By

KHAIRUL NAIM BIN MD PADZIL

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy**

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January 2024

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In the food industry, the foodborne bacteria and spoilage microorganisms have capabilities to cause foodborne diseases and spoilage. This has been prevented with utilization of synthetic antimicrobial agents in food products. However, its application has long term affected human health. This concern needs to be controlled which involves the utilization of safe natural plant such as Kayu Manis Hutan [*Cinnamomum iners* (Reinw. ex Nees and T. Nees) Blume] which reported to have antimicrobial and antioxidant activities. The study aims to determine the antimicrobial activity of ethanolic young and old *C. iners* leaves extracts against selected pathogenic microorganisms namely *Bacillus cereus*, *B. megaterium*, *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Aspergillus flavus*, *A. niger*, *Rhizopus oligosporus* and *R. oryzae*. The result of antibacterial activity has exhibited inhibition zone at range

between 9.00 to 10.00 mm against all tested bacteria strains. The tested bacteria growths were inhibited at MICs ranged from 0.16 to 2.50 mg/mL, and were killed at MBCs ranged from 1.25 to 5.00 mg/mL. Time kills curve study showed that all tested bacteria can be killed at 4× MIC within 4 h. The antifungal activity of the extracts has exhibited zones of inhibition at range of 6.50 to 7.25 mm against all tested food spoilage microorganisms. The MICs of the extracts ranged from 0.63 to 5.00 mg/mL, and the MFCs at ranges of 1.25 to 5.00 mg/mL. In qualitative analysis, *A. niger* and *R. oryzae* showed no growth after being treated with young leaves extracts at 4× MIC while *R. oligosporus* and *A. flavus* at 2× MIC for 14 days. Meanwhile, no growth was shown on all tested fungi after being treated with old leaves extracts at 2× MIC for 14 days. In quantitative analysis, using 4× MIC for 24 h, the conidia germination was fully inhibited for all tested fungi. The antimicrobial activities of both extracts were shown not significantly different after being treated at different pHs and temperatures. The SEM analysis showed morphological features in selected treated microorganisms namely, *B. cereus*, *K. pneumoniae* and *A. flavus* were changed in the cell wall shape, ruptured, and the cytoplasm leaked. Both leaves extracts have exhibited high total phenolic content and antioxidant activities. The toxicity study demonstrated that both extracts was not toxic to *Artemia salina* with LC₅₀ at 19.62 mg/mL and 13.20 mg/mL, respectively. GC-MS analysis has identified the presence of active volatile compounds in both extracts included phytol, hexadecanoic acid, dihydro(-)-neoclovene-(II), 14-β-pregnane and 2-hexadecenal. LC-MS have detected coniferyl alcohol, isorhamnetin-3-O-rutinoside in young leaf extract while α-terpineol, and 4-methylcatechol in old leaf extract. The decrease of 3

Log₁₀ CFU/g of total plate count, total viable mold count, *B. cereus*, *E. coli*, and *K. pneumoniae* count on raw chicken meat sample was observed when exposed to 5.00% (v/v) of *C. iners* extracts and maintain its microbiological quality with longer exposure within 21 days in different storage conditions. In conclusion, *C. iners* extracts and its compounds have shown potential of antimicrobial and antioxidant activities against foodborne pathogens and spoilage microorganism and thus can be developed as natural antimicrobial agent for food preservatives.

Keywords: *Cinnamomum iners* leaf extracts, antibacterial, antioxidant, toxicity, microbial population

SDG: Goal 3 - Good Health and Well Being

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**BIOAKTIVITI DAN ANALISIS FITOKIMIA DAN KETOKSIKAN EKSTRAK
DAUN KAYU MANIS HUTAN [*Cinnamomum iners* (Reinw. Ex Nees dan
T. Nees) Blume] DAN PENGGUNAANNYA DALAM MAKANAN**

Oleh

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Dalam industri makanan, bakteria bawaan makanan dan mikroorganisma perosak mempunyai keupayaan untuk menyebabkan penyakit bawaan makanan. Ini telah dicegah dengan penggunaan agen antimikrob sintetik dalam produk makanan. Walau bagaimanapun, penggunaannya telah menjejaskan kesihatan manusia jangka panjang. Kebimbangan ini perlu dikawal dengan menggunakan produk tumbuhan semulajadi yang selamat seperti Kayu Manis Hutan [*Cinnamomum iners* (Reinw. Ex Nees and T. Nees) Blume] yang dilaporkan mempunyai aktiviti antimikrob dan antioksidan. Matlamat kajian ini adalah untuk menentukan aktiviti antimikrob ekstrak daun *C. iners* muda dan tua dalam etanol terhadap bakteria bawaan makanan terpilih iaitu *Bacillus cereus*, *B. megaterium*, *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Aspergillus flavus*, *A. niger*, *Rhizopus*

oligosporus dan *R. oryzae*. Hasil daripada aktiviti antibakteria telah menunjukkan zon perencatan pada julat antara 9.00 hingga 10.00 mm terhadap semua bakteria yang diuji. Pertumbuhan bakteria yang diuji telah terhalang pada MIC berjulat 0.16 hingga 2.50 mg/mL, dan dibunuh dengan MBC pada nilai antara 1.25 hingga 5.00 mg/mL. Kajian pembunuhan masa menunjukkan bahawa semua bakteria yang diuji boleh dibunuh oleh ekstrak pada 4× MIC dalam tempoh 4 jam. Aktiviti antikulat oleh ekstrak menunjukkan penghalang kawasan perencatan yang berkisar antara 6.50 hingga 7.25 mm terhadap semua spesies kulat berfilamen yang diuji serta MIC ekstrak berjulat antara 0.63 hingga 5.00 mg/mL, dan MFC pada julat 1.25 hingga 5.00 mg/mL. Dalam analisis kualitatif, *A. niger* dan *R. oryzae* tidak menunjukkan pertumbuhan selepas dirawat dengan ekstrak daun muda pada 4× MIC manakala *R. oligosporus* dan *A. flavus* pada 2× MIC selama 14 hari. Sementara itu, tiada pertumbuhan ditunjukkan pada semua kulat yang diuji selepas dirawat dengan ekstrak daun tua pada 2× MIC selama 14 hari. Dalam analisis kuantitatif, dengan menggunakan 4× MIC selama 24 jam, percambahan konidia telah dihalang sepenuhnya untuk semua kulat yang diuji. Aktiviti antimikrob bagi kedua-dua ekstrak tidak menunjukkan perbezaan yang signifikan selepas dirawat pada pH dan suhu yang berbeza. Analisis SEM menunjukkan bahawa ciri morfologi dalam mikroorganisma *B. cereus*, *K. pneumoniae* dan *A. flavus* mengalami perubahan dalam bentuk dinding sel, kerosakan, dan kebocoran sitoplasma. Kedua-dua ekstrak telah mempamerkan jumlah kandungan fenolik yang tinggi dan aktiviti antioksidan. Kajian ketoksikan menunjukkan kedua-dua ekstrak tidak beracun kepada *Artemia salina* dengan nilai LC₅₀ pada 19.62 mg/mL dan 13.20 mg/mL. Analisis

GC-MS telah mengenal pasti kewujudan sebatian meruap yang aktif dalam kedua-dua ekstrak termasuk fitol, asid heksadekanoik, dihydro(-)-neoklovin-(II), 14- β -pregnan dan 2-heksadesenal. LC-MS telah mengesan koniferil alkohol, isorhamnetin-3-O-rutinosid dalam ekstrak daun muda manakala α -terpineol, 4-methylkatekol dalam ekstrak daun tua. Penurunan 3 Log₁₀ CFU/g dalam jumlah kiraan piring, jumlah kiraan kulat hidup, jumlah *B. cereus*, *E. coli* dan *K. pneumoniae* pada daging ayam mentah diperhatikan apabila terdedah pada 5.00% (v/v) ekstrak *C. iners* dan kualiti mikrobiologi dapat dikekalkan dengan pendedahan yang lebih lama selama 21 hari dalam keadaan penyimpanan yang berbeza. Kesimpulannya, ekstrak *C. iners* dan sebatian-sebatian dalamnya telah menunjukkan potensi aktiviti antimikrob dan antioksidan terhadap bakteria bawaan makanan dan mikroorganisma perosak, dan oleh itu ia boleh dibangunkan sebagai agen antimikrob semulajadi untuk pengawet makanan.

Kata Kunci: Ekstrak daun *Cinnamomum iners*, antibakteria, antioksidan, ketoksikan, populasi mikrob

SDG: Matlamat 3 - Kesihatan Baik dan Kesejahteraan

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LIST OF ABBREVIATIONS

ABTS	2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)
CFU	Colony Forming Unit
CHX	Clorhexidine
CLSI	Clinical and Laboratory Standards Institute
DMSO	Dimethyl sulfoxide
DPPH	1,1-diphenyl-2-picryl-hydrazyl
EO	Essential oil
FRAP	Ferric reducing antioxidant potential
GAE	Gallic acid equivalent
GC-MS	Gas Chromatography – Mass Spectrometry
IC ₅₀	Inhibition Concentration at 50 percent
LC ₅₀	Lethal Concentration at 50 percent
LC-MS	Liquid Chromatography – Mass Spectrometry
<i>m/z</i>	mass/charge ratio
MFC	Minimum Fungicidal Concentration
MHA	Mueller Hinton agar
MHB	Mueller Hinton broth
MHz	Megahertz
NIST	National Institute of Standards and Technology
PBS	Phosphate Buffer Saline
SEM	Scanning Electron Microscopy
TE	Trolox equivalent

CHAPTER 1

INTRODUCTION

1.1 Background

Foodborne diseases represent a global health threat encountered by the food industry. It is caused by ingesting food that are deteriorated and contaminated by microorganisms, such as bacteria and fungi, as well as natural toxins (mycotoxins), chemicals, and physical agents (Sharif et al., 2018). Pathogenic bacteria such *Salmonella Typhi*, *Escherichia coli*, *Staphylococcus aureus*, *Vibrio cholera* and *Listeria monocytogenes* are the most frequent cause of outbreaks (Gourama, 2020). These pathogens and their toxins can enter the body through different modes and cause foodborne illness that may include abdominal cramps, nausea, vomiting, dehydration, diarrhea, and fever (Switaj et al., 2015). Foodborne bacteria also can be transmitted during food preparation, for example at the farm; the animals have already been infected by pathogenic microorganisms such as *E. coli*, *Bacillus cereus* and *Proteus mirabilis* (Abdul-Mutalib et al., 2015). As a result, without prevention and preservation, food could transmit foodborne and spoilage contamination in food industry.

In order to avoid the growth of harmful bacteria in food, the use of synthetic or chemical antimicrobial compounds is involved in a number of techniques for inhibiting the growth of microorganisms (Uka et al., 2020). However, the threat posed by the quick emergence and spread of bacterial resistance caused by

synthetic antimicrobial treatments becoming more concerned. The synthetic antimicrobial has serious impacts on human health as their long run use causes liver damage, asthma, many allergic reactions, and cancer. Due to negative consumer perception of synthetic antimicrobial agents, the demand for safe natural antimicrobial has increased for food products (Ibrahim et al., 2021).

For many years, natural products have been recognized as an essential source of active substances (Mlala et al., 2019) with more than 200,000 chemical compounds are synthesized to possess medicinal importance (Yang et al., 2016). Plant derived antimicrobial often used to improve health in communities, providing health benefits, prevention diseases and numerous nutritional value (Santini et al., 2017). Worldwide, about 70% plant-based preparations are used as traditional medicines and also facilitate the base for 50% of prescription and/or over the counter drugs used in the Western-type practice of medicine (Pandey, 2019). Additionally, traditional medicines practices and herbal remedies has preference for using young leaves than old leaves of herb due to several factor, including the concentration of active compounds, taste, and safety considerations (Alamgir, 2017). One of the potential plants is *Cinnamomum iners* (Kayu Manis Hutan). It is commonly distributed in the tropical and subtropical regions, Asia and South and Central America, and Australia (Wuu-Kuang, 2011). It originated from Sri Lanka and widely distributed in Myanmar, Sumatra, India, Indonesia (Java), Malaysia, Thailand, Vietnam and Sulawesi (Udayaprakash et al., 2015). According to botanical classification, *C. iners* is one of the 250 species from the genus

Cinnamomum commonly known as wild cinnamon, kayu manis hutan, pokok teja, medang kemangi (Malaysia), teja lawang (Indonesia) and deya gui (China) (Ghalib et al., 2012; Wahab and Hussain, 2021).

The plant *C. iners* is used as a traditional medicine to relieve headaches, breathing and appetite problems, illnesses with bacterial symptoms such as fevers, digestive ailments, coughs, diarrhea and dysentery (Mustaffa et al., 2011). An ethnobotanical report reported that *C. iners* has medicinal properties of antiplasmodial, cytotoxicity, amylase inhibitor, anti-inflammatory, antimicrobial, antioxidant, anti-diarrheal, analgesic and carminative activity (Vigila et al., 2018). According to Mustaffa et al. (2011), xanthorrhizol is known as the active compound isolated from *C. iners* methanolic leaves extract, and was shown to have antimicrobial activity against methicillin resistance *S. aureus* (MRSA). The extracts of *C. iners* contain numerous bioactive compounds such as cinnamic aldehyde (aldehyde), (*E*)-phytol acetate (acetate), linoleic acid (fatty acid) and linalool; β -pinene; β -selinene; α -humulene; α -pinene; α -selinene (terpinenes) (Kumar et al., 2019). Different leaf maturity exhibited different polyphenolic compound and bioactivity (Dziadek et al., 2019). The dried mature leaves of *C. iners* used as spices includes in flavoring stews, pilafs, curries, cornmeal porridge and jerk marinades. This study investigates the antimicrobial and antioxidant activities, phytochemical and toxicity analysis of ethanolic kayu manis hutan (*Cinnamomum iners*) leaves extract against foodborne pathogens and application in food.

1.2 Problem statements

The current existing methods of controlling food-borne illness are the misuse and overuse of antibiotics in food production which contribute to the development of antibiotic-resistant bacteria. Due to the increasing prevalence of this life-threatening microorganism infection and the ability of these pathogens to develop resistance to antibiotic, there is a great need to find new compounds to combat those (Stan et al., 2021). Herbs demonstrate a varied range of biological activities for managing diseases, including foodborne pathogenic microbes with the help of essential oils, phenolic compounds, flavonoids, alkaloids, and other bioactive molecules (Gupta and Birdi, 2017). By incorporating plant-derived antimicrobial agents into food products, these antimicrobial agents can inhibit the growth of spoilage-causing microorganisms, thereby extending the expiration date of the food products (Nogueira et al., 2020). This has inspired the present study to determine the antimicrobial activity of plants which have a wide range of medicinal properties.

The extracts of *C. iners* leaf show significant antibacterial () and antifungal activity against foodborne bacteria (Vigila et al., 2018; Anis et al., 2012). The antimicrobial activity was achieved through various combinations of phenols, triterpenoids, lignins, alkaloids, saponins, tannins and flavonoids (Buru et al., 2014). Unfortunately, the antibacterial and antifungal activities of ethanolic young and old *C. iners* crude leaves extracts against foodborne pathogens and spoilage microorganisms have not been assessed. The stability of *C. iners* extracts on different parameters such as pHs and temperatures against

selected foodborne pathogens and spoilage microorganisms were lacking in finding. To the best of the researcher's knowledge, the phytochemicals compounds in the ethanolic young and old *C. iners* crude leaves extracts that are responsible for the antibacterial and the antifungal activities against foodborne pathogens and spoilage microorganisms have never been examined. The extracts of *C. iners* have been reported to possess medicinal properties such as antioxidant activity (Mustaffa et al., 2016). Unfortunately, the total phenolic content and antioxidant assay of ethanolic young and old *C. iners* crude leaves extracts are not yet reported. The determination of toxicity of ethanolic young and old *C. iners* crude leaves extracts using brine shrimp are also not yet reported. During the storage period, poultry meat undergoes many undesirable changes due to microbial growth that leads to spoilage (Saleh et al., 2020). However, reduction of microbial population in poultry meat by ethanolic young and old *C. iners* crude leaf extracts is not yet reported. Thus, the general objective of the present study is to assess the antimicrobial and antioxidant activities, phytochemical and toxicity analyses of ethanolic kayu manis hutan (*Cinnamomum iners*] leaf extract and application in food.

1.3 Objectives

General of objective of this study is to discover the potential of ethanolic *C. iners* leaf extracts on antimicrobial activity against several type of pathogenic and spoilage microorganism, its phytochemical constituents, the toxicity activity, the antioxidant activity and to determine the ability of this plant extract as natural food preservatives. The specific objectives were:

1. To determine the antibacterial activity of young and old *C. iners* crude leaves extracts on *B. cereus*, *B. megaterium*, *E. coli*, *K. pneumoniae* and *P. mirabilis*, followed with extract stability of different pHs and temperatures and its mode of action by SEM.
2. To determine the antifungal activity of young and old *C. iners* crude leaves extracts on *A. flavus*, *A. niger*, *R. oligosporus* and *R. oryzae*, followed with extract stability of different pHs and temperatures and its mode of action by SEM.
3. To identify the bioactive compounds in young and old *C. iners* crude leaves extracts through GC-MS and LC-MS analyses.
4. To evaluate the toxicity of young and old *C. iners* crude leaves extracts by using brine shrimp (*Artemia salina*) lethality assay, the total phenolic content and the antioxidant activities in term of DPPH, FRAP and ABTS assay.
5. To evaluate the effect of young and old *C. iners* crude leaves extracts on microorganisms in raw meat chicken samples.

