



**TRACING THE DOMESTICATION HISTORY OF MANGOSTEEN
(*Garcinia mangostana* L.) USING MORPHOLOGICAL AND GENETIC
APPROACHES**

By

YAO TZE LEONG

**Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia and University of Montpellier
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

November 2023

FPAS 2023 16

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia and University of Montpellier, in fulfilment of the requirement for the degree of Doctor of Philosophy

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Chairperson : Associate Professor Mohd Nazre Saleh, PhD
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Rainforests in Southeast Asia harbour a great diversity of edible fruits. One of the most emblematic is mangosteen (*Garcinia mangostana* L. var. *mangostana*; Clusiaceae), popularly known as the queen of fruits for its exquisite sweet-sour taste and aesthetic appearance. Previous cytological and phylogenetics studies reported contrasting results on the biological origin of the cultivated mangosteen. The more recent findings suggested the wild populations of *G. mangostana* var. *malaccensis* as the most probable sole progenitor of the cultivated mangosteen but the overview of its domestication history is far from complete. Against the backdrop of the contradictory theories of its biological origin, the current study set to trace back the domestication history of cultivated mangosteen. Our textual review revealed the earliest written records available indicated that fruits of mangosteen have been sold in marketplaces in the major seaports in the Malay Archipelago for at least 600 years. To elucidate the phenotypic affinities among the cultivated mangosteen and wild relatives, we used morphology as a proxy. In the morphometric analyses, we included 124 specimens consisting of cultivated and wild mangosteens. A total of 38 characters are included in this study. We used Principal Coordinates Analysis (PCoA) and Ascendent Hierarchical Classification (AHC) to evaluate the morphological characters used in species delimitation. Our findings showed that wild mangosteen species can be clearly delineated from cultivated mangosteen using morphological characters. However, we could not find character or combination of characters with statistical support that delimitate taxonomic varieties in *G. mangostana*. We employed cpDNA SNPs as the genetic markers to examine phylogeny and to infer haplotypes. We selected 192 individuals for genomic library construction. Raw reads were generated applying high-throughput shotgun sequencing. We plotted cladogram and haplotype network diagram to illustrate the genealogy of cultivated and wild mangosteen. The haplotype network consists of populations distributed in East and West Coasts of Peninsular Malaysia, Singapore, Sarawak, Sabah and Africa. This network enables us to interpret the effects of seed dispersals on geneflows in the cultivated and wild populations, a congregate haplotype consists of cultivated mangosteen from a few

regions exemplified that cultivated mangosteen is assumably disseminated by cultivators over long time. Among wild populations of var. *borneensis* and var. *malaccensis*, the gene flow is relatively localised. The localised gene flow is most probably the consequences of dispersals by primates. Considering the findings from morphometric and genetic analyses, we conclude that a distinction between the wild and cultivated compartments of mangosteen is not attainable. Regarding the domestication history, our findings demonstrated that the cultivated var. *mangostana* of Peninsular Malaysia and Singapore arose from the wild populations of var. *malaccensis*. We hope this study would spur interest among researchers across the Southeast Asian countries to further clarify the taxonomy, phylogeny and genetic diversities of wild *Garcinia* congeners, as well as other poorly understood cultivated trees at regional scope.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia dan Universiti Montpellier, sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**MENJEJAKI SEJARAH DOMESTIKASI MANGGIS (*Garcinia mangostana* L.)
MENGUNAKAN PENDEKATAN MORFOLOGI DAN GENETIK**

Oleh

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November 2023

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Hutan hujan di Asia Tenggara kaya dengan kepelbagaian spesis pokok buah. Salah satu spesis yang paling terkenal ialah manggis (*Garcinia mangostana* L. var. *mangostana*; Clusiaceae), juga dikenali sebagai ratu buah-buahan kerana rasa manis-masam yang enak dan penampilan yang estetik. Kajian sitologi dan filogenetik sebelum ini melaporkan keputusan yang berbeza mengenai asal biologi manggis yang ditanam. Penemuan yang lebih terkini mencadangkan populasi liar *G. mangostana* var. *malaccensis* sebagai moyang yang paling berkemungkinan bagi manggis tetapi gambaran keseluruhan sejarah domestikasi masih jauh dari lengkap. Berlatarbelakangkan teori-teori yang bercanggah tentang asal usul biologinya, kajian ini bertujuan untuk menjejaki sejarah domestikasi manggis. Kajian tekstual kami mendapati rekod bertulis terawal menunjukkan bahawa buah manggis telah dijual di pasaran di pelabuhan utama di sekitar Kepulauan Melayu selama 600 tahun sekurang-kurangnya. Untuk memperjelaskan pertalian fenotip antara manggis dan saudara liar, kami menggunakan morfologi sebagai proksi. Dalam analisis morfometrik, kami melibatkan 124 spesis yang terdiri daripada manggis kampung dan taksa liar. Sebanyak 38 karakter morfologi diambil kira dalam kajian ini. Kami menggunakan *Principal Coordinate Analysis* (PCoA) dan *Ascending Hierarchical Classification* (AHC) untuk menilai karakter morfologi yang digunakan dalam persempadanan semula spesies. Penemuan kami menunjukkan bahawa spesies manggis liar boleh ditakrifkan dengan jelas berbeza daripada manggis menggunakan karakter morfologi. Walau bagaimanapun, kami tidak menemui karakter atau gabungan karakter dengan sokongan statistik yang membatasi jenis variasi taksonomi dalam *G. mangostana*. Kami menggunakan SNP cpDNA sebagai penanda genetik untuk kajian filogeni dan haplotip. Kami memilih 192 individu untuk *genomic library construction*. Bacaan jujukan dijana menggunakan teknologi *high-throughput shotgun sequencing*. Kami memplot rajah rangkaian kladogram dan haplotaip untuk menggambarkan silsilah manggis dan spesis liar. Rangkaian haplotaip terdiri daripada populasi yang bertaburan di Pantai Timur dan Barat Semenanjung Malaysia, Singapura, Sarawak, Sabah dan Afrika. Rangkaian ini membolehkan kami mentafsir kesan penyebaran benih pada aliran gen dalam populasi

yang ditanam dan liar, haplotaip berkumpul terdiri daripada manggis yang ditanam dari beberapa kawasan yang menunjukkan bahawa manggis yang ditanam mungkin disebarkan oleh penanam dalam jangka masa yang lama. Antara populasi liar var. *borneensis* dan var. *malaccensis*, aliran gen agak setempat. Aliran gen setempat kemungkinan besar adalah akibat daripada penyebaran oleh haiwan primata. Memandangkan dapatan daripada analisis morfometrik dan genetik, kami menyimpulkan bahawa perbezaan antara manggis liar dan ditanam tidak dapat dicapai. Mengenai sejarah domestikasi, penemuan kami menunjukkan bahawa var. *mangostana* Semenanjung Malaysia dan Singapura muncul daripada populasi liar var. *malaccensis* setempat. Kami berharap kajian ini menyemarakkan minat di kalangan penyelidik di seluruh negara Asia Tenggara untuk memperdalamkan kajian taksonomi, filogeni dan kepelbagaian genetik spesies *Garcinia* liar, serta spesies pokok-pokok buah lain yang kurang difahami pada skop serantau.

ACKNOWLEDGEMENTS

“It took a village to raise a child” a proverb goes, and I would say “it took the whole world to complete a PhD study”! Thus, I humbly take this opportunity to thank all authorities, organisations and individuals that made everything related to this PhD project possible.

I gratefully acknowledge funding from Labex AGRO 2011-LABX-002 (under I-Site MUSE framework) coordinated by Agropolis Fondation, IRD-ARTS fellowship, Universiti Putra Malaysia (UPM), the Institute of Tropical Agriculture and Food Security (ITAFoS, UPM), and Southeast Asian Regional Center for Graduate Study and Research in Agriculture (SEARCA). I thank Forest Research Institute Malaysia generously granted a four-year full salary study leave and International Association for Plant Taxonomy (IAPT) for an IAPT Dan Nicolson Grant that enabled field trips and herbaria visits.

The following authorities of three nations are acknowledged for the permits granted to carry out sampling fieldworks in the territory under their jurisdiction: INDONESIA: Direktorat Tata Kelola Perizinan Riset dan Inovasi dan Otoritas Ilmiah, Indonesia: surat izin penelitian (letter of research permit, nomor: 101/SIP/IV/FR/7/2022); MALAYSIA: Jabatan Perhutanan Semenanjung Malaysia, no. rujukan surat JH/100Jld.23(15); Sabah Access License Ref. Number JKM/MBS.1000-2/2 JLD.10 (116); Sabah license for transfer of biological resources License Ref. Number JKM/MBS.1000-2/3 JLD.4 (61); Jabatan Perhutanan Negeri Terengganu, permit masuk hutan no. 1067/2020; Taman Pertanian Universiti Putra Malaysia, no. permohonan 009/2020, ruj. ‘UPM/FH/JPUH/1A-09’; Pejabat Hutan Daerah Terengganu Utara Ruj. ‘bil.(4)d/m.PHDTU.1/15/2 BHG.6; Hutan Simpan Ayer Hitam, Puchong, ruj. ‘UPM/FH/SHL/100-3/5’; SINGAPORE: Singapore National Parks Board, Permit number NP/RP20-004.

I thank the curators and managers of herbaria at Naturalis Biodiversity Centre, Netherlands (AMS, L, U); Bogor, Indonesia (BO); Institut National de la Recherche Agronomique and Parc National de Guadeloupe (GUAD); Royal Botanic Garden Kew, UK (K); Forest Research Institute Malaysia, Kepong (KEP); Malaya University, Malaysia (KLU); Malaysian Agricultural Research and Development Institute (MDI); University of Montpellier, France (MPU); National Museum of Natural History, France (P); Sabah Forestry Department, Malaysia (SAN), Sarawak Forestry Department, Malaysia (SAR), Singapore Botanic Garden (SING); Smithsonian Institution, United States of America (US); and Institute of Research and Development for Natural Resource Conservation Technology, Wanariset, East Kalimantan, Indonesia (WAN) for allowing me to examine the specimens under their care, sending specimens on loan, or making the images of their specimens available for this study. Also, curators of KEP, KLU, P, SAN, SAR, WAN are gratefully acknowledged for consenting permission to me to extract leaf tissues from herbarium specimens for genetic analyses.

My heartfelt gratitude goes to my amazing thesis co-directors of the cotutelle studentship, Dr. Jérôme Duminil and Assoc. Prof. Dr Mohd Nazre Saleh. Their inspiring academic guidance and solid administrative support are vital to this PhD project.

Comprehensive examination examiners at UPM, Dr Christina Yong Seok Yien and Dr Razak Terhem; and committee members of three UM *Comité de suivi* evaluations in years 2020, 2021 and 2022, Prof. Dr Doyle McKey, Dr Yves Vigouroux, Prof. Dr Olivier Hardy, Dr Jérôme Munzinger and Dr Riina Jalonen are gratefully acknowledged for their comments and suggestion in keeping this PhD project on the right track.

Many individuals have helped in facilitating the sampling work and herbarium visits: INDONESIA: Dr Deden Derajat Matra, Mr Wahyu, Mr Adrian, Mr Agus (Institut Pertanian Bogor University), Mr Gregori Garnadi Hambali (Yatazawa Garden), Dr Abdulrokhman Kartonegoro (BO), Mr Harry Wiriadinata (previously BO); Mr Nanang (Koperasi Pekebun Cengal), Mr Sugema (Kelompok Tani Wanayasa), Mr Sumpena (Kelompok Tani Puspahiang), Mdm Mira Kumala Ningsih (WAN), Mr Ambriansyah, Mr Agus Dien (Kawasan Wisata Pendidikan Lingkungan Hidup, Balikpapan); MALAYSIA: Dr Yong Khien Thye (Rimba Ilmu, Universiti Malaya), Mr Foo Soh Yong (Bandar Seri Coalfields Central Park, Malaysia), Mr Kamarulizwan Kamaruddin (Sultan Idris Shah Forestry Education Centre), Mr Lee and Ms Molly, private home gardens at Sg. Mati), Mr Mohd Noor Azizul bin Akbarruddin, Mr Norhaimanshah bin Ismail, Mr Ahmad Firdaus bin Mokhtar, Mrs Hafidzah binti Nor Ismail, Mr Roslin bin Ismail (Taman Pertanian Universiti, UPM), Ms Sheila Ramasamy (Tunas Harapan Nursery), Mdm Nur Aida, Mr Azwan, Dr Nor Faizal Ghazali (MARDI), Ms Lem Ming Siang (Department of Agriculture), Mr Abdul Halim bin Sulong, Mdm Nor Azmira binti Tukiran (Bukit Goh Agricultural Centre); Mr Musa, Mr Ayau Kanir, (indigenous orchards at Bukit Lagong), Mr Muhamad Hatta, Mr Norazmi, Mr Ahmad Raffizi, Zarul Zurhaidy (FRIM), Dr Joan Pereira, Dr Ruben Nilus, Mdm Suzana Sabri, Mr Postar Miun, Mr Jeisin Jumian, Mr Juhar Aribin (SAN), Mr Jain Linton, Mr Ramani Lajuin (Sabah Agricultural Park), Mdm Mohizah Mohamad, Mdm Julia Sang, Mdm Ling Choi Ying (SAR); SINGAPORE: Dr Ngo Kang Min and Dr Lim Jun Ying (previously Nanyang Technological University, Singapore), Dr David Middleton (previously Singapore Botanic Garden), Mr Elango Velautham, Dr Chong Kwek Yan, Mr Paul Leong, Mr Koh Sin Lan, Mr Lim Wei Hao, Mr Athen Parusamy, Mr Edward Chia, Ms Serena Lee (Singapore Botanic Garden). To them I owed a huge debt for their facilitation in collecting the materials for morphological and genetic analyses.

To my colleagues and friends at IRD, Montpellier, France; IRD Jakarta Office, Indonesia; Faculty of Forestry and Environment, UPM and Forest Research Institute Malaysia, I extend my big thanks for your convivial companionship and professional supports.

Last but not least, I thank my beloved family for their unconditional love and unwavering support towards me throughout my PhD journey.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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Date: 14 March 2024

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF APPENDICES	xviii
LIST OF ABBREVIATIONS	xix
 CHAPTER	
 1 GENERAL INTRODUCTION	 1
1.1 Introduction to cultivated mangosteen and its wild relatives	1
1.1.1 The cultivated mangosteen	1
1.1.2 The wild varieties	5
1.1.3 The geographical distribution patterns and its determining factors	7
1.1.4 Breeding biology	11
1.1.5 Polyploidy	13
1.2 Fruit tree domestication	14
1.2.1 Domestication: a bird's eye view	14
1.2.2 The domestication of fruit trees	16
1.2.3 Domestication of fruit tree in the tropics	17
1.2.4 Domestication of fruit trees in Southeast Asia	19
1.2.5 Exceptional diversity in edible fruit species	19
1.2.6 Paucity of historical and archaeological records	19
1.2.7 Developmental pathways of fruit tree domestication and fruit production	20
1.3 Research problems and plausible solutions	21
1.3.1 The origin of the cultivated mangosteen: a contentious conundrum	21
1.3.2 Deducing the domestication history: suitable approaches for <i>G. mangostana</i>	22
1.3.3 Archaeobotanical study	23
1.3.4 Examination and interpretation of historical records	23
1.3.5 Ethnobotanical survey	24
1.3.6 Morphological analysis	24
1.3.7 Genetic analysis	24
1.4 Objectives	25
1.5 Thesis work plan	26
1.5.1 Sampling campaign plan	26
1.5.2 Morphometric analyses	28

1.5.3	Genetics: phylogenomic and population genetic approaches	28
1.6	Challenges and limitations	28
2	THE ORIGIN OF CULTIVATED MANGOSTEEN (<i>GARCINIA MANGOSTANA</i> L. VAR. <i>MANGOSTANA</i>): CRITICAL ASSESSMENTS AND AN EVOLUTIONARY-ECOLOGICAL PERSPECTIVE	40
2.1	Introduction	41
2.2	The taxonomy and distribution of <i>Garcinia</i> sect. <i>Garcinia</i>	42
2.3	Domestication of <i>G. mangostana</i>	48
2.3.1	What can the historical record and linguistic evidence tell us about the origin of cultivated mangosteen?	49
2.4	What is the wild ancestor of <i>G. mangostana</i> var. <i>mangostana</i> ?	51
2.4.1	Mangosteen is a hybrid of two wild relatives	51
2.4.2	Mangosteen derived solely from <i>G. malaccensis</i>	54
2.4.3	The possible domestication syndromes of mangosteen	55
2.4.4	<i>Do male trees occur in mangosteen?</i>	55
2.4.5	Apomixis	56
2.4.6	Polyploidy	57
2.4.7	Genetic variation	58
2.5	Hypothesis: Domestication at the forest- <i>dusun</i> interface	60
2.6	Synthesis	62
3	MULTIVARIATE MORPHOMETRIC ANALYSIS OF MANGOSTEEN (<i>GARCINIA MANGOSTANA</i> VAR. <i>MANGOSTANA</i>, <i>CLUSIACEAE</i>) AND ITS WILD RELATIVES	74
3.1	Introduction	75
3.2	Materials and methods	78
3.2.1	Specimens and taxa included	78
3.2.2	Characters and character states selection	79
3.2.3	Algorithms in PCoA, AHC and CH index	85
3.3	Results	85
3.3.1	PCoA	90
3.3.2	AHC	91
3.4	Discussion	92
3.4.1	The efficacy of morphological characters in defining the closely related taxa	92
3.4.2	Assessment on characters combinations data subset	94
3.5	Conclusion	95

4	PHYLOGENY AND HAPLOTYPES INFERRED FROM CHLOROPLAST SINGLE NUCLEOTIDE POLYMORPHISMS: A PRELIMINARY LOOK INTO THE DOMESTICATION HISTORY OF MANGOSTEENS	101
4.1	Introduction	102
4.2	Material and method	104
4.2.1	Plant material	104
4.2.2	Genomic library construction and whole genome sequencing	104
4.2.3	Chloroplast genome reconstruction and variant calling	107
4.2.4	Phylogeny analyses	107
4.2.5	Haplotypic analyses	107
4.3	Results	108
4.3.1	The phylogeny of <i>Garcinia</i> focusing on cultivated and wild mangosteens	108
4.3.2	The haplotype network of the cultivated mangosteens and wild relatives	110
4.4	Discussion	112
4.5	Conclusion	115
5	GENERAL DISCUSSION, CONCLUSION AND PERSPECTIVE	122
5.1	General discussion	122
5.1.1	Synthesis of the previous chapters	122
5.1.2	Discordance between morphotypes and haplotypes	122
5.1.3	Advantages and disadvantages of cpDNA SNPs	123
5.1.4	Gene flow through seed dispersals	124
5.1.5	Disadvantaged without SSRs dataset	124
5.1.6	Ecotypes in wild populations?	124
5.1.7	Domestication history	125
5.1.8	Alternative theories of the forest- <i>dusun</i> interface	125
5.2	Conclusion	126
5.3	Perspective	126
5.3.1	Additional nrDNA SNPs needed	126
5.3.2	Long-term field observations and garden experiments	127
5.3.3	Naming of cultivars	127
5.3.4	Conservation of genetic resources in the natural forests	127
6	SYNTHESE EN FRANÇAIS	133
6.1	Introduction	133
6.2	Méthodologie	134
6.2.1	Analyses morphométriques	134
6.2.2	Analyses génétiques	135
6.3	Principaux résultats	135
6.3.1	Examen des textes	135
6.3.2	Analyses morphométriques	135
6.3.3	Analyses génétiques	136

6.4	Discussion générale	136
6.4.1	Synthèse des chapitres précédents	136
6.4.2	Histoire de la domestication	138
6.4.3	Théories alternatives de l'interface forêt- <i>dusun</i>	139
6.5	Conclusion	139
REFERENCES		141
APPENDICES		145
BIODATA OF STUDENT		152
LIST OF PUBLICATIONS		153



LIST OF TABLES

Table	Page
1.1 Main features of <i>G. mangostana</i> varieties	5
1.2 Chromosome counts in <i>G. mangostana</i> var. <i>mangostana</i> and <i>malaccensis</i>	14
1.3 Phasic framework of mutual progress between production systems and cultivated trees in exploitation of tree crops	21
2.1 List of taxa of sect. <i>Garcinia</i> and their synonyms	43
2.2 Comparison of the varieties of <i>Garcinia mangostana</i>	46
2.3 Summary of proposed hypotheses on the origin of cultivated mangosteen, listed chronologically	51
2.4 Chromosome counts of taxa in sect. <i>Garcinia</i> species	52
3.1 Number of specimens analysed with details of taxa and specimen-subsets	79
3.2 Characters and character states examined and analysed	81
3.3 Calinski-Harabasz indices of six data subsets tested with 2–10 partitions	86
3.4 Summary of eigenvalues calculated in PCoA	91
3.5 The effect of additional characters considering the findings from PCoA, AHC and CH analyses	95

LIST OF FIGURES

Figure	Page
1.1 Cultivated mangosteen trees in Taman Pertanian Universiti Ladang Puchong, Universiti Putra Malaysia planted at 12 by 12 m distance	1
1.2 The salient characters of mangosteen	3
1.3 The fruits of mangosteen	4
1.4 Geographical distribution of <i>G. mangostana</i> wild varieties: var. <i>borneensis</i> and var. <i>malaccensis</i> .	6
1.5 Map of Southeast Asia	8
1.6 Climate pattern in Southeast Asia (and Papua New Guinea) according to Köppen climate classification	9
1.7 Sub-biogeographical regions of the Malesia. Malesia is a floristic biogeographical region that comprises Maritime Southeast Asia and Papua New Guinea	10
1.8 Extent and distribution of vegetation types on Sunda Shelf during the Last Glacial Maximum as inferred from distribution modelling by Cannon et al. (2009)	13
1.9 Map depicting Harlan's centres and non-centres of agricultural origins	18
1.10 Forest- <i>Dusun</i> Interface. This evolutionary-ecological theory put forward two possible scenarios regarding the inception of apomixis reproduction and polyploidy in cultivated mangosteen	22
1.11 The distribution of all samples acquired for this study	27
1.12 The sampling coverage of two wild varieties	27

2.1	A. Schematic phylogenetic tree adapted from Figure 5.3 in Nazre (2006) depicting the affinity between the taxa discussed in the text. Names in box denotes taxa with multiple accessions. B. Photographs of live plants and herbarium specimens of selected taxa. <i>G. celebica</i> : (i) male flower (YTL00014, MPU) top view, (ii) side view, (iii) fruits (YTL00016, MPU) in transverse (TS) and longitudinal sections (LS), (iv) persistent stigma top view; <i>G. mangostana</i> var. <i>mangostana</i> : (v) fruits (SAN161001, SAN), exocarps and mesocarps dissected in TS and LS exposing the white pulpy edible endocarp, (vi) persistent stigma top view; <i>G. mangostana</i> var. <i>malaccensis</i> : (vii) male flower (Ridley 6197, SING) side view, (viii) female flower (FRI72021, KEP) top view, (ix) young fruit (FRI72021, KEP) persistent stigma top view; <i>G. mangostana</i> var. <i>borneensis</i> : (x) male flowers (SAN61164, KEP) side view, (xi) fruit (AA1004, WAN) persistent stigma top view; <i>G. penangiana</i> : (xii) male flowers (La Frankie 2120, KEP) side view, (xiii) fruits (FRI27009, KEP) persistent stigma tangential view. Photo credits: (viii) and (ix) by P. Wilkie (Royal Botanic Garden, Edinburgh), all others by the first author	44
2.2	Geographical distribution of taxa of <i>Garcinia</i> sect. <i>Garcinia</i> . Bioregions appear in bold typeface	45
3.1	The geographical distribution of samples examined and analysed	77
3.2	Scatter plots illustrating the results of PCoA analyses of six data subsets labelled A–F	86
3.3	AHC dendrograms of all specimens	87
3.4	AHC dendrograms of vegetative and fruit specimens subset	88
3.5	AHC dendrograms of vegetative and male flower specimens subset	89
3.6	Bar plots illustrating the results of CH indices for six data subsets	90
3.7	The continuum character states of persistent stigma plates surface of <i>G. mangostana</i> varieties	93
4.1a	The geographical distribution of all samples included in the genomic library construction	105
4.1b	The geographical distribution of samples located in Southeast Asia included in the genomic library construction	106
4.2	Cladogram of <i>Garcinia</i> spp. based on chloroplast SNPs	109
4.3	Haplotype network diagram illustrating possible haplotypic relationships between cultivated mangosteen and wild varieties based on chloroplast SNPs	111

LIST OF APPENDICES

Appendix	Page
3. 1 List of specimens included in this study. In specimens without a collection number, we provide barcodes or QR codes	145
4. 1 Samples (n=97) included for genomic library construction and downstream genetic analyses	148
4. 2 The list of individuals comprised in HP1–3	151



LIST OF ABBREVIATIONS

AFLP	Amplified fragment length polymorphism
AHC	Ascendent hierarchical classification
BCE	Before common era
CE	Common era
CH index	Calinski–Harabasz index
cpDNA	Chloroplast DNA
DNA	Deoxyribonucleic acid
DYNADIV	Plant Biodiversity and Adaptation Research Group
FAO	Food and Agriculture Organization
FCM	Flow cytometry
I.F.A.C.	<i>Institut des fruits et agrumes coloniaux</i>
IRD	French National Research Institute for Sustainable Development
ISSR	Inter simple sequence repeats
ITS	Internal transcribed spacer
LGM	Last Glacial Maximum
MATAB	Mixed alkyltrimethylammonium bromide
nrDNA	Nuclear DNA
PCoA	Principal coordinates analysis
Pen. M'sia	Peninsular Malaysia
RAF	Randomly amplified DNA fingerprinting
RAPD	Random amplified polymorphic DNA
SEA	Southeast Asia
SNPs	Single nucleotide polymorphisms
SSRs	Simple sequence repeats
WGS	Whole genome sequencing
WMO	World Meteorological Organization

CHAPTER 1

GENERAL INTRODUCTION

1.1 Introduction to cultivated mangosteen and its wild relatives

1.1.1 The cultivated mangosteen

Mangosteen (*Garcinia mangostana*, Clusiaceae) stands as an emblematic tropical fruit indigenous to Southeast Asia, renowned for its consumption in its fresh state and widespread popularity among the local population. Distinguished by its exquisite taste and aesthetically pleasing appearance, mangosteen has earned the epithet “queen of fruits”. Globally, mangosteen holds significance as a key export within the commodity cluster “Mango, mangosteen and guava” (FAO, 2020). While it has been introduced throughout the humid tropics, Southeast Asia remains the major production region for mangosteen (Osman & Milan, 2006). Presently, cultivation of mangosteen has expanded to encompass the American and African tropics as well (Lim, 2012; Murthy et al., 2018).

