



UNIVERSITI PUTRA MALAYSIA

**PROBIOTIC EFFECTS OF *LACTOBACILLUS* STRAINS ON CECAL
MICROBIOTA AND FERMENTATION, INTESTINAL DEVELOPMENT
AND RESPONSE TO HUMAN CONTACT IN BROILER CHICKENS**

**AMIR MEIMANDIPOUR
FSTM 2010 4**





**PROBIOTIC EFFECTS OF *LACTOBACILLUS* STRAINS ON CECAL
MICROBIOTA AND FERMENTATION, INTESTINAL DEVELOPMENT AND
RESPONSE TO HUMAN CONTACT IN BROILER CHICKENS**

By

AMIR MEIMANDIPOUR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of
Philosophy**

March 2010



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

PROBIOTIC EFFECTS OF *LACTOBACILLUS* STRAINS ON CECAL MICROBIOTA AND FERMENTATION, INTESTINAL DEVELOPMENT AND RESPONSE TO HUMAN CONTACT IN BROILER CHICKENS

By

AMIR MEIMANDIPOUR

March 2010

Chairman: Professor Mohd Yazid Abdul Manap, PhD

Faculty: Food Science and Technology

The aim of this study was to evaluate the probiotic effects of *Lactobacillus agilis* JCM 1048 and *Lactobacillus salivarius* subsp. *salicinius* JCM 1230 under *in vitro* and *in vivo* conditions. In the first *in vitro* experiment, batch bioreactor system was used to assess the effects of two *Lactobacillus* strains on selected bacterial groups, short chain fatty acids (SCFAs) and lactate concentrations. In this experiment, 24 h incubation was done in a simulated chicken cecum, at two pH values. The addition of *L. agilis* JCM 1048 and *L. salivarius* subsp. *salicinius* JCM 1230 significantly ($P < 0.05$) increased number of total anaerobes (19%), lactobacilli (18%) and bifidobacteria (27%) at pH 5.8 while decreased those of aerobes (pH=5.8, 13% & pH=6.7, 9%) and streptococci (22% in both pH values) in (Cc+L) treatment group after 24 h incubation as compared to (Cc) group (Table 3.1). The supplementation with lactobacilli was found to increase the



production of lactate, propionate and butyrate (pH=5.8, 256% & pH=6.7, 246%), while suppressed the concentration of acetate (Table 3.3). Furthermore, pH did not change the formation of butyrate, while the production of acetate and propionate decreased at pH 5.8. The second *in vitro* experiment was also conducted using a batch bioreactor. The microbial groups were determined by using real time polymerase chain reaction (PCR) and SCFAs concentrations were monitored over 24 h batch culture incubation at pH 5.8. The supplementation of cecal content with two strains of *Lactobacillus* significantly ($P<0.05$) increased the number of lactobacilli (8%), bifidobacteria (10%) and *Faecalibacterium prausnitzii* (10%) compared to (Cc+L) group (Table 4.6). *Lactobacillus* supplementation did not affect ($P<0.05$) the number of *E.coli* and *Clostridium butyricum*; however, it significantly ($P<0.05$) decreased those of *Salmonella* (14%). Inclusion of *Lactobacillus* in the vessel containing cecal material significantly ($P<0.05$) increased the production of propionate and butyrate (261%), whereas decreased the amount of acetate (Table 4.8).

The *in vivo* study was conducted to assess the effects of two *Lactobacillus* strains on broiler performance, small intestine development and SCFAs concentration in various segments of the GIT in the chickens under unpleasant physical contact (UPC). The following treatments were applied from day one: (1) chicks exposed to normal human contact fed basal diet