REFERENCES

- Abd El-Aziz, D. M. (2013). Detection of *Salmonella* Typhimurium in retail chicken meat and chicken giblets. *Asian Pacific Journal of Tropical Biomedicine*, 3(9), 678-681.
- Abdel-Aziz, S. M., Asker, M. M., Keera, A. A., and Mahmoud, M. G. (2016). Microbial food spoilage: control strategies for shelf life extension. *Microbes in Food and Health*, 1; 239-264.
- Abdeldaiem, M. H., Ali, H. G. M., and Foda, M. I. (2017). Improving the quality of minced beef by using mulberry leaves extract. *Journal of Food Measurement and Characterization*, 11, 1681-1689.
- Abdel-Mageed, W. M., Bayoumi, S. A. L. H., Salama, A. A. R., Salem-Bekhit, M. M., Abd-Alrahman, S. H., and Sayed, H. M. (2014). Antioxidant lipoxygenase inhibitors from the leaf extracts of *Simmondsia chinensis*. *Asian Pacific Journal of Tropical Medicine*, 7, 521-526.
- Abdelwahab, S. I., Mariod, A. A., Taha, M. M. E., Zaman, F. Q., Abdelmageed, A. H. A., Khamis, S., Sivasothy, Y. and Awang, K. (2017). Chemical composition and antioxidant properties of the essential oil of *Cinnamomum altissimum* Kosterm. (Lauraceae). *Arabian Journal of Chemistry*, 10(1), 131-135.
- Abdullah, B. M., Mehdi, M. A. H., Khan, A. R., and Pathan, J. M. (2020). Gas chromatography-mass spectrometry (GC-MS) analysis of ajwain (*Trachyspermum ammi*) seed extract. *International Journal of Pharmaceutical Quality Assurance*, 11(2), 228-231.
- Abdul-Mutalib, N. A., Syafinaz, A. N., Sakai, K., and Shirai, Y. (2015). An overview of foodborne illness and food safety in Malaysia. *International Food Research Journal*, 22(3), 896.
- Abebe, E., Gugsa, G., and Ahmed, M. (2020). Review on major food-borne zoonotic bacterial pathogens. *Journal of Tropical Medicine*, 2020, 1-19
- Abeysekera, W. P. K. M., Arachchige, S. P. G., Abeysekera, W. K. S. M., Ratnasooriya, W. D. and Medawatta, H. M. U. I. (2019). Antioxidant and glycemic regulatory properties potential of different maturity stages of leaf of ceylon cinnamon (*Cinnamomum zeylanicum* Blume) *in vitro*. *Evidence-Based Complementary and Alternative Medicine*, 1, 1-10.
- Abu Bakar, F. I., Abu Bakar, M. F., Rahmat, A., Abdullah, N., Sabran, S. F., and Endrini, S. (2018). Anti-gout potential of Malaysian medicinal plants. *Frontiers in Pharmacology*, 9; 261-269.

- Abubacker, M. N., and Deepalakshmi, T. (2013). *In vitro* antifungal potential of bioactive compound methyl ester of hexadecanoic acid isolated from *Annona muricata* linn (annonaceae) leaves. *Bioscience Biotechnology Resesearch Asia*, 10(2), 879-884.
- Adenike, A. O. (2021). Assessment of medicinal value of (G. Don) Ex Hoyle (*Massularia Acuminate*) stem as a potent chew stick using GC-MS analysis. *British Journal of Medical and Health Sciences*, 3(6), 1-3
- Adhikari, P., and Pandey, A. (2021). Antimicrobial compound production by pigment producing endophytic bacterium (*Burkholderia* sp.) isolated from *Taxus wallichiana* Zucc. *Biologia*, 76(11), 3567-3578.
- Adjimani, J. P. and Asare, P. (2015). Antioxidant and free radical scavenging activity of iron chelators. *Toxicology Reports*, 2, 721-728.
- Adumanya, O. C. U., Uwakwe, A. A. and Essien, E. B. (2014). Essential oil composition (terpenes) of *Salacia senegalensis* Lam (DC) leaf. *British Journal of Research*, 1(2); 26-34
- Agrawal, A., Kaushik, N., and Biswas, S. (2014). Derivatives and applications of lignin—an insight. *The SciTech Journal*, 1(7), 30-36.
- Aguoru, C. U., Bashayi, C. G., and Ogbonna, I. O. (2017). Phytochemical profile of stem bark extracts of *Khaya senegalensis* by Gas Chromatography-Mass Spectrometry (GC-MS) analysis. *Journal of Pharmacognosy and Phytotherapy*, 9(3), 35-43.
- Akbar, A., Sitara, U., Ali, I., Khan, M. I., Phadungchob, T., and Anal, A. K. (2014). Presence of *Escherichia coli* in poultry meat: A potential food safety threat. *International Food Research Journal*, 21(3), 941.
- Akter, J., Hossain, M. A., Takara, K., Islam, M. Z., and Hou, D. X. (2019). Antioxidant activity of different species and varieties of turmeric (*Curcuma* spp): Isolation of active compounds. *Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology*, 215, 9-17.
- Alamgir, A. N. M. (2017). Herbal drugs: their collection, preservation, and preparation; evaluation, quality control, and standardization of herbal drugs. *Therapeutic Use of Medicinal Plants and Their Extracts: Volume 1*:
- Alara, O. R., Abdurahman, N. H., & Ukaegbu, C. I. (2021). Extraction of phenolic compounds: A review. *Current Research in Food Science*, 4, 200-214.
- Al-Fekaiki, D. F., Niamah, A. K., and Al-Sahlany, S. T. G. (2017). Extraction and identification of essential oil from *Cinnamomum zeylanicum* barks and study the antibacterial activity. *The Journal of Microbiology, Biotechnology and Food Sciences*, 7(3), 312.

- Alirezalu, K., Pateiro, M., Yaghoubi, M., Alirezalu, A., Peighambardoust, S. H., and Lorenzo, J. M. (2020). Phytochemical constituents, advanced extraction technologies and techno-functional properties of selected Mediterranean plants for use in meat products. A comprehensive review. *Trends in Food Science and Technology*, 100, 292-306.
- Aljamali, N. M. (2021). Review on food poisoning (types, causes, symptoms, diagnosis, treatment). *Global Academic Journal of Pharmacy and Drug Research*, 3(4), 54-61.
- Allocati, N., Masulli, M., Alexeyev, M. F., and Di Ilio, C. (2013). *Escherichia coli* in Europe: an overview. *International Journal of Environmental Research and Public Health*, 10(12), 6235-6254.
- Al-Qudah, M. A. (2013). Chemical composition and antimicrobial activity of the essential oil of *Linum pubescens* growing wild in Jordan. *Natural product research*, 27(12), 1141-1144.
- Alshannaq, A., and Yu, J. H. (2017). Occurrence, toxicity, and analysis of major mycotoxins in food. *International Journal of Environmental Research and Public Health*, 14(6), 632-652.
- Al-Snafi, A. E. (2019). Constituents and pharmacological effects of *Leontice leontopetalum* - a review. *To Chemistry Journal*, 3, 103-109.
- Alvarez-Ordóñez, A., Fernández, A., Bernardo, A. and Lopez, M. (2010). Acid tolerance in *Salmonella* Typhimurium induced by culturing in the presence of organic acids at different growth temperatures. *Food Microbiology*. 27: 44-49.
- Alves, F. S., Cruz, J. N., de Farias Ramos, I. N., do Nascimento Brandão, D. L., Queiroz, R. N., da Silva, G. V., Dolabela, M. F., Coasta, M. L., Khayat, A. S., Rego, J. A. R. and do Socorro Barros Brasil, D. (2022). Evaluation of antimicrobial activity and cytotoxicity effects of extracts of *Piper nigrum* L. and piperine. *Separations*, 10(1), 21, 1-10
- Alyaseen, F. F., Hassan, B. A., and Abdulhussein, H. S. (2018). Extraction, isolation and chemical identification of piperine alkaloid from black pepper seeds and its antibacterial activity. *Plant Archives*, 18(2), 2171-2176.
- Amaegberi, N. V., Semenкова, G. N., Lisovskaya, A. G., Kvacheva, Z. B., and Shadyro, O. I. (2019). Modification of redox processes in C6 glioma cells by 2-hexadecenal, the product of sphingolipid destruction. *Biophysics*, 64, 424-430.
- Amaiike, S., and Keller, N. P. (2011). *Aspergillus flavus*. *Annual Review of Phytopathology*, 49, 107-133.
- Amalraj, S., Mariyammal, V., Murugan, R., Gurav, S. S., Krupa, J., and Ayyanar, M. (2021). Comparative evaluation on chemical composition, in

- vitro antioxidant, antidiabetic and antibacterial activities of various solvent extracts of *Dregea volubilis* leaves. *South African Journal of Botany*, 138, 115-123.
- Anand, U., Jacobo-Herrera, N., Altemimi, A. and Lakhssassi, N. (2019). A comprehensive review on medicinal plants as antimicrobial therapeutics: potential avenues of biocompatible drug discovery. *Metabolites*, 9(11), 258-266.
- Androutsopoulos, V. P., Ruparelia, K. C., Papakyriakou, A., Filippakis, H., Tsatsakis, A. M., and Spandidos, D. A. (2011). Anticancer effects of the metabolic products of the resveratrol analogue, DMU-212: structural requirements for potency. *European Journal of Medicinal Chemistry*, 46(6), 2586-2595.
- Anis, Z., Hashim, R., Hasan Mehdi, S., and Murad Ghalib, R. (2012). Radical scavenging activity, total phenol content and antifungal activity of *Cinnamomum iners* wood. *Iranica Journal of Energy and Environment*, 3(5); 74-78.
- Annegowda, H. V., Gooi, T. S., Awang, S. H. H., Alias, N. A. and Mordi, M. N. (2012). Evaluation of analgesic and antioxidant potent of various extracts of *Cinnamomum iners* bark. *International Journal of Pharmacology*, 8(3), 198-203.
- Applava, L., Karlíčková, J., Warncke, P., Macáková, K., Hrubša, M., Macháček, M., Tvrđy, V., Fischer, D. and Mladěnka, P. (2019). 4-Methylcatechol, a flavonoid metabolite with potent antiplatelet effects. *Molecular Nutrition and Food Research*, 63(20), 1900261, 1-11
- Arasu, M. V., Viayaraghavan, P., Ilavenil, S., Al-Dhabi, N. A., and Choi, K. C. (2019). Essential oil of four medicinal plants and protective properties in plum fruits against the spoilage bacteria and fungi. *Industrial Crops and Products*, 133, 54-62.
- Arifullah, M., Vikram, P., Chiruvella, K. K., Shaik, M. M. and Ripain, I. H. B. A. (2014). A review on Malaysian plants used for screening of antimicrobial activity. *Annual Research and Review in Biology*, 4 (13), 2088-2132.
- Arora, S., and Kumar, G. (2018). Phytochemical screening of root, stem and leaves of *Cenchrus biflorus* Roxb. *Journal of Pharmacognosy and Phytochemistry*, 7(1), 1445-1450.
- Arrais, A., Testori, F., Calligari, R., Gianotti, V., Roncoli, M., Caramaschi, A., Todeschini, A., Massa, N. and Bona, E. (2022). Extracts from cabbage leaves: preliminary results towards a “universal” highly-performant antibacterial and antifungal natural mixture. *Biology*, 11(7), 1080-1087
- Artés-Hernández, F., Castillejo, N., and Martínez-Zamora, L. (2022). UV and visible spectrum led lighting as abiotic elicitors of bioactive compounds in

sprouts, microgreens, and baby leaves - A comprehensive review including their mode of action. *Foods*, 11(3), 265, 1-15

Awasti, N., and Anand, S. (2020). The role of yeast and molds in dairy industry: An update. *Dairy Processing: Advanced Research to Applications*, 243-262.

Baliyan, S., Mukherjee, R., Priyadarshine, A., Vibhuti, A., Gupta, A., Pandey, R.P. and Chang, C.M. (2022). Determination of antioxidants by DPPH radical scavenging activity and quantitative phytochemical analysis of *Ficus religiosa*. *Molecules*, 27(4), 1326-1345.

Banglao, W., Thongmee, A., Sukplang, P. and Wanakhachornkrai, O. (2020). Determination of antioxidant, anti-aging and cytotoxicity activity of the essential oils from *Cinnamomum zeylanicum*. *Journal of Microbiology, Biotechnology and Food Sciences*, 10 (3), 436-440.

Batiha, G. E. S, Alkazmi, L. M., Wasef, L. G., Beshbishy, A. M., Nadwa, E. H., and Rashwan, E. K. (2020). *Syzygium aromaticum* L.(Myrtaceae): traditional uses, bioactive chemical constituents, pharmacological and toxicological activities. *Biomolecules*, 10(2), 202-209

Bencheraiet, R., Kabouche, A., Kabouche, Z., Touzani, R., and Jay, M. (2012). Flavonol 3-O-Glycosides from three Algerian *Bupleurum* Species. *Records of Natural Products*, 6(2), 11-16

Benedetto, C., D'Auria, M., Mecca, M., Prasad, P., Singh, P., Singh, S., Sinisgalli, C. and Milella, L. (2019). Chemical and biological evaluation of essential oil from *Saussurea costus* (Falc.) Lipsch. from Garhwal Himalaya collected at different harvesting periods. *Natural product research*, 33(16), 2355-2358.

Benjamin, L., Atwill, E.R., Jay-Russell, M., Cooley, M., Carychao, D., Gorski, L. and Mandrell, R.E. (2013). Occurrence of generic *Escherichia coli*, *E. coli* O157 and *Salmonella* spp. in water and sediment from leafy green produce farms and streams on the Central California coast. *International Journal of Food Microbiology*. 165: 65-76.

Bhagwat, V. R. (2019). Safety of water used in food production. In *Food Safety and Human Health* (pp. 219-247). Academic Press.

Bhaskar, A., Sekhar, S., and Javaraiah, R. (2020). Antibiofilm and anti-inflammatory potential of lignans with collegial effect: Secoisolariciresinol diglucoside and sesamin as antimicrobial sources. *Journal of Applied Biology and Biotechnology*, 8(4), 45-51.

Bhat, M.H., Jain, A.K., and Fayaz, M. (2018). Indian herbal drug industry: challenges and future prospects. *Ethnobotany and Physiology*, 1; 1-10

- Bhattacharya, T., Dey, P. S., Akter, R., Kabir, M. T., Rahman, M. H., and Rauf, A. (2021). Effect of natural leaf extracts as phytomedicine in curing geriatrics. *Experimental Gerontology*, 150; 341-352.
- Bintsis, T. (2017). Foodborne pathogens. *AIMS Microbiology*, 3(3): 529–563.
- Birben, E., Sahiner, U.M., Sackesen, C., Erzurum, S. and Kalayci, O. (2012). Oxidative stress and antioxidant defence. *World Allergy Organization Journal*, 5, 9-19.
- Blaszczyk, N., Rosiak, A. and Kałużna-Czaplińska, J. (2021). The potential role of cinnamon in human health. *Forest*, 12(5), 648-655.
- Boateng, L., Ansong, R., Owusu, W., and Steiner-Asiedu, M. (2016). Coconut oil and palm oil's role in nutrition, health and national development: A review. *Ghana Medical Journal*, 50(3), 189-196.
- Bocchi, M. B., Perna, A., Cianni, L., Vitiello, R., Greco, T., Maccauro, G., and Perisano, C. (2020). A rare case of *Bacillus megaterium* soft tissues infection. *Acta Bio Medica: Atenei Parmensis*, 91(14), 1-10.
- Bora, K. S. and Sharma, A. (2011). Evaluation of antioxidant and free-radical scavenging potential of *Artemisia absinthium*. *Pharmaceutical Biology*, 49(12), 1216-1223.
- Bordoloi, M., Saikia, S., Bordoloi, P. K., Kolita, B., Dutta, P. P., Bhuyan, P. D., Dutta, S. C. and Rao, P. G. (2017). Isolation, characterization and antifungal activity of very long chain alkane derivatives from *Cinnamomum obtusifolium*, *Elaeocarpus lanceifolius* and *Baccaurea sapida*. *Journal of Molecular Structure*, 1142, 200-210.
- Botey-Salo, P., Anyogu, A., Varnam, A.H. and Sutherland, J.P. (2012). Survival of inoculated *Salmonella* on the shell of hens' eggs and its potential significance. *Food Control*. 28: 463-469.
- Bottone, E. J. (2010). *Bacillus cereus*, a volatile human pathogen. *Clinical Microbiology Reviews*, 23(2), 382-398.
- Bourhia, M., Laasri, F. E., Moussa, S. I., Ullah, R., Bari, A., Saeed Ali, S., Kaoutar, A., Said, A. A. H., Mzibri, M. E., Said, G., Khilil, N. and Benbacer, L. (2019). Phytochemistry, antioxidant activity, antiproliferative effect, and acute toxicity testing of two Moroccan *Aristolochia* species. *Evidence-Based Complementary and Alternative Medicine*, 2019.
- Brahmachari, G., Das, S., Kumar, A., Misra, N., Sharma, S., and Gupta, V. K. (2017). Structural confirmation, single X-ray crystallographic behavior, molecular docking and other physico-chemical properties of gerberinol, a natural dimethyl dicoumarol from *Gerbera lanuginosa* Benth. (Compositae). *Journal of Molecular Structure*, 1136, 214-221.

- Brodowska, K. M., Brodowska, A. J., Smigielski, K. and Lodyga-Chruscinska, E. (2016). Antioxidant profile of essential oils and extracts of cinnamon bark (*Cinnamomum cassia*). *European Journal of Biological Research*, 6(4), 310-316.
- Burenjargal, M., Narangerel, T., Batmunkh, T., Dong, A., and Idesh, S. (2023). A review of the bioactive properties of Mongolian plants, with a focus on their potential as natural food preservatives. *Food Science and Nutrition*, 1; 1-17
- Burri, S. C., Ekholm, A., Håkansson, Å., Tornberg, E., and Rumpunen, K. (2017). Antioxidant capacity and major phenol compounds of horticultural plant materials not usually used. *Journal of Functional Foods*, 38, 119-127.
- Buru, A. S., Pichika, M. R., Neela, V., and Mohandas, K. (2014). In vitro antibacterial effects of *Cinnamomum* extracts on common bacteria found in wound infections with emphasis on methicillin-resistant *Staphylococcus aureus*. *Journal of Ethnopharmacology*, 153(3), 587-595.
- Butkhup, L., and Samappito, S. (2011). Changes in physico-chemical properties, polyphenol compounds and antiradical activity during development and ripening of maoluang (*Antidesma bunius* L. Spreng) fruits. *Journal of Fruit and Ornamental Plant Research*, 19(1), 85-99.
- Cáceres, L. A., McGarvey, B. D., Briens, C., Berruti, F., Yeung, K. K. C., and Scott, I. M. (2015). Insecticidal properties of pyrolysis bio-oil from greenhouse tomato residue biomass. *Journal of Analytical and Applied Pyrolysis*, 112, 333-340.
- Cai, S., Gao, F., Zhang, X., Wang, O., Wu, W., Zhu, S., Zhang, D., Zhou, F. and Ji, B. (2014). Evaluation of γ -aminobutyric acid, phytate and antioxidant activity of tempeh-like fermented oats (*Avena sativa* L.) prepared with different filamentous fungi. *Journal of Food Science and Technology*, 51, 2544-2551.
- Cao, H., Saroglu, O., Karadag, A., Diaconeasa, Z., Zoccatelli, G., Conte-Junior, C. A., Gonzalez-Aguilar, G. A., O, J., Bai, W., Zamarioli, C. M., Freitas, L. A., Shpigelman, A., Campelo, P. H., Capanoglu, E., Hii, C. L., Jafari, S. M., Qi, Y., Liao, P., Wang, M., Zou, L., Borke, P., Simal-Gandara, J. and Xiao, J. (2021). Available technologies on improving the stability of polyphenols in food processing. *Food Frontiers*, 2(2), 109-139.
- Carballo, A. I., Martínez, A. L., González-Trujano, M. E., Pellicer, F., Ventura-Martínez, R., Díaz-Reval, M. I., and López-Muñoz, F. J. (2010). Antinociceptive activity of *Annona diversifolia* Saff. leaf extracts and palmitone as a bioactive compound. *Pharmacology Biochemistry and Behavior*, 95(1), 6-12.
- Carrillo-Marquez, M. A. (2016). Botulism. *Pediatrics in Review*, 37(5), 183-192.

