



**EVALUATION OF FERMENTED PLANT EXTRACTS AND WAX
DEGRADING BACTERIA ON *Phenacoccus solenopsis* Tinsley OF *Hibiscus* sp.**

By

SULTAN AHMMED

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

January 2023

FP 2023 22

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior and written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

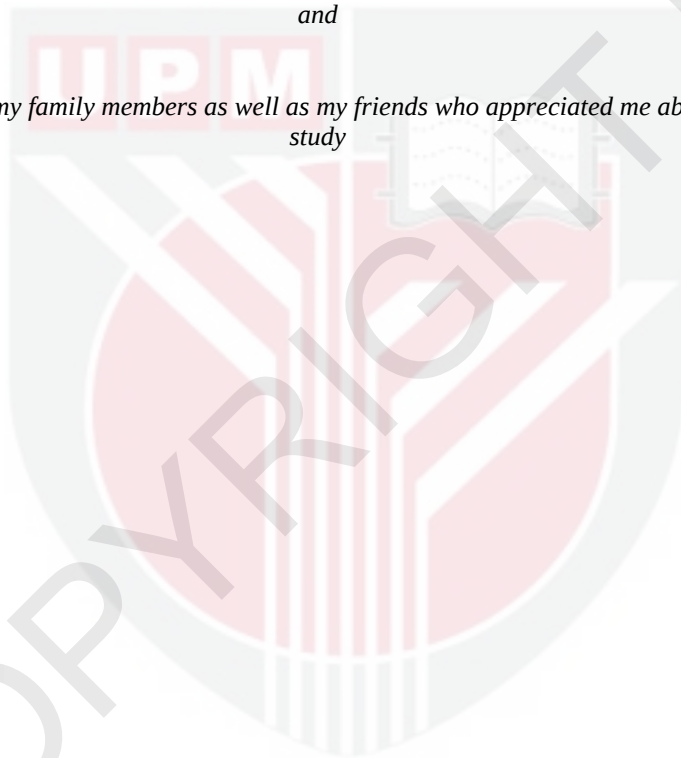
*I dedicated my thesis to beloved my mother, my wife and my son for their prayer,
patient and support*

During my study to achieve my goal

Grateful to Almighty Allah

and

*Thanks to my family members as well as my friends who appreciated me about my
study*



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in partial fulfilment of the requirement for the degree of Doctor of Philosophy

**EVALUATION OF FERMENTED PLANT EXTRACTS AND WAX
DEGRADING BACTERIA ON *Phenacoccus solenopsis* Tinsley OF *Hibiscus* sp.**

By

SULTAN AHMMED

January 2023

Chairperson : Associate Professor Lau Wei Hong, PhD
Faculty : Agriculture

The mealybugs collected from hibiscus plants were confirmed as *Phenacoccus solenopsis*, *Maconellicoccus hirsurtus*, and *Paracoccus marginatus* based on their morphological characteristics and genes analysis such as 18S and 28S ribosomal, and COI mitochondrial genes. The percentage infestation of *P. solenopsis* was 40%, followed by *P. marginatus* (30%) and *M. hirsurtus* (15%) on *Hibiscus rosa sinensis* plants during specimen collection. *Phenacoccus solenopsis* was selected for further study based on the infestation level. It is one of the noxious sucking polyphagous pests causing severe losses and its control has been an issue of significance in pest management. Management of mealybug is challenging because of its wide host range, presence of a waxy coating on the body, and high reproductive potentiality. Fermented plant extracts (FPEs) and their wax degrading bacteria were screened for their relative toxicity against *P. solenopsis*. A total of 13 plant materials namely peppermint, Mexican mint, onion, garlic, turmeric, lemongrass, variegated mint, kaffir lime, lime, neem, galangal, mahogany, and ficus were selected for FPEs study based on their insecticidal properties. The FPEs of kaffir lime, turmeric, and ficus were effective in controlling *P. solenopsis* and scored the lowest LC₅₀ value among the tested FPEs. All tested FPEs did not induce phytotoxicity to hibiscus leaves below 10% concentration (w/v) after 120 h postexposure, however, FPE of variegated mint and ficus had induced severe phytotoxic effect at >20% concentration to hibiscus leaves after 72 h post exposures. All FPEs were also evaluated for their repellency potential. Among the tested FPEs, fermented peppermint extract demonstrated the best repellent action against *P. solenopsis* after 1 h post exposure and it contained hydroxy-alpha-terpenyl acetate, 2-hydroxy methyl ester tetradecanoic acid, nonanol, and 2-heptanol attributed to repellent property while dropped after 24 h post exposure. The FPE of kaffir lime, turmeric, and ficus had less repellent action after 1 h and 3 h post exposure, however, fermented ficus extract could repel more than 50% *P. solenopsis* after 24 h post exposure. Based on repellent activity FPEs of pepper mint, Mexican mint, variegated mint, kaffir lime, galangal, lemongrass, and mahogany belonged to Class II (low repellent) after 24 h post exposure. FPEs of onion, turmeric, garlic and neem belonged to Class III (moderate repellent) and only fermented ficus extract belonged to Class IV

(high repellent) after 24 h post exposure. On contrary, FPEs of pepper mint, Mexican mint, variegated mint, lemongrass, lime, and mahogany exhibited decreased repellent activity while FPEs of ficus, onion, turmeric, garlic, neem, kaffir lime, and galangal showed increasing repellent activity with exposure period. All the FPEs contained phenolic and alcoholic compounds in very trace amounts while butanoic acid was the highest abundant in the FPEs of ficus attributed to the highest repellent activity (Class IV) for *P. solenopsis*. Based on all the test results, fermented ficus extract was selected for waxdegrading bacteria screening. A total of 10 bacteria were isolated from the fermented ficus extract and confirmed by morphological and molecular characterization. Among the tested bacteria isolates, *Klebsiella pneumoniae* 0F1B1 was highly disruptive (83 % mortality) and inhibited the growth of *P. solenopsis* compared to other isolates. *Bacillus* sp. 42F1B3 exhibited the best mealybug wax degrading ability. In the glasshouse trial, fermented ficus extract performed the best in terms of the percentage reduction of *P. solenopsis* without any remarkable phytotoxicity on *H. rosa sinensis*, followed by kaffir lime and turmeric. It was noticed that *H. cannabinas* was more prone to phytotoxicity damage by FPEs than *H. rosasinensis*. Plant growth and photosynthesis were more inhibited in the FPE of turmeric-treated plants than the FPEs of kaffir lime and ficus-treated plants during the glasshouse trial, but all affected plants recovered after 15 days post treatment. The findings of the current study suggest the fermented kaffir lime extract may have beneficial effects on the growth of hibiscus plants along with mealybug control. In conclusion, FPE of ficus and *Klebsiella pneumoniae* 0F1B1 as wax degrading bacterium are good candidates to control *P. solenopsis* either as an insecticide or repellent. Different plants may have different responses to the phytotoxicity effect of FPEs. Therefore, the FPE of kaffir lime is suggested as a good candidate for the management of *P. solenopsis* in hibiscus plants.

Keywords: Fermentation, plant extract, *Phenococcus solenopsis*, wax degrading bacteria

SDG: Responsible consumption and production, zero hunger, no poverty

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENILAIAN EKSTRAK FERMENTASI TUMBUHAN DAN BAKTERIA
PENGURAI LILIN TERHADAP *Phenacoccus solenopsis* Tinsley *Hibiscus* sp.**

Oleh

SULTAN AHMMED

Januari 2023

Pengerusi : Profesor Madya Lau Wei Hong, PhD
Fakulti : Pertanian

Koya yang dikumpul daripada pokok bunga raya telah disahkan sebagai *Phenacoccus solenopsis*, *Maconellicoccus hirsurtus* dan *Paracoccus marginatus* berdasarkan ciri-ciri morfologi dan analisis gen seperti 18S dan 28S ribosom, dan gen mitokondria COI. Peratusan serangan *P. solenopsis* ialah 40%, diikuti dengan *P. marginatus* (30%) dan *M. hirsurtus* (15%) pada pokok *Hibiscus rosa sinensis* semasa pengumpulan spesimen. *Phenacoccus solenopsis* telah dipilih untuk kajian selanjutnya berdasarkan kepada kadar serangan. *Phenacoccus solenopsis* merupakan salah satu jenis perosak penghisap polifagus yang berbahaya yang boleh menyebabkan kerugian teruk dan kawalannya telah menjadi satu isu penting dalam pengurusan perosak. Pengurusan koya adalah mencabar kerana julat perumahannya yang luas, kehadiran salutan berlilin pada badan dan pembiakan yang tinggi. Ekstrak fermentasi tumbuhan (FPE) dan bakteria yang mengurangkan lilinnya telah disaring untuk kajian ketoksikan relatifnya terhadap *P. solenopsis*. Sebanyak 13 bahan tumbuhan iaitu pudina, pudina Mexico, bawang merah, bawang putih, kunyit, serai, pudina bervariasi, limau purut, limau, mambu, lengkuas, mahogani dan ficus telah dipilih untuk kajian FPE berdasarkan kepada sifat keracunan terhadap serangga. FPE limau purut, kunyit dan ficus didapati berkesan dalam pengawalan *P. solenopsis* dan mencapai nilai LC_{50} terendah berbanding dengan FPE lain yang diuji. Semua FPE yang diuji tidak menyebabkan fitotoksiti terhadap daun bunga raya di bawah kepekatan 10% (w/v) selepas 120 jam pasca pendedahan. Walau bagaimanapun, FPE pudina bervariasi dan ficus telah menyebabkan kesan fitotoksik yang teruk pada kepekatan >20% pada daun bunga raya selepas 72 jam pasca pendedahan. Semua FPE telah dinilai untuk potensi pencegahannya. Antara FPE yang diuji, ekstrak fermentasi pudina telah menunjukkan tindakan pencegahan serangga yang terbaik terhadap *P. solenopsis* selepas pendedahan 1 jam pasca pendedahan dan ia mengandungi hidroksi-alfa-terpenil asetat, asid tetradekanoik 2-hidroksi metil ester, nonanol dan 2-heptanol yang bersifat pencegah serangga. Walau bagaimanapun, aktiviti pencegahan serangga oleh ekstrak fermentasi pudina menurun selepas 24 jam pasca pendedahan. FPE limau purut, kunyit dan ficus mempunyai tindakan pencegahan serangga yang menurun selepas 1 jam dan 3 jam pasca pendedahan manakala ekstrak fermentasi ficus boleh mencegah lebih daripada 50% *P. solenopsis* selepas 24 jam

pasca pendedahan. Berdasarkan kepada aktiviti pencegahan serangga oleh FPE pudina, pudina Mexico, pudina bervariasi, limau purut, lengkuas, serai dan mahogani tergolong dalam Kelas II (pencegah serangga rendah) selepas 24 jam pasca pendedahan. FPE bawang merah, kunyit, bawang putih dan mambu tergolong dalam Kelas III (pencegah serangga sederhana) dan hanya ekstrak fermentasi ficus tergolong dalam Kelas IV (pencegah serangga tinggi) selepas 24 jam pasca pendedahan. Sebaliknya, FPE pudina, pudina Mexico, pudina bervariasi, serai, limau dan mahogani menunjukkan penurunan aktiviti mencegah serangga manakala FPE ficus, bawang, kunyit, bawang putih, mambu, limau purut dan lengkuas menunjukkan peningkatan aktiviti pencegahan serangga dengan tempoh pendedahan yang meningkat. Semua FPE mengandungi sebatian fenol dan alkohol dalam jumlah yang sangat sedikit manakala asid butanoik adalah sebatian yang paling banyak dalam FPE ficus yang memberi kesan pencegahan serangga yang tertinggi (Kelas IV) terhadap *P. solenopsis*. Berdasarkan kepada semua keputusan ujian yang dijalankan, ekstrak fermentasi ficus dipilih untuk menyaring bakteria yang boleh mengurai lilin. Sebanyak 10 bakteria telah dipencilkan daripada ekstrak fermentasi ficus dan disahkan dengan pencirian morfologi dan molekul. Antara bakteria yang diuji, *Klebsiella pneumoniae* 0F1B1 adalah sangat berkesan (83 % kematian) dan menghalang pertumbuhan *P. solenopsis* berbanding dengan bakteria yang lain. *Bacillus* sp. 42PB3 mempamerkan keupayaan mengurai lilin koya yang terbaik. Dalam kajian di rumah kaca, ekstrak fermentasi ficus menunjukkan prestasi terbaik dari segi pengurangan peratusan *P. solenopsis* tanpa sebarang fitotoksiti pada *H. rosa sinensis*, diikuti dengan limau purut dan kunyit. *Hibiscus cannabinas* lebih cepat menunjukkan kerosakan fitotoksiti oleh FPE berbanding dengan *H. rosa sinensis*. Pertumbuhan tumbuhan dan fotosintesis boleh dihalang dengan rawatan FPE kunyit berbanding dengan FPE limau purut dan ficus semasa percubaan di rumah kaca, tetapi semua tumbuhan yang terjejas telah pulih selepas 15 hari pasca rawatan. Penemuan kajian ini mencadangkan ekstrak fermentasi limau purut memberi manfaat kepada pertumbuhan pokok bunga raya bersama dengan kawalan koya. Kesimpulannya, FPE ficus dan *K. pneumoniae* 0F1B1 adalah calon yang baik untuk mengawal *P. solenopsis* sama ada sebagai racun atau pencegah serangga. Tumbuhan yang berbeza mungkin mempunyai tindak balas yang berbeza terhadap kesan fitotoksiti FPE. Oleh itu, FPE limau purut dicadangkan sebagai calon yang baik untuk pengawalan *P. solenopsis* pada pokok bunga raya.

Kata kunci: Fermentasi, ekstrak tumbuhan, *Phenococcus solenopsis*, bakteria pelupus lilin

SDG: Penggunaan dan pengeluaran yang bertanggungjawab, sifar kelaparan, tiada kemiskinan

ACKNOWLEDGEMENTS

First of all, I am most grateful to Almighty Allah for giving me the strength and courage to complete the writing of this research thesis. I would like to express my sincere gratitude to dear supervisor, Associate Professor Dr. Lau Wei Hong for her intellectual guidance and supervision, productive discussion, constructive criticisms, advice and continuous encouragement throughout the study. I am very much grateful and indebted to supervisory committee members, Professor Dr. Uma Rani a/p Sinniah and Associate professor Dr. Nur Azura binti Adam for their constructive suggestions and guidance in all phases of this research project. I express my deep sense of respect to all of the lecturers and professors in the Department of Plant protection for their valuable suggestions and constructive comments. I would like to thank my lab mates, especially Vinailosni A/P Amirthalingam, Mr. Amali, Mrs. Erise Anggraini, Mr. Ahad Gul Khadem and Khairul Anuar Muhammad Issa for their valuable support in my research work, I am grateful to all the technical laboratory staff of the Department of Plant Protection, Faculty of Agriculture.

I am indebted to my mother Jamila Begum, wife Mumtahana Yasmin, son Muttmain Ahmad and brothers for their prayer, encouragement and tolerance during the whole period of the study. A special thanks to my dear Father in-law Professor Dr. Abdullahel Baqui and Mother in-law Nurun Naher Begum for their prayer and encouragement throughout the study.

Thankfully acknowledge the assistance of the National Agricultural Technology Program- Phase 2 project (NATP-2), Bangladesh Agricultural Research Council (BARC), Farmgate, Dhaka-1215, Bangladesh, for financial support and facilitating my study and University Putra Malaysia for providing research facilities under the Department of Plant Protection, Faculty of Agriculture.

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements of the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Lau Wei Hong, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Uma Rani a/p Sinniah, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Nur Azura binti Adam, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 13 June 2024

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xviii
LIST OF ABBREVIATIONS	xxiii
 CHAPTER	
 1 INTRODUCTION	 1
 2 LITERATURE REVIEW	 4
2.1 Hibiscus (<i>Hibiscus rosa-sinensis</i>)	4
2.2 Kenaf (<i>Hibiscus cannabinus</i>)	5
2.3 Mealybugs	5
2.3.1 <i>Phenacoccus solenopsis</i> (Tinsley)	6
2.3.2 <i>Maconellicoccus hirsutus</i> (Green)	8
2.3.3 <i>Paracoccus marginatus</i> (Williams and Granara deWillink)	9
2.3.4 Approaches in Mealybug identification	11
2.3.5 Distribution of mealybugs	12
2.3.6 Host range	13
2.3.7 Scenario of mealybug infection on <i>Hibiscus</i> sp.	14
2.3.8 Nature of damage	15
2.3.9 Management of mealybugs	16
2.4 Garbage enzyme or fermented plant extract	24
2.4.1 Fermentation process	25
2.4.2 Microbes in fermentation	25
2.4.3 Applications of fermented plant extract	26
2.4.4 Wax degrading bacteria	28
 3 MORPHOLOGICAL AND MOLECULAR CHARACTERIZATION OF MEALYBUGS INFESTED ON <i>Hibiscus rosa sinensis</i>	 30
3.1 Introduction	30
3.2 Materials and methods	31
3.2.1 Sample collection	31
3.2.2 Morphological study	32
3.2.3 DNA extraction	32
3.2.4 Gene Amplification by PCR	33
3.2.5 Sequence analysis	33
3.2.6 Phylogenetic analysis	34
3.3 Results	36
3.3.1 Morphological characterization	36

3.3.2	Molecular analysis	41
3.4	Discussion	44
3.5	Conclusion	46
4	EVALUATION OF FERMENTED PLANT EXTRACTS AGAINST MEALYBUGS ON HIBISCUS AT LABORATORY CONDITION	47
4.1	Introduction	47
4.2	Materials and methods	48
4.2.1	Sample collection	48
4.2.2	Preparation of host plant for mealybug rearing	48
4.2.3	Rearing of mealybugs	48
4.2.4	Fermented plant extracts (FPEs)	49
4.2.5	Bioassay of FPEs on <i>P. solenopsis</i>	52
4.2.6	Phytotoxicity of FPE on <i>Hibiscus rosa sinensis</i> leaves	53
4.3	Result	54
4.3.1	Effect of FPE on the mortality of <i>P. solenopsis</i> after 24 h post treatment	54
4.3.2	Effect of FPE on the mortality of <i>P. solenopsis</i> after 72 h post treatment	54
4.3.3	Effect of FPE on the mortality of <i>P. solenopsis</i> after 120 h post treatment	55
4.3.4	Mean lethal concentration of FPE against <i>P. solenopsis</i>	57
4.3.5	Phytotoxicity effect of FPEs on hibiscus leaf	57
4.4	Discussion	60
4.5	Conclusion	63
5	CHEMICAL PROFILING AND REPELLENCY RESPONSE OF FERMENTED PLANT EXTRACTS AGAINST PHENACOCOCCUS SOLENOPSIS	64
5.1	Introduction	64
5.2	Materials and methods	64
5.2.1	Study area	64
5.2.2	Collection and rearing of <i>Phenacoccus solenopsis</i>	65
5.2.3	Fermented Plant Extract	65
5.3	Results	67
5.3.1	Repellent activity of FPEs on <i>P. solenopsis</i> after 1h post treatment	67
5.3.2	Repellent activity of FPEs on <i>P. solenopsis</i> after 3 h post treatment	68
5.3.3	Repellent activity of FPEs on <i>P. solenopsis</i> after 6 h post treatment	69
5.3.4	Repellent activity of FPEs on <i>P. solenopsis</i> after 24 h post treatment	70
5.3.5	Repellent activity of FPEs on <i>P. solenopsis</i> versus time of post treatment	71
5.3.6	Volatile bioactive compounds of FPEs	72
5.4	Discussion	89
5.5	Conclusion	94

