



**ANTIBACTERIAL AND ANTIOXIDANT ACTIVITIES, PHYTOCHEMICAL
AND TOXICITY ANALYSES OF *MELINJO* (*Gnetum gnemon* L.) LEAF
EXTRACT**

By

TRISHA MANSURA RAHMAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

January 2024

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DEDICATION

This thesis is dedicated to my beloved family, supervisors, and friends



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

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January 2024

Chairman : Associate Professor Yaya Rukayadi, PhD
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Global concern about foodborne pathogens is heightened due to their potential to cause widespread illness, economic loss, and worldwide undermining of food safety and security measures. Plant-derived antimicrobial compounds typically have a wider range of efficacy against foodborne pathogens than synthetic alternatives, in addition to being biodegradable and eco-friendly. Incorporating natural antimicrobial agents is crucial to mitigate the threat of antimicrobial resistance, promote sustainable practices, and lessen the possible adverse effects linked to synthetic substances. *Melinjo* (*Gnetum gnemon* L.) is one of the possible plant options. In Malaysia and Indonesia, several parts of the *Melinjo* plant, including leaves, seeds, and fruits, are utilized in culinary practices.

The antibacterial activities of ethanolic *Melinjo* leaf extract were determined using standard methods of CLSI, in terms of well diffusion assay (WDA),

minimum inhibitory concentration (MIC), minimum bactericidal concentration (MBC), and time-kill analysis. Total phenolic content (TPC) and antioxidant activities were performed using 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay, ferric reducing antioxidant power (FRAP) assay and 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid (ABTS) assay. *Melinjo* showed antibacterial activities against tested foodborne bacteria. The inhibition zone in WDA ranged from 8.50 to 13.50 mm, MIC ranged from 6.25 to 50.00 mg/mL and MBC values were between 12.5 to >50 mg/mL. Results from time-kill analysis showed that at 4×MIC bacterial strains were completely killed within 1 to 4 h incubation time. Total phenolic content was demonstrated at 39.4 ± 2.69 mg GAE/g, and DPPH and ABTS assay showed scavenging activity with the IC₅₀ value of 2.51 ± 0.26 mg/mL and 5.02 ± 0.30 mg/mL, respectively. Ferric-reducing antioxidant power activity was determined at 696.67 ± 13.56 µmol TEAC/g DW. The LC-MS analysis revealed the presence of various non-volatile compounds like anonaine, diosmetin 7-rutinoside, rhapontigenin, hamamelitannin, heteroflavanone C, tryptophan, [10]-gingerdiol. GC-MS analysis identified several volatile compounds such as phytol, vitamin E, ascorbic acid 2,6-dihexadecanoate, linoleic acid, gamma-sitosterol, and gamma-tocopherol. These identified compounds may have contributed to the antibacterial and antioxidant activities of *Melinjo* leaf extract. The LC₅₀ of leaf extract was found to be 41.08 mg/mL which is more than 1 mg/mL; meaning that the extract is non-toxic and safe for human consumption.

In conclusion, ethanolic *Melinjo* leaf extract exhibits antibacterial activities against selected foodborne bacteria, and antioxidant potential due to the

presence of bioactive compounds. It also is safe for consumption. Therefore, ethanolic *G. gnemon* L. extract leaf extract can be established as a natural anti-bacterial agent to be used in the development of natural food preservatives.

Keywords: Antibacterial, Antioxidant, *Gnetum Gnemon* L., Phytochemical, Toxicity.

SDG: GOAL 2: Zero Hunger, GOAL 3: Good Health and Well-Being



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**AKTIVITI ANTIBAKTERIA DAN ANTI-OKSIDAN, ANALISIS FITOKIMIA
DAN TOKSISITI EKSTRAK DAUN *MELINJO* (*Gnetum gnemon* L.)**

Oleh

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Kebimbangan global tentang patogen bawaan makanan meningkat kerana potensinya menyebabkan penyakit yang meluas, kehilangan ekonomi, dan menjejaskan keselamatan makanan dan langkah keselamatan di seluruh dunia. Sebatian antimikrob yang berasal dari tumbuhan biasanya mempunyai julat keberkesanan yang lebih luas terhadap patogen bawaan makanan daripada alternatif sintetik, selain boleh terbiodegradasi dan mesra alam. Menggabungkan agen antimikrob semulajadi adalah penting untuk mengurangkan ancaman rintangan antimikrob, menggalakkan amalan mampan, dan mengurangkan kemungkinan kesan buruk yang dikaitkan dengan bahan sintetik. *Melinjo* (*Gnetum gnemon* L.) adalah salah satu pilihan tumbuhan yang mungkin. Di Malaysia dan Indonesia, beberapa bahagian tumbuhan *Melinjo*, termasuk daun, biji, dan buah-buahan, digunakan dalam amalan masakan.

Aktiviti antibakteria ekstrak daun *Melinjo* etanolik ditentukan menggunakan kaedah standard CLSI, dari segi ujian resapan telaga (WDA), kepekatan perencatan minimum (MIC), kepekatan bakteria minimum (MBC), dan analisis bunuh masa. Jumlah kandungan fenolik (TPC) dan aktiviti antioksidan dilakukan menggunakan ujian 2,2-difenil-1-picrylhydrazyl (DPPH), ujian kuasa antioksidan pengurangan ferik (FRAP) dan ujian 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic asid (ABTS). *Melinjo* menunjukkan aktiviti antibakteria terhadap bakteria bawaan makanan yang diuji. Zon perencatan dalam WDA adalah antara 8.50 hingga 13.50 mm, MIC berkisar antara 6.25 hingga 50.00 mg/mL dan nilai MBC adalah antara 12.5 hingga >50 mg/mL. Keputusan daripada analisis bunuh masa menunjukkan bahawa pada 4×MIC strain bakteria terbunuh sepenuhnya dalam masa inkubasi 1 hingga 4 h. Jumlah kandungan fenolik ditunjukkan pada 39.4 ± 2.69 mg GAE/g, dan ujian DPPH dan ABTS menunjukkan aktiviti mengais dengan nilai IC_{50} masing-masing 2.51 ± 0.26 mg/mL dan 5.02 ± 0.30 mg/mL. Aktiviti kuasa antioksidan yang mengurangkan ferrik telah ditentukan pada 696.67 ± 13.56 mikron TEAC/g DW. Analisis LC-MS mendedahkan kehadiran pelbagai sebatian tidak meruap seperti anonaine, diosmetin 7-rutinoside, rhapontigenin, hamamelitannin, heteroflavanone C, tryptophan, [10]-gingerdiol. Analisis GC-MS mengenal pasti beberapa sebatian meruap seperti phytol, vitamin E, asid askorbik 2,6-dihexadecanoate, asid linoleik, gamma-sitosterol, dan gamma-tokoferol. Sebatian yang dikenal pasti ini mungkin menyumbang kepada aktiviti antibakteria dan antioksidan ekstrak daun *Melinjo*.

LC₅₀ ekstrak daun didapati 41.08 mg/mL yang lebih daripada 1 mg/mL; bermakna ekstrak itu tidak toksik dan selamat untuk kegunaan manusia. Sebagai kesimpulan, ekstrak daun *Melinjo* etanolik mempamerkan aktiviti antibakteria terhadap bakteria bawaan makanan terpilih, dan potensi antioksidan kerana kehadiran sebatian bioaktif. Ia juga selamat untuk dimakan. Oleh itu, ekstrak daun ekstrak *G. gnemon* L. etanol boleh ditubuhkan sebagai agen anti-bakteria semula jadi untuk digunakan dalam pembangunan pengawet makanan semula jadi.

Kata kunci: Antibakteria, Antioksidan, *Gnetum Gnemon* L., Fitokimia, Ketoksikan.

SDG: MATLAMAT 2: Sifar Kelaparan, MATLAMAT 3: Kesihatan dan Kesejahteraan Yang Baik

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LIST OF ABBREVIATIONS

ATTC	American Type Culture Collection
CDC	Centers for Disease Control and Prevention
CFU	Colony Forming Unit
CHX	Chlorhexidine
CLSI	Clinical and Laboratory Standards Institute
DMSO	Dimethyl sulfoxide
EMLE	Ethanollic <i>Melinjo</i> Leaf Extract
FAO	Food and Agriculture Organization
GC-MS	Gas Chromatography-Mass Spectrometry
GRAS	Generally Recognised as Safe
HPLC	High-Performance Liquid Chromatography
IC ₅₀	Half maximal inhibitory concentration
LC ₅₀	Median Lethality Concentration
LC-MS	Liquid Chromatography-Mass Spectrometry
<i>m/z</i>	mass/charge ratio
MBC	Minimum Bactericidal Concentration
MHA	Mueller Hinton agar
MHB	Mueller Hinton broth
MIC	Minimum Inhibitory Concentration
MPa	Mega Pascal
NIST	National Institute of Standards and Technology
PG	Peptidoglycan

CHAPTER 1

INTRODUCTION

1.1 Background

The variety of edible plants found in nature has sparked numerous breakthroughs in the research domain. The significant abundance of bioactive compounds within these plants plays a pivotal role in their application in the pharmaceutical and food industries as valuable resources. Edible plants are plants that are safe for human consumption and primarily used as food for their nutritional value and taste. They provide essential nutrients and can be part of a regular diet (Panda et al., 2019). Fruits, vegetables, grains, nuts, seeds, and certain herbs are examples of edible plants.

On the other hand, medicinal plants are those that have properties or compounds with potential health benefits and are used for medicinal purposes. These plants may have specific therapeutic properties and are often utilized to treat or alleviate various health conditions. Medicinal plants can be consumed as tea, extracts, and capsules, or used topically as ointments or poultices (Kurhekar, 2021). *Gnetum gnemon* L. has been used as both an edible and medicinal plant for therapeutic purposes among local and tribal people native to Southeast Asia and the western Pacific Ocean islands, ranging from Assam to Indonesia, Malaysia, Philippines, and Fiji (Barua et al., 2015).

Gnetum gnemon L. is nutritionally valuable, boasting a wealth of minerals and proteins. It is a promising candidate for providing healthy sustenance in underprivileged regions with limited access to protein-rich foods (Bhat & Binti, 2014). Moreover, this plant is known to be abundant in various beneficial compounds, including antioxidants, tyrosinase inhibitors, antimicrobial agents, anti-aging substances, and other bioactive compounds (Barua et al., 2015; Agarwal et al., 2020). *G. gnemon* L. holds significant importance as a leafy vegetable, particularly among tribal communities within Arunachal Pradesh, India. Additionally, the young twigs of this plant are readily available in local markets (Bharali et al., 2018). Despite the potential of different plants to treat various pathological conditions, a significant portion remains understudied, with only 15% having been analyzed for their phytochemical composition (Thomford et al., 2018).

It is widely recognized that numerous antimicrobial compounds derived from plants exhibit high efficacy against both Gram-positive and Gram-negative bacteria (Hochma et al., 2021). Over the period from 1981 to 2019, a total of 162 new antimicrobial drugs received approval, and intriguingly, 94 of them were derived from plant sources (Newman & Cragg, 2020). Edible plants are abundant sources of bioactive compounds that exert various physiological effects, encompassing anticancer, antioxidant, anti-inflammatory, and antimicrobial activities (Aisa et al., 2019). These natural plant extracts find frequent application in extending the shelf life of both fresh and processed foods, ensuring their quality and safety are preserved.

Phytochemical investigations into these extracts and their diverse biological activities are of significant importance in the food and human nutrition industries (Rapoport & Drausal, 2013). Antibiotics are essential in combating bacterial infections, but their effectiveness is being compromised by antibiotic resistance. Overuse and misuse of antibiotics have led to increased residual antibiotics in food and environments, causing the rapid spread of antibiotic-resistant bacteria (Ben et al., 2019). This poses a significant threat to public health, as infections become harder to treat, leading to longer recovery times, increased healthcare costs, and higher mortality rates.

The emergence of antibiotic-resistant bacteria, particularly multidrug resistance, is a serious problem in medical care, animal husbandry, food industry, and public health (Monger et al., 2021). The development of new antibiotics is lagging behind the spread of multidrug-resistant bacteria, requiring significant investment in research and development. Addressing antibiotic resistance and promoting responsible antibiotic use is crucial for public health (Kim & Ahn, 2022).

1.2 Problem Statements

Food is an essential requirement for sustaining human life nutritionally. Undesirable microbes can contaminate food products, food processing facilities, or manufacturing environments, posing a risk of human diseases (Yousef & Abdelhamid, 2019; Abdelhamid & El-Dougdoug, 2020).

Atabila et al. (2018) stated that access to safe and nutritious food is crucial for preserving life and improving health. They stated that contaminated meals including microbial infections and chemicals increase global illness and death rates. Food pathogens are microorganisms such as bacteria, viruses, or parasites present in food, beverages, or water that can lead to infections, intoxication, or toxin-mediated illnesses. They are categorized according to the categories of foods they contaminate (Madilo et al., 2023). Pathogens such as *Campylobacter*, *Salmonella*, *Shigella*, *Escherichia coli*, *Clostridium botulinum*, *Clostridium perfringens*, *Vibrio cholera*, *Staphylococcus aureus*, *Enterobacter*, *Bacillus* species, and *Listeria monocytogenes* are associated with foodborne illnesses.