- Chala, B., and Hamde, F. (2021). Emerging and re-emerging vector-borne infectious diseases and the challenges for control: A review. *Frontiers in Public Health*, 9, 715759, 1-14
- Chanda, S. and Ramachandra, T. V. (2019). A review on some therapeutic aspects of phytochemicals present in medicinal plants. *International Journal of Pharmacy and Life Sciences*, 10(1); 6052-6058.
- Chanda, S., and Rakholiya, K. (2011). Combination therapy: Synergism between natural plant extracts and antibiotics against infectious diseases. *Microbiol Book Series*, 1; 520-529.
- Chandni, T., Priti, M., and Manodeep, S. (2014). Antibacterial compounds in different species of *Datura*: A review. *African Journal of Microbiology Research*, 8(21); 2090-2098.
- Chandra, H., Bishnoi, P., Yadav, A., Patni, B., Mishra, A. P., and Nautiyal, A. R. (2017). Antimicrobial resistance and the alternative resources with special emphasis on plant-based antimicrobials - a review. *Plants*, 6(2); 16-24.
- Chandrasekaran, M., Senthilkumar, A., and Venkatesalu, V. (2011). Antibacterial and antifungal efficacy of fatty acid methyl esters from the leaves of *Sesuvium portulacastrum* L. *European Review for Medical and Pharmacological Sciences*, 15(7), 36-47
- Chang, S. W., Lee, J. S., Lee, J. H., Kim, J. Y., Hong, J., Kim, S. K., Lee, D. and Jang, D. S. (2021). Aromatic and aliphatic apiuronides from the bark of *Cinnamomum cassia*. *Journal of Natural Products*, 84(3), 553-561.
- Chattha, F. A., Munawar, M. A., Nisa, M., Ashraf, M., Kousar, S., and Arshad, S. (2015). Potential antibacterial activity of coumarin and coumarin-3-acetic acid derivatives. *Pakistan Journal of Pharmaceutical Sciences*, 28(3); 819–823.
- Chaturvedi, S., and Sarethy, I. P. (2021). Virtual screening of Compounds from microcolonial fungal strain TD-062 obtained from the Thar Desert of India. *Current Trends in Biotechnology and Pharmacy*, 15(6), 62-66.
- Chelliah, R., Ramakrishnan, S., and Antony, U. (2017). Nutritional quality of *Moringa oleifera* for its bioactivity and antibacterial properties. *International Food Research Journal*, 24(2), 825.
- Chen, H. J., Awakawa, T., Sun, J. Y., Wakimoto, T., and Abe, I. (2013). Epigenetic modifier-induced biosynthesis of novel fusaric acid derivatives in endophytic fungi from *Datura stramonium* L. *Natural Products and Bioprospecting*, 3, 20-23.
- Chen, P. Y., Yu, J. W., Lu, F. L., Lin, M. C. and Cheng, H. F. (2016). Differentiating parts of *Cinnamomum cassia* using LC-qTOF-MS in

conjunction with principal component analysis. *Biomedical Chromatography*, 30(9), 1449-1457.

Chen, T. Y., Tai, S. H., Lee, E. J., Huang, C. C., Lee, A. C., Huang, S. Y., and Wu, T. S. (2011). Cinnamophilin offers prolonged neuroprotection against gray and white matter damage and improves functional and electrophysiological outcomes after transient focal cerebral ischemia. *Critical care medicine*, 39(5), 1130-1137.

Chen, X. M., Ma, Z., & Kitts, D. D. (2018). Effects of processing method and age of leaves on phytochemical profiles and bioactivity of coffee leaves. *Food chemistry*, 249, 143-153.

Chen, X., Ying, X., Chen, L., Zhang, W., and Zhang, Y. (2015). Protective effects of sesamin on liver fibrosis through antioxidative and anti-inflammatory activities in rats. *Immunopharmacology and Immunotoxicology*, 37(5), 465-472.

Chen, X., Zhao, X., Deng, Y., Bu, X., Ye, H., and Guo, N. (2019). Antimicrobial potential of myristic acid against *Listeria monocytogenes* in milk. *The Journal of Antibiotics*, 72(5), 298-305.

Cheng, K. C., Hsueh, M. C., Chang, H. C., Lee, A. Y. L., Wang, H. M., and Chen, C. Y. (2010). Antioxidants from the leaves of *Cinnamomum kotoense*. *Natural Product Communications*, 5(6), 911-912

Chew, K. K., Khoo, M. Z., Ng, S. Y., Thoo, Y. Y., Aida, W. W., and Ho, C. W. (2011). Effect of ethanol concentration, extraction time and extraction temperature on the recovery of phenolic compounds and antioxidant capacity of *Orthosiphon stamineus* extracts. *International Food Research Journal*, 18(4), 1427-1436

Chniguir, A., Zioud, F., Marzaoli, V., El-Benna, J., and Bachoual, R. (2019). *Syzygium aromaticum* aqueous extract inhibits human neutrophils myeloperoxidase and protects mice from LPS-induced lung inflammation. *Pharmaceutical Biology*, 57(1), 55-63.

Cho, H. J., Son, S. H., Chen, W., Son, Y. E., Lee, I., Yu, J. H., and Park, H. S. (2022). Regulation of Conidiogenesis in *Aspergillus flavus*. *Cells*, 11(18), 2796.

Chon, J. W., Kim, Y. J., Kim, D. H., Song, K. Y., Kim, H., and Seo, K. H. (2019). Supplementation of modified mannitol-yolk-polymyxin B agar with cefuroxime for quantitative detection of *Bacillus cereus* in food. *Journal of Food Science*, 84(1), 133-137.

Choudhary, M. I., Musharraf, S. G., Nawaz, S. A., Anjum, S., Parvez, M., and Fun, H. K. (2005). Microbial transformation of isolongifolol and butyrylcholinesterase inhibitory activity of transformed products. *Bioorganic and Medicinal Chemistry*, 13(6), 1939-1944.

- Clarelli, F., Palmer, A., Singh, B., Storflor, M., Lauksund, S., Cohen, T., Abel, S. and Abel zur Wiesch, P. (2020). Drug-target binding quantitatively predicts optimal antibiotic dose levels in quinolones. *PLOS Computational Biology*, 16(8), e1008106, 1-17
- Correa, M.G. and Couto, J.S. (2016). Anticancer properties of *Psidium guajava* - a mini-review. *Asian Pacific Journal of Cancer Prevention*, 17(9), 4199-4204.
- Cortese, M., Gigliobianco, M. R., Magnoni, F., Censi, R., and Di Martino, P. (2020). Compensate for or minimize matrix effects? Strategies for overcoming matrix effects in liquid chromatography-mass spectrometry technique: a tutorial review. *Molecules*, 25(13), 3047.
- Daniel, D.S., Ng, Y.K., Chua, E.L., Arumugam, Y., Wong, Y.L and Kumaran, J.V. (2013). Isolation and identification of gastrointestinal microbiota from the short-nosed fruit bat *Cynopterus brachyotis*. *Microbiology Research*. 168(8): 485-496.
- Daniels, A. O., Temikotan, T., and Ibiyemi, D. A. (2021). Identification and characterization of fatty acids, phytochemical properties and antibacterial effect of the ethyl acetate extract of *Piliostigma reticulatum*. *Journal of Biotechnology and Bioengineering*, 5, 30-40.
- Dar, R. A., Shahnawaz, M. and Qazi, P. H. (2017). General overview of medicinal plants: A review. *The Journal of Phytopharmacology*, 6(6); 349-351
- Das, A. K., Nanda, P. K., Das, A., and Biswas, S. (2019). Hazards and safety issues of meat and meat products. In *Food Safety and Human Health* (pp. 145-168). Academic Press.
- Davis, G. S., and Price, L. B. (2016). Recent research examining links among *Klebsiella pneumoniae* from food, food animals, and human extraintestinal infections. *Current Environmental Health Reports*, 3, 128-135.
- de Jonghe, V., Coorevits, A., Van Hoorde, K., Messens, W., Van Landschoot, A., De Vos, P. and Heyndrickx, M. (2010). Influence of storage conditions on the growth of *Pseudomonas* species in refrigerated raw milk. *Applied and Environmental Microbiology*. 77(2): 460-470.
- De Kok, R. P. J. (2019). A revision of *Cinnamomum* Schaeff. (Lauraceae) for peninsular Malaysia and Singapore. *Gardens' Bulletin Singapore*, 71, 89-139.
- Dehpour, A. A., Yousefian, M., Jafary Kelarjani, S. A., Koshmoo, M., Mirzanegad, S., Mahdavi, V., and Javad Bayani, M. J. (2012). Antibacterial activity and composition of essential oils of flower *Allium rotundum*. *Advances in Environmental Biology*, 6(3), 1020-1025.

- Deng, Q., Zinoviadou, K. G., Galanakis, C. M., Orlie, V., Grimi, N., Vorobiev, E., Lebovka, N. and Barba, F. J. (2015). The effects of conventional and non-conventional processing on glucosinolates and its derived forms, isothiocyanates: extraction, degradation, and applications. *Food Engineering Reviews*, 7, 357-381.
- Devi, K. P., Sakthivel, R., Nisha, S. A., Suganthi, N., and Pandian, S. K. (2013). Eugenol alters the integrity of cell membrane and acts against the nosocomial pathogen *Proteus mirabilis*. *Archives of Pharmacal Research*, 36, 282-292.
- Dhalaria, R., Verma, R., Kumar, D., Puri, S., Tapwal, A., Kumar, V., Nepovimova, E. and Kuca, K. (2020). Bioactive compounds of edible fruits with their anti-aging properties: A comprehensive review to prolong human life. *Antioxidants*, 9(11), 1123-1133
- Dharmayani, N. K. T., Yoshimura, T., Hermawati, E., Juliawaty, L. D., and Syah, Y. M. (2020). Antibacterial and antifungal two phenolic sesquiterpenes from *Dysoxylum densiflorum*. *Zeitschrift für Naturforschung C*, 75(1-2), 1-5.
- Di Sotto, A., Vitalone, A., and Di Giacomo, S. (2020). Plant-derived nutraceuticals and immune system modulation: an evidence-based overview. *Vaccines*, 8(3), 468-477
- Diao, M., Qi, D., Xu, M., Lu, Z., Lv, F., Bie, X., Zhang, C. and Zhao, H. (2018). Antibacterial activity and mechanism of monolauryl-galactosylglycerol against *Bacillus cereus*. *Food Control*, 85, 339-344.
- Dogan, A., Otlu, S., Çelebi, Ö., Aksu, P., Sağlam, A. G., Doğan, A. N. C., and Mutlu, N. (2017). An investigation of antibacterial effects of steroids. *Turkish Journal of Veterinary and Animal Sciences*, 41(2), 302-305.
- Doyle, M. E., Hartmann, F. A. and Wong, A. C. L. (2011). White paper on sources of methicillin-resistant *Staphylococcus aureus* (MRSA) and other methicillin-resistant staphylococci: Implications for our food supply?. *Fri Food Safety Reviews*. Food Research Institute, Madison.
- Driel, B. O. V., Schuldt, M., Algül, S., Levin, E., Güclü, A., Germans, T., Rossum, A. C., Pei, J., Harakalova, M., Baas, A., Jans, J. J. M. and van der Velden, J. (2021). Metabolomics in severe aortic stenosis reveals intermediates of nitric oxide synthesis as most distinctive markers. *International Journal of Molecular Sciences*, 22(7), 3569-3579
- Dumanovic, J., Nepovimova, E., Natic, M., Kuca, K. and Jacevic, V. (2021). The significance of reactive oxygen species and antioxidant defense system in plants: a concise overview. *Frontiers in Plant Science*, 11, 1-13.

- Dziadek, K., Kopeć, A., and Tabaszewska, M. (2019). Potential of sweet cherry (*Prunus avium* L.) by-products: bioactive compounds and antioxidant activity of leaves and petioles. *European Food Research and Technology*, 245(3), 763-772.
- Elamary, R. B., Albarakaty, F. M., and Salem, W. M. (2020). Efficacy of *Acacia nilotica* aqueous extract in treating biofilm-forming and multidrug resistant uropathogens isolated from patients with UTI syndrome. *Scientific Reports*, 10(1), 11125.
- Essid, R., Hammami, M., Gharbi, D., Karkouch, I., Hamouda, T. B., Elkahoui, S., Limam, F. and Tabbene, O. (2017). Antifungal mechanism of the combination of *Cinnamomum verum* and *Pelargonium graveolens* essential oils with fluconazole against pathogenic *Candida* strains. *Applied Microbiology and Biotechnology*, 101(18), 6993-7006.
- Farag, S. M., Essa, E. E., Alharbi, S. A., Alfarraj, S., and El-Hassan, G. A. (2021). Agro-waste derived compounds (flax and black seed peels): Toxicological effect against the West Nile virus vector, *Culex pipiens* L. with special reference to GC–MS analysis. *Saudi Journal of Biological Sciences*, 28(9), 5261-5267.
- Fernandez-Delgado, M., Garcia-Amado, M.A., Contretras, M., Incani, R. N., Chirinos, H., Rojas, H. and Suarez, P. (2015). Survival, induction and resuscitation of *Vibrio cholerae* from the viable but non-culturable state in the Southern Caribbean Sea. *Revista De Medicina Tropical De Sao Paulo*. 57(1): 21-26.
- Fielding, B. C., Mnabisa, A., Gouws, P. A., and Morris, T. (2012). Antimicrobial-resistant *Klebsiella* species isolated from free-range chicken samples in an informal settlement. *Archives of Medical Science*, 8(1), 39-42.
- Figueiredo, A. C., Barroso, J. G., Pedro, L. G., and Scheffer, J. J. (2008). Factors affecting secondary metabolite production in plants: volatile components and essential oils. *Flavour and Fragrance Journal*, 23(4), 213-226.
- Fosgerau, K., and Hoffmann, T. (2015). Peptide therapeutics: current status and future directions. *Drug Discovery Today*, 20(1), 122-128.
- Founou, L. L., Founou, R. C., and Essack, S. Y. (2021). Antimicrobial resistance in the farm-to-plate continuum: More than a food safety issue. *Future science OA*, 7(5), 3-11
- Furneri, P. M., and Bisignano, G. (2010). Usage of hydroxytyrosol for antimycoplasmal activity. In *Olives and olive oil in health and disease prevention*; 1283-1288. Academic Press.