6	IDENTIFICATION OF WAX DEGRADING BACTERIA FROM FERMENTED PLANT EXTRACTS AND EVALUATION AGAINST <i>Phenacoccus solenopsis</i>	95
6.1	Introduction	95
6.2	Materials and methods	96
6.2.1	Study Area	96
6.2.2	Maintenance of mealybug rearing	96
6.2.3	Preparation of fermented plant extracts	96
6.2.4	Preparation of culture media	96
6.2.5	Wax extraction from mealybug	96
6.2.6	Isolation of wax degrading bacteria from fermented ficus extract	97
6.2.7	Gram staining of bacteria	97
6.2.8	Microscopic analysis of bacteria	97
6.2.9	Molecular identification of bacteria	98
6.2.10	Wax degradation (Turbidimetry) screening with redox indicator 2,6-dichlorophenol indophenol (DCPIP)	101
6.2.11	Preparation of bacterial culture for bioassay	101
6.2.12	Insecticidal bioassay	102
6.2.13	Mealybug wax removal test	103
6.2.14	Effect of bacterial cell suspension on the body weight of mealybugs	104
6.2.15	Effect of bacterial cell suspension on population growth of mealybugs	105
6.3	Result	105
6.3.1	pH versus bacterial colony of fermented ficus extract	105
6.3.2	Morphological characterization of bacteria isolated from fermented ficus extract	107
6.3.3	Molecular characterization	110
6.3.4	Bioassay of bacteria on <i>P. solenopsis</i>	119
6.3.5	Wax degradation test	126
6.3.6	Effect of bacteria on offspring production and body weight of malybugs	129
6.3.7	Detection of hydrocarbon degrading genes	129
6.4	Discussion	132
6.5	Conclusion	136
7	EFFICACY OF FERMENTED PLANT EXTRACTS against <i>Phenacoccus solenopsis</i> ON <i>Hibiscus</i> spp. UNDER GLASS HOUSE CONDITION	137
7.1	Introduction	137
7.2	Materials and methods	138
7.2.1	Study area	138
7.2.2	Cultivation of <i>Hibiscus</i> species	138
7.2.3	Rearing of <i>Phenacoccus solenopsis</i>	139
7.2.4	Infestation of plants with <i>P. solenopsis</i>	139
7.2.5	Application of fermented plant extract (FPEs)	139
7.2.6	Measurement of gas exchange	140
7.2.7	Relative chlorophyll content	141

7.2.8	Growth and biomass measurement	142
7.2.9	Phytotoxicity test	142
7.2.10	Data analysis	143
7.3	Result	143
7.3.1	FPE against <i>P. solenopsis</i> on <i>H. cannabinas</i>	143
7.3.2	FPE against <i>P. solenopsis</i> on <i>Hibiscus rosa sinensis</i>	144
7.3.3	Physiological response	146
7.3.4	Effect of FPE on the plant growth of <i>H. rosa sinensis</i>	150
7.3.5	Effect of FPE on the plant growth of <i>Hibiscus cannabinus</i>	151
7.3.6	Phytotoxic effect of FPEs on <i>Hibiscus</i> spp. leaves	155
7.4	Discussion	157
7.5	Conclusion	161
8	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS	162
	REFERENCES	165
	APPENDICES	217
	BIODATA OF STUDENT	289
	LIST OF PUBLICATIONS	290

LIST OF TABLES

Table	Page
2.1 The mode of action of the chemical insecticides used in mealybug control	17
2.2 Common botanicals used in mealybug control	19
2.3 Biological management of mealybugs	24
3.1 Molecular markers used in the identification of mealybugs found on <i>H. rosa sinensis</i>	34
3.2 Reference sequences used in the phylogenetic tree construction	35
3.3 ID of mealybug species collected from <i>Hibiscus rosa sinensis</i> in Malaysia	42
4.1 List of plants and plant parts selected for preparation of FPEs	49
4.2 Mass of the freeze-dried material of FPEs	52
4.3 Phytotoxicity rating scale on leaves of <i>Hibiscus rosa sinensis</i> due to the treatment of FPEs	53
4.4 Response of <i>P. solenopsis</i> to different FPEs at different concentrations after different time of exposure. Values express in Mean $\pm$ S.E	56
4.5 LC ₅₀ of FPEs against 3rd instars nymphs of <i>P. solenopsis</i>	57
4.6 Phytotoxic grade of FPE on <i>H. rosa sinensis</i> leaf	59
5.1 Repellency rating scale due to response of FPEs to <i>Hibiscus rosa sinensis</i>	66
5.2 Mean percentage repellency of FPEs on <i>P. solenopsis</i> after 1h post treatment	68
5.3 Mean percentage repellency of FPEs on <i>P. solenopsis</i> after 3h post treatment	69
5.4 Mean percentage repellency of FPEs on <i>P. solenopsis</i> after 6 h post treatment	70
5.5 Mean percentage repellency of FPEs <i>P. solenopsis</i> after 24 h post treatment	71

5.6	Class of FPEs at 25% concentration within 24 h post treatment	72
5.7	Volatile bioactive compounds attributed to repellent activity	74
6.1	Universal molecular markers used in bacteria identification	99
6.2	Molecular markers used in the screening of wax-degrading bacteria isolated from fermented ficus extract	99
6.3	Isolation of bacteria from fermented ficus extract at different interval of fermentation time	106
6.4	Morphological characteristics of bacteria isolated from fermented ficus extract	108
6.5	Molecular identity of wax degrading bacteria isolated from fermented ficus extract using <i>16S rRNA</i> gene	111
6.6	Molecular identity of wax degrading bacteria isolated from fermented ficus extract using <i>motB</i> gene	113
6.7	Molecular identity of wax degrading bacteria isolated from fermented ficus extract using <i>ropB</i> gene	115
6.8	Molecular identity of wax degrading bacteria isolated from fermented plant extract using <i>recQ</i> gene	117
6.9	Output of molecular identification of bacteria isolates by using different markers	119
6.10	Mean percentage mortality of <i>Phenacoccus solenopsis</i> (leaf dipping method) after exposure to different concentrations of bacteria isolated from fermented ficus extract	121
6.11	Mean percentage mortality of <i>Phenacoccus solenopsis</i> (Spraying method) after exposure to different concentrations of bacteria isolated from fermented ficus extract	123
6.12	Mean percentage nymphal mortality of <i>Phenacoccus solenopsis</i> after exposure to different bacterial supernatant	125
6.13	<i>In vitro</i> wax degradation performance of bacterial isolates	127
6.14	Effect of bacterial suspension on the offspring production and body weight of <i>Phenacoccus solenopsis</i> at 10^5 CFU/mL. Values express (Mean $\pm$ S.E)	129
6.15	Molecular identity of wax degrading bacteria isolated from fermented ficus extract using <i>Catechol-2,3-dioxygenase</i> and <i>almA</i> genes	131

7.1	Mean percent cumulative reduction of <i>P. solenopsis</i> after treated with fermented plant extracts on <i>H. cannabinus</i> . Values express (Mean $\pm$ S.E)	145
7.2	Mean percent cumulative reduction of <i>P. solenopsis</i> after treated with fermented plants extract on <i>H. rosa sinensis</i> . Values express (Mean $\pm$ S.E)	145
7.3	Pearson correlation between photosynthetic rate, stomatal conductance and transpiration rate	150
7.4	Effect of fermented plant extracts on the plant growth of <i>H. rosa sinensis</i> (Mean $\pm$ S.E)	152
7.5	Effect of fermented plant extracts on the plant growth of <i>H. cannabinas</i> (Mean $\pm$ S.E)	152

LIST OF FIGURES

Figure	Page
2.1 Different stages of <i>Phenacoccus solenopsis</i> . A) egg, B) first instar, (crawler), C) second instar, D) third instar, E) adult female, F) oviposition	6
2.2 Different stages of <i>maconellicoccus hirsutus</i> . A) egg, B) first instar, C) second instar, D) third instar, E) fourth instar, F) gravid female, G) ovisac, H) adult male	9
2.3 Different stages of <i>Paracoccus marginatus</i>	10
2.4 Oxidation pathways for the metabolism of long chain hydrocarbons waxes	29
3.1 Mealybug samples collected from Infested <i>Hibiscus rosa sinensis</i> plant. A) <i>Phenacoccus solenopsis</i> , B) <i>Maconellicoccus hirsutus</i> , and C) <i>Paracoccus marginatus</i> .	31
3.2 Different mealybug species infested hibiscus plant. A) <i>Phenacoccus solenopsis</i> , B) <i>Maconellicoccus hirsutus</i> , and C) <i>Paracoccus marginatus</i> .	37
3.3 SEM view of <i>P. solenopsis</i> . A) 9-segmented antenna, B) leg, C) digitules, D) ostiole, E) cerarius with trilocular pore and only two conical setae, F) circulus, G) multilocular pore, H) trilocular pore, I) glassy wax rods, J) tribular duct, K) anal bar and L) waxy structure	38
3.4 SEM view of <i>M. hirsutus</i> . A) Dorsal view of adult female, B) ventral view of adult female, C) 9- segmented antenna, D) Well developed hindleg E) ostiole, F) trilocular pore (T), G) tibular duct, H) circulus, I) multilocular pore (M) and J) anal bar. S: setae	39
3.5 SEM view of <i>P. marginatus</i> . A) 8-segmented antenna, B) translucent pores on the hind coxa, C) conical seta with trilocular pore, D) digitules, E) ostiole, F) trilocular pore, G) vulva, H) multilocular pore, I) tibular duct	40
3.6 Amplification of 18S, 28S and CO1 genes of mealybugs. Lane 1: CSL- MDNA-1KbPLUS DNA Ladder RTU; Lane 2-4: primer 18S; Lane 5-7: primer 28S; Lane 8-10: primer CO1(1718); and Lane 11-13: primer CO1(2183). PS: <i>Phenacoccus solenopsis</i> Tinsley; MH: <i>Maconellicoccus hirsutus</i> ; PM: <i>Paracoccus marginatus</i> . PCR products were electrophoresed on 1% (w/v) agarose gels	41

3.7	Phylogenetic tree based on <i>18S</i> ribosomal haplotypes using Maximum Likelihood method	43
3.8	Phylogenetic tree based on <i>COI</i> (<i>mtDNA</i>) mitochondrial haplotypes using Maximum Likelihood method	43
3.9	Phylogenetic tree based on <i>28S</i> ribosomal haplotypes using Maximum Likelihood method	44
4.1	Mealybug collection and rearing. (A) Mealybug on <i>Hibiscus rosa sinensis</i> plants, (B) mealybug rearing on green okra fruits, and (C) mealybug rearing on sprouted potatoes	48
4.2	Plant materials for the FPE preparation. a) peppermint, b) Mexican mint, c) Variegated mint, d) lemongrass, e) ficus, f) galanga, g) neem, h) mahogany, i) garlic, j) onion, K) turmeric, L) kaffir lime and M) lime	50
4.3	Preparation of plant materials for fermentation. (a) and (b) cutting plant materials by knife, and (c) mixture of FPE in plastic containers	51
4.4	Filtration of FP	51
4.5	Phytotoxicity effect of fermented variegated mint extract at different concentrations after 120 h post treatment. G1T1: 5% FPE, G2T8: 10% FPE, G3T2: 15% FPE, G4T2: 20% FPE, and G5T2: 25% FPE	60
5.1	Repellency test of FPEs against <i>P. solenopsis</i> . (a) Air drying of treated and untreated portions of filter papers, and (b) releasing of <i>P. solenopsis</i> on the filter paper	66
5.2	Repellent response of FPEs to <i>P. solenopsis</i> at different exposure time	72
5.3	GCMS Chromatogram of fermented ficus extract	76
5.4	GCMS Chromatogram of fermented onion extract	77
5.5	GCMS Chromatogram of fermented garlic extract	78
5.6	GCMS Chromatogram of fermented turmeric extract	79
5.7	GCMS Chromatogram of fermented lime extract	80
5.8	GCMS Chromatogram of fermented neem extract	81
5.9	GCMS Chromatogram of fermented peppermint extract	82
5.10	GCMS Chromatogram of fermented kaffir lime extract	83

5.11	GCMS Chromatogram of fermented galangal extract	85
5.12	GCMS Chromatogram of fermented lemongrass extract	86
5.13	GCMS Chromatogram of fermented variegated mint extract	87
5.14	GCMS Chromatogram of fermented Mexican mint extract	88
5.15	GCMS Chromatogram in fermented mahogany extract	89
6.1	Treated hibiscus leaf with mealybug in a 9 cm petri dish	102
6.2	pH record during fermentation of ficus leaves	106
6.3	Wax-degrading bacteria isolated from fermented ficus extract	107
6.4	Colony characteristics of fermented ficus extract bacterial isolates. (1) 0F1B1, (2) 0F1S1, (3) 0F1S2, (4) 7F2A4, (5) 14F1C3, (6) 21F1E3, (7) 35F2B1, (8) 35F2B5 (9) 35F1C1, (10) 42F2B2, (11) 42F1B3, (12) 49F1B1, and (13) 49F1C1. Magnification 40X	109
6.5	Gram staining of bacteria isolated from fermented ficus extract. Magnification 100X	109
6.6	Amplification of 16S <i>rRNA</i> gene of bacteria isolates. Lane L: 10 kb DNA Ladder, Lane 1: 0F1B1, lane 2: 7F2A4, lane 3: 14F1C3, lane 4: 21F1E3, lane 5: 35F3B1, lane 6: 35F2B5, lane 7: 35F1C1, lane 8: 42F2B2, lane 9: 49F1B1, lane 10: 49F1C1, lane 11: 42F1B3, lane 12: 0F1S1 and lane 13: 0F1S2	110
6.7	Neighbor-joining analysis of bacterial sequences isolated from fermented ficus extract based on 16S <i>gene</i>	112
6.8	Amplification of <i>motB</i> gene of bacteria isolates. Lane L: 10 kb DNA Ladder, Lane 1: 0F1B1, lane 2: 7F2A4, lane 3: 14F1C3, lane 4: 21F1E3, lane 5: 35F3B1, lane 6: 35F2B5, lane 7: 35F1C1, lane 8: 42F2B2, lane 9: 49F1B1, lane 10: 49F1C1, lane 11: 42F1B3, lane 12: 0F1S1 and lane 13: 0F1S2	113
6.9	Neighbor-joining analysis of bacterial sequences isolated from fermented ficus extract based on <i>motB</i> gene	114
6.10	Amplification of <i>ropB</i> gene of bacteria isolates. Lane L: 10 kb DNA Ladder, Lane 1: 0F1B1, lane 2: 7F2A4, lane 3: 14F1C3, lane 4: 21F1E3, lane 5: 35F3B1, lane 6: 35F2B5, lane 7: 35F1C1, lane 8: 42F2B2, lane 9: 49F1B1, lane 10: 49F1C1, lane 11: 42F1B3, lane 12: 0F1S1 and lane 13: 0F1S2	115
6.11	Neighbor-joining analysis of bacterial sequences isolated from fermented ficus extract based on <i>ropB</i> gene	116

6.12	Amplification of <i>recQ</i> gene of bacteria isolates. Lane L: 10 kb DNA Ladder, Lane 1:0F1B1, lane 2: 7F2A4, lane 3: 141F1C3, lane 4: 21F1E3, lane 5: 35F3B1, lane 6: 35F2B5, lane 7: 35F1C1, lane 8: 42F2B2, lane 9: 49F1B1, lane 10: 49F1C1, lane 11:42F1B3, lane 12: 0F1S1 and lane 13: 0F1S2	117
6.13	Neighbor-joining analysis of bacterial sequences isolated from fermented ficus extract using on <i>recQ</i> gene	118
6.14	<i>In vitro</i> wax degradation trend by different bacteria isolates (0F1B1= <i>K. pneumoniae</i> , 21F1E3= <i>Paenibacillus</i> sp., 35F1C1= <i>Bacillus</i> sp.1=35F1C1 , <i>Bacillus</i> sp. 2=42F1B3, Control= Water)	126
6.15	Mealybug wax degradation screening by DCPIP for 288 h	127
6.16	Paraffin wax degradation screening by DCPIP for 288 h	127
6.17	Percent wax removal from mealybug body. (0F1B1= <i>K. pneumoniae</i> , 21F1E3= <i>Paenibacillus</i> sp., 35F1C1= <i>Bacillus</i> sp.1=35F1C1 , <i>Bacillus</i> sp. 2=42F1B3, Control= Water)	128
6.18	Bacteria treated mealybugs after 48 h. (1) <i>K. pneumoniae</i> 0F1B1, (2) <i>Paenibacillus</i> sp. 21F1E3, (3) <i>Bacillus</i> sp. 1-35F1C1, (4) <i>Bacillus</i> sp. 2-42F1B3, and (5) control (distilled water)	128
6.19	Amplification of <i>Catechol-2,3-dioxygenase</i> gene. Lane L: 10 kb DNA Ladder, Lane 1:0F1B1, lane 2: 7F2A4, lane 3: 141F1C3, lane 4: 21F1E3, lane 5: 35F1C1, lane 6: 42F2B2, lane 7: 49F1B1, lane 8:42F1B3, lane 9: 0F1S1 and lane 10: 0F1S2	130
6.20	Amplification of <i>almA</i> hydrocarbon degrading gene. Lane L: 10 kb DNA Ladder, Lane 1:0F1B1, lane 2: 7F2A4, lane 3: 141F1C3, lane 4: 21F1E3, lane 5: 35F1C1, lane 6: 42F2B2, lane 7: 49F1B1, lane 8:42F1B3, lane 9: 0F1S1 and lane 10: 0F1S2	130
6.21	Neighbor-joining analysis of 0F1B1-c-UPM and 21F1E3-c-UPM isolated from fermented ficus extract by <i>catechol-2,3-dioxygenase</i> gene	131
6.22	Neighbor-joining analysis of 0F1B1-a-UPM and 21F1E3-a-UPM isolated from fermented ficus extract by <i>almA</i> gene	132
7.1	Device for photosynthesis and chlorophyll content measurement. A) Measurement of relative chlorophyll content (SPAD value), B) portable photosynthesis system (LiCOR Inc., Lincoln, Nebraska, USA)	141
7.2	<i>Hibiscus rosa sinensis</i> plant with flower (a) and <i>Hibiscus cannabinas</i> with pod (b)	142

- 7.3 Mean $\pm$ SE photosynthetic rate ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$) of *H. rosa sinensis* before and after treated with three different fermented plant extracts and one commercial botanical. T₁: mealybug infested plants treated with fermented turmeric extract, T₂: mealybug infested plants treated with fermented kaffir lime, T₃: mealybug infested plants treated with fermented ficus extract, T₄: mealybug infested plants treated with A-force as positive control, T₅: water treated plants as negative control, T₆: non-infested plants treated with fermented turmeric extract, T₇: non-infested plants treated with fermented kaffir lime extract, T₈: non-infested plants treated with fermented ficus extract. T₉: non-infested plants treated with A-force, TDW: non-infested plants treated with distilled water 147
- 7.4 Relative chlorophyll content (Mean $\pm$ SE) of *Hibiscus rosa sinensis* before and after treated with three different fermented plant extracts and one commercial botanical. T₁: mealybug infested plants treated with fermented turmeric extract, T₂: mealybug infested plants treated with fermented kaffir lime, T₃: mealybug infested plants treated with fermented ficus extract, T₄: mealybug infested plants treated with A-force as positive control, T₅: water treated plants as negative control, T₆: non-infested plants treated with fermented turmeric extract, T₇: non-infested plants treated with fermented kaffir lime extract, T₈: non-infested plants treated with fermented ficus extract. T₉: non-infested plants treated with A-force, TDW: non-infested plants treated with distilled water 148
- 7.5 Effect of fermented plant extracts on *Hibiscus rosa sinensis* plant after 30 days post treatment. T₁: mealybug infested plants treated with fermented turmeric extract, T₂: mealybug infested plants treated with fermented kaffir lime, T₃: mealybug infested plants treated with fermented ficus extract, T₄: mealybug infested plants treated with A-force as positive control, T₅: water treated plants as negative control, T₆: non-infested plants treated with fermented turmeric extract, T₇: non-infested plants treated with fermented kaffir lime extract, T₈: non-infested plants treated with fermented ficus extract. T₉: non-infested plants treated with A-force, TDW: non-infested plants treated with distilled water 149
- 7.6 Response of *H. rosa sinensis* plants after infested with *P. solenopsis* and treated with FPEs after 30 days post treatment. T₁: mealybug infested plants treated with fermented turmeric extract, T₂: mealybug infested plants treated with fermented kaffir lime, T₃: mealybug infested plants treated with fermented ficus extract, T₄: mealybug infested plants treated with A-force as positive control, and T₅: distilled water treated plants as negative control 153

7.7	Response of <i>H. cannabinas</i> plants after infested with <i>P. solenopsis</i> and treated with FPEs after 55 days post treatment. T ₁ : mealybug infested plants treated with fermented turmeric extract, T ₂ : mealybug infested plants treated with fermented kaffir lime, T ₃ : mealybug infested plants treated with fermented ficus extract, T ₄ : mealybug infested plants treated with A-force as positive control, and T ₅ : distilled water treated plants as negative control	154
7.8	Phytotoxicity (% leaf area damage) of three FPEs on <i>H. cannabinas</i> leaves at laboratory condition	155
7.9	% Phytotoxicity (%leaf area damage) of FPE on <i>H. cannabinas</i> plant at glasshouse	155
7.10	Phytotoxicity effect of fermented plant extracts on <i>H. cannabinas</i> plants after 24 h post treatment. T ₁ : plants treated with fermented turmeric extract, T ₂ : plants treated with fermented kaffir lime, T ₃ : plants treated with fermented ficus extract, T ₄ : plants treated with A-force (positive control), and T ₅ : distilled water treated plants as negative control	156
7.11	Phytotoxicity effect of fermented plant extracts on <i>H. rosa sinensis</i> plants after 24 h post treatment. T ₁ : plants treated with fermented turmeric extract, T ₂ : plants treated with fermented kaffir lime, T ₃ : plants treated with fermented ficus extract, T ₄ : plants treated with A-force (positive control), and T ₅ : distilled water treated plants as negative control	157

LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
CRD	Completely Randomized Design
DMRT	Duncan's Multiple Range Test
dH ₂ O	Distilled Water
FPE	Fermented Plant Extract
GCMS	Gas Chromatography-Mass Spectrometry
g	Gram
h	Hour
LC ₅₀	Lethal Concentration 50
LDM	Leaf Dipping Method
LIP	Laboratory of Insect Pathology
mg/L	Milligram Per Litre
min.	Minute
mL	Milliliter
mm	Millimeter
nm	Nanometer
p. t	Post Treatment
SAS	Statistical Analyses System
SEM	Scanning Electron Microscopy
SPAD	Soil Plant Analysis Development
rpm	Rotation Per Minute
RT	Retention Time
RCBD	Randomized Complete Block Design
TAM	Top Application Method

UPM Universiti Putra Malaysia

μl Microliter

w/v Weight to Volume



CHAPTER 1

INTRODUCTION

The genus *Hibiscus* belongs to Malvaceae family which comprises of several hundred species woody shrubs, small trees, and both annual and perennial herbaceous plants (Fatima et al., 2016; Taylo and Magdalita, 2021). *Hibiscus* are used for landscaping, construction, rope, fibre and paper production, food and beverage, folk medicine, symbolism and culture, national and state symbol, etc. Several species notably *H. syriacus* L. and *H. rosa-sinensis* L. are widely grown as ornamental plants (El Mokni and Iamónico, 2020). *Hibiscus rosa-sinensis* L. is one of the most widely cultivated flowering plants in the tropical and sub-tropical places including Asia (Magdalita et al., 2010). It is the national flower of Malaysia and commonly grown in Malaysia (Bhaskar, 2011).