Osman & Milan (2006) have presented a comprehensive account on the husbandry of mangosteen. Cultivated mangosteen trees in orchards, featuring well-spaced planting distances, develop a roundish crown with dense foliage (**Figure 1.1**). Exudates from wounded sapwood appear as a mustard yellow and sticky substance (**Figure 1.2A**). The twigs and leaves of the mangosteen exhibit an opposite arrangement (**Figure 1.2B**), while the flowers emerge singly from the apical end of twigs (**Figure 1.2C**).



Figure 1.1 : Cultivated mangosteen trees in Taman Pertanian Universiti Ladang Puchong, Universiti Putra Malaysia planted at 12 by 12 m distance. The trees are 6–8 m tall and the tree trunks measured 30–40 cm diameter at breast height

The mangosteen fruit exhibits an oblate-globular shape with a measured diameter ranging 4–7 cm (**Figure 1.3**). The outermost layer of the mangosteen, commonly referred to as the rind, is thick and softly liginous. The colour of the outer surface of the rind changes through ripening process, transitioning from white to pink, mauve and eventually to purple-black. Encased within the rind are the edible flesh, organised into four to eight unequal segments. Notably, the count of flesh segments aligns consistently with the lobes of persistent stigma plate. The glistening white flesh is both delicate and juicy, offering a mild sweetness with a subtle sourish tinge. Within the fruit, one might encounter one to three seeds in the larger flesh segments, while the smaller segments are typically seedless. It is noteworthy that completely seedless fruits are not a common occurrence.

Mangosteen has garnered substantial attention within the realm of research, mainly owing to its medicinal properties and unusual breeding system. While recent reviews of the previous have explored the medicinal aspects studies regarding the medicinal properties of mangosteen (Ovalle-Magallanes et al., 2017) and the origin of cultivated mangosteen (Yao et al., 2023), a critical gap remains in the understanding of its domestication history. Consequently, the present study aims to address this research gap by delving into the domestication history of cultivated mangosteen, employing population genetic studies that encompass both cultivated and wild varieties. To complement the population genetic approach, morphometric analyses have been incorporated, providing additional supporting information to augment the comprehensive exploration of mangosteen's domestication history.

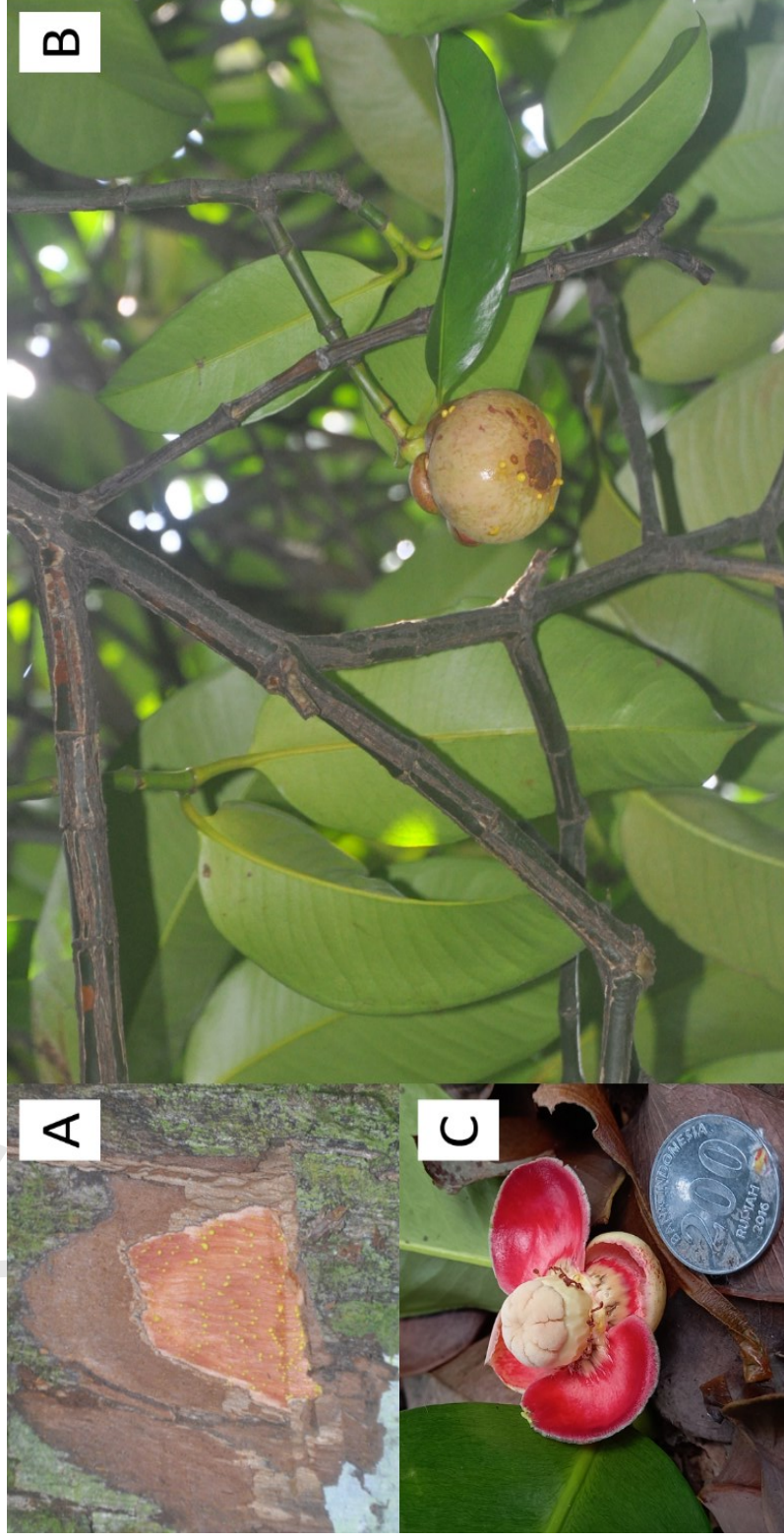


Figure 1.2 : The salient characters of mangosteen. A. sticky, mustard yellow droplets of exudates observed on the wounded sapwood; B. oppositely arranged twigs and leaves; C. an overmature flower without petals exposing the adaxial surface of sepals in red, ovary and stigma are white and numerous staminodes encircle the ovary base

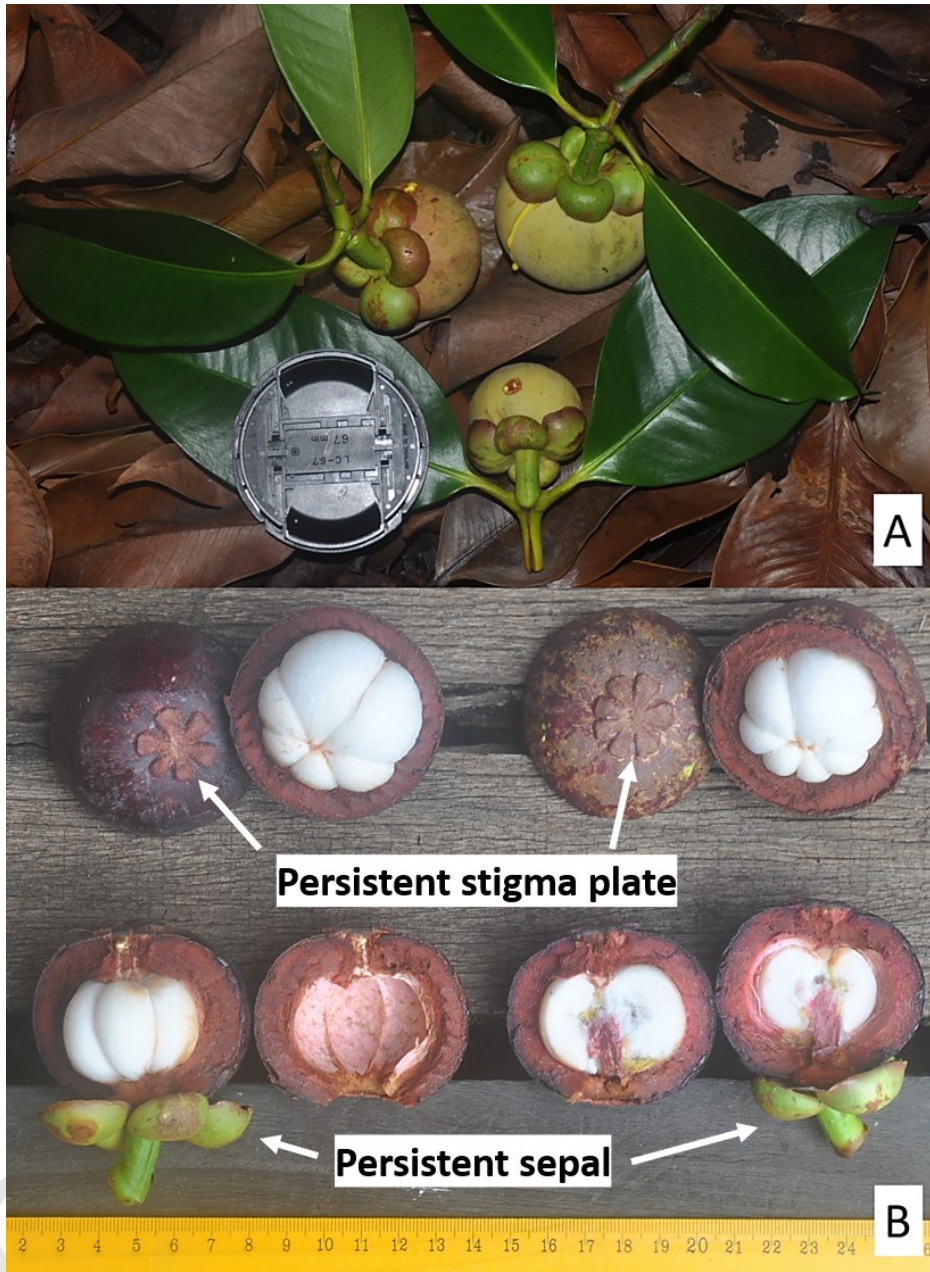


Figure 1.3 : The fruits of mangosteen. A. fruits issued from the apical ends of the twigs, young fruits are pale yellowish green; B. mature fruits with the rinds cut longitudinally (two fruits above) and vertically (two fruits below), pried apart to expose the flesh segments. Note that the number of flesh segments and lobes of persistent stigma plate are always count the same

1.1.2 The wild varieties

Within taxonomic classification, mangosteen is the type species of *Garcinia* L. sect. *Garcinia*. This taxonomic section has undergone a recent revision, resulting in its circumscription to encompass a total of 13 species (Nazre et al., 2018). Notably, *G. mangostana*, the focus of this study, exhibits two taxonomic varieties within its natural habitat, namely (i) *G. mangostana* var. *borneensis* and (ii) var. *malaccensis*. In a phylogenetic analysis conducted by Nazre (2014), *G. mangostana* var. *mangostana* and var. *malaccensis* collectively form a paraphyletic clade, with var. *borneensis* emerging as the sister taxon to this paraphyletic clade. Key characteristics of the *G. mangostana* varieties are succinctly summarised in **Table 1.1**.

Table 1.1 : Main features of *G. mangostana* varieties

Taxon	Presence of pistilodes in male flowers	Fruit shape	Fruit size (across the widest dimension)	Fruit apex; persistent stigma plate shape, and surface texture	Cultivated or wild	Natural geographical distribution
<i>mangostana</i>	present*, c. 5 mm long	ovoid or globose	4–7 cm	rounded; circle, and smooth	cultivated; feral populations might exist	Malaysia and Indonesia
<i>malaccensis</i>	absent, or if present c. 2 mm	globose, ellipsoid or ovoid	3–4 cm	acute to attenuate; circle, and rugose	cultivated and wild; cultivated populations are called <i>mesta</i> in Peninsular Malaysia	Sumatra; Malay Peninsula; Borneo: Southwest Sarawak and Brunei
<i>borneensis</i>	absent, or if present c. 2 mm	globose	3–4 cm	rounded; circle or oval, and smooth to slightly rugose	wild	Borneo: East coasts of Sabah and Kalimantan

*Hambali and Natawijaya (2016) mentioned that the male flowers of var. *mangostana* described by Idris & Rukayah (1987) fell within the morphological plasticity of male flowers they observed in Sumatran *G. mangostana* var. *malaccensis* populations. (Adapted from Nazre et al. (2018))

The distribution of *Garcinia mangostana* var. *borneensis* encompasses the region of east Borneo, whereas var. *malaccensis* is found in Sumatra, Peninsular Malaysia, and west Borneo (**Figure 1.4**). Notably, these taxonomic varieties serve as the primary candidates for the wild progenitors of the cultivated mangosteens.

The vernacular names “*manggis*” and “*manggis kampung*” are commonly attributed to *G. mangostana* var. *mangostana* in Peninsular Malaysia. The wild var. *borneensis* occurs within the ecological research plots in natural forests in Kabili-Sepilok Forest Reserve, Sabah, Malaysia is referred to as “*manggis*”. Meanwhile, the wild *G. mangostana* var. *malaccensis* is denoted as “*manggis hutan*” and known as “*mesta*” for the cultivated populations in Peninsular Malaysia.

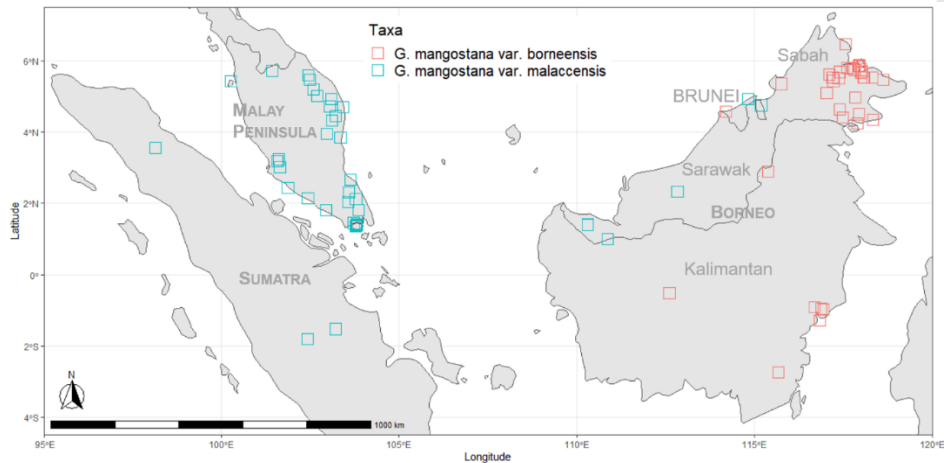


Figure 1.4 : Geographical distribution of *G. mangostana* wild varieties: var. *borneensis* and var. *malaccensis*

In West Java, distinct local names are employed to designate the cultivated *G. mangostana* var. *mangostana*, specifically identifies with their respective cultivation areas in West Java. These include (i) “*manggis Cengal*”, (ii) “*manggis Puspahiang*” and (iii) “*manggis Wanayasa*”.

In Sumatra, various local names are assigned to the cultivated mangosteen, including (i) “*manggis Ratu Tembilahan*” from Riau; (ii) “*manggis puting susu*”, (iii) “*manggis curup*”, (iv) “*manggis burik*”, (v) “*manggis kutu*”, (vi) “*manggis hijau*”, (vii) “*manggis kuning*”, (viii) “*manggis jumbo*” and (ix) “*manggis isi banyak / kipas*” from Bengkulu and Bangka-Belitung (Mansyah et al., 2007). The absence of references to herbarium specimens or deposited sequences in publicly accessible database for the aforementioned names impedes further research on these cultivated mangosteens due to the lack of referencing materials.

In North Kalimantan (Borneo), “*manggis Malinau*” likely represent a taxon differentiated from *G. mangostana*, as indicated by both morphological and phylogenetic findings (Santoso et al., 2019). While I have enumerated the aforementioned names from West Java, Sumatra and North Kalimantan, it is acknowledged that there may exist additional vernacular names in these regions that remain localised and have not yet been scientifically documented. Moreover, some local names might potentially denote

the same cultivated group. Notably, a comprehensive characterization of the morphological and genetic aspects of these cultivated groups is yet to be established.

1.1.3 The geographical distribution patterns and its determining factors

Cultivated mangosteen is extensively cultivated in Southeast Asia (**Figure 1.5**) including Indonesia, Malaysia, Thailand and Vietnam. In contrast, wild mangosteen varieties, namely var. *borneensis* and var. *malaccensis* are indigenous to Brunei, Indonesia, Malaysia and Singapore (**Figure 1.4**). The determination of the geographical distribution of these wild mangosteens varieties is imperative for the planning of sampling campaign.

To comprehend the distribution patterns of *G. mangostana* varieties, a thorough examination of contributing factors becomes essential. These factors encompass climatic zones, biogeography, historic climate trends and the prevalence and extent of rainforests refugia. Investigating these elements is fundamental for gaining insights into the intricacies of the distribution patterns of *G. mangostana* varieties.

Southeast Asia (SEA hereafter) is divided into two major subregions: Mainland SEA and Maritime SEA. Mainland SEA is positioned in between India to the west and China to the northeast. Historically, Cambodia, Laos and Vietnam were collectively referred to as Indochina, a term now commonly used in the context of biogeography to encompass these three countries.

Maritime SEA is widely recognised as the Malay Archipelago. This region includes Peninsular Malaysia, and among its larger islands are Sumatra, Java and Sulawesi in Indonesia; Luzon, Mindanao, Samar and Palawan in the Philippines. Borneo Island, located at the centre of Maritime SEA, has international borders shared among three countries: Brunei, Indonesia (Kalimantan), and Malaysia (Sabah and Sarawak).

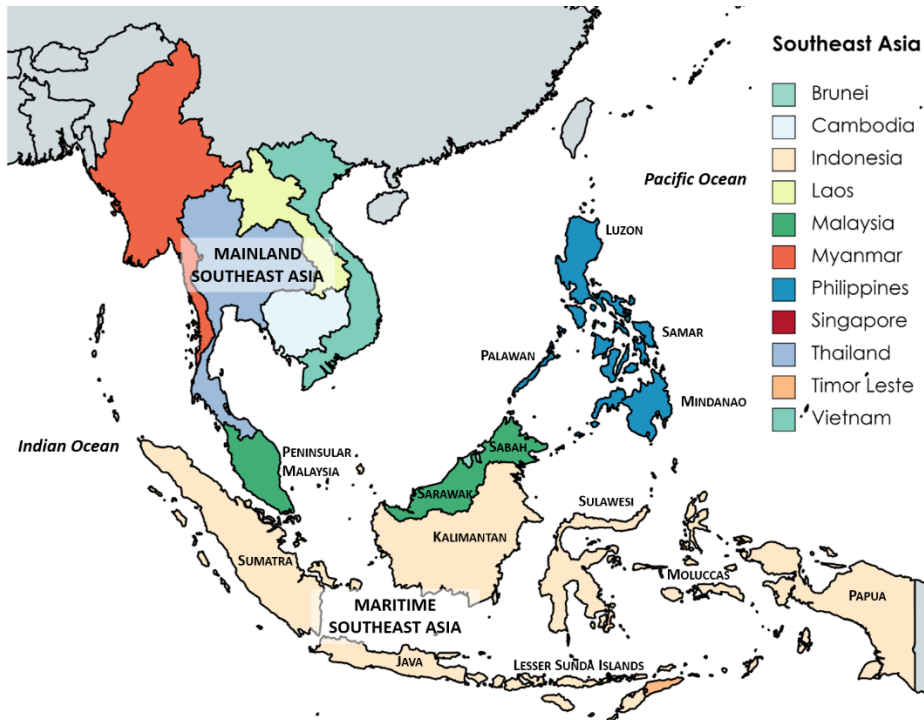


Figure 1.5 : Map of Southeast Asia. Southeast Asia comprises 11 countries that commonly categorised into Mainland Southeast Asia that comprises five countries: Cambodia, Laos, Myanmar, Thailand, Vietnam; and Maritime Southeast Asia that comprises six countries: Brunei, Indonesia, Malaysia, Philippines, Singapore, Timor Leste. Map created with mapchart.net

In accordance with the Köppen climate classification (Beck et al., 2018), noticeable distinctions in climate exist between Mainland SEA and Maritime SEA (**Figure 1.6**). Mainland SEA predominantly experiences a monsoon climate. Conversely, an equatorial climate prevails in Malaya, Sumatra and Borneo, coinciding with the natural distribution of *G. mangostana* wild varieties (**Figure 1.4**). In Java, equatorial climate is limited to a small area in the western part and the central highlands of the island, while monsoon and tropical savannah climates dominate other regions.

Southeast Asia map of Köppen climate classification

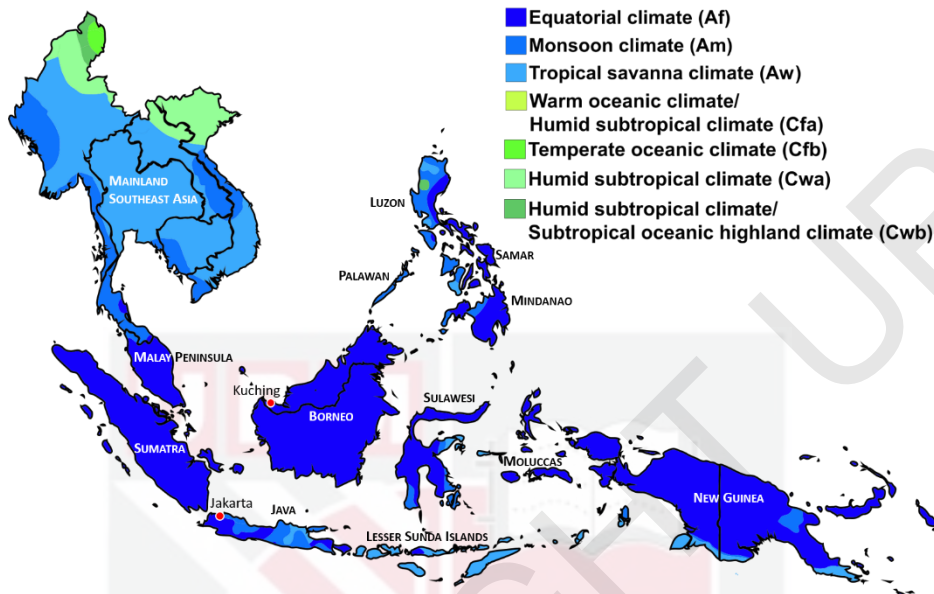


Figure 1.6 : Climate pattern in Southeast Asia (and Papua New Guinea) according to Köppen climate classification. Map downloaded from Wikipedia (Ali Zifan, 2016)

A significant distinction between equatorial climate and monsoon climate lies in the presence of a well-defined annual dry season in the area characterised by a monsoon climate, whereas equatorial climates lack a pronounced dry season. A dry season in Tropical Asia is identified by an average monthly rainfall below 100 mm for three consecutive months or more.

Illustrating this contrast, consider Jakarta, the capital city of Indonesia situated in northwest Java, which falls between equatorial and monsoon climate zones. Using Jakarta as an example, the annual precipitation is approximately 1700 mm, with average monthly precipitations falling below 50 mm for four consecutive months from July to October (WMO, 2023).

In contrast, Kuching, the capital of Sarawak in Northwest Borneo, located in the equatorial climate zone, exhibit a different pattern. The average annual rainfall in Kuching is slightly above 4000 mm, with July recording the lowest precipitation at about 185 mm. However, in the remaining 11 months, average monthly rainfalls range between 220–685 mm, exemplifying the consistent and comparatively higher precipitation levels in equatorial climates.

The wild mangosteen varieties are restricted to evergreen rainforests in Sumatra, Peninsular Malaysia, and Borneo (Hambali & Natawijaya, 2016; Nazre et al., 2018). Field observations on the wild populations of var. *borneensis* revealed that these trees are typically situated along ephemeral streams in low-lying flat land (Yao et al., 2023).

It is noteworthy that cultivated mangosteen thrives in environments with minimal water stress, particularly during the seedling and sapling stages. In instances of dry seasons, irrigation may be necessary for survival (Osman & Milan, 2006). Therefore, one can infer that the mangosteen varieties in their natural habitats are predominantly found in regions devoid of a monsoon climate.

Malesia constitutes a botanical region encompassing Maritime SEA and New Guinea (Figure 1.7). The recognition of this region dates back to Heinrich Zollinger in 1857, with the concept subsequently refined and adopted by Cornelius van Steenis for “Flora Malesiana” project in mid twentieth century (Johns, 1995). Welzen et al. (2011) proposed the division of Malesia into three subregions: (i) the ever-wet Sundaland (Sunda), that comprises Sumatra, Malaya and Borneo; (ii) the Wallacea, encompassing Java, the Philippines, Sulawesi, Lesser Sunda Islands and Moluccas; and (iii) the New Guinea (Sahul). Welzen et al. (2011) also noted that Java Island straddles across Sunda and Wallacea floristic regions, reflecting the contrast between the flora in the ever-wet western part and areas with pronounced dry monsoon in the central and eastern parts. This regional division provides distinction of the subdivisions of flora within Malesia.