- Gao, Y., Tang, W., Gao, H. E., Chan, E., Lan, J., Li, X., and Zhou, S. (2005). Antimicrobial activity of the medicinal mushroom *Ganoderma*. *Food Reviews International*, 21(2), 211-229.
- Garmana, A. N., Sukandar, E. Y., and Fidrianny, I. (2014). Activity of several plant extracts against drug-sensitive and drug-resistant microbes. *Procedia Chemistry*, 13, 164-169.
- Garnier, L., Valence, F. and Mounier, J. (2017). Diversity and control of spoilage fungi in dairy products: An Update. *Microorganisms*, 5(42): 1-33.
- Ge, Y. C., Zhang, H. J., Wang, K. W., and Fan, X. F. (2018). Aporphine alkaloids from *Illigera aromatica* from guangxi province, China. *Phytochemistry*, 154, 73-76.
- Geetha, R. V. and Roy, A. (2014). Essential oil repellents – a short review. *International Journal of Drug Development and Research*, 6(2); 20-27
- Ghalib, R. M., Hashim, R., Sulaiman, O., Mehdi, S. H., Anis, Z., Rahman, S. Z., Ahame, K. B. M. and Majid, A. M. S. A. (2012). Phytochemical analysis, cytotoxic activity and constituent's activity relationships of the leaves of *Cinnamomum iners* (Reinw. ex Blume Lauraceae). *Natural Product Research*, 26 (22); 2155-2158
- Ghaneian, M. T., Ehrampoush, M. H., Jebali, A., Hekmatimoghaddam, S., and Mahmoudi, M. (2015). Antimicrobial activity, toxicity and stability of phytol as a novel surface disinfectant. *Environmental Health Engineering and Management Journal*, 2(1), 13-16.
- Ghatar, J. M., Ehterami, A., Nazarnezhad, S., Hassani, M. S., Rezaei Kolarijani, N., Mahami, S., and Salehi, M. (2023). A novel hydrogel containing 4-methylcatechol for skin regeneration: in vitro and in vivo study. *Biomedical Engineering Letters*, 1-11.
- Gholamipourfard, K., Salehi, M., & Banchio, E. (2021). *Mentha piperita* phytochemicals in agriculture, food industry and medicine: Features and applications. *South African Journal of Botany*, 141, 183-195.
- Ghosh, T., Basu, A., Adhikari, D., Roy, D., and Pal, A. K. (2015). Antioxidant activity and structural features of *Cinnamomum zeylanicum*. *3 Biotech*, 5, 939-947.
- Glare, T.R., Gartrell, B.D., Brookes, J.J. and Perrot, J.K. (2014). Isolation and identification of *Aspergillus* spp. from brown kiwi (*Apteryx mantelli*) nocturnal house in New Zealand. *Avian Diseases*. 58: 16-24.
- Goh, S. G., Leili, A. H., Kuan, C. H., Loo, Y. Y., Lye, Y. L., San Chang, W., Soopna, P., Najwa, M. S., Tang, J. Y. H., Yaya, R., Nischibuchi, M., Nakaguchi, Y. and Son, R. (2014). Transmission of *Listeria*

- monocytogenes* from raw chicken meat to cooked chicken meat through cutting boards. *Food Control*, 37, 51-55.
- Gonelimali, F. D., Lin, J., Miao, W., Xuan, J., Charles, F., Chen, M., and Hatab, S. R. (2018). Antimicrobial properties and mechanism of action of some plant extracts against food pathogens and spoilage microorganisms. *Frontiers in Microbiology*, 9, 1639-1649
- Goraya, R.K. and Bajwa, U. (2015). Enhancing the functional properties and nutritional quality of ice cream with processed amla (Indian gooseberry). *Journal of Food Science and Technology*, 52(12), 7861-7871.
- Gorniak, I., Bartoszewski, R., and Króliczewski, J. (2019). Comprehensive review of antimicrobial activities of plant flavonoids. *Phytochemistry Reviews*, 18(1); 241-272.
- Gourama, H. (2020). Foodborne pathogens. *Food Safety Engineering*, 25-49.
- Guo, Y., Zhou, H., Qin, L., Pang, Z., Qin, T., Ren, H., Pan, Z. and Zhou, K. (2016). Frequency, antimicrobial resistance and genetic diversity of *Klebsiella pneumoniae* in food samples. *Plos One*. 11(4): 1-13.
- Gupta, A. and Pandey, A. K. (2020). Antibacterial lead compounds and their targets for drug development. In *Phytochemicals as Lead Compounds for New Drug Discovery*, (pp. 275-292). Elsevier
- Gupta, A., Naraniwal, M., and Kothari, V. (2012). Modern extraction methods for preparation of bioactive plant extracts. *International Journal of Applied and Natural Sciences*, 1(1), 8-26.
- Gupta, P. D., and Birdi, T. J. (2017). Development of botanicals to combat antibiotic resistance. *Journal of Ayurveda and Integrative Medicine*, 8(4), 266-275.
- Habbal, O., Hasson, S. S., El-Hag, A. H., Al-Mahrooqi, Z., Al-Hashmi, N., Al-Bimani, Z., and Al-Jabri, A. A. (2011). Antibacterial activity of *Lawsonia inermis* Linn (Henna) against *Pseudomonas aeruginosa*. *Asian Pacific Journal of Tropical Biomedicine*, 1(3), 173-176.
- Hamada, H. M., Awad, M., El-Hefny, M., and Moustafa, M. A. M. (2018). Insecticidal activity of garlic (*Allium sativum*) and ginger (*Zingiber officinale*) oils on the cotton leafworm (*Spodoptera littoralis*) (Boisd.) (Lepidoptera: Noctuidae). *African Entomology*, 26(1), 84-94.
- Hamidi, R., Jovanova, B. and Panovska, T.K. (2014). Toxicological evaluation of the plant products using brine shrimp (*Artemia salina* L.) model. *Macedonian Pharmaceutical Bulletin*, 60(1), 9-18.

- Haq, I. U., Imran, M., Nadeem, M., Tufail, T., Gondal, T. A., and Mubarak, M. S. (2021). Piperine: A review of its biological effects. *Phytotherapy Research*, 35(2), 680-700.
- Haque, M. H., Sarker, S., Islam, M. S., Islam, M. A., Karim, M. R., Kayesh, M. E. H., Shiddiky, M. J. A. & Anwer, M. S. (2020). Sustainable antibiotic-free broiler meat production: Current trends, challenges, and possibilities in a developing country perspective. *Biology*, 9(11), 411.
- Hartantyo, S. H. P., Chau, M. L., Koh, T. H., Yap, M., Yi, T., Cao, D. Y. H., Gutiérrez, R. A. and Ng, L. C. (2020). Foodborne *Klebsiella pneumoniae*: virulence potential, antibiotic resistance, and risks to food safety. *Journal of food Protection*, 83(7), 1096-1103.
- Hassanpour, S., Maherisis, N., and Eshratkhah, B. (2011). Plants and secondary metabolites (tannins): a review. *International Journal of Forest, Soil and Erosion*, 1;1-10
- Henry, G. E., Momin, R. A., Nair, M. G., and Dewitt, D. L. (2002). Antioxidant and cyclooxygenase activities of fatty acids found in food. *Journal of Agricultural and Food Chemistry*, 50(8), 2231-2234.
- Hifnawy, M. S., Salam, R. M. A., Rabeh, M. A., and Aboseada, M. A. (2013). Glucosinolates, glycosidically bound volatiles and antimicrobial activity of *Brassica oleraceae* var. botrytis,(sultany cultivar). *Archives of Biochemistry and Biophysics*, 3, 66-81.
- Hikal, D. M. (2018). Antibacterial activity of piperine and black pepper oil. *Biosciences Biotechnology Research Asia*, 15(4), 877-884
- Hintz, T., Matthews, K. K., and Di, R. (2015). The use of plant antimicrobial compounds for food preservation. *BioMed Research International*, 2015, 1-10
- Hong, L. S., Ibrahim, D., Kassim, J., and Sulaiman, S. (2011). Gallic acid: an anticandidal compound in hydrolysable tannin extracted from the barks of *Rhizophora apiculata* Blume. *Journal of Applied Pharmaceutical Science*, 1(6), 75-79.
- Hosseinzadeh, S., Jafarikukhdan, A., Hosseini, A., and Armand, R. (2015). The application of medicinal plants in traditional and modern medicine: a review of *Thymus vulgaris*. *International Journal of Clinical Medicine*, 6(09), 635-641.
- Hsiao, G., Teng, C. M., Sheu, J. R., Cheng, Y. W., Lam, K. K., Lee, Y. M., We, T. S and Yen, M. H. (2001). Cinnamophilin as a novel antiperoxidative cytoprotectant and free radical scavenger. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1525(1-2), 77-88.

- Huang, Y. C., Wu, B. H., Chu, Y. L., Chang, W. C., and Wu, M. C. (2018). Effects of tempeh fermentation with *Lactobacillus plantarum* and *Rhizopus oligosporus* on streptozotocin-induced type II diabetes mellitus in rats. *Nutrients*, 10(9), 1143-1152
- Hussain, H. I., Kasinadhuni, N., and Arioli, T. (2021). The effect of seaweed extract on tomato plant growth, productivity and soil. *Journal of Applied Phycology*, 33(2), 1305-1314.
- Hussain, S. Z., and Maqbool, K. (2014). GC-MS: principle, technique and its application in food science. *International Journal of Current Science*, 13, 116-126.
- Hussein, S., Abdel-Aal, S., and Emam, S. (2019). Phytochemical analysis and Antiproliferative activity of *Lotus peregrinus* L. against MCF7 (breast carcinoma cell line). *Benha Veterinary Medical Journal*, 36(1), 351-360.
- Ibrahim, S. A., Ayivi, R. D., Zimmerman, T., Siddiqui, S. A., Altemimi, A. B., Fidan, H., Esatbeyoglu, T. and Bakhshayesh, R. V. (2021). Lactic acid bacteria as antimicrobial agents: Food safety and microbial food spoilage prevention. *Foods*, 10(12), 3131-3140
- Ifeanyi, O.E. (2018). A review on free radicals and antioxidants. *International Journal of Current Research in Medical Sciences*, 4(2), 123-133.
- Inkson, B. J. (2016). Scanning electron microscopy (SEM) and transmission electron microscopy (TEM) for materials characterization. In *Materials characterization using nondestructive evaluation (NDE) methods* (pp. 17-43). Woodhead publishing.
- Jaganathan, S. K., and Supriyanto, E. (2012). Antiproliferative and molecular mechanism of eugenol-induced apoptosis in cancer cells. *Molecules*, 17(6), 6290-6304.
- Jain, C., Khatana, S. and Vijayvergia, R. (2019). Bioactivity of secondary metabolites of various plants: a review. *International Journal of Pharmaceutical Sciences and Research*. 10(2). 494-498
- Jalalvand, A. R., Zhaleh, M., Goorani, S., Zangeneh, M. M., Seydi, N., Zangeneh, A., and Moradi, R. (2019). Chemical characterization and antioxidant, cytotoxic, antibacterial, and antifungal properties of ethanolic extract of *Allium saralicum* RM Fritsch leaves rich in linolenic acid, methyl ester. *Journal of Photochemistry and Photobiology B: Biology*, 192, 103-112.
- Jamil, N. and Pa'ee, F. (Eds.). (2018). Proceedings from *AIP Conference Proceedings*. Antimicrobial activity from leaf, flower, stem, and root of *Clitoria ternatea* – a review.

- Jang, J., Hur, H. G., Sadowsky, M. J., Byappanahalli, M. N., Yan, T., and Ishii, S. (2017). Environmental *Escherichia coli*: ecology and public health implications - a review. *Journal of Applied Microbiology*, 123(3), 570-581.
- Jiang, S., Liu, H. and Li, C. (2021). Dietary regulation of oxidative stress in chronic metabolic diseases. *Foods*, 10(8), 1854-1872.
- Jirakitticharoen, S., Wisuitiprot, W., Jitareerat, P., and Wongs-Aree, C. (2022). Phenolics, antioxidant and antibacterial activities of immature and mature *Blumea balsamifera* leaf extracts eluted with different solvents. *Journal of Tropical Medicine*, 2022, 1-10
- Jokić, S., Nagy, B., Zeković, Z., Vidović, S., Bilić, M., Velić, D., and Simándi, B. (2012). Effects of supercritical CO₂ extraction parameters on soybean oil yield. *Food and Bioproducts Processing*, 90(4), 693-699.
- Jualang, A. G., Azlinah, M., Lee, P. C., and How, S. E. (2013). Antibacterials and phytochemicals investigations of *Chromolaena odorata* (L.F) King and Robinson (asteraceae) from Sabah, Malaysia. *Journal of Environmental Microbiology and Toxicology*, 1(1), 18-24.
- Kaczmarek, M., Avery, S. V., and Singleton, I. (2019). Microbes associated with fresh produce: Sources, types and methods to reduce spoilage and contamination. In *Advances in Applied Microbiology* (107), (pp. 29-82). Academic Press.
- Kaderi, K. F., Islam, M. S., Sarker, T., Islam, S. S., Mondol, D., Faruk, M. O., and Biswas, G. (2023). Study on the efficacy of green tea extract on *Staphylococcus aureus* and *Klebsiella pneumoniae* of raw chicken meat. *Meat Research*, 3(1), 1-10.
- Kai, M. (2020). Diversity and distribution of volatile secondary metabolites throughout *Bacillus subtilis* isolates. *Frontiers in Microbiology*, 11, 559-565
- Kamaliroosta, L., Gharachorloo, M., Kamaliroosta, Z., and Alimohammad Zadeh, K. H. (2012). Extraction of cinnamon essential oil and identification of its chemical compounds. *Journal Medicinal Plants Research*, 6(4), 609-614.
- Kamble, A., Naik, S., Talathi, M., Jadhav, D., Pingale, S., and Kaul-Ghanekar, R. (2022). Cervicovaginal microflora isolated from healthy women exhibited probiotic properties and antimicrobial activity against pathogens isolated from cervical cancer patients. *Archives of Microbiology*, 204(8), 491.
- Kanatt, S. R. (2020). Development of active/intelligent food packaging film containing *Amaranthus* leaf extract for shelf-life extension of chicken/fish during chilled storage. *Food Packaging and Shelf Life*, 24, 100506, 1-15.

- Kebede, M. and Emire, S.A. (2019). Application of antioxidant in food processing, industry: options to improve the extraction yields and market value of natural products. *Advances in Food Technology and Nutritional Sciences*, 5(2), 38-48.
- Khameneh, B., Iranshahy, M., Soheili, V., and Fazly Bazzaz, B. S. (2019). Review on plant antimicrobials: a mechanistic viewpoint. *Antimicrobial Resistance and Infection Control*, 8(1), 1-28.
- Khan, Z. A., Siddiqui, M. F., and Park, S. (2019). Current and emerging methods of antibiotic susceptibility testing. *Diagnostics*, 9(2), 49-56
- Khaneghah, A. M., Abhari, K., Eş, I., Soares, M. B., Oliveira, R. B., Hosseini, H., Mohammad Rezaei, Balthazar, C. F., Silva, R., Cruz, A. G., Ranadheera, C. S and Sant'Ana, A. S. (2020). Interactions between probiotics and pathogenic microorganisms in hosts and foods: A review. *Trends in Food Science and Technology*, 95, 205-218.
- Kim, M. J., Moon, Y., Tou, J. C., Mou, B., and Waterland, N. L. (2016). Nutritional value, bioactive compounds and health benefits of lettuce (*Lactuca sativa* L.). *Journal of Food Composition and Analysis*, 49, 19-34.
- Kim, W. J., Lee, K. A., Kim, K. T., Chung, M. S., Cho, S. W., and Paik, H. D. (2011). Antimicrobial effects of onion (*Allium cepa* L.) peel extracts produced via subcritical water extraction against *Bacillus cereus* strains as compared with ethanolic and hot water extraction. *Food Science and Biotechnology*, 20, 1101-1106.
- Kirakosyan, A., Seymour, E. M., Noon, K. R., Llanes, D. E. U., Kaufman, P. B., Warber, S. L., and Bolling, S. F. (2010). Interactions of antioxidants isolated from tart cherry (*Prunus cerasus*) fruits. *Food Chemistry*, 122(1), 78-83.
- Kiran, V., Vivek, T., and Joshi, V. K. (2018). Gas chromatography-mass spectroscopic (GC-MS) analysis of Gandharvahasta-taila: a popular compound formulation of Yurveda. *International Journal of Current Research in Life Sciences*, 7(03), 2024-2028.
- Kirillov, V., Stikhareva, T., Suleimen, Y., Serafimovich, M., Kabanova, S., and Mukanov, B. (2017). Chemical composition of the essential oil from carnation coniferous (*Dianthus acicularis* Fisch. ex Ledeb) growing wild in Northern Kazakhstan. *Natural Product Research*, 31(1), 117-123.
- Kiryu, M., Hamanaka, M., Yoshitomi, K., Mochizuki, S., Akimitsu, K., and Gomi, K. (2018). Rice terpene synthase 18 (OsTPS18) encodes a sesquiterpene synthase that produces an antibacterial (E)-nerolidol against a bacterial pathogen of rice. *Journal of General Plant Pathology*, 84, 221-229.

- Kohlerschmidt, D. J., Mingle, L. A., Dumas, N. B., and Nattanmai, G. (2021). Identification of aerobic Gram-negative bacteria. In *Practical handbook of microbiology* (pp. 59-70). CRC Press.
- Kristiningrum, N., Cahyanti, Y. N., and Wulandari, L. (2017). Determination of total phenolic content and antioxidant activity in methanolic extract of Robusta and Arabica coffee leaves. *UNEJ e-Proceeding*, 96-99.
- Kumar, A. (2018). Important for life '*Psidium guajava*'. *International Journal of Pharmacological and Bio Sciences*, 2(1): 53-69.
- Kumar, S., and Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*, 2013, 1-14
- Kumar, S., Arif, M., Jawaid, T., Al-Khamees, O. A., Anjum, A., Shafi, S., Thirunavukkarasu, V., Josephine, S. P., Muteeb, G., Singh, K. and Qadir, A. (2023). Antiplatelet and thrombolytic activity of phenolic-insistent fractions from the new-fangled stem of *Ficus benghalensis* with concurrent GC-MS analysis. In *Intelligent Pharmacy* (pp. 34-65). KeAi Press.
- Kumar, S., Kumari, R. and Mishra, S. (2019). Pharmacological properties and their medicinal uses of *Cinnamomum*; a review. *Journal of Pharmacy and Pharmacology*, 71; 1735-1761
- Kurek, A. N. N. A., Grudniak, A. M., Kraczkiewicz-Dowjat, A., and Wolska, K. I. (2011). New antibacterial therapeutics and strategies. *Polish Journal of Microbiology*, 60(1), 3-11.
- Kwan (2019). www.natureloveyou.sg/Cinnamomum%20iners/Main.html. Received date: 20 July 2023
- Langat, M. K., Mayowa, Y., Sadgrove, N., Danyaal, M., Prescott, T. A., Kami, T., Schwikkard, S., Barker, J. and Cheek, M. (2022). Multi-layered antimicrobial synergism of (E)-caryophyllene with minor compounds, tecleanatalensine B and normelicopine, from the leaves of *Vepris gossweileri* (l. Verd.) Mziray. *Natural Product Research*, 36(10), 2447-2457.
- Lee, H. Y., Park, S. H., Lee, M., Kim, H. J., Ryu, S. Y., Kim, N. D., Hwang, B. Y., Hong, J. T., Han, S. B. and Kim, Y. (2012). 1-Dehydro-[10]-gingerdione from ginger inhibits IKK β activity for NF- κ B activation and suppresses NF- κ B-regulated expression of inflammatory genes. *British Journal of Pharmacology*, 167(1), 128-140.
- Lee, S. H., Chang, K. S., Su, M. S., Huang, Y. S., and Jang, H. D. (2007). Effects of some Chinese medicinal plant extracts on five different fungi. *Food Control*, 18(12), 1547-1554.