Several researchers (Fatima et al., 2016; Taylo and Magdalita, 2021) reported a high percent incidence of thrips, flea beetle, mealybugs, and soft scale insects on *Hibiscus* spp. Till date, five mealybug species namely *Ferrisia sylirii* (Cockerell), *Maconellicoccus hirsutus* (Green), *Paracoccus marginatus* (Williams and Granara de Willink), *Phenacoccus solenopsis* (Tinsley), and *Pseudococcus jackbeardsleyi* (Gimpel and Miller) have been reported infesting the hibiscus plant such as *Hibiscus rosa-sinensis* L. (national flower), *H. liliiflorus* L. and hibiscus hybrids in Malaysia (Williams, 2004; Sartiami et al., 2016). *Phenacoccus solenopsis* (Tinsley) is one of the most destructive mealybug species commonly found on *Hibiscus* sp. It has been reported as a serious and invasive pest of *H. rosa-sinensis* L. in Pakistan, India (Hodgson et al., 2008), Nigeria (Akintola and Ande, 2008) and China (Wang et al., 2009; Wu and Zhang, 2009). Beside *H. rosa-sinensis* L., *P. solenopsis* (Tinsley) also attacked other *Hibiscus* species such as *H. cannabinas* L. (Kenaf) which is cultivated as a cash crop in Malaysia (Roslan et al., 2011; Hanim, 2012; Kamal et al., 2012). This crop has been cultivated as an alternative material to produce bio-composite and used for production of furniture, pulp, paper, fibre board, animal feed and materials for construction and automotive. Kyoto Protocol also recognized it as an eco-friendly industrial organic material to be effective in reducing global warming (Mohd Rifaat, 2008). Among the kenaf insect pests, mealybug is one of the most destructive pests on kenaf in Indian subcontinent (Vennila et al., 2011). The highest level of mealybug infestation (47.58%) was reported in BJRI Kenaf-3 (Bot kenaf) causing 47.82% fibre yield loss in Bangladesh (BJRI Annual report, 2017). In Malaysia, *P. solenopsis* (Tinsley) was observed on kenaf (Malisa et al., 2011; Roslan et al., 2011). Infestation of mealybug could cause yellowing of leaves, defoliation, flower buds that failed to open, ceasing the natural process of photosynthesis, ultimately reduced growth, wrinkled and curling younger leaves, bunchy top and death of plants malformation of shoots, leaves, flowers and fruits (Dhawan, 1980; Culik and Gulan, 2005; Chong et al., 2008; Nagrare et al., 2011; Sartiami et al., 2016), toxin injection into plant tissue and virus transmission (Ben-Dov, 2005; Hoffman et al., 2011).

There are several control measures available, but none is absolute. If there is no proper control method on this insect pest, the crop production in Malaysia will be affected as

well. Although the insecticides have provided high efficacy in insect pest control in general, but it may induce resistance in mealybugs due to their high reproductive potentiality and multiple generations each year (Mark and Gullan, 2005). Sometimes, the efficacy of conventional insecticides against mealybugs has been proven unsatisfactory and ineffective because of the waxy covering on body that hinders penetration of the active ingredients of insecticides (Sparks et al., 1996; Joshi et al., 2010; Nagrare et al., 2011). Besides, indiscriminate insecticide application will disrupt the potential natural enemies and pest resurgence of new insect leading to additional insecticides application to control these pests. An increased in field resistance instances to insecticides have been documented due to detrimental effects on the environment rendering insecticide-based control techniques ineffective (Depickère et al., 2012). These drawbacks led to explore new eco-friendly alternatives as a part of integrated control strategy for mealybugs.

Fermented plant extracts (FPEs) are produced by fermenting food of kitchen waste/ fruits peel/ garden waste, water, and molasses/brown sugar. The microorganisms present in the waste metabolize the food waste into acetic acid or “vinegar”, ethanol and propionic acid. High concentration of acetic acid at low pH is an important factor of the FPEs to be effective in reducing odor, cleaning, and avoiding blockage of drainage, controlling insect etc. Besides the acetic acid, alcohol and propionic acid can resolve severe insecticidal resistance situations and suitable to offer alternative to the chemical control of insect pests (Prakash, 2011). According to Drecampbell (2021), the acidic solutions such as apple cider vinegar could kill mealybugs successfully. The byproduct of FPEs could offer a cheaper source of acidic solutions for mealybug management. Dar et al. (2019) stated that FPEs seem to appear as a promising alternative to replace conventional chemical control and could get around the worst situations of insecticide resistance.

During the fermentation process, the fermenting microbes such as bacteria, yeasts, and molds are involved in the decomposition of plant materials and wax degradation (Doyle and Meng, 2006; Arunkumar et al., 2017a; Mokoena, 2017). A number of bacteria such as *Acinetobacter*, *Actinomycetes*, *Alcaligenes*, *Arthrobacter*, *Bacillus*, *Corynebacteria*, *Flavobacterium*, *Micrococcus*, *Neisseria*, *Nocardia*, *Plesiomonas*, *Proteus*, *Pseudomonas*, *Rhodococcus*, *Xanthomonas*, *Zoogloea* are involved in the wax degradation (Ridgway et al., 1990; Roper 2004; Xu et al., 2013). They degrade wax by utilizing it as the sole carbon source and energy for their growth (Arunkumar et al., 2017a). Salunkhe et al. (2013) and Arunkumar et al. (2017a) demonstrated the potential promising wax degrading result for bacteria against mealybug. Ibrahim et al. (2020) had isolated *B. cereus*, *B. subtilis*, *B. licheniformis* and *Pseudomonas putida* from *Plenococcus citri* (Risso). Krishnamoorthy et al. (2020) also reported *Bacillus* sp isolated from *Paracoccus marginatus* as efficient biocontrol agent for mealybugs. The application of FPE in crop protection program will reduce the usage of synthetic pesticides in agricultural insect pest.

Regarding the waxy coating body *P. solenopsis* (Tinsley) along with the drawbacks of traditional insecticides, it is challenging to explore eco-friendly management alternatives. However, several researchers have reported that FPEs could serve as an anti-microbial agent or insecticide, and they have a sour smell like vinegar that acts as

repellant to insects (Burnette, 2010; Cortesia et al., 2014; Rahmat et al., 2104; Nazim, 2017). A few number of researches on the efficacy of FPEs have been carried out to control aphids and white flies (Nzanza and Mashela, 2012), 28-spotted beetles (Baloc and Bulong, 2015), cutworm (Sahayaraj, 2011, Subash and Raju, 2014), and white grub (Ayub et al., 2021). Recently, Khadem et al. (2022) have reported the use of FPEs as a promising candidate to repel and to control *Planacoccus citri* (Risso). In fact, there was not any attempt to test the insecticidal activity of FPE and wax degrading bacteria associated with fermentation against *P. solenopsis* (Tinsley). Therefore, this study was undertaken to discover a safer and effective pest control approach to control the mealybugs infesting the *H. rosa sinensis* L. and *H. cannabinus* L. under laboratory and field conditions with the following objectives:

1. Identification of mealybugs collected from *Hibiscus rosa sinensis*.
2. Evaluation of fermented plant extracts against mealybugs under laboratory condition.
3. Evaluation of wax-degrading bacteria against mealybugs under laboratory condition.
4. Field evaluation of fermented plant extracts against mealybugs on *H. rosa sinensis* and *H. cannabinus*.

REFERENCES

- Abbas, G., Arif, M. J., Ashfaq, M., Aslam, M. and Saeed, S. (2010). The impact of some environmental factors on the fecundity of *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae): a serious pest of cotton and other crops. *Pakistan Journal of Agricultural Sciences*, 47(4): 321-325.
- Abbas, G., Arif, M. J., Saeed, S. and Karar, H. (2008). Important aspects of the biology of an undescribed species *Phenacoccus gossypiphilous* nov. nud. *Pseudococcidae: Hemiptera*) a serious pest of cotton in Pakistan, 244.
- Abbas, G., Arif, M. J., Saeed, S. and Karar, H. (2009). A new invasive species of genus *Phenacoccus* Cockerell attacking cotton in Pakistan. *International Journal of Agriculture and Biology*, 11(1): 54-58.
- Abdel-Megeed, A., Al-Harbi, N. and Al-Deyab, S. (2010). Hexadecane degradation by bacterial strains isolated from contaminated soils. *African Journal of Biotechnology*, 9(44): 7487-7494.
- Abdel-Reheem, S., Belal, M. H. and Gupta, G. (1991). Photosynthesis inhibition of soybean leaves by insecticides. *Environmental Pollution*, 74(3): 245-250.
- Abd-Rabou, S., Shalaby, H., Germain, J. F., Ris, N., Kreiter, P. and Malausa, T. (2012). Identification of mealybug pest species (Hemiptera: Pseudococcidae) in Egypt and France, using a DNA barcoding approach. *Bulletin of entomological Research*, 102(5): 515-523.
- Abiy, E., Gebre-Michael, T., Balkew, M. and Medhin, G. (2015). Repellent efficacy of DEET, MyggA, neem (*Azadirachta indica*) oil and chinaberry (*Melia azedarach*) oil against *Anopheles arabiensis*, the principal malaria vector in Ethiopia. *Malaria Journal*, 14(1): 1-6.
- Aboelhadid, S. M., Kamel, A. A., Arafa, W. M. and Shokier, K. A. (2013). Effect of *Allium sativum* and *Allium cepa* oils on different stages of *Boophilus annulatus*. *Parasitology Research*, 112(5): 1883-1890.
- Acharya, V. S., Prasad, H. and Kappor, C. J. (2008). Record of Mealybug, *Maconellicoccus* Sp. on Bt Cotton in Northern Rajasthan. *Insect Environ*, 14(1): 18.
- Adegoke, A. S., Jerry, O. V. and Ademola, O. G. (2019). GC-MS analysis of phytochemical constituents in methanol extract of wood bark from *Durio zibethinus* Murr. *International Journal of Medicinal Plants and Natural Products*, 5(3): 1-11.
- Adelskov, J. and Patel, B. K. (2016). A molecular phylogenetic framework for *Bacillus subtilis* using genome sequences and its application to *Bacillus subtilis*

subspecies *stecoris* strain D7XPN1, an isolate from a commercial food-waste degrading bioreactor. *3 Biotechnology*, 6(1): 1-16.

Agostini-Costa, T.S., Vieira, R.F., Bizzo, H.R., Silveira, D. and Gimenes, M.A. (2012) Chromatography and its applications. In *Plant secondary metabolites*; Dhanarasu, S., Ed.; In Tech Publisher: Rijeka, Croatia, 131–164.

Aheer, G. M., Shah, Z. and Saeed, M. (2009). Seasonal history and biology of cotton mealy bug, *Phenacoccus Solenopsis* Tinsley. *Journal of Agricultural Research* (03681157), 47(4).

Ahmad Nazarudin, M. R. (2012). Plant growth retardants effect on growth and flowering of potted *Hibiscus rosa-sinensis* L. *Journal of Tropical Plant Physiology*, 4: 29-40.

Ahmadi, M., Amiri-Besheli, B. and Hosieni, S. Z. (2012). Evaluating the effect of some botanical insecticides on the citrus mealybug *Planococcus citri* (Risso) (Hemiptera: Pseudococcidae). *African Journal of Biotechnology*, 11(53): 11620-11624.

Ahmed, A.M., Jun, M.A., Qiu, B.L., Ri-Rong, H.E., Mu-Tao, W.U., Liang, F., Zhao, J.P., Lin, L., Hu, X.N. and Lv, L.H. (2015). Genetic record for a recent invasion of *Phenacoccus solenopsis* in Asia. *Environmental entomology*, 44(3): 907-918.

Ahmed, M. T., Begum, M. and Zaman, M. W. (2014). The Repellent Effect of Some Indigenous Plant Extracts Against Pulse Beetle (*Callosobruchus chinensis*). *Journal of Environmental Science and Natural Resources*, 7(2): 151-154.

Akintola, A. J. and Ande, A. T. (2008). First record of *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) on *Hibiscus rosa-sinensis* in Nigeria. *Agricultural Journal*, 3(1): 1-3.

Akinyemi, A. J., Faboya, L., Awonegan, A., Olayide, I., Anadozie, S. and Oluwasola, T. (2018). Antioxidant and anti-Acetylcholinesterase activities of essential oils from garlic (*Allium sativum*) Bulbs. *International Journal of Plant Research*, 31.

Akpan, G. A. (2000). Cytogenetic characteristics and the breeding system in six *Hibiscus* species. *Theoretical and Applied genetics*, 100(2): 315-318.

Aktar, W., Sengupta, D. and Chowdhury, A. (2009). Impact of pesticides use in agriculture: their benefits and hazards. *Interdisciplinary toxicology*, 2(1): 1-12.

Alam, S. F., Seni, A. and Sahoo, A. K. (2011). Biology of the mealybug, *Phenacoccus solenopsis* Tinsley (Pseudococcidae: Hemiptera) on sprouted potato and brinjal plant. *The Journal of Plant Protection Sciences*, 3(2): 32-36.

Ali, A., Cantrell, C. L., Bernier, U. R., Duke, S. O., Schneider, J. C., Agramonte, N. M. and Khan, I. (2014). *Aedes aegypti* (Diptera: Culicidae) biting deterrence: structure-activity relationship of saturated and unsaturated fatty acids. *Journal of medical entomology*, 49(6): 1370-1378.

- Ali, E. A., Ibrahim, H. A. K., Abd ell Razzik, M. I. and Balboul, O. A. (2017). Biocidal effect of some bacterial isolates against mealy bug insect *Phenacoccus parvus* (Hemiptera: Pseudococcidae). *Middle East Journal of Applied Sciences*, 7: 1066–1079.
- Ali, M. and Chaudhary, N. (2011). *Ficus hispida* Linn.: A review of its pharmacognostic and ethnomedicinal properties. *Pharmacognosy reviews*, 5(9): 96–102.
- Aloys, N. and Angeline, N. (2009). Traditional fermented foods and beverages in Burundi. *Food Research International*, 42: 588-594.
- Amarasekare, K. G., Chong, J. H., Epsky, N. D. and Mannion, C. M. (2008). Effect of temperature on the life history of the mealybug *Paracoccus marginatus* (Hemiptera: Pseudococcidae). *Journal of Economic Entomology*, 101(6): 1798-1804.
- Ambarish, S., Biradar, A. P. and Jagginavar, S. B. (2017). Phytotoxicity and their bio-efficacy of pesticides against key insect pests of Rabi sorghum [*Sorghum bicolor* (L.) Moench]. *Journal of Entomology and Zoology Studies*, 5(2): 716-720.
- Amer, A. and Mehlhorn, H. (2006). Repellency effect of forty-one essential oils against *Aedes*, *Anopheles*, and *Culex* mosquitoes. *Parasitology research*, 99(4): 478-490.
- Ammar, E. D., Alessandro, R. T. and Hall, D. G. (2013). Ultrastructural and chemical studies on waxy secretions and wax-producing structures on the integument of the woolly oak aphid *Stegophylla brevirostris* Quednau (Hemiptera: Aphididae). *Journal of Microscopy and Ultrastructure*, 1(1-2): 43-50.
- Amusan, A. A. S., Idowu, A. B. and Arowolo, F. S. (2005). Comparative toxicity effect of bush tea leaves (*Hyptis suaveolens*) and orange peel (*Citrus sinensis*) oil extract on larvae of the yellow fever mosquito *Aedes aegypti*. *Tanzania Journal of Health Research*, 7(3): 174-178.
- Andersen, P. C., Mizell, R. F., French, W. J. and Aldrich, J. H. (1986). Effect of multiple applications of pesticides on leaf gas exchange of peach. *Hort Science*, 21(3): 508-510.
- Anita, S., Ruchika, K. and Dolly, K. (2012). Repellence property of traditional plant leaf extracts against *Aphis gossypii* Glover and *Phenacoccus solenopsis* Tinsley. *African Journal of Agricultural Research*, 7(11): 1623-1628.
- Ansari, M. A. and Razdan, R. K. (1994). Repellent action of *Cymbopogon martinii martinii* Stapf var. *sofia* oil against mosquitoes. *Indian journal of malariology*, 31(3): 95-102.

- Ansari, M. A., Mittal, P. K., Razdan, R. K. and Sreehari, U. (2005). Larvicidal and mosquito repellent activities of Pine (*Pinus longifolia*, Family: Pinaceae) oil. *Journal of vector borne diseases*, 42(3): 95.
- Arain, M. I. (2009). Effect of Botanical pesticides against mealy bug on cotton. <http://agris.fao.org/agris-search/search.do?recordID=PK2011000066>.
- Arici, M. and Daglioglu, O. (2002). Boza: a lactic acid fermented cereal beverage as a traditional Turkish food. *Food Reviews International*, 18: 39-48.
- Arif, M. I., Muhammad, R. and Ghaffar, A. (2009). Host plants of cotton mealybug (*Phenacoccus solenopsis*): a new menace to cotton agroecosystem of Punjab, Pakistan. *International Journal of Agriculture and Biology*, 11(2):163-167.
- Aristizábal, L. F., Mannion, C., Bergh, C. and Arthurs, S. (2012). Life history of pink hibiscus mealybug, *Maconellicoccus hirsutus* (Hemiptera: Pseudococcidae) on three *Hibiscus rosa-sinensis* cultivars. *Florida Entomologist*, 95(1): 89-94.
- Arumugam, G., Swamy, M. K. and Sinniah, U. R. (2016). *Plectranthus amboinicus* (Lour.) Spreng: botanical, phytochemical, pharmacological and nutritional significance. *Molecules*, 21(4): 369.
- Arun, C. and Sivashanmugam, P. (2015). Investigation of biocatalytic potential of garbage enzyme and its influence on stabilization of industrial waste activated sludge. *Process Safety and Environmental Protection*, 94: 471-478.
- Arunkumar, N., Banu, J. G., Gopalakrishnan, N. and Prakash A.H. (2017b). Isolation and characterization of wax degrading bacteria isolated from cotton mealy bugs for their application in sustainable agriculture production. Extended abstract for Fifth International Conference on Sustainable Utilization of Tropical Plant Biomass: *Bioproducts, Biocatalysts and Biorefinery* (SutB4), TNAU, Coimbatore. pp.153-155.
- Arunkumar, N., Banu, J. G., Gopalakrishnan, N. and Prakash, A. H. (2021). Efficacy of Lipase-Producing, Wax-degrading Bacteria against the Solenopsis Mealybug, *Phenacoccus solenopsis* Tinsley and the Striped Mealybug, *Ferrisia virgata* Cockerell (Homoptera: Pseudococcidae) on Cotton. *Pakistan Journal of Zoology*, 1-10.
- Arunkumar, N., Gulsar Banu, J., Gopalakrishnan, N. and Prakash, A. H. (2017a). Wax degrading bacteria: Scope and applications in agriculture. *International Journal of Current Microbiology and Applied Sciences*, 6(2): 649-664.
- Arunrangi, T. and Songsrirote, K. (2013). Effects of biocides on chlorophyll contents of detached basil leaves. *Songklanakarin Journal of Science and Technology*, 35(3).
- Asadollahi, A., Khoobdel, M., Zahraei-Ramazani, A., Azarmi, S. and Mosawi, S. H. (2019). Effectiveness of plant-based repellents against different *Anopheles* species: a systematic review. *Malaria Journal*, 18(1): 1-20.