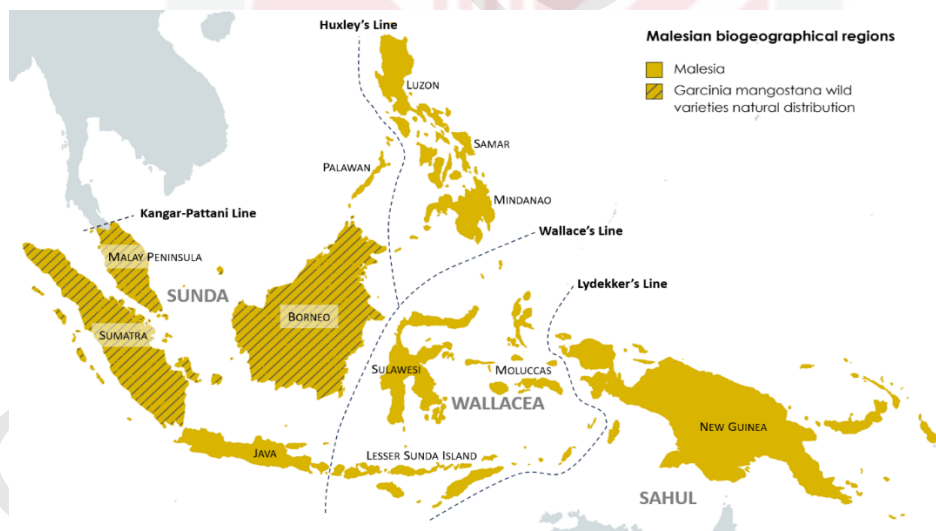


Figure 1.7 : Sub-biogeographical regions of the Malesia. Malesia is a floristic biogeographical region that comprises Maritime Southeast Asia and Papua New Guinea. Kangar-Pattani, Huxley's, Wallace's and Lydekker's Lines are the hypothetical biogeography boundaries proposed by naturalists based on their observations on faunistic and floristic distribution patterns. *Garcinia mangostana* varieties are naturally distributed in Malaya, Sumatra and Borneo. Base map created with mapchart.net

During past glacial maximums occurring approximately 17,000, 150,000 and 250,000 years before the present, when the sea level was about 120 m lower than the present sea level, two land masses appeared above sea level in the Malesia region, namely Sunda Shelf and Sahul Shelf (Voris, 2000), respectively labelled Sunda and Sahul in **Figure 1.7**. The islands in Wallacea remained separated by sea surfaces during these glacial periods.

Periodic unions of Malay Peninsula, Sumatra, Borneo, Palawan, Java and Bali occurred on the Sunda Shelf during past glacial maximums, facilitating the dispersal of plants, animals, and human migrations across the landmass. Flora and fauna occurring in the Sunda region are known as Sundaic elements, and the varieties of *Garcinia mangostana* are considered Sundaic elements.

However, the extent and distribution of the evergreen lowland forests were different during the last glacial maximum (LGM) (Cannon et al., 2009) (**Figure 1.8**). The lowland evergreen forest covered a substantial area currently submerged under the South China Sea, Java Sea and Strait of Malacca. When overlaying the extent and distribution of the evergreen lowland forests during the LGM on the current landmass, it can be inferred that the present Sundaic rainforests, especially in the lowland area of Sumatra, the Malay Peninsula, and Borneo, represent refugial dispersions of the past Sunda Shelf evergreen forests that migrated from adjacent areas now submerged.

Significantly, it is evident that the area where Java Island located was completely devoid of, and considerably distant from evergreen lowland rainforest during the LGM. This suggest that the predecessors of *G. mangostana* would not have survived in Java and adjacent areas during this period. Considering common factors influencing the distribution pattern of tree species such as current climate and historic vegetation distribution, one can infer that the natural distribution of wild *G. mangostana* varieties is confined to the evergreen Sundaic forests in Sumatra, the Malay Peninsula and Borneo, which are situated within the equatorial climate zone.

1.1.4 Breeding biology

Cultivated *G. mangostana* var. *mangostana* is a polyploid species primarily propagated asexually through seeds generated via adventitious embryogenesis. As a result, cultivated mangosteens exhibit uniform phenotypic appearance. The genotypes of these mangosteen, inclusive of heritable somatic mutations, remained stable across generations within cultivated lineages. The subsequent paragraphs provide a brief discussion of the key aspects of the distinctive breeding biology exhibited in cultivated mangosteen.

Dioecy is commonly observed in *Garcinia* species within the rainforests of Southeast Asia (Ha et al., 1988; Thomas, 1997). The presence of male trees in *G. mangostana* var. *mangostana* has been a subject of contention since the 1830s (Horn, 1940; King, 1890; Ridley, 1922; Roxburgh, 1832). Idris & Rukayah (1987) reported the identification of a male mangosteen tree in Peninsular Malaysia. They described the functional male

flowers characterised by a stamens mass surrounding a pistillode. Although they detailed the morphology of the pollen grains and provided scanning electron microscope images, no information on their viability was furnished.

Staminodes are found in the functionally female flowers of mangosteen, where they produce no viable pollen (Lim, 1984). While labile sex expression is recognised in angiosperms (Käfer et al., 2022), the current understanding of *Garcinia* species is limited, hampering a comprehensive discussion. Scarce data are available regarding the sex ratio in wild populations of *Garcinia* species, with a notable lack of information on the sex ratio in *G. mangostana* var. *malaccensis* and var. *borneensis*.

Somatic embryogenesis in cultivated mangosteen was demonstrated over a century ago (A. L. Lim, 1984; Sprecher, 1919a, 1919b). The formation of asexual seeds in cultivated mangosteen results from adventitious embryo originating from a somatic cell of the ovary (Sprecher, 1919a, 1919b), and such asexual seed formation may be a common phenomenon in *Garcinia* species (Sweeney, 2008). While somatic embryogenesis is recognised as an efficient propagation pathway and is exploited in horticultural practices (Ramírez-Mosqueda, 2022), information regarding its natural occurrence is limited.

Hambali & Natawijaya (2016) successfully pollinated *G. mangostana* var. *mangostana* female flowers with var. *malaccensis* pollens in garden experiments. Their efforts resulted in obtaintion of viable seeds, which developed into triploid seedlings (Hambali & Natawijaya, 2016). These results show that mangosteen is a facultative apomictic species that is capable of sexual reproduction when a suitable source of viable pollen is available. From the same experiments, one knows that at least some var. *malaccensis* produce viable pollen.

In the case of *Mesta*, a cultivated variety of var. *malaccensis*, field observations and discussion with growers in Peninsular Malaysia indicated the likelihood of apomictic fruit production, given the apparent absence of male individuals in cultivation.

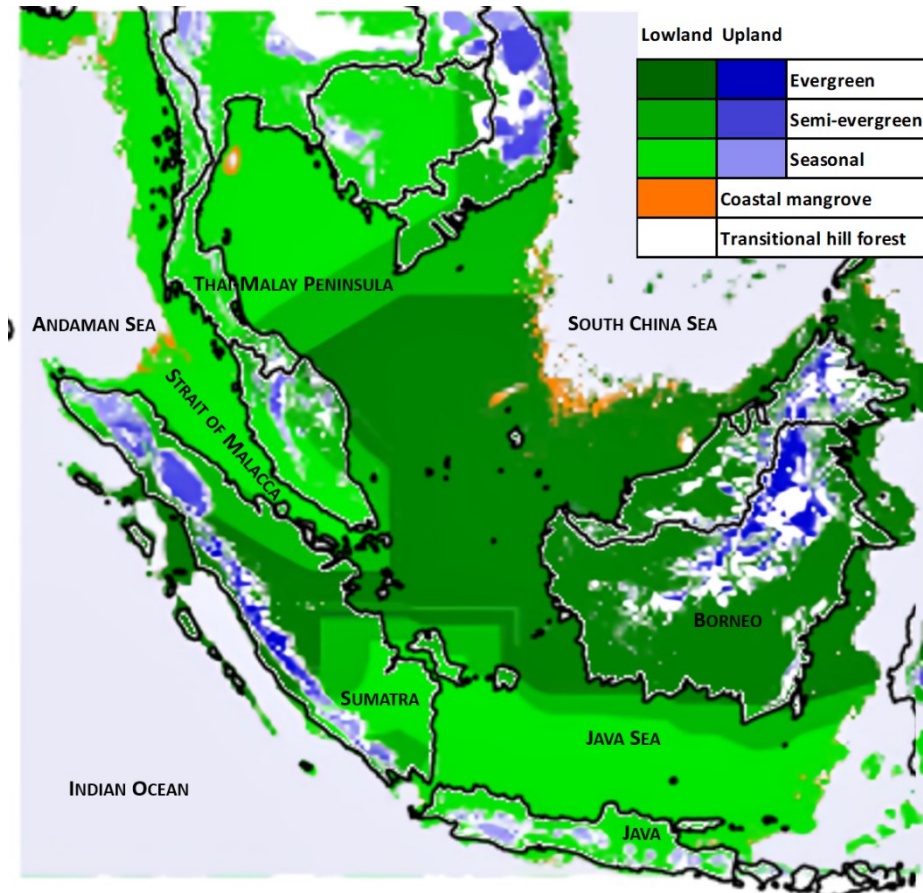


Figure 1.8 : Extent and distribution of vegetation types on Sunda Shelf during the Last Glacial Maximum as inferred from distribution modelling by Cannon et al. (2009). Adapted from Figure 1A, Cannon et al. (2009) depicting the maximum extent of evergreen vegetation model. Boundaries of the current land areas and labels of the regions are added

1.1.5 Polyploidy

The chromosomes of *Garcinia* present a challenge for enumeration due to their small size (Midin et al., 2018; Richards, 1990). Polyploidy has been identified in *G. mangostana* (Table 1.2): investigation reveal that var. *mangostana* is tetraploid (Matra et al., 2016; Midin, 2018), while diploids, triploids (Hambali & Natawijaya, 2016) and tetraploids (Midin, 2018) are observed in var. *malaccensis*. The chromosome number for var. *borneensis* remained unreported. Midin et al. (2018) demonstrated that dysploidy/aneuploidy in var. *mangostana*, revealing highly variable chromosome numbers ranging from 74 to 110 in an analysis of 20 metaphase spreads.

The maintenance of polyploidy in *G. mangostana* is facilitated by apomictic reproduction, mitigating potential difficulties with chromosome pairing during meiosis. Initial study proposed *G. mangostana* var. *mangostana* as allopolyploid (Richards, 1990), while subsequent studies categorise it as autopolyploidy (Hambali & Natawijaya, 2016; Nazre, 2014).

Table 1.2 : Chromosome counts in *G. mangostana* var. *mangostana* and *malaccensis*

var.	2n	Reference	Region
<i>mangostana</i>	c. 76	Krishnaswamy & Raman, 1949	Kerala, India
	96	Chennaveeraiah & Razdan, 1975; Tixier, 1960	India(?); Laos
	c. 88–90	Ha, 1978	Pen. Malaysia
	96	Hambali & Natawijaya, 2016	Java
	74–110	Midin, 2018	Pen. Malaysia
<i>malaccensis</i>	48, 72	Hambali & Natawijaya, 2016	Sumatra

1.2 Fruit tree domestication

1.2.1 Domestication: a bird's eye view

The domestication of plants and animals stands as arguably the most significant revolution of human history, propelling the emergence of human civilizations and, ultimately, shaping the contemporary world in which we live today (Diamond, 2002). Domesticated plants and animals have bestowed upon humankind a more stable and reliable food supply. Additionally, the domestication of animals also provided humans with transportation, labour, and companionship. Domestication practices exemplify how humans coexist and engage with the natural world, particularly in terms of modifying the natural environment and managing biological diversity. Over the past 12,000 years, humans have brought thousands of plant species into domestication (Meyer & Purugganan, 2013).

In the realm of plant domestication, early written texts addressing useful plants and their cultivation trace back to classical antiquity, exemplified prominently by “*Historia plantarum*” (Theophrastus, 1916), written in between c. 350–287 BCE. Another seminal work, “*The origin of cultivated plants*” (de Candolle, 2011 [originally published in 1886]) culminated from the first systematic investigation into plant domestication at a global scale. “*Origin and Geography of Cultivated Plants*” (Vavilov & Dorofeev, 1992) signified the importance of genetic diversity for crop improvement, influencing the subsequent breeding programmes and the development of high-yielding, disease-resistant crop varieties. These publications laid the groundwork for understanding the origins, diversity, and conservation of cultivated plants, as well as their significance for global agriculture.

In recent decades, the field of archaeobotany and genetic studies has witnessed significant advancements, contributing profound insights into the origin and dissemination of agriculture and crops. Concurrently, the acceleration in the loss of

traditional landraces and wild relatives, has resulted in a drastic decline in agrobiodiversity. Against this backdrop, Harlan (1992) undertook the task of synthesizing these developments in “Crop and man”, thereby presenting a comprehensive perspective on the relationship between humans and crops. The work integrates findings of anthropology, archaeology, botany, and agronomy, employing an interdisciplinary approach to illuminate various facets of this dynamic interaction.

Harlan's work not only serves as a bridge between diverse academic disciplines but also furnishes vivid examples illustrating how various socio-cultural human societies have actively influenced crop diversity and the evolving landscapes of agriculture over time. Furthermore, the author underscores the significance of crop diversity as an indispensable resource for ensuring food security and promoting agricultural sustainability. In doing so, Harlan's synthesis emerges as a crucial resource for scholars and researchers seeking a nuanced understanding of the intricate interplay between humanity and the cultivation of crops.

From the perspective of contemporary scientific inquiry, domestication is defined as an evolutionary process characterised by directional selection of traits that are favourable to humans (Kantar et al., 2017). In elucidating this phenomenon, the “modern synthesis” offers a robust conceptual framework, particularly in examination to study the population genetics concerning domesticated plants and animals (Zeder, 2017).

The modern synthesis represents an integrative approach that combined insights from various disciplines, including anthropology, archaeology, evolutionary biology, genetics and phenotypic changes. This comprehensive methodology aims to elucidate the process of domestication by scrutinising the transformations undergone by plants and animals during their transition from wild to domesticated forms. Utilizing archaeological studies, researchers uncover samples that facilitate the interpretation of phenotypic changes in the domesticates. Moreover, these archaeological samples serve as a source of DNA materials for conducting ancient DNA analyses (Zeder et al., 2006).

In instances of exceptional circumstances, researchers may acquire resurrection DNA by germinating ancient seeds found at archaeological sites, thereby enabling in-depth genetic studies (Gros-Balthazard et al., 2021). Within the realm of genetics, ongoing advancements in DNA sequencing and genetic analysis technologies have empowered researchers to compare the genomes of wild and domesticated species. This comparative analysis is instrumental in deducing the domestication history, as demonstrated in the studies examining apricot (Groppi et al., 2021), olive (Besnard et al., 2013) and pears (Yue et al., 2018). These scientific endeavors showcase the pivotal role played by modern genetic tools in unraveling the complex narrative of domestication across various plant and animal species.

In instances where ancient DNA from the progenitors is accessible, researchers can conduct genomic comparisons between these ancestral specimens and contemporary cultigens (Cappellini et al., 2010; Cohen et al., 2023). This comparative analysis, which involves assessing the genetic material of both wild and ancient progenitors alongside that of cultigens, serves as a tool for identifying specific genes or genomic regions that

have undergone selective changes during the process of domestication. Such investigations provide crucial insights into the molecular basis of domestication.

Although domesticated plants benefit human in various ways, the domestication processes, particularly those involving selection and adaptations to man-made environments, have led to a reduction in genetic diversity within cultigens (Smýkal et al., 2018). The intentional breeding and cultivation practices have often favored specific traits, resulting in a narrowing of the genetic variation present in domesticated crops. This reduction in diversity can have implications for the resilience and adaptability of cultivated species in the face of environmental challenges.

Conversely, wild relatives of domesticated plants serve as a genetic reservoir for cultigens, harboring a reservoir of potential genes that can contribute to crop improvements and enhance adaptability (Majeed et al., 2021). The genetic diversity found in wild relatives represents a valuable resource for breeding programs, offering a pool of traits that may resist pests, diseases, and environmental stresses. Therefore, understanding and preserving the genetic diversity present in wild relatives is essential for ensuring the long-term sustainability and resilience of domesticated crops.

The modern synthesis serves as a solid foundation for exploring the origins and genetic constituents of domesticates species, especially in the case of understudied tropical fruit trees such as the cultivated mangosteen, which is the focal of the present study. In the subsequent sections, I delve into topics pertinent to fruit tree domestication, with a specific emphasis on tropical regions. This investigation aims to contribute to our understanding of the complex processes involved in the domestication of tropical fruit trees.

1.2.2 The domestication of fruit trees

The early domestication of fruit trees is well exemplified by the cultivation of fig (*Ficus carica* L.) and olive (*Olea europaea* L.) in the Near East (Zohary & Spiegel-Roy, 1975), as documented in early records dating back to 3000–2000 and 2000–1000 BCE, respectively. Beyond the near east, citruses such as *Citrus aurantium* L. and *C. maxima* (Burm.) Merr. were domesticated in China before 2200 BCE (Spiegel-Roy, 1986) and the cultivation of mango (*Mangifera indica* L.) is noted to have started around 2000 BCE in the Indian subcontinent (Mukherjee, 1972).

Domestication syndrome is generally defined as a suite of traits that characterise a divergence from the wild progenitor(s) in domesticated plants. This suite comprises traits desirable to humans, though not necessarily beneficial to the plant (Meyer et al., 2012). A review on the domestication syndrome in historically cultivated fruit trees highlighted specific trait changes compared to their wild progenitors. Notable major changes include self-fertility in peach and sour cherry, the elimination of bitterness in almond nuts, reduced resin content in mango fruits, and the development of thornlessness in citruses (Spiegel-Roy, 1986). However, the review emphasized these trait changes in

domesticated fruit trees, in comparison to their progenitors, may not always be pronounced.

Unlike domesticated annuals, especially grain crops, tree crops often exhibit fewer domestication syndromes. An analysis of major and minor crops over domestication time, encompassing species (Kantar et al., 2017), included only two fruit tree crops: olive and Brazil nuts (*Bertholletia excelsa* Bonpl.). Olive and sunflower were introduced into domestication around the same period. However, while six domestication syndromes are recognised in sunflower, only two are identified in the case of olive. The longer generation time of tree crops emerges as an obvious factor contributing to this disparity.

In the context of genetic considerations, domesticated perennial fruit crops exhibit weaker genetic bottleneck, retaining approximately 95% of the ancestral genetic variations. This contrast with annual fruit crops, which retain only around 60% of the original genetic diversity (Miller & Gross, 2011). The genetic profile of perennial fruit crops is also enriched through sexual reproduction cycles and the fixation of somatic mutations in clonal cultivars (Gaut, Díez & Morrell, 2015).

1.2.3 Domestication of fruit tree in the tropics

Harlan (1971) recognised four regions of agricultural origin across the tropical belt, as illustrated in **Figure 1.9**. These include three ‘non-centres’: (i) African (A2), (ii) Southeast Asian and South Pacific (B2), and (iii) South American (C2). Additionally, there is the Mesoamerican ‘centre’ (C1). Notably, among the widely cultivated species of fruit and nut trees, such as *Artocarpus* spp. (breadfruits), *Citrus* spp. (citruses), *Mangifera* spp. (mangoes), *Nephelium* spp. (rambutans) and *Syzygium* spp. (wax apple), their origins can be traced back to Southeast Asia and South Pacific.

Emblematic fruit tree species of tropical America origin include *Bertholletia excelsa*, *Theobroma cacao* L. (cacao), *Pouteria caimito* Radlk. (abiu), *Inga edulis* Mart. (ice-cream bean) and *Pourouma cecropiifolia* Mart. (Amazon tree-grape). Their traces of domestication are evidenced by being hyper-dominant in Amazonian forests, particularly in the vicinities of archaeological sites (Levis et al., 2017). These species also exhibit a broader distribution than other dominant species, suggesting human mediated dispersal.

In addition to the aforementioned iconic fruit tree species, numerous other species are cultivated in the tropics for fruits and nuts (Clement, 1999), with their cultivation status spanning a continuum from incipient change to fully domesticated. The survival of these species is entirely dependent on the intervention of farmers (Clement et al., 2015). Due to their reliance on human tending, serious genetic diversity loss occurred in these fruit tree species during the drastic human population decline in the Amazonian forests around 1500’s (Clement, 1999), attributed to Portuguese and Spanish colonisation. The population decline was so severe that some settlements were completely wiped out, leaving the domesticated trees unattended and eventually decimated. Recent forest degradation (Escobar, 2019) has further exacerbated the loss.

Among the tree crops originating from tropical Africa, *Coffea arabica* L. and *C. canephora* Pierre ex A.Froehner are perhaps the most extensively studied global commodities concerning their origin and genetic diversity (Bawin et al., 2021; Davis et al., 2023; Ngugi & Aluka, 2019). Nevertheless, the domestication of other fruit tree species in Tropical Africa has remained understudied despite their socioeconomic significance (Rimlinger et al., 2019). *Dacryodes edulis* (G.Don.) H.J.Lam (*safoutier*, African plum), an iconic species with an oily endocarp eaten as a vegetable, is relatively well-studied. Human selection has revealed a domestication syndrome, including a higher percentage of male and male-hermaphrodite sexual types in the wild compartment and a lower fruit and seed mass in wild individuals compared to cultivated ones (Mboujda et al., 2022). Highly variable and transferrable microsatellites have been developed to investigate the population structure of the species (Rimlinger et al., 2020).

Cola nuts (*Cola acuminata* (P.Beauv.) Schott & Endl. and *C. nitida* (Vent.) Schott & Endl.), and seed of *Garcinia kola* Heckel are used as stimulant. Research into their genetic diversity and domestication is still in its infancy (Yogom et al., 2023).

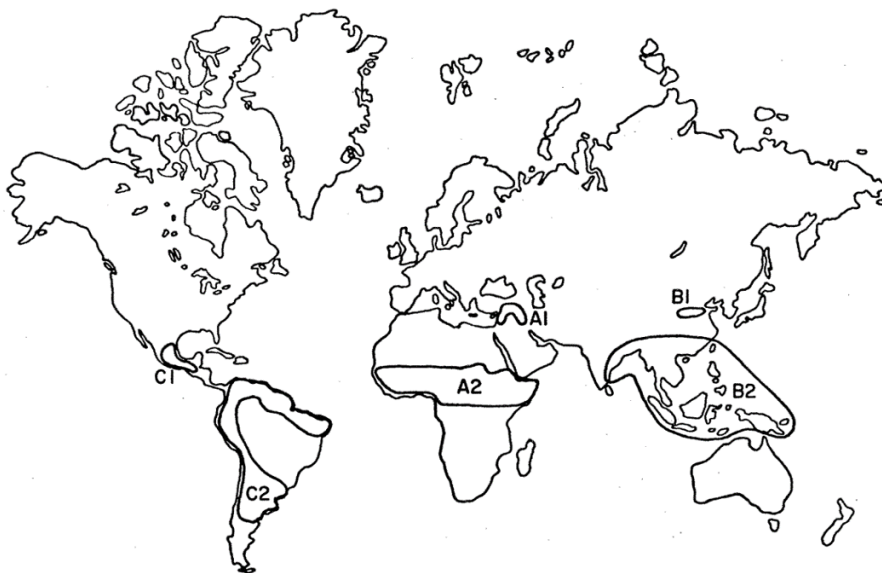


Figure 1.9 : Map depicting Harlan's centres and non-centres of agricultural origins (adapted from Figure 6, Harlan, 1971). A1, Near East centre; A2, African non-centre; BI, North Chinese centre; B2, Southeast Asian and South Pacific non-centre; CI, Mesoamerican centre; C2, South American non-centre

In contrast to the major fruit tree species in Eurasia such as apple and pears, which dispersed over vast land areas, sea routes play a crucial role in the spread of tropical fruits. Breadfruit (*Artocarpus altilis* (Parkinson) Fosberg) is recognised to have been disseminated through human migrations between archipelagos in the Pacific Ocean (Zerega et al., 2004). Among the three tropical regions, transregional exchanges of many

of these fruit tree species occurred during the early part of the Age of Exploration, particularly through the efforts of the Portuguese and Spanish (Blench, 2007).

In contemporary times, approximately 2700 species of tropical fruits are available in marketplaces. These fruits not only serve as an important source of nutrition for the local populations but also contribute to the incomes for farmers engaged in production for export markets (FAO, 2023). Noteworthy is the fact that nearly 99 % of tropical fruit production originates in developing countries, playing a vital role in sustaining local subsistence economies and livelihoods.

1.2.4 Domestication of fruit trees in Southeast Asia

Cultivated mangosteen and its wild relatives originated in Southeast Asia. Hence, the domestication of fruit trees in Southeast Asia is explored in a more in-depth manner. I have considered topics across broader aspects to offer a comprehensive perspective for the current study.

1.2.5 Exceptional diversity in edible fruit species

The lowland forests in Southeast Asia host an extraordinary diversity of edible fruit tree species (Harlan, 1992). An exploration into the species composition of a 50-ha permanent tree demography plot at Pasoh, Negeri Sembilan, Peninsular Malaysia provided insight into the diversity of edible wild fruits (Saw et al., 1991). The study revealed that several genera are especially species-rich such as *Artocarpus* (Moraceae) with 10 species; *Garcinia* (Clusiaceae) with 13 species, *Mangifera* (Anacardiaceae) with 12 species, and *Nephelium* (Sapindaceae) with 5 species. These invaluable natural resources gave rise to many of the fruits that have already been brought into cultivation and continue to serve as a potential reservoir of agrobiodiversity.

A comprehensive compilation of edible fruits and nuts occurring in Southeast Asia is presented in the “Edible Fruits and Nuts” in the “Plant Resources of South-East Asia” series (Verheij & Coronel, 1991). This compendium features 120 major species with detailed monographs and illustrations in line drawings, along with summary information on an additional 275 minor species.

1.2.6 Paucity of historical and archaeological records

The historical textual record on plant domestication in Southeast Asia is sparse. Contacts between the region and Indian and Arabian traders through the Indian Ocean Trade routes date back to ancient times. However, it was not until the early 15th century that the first written records of natural products with explicit details of the locations appeared (Ma, 2019 [1416]). Blench (2007) employed a linguistic approach, analysing loanwords to infer the routes of the spread of fruit cultivation in mainland Southeast Asia. Linguistic analyses suggest common exchanges of fruits between Southeast Asia, India and China.

However, many characteristic Southeast Asian fruits have only recently spread beyond the region.

Similarly, archaeological findings recovering food residuals, especially fruits, are limited (Paz, 2005). Traces of starchy vegetables and spices, such as sago, purple yam and nutmeg, were evident on potsherds dated around 1500–300 BCE, excavated from an archaeological site in Pulau Ai, South Moluccas, Indonesia (Lape et al., 2018). Charred rind fragments of citrus fruits were recovered in flotation samples dated around 400–1 BCE at Khao Sam Kaeo, Peninsular Thailand (Fuller et al., 2018). Among local fruits, durian (*Durio zibethinus* L.) and jackfruit (*Artocarpus* spp.) produce edible seeds covered by a thin seed coat. These factors limit archaeobotanical record of persevered seeds. In contrast, hard protective structures, such as the endocarps enveloping the seeds of *Canarium* and *Terminalia* species, are more likely to be found as charred archaeological plant macrofossils (Fairbairn & Florin, 2022).