- Lee, W., Woo, E. R., and Lee, D. G. (2016). Phytol has antibacterial property by inducing oxidative stress response in *Pseudomonas aeruginosa*. *Free Radical Research*, 50(12), 1309-1318.
- Li, J., Zhou, L. U., Long, C., Fang, L., Chen, Q., Chen, Q., Liang, J., Yang, Y., Zhu, H., Chen, Z., Gao, S., Li, Z., Li, Q., Huang, Q, and Zhang, Y. (2019). An investigation of bongkreik acid poisoning caused by consumption of a nonfermented rice noodle product without noticeable signs of spoilage. *Journal of Food Protection*, 82(10), 1650-1654.
- Li, W., Jiang, Y., Hu, C., Liu, G., Li, Y., and Wang, S. (2023). Identification, pathogenic mechanism and control of *Rhizopus oryzae* causing postharvest fruit rot in pumpkin. *Postharvest Biology and Technology*, 204, 112460.
- Li, Y., Kong, D., Fu, Y., Sussman, M. R., and Wu, H. (2020). The effect of developmental and environmental factors on secondary metabolites in medicinal plants. *Plant Physiology and Biochemistry*, 148, 80-89.
- Liang, D., Feng, B., Li, N., Su, L., Wang, Z., Kong, F. and Bi, Y. (2022). Preparation, characterization, and biological activity of *Cinnamomum cassia* essential oil nano-emulsion. *Ultrasonics Sonochemistry*, 86, 106009, 1-10
- Lim, T. K. (2013). *Lonicera japonica*. In *Edible Medicinal and Non-Medicinal Plants: Volume 7, Flowers* (pp. 656-680). Dordrecht: Springer Netherlands.
- Limna Mol, V. P., Abdulaziz, A., Sneha, K. G., Praveen, P. J., Raveendran, T. V., and Parameswaran, P. S. (2020). Inhibition of pathogenic *Vibrio harveyi* using calamenene, derived from the Indian gorgonian *Subergorgia reticulata*, and its synthetic analog. *3 Biotech*, 10, 1-7.
- Liu, C. H., and Huang, H. Y. (2012). Antimicrobial activity of curcumin-loaded myristic acid microemulsions against *Staphylococcus epidermidis*. *Chemical and Pharmaceutical Bulletin*, 60(9), 1118-1124.
- Liu, T. Z., Cheng, J. T., Yiin, S. J., Chen, C. Y., Chen, C. H., Wu, M. J., and Chern, C. L. (2008). Isoobtusilactone A induces both caspase-dependent and-independent apoptosis in Hep G2 cells. *Food and Chemical Toxicology*, 46(1), 321-327.
- Londoño-Hernández, L., Ramírez-Toro, C., Ruiz, H. A., Ascacio-Valdés, J. A., Aguilar-Gonzalez, M. A., Rodríguez-Herrera, R., and Aguilar, C. N. (2017). *Rhizopus oryzae*—Ancient microbial resource with importance in modern food industry. *International Journal of Food Microbiology*, 257, 110-127.
- Lopez-Brea, S. G., Gómez-Torres, N., and Arribas, M. Á. (2017). Spore-forming bacteria in dairy products. *Microbiology in Dairy Processing: Challenges and Opportunities*, 11-36.

- López-Campos, G., Martínez-Suárez, J. V., Aguado-Urda, M., and López-Alonso, V. (2012). Detection, identification, and analysis of foodborne pathogens. *Microarray Detection and Characterization of Bacterial Foodborne Pathogens*, 13-32
- Lopez-Valladares, G., Danielsson-Tham, M. L., and Tham, W. (2018). Implicated food products for listeriosis and changes in serovars of *Listeria monocytogenes* affecting humans in recent decades. *Foodborne Pathogens and Disease*, 15(7), 387-397.
- Lourenco, S.C., Moldao-Martins, M. and Alves, V.D. (2019). Antioxidants of natural plant origins: from sources to food industry applications. *Molecules*, 24(22), 4132-4168.
- Lu, Z., Zhu, L., Lu, J., Shen, N., Wang, L., Liu, S., Wang, Q., Yu, W., kato-Naguchi, H., Li, W., Jin, B., Wang, L. & Lin, J. (2022). Rejuvenation increases leaf biomass and flavonoid accumulation in *Ginkgo biloba*. *Horticulture Research*, 9, uhab018.
- Lübeck, M., and Lübeck, P. S. (2022). Fungal cell factories for efficient and sustainable production of proteins and peptides. *Microorganisms*, 10(4), 753.
- Lv, Y., Cheng, X., Wu, D., Du, G., Zhou, J., and Chen, J. (2018). Improving bioconversion of eugenol to coniferyl alcohol by in situ eliminating harmful H₂O₂. *Bioresource technology*, 267, 578-583.
- Ma, M., Wen, X., Xie, Y., Guo, Z., Zhao, R., Yu, P., Gong, D., Deng, S. and Zeng, Z. (2018). Antifungal activity and mechanism of monocaprin against food spoilage fungi. *Food Control*, 84, 561-568.
- Maffei, M. E., Gertsch, J., and Appendino, G. (2011). Plant volatiles: production, function and pharmacology. *Natural Product Reports*, 28(8), 1359-1380.
- Mailafiya, M. M., Yusuf, A. J., Abdullahi, M. I., Aleku, G. A., Ibrahim, I. A., Yahaya, M., Abubakar, H., Sanusi, A., Adamu, H. W. and Alebiosu, C. O. (2018). Antimicrobial activity of stigmasterol from the stem bark of *Neocarya macrophylla*. *Journal of Medicinal Plants for Economic Development*, 2(1), 1-5.
- Mailoa, M. N., Mahendradatta, M., Laga, A., and Djide, N. (2014). Antimicrobial activities of tannins extract from guava leaves (*Psidium guajava* L.) on pathogens microbial. *International Journal of Scientific and Technology Research*, 3(1), 236-241.
- Majdalawieh, A. F., Yousef, S. M., Abu-Yousef, I. A., and Nasrallah, G. K. (2022). Immunomodulatory and anti-inflammatory effects of sesamin: Mechanisms of action and future directions. *Critical Reviews in Food Science and Nutrition*, 62(18), 5081-5112.

- Makwana, S., Choudhary, R., Haddock, J., and Kohli, P. (2015). *In-vitro* antibacterial activity of plant based phenolic compounds for food safety and preservation. *LWT-Food Science and Technology*, 62(2), 935-939.
- Mandal, S. and Shi, S. Q. (2020). Agricultural plants and their antimicrobial activities - a mini review. *American Journal of Biomedical Science and Research*, 11(2); 179-181
- Manges, A. R. (2016). *Escherichia coli* and urinary tract infections: the role of poultry-meat. *Clinical Microbiology and Infection*, 22(2), 122-129.
- Manousi, N., Sarakatsianos, I., and Samanidou, V. (2019). Extraction techniques of phenolic compounds and other bioactive compounds from medicinal and aromatic plants. *In Engineering tools in the beverage industry* (pp. 283-314). Woodhead Publishing.
- Mariyammal, V., Sathiageetha, V., Amalraj, S., Gurav, S. S., Amiri-Ardekani, E., Jeeva, S., and Ayyanar, M. (2023). Chemical profiling of *Aristolochia tagala* Cham. leaf extracts by GC-MS analysis and evaluation of its antibacterial activity. *Journal of the Indian Chemical Society*, 100(1), 1008, 1-10
- Maroszynska, M., Kunicka-Styczynska, A., Rajkowska, K., Maroszynska, I. (2013). Antibiotics sensitivity of *Candida* clinical and food-borne isolates. *Acta Biochimica Polonica*. 60(4): 719-724.
- Marrez, D. A., Naguib, M. M., Sultan, Y. Y., and Higazy, A. M. (2019). Antimicrobial and anticancer activities of *Scenedesmus obliquus* metabolites. *Heliyon*, 5(3), 1-11.
- Mathew, S., Abraham, T.E. and Zakaria, Z.A. (2015). Reactivity of phenolic compounds towards free radicals under *in vitro* conditions. *Journal of Food Science and Technology*, 52, 5790-5798.
- Mathur, A., Prasad, G. B. K. S., Rao, N., Babu, P., and Dua, V. K. (2011). Isolation and identification of antimicrobial compounds from *Mentha piperita*. *Rasayan Journal of Chemistry*, 4(1), 36-42.
- Mehdizadeh, T., and Langroodi, A. M. (2019). Chitosan coatings incorporated with propolis extract and *Zataria multiflora* Boiss oil for active packaging of chicken breast meat. *International Journal of Biological Macromolecules*, 141, 401-409.
- Mendiola, J. A., Santoyo, S., Cifuentes, A., Reglero, G., Ibanez, E., and Señoráns, F. J. (2008). Antimicrobial activity of sub-and supercritical CO₂ extracts of the green alga *Dunaliella salina*. *Journal of Food Protection*, 71(10), 2138-2143.
- Mendonca, A., Thomas-Popo, E., and Gordon, A. (2020). Microbiological considerations in food safety and quality systems implementation. In *Food*

safety and quality systems in developing countries (pp. 185-260). Academic Press.

Mesele, F., and Abunna, F. (2019). *Escherichia coli* O157:H7 in foods of animal origin and its food safety implications. *Advance Biological Research*, 13(4), 134-145.

Messele, Y. E., Abdi, R. D., Yalew, S. T., Tegegne, D. T., Emeru, B. A., and Werid, G. M. (2017). Molecular determination of antimicrobial resistance in *Escherichia coli* isolated from raw meat in Addis Ababa and Bishoftu, Ethiopia. *Annals of Clinical Microbiology and Antimicrobials*, 16(1), 1-9.

Miksusanti, H., and Masril, K. I. (2013). Antibacterial and antioxidant of uwi (*Dioscorea alata* L.) starch edible film incorporated with ginger essential oil. *International Journal of Bioscience, Biochemistry and Bioinformatics*, 3(4), 354-356.

Milaciu, M. V., Ciumărnean, L., Orășan, O. H., Para, I., Alexescu, T., and Negrean, V. (2016). Semiology of food poisoning. *Human and Veterinary Medicine*, 8(2), 108-113.

Milind, P., and Deepa, K. (2011). Clove: a champion spice. *International Journal of Ayurveda Research*, 2(1), 47-54.

Mirabile, G., Bella, P., Vella, A., Ferrantelli, V. and Torta, L. (2021). Fungal contaminants and mycotoxins in nuts. *Nuts and Nut Products in Human Health and Nutrition*, 146-165.

Mirza, S. K., Asema, U. K., and Kasim, S. S. (2017). To study the harmful effects of food preservatives on human health. *Journal Medicinal Chemistry and Drug Discovery*, 2, 610-616.

Mitomo, S., Hirota, M., and Fujita, T. (2019). New xanthine oxidase inhibitors from the fruiting bodies of *Tyromyces fissilis*. *Bioscience, Biotechnology, and Biochemistry*, 83(5), 813-823.

Mittal, M., Gupta, N., Parashar, P., Mehra, V., and Khatri, M. (2014). Phytochemical evaluation and pharmacological activity of *Syzygium aromaticum*: a comprehensive review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(8), 67-72.

Mlala, S., Oyediji, A. O., Gondwe, M., and Oyediji, O. O. (2019). Ursolic acid and its derivatives as bioactive agents. *Molecules*, 24(15), 2751.

Mohammed, E. T. and Mustafa, Y. F (2020). Coumarins from Red Delicious apple seeds: extraction, phytochemical analysis, and evaluation as antimicrobial agents. *Systematic Reviews in Pharmacy*, 11(2): 64-70

Moini Jazani, A., Nasimi Doost Azgomi, H., Nasimi Doost Azgomi, A., and Nasimi Doost Azgomi, R. (2019). A comprehensive review of clinical

- studies with herbal medicine on polycystic ovary syndrome (PCOS). *DARU Journal of Pharmaceutical Sciences*, 27, 863-877.
- Monkai, J., Chukeatirote, E., Chamyuang, S., Synytsya, A., Ruml, T., and Hyde, K. D. (2013). Antimicrobial activity of some saprobic fungi isolated from *Magnolia liliifera* and *Cinnamomum iners* leaves. *Mycology*, 4(2), 82-84.
- Monkolval, P., Insuwan, W., Nanuam, J., Bunjan, K., Phosai, W., Thanomsit, C., Wattanakornsiri A and Phimchan, P. (2020). The phytochemical and protein extraction of Golden shower tree (*Cassia fistula*). *Naresuan University Journal: Science and Technology*, 29(1), 21-35.
- Mostafa, A. A., Al-Askar, A. A., Almaary, K. S., Dawoud, T. M., Sholkamy, E. N., and Bakri, M. M. (2018). Antimicrobial activity of some plant extracts against bacterial strains causing food poisoning diseases. *Saudi Journal of Biological Sciences*, 25(2), 361-366
- Mouahid, A., Dufour, C., and Badens, E. (2017). Supercritical CO₂ extraction from endemic Corsican plants; comparison of oil composition and extraction yield with hydrodistillation method. *Journal of CO₂ Utilization*, 20, 263-273.
- Mrityunjoy, A., Kaniz, F., Fahmida J., Shanzida, J.S., Md. Aftab, U. and Rashed, N. Prevalence of *Vibrio cholerae* in different food samples in the city of Dhaka, Bangladesh. *International Food Research Journal*. 20(2): 1017-1022.
- Mughini-Gras, L., Enserink, R., Friesema, I., Heck, M., van Duynhoven, Y., and van Pelt, W. (2014). Risk factors for human salmonellosis originating from pigs, cattle, broiler chickens and egg laying hens: a combined case-control and source attribution analysis. *PloS one*, 9(2), e87933.
- Muhammad, D. R. A., and Dewettinck, K. (2017). Cinnamon and its derivatives as potential ingredient in functional food—A review. *International Journal of Food Properties*, 20(2), 2237-2263.
- Mukherjee, S., and Ghorai, S. (2023). Fungal biology. In *Current Developments in Biotechnology and Bioengineering* (pp. 67-104). Elsevier.
- Mukherji, R., and Prabhune, A. (2015). A new class of bacterial quorum sensing antagonists: glycomonoterpenols synthesized using linalool and alpha terpineol. *World Journal of Microbiology and Biotechnology*, 31, 841-849.
- Mustaffa, F., Hassan, Z. and Asmawi, M. Z. (2016). *Cinnamomum iners* leaves as an alternative therapy for diabetes. *Asian Journal of Biochemistry*, 11: 44-52.

- Mustaffa, F., Hassan, Z., Yusof, N. A., Razak, K. N. A. and Asmawi, M. Z. (2014). Antidiabetic mechanism of standardized extract, fraction and subfraction of *Cinnamomum iners* leaves. *International Journal of Pharmaceutical Sciences Review and Research*, 26(2); 209-214
- Mustaffa, F., Indurkar, J., Ismail, S., Mordi, M.N., Ramanathan, S., and Mansor, S.M. (2010). Antioxidant capacity and toxicity screening of *Cinnamomum iners* standardized leaves methanolic extract. *International Journal of Pharmacology*, 6, 888–895.
- Mustaffa, F., Indurkar, J., Ismail, S., Shah, M. and Mansor, S.M. (2011). An antimicrobial compound isolated from *Cinnamomum iners* leaves with activity against methicillin-resistant *Staphylococcus aureus*. *Molecules*, 16(4), 3037-3047.
- Mustaffa, F., Indurkar, J., Shah, M., Ismail, S. and Mansor, S. M. (2012). Review on pharmacological activities of *Cinnamomum iners* Reinw. ex Blume. *Natural Product Research*; 27(10); 888-895
- Mustaffa, I. N., Ramle, S. F. M., Adenam, N. M., Awalludin, M. F., Zaudin, N. A. C., Hamid, Z. A. A. and Hermawan, A. (2020). Potential of *Cinnamomum iners* wood as antimicrobial agent. *International Conference on Science and Technology*, 596; 1-10
- N'guessan, E., Bakayoko, S., Cisse, M., Dalie, W., and Sindic, M. (2019). Prevalence of *Bacillus cereus* and emetic strains detection from Ivory Coast local flours. *Agronomie Africaine*, 8(1), 151-159.
- Naik, B., Maurya, V. K., Kumar, V., Kumar, V., Upadhyay, S., and Gupta, S. (2021). Phytochemical analysis of *Diplazium esculentum* reveals the presence of medically important components. *Current Nutrition and Food Science*, 17(2), 210-215.
- Nam, K. C., Jo, C., and Lee, M. (2010). Meat products and consumption culture in the East. *Meat Science*, 86(1), 95-102.
- Narwade, D. A., and Aher, A. N. (2019). A review on: phytochemical and pharmacological study of *Annona Squamosa*. *PharmaTutor*, 7(1), 5-10.
- Naz, G., Anjum, A. A., Nawaz, M., Iqbal, S., Azeem, S., Ali, T., and Manzoor, R. (2023). Evaluation of *Cinnamomum verum* essential oils against ochratoxin a-producing *Aspergillus parasiticus* in stored wheat, maize and rice. *Polish Journal of Environmental Studies*, 32(1), 1-12
- Nazzaro, F., Fratianni, F., Coppola, R., and De Feo, V. (2017). Essential oils and antifungal activity. *Pharmaceuticals*, 10(4), 86-94.
- Nelson, E.J., Harris, J.B., Jr, J.G., Calderwood, S.B. and Camilli, A. (2013). Cholera transmission: the host, pathogen and bacteriophage dynamic. *Nature Review Microbiology*. 7(10): 1-24.

- Newell, D. G., Koopmans, M., Verhoef, L., Duizer, E., Aidara-Kane, A., Sprong, H., Opsteegh, M., Langelaar, M., Threlfall, J., Scheutz, F., Giessen J. and Kruse, H. (2010). Food-borne diseases - the challenges of 20 years ago still persist while new ones continue to emerge. *International Journal of Food Microbiology*, 139, 3-15.
- Ngamakeue, N., and Chitprasert, P. (2016). Encapsulation of holy basil essential oil in gelatin: Effects of palmitic acid in carboxymethyl cellulose emulsion coating on antioxidant and antimicrobial activities. *Food and Bioprocess Technology*, 9, 1735-1745.
- Nieto, G. (2017). Biological activities of three essential oils of the Lamiaceae family. *Medicines*, 4(3), 63-74
- Niksic, H., Becic, F., Koric, E., Gusic, I., Omeragic, E., Muratovic, S., Miladinovic, B. and Duric, K. (2021). Cytotoxicity screening of *Thymus vulgaris* L. essential oil in brine shrimp nauplii and cancer cell lines. *Scientific Reports*, 11(1), 13178-11188
- Nishad, J. (2022). Stability of plant extracts. In *Plant Extracts: Applications in the Food Industry* (pp. 89-126). Academic Press.
- Nishiumi, S., Miyamoto, S., Kawabata, K., Ohnishi, K., Mukai, R., Murakami, A. Ashida, H. and Terao, J. (2011). Dietary flavonoids as cancer-preventive and therapeutic biofactors. *Frontiers in Bioscience*, 3, 1332-62.
- Nobosse, P., Fombang, E.N. and Mbofung, C.M.F. (2018). Effects of age and extraction solvent on phytochemical content and antioxidant activity of fresh *Moringa oleifera* L. leaves. *Food Science and Nutrition*, 6(8), 2188-2198.
- Nogueira, G. F., Oliveira, R. A. D., Velasco, J. I. and Fakhouri, F. M. (2020). Methods of incorporating plant-derived bioactive compounds into films made with agro-based polymers for application as food packaging: A brief review. *Polymers*, 12(11), 2518-2526
- Nonglang, F. P., Khale, A., Wankhar, W., and Bhan, S. (2022). Pharmacognostic evaluation of *Eranthemum indicum* extracts for its in-vitro antioxidant activity, acute toxicology, and investigation of potent bioactive phytochemicals using HPTLC and GCMS. *Beni-Suef University Journal of Basic and Applied Sciences*, 11(1), 1-17.
- Noorlis, A., Ghazali, F. M., Cheah, Y. K., Tuan Zainazor, T. C., Ponniah, J., Tunung, R., Tang, J. Y. H., Nishibuchi, M., Nakaguchi, Y. and Son, R. (2012). Prevalence and quantification of *Vibrio* species and *Vibrio parahaemolyticus* in freshwater fish at hypermarket level. *International Food Research Journal*. 18: 689-695.
- Nugraha, A., & Nandiyanto, A. B. D. (2021). How to read and Interpret GC/MS spectra. *Indonesian Journal of Multidisciplinary Research*, 1(2), 171-206.