(control); (2) chicks were exposed to UPC and fed basal diet (UPC-BD); (3) chicks were exposed to UPC and fed basal diet supplemented with *Lactobacillus* (UPC-BDL). From day 1 to day 21, the chicks in UPC groups were caught using both hands, placed in plastic crates and moved to a separate room. The chicks were picked up in a group, suspended by legs and swung gently for 30 s once daily. Unpleasant physical contact significantly ($P<0.05$) increased blood plasma concentration in UPC-BD and UPC-BDL groups, respectively 37% and 27% at 2 weeks of age (Table 5.2). Broilers in UPC-BDL treatment group showed significantly ($P<0.05$) higher body weight gain (10%) and better feed conversion ratio (7.4%) as compared with UPC-BD treatment group at 7 days of age (Table 5.4). Duodenal villi height was numerically suppressed (7.4%) in UPC-BD treatment as compared with the control group at 14 days of age (Table 5.5). Dietary *Lactobacillus* profoundly increased (11%) duodenal villi height in UPC-BDL birds at 28 days of age. Broilers in UPC-BD treatment showed a significant ($P<0.05$) lower concentration of duodenal lactate (22%), ileal acetate (51%), cecal lactate (16%), cecal acetate (41%), and cecal butyrate (72%) as compared to the control groups at 14 days of age (Table 5.9). However, lactobacilli administration significantly ($P<0.05$) increased the concentration of duodenal lactate (21%), duodenal propionate (23%), cecal lactate (16%), and cecal butyrate (112%) in UPC-BDL treatment group compared to UPC-BD counterparts at similar age. Lactobacilli



supplementation profoundly suppressed ileal and cecal acetate concentration, respectively 51% and 42% in 14 days UPC-BDL treatment group compared with UPC-BD birds. Both duodenal and jejunal pH values significantly ($P<0.05$) increased (6%) in UPC-BD treatment compared with the control group at 14 days of age (Table 5.14). Adding *Lactobacillus* to the diet, significantly ($P<0.05$) lowered duodenal and jejunal pH values in UPC-BDL group as compared with UPC-BD group at 14 and 28 days of age.

In conclusion, the results of the current study suggested that *L. agilis* JCM 1048 and *L. salivarius* subsp. *salicinius* JCM 1230 have the ability to re-establish proper microbial balance in the chicken cecum. *Lactobacillus* supplementation profoundly reduced total number of aerobes, *Streptococcus*, and *Salmonella* in favor of lactobacilli, bifidobacteria, and *F. prausnitzii* which was coincided by high formation of lactate, butyrate as well as propionate. Although, UPC was not too aversive to decrease broiler performance, but it negatively affected duodenal structural growth in broilers of UPC-BD group at 14 days of age. Furthermore, UPC negatively changed the GIT bacterial metabolic end products during unpleasant handling. Dietary supplementation with *Lactobacillus* ameliorated detrimental effects of UPC on broilers over the course of the experimental period.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah PhD

**KESAN PROBIOTIK STRAIN *LACTOBACILLUS* PADA CECAL
MIKROBIOTA DAN FERMENTASI, PERKEMBANGAN USUS KECIL
DAN RESPON AYAM PEDAGING TERHADAP MANUSIA**

Oleh

AMIR MEIMANDIPOUR

Mac 2010

Pengerusi: Professor Mohd Yazid Abdul Manap, PhD

Fakulti: Sains dan Teknologi Makanan

Tujuan kajian ini adalah untuk menilai kesan-kesan probiotik *Lactobacillus agilis* JCM 1048 dan *L. salivarius* subsp. *salicinius* JCM 1230 dalam keadaan *in vitro* dan *in vivo*. Dalam eksperimen *in vitro* yang pertama, sistem bioreaktor berkelompok telah digunakan untuk menilai pengaruh dua strain *Lactobacillus* terhadap kumpulan bakteria yang terpilih, asid lemak rantai pendek (SCFAs) dan kepekatan laktat. Dalam kajian ini, inkubasi selama 24 jam dalam simulasi cecum ayam telah dijalankan pada dua nilai pH. Penambahan *L. agilis* JCM 1048 dan *L. salivarius* subsp. *salicinius* JCM 1230 telah meningkatkan jumlah bilangan koloni pada keseluruhan anaerob (19%), *Lactobacilli* (18%) dan *Bifidobakteria* (27%) pada pH 5.8 dan juga mengurangkan bilangan aerob (pH=5.8, 13% dan pH=6.7, 9%) dan *Streptococci* (22% pada kedua-dua nilai pH) dalam kumpulan rawatan (Cc+L) selepas inkubasi selama 24 jam jika dibandingkan kepada