- Ashbell, G., Pahlow, G., Dinter, B. and Weinberg, Z. G. (1987). Dynamics of orange peel fermentation during ensilage. *Journal of applied bacteriology*, 63(4): 275-279.
- Ashfaq, M., Noor, A. R. and Mansoor, S. (2010). DNA-based characterization of an invasive mealybug (Hemiptera: Pseudococcidae) species damaging cotton in Pakistan. *Applied Entomology and Zoology*, 45(3): 395–404.
- Asmanizar, M. A. S. and Maimunah, F. (2016). Studies on the efficacy of some rhizome powders in protecting stored mung bean *Phaseolus radiatus* against pulse beetle *Callosobruchus chinensis* (coleoptera: bruchidae). *Serangga*, 21(1): 61-69.
- Ateyyat, M. A., Shatnawi, M. and Al-Mazra'awi, M. (2010). Isolation and identification of culturable forms of bacteria from the sweet potato whitefly *Bemisia tabaci* Genn.(Homoptera: Aleyrodidae) in Jordan. *Turkish Journal of Agriculture and Forestry*, 34(3): 225-234.
- Avise, J.C. (2004). Molecular markers, natural history and evolution, 2nd ed. by Sinauer Associates, Sunderland, Massachusetts. pp. 684.
- Avoseh, O., Oyedeji, O., Rungqu, P., Nkeh-Chungag, B. and Oyedeji, A. (2015). Cymbopogon species; ethnopharmacology, phytochemistry and the pharmacological importance. *Molecules*, 20(5): 7438-7453.
- Axelsson, L. (2004). Lactic acid bacteria: classification and physiology. *Food Science and Technology-New York-Marcel Dekker*, 139: 1-66.
- Ayub, M., Hussain, M., Ahmad, S., Rizvi, S. A. H., Hussain, S., Qasim, M. and Mastoi, M. I. (2021). Insecticidal Potential of some Fermented Native Plant Extracts for the Management of White Grub (Scarabaeidae: Coleoptera) Infestation on Potato Tubers in Skardu Baltistan. *Pakistan Journal of Agricultural Research*, 34(2).
- Azad, M. A. K., Yesmin, M. N. and Islam, M. S. (2012). Effect of botanical extract on pest control in brinjal field. *Journal of Environmental Science and Natural Resources*, 5(2): 173-176.
- Azokpota, P., Hounhouigan, D. J. and Nago, M. C. (2006). Microbiological and chemical changes during the fermentation of African locust bean (*Parkia biglobosa*) to produce afitin, iru and sonru, three traditional condiments produced in Benin. *International journal of food microbiology*, 107(3): 304-309.
- Babu, S. R. and Meghwal, M. L. (2014). Population dynamics and monitoring of sucking pests and bollworms on Bt cotton in humid zone of southern Rajasthan. *The Bioscan*, 9(2): 629-632.
- Badshah, H., Ullah, F., Farid, A., Calatayud, P. A. and Crickmore, N. (2015). Toxicity of Neem Seed *Azadirachta indica* Juss (Meliaceae) Different Solvents Extracts against Cotton Mealybug *Phenacoccus solenopsis* Tinsley (Sternorrhyncha:

Pseudococcidae) under Laboratory Conditions. *Journal of Entomology and Zoology Studies*, 3: 45-49.

- Bakar, K. B. B. (2010). *Garbage enzyme as an alternative method in treatment of sullage* (Doctoral dissertation, Master thesis, University Technology Malaysia, Malaysia).
- Baker, K.M. (1994). Bioremediation. *Mc Graw Hill.*, New York, p. 31.
- Bakkali, F., Averbeck, S., Averbeck, D. and Idaomar, M. (2008). Biological effects of essential oils—a review. *Food and chemical toxicology*, 46(2): 446-475.
- Ball, S.L., and Armstrong, K.F. (2006). DNA barcodes for insect pest identification. *Canadian Journal of Forest Research*, 36(2): 337-350.
- Baloc, H. A. and Bulong, M. P. (2015). Efficacy of fermented botanical plant extracts in the management of white flies and 28-Spotted beetles in tomato. *International Journal of Science Researsch*, 4(7): 2566-2569.
- Balogun, S. A., Shofola, T. C., Okediji, A. O. and Ayangbenro, A. S. (2015). Screening of hydrocarbonoclastic bacteria using Redox indicator 2, 6-dichlorophenol indophenol. *Global NEST Journal*, 17(3): 565-573.
- Bamaiyi, L. J., Ndams, I. S., Toro, W. A. and Odekina, S. (2007). Laboratory Evaluation of Mahogany (*Khaya senegalensis*) (Desv.) Seed Oil and Seed Powder for the Control of *Callosobruchus maculatus* (Fab.) (Coleoptera: Bruchidae) on Stored Cowpea. *Journal of Entomology*, 4: 237-242.
- Banu, J. G., Suruliveru, T., Amutha, M. and Gapalakrishnan, N. (2010). Susceptibility of Cotton mealybug, *Paracoccus marginatus* to Entomopathogenic fungi. *Annals of Plant Protection Sciences*, 18(1): 247-248.
- Baran, B., Krzyżowski, M., Cup, M., Janiec, J., Grabowski, M. and Francikowski, J. (2018). Repellent effect of volatile fatty acids on lesser mealworm (*Alphitobius diaperinus*). *Insects*, 9(1): 35.
- Barnard, D. R. (1999). Repellency of essential oils to mosquitoes (*Diptera: Culicidae*). *Journal of Medical Entomology*, 36(5): 625-629.
- Basri, M. H. A., Abdu, A., Junejo, N., Hazandy, A. H. and Ahmed, K. (2014). Journey of kenaf in Malaysia: A Review. *Scientific Research and Essays*, 9(11): 458-470.
- Beardsley, J.W. (1985). Notes and exhibitions. *Maconellicoccus hirsutus* (Green). *Proceedings of the Hawaiian Entomological Society*, 25: 27.
- Beaumont, M. (2002). Flavouring composition prepared by fermentation with *Bacillus* spp. *International Journal of Food Microbiology*, 75(3): 189-196.

- Beevi, N. D., Janarthanan, R. and Natarajan, K. (1992). Efficacy of some insecticides against eggs of *Maconellicoccus hirsutus* Green on mulberry *Morus alba* L. *Journal of Insect Science*. 5(1): 114.
- Beltrà, A., Soto, A. and Malausa T. (2012). Molecular and morphological characterisation of *Pseudococcidae* surveyed on crops and ornamental plants in Spain. *Bulletin of Entomological Research*, 102(2): 165-172.
- Ben-Dov, Y. (1994). *A systematic catalogue of the mealybugs of the world (Insecta: Homoptera: Coccoidea: Pseudococcidae and Putoidae) with data on geographical distribution, host plants, biology and economic importance*. Intercept Limited. Andover, MA
- Ben-Dov, Y. (2005). ScaleNet, *Pseudococcidae elisae* catalogue. (<http://www.sel.barc.usda.gov/catalogs/pseudoco/Pseudococcuselisae.htm>) (accessed on 1 August 2015).
- Ben-Dov, Y. (2008). Scale Net, *Paracoccus marginatus*. Available from <http://www.sel.barc.usda.gov/catalogs/pseudoco/Paracoccusmarginatus.htm> [10 April 2010].
- Ben-Dov, Y., Miller, D.R. and Gibson, G.A.P. (2006) ScaleNet: a database of the scale insects of the world. United States Department of Agriculture (USDA), Beltsville, MD. Available online at <http://www.sel.barc.usda.gov/scalenet/scalenet.htm> (accessed 9 December 2010).
- Ben-Dov, Y., Miller, D.R. and Gibson, G.A.P. (2010). ScaleNet, Scales in a Family/Genus Query Results. Available online at <http://www.sel.barc.usda.gov/scalecgi/chklist.exe?Family=Pseudococcidae&genus> (accessed 31 August 2011)
- Benelli, G., Canale, A., Toniolo, C., Higuchi, A., Murugan, K., Pavela, R. and Nicoletti, M. (2017). Neem (*Azadirachta indica*): towards the ideal insecticide?. *Natural product research*, 31(4): 369-386.
- Bharathi, K. and Muthukrishnan, N. (2017). Survey and records of mealybugs species on cotton and alternate host of key mealybug *Phenacoccus solenopsis* Tinsley and its natural enemies complex in major cotton growing areas of South Tamil Nadu, India. *International Journal of Current Microbiology and Applied Science*, 6(12): 1047-1054.
- Bharathithasan, M., Ravindran, D. R., Rajendran, D., Chun, S. K., Abbas, S. A., Sugathan, S. and Ravi, R. (2021). Analysis of chemical compositions and larvicidal activity of nut extracts from *Areca catechu* Linn against *Aedes* (Diptera: Culicidae). *PloS one*, 16(11):026-0281.
- Bhaskar, A. (2011). Evaluation of hypolipidaemic activity of *Hibiscus rosinensis* L. *Journal of Pharmaceutical Research*, 4(10): 3293-3294.

- Bidoia, E. D., Montagnolli, R. N. and Lopes, P. R. M. (2010). Microbial biodegradation potential of hydrocarbons evaluated by colorimetric technique: a case study. *Applied Microbiology and Biotechnology*, 7: 1277-1288.
- Bilal, H., Akram, W., Khan, H. A. A., Hassan, S. A. and Khan, I. A. (2012). Toxicity of selected indigenous plant extracts against *Aedes albopictus* (Diptera: Culicidae): A potential Dengue vector in dengue positive areas. *Pakistan Journal of Zoology*, 44(2).
- Bina, S., Javadi, I. and Iravani, O. (2017). Evaluation of the Repellency Effect of Neem (*Melia azedarach*) Plant Extracts Based on the Mittler and Dadd Method. *Journal of Agricultural Chemistry and Environment*, 6(04): 165.
- Bisevac, L. (1997). A new method for mounting thrips (Thysanoptera) on slides. *Australian Journal of Entomology*, 36(3): 220-220.
- BJRI Annual report (2017). *Study on varietal assessment and management of mealybug of kenaf*. Entomology department, Bangladesh Jute Research Institute.
- Blackburn, M., Golubeva, E., Bowen, D. and Ffrench-Constant, R. H. (1998). A novel insecticidal toxin from *Photorhabdus luminescens*, toxin complex a (Tca), and its histopathological effects on the midgut of *Manduca sexta*. *Applied and Environmental Microbiology*, 64(8): 3036-3041.
- Blandino, A., Al-Aseeri, M., Pandiella, S., Cantero, D. and Webb, C. (2003). Cereal-based fermented foods and beverages. *Food Research International*, 36: 527-543.
- Bligny, R., Roby, C. and Douce, R. (1989). Phosphorus-31 nuclear magnetic resonance studies in higher plant cells. *Nuclear magnetic resonance in agriculture*, 71-89.
- Boodram, R. and Khan, A. (2019). Bioactivity of *Citrus aurantifolia*, *Citrus limon* and *Piper nigrum* essential oils on *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). *Journal of Biopesticides*, 12(1): 76-82.
- Boyer, S., Blakemore, R.J. and Wratten, S.D. (2011). An integrative taxonomic approach to the identification of three new New Zealand endemic earthworm species (Acanthodrilidae, Octochaetidae: Oligochaeta). *Zootaxa*, 2994(1): 21-32.
- Brown, M. and Hebert, A. A. (1997). Insect repellents: an overview. *Journal of the American Academy of Dermatology*, 36(2): 243-249.
- Bučková, M., Puškarová, A., Chovanová, K., Kraková, L., Ferianc, P. and Pangallo, D. (2013). A simple strategy for investigating the diversity and hydrocarbon degradation abilities of cultivable bacteria from contaminated soil. *World Journal of Microbiology and Biotechnology*, 29(6): 1085-1098.
- Burkill, I. H. (1935). A dictionary of the economic products of the Malay Peninsula. *Volume II (IZ)*. London: Crown Agents for the Colonies.

- Burnette, R. (2010). An introduction to wood vinegar. *ECHO Asia Regional Office*, (<http://c.ymcdn.com/sites/www.echocommunity.org>), Accessed February, 13, 2013.
- Cabizza, M., Angioni, A., Melis, M., Cabras, M., Tuberoso, C. V. and Cabras, P. (2004). Rotenone and rotenoids in cube resins, formulations, and residues on olives. *Journal of agricultural and food chemistry*, 52(2): 288-293.
- Cabras, P., Caboni, P., Cabras, M., Angioni, A. and Russo, M. (2002). Rotenone residues on olives and in olive oil. *Journal of Agricultural and Food Chemistry*, 50(9): 2576-2580.
- Catling, D. and Grayson J. (1982). Identification of vegetable fibers. Chapter 8 Kenafi chapter 9 Kose He. Chapman and Hall, New York.
- Cavalieri, V., Mazzeo, G., Garzia, G. T., Buonocore, E. and Russo, A. (2008). Identification of *Planococcus ficus* and *Planococcus citri* (Hemiptera: Pseudococcidae) by PCR-RFLP of COI gene. *Zootaxa*, 1816(1): 65-68.
- Chakraborty, D. P., Basak, S. P., Das, B. C. and Beugelmans, R. (1971). Cyclomahogenol, a new tetracyclic triterpene from *Swietenia mahagoni*. *Phytochemistry*, 10(6): 1367-1372.
- Chalermisan, Y. and Peerapan, S. (2009). Wood vinegar: by-product from rural charcoal kiln and its role in plant protection. *Asian Journal of Food and Agro-Industry*, 2.
- Chatzidimitriou, E., Simonato, M., Watson, G. W., Martinez-Sañudo, I., Tanaka, H., Zhao, J. and Pellizzari, G. (2016). Are *Phenacoccus solani* Ferris and *P. defectus* Ferris (Hemiptera: Pseudococcidae) distinct species. *Zootaxa*, 4093(4): 539-551.
- Chavan, B. P. and Kadam, J. R. (2009). Effect of combination of adjuvants on liquid formulations of *Verticillium lecanii* (Zimmermann) Viegas and their efficacy. *Journal of Biological Control*, 23(1): 73-77.
- Chen, Y., Huang, B., Huang, M. and Cai, B. (2011). On the preparation and characterization of activated carbon from mangosteen shell. *Journal of the Taiwan Institute of Chemical Engineers*, 42(5): 837-842.
- Cheng, S. S., Huang, C. G., Chen, Y. J., Yu, J. J., Chen, W. J. and Chang, S. T. (2009). Chemical compositions and larvicidal activities of leaf essential oils from two eucalyptus species. *Bioresource technology*, 100(1): 452-456.
- Chikere, C. B. and Obieze, C. C. (2018). Effect of hydrocarbon pollution on microbial diversity and implication for bioremediation. *Proceedings of the 2018 International Women in Science Without Borders (WISWB)-Indaba*, 10-11.
- Chikere, C. B., Okoye, A. U. and Okpokwasili, G. C. (2016). Microbial Community Profiling of Active Oleophilic Bacteria involved in Bioreactor-based Crude-oil

Polluted Sediment Treatment. *Journal of Applied and Environmental Microbiology*, 4 (1): 1–20.

Choochote, W., Chaithong, U., Kamsuk, K., Jitpakdi, A., Tippawangkosol, P., Tuetun, B. and Pitasawat, B. (2007). Repellent activity of selected essential oils against *Aedes aegypti*. *Fitoterapia*, 78(5): 359-364.

Christenhusz, M. J. M. and Byng, J. W. (2016). The number of known plants species in the world and its annual increase. *Phytotaxa*, 261: 201–217.

Christopher, B. and Zuk Judith, D. (1997). The American Horticultural Society: *AZ Encyclopedia of Garden Plants*.

Cloyd, R. A. (2011). *Mealybug: Management in Greenhouses and Interiorscapes*. Agricultural Experiment Station and Cooperative Extension Service, Kansas State University. Free online at: [http:// www.bookstore. ksre.ksu.edu/pubs/ mf3001.pdf](http://www.bookstore.ksre.ksu.edu/pubs/mf3001.pdf)

Cobley, L. S. (1976). *An introduction to the botany of tropical crops*. 2nd ed. revised by WM Steele. Longman Group Limited.

Copping, L. G. and Menn, J. J. (2000). Biopesticides: a review of their action, applications and efficacy. *Pest Management Science: Formerly Pesticide Science*, 56(8): 651-676.

Correa, M., Aguirre, C., Germain, J. F., Hinrichsen, P., Zaviezo, T., Malausa, T. and Prado, E. (2011). A new species of *Pseudococcus* (Hemiptera: Pseudococcidae) belonging to the '*Pseudococcus maritimus*' complex from Chile: molecular and morphological description. *Zootaxa*, 2926(1): 46–54.

Cortesia, C., Vilchèze, C., Bernut, A., Contreras, W., Gómez, K., de Waard, J., Jacobs, W.R., Kremer, L. and Takiff, H. (2014). Acetic acid, the active component of vinegar, is an effective tuberculocidal disinfectant. *MBio*, 5(2): 13-15.

Coudriet, D. L., Prabhaker, N. and Meyerdirk, D. E. (1985). Sweet potato whitefly (Homoptera: Aleyrodidae): Effects of neem-seed extract on oviposition and immature stages. *Environmental entomology*, 14(6): 776-779.

Cox, J.M. and Pearce, M.J. (1983). Wax produced by dermal pores in three species of mealybug (Homoptera: Pseudococcidae). *International journal of insect morphology and embryology*, 12(4): 235-248.

Culik, M.P. and Gullan, P.J. (2005). A new pest of tomato and other records of mealybug (Hemiptera: Pseudococcidae) from Espírito Santo, Brazil. *Zootaxa*, 964: 1–8.

Dahllöf, I., Baillie, H. and Kjelleberg, S. (2000). rpoB-based microbial community analysis avoids limitations inherent in 16S rRNA gene intraspecies heterogeneity. *Applied and environmental microbiology*, 66(8): 3376-3380.

- Dahmana, H., Raoult, D., Fenollar, F. and Mediannikov, O. (2020). Insecticidal Activity of Bacteria from Larvae Breeding Site with Natural Larvae Mortality: Screening of Separated Supernatant and Pellet Fractions. *Pathogens (Basel, Switzerland)*, 9(6): 486.
- Damalas, C. A. (2011). Potential uses of turmeric (*Curcuma longa*) products as alternative means of pest management in crop production. *Plant Omics*, 4(3): 136-141.
- Daniels, C. H. and Fults, J. (2002). Fact sheet for vinegar/acetic acid recommendations. *Washington State University, Fact Sheet PIC-01002*.
- Daniels, C. H. and Miller, T. W. (2016). Pesticide ingredient: acetic acid/vinegar. *Washington state University Extension Fact Sheet*.at <http://pubs.wsu.edu>.
- Dar, S. A., Khan, Z. H., Khan, A. A. and Ahmad, S. B. (2019). Biopesticides–Its Prospects and Limitations: An Overview. *Perspective in Animal Ecology and Reproduction*. New Delhi, India: Astral International (P) Ltd, 296-314.
- Das, S. N. and Soumyesh, M. (2014). Green agriculture: evaluation of plant extracts for effectiveness against mealy bug. In *Proceedings of the International Conference on Forests, Soil and Rural Livelihoods in a Changing Climate, Kathmandu, Nepal, 27-30 September 2014* (pp. 211-219). Kathmandu University.
- Dash, P. R., Mou, K. M., Erina, I. N., Ripa, F. A., Al Masud, K. N. and Ali, M. S. (2017). Study of anthelmintic and insecticidal activities of different extracts of *Kaempferia galanga*. *International Journal of Pharmaceutical Sciences and Research*, 8(2): 729-733.
- David, H. and V. Nandagopal (1986). Pests of sugarcane: Distribution symptomology attack and identification. In: *Sugarcane Entomology in India* (Eds.), 1986: 1-29.
- David, P. M. M. ; Rajkumar, K., Razak, T. A., Nelson, S. J., Nainar, P., Baskaran, R. K. M. and Rajavel, D. S. (2010). Efficacy of castor oil-based soft soaps against cotton mealybug, *Phenacoccus solenopsis* Tinsley on brinjal. *Karnataka Journal of Agricultural Sciences*. 23 (1):169-170.
- Davidson, E. W. (1973). Ultrastructure of American foulbrood disease pathogenesis in larvae of the worker honeybee, *Apis mellifera*. *Journal of Invertebrate Pathology*, 21(1): 53-61.
- De Clerck, E., Vanhoutte, T., Hebb, T., Geerinck, J., Devos, J. and De Vos, P. (2004). Isolation, characterization, and identification of bacterial contaminants in semifinal gelatin extracts. *Applied and Environmental Microbiology*, 70(6): 3664-3672.
- Debra, K. R. and Misheck, D. (2014). Onion (*Allium cepa*) and garlic (*Allium sativum*) as pest control intercrops in cabbage-based intercrop systems in Zimbabwe. *IOSR Journal of Agriculture and Veterinary Science*, 7(2): 13-17.