- Nyati, K. K., and Nyati, R. (2013). Role of *Campylobacter jejuni* infection in the pathogenesis of Guillain-Barré syndrome: an update. *BioMed Research International*, 2013, 1-12
- Nyenje, M.E., Odjadjare, C.E., Tanih, N.F., Green, E. and Ndip, R.N. (2012). Foodborne pathogens recovered from ready-to-eat foods from roadside cafeterias and retail outlets in Alice, Eastern Cape Province, South Africa: public health implications. *International Journal of Environmental Research Public Health*. 9: 2608-2619.
- Nyirimigabo, E., Xu, Y., Li, Y., Wang, Y., Agyemang, K., and Zhang, Y. (2015). A review on phytochemistry, pharmacology and toxicology studies of *Aconitum*. *Journal of Pharmacy and Pharmacology*, 67(1), 1-19.
- Odeyemi, O. A. (2016). Incidence and prevalence of *Vibrio parahaemolyticus* in seafood: a systematic review and meta-analysis. *Springer Plus*. 5(464): 1-17.
- Oh, M., Park, H. S., Um, S., Yang, T. J., and Kim, S. H. (2022). A comparative phytochemical study of nine Lauraceae species by using chemometric data analysis. *PloS one*, 17(9), e0273616, 1-14
- Olajuyigbe, O. O. and Afolayan, A. J. (2012). In vitro antibacterial and time-kill evaluation of the *Erythrina caffra* Thunb. extract against bacteria associated with diarrhoea. *The Scientific World Journal*, 2012, 1-9
- Olszowy, M. (2019). What is responsible for antioxidant properties of polyphenolic compounds from plants? *Plant Physiology and Biochemistry*, 144, 135-143.
- Omojate Godstime, C., Enwa Felix, O., Jewo Augustina, O. and Eze Christopher, O. (2014). Mechanisms of antimicrobial actions of phytochemicals against enteric pathogens—a review. *Journal of Pharmaceutical and Biological Sciences*, 2(2), 77-85.
- Onyebuchi, C., and Kavaz, D. (2020). Effect of extraction temperature and solvent type on the bioactive potential of *Ocimum gratissimum* L. extracts. *Scientific Reports*, 10(1), 21760-21769
- Orihara, Y., Hamamoto, H., Kasuga, H., Shimada, T., Kawaguchi, Y., and Sekimizu, K. (2008). A silkworm–baculovirus model for assessing the therapeutic effects of antiviral compounds: characterization and application to the isolation of antivirals from traditional medicines. *Journal of General Virology*, 89(1), 188-194.
- Otero, C., Miranda-Rojas, S., Llancalahuén, F. M., Fuentes, J. A., Atala, C., González-Silva, G., Verdugo, D., Sierra-Rosales, P., Moreno, A. and Gordillo-Fuenzalida, F. (2022). Biochemical characterization of *Peumus boldus* fruits: Insights of its antioxidant properties through a theoretical approach. *Food Chemistry*, 370, 131012-131028

- Othman, L., Sleiman, A. and Abdel-Massih, R. M. (2019). Antimicrobial activity of polyphenols and alkaloids in middle eastern plants. *Frontiers in Microbiology*, 10, 911-921.
- Over, K. F., Hettiarachchy, N., Johnson, M. G., and Davis, B. (2009). Effect of organic acids and plant extracts on *Escherichia coli* O157: H7, *Listeria monocytogenes*, and *Salmonella* Typhimurium in broth culture model and chicken meat systems. *Journal of Food Science*, 74(9), 515-521.
- Ozcelik, B., Kartal, M. and Orhan, I. (2011). Cytotoxicity, antiviral and antimicrobial activities of alkaloids, flavonoids, and phenolic acids. *Pharmaceutical Biology*, 49(4), 396-402.
- Ozogul, Y., Boğa, E. K., Akyol, I., Durmus, M., Ucar, Y., Regenstein, J. M., and Köşker, A. R. (2020). Antimicrobial activity of thyme essential oil nanoemulsions on spoilage bacteria of fish and food-borne pathogens. *Food Bioscience*, 36, 100635-100644
- Padzil, K. N. M., Ayob, N. M., Alzabt, A. M., and Rukayadi, Y. (2021). Antibacterial activity of taro (*Colocasia esculenta* L.) leaves extracts against *Staphylococcus aureus* and *Vibrio parahaemolyticus* and its effect on microbial population in sardine (*Sardinella longiceps* Val.). *Food Research*, 5(2), 88-97.
- Pakbin, B., Brück, W. M., Brück, T. B., Allahyari, S., and Ashrafi Tamai, I. (2023). A quantitative prevalence of *Escherichia coli* O157 in different food samples using real-time qPCR method. *Food Science and Nutrition*, 11(1), 228-235.
- Pal, M., Asefa, M., Deressa, A., and Muzein, R. (2014). Processed foods and *Bacillus cereus* poisoning. *Beverage and Food World*, 41(12), 41-43.
- Pal, M., Ayele, Y., Hadush, M., Panigrahi, S., and Jadhav, V. J. (2018). Public health hazards due to unsafe drinking water. *Air and Water Borne Diseases Journal*, 7(2), 138-147.
- Palchykov, V. A., Zazharskyi, V. V., Brygadyrenko, V. V., Davydenko, P. O., Kulishenko, O. M., and Borovik, I. V. (2020). Chemical composition and antibacterial effect of ethanolic extract of *Buxus sempervirens* on cryogenic strains of microorganisms *in vitro*. *Chemical Data Collections*, 25, 100323-100328
- Pandey, S. (2019). Review on medicinal importance of *Vigna* genus. *Plant Science Today*, 6(4), 450-456.
- Pang, K. L., Thong, W. L. and How, S. E. (2009). *Cinnamomum iners* as Mitogen-Activated Protein Kinase (MKK1) inhibitor. *International Journal of Engineering and Technology*, 1(4,); 310-313

- Papuc, C., Goran, G. V., Predescu, C. N., Nicorescu, V., and Stefan, G. (2017). Plant polyphenols as antioxidant and antibacterial agents for shelf-life extension of meat and meat products: Classification, structures, sources, and action mechanisms. *Comprehensive Reviews in Food Science and Food Safety*, 16(6), 1243-1268.
- Pariza, M. W., Gillies, K. O., Kraak-Ripple, S. F., Leyer, G., and Smith, A. B. (2015). Determining the safety of microbial cultures for consumption by humans and animals. *Regulatory Toxicology and Pharmacology*, 73(1), 164-171.
- Pateiro, M., Munekata, P. E., Sant'Ana, A. S., Domínguez, R., Rodríguez-Lázaro, D., and Lorenzo, J. M. (2021). Application of essential oils as antimicrobial agents against spoilage and pathogenic microorganisms in meat products. *International Journal of Food Microbiology*, 337, 108966, 1-13
- Patel, M. K., Mishra, A., and Jha, B. (2016). Non-targeted metabolite profiling and scavenging activity unveil the nutraceutical potential of psyllium (*Plantago ovata* Forsk). *Frontiers in Plant Science*, 7, 431, 1-11
- Patra, J. K., and Mohanta, Y. K. (2014). Antimicrobial compounds from mangrove plants: A pharmaceutical prospective. *Chinese Journal of Integrative Medicine*, 20(4), 311-320.
- Peixoto, L. R., Rosalen, P. L., Ferreira, G. L. S., Freires, I. A., de Carvalho, F. G., Castellano, L. R., and de Castro, R. D. (2017). Antifungal activity, mode of action and anti-biofilm effects of *Laurus nobilis* Linnaeus essential oil against *Candida* spp. *Archives of Oral Biology*, 73, 179-185.
- Pejin, B., Savic, A., Sokovic, M., Glamoclija, J., Ciric, A., Nikolic, M., Radotic, K. and Mojovic, M. (2014). Further in vitro evaluation of antiradical and antimicrobial activities of phytol. *Natural Product Research*, 28(6), 372-376.
- Perrone, G., and Gallo, A. (2017). Aspergillus species and their associated mycotoxins. *Mycotoxigenic fungi: Methods and Protocols*, 33-49.
- Phuah, E. T., Yap, J. W. L., Lau, C. W., Lee, Y. Y., and Tang, T. K. (2022). Vegetable Oils and Animal Fats Animal fats: Sources, Properties and Recovery Recovery. *Recent Advances in Edible Fats and Oils Technology: Processing, Health Implications, Economic and Environmental Impact*, 5, 1-26.
- Piekarska-Radzik, L., & Klewicka, E. (2021). Mutual influence of polyphenols and Lactobacillus spp. bacteria in food: A review. *European Food Research and Technology*, 247(1), 9-24.
- Pinto, T., Aires, A., Cosme, F., Bacelar, E., Morais, M. C., Oliveira, I., Cardoso, J. F., Anjos, R., Vilela, A. and Gonçalves, B. (2021). Bioactive (poly)

phenols, volatile compounds from vegetables, medicinal and aromatic plants. *Foods*, 10(1), 106.

Pirzadeh, M., Caporaso, N., Rauf, A., Shariati, M. A., Yessimbekov, Z., Khan, M. U., Imran, M. and Mubarak, M. S. (2021). Pomegranate as a source of bioactive constituents: A review on their characterization, properties and applications. *Critical Reviews in Food Science and Nutrition*, 61(6), 982-999.

Plenagl, N., Seitz, B. S., Duse, L., Pinnapireddy, S. R., Jedelska, J., Bruessler, J., and Bakowsky, U. (2019). Hypericin inclusion complexes encapsulated in liposomes for antimicrobial photodynamic therapy. *International Journal of Pharmaceutics*, 570, 118666, 1-15

Prakash, S., and Shelke, A. U. (2014). Role of Triphala in dentistry. *Journal of Indian Society of Periodontology*, 18(2), 132.

Pulikottil, S. J., and Nath, S. (2015). Potential of clove of *Syzygium aromaticum* in development of a therapeutic agent for periodontal disease.: a review. *South African Dental Journal*, 70(3), 108-115.

Qi, J., Li, X., Zhang, W., Wang, H., Zhou, G., and Xu, X. (2018). Influence of stewing time on the texture, ultrastructure and *in vitro* digestibility of meat from the yellow-feathered chicken breed. *Animal Science Journal*, 89(2), 474-482.

Qu, S., Yang, K., Chen, L., Liu, M., Geng, Q., He, X., Li, Y., Liu, Y. and Tian, J. (2019). Cinnamaldehyde, a promising natural preservative against *Aspergillus flavus*. *Frontiers in Microbiology*, 10, 2895-2901

Racchi, M. L. (2013). Antioxidant defenses in plants with attention to *Prunus* and *Citrus* spp. *Antioxidants*, 2(4), 340-369.

Ragasa, C. Y., Tsai, P. W., and Shen, C. C. (2009). Antimicrobial terpenoids from *Erigeron sumatrensis*. *NRCP Research Journal*, 10(1), 27-32.

Rahayu, A. (2020). Analysis of *Cinnamomum* land suitability and the conservation efforts at Eka Karya Bali Botanic Garden. *Agrikultura*, 31(3), 214-221.

Rahimah, S. B., Djunaedi, D. D., Soeroto, A. Y., and Bisri, T. (2019). The phytochemical screening, total phenolic contents and antioxidant activities *in vitro* of white oyster mushroom (i) preparations. *Macedonian Journal of Medical Sciences*, 7(15), 2404-2420

Ramli, S., Son, R., Shaari, K. and Rukayadi, Y. (2020). Toxicity analysis of *Syzygium polyanthum* (Wight) Walp. leaves extract and its stability against different pH and temperature. *Emirates Journal of Food and Agriculture*, 32(6), 461-468.

- Ramya, K. S., Kanimathi, P., and Radha, A. (2019). GC–MS analysis and antimicrobial activity of various solvent extracts from *Simarouba glauca* leaves. *Journal of Pharmacognosy and Phytochemistry*, 8(2), 166-171.
- Rane, S. (2011). Street vended food in developing world: hazard analyses. *Indian Journal of Microbiology*, 51(1), 100-106.
- Rangsinth, P., Prasansuklab, A., Duangjan, C., Gu, X., Meemon, K., Wink, M., and Tencomnao, T. (2019). Leaf extract of *Caesalpinia mimosoides* enhances oxidative stress resistance and prolongs lifespan in *Caenorhabditis elegans*. *BMC Complementary and Alternative Medicine*, 19(1), 1-13.
- Raphael, E. C. (2011). Traditional medicine in Nigeria; current status and the future. *Research Journal of Pharmacology*, 5(6); 90-94
- Ravimannan, N., Sevel, P., and Saarutharshan, S. (2016). Study on fungi associated with spoilage of bread. *International Journal of Advanced Research in Biological Sciences*, 3(4), 165-167.
- Rawat, M., Varshney, A., Rai, M., Chikara, A., Pohty, A. L., Joshi, A., Binjola, A., Singh, C. P., Rawat, K., Rather, M. A. & Gupta, A. K. (2023). A comprehensive review on nutraceutical potential of underutilized cereals and cereal-based products. *Journal of Agriculture and Food Research*, 100619.
- Redondo, L. M., Dominguez, J. E., Rabinovitz, B. C., Redondo, E. A., and Miyakawa, M. F. (2015). Hydrolyzable and condensed tannins resistance in *Clostridium perfringens*. *Anaerobe*, 34, 139-145.
- Rekha, C., Poornima, G., Manasa, M., Abhipsa, V., Devi, J.P., Kumar, H.T.V. and Kekuda, T.R.P. (2012). Ascorbic acid, total phenol content and antioxidant activity of fresh juices of four ripe and unripe citrus fruits. *Chemical Science Transaction*, 1(2), 303-310.
- Ribes, S., Fuentes, A., Talens, P., Barat, J. M., Ferrari, G., and Donsì, F. (2017). Influence of emulsifier type on the antifungal activity of cinnamon leaf, lemon and bergamot oil nanoemulsions against *Aspergillus niger*. *Food Control*, 73, 784-795.
- Ricardo, E., Miranda, I. M., Faria-Ramos, I., Silva, R. M., Rodrigues, A. G. and Pina-Vaz, I. (2014). *In vivo* and *in vitro* acquisition of resistance to voriconazole by *Candida krusei*. *Antimicrobial Agents and Chemotherapy*. 58(8): 4604-4611.
- Rio, J. C., Rencoret, J., Gutiérrez, A., Elder, T., Kim, H., and Ralph, J. (2020). Lignin monomers from beyond the canonical monolignol biosynthetic pathway: another brick in the wall. *ACS Sustainable Chemistry and Engineering*, 8(13), 4997-5012.

- Robert-Pillot, A., Copin, S., Himber, C., Gay, M., and Quilici, M. L. (2014). Occurrence of the three major *Vibrio* species pathogenic for human in seafood products consumed in France using real-time PCR. *International Journal of Food Microbiology*, 189, 75-81.
- Rochmat, A., Aditya, G., Kusmayanti, N., Kustiningsih, I., Hariri, A., and Rezaldi, F. (2022). *In vitro* activity and docking approach in silico leaf extract *Syzygium polyanthum* (wight) walp. as a *Salmonella* Typhi inhibitor. *Trends in Sciences*, 19(16), 5654-5654.
- Rosa, L. A., Moreno-Escamilla, J. O., Rodrigo-García, J., and Alvarez-Parrilla, E. (2019). Phenolic compounds. In *Postharvest physiology and biochemistry of fruits and vegetables* (pp. 253-271). Woodhead publishing.
- Roy, A. (2017). A review on the alkaloids an important therapeutic compound from plants. *International Journal of Plant Biology*, 3(2), 1-9.
- Rukayadi, Y., Lau, K.Y., Zainin, N.S., Zakaria, M., and Abas, F. (2013). Screening antimicrobial activity of tropical edible medicinal plant extracts against five standard microorganisms for natural food preservative. *International Food Research Journal*, 20(5), 2905-2910.
- Russo, S., Cabrera, N., Chludil, H., Yaber-Grass, M., & Leicach, S. (2015). Insecticidal activity of young and mature leaves essential oil from *Eucalyptus globulus* Labill. against *Tribolium confusum* Jacquelin du Val (Coleoptera: Tenebrionidae). *Chilean journal of agricultural research*, 75(3), 375-379.
- Sahoo, P. M. S., Behera, S., Behura, R., Acharya, A., Biswal, D., Suna, S. K., and Jali, B. R. (2022). A brief review: antibacterial activity of Quinone derivatives. *Biointerface Research in Applied Chemistry*, 12, 3247-3258.
- Sahu, M., and Bala, S. (2017). Food processing, food spoilage and their prevention: An overview. *International Journal of Life Sciences Research*, 3(1), 753-759.
- Saide, A., Lauritano, C., and Ianora, A. (2020). Pheophorbide a: State of the Art. *Marine Drugs*, 18(5), 257-270
- Saikia, P., and Joshi, S. R. (2010). Retail market poultry meats of North-East India—a microbiological survey for pathogenic contaminants. *Research Journal Microbiology*, 5(1), 36-43.
- Sajjad, W., Din, G., Rafiq, M., Iqbal, A., Khan, S., Zada, S., Ali, B. and Kang, S. (2020). Pigment production by cold-adapted bacteria and fungi: colorful tale of cryosphere with wide range applications. *Extremophiles*, 24, 447-473.