kumpulan (Cc). Penambahan *Lactobacilli* telah menunjukkan peningkatan terhadap penghasilan laktat, propionat dan butyrate (pH=5.8, 256% and pH=6.7, 246%) dan merendahkan kepekatan asetat. Tambahan lagi, pH tidak mengubah pembentukan butir, manakala penghasilan asetat dan propionat menurun pada pH 5.8. Eksperimen *in vitro* yang kedua juga dijalankan dengan menggunakan bioreaktor berkelompok. Kumpulan mikrob ditentukan dengan kaedah 'real time' Polymerase Chain Reaction (PCR) dan kepekatan asid lemak rantai pendek (SCFAs) dipantau selepas 24 jam inkubasi kultur berkelompok pada pH 5.8. Penambahan dua strain *Lactobacillus* dalam kandungan cecal telah meningkatkan bilangan *Lactobacillus* (8%), *Bifidobacteria* (10%) dan *Faecalibacterium prausnitzii* (10%) secara signifikan ($P<0.05$) berbanding kumpulan kawalannya (Cc+L). Penambahan *Lactobacillus* tidak mempengaruhi ($P<0.05$) bilangan *E.coli* dan *Clostridium butyricum*; bagaimanapun, ia mengurangkan bilangan *Salmonella* (14%) secara signifikan ($P<0.05$). Rangkuman *Lactobacillus* di dalam vessel mengandungi material cecal telah meningkatkan penghasilan propionat dan butyrat (261%) secara signifikan ($P<0.05$), bagaimanapun telah mengurangkan jumlah asetat.

Kajian secara *in vivo* telah dijalankan untuk menentukan kesan dua strain *Lactobacillus* terhadap prestasi ayam ternak, perkembangan usus kecil dan kepekatan SCFAs pada pelbagai segmen GIT pada ayam dibawah

interaksi fizikal tidak menyenangkan (UPC). Rawatan berikutnya dijalankan sejak hari pertama: (1) anak-anak ayam didedahkan kepada diet makanan asas dan interaksi dengan manusia secara biasa (kawalan); (2) anak-anak ayam didedahkan dengan UPC dan diet makanan asas (UPC-BD); (3) anak-anak ayam didedahkan kepada UPC dan diet makanan asas ditambah dengan *Lactobacillus* (UPC-BDL). Dari hari pertama sehingga hari ke 21, anak-anak ayam yang dalam kumpulan UPC ditangkap menggunakan dua belah tangan, seterusnya disimpan di dalam tong plastik dan dipindahkan ke bilik berasingan. Ayam tersebut di ambil secara berkumpulan, digantung pada bahagian kaki dan dihayun secara perlahan selama 30 saat sekali setiap sehari. UPC telah menyebabkan peningkatan secara signifikan 37% dan 27% pada kepekatan plasma darah pada kumpulan ayam rawatan UPC-BD dan UPC-BDL masing-masing pada umur yang ke-14 hari. Ayam ternakan dalam kumpulan UPC-BDL menunjukkan penambahan berat badan yang tinggi (10%) secara signifikan ($P < 0.05$) dan pertukaran catuan makanan yang lebih baik (7.4%) jika dibandingkan pada kumpulan UPC-BD pada umur hari yang ke-7. Ketinggian vilus duodenal telah disekat dari segi bilangan (7.4%) di dalam ayam-ayam dalam kumpulan olahan UPC-BD jika dibandingkan pada kumpulan kawalan pada usia yang ke-14 hari. Diet *Lactobacillus* meningkatkan ketinggian vilus duodenal (11%) pada kumpulan ayam rawatan UPC-BDL pada usia 28 hari. Ayam- ayam ternak dalam rawatan