These pathogens can be found in various raw or undercooked foods, meat, poultry, eggs, under-pasteurized milk, improperly canned foods, unpasteurized milk, as well as soft cheese (Havelaar et al., 2015). At present, food safety concerns have escalated globally because of the rise of foodborne illness due to microbial contamination (World Health Organization, 2021). Worldwide, almost 600 million individuals suffer foodborne diseases from consuming contaminated food, resulting in roughly 420,000 deaths daily (Szakály et al., 2020). To combat this situation, natural antimicrobial agents derived from plants could be the safest alternative to chemical antimicrobial agents and antibiotics.

Parhusip and Sitanggang (2011) reported the antimicrobial activity of *Melinjo* seed and peel extract against some pathogenic bacteria including *Bacillus*

cereus ATCC10876, *Staphylococcus aureus* ATCC25953, and *Enterobacter aerogenes* ATCC13048. Another study conducted by Dayoh et al. (2021) stated that ethanolic *Melinjo* leaf extract has antibacterial activity against *S. aureus* ATCC25923. One of the members of the *Gnetum* family is *G. africanum* Welw. which is a native plant of Africa. Ilodibia et al. (2015) reported antibacterial and anti-fungal activity of aquas and ethanolic *G. africanum* leaf extract against selected pathogens such as *S. aureus* (NR 201), *Escherichia coli* (NR 202), *Salmonella* Typhi (NR 204) and fungal strain *Aspergillus niger* (NR 241) and *Candida albican* (NR 242).

The antibacterial activity of *G. gnemon* L. leaf extract against various foodborne bacteria has not been previously reported. The current study aimed to address this gap in knowledge by investigating the potential of *G. gnemon* L. leaf extract as a natural antimicrobial agent. Due to the increasing concerns over the development of antibiotic resistance and the limitations associated with synthetic preservatives, there is a growing interest in exploring natural alternatives. The findings of this study can contribute to this area of research by suggesting that *G. gnemon* L. leaf extract may possess antibacterial properties against a range of foodborne pathogens.

Kato et al. (2011) and Santoso et al. (2010) documented the presence of many bioactive substances, such as saponins, tannins, and flavonoids, inside *G. gnemon* L. The *Melinjo* seed is known for its high content of resveratrol, a specific type of stilbenoid that is predominantly found in the form of dimers, namely gnetin C. This unique dimer form of resveratrol has been the subject

of research for over a decade, with particular focus and contributions from Japanese scientists (Barua et al., 2015; Wazir et al., 2011) who analyzed antioxidant activities of different parts of *G. gnemon* L. to determine the total phenolic and antioxidants activity of the plant. The quantification of total phenolic content in the plant extracts was conducted using the Folin-Ciocalteu method. Additionally, the assessment of antioxidant activities in the plant extracts was performed using three different assays: 2,2-diphenyl-1-picrylhydrazyl (DPPH), ferric reducing antioxidant power (FRAP), and 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid (ABTS) assays.

To the best of our knowledge, no previous study was done to determine the ABTS scavenging activity of *Melinjo* leaf extract and DPPH, FRAP, and total phenolic content (TPC) of ethanolic *Melinjo* leaf extract at the concentration used in this current study compared to the previous studies. Our findings not only shed light on the antioxidant potential of *Melinjo* leaf extract but also pave the way for future investigations into its application in various food products. Utilizing the antioxidant properties of plant extracts offers the possibility of reducing reliance on synthetic antioxidants, thereby meeting consumer preferences for clean-label and natural food products.

Phytochemical compounds are chemical substances found in plants that exhibit pharmacological and biological activities. These compounds are naturally produced by plants as part of their metabolic processes and can have various effects on living organisms. Phytochemicals are divided into two categories, namely primary and secondary metabolites, based on their roles

in plant metabolism. Primary metabolites, essential for plant survival, encompass carbohydrates, amino acids, proteins, lipids, purines, and pyrimidines found in nucleic acids.

On the other hand, secondary metabolites comprise the remaining plant chemicals that are produced through metabolic pathways derived from the primary ones (Bone and Mills, 2013; Hussein and El-Anssary, 2019). These secondary metabolites play crucial roles in protecting plants from pathogens as they possess antiviral, antifungal, and antibiotic properties. Additionally, they serve as essential UV-absorbing agents, safeguarding leaves from severe damage caused by light exposure (Bourgaud et al., 2001). Throughout history, these biologically active plant secondary metabolites have been utilized in traditional medicine, and the medicinal effects of plants can be attributed to these molecules (Jamwal et al., 2018). Multiple experiments were undertaken to investigate the presence of phytochemical substances in *G. gnemon* L. Previous research has identified various phytochemical compounds in multiple parts of *G. gnemon* L. plants, including stilbenoids, resveratrol, saponins, flavonoids, tannins, 3,4-dimethoxychlorogenic acid, gnemonols K and L, vitamin C, beta-carotene, lutein, polyphenols, and vitamin E (Anisong et al., 2022).

The present study represented the first attempt to analyze the ethanolic *Melinjo* leaf extract using LC-MS and GC-MS techniques. Despite the extensive research on phytochemical compounds in *Melinjo*, no prior investigations have been conducted using these analytical methods.

Over a significant period, individuals have incorporated *Melinjo* into their diets as both a leafy vegetable and for its medicinal attributes. *Melinjo* leaf has been included in culinary practices and utilized as remedies for health-related issues (Barua et al., 2015). This longstanding customary application suggests a reasonable level of safety. Nevertheless, there exists an absence of empirical evidence concerning the potential toxicity of *Melinjo* leaf, specifically when evaluated using scientific techniques such as the brine shrimp lethality assay. Therefore, it is crucial to undertake a systematic analysis of toxicity to substantiate the safety of *Melinjo* leaf, bridging the traditional usage with scientific validation.

1.3 Objectives

The general goals of this study were to explore the antibacterial and antioxidant potential of *Melinjo* (*Gnetum gnemon* L.) leaf extract. The specific objectives are:

1. To determine the antibacterial activity of *G. gnemon* L. leaf extract on foodborne pathogens in terms of well diffusion assay (WDA), minimum inhibition concentration (MIC), minimum bactericidal concentration (MBC), and killing time curve analysis.
2. To determine the total phenolic content (TPC) and antioxidant activity of *G. gnemon* L. leaf extract.
3. To identify the bioactive compounds in *G. gnemon* L. leaf extract which are responsible for antimicrobial and antioxidant activity by Gas Chromatography-Mass Spectrometry (GC-MS) and Liquid Chromatography-Mass Spectrometry (LC-MS).
4. To evaluate the toxicity of *G. gnemon* L. leaf extract using brine shrimp (*Artemia salina*) lethality assay.

1.4 Research Questions

This study explores four research questions to investigate its core objectives.

The research questions are:

1. Does the leaf extract of *Gnetum gnemon* L. exhibit antibacterial activity against foodborne pathogens when assessed through well diffusion assay (WDA), minimum inhibition concentration (MIC), minimum bactericidal concentration (MBC), and killing time curve analysis?
2. Does *Gnetum gnemon* L. leaf contain phenolic compounds and possess antioxidant activity? If so, what are their concentrations?
3. What are the bioactive compounds present in *Gnetum gnemon* L. leaf extract that contribute to its antimicrobial and antioxidant activity, as identified by Gas Chromatography-Mass Spectrometry (GC-MS) and Liquid Chromatography-Mass Spectrometry (LC-MS)?
4. What is the toxicity level of *Gnetum gnemon* L. leaf extract as determined by the brine shrimp (*Artemia salina*) lethality assay?

1.5 Hypothesis

This study investigates its research questions through four hypotheses. The hypotheses are:

1. The leaf extract of *Gnetum gnemon* L. has significant antibacterial activity against foodborne pathogens, evidenced by the inhibition zones observed in the well diffusion assay, low MIC and MBC values, and a notable decrease in bacterial population over time in the killing time curve analysis.
2. *Gnetum gnemon* L. leaf contains significant levels of phenolic contents and has antioxidant activity.
3. There are bioactive compounds present in *Gnetum gnemon* L. leaf extract which may contribute to its antimicrobial and antioxidant activity, as detected by Gas Chromatography-Mass Spectrometry (GC-MS) and Liquid Chromatography-Mass Spectrometry (LC-MS).
4. *Gnetum gnemon* L. leaf extract exhibits no toxicity in the brine shrimp (*Artemia salina*) lethality assay.

REFERENCES

- Abdelhamid, A. G., and El-DougDoug, N. K. (2020). Controlling foodborne pathogens with natural antimicrobials by biological control and antivirulence strategies. *Heliyon*, 6(9), e05020.
- Abdul Hussein, T. M., Allaithi, S. A., Mahdi, A. J., and Mouhsen, Z. M. (2017). Evaluation of inhibitory efficiency for some aqueous and alcoholic extracts of cumin, fenugreek, sweet fennel and black cumin in growth of *Escherichia coli* and *Staphylococcus aureus* bacteria. *Ibn Al-Haitham Journal for Pure and Applied Sciences*, 30(1), 279-287.
- Abebe, E., Gugsu, G., and Ahmed, M. (2020). Review on major food-borne zoonotic bacterial pathogens. *Journal of Tropical Medicine*, 4674235, 1–19.
- Abubakar, A. R., & Haque, M. (2020). Preparation of medicinal plants: Basic extraction and fractionation procedures for experimental purposes. *Journal of Pharmacy and Bioallied Sciences*, 12(1), 1.
- Abunna, F., Abriham, T., Gizaw, F., Beyene, T., Feyisa, A., Ayana, D., and Duguma, R. (2016). *Staphylococcus*: isolation, identification and antimicrobial resistance in dairy cattle farms, municipal abattoir and personnel in and around Asella, Ethiopia. *Journal of Veterinary Science and Technology*, 7(6), 1-7.
- Addis, M., and Sisay, D. (2015). A review on major food borne bacterial illnesses. *Journal of Tropical Diseases*, 3(4), 1-7.
- Agarwal, H., Dowarah, B., Baruah, P. M., Bordoloi, K. S., Krishnatraya, D. B., and Agarwala, N. (2020). Endophytes from *Gnetum gnemon* L. can protect seedlings against the infection of phytopathogenic bacterium *Ralstonia solanacearum* as well as promote plant growth in tomato. *Microbiological Research*, 238, 126503.
- Ahmat, N., Kamarozaman, A. S., Johari, M. S. M., Abas, F., Mohamad, S. A. S., and Yunoh, S. M. M. (2022). Screening of phytochemicals from the ethanolic extracts of *Gnetum gnemon*, *Gnetum latifolium* and *Cynometra malaccensis* of Kuala Keniam, Pahang. *IOP Conference Series: Earth and Environmental Science*, 1019(1), 012001.
- Aisa, H. A., Xin, X., and Tang, D. (2019). *Nigella sativa*: a medicinal and edible plant that ameliorates diabetes. *Bioactive Food as Dietary Interventions for Diabetes*, 2(2), 629-640.
- Aiyegoro, O. A., Afolayan, A. J., Okoh, and A. I. (2008). *In vitro* time kill assessment of crude methanol extract of *Helichrysum pedunculatum* leaves. *African Journal of Biotechnology*, 7(11), 1684–1688.