1.2.7 Developmental pathways of fruit tree domestication and fruit production

When studying the developmental pathways of fruit tree domestication and fruit production in the region, two key points to consider. Firstly, the fruiting onset is a crucial factor. Well-tended trees in modern cultivation only begin to set fruit at seven year-old in both durian (National Research Council, 1975) and mangosteen (Almeyda & Martin, 1976). The initial planting stocks introduced into cultivation by early cultivators would require an even longer period.

Secondly, the adaptability of fruit trees seedlings and saplings to establish themselves in man-made environments is significant. Using durian and mangosteen as examples once again, their seedlings and saplings exhibit shade tolerance and would not survive without intensive care in a man-made environment. In contrast, the natural environment of a pristine forest with a closed canopy and high humidity proves ideal for the survival and growth of seedlings and saplings.

The cultivation of trees with a delayed onset of production represents a substantial long-term investment. The husbandry practices involving the introduction of forest species into a human modified environments, as outlined by Wiersum (2008) (**Table 1.3**), were likely inculcated through heuristic learning among early cultivators. Wiersum (2008) framed the outline of mutual development between production systems and cultivated trees into phasic orders. These phasic orders advance along a gradient of increasing management intensity, emphasising the coevolution of niches and cultivated trees. Wiersum's framework was formulated with specific reference to Southeast Asian scenarios, supported by examples from the Neotropics (Anderson, 1990).

The progression unfolds as follows: initially, local communities gathered fruits from wild trees in the natural forests. This was followed by a gradient of forest management regimes aimed at enhancing the presence of fruit trees and increasing fruit yield. Ultimately, cultigens were introduced into man-made environments (**Table 1.3**). These

phases differ in the intensity of human intervention and the environment settings involved.

Table 1.3 : Phasic framework of mutual progress between production systems and cultivated trees in exploitation of tree crops; after Table 4.2

Phase	<i>modus operandi</i>
I. Acquiring of wild tree products by gathering/collection	range from uncontrolled, open access gathering of forest products to controlled gathering of wild tree products
II. Initial production of wild tree products	range from systematic collection of wild tree products with protective tending of valued tree species to selective cultivation of valued trees by artificial <i>in situ</i> regeneration of native tree
III. Cultivation of wild trees	cultivated of selected native tree species in artificially established plantations
IV. Cultivation of fully domesticated trees	cultivation of genetically selected or improved tree crops in intensively managed plantations

(Adapted from Wiersum 2008)

1.3 Research problems and plausible solutions

1.3.1 The origin of the cultivated mangosteen: a contentious conundrum

The origin of the cultivated mangosteen has been a contentious topic for decades, marked by incongruous findings from cytological and phylogenetics studies. Candidate progenitors, including *Garcinia celebica*, *G. diospyrifolia*, *G. mangostana* var. *malaccensis* and *G. penangiana* have been proposed in various studies (Richards, 1990; Abdullah et al., 2012; Nazre, 2014). More recent findings, particularly from Hambali & Natawijaya (2016) and Nazre (2014), suggest that var. *malaccensis* is the most probable sole progenitor of the cultivated mangosteen.

Despite there previous studies, there has been limited study on *G. mangostana* var. *borneensis*, especially its genetic affinity to var. *mangostana*. The inclusion of *G. mangostana* var. *borneensis* in this study is, therefore, indispensable to clarify the tokogeny between *G. mangostana* varieties.

Nazre (2014) and Hambali & Natawijaya (2016) have provided indications on how the cultivated mangosteen emerged from var. *malaccensis*, but the overview of its domestication history remains incomplete. *Garcinia mangostana* var. *malaccensis* exhibits distribution throughout the Malay Peninsula, Sumatra and West Borneo. However, a crucial question arises: does the domentsitcation history in mangosteen show any geographical-signal? For example, *G. mangostana* var. *borneensis* is restricted to East Borneo, particularly common in the east coast of Sabah. Are the mangosteens cultivated in East Borneo more closely related to var. *borneensis* when compared to var. *malaccensis*? Alternatively, did the modern cultivated mangosteens originate from a few

polyploid apomictic ancestors in a particular region, and was the lineage propagated and disseminated through human-mediated dispersal? These questions, as of now, remained unanswered.

Considering the effects of domestication syndrome on the breeding biology, particularly apomixis in mangosteen, Yao et al. (2023) have proposed an evolutionary-ecological theory named “Forest-*Dusun* Interface” (**Figure 1.10 & Chapter 2**), which aligned with Wiersum’s (2008) phasic framework. In this context, “*Dusun*” (Malay) refers to subsistence orchards. The Forest-*Dusun* Interface presents two possible scenarios.

In the first scenario, apomixis evolved during domestication. Upon the initial and unbiased introduction of planting stocks into the man-made environments, the culling of individuals that did not bear fruits might have occurred, leading to the positive selection of apomictic females. Polyploidy might have evolved concurrently with or independently of apomixis. The second scenario is simpler: apomixis and polyploidy were already present in wild progenitors and were introduced into cultivation by the early cultivators.

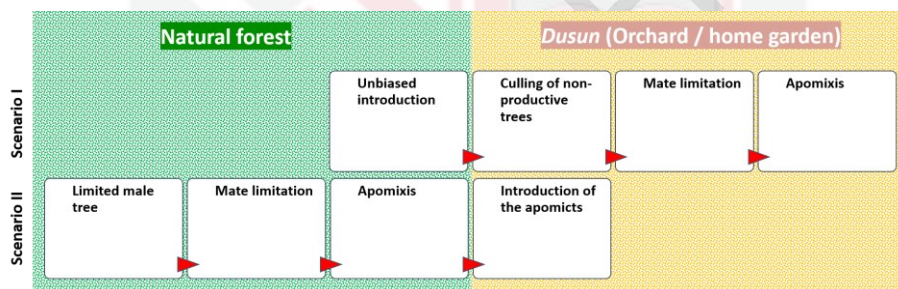


Figure 1.10 : Forest-*Dusun* Interface. This evolutionary-ecological theory put forward two possible scenarios regarding the inception of apomixis reproduction and polyploidy in cultivated mangosteen

1.3.2 Deducing the domestication history: suitable approaches for *G. mangostana*

The current understanding of the cultivated mangosteen and its wild relatives prompts new consequential research questions:

- 1) Is the varietal delimitation of the cultivated and wild varieties well defined based on morphological characters?
- 2) What is the intraspecific tokogeny between the cultivated populations and wild relatives?
- 3) Given that var. *malaccensis* is distributed in Sumatra, Malay Peninsula and western Borneo, and var. *borneensis* is distributed in eastern Borneo, is there

a recoverable geographical signal between the cultivated populations and the wild relatives in these respective regions?

Addressing these questions will contribute to a more comprehensive understanding of the mangosteen's domestication history, varietal distinctions, and the relationships between cultivated and wild populations across different geographical regions.

Several methods are commonly employed to trace the domestication history of a cultivated trees. These methods, as outlined by Herniter et al. (2020) and Warschefsky & von Wettberg (2019), include (i) archaeobotanical studies; (ii) examination and interpretation of historical records; (iii) ethnobotanical surveys; (v) morphological analyses and (v) genetic analyses.

Realistically, a combination of methods, tailored to the available data and research context, is often the most effective approach. Integrating findings from multiple sources can provide a more comprehensive understanding of mangosteen's domestication history.

1.3.3 Archaeobotanical study

Archaeobotanical studies involve the analysis of plant remains preserved, and recovered from archaeological sites. The examination of the carbonised or charred plant remains, such as ancient stony seeds and hard fruits capsules, allows researchers to identify and compare morphological characteristics of the ancient domesticated species with modern domesticated species and their wild relatives. By interpreting these findings, researchers can infer when and where the domestication originated. Unfortunately, representations of plant remains, especially tree fruit, in archaeobotanical studies of Southeast Asia are scarce and restricted to a few species (Fairbairn & Florin, 2022; Paz, 2005). Regrettably, to date, archaeobotanical approaches have not provided any information on the cultivation history of mangosteen.

1.3.4 Examination and interpretation of historical records

Examination and interpretation of historical records, such as ancient texts, manuscripts and illustrations such as cave paintings and ancient building reliefs, offer insightful information into the domestication history of species. These records may describe cultivation practices, natural products available in the marketplace, trade routes, and the cultural significance of specific domesticated trees. Such information provides useful insights into the timeline of domestication and the geographical spread of domesticated tree species.

In the case of Southeast Asia, historical texts on the geography, culture and natural products often find their first records in Chinese texts (Gong, 2019 [originally published in 1434]; I-ching, 1896 [originally written between 671–695]; Ma, 2019 [originally published in 1416]). These historical texts are written in archaic languages and can be

difficult to interpret without translated, or annotated versions. Historical records suggest that mangosteen was among the fruits sold in the marketplaces in East Java and North Sumatra (Gong, 2019; Ma, 2019). However, there is no indication of whether the fruits were collected from forests or cultivated in orchards or home gardens (Yao et al., 2023).

1.3.5 Ethnobotanical survey

Ethnobotanical surveys are invaluable for gathering information on cultivation practices, providing insights into the selection of desirable traits (McKey et al., 2010). Recent ethnobotanical studies conducted in Southeast Asia have primarily focused on the tree structure and species composition of the agroforests and home gardens (Aumeeruddy, 1993; Michon et al., 1983, 1986). In contrast, older studies such as Freeman (1955) focused on grain crops such as hill rice. However, as of my best knowledge, I am not aware of any specific ethnobotanical study in the region that specifically focuses on how farmers' practices shaped the variation in fruit tree produce, as exemplified in Rimlinger et al. (2019).

1.3.6 Morphological analysis

Morphological analyses serve as the initial steps in identifying the probable wild relatives of a cultivated species, providing a direct observation on phenotypes. Through comparisons on morphological traits in vegetative (twigs and leaves) and reproductive organs (flowers and fruits) between the domesticated and wild populations, researchers can identify changes that occurred during domestication, characterizing the domestication syndrome. Botanical institutes offer excellent opportunities for conducting morphological studies.

Herbarium specimens of *G. mangostana* wild relatives are well-represented in regional herbaria, including Kepong, Forest Research Institute Malaysia (KEP); Sandakan, Forest Research Centre, Sabah (SAN); Singapore, Singapore Botanic Garden (SING); and Wanariset, Balai Penerapan Standar Instrumen Lingkungan Hidup dan Kehutanan Samboja, East Kalimantan (WAN). Geographical distributions and morphological variations of var. *malaccensis* and var. *borneensis* are primarily documented through herbarium records. Unfortunately, the full range of variations in cultivated mangosteen is not comprehensively represented in herbaria.

1.3.7 Genetic analysis

Genetic analyses play a crucial role in tracing the domestication history of cultivated species. Depending on the availability of reference genomes and the selection of the genetic markers, researchers can address different research questions. Presently, high-throughput next-generation sequencing technologies enable the generation of large datasets of Single Nucleotide Polymorphisms (SNPs) at a manageable cost. Simple Sequence Repeats (SSRs) continue to be an effective marker for examining population genetic variation.

By comparing SNPs and SSRs between populations of cultivated and wild relatives, researchers can characterise population genetic variation and identify genotypes. This information aids in understanding the relationships, diversity, and evolutionary dynamics between cultivated and wild populations, contributing to a more comprehensive understanding of the domestication history of the species in question.

The chloroplast genome is known for its higher conservation in both gene content and physical structure compared to nuclear and mitochondrial genomes (Jansen et al., 2005). The nucleotide mutation rate of chloroplast genes is moderate, falling between the rates of nuclear and mitochondrial genes (Dong et al., 2020). Chloroplast markers have long been proven suitable for inferring the population tokogeny of different plant groups (D. E. Soltis et al., 1992) and enable the differentiation of lineage sorting from polymorphic ancestry (Lavin et al., 1991).

Studies by Miller & Schaal (2005) and (Yue et al., 2018) successfully identified haplotypes in fruit tree species using chloroplast markers. These studies were able to infer the domestication history and geographical origin of cultivated fruit. Additionally, the chloroplast genome is maternally inherited, haploid and non-recombining (Birky, 2001). These characteristics makes chloroplast genome SNPs ideal markers for reconstructing the demographic history of plant species.

Given the scarcity of archaeobotanical findings, historical records, and ethnobotanical studies focused on cultivated trees in Southeast Asia, the adaption of comparative morphological and genetic studies appeared as the most viable and realistic approaches for the current study

1.4 Objectives

The overarching goal of this study is to deduce the domestication history of the cultivated mangosteen and contribute to the conservation, sustainable management, and utilisation of genetic resources in both cultivated and wild mangosteen populations. The specific objectives identified to achieve this overarching goal are outlined below:

- (I) To clarify the relationship between and within the cultivated mangosteen and wild relatives through morphological and genetics studies;
- (II) To elucidate the origin of domesticated and evolutionary history of cultivated mangosteen through genetic analyses;
- (III) To characterise the geographical distribution of genetic diversity among the wild populations for the formulation of conservation plan.

1.5 Thesis work plan

The current study adopts a multiple-faceted approach, incorporating both botanical and genetic methods, to fulfil its objectives. **Chapter 2** presents an extensive literature review with a specific emphasis on the origin of cultivated mangosteen and its breeding biology. This review aids in establishing a coherent theoretical framework and defining the geographical boundaries for sampling. The sampling scope, and overview of morphology and genetic studies, are outlined below.

1.5.1 Sampling campaign plan

Given the postulation from previous studies that the cultivated mangosteen originated in a region spanning across Sumatra, Peninsular Malaysia, Singapore and Borneo, strategic sampling efforts are essential. To elucidate the origin of cultivated mangosteen and gain a comprehensive perspective of the domestication history, sampling should cover populations across Indonesia, Malaysia and Singapore. This geographical approach aims to capture the diversity of mangosteen populations within the suggested origin region.

In term of taxonomic representation, it is crucial to conduct sampling not only of cultivated mangosteen across the region but also of its wild relatives, namely *G. mangostana* var. *borneensis* and var. *malaccensis* are needed. This inclusive approach ensures a thorough examination of the entire species complex, facilitating a more accurate analysis of the relationships between cultivated and wild mangosteen populations.

Sampling of the cultivated *G. mangostana* var. *mangostana* and var. *malaccensis* from orchards and agricultural centres represents a relatively easy task. However, the challenge lies in the sampling of wild relatives, especially in acquiring at least 20 individuals per population for population genetic study. Wild populations of *G. mangostana* var. *borneensis* and var. *malaccensis* are located in remote forested areas. Locating and collecting them is logistically challenging. Additionally, obtaining the necessary permissions for fieldworks across the entire region add another layer of complexity. Extracting leaf tissues from herbarium specimens is an alternative sampling method for the wild relatives.

Species of congeners, especially in the *Garcinia* sect. *Garcinia* are also collected when they were encountered during the fieldworks. Additionally, other *Garcinia* species not native to Southeast Asia were collected from botanical gardens, agriculture centres, and private gardens. In total, the study gathered 545 accessions of leaf tissues. These accessions, obtained from both field collections and the extraction of tissues from herbarium specimens, cover the entire region of origin. The localities of all samples gathered in this study are mapped in **Figure 1.11**.

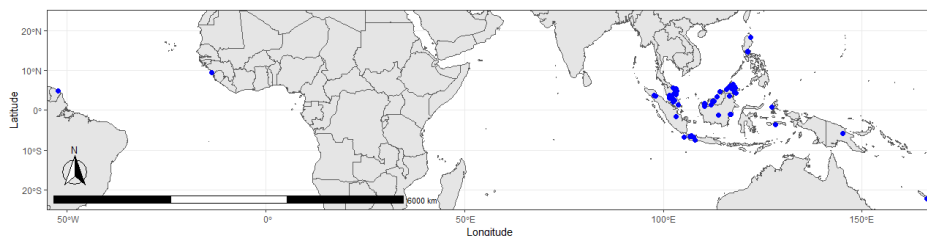


Figure 1.11 : The distribution of all samples acquired for this study

Field trips were conducted for three main purposes: (i) to supplement herbarium specimens intended for morphometric analyses; (ii) to make observations on field characters and niche preferences; and (iii) to collect leaf tissues for subsequent genetic analyses. The sampling fieldwork spanned various countries and regions, specifically Peninsular Malaysia, Singapore, West Java, Sabah and East Kalimantan. The sampling coverage of the two wild varieties is mapped in **Figure 1.12**. It is crucial to emphasize that the majority of the collected accessions originated from sterile trees encountered during the fieldwork. Additionally, some specimens were obtained from young saplings, and attempts to locate the supposed mother trees were unsuccessful. Consequently, the taxon identification of these samples relied on vegetative characters.

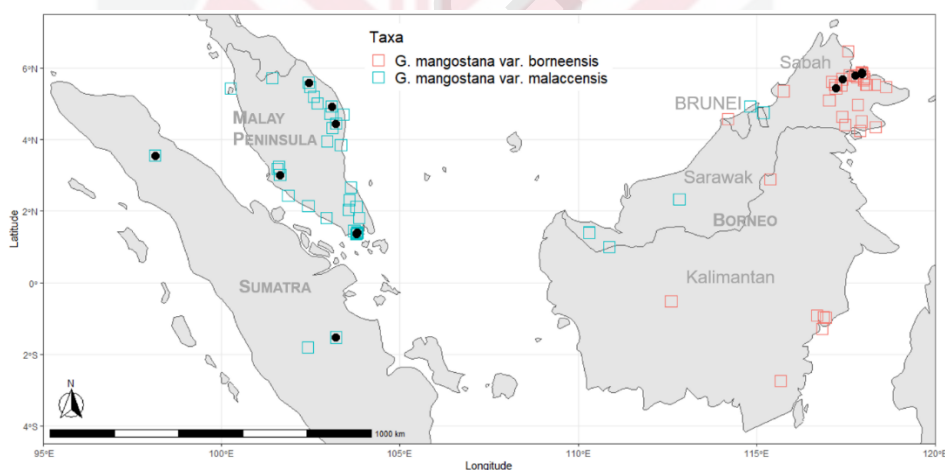


Figure 1.12 : The sampling coverage of two wild varieties. The bullets (•) denote the sites where samples were acquired for this study. Note that the Sumatran samples were collected *ex-situ* from private gardens in West Java

1.5.2 Morphometric analyses

Cultivated mangosteens were classified under *G. mangostana* var. *mangostana*, while var. *malaccensis* and var. *borneensis* were recently defined to accommodate the two wild relatives (Nazre et al., 2018). However, the morphological characters used to delimitate these two new taxa have not undergone rigorous statistical analyses to assess their efficacy. In order to clarify the varietal distinction within *Garcinia mangostana* and distinguish them from two other morphologically similar species, namely *G. penangiana* and *G. venulosa*, morphometric analyses were employed. A total of 124 specimens were examined, including *G. mangostana* var. *mangostana* (35), var. *borneensis* (16), var. *malaccensis* (23), *G. penangiana* (30) and *G. venulosa* (20). The results of the morphometric study, outlined in **Chapter 3**, are essential for determining whether the morphological phenotypes align with the genotypes. These findings would contribute to validating the consistency between morphological phenotypes and genetic makeup.

1.5.3 Genetics: phylogenomic and population genetic approaches

Laboratory work was conducted at “Plant Biodiversity and Adaptation Research Group (DYNADIV)” plant genetics laboratory in *Institut de Recherche pour le Développement* (IRD), Montpellier, France.

A total of 192 samples were chosen for genetic library construction. The High-seq Illumina technology was employed to conduct the sequencing of the genetics datasets. The two datasets generated from the library encompassed: (i) untargeted whole genome sequencing (WGS) sequences, and (ii) targeted sequencing using universal Angiosperms353 probes. In case of WGS sequences, alignment with a published chloroplast genome facilitated the identification of Single Nucleotide Polymorphisms (SNPs) from the mapped sequences. The chloroplast genomic SNPs were instrumental in unraveling the phylogenetic relationships among all studied taxa, as well as elucidating haplotypes within cultivated and wild mangosteen populations. The genetic study is presented in **Chapter 4**.

Notably, at the time of thesis submission, the bioinformatics analysis of the Angiosperms353 dataset had been completed up to the re-pairing stage. The presence of polyploidy in cultivated mangosteen necessitated the implementation of more technically-demanding and time-consuming procedures to eliminate paralogs. Consequently, the Angiosperms353 dataset was excluded from the genetic analyses.

1.6 Challenges and limitations

The COVID-19 pandemic, while no longer a current public health issue, continues to exert adverse effect to the post-graduate study projects initiated during the public health emergency. This PhD studentship is a cotutelle programme jointly registered at both the University of Montpellier and Universiti Putra Malaysia. The initial project plan

involved field trips for samplings in Southeast Asia, laboratory analyses in Montpellier, and inter-university travel. Additionally, extensive sampling across the natural distribution of study objects in Southeast Asia is imperative. However, travel restriction imposed due to the pandemic impeded the progress of the project.

The natural distribution of both origin cultigens and wild relatives spans multiple countries in Southeast Asia. Securing collection and export permits from various countries across the region is inherently challenging even under ordinary circumstances, let alone during a global pandemic. The travel restrictions, therefore, presented a challenge to the timely execution of the project.



References

- Abdullah, N. A. P., Richards, A. J., & Wolff, K. (2012). Molecular evidence in identifying parents of *Garcinia mangostana* L. *Pertanika Journal of Tropical Agricultural Science*, 35(2), 257–270.
- Ali Zifan. (2016, February 20). *File: Southeast Asia map of Köppen climate classification.svg*—Wikipedia.
https://commons.wikimedia.org/wiki/File:Southeast_Asia_map_of_K%C3%B6ppen_climate_classification.svg
- Almeyda, N., & Martin, F. M. (1976). *Cultivation of neglected tropical fruits with promise: Part 1. The mangosteen*. Agricultural Research Service, USDA.
<https://ia600209.us.archive.org/17/items/cultivationofneg155unit/cultivationofneg155unit.pdf>
- Anderson, A. B. (1990). Extraction and forest management by rural inhabitants in the Amazon estuary. In A. B. Anderson (Ed.), *Alternatives to deforestation: Steps toward sustainable use of the Amazon Rain Forest*. Columbia University Press.
<https://www.osti.gov/biblio/6339471>
- Arias, R. S., Ballard, L. L., Duke, M. V., Simpson, S. A., Liu, X. F., Orner, V. A., Sobolev, V. S., Scheffler, B. E., & Martinez-Castillo, J. (2020). Development of nuclear microsatellite markers to facilitate germplasm conservation and population genetics studies of five groups of tropical perennial plants with edible fruits and shoots: Rambutan (*Nephelium lappaceum* L.), sapodilla (*Manilkara zapota* (L.) P. Royen), lychee (*Litchi chinensis* Sonn.), mangosteen (*Garcinia mangostana* Linn. and *Garcinia cochinchinensis* (Lour.) Choisy) and bamboo (*Bambusa vulgaris* Schrad. ex J.C. Wendl and *Guadua angustifolia* Kunth). *Genetic Resources and Crop Evolution*, 67(7), 1715–1731.
<https://doi.org/10.1007/s10722-020-00965-w>
- Aumeeruddy, Y. (1993). *Agroforêts et aires de forêts protégées. Représentations et pratiques agroforestières paysannes du Parc National Kerinci Seblat, Sumatra, Indonésie* [Doctoral dissertation]. Université de Montpellier 2.
- Bawin, Y., Ruttink, T., Staelens, A., Haegeman, A., Stoffelen, P., Mwanga Mwanga, J.-C. I., Roldán-Ruiz, I., Honnay, O., & Janssens, S. B. (2021). Phylogenomic analysis clarifies the evolutionary origin of *Coffea arabica*. *Journal of Systematics and Evolution*, 59(5), 953–963. <https://doi.org/10.1111/jse.12694>
- Beck, H. E., Zimmermann, N. E., McVicar, T. R., Vergopolan, N., Berg, A., & Wood, E. F. (2018). Present and future Köppen-Geiger climate classification maps at 1-km resolution. *Scientific Data*, 5(1), Article 1.
<https://doi.org/10.1038/sdata.2018.214>
- Besnard, G., Khadari, B., Navascués, M., Fernández-Mazuecos, M., El Bakkali, A., Arrigo, N., Baali-Cherif, D., Brunini-Bronzini de Caraffa, V., Santoni, S., Vargas, P., & Savolainen, V. (2013). The complex history of the olive tree: From Late Quaternary diversification of Mediterranean lineages to primary domestication in the northern Levant. *Proceedings of the Royal Society B*:

- Birky, C. W. (2001). The inheritance of genes in mitochondria and chloroplasts: Laws, mechanisms, and models. *Annual Review of Genetics*, 35, 125–148. <https://doi.org/10.1146/annurev.genet.35.102401.090231>
- Blench, R. (2007). *A history of fruits on the SE Asian mainland*. EurASEAA 11, Bougon, France.
- Brunner, B. R., & Morales-Payan, J. P. (2010). Mangosteen and Rambutan. In *Soils, Plant Growth and Crop Production* (p. 26). UNESCO.
- Candolle, A. de. (2011). *The Origin of Cultivated Plants*. Cambridge University Press. <https://doi.org/10.1017/CBO9781139107365>
- Cannon, C. H., Morley, R. J., & Bush, A. B. G. (2009). The current refugial rainforests of Sundaland are unrepresentative of their biogeographic past and highly vulnerable to disturbance. *Proceedings of the National Academy of Sciences*, 106(27), 11188–11193. <https://doi.org/10.1073/pnas.0809865106>
- Cappellini, E., Gilbert, M. T. P., Geuna, F., Fiorentino, G., Hall, A., Thomas-Oates, J., Ashton, P. D., Ashford, D. A., Arthur, P., Campos, P. F., Kool, J., Willerslev, E., & Collins, M. J. (2010). A multidisciplinary study of archaeological grape seeds. *Naturwissenschaften*, 97(2), 205–217. <https://doi.org/10.1007/s00114-009-0629-3>
- Chennaveeraiah, M. S., & Razdan, M. K. (1975). The basic chromosome number in the genus *Garcinia* L. (Guttiferae). *Proc. Indian Sci. Congr. Assoc.*, 62, 127.
- Clement, C. R. (1999). 1492 and the loss of amazonian crop genetic resources. I. The relation between domestication and human population decline. *Economic Botany*, 53(2), 188–202. <https://doi.org/10.1007/BF02866498>
- Clement, C. R., Denevan, W. M., Heckenberger, M. J., Junqueira, A. B., Neves, E. G., Teixeira, W. G., & Woods, W. I. (2015). The domestication of Amazonia before European conquest. *Proceedings of the Royal Society B: Biological Sciences*, 282(1812), 20150813. <https://doi.org/10.1098/rspb.2015.0813>
- Cohen, P., Bacilieri, R., Ramos-Madriral, J., Privman, E., Boaretto, E., Weber, A., Fuks, D., Weiss, E., Erickson-Gini, T., Bucking, S., Tepper, Y., Cvikel, D., Schmidt, J., Gilbert, M. T. P., Wales, N., Bar-Oz, G., & Meiri, M. (2023). Ancient DNA from a lost Negev Highlands desert grape reveals a Late Antiquity wine lineage. *Proceedings of the National Academy of Sciences*, 120(17), e2213563120. <https://doi.org/10.1073/pnas.2213563120>
- Davis, A. P., Kiwuka, C., Faruk, A., Mulumba, J., & Kalema, J. (2023). A review of the indigenous coffee resources of Uganda and their potential for coffee sector sustainability and development. *Frontiers in Plant Science*, 13. <https://www.frontiersin.org/articles/10.3389/fpls.2022.1057317>