UPC-BD secara signifikan ($P<0.05$) mempunyai kepekatan rendah pada duodenal laktat (22%), ileal asetat (51%), cecal laktat (16%), cecal asetat (41%), dan cecal butirat (72%) jika dibandingkan pada kumpulan kawalan pada usia yang ke-14 hari. Bagaimanapun, pelaksanaan lactobacilli dalam kajian ini secara signifikan telah meningkatkan kepekatan duodenal laktat (21%), duodenal propionat (23%), cecal laktat (16%), dan cecal butirat (112%) dalam kumpulan rawatan UPC-BDL jika dibandingkan pada kumpulan rawatan UPC-BD pada usia yang sama. Penambahan *Lactobacilli* ternyata telah menyekat kepekatan ileal dan cecal asetat masing-masing pada 51% dan 42% pada usia 14 hari dalam kumpulan rawatan UPC-BDL jika dibandingkan pada kumpulan ayam rawatan UPC-BD. Nilai pH pada kedua-dua duodenal dan jejunal meningkat (6%) secara signifikan ($P<0.05$) pada kumpulan rawatan UPC-BD jika dibandingkan pada kumpulan kawalan pada usia 14 hari. Penambahan *Lactobacillus* didalam diet makanan ayam secara signifikan ($P<0.05$) telah merendahkan nilai pH pada duodenal dan jejunal pada kumpulan UPC-BDL jika dibandingkan kepada kumpulan UPC-BD pada usia 14 dan 28 hari.

Kesimpulannya, keputusan-keputusan dalam kajian terbaru ini mencadangkan bahawa dua *L. agilis* JCM 1048 dan *L. salivarius* subsp. *salicinius* JCM 1230 mempunyai kemampuan untuk memantapkan semula keseimbangan mikrob yang sempurna dengan pembentukan laktat dan

juga propionat, dan merangsang bakteria penghasil butyrate terutamanya *F. prausnitzii* untuk menghasilkan butyrat dalam cecum ayam. Sungguhpun kaedah UPC tidak begitu di gemari dalam mengurangkan prestasi ayam ternak, tetapi ianya memberi kesan negatif terhadap pertumbuhan struktur duodenal dalam ayam-ayam ternak kumpulan UPC-BD pada usia 14 hari. Tambahan lagi, UPC mengubah secara negatif terhadap hasil akhir metabolik bakteria GIT semasa pengendalian yang kurang menyenangkan. Penambahan diet makanan dengan *Lactobacillus* memulihkan kesan penjejasan UPC pada ternakan-ternakan sepanjang tempoh eksperimen.



ACKNOWLEDGEMENTS

I would first like to thank God for giving me this opportunity to continue my studies in a field that I enjoy so much. I would like to thank the people who made this project possible: my supervisor Prof. Mohd Yazid Abd Manap for his guidance and support during all these years. You have been always been there for me. I appreciate your trust and friendship. You have been truly a tremendous role model and influence in my career. I admire you. I would like also to acknowledge the members of my committee, Associated Professor Dr. Shuhaimi Bin. Mustafa, Professor Dr. Mohd Hair Bejo, and Associate Professor Dr. Azhar Kasim for all their advice and contributions throughout this project.

I would like to thank Mr. Abdul Halim Abdul Rahman and Mr. Saipuzaman Ali for assisting in the HPLC and morphology analysis, respectively. The assistance of broiler house technicians, Mr. Mazlan Hamzah and Mr. Ponnusamy Muniandy at the Department of Animal Science was greatly appreciated.

This thesis is dedicated to my loving parents, whose love and support allowed me to complete this education and for their continuing encouragement and support all these years. It is also dedicated to my wife



and the rest of my family who provided the support that helped me to make it through the tough times.



I certify that a Thesis Examination Committee has met on 10 of March to conduct the final examination of Amir Meimandipour on his thesis entitled “**Probiotic effects of *Lactobacillus* strains on cecal microbiota and fermentation, intestinal development and response to human contact in broiler chickens**” in accordance with the Universities and University colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Son Radu, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Saleha Abdul Aziz, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Zulkifli Idrus, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Marilena Marino, PhD

Assistant professor
Department of Food Science
Universita degli Studi di Udine
Italy
(External Examiner)

BUJANG BIN KIM HUAT, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:



This thesis was submitted to the senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Yazid Abd Manap PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Mohd Hair Bejo, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Shuhaimi Bin Mustafa, PhD

Associated Professor
Faculty of Biotechnology and Molecular Science
Universiti Putra Malaysia
(Member)

Azhar Kasim, PhD

Associated Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

HASANAH MOHD GHAZALI, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 13 May 2010



DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not currently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