- Aklilu, A., Kahase, D., Dessalegn, M., Tarekegn, N., Gebremichael, S., Zenebe, S., Desta, K., Mulugeta, G., Mamuye, Y., and Mama, M. (2015). Prevalence of intestinal parasites, *Salmonella* and *Shigella* among apparently health food handlers of Addis Ababa University student's cafeteria, Addis Ababa, Ethiopia. *BMC Research Notes*, 8(1), 1-6.
- Alayande, K. A., Pohl, C. H., and Ashafa, A. O. T. (2017). Time-kill kinetics and biocidal effect of *Euclea crispa* leaf extracts against microbial membrane. *Asian Pacific Journal of Tropical Medicine*, 10(4), 390-399.
- Al-Garadi, M., Qaid, M., Alqhtani, A., Alhaji, M., Al-abdullatif, A., and Al-Mufarrej, S. (2023). *In vitro* antimicrobial efficacy assessment of ethanolic and aqueous extracts of *Cinnamon* (*Cinnamomum verum*) bark against selected microbes. *Brazilian Journal of Poultry Science*, 25(1). <https://doi.org/10.1590/1806-9061-2022-1682>
- Ali, F., Assanta, M. A., and Robert, C. (2011). *Gnetum africanum*: a wild food plant from the African forest with many nutritional and medicinal properties. *Journal of medicinal food*, 14(11), 1289-1297.
- Alimi, B. A., Lawal, R., and Odetunde, O. N. (2022). Food safety and microbiological hazards associated with retail meat at butchery outlets in north-central Nigeria. *Food Control*, 139, 109061.
- Almoulah, N. F., Voynikov, Y., Gevrenova, R., Schohn, H., Tzanova, T., Yagi, S., ... & Laurain-Mattar, D. (2017). Antibacterial, antiproliferative and antioxidant activity of leaf extracts of selected *Solanaceae* species. *South African Journal of Botany*, 112(1), 368-374.
- Al-Salih, D. A. A. K., Aziz, F. M., Mshimesh, B. A. R., and Jehad, M. T. (2013). Antibacterial effects of Vitamin E: *In vitro* study. *Journal of Biotechnology Research Center*, 7(2), 17–23.
- Andersson, A., Rönner, U., and Granum, P. E. (1995). What problems does the food industry have with the spore-forming pathogens *Bacillus cereus* and *Clostridium perfringens*? *International Journal of Food Microbiology*, 28(2), 145–155.
- Anisong, N., Siripongvutikorn, S., Wichienchot, S., and Puttarak, P. (2022). A comprehensive review on nutritional contents and functional properties of *Gnetum gnemon* Linn. *Food Science and Technology*, 42(1), e100121.
- Anthony, J.S., W. Zuchowski and W.N.Setzer, (2009). Composition of the floral essential oil of *Brugmansia suaveolens*. *Records of Natural Products*, 3(2): 76-81
- Apriliyanti, M. W., Ardiyansyah, and Handayani, A. M. (2018). Antioxidant activity, total phenol, and sensory properties of *Melinjo* peel tea with pre-treatment. *IOP Conference Series*, 207, 012044.

- Arabski, M., Wąsik, S., Dworecki, K., and Kaca, W. (2009). Laser interferometric and cultivation methods for measurement of colistin/ampicillin and saponin interactions with smooth and rough of *Proteus mirabilis* lipopolysaccharides and cells. *Journal of Microbiological Methods*, 77(2), 178–183.
- Argaw, S., and Addis, M. (2015). A review on staphylococcal food poisoning. *Food Science and Quality Management*, 40(1), 59-72.
- Arnesen LPS, Fagerlund A, Granum PE. (2008). From soil to gut: *Bacillus cereus* and its food poisoning toxins. *FEMS Microbiol Review*. 32(1),579–606.
- Ashurst, J. V. (2023). *Klebsiella pneumonia*. StatPearls - NCBI Bookshelf.
- Atabila, A., Phung, D. T., Hogarh, J. N., Sadler, R., Connell, D., & Chu, C. (2018). Health risk assessment of dermal exposure to chlorpyrifos among applicators on rice farms in Ghana. *Chemosphere*, 203, 83–89.
- Atwaa, E. S. H., Shahein, M. R., Radwan, H. A., Mohammed, N. S., Aloraini, M.A., Albezrah, N. K. A., Alharbi, M. A., Sayed, H. H., Daoud, M. A., and Elmahallawy, E. K. (2022). Antimicrobial activity of some plant extracts and their applications in homemade tomato paste and pasteurized cow milk as natural preservatives. *Fermentation*, 8(9), 428.
- Ayana, Z., Yohannis, M., and Abera, Z. (2015). Food-borne bacterial diseases in Ethiopia. *Journal of the Academy of Nutrition and Dietetics*, 4(1), 62-76.
- Aziz, M., and Karboune, S. (2018). Natural antimicrobial/antioxidant agents in meat and poultry products as well as fruits and vegetables: A review. *Critical Reviews in Food Science and Nutrition*, 58(3), 486-511.
- Baba, S. A., and Parah, S. A. (2015). Determination of total phenolic and flavonoid content, antimicrobial and antioxidant activity of a root extract of *Arisaema jacquemontii* Blume. *Journal of Taibah University for Science*, 9(4), 449–454.
- Bacon, R. T., and Sofos, J. N. (2003). Characteristics of biological hazards in foods. *Food safety handbook*, 10, 157-95.
- Balpetek, D., and Gürbüz, Ü. (2010). Investigations on the presence of *E. coli* O157: H7 in some meat products. *Eurasian Journal of Veterinary Sciences*, 26(1), 25-31.
- Bansal, A., and Priyadarsini, C. (2021). Medicinal properties of phytochemicals and their production. In Intech Open eBooks. *Natural Drugs from Plants*, pp.151-166, IntechOpen.

- Barbieri, R., Coppo, E., Marchese, A., Daglia, M., Sobarzo-Sánchez, E., Nabavi, S. F., and Nabavi, S. M. (2017). Phytochemicals for human disease: An update on plant-derived compounds antibacterial activity. *Microbiological Research*, 196, 44–68.
- Ben, Y., Fu, C., Hu, M., Liu, L., Wong, M. H., & Chen, K. (2019). Human health risk assessment of antibiotic resistance associated with antibiotic residues in the environment: A review. *Environmental Research*, 169, 483–493.
- Besser T, Goldoft M, Pritchett L, Khakhria R, Hancock D, Rice D, Gay J, Johnson W, and Gay C. (...) Multiresistant *Salmonella* Typhimurium DT104 infections of humans and domestic animals in the Pacific Northwest of the United States. *Epidemiology and Infection*, 124(2), 193-200.
- Beveridge, T. J. (1999). Structures of gram-negative cell walls and their derived membrane vesicles. *Journal of Bacteriology*, 181(16), 4725-4733.
- Bharali, P., Dutta, P., Kalita, M. C., Das, A. K., Tag, H., and Baruah, A. M. (2018). Evaluation of antioxidant and proximate compositions of the leaf extract of *Gnetum gnemon* L. *International Research Journal of Pharmacy*, 9(10), 101–105.
- Bhat, R., and Yahya, N. B. (2014). Evaluating *Belinjau* (*Gnetum gnemon* L.) seed flour quality as a base for development of novel food products and food formulations. *Food Chemistry*, 156, 42–49.
- Bhattachar, S. (2011). Natural antimutagens: A review. *Research Journal of Medicinal Plant*, 5(2), 116–126.
- Bhunja, A. K. (2018). Biology of microbial pathogens. *Foodborne Microbial Pathogens*, 25–42.
- Bi, X., and Xu, W. Y. (2013). An investigation of food poisoning caused by *Klebsiella pneumoniae*. *Chinese Practical Medicine* 8, 275–276.
- Bintsis, T. (2017). Foodborne pathogens. *AIMS Microbiology*, 3(3), 529-563.
- Bone, K., and Mills, S. (2012). Principles and practice of phytotherapy: Modern herbal medicine. *Elsevier Health Sciences*.
- Bonnet, M., Lagier, J., Raoult, D., & Khelaifia, S. (2020). Bacterial culture through selective and non-selective conditions: the evolution of culture media in clinical microbiology. *New Microbes and New Infections*, 34, 100622.
- Bourgaud, F., Gravot, A., Milesi, S., and Gontier, E. (2001). Production of plant secondary metabolites: a historical perspective. *Plant Science*, 161(5), 839–851.

- Breijyeh, Z., Jubeh, B., and Karaman, R. (2020). Resistance of Gram-negative bacteria to current antibacterial agents and approaches to resolve it. *Molecules*, 25(6), 1340.
- Brisse, S., Fevre, C., Passet, V., Issenhuth-Jeanjean, S., Tournebize, R., Diancourt, L., and Grimont, P. (2009). Virulent clones of *Klebsiella pneumoniae*: identification and evolutionary scenario based on genomic and phenotypic characterization. *PLoS ONE*, 4(3), e4982.
- Brovko, L. Y., Anany, H., and Griffiths, M. W. (2012). Bacteriophages for detection and control of bacterial pathogens in food and food-processing environment. *Advances in Food and Nutrition Research*, 67, 241-288.
- Brul, S., van Beilen, J., Caspers, M., O'Brien, A., de Koster, C., Oomes, S., Smelt, J., Kort, R., and Ter Beek, A. (2011). Challenges and advances in systems biology analysis of *Bacillus* spore physiology; molecular differences between an extreme heat resistant spore forming *Bacillus subtilis* food isolate and a laboratory strain. *Food Microbiology*, 28(2), 221–227.
- Bunk, B., Schulz, A., Stammen, S., Münch, R., Warren, M. J., Jahn, D., and Biedendieck, R. (2010). A short story about a big magic bug. *Bioengineered Bugs*, 1(2), 85–91.
- Burt, S., and Reinders, R. (2003). Antibacterial activity of selected plant essential oils against *Escherichia coli* O157:H7. *Letters in Applied Microbiology*, 36(3), 162–167.
- Cabral, A. B., Melo, R. D. C. D. A., Maciel, M. A. V., and Lopes, A. C. S. (2012, October). Multidrug resistance genes, including blaKPC and blaCTX-M-2, among *Klebsiella pneumoniae* isolated in Recife, Brazil. *Revista Da Sociedade Brasileira De Medicina Tropical*, 45(5), 572–578.
- Câmara, J., Herbert, P., Marques, J., and Alves, M. (2004). Varietal flavour compounds of four grape varieties producing madeira wines. *Analytica Chimica Acta*, 513(1), 203–207.
- Cao, X., Xu, X., Zhang, Z., Shen, H., Chen, J., and Zhang, K. (2014). Molecular characterization of clinical multidrug-resistant *Klebsiella pneumoniae* isolates. *Annals of Clinical Microbiology and Antimicrobials*, 13(1), 1-5.
- Carrique-Mas, J. J., and Bryant, J. E. (2013). A review of foodborne bacterial and parasitic zoonoses in Vietnam. *Ecohealth*, 10(4), 465-489.
- Catania, A. M., Civera, T., Di Ciccio, P. A., Grassi, M. A., Morra, P., and Dalmaso, A. (2021). Characterization of vegetative *Bacillus cereus* and *Bacillus subtilis* strains isolated from processed cheese products in an Italian dairy plant. *Foods*, 10(11), 2876.

- Ceyhan-Güvensen, N., and Keskin, D. (2016). Chemical content and antimicrobial properties of three different extracts of *Mentha pulegium* leaf from Mugla Region, Turkey. *Journal of Environmental Biology*, 37(6), 1341-1346.
- Charakida, A., Charakida, M., and Chu, A. (2007). Double-blind, randomized, placebo-controlled study of a lotion containing triethyl citrate and ethyl linoleate in the treatment of acne vulgaris. *British Journal of Dermatology*, 157(3), 569–574.
- Chen T. R., Wei Q. K., Chen Y. J. (2011). *Pseudomonas* spp. and *Hafnia alvei* growth in UHT milk at cold storage. *Food Control*, 22, 697–701.
- Chorianopoulos, N., Giaouris, E., Skandamis, P., Haroutounian, S., and Nychas, G. J. (2008). Disinfectant test against monoculture and mixed-culture biofilms composed of technological, spoilage and pathogenic bacteria: bactericidal effect of essential oil and hydrosol of *Satureja thymbra* and comparison with standard acid–base sanitizers. *Journal of Applied Microbiology*, 104(6), 1586–1596.
- Chuang, P. H., Lee, C. W., Chou, J. Y., Murugan, M., Shieh, B. J., and Chen, H. M. (2007). Anti-fungal activity of crude extracts and essential oil of *Moringa oleifera* Lam. *Bioresource Technology*, 98(1), 232–236.
- Cicerale, S., Lucas, L., and Keast, R. (2012). Antimicrobial, antioxidant and anti-inflammatory phenolic activities in extra virgin olive oil. *Current Opinion in Biotechnology*, 23(2), 129–135.
- CN110590522A - Synthesis method of megastigmatrienone - Google Patents. (2019). CN110590522A - Synthesis method of megastigmatrienone - google patents. <https://patents.google.com/patent/CN110590522A/en>
- Cobrado, L., Azevedo, M. M., Silva-Dias, A., Ramos, J. P., Pina-Vaz, C., and Rodrigues, A. G. (2012). Cerium, chitosan and hamamelitannin as novel biofilm inhibitors? *Journal of Antimicrobial Chemotherapy*, 67(5), 1159–1162.
- Cochrane, S. A., and Vederas, J. C. (2016). Lipopeptides from *Bacillus* and *Paenibacillus* spp.: A Gold Mine of Antibiotic Candidates. *Medicinal Research Reviews*, 36(1), 4–31.
- Coico, R. (2006). Gram staining. *Current Protocols In Microbiology*, (1), A-3C.
- Conlan, J. V., Sripa, B., Attwood, S., and Newton, P. N. (2011). A review of parasitic zoonoses in a changing Southeast Asia. *Veterinary Parasitology*, 182(1), 22-40.
- Cruz, A. G., Faria, J. A., Pollonio, M. A., Bolini, H. M., Celeghini, R. M., Granato, D., and Shah, N. P. (2011). Cheeses with reduced sodium content: effects on functionality, public health benefits and sensory properties. *Trends in Food Science & Technology*, 22(6), 276-291.