- Dempsey, J. M. (1975). *Fiber crops*. Univ. Presses of Florida.
- Den Belder, E., Elderson, J. and Agca, I. (2001). Is olfactory orientation of *Thrips tabaci* disrupted by a non-host plant. In *Proc. Exp. Appl. Entomol. NEV Amsterdam*, 12: 61-64.
- Deng, M. C., Li, J., Liang, F. R., Yi, M., Xu, X. M., Yuan, J. P. and Wang, J. H. (2014). Isolation and characterization of a novel hydrocarbon-degrading bacterium *Achromobacter* sp. HZ01 from the crude oil-contaminated seawater at the Daya Bay, southern China. *Marine Pollution Bulletin*, 83(1): 79-86.
- Denloye, A. A. (2010). Bioactivity of powder and extracts from garlic, *Allium sativum* L. (Alliaceae) and spring onion, *Allium fistulosum* L. (Alliaceae) against *Callosobruchus maculatus* F. (Coleoptera: Bruchidae) on cowpea, *Vigna unguiculata* (L.) Walp (Leguminosae) seeds. *Psyche*, 2010.
- Denloye, A., Makanjuola, W., Teslim, O., Alafia, O., Kasali, A. and Eshilokun, A. (2010). Toxicity of *Chenopodium ambrosioides* L. (Chenopodiaceae) products from Nigeria against three storage insects. *Journal of Plant Protection Research*, 50(3): 379–384
- Depickère, S., Buitrago, R., Siñani, E., Baune, M., Monje, M., Lopez, R. and Brenière, S. F. (2012). Susceptibility and resistance to deltamethrin of wild and domestic populations of *Triatoma infestans* (Reduviidae: Triatominae) in Bolivia: new discoveries. *Memórias do Instituto Oswaldo Cruz*, 107: 1042-1047.
- Deshmukh, A.J., Vennila, S., Pinjarakar, D.B., Ghodki, B.S. and Kranthi, K.R. (2009). Host range of mealybug *Phenacoccus solenopsis* Tinsley in cotton + pigeon pea cropping system of Central India. *Proceedings of the national symposium on "Bt Cotton: opportunities and prospects*, CICR, Nagpur. p. 127.
- Devi, S., Saini, H. S. and Kaur, S. (2022). Insecticidal and growth inhibitory activity of gut microbes isolated from adults of *Spodoptera litura* (Fab.). *BMC microbiology*, 22(1): 71.
- Dhaliwal, G. S., Singh, R. and Chhillar, B. S. (2006). *Essentials of agricultural entomology*. Kalyani Publishers. New Delhi, India.
- Dhawan, A. K. and Saini, S. (2009, January). Study on biology of mealybug, *Phenacoccus solenopsis* Tinsley on cotton in Punjab. In *Symposium Abstracts. Proceedings of the National Symposium on IPM Strategies to Combat Emerging Pests in the Current Scenario of Climate Change*, 35-36.
- Dhawan, A. K., Joginder, S. and Sidhu, A. S. (1980). *Maconellicoccus* sp. attacking arboreum cotton in Punjab. *Science and culture*, 46(7):258.
- Di Muccio, T., Marinucci, M., Frusteri, L., Maroli, M., Pesson, B. and Gramiccia, M. (2000). Phylogenetic analysis of *Phlebotomus* species belonging to the subgenus *Larrousius* (Diptera, Psychodidae) by ITS2 rDNA sequences. *Insect Biochemistry and Molecular Biology*, 30(5): 387-393.

- Díaz-Fleischer, F., Arredondo, J., Lasa, R., Bonilla, C., Debernardi, D., Pérez-Staples, D. and Williams, T. (2019). Sickly sweet: insecticidal polyols induce lethal regurgitation in dipteran pests. *Insects*, 10(2): 53.
- Dimetry, N. Z. and Schmidt, G. H. (1992). Efficacy of Neem-Azal S and Margosan-0 against the bean aphid, *Aphis fabae* Scop. *Anzeiger für Schädlingskunde, Pflanzenschutz, Umweltschutz*, 65(4): 75-79.
- Dimetry, N. Z., Gomaa, A. A., Salem, A. A. and Abd-El-Moniem, A. S. H. (1996). Bioactivity of some formulations of neem seed extracts against the whitefly *Bemisia tabaci* (Genn.). *Anzeiger für Schädlingskunde, Pflanzenschutz, Umweltschutz*, 69(6): 140-141.
- Dimitri, M. (1987). *Enciclopedia Argentina de Agricultura Jardinería*. Tomo I. Descripción de plantas cultivadas. Buenos Aires:Editorial ACME S.A.C.I.
- Divya, S. and Kalyanasundaram, M. (2019). Composition of the wax particles and surface waxes of the adult mealybug species. *Journal of Experimental Zoology, India*, 22(2): 747-751.
- Downie, D.A. and Gullan, P.J. (2004). Phylogenetic analysis of mealybugs (Hemiptera: Coccoidea: Pseudococcidae) based on DNA sequences from three nuclear genes, and a review of the higher classification. *Systematic Entomology*, 29: 238–259.
- Doyle, M. P. and Meng, J. I. A. N. G. H. O. N. G. (2006). Bacteria in food and beverage production. *Prokaryotes*. Springer, New York, 797-811.
- Drancourt, M. and Raoult, D. (2002). rpoB gene sequence-based identification of *Staphylococcus* species. *Journal of Clinical Microbiology*, 40(4): 1333-1338.
- Drecampbell (2021). com/natural-ways-get-rid-mealybugs respond.
- Dreistadt, S. H. (2001). *Integrated pest management for floriculture and nurseries* (Vol. 3402). University of California Agriculture and Natural Resources, 3402:422.
- EcoWalktheTalk, “The effective are Garbage Enzymes” [Online], Available from HTTP:<http://www.ecowalkthehealth.com> [6 Dec 2012]
- Edeerozey, A. M., Akil, H. M., Azhar, A. B. and Ariffin, M. Z. (2007). Chemical modification of kenaf fibers. *Materials letters*, 61(10): 2023-2025.
- Effiom, O. E., Avoaja, D. A. and Ohaeri, C. C. (2012). Mosquito repellent activity of phytochemical extracts from peels of citrus fruit species. *Global Journal of Science Frontier Research*, 12(1): 1-8.

- El Mokni, R., and Iamonico, D. (2020). New aliens in Malvaceae for the North African flora, with nomenclatural notes. *Collectanea Botanica*, 39(1): 10-3989.
- El-Ashram, D., Abd El-Mageed, S. A. M. and Shaaban, A. R. (2020). Potential toxicity of some essential oils on mealybug, *Maconellicoccus hirsutus* (Hemiptera: Pseudococcidae). *Egyptian Journal of Plant Protection Research Institute*, 3(2): 794-803.
- Elisa, D., Boeckelmann, U., Braun, B., Diehl, D., Schaumann, G. E. and Grohmann, E. (2006). Wax-degrading bacteria influence water repellency of soil. *Bioresource Technology*, 50: 1-12.
- El-Zahi, E. Z. S., Aref, S. A. E. S. and Korish, S. K. M. (2016). The cotton mealybug, *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) as a new menace to cotton in Egypt and its chemical control. *Journal of plant protection research*, 56(2): 111-115.
- El-Zahi, E.S. and Farag, S. A.E. (2017). Population dynamic of *Phenacoccus solenopsis* Tinsley on cotton plants and its susceptibility to some insecticides in relation to the exposure method. *Alexandria science exchange journal*, 38(April-June): 231-237.
- Erler, F. and Ates, A. O. (2015). Potential of two entomopathogenic fungi, *Beauveria bassiana* and *Metarhizium anisopliae* (Coleoptera: Scarabaeidae), as biological control agents against the June beetle. *Journal of Insect Science*, 15(1): 44.
- Evans, S. C., Shaw, E. M. and Rypstra, A. L. (2010). Exposure to a glyphosate-based herbicide affects agrobiont predatory arthropod behaviour and long-term survival. *Ecotoxicology*, 19(7): 1249-1257.
- Fairolniza, M., Leow, T., Mukred, A.D., Bakar, A., Basri, M., Raja, R. and Rahman, A. (2007). Production of L2 lipase by *Bacillus* sp. strain L2: nutritional and physical factors. *Journal of Basic Microbiology*, 47(5): 406-412.
- Fan, B., Blom, J., Klenk, H. P. and Borriss, R. (2017). *Bacillus amyloliquefaciens*, *Bacillus velezensis*, and *Bacillus siamensis* form an “operational group *B. amyloliquefaciens*” within the *B. subtilis* species complex. *Frontiers in microbiology*, 8: 22.
- Fan, S. L., Rita, M. A., Dzolkhifli, O. and Mawardi, R. (2011). Insecticidal properties of *Citrus hystrix* DC leaves essential oil against *Spodoptera litura* fabricius. *Journal of Medicinal Plants Research*, 5(16): 3739-3744.
- Fatima, S., M. H., Malik, M. F., Noureen, N., ul Ane, N. and Abbas, Z. (2016). Field efficacy of some insecticides against hibiscus mealybug, *Maconellicoccus hirsutus* (Hemiptera: Pseudococcidae). *Journal of Entomology and Zoology Studies*, 4(1): 240-244.
- Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. *Evolution*, 39(4): 783-791.

- Feng, Y., Zhang, M., Mujumdar, A. S. and Gao, Z. (2017). Recent research process of fermented plant extract: A review. *Trends in Food Science and Technology*, 65: 40-48.
- Ferree, D. C., Myers, S. C., Rom, C. R. and Taylor, B. H. (1983, July). Physiological aspects of summer pruning. In *International Workshop on Controlling Vigor in Fruit Trees*, 146: 243-252.
- Fuchs, T. W., Stewart, J. W., Minzenmayer, R. and Rose, M. (1991). First record of *Phenacoccus solenopsis* Tinsley in cultivated cotton in the United States. *Southwestern Entomologist*, 16(3): 215-221.
- García Morales, M., Denno, B. D., Miller, D. R., Miller, G. L., Ben-Dov, Y. and Hardy, N. B. (2018). 'ScaleNet: A literature-based model of scale insect biology and systematics'. Retrieved from <http://scalenet.info>
- García, M., Donadel, O. J., Ardanaz, C. E., Tonn, C. E. and Sosa, M. E. (2005). Toxic and repellent effects of *Baccharis salicifolia* essential oil on *Tribolium castaneum*. *Pest Management Science: formerly Pesticide Science*, 61(6): 612-618.
- Gariepy, T. D., Kuhlmann, U., Gillott, C. and Erlandson, M. (2007). Parasitoids, predators and PCR: the use of diagnostic molecular markers in biological control of Arthropods. *Journal of applied entomology*, 131(4): 225-240.
- Ghananand, T., Prasad, C. S. and Lok, N. (2011). Effect of insecticides, bio-pesticides and botanicals on the population of natural enemies in Brinjal ecosystem. *Vegetos*, 24(2): 40-44.
- Ghanem, I., Orfi, M. and Shamma, M. (2007). Biodegradation of chlorpyrifos by *Klebsiella* sp. isolated from an activated sludge sample of waste water treatment plant in damascus. *Folia Microbiologica*, 52(4): 423-427.
- Gharsan, F. N. (2019). A review of the bioactivity of plant products against *Aedes aegypti* (Diptera: Culicidae). *Journal of Entomological Science*, 54(3): 256-274.
- Ghose, S. K. (1971). Morphology of various instars of both sexes of the mealybug, *Maconellicoccus hirsutus* (Green) (Pseudococcidae: Hemiptera). *Indian Journal of Agricultural Sciences*, 41(7): 602-611.
- Ghosh, R., Sharatchandra, K. H., Rita, S. and Thokchom, I. S. (2004). Hypoglycemic activity of *Ficus hispida* (bark) in normal and diabetic albino rats. *Indian Journal of pharmacology*, 36(4): 222-225.
- Giglio, A., Giulianini, P. G., Zetto, T. and Talarico, F. (2011). Effects of the pesticide dimethoate on a non-target generalist carabid, *Pterostichus melas italicus* (Dejean, 1828)(Coleoptera: Carabidae). *Italian Journal of Zoology*, 78(4): 471-477.

- Giménez–Moolhuyzen, M., van der Blom, J., Lorenzo–Mínguez, P., Cabello, T. and Crisol–Martínez, E. (2020). Photosynthesis inhibiting effects of pesticides on sweet pepper leaves. *Insects*, 11(2): 69.
- Gitahi, S. M., Piero, M. N., Mburu, D. N. and Machochi, A. K. (2021). Repellent effects of selected organic leaf extracts of *Tithonia diversifolia* (Hemsl.) A. Gray and *Vernonia lasiopus* (O. Hoffman) against *Sitophilus zeamais* Motschulsky (Coleoptera: Curculionidae). *The Scientific World Journal*, 2021.
- Gojo, C., Nunez, P. H., Anulacion, R. L. and Gregorio, N. A. (2018). Insecticidal property and phytochemical screening of mahogany (*Swietenia macrophylla*) leaves, barks and seeds as an alternative insecticide against fungi growing termites (*Macrotermes gilvus*). *International Journal of Biology, Pharmacy and Allied Science*, 7(8): 1521-1528.
- Goolsby, J. A., Kirk, A. A. and Meyerdirk, D. E. (2002). Seasonal phenology and natural enemies of *Maconellicoccus hirsutus* (Hemiptera: Pseudococcidae) in Australia. *Florida Entomologist*, 85(3): 494-498.
- Gosalwad, S. S., Bhosle, B. B., Wadnerkar, D. W., Ilyas, M. and Khan, F. S. (2009). Biology and feeding potential of *Cryptolaemus montrouzieri* mulsant (Coccinellidae: Coleoptera) on *Maconellicoccus hirsutus* and *Phenacoccus solenopsis*. *Journal of Plant Protection and Environment*, 6(2): 73-77.
- Gotyal, B. S., Satpathy, S., Selvaraj, K., Meena, P. N., and Naik, R. K. (2014). Mealybug (*Phenacoccus solenopsis* Tinsley) -A New Pest in Jute Crop: Management Strategies. *Pop. Kheti*, 2(3): 123-125.
- Granara de Willink, M.C. (2003). New records and host plants of *Phenacoccus* for Argentina (Hemiptera: Pseudococcidae). *Revista de la Sociedad Entomológica Argentina*, 62: 80–82.
- Gowda, D. K. S., Manjunath, D., Pradip, K. and Datta, R. K. (1996). Spalgisepius Westwood (Lepidoptera: Lycaenidae)-a potential predator of mulberry mealybug, *Maconellicoccus hirsutus*. *Insect Environment*, 2(3): 87-88.
- Gross, S., Dreishpoun, Y., Blachinski, D., Shmueli, S., Steinberg, S. and Mendel, Z. (1999). Cork scars on fruits of the citrus variety 'Sweetie' as related to infestation by the citrus mealybug. *Alon Hanotea*, 53(11): 463-468.
- Gullan, P. J. and Kosztarab, M. (1997). Adaptations in scale insects. *Annual Review of Entomology*, 42(1), 23-50.
- Gullan, P.J., Downie, D.A. and Steffan, S.A. (2003). A new pest species of the mealybug genus *Ferrisia* Fullaway (Hemiptera: Pseudococcidae) from the United States. *Annals of the Entomological Society of America*, 96(6): 723-737.
- Gulsarbanu, J., Surlivelu, T., Balamurugan, M., Amutha, M. and Gopalkrishnan, N., (2009). Natural occurrence of entomopathogenic fungi in mealybug, *Phenacoccus solenopsis* Tinsley (Hemiptera: Psuedococcidae) in India.

Proceedings of the national symposium on "Bt Cotton: opportunities and prospects, CICR, Nagpur. p.121.

- Guo, T. C., Fang, B. T. and Wang, C. Y. (2005). Effects of water regulations on the kinetic parameters of chlorophyll fluorescence in wheat flag leaves as well as wheat yield. *Agricultural Research in the Arid Areas*, 23(2): 6-10.
- Gupta, S. K., Biswas, H. and Das, S. N. (2007). Bioeffectiveness of some plant extracts towards causing mortality of *Brevipalpus phoenicis* (Geij)(Acari: Tenuipalpidae), A new pest of medicinal plant, Basak, *Justicia adhatoda* L. Nessn (Acanthaceae). *Bioprospecting and application of medicinal plants in common ailments* (Edition Eds. Gupta SK and Maitra, BR), RK Mission, Narendrapur, 121-126.
- Gupta, S.S., Jafar, V., Jaiwal Raman, P., Singh, M. and Maithani (2011). Review on titrimetric analysis. *International Journal of Comprehensive Pharmacy*, 2(5):1-6.
- Haba, E., Bresco, O., Ferrer, C., Marques, A., Busquets, M. and Manresa, A. (2000). Isolation of lipase-secreting bacteria by deploying used frying oil as selective substrate. *Enzyme and microbial technology*, 26(1): 40-44.
- Haile, F. J., Peterson, R. K. and Higley, L. G. (1999). Gas-exchange responses of alfalfa and soybean treated with insecticides. *Journal of economic entomology*, 92(4): 954-959.
- Hajibabaei, M., Janzen, D. H., Burns, J. M., Hallwachs, W. and Hebert, P. D. N. (2006). DNA barcodes distinguish species of tropical Lepidoptera. *Proceedings of the National Academy of Sciences*, 103(4): 968-971.
- Hall, T.A. (2010). BioEdit, version 7.2.5th ed. for Windows 95/98/NT. Carlsbad, California.
- Hameed, A., Aziz, M. A. and Aheer, G. M. (2012). Impact of ecological conditions on biology of cotton mealy bug, *Phenacoccus solenopsis* (Hemiptera: Pseudococcidae) in laboratory. *Zoological society of Pakistan*, 44(3): 685-690.
- Hamon, A. B., Lambdin, P. L. and Kosztarab, M. (1975). Eggs and wax secretion of *Kermes kingi*. *Annals of the Entomological Society of America*, 68(6): 1077-1078.
- Haniff, M. H. (2006). Gas exchange of excised oil palm (*Elaeis guineensis*) fronds. *Asian Journal of Plant Sciences*, 5(1): 9-13.
- Hanim, A. (2012). Nothing positive about tobacco farming. In: Star Online, 4 September. www.thestar.com.my/business/business-news/2012/09/04/nothing-positive-about-tobacco-farming [05.10.2017]

- Hardy, N.B., Gullan, P.J. and Hodgson, C.J. (2008). A subfamilylevel classification of mealybugs (Hemiptera: Pseudococcidae) based on integrated molecular and morphological data. *Systematic Entomology* 33: 51–71.
- Harley P. J. and Prescott L. M. (1996). *Laboratory exercises in microbiology 3rd Ed.* Times mirror Higher Education Group, Inc. p. 24.
- Harley, R. M., Atkins, S., Budantsev, A. L., Cantino, P. D., Conn, B. J., Grayer, R. J., Harley, M.M., de Kok, R., Krestovskaja, T., Morales, R. and Paton, A. J. (2004). Labiatae pages 167275. *Joachim W. Kadereit, (volume ed.), The Families and Genera of Vascular Plants*, 7: 2275-2283.
- Harrigan, W. F. and McCance, M. E. (1976). *Laboratory methods in food and dairy microbiology*. Academic Press Inc. (London) Ltd. pp389
- He, Y., Qin, Q., DiLegge, M. J. and Vivanco, J. M. (2019). Isolation of *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* from entomopathogenic nematode-insect host relationship to examine bacterial pathogenicity on *Trichoplusia ni*. *Microbial pathogenesis*, 135: 103606.
- Hebert, P. D., Cywinska, A., Ball, S. L. and DeWaard, J. R. (2003). Biological identifications through DNA barcodes. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 270(1512): 313-321.
- Helmy, N., Bakr, R. F., Nawwar, G. A. and Helmy, O. M. (2010). Biochemical effects of some agricultural waste extracts against *Culex pipiens* (Diptera: Culicidae). *Egyptian Academic Journal of Biological Sciences. C, Physiology and Molecular Biology*, 2(2): 75-81.
- Henderson, C. F. and Tilton, E. W. (1955). Tests with acaricides against the brown wheat mite. *Journal of economic entomology*, 48(2): 157-161.
- Heo, J., Kim, J. S., Hong, S. B., Park, B. Y., Kim, S. J. and Kwon, S. W. (2019). Genetic marker gene, recQ, differentiating *Bacillus subtilis* and the closely related *Bacillus* species. *FEMS Microbiology Letters*, 366(16):172.
- Heu, R. A., Fukada, M. T. and Conant, P. (2007). Papaya mealybug, *Paracoccus marginatus* Williams and Granara de Willink (Hemiptera: Pseudococcidae). State of Hawaii Deptment of Agriculture, Honolulu, HI.
- Heywood, V. H., Brummitt, R. K., Culham, A. and Seberg, O. (2007). *Flowering plant families of the world* (Vol. 88). Ontario: Firefly Books.
- Hikal, W. M., Baeshen, R. S. and Said-Al Ahl, H. A. (2017). Botanical insecticide as simple extractives for pest control. *Cogent Biology*, 3(1): 1404274.
- Hillis, D. M. and Bull, J. J. (1993). An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. *Systematic biology*, 42(2): 182-192.