AMIR MEIMANDIPOUR
Date: 10 March 2010



TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	vi
ACKNOWLEDGEMENTS	xi
APPROVAL	xiii
DECLARATION	xv
LIST OF TABLES	xix
LIST OF FIGURES	xxii
LIST OF ABBREVIATIONS	xxvi
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	7
2.1 Chicken GIT Compartments	7
2.2 Chicken GIT Microbiota	9
2.3 Acquisition and Succession of GIT Microbiota	11
2.4 Biological Community of Microbiota	13
2.5 Inter-species Differences in Intestinal Microbiota	14
2.6 Spatial Distribution of Chicken GIT Microbiota	15
2.7 Factors Influencing the Intestinal Microbiota	18
2.7.1 Interaction Between Bacteria	18
2.7.2 Diet	20
2.7.3 Stressful Condition	25
2.7.4 Using Probiotic as a Strategy to Combat the Deleterious Effects of Stress on the GIT	28
2.7.5 Probiotics	28
2.7.6 The Effects of Probiotics in Poultry	29
2.7.7 Possible Mode of Action for Probiotic	32
2.7.7.1 Enhancement of Immune System	32
2.7.7.2 Competition for Colonization Sites	35
2.7.7.3 Production of Anti-microbial Substances	36
2.7.7.4 Micro-environmental Alterations	37
2.8 Intestinal Epithelium Development and Enterocyte Maturity	38
2.9 Effect of SCFAs on Enterocyte Metabolism	41
2.9.1 Uptake of SCFAs	42
2.9.2 SCFAs Utilization	44
2.9.3 Effect of SCFAs on Cytokinetics	45



2.10	Molecular Techniques for the Analysis of the GIT Microbiota	47
2.10.1	Real Time PCR	48
3	IN VITRO FERMENTATION OF BROILER CECAL CONTENT: THE ROLE OF LACTOBACILLI AND pH VALUE ON THE COMPOSITION OF MICROBIOTA AND THIER METABOLIC END PRODUCTS	55
3.1	Introduction	55
3.2	Materials and Methods	56
3.2.1	Cecal Sample	56
3.2.2	Bacterial Strains and Culture Conditions	57
3.2.3	Basal Nutrient Medium Preparation	58
3.2.4	Batch Culture Fermentations	59
3.2.5	Bacterial Enumeration	60
3.2.6	SCFAs and Lactate Analysis	61
3.2.7	Statistical Analysis	62
3.3	Results	62
3.3.1	Changes in the Bacterial Population Numbers	62
3.3.2	Changes in SCFAs and Lactate Concentrations	64
3.4	Discussion	66
3.5	Conclusion	70
4	THE STIMULATION OF BUTYRIC PRODUCING BACTERIA PROLIFERATION IN CHICKEN CECUM BY LACTOBACILLUS TREATMENT	73
4.1	Introduction	71
4.2	Materials and Methods	72
4.2.1	Cecal Sample	72
4.2.2	<i>Lactobacillus</i> Strains Used as Inoculants and Their Culture Conditions	73
4.2.3	Batch Culture Fermentations	73
4.2.4	SCFAs and Lactate Analysis	74
4.2.5	Bacterial Strains and Culture Conditions	75
4.2.5.1	Anaerobic Culture Condition	76
4.2.6	DNA Extraction From Bacterial Cultures	76
4.2.7	DNA Extraction From Cecal Content	77
4.2.8	Primers and PCR Amplification	78
4.2.9	Cloning of Target Sequence into the Competent Cell	81
4.2.10	Plasmid DNA Standards	82
4.2.11	Generation of Standard Curves	82
4.2.12	Real-Time PCR	83