- Diamond, J. (2002). Evolution, consequences and future of plant and animal domestication. *Nature*, 418(6898), Article 6898. <https://doi.org/10.1038/nature01019>
- Dong, W., Xu, C., Wen, J., & Zhou, S. (2020). Evolutionary directions of single nucleotide substitutions and structural mutations in the chloroplast genomes of the family Calycanthaceae. *BMC Evolutionary Biology*, 20(1), 96. <https://doi.org/10.1186/s12862-020-01661-0>
- Escobar, H. (2019). Amazon fires clearly linked to deforestation, scientists say. *Science*, 365(6456), 853–853. <https://doi.org/10.1126/science.365.6456.853>
- Fairbairn, A. S., & Florin, S. A. (2022). Archaeological identification of fragmented nuts and fruits from key Asia-Pacific economic tree species using anatomical criteria: Comparative analysis of *Canarium*, *Pandanus* and *Terminalia*. *Archaeology in Oceania*, 57(3), 160–188. <https://doi.org/10.1002/arco.5273>
- FAO. (2020). *Major Tropical Fruits Market Review February 2020 snapshot*. FAO. <http://www.fao.org/3/ca9213en/ca9213en.pdf>
- FAO. (2023). Tropical fruits. *Commodities: Tropical Fruits*. <https://www.fao.org/markets-and-trade/commodities/en/>
- Freeman, D. (1955). *Iban agriculture a report on the shifting cultivation of hill rice by the Iban of Sarawak*. H.M. Stationery Off.
- Fuller, D. Q., Castillo, C., Kingwell-Banham, E., Qin, L., & Weisskopf, A. (2018). Charred pummelo peel, historical linguistics and other tree crops: Approaches to framing the historical context of early Citrus cultivation in East, South and Southeast Asia. In G. Fiorentino & V. Zech-Matterne (Eds.), *AGRUMED: Archaeology and history of citrus fruit in the Mediterranean: Acclimatization, diversifications, uses*. Publications du Centre Jean Bérard. <https://doi.org/10.4000/books.pcbj.2173>
- Gaut B. S., Díez, C. M., & Morrell, P. L. (2015). Genomics and the Contrasting Dynamics of Annual and Perennial Domestication. *Trends in Genetics*, 31(12), 709–719. <https://doi.org/10.1016/j.tig.2015.10.002>
- Gong, Z. (2019). 《西洋番国志 校注本》(*Collated and annotated Xī Yáng Fān Guó Zhì [1434], 'The Overall Survey of the Ocean's Shores'*) (D. Xiang, Ed.). Hua Wen Publisher.
- Groppi, A., Liu, S., Cornille, A., Decroocq, S., Bui, Q. T., Tricon, D., Cruaud, C., Arribat, S., Belser, C., Marande, W., Salse, J., Huneau, C., Rodde, N., Rhalloussi, W., Cauet, S., Istace, B., Denis, E., Carrère, S., Audergon, J.-M., ... Decroocq, V. (2021). Population genomics of apricots unravels domestication history and adaptive events. *Nature Communications*, 12(1), Article 1. <https://doi.org/10.1038/s41467-021-24283-6>
- Gros-Balthazard, M., Flowers, J. M., Hazzouri, K. M., Ferrand, S., Aberlenc, F., Sallon, S., & Purugganan, M. D. (2021). The genomes of ancient date palms

- germinated from 2,000 y old seeds. *Proceedings of the National Academy of Sciences*, 118(19), e2025337118. <https://doi.org/10.1073/pnas.2025337118>
- Ha, C. O. (1978). *Embryological and cytological aspects of the reproductive biology of some understorey rainforest trees*. Unpublished PhD. thesis. University of Malaya.
- Ha, C. O., Sands, V. E., Soepadmo, E., & Jong, K. (1988). Reproductive patterns of selected understorey trees in the Malaysian rain forest: The apomictic species. *Botanical Journal of the Linnean Society*, 97(3), 317–331. <https://doi.org/10.1111/j.1095-8339.1988.tb01586.x>
- Hambali, G. G., & Natawijaya, A. (2016). A preliminary study of *Garcinia malaccensis* collection in Taman Buah Mekarsari, Cileungsi, Bogor, Indonesia. *Contribution of Breeding Research for Sustainable Agricultural Production under Changing Environment for Food Security in Asia and Oceania*, 31–36.
- Harlan, J. R. (1971). Agricultural Origins: Centers and Noncenters. *Science*, 174(4008), 468–474.
- Harlan, J. R. (1992). *Crops and Man*. American Society of Agronomy : Crop Science Society of America. <https://dl.sciencesocieties.org/publications/books/tocs/acsesspublicati/cropsandman>
- Herniter, I. A., Muñoz-Amatriain, M., & Close, T. J. (2020). Genetic, textual, and archeological evidence of the historical global spread of cowpea (*VIGNA UNGUICULATA* [L.] Walp.). *Legume Science*, 2(4). <https://doi.org/10.1002/leg3.57>
- Horn, C. L. (1940). Existence of only one variety of cultivated mangosteen explained by asexually formed ‘seed’. *Science*, 92(2385), 237–238. <https://doi.org/10.1126/science.92.2385.237>
- I-ching. (1896). *A record of the Buddhist religion as practised in India and the Malay archipelago*. Oxford, The Clarendon press. <http://archive.org/details/recordofbuddhist00ichi>
- Idris, S., & Rukayah, A. (1987). Description of the male mangosteen (*Garcinia mangostana* L.) discovered in Peninsular Malaysia. *MARDI Research Bulletin*, 15(1), 63–66.
- Jansen, R. K., Raubeson, L. A., Boore, J. L., dePamphilis, C. W., Chumley, T. W., Haberle, R. C., Wyman, S. K., Alverson, A. J., Peery, R., Herman, S. J., Fourcade, H. M., Kuehl, J. V., McNeal, J. R., Leebens-Mack, J., & Cui, L. (2005). Methods for Obtaining and Analyzing Whole Chloroplast Genome Sequences. In *Methods in Enzymology* (Vol. 395, pp. 348–384). Academic Press. [https://doi.org/10.1016/S0076-6879\(05\)95020-9](https://doi.org/10.1016/S0076-6879(05)95020-9)
- Johns, R. J. (1995). Malesia – an Introduction. *Curtis’s Botanical Magazine*, 12(2), 52–62.

- Jones, C. J., Edwards, K. J., Castaglione, S., Winfield, M. O., Sala, F., van de Wiel, C., Bredemeijer, G., Vosman, B., Matthes, M., Daly, A., Brettschneider, R., Bettini, P., Buiatti, M., Maestri, E., Malcevski, A., Marmioli, N., Aert, R., Volckaert, G., Rueda, J., ... Karp, A. (1997). Reproducibility testing of RAPD, AFLP and SSR markers in plants by a network of European laboratories. *Molecular Breeding*, 3(5), 381–390. <https://doi.org/10.1023/A:1009612517139>
- Käfer, J., Méndez, M., & Mousset, S. (2022). Labile sex expression in angiosperm species with sex chromosomes. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 377(1850), 20210216. <https://doi.org/10.1098/rstb.2021.0216>
- Kantar, M. B., Nashoba, A. R., Anderson, J. E., Blackman, B. K., & Rieseberg, L. H. (2017). The Genetics and Genomics of Plant Domestication. *BioScience*, 67(11), 971–982. <https://doi.org/10.1093/biosci/bix114>
- King, G. (1890). VIII. Materials for a Flora of Malayan Peninsula. *The Journal of the Asiatic Society of Bengal*, 59(2), 113–206.
- Krishnaswamy, N., & Raman, V. S. (1949). A note on the chromosome numbers of some economic plants of India. *Current Science*, 10, 376–378.
- Lape, P., Peterson, E., Tanudirjo, D., Shiung, C.-C., Lee, G.-A., Field, J., & Coster, A. (2018). New Data from an Open Neolithic Site in Eastern Indonesia. *Asian Perspectives*, 57(2), 222–243. <https://doi.org/10.1353/asi.2018.0015>
- Lavin, M., Mathews, S., & Hughes, C. (1991). Chloroplast DNA Variation in *Gliricidia sepium* (Leguminosae): Intraspecific Phylogeny and Tokogeny. *American Journal of Botany*, 78(11), 1576–1585. <https://doi.org/10.2307/2444983>
- Levis, C., Costa, F. R. C., Bongers, F., Peña-Claros, M., Clement, C. R., Junqueira, A. B., Neves, E. G., Tamanaha, E. K., Figueiredo, F. O. G., Salomão, R. P., Castilho, C. V., Magnusson, W. E., Phillips, O. L., Guevara, J. E., Sabatier, D., Molino, J.-F., López, D. C., Mendoza, A. M., Pitman, N. C. A., ... ter Steege, H. (2017). Persistent effects of pre-Columbian plant domestication on Amazonian forest composition. *Science*, 355(6328), 925–931. <https://doi.org/10.1126/science.aal0157>
- Lim, A. L. (1984). The embryology of *Garcinia mangostana* L. (Clusiaceae). *Gardens' Bulletin Singapore*, 37(1).
- Lim, T. K. (2012). *Garcinia malaccensis*. In T. K. Lim (Ed.), *Edible Medicinal And Non-Medicinal Plants: Volume 2, Fruits* (pp. 80–82). Springer Netherlands. https://doi.org/10.1007/978-94-007-1764-0_14
- Ma, H. (2019). 《瀛涯勝覽 校注本》(Collated and annotated *Ying Ya Sheng Lan* [1416], 'The Overall Survey of the Ocean's Shores') (C.-C. Feng, Ed.). Hua Wen Publisher.
- Majeed, S., Chaudhary, M. T., Hulse-Kemp, A. M., & Azhar, M. T. (2021). Chapter 1 - Introduction: Crop Wild Relatives in Plant Breeding. In M. T. Azhar & S. H.

- Wani (Eds.), *Wild Germplasm for Genetic Improvement in Crop Plants* (pp. 1–18). Academic Press. <https://doi.org/10.1016/B978-0-12-822137-2.00001-1>
- Mansyah, E., Jawal, M. A. S., Muas, I., Hendri, & Usman, F. (2007). Identifikasi dan karakterisasi manggis di Provinsi Bengkulu dan Banka-Belitung. *Jurnal Hortikultura*, 17(2), 118–126.
- Matra, D. D., Poerwanto, R., Santosa, E., Sobir, Higashio, H., Anzai, H., & Inoue, E. (2016). Analysis of Allelic Diversity and Genetic Relationships Among Cultivated Mangosteen (*Garcinia mangostana* L.) in Java, Indonesia Using Microsatellite Markers and Morphological Characters. *Tropical Plant Biology*, 9(1), 29–41. <https://doi.org/10.1007/s12042-016-9161-8>
- Mboujda, F. M. M., Avana-Tientcheu, M.-L., Momo, S. T., Ntongme, A. M., Vaissayre, V., Azandi, L. N., Dussert, S., Womeni, H., Onana, J.-M., Sonké, B., Tankou, C., & Duminil, J. (2022). Domestication Syndrome in *Dacryodes edulis* (Burseraceae): Comparison of Morphological and Biochemical Traits between Wild and Cultivated Populations. *Plants*, 11(19), Article 19. <https://doi.org/10.3390/plants11192496>
- McKey, D., Elias, M., Pujol, B., & Duputié, A. (2010). The evolutionary ecology of clonally propagated domesticated plants: Tansley review. *New Phytologist*, 186(2), 318–332. <https://doi.org/10.1111/j.1469-8137.2010.03210.x>
- Meyer, R. S., DuVal, A. E., & Jensen, H. R. (2012). Patterns and processes in crop domestication: An historical review and quantitative analysis of 203 global food crops: Tansley review. *New Phytologist*, 196(1), 29–48. <https://doi.org/10.1111/j.1469-8137.2012.04253.x>
- Meyer, R. S., & Purugganan, M. D. (2013). Evolution of crop species: Genetics of domestication and diversification. *Nature Reviews. Genetics*, 14(12), 840–852. <https://doi.org/10.1038/nrg3605>
- Michon, G., Bompard, J., Hecketsweiler, P., & Ducatillion, C. (1983). Tropical forest architectural analysis as applied to agroforests in the humid tropics: The example of traditional village-agroforests in West Java. *Agroforestry Systems*, 1(2), 117–129. <https://doi.org/10.1007/BF00596353>
- Michon, G., Mary, F., & Bompard, J. (1986). Multistoried agroforestry garden system in West Sumatra, Indonesia. *Agroforestry Systems*, 4(4), 315–338. <https://doi.org/10.1007/BF00048106>
- Midin, M. R. (2018). *Cytogenetics study of Garcinia mangostana* [Doctoral dissertation, International Islamic University Malaysia]. <https://lib.iium.edu.my/mom/services/mom/document/getFile/Xk6MR5OemyO2SgXqQZvLcwWnaeRK16Ry20190410115700600>
- Midin, M. R., Nordin, M. S., Madon, M., Nazre, S., Goh, H.-H., & Normah, M. N. (2018). Determination of the chromosome number and genome size of *Garcinia mangostana* L. via cytogenetics, flow cytometry and k-mer analyses. *Caryologia*, 71(1), 35–44. <https://doi.org/10.1080/00087114.2017.1403762>

- Miller, A., & Gross, B. L. (2011). From forest to field: Perennial fruit crop domestication. *American Journal of Botany*, 98(9), 1389–1414. <https://doi.org/10.3732/ajb.1000522>
- Miller, A., & Schaal, B. (2005). Domestication of a Mesoamerican cultivated fruit tree, *Spondias purpurea*. *Proceedings of the National Academy of Sciences*, 102(36), 12801–12806. <https://doi.org/10.1073/pnas.0505447102>
- Mukherjee, S. K. (1972). Origin of mango (*Mangifera indica*). *Economic Botany*, 26(3), 260–264. <https://doi.org/10.1007/BF02861039>
- Murthy, H. N., Dandin, V. S., Dalawai, D., Park, S.-Y., & Paek, K.-Y. (2018). Breeding of *Garcinia* spp. In J. M. Al-Khayri, S. M. Jain, & D. V. Johnson (Eds.), *Advances in Plant Breeding Strategies: Fruits* (pp. 773–809). Springer International Publishing. https://doi.org/10.1007/978-3-319-91944-7_19
- National Research Council. (1975). *Underexploited Tropical Plants with Promising Economic Value*. National Academy of Sciences.
- Nazre, S. (2014). New evidence on the origin of mangosteen (*Garcinia mangostana* L.) based on morphology and ITS sequence. *Genetic Resources and Crop Evolution*, 61(6), 1147–1158. <https://doi.org/10.1007/s10722-014-0097-2>
- Nazre, S., Newman, M. F., Pennington, R. T., & Middleton, D. J. (2018). Taxonomic revision of *Garcinia* Section *Garcinia* (Clusiaceae). *Phytotaxa*, 373(1), 1–52. <https://doi.org/10.11646/phytotaxa.373.1.1>
- Ngugi, K., & Aluka, P. (2019). Genetic and Phenotypic Diversity of Robusta Coffee (*Coffea canephora* L.). In A. M. Grumezescu & A. M. Holban (Eds.), *Caffeinated and Cocoa Based Beverages* (pp. 89–130). Woodhead Publishing. <https://doi.org/10.1016/B978-0-12-815864-7.00003-9>
- Osman, M., & Milan, A. R. (2006). *Mangosteen Garcinia mangostana* L. (J. T. Williams, R. W. Smith, N. Haq, & Z. Dunsiger, Eds.). Southampton Centre for Underutilised Crops, University of Southampton.
- Ovalle-Magallanes, B., Eugenio-Pérez, D., & Pedraza-Chaverri, J. (2017). Medicinal properties of mangosteen (*Garcinia mangostana* L.): A comprehensive update. *Food and Chemical Toxicology*, 109, 102–122. <https://doi.org/10.1016/j.fct.2017.08.021>
- Paz, V. (2005). Rock Shelters, Caves, and Archaeobotany in Island Southeast Asia. *Asian Perspectives*, 44(1), 107–118.
- Ramírez-Mosqueda, M. A. (2022). Overview of Somatic Embryogenesis. *Methods in Molecular Biology (Clifton, N.J.)*, 2527, 1–8. https://doi.org/10.1007/978-1-0716-2485-2_1
- Richards, A. J. (1990). Studies in *Garcinia*, dioecious tropical forest trees: The origin of the mangosteen (*G. Mangostana* L.). *Botanical Journal of the Linnean Society*, 103(4), 301–308. <https://doi.org/10.1111/j.1095-8339.1990.tb00191.x>

- Ridley, H. N. (1922). Guttiferae. In *Flora of Malay Peninsula* (Vol. 1, pp. 166–192). L. Reeve & Co.
- Rimlinger, A., Carrière, S. M., Avana, M.-L., Nguengang, A., & Duminil, J. (2019). The Influence of Farmers' Strategies on Local Practices, Knowledge, and Varietal Diversity of the Safou Tree (*Dacryodes edulis*) in Western Cameroon. *Economic Botany*, 73(2), 249–264. <https://doi.org/10.1007/s12231-019-09455-2>
- Rimlinger, A., Marie, L., Avana, M.-L., Bouka, G. U. D., Zekraoui, L., Mariac, C., Carrière, S. M., & Duminil, J. (2020). New microsatellite markers for *Dacryodes edulis* (Burseraceae), an indigenous fruit tree species from Central Africa. *Molecular Biology Reports*, 47(3), 2391–2396. <https://doi.org/10.1007/s11033-020-05270-4>
- Roxburgh, W. (1832). *Garcinia*. In *Flora Indica* (Vol. 2, pp. 618–630).
- Santoso, P. J., Mansyah, E., & Pancoro, A. (2019). Klarifikasi Keanggotaan Manggis 'Malinau' Dalam Species *Garcinia mangostana* Menggunakan Analisis Filogenik. *Jurnal Pertanian Tropik*, 6(1), 42–48. <https://doi.org/10.32734/jpt.v6i1.3038>
- Saw, L. G., LaFrankie, J. V., Kochummen, K. M., & Yap, S. K. (1991). Fruit trees in a Malaysian rain forest. *Economic Botany*, 45(1), 120–136. <https://doi.org/10.1007/BF02860057>
- Smýkal, P., Nelson, M. N., Berger, J. D., & Von Wettberg, E. J. B. (2018). The Impact of Genetic Changes during Crop Domestication. *Agronomy*, 8(7), Article 7. <https://doi.org/10.3390/agronomy8070119>
- Soltis, D. E., Soltis, P. S., & Milligan, B. G. (1992). Intraspecific Chloroplast DNA Variation: Systematic and Phylogenetic Implications. In P. S. Soltis, D. E. Soltis, & J. J. Doyle (Eds.), *Molecular Systematics of Plants* (pp. 117–150). Springer US. https://doi.org/10.1007/978-1-4615-3276-7_6
- Spiegel-Roy, P. (1986). Domestication of Fruit Trees. In C. Barigozzi (Ed.), *The origin and domestication of cultivated plants* (Vol. 16, pp. 201–211). Elsevier. <https://doi.org/10.1016/B978-0-444-42703-8.50017-8>
- Sprecher, A. (1919a). Étude sur la semence et al germination du *Garcinia mangostana* L. *Revue Generale de Botanique*, 31, 513–531.
- Sprecher, A. (1919b). Étude sur la semence et al germination du *Garcinia mangostana* L. (fin). *Revue Generale de Botanique*, 31, 609–633.
- Sweeney, P. W. (2008). Phylogeny and floral diversity in the genus *Garcinia* (Clusiaceae) and relatives. *International Journal of Plant Sciences*, 169(9), 1288–1303. <https://doi.org/10.1086/591990>
- Theophrastus. (1916). *Enquiry into Plants, Volume I: Books 1-5*: (A. F. Hort, Trans.; Vol. 1). Harvard University Press.

- Thomas, S. C. (1997). Geographic parthenogenesis in a tropical forest tree. *American Journal of Botany*, 84(8), 1012–1015.
- Tixier, P. (1960). Données cytologiques sur quelques Guttiférales récoltées au Laos. *Revue de Cytologie et de Biologie Végétales*, 22, 65–70.
- Vavilov, N. I., & Dorofeev, V. F. (1992). *Origin and Geography of Cultivated Plants*. Cambridge University Press.
- Verheij, E. W. M., & Coronel, R. E. (1991). *Edible Fruits and Nuts*. Pudoc.
- Voris, H. K. (2000). Maps of Pleistocene sea levels in Southeast Asia: Shorelines, river systems and time durations. *Journal of Biogeography*, 27(5), 1153–1167. <https://doi.org/10.1046/j.1365-2699.2000.00489.x>
- Warschefsky, E. J., & von Wettberg, E. J. B. (2019). Population genomic analysis of mango (*Mangifera indica*) suggests a complex history of domestication. *New Phytologist*, 222(4), 2023–2037. <https://doi.org/10.1111/nph.15731>
- Welzen, P. C. van, Parnell, J. A. N., & Slik, J. W. F. (2011). Wallace's Line and plant distributions: Two or three phytogeographical areas and where to group Java?: WALLACE'S LINE AND PLANT DISTRIBUTIONS. *Biological Journal of the Linnean Society*, 103(3), 531–545. <https://doi.org/10.1111/j.1095-8312.2011.01647.x>
- Wiersum, K. F. (2008). Domestication of trees or forests: Development pathways for fruit tree production in South-east Asia. In *Indigenous fruit trees in the tropics; domestication, utilization and commercialization* (pp. 70–83). CABI. <https://research.wur.nl/en/publications/domestication-of-trees-or-forests-development-pathways-for-fruit->
- WMO. (2023). *World Weather Information Service*. World Weather Information Service. <https://worldweather.wmo.int/en/city.html?cityId=310>
- Yao, T. L., Nazre, M., McKey, D., Jalonen, R., & Duminil, J. (2023). The origin of cultivated mangosteen (*Garcinia mangostana* L. var. *mangostana*): Critical assessments and an evolutionary-ecological perspective. *Ecology and Evolution*, 13(3), e9792. <https://doi.org/10.1002/ece3.9792>
- Yogom, B. T., Avana-Tientcheu, M.-L., Monthé, F. K., Bissiengou, P., Loumeto, J. J., Zekraoui, L., Hervouet, C., Fonkou, T., Mariac, C., & Duminil, J. (2023). Genetic diversity and structure in wild and cultivated populations of an emblematic African tree species, *Garcinia kola* (Clusiaceae). *Tree Genetics & Genomes*, 19(5), 39. <https://doi.org/10.1007/s11295-023-01614-w>
- Yue, X., Zheng, X., Zong, Y., Jiang, S., Hu, C., Yu, P., Liu, G., Cao, Y., Hu, H., & Teng, Y. (2018). Combined Analyses of Chloroplast DNA Haplotypes and Microsatellite Markers Reveal New Insights Into the Origin and Dissemination Route of Cultivated Pears Native to East Asia. *Frontiers in Plant Science*, 9. <https://www.frontiersin.org/articles/10.3389/fpls.2018.00591>

- Zeder, M. A., Emshwiller, E., Smith, B. D., & Bradley, D. G. (2006). Documenting domestication: The intersection of genetics and archaeology. *Trends in Genetics*, 22(3), 139–155. <https://doi.org/10.1016/j.tig.2006.01.007>
- Zerega, N. J. C., Ragone, D., & Motley, T. J. (2004). Complex origins of breadfruit (*Artocarpus altilis*, Moraceae): Implications for human migrations in Oceania. *American Journal of Botany*, 91(5), 760–766. <https://doi.org/10.3732/ajb.91.5.760>
- Zohary, D., & Spiegel-Roy, P. (1975). Beginnings of Fruit Growing in the Old World. *Science*, 187(4174), 319–327. <https://doi.org/10.1126/science.187.4174.319>



References

- Abdullah, N. A. P., A. J. Richards, and K. Wolff. 2012. Molecular evidence in identifying parents of *Garcinia mangostana* L. *Pertanika Journal of Tropical Agricultural Science* 35(2):257–70.
- Abu Bakar, S., Sampathrajan, S., Loke, K.-K., Goh, H.-H., and Mohd Noor, N. 2016. DNA-seq analysis of *Garcinia mangostana*. *Genomics Data*, 7, 62–63. <https://doi.org/10.1016/j.gdata.2015.11.018>
- Almeyda, N. and F. M. Martin. 1976. Cultivation of neglected tropical fruits with promise: Part 1. The Mangosteen. Agricultural Research Service, USDA.
- Anderson, T. 1874. Guttiferae. Pp. 258–78 in *Flora of British India*. Vol. 1, pt. 2, edited by J. D. Hooker. London: L. Reeve & Co.
- Ashton, P. S. 1988. *Manual of the Non-Dipterocarp Trees of Sarawak*, Volume II. Kuala Lumpur: Dewan Bahasa dan Pustaka.
- Baleshwar Sharma, P.H., P. J. Handique, and H. Sunitibala Devi. 2013. A historical and taxonomic overview of *Garcinia* L. and its reproductive ecology. *Folia Malaysiana*, 14(1), 63–76.
- Balloux, F., L. Lehmann, and T. de Meeüs. 2003. The population genetics of clonal and partially clonal diploids. *Genetics* 164(4):1635–44.
- Bauhin, C. 1623. *Pinax Theatri Botanici*. Basileae Helvet: Sumptibus & typis Ludovici Regis.
- Bauhin, J., D. Chabrey, J. H. Cherler, and F. L. von Graffenried. 1650. *Historia Plantarum Universalis, Nova, et Absolutissima: cum Consensu et Dissensu circa Eas*. Vol. 1. Ebroduni [Yverdon].
- Beze, C. de. 1692. De quelques arbres et de quelques plantes de Malaque. *Memoires de Mathematique & de Physique de l'academie Royale des Sciences de Paris* 433–41.
- Bondt, J. de. 1658. *Historiae Naturalis & Medicae Indiae Orientalis Libri Sex: Commentarii, Quos Auctor, Morte in Indiis Praeventus, Indigestos Reliquit, a Gvlielmo Pisone. Apud Ludovicum et Danielelem Elzevirios*.
- Burkill, I. H. 1966. *Garcinia*. Pp. 1063–74 in *A Dictionary of the Economic Products of the Malay Peninsula*. Vol. 1 (A-H). Kuala Lumpur: Governments of Malaysia and Singapore by the Ministry of Agriculture and cooperatives.
- Cardim, A. F. 1645. *Relation de la province du Japon*. Adrien Quinqué.
- Carman, J. G. 1997. Asynchronous expression of duplicate genes in Angiosperms may cause apomixis, bispory, tetraspory, and polyembryony. *Biological Journal of the Linnean Society* 61(1):51–94. doi: 10.1111/j.1095-8312.1997.tb01778.x.