- Salazar-Ramiro, A., Ramirez-Ortega, D., Cruz, V.P., Hernandez-Pedro, N.Y., Gonzalez-Esquivel, D.F., Sotelo, J. and Pineda, B. (2016). Role of redox status in development of glioblastoma. *Frontiers in Immunology*, 7, 1-15.
- Saleh, E., Morshdy, A. E., El-Manakhly, E., Al-Rashed, S., F. Hetta, H., Jeandet, P., Yahia, R., Batiha, G. E. S. and Ali, E. (2020). Effects of olive leaf extracts as natural preservative on retailed poultry meat quality. *Foods*, 9(8), 1017-1022
- Salem, A. M., Mohamed, E. F., and Selim, E. M. (2018). Antimicrobial effect of some natural oils on *Bacillus cereus* in minced beef. *Benha Veterinary Medical Journal*, 34(2), 149-156.
- Sambu, S., Hemaram, U., Murugan, R., and Alsofi, A. A. (2022). Toxicological and teratogenic effect of various food additives: an updated review. *BioMed Research International*, 2022, 1-10
- Sanchez, C. (2017). Reactive oxygen species and antioxidant properties from mushrooms. *Synthetic and Systems Biotechnology*, 2(1), 13-22.
- Santini, A., Tenore, G. C., and Novellino, E. (2017). Nutraceuticals: A paradigm of proactive medicine. *European Journal of Pharmaceutical Sciences*, 96, 53-61.
- Sarah, Q.S., Anny, F.C. and Misbahuddin, M. (2017). Brine shrimp lethality assay. *Bangladesh Journal of Pharmacology*, 12, 186-189.
- Sargiya, H., Goyal, P. K., and Vyas, B. (2017). Antimicrobial drugs of herbal origin. *International Journal of Pharmaceutical Erudition*, 7(1), 1-10.
- Sarkic, A., Stappen, I. (2018). Essential oils and their single compounds in cosmetics—A critical review. *Cosmetics*, 5(1), 11.
- Saxena, M., Saxena, J., Nema, R., Singh, D., and Gupta, A. (2013). Phytochemistry of medicinal plants. *Journal of Pharmacognosy and Phytochemistry*, 1(6), 168-182.
- Scharff, R. L. (2012). Economic burden from health losses due to foodborne illness in the United States. *Journal of Food Protection*, 75(1), 123-131.
- Scherlach, K., Graupner, K., and Hertweck, C. (2013). Molecular bacteria-fungi interactions: effects on environment, food, and medicine. *Annual Review of Microbiology*, 67, 375-397.
- Schmitzer, V., Veberic, R., and Stampar, F. (2012). European elderberry (*Sambucus nigra* L.) and American Elderberry (*Sambucus canadensis* L.): Botanical, chemical and health properties of flowers, berries and their products. *Berries Prop. Consum. Nutr*, 2012, 127-144.

- Schweitzer, B., Balázs, V. L., Molnár, S., Szögi-Tatár, B., Böszörményi, A., Palkovics, T., Horvath, G. and Schneider, G. (2022). Antibacterial effect of lemongrass (*Cymbopogon citratus*) against the aetiological agents of pitted keratolysis. *Molecules*, 27(4), 1423-1434
- Selvarajan, E., Veena, R., and Manoj Kumar, N. (2018). Polyphenol oxidase, beyond enzyme browning. In *Microbial Bioprospecting for Sustainable Development* (pp. 203-222). Springer
- Sen, A., and Batra, A. (2012). Evaluation of antimicrobial activity of different solvent extracts of medicinal plant: *Melia azedarach* L. *International Journal of Current Pharmaceutical Research*, 4(2), 67-73.
- Sephton-Clark, P. C., and Voelz, K. (2018). Spore germination of pathogenic filamentous fungi. In *Advances in Applied Microbiology* (102), pp. 117-157. Academic Press.
- Shaaban, M. T., Ghaly, M. F., and Fahmi, S. M. (2021). Antibacterial activities of hexadecanoic acid methyl ester and green-synthesized silver nanoparticles against multidrug-resistant bacteria. *Journal of Basic Microbiology*, 61(6), 557-568.
- Shah, A., & Smith, D. L. (2020). Flavonoids in agriculture: Chemistry and roles in, biotic and abiotic stress responses, and microbial associations. *Agronomy*, 10(8), 1209.
- Shahwar, M. K., El-Ghorab, A. H., Anjum, F. M., Butt, M. S., Hussain, S., and Nadeem, M. (2012). Characterization of coriander (*Coriandrum sativum* L.) seeds and leaves: volatile and non-volatile extracts. *International Journal of Food Properties*, 15(4), 736-747.
- Shakya, A. K. (2016). Medicinal plants: future source of new drugs. *International Journal of Herbal Medicine*, 4(4); 59-64.
- Sharif, M. K., Javed, K., and Nasir, A. (2018). Foodborne illness: threats and control. In *Foodborne diseases* (pp. 501-523). Academic Press.
- Sharifi-Rad, J., Sureda, A., Tenore, G. C., Daglia, M., Sharifi-Rad, M., Valussi, M., Tundis, R., Sharifi-Rad, M., Loizzo, M. R., Ademiluyi, A. O., Sharifi-Rad, R., Ayatollahi, S. A. and Iriti, M. (2017). Biological activities of essential oils: From plant chemoecology to traditional healing systems. *Molecules*, 22(1), 70-81
- Sharma, P., Wichaphon, J., and Klangpetch, W. (2020). Antimicrobial and antioxidant activities of defatted *Moringa oleifera* seed meal extract obtained by ultrasound-assisted extraction and application as a natural antimicrobial coating for raw chicken sausages. *International Journal of Food Microbiology*, 332, 108770, 1-15

- Sharpe, R. A., Bearman, N., Thornton, C. R., Husk, K., and Osborne, N. J. (2015). Indoor fungal diversity and asthma: a meta-analysis and systematic review of risk factors. *Journal of Allergy and Clinical Immunology*, 135(1), 110-122.
- Sherikar, A.S. and Mahanthesh, M.C. (2015). Evaluation of aqueous and methanolic extract of leaves of *Epipremnum aureum* for radical scavenging activity by DPPH method, total phenolic content, reducing capacity assay and FRAP assay. *Journal of Pharmacognosy and Phytochemistry*, 4(4), 36-40.
- Shibano, M., Kakutani, K., Taniguchi, M., Yasuda, M., and Baba, K. (2008). Antioxidant constituents in the dayflower (*Commelina communis* L.) and their α -glucosidase-inhibitory activity. *Journal of Natural Medicines*, 62, 349-353.
- Shirsath, S. R., Sonawane, S. H., and Gogate, P. R. (2012). Intensification of extraction of natural products using ultrasonic irradiations - A review of current status. *Chemical Engineering and Processing: Process Intensification*, 53, 10-23.
- Shohayeb, M., Abdel-Hameed, E., and Bazaid, S. (2013). Antimicrobial activity of tannins and extracts of different parts of *Conocarpus erectus* L. *International Journal of Pharma and Bio Sciences*, 3(2), 544-553.
- Shreaz, S., Wani, W. A., Behbehani, J. M., Raja, V., Irshad, M., Karched, M., Ali, I, Sidiqi, W. A. and Hun, L. T. (2016). Cinnamaldehyde and its derivatives, a novel class of antifungal agents. *Fitoterapia*, 112, 116-131.
- Siddiky, N. A., Sarker, M. S., Khan, M. S. R., Begum, R., Kabir, M. E., Karim, M. R., Rahman, M. T., Mahmud, A. and Samad, M. A. (2021). Virulence and antimicrobial resistance profiles of *Salmonella enterica* serovars isolated from chicken at wet markets in Dhaka, Bangladesh. *Microorganisms*, 9(5), 952.
- Silva, R. O., Sousa, F. B. M., Damasceno, S. R., Carvalho, N. S., Silva, V. G., Oliveira, F. R. M. A., Sousa, D. P., Aragao, K. S., Barbosa, A. L. R., Freitas, R. M. and Medeiros, J. V. R. (2014). Phytol, a diterpene alcohol, inhibits the inflammatory response by reducing cytokine production and oxidative stress. *Fundamental and Clinical Pharmacology*, 28(4), 455-464.
- Singh, N., and Gaur, S. (2021). GRAS fungi: A new horizon in safer food product. *Fungi in Sustainable Food Production*, 27-37.
- Singh, R., and Jawaid, T. (2012). *Cinnamomum camphora* (Kapur). *Pharmacognosy Journal*, 4(28), 1-5.
- Sirtori, F., Dimauro, C., Bozzi, R., Aquilani, C., Franci, O., Calamai, L., Pezzati, A. and Pugliese, C. (2020). Evolution of volatile compounds and physical, chemical and sensory characteristics of Toscano PDO ham from fresh to

- dry-cured product. *European Food Research and Technology*, 246, 409-424.
- Siyumbwa, S. N., Ekeuku, S. O., Amini, F., Emerald, N. M., Sharma, D., and Okechukwu, P. N. (2019). Wound healing and antibacterial activities of 2-Pentadecanone in streptozotocin-induced Type 2 diabetic rats. *Pharmacognosy Magazine*, 15(1), 71-77.
- Smith, J. L., and Fratamico, P. M. (2017). *Escherichia coli* as a Pathogen. In *Foodborne diseases* (pp. 189-208). Academic Press.
- Snyder, A. B., and Worobo, R. W. (2018). Fungal spoilage in food processing. *Journal of Food Protection*, 81(6), 1035-1040.
- Sohn, H. R., Bae, J. H., Hou, C. T., and Kim, H. R. (2013). Antibacterial activity of a 7, 10-dihydroxy-8 (E)-octadecenoic acid against plant pathogenic bacteria. *Enzyme and Microbial Technology*, 53(3), 152-153.
- Somai, B. M., and Belewa, V. (2011). Aqueous extracts of *Tulbaghia violacea* inhibit germination of *Aspergillus flavus* and *Aspergillus parasiticus* conidia. *Journal of Food Protection*, 74(6), 1007-1011.
- Souza, W. R. (2013). Microbial degradation of lignocellulosic biomass. *Sustainable Degradation of Lignocellulosic Biomass-Techniques, Applications and Commercialization*, 15, 207-247.
- Sowjanya, R., Shankar, M., Sireesha, B., Naik, A. E., Yudharaj, P., and Priyadarshini, R. R. (2017). An overview on inflammation and plant having anti-inflammatory activity. *International Journal of Phytopharmacy*, 7, 25-32.
- Srivastava, B., Sagar, A., and Dubey, N. K. (2011). Evaluation of *Cinnamomum tamala* oil and its phenylpropanoid eugenol for their antifungal and antiaflatoxigenic activity. *Food Analytical Methods*, 4, 347-356.
- Stan, D., Enciu, A. M., Mateescu, A. L., Ion, A. C., Brezeanu, A. C., Stan, D., and Tanase, C. (2021). Natural compounds with antimicrobial and antiviral effect and nanocarriers used for their transportation. *Frontiers in Pharmacology*, 12, 723233, 1-10
- Suarez-Estrella, D., Torri, L., Pagani, M. A., and Marti, A. (2018). Quinoa bitterness: Causes and solutions for improving product acceptability. *Journal of the Science of Food and Agriculture*, 98(11), 4033-4041.
- Suciu, F., Stoicescu, I., Lupu, E. C., Musuc, A. M., Popescu, A., Mititelu, M., Rosca, A. C., Dumitrescu, D. E., Badea, F. C., Caraiane, A. and Badea, V. (2023). HPLC analysis of polyphenolic compounds in *Lysimachia*

- nummularia* L. and comparative determination of antioxidant capacity. *Applied Sciences*, 13(4), 2159.
- Suhaimi, A. T., Ariffin, Z. Z., Ali, N. A. M., Halim, M. I. A., Mahat, M. M. and Safian, M. F. (2017). Essential oil chemical constituent analysis of *Cinamomum iners*. *Journal of Engineering and Applied Sciences*, 12(21); 5369-5372
- Sujatha, S., Anand, S., Sangeetha, K. N., Shilpa, K., Lakshmi, J., Balakrishnan, A., and Lakshmi, B. S. (2010). Biological evaluation of (3 β)-stigmast-5-en-3-ol as potent anti-diabetic agent in regulating glucose transport using *in vitro* model. *International Journal of Diabetes Mellitus*, 2(2), 101-109.
- Supardy, N. A., Ibrahim, D., Sulaiman, S. F., and Zakaria, N. A. (2012). Inhibition of *Klebsiella pneumoniae* ATCC13883 cells by hexane extract of *Halimeda discoidea* (Decaisne) and the identification of its potential bioactive compounds. *Journal of Microbiology and Biotechnology*, 22(6), 872-881.
- Surai, P.F., Kochish, I.I., Fisinin, V.I. and Kidd, M.T. (2019). Antioxidant defence systems and oxidative stress in poultry biology; an update. *Antioxidants*, 8(7), 235-271.
- Suryawanshi, V.S., Yadav, A.R. and Mohite, S.K. (2020). Toxicological assessment using brine shrimp lethality assay and antimicrobial activity of *Capparis grandis*. *Journal of University of Shanghai for Science and Technology*, 22(11), 746-760.
- Swallah, M.S., Sun, H., Affoh, R., Fu, H. and Yu, H. (2020). Antioxidant potential overviews of secondary metabolites (polyphenols) in fruits. *International Journal of Food Science*, 1, 1-8.
- Switaj, T. L., Winter, K. J., and Christensen, S. (2015). Diagnosis and management of foodborne illness. *American Family Physician*, 92(5), 358-365.
- Syed, J. H., Alamdar, A., Mohammad, A., Ahad, K., Shabir, Z., Ahmed, H., Ali, S. M., Sani, S. G. A. S., Bokhari, H., Gallagher, K. D., Ahmad, I. and Eqani, S. A. M. A. S. (2014). Pesticide residues in fruits and vegetables from Pakistan: a review of the occurrence and associated human health risks. *Environmental Science and Pollution Research*, 21, 13367-13393.
- Syeda, A. M., and Riazunnisa, K. (2020). Data on GC-MS analysis, *in vitro* anti-oxidant and anti-microbial activity of the *Catharanthus roseus* and *Moringa oleifera* leaf extracts. *Data in Brief*, 29, 105258-105269.
- Sylvester, W. S., Son, R., Lew, K. F., and Rukayadi, Y. (2015). Antibacterial activity of Java turmeric (*Curcuma xanthorrhiza* Roxb.) extract against

Klebsiella pneumoniae isolated from several vegetables. *International Food Research Journal*, 22(5).

Tabatabaei, S. M., Badalzadeh, R., Mohammadnezhad, G. R., and Balaei, R. (2015). Effects of Cinnamon extract on biochemical enzymes, TNF- α and NF- κ B gene expression levels in liver of broiler chickens inoculated with *Escherichia coli*. *Pesquisa Veterinária Brasileira*, 35, 781-787.

Talla, R. M., Jouda, J. B., Mbazoa, C. D., Kapche, G. D. W. F., Lenta, B. N., Sewald, N., and Wandji, J. (2021). Pentacyclic triterpenoids and other constituents isolated from the leaves of *Gambeya lacourtiana* and their antibacterial activity. *Biochemical Systematics and Ecology*, 98, 104322-104329

Tan, J.B.L. and Lim, Y.Y. (2015). Critical analysis of current methods for assessing the *in vitro* antioxidant and antibacterial activity of plant extracts. *Food Chemistry*, 172, 814-822.

Tanaka, K., Li, F., Tezuka, Y., Watanabe, S., Kawahara, N., and Kida, H. (2013). Evaluation of the quality of Chinese and Vietnamese cassia using LC-MS and multivariate analysis. *Natural Product Communications*, 8(1), 75-78

Tanapichatsakul, C., Khruengsai, S., Monggoot, S., and Pripdeevech, P. (2019). Production of eugenol from fungal endophytes *Neopestalotiopsis* sp. and *Diaporthe* sp. isolated from *Cinnamomum loureiroi* leaves. *PeerJ*, 7, e6427.

Tanase, C., Coşarcă, S., and Muntean, D. L. (2019). A critical review of phenolic compounds extracted from the bark of woody vascular plants and their potential biological activity. *Molecules*, 24(6), 1182-1192

Tello, J.C.C., Flores, H.E.M., Romo, M.G.G., Ramirez, J.S.P., Molina, A.S., Cortez, O.A., Camacho, M.C.B. and Lopez, J.O.R. (2018). Quantification 151 of phenolic compounds and *in vitro* radical scavenging abilities with leaf extracts from two varieties of *Psidium guajava* L. *Antioxidant*, 7, 34-46.

Teng, Y. N., Huang, B. H., Huang, S. Y., Wu, I. T., Wu, T. S., Lee, T. E., and Hung, C. C. (2021). Cinnamophilin overcomes cancer multi-drug resistance via allosterically modulating human P-glycoprotein on both drug binding sites and ATPase binding sites. *Biomedicine and Pharmacotherapy*, 144, 112379-112388

Tewari, A., and Abdullah, S. (2015). *Bacillus cereus* food poisoning: international and Indian perspective. *Journal of Food Science and Technology*, 52, 2500-2511.