- Cusato, S., Gameiro, A. H., Corassin, C. H., Sant'Ana, A. S., Cruz, A. G., Faria, J. D. A. F., and de Oliveira, C. A. F. (2013). Food safety systems in a small dairy factory: Implementation, major challenges, and assessment of systems' performances. *Foodborne Pathogens and Disease*, 10(1), 6-12.
- Daglia, M. (2012). Polyphenols as antimicrobial agents. *Current Opinion in Biotechnology*, 23(2), 174–181.
- Damaceno, H. F. B., de Freitas Junior, C. V., Marinho, I. L., Cupertino, T. R., Costa, L. E. D. O., and Nascimento, J. D. S. (2015). Antibiotic resistance versus antimicrobial substances production by Gram-negative foodborne pathogens isolated from minas frescal cheese: heads or tails?. *Foodborne Pathogens and Disease*, 12(4), 297-301.
- Das, S., Chaudhari, A. K., Singh, A., Singh, V. K., Dwivedy, A. K., and Dubey, N. K. (2020). Foodborne microbial toxins and their inhibition by plant-based chemicals. In *Functional And Preservative Properties of Phytochemicals* (pp.165-207). Academic Press.
- Datta, P., and Gupta, V. (2019). Next-generation strategy for treating drug resistant bacteria: Antibiotic hybrids. *Indian Journal of Medical Research*, 149(2), 97.
- Davis, G. S., and Price, L. B. (2016). Recent research examining links among *Klebsiella pneumoniae* from food, food animals, and human extraintestinal infections. *Current Environmental Health Reports*, 3(2), 128–135.
- Dayoh, Paul and Isbandiati, Endang and Rahayu, Titien. (2021). Antibacterial effect of *Gnetum gnemon* L. leaf extract on *Staphylococcus aureus*. *Journal Widya Medika Junior*. 3(1), 122-130.
- De Vos, P., Garrity, G., Jones, D., Krieg, N., Ludwig, w., Rainey, F., Schleifer, K., and Whitman, W. (2009). Bergey's manual of systematic bacteriology: Volume 3: *The Firmicutes* (Vol. 3). Springer Science and Business Media.
- Del Nobile, M., Corbo, M., Speranza, B., Sinigaglia, M., Conte, A., and Caroprese, M. (2009) Combined effect of MAP and active compounds on fresh blue fish burger. *International Journal of Food Microbiology*, 135(3), 281–287.
- Delazar, Abbas and Nazifi, Ehsan and Movafeghi, Ali and Nahar, Dr Lutfun and Nazemiyeh, Hossein and Moghadam, S. and Asnaashari, Solmaz and Sarker, Satyajit. (2009). GC-MS analyses of *Ornithogalum procerum*. *DARU Journal of Pharmaceutical Sciences*, 17(1), 33-36.
- Department of Health and Human Services, U. S., and Food and Drug Administration, U. S. (2019). Bad Bug Book. *Handbook of Foodborne Pathogenic Microorganisms and Natural Toxins (2nd Edition)*.

- Desta Sisay, M. A. (2015). A review on major food borne bacterial illnesses. *Journal of Tropical Diseases*, 03(4), 55-65.
- Di Martino, P. (2022). Antimicrobial agents and microbial ecology. *AIMS Microbiology*, 8(1), 1–4.
- Diarra, M. S., Hassan, Y. I., Block, G. S., Drover, J. C., Delaquis, P., and Oomah, B. D. (2020). Antibacterial activities of a polyphenolic-rich extract prepared from American cranberry (*Vaccinium macrocarpon*) fruit pomace against *Listeria* spp. *LWT*, 123, 109056.
- Dietrich, R., Jessberger, N., Ehling-Schulz, M., Märklbauer, E., and Granum, P. E. (2021). The food poisoning toxins of *Bacillus cereus*. *Toxins*, 13(2), 98.
- Dilika, F., Bremner, P., and Meyer, J. (2000). Antibacterial activity of linoleic and oleic acids isolated from *Helichrysum pedunculatum*: a plant used during circumcision rites. *Fitoterapia*, 71(4), 450–452.
- Divyadharsini, V., Maheswari, T. U., Rajeshkumar, S., and Umamaheswari, T. N. (2023). Assessment of antimicrobial activity of lycopene, vitamin e, and lycopene-Vitamin E combination against *Staphylococcus aureus*, *Streptococcus mutans*, *Enterococcus faecalis*, and *Candida albicans*: An *in vitro* study. *Cureus*, 15(7), 14-16.
- Do Prado-Silva, L., Brancini, G. T., Braga, G. U., Liao, X., Ding, T., and Sant'Ana, A. S. (2022). Antimicrobial photodynamic treatment (apdt) as an innovative technology to control spoilage and pathogenic microorganisms in agri-food products: an updated review. *Food Control*, 132, 108527.
- Donnenberg, M. (Ed.). (2013). *Escherichia coli*: pathotypes and principles of pathogenesis. In *International Encyclopedia of Public Health* pp. 585-593, Academic Press.
- Dorman, H. J. D., and Deans, S. G. (2000). Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *Journal of Applied Microbiology*, 88(2), 308–316.
- Doyle, M. E., and Glass, K. A. (2010). Sodium reduction and its effect on food safety, food quality, and human health. *Comprehensive Reviews In Food Science And Food Safety*, 9(1), 44-56.
- Du, X., Finn, C. E., and Qian, M. C. (2010). Bound volatile precursors in genotypes in the pedigree of 'marion' blackberry (*Rubus* Sp.). *Journal of Agricultural and Food Chemistry*, 58(6), 3694–3699.
- Dunphy, L. J., Yen, P., & Papin, J. A. (2019). Integrated experimental and computational analyses reveal differential metabolic functionality in antibiotic-resistant *Pseudomonas aeruginosa*. *Cell Systems*, 8(1), 3-14.e3.

- Dutta, P. P., Bordoloi, M., Roy, S., Narzary, B., Gogoi, K., Bhattacharyya, D. R., Mohapatra, P. K., and Mazumder, B. (2018). Antiplasmodial activity of *Gnetum gnemon* leaf and compounds isolated from them. *Natural Product Communications*, 13(10), 1934578X1801301.
- Ekici, G., and Dümen, E. (2019). *Escherichia coli* and food safety. In *The universe of Escherichia coli*, pp.69-87. IntechOpen.
- Exner, M., Bhattacharya, S., Christiansen, B., Gebel, J., Goroncy-Bermes, P., Hartemann, P., and Trautmann, M. (2017). Antibiotic resistance: What is so special about multidrug-resistant Gram-negative bacteria?. *GMS Hygiene and Infection control*, 12.
- Ezekwe, A., Ugwueze, P., Nwankpa, P., Egwurugwu, J., Ekweogu, C., Emengaha, F., and Akukwu, D. (2020). Qualitative phytochemical screening, gcms studies and in-vitro anti-oxidative properties of aqueous leaf extract of *Gnetum africanum*. *Journal of Drug Delivery and Therapeutics*, 10(1), 11–17.
- F Gomes-Rochette, N., Da Silveira Vasconcelos, M., M Nabavi, S., F Mota, E., CS Nunes-Pinheiro, D., Daglia, M., and F De Melo, D. (2016). Fruit as potent natural antioxidants and their biological effects. *Current Pharmaceutical Biotechnology*, 17(11), 986-993.
- Falk, J., and Munné-Bosch, S. (2010). Tocochromanol functions in plants: antioxidation and beyond. *Journal of Experimental Botany*, 61(6), 1549–1566.
- Fernandes, R. P., Trindade, M. A., Tonin, F. G., Lima, C., Pugine, S. M. P., Munekata, P. E., Lorenzo, J. M., and De Melo, M. P. (2015). Evaluation of antioxidant capacity of 13 plant extracts by three different methods: cluster analyses applied for selection of the natural extracts with higher antioxidant capacity to replace synthetic antioxidant in lamb burgers. *Journal of Food Science and Technology*, 53(1), 451–460.
- Fernández-No, I., Guarddon, M., Böhme, K., Cepeda, A., Calo-Mata, P., and Barros-Velázquez, J. (2011). Detection and quantification of spoilage and pathogenic *Bacillus cereus*, *Bacillus subtilis* and *Bacillus licheniformis* by real-time PCR. *Food Microbiology*, 28(3), 605–610.
- Fluit, A. C. (2005). Towards more virulent and antibiotic-resistant *Salmonella*? *FEMS Immunology and Medical Microbiology*, 43(1), 1–11.
- Forbes-Hernández, T. Y., Giampieri, F., Gasparini, M., Mazzoni, L., Quiles, J. L., Alvarez-Suarez, J. M., and Battino, M. (2014). The effects of bioactive compounds from plant foods on mitochondrial function: A focus on apoptotic mechanisms. *Food and Chemical Toxicology*, 68, 154-182.
- Foster, T. J. (2002). *Staphylococcus aureus*. *Molecular Medical Microbiology*, 839-888.

- Freitas, R., Brito, M. A. V. P., Nero, L. A., and de Carvalho, A. F. (2013). Microbiological safety of Minas Frescal Cheese (MFC) and tracking the contamination of *Escherichia coli* and *Staphylococcus aureus* in MFC processing. *Foodborne Pathogens and Disease*, 10(11), 951-955.
- From, C., Hormazabal, V., and Granum, P. E. (2007). Food poisoning associated with pumilacidin-producing *Bacillus pumilus* in rice. *International Journal of Food Microbiology*, 115(3), 319–324.
- Galán, J. E. (2021). *Salmonella* Typhimurium and inflammation: a pathogen-centric affair. *Nature Reviews Microbiology*, 19(11), 716–725.
- Gao, X., Li, C., He, R., Zhang, Y., Wang, B., Zhang, Z. H., and Ho, C. T. (2023). Research advances on biogenic amines in traditional fermented foods: emphasis on formation mechanism, detection and control methods. *Food Chemistry*, 405, 134911.
- Garrity, G. M., Brenner, D. J., Krieg, N. R., Staley, J. R., and Manual, B. S. (2005). Systematic bacteriology. *The Proteobacteria, Part C: The Alpha-, Beta-, Delta-, and Epsilonproteobacteria*, *Bergey's Manual Trust, Department of Microbiology and Molecular Genetics*, 2.
- Giraffa, G., Chanishvili, N., and Widyastuti, Y. (2010). Importance of *Lactobacilli* in food and feed biotechnology. *Research in Microbiology*, 161(6), 480-487.
- Girish, H. V., and Satish, S. (2008). Antibacterial activity of important medicinal plants on human pathogenic bacteria-a comparative analysis. *World Applied Sciences Journal*, 5(3), 267-271.
- Gonelimali, F. D., Lin, J., Miao, W., Xuan, J., Charles, F., Chen, M., and Hatab, S. R. (2018). Antimicrobial properties and mechanism of action of some plant extracts against food pathogens and spoilage microorganisms. *Frontiers in Microbiology*, 9, 1639.
- Górniak, I., Bartoszewski, R. & Króliczewski, J. (2019). Comprehensive review of antimicrobial activities of plant flavonoids. *Phytochem Review* 18, 241–272
- Guo, Y., Wang, S., Zhan, L., Jin, Y., Duan, J., Hao, Z., and Yu, F. (2017). Microbiological and clinical characteristics of hypermucoviscous *Klebsiella pneumoniae* isolates associated with invasive infections in China. *Frontiers In Cellular and Infection Microbiology*, 7(1), 24.
- Gutbrod, P., Yang, W., Grujicic, G. V., Peisker, H., Gutbrod, K., Du, L. F., and Dörmann, P. (2021). Phytol derived from chlorophyll hydrolysis in plants is metabolized via phytenal. *Journal of Biological Chemistry*, 296, 100530.
- Gutierrez, J., Barry-Ryan, C., and Bourke, P. (2008). The antimicrobial efficacy of plant essential oil combinations and interactions with food ingredients. *International Journal of Food Microbiology*, 124(1), 91–97.