- Hodgson, C.J., Abbas, G., Arif, M.J., Saeed, S. and Karar, H. (2008). *Phenacoccus solenopsis* Tinsley (Sternorrhyncha:Coccoidea: Pseudococcidae), an invasive mealybug damaging cotton in Pakistan and India with a discussion on seasonal morphological variation. *Zootaxa*, 1913: 1-35.
- Hoffman, M.T., Duan, Y., Zhou, L., Stocks, I. and Hall, D. (2011). Is the striped mealybug, *Ferrisia virgata*, a vector of huanglongbing bacterium *Candidatus Liberibacter asiaticus*. *Phytopathology*, 101.
- Hori, M. (1996). Settling inhibition and insecticidal activity of garlic and onion oils against *Myzus persicae* (Sulzer)(Homoptera:Aphididae). *Applied entomology and zoology*, 31(4): 605-612.
- Hou, X. Q., Zhang, D. D., Powell, D., Wang, H. L., Andersson, M. N. and Löfstedt, C. (2021). Ionotropic receptors in the turnip moth *Agrotis segetum* respond to repellent medium-chain fatty acids. *biRxiv*, 27: 1-22.
- Howell, J. (2018). 5 Natural Ways to De-bug Your Home. *Homeowners Tips*.
- Huang, S. H., Xian, J. D., Kong, S. Z., Li, Y. C., Xie, J. H., Lin, J. and Su, Z. R. (2014). Insecticidal activity of pogostone against *Spodoptera litura* and *Spodoptera exigua* (Lepidoptera: Noctuidae). *Pest management science*, 70(3): 510-516.
- Huang, X., Zhang, N., Yong, X., Yang, X. and Shen, Q. (2012). Biocontrol of *Rhizoctonia solani* damping-off disease in cucumber with *Bacillus pumilus* SQR-N43. *Microbiological Research*, 167(3): 135-143.
- Huang, Y., Chen, S. X. and Ho, S. H. (2000). Bioactivities of methyl allyl disulfide and diallyl trisulfide from essential oil of garlic to two species of stored-product pests, *Sitophilus zeamais* (Coleoptera:Curculionidae) and *Tribolium castaneum* (Coleoptera:Tenebrionidae). *Journal of Economic Entomology*, 93(2): 537-543.
- Husain, D. R., Goutx, M., Bezac, C., Gilewicz, M. and Bertrand, J. C. (1997). Morphological adaptation of *Pseudomonas nautica* strain 617 to growth on eicosane and modes of eicosane uptake. *Letters in applied Microbiology*, 24(1): 55-58.
- Hussain, F., Hameed, I., Dastagir, G., Khan, I. and Ahmad, B. (2010). Cytotoxicity and phytotoxicity of some selected medicinal plants of the family Polygonaceae. *African Journal of Biotechnology*, 9(5):28.
- Huws, S. A., Edwards, J. E., Kim, E. J. and Scollan, N. D. (2007). Specificity and sensitivity of eubacterial primers utilized for molecular profiling of bacteria within complex microbial ecosystems. *Journal of Microbiological Methods*, 70(3): 565-569.
- Ibáñez, M. D. and Blázquez, M. A. (2019). Ginger and turmeric essential oils for weed control and food crop protection. *Plants*, 8(3): 59.

- Ibargutxi, M. A., Muñoz, D., de Escudero, I. R. and Caballero, P. (2008). Interactions between Cry1Ac, Cry2Ab, and Cry1Fa *Bacillus thuringiensis* toxins in the cotton pests *Helicoverpa armigera* (Hübner) and *Earias insulana* (Boisduval). *Biological Control*, 47(1): 89-96.
- Ibrahim, S. S., Moharum, F. A. and Abd El-Ghany, N. M. (2015). The cotton mealybug *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) as a new insect pest on tomato plants in Egypt. *Journal of plant protection research*, 55(1): 48-51.
- Ibrahim, S., Gupta, R. K. and Kour, R. (2020). Isolation and characterization of a novel paraffin wax and solid wax degrading endosymbiotic bacteria from Citrus mealybug (*Planococcus citri*). *International Journal of Chemical Studies*, 8(5): 1345-1349.
- Ignacimuthu, S. and Jayaraj, S. (Eds.). (2005). *Green pesticides for insect pest management*. Alpha Science Int'l Ltd..
- Ijah, U. J. J. and Antai, S. P. (2003). The potential use of chicken-drop micro-organisms for oil spill remediation. *Environmentalist*, 23(1): 89-95.
- Indrayani, Y., Oramahi, H.A. and Nurhaida. (2010). Evaluation of Liquid Smoke as Bio-Pesticide to Control Subterranean Termites *Cryptotermes* sp. *Jurnal Ketawang Fakultas Kehutanan Universitas Tanjungpura Pontianak*. Hal, 87-96.
- Innocent, E., Gikonyo, N. K. and Nkunya, M. H. (2008). Repellency property of long chain aliphatic methyl ketones against *Anopheles gambiae* ss. *Tanzania journal of health research*, 10(1): 50-54.
- Insua, J. L., Llobet, E., Moranta, D., Pérez-Gutiérrez, C., Tomás, A., Garmendia, J. and Bengoechea, J. A. (2013). Modeling *Klebsiella pneumoniae* pathogenesis by infection of the wax moth *Galleria mellonella*. *Infection and immunity*, 81(10): 3552-3565.
- Ishii, T., Matsuzawa, H. and Vairappan, C. S. (2010). Repellent activity of common spices against the rice weevil, *Sitophilus zeamais* Motsch (Coleoptera, Curculionidae). *Journal of Tropical Biology and Conservation (JTBC)*, 7.
- Islam, S. and Ashraful, M. (2015). *Evaluatuon of plant extract and mechanical band against mango mealybug (Drosicha mangiferae)* (Doctoral dissertation, Department of Entomology, Sher-e-Bangla Agricultural Universty Dhaka).
- Isman, M. B. (1995). Leads and prospects for the development of new botanical insecticides. *Reviews of environmental contamination and toxicology*, 3: 1-20.
- Isman, M. B. (2006). Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annual Review of Entomology*, 51: 45-66.

- Isman, M. B. (2008). Botanical insecticides: for richer, for poorer. *Pest Management Science: formerly Pesticide Science*, 64(1): 8-11.
- Jadhav, S.S. (1993). Life history of grape mealy bug, *Maconellicoccus hirsutus* (Green) (Hemiptera: Pseudococcidae) at different temperatures. *Maharashtra Journal of Horticulture*, 7(1): 16-29.
- Jagadish, K. S., Shadhanaikural, A., Chandru, R. and Shadakshari, Y. (2009). Biochemical and morphological changes due to mealybug *Phenacoccus solenopsis* Tinsley (Homoptera: Pseudococcidae) infestation on sunflower (*Helianthus annuus* L.). *Insect Environment*, 15(1): 28-30.
- Janda, J. M. and Abbott, S. L. (2007). 16S rRNA gene sequencing for bacterial identification in the diagnostic laboratory: pluses, perils, and pitfalls. *Journal of clinical microbiology*, 45(9): 2761-2764.
- Jat, B. L., Mehta, D. M., Ghetiya, L. V., Patil, R. A. and Tatarwal, A. S. (2014). Seasonal incidence of mealy bug, *Phenacoccus solenopsis* in bidi tobacco. *Indian Journal of Plant Protection*, 42(3): 294-296.
- Jhala, R. C., Bharpoda, T. M. and Patel, M. G. (2008). *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae), the mealy bug species recorded first time on cotton and its alternate host plants in Gujarat, India. *Uttar Pradesh J. Zoology*, 28(3): 403-406.
- Jiménez, M. P. and Meneses, M.B. (2016). First study about morphological and molecular identification of *Pseudococcus elisae* (Hemiptera: Pseudococcidae) in Costa rica. *International Journal of Current Research*, 8(05):31791-31800.
- Johnson, M. W., Welter, S. C., Toscano, N. C., Iwata, Y. and Ting, P. (1983). Lettuce yield reductions correlated with methyl parathion use. *Journal of economic entomology*, 76(6): 1390-1394.
- Jones, V. P., Youngman, R. R. and Parrella, M. P. (1983). Effect of selected acaricides on photosynthetic rates of lemon and orange leaves in California. *Journal of economic entomology*, 76(5): 1178-1180.
- Joshi, M. D., Butani, P. G., Patel, V. N. and Jeyakumar, P. (2010). Cotton mealy bug, *Phenacoccus solenopsis* Tinsley-a review. *Agricultural Reviews*, 31(2): 113-119.
- Judoamidjojo, R.M., Said, E.G. and Hartono. L. (1989). Biokonversi. *Bogor: Departemen Pendidikan dan Kebudayaan Direktorat Jenderal Pendidikan Tinggi, Pusat Antar Universitas Bioteknologi Institut Pertanian Bogor*.
- Kaila, L. and Stahls, G. (2006). DNA barcodes: evaluating the potential of COI to differentiate closely related species of *Elachista* (Lepidoptera: Gelechioidea: Elachistidae) from Australia. *Zootaxa*, 1170(1): 1-26.

- Kairo, M. T., Pollard, G. V., Peterkin, D. D. and Lopez, V. F. (2000). Biological control of the hibiscus mealybug, *Maconellicoccus hirsutus* Green (Hemiptera: Pseudococcidae) in the Caribbean. *Integrated Pest Management Reviews*, 5(4): 241-254.
- Kamal, I., Thirmizir, M. Z. Beyer, G., Saad, M. J., Rashid, N. A. and Kadir, Y. A. (2012). Kenaf For Biocomposite: An Overview. *Journal of Science and Technology*, 6(2): 41-66.
- Kanjanapothi, D., Panthong, A., Lertprasertsuke, N., Taesotikul, T., Rujjanawate, C., Kaewpinit, D., Sudthayakorn, R., Choochote, W., Chaithong, U, Jitpakdi, A. and Pitasawat, B. (2004). Toxicity of crude rhizome extract of *Kaempferia galanga* L.(Proh Hom). *Journal of Ethnopharmacology*, 90(2-3): 359-365.
- Kant, R. and Kumar, V. (2007). Impact of Climatic Conditions on the Infestation of Major Pests of Mulberry (*Morus* Spp.) in Doon Valley. *Journal of the Entomological Research Society*, 9(2):17-30.
- Karamaouna, F., Kimbaris, A., Michaelakis, A., Papachristos, D., Polissiou, M., Papatsakona, P. and Miller, T. (2013). Insecticidal activity of plant essential oils against the vine mealybug, *Planococcus ficus*. *Journal of Insect Science*, 13(1): 142.
- Kasuga, I., Nakajima, F. and Furumai, H. (2007). Diversity of catechol 2, 3-dioxygenase genes of bacteria responding to dissolved organic matter derived from different sources in a eutrophic lake. *FEMS microbiology ecology*, 61(3): 449-458.
- Kaur, H. and Virk, J. S. (2012). Feeding potential of *Cryptolaemus montrouzieri* against the mealybug *Phenacoccus solenopsis*. *Phytoparasitica*, 40(2): 131-136.
- Kaur, S. (2015). DNA barcoding and its applications. *International Journal of Engineering Research and General Science*, 3(2): 602-604.
- Keane, S. and Ryan, M. F. (1999). Purification, characterisation, and inhibition by monoterpenes of acetylcholinesterase from the waxmoth, *Galleria mellonella* (L.). *Insect biochemistry and molecular biology*, 29(12): 1097-1104.
- Keat, S. P. (2010). Determination of acetic acid in garbage enzyme property associated with improving water quality of recreational lake (BSc dissertation, Tunku Abdul Rahman College, Malaysia).
- Kedia, A., Prakash, B., Mishra, P. K., Singh, P. and Dubey, N. K. (2015). Botanicals as eco-friendly biorational alternatives of synthetic pesticides against *Callosobruchus* spp. (Coleoptera: Bruchidae)—a review. *Journal of Food Science and Technology*, 52(3): 1239-1257.
- Khadem, A. G., Chin, N. L., Mokhtar, A. S. and Hong, L. W. (2022). Potential Use of Fermented Plant Extracts as Biological Control Agents for Citrus Mealy Bugs, *Planococcus citri*. *Annals of Agri-Bio Research*, 27(1): 79-86.

- Khan, M. A., Ranjha, M. and Khan, J. (2013). Repellency effects of different plant extracts to cotton mealy bug, Tinsley (Hemiptera: Pseudococcidae) *Phenococcus solenopsis*. *Pakistan Journal of Agricultural Research*, 26(3): 213-219.
- Khan, R. A. and Ashfaq, M. (2004). Funnel Type Slippery Trap: A mechanical device to control mango mealy bug. *SAIC Newsletter, Bangladesh*, 14(3): 2-9.
- Khater, H. F., Ramadan, M. Y. and El-Madawy, R. S. (2009). Lousicidal, ovicidal and repellent efficacy of some essential oils against lice and flies infesting water buffaloes in Egypt. *Veterinary Parasitology*, 164(2-4): 257-266.
- Khuhro, S. N., Lohar, M. K., Nizamani, S. M. and Abro, G. H. (2013). Prey searching ability of *Brumus suturalis* (Fabricius)(Coleoptera: Coccinellidae) on cotton mealybug under laboratory and field conditions. *Pakistan Journal of Agriculture, Agricultural Engineering, Veterinary Sciences*, 29(1): 70-76.
- Kim, D. H., Seo, H. E., Lee, S. C. and Lee, K. Y. (2008). Effects of wood vinegar mixed with insecticides on the mortalities of *Nilaparvata lugens* and *Laodelphax striatellus* (Homoptera: Delphacidae). *Animal Cells and Systems*, 12(1): 47-52.
- Kim, J. K., Kang, C. S., Lee, J. K., KIM, Y. R., HAN, H. Y. and YUN, H. K. (2005). Evaluation of repellency effect of two natural aroma mosquito repellent compounds, citronella and citronellal. *Entomological Research*, 35(2): 117-120.
- Kimutai, A., Ngeiywa, M., Mulaa, M., Njagi, P. G., Ingonga, J., Nyamwamu, L. B., Ombati, C. and Ngumbi, P. (2017). Repellent effects of the essential oils of *Cymbopogon citratus* and *Tagetes minuta* on the sandfly, *Phlebotomus duboscqi*. *BMC Research Notes*, 10(1): 1-9.
- Klein, D. W., Prescott, L. M. and Harley, J. (2005). *Microbiology*. New York: McGraw-Hill.
- Klingeman, W. E., Buntin, G. D., van Iersel, M. W. and Braman, S. K. (2000). Whole-plant gas exchange, not individual-leaf measurements, accurately assesses azalea response to insecticides. *Crop Protection*, 19(6): 407-415.
- Koli, H. R. (2003). *Seasonal incidence and management of grape mealy bug, Maconellicoccus hirsutus* (Green). M. Sc. (Agri.) thesis, Mahatma Phule Krishi Vidhyapeeth, Rahuri, Maharashtra (India).
- Kondo, T., Penny, J. and Williams, D. (2008). Coccidology. The study of scale insects (Hemiptera: Sternorrhyncha: Coccoidea). *Ciencia y Tecnología Agropecuaria*, 9(2): 55-61.
- Kranthi, K. R., Jadhav, D. R., Kranthi, S., Wanjari, R. R., Ali, S. S. and Russell, D. A. (2002). Insecticide resistance in five major insect pests of cotton in India. *Crop protection*, 21(6): 449-460.

- Krause, G. H. and Weis, E. (1991). Chlorophyll fluorescence and photosynthesis: the basics. *Annual review of plant biology*, 42(1): 313-349.
- Krishnamoorthy, R., Arul Jose, P., Janahiraman, V., Indira Gandhi, P., Gandhi Gracy, R., Jalali, S. K. and Anandham, R. (2020). Function and insecticidal activity of bacteria associated with papaya mealybug, *Paracoccus marginatus* Williams and Granara de Willink (Hemiptera: Pseudococcidae). *Biocontrol Science and Technology*, 30(8): 762-778.
- Krugh, B. W. and Miles, D. (1996). Monitoring the effects of five “nonherbicidal” pesticide chemicals on terrestrial plants using chlorophyll fluorescence. *Environmental Toxicology and Chemistry: An International Journal*, 15(4): 495-500.
- Kubota, F. and Hamid, A. (1992). Comparative analysis of dry matter production and photosynthesis between mungbean (*Vigna radiata* (L.) and black gram (*V. mungo* (L.) Hepper grown in different light intensities.
- Kumar, A. and Singh A. (2012). Review on *Hibiscus rosa sinensis*. *International Journal of Research in Pharmaceutical and Biomedical Sciences*, 3(2): 534-538.
- Kumar, L.S., Sawant, A.S., Gupta, V.S. and Ranjekar, P.K. (2001). Genetic variation in Indian populations of *Scirpophaga incertulas* as revealed by RAPD-PCR analysis. *Biochemistry Genetic*, 39(1): 43–57.
- Kumar, P. and Poehling, H. M. (2006). Persistence of soil and foliar azadirachtin treatments to control sweet potato whitefly *Bemisia tabaci* Gennadius (Homoptera: Aleyrodidae) on tomatoes under controlled (laboratory) and field (netted greenhouse) conditions in the humid tropics. *Journal of Pest Science*, 79(4): 189-199.
- Kumar, P., Mishra, S., Malik, A. and Satya, S. (2011). Insecticidal properties of *Mentha* species: A review. *Industrial Crops and Products*. 34(1): 802-817.
- Kumar, R., Nitharwal, M., Chauhan, R., Pal, V. and Kranthi, K. R. (2012). Evaluation of ecofriendly control methods for management of mealybug, *Phenacoccus solenopsis* Tinsley in cotton. *Journal of Entomology*, 9(1): 32–40.
- Kumar, S., Singh, K. and Dwivedi, K. N. (2017). Potential of Indian traditional medicinal plant turmeric as insecticide Antifeedant and insect repellent against household, museum and library insect pests. *International Journal of Entomology Research*. 2(3): 42-46.
- Kumar, S., Stecher, G., Li, M., Knyaz, C. and Tamura, K. (2018). MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular biology and evolution*, 35(6): 1547.
- Kumar, V., Tewari, S.K. and Datta, R.K. (1997). Dermal pores and wax secretion in mealybug *Maconellicoccus hirsutus* (Hemiptera, Pseudococcidae). A pest of mulberry. *Italian Journal of Zoology*, 64(4): 307-311.