4.3	Statistical Analysis	86
4.4	Results	86
4.4.1	Cloning Assay	86
4.4.2	Standard Curve and Melting Curve Analysis	86
4.4.3	Changes in the Bacterial Gene Copy Numbers	88
4.4.4	Changes in the SCFAs and Lactate Concentrations	89
4.5	Discussion	92
4.6	Conclusion	95
5	EFFECTS OF UNPLEASANT PHYSICAL CONTACT ON THE GIT MORPHOLOGY, BLOOD CORTICOSTERONE, SCFAs CONCENTRATION AND MODULATING ROLE OF TWO <i>LACTOBACILLUS</i> STRAINS IN BROILERS	96
5.1	Introduction	96
5.2	Materials and Methods	98
5.2.1	Preparation of Inoculant and Supplement Feed	98
5.2.2	Animals and Diets	98
5.2.3	Unpleasant Physical Contact Condition	100
5.2.4	Body Weight and Feed Consumption	101
5.2.5	Blood Samples	101
5.2.6	Blood Corticosterone Assay	101
5.2.7	Tissue and GIT Content Samples	102
5.2.8	SCFAs and Lactate Analysis	103
5.2.9	Experimental Design and Data Analysis	103
5.3	Results	104
5.3.1	Blood Corticosterone Concentration	104
5.3.2	Broiler Performance	106
5.3.3	Histologic Findings	108
5.3.4	Gastrointestinal Tract SCFAs and Lactate Concentration	111
5.3.5	Gastrointestinal Tract pH Value	119
5.4	Discussion	121
5.5	Conclusion	135
6	SUMMARY AND CONCLUSION	136
	REFERENCES	145
	APPENDICES	167
	BIODATA OF STUDENT	185
	LIST OF PUBLICATIONS	186



LIST OF TABLES

Table		Page
2.1	The effects of probiotics in poultry	31
3.1	Bacterial population in stirred, PH-controlled batch culture fermentations	63
3.2	Concentrations of SCFA and lactate (mmoL/L) at 0 and after 24 h of fermentation	65
3.3	Production of SCFA and lactate (mmoL/L) by cecal microbiota of broiler chicken after 24 h of fermentation	66
4.1	Types of bacteria, culture media and their environmental growth condition	76
4.2	Primer sets used in this study	80
4.3	PCR components, its volume and concentration for each PCR tube	81
4.4	The amount of components in ligation mixture for ligation of purified product to plasmid vector	82
4.5	The amount of components in each tube for real time PCR assays	85
4.6	Real-time PCR quantitation of 16S rRNA genes in stirred, pH-controlled batch culture fermentations	89
4.7	Concentrations of SCFAs at 0 and after 24 h of fermentation	91
4.8	Production of acetate, propionate, butyrate and lactate by cecal microbiota of broiler chicken after 24 h of fermentation	92
5.1	Composition of the experimental diets (%)	100
5.2	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on the concentration of corticosterone over the course of the experiment of	105



	broiler chicken	
5.3	Blood corticosterone concentration (ng/mL) over the course of the experiment	105
5.4	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on growth performance, feed intake and feed conversion ratio of broiler chicken	107
5.5	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on villi height in the different small intestinal regions of broiler chicken	109
5.6	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on crypt depth in the different small intestinal regions of broiler chicken	110
5.7	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on villi height: crypt depth ratio in the different small intestinal regions of broiler chicken	111
5.8	Concentration of SCFA and lactate in different parts of the GIT during the experimental period	114
5.9	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on the concentration of SCFA and lactate in the different GIT regions of broiler chicken at 14 days of age	115
5.10	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on the concentration of SCFA and lactate in the different GIT regions of broiler chicken at 28 days of age	116
5.11	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on the concentration of SCFA and lactate in the different GIT regions of broiler chicken at 35 days of age	117
5.12	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on the concentration of SCFA and lactate in the different GIT regions of broiler chicken at 42 days of age	118

5.13	The pH values in the different GIT regions of broiler chicken during the experiment	119
5.14	Influence of <i>Lactobacillus</i> supplemented diet and unpleasant physical contact on the pH values in the different GIT regions of broiler chicken	120