- Tian, J., Zeng, X., Zeng, H., Feng, Z., Miao, X., and Peng, X. (2013). Investigations on the antifungal effect of nerol against *Aspergillus flavus* causing food spoilage. *The Scientific World Journal*, 2013, 1-14
- Tian, Y., Jia, X., Wang, Q., Lu, T., Deng, G., Tian, M., and Zhou, Y. (2022). Antioxidant, antibacterial, enzyme inhibitory, and anticancer activities and chemical composition of *Alpinia galanga* flower essential oil. *Pharmaceuticals*, 15(9), 1069-1085
- Tiku, A. R. (2018). Antimicrobial compounds and their role in plant defense. In *Molecular Aspects of Plant-Pathogen Interaction* (pp. 283-307). Springer
- Tinikul, R., and Chaiken, P. (2016). Structure, mechanism, and mutation of bacterial luciferase. *Bioluminescence: Fundamentals and Applications in Biotechnology*, 3, 47-74.
- Todd, E. C., Michaels, B. S., Greig, J. D., Smith, D., Holah, J., and Bartleson, C. A. (2010). Outbreaks where food workers have been implicated in the spread of foodborne disease. Part 7. Barriers to reduce contamination of food by workers. *Journal of Food Protection*, 73(8), 1552-1565.
- Trajano, V. N., Lima, E. D. O., and Souza, F. S. D. (2012). Antifungal activity of the essential oil of *Cinnamomum zeylanicum* Blume and eugenol on *Aspergillus flavus*. *Journal of Essential Oil-Bearing Plants*, 15(5), 785-793.
- Tsay, G. J., Lin, Y. T., Hsu, C. H., Tang, F. Y., Kuo, Y. H., and Chao, C. Y. (2021). Adlay hull extracts attenuate β -amyloid-induced neurotoxicity and oxidative stress in PC12 cells through antioxidative, anti-inflammatory, and antiapoptotic activities. *Biochemistry and Biophysics Reports*, 26, 101020-101029
- Tulini, F. L., Souza, V. B., Thomazini, M., Silva, M. P., Massarioli, A. P., Alencar, S. M., Pallone, E. M. J. A., Genovese, M. and Favaro-Trindade, C. S. (2017). Evaluation of the release profile, stability and antioxidant activity of a proanthocyanidin-rich cinnamon (*Cinnamomum zeylanicum*) extract co-encapsulated with α -tocopherol by spray chilling. *Food Research International*, 95, 117-124.
- Tzortzakis, N. G. (2009). Impact of cinnamon oil-enrichment on microbial spoilage of fresh produce. *Innovative Food Science and Emerging Technologies*, 10(1), 97-102.
- Udayaprakash, N. K., Ranjithkumar, M., Deepa, S., Sripriya, N., Al-Arfaj, A. A. and Bhuvaneshwari, S. (2015). Antioxidant, free radical scavenging and GC-MS composition of *Cinnamomum iners* Reinw. ex Blume. *Industrial Crops and Products*, 69, 175-179.
- Uddin, M. A., Shahinuzzaman, M., Rana, M. S., and Yaakob, Z. (2017). Study of chemical composition and medicinal properties of volatile oil from clove

buds (*Eugenia caryophyllus*). *International Journal of Pharmaceutical Sciences and Research*, 8(2), 895.

Udoh. I.P., Eleazar, C.I., Ogeneh, B.O. and Ohanu, M.E. (2015). Studies on fungi responsible for the spoilage/deterioration of some edible fruits and vegetables. *Advances in Microbiology*, 5: 285-290.

Uka, V., Cary, J. W., Lebar, M. D., Puel, O., De Saeger, S., and Diana Di Mavungu, J. (2020). Chemical repertoire and biosynthetic machinery of the *Aspergillus flavus* secondary metabolome: A review. *Comprehensive Reviews in Food Science and Food Safety*, 19(6), 2797-2842.

Ulbricht, C., Seamon, E., Windsor, R. C., Armbruster, N., Bryan, J. K., Costa, D., Giese, N., Gruenwalk, J., Lovin. R., Isaac, R., Serrano, J. M. G., Tanguay-Colucci, S., Weissner, W., Yoon, H. and Zhang, J. (2011). An evidence-based systematic review of cinnamon (*Cinnamomum* spp.) by the Natural Standard Research Collaboration. *Journal of Dietary Supplements*, 8(4), 378-454.

Ul-Hamid, A. (2018). *A beginners' guide to scanning electron microscopy*, 1, (pp. 402). Cham, Switzerland: Springer International Publishing.

Unsal, V., Dalkiran, T., Cicek, M. and Kolukcu, E. (2020). The role of natural antioxidants against reactive oxygen species produced by cadmium toxicity: a review. *Advanced Pharmaceutical Bulletin*, 10(2), 184-202.

Vaishnavi, J., Reddy, S., Narmadha, S., and Osborne, W. J. (2021). Detection and purification of microbial volatile organic compounds. In *Volatiles and Metabolites of Microbes* (pp. 51-64). Academic Press.

Valavanidis, A., Vlachogianni, T., Fiotakis, K., and Loridas, S. (2013). Pulmonary oxidative stress, inflammation and cancer: respirable particulate matter, fibrous dusts and ozone as major causes of lung carcinogenesis through reactive oxygen species mechanisms. *International Journal of Environmental Research and Public Health*, 10(9), 3886-3907.

Valdes, A., Ramos, M., Beltrán, A., Jiménez, A., and Garrigós, M. C. (2017). State of the art of antimicrobial edible coatings for food packaging applications. *Coatings*, 7(4), 56-66

Varga, J., Kocsubé, S., Péteri, Z., Vágvolgyi, C., and Tóth, B. (2010). Chemical, physical and biological approaches to prevent ochratoxin induced toxicoses in humans and animals. *Toxins*, 2(7), 1718-1750.

Vieira de Moraes, D., Rosalen, P. L., Ikegaki, M., de Souza Silva, A. P., Massarioli, A. P., and de Alencar, S. M. (2021). Active antioxidant phenolics from Brazilian red propolis: An optimization study for their recovery and identification by LC-ESI-QTOF-MS/MS. *Antioxidants*, 10(2), 297.

- Vigila, A. G., Sahayaraj, K. and Baskaran, X. (2018). In vitro antimicrobial activities of *Cinnamomum iners* leaf and bark extracts against pathogens of food borne diseases. *Approaches in Poultry, Dairy and Veterinary Sciences*, 3(4); 1-5.
- Vlaicu, P. A., Untea, A. E., Panaite, T. D., and Turcu, R. P. (2020). Effect of dietary orange and grapefruit peel on growth performance, health status, meat quality and intestinal microflora of broiler chickens. *Italian Journal of Animal Science*, 19(1), 1394-1405.
- Wahab, I. R. A. and Hussain, M. H. (2021). Brine shrimp lethality test of various *Cinnamomum iners* (Lauraceae) barks extracts. *Journal of Tropical Resources and Sustainable Science*, 6; 109-113
- Waldorf, A. R., and Diamond, R. D. (2020). Aspergillosis and mucormycosis. In *Immunology of the Fungal Diseases* (pp. 29-55). CRC Press.
- Wang, C., Han, J., Pu, Y., & Wang, X. (2022). Tea (*Camellia sinensis*): a review of nutritional composition, potential applications, and Omics Research. *Applied Sciences*, 12(12), 5874.
- Wang, D., Cui, L., Ren, H., Wang, Y., Long, D., and Niu, Y. (2021a). Anti-fungal activity and preliminary active components separation from ethanol extracts in Saffron (*Crocus sativus* L.) lateral buds. *Industrial Crops and Products*, 173, 114081.
- Wang, M., Huang, W., Hu, Y., Zhang, L., Shao, Y., Wang, M., Zhang, F., Zhao, Z., Mei, X., Li, T., Wang, D., Liang, Y., Li, J., Huang, Y., Zhang, L., Xu, T., Song, H., Zhong, Y. and Lu, B. (2018). Phytosterol profiles of common foods and estimated natural intake of different structures and forms in China. *Journal of Agricultural and Food Chemistry*, 66(11), 2669-2676
- Wang, Y., Zhao, Y., Xue, F., Nan, X., Wang, H., Hua, D., Liu, J., Yang, L., Jiang, L. and Xiong, B. (2020). Nutritional value, bioactivity, and application potential of Jerusalem artichoke (*Helianthus tuberosus* L.) as a neotype feed resource. *Animal Nutrition*, 6(4), 429-437.
- Wason, S., Verma, T., Irmak, S., & Subbiah, J. (2022). In-package pasteurization of dried basil leaves using radiofrequency heating. *Food Control*, 141, 109179.
- Welch, A. A., Shakya-Shrestha, S., Lentjes, M. A., Wareham, N. J., and Khaw, K. T. (2010). Dietary intake and status of n-3 polyunsaturated fatty acids in a population of fish-eating and non-fish-eating meat-eaters, vegetarians, and vegans and the precursor-product ratio of α -linolenic acid to long-chain n-3 polyunsaturated fatty acids: results from the EPIC-Norfolk cohort. *The American Journal of Clinical Nutrition*, 92(5), 1040-1051.

- Wong, J. X., and Ramli, S. (2021). Antimicrobial activity of different types of *Centella asiatica* extracts against foodborne pathogens and food spoilage microorganisms. *LWT*, 142, 111026-111031
- Wong, J. X., Ramli, S., and Chen, S. N. (2021). Assessment of toxic effect of *Centella asiatica* extract and its application as natural preservative in fresh-cut mango, pear and cabbage. *Journal of Food Processing and Preservation*, 45(10), 1-35.
- Wong, J. X., Ramli, S., Rukayadi, Y., Juhari, N. K. K., Radu, S., and Abd Wahid, N. B. (2022). Antifungal activity of ethanolic extract of *Syzygium polyanthum* (Wight) Walp. leaves extract against several types of filamentous fungi and *Candida* species. *Malaysian Journal of Microscopy*, 18(1), 21-36
- Wong, M. H. Y., Wan, H. Y., and Chen, S. (2013). Characterization of multidrug-resistant *Proteus mirabilis* isolated from chicken carcasses. *Foodborne Pathogens and Disease*, 10(2), 177-181.
- World Health Organisation, 2008. Media Centre, Traditional Medicine, 20.
- Wu, H., Song, Z., Niu, X., Liu, J., Jiang, J., & Li, Y. (2022). Classification of toona sinensis young leaves using machine learning and UAV-borne hyperspectral imagery. *Frontiers in Plant Science*, 13, 940327.
- Wu, M., Ni, L., Lu, H., Xu, H., Zou, S., and Zou, X. (2020). Terpenoids and their biological activities from *Cinnamomum*: a review. *Journal of Chemistry*, 2020, 1-14.
- Wu, S., Wu, Q., Zhang, J., Chen, M. and Yan, Z. (2015). Prevalence, antibiotic resistance and genetic diversity of *Listeria monocytogenes* isolated from retail ready-to-eat foods in China. *Food Control*. 47: 340-347.
- Wuu-Kuang, S. (2011). Taxonomic revision of *Cinnamomum* (Lauraceae) in Borneo. *Blumea-Biodiversity, Evolution and Biogeography of Plants*, 56(3), 241-264.
- Xie, Y., Peng, Q., Ji, Y., Xie, A., Yang, L., Mu, S., Li, Z., He, T., Xiao, Y., Zhao, J. and Zhang, Q. (2021). Isolation and identification of antibacterial bioactive compounds from *Bacillus megaterium* L2. *Frontiers in Microbiology*, 12, 645484, 1-10
- Xing, Y., Li, X., Xu, Q., Yun, J., and Lu, Y. (2010). Antifungal activities of cinnamon oil against *Rhizopus nigricans*, *Aspergillus flavus* and *Penicillium expansum* in vitro and in vivo fruit test. *International Journal of Food Science and Technology*, 45(9), 1837-1842.
- Yaashikaa, P.R., Saravanan, A. and Kumar, P.S. (2016). Isolation and identification of *Vibrio cholerae* and *Vibrio parahaemolyticus* from prawn

(*Penaeus monodon*) seafood: Preservation strategies. *Microbial Pathogenesis*, 99: 5-13.

Yaghoubi, M., Ayaseh, A., Alirezalu, K., Nemati, Z., Pateiro, M., and Lorenzo, J. M. (2021). Effect of chitosan coating incorporated with *Artemisia fragrans* essential oil on fresh chicken meat during refrigerated storage. *Polymers*, 13(5), 716-719

Yahaya, M. F., Osemeahon, S. A., Shagal, M. H., Maitera, O. N., Dass, P. M., and Yelwa, J. M. (2021). Antimicrobial, antioxidant, cytotoxicity profiles and chemical compositions of ethanolic extracts of *Ficus polita* and *Ficus thonningii* plant. *Journal of Research in Chemistry*, 2(1), 4-10

Yalcin, H. and Capar, T. D. (2017). Bioactive compounds of fruits and vegetables. *Minimally Processed Refrigerated Fruits and Vegetables*, 1, 723–745

Yang, C. H., Li, R.X. and Chuang, L.Y. (2012). Antioxidant activity of various parts of *Cinnamomum cassia* extracted with different extraction methods. *Molecules*, 17(6), 7294-7304.

Yang, L., Yang, C., Li, C., Zhao, Q., Liu, L., Fang, X., and Chen, X. Y. (2016). Recent advances in biosynthesis of bioactive compounds in traditional Chinese medicinal plants. *Science Bulletin*, 61(1), 3-17.

Yang, S. C., Lin, C. H., Aljuffali, I. A., and Fang, J. Y. (2017). Current pathogenic *Escherichia coli* foodborne outbreak cases and therapy development. *Archives of Microbiology*, 199, 811-825.

Yang, W., Chen, X., Li, Y., Guo, S., Wang, Z., and Yu, X. (2020). Advances in pharmacological activities of terpenoids. *Natural Product Communications*, 15(3), 1-10

Yang, Y. L., Al-Mahdy, D. A., Wu, M. L., Zheng, X. T., Piao, X. H., Chen, A. L., Wang, S. M., and Yang, Q. and Ge, Y. W. (2022). LC-MS-based identification and antioxidant evaluation of small molecules from the cinnamon oil extraction waste. *Food Chemistry*, 366, 130576.

Yap, P. S. X., Krishnan, T., Chan, K. G., and Lim, S. H. E. (2015). Antibacterial mode of action of *Cinnamomum verum* bark essential oil, alone and in combination with piperacillin, against a multi-drug-resistant *Escherichia coli* strain. *Journal of Microbiology and Biotechnology*, 25(8), 1299-1306.

Yong, Z., Yanan, Y., Zhi, Z., Yinfeng, T., Lin, D., Yiying, L., Weiying, L., Chongming, W. and Xiaopo, Z. (2022). Laurolitsine ameliorates type 2 diabetes by regulating the hepatic LKB1-AMPK pathway and gut microbiota. *Phytomedicine*, 106, 154423, 1-8

- Yu, X., Zhao, M., Liu, F., Zeng, S., and Hu, J. (2013). Identification of 2, 3-dihydro-3, 5-dihydroxy-6-methyl-4H-pyran-4-one as a strong antioxidant in glucose–histidine Maillard reaction products. *Food Research International*, 51(1), 397-403.
- Yusoff, S. F., Haron, F. F., Tengku Muda Mohamed, M., Asib, N., Sakimin, S. Z., Abu Kassim, F., and Ismail, S. I. (2021). Antifungal activity and phytochemical screening of *Vernonia amygdalina* extract against *Botrytis cinerea* causing gray mold disease on tomato fruits. *Biology*, 9(9), 286-293
- Zahid, M., Arif, M., Rahman, M. A., Singh, K., and Mujahid, M. (2018). Solvent extraction and gas chromatography–mass spectrometry analysis of *Annona squamosa* L. seeds for determination of bioactives, fatty acid/fatty oil composition, and antioxidant activity. *Journal of Dietary Supplements*, 15(5), 613-623.
- Zaki, A. A., Xu, X., Wang, Y., Shie, P. H., and Qiu, L. (2021). A new anti-inflammatory flavonoid glycoside from *Tetraena aegyptia*. *Natural Product Research*, 35(12), 1985-1990.
- Zaouri, N., Cui, Z., Peinetti, A. S., Lu, Y., and Hong, P. Y. (2019). DNAzyme-based biosensor as a rapid and accurate verification tool to complement simultaneous enzyme-based media for *E. coli* detection. *Environmental Science: Water Research and Technology*, 5(12), 2260-2268.
- Zargoosh, Z., Ghavam, M., Bacchetta, G., and Tavili, A. (2019). Effects of ecological factors on the antioxidant potential and total phenol content of *Scrophularia striata* Boiss. *Scientific Reports*, 9(1), 16021-16024
- Zaynab, M., Fatima, M., Abbas, S., Sharif, Y., Umair, M., Zafar, M. H., and Bahadar, K. (2018). Role of secondary metabolites in plant defense against pathogens. *Microbial Pathogenesis*, 124, 198-202.
- Zeki, Ö. C., Eylem, C. C., Reçber, T., Kır, S., and Nemutlu, E. (2020). Integration of GC–MS and LC–MS for untargeted metabolomics profiling. *Journal of Pharmaceutical and Biomedical Analysis*, 190, 113509, 1-7
- Zhang, A., Sun, H., Wang, P., Han, Y., and Wang, X. (2012). Modern analytical techniques in metabolomics analysis. *Analyst*, 137(2), 293-300.
- Zhang, G., Yan, X., Xia, J., Zhao, J., Ma, M., Yu, P., Gong, D. and Zeng, Z. (2021). Assessment of the effect of ethanol extracts from *Cinnamomum camphora* seed kernel on intestinal inflammation using simulated gastrointestinal digestion and a Caco-2/RAW264.7 co-culture system. *Food and Function*, 12(19), 9197-9210.
- Zhang, H., He, J., Li, B., Xiong, H., Xu, W., and Meng, X. (2011). Aflatoxin contamination and research in China. *Aflatoxins-Detection, Measurement and Control*, 21-36.

- Zhao, K. (2013). Chemical constituents from barks of *Cinnamomum cassia* growing in China. *Chinese Traditional and Herbal Drugs*, 2358-2363.
- Zhu, G., Zheng, X., Wang, Z., and Xu, X. (2022). Post-translational modifications by lipid metabolites during the DNA damage response and their role in cancer. *Biomolecules*, 12(11), 1655-1660
- Zmejkoski, D., Spasojević, D., Orlovska, I., Kozyrovska, N., Soković, M., Glamočlija, J., Dmitrovic, S., Matovic, B., Tasic, N., Maksiovic, V., Sosnin, M. and Radotić, K. (2018). Bacterial cellulose-lignin composite hydrogel as a promising agent in chronic wound healing. *International Journal of Biological Macromolecules*, 118, 494-503.
- Zong, L., Yang, Y., Zhang, J., Dai, L., Luo, Y., Yang, J., and Luo, X. (2021). *Rhododendron molle* G. Don extract induces apoptosis and inhibits migration in human colorectal cancer cells and potential anticancer components analysis. *Molecules*, 26(10), 2990-2998.
- Zulfa, Z., Chia, C. T., and Rukayadi, Y. (2016). In vitro antimicrobial activity of *Cymbopogon citratus* (lemongrass) extracts against selected foodborne pathogens. *International Food Research Journal*, 23(3), 1262-1273.