- Chennaveeraiah, M. S. and M. K. Razdan. 1975. The basic chromosome number in the genus *Garcinia* L. (Guttiferae). Proceedings of the Indian Science Congress 62:127.
- Chennaveeraiah, M. S. and M. K. Razdan. 1980. Karyomorphological and phytochemical studies in evaluating species relationships in *Garcinia* Linn. and systematic position of *G. xanthochymus* complex. Journal of the Indian Botanical Society 59:251–62.
- Chevalier, A. 1919. Les cultures fruitieres en Indochine. Bulletin Agricole de l'institut Scientifique de Saigon 1(4):97–111.
- Clement, C. R. (1999). 1492 and the loss of Amazonian crop genetic resources. I. The relation between domestication and human population decline. Economic Botany, 53(2), 188–202. <https://doi.org/10.1007/BF02866498>
- Clusius, C. 1605. Exoticorum Libri Decem: Quibus Animalium, Plantarum, Aromatum, Aliorumque Peregrinorum Fructuum Historiae Describuntur; Item Petri Belloni Observationes, Eodem Carolo Clusio Interprete ex officina Plantiniana Raphelengii.
- Corner, E. J. H. 1952. Wayside Trees of Malaya. Vol. 1. 2nd ed. Kuala Lumpur: Malayan Nature Society.
- Corner, E. J. H. 1997. Wayside Trees of Malaya. Vol. 1. 4th ed. Kuala Lumpur: Malayan Nature Society.
- Cruz, F.S.D.J. 2001. Status report on genetic resources of Mangosteen (*Garcinia mangostana* L.) in Southeast Asia. IPGRI Office for South Asia, Delhi.
- Dahlgren, B. E. 1947. Tropical and Subtropical Fruits. Chicago: Chicago Natural History Museum.
- Davies, S. J., S. Tan, J. V. LaFrankie, and M. D. Potts. 2005. Soil-related floristic variation in a hyperdiverse dipterocarp forest. Pp. 22–34 in pollination ecology and the rain forest: Sarawak studies, ecological studies, edited by D. W. Roubik, S. Sakai, and A. A. Hamid Karim. New York: Springer.
- Dewan Bahasa dan Pustaka. 2021. Carian umum. Retrieved on 8 January 2023 (<https://prpm.dbp.gov.my/cari1?keyword=dusun>).
- Engler, A. 1893. Guttiferae. In: Engler, A. & Prantl, K.A.E. (Eds.) Naturlichen pflanzenfamilien. Engelman, Leipzig, pp. 194–242.
- Engler, A. 1925. Guttiferae. In: Engler, A. & Prantl, K.A.E. (Eds.) Naturlichen pflanzenfamilien 2nd. ed. Engelman, Leipzig, pp. 154–237.
- Fairchild, D. 1915. The mangosteen. The Journal of Heredity 6(8):339–47.
- Garcia, S., Kovařík, A., Leitch, A. R., and Garnatje, T. 2017. Cytogenetic features of rDNA genes across land plants: analysis of the Plant rDNA database. The Plant Journal 89(5):1020–1030. doi: 10.1111/tpj.13442

- Garcin, L. 1733. The settling of a new genus of plants, called after the Malayans, mangostans; by Laurentius Garcin, M. D. and F. R. S. Translated from the French by Mr. Zollman, F. R. S. Philosophical Transactions of the Royal Society of London 38(431):232–42. doi: 10.1098/rstl.1733.0038.
- Gianno, R. and K. J. Bayr. 2009. Semelai agricultural patterns: toward an understanding of variation among indigenous cultures in southern Peninsular Malaysia. *Journal of Southeast Asian Studies* 40(1):153–85.
- Gomperts, A., Arnoud Haag, and P. Carey. 2012. Mapping Majapahit: Wardenaar's archaeological survey at Trowulan in 1815'. *Indonesia* 93:177. doi: 10.5728/indonesia.93.0177.
- Gong, Z. 2019 [originally published in 1434]. 《西洋番国志 校注本》(Collated and annotated Xī Yáng Fān Guó Zhì [1434], The Overall Survey of the Ocean's Shores). edited by D. Xiang. Beijing: Hua Wen Publisher.
- Grimanelli, D., O. Leblanc, E. Perotti, and U. Grossniklaus. 2001. Developmental genetics of gametophytic apomixis. *Trends in Genetics* 17(10):597–604. doi: 10.1016/S0168-9525(01)02454-4.
- Ha, C. O. 1978. Embryological and cytological aspects of the reproductive biology of some understorey rainforest trees. Ph.D. Thesis, University of Malaya.
- Ha, C. O., V. E. Sands, E. Soepadmo, and K. Jong. 1988. Reproductive patterns of selected understorey trees in the Malaysian rain forest: the apomictic species. *Botanical Journal of the Linnean Society* 97(3):317–31. doi: 10.1111/j.1095-8339.1988.tb01586.x.
- Halkett, F., J. C. Simon, and F. Balloux. 2005. Tackling the population genetics of clonal and partially clonal organisms. *Trends in Ecology & Evolution* 20(4):194–201. doi: 10.1016/j.tree.2005.01.001.
- Hambali, G. G. and A. Natawijaya. 2016. A preliminary study of *Garcinia malaccensis* collection in Taman Buah Mekarsari, Cileungsi, Bogor, Indonesia. Pp. 31–36 in *Contribution of Breeding Research for Sustainable Agricultural Production under Changing Environment for Food Security in Asia and Oceania*. Bogor: Penerbit IPB Press.
- Hand, M. L. and A. M. G. Koltunow. 2014. The genetic control of apomixis: asexual seed formation. *Genetics* 197(2):441–50. doi: 10.1534/genetics.114.163105.
- Hardy, O.J., Charbonnel, N., Fréville, H. and Heuertz, M. 2003. Microsatellite allele sizes: a simple test to assess their significance on genetic differentiation. *Genetics* 163(4):1467–1482. doi: 10.1093/genetics/163.4.1467
- Harlan, J. R. 1992. *Crops and Man*. Madison, Wis., USA: American Society of Agronomy: Crop Science Society of America.
- He, P., L. Li, L. Cheng, H. Wang, and Y. Chang. 2018. Variation in ploidy level and morphological traits in the progeny of the triploid apple variety Jonagold. *Czech*

- Journal of Genetics and Plant Breeding 54(No. 3):135–42. doi: 10.17221/201/2016-CJGPB.
- Hojsgaard, D. and Hörandl, E. 2019. The rise of apomixis in natural plant populations. *Frontiers in Plant Science* 10. doi: 10.3389/fpls.2019.00358
- Horn, C. L. 1940. Existence of only one variety of cultivated mangosteen explained by asexually formed “seed”. *Science* 92(2385):237–38. doi: 10.1126/science.92.2385.237.
- Hörandl, E. and Hojsgaard, D. 2012. The evolution of apomixis in angiosperms: A reappraisal. *Plant Biosystems* 146(3):681–693.
- Idris, S. and A. Rukayah. 1987. Description of the male mangosteen (*Garcinia mangostana* L.) discovered in Peninsular Malaysia. *MARDI Research Bulletin* 15(1):63–66.
- Jones, S. W. 1980. Morphology and major taxonomy of *Garcinia* (Guttiferae). Thesis, University of Leicester.
- King, G. 1890. VIII. Materials for a Flora of Malayan Peninsula. *The Journal of the Asiatic Society of Bengal* 59(2):113–206.
- Kirst, M., C. M. Cordeiro, G. D. S. P. Rezende, and D. Grattapaglia. 2005. Power of microsatellite markers for fingerprinting and parentage analysis in *Eucalyptus grandis* breeding populations. *Journal of Heredity* 96(2):161–66. doi: 10.1093/jhered/esi023.
- Kochummen, K. M. and T. C. Whitmore. 1973. Notes on the systematy of Malayan phanerogams XVII-XXII. *Gardens’ Bulletin Singapore* 26(2):269–87.
- Krishnaswamy, N. and V. S. Raman. 1949. A note on the chromosome numbers of some economic plants of India. *Current Science* 10:376–78.
- Lamb, A. 2019. *A Guide to Wild Fruits of Borneo*. edited by L. Neo and K. Lyons. Kota Kinabalu, Sabah: Natural History Publications (Borneo).
- Landis, J. B., D. E. Soltis, Z. Li, H. E. Marx, M. S. Barker, D. C. Tank, and P. S. Soltis. 2018. Impact of whole-genome duplication events on diversification rates in Angiosperms. *American Journal of Botany* 105(3):348–63. doi: 10.1002/ajb2.1060.
- León, J. 1982. *Genetic Resources of Tropical Crops*. Turrialba: Biblioteca Orton IICA / CATIE.
- Léotard, G., Debout, G., Dalecky, A., Guillot, S., Gaume, L., McKey, D. and Kjellberg, F. 2009. Range expansion drives dispersal evolution in an equatorial three-species symbiosis. *PLoS One* 4(4). doi: 10.1371/journal.pone.0005377.
- Lim, A. L. 1984. The embryology of *Garcinia mangostana* L. (Clusiaceae). *Gardens’ Bulletin Singapore* 37(1):93–103.

- Lim T.K. (2012) *Garcinia mangostana*. In: Edible medicinal and non-medicinal plants. Springer, Dordrecht, pp 83–108.
- Linnaeus, C. 1753. Species Plantarum: Exhibentes Plantas Rite Cognitas Ad Genera Relatas, Cum Differentiis Specificis, Nominibus Trivialibus, Synonymis Selectis, Locis Natalibus, Secundum Systema Sexuale Digestas. Vol. 1. Stockholm: L. Salvius.
- Linnaeus, C. 1754. Genera Plantarum. Ed. 5th. Holmiæ (Stockholm): Impensis Laurentii Salvii.
- Linschoten, J. H. van, A. C. Burnell, and P. A. Tiele. 1885. The voyage of John Huyghen van Linschoten to the East Indies: from the old English translation of 1598: the first book, containing his description of the East. London: Printed for the Hakluyt Society.
- Ma, Huan. 2019 [originally published in 1416]. 《瀛涯勝覽 校注本》 (Collated and annotated Yíng Yá Shèng Lǎn [1416], 'The Overall Survey of the Ocean's Shores'). edited by C. Feng. Beijing: Hua Wen Publisher.
- Mandelslo, J. A. de. 1669. The Travels of John Albert de Mandelslo (a Gentleman belonging to the Embassy) from Persia, into the East-Indies. Vol. 2. London: Printed for John Starkey, and Thomas Basset.
- Mansyah, E., A. Baihaki, R. Setiamihardja, Juliati S. Darsa, and Sobir. 2003. Analisis variabilitas genetik manggis (*Garcinia mangostana* L.) di Jawa dan Sumatera Barat menggunakan teknik RAPD'. Zuriat 14(1):35–44. doi: 10.24198/zuriat.v14i1.6813.
- Matra, D. D., R. Poerwanto, E. Santosa, Sobir, H. Higashio, H. Anzai, and E. Inoue. 2016. Analysis of allelic diversity and genetic relationships among cultivated mangosteen (*Garcinia mangostana* L.) in Java, Indonesia using microsatellite markers and morphological characters. Tropical Plant Biology 9(1):29–41. doi: 10.1007/s12042-016-9161-8.
- McConkey, K. R., W. Y. Brockelman, C. Saralamba, and A. Nathalang. 2015. Effectiveness of primate seed dispersers for an “oversized” fruit, *Garcinia benthamii*. Ecology 96(10):2737–47. doi: 10.1890/14-1931.1.
- McKey, D., M. Elias, B. Pujol, and A. Duputié. 2010. The evolutionary ecology of clonally propagated domesticated plants. New Phytologist 186(2):318–32. doi: 10.1111/j.1469-8137.2010.03210.x.
- Michon, G., F. Mary, and J. Bompard. 1986. Multistoried agroforestry garden system in West Sumatra, Indonesia. Agroforestry Systems 4(4):315–38. doi: 10.1007/BF00048106.
- Midin, M. R. 2018. Cytogenetics study of *Garcinia mangostana*. Ph.D. Thesis, International Islamic University Malaysia.
- Midin, M. R., M. S. Nordin, M. Madon, M. Nazre, H. H. Goh, and M. N. Normah. 2018. Determination of the chromosome number and genome size of *Garcinia*

- mangostana* L. via cytogenetics, flow cytometry and k-mer analyses. *Caryologia* 71(1):35–44. doi: 10.1080/00087114.2017.1403762.
- Murthy, H. N., Dandin, V. S., Dalawai, D., Park, S.-Y., and Paek, K.-Y. 2018. Breeding of *Garcinia* spp. In J. M. Al-Khayri, S. M. Jain, & D. V. Johnson (Eds.), *Advances in Plant Breeding Strategies: Fruits* (pp. 773–809). Springer International Publishing. doi: 10.1007/978-3-319-91944-7_19
- Nazre, M. 2006. Taxonomic revision and molecular studies of *Garcinia* sect. *Garcinia* (Guttiferae). Thesis, University of Edinburgh.
- Nazre, M. 2010. Historical review and notes on the correct scientific name for seashore mangosteen. *Genetic Resources and Crop Evolution* 57(8):1249–59. doi: 10.1007/s10722-010-9588-y.
- Nazre, M. 2014. New evidence on the origin of mangosteen (*Garcinia mangostana* L.) based on morphology and ITS sequence. *Genetic Resources and Crop Evolution* 61(6):1147–58. doi: 10.1007/s10722-014-0097-2.
- Nazre, M., M. F. Newman, R. T. Pennington, and D. J. Middleton. 2018. Taxonomic revision of *Garcinia* sect. *Garcinia* (Clusiaceae). *Phytotaxa* 373(1):1–52. doi: 10.11646/phytotaxa.373.1.1.
- Orta, G. de, C. McKew Parr, R. Parr, and F. M. de Melo Ficalho. 1913 [originally published in 1563]. *Colloquies on the simples & drugs of India*. London, H. Sotheran and co.
- Orwa, C., Mutua, A., Kindt, R., Jamnadass, R., and Anthony, S. 2009. *Garcinia mangostana* http://apps.worldagroforestry.org/treedb2/AFTPDFS/Garcinia_mangostana.PD F. Retrieved on 27 December 2022.
- Osman, M. and A. R. Milan. 2006. *Mangosteen (Garcinia mangostana L.)* edited by J. T. Williams, R. W. Smith, N. Haq, and Z. Dunsiger. Southampton: Southampton Centre for Under-utilised Crops, University of Southampton.
- Othman, Y. and H. D. Tindall. 1995. *Mangosteen cultivation*. Vol. 129. Rome: Food and Agriculture Organization of the United Nations.
- Pierre, J. B. L. 1882. *Flore Forestiere de La Cochinchine* [4], plates 49-64 in Fasc. 4. Paris: O. Doin.
- Ramage, C. M., L. Sandø, C. P. Peace, B. J. Carroll, and R. A. Drew. 2004. Genetic diversity revealed in the apomictic fruit species *Garcinia mangostana* L. (mangosteen). *Euphytica* 136(1):1–10. doi: 10.1023/B:EUPH.0000019456.06040.eb.
- Ramsey, J. and T. S. Ramsey. 2014. Ecological studies of polyploidy in the 100 years following its discovery. *Philosophical Transactions of the Royal Society B: Biological Sciences* 369(1648):20130352. doi: 10.1098/rstb.2013.0352.

- Renner, S. S. 2014. The relative and absolute frequencies of angiosperm sexual systems: dioecy, monoecy, gynodioecy, and an updated online database. *American Journal of Botany* 101(10):1588–96. doi: 10.3732/ajb.1400196.
- Richards, A. J. 1990a. Studies in *Garcinia*, dioecious tropical forest trees: agamospermy. *Botanical Journal of the Linnean Society* 103(3):233–50. doi: 10.1111/j.1095-8339.1990.tb00186.x.
- Richards, A. J. 1990b. Studies in *Garcinia*, dioecious tropical forest trees: the origin of the mangosteen (*G. mangostana* L.). *Botanical Journal of the Linnean Society* 103(4):301–8. doi: 10.1111/j.1095-8339.1990.tb00191.x.
- Richards, A. J. 1996. Breeding systems in flowering plants and the control of variability. *Folia Geobotanica & Phytotaxonomica* 31(3):283–93.
- Richards, A. J. 2003. Apomixis in flowering plants: an overview. *Philosophical Transactions: Biological Sciences* 358(1434):1085–93.
- Ridley, H. N. 1893. On the flora of the Eastern Coast of the Malay Peninsula. *Transactions of the Linnean Society of London. 2nd Series: Botany* 3(9):267–408.
- Ridley, H. N. 1922. Guttiferae. Pp. 166–92 in *Flora of Malay Peninsula*. Vol. 1. London: L. Reeve & Co.
- Roxburgh, W. 1832. *Garcinia*. In *Flora Indica* Vol. 2, pp. 618–630.
- Rumphius, G. E. 2011 [originally published in 1741]. *The Ambonese Herbal*. Vol. 1. Introduction and Book I: Containing all sorts of trees, that bear edible fruits, and are husbanded by people. New Haven: Yale University Press & National Tropical Botanical Garden.
- Samsir, S. A., H. Bunawan, C. Y. Choong, and M. N. Normah. 2016. Dataset of SSR markers for ISSR-Suppression-PCR to detect genetic variation in *Garcinia mangostana* L. in Peninsular Malaysia. *Data in Brief* 8:1438–42. doi: 10.1016/j.dib.2016.08.016.
- Sandø, L., C. Peace, C. Ramage, B. J. Carrol, and R. Drew. 2005. Assessment of genetic diversity in Australian-grown mangosteen (*Garcinia mangostana* L.) and its wild relatives. *Acta Horticulturae* (692):143–52. doi: 10.17660/ActaHortic.2005.692.19.
- Sinaga, S., Sobir, R. Poerwanto, H. Aswidinnoor, and D. Duryadi. 2007. Genetic variability analysis on apomictic mangosteen (*Garcinia mangostana*) in Indonesian and its close related species by using RAPD Markers. *Floribunda* 3(3):77–84.
- Shaharudin, M. A., Zaki, N. M., Saleh, M. N., Mohd Noor, N. and Midin, M. R. 2022. The origin of mangosteen: A review. *Genetic Resources and Crop Evolution* 69(7): 2291–2299. doi: 10.1007/s10722-022-01426-2

- Sobir, S. Sinaga, R. Poerwanto, and R. Lukman. 2009. comparison analysis of genetic diversity of Indonesian mangosteens (*Garcinia mangostana* L.) and related species by means of isozymes and AFLP markers. *BIOVERSITAS* 10(4):163–67.
- Soepadmo, E. 1989. Contribution of reproductive biological studies towards the conservation and development of Malaysian plant genetic resources. Pp. 1–41 in *Genetic resources of under-utilised plants in Malaysia*, edited by A. H. Zakri. Kuala Lumpur: Malaysian National Committee of Plant Genetic Resources.
- Sosef, M. and G. Dauby. 2012. Contribution to the taxonomy of *Garcinia* (Clusiaceae) in Africa, including two new species from Gabon and a key to the Lower Guinean species. *PhytoKeys* 17:41–62. doi: 10.3897/phytokeys.17.3114.
- Sprecher, A. 1919a. Étude sur la semence et la germination du *Garcinia mangostana* L. *Revue Generale de Botanique* 31:513–31.
- Sprecher, A. 1919b. Étude sur la semence et la germination du *Garcinia mangostana* L. (fin). *Revue Generale de Botanique* 31:609–33.
- Steenis-Kruseman, M. J. van. 1950. Chapter VI. Important sources of information used in compiling the list of collectors; special part: alphabetical list of collectors. Pp. 1–606 in *Flora Malesiana*. Vol. 1. Malaysian plant collectors and collections, I, Spermatophyta, edited by C. G. G. J. van Steenis. Djakarta: Noordhoff-Kolff N.V.
- Stevens, P. F. 2001. Angiosperm Phylogeny Website. Version 14, July 2017 [and more or less continuously updated since]. Retrieved on 27 December 2022 (<http://www.mobot.org/MOBOT/research/APweb/welcome.html>).
- Stevens P.F. 2007. Clusiaceae-Guttiferae. In: Kubitzki K. (Ed). *The families and genera of vascular plants IX. Flowering Plants Eudicots*. Springer Verlag, Berlin: 48–66. doi: 10.1007/978-3-540-32219-1_10
- Sulassih, Sobir, and E. Santosa. 2013. Phylogenetic analysis of mangosteen (*Garcinia mangostana* L.) and its relatives based on morphological and Inter Simple Sequence Repeat (ISSR) markers. *SABRAO Journal of Breeding and Genetics* 45(3):478–90.
- Sulistiono, B. 2015. Menelusuri tinggalan arkeologi Kesultanan Samudera Pasai. Bayt Al-Qur'an dan Museum Istiqlal, Jakarta.
- Sweeney, P. W. 2008. Phylogeny and floral diversity in the genus *Garcinia* (Clusiaceae) and relatives. *International Journal of Plant Sciences* 169(9):1288–1303. doi: 10.1086/591990.
- Thomas, S. C. 1997. Geographic parthenogenesis in a tropical forest tree. *American Journal of Botany* 84(8):1012–15.
- Tixier, P. 1960. Données cytologiques sur quelques Guttiférales récoltées au Laos. *Revue de Cytologie et de Biologie Végétales* 22:65–70.

- Treub, M. 1911. Le sac embryonnaire et l'embryon dans les Angiospermes, nouvelle série de recherches. Annales du Jardin botanique de Buitenzorg 24.
- Verheij, E. W. M. 1991. *Garcinia*. Pp. 175–81 in Edible Fruits and Nuts, Plant Resources of South-East Asia, edited by E. W. M. Verheij and R. E. Coronel. Wageningen: Pudoc.
- Wee, C.-C., Nor Muhammad, N. A., Subbiah, V. K., Arita, M., Nakamura, Y., and Goh, H.-H. 2022. Plastomes of *Garcinia mangostana* L. and comparative analysis with other *Garcinia* species. bioRxiv. doi: 10.1101/2022.02.22.481552.
- Whitmore, T. C. 1973. Tree Flora of Malaya. Vol. 2. Kuala Lumpur & London: Longman.
- Yapwattanaphun, C., S. Subhadrabandhu, C. Honsho, and K. Yonemori. 2004. phylogenetic relationship of mangosteen (*Garcinia mangostana*) and several wild relatives (*Garcinia* spp.) revealed by ITS Sequence Data. Journal of the American Society for Horticultural Science 129(3):368–73. doi: 10.21273/JASHS.129.3.0368.
- Yule, H. and A. C. Burnell. 1886. Hobson-Jobson: Being a glossary of Anglo-India Colloquial Words and Phrases, and of Kindred Terms, Etymological, Historical, Geographical and Discursive. London: Murray.
- Zeven, A. C. and J. M. J. de Wet. 1982. Dictionary of Cultivated Plants and Their Regions of Diversity: excluding most Ornamentals, Forest Trees and Lower Plants. 2. ed., rev. Wageningen: Centre for Agricultural Publishing and Documentation.
- Zhang, K., X. Wang, and F. Cheng. 2019. Plant polyploidy: origin, evolution, and its influence on crop domestication. Horticultural Plant Journal 5(6):231–39. doi: 10.1016/j.hpj.2019.11.003.