- Kunwar, R. M. and Busmann, R. W. (2006). Ficus (Fig) species in Nepal: a review of diversity and indigenous uses. *Lyonia*, 11(1): 85-97.
- Landge, S.A. (2014). Studies on Biology, Mass Multiplication and Management of Cotton Mealybug, *Phenacoccus Solenopsis* (Tinsley) and Its Parasitoid” Phd Thesis, Department of Agricultural Entomology, Post Graduate Institute, Mahatma Phule Krishi Vidyapeeth, Ahmednagar, maharashtra, india.
- Langhoff, P., Authier, A., Buckley, T. R., Dugdale, J. S., Rodrigo, A. and Newcomb, R. D. (2009). DNA barcoding of the endemic New Zealand leafroller moth genera, *Ctenopseustis* and *Planotortrix*. *Molecular Ecology Resources*, 9(3): 691-698.
- Lanjar, A. G., Rustamani, M. A. and Solangi, A. W. (2015). Effect of Botanical Extracts against mango mealybug *Drosicha mangiferae* (Green). *Science International*, 27(1): 343-346.
- LaPre, L. F., Sances, F. V., Toscano, N. C., Oatman, E. R., Voth, V. and Johnson, M. W. (1982). The effects of acaricides on the physiology, growth, and yield of strawberries. *Journal of economic entomology*, 75(4): 616-619.
- Le, C., Ruz-Febles, N.M., Alvarado-Canché, A.R., Canul-Solís, J.R., LópezCobá, E. and Campos-Navarrete, M.J. (2018). Ethanolic extracts of *Brosimum alicastrum* and *Plectranthus amboinicus* for the control of *Raoiella indica*. *Journal of Entomology and Zoology Studies*, 6(5): 625-628.
- Lee, H. S., Shin, W. K., Song, C., Cho, K. Y. and Ahn, Y. J. (2001). Insecticidal activities of ar-turmerone identified in *Curcuma longa* rhizome against *Nilaparvata lugens* (Homoptera: Delphacidae) and *Plutella xylostella* (Lepidoptera: Yponomeutidae). *Journal of Asia-Pacific Entomology*, 4(2): 181-185.
- Leegood, R. C., Labate, C. A., Huber, S. C., Neuhaus, H. E. and Stitt, M. (1988). Phosphate sequestration by glycerol and its effects on photosynthetic carbon assimilation by leaves. *Planta*, 176(1): 117-126.
- LeMahieu, P.J., Oplinger, E.S. and Putnam, D.H. (2011). Kenaf, Alternative Field Corps Manual, online:<http://www.hort.purdue.edu/newcrop/afcm/kenaf.html>. Accessed on 2011 Oct19
- Lengai, G. M. and Muthomi, J. W. (2018). Biopesticides and their role in sustainable agricultural production. *Journal of Biosciences and Medicines*, 6(06): 7-41
- LeOra, S. (2003). Poloplus, a user's guide to probit or logit analysis. *LeOra Software, Berkeley, CA*.
- Levin, R. A., Raguso, R. A. and McDade, L. A. (2001). Fragrance chemistry and pollinator affinities in Nyctaginaceae. *Phytochemistry*, 58(3): 429-440.

- Lewis, S. (2020). Top essential oils that repel bugs+bug spray recipe, diffuser blends, and more dry recipes to naturally keep bugs away.
- Li, H. M., Deng, R. Q., Wang, J. W., Chen, Z. Y., Jia, F. L. and Wang, X. Z. (2005). A preliminary phylogeny of the *Pentatomomorpha* (Hemiptera: Heteroptera) based on nuclear 18S rDNA and mitochondrial DNA sequences. *Molecular phylogenetics and evolution*, 37(2): 313-326.
- Li, Y. and Mai, Y. W. (2006). Interfacial characteristics of sisal fiber and polymeric matrices. *The journal of adhesion*, 82(5): 527-554.
- Lin, C.P. and Danforth, B.N. (2004). How do insect nuclear and mitochondrial gene substitution patterns differ? *Molecular Phylogenetics and Evolution*, 30(3): 686–702.
- Lin, P., Lin, L., Wu, J., Lin, N., Cai, J. and Qi, J. (2004). Breeding of FuHong4, a kenaf variety with high yielding and resistance. *Plant Fiber Production*, 26: 1-4.
- Linares, M. C., Soto-Calderón, I. D., Lees, D. C. and Anthony, N. M. (2009). High mitochondrial diversity in geographically widespread butterflies of Madagascar: a test of the DNA barcoding approach. *Molecular Phylogenetics and Evolution*, 50(3): 485-495.
- Liu, J., Li, Q., Kong, L., Yu, H. and Zheng, X. (2011). Identifying the true oysters (Bivalvia: Ostreidae) with mitochondrial phylogeny and distance-based DNA barcoding. *Molecular Ecology Resources*, 11(5): 820–830.
- Liu, T. X. and Stansly, P. A. (1995). Deposition and bioassay of insecticides applied by leaf dip and spray tower against *Bemisia argentifolii* nymphs (Homoptera: Aleyrodidae). *Pesticide Science*, 44(4): 317-322.
- Liu, X. C., Liang, Y., Shi, W. P., Liu, Q. Z., Zhou, L. and Liu, Z. L. (2014). Repellent and insecticidal effects of the essential oil of *Kaempferia galanga* rhizomes to *Liposcelis bostrychophila* (Psocoptera: Liposcelidae). *Journal of economic entomology*, 107(4): 1706-1712.
- Loh, F. S., Awang, R. M., Omar, D. and Rahmani, M. (2011). Insecticidal properties of *Citrus hystrix* DC leaves essential oil against *Spodoptera litura* fabricius. *Journal of Medicinal Plants Research*, 5(16): 3739-3744.
- López, M. D. and Pascual-Villalobos, M. J. (2010). Mode of inhibition of acetylcholinesterase by monoterpenoids and implications for pest control. *Industrial Crops and Products*, 31(2): 284-288.
- Luscombe, B. M., Pallett, K. E., Loubiere, P., Millet, J. C., Garejo, J. and Vrabel, T. E. (1995). A novel herbicide for broad leaf cane and grass weed control in maize and sugar cane. *Proceedings of the Brighton crop Protect*, 2: 35-42.

- Lynch, J. M. and Hobbie, J. E. (1988). Microflora of aerial plants parts. *Concepts and applications in Microbial Ecology (Eds) JM Lynch and JE Hobbie. Blackwell Scientific Publishers. Oxford*, 222-230.
- Magdalita P.M. and Pimentel R.B. (2010). *Hibiscus 2010: Hybrids and Philippines' women achievers*. National Academy of Science and Technology, Bicutan, Taguig City, Metro Manila, Philippines. 58p.
- Maheswari, M. and Govindaiah, G. (2017). Evaluation of different plant extracts for management of mealy bugs and leaf rollers in the mulberry field. *Screening*, 11: 12.
- Maia, M. F. and Moore, S. J. (2011). Plant-based insect repellents: a review of their efficacy, development and testing. *Malaria journal*, 10(1): 1-15.
- Majeed, M. Z., Nawaz, M. I., Khan, R. R., Farooq, U. and Ma, C. S. (2018). Insecticidal effects of acetone, ethanol and aqueous extracts of *Azadirachta indica* (A. Juss), *Citrus aurantium* (L.), *Citrus sinensis* (L.) and *Eucalyptus camaldulensis* (Dehnh.) against mealybugs (Hemiptera: Pseudococcidae). *Tropical and Subtropical Agroecosystems*, 21(3).
- Malausa, T., Fenis, A., Warot, S., Germain, J.F. and Ris, N. (2011). DNA markers to disentangle complexes of cryptic taxa in mealybugs (Hemiptera: Pseudococcidae). *Journal of applied Entomology*, 135(12): 142-155.
- Malisa, M. N., Hamdan, J. and Husni, M. H. A. (2011). Yield response of Kenaf (*Hibiscus cannabinus* L.) to different rates of charcoal and nitrogen fertilizer on BRIS soils in Malaysia. *Middle-East Journal of Scientific Research*, 10(1): 54-59.
- Mamoon-ur-Rashid, M., Khattak, M. K. and Abdullah, K. (2012). Phenological response of cotton mealybug, *Phenacoccus solenopsis* Tinsley (Sternorrhyncha: Pseudococcidae) to three prominent host plants. *Pakistan Journal of Zoology*, 44(2): 341-346.
- Mandal, D., Bhowmik, P., Halder, P. and Chatterjee, M. L. (2013). Determination of relative toxicity and base line data of different insecticides against cotton mealybug (*Phenacoccus solenopsis* Tinsley). *The Journal of Plant Protection Sciences*, 5(1): 26-31.
- Mandal, S. C. and Kumar, C. A. (2002). Studies on anti-diarrhoeal activity of *Ficus hispida*. Leaf extract in rats. *Fitoterapia*, 73(7-8): 663-667.
- Mani, M. (1986). *Distribution, bioecology and management of the grape mealybug, Maconellicoccus hirsutus (Green) with special reference to its natural enemies* (Doctoral dissertation, University of Agricultural Sciences, GKVK).
- Mani, M. (1989). A review of the pink mealybug, *Maconellicoccus hirsutus* (Green). *International Journal of Tropical Insect Science*, 10(2): 157-167.

- Mani, M. and Shivaraju, C. (Eds.). (2016). *Mealybugs and their management in agricultural and horticultural crops* (pp. 209-222). New Delhi, India::Springer.
- Mani, M., Shivaraju, C. and Shylesha, A. N. (2012). *Paracoccus marginatus*, an invasive mealybug of papaya and its biological control –An overview. *Journal of Biological Control*. 26(3): 201–216.
- Manzanares, M., Tenorio, J. L. and Ayerbe, L. (1997). Sowing time, cultivar, plant population and application of N fertilizer on Kenaf in Spain's central plateau. *Biomass and Bioenergy*, 12(4): 263-271.
- Mariano, A. P., Bonotto, D. M., Angelis, D. D. F. D., Pirôllo, M. P. S. and Contiero, J. (2008). Biodegradability of commercial and weathered diesel oils. *Brazilian Journal of Microbiology*, 39: 133-142.
- Mark, P.C. and Gullan, P.J. (2005). A new pest of tomato and other records of mealybugs (Hemiptera: Pseudococcidae) from Espírito Santo, Brazil. *Zootaxa*, 964: 1-8.
- Marsaro Júnior, A. L., Peronti, A. L. B. G., Penteado-Dias, A. M., Morais, E. G. F. and Pereira, P. D. S. (2013). First report of *Maconellicoccus hirsutus* (Green, 1908)(Hemiptera: Coccoidea: Pseudococcidae) and the associated parasitoid *Anagyrus kamali* Moursi, 1948 (Hymenoptera: Encyrtidae), in Brazil. *Brazilian Journal of Biology*, 73: 413-418.
- Marsh, A. J., Hill, C., Ross, R. P. and Cotter, P. D. (2014). Fermented beverages with health-promoting potential: past and future perspectives. *Trends in Food Science and Technology*, 38: 113-124.
- De Martino, L. D., Mancini, E., de Almeida, L. F. R. and Feo, V. D. (2010). The antigerminative activity of twenty-seven monoterpenes. *Molecules*, 15(9): 6630-6637.
- Martins, D. D. S., Fornazier, M. J., Peronti, A. L. B., Culik, M. P., Souza, C. A. S., Taques, R. C. and Queiroz, R. B. (2019). *Maconellicoccus hirsutus* (Hemiptera: Pseudococcidae) in Brazil: recent spread, natural enemies, and new hosts. *Florida Entomologist*, 102(2): 438-443.
- Mastoi, M. I., Azura, A. N., Muhammad, R., Idris, A. B. and Ibrahim, Y. (2011). First report of papaya mealybug *Paracoccus marginatus* (Hemiptera: Pseudococcidae) from Malaysia. *Australian Journal of Basic and Applied Sciences*, 5(7): 1247-1250.
- Mc Donnell, R.J., Rugman-Jones, P., Backeljau, T., Breugelmans, K., Jordaens, K., Stouthamer, R., Paine, T. and Gormally, M. (2011). Molecular identification of the exotic slug *Arion subfuscus* sensu stricto (Gastropoda: Pulmonata) in California, with comments on the source location of introduced populations. *Biological Invasions*, 13(1): 61-66.

- McKenzie, H. L. (1967). *Mealybugs of California*. University of California Press, Berkeley, CA, 525 pp
- Mendoza-García, E. E., Ortega-Arenas, L. D., Serrato-Cruz, M. Á., Villanueva-Jiménez, J. A., López-Arroyo, J. I. and Pérez-Pacheco, R. (2019). Chemical composition, toxicity, and repellence of plant essential oils against *Diaphorina citri* (Hemiptera: Liviidae). *Chilean journal of agricultural research*, 79(4): 636-647.
- Mensah, F. A., Inkum, I. E., Agbale, C. M. and Eric, A. (2014). Comparative evaluation of the insecticidal and insect repellent properties of the volatile oils of *Citrus aurantifolia* (Lime), *Citrus sinensis* (Sweet orange) and *Citrus limon* (Lemon) on *Camponotus nearcticus* (Carpenter ants). *International Journal of Novel Research in Interdisciplinary Studies*, 1: 19-25.
- Mersha, W., Ayele, N., Fentahun, G., Getinet, M., Kassu, K. and Nagappan, R. (2014). Repellent and insecticidal activity of *Mentha piperita* (L.) plant extracts against cabbage aphid [*Brevicoryne brassicae* Linn. (Homoptera: Aphididae)]. *American-Eurasian Journal of Scientific Research*, 9(6): 150-156.
- Meyerdirk, D. E. (1996). Hibiscus or Pink mealybug. *USDA APHIS Plant Protection and quarantine*, pp. 3
- Meyerdirk, D. E., Muniappan, R., Warkentin, R., Bamba, J. and Reddy, G. V. P. (2004). Biological control of the papaya mealybug, *Paracoccus marginatus* (Hemiptera: Pseudococcidae) in Guam. *Plant Protection Quarterly*, 19: 110-114.
- Michaelakis, A., Papachristos, D., Kimbaris, A., Koliopoulos, G., Giatropoulos, A. and Polissiou, M. G. (2009). Citrus essential oils and four enantiomeric pinenes against *Culex pipiens* (Diptera: Culicidae). *Parasitology research*, 105(3): 769-773.
- Miller DR. (1999). Identification of the pink hibiscus mealybug, *Maconellicoccus hirsutus* (Green) (Hemiptera: Sternorrhyncha: Pseudococcidae). *Insecta mundi*, 189-203.
- Miller, D. R. and Miller, G. L. (2002). Redescription of *Paracoccus marginatus* Williams and Granara de Willink (Hemiptera: Coccoidea: Pseudococcidae), including descriptions of the immature stages and adult male. *Proceedings of the Entomological Society of Washington*, 104(1): 1-23.
- Miller, D. R., Miller, G. L. and Watson, G. W. (2002). Invasive species of mealybugs (Hemiptera: Pseudococcidae). *Proceedings of Entomological Society of Washington*, 104: 825-836.
- Miller, D. R., Rung, A. and Parikh, G. (2014). Scale Insects, edition 2, a tool for the identification of potential pest scales at USA ports-of-entry (Hemiptera, Sternorrhyncha, Coccoidea). *ZooKeys*, (431): 61.

- Miller, D.R. (2005). Selected scale insect groups (Hemiptera: Coccoidea) in the southern region of the United States. *Florida Entomologist*, 88: 482–501.
- Miller, D.R., Rung, A., Venable, G.L. and Gill, R.J. (2007). ScaleNet. Scale Insects: Identification tools, images, and diagnostic information for species of quarantine significance, Systematic Entomology Laboratory, USDAARS. [in USA]
- Miller, G. T. (2004). Sustaining the earth, Thompson learning. Inc. *Pacific Grove, California*, 9: 211-216.
- Miresmailli, S. and Isman, M. B. (2006). Efficacy and persistence of rosemary oil as an acaricide against two spotted spider mites (Acari: Tetranychidae) on greenhouse tomato. *Journal of economic entomology*, 99(6): 2015-2023.
- Mishra, V., Srivastava, G., Prasad, S. M. and Abraham, G. (2008). Growth, photosynthetic pigments and photosynthetic activity during seedling stage of cowpea (*Vigna unguiculata*) in response to UV-B and dimethoate. *Pesticide Biochemistry and Physiology*, 92(1): 30-37.
- Mitchell, P. L., Gupta, R., Singh, A. K. and Kumar, P. (2004). Behavioral and developmental effects of neem extracts on *Clavigralla scutellaris* (Hemiptera: Heteroptera: Coreidae) and its egg parasitoid, *Gryon fulviventre* (Hymenoptera: Scelionidae). *Journal of Economic Entomology*, 97(3): 916-923.
- Mitu, M. H. (2017). *Phytochemical and Pharmacological Investigations on Dichloromethane (Dcm) Fraction Of Ficus Hippida Leaf Extract* (Doctoral dissertation, East West University).
- Mobaiyen, H., Khosro, S., Alireza, D., Abdol-Hosen, N., Farhad, A. P. and Naser, M. S. (2013). Study on liquid paraffin decomposer microorganisms isolated from soils of Julfa-Iran. *African Journal of Microbiology Research*, 7(3): 260-265.
- Mohamed, A., Bhardwaj, H., Hamama, A. and Webber Iii, C. (1995). Chemical composition of kenaf (*Hibiscus cannabinus* L.) seed oil. *Industrial crops and products*, 4(3): 157-165.
- Mohamedova, M. S., Valcheva, I. S., Draganova, D. G., Naydenov, M. K. and Borisov, Y. B. (2017). Effect of *Bacillus amyloliquefaciens* A1, *Paenibacillus polymyxa* AB3 and *Providencia rettgeri* K10 on the citrus mealybug, *Planococcus citri* (Risso) (Hemiptera: Pseudococcidae). *Egyptian Journal of Biological Pest Control*, 27: 41–47.
- Mohammad A.K. (2008) Mealybug an emerging threat to cotton crop. www.pakistan.com.
- Mohd Rifaat, A.H. (2008). Kenaf ganti tembakau, Berita Harian, online: <http://www.mtib.gov.my/repository/stayinform/kenaf-ganti-tembakau.pdf>, accessed on 15th August 2012.

- Mokoena, M. P. (2017). Lactic acid bacteria and their bacteriocins: classification, biosynthesis and applications against uropathogens: a mini review. *Molecules*, 22(8): 1255.
- Moniruzzaman, M., Yaakob, Z., Khatun, R. and Awang, N. (2017). Mealybug (Pseudococcidae) infestation and organic control in fig (*Ficus carica*) orchards of Malaysia. *Biology and Environment: Proceedings of the Royal Irish Academy*.
- Moore, S. J., Lenglet, A. and Hill, N. (2006). Plant-based insect repellents. In: Debboun M, Frances SP, Strickman D, editors. *Insect repellents: principles methods, and use*. 1st ed. Boca Raton: CRC Press. p. 275–304.
- Morimoto, M., Urakawa, M., Fujitaka, T. and Komai, K. (2014). Structure-activity relationship for the insect antifeedant activity of benzofuran derivatives. *Bioscience, biotechnology, and biochemistry*, 63(5): 840-846.
- Mostafa, M. E., Youssef, N. M. and Abaza, A. M. (2018). Insecticidal activity and chemical composition of plant essential oils against cotton mealybug, *Phenacoccus solenopsis* (Tinsley)(Hemiptera: Pseudococcidae). *Mortality*, 100, 7.
- Moustafa, M. A., Awad, M., Amer, A., Hassan, N. N., Ibrahim, E. D. S., Ali, H. M. and Salem, M. Z. (2021). Insecticidal activity of lemongrass essential oil as an eco-friendly agent against the black Cutworm *Agrotis ipsilon* (Lepidoptera: Noctuidae). *Insects*, 12(8): 737.
- Mowry, T. M. and Barbour, J. D. (2004). Distinguishing the parasitic wasp, *Peristenus howardi*, from some of its congeners using polymerase chain reaction and restriction endonuclease digestion. *Journal of Insect Science*, 4(1): 25–31.
- Muniappan, R., Meyerdirk, D. E., Sengebau, F. M., Berringer, D. D. and Reddy, G. V. P. (2006). Classical biological control of the papaya mealybug, *Paracoccus marginatus* (Hemiptera: Pseudococcidae) in the Republic of Palau. *Florida Entomologist*, 212-217.
- Muniappan, R., Shepard, B. M., Watson, G. W., Carner, G. R., Rauf, A., Sartiami, D. and Rahman, A. Z. (2009). New records of invasive insects (hemiptera: Sternorrhyncha) in Southeast Asia and West Africa. *Journal of Agricultural and Urban Entomology*, 26(4): 167-174.
- Muniappan, R., Shepard, B. M., Watson, G. W., Carner, G. R., Sartiami, D., Rauf, A. and Hammig, M. D. (2008). First report of the papaya mealybug, *Paracoccus marginatus* (Hemiptera: Pseudococcidae), in Indonesia and India. *Journal of Agricultural and Urban Entomology*, 25(1): 37-40.
- Muniappan, R., Watson, G. W., Vaughan, L., Gilbertson, R. and Noussourou, M. (2012). New records of mealybugs, scale insects, and whiteflies (Hemiptera:

Sternorrhyncha) from Mali and Senegal. *Journal of Agricultural and Urban Entomology*, 28(1), 1-7.

Muqaddim, M. (2017). Phytochemical and biological investigation of *Kaempferia galangal* laves. Bachelor Thesis. The Deaprtment of Pharmacy, BRAC University, Bangladesh.

Murthy, C. S. H. N. (1983). Effects of pesticides on photosynthesis. In *Residue Reviews* (pp. 107-129). Springer, New York, NY.