- Gutierrez, J., Barry-Ryan, C., and Bourke, P. (2009). Antimicrobial activity of plant essential oils using food model media: efficacy, synergistic potential and interactions with food components. *Food Microbiology*, 26(2), 142-150.
- Guyot-Declerck, C., Chevance, F., Lermusieau, G., and Collin, S. (2000). Optimized extraction procedure for quantifying norisoprenoids in honey and honey food products. *Journal of Agricultural and Food Chemistry*, 48(12), 5850–5855.
- Gyawali, R., and Ibrahim, S. A. (2014). Natural products as antimicrobial agents. *Food Control*, 46, 412–429.
- Halim, H. H., Pak Dek, M. S., Hamid, A. A., Saari, N., Mohd Lazim, M. I., Abas, F., Ngalim, A., Ismail, A., and Jaafar, A. H. (2023). Novel sources of bioactive compounds in coconut (*Cocos nucifera* L.) water from different maturity levels and varieties as potent skin anti-aging strategies and anti-fatigue agents. *Food Bioscience*, 51, 102326.
- Hariri, S. (2022). Detection of *escherichia coli* in food samples using culture and polymerase chain reaction methods. *Cureus*, 14(12), e32808
- Hartmann, M. S., Mousavi, S., Bereswill, S., and Heimesaat, M. M. (2020). Vitamin E as promising adjunct treatment option in the combat of infectious diseases caused by bacterial including multi-drug resistant pathogens –results from a comprehensive literature survey. *European Journal of Microbiology and Immunology*, 10(4), 193–201.
- Haryani, Y., Noorzaleha, A., Fatimah, A., Noorjahan, B., Patrick, G., Shamsinar, A., Laila, R., and Son, R. (2007). Incidence of *Klebsiella pneumonia* in street foods sold in Malaysia and their characterization by antibiotic resistance, plasmid profiling, and RAPD–PCR analysis. *Food Control*, 18(7), 847–853.
- Havelaar, A. H., Kirk, M., Torgerson, P. R., Gibb, H. J., Hald, T., Lake, R., Praet, N., Bellinger, D. C., De Silva, N. R., Gargouri, N., Speybroeck, N., Cawthorne, A., Mathers, C., Stein, C., Angulo, F. J., & Devleesschauwer, B. (2015). World Health Organization Global Estimates and regional comparisons of the burden of foodborne disease in 2010. *PLOS Medicine*, 12(12), e1001923.
- Helal, G. A., Sarhan, M. M., Shahla, A. N. K. A., & El-Khair, E. K. A. (2006). Effects of *Cymbopogon citratus* L. essential oil on the growth, lipid content and morphogenesis of *Aspergillus niger* ML2-strain. *Journal of Basic Microbiology*, 46(6), 456–469.
- Hemalata, V. B., and Virupakshaiah, D. B. M. (2016). Isolation and identification of food borne pathogens from spoiled food samples. *International Journal of Current Microbiology and Applied Sciences*, 5(6), 1017-1025.

- Heredia, N., and García, S. (2018). Animals as sources of food-borne pathogens: a review. *Animal Nutrition*, 4(3), 250–255.
- Hiko, A., Asrat, D., and Zewde, G. (2008). Occurrence of *Escherichia coli* O157: H7 in retail raw meat products in Ethiopia. *The Journal of Infection in Developing Countries*, 2(05), 389-393.
- Hiramoto, K., Nasuhara, A., Michikoshi, K., Kato, T., and Kikugawa, K. (1997). DNA strand-breaking activity and mutagenicity of 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-pyran-4-one (DDMP), a Maillard reaction product of glucose and glycine. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 395(1), 47–56.
- Hochma, E., Yarmolinsky, L., Khalfin, B., Nisnevitch, M., Ben-Shabat, S., and Nakonechny, F. (2021). Antimicrobial effect of phytochemicals from edible plants. *Processes*, 9(11), 2089.
- Høiby, N., Bjarnsholt, T., Givskov, M., Molin, S., & Ciofu, O. (2010). Antibiotic resistance of bacterial biofilms. *International Journal of Antimicrobial Agents*, 35(4), 322–332.
- Huang, C. B., George, B., and Ebersole, J. L. (2010). Antimicrobial activity of n-6, n-7 and n-9 fatty acids and their esters for oral microorganisms. *Archives of Oral Biology*, 55(8), 555–560.
- Huh, S., Kim, Y. S., Jung, E., Lim, J., Jung, K. S., Kim, M. O., Lee, J., and Park, D. (2010). Melanogenesis Inhibitory Effect of Fatty Acid Alkyl Esters Isolated from *Oxalis triangularis*. *Biological and Pharmaceutical Bulletin*, 33(7), 1242–1245.
- Humphrey, T. (2004). Salmonella, stress responses and food safety. *Nature Reviews Microbiology*, 2(6), 504–509.
- Húngaro, H., Peña, W., Silva, N., Carvalho, R., Alvarenga, V., and Sant'Ana, A. (2013). Food Microbiology. *Encyclopedia of Agriculture and Food Systems*, 2(1), 213-231.
- Hussain, M. (2016). Food Contamination: Major Challenges of the Future. *Foods*, 5(4), 21.
- Hussein, R. A., and El-Anssary, A. A. (2019). Plants secondary metabolites: the key drivers of the pharmacological actions of medicinal plants. *Herbal Medicine*, 1(3), 11-31.
- Hutt, P. B. (1984) Government regulation of the integrity of the food supply. *Annual Review of Nutrition*, 4(1), 1–21.
- Hygreeva, D., Pandey, M., and Radhakrishna, K. (2014). Potential applications of plant based derivatives as fat replacers, antioxidants and antimicrobials in fresh and processed meat products. *Meat Science*, 98(1), 47–57.

- Ibrahim, D., Hong, L. S., and Kuppan, N. (2013). Antimicrobial activity of crude methanolic extract from *Phyllanthus niruri*. *Natural Product Communications*, 8(4), 1934578X1300800.
- Iliya, I., Ali, Z., Tanaka, T., Iinuma, M., Furasawa, M., Nakaya, K. I., and Ubukata, M. (2003). Three new trimeric stilbenes from *Gnetum gnemon*. *Chemical and Pharmaceutical Bulletin*, 51(1), 85-88.
- Iliya, I., Ali, Z., Tanaka, T., Iinuma, M., Furasawa, M., Murata, J., and Darnaedi, D. (2002). Four new stilbene oligomers in the root of *Gnetum gnemon*. *Helvetica Chimica Acta*, 85(8), 2538-2546.
- Ilodibia, C. V., Ugwu, R. U., Nwokolo, O. L., Chukwuma, M., and Akachukwu, E. (2015). Phytochemical screening, antifungal and antibacterial activity of aqueous and ethanolic leaf and stem extracts of *Gnetum africanum* Welw. *Research Journal of Medicinal Plant*, 9(6), 275–283.
- Indu, M., Hatha, A., Abirosh, C., Harsha, U., and Vivekanandan, G. (2006). Antimicrobial activity of some of the South-indian spices against serotypes of *Escherichia coli*, *Salmonella*, *Listeria monocytogenes* and *Aeromonas hydrophila*. *Brazilian Journal of Microbiology*, 37(2), 153–158.
- International commission on microbiological specifications for food (1996). *Microorganisms in foods 5: Characteristics of microbial pathogens* (Vol. 5). Springer Science and Business Media.
- Ioannidis, A., Magana, M., Bologa, C., Oprea, T. I., Paulsen, I. T., & Tegos, G. P. (2015). Defining the microbial effluxome in the content of the host-microbiome interaction. *Frontiers in Pharmacology*, 6.
- Jafari-sales, A., Jafari, B., Khaneshpour, H. and Pashazadeh, M. (2020). Antibacterial effect of methanolic extract of *rosa damascena* on standard bacteria *Staphylococcus aureus*, *Bacillus cereus*, *Escherichia coli* and *Pseudomonas aeruginosa* in vitro. *International Journal of Nature and Life Sciences*, 4 (1), 40-46.
- Jaffee, S., Henson, S., Unnevehr, L., Grace, D., and Cassou, E. (2018). The safe food imperative: *Accelerating progress in low-and middle-income countries*. World Bank Publications.
- Jamwal, K., Bhattacharya, S., and Puri, S. (2018). Plant growth regulator mediated consequences of secondary metabolites in medicinal plants. *Journal of Applied Research on Medicinal and Aromatic Plants*, 9(1), 26–38.
- Jubeh, B., Breijyeh, Z., and Karaman, R. (2020). Resistance of Gram-positive bacteria to current antibacterial agents and overcoming approaches. *Molecules*, 25(12), 2888.

- Jubeh, B., Breijyeh, Z., and Karaman, R. (2020). Resistance of Gram-positive bacteria to current antibacterial agents and overcoming approaches. *Molecules*, 25(12), 2888.
- Kahkashan, P., Bokhari, N. A., Iffat, S., and Wahibi, M. S. (2014). *In vitro* antimicrobial activity of some medicinal plants found in Saudi Arabia. *Journal of Pure and Applied Microbiology*, 8(3), 2129-2134.
- Kato, H., Samizo, M., Kawabata, R., Takano, F., and Ohta, T. (2011). Stilbenoids from the *Melinjo* (*Gnetum gnemon* L.) fruit modulate cytokine production in murine peyer's patch cells ex vivo. *Planta Medica*, 77(10), 1027-1034.
- Khameneh, B., Iranshahy, M., Soheili, V., and Fazly Bazzaz, B. S. (2019). Review on plant antimicrobials: a mechanistic viewpoint. *Antimicrobial Resistance and Infection Control*, 8(1), 1-28.
- Khan, M. I., Ahhmed, A., Shin, J. H., Baek, J. S., Kim, M. Y., and Kim, J. D. (2018). Green tea seed isolated saponins exerts antibacterial effects against various strains of Gram-positive and Gram-negative bacteria, a comprehensive study *in vitro* and *in vivo*. *Evidence-based Complementary and Alternative Medicine: ECAM*, 2018.
- Kim, H. S., Chon, J. W., Kim, Y. J., Kim, D. H., Kim, M. S., and Seo, K. H. (2015). Prevalence and characterization of extended-spectrum- β -lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* in ready-to-eat vegetables. *International Journal of Food Microbiology*, 207(1), 83–86.
- Kim, J., & Ahn, J. (2022). Emergence and spread of antibiotic-resistant foodborne pathogens from farm to table. *Food Science and Biotechnology*, 31(12), 1481–1499.
- Kim, S., Lee, M., Park, T., and Rhee, M. (2013). A combined intervention using fermented ethanol and supercritical carbon dioxide to control *Bacillus cereus* and *Bacillus subtilis* in rice. *Food Control*, 32(1), 93–98.
- King, Thea, Martin Cole, Jeffrey M. Farber, Gerhard Eisenbrand, Dimitrios Zabaras, Edward M. Fox, and Jeremy P. Hill (2017): "Food safety for food security: relationship between global megatrends and developments in food safety." *Trends in Food Science & Technology* 68, 160-175.
- Ko, A., and Cho, S. K. (2018). Ethyl linoleate inhibits α -MSH-induced melanogenesis through Akt/gSK3 β / β -catenin signal pathway. *The Korean Journal of Physiology and Pharmacology: Official Journal of the Korean Physiological Society and the Korean Society of Pharmacology*, 22(1), 53-61.
- Kongkachuichai, R., Charoensiri, R., Yakoh, K., Kringkasemsee, A., and Insung, P. (2015). Nutrients value and antioxidant content of indigenous vegetables from Southern Thailand. *Food Chemistry*, 173, 838–846.

- Koohsari, H., Ghaemi, E. A., Sheshpoli, M. S., Jahedi, M., and Zahiri, M. (2015). The investigation of antibacterial activity of selected native plants from north of Iran. *Journal of Medicine and Life*, 8(2), 38.
- Kouadio, Y., Nanga Yessé, Z., Guichard, B., Kra Athanase, K., Amian, K., Luc Kouakou, K., ... and Guillaume Yao, L. (2012). Occurrence of *Enterobacter sakazakii* (Cronobacter) and other *Enterobacteriaceae* in commercial powdered infant formula in Abidjan, Ivory Coast. *Food and Nutrition Sciences*, 3(6), 200-205.
- Kumari and Sharma (2022). GC-MS analysis of the methanol fraction of *Ailanthus excelsa* roxb. Fruit. *Asian Journal of Pharmaceutical and Clinical Research*, 6(4), 51–55.
- Kurhekar, J. V. (2021). Ancient and modern practices in phytomedicine. In *preparation of phytopharmaceuticals for the management of disorders* (pp. 55-75). Academic Press.
- Kyahar, F. I., Onwuliri, E. A., Ehinmidu, J. O., and Oladosu, P. O. (2021). Time-kill kinetics and antibacterial activity of root extract of *Adenodolichos paniculatus* (Hua) Hutch & Dalz (Fabaceae). *Journal of Pharmacy & Bioresources*, 18(2), 95-102.
- Kyslychenko, V., Karpiuk, U., Ia, D., and Abu-Darwish, M. S. (2010). Phenolic compounds and terpenes in the green parts of *Glycine hispida*. *Advances in Environmental Biology*, 2(1) 490-495.
- Laslo, and György. (2018). Evaluation of the microbiological quality of some dairy products. *Acta Universitatis Sapientiae, Alimentaria*, 11(1), 27–44.
- Laslo, É., and György, É. (2018). Evaluation of the microbiological quality of some dairy products. *Acta Universitatis Sapientiae, Alimentaria*, 11(1), 27-44.
- Lee, J. Y., Kim, Y. S., and Shin, D. H. (2002). Antimicrobial synergistic effect of linolenic acid and monoglyceride against *Bacillus cereus* and *Staphylococcus aureus*. *Journal of Agricultural and Food Chemistry*, 50(7), 2193–2199.
- Lee, N. K., and Paik, H. D. (2016). Status, antimicrobial mechanism, and regulation of natural preservatives in livestock food systems. *Korean Journal for Food Science of Animal Resources*, 36(4), 547–557.
- Lee, S., M. Seo, M. Riu, J. Cotta, D. Block, N. Dokoozlian and S. Ebeler, (2007). Vine microclimate and norisoprenoid concentration in cabernet sauvignon grapes and wines. *American Journal of Enology and Viticulture*, 58(3), 291-301.
- Leff, J. W., and Fierer, N. (2013). Bacterial communities associated with the surfaces of fresh fruits and vegetables. *PLoS ONE*, 8(3), e59310.