References

- Abu Bakar S, Sampathrajan S, Loke KK, Goh HH, Mohd Noor N. 2016. DNA-seq analysis of *Garcinia mangostana*. Genomics Data. 7:62–63. <https://doi.org/10.1016/j.gdata.2015.11.018>
- Anderson T. 1874. Guttiferae. In: Hooker JD, editor. Flora of British India. Vol. 1, pt. 2. London: L. Reeve & Co. pp. 258–278.
- Beentje HJ. 2010. The Kew Plant Glossary. An illustrated dictionary of plant terms. London: Kew Publishing.
- Blanco FM. 1837. Flora de Filipinas: Segun el sistema sexual de Linneo. C. Lopez.
- Borkent A. 2021. Diagnosing diagnoses – can we improve our taxonomy? ZooKeys. 1071:43–48. <https://doi.org/10.3897/zookeys.1071.72904>
- Caliński T, Harabasz J. 1974. A dendrite method for cluster analysis. Communications in Statistics. 3(1):1–27. <https://doi.org/10.1080/03610927408827101>
- Chitwood DH, Mullins J, Migicovsky Z, Frank M, VanBuren R, Londo JP. 2021. Vein-to-blade ratio is an allometric indicator of leaf size and plasticity. American Journal of Botany. 108(4):571–579. <https://doi.org/10.1002/ajb2.1639>
- Corner EJH. 1997. *Wayside Trees of Malaya*. 4th ed., Vol. 1. Kuala Lumpur: Malayan Nature Society.
- Dallwitz MJ. 1980. A General System for Coding Taxonomic Descriptions. Taxon. 29(1):41–46. <https://doi.org/10.2307/1219595>
- Dallwitz MJ, Paine TA, Zurcher EJ. 1999. User's guide to the DELTA Editor. Delta-intkey.com.
- Davies SJ. 2005. Soil-related floristic variation in a hyperdiverse dipterocarp forest. In Roubik D, Sakai S, Hamid AA, editors. Pollination ecology and the rain forest: Sarawak studies. Springer-Verlag. pp. 22–34. <https://doi.org/10.1007/b138701>
- Dray S, Dufour AB. 2007. The **ade4** package: implementing the duality diagram for ecologists. Journal of Statistical Software. 22(4). <https://doi.org/10.18637/jss.v022.i04>
- George ST, Latha AKB, Mathew KL, Geetha CK. 1992. Pattern of flowering and flower development in kodapuli (*Garcinia cambogia* Desr.). Indian Cocoa, Arecanut and Spices Journal. 16(2):68–70.
- Goh HH, Abu Bakar S, Kamal Azlan ND, Zainal Z, Mohd Noor N. 2019. Transcriptional reprogramming during *Garcinia*-type recalcitrant seed germination of *Garcinia mangostana*. Scientia Horticulturae. 257:108727. <https://doi.org/10.1016/j.scienta.2019.108727>
- Gower JC. 1971. A general coefficient of similarity and some of its properties. Biometrics. 27(4):857–871. <https://doi.org/10.2307/2528823>

- Ha CO, Sands VE, Soepadmo E, Jong K. 1988. Reproductive patterns of selected understorey trees in the Malaysian rain forest: The apomictic species. *Botanical Journal of the Linnean Society*. 97(3):317–331. <https://doi.org/10.1111/j.1095-8339.1988.tb01586.x>
- Hambali GG, Natawijaya A. 2016. A preliminary study of *Garcinia malaccensis* collection in Taman Buah Mekarsari, Cileungsi, Bogor, Indonesia. *Contribution of Breeding Research for Sustainable Agricultural Production under Changing Environment for Food Security in Asia and Oceania*. Bogor: Penerbit IPB Press. pp: 31–36.
- Hamilton CW, Reichard SH. 1992. Current practice in the use of subspecies, variety, and forma in the classification of wild plants. *Taxon*. 41(3):485–498. <https://doi.org/10.2307/1222819>
- Harlan JR, de Wet MJ. 1971. Toward a rational classification of cultivated plants. *Taxon*. 20(4):509–517. <https://doi.org/10.2307/1218252>
- Joseph KS, Murthy HN. 2015. Sexual system of *Garcinia indica* Choisy: geographic variation in trioecy and sexual dimorphism in floral traits. *Plant Systematics and Evolution*. 301(3):1065–1071.
- Kochummen KM. 1997. *Tree Flora of Pasoh*. Kepong: Forest Research Institute Malaysia.
- Kochummen KM, Whitmore TC. 1973. Notes on the systematy of Malayan phanerogams XVII–XXII. *Gardens' Bulletin Singapore*. 26(2):269–287.
- Le Moguedec G. 2020. R resources improve dendrograms [Data set]. Portail Data INRAE. <https://doi.org/10.15454/DAAUP1>
- Leal DO, Benevides CR, Silva RCP, Santiago-Fernandes LDR, Sá-Haiad B, Lima HA. 2013. *Garcinia brasiliensis*: Insights into reproductive phenology and sexual system in a Neotropical environment. *Plant Systematics and Evolution*. 299(8):1577–1585. <https://doi.org/10.1007/s00606-013-0833-7>
- Legendre P, Legendre LFJ. 1998. *Numerical Ecology*. Elsevier Science.
- Linnaeus C. 1753. *Species plantarum: Exhibentes plantas rite cognitatas ad genera relatas, cum differentiis specificis, nominibus trivialibus, synonymis selectis, locis natalibus, secundum systema sexuale digestas*. Vol. 1. L. Salvius.
- Maechler M, Rousseeuw P, Struyf A, Hubert M, Hornik K. 2022. *Cluster: cluster analysis basics and extensions*. R package version 2.1.4.
- Matra DD, Kozaki T, Ishii K, Poerwanto R, Inoue E. 2019. Comparative transcriptome analysis of translucent flesh disorder in mangosteen (*Garcinia mangostana* L.) fruits in response to different water regimes. *PLOS ONE*. 14(7):e0219976. <https://doi.org/10.1371/journal.pone.0219976>

- Matra DD, Poerwanto R, Santosa E, Sobir, Higashio H, Anzai H, Inoue E. 2016. Analysis of allelic diversity and genetic relationships among cultivated mangosteen (*Garcinia mangostana* L.) in Java, Indonesia using microsatellite markers and morphological characters. *Tropical Plant Biology*. 9(1):29–41. <https://doi.org/10.1007/s12042-016-9161-8>
- Microsoft Corporation. 2018. Microsoft Excel (2019 (16.0)). <https://office.microsoft.com/excel>
- Midin MR, Nordin MS, Madon M, Nazre S, Goh HH, Normah MN. 2018. Determination of the chromosome number and genome size of *Garcinia mangostana* L. via cytogenetics, flow cytometry and k-mer analyses. *Caryologia*. 71(1):35–44. <https://doi.org/10.1080/00087114.2017.1403762>
- Morel J, Le Moguédec G, Munzinger J. 2021. Multivariate morphometric analysis supported by an anatomical approach to assess species delimitation in *Xyris* (Xyridaceae) in New Caledonia. *Botanical Journal of the Linnean Society*. <https://doi.org/10.1093/botlinnean/boaa108>
- Murthy HN, Dandin VS, Dalawai D, Park SY, Paek KY. 2018. Breeding of *Garcinia* spp. In Al-Khayri JM, Jain SM, Johnson DV, editors. *Advances in Plant Breeding Strategies: Fruits*. Springer International Publishing. pp. 773–809. <https://doi.org/10.1007/978-3-319-91944-7>
- Nazre S. 2006. Taxonomic revision and molecular studies of *Garcinia* section *Garcinia* (Guttiferae). PhD Thesis. University of Edinburgh.
- Nazre S. 2014. New evidence on the origin of mangosteen (*Garcinia mangostana* L.) based on morphology and ITS sequence. *Genetic Resources and Crop Evolution*. 61(6):1147–1158. <https://doi.org/10.1007/s10722-014-0097-2>
- Nazre S, Newman MF, Pennington RT, Middleton DJ. 2018. Taxonomic revision of *Garcinia* Section *Garcinia* (Clusiaceae). *Phytotaxa*. 373(1):1–52. <https://doi.org/10.11646/phytotaxa.373.1.1>
- Oksanen J, Simpson GL, Guillaume Blanchet F, Kindt R, Legendre P, Minchin PR, O'Hara RB, Solymos P, Stevens MHH, Szoecs E, Wagner H, Barbour M, Bedward M, Bolker B, Borcard D, Carvalho G, Chirico M, De Caceres M, Durand S, ... Weedon J. 2022. *Vegan: Community Ecology Package*. R package version 2.6-2. <https://CRAN.R-project.org/package=vegan>
- Pangsuban S, Bamroongrugs N, Kanchanapoom K, Nualsri C. 2007. An evaluation of the sexual system of *Garcinia atroviridis* L. (Clusiaceae), based on reproductive features. *Songklanakarin Journal of Science and Technology*. 29(6):13.
- Pierre AH, Le Moguédec G, Lowry PP, Munzinger J. 2014. Multivariate morphometric analysis and species delimitation in the endemic New Caledonian genus *Storthocalyx* (Sapindaceae): morphometric analysis of *Storthocalyx*. *Botanical Journal of the Linnean Society*. <https://doi.org/10.1111/boj.12199>
- Pierre JBL. 1883. *Flore forestiere de la Cochinchine* Fasc. 6:plates 84–96. O. Doin.

- Pipoly JJ. 1987. A systematic revision of the genus *Cybianthus*, subgenus *Grammadenia* (Myrsinaceae). New York: New York Botanical Garden.
- Posit team. 2022. RStudio: integrated development environment for R. Posit Software, PBC. <http://www.posit.co>
- R Core Team. 2021. R: A language and environment for statistical computing. R Foundation for Statistical Computing. <https://www.R-project.org/>
- Schneider CA, Rasband WS, Eliceiri KW. 2012. NIH Image to ImageJ: 25 years of image analysis. *Nature Methods*. 9(7):671–675. <https://doi.org/10.1038/nmeth.2089>
- Soepadmo E. 1989. Contribution of reproductive biological studies towards the conservation and development of Malaysian plant genetic resources. In Zakri AH, editor. Genetic resources of under-utilised plants in Malaysia. Kuala Lumpur: Malaysian National Committee of Plant Genetic Resources. pp. 1–41.
- Sosef M, Dauby G. 2012. Contribution to the taxonomy of *Garcinia* (Clusiaceae) in Africa, including two new species from Gabon and a key to the Lower Guinean species. *PhytoKeys*. 17:41–62. <https://doi.org/10.3897/phytokeys.17.3114>
- Sprecher A. 1919. Étude sur la semence et al germination du *Garcinia mangostana* L. (fin). *Revue Generale de Botanique*. 31:609–633.
- Steenis-Kruseman MJ van. 1950. Chapter VI. Important sources of information used in compiling the list of collectors; special part: alphabetical list of collectors. In Steenis CGGJ van, editor. *Flora Malesiana*. Serie 1, vol. 1. Noordhoff-Kolff N.V. pp. 1–606.
- Stevens PF. 2007. Clusiaceae-Guttiferae. In Kubitzki K, editor. Flowering Plants, Eudicots: Berberidopsidales, Buxales, Crossosomatales, Fabales p.p., Geraniales, Gunnerales, Myrtales p.p., Proteales, Saxifragales, Vitales, Zygophyllales, Clusiaceae Alliance, Passifloraceae Alliance, Dilleniaceae, Huaceae, Picramniaceae, Sabiaceae. Springer. pp. 48–66. https://doi.org/10.1007/978-3-540-32219-1_10
- Stevens PF. 2001 onwards. Angiosperm Phylogeny Website. Version 14, July 2017 [and more or less continuously updated since]. <http://www.mobot.org/MOBOT/research/APweb/welcome.html>
- Thiers BM. 2023. Index Herbariorum: a global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium. http://sweetgum.nybg.org/science/ih/?_ga=2.67792151.720125219.1601879821-1637336645.1601879821
- Thomas SC. 1997. Geographic parthenogenesis in a tropical forest tree. *American Journal of Botany*. 84(8):1012–1015.
- van den Berg ME. 1979. Revisão das espécies brasileiras do gênero *Rheedia* L. (Guttiferae). *Acta Amazonica*. 9(1):43–74.

- Vichiato MR de M, Vichiato M, Pasqual M, Rodrigues FA, Castro DM de. 2014. Morphological effects of induced polyploidy in *Dendrobium nobile* Lindl. (Orchidaceae). *Crop Breeding and Applied Biotechnology*. 14:154–159. <https://doi.org/10.1590/1984-70332014v14n3a23>
- Ward JH. 1963. Hierarchical grouping to optimize an objective function. *Journal of the American Statistical Association*. 58(301):236–244. <https://doi.org/10.2307/2282967>
- Wee CC, Nor Muhammad NA, Subbiah VK, Arita M, Nakamura Y, Goh HH. 2022. Mitochondrial genome of *Garcinia mangostana* L. variety mesta. *Scientific Reports*. 12:9480. <https://doi.org/10.1038/s41598-022-13706-z>
- Wee CC, Nor Muhammad NA, Subbiah VK, Arita M, Nakamura Y, Goh HH. 2023. Plastomes of *Garcinia mangostana* L. and comparative analysis with other *Garcinia* species. *Plants*. 12(4). <https://doi.org/10.3390/plants12040930>
- Whitmore TC. 1973. *Tree Flora of Malaya*. Vol. 2. Kuala Lumpur: Longman.
- Yao TL, Nazre M, McKey D, Jalonen R, Duminil J. 2023. The origin of cultivated mangosteen (*Garcinia mangostana* L. var. *mangostana*): critical assessments and an evolutionary-ecological perspective. *Ecology and Evolution*. 13(3). <https://doi.org/10.1002/ece3.9792>
- Yapwattanaphun C, Subhadrabandhu S, Honsho C, Yonemori K. 2004. Phylogenetic relationship of mangosteen (*Garcinia mangostana*) and several wild relatives (*Garcinia* spp.) revealed by its sequence data. *Journal of the American Society for Horticultural Science*. 129(3):368–373. <https://doi.org/10.21273/JASHS.129.3.0368>

References

- Abdullah, N. A. P., Richards, A. J., & Wolff, K. (2012). Molecular evidence in identifying parents of *Garcinia mangostana* L. *Pertanika Journal of Tropical Agricultural Science*, 35(2), 257–270.
- Baucom, R. S., Estill, J. C., & Cruzan, M. B. (2005). The effect of deforestation on the genetic diversity and structure in *Acer saccharum* (Marsh): Evidence for the loss and restructuring of genetic variation in a natural system. *Conservation Genetics*, 6(1), 39.
- Charif, D., & Lobry, J. R. (2007). SeqinR 1.0-2: A Contributed Package to the R Project for Statistical Computing Devoted to Biological Sequences Retrieval and Analysis. In U. Bastolla, M. Porto, H. E. Roman, & M. Vendruscolo (Eds.), *Structural Approaches to Sequence Evolution: Molecules, Networks, Populations* (pp. 207–232). Springer. https://doi.org/10.1007/978-3-540-35306-5_10
- Clement, M., Posada, D., & Crandall, K. A. (2000). TCS: A computer program to estimate gene genealogies. *Molecular Ecology*, 9(10), 1657–1659. <https://doi.org/10.1046/j.1365-294x.2000.01020.x>
- Cubry, P. (2017). *VCF2fasta_dev2* [R]. IRD.
- Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., Handsaker, R. E., Lunter, G., Marth, G. T., Sherry, S. T., McVean, G., Durbin, R., & 1000 Genomes Project Analysis Group. (2011). The variant call format and VCFtools. *Bioinformatics*, 27(15), 2156–2158. <https://doi.org/10.1093/bioinformatics/btr330>
- Danecek, P., Bonfield, J. K., Liddle, J., Marshall, J., Ohan, V., Pollard, M. O., Whitwham, A., Keane, T., McCarthy, S. A., Davies, R. M., & Li, H. (2021). Twelve years of SAMtools and BCFtools. *GigaScience*, 10(2), giab008. <https://doi.org/10.1093/gigascience/giab008>
- FAO. (2020). *Major Tropical Fruits Market Review February 2020 snapshot*. FAO. <http://www.fao.org/3/ca9213en/ca9213en.pdf>
- Greiner, S., Sobanski, J., & Bock, R. (2015). Why are most organelle genomes transmitted maternally?: Prospects & Overviews. *BioEssays*, 37(1), 80–94. <https://doi.org/10.1002/bies.201400110>
- Ha, C. O., Sands, V. E., Soepadmo, E., & Jong, K. (1988). Reproductive patterns of selected understorey trees in the Malaysian rain forest: The apomictic species. *Botanical Journal of the Linnean Society*, 97(3), 317–331. <https://doi.org/10.1111/j.1095-8339.1988.tb01586.x>
- Hambali, G. G., & Natawijaya, A. (2016). A preliminary study of *Garcinia malaccensis* collection in Taman Buah Mekarsari, Cileungsi, Bogor, Indonesia. *Contribution of Breeding Research for Sustainable Agricultural Production under Changing Environment for Food Security in Asia and Oceania*, 31–36.

- Hoang, D. T., Chernomor, O., von Haeseler, A., Minh, B. Q., & Vinh, L. S. (2018). UFBoot2: Improving the Ultrafast Bootstrap Approximation. *Molecular Biology and Evolution*, 35(2), 518–522. <https://doi.org/10.1093/molbev/msx281>
- Idris, S., & Rukayah, A. (1987). Description of the male mangosteen (*Garcinia mangostana* L.) discovered in Peninsular Malaysia. *MARDI Research Bulletin*, 15(1), 63–66.
- Inkscape Project. (2022). *Inkscape* (Inkscape 1.2.2 (732a01da63, 2022-12-09)) [Computer software]. <https://inkscape.org>
- Jones, S. W. (1980). *Morphology and major taxonomy of Garcinia (Guttiferae)* [Doctoral Thesis, University of Leicester]. <http://hdl.handle.net/2381/35319>
- Kalyaanamoorthy, S., Minh, B. Q., Wong, T. K. F., von Haeseler, A., & Jermini, L. S. (2017). ModelFinder: Fast model selection for accurate phylogenetic estimates. *Nature Methods*, 14(6), Article 6. <https://doi.org/10.1038/nmeth.4285>
- Koboldt, D. C., Zhang, Q., Larson, D. E., Shen, D., McLellan, M. D., Lin, L., Miller, C. A., Mardis, E. R., Ding, L., & Wilson, R. K. (2012). VarScan 2: Somatic mutation and copy number alteration discovery in cancer by exome sequencing. *Genome Research*, 22(3), 568–576. <https://doi.org/10.1101/gr.129684.111>
- Leigh, J. W., & Bryant, D. (2015). popart: Full-feature software for haplotype network construction. *Methods in Ecology and Evolution*, 6(9), 1110–1116. <https://doi.org/10.1111/2041-210X.12410>
- Li, H., & Durbin, R. (2009). Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics*, 25(14), 1754–1760. <https://doi.org/10.1093/bioinformatics/btp324>
- Lim, A. L. (1984). The embryology of *Garcinia mangostana* L. (Clusiaceae). *Gardens' Bulletin Singapore*, 37(1).
- Lim, T. K. (2012). *Garcinia malaccensis*. In T. K. Lim (Ed.), *Edible Medicinal And Non-Medicinal Plants: Volume 2, Fruits* (pp. 80–82). Springer Netherlands. https://doi.org/10.1007/978-94-007-1764-0_14
- Maillol, V. (2013). *demultadapt: UMR AGAP, Equipe Diversité et Adaptation de la Vigne et des Espèces Méditerranéennes, INRA, 2 Place Viala, Montpellier 34060, France* [Python]. <https://github.com/Maillol/demultadapt>
- Mariac, C., Robert, T., Allinne, C., Remigereau, M. S., Luxereau, A., Tidjani, M., Seyni, O., Bezancon, G., Pham, J. L., & Sarr, A. (2006). Genetic diversity and gene flow among pearl millet crop/weed complex: A case study. *Theoretical and Applied Genetics*, 113(6), 1003–1014. <https://doi.org/10.1007/s00122-006-0360-9>
- Mariac, C., Scarcelli, N., Pouzadou, J., Barnaud, A., Billot, C., Faye, A., Kougbadjjo, A., Maillol, V., Martin, G., Sabot, F., Santoni, S., Vigouroux, Y., & Couvreur,

- T. L. P. (2014). Cost-effective enrichment hybridization capture of chloroplast genomes at deep multiplexing levels for population genetics and phylogeography studies. *Molecular Ecology Resources*, 14(6), 1103–1113. <https://doi.org/10.1111/1755-0998.12258>
- Martin, M. (2011). Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet.Journal*, 17(1), Article 1. <https://doi.org/10.14806/ej.17.1.200>
- Matra, D. D., Poerwanto, R., Santosa, E., Sobir, Higashio, H., Anzai, H., & Inoue, E. (2016). Analysis of Allelic Diversity and Genetic Relationships Among Cultivated Mangosteen (*Garcinia mangostana* L.) in Java, Indonesia Using Microsatellite Markers and Morphological Characters. *Tropical Plant Biology*, 9(1), 29–41. <https://doi.org/10.1007/s12042-016-9161-8>
- Medellín Zabala, D. M. (2016). *Sistématica del género Garcinia (Clusiaceae): Revisión taxonómica para Colombia y filogenia de las especies Neotropicales* [PhD, Universidad Nacional de Colombia]. <https://repositorio.unal.edu.co/handle/unal/56032>
- Midin, M. R. (2018). *Cytogenetics study of Garcinia mangostana* [Doctoral dissertation, International Islamic University Malaysia]. <https://lib.iiu.edu.my/mom/services/mom/document/getFile/Xk6MR5OemyO2SgXqQZvLcwWnaeRKl6Ry20190410115700600>
- Midin, M. R., Nordin, M. S., Madon, M., Nazre, S., Goh, H.-H., & Normah, M. N. (2018). Determination of the chromosome number and genome size of *Garcinia mangostana* L. via cytogenetics, flow cytometry and k-mer analyses. *Caryologia*, 71(1), 35–44. <https://doi.org/10.1080/00087114.2017.1403762>
- Murthy, H. N., Dandin, V. S., Dalawai, D., Park, S.-Y., & Paek, K.-Y. (2018). Breeding of *Garcinia* spp. In J. M. Al-Khayri, S. M. Jain, & D. V. Johnson (Eds.), *Advances in Plant Breeding Strategies: Fruits* (pp. 773–809). Springer International Publishing. https://doi.org/10.1007/978-3-319-91944-7_19
- Nazre, S. (2014). New evidence on the origin of mangosteen (*Garcinia mangostana* L.) based on morphology and ITS sequence. *Genetic Resources and Crop Evolution*, 61(6), 1147–1158. <https://doi.org/10.1007/s10722-014-0097-2>
- Nazre, S., Clyde, M. M., & Latiff, A. (2007). Phylogenetic relationships of locally cultivated *Garcinia* species with some wild relatives. *Malaysian Applied Biology*, 36(1), 31–40.
- Nazre, S., Newman, M. F., Pennington, R. T., & Middleton, D. J. (2018). Taxonomic revision of *Garcinia* Section *Garcinia* (Clusiaceae). *Phytotaxa*, 373(1), 1–52. <https://doi.org/10.11646/phytotaxa.373.1.1>
- Ngernsaengsaruy, C. (2022). Lectotypifications of Three Names in *Garcinia*, Synonymy of *Garcinia pedunculata* and Detailed Descriptions of Three Species in *Garcinia* Section *Brindonia* (Clusiaceae). *Diversity*, 14(7), Article 7. <https://doi.org/10.3390/d14070556>

- Nguyen, L.-T., Schmidt, H. A., von Haeseler, A., & Minh, B. Q. (2015). IQ-TREE: A Fast and Effective Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies. *Molecular Biology and Evolution*, 32(1), 268–274. <https://doi.org/10.1093/molbev/msu300>
- Osman, M., & Milan, A. R. (2006). *Mangosteen Garcinia mangostana L.* (J. T. Williams, R. W. Smith, N. Haq, & Z. Dunsiger, Eds.). Southampton Centre for Underutilised Crops, University of Southampton.
- Ovalle-Magallanes, B., Eugenio-Pérez, D., & Pedraza-Chaverri, J. (2017). Medicinal properties of mangosteen (*Garcinia mangostana L.*): A comprehensive update. *Food and Chemical Toxicology*, 109, 102–122. <https://doi.org/10.1016/j.fct.2017.08.021>
- Paradis, E., Claude, J., & Strimmer, K. (2004). APE: Analyses of Phylogenetics and Evolution in R language. *Bioinformatics*, 20(2), 289–290. <https://doi.org/10.1093/bioinformatics/btg412>
- Rambaut, A. (2018). *FigTree v1.4.4* [Computer software]. Institute of Evolutionary Biology, University of Edinburgh. <http://tree.bio.ed.ac.uk/software/figtree/>
- Richards, A. J. (1990). Studies in *Garcinia*, dioecious tropical forest trees: The origin of the mangosteen (*G. Mangostana L.*). *Botanical Journal of the Linnean Society*, 103(4), 301–308. <https://doi.org/10.1111/j.1095-8339.1990.tb00191.x>
- Samsir, S. A., Bunawan, H., Choong, C. Y., & Normah, M. N. (2016). Dataset of SSR markers for ISSR-Suppression-PCR to detect genetic variation in *Garcinia mangostana L.* in Peninsular Malaysia. *Data in Brief*, 8, 1438–1442. <https://doi.org/10.1016/j.dib.2016.08.016>
- Sarah, G. (2014). *Filter FASTQ reads on mean quality* [Perl]. INRA-CIRAD. https://github.com/SouthGreenPlatform/arcad-hts/blob/master/scripts/arcad_hts_2_Filter_Fastq_On_Mean_Quality.pl
- Soltis, D. E., Soltis, P. S., & Milligan, B. G. (1992). Intraspecific Chloroplast DNA Variation: Systematic and Phylogenetic Implications. In P. S. Soltis, D. E. Soltis, & J. J. Doyle (Eds.), *Molecular Systematics of Plants* (pp. 117–150). Springer US. https://doi.org/10.1007/978-1-4615-3276-7_6
- Sprecher, A. (1919a). Étude sur la semence et al germination du *Garcinia mangostana L.* *Revue Generale de Botanique*, 31, 513–531.
- Sprecher, A. (1919b). Étude sur la semence et al germination du *Garcinia mangostana L.* (fin). *Revue Generale de Botanique*, 31, 609–633.
- Sweeney, P. W. (2008). Phylogeny and floral diversity in the genus *Garcinia* (Clusiaceae) and relatives. *International Journal of Plant Sciences*, 169(9), 1288–1303. <https://doi.org/10.1086/591990>
- Thomas, S. C. (1997). Geographic parthenogenesis in a tropical forest tree. *American Journal of Botany*, 84(8), 1012–1015.