Murugan, K., Kalimuthu, K., Kumar, P. M., Hwang, J. S. and Nicoletti, M. (2013). Larval and pupal toxicity effects of *Plectranthus amboinicus*, *Bacillus sphaericus* and predatory copepods for the control of the dengue vector, *Aedes aegypti*. *Phytoparasitica*, 41(3): 307-316.

Mya, M. M., Aung, Z. Z., New, C. T., Oo, A. W., Htay, T. M. and Thauang, S. (2017). Maung Larvicidal, Ovicidal and repellent effect of *Citrus hystrix* DC (Kaffir lime) fruit, peel and internal materials extracts on *Aedes aegypti* mosquitoes. *Journal of Biological Engineering Research and Review*, 4(1): 34-43.

Nagrare, V. S., Kranthi, S., Kranthi, K. R., Naik, V. C. B., Deshmukh, V., Naikwadi, B. and Dahekar, A. (2016). Relative toxicity of insecticides against cotton mealybug *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) and its fortuous parasitod *Aenasius bambawalei* Hayat (Hymenoptera: Encyrtidae). *Journal of Applied and Natural Science*, 8(2): 987-994.

Nagrare, V. S., Kranthi, S., Kumar, R., Jothi, B. D., Amutha, M., Deshmukh, A. J. and Kranthi, K. R. (2011). Compendium of cotton mealybugs. *Technical Bulletin*, Central Institute for Cotton Research, Nagpur, P 42

Naik, M. J. and Naik, A. S. (2015). Impact of botanical extracts on histopathology of silkworm (*Bombyx mori* L.). *Journal of Experimental Biology and Agricultural Sciences*, 3(3): 281-287.

Naik, M.J. and Naik, S. (2012). Evaluation of Effects of botanical extracts against the pink mealybug (*Maconellicoccus hirsutus* Green) on aminotransferases. *International Journal of Science Innovations and Discoveries*, 2(5): 491-497.

Nalini, T. (2015). Field incidence of mealybugs and its parasitization by encyrtids in various crop ecosystems. *Plant Archives*, 15(2): 671-676.

Nararak, J., Sathantriphop, S., Kongmee, M., Bangs, M. J. and Chareonviriyaphap, T. (2017). Excito-repellency of *Citrus hystrix* DC leaf and peel essential oils against *Aedes aegypti* and *Anopheles minimus* (Diptera: Culicidae), vectors of human pathogens. *Journal of Medical Entomology*, 54(1): 178-186.

Nathanson, J. A., Hunnicutt, E. J., Kantham, L. and Scavone, C. (1993). Cocaine as a naturally occurring insecticide. *Proceedings of the National Academy of Sciences*, 90(20): 9645-9648.

- Nazim, F. and Meera, V. (2017). Comparison of treatment of greywater using garbage and citrus enzymes. *International Journal of Innovative Research in Science, Engineering and Technology*, 6(4): 49-54.
- Nerio, L. S., Olivero-Verbel, J. and Stashenko, E. (2010). Repellent activity of essential oils: a review. *Bioresource technology*, 101(1): 372-378.
- Neupane, K. and Khadka, R. (2019). Production of Garbage Enzyme from Different Fruit and Vegetable Wastes and Evaluation of its Enzymatic and Antimicrobial Efficacy. *Tribhuvan University Journal of Microbiology*, 6(1): 113-118.
- Nguyen, V. B., Nguyen, A. D. and Wang, S. L. (2017). Utilization of fishery processing by-product squid pens for α -glucosidase inhibitors production by *Paenibacillus* sp. *Marine Drugs*, 15(9): 274.
- Nikam, N. D., Patel, B. H. and Korat, D. M. (2010). Biology of invasive mealybug, *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) on cotton. *Karnataka Journal of Agricultural Sciences*, 23(4): 649-651.
- Nisha, R. and Kennedy, J. S. (2017). Life cycle of papaya mealybug *Paracoccus marginatus* Williams and Granara de Willink on different host plants vis-à-vis divergent natural selection. *Journal of Entomology and Zoology Studies*, 5: 91-102.
- Nishimura, N., Izumi, A. and Kuroda, K. I. (2002). Structural characterization of kenaf lignin: differences among kenaf varieties. *Industrial Crops and Products*, 15(2): 115-122.
- Nitschke, M., Ferraz, C. and Pastore, G. M. (2004). Selection of microorganisms for biosurfactant production using agro-industrial wastes. *Brazilian Journal of Microbiology*, 35: 81-85.
- Noureen, N., Hussain, M., Fatima, S. and Ghazanfar, M. (2016). Cotton mealybug management: a review. *Journal of Entomology and Zoology Studies*, 4(4): 657-663.
- Nzanza, B. and Mashela, P. W. (2012). Control of whiteflies and aphids in tomato (*Solanum lycopersicum* L.) by fermented plant extracts of neem leaf and wild garlic. *African Journal of Biotechnology*, 11(94): 16077-16082.
- Obiakalaje, U. M., Makinde, O. A. and Amakoromo, E. R. (2015). Bioremediation of crude oil polluted soil using animal waste. *International Journal of Environmental Bioremediation and Biodegradation*, 3(3): 79-85.
- OEPP/EPPO (2005). Data sheets on quarantine pests *Maconellicoccus hirsutus*. *Bulletin OEPP/EPPO Bulletin*, 35, 413-415.

- OEPP/EPPO. (2006). EPPO Standards PM 7/77(1) diagnostic protocol for *Maconellicoccus hirsutus*. OEPP/EPPO, *Bulletin OEPP/EPPO Bulletin*, 36: 167–169.
- Okolle, J. N., Lum, N. M.-C., Ngosong, C., Nanganoa, L. T., Clovis, T. and Peter, N. (2018). Efficacy of four plant emulsions for managing long tail mealybugs (*Pseudococcus longispinus*) infesting banana plantations in Cameroon. *Journal of Entomology and Zoology Studies*, 6(3): 837–843.
- Okoye, A. U., Chikere, C. B. and Okpokwasili, G. C. (2019). Characterization of potential paraffin wax removing bacteria for sustainable biotechnological application. In *SPE Nigeria Annual International Conference and Exhibition*. OnePetro.
- Oliveira, E. R. D., Alves, D. S., Carvalho, G. A., Oliveira, B. M. R. G. D., Aazza, S. and Bertolucci, S. K. V. (2018). Toxicity of *Cymbopogon flexuosus* essential oil and citral for *Spodoptera frugiperda*. *Ciência e Agrotecnologia*, 42: 408-419.
- Oliwa-Stasiak, K., Molnar, C. I., Arshak, K., Bartoszcze, M. and Adley, C. C. (2010). Development of a PCR assay for identification of the *Bacillus cereus* group species. *Journal of applied microbiology*, 108(1): 266-273.
- Ong, H. G. and Kim, Y. D. (2014). Quantitative ethnobotanical study of the medicinal plants used by the Ati Negrito indigenous group in Guimaras island, Philippines. *Journal of ethnopharmacology*, 157: 228-242.
- Özkara, A., Akyl, D. and Konuk, M. (2016). Pesticides, environmental pollution, and health. In *Environmental health risk-hazardous factors to living species*. IntechOpen. 10.5772/61472(Chapter 1), –. doi:10.5772/63094
- Pacheco da Silva, V.C., Bertin, A., Blin, A., Germain, J.F., Bernardi, D., Rignol, G., Botton, M. and Malausa, T. (2014). Molecular and Morphological Identification of Mealybug Species (Hemiptera: Pseudococcidae) in Brazilian Vineyards. *PLoS ONE*, 9(7): e103267.
- Pangnakorn, U. and Chuenchooklin, S. (2018). Toxicity of essential oils to stored product pest and application to extrusion coating film for extend rice storage life. *International Journal of Environmental Monitoring and Analysis*, 6(2): 65-70.
- Paridah, M. T., Abdelrhman, A. H. and Shahwahid, M. (2017). Cost benefit analysis of kenaf cultivation for producing fiber in Malaysia. *Arabian Journal of Business and Management Review*, 7(5).
- Park, H.J., Jin, G. and Nakhleh, L. (2010). Bootstrap-based support of HGT Inferred by maximum parsimony. *BMC Evolutionary Biology*, 10(1): 1-11.
- Park, I. K., Choi, K. S., Kim, D. H., Choi, I. H., Kim, L. S., Bak, W. C., Choi, J.W. and Shin, S. C. (2006). Fumigant activity of plant essential oils and components from horseradish (*Armoracia rusticana*), anise (*Pimpinella anisum*) and garlic

(*Allium sativum*) oils against *Lycoriella ingenua* (Diptera: Sciaridae). *Pest Management Science: formerly Pesticide Science*, 62(8): 723-728.

- Parvin, S., Zeng, X. N. and Islam, T. (2012). Bioactivity of Indonesian mahogany, *Toona sureni* (Blume)(Meliaceae), against the red flour beetle, *Tribolium castaneum* (Coleoptera, Tenebrionidae). *Revista Brasileira de Entomologia*, 56: 354-358.
- Patel, D. D. and Lakshmi, B. (2016). Study on the role of *Nocardia farcinica* in enhancing the flow rate of crude oil. *Bioremediation Journal*, 20(3): 224-232.
- Patel, M. G., Jhala, R. C., Vaghela, N. M. and Chauhan, V. R. (2010). Bio-efficacy of buprofezin against mealy bug, *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) an invasive pest of cotton. *Karnataka Journal of Agricultural Sciences*, 23(1): 14-18.
- Pavela, R. (2011). Insecticidal properties of phenols on *Culex quinquefasciatus* Say and *Musca domestica* L. *Parasitology research*, 109(6): 1547-1553.
- Pawar, S. R., Desai, H. R., Bhandari, G. R. and Patel, C. J. (2017). Biology of the mealybug, *Phenacoccus solenopsis* Tinsley infesting Bt cotton. *International Journal of Current Microbiology and Applied Science*, 6: 1287-1297.
- Peeyush, K., Sapna, M., Anushree, M. and Santosh, S. (2011). Insecticidal properties of *Mentha* species: a review. *Industrial Crops and Products*, 34(1): 802-817.
- Pencinta Alam, "Garbage Enzyme", Newsletter of the Malaysian Nature Society, [Online], Available at HTTP: <http://reviews.ebay.com.sg> [22 Oct 2012].
- Penkhrue, W., Jendrossek, D., Khanongnuch, C., Pathom-Aree, W., Aizawa, T., Behrens, R. L. and Lumyong, S. (2020). Response surface method for polyhydroxybutyrate (PHB) bioplastic accumulation in *Bacillus drentensis* BP17 using pineapple peel. *PloS one*, 15(3): e0230443.
- Peraza-Sánchez, S. R., Chai, H. B., Shin, Y. G., Santisuk, T., Reutrakul, V., Farnsworth, N. R. and Kinghorn, A. D. (2002). Constituents of the leaves and twigs of *Ficus hispida*. *Planta medica*, 68(02): 186-188.
- Perry, J. J. and Scheld, H. W. (1968). Oxidation of hydrocarbons by microorganisms isolated from soil. *Canadian Journal of Microbiology*, 14(4): 403-407.
- Peterson, C. and Coats, J. (2001). Insect repellents-past, present and future. *Pesticide Outlook*, 12(4): 154-158.
- Pinang, P.S. 2012 "Change climate" [Online], Available at HTTP:<http://www.enzymesos.com> [12 Nov 2012].
- Pinkston, K. (1994). *Insects and Mites Affecting Ornamentals*. Oklahoma Cooperative Extension Service, Oklahoma State University. p. 53-54.

- Plata-Rueda, A., Martínez, L. C., Santos, M. H. D., Fernandes, F. L., Wilcken, C. F., Soares, M. A. and Zanuncio, J. C. (2017). Insecticidal activity of garlic essential oil and their constituents against the mealworm beetle, *Tenebrio molitor* Linnaeus (Coleoptera: Tenebrionidae). *Scientific reports*, 7(1): 1-11.
- Plata-Rueda, A., Rolim, G. D. S., Wilcken, C. F., Zanuncio, J. C., Serrão, J. E. and Martínez, L. C. (2020). Acute toxicity and sublethal effects of lemongrass essential oil and their components against the granary weevil, *Sitophilus granarius*. *Insects*, 11(6): 379.
- Prajapati, V., Tripathi, A. K., Aggarwal, K. K. and Khanuja, S. P. S. (2005). Insecticidal, repellent and oviposition-deterrent activity of selected essential oils against *Anopheles stephensi*, *Aedes aegypti* and *Culex quinquefasciatus*. *Bioresource technology*, 96(16): 1749-1757.
- Prakash, A. and Rao, J. (1997). *Botanical Pesticides in Agriculture*. 1st Edn., CRC Press Inc., Baton Rouge, Florida Pages: 461.
- Prakash, B. (2011). How to make and use garbage enzyme. Retrieved, 2(09), 2012. Available online: <http://www.ecowalkthetalk.com/> (accessed on 7 September 2018).
- Prakash, C., Kumarand, S. K. S. and Sree, P. S. (2019). Effect of Repellency on Mosquito Nets using Citrus Peel Extract. *International Journal of Innovative Technology and Exploring Engineering*, 8(11): 1531-1535.
- Prasad, Y. G., Prabhakar, M., Sreedevi, G., Rao, G. R. and Venkateswarlu, B. (2012). Effect of temperature on development, survival and reproduction of the mealybug, *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) on cotton. *Crop protection*, 39: 81-88.
- Prawira, H., Oramahi, H. A., Setyawati, D. and Diba, F. (2014). Application of liquid smoke *Vitex pubescens* Vahl wood for preservation rubber wood. *J. Hutan Lestari*, 1(1), 16-22.
- Price, D. N. and Berry, M. S. (2006). Comparison of effects of octopamine and insecticidal essential oils on activity in the nerve cord, foregut, and dorsal unpaired median neurons of cockroaches. *Journal of insect physiology*, 52(3): 309-319.
- Prishanthini, M. and Vinobaba, M. (2009). First record of new exotic Mealybug species, *Phenacoccus solenopsis* Tinsley, 1898 (Hemiptera: Pseudococcidae), its Host range and abundance in the Eastern Sri Lanka. *JSc-EUSL*, 6: 88-100.
- Prishanthini, M. and Vinobaba, M. (2014). Efficacy of some selected botanical extracts against the Cotton mealybug *Phenacoccus solenopsis* (Tinsley) (Hemiptera: Pseudococcidae). *International Journal of Scientific and Research Publications*, 4(3): 1-6.

- Prowse, G. M., Galloway, T. S. and Foggo, A. (2006). Insecticidal activity of garlic juice in two dipteran pests. *Agricultural and Forest Entomology*, 8(1), 1-6.
- Pumnuan, J. and Insung, A. (2016). Fumigant toxicity of plant essential oils in controlling thrips, *Frankliniella schultzei* (Thysanoptera: Thripidae) and mealybug, *Pseudococcus jackbeardsleyi* (Hemiptera: Pseudococcidae). *Journal of Entomological Research*, 40(1): 1–10.
- Pushpanathan, T., Jebanesan, A. and Govindarajan, M. (2006). Larvicidal, ovicidal and repellent activities of *Cymbopogon citratus* Stapf (Graminae) essential oil against the filarial mosquito *Culex quinquefasciatus* (Say)(Diptera: Culicidae). *Trop Biomed*, 23(2): 208-212.
- Puupponen-Pimiä, R., Nohynek, L., Ammann, S., Oksman-Caldentey, K. M. and Buchert, J. (2008). Enzyme-assisted processing increases antimicrobial and antioxidant activity of bilberry. *Journal of Agricultural and Food Chemistry*, 56(3): 681-688.
- Qiao-Li, L., Guang-Wen, L. and Yong-Yue, L. (2014). Toxicity of 14 insecticides to cotton mealybug *Phenacoccus solenopsis* Tinsely with leaf dipping method. *Guangdong Agricultural Sciences*, 6.
- Quisumbing, E. (1951). Medicinal plants of the Philippines. *Department of Agriculture and Commerce, Philippine Islands Technical Bulletin.*, (16).
- Radhakrishnan, J., Teasdale, J. R. and Coffman, C. B. (2002). Vinegar as a potential herbicide for organic agriculture. In *Proceedings of Northeastern Weed Science Society*, 56: 100.
- Ragasa C.Y. and Rufino L.A.A. (2011). Antimicrobial flavonoid from *Hibiscus rosa-sinensis* Linn. *The Manila Journal of Science*, 7(1): 12-18.
- Raguso, R. A. and Roy, B. A. (1998). ‘Floral’ scent production by Puccinia rust fungi that mimic flowers. *Molecular Ecology*, 7(9): 1127-1136.
- Rahman, K. A. and Latif, M. A. (1944). Description, bionomics and control of the giant mealybug, *Drosicha stebbingi*, Green (Homoptera: Coccidae). *Bulletin of Entomological Research*, 35(2): 197-209.
- Rahmat, B., Pangesti, D., Natawijaya, D. and Sufyadi, D. (2014). Generation of wood-waste vinegar and its effectiveness as a plant growth regulator and pest insect repellent. *BioResources*, 9(4): 6350-6360.
- Raja, R. R. (2012). Medicinally potential plants of Labiatae (*Lamiaceae*) family: an overview. *Research Journal of Medicinal Plant*, 6(3): 203-213.
- Rajendran, S. and Sriranjini, V. (2008). Plant products as fumigants for stored-product insect control. *Journal of stored products Research*, 44(2): 126-135.

- Ram, P. and Saini, R. K. (2010). Biological control of solenopsis mealybug, *Phenacoccus solenopsis* Tinsley on cotton: a typical example of fortuitous biological control. *Journal of Biological Control*, 24(2): 104-109.
- Rani, G. A., Padmalatha, C., Raj, R. S. and Singh, A. R. (2011). Impact of supplementation of amway protein on the economic characters and energy budget of silkworm *Bombyx mori* L. *Asian Journal of Animal Sciences*, 5(3): 190-195.
- Rao, C. N., Shivankar, V. J. and Singh, S. (2006). Citrus mealybug (*Planococcus citri* Risso) management-a review. *Agricultural Reviews*, 27(2): 142-146.
- Rashid, M. M. U., Khattak, M. K., Abdullah, K., Amir, M., Tariq, M. and Nawaz, S. (2012). Feeding potential of *Chrysoperla carnea* and *Cryptolaemus montrouzieri* on cotton mealybug, *Phenacoccus solenopsis*. *The Journal of Animal and Plant Sciences*, 22(3): 639-64.
- Rathod, P. K., Mane, P. N., Lande, G. K. and Sable, Y. R. (2008). First record of mealybug, *Maconellicoccus hirsutus* (Green) on sunflower in western Vidharbha (MS). *Insect Environ*, 14(1): 18.
- Ratnasekera, D. and Rajapakse, R. H. S. (2010). Repellent properties of plant oil vapours on pulse beetle (*Callasobruchus maculatus* L.)(Coleoptera: Bruchidae) in stored green gram (*Vigna radiata* Walp.). *Tropical Agricultural Research and Extension*, 12(1):13-16.
- Ravikumar (2004). Evaluation of organic and indigenous products for the management of *Helicoverpa armigera* (Hubner) in Chilli. M.Sc. (Agri.). Thesis, University. Agriculter. Science. Dharwad (India).
- Ray, R. C., and Sivakumar, P. S. (2009). Traditional and novel fermented foods and beverages from tropical root and tuber crops: review. *International Journal of Food Science and Technology*, 44: 1073-1087.
- Reddy, K. R. and Matcha, S. K. (2010). Quantifying nitrogen effects on castor bean (*Ricinus communis* L.) development, growth, and photosynthesis. *Industrial Crops and Products*, 31(1): 185-191.
- Regnault-Roger, C., Vincent, C. and Arnason, J. T. (2012). Essential oils in insect control: low-risk products in a high-stakes world. *Annual review of entomology*, 57: 405-424.
- Retief, E. (2000). Lamiaceae (Labiatae). In Seed Plants of Southern Africa;Leistner, O.A., Ed.; National Botanical Institute: Cape Town, South Africa; pp. 323–334.
- Ricupero, M., Biondi, A., Russo, A., Zappalà, L. and Mazzeo, G. (2021). The Cotton Mealybug Is Spreading along the Mediterranean: First Pest Detection in Italian Tomatoes. *Insects*, 12(8): 675.

- Ridgway, H. F., Safarik, J., Phipps, D., Carl, P. and Clark, D. (1990). Identification and catabolic activity of well-derived gasoline-degrading bacteria from a contaminated aquifer. *Applied and environmental microbiology*, 56(11): 3565-3575.
- Rishikumar, P., V., chauhan R. and Laljat, S. (2009). Bio-ecological studies on solenopsis mealybug, *