- Li, A., Li, S., Zhang, Y., Xu, X., Chen, Y., & Li, H. (2014). Resources and biological activities of natural polyphenols. *Nutrients*, 6(12), 6020–6047.
- Li, H. T., Wu, H. M., Chen, H. L., Liu, C. M., and Chen, C. Y. (2013). The pharmacological activities of (–)-anonaine. *Molecules*, 18(7), 8257–8263.
- Li, W. R., Shi, Q. S., Liang, Q., Xie, X. B., Huang, X. M., and Chen, Y. B. (2014). Antibacterial activity and kinetics of *Litsea cubeba* oil on *Escherichia coli*. *PLoS ONE*, 9(11), e110983.
- Li, X., Gu, N., Huang, T. Y., Zhong, F., and Peng, G. (2023). *Pseudomonas aeruginosa*: a typical biofilm forming pathogen and an emerging but underestimated pathogen in food processing. *Frontiers in Microbiology*, 13, 1114199.
- Lim, T. K., (2012). *Gnetum gnemon*. *Edible Medicinal and Non-Medicinal Plants*: 3(1), 45-50.
- Lister, P., Wolter, D. J., & Hanson, N. D. (2009). Antibacterial-resistant *Pseudomonas aeruginosa*: clinical impact and complex regulation of chromosomally encoded resistance mechanisms. *Clinical Microbiology Reviews*, 22(4), 582–610.
- Liu, J., Willems, H. M. E., Sansevere, E. A., Allert, S., Barker, K. S., Lowes, D. J., Dixon, A. C., Xu, Z., Miao, J., DeJarnette, C., Tournu, H., Palmer, G. E., Richardson, J. P., Barrera, F. N., Hube, B., Naglik, J. R., and Peters, B. M. (2021). A variant ECE1 allele contributes to reduced pathogenicity of *Candida albicans* during vulvovaginal candidiasis. *PLOS Pathogens*, 17(9), e1009884.
- Loo, Y. Y., Rukayadi, Y., Kuan, C. H., Chieng, B. W., Nishibuchi, M., & Radu, S. (2018). *In vitro* antimicrobial activity of green synthesized silver nanoparticles against selected Gram-negative foodborne pathogens. *Frontiers in Microbiology*, 9, 379304.
- Lorenzo, J. M., Muneke, P. E., Dominguez, R., Pateiro, M., Saraiva, J. A., and Franco, D. (2018). Main groups of microorganisms of relevance for food safety and stability: General aspects and overall description. In *Innovative Technologies for Food Preservation* (pp. 53-107). Academic Press.
- Lou, Z., Wang, H., Lv, W., Ma, C., Wang, Z., & Chen, S. (2010). Assessment of antibacterial activity of fractions from burdock leaf against food-related bacteria. *Food Control*, 21(9), 1272-1278.
- Madilo, F. K., Islam, M. N., Letsyo, E., Roy, N., Klutse, C. M., Quansah, E., Darku, P. A., & Amin, M. B. (2023). Foodborne pathogens awareness and food safety knowledge of street-vended food consumers: A case of university students in Ghana. *Heliyon*, 9(7), e17795.

- Maffei, D. F., Batalha, E. Y., Landgraf, M., Schaffner, D. W., and Franco, B. D. (2016). Microbiology of organic and conventionally grown fresh produce. *Brazilian Journal of Microbiology*, 47, 99–105.
- Mahmoudi, S., Nasiri, R., and Jafari Sales, A. (2019). In-vitro antibacterial effects of methanolic extract of peppermint (*Mentha Piperita* Lamiaceae) on standard *Staphylococcus aureus*, *Bacillus cereus*, *Escherichia coli* and *Pseudomonas aeruginosa* strain. *Jorjani Biomedicine Journal*, 7(4), 4-10.
- Majumdar, A., Pradhan, N., Sadasivan, J., Acharya, A., Ojha, N., Babu, S., and Bose, S. (2018). Food degradation and foodborne diseases: a microbial approach. In *Microbial Contamination and Food Degradation* (pp. 109-148). Academic Press.
- Makwana, S., Choudhary, R., Haddock, J., and Kohli, P. (2015). In-vitro antibacterial activity of plant based phenolic compounds for food safety and preservation. *LWT - Food Science and Technology*, 62(2), 935–939.
- Mara, D., and Horan, N. J. (Eds.). (2003). Handbook of water and wastewater microbiology, pp 57-68. *Elsevier*.
- Martino, P. D. (2022). Antimicrobial agents and microbial ecology. *AIMS Microbiology*, 8(1), 1-4.
- Martinović, T., Andjelković, U., Gajdošik, M. Š., Rešetar, D., and Josić, D. (2016). Foodborne pathogens and their toxins. *Journal of Proteomics*, 147(1), 226-235.
- Masango, P. (2005). Cleaner production of essential oils by steam distillation. *Journal of Cleaner Production*, 13(8), 833–839.
- Mathew, S., and Abraham, T. E. (2006). In vitro antioxidant activity and scavenging effects of *Cinnamomum verum* leaf extract assayed by different methodologies. *Food and Chemical Toxicology*, 44(2), 198–206.
- Mathews, A., Arbal, A. V., Kaarunya, A., Jha, P. K., Le-Bail, A., & Rawson, A. (2024). Conventional vs modern extraction techniques in the food industry. In *Elsevier eBooks* (pp. 97–146).
- Meesilp, N., and Mesil, N. (2019). Effect of microbial sanitizers for reducing biofilm formation of *Staphylococcus aureus* and *Pseudomonas aeruginosa* on stainless steel by cultivation with UHT milk. *Food Science and Biotechnology*, 28, 289-296.
- Membre, J. M., and Guillou, S. (2016). Latest developments in foodborne pathogen risk assessment. *Current Opinion in Food Science*, 8(1), 120-126.

- Mendoza, N., and Silva, E. M. E. (2018). Introduction to phytochemicals: secondary metabolites from plants with active principles for pharmacological importance. *Phytochemicals: Source of Antioxidants and Role in Disease Prevention*, 25, 1-5.
- Miklasińska-Majdanik, M., Kępa, M., Wojtyczka, R. D., Idzik, D., and Wąsik, T. J. (2018). Phenolic compounds diminish antibiotic resistance of *Staphylococcus aureus* clinical strains. *International Journal of Environmental Research and Public Health*, 15(10), 2321.
- Miller, S. I. (2016). Antibiotic resistance and regulation of the Gram-negative bacterial outer membrane barrier by host innate immune molecules. *MBio*, 7(5), 10-1128.
- Miranda, R. C., and Schaffner, D. W. (2016). Longer contact times increase cross-contamination of *Enterobacter aerogenes* from surfaces to food. *Applied and Environmental Microbiology*, 82(21), 6490–6496.
- Mladenović, K. G., Grujović, M. Ž., Kiš, M., Furmeg, S., Tkalec, V. J., Stefanović, O. D., and Kocić-Tanackov, S. D. (2021). *Enterobacteriaceae* in food safety with an emphasis on raw milk and meat. *Applied Microbiology and Biotechnology*, 105(1), 8615–8627.
- Moi, I. M., Ibrahim, Z., Abubakar, B. M., Katagum, Y. M., Abdullahi, A., Yiga, G. A., and Ayuba, I. (2022). Properties of foodborne pathogens and their diseases. *Foodborne Pathogens-Recent Advances in Control and Detection*, IntechOpen.
- Moloney, M. G. (2016). Natural products as a source for novel antibiotics. *Trends in Pharmacological Sciences*, 37(8), 689–701.
- Monger, X. C., Gilbert, A., Saucier, L., and Vincent, A. T. (2021). Antibiotic resistance: From pig to meat. *Antibiotics*, 10(10), 1209.
- Moyes, R. B., Reynolds, J., and Breakwell, D. P. (2009). Differential staining of bacteria: gram stain. *Current Protocols in Microbiology*, 15(1), A-3C.
- Muthukumaran, P., Padmapriya, P., Salomi, S., Umamaheshwari, R., Kalaiarasan, P., and Malarvizhi, C. (2011). *In vitro* anti-microbial activity of leaf powder. *Asian Journal of Pharmaceutical Research*, 1(4), 108-110.
- Naguib, M. M., and Valvano, M. A. (2018). Vitamin E increases antimicrobial sensitivity by inhibiting bacterial lipocalin antibiotic binding. *Mosphere*, 3(6), e00564-18.
- Nawawi, D. S., Syafii, W., Akiyama, T., and Matsumoto, Y. (2016). Characteristics of guaiacyl-syringyl lignin in reaction wood in the gymnosperm *Gnetum gnemon* L. *Holzforschung*, 70(7), 593–602.

- Negi, P. S., Chauhan, A. S., Sadia, G. A., Rohinishree, Y. S., and Ramteke, R. S. (2005). Antioxidant and antibacterial activities of various seabuckthorn (*Hippophae rhamnoides* L.) seed extracts. *Food Chemistry*, 92(1), 119-124.
- Newman, D. J., and Cragg, G. M. (2020). Natural products as sources of new drugs over the nearly four decades from 01/1981 to 09/2019. *Journal of Natural Products*, 83(3), 770–803.
- Nicorescu, I., Nguyen, B., Moreau-Ferret, M., Agoulon, A., Chevalier, S., and Orange, N. (2013). Pulsed light inactivation of *Bacillus subtilis* vegetative cells in suspensions and spices. *Food Control*, 31(1), 151–157.
- Norajit, K., Laohakunjit, N., and Kerdchoechuen, O. (2007). Antibacterial effect of five *Zingiberaceae* essential oils. *Molecules*, 12(8), 2047–2060.
- Noreen, H., Semmar, N., Farman, M., & McCullagh, J. S. (2017). Measurement of total phenolic content and antioxidant activity of aerial parts of medicinal plant *Coronopus didymus*. *Asian Pacific Journal of Tropical Medicine*, 10(8), 792-801.
- Ogunmoye, A. O., Olubomehin, O. O., Atewolara-Odule, C. O., Ogundare, S. A., and Yussuf, S. T. (2020). GC-MS analysis of the volatile constituents from the air-dried leaf of *Terminalia catappa* (Linnaeus). *FUW Trends in Science and Technology Journal*, 5(3), 948-951.
- Ogunsina, O. I. (2020). Antibacterial activities and time-killing kinetics of *Lannea acida* extracts against selected microbes. *Agricultural and Biological Sciences Journal*, 6(3), 129-136.
- Oonaka, K., Furuhashi, K., Hara, M., and Fukuyama, M. (2010). Powder infant formula milk contaminated with *Enterobacter sakazakii*. *Japanese Journal of Infectious Diseases*, 63(2), 103-107.
- Overdevest, I., Heck, M., van der Zwaluw, K., Huijsdens, X., van Santen, M., Rijnsburger, M., Eustace, A., Xu, L., Hawkey, P., Savelkoul, P., Vandenbroucke-Grauls, C., Willemsen, I., van der Ven, J., Verhulst, C., and Kluytmans, J. (2014). Extended-spectrum β -lactamase producing *Klebsiella* spp. in chicken meat and humans: a comparison of typing methods. *Clinical Microbiology and Infection*, 20(3), 251–255.
- Panda, S. K., Mohanta, Y. K., Padhi, L., and Luyten, W. (2019). Antimicrobial activity of select edible plants from Odisha, India against food-borne pathogens. *Lwt*, 113, 108246.
- Pang, Z., Raudonis, R., Glick, B. R., Lin, T. J., and Cheng, Z. (2019). Antibiotic resistance in *Pseudomonas aeruginosa*: mechanisms and alternative therapeutic strategies. *Biotechnology Advances*, 37(1), 177–192.
- Parhusip, A. J. N., and Sitanggang, A. B. (2011). Antimicrobial activity of *Melinjo* seed and peel extract (*Gnetum gnemon*) against selected pathogenic bacteria. *Microbiology Indonesia*, 5(3), 2-2.