LIST OF FIGURES

Figure		Page
2.1	The monogastric intestinal tract of a chicken	8
2.2	The subdivision of the avian GIT in relation to its microbial population: crop, proventriculus, ileum, ceca, colon and cloaca	10
2.3	Changes in composition of the GIT microbiota due to stress, transportation and other factors	26
2.4	Villi and crypt structure	38
2.5	The TaqMan 5' exonuclease assay	50
2.6	DNA polymerase 5' exonuclease activity	51
2.7	Representation of real time PCR with SYBR Green	52
3.1	Biostat Q multi-fermentor system (stirred tank bioreactor)	59
4.1	Mastercycler gradient PCR machine, Eppendorf, Hamburg, Germany	79
4.2	Computer operated Rotorgene 3000 Real time-PCR system (Corbett Research, Australia)	84
A.1	Gel electrophoresis image of bacterial DNA after extraction of cells	168
A.2	Gel electrophoresis image of PCR product from plasmid DNA that carry target sequence of <i>C. butyricum</i> (wells containing the same PCR products which run in different annealing temperatures to optimize it)	168
A.3	Gel electrophoresis image of PCR product from plasmid DNA that carry target sequence of <i>E.coli</i> (wells containing the same PCR products which run in different annealing temperatures to optimize it)	169
A.4	Gel electrophoresis image of PCR product from plasmid	170



	DNA that carry target sequences of <i>Lactobacillus</i> , <i>Bifidobacterium</i> and <i>F. parazinitzi</i> (wells containing the same PCR products in each section which run in different annealing temperatures to optimize it)	
A.5	Gel electrophoresis image of PCR product from plasmid DNA that carry target sequence of <i>Salmonella</i> (wells containing the same PCR products which run in different annealing temperatures to optimize it)	171
A.6	Post-amplification melting curve analysis of <i>Bifidobacterium</i> . The rate of change of fluorescence against temperature shows the specificity of the amplification	172
A.7	Amplification plot with threshold line of <i>Bifidobacterium</i>	172
A.8	<i>Bifidobacterium</i> standard curve generated by real time pcr machine (relation between the threshold cycle and number of <i>Bifidobacterium</i> 16s rRNA gene per microliter)	173
A.9	Post-amplification melting curve analysis of <i>Clostridium butyricum</i> . The rate of change of fluorescence against temperature shows the specificity of the amplification standard melting curve	173
A.10	Amplification plot with threshold line of <i>C. butyricum</i>	174
A.11	<i>Clostridium butyricum</i> standard curve generated by real time PCR (relation between the threshold cycle and number of <i>C. butyricum</i> 16s rRNA gene per microliter)	174
A.12	Post-amplification melting curve analysis of <i>E.coli</i> . The rate of change of fluorescence against temperature shows the specificity of the amplification standard melting curve	175
A.13	Amplification plot with threshold line of <i>E.coli</i>	175
A.14	<i>E.coli</i> standard curve generated by real time PCR (relation between the threshold cycle and number of <i>E.coli</i> 16s rRNA gene per microliter)	176



A.15	Post-amplification melting curve analysis of <i>Faecalibacterium parazinitzi</i> . The rate of change of fluorescence against temperature shows the specificity of the amplification standard melting curve	176
A.16	Amplification plot with threshold line of <i>F. parazinitzi</i>	177
A.17	<i>F. parazinitzi</i> standard curve generated by real time PCR (relation between the threshold cycle and number of <i>F. parazinitzi</i> 16s rRNA gene per microliter)	177
A.18	Post-amplification melting curve analysis of <i>Lactobacillus</i> . The rate of change of fluorescence against temperature shows the specificity of the amplification standard melting curve	178
A.19	Amplification plot with threshold line of <i>Lactobacillus</i>	178
A.20	<i>Lactobacillus</i> standard curve generated by real time PCR (relation between the threshold cycle and number of <i>Lactobacillus</i> 16s rRNA gene per microliter)	179
A.21	Post-amplification melting curve analysis of <i>Salmonella</i> . The rate of change of fluorescence against temperature shows the specificity of the amplification standard melting curve	179
A.22	Amplification plot with threshold line of <i>Salmonella</i>	180
A.23	<i>Salmonella</i> standard curve generated by real time PCR (relation between the threshold cycle and number of <i>Salmonella</i> 16s rRNA gene per microliter)	180
A.24	Duodenal villi height (VH) and crypt depth (CD) in broiler chicken	181
A.25	Jejunal villi height (VH) and crypt depth (CD) in broiler chicken	182
A.26	Ileal villi height (VH) and crypt depth (CD) in broiler Chicken	183
A.27	Ethical approval for project "Effects of unpleasant	184