- Tonti-Filippini, J., Nevill, P. G., Dixon, K., & Small, I. (2017). What can we do with 1000 plastid genomes? *The Plant Journal*, 90(4), 808–818. <https://doi.org/10.1111/tpj.13491>
- Villanueva-Corrales, S., García-Botero, C., Garcés-Cardona, F., Ramírez-Ríos, V., Villanueva-Mejía, D. F., & Álvarez, J. C. (2021). The Complete Chloroplast Genome of *Plukenetia volubilis* Provides Insights Into the Organelle Inheritance. *Frontiers in Plant Science*, 12. <https://www.frontiersin.org/articles/10.3389/fpls.2021.667060>
- Wee, C.-C., Nor Muhammad, N. A., Subbiah, V. K., Arita, M., Nakamura, Y., & Goh, H.-H. (2023). Plastomes of *Garcinia mangostana* L. and Comparative Analysis with Other *Garcinia* Species. *Plants*, 12(4), Article 4. <https://doi.org/10.3390/plants12040930>
- Yao, T. L., Nazre, M., Duminil, J., Loup, C., & Munzinger, J. (2024). Multivariate morphometric analysis of mangosteen (*Garcinia mangostana* var. *Mangostana*, Clusiaceae) and its wild relatives. *Edinburgh Journal of Botany*. *Accepted*.
- Yao, T. L., Nazre, M., McKey, D., Jalonen, R., & Duminil, J. (2023). The origin of cultivated mangosteen (*Garcinia mangostana* L. var. *mangostana*): Critical assessments and an evolutionary-ecological perspective. *Ecology and Evolution*, 13(3), e9792. <https://doi.org/10.1002/ece3.9792>
- Yapwattanaphun, C., Subhadrabandhu, S., Honsho, C., & Yonemori, K. (2004). Phylogenetic Relationship of Mangosteen (*Garcinia mangostana*) and Several Wild Relatives (*Garcinia* spp.) Revealed by ITS Sequence Data. *Journal of the American Society for Horticultural Science*, 129(3), 368–373. <https://doi.org/10.21273/JASHS.129.3.0368>
- Yue, X., Zheng, X., Zong, Y., Jiang, S., Hu, C., Yu, P., Liu, G., Cao, Y., Hu, H., & Teng, Y. (2018). Combined Analyses of Chloroplast DNA Haplotypes and Microsatellite Markers Reveal New Insights Into the Origin and Dissemination Route of Cultivated Pears Native to East Asia. *Frontiers in Plant Science*, 9. <https://www.frontiersin.org/articles/10.3389/fpls.2018.00591>
- Yue, Y., Li, J., Sun, X., Li, Z., & Jiang, B. (2023). Polymorphism analysis of the chloroplast and mitochondrial genomes in soybean. *BMC Plant Biology*, 23(1), 15. <https://doi.org/10.1186/s12870-022-04028-3>
- Zhang, Q., Liu, Y., & Sodmergen, null. (2003). Examination of the cytoplasmic DNA in male reproductive cells to determine the potential for cytoplasmic inheritance in 295 angiosperm species. *Plant & Cell Physiology*, 44(9), 941–951. <https://doi.org/10.1093/pcp/pcg121>

References

- Arias, R. S., Ballard, L. L., Duke, M. V., Simpson, S. A., Liu, X. F., Orner, V. A., Sobolev, V. S., Scheffler, B. E., & Martinez-Castillo, J. (2020). Development of nuclear microsatellite markers to facilitate germplasm conservation and population genetics studies of five groups of tropical perennial plants with edible fruits and shoots: Rambutan (*Nephelium lappaceum* L.), sapodilla (*Manilkara zapota* (L.) P. Royen), lychee (*Litchi chinensis* Sonn.), mangosteen (*Garcinia mangostana* Linn. and *Garcinia cochinchinensis* (Lour.) Choisy) and bamboo (*Bambusa vulgaris* Schrad. ex J.C. Wendl and *Guadua angustifolia* Kunth). *Genetic Resources and Crop Evolution*, 67(7), 1715–1731. <https://doi.org/10.1007/s10722-020-00965-w>
- Aumeeruddy-Thomas, Y., Younes, H., Ater, M., & Khadari, B. (2014). *Beyond the Divide Between Wild and Domesticated: Spatiality, Domesticity and Practices Pertaining to Fig (Ficus carica L.) and Olive (Olea europaea L.) Agroecosystems among Jbala Communities in Northern Morocco* (pp. 191–197).
- Bird, M. I., Taylor, D., & Hunt, C. (2005). Palaeoenvironments of insular Southeast Asia during the Last Glacial Period: A savanna corridor in Sundaland? *Quaternary Science Reviews*, 24(20), 2228–2242. <https://doi.org/10.1016/j.quascirev.2005.04.004>
- Bowers, J. E., Chapman, B. A., Rong, J., & Paterson, A. H. (2003). Unravelling angiosperm genome evolution by phylogenetic analysis of chromosomal duplication events. *Nature*, 422(6930), Article 6930. <https://doi.org/10.1038/nature01521>
- Brickell, C. (Ed.). (2016). *International code of nomenclature for cultivated plants: (ICNCP or Cultivated Plant Code); incorporating the rules and recommendations for naming plants in cultivation; adopted by the International Union of Biological Sciences, International Commission for the Nomenclature of Cultivated Plants* (Ninth Edition). International Society for Horticultural Science.
- Caillon, S., Quero-Garcia, J., Lescure, J.-P., & Lebot, V. (2006). Nature of taro (*Colocasia esculenta* (L.) Schott) genetic diversity prevalent in a Pacific Ocean island, Vanua Lava, Vanuatu. *Genetic Resources and Crop Evolution*, 53(6), 1273–1289. <https://doi.org/10.1007/s10722-005-3877-x>
- Camus, M. F., Alexander-Lawrie, B., Sharbrough, J., & Hurst, G. D. D. (2022). Inheritance through the cytoplasm. *Heredity*, 129(1), Article 1. <https://doi.org/10.1038/s41437-022-00540-2>
- Chanderbali, A. S., Berger, B. A., Howarth, D. G., Soltis, D. E., & Soltis, P. S. (2017). Evolution of floral diversity: Genomics, genes and gamma. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 372(1713), 20150509. <https://doi.org/10.1098/rstb.2015.0509>

- Chennaveeraiah, M. S., & Razdan, M. K. (1980). Karyomorphological and phytochemical studies in evaluating species relationships in *Garcinia* Linn. And systematic position of *G. Xanthochymus* complex. *J. Indian Bot. Soc.*, 59, 251–262.
- Cornille, A., Gladieux, P., Smulders, M. J. M., Roldán-Ruiz, I., Laurens, F., Le Cam, B., Nersesyan, A., Clavel, J., Olonova, M., Feugey, L., Gabrielyan, I., Zhang, X.-G., Tenaillon, M. I., & Giraud, T. (2012). New Insight into the History of Domesticated Apple: Secondary Contribution of the European Wild Apple to the Genome of Cultivated Varieties. *PLoS Genetics*, 8(5), e1002703. <https://doi.org/10.1371/journal.pgen.1002703>
- Dong, Y., Duan, S., Xia, Q., Liang, Z., Dong, X., Margaryan, K., Musayev, M., Goryslavets, S., Zdunić, G., Bert, P.-F., Lacombe, T., Maul, E., Nick, P., Bitskinashvili, K., Bisztray, G. D., Drori, E., De Lorenzis, G., Cunha, J., Popescu, C. F., ... Chen, W. (2023). Dual domestications and origin of traits in grapevine evolution. *Science*, 379(6635), 892–901. <https://doi.org/10.1126/science.add8655>
- Ferguson, A. R. (2013). Kiwifruit: The wild and the cultivated plants. *Advances in Food and Nutrition Research*, 68, 15–32. <https://doi.org/10.1016/B978-0-12-394294-4.00002-X>
- Gaisberger, H., Fremout, T., Kettle, C. J., Vinceti, B., Kemalasari, D., Kanchanarak, T., Thomas, E., Serra-Diaz, J. M., Svenning, J.-C., Slik, F., Eiadthong, W., Palanisamy, K., Ravikanth, G., Bodos, V., Sang, J., Warriar, R. R., Wee, A. K. S., Elloran, C., Ramos, L. T., ... Jalonen, R. (2022). Tropical and subtropical Asia's valued tree species under threat. *Conservation Biology*, 36(3), e13873. <https://doi.org/10.1111/cobi.13873>
- Grover, C. E., Arick, M. A., II, Thrash, A., Sharbrough, J., Hu, G., Yuan, D., Snodgrass, S., Miller, E. R., Ramaraj, T., Peterson, D. G., Udall, J. A., & Wendel, J. F. (2022). Dual Domestication, Diversity, and Differential Introgression in Old World Cotton Diploids. *Genome Biology and Evolution*, 14(12), evac170. <https://doi.org/10.1093/gbe/evac170>
- Ha, C. O., Sands, V. E., Soepadmo, E., & Jong, K. (1988). Reproductive patterns of selected understorey trees in the Malaysian rainforest: Apomictic species. *Bot. J. Linn. Soc.*, 97, 317–331.
- Hambali, G. G., & Natawijaya, A. (2016). A preliminary study of *Garcinia malaccensis* collection in Taman Buah Mekarsari, Cileungsi, Bogor, Indonesia. *Contribution of Breeding Research for Sustainable Agricultural Production under Changing Environment for Food Security in Asia and Oceania*, 31–36.
- Hmimsa, Y., Aumeeruddy-Thomas, Y., & Ater, M. (2012). Vernacular Taxonomy, Classification and Varietal Diversity of fig (*Ficus carica* L.) Among Jbala cultivators in Northern Morocco. *Human Ecology*, 40. <https://doi.org/10.1007/s10745-012-9471-x>

- Jaillon, O., Aury, J.-M., Noel, B., Policriti, A., Clepet, C., Casagrande, A., Choisne, N., Aubourg, S., Vitulo, N., Jubin, C., Vezzi, A., Legeai, F., Hugueney, P., Dasilva, C., Horner, D., Mica, E., Jublot, D., Poulain, J., Bruyère, C., ... The French–Italian Public Consortium for Grapevine Genome Characterization. (2007). The grapevine genome sequence suggests ancestral hexaploidization in major angiosperm phyla. *Nature*, 449(7161), Article 7161. <https://doi.org/10.1038/nature06148>
- Johnson, M. G., Pokorny, L., Dodsworth, S., Botigué, L. R., Cowan, R. S., Devault, A., Eiserhardt, W. L., Epitawalage, N., Forest, F., Kim, J. T., Leebens-Mack, J. H., Leitch, I. J., Maurin, O., Soltis, D. E., Soltis, P. S., Wong, G. K., Baker, W. J., & Wickett, N. J. (2019). A Universal Probe Set for Targeted Sequencing of 353 Nuclear Genes from Any Flowering Plant Designed Using k-Medoids Clustering. *Systematic Biology*, 68(4), 594–606. <https://doi.org/10.1093/sysbio/syy086>
- Matra, D. D., Poerwanto, R., Santosa, E., Sobir, Higashio, H., Anzai, H., & Inoue, E. (2016). Analysis of Allelic Diversity and Genetic Relationships Among Cultivated Mangosteen (*Garcinia mangostana* L.) in Java, Indonesia Using Microsatellite Markers and Morphological Characters. *Tropical Plant Biology*, 9(1), 29–41. <https://doi.org/10.1007/s12042-016-9161-8>
- McKey, D., Elias, M., Pujol, B., & Duputié, A. (2010). The evolutionary ecology of clonally propagated domesticated plants: Tansley review. *New Phytologist*, 186(2), 318–332. <https://doi.org/10.1111/j.1469-8137.2010.03210.x>
- Nazre, S. (2014). New evidence on the origin of mangosteen (*Garcinia mangostana* L.) based on morphology and ITS sequence. *Genetic Resources and Crop Evolution*, 61(6), 1147–1158. <https://doi.org/10.1007/s10722-014-0097-2>
- Nge, F. J., Biffin, E., Thiele, K. R., & Waycott, M. (2021). Reticulate Evolution, Ancient Chloroplast Haplotypes, and Rapid Radiation of the Australian Plant Genus *Adenanthos* (Proteaceae). *Frontiers in Ecology and Evolution*, 8. <https://www.frontiersin.org/articles/10.3389/fevo.2020.616741>
- Petit, R. J., Duminil, J., Fineschi, S., Hampe, A., Salvini, D., & Vendramin, G. G. (2004). INVITED REVIEW: Comparative organization of chloroplast, mitochondrial and nuclear diversity in plant populations. *Molecular Ecology*, 14(3). <https://doi.org/10.1111/j.1365-294x.2004.02410.x>
- Purseglove, J. W. (1968). The origin and spread of tropical crops. In *Tropical Crops, Dicotyledon I* (Vol. 1).
- Samsir, S. A., Bunawan, H., Choong, C. Y., & Normah, M. N. (2016). Dataset of SSR markers for ISSR-Suppression-PCR to detect genetic variation in *Garcinia mangostana* L. in Peninsular Malaysia. *Data in Brief*, 8, 1438–1442. <https://doi.org/10.1016/j.dib.2016.08.016>
- The IUCN Red List of Threatened Species*. (n.d.). IUCN Red List of Threatened Species. Retrieved 11 September 2023, from <https://www.iucnredlist.org/en>

- Turland, N. J., Wiersema, J. H., Barrie, F. R., Greuter, W., Hawksworth, D. L., Herendeen, P. S., Knapp, S., Kusber, W.-H., Li, D.-Z., Marhold, K., May, T. W., McNeill, J., Munro, A. M., Prado, J., Price, M. J., Smith, G., & International Association for Plant Taxonomy (Eds.). (2018). *International code of nomenclature for algae, fungi, and plants (Shenzhen Code): Adopted by the nineteenth International Botanical Congress, Shenzhen, China, July, 2017*. Koeltz Botanical Books.
- Vinceti, B., Loo, J., Gaisberger, H., van Zonneveld, M. J., Schueler, S., Konrad, H., Kadu, C. A. C., & Geburek, T. (2013). Conservation Priorities for *Prunus africana* Defined with the Aid of Spatial Analysis of Genetic Data and Climatic Variables. *PLoS ONE*, 8(3), e59987. <https://doi.org/10.1371/journal.pone.0059987>
- Wee, C.-C., Nor Muhammad, N. A., Subbiah, V. K., Arita, M., Nakamura, Y., & Goh, H.-H. (2023). Plastomes of *Garcinia mangostana* L. and Comparative Analysis with Other *Garcinia* Species. *Plants*, 12(4), Article 4. <https://doi.org/10.3390/plants12040930>
- Wong, W. W. W., & Lamb, A. (1993). *Fruits, nuts and spices*. In-house Seminar and Workshop on Fruits, Nuts, and Spices (1990 : Agriculture Research Station). <https://agris.fao.org/agris-search/search.do?recordID=US201300039756>
- Yao, T. L., Nazre, M., McKey, D., Jalonen, R., & Duminil, J. (2023). The origin of cultivated mangosteen (*Garcinia mangostana* L. var. *mangostana*): Critical assessments and an evolutionary-ecological perspective. *Ecology and Evolution*, 13(3), e9792. <https://doi.org/10.1002/ece3.9792>
- Yapwattanaphun, C., Subhadrabandhu, S., Honsho, C., & Yonemori, K. (2004). Phylogenetic Relationship of Mangosteen (*Garcinia mangostana*) and Several Wild Relatives (*Garcinia* spp.) Revealed by ITS Sequence Data. *Journal of the American Society for Horticultural Science*, 129(3), 368–373. <https://doi.org/10.21273/JASHS.129.3.0368>
- Yogom, B. T., Avana-Tientcheu, M.-L., Monthé, F. K., Bissiengou, P., Loumeto, J. J., Zekraoui, L., Hervouet, C., Fonkou, T., Mariac, C., & Duminil, J. (2023). Genetic diversity and structure in wild and cultivated populations of an emblematic African tree species, *Garcinia kola* (Clusiaceae). *Tree Genetics & Genomes*, 19(5), 39. <https://doi.org/10.1007/s11295-023-01614-w>
- Záveská Drábková, L. (2021). Herbarium Specimens: A Treasure for DNA Extraction, an Update. *Methods in Molecular Biology (Clifton, N.J.)*, 2222, 69–88. https://doi.org/10.1007/978-1-0716-0997-2_4

REFERENCES

- Abdullah, N. A. P., Richards, A. J., & Wolff, K. (2012). Molecular evidence in identifying parents of *Garcinia mangostana* L. *Pertanika Journal of Tropical Agricultural Science*, 35(2), 257–270.
- Aumeeruddy-Thomas, Y., Younes, H., Ater, M., & Khadari, B. (2014). *Beyond the Divide Between Wild and Domesticated: Spatiality, Domesticity and Practices Pertaining to Fig (Ficus carica L.) and Olive (Olea europaea L.) Agroecosystems among Jbala Communities in Northern Morocco* (pp. 191–197).
- Beentje, H. J. (2010). *The Kew Plant Glossary. An illustrated dictionary of plant terms*. Kew Publishing.
- Bird, M. I., Taylor, D., & Hunt, C. (2005). Palaeoenvironments of insular Southeast Asia during the Last Glacial Period: A savanna corridor in Sundaland? *Quaternary Science Reviews*, 24(20), 2228–2242. <https://doi.org/10.1016/j.quascirev.2005.04.004>
- Bowers, J. E., Chapman, B. A., Rong, J., & Paterson, A. H. (2003). Unravelling angiosperm genome evolution by phylogenetic analysis of chromosomal duplication events. *Nature*, 422(6930), Article 6930. <https://doi.org/10.1038/nature01521>
- Caillon, S., Quero-Garcia, J., Lescure, J.-P., & Lebot, V. (2006). Nature of taro (*Colocasia esculenta* (L.) Schott) genetic diversity prevalent in a Pacific Ocean island, Vanua Lava, Vanuatu. *Genetic Resources and Crop Evolution*, 53(6), 1273–1289. <https://doi.org/10.1007/s10722-005-3877-x>
- Caliński, T., & Harabasz, J. (1974). A dendrite method for cluster analysis. *Communications in Statistics*, 3(1), 1–27. <https://doi.org/10.1080/03610927408827101>
- Camus, M. F., Alexander-Lawrie, B., Sharbrough, J., & Hurst, G. D. D. (2022). Inheritance through the cytoplasm. *Heredity*, 129(1), Article 1. <https://doi.org/10.1038/s41437-022-00540-2>
- Chanderbali, A. S., Berger, B. A., Howarth, D. G., Soltis, D. E., & Soltis, P. S. (2017). Evolution of floral diversity: Genomics, genes and gamma. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 372(1713), 20150509. <https://doi.org/10.1098/rstb.2015.0509>
- Chennaveeraiah, M. S., & Razdan, M. K. (1980). Karyomorphological and phytochemical studies in evaluating species relationships in *Garcinia* Linn. And systematic position of *G. Xanthochymus* complex. *J. Indian Bot. Soc.*, 59, 251–262.
- Clement, M., Posada, D., & Crandall, K. A. (2000). TCS: A computer program to estimate gene genealogies. *Molecular Ecology*, 9(10), 1657–1659. <https://doi.org/10.1046/j.1365-294x.2000.01020.x>

- Dong, Y., Duan, S., Xia, Q., Liang, Z., Dong, X., Margaryan, K., Musayev, M., Goryslavets, S., Zdunić, G., Bert, P.-F., Lacombe, T., Maul, E., Nick, P., Bitskinashvili, K., Bisztray, G. D., Drori, E., De Lorenzis, G., Cunha, J., Popescu, C. F., ... Chen, W. (2023). Dual domestications and origin of traits in grapevine evolution. *Science*, 379(6635), 892–901. <https://doi.org/10.1126/science.add8655>
- FAO. (2020). *Major Tropical Fruits Market Review February 2020 snapshot*. FAO. <http://www.fao.org/3/ca9213en/ca9213en.pdf>
- Ferguson, A. R. (2013). Kiwifruit: The wild and the cultivated plants. *Advances in Food and Nutrition Research*, 68, 15–32. <https://doi.org/10.1016/B978-0-12-394294-4.00002-X>
- Gong, Z. (2019). 《西洋番国志 校注本》 (*Collated and annotated Xī Yáng Fān Guó Zhì [1434], 'The Overall Survey of the Ocean's Shores'*) (D. Xiang, Ed.). Hua Wen Publisher.
- Gower, J. C. (1971). A General Coefficient of Similarity and Some of Its Properties. *Biometrics*, 27(4), 857–871. <https://doi.org/10.2307/2528823>
- Grover, C. E., Arick, M. A., II, Thrash, A., Sharbrough, J., Hu, G., Yuan, D., Snodgrass, S., Miller, E. R., Ramaraj, T., Peterson, D. G., Udall, J. A., & Wendel, J. F. (2022). Dual Domestication, Diversity, and Differential Introgression in Old World Cotton Diploids. *Genome Biology and Evolution*, 14(12), evac170. <https://doi.org/10.1093/gbe/evac170>
- Ha, C. O., Sands, V. E., Soepadmo, E., & Jong, K. (1988). Reproductive patterns of selected understorey trees in the Malaysian rainforest: Apomictic species. *Bot. J. Linn. Soc.*, 97, 317–331.
- Hambali, G. G., & Natawijaya, A. (2016). A preliminary study of *Garcinia malaccensis* collection in Taman Buah Mekarsari, Cileungsi, Bogor, Indonesia. *Contribution of Breeding Research for Sustainable Agricultural Production under Changing Environment for Food Security in Asia and Oceania*, 31–36.
- Idris, S., & Rukayah, A. (1987). Description of the male mangosteen (*Garcinia mangostana* L.) discovered in Peninsular Malaysia. *MARDI Research Bulletin*, 15(1), 63–66.
- Jaillon, O., Aury, J.-M., Noel, B., Policriti, A., Clepet, C., Casagrande, A., Choisne, N., Aubourg, S., Vitulo, N., Jubin, C., Vezzi, A., Legeai, F., Huguency, P., Dasilva, C., Horner, D., Mica, E., Jublot, D., Poulain, J., Bruyère, C., ... The French–Italian Public Consortium for Grapevine Genome Characterization. (2007). The grapevine genome sequence suggests ancestral hexaploidization in major angiosperm phyla. *Nature*, 449(7161), Article 7161. <https://doi.org/10.1038/nature06148>
- Jones, S. W. (1980). *Morphology and major taxonomy of Garcinia (Guttiferae)* [Doctoral Thesis, University of Leicester]. <http://hdl.handle.net/2381/35319>

- Legendre, P., & Legendre, L. F. J. (1998). *Numerical Ecology*. Elsevier Science.
- Leigh, J. W., & Bryant, D. (2015). popart: Full-feature software for haplotype network construction. *Methods in Ecology and Evolution*, 6(9), 1110–1116. <https://doi.org/10.1111/2041-210X.12410>
- Lim, A. L. (1984). The embryology of *Garcinia mangostana* L. (Clusiaceae). *Gardens' Bulletin Singapore*, 37(1).
- Lim, T. K. (2012). *Garcinia malaccensis*. In T. K. Lim (Ed.), *Edible Medicinal And Non-Medicinal Plants: Volume 2, Fruits* (pp. 80–82). Springer Netherlands. https://doi.org/10.1007/978-94-007-1764-0_14
- Ma, H. (2019). 《瀛涯勝覽 校注本》(*Collated and annotated Ying Ya Sheng Lǎn [1416], 'The Overall Survey of the Ocean's Shores'*) (C.-C. Feng, Ed.). Hua Wen Publisher.
- Matra, D. D., Poerwanto, R., Santosa, E., Sobir, Higashio, H., Anzai, H., & Inoue, E. (2016). Analysis of Allelic Diversity and Genetic Relationships Among Cultivated Mangosteen (*Garcinia mangostana* L.) in Java, Indonesia Using Microsatellite Markers and Morphological Characters. *Tropical Plant Biology*, 9(1), 29–41. <https://doi.org/10.1007/s12042-016-9161-8>
- McKey, D., Elias, M., Pujol, B., & Duputié, A. (2010). The evolutionary ecology of clonally propagated domesticated plants: Tansley review. *New Phytologist*, 186(2), 318–332. <https://doi.org/10.1111/j.1469-8137.2010.03210.x>
- Midin, M. R. (2018). *Cytogenetics study of Garcinia mangostana* [Doctoral dissertation, International Islamic University Malaysia]. <https://lib.iium.edu.my/mom/services/mom/document/getFile/Xk6MR5OemyO2SgXqQZvLcwWnaeRKl6Ry20190410115700600>
- Morel, J., Le Mogueédec, G., & Munzinger, J. (2021). Multivariate morphometric analysis supported by an anatomical approach to assess species delimitation in *Xyris* (Xyridaceae) in New Caledonia. *Botanical Journal of the Linnean Society*, boaa108. <https://doi.org/10.1093/botlinnean/boaa108>
- Murthy, H. N., Dandin, V. S., Dalawai, D., Park, S.-Y., & Paek, K.-Y. (2018). Breeding of *Garcinia* spp. In J. M. Al-Khayri, S. M. Jain, & D. V. Johnson (Eds.), *Advances in Plant Breeding Strategies: Fruits* (pp. 773–809). Springer International Publishing. https://doi.org/10.1007/978-3-319-91944-7_19
- Nazre, S. (2014). New evidence on the origin of mangosteen (*Garcinia mangostana* L.) based on morphology and ITS sequence. *Genetic Resources and Crop Evolution*, 61(6), 1147–1158. <https://doi.org/10.1007/s10722-014-0097-2>
- Nazre, S., Newman, M. F., Pennington, R. T., & Middleton, D. J. (2018). Taxonomic revision of *Garcinia* Section *Garcinia* (Clusiaceae). *Phytotaxa*, 373(1), 1–52. <https://doi.org/10.11646/phytotaxa.373.1.1>

- Nguyen, L.-T., Schmidt, H. A., von Haeseler, A., & Minh, B. Q. (2015). IQ-TREE: A Fast and Effective Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies. *Molecular Biology and Evolution*, 32(1), 268–274. <https://doi.org/10.1093/molbev/msu300>
- Pierre, A.-H., Le Moguédec, G., Lowry, P. P., & Munzinger, J. (2014). Multivariate morphometric analysis and species delimitation in the endemic New Caledonian genus *Storthocalyx* (Sapindaceae): Morphometric Analysis of *Storthocalyx*. *Botanical Journal of the Linnean Society*, n/a-n/a. <https://doi.org/10.1111/boj.12199>
- Rambaut, A. (2018). *FigTree v1.4.4* [Computer software]. Institute of Evolutionary Biology, University of Edinburgh. <http://tree.bio.ed.ac.uk/software/figtree/>
- Richards, A. J. (1990). Studies in *Garcinia*, dioecious tropical forest trees: The origin of the mangosteen (*G. Mangostana* L.). *Botanical Journal of the Linnean Society*, 103(4), 301–308. <https://doi.org/10.1111/j.1095-8339.1990.tb00191.x>
- Sprecher, A. (1919a). Étude sur la semence et al germination du *Garcinia mangostana* L. *Revue Generale de Botanique*, 31, 513–531.
- Sprecher, A. (1919b). Étude sur la semence et al germination du *Garcinia mangostana* L. (fin). *Revue Generale de Botanique*, 31, 609–633.
- Verheij, E. W. M., & Coronel, R. E. (1991). *Edible Fruits and Nuts*. Pudoc.
- Ward, J. H. (1963). Hierarchical Grouping to Optimize an Objective Function. *Journal of the American Statistical Association*, 58(301), 236–244. <https://doi.org/10.2307/2282967>
- Yao, T. L., Nazre, M., McKey, D., Jalonen, R., & Duminil, J. (2023). The origin of cultivated mangosteen (*Garcinia mangostana* L. var. *mangostana*): Critical assessments and an evolutionary-ecological perspective. *Ecology and Evolution*, 13(3), e9792. <https://doi.org/10.1002/ece3.9792>
- Yogom, B. T., Avana-Tientcheu, M.-L., Monthé, F. K., Bissiengou, P., Loumeto, J. J., Zekraoui, L., Hervouet, C., Fonkou, T., Mariac, C., & Duminil, J. (2023). Genetic diversity and structure in wild and cultivated populations of an emblematic African tree species, *Garcinia kola* (Clusiaceae). *Tree Genetics & Genomes*, 19(5), 39. <https://doi.org/10.1007/s11295-023-01614-w>