- Park, M., Bae, J., and Lee, D. S. (2008). Antibacterial activity of [10]-gingerol and [12]-gingerol isolated from ginger rhizome against periodontal bacteria. *Phytotherapy Research*, 22(11), 1446–1449.
- Park, S. Y., Seetharaman, R., Ko, M. J., Kim, D. Y., Kim, T. H., Yoon, M. K., Kwak, J. H., Lee, S. J., Bae, Y. S., and Choi, Y. W. (2014). Ethyl linoleate from garlic attenuates lipopolysaccharide-induced pro-inflammatory cytokine production by inducing heme oxygenase-1 in RAW264.7 cells. *International Immunopharmacology*, 19(2), 253–261.
- Pavic, S., Brett, M., Petric, I., Lastre, D., Smoljanovic, M., and Atkinson, M. (2005). An outbreak of food poisoning in a kindergarten caused by milk powder containing toxigenic *Bacillus subtilis* and *Bacillus licheniformis*. *Archiv für Lebensmittelhygiene*, 56(1), 20-22.
- Phuyal, N., Jha, P. K., Raturi, P. P., and Rajbhandary, S. (2020). Total phenolic, flavonoid contents, and antioxidant activities of fruit, seed, and bark extracts of *Zanthoxylum armatum* DC. *The Scientific World Journal*, 2(2), 79-86.
- Piggot, P. (2008). *Bacillus Subtilis*. *Encyclopedia of Microbiology*, 3(1), 45-56.
- Pisoschi, A. M., Cimpeanu, C., and Predoi, G. (2015). Electrochemical methods for total antioxidant capacity and its main contributor's determination: a review. *Open Chemistry*, 13(1), 000010151520150099.
- Pisoschi, A. M., Pop, A., Cimpeanu, C., and Predoi, G. (2016). Antioxidant capacity determination in plants and plant-derived products: a review. *Oxidative Medicine and Cellular Longevity*, 1–36.
- Pisoschi, A. M., Pop, A., Georgescu, C., Turcuş, V., Olah, N. K., and Mathe, E. (2018). An overview of natural antimicrobials' role in food. *European Journal of Medicinal Chemistry*, 143(1), 922–935.
- Podolak, R., Enache, E., Stone, W., Black, D. G., and Elliott, P. H. (2010). Sources and risk factors for contamination, survival, persistence, and heat resistance of salmonella in low-moisture foods. *Journal of Food Protection*, 73(10), 1919–1936.
- Poirel, L., Madec, J. Y., Lupo, A., Schink, A. K., Kieffer, N., Nordmann, P., and Schwarz, S. (2018). Antimicrobial resistance in *Escherichia coli*. *Microbiology Spectrum*, 6(4), 6-4.
- Potorti, A. G., Tropea, A., Lo Turco, V., Pellizzeri, V., Belfita, A., Dugo, G., and Di Bella, G. (2019). Mycotoxins in spices and culinary herbs from Italy and Tunisia. *Natural Product Research*, 34(1), 167–171.
- Puspanadan, S., Afsahhejri, L., Loo, Y. Y., Nillian, E., Kuan, C. H., Goh, S. G., et al. (2012). Detection of *Klebsiella pneumoniae* in raw vegetables using most probable number-polymerase chain reaction (MPN-PCR). *International Food Research Journal*, 19(4), 1757–1762.

- Rabizadeh, F., Mirian, M. S., Doosti, R., Kiani-Anbouhi, R., and Eftekhari, E. (2022). Phytochemical classification of medicinal plants used in the treatment of kidney disease based on traditional Persian medicine. *Evidence-Based Complementary and Alternative Medicine*, 1–13.
- Rahman, M. M., Islam, M. B., Biswas, M., and Khurshid Alam, A. H. M. (2015). *In vitro* antioxidant and free radical scavenging activity of different parts of *Tabebuia pallida* growing in Bangladesh. *BMC Research Notes*, 8(1), 1–9.
- Rahman, M. M., Sheikh, M. M. I., Sharmin, S. A., Islam, M. S., Rahman, M. A., Rahman, M. M., and Alam, M. F. (2009). Antibacterial activity of leaf juice and extracts of *Moringa oleifera* Lam. against some human pathogenic bacteria. *CMU Journal of Natural Science*, 8(2), 219.
- Rajagopal, M., and Walker, S. (2015). Envelope structures of Gram-positive bacteria. *Current Topics in Microbiology and Immunology*, 1–44.
- Rajkowski, K. T., and Smith, J. L. (2001). Update: food poisoning and other diseases induced by *Bacillus cereus*. In *Foodborne Disease Handbook* (Vol. 1, pp. 61–76). Marcel Dekker, Inc. New York.
- Rajurkar, N., and Hande, S. (2011). Estimation of phytochemical content and antioxidant activity of some selected traditional Indian medicinal plants. *Indian Journal of Pharmaceutical Sciences*, 73(2), 146.
- Rapoport, E. H., and Drausal, B. S. (2013). Edible Plants. *Encyclopedia of Biodiversity*, 2(2), 127–132.
- Rauha, J. P., Remes, S., Heinonen, M., Hopia, A., Kähkönen, M., Kujala, T., Pihlaja, K., Vuorela, H., and Vuorela, P. (2000). Antimicrobial effects of Finnish plant extracts containing flavonoids and other phenolic compounds. *International Journal of Food Microbiology*, 56(1), 3–12.
- Remington, J. S., and Schimpff, S. C. (1981). Please don't eat the salads. *New England Journal of Medicine*, 304(7), 433–435.
- Reygaert, W. (2018). An overview of the antimicrobial resistance mechanisms of bacteria. *AIMS Microbiology*, 4(3), 482–501.
- Rico, D., Martín-Diana, A. B., Baviera, J. M. B., and Barry-Ryan, C. (2007). Extending and measuring the quality of fresh-cut fruit and vegetables: a review. *Trends in Food Science and Technology*, 18(7), 373–386.
- Rønning, T. G., Aas, C. G., Støen, R., Bergh, K., Afset, J. E., Holte, M. S., and Radtke, A. (2018). Investigation of an outbreak caused by antibiotic-susceptible *Klebsiella oxytoca* in a neonatal intensive care unit in Norway. *Acta Paediatrica*, 108(1), 76–82.
- Rossiter, S. E., Fletcher, M. H., and Wuest, W. M. (2017). Natural products as platforms to overcome antibiotic resistance. *Chemical Reviews*, 117(19), 12415–12474.

- Rota, M. C., Herrera, A., MartíNez, R., Sotomayor, J. A., and Jordán, M. J. (2008). Antimicrobial activity and chemical composition of *Thymus vulgaris*, *Thymus zygis* and *Thymus hyemalis* essential oils. *Food Control*, 19(7), 681–687.
- Rowan N. J., Deans K., Anderson J. G., Gemmell C. G., Hunter I. S., and ChaithongT. (2001). Putative virulence factor expression by clinical and food isolates of *Bacillus* spp. after growth in reconstituted infant milk formulae. *Applied and Environmental Microbiology*, 67(9), 3873–3881.
- Rukayadi, Y., Lau, K.Y., Zainin, N.S., Zakaria, M. and Abas, F. (2013). Screening antimicrobial activity of tropical edible medicinal plant extracts against five standard microorganisms for natural food preservative. *International Food Research Journal*, 20(5): 2905-2910.
- Rukayadi, Y., Lee, K. H., Han, S. H., Kim, S. K., and Hwang, J. K. (2009). Antibacterial and sporicidal activity of macelignan isolated from nutmeg (*Myristica fragrans* Houtt.) against *Bacillus cereus*. *Food Science and Biotechnology*, 18(5), 1301-1304.
- S. A. Mustafa and L. A. Inanc, (2018). Antibiotic resistance of *Escherichia coli* O157: H7 isolated from chicken meats. *KSÜ Doğa Bilimleri Dergisi*, 21(1), 7-12.
- Sánchez-Hernández, E., Balduque-Gil, J., González-García, V., Barriuso-Vargas, J. J., Casanova-Gascón, J., Martín-Gil, J., and Martín-Ramos, P. (2023). Phytochemical profiling of *Sambucus nigra* L. Flower and leaf extracts and their antimicrobial potential against almond tree pathogens. *International Journal of Molecular Sciences*, 24(2), 1154.
- Sánchez-Hernández, E., Buzón-Durán, L., Cuchí-Oterino, J. A., Martín-Gil, J., Lorenzo-Vidal, B., and Martín-Ramos, P. (2022). Dwarf pomegranate (*punica granatum* L. Var. Nana): source of 5-hmf and bioactive compounds with applications in the protection of woody crops. *Plants*, 11(4), 550.
- Sánchez-Hernández, E., González-García, V., Palacio-Bielsa, A., Casanova-Gascón, J., Navas-Gracia, L. M., Martín-Gil, J., and Martín-Ramos, P. (2023). Phytochemical constituents and antimicrobial activity of *Euphorbia serrata* L. extracts for *Borago officinalis* L. crop protection. *Horticulturae*, 9(6), 652.
- Sani, N. A., and Yi, L. Y. (2011). *Enterobacteriaceae*, *Cronobacter* (*Enterobacter*) sakazakii and microbial population in infant formula products in the Malaysian market. *Sains Malaysiana*, 40(4), 345-351.
- Santoso, M., Naka, Y., Angkawidjaja, C., Yamaguchi, T., Matoba, T., and Takamura, H. (2010). Antioxidant and DNA damage prevention activities of the edible parts of *Gnetum gnemon* and their changes upon heat treatment. *Food Science and Technology Research*, 16(6), 549-556.

- Sari, M., Rahmawati, S. I., Izzati, F. N., and Putra, M. Y. (2023). Antioxidant activity of ethanolic extract of peel and seed *Melinjo* (*Gnetum gnemon*) based on color variations. In *1st international conference for health research–brin* (ICHR 2022) (pp. 255-265). Atlantis Press.
- Sauer, K., Camper, A. K., Ehrlich, G. D., Costerton, J. W., and Davies, D. G. (2002). *Pseudomonas aeruginosa* displays multiple phenotypes during development as a biofilm. *Journal of Bacteriology*, 184(4), 1140-1154.
- Scallan, E., Hoekstra, R. M., Angulo, F. J., Tauxe, R. V., Widdowson, M. A., Roy, S. L., Jones, J. L., and Griffin, P. M. (2011). Foodborne illness acquired in the United States—major pathogens. *Emerging Infectious Diseases*, 17(1), 7-15.
- Sen, C. K., Khanna, S., and Roy, S. (2007). Tocotrienols in health and disease: the other half of the natural vitamin E family. *Molecular Aspects of Medicine*, 28(5-6), 692-728.
- Shaheed, K. A., Alsirraj, M. A., Allaith, S. A., Noori, N. A., Obaid, M. H., and Mouhsan, Z. M. (2018). The biological activities of seeds extract for fenugreek and black cumin and its inhibitory influences towards some pathogens. *Iraq Medical Journal*, 2(2), 116-129.
- Shahidi, F., Pinaffi-Langley, A. C. C., Fuentes, J., Speisky, H., and de Camargo, A. C. (2021). Vitamin E as an essential micronutrient for human health: common, novel, and unexplored dietary sources. *Free Radical Biology and Medicine*, 176(1), 312-321.
- Sharifa, A. A., Neoh, Y. L., Iswadi, M. I., Khairul, O., Abdul Halim, M., Jamaludin, M., and Hing, H. L. (2008). Effects of methanol, ethanol and aqueous extract of *Plantago major* on gram positive bacteria, gram negative bacteria and yeast. *Ann Microsc*, 8(1), 42-44.
- Shimoi, K., Masuda, S., Shen, B., Furugori, M., and Kinae, N. (1996). Radioprotective effects of antioxidative plant flavonoids in mice. *Mutation Research: Fundamental and Molecular Mechanisms of Mutagenesis*, 350(1), 153-161.
- Shwaish, T., Al-Imarah, F. J., and Jasim, F. A. (2019). Wound healing capacity, antibacterial activity, and Gc-ms analysis of *Bienertia sinuspersici* leaf Extract. *Journal of Physics: Conference Series*, 1294(6), 062056.
- Sieberi, B. M., Omwenga, G., Wambua, R. K., Samoei, J. C., & Ngugi, M. P. (2020). Screening of the dichloromethane: methanolic extract of *Centella asiatica* for antibacterial activities against *Salmonella typhi*, *Escherichia coli*, *Shigella sonnei*, *Bacillus subtilis*, and *Staphylococcus aureus*. *The Scientific World Journal*, 2020(1), 1-8.
- Silhavy, T. J., Kahne, D., and Walker, S. (2010). The bacterial cell envelope. *Cold Spring Harbor Perspectives in Biology*, 2(5), a000414-a000414.

- Singh, R., Dar, S. A., and Sharma, P. (2012). Antibacterial activity and toxicological evaluation of semi purified hexane extract of *Urtica dioica* leaf. *Research Journal of Medicinal Plant*, 6(2), 123-135.
- Sizar, O., Leslie, S. W., and Unakal, C. G. (2023). Gram-positive Bacteria. In *Stat Pearls*. Stat Pearls Publishing.
- Skroza, D., Šimat, V., Smole Možina, S., Katalinić, V., Boban, N., & Generalić Mekinić, I. (2019). Interactions of resveratrol with other phenolics and activity against food-borne pathogens. *Food Science & Nutrition*, 7(7), 2312-2318.
- Sofos, J. N. (2008). Challenges to meat safety in the 21st century. *Meat Science*, 78(1-2), 3-13.
- Soobrattee, M. A., Neergheen, V. S., Luximon-Ramma, A., Aruoma, O. I., and Bahorun, T. (2005). Phenolics as potential antioxidant therapeutic agents: mechanism and actions. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 579(1-2), 200-213.
- Sozen, E., Demirel, T., and Ozer, N. K. (2019). Vitamin E: regulatory role in the cardiovascular system. *IUBMB Life*, 71(4), 507-515.
- Srisamran, J., Atwill, E. R., Chuanchuen, R., and Jeamsripong, S. (2022). Detection and analysis of indicator and pathogenic bacteria in conventional and organic fruits and vegetables sold in retail markets. *Food Quality and Safety*, 6, fyac013.
- Steele, M., and Odumeru, J. (2004). Irrigation water as source of foodborne pathogens on fruit and vegetables. *Journal of Food Protection*, 67(12), 2839-2849.
- Stenfors Arnesen, L. P., Fagerlund, A., and Granum, P. E. (2008). From soil to gut: *Bacillus cereus* and its food poisoning toxins. *FEMS Microbiology Reviews*, 32(4), 579-606.
- Stéphane, F. F. Y., Jules, B. K. J., Batiha, G. E., Ali, I., & Bruno, L. N. (2022). Extraction of Bioactive Compounds from Medicinal Plants and Herbs. In *IntechOpen eBooks*.
- Subramanian, G., Ram, M. S., Mongia, S., Singh, V., Ilavazhagan, G., and Sawhney, R. C. (2003). Evaluation of antioxidant activity of leaf extract of Seabuckthorn (*Hippophae rhamnoides* L.) on chromium(VI) induced oxidative stress in albino rats. *Journal of Ethnopharmacology*, 87(2-3), 247-251.
- Suman, T., Elumalai, D., Kaleena, P., and Rajasree, S. R. (2013). GC–MS analysis of bioactive components and synthesis of silver nanoparticle using *Ammannia baccifera* aerial extract and its larvicidal activity against malaria and filariasis vectors. *Industrial Crops and Products*, 47(1), 239-245.

- Sun, F., Wu, D., Qiu, Z., Jin, M., wang, X., and Li, J. (2010). Development of real-time PCR systems based on SYBR Green for the specific detection and quantification of *Klebsiella pneumoniae* in infant formula. *Food Control*, 21(4), 487-491.
- Suominen, I., Andersson, M. A., Andersson, M. C., Hallaksela, A. M., Kämpfer, P., Rainey, F. A., and Salkinoja-Salonen, M. (2001). Toxic *Bacillus pumilus* from indoor air, recycled paper pulp, norway spruce, food poisoning outbreaks and clinical samples. *Systematic and Applied Microbiology*, 24(2), 267-276.
- Sutherland, J., Miles, M., Hedderley, D., Li, J., Devoy, S., Sutton, K., and Lauren, D. (2009). Invitroeffects of food extracts on selected probiotic and pathogenic bacteria. *International Journal of Food Sciences and Nutrition*, 60(8), 717-727.
- Szakály, Z., Soós, M., Balsa-Budai, N., Kovács, S., & Kontor, E. (2020). The effect of an evaluative label on consumer perception of cheeses in hungary. *Foods* (Basel, Switzerland), 9(5), 563.
- Takó, M., Kerekes, E. B., Zambrano, C., Kotogán, A., Papp, T., Krisch, J., & Vágvolgyi, C. (2020). Plant phenolics and phenolic-enriched extracts as antimicrobial agents against food-contaminating microorganisms. *Antioxidants*, 9(2), 165.
- Talon, R., and Zagorec, M. (2017). Beneficial microorganisms for food manufacturing-fermented and biopreserved foods and beverages. *Microorganisms*, 5(4), 71.
- Taylor, J. M., Sutherland, A. D., Aidoo, K. E., and Logan, N. A. (2005). Heat-stable toxin production by strains of *Bacillus cereus*, *Bacillus firmus*, *Bacillus megaterium*, *Bacillus simplex* and *Bacillus licheniformis*. *FEMS Microbiology Letters*, 242(2), 313-317.
- Thampi, N., and Jeyadoss, V. S. (2015). In vitro time-kill and antiradical assays on green onion and garlic against specific diarrheagenic pathogens. *The Scitech Journal*, 2(03), 28-38.
- Thomford, N. E., Senthebane, D. A., Rowe, A., Munro, D., Seele, P. P., Maroyi, A., and Dzobo, K. (2018). Natural products for drug discovery in the 21st century: innovations for novel drug discovery. *International Journal of Molecular Sciences*, 19(6), 1578.
- Thompson, J. (2004). Warning of contamination of powdered infant formula. *Community Practitioner*, 77(4), 155.
- Tiwari, B. K., Valdramidis, V. P., O' Donnell, C. P., Muthukumarappan, K., Bourke, P., and Cullen, P. J. (2009). Application of natural antimicrobials for food preservation. *Journal of Agricultural and Food Chemistry*, 57(14), 5987-6000.

- Tropea, A. (2022). Microbial contamination and public health: an overview. *International Journal of Environmental Research and Public Health*, 19(12), 7441.
- Uçar G, Yörük NG, Güner A(2015). *Escherichia coli* infections. *Türkiye Klinikleri Journals Food Hygiene Technology*, 1(3), 22-29
- Ulusoy, S., Boşgelmez-Tınaz, G., and Seçilmiş-Canbay, H. (2009). Tocopherol, carotene, phenolic contents and antibacterial properties of rose essential oil, hydrosol and absolute. *Current Microbiology*, 59(5), 554-558.
- Vary, P. S., Biedendieck, R., Fuerch, T., Meinhardt, F., Rohde, M., Deckwer, W. D., and Jahn, D. (2007). *Bacillus megaterium*—from simple soil bacterium to industrial protein production host. *Applied Microbiology and Biotechnology*, 76(5), 957-967.
- Velu, G., Palanichamy, V., and Rajan, A. P. (2018). Phytochemical and pharmacological importance of plant secondary metabolites in modern medicine. *Bioorganic Phase in Natural Food: An Overview*, 135–156.
- Verpoorte, R. (2000). Pharmacognosy in the new millennium: leadfinding and biotechnology. *Journal of Pharmacy and Pharmacology*, 52(3), 253-262.
- Voda, K., Boh, B., Vrtačnik, M., and Pohleven, F. (2003). Effect of the antifungal activity of oxygenated aromatic essential oil compounds on the white-rot *Trametes versicolor* and the brown-rot *Coniophora puteana*. *International Biodeterioration and Biodegradation*, 51(1), 51–59.
- Vu, T. T., Kim, H., Tran, V. K., Le Dang, Q., Nguyen, H. T., Kim, H., Kim, I. S., Choi, G. J., and Kim, J. C. (2015). *In vitro* antibacterial activity of selected medicinal plants traditionally used in Vietnam against human pathogenic bacteria. *BMC Complementary and Alternative Medicine*, 16(1), 1-6.
- Vugia, D., Samuel, M., Farley, M., Marcus, R., Shiferaw, B., Shallow, S., Smith, K., and Angulo, F. (2004). Invasive *Salmonella* infections in the united states, foodnet, 1996–1999: incidence, serotype distribution, and outcome. *Clinical Infectious Diseases*, 38(s3), S149-S156.
- Wanakhachornkrai, O., Banglao, W., Thongee, A., and Sukplang, P. (2020). Determination of antioxidant, anti-aging and cytotoxicity activity of the essential oils from *Cinnamomum zeylanicum*. *Journal Of Microbiology, Biotechnology and Food Sciences*, 10(3), 436-440.
- Wazir, D., Ahmad, S., Muse, R., Mahmood, M., and Shukor, M. Y. (2011). Antioxidant activities of different parts of *Gnetum gnemon* L. *Journal of Plant Biochemistry and Biotechnology*, 20(2), 234-240.
- Williamson, J. (2016). The Oxford Companion to Wine. *Reference Reviews*, 30(4), 29-37.

- Wong, J. X., Ramli, S., and Chen, S. N. (2021). Assessment of toxic effect of *Centella asiatica* extract and its application as natural preservative in fresh-cut mango, pear and cabbage. *Journal of Food Processing and Preservation*, 45(10), e15824.
- World Health Organization. (2015). Food Safety: What you should know (No. SEA-NUT-196). World Health Organization.
- World Health Organization. (2021). World Health Organization model list of essential medicines: 22nd list (2021). (World Health Organization).
- Xiao, F., Xu, T., Lu, B., and Liu, R. (2020). Guidelines for antioxidant assays for food components. *Food Frontiers*, 1(1), 60-69.
- Xu, Z., Xie, J., Soteyome, T., Peters, B. M., Shirliff, M. E., Liu, J., and Harro, J. M. (2019). Polymicrobial interaction and biofilms between *Staphylococcus aureus* and *Pseudomonas aeruginosa*: an underestimated concern in food safety. *Current Opinion in Food Science*, 26(1), 57-64.
- Yahya, M., Ginting, B., and Saidi, N. (2021). *In-Vitro* screenings for biological and antioxidant activities of water extract from theobroma *Cacao L.* pod husk: potential utilization in foods. *Molecules*, 26(22), 6915-7015.
- Yang, D. C., Blair, K. M., and Salama, N. R. (2016). Staying in shape: the impact of cell shape on bacterial survival in diverse environments. *Microbiology and Molecular Biology Reviews*, 80(1), 187-203.
- Yassin, N. A., and Ahmad, A. M. (2012). Incidence and resistotyping profiles of *Bacillus subtilis* isolated from azadi teaching hospital in Duhok city, Iraq. *Materia Socio-medica*, 24(3), 194-197.
- Yousef A.E., Abdelhamid A.G. Behavior of microorganisms in food: growth, survival, and death. In: Doyle M.P., Diez-Gonzalez F., Hill C., Doyle M., Diez-Gonzalez F., Hill C., editors. *Food Microbiology: Fundamentals and Frontiers*. fifth ed. ASM Press; Washington, DC: 2019. pp. 3–21. ASM Press, Washington, DC, USA.
- Yu, X., Zhao, M., Liu, F., Zeng, S., and Hu, J. (2013). Identification of 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-pyran-4-one as a strong antioxidant in glucose–histidine maillard reaction products. *Food Research International*, 51(1), 397-403.
- Zainin, N.S., Lau, K.Y., Zakaria, M., Son, R., Abdull Razis, A.F. and Rukayadi, Y. (2013). Antibacterial activity of *Boesenbergia rotunda* (L.) Mansf. A. extract against *Escherichia coli*. *International Food Research Journal*, 20(6), 3319-3323.
- Zhang, Q., Lin, Y., Shen, G., Zhang, H., and Lyu, S. (2022). Siderophores of *Bacillus pumilus* promote 2-keto-L-gulonic acid production in a vitamin C microbial fermentation system. *Journal of Basic Microbiology*, 62(7), 833-842.

- Zhang, R., Huo, W., Zhu, W., and Mao, S. (2015). Characterization of bacterial community of raw milk from dairy cows during subacute ruminal acidosis challenge by high-throughput sequencing. *Journal of the Science of Food and Agriculture*, 95(5), 1072-1079.
- Zhang, S., Yang, G., Ye, Q., Wu, Q., Zhang, J., and Huang, Y. (2018). Phenotypic and genotypic characterization of *Klebsiella pneumoniae* isolated from retail foods in china. *Frontiers in Microbiology*, 9(1), 110-119.
- Zheleva-Dimitrova, D., Nedialkov, P., & Kitanov, G. (2010). Radical scavenging and antioxidant activities of methanolic extracts from *Hypericum* species growing in Bulgaria. *Pharmacognosy Magazine*, 6(22), 74.
- Zheng, C. J., Yoo, J. S., Lee, T. G., Cho, H. Y., Kim, Y. H., and Kim, W. G. (2005). Fatty acid synthesis is a target for antibacterial activity of unsaturated fatty acids. *FEBS Letters*, 579(23), 5157-5162.
- Zhu, X., Gao, Y., Chen, Z., and Su, Q. (2009). Development of a chromatographic fingerprint of tobacco flavor by use of GC and GC-MS. *Chromatographia*, 69(1), 735-742.
- Zingg, J. M. (2007). Modulation of signal transduction by vitamin E. *Molecular Aspects of Medicine*, 28(5-6), 481-506.
- Zizovic, I., Stamenić, M., Orlović, A., and Skala, D. (2005). Supercritical carbon dioxide essential oil extraction of *Lamiaceae* family species: Mathematical modelling on the micro-scale and process optimization. *Chemical Engineering Science*, 60(23), 6747-6756.
- Zwietering, M. H., Jacxsens, L., Membré, J. M., Nauta, M., and Peterz, M. (2016). Relevance of microbial finished product testing in food safety management. *Food Control*, 60, 31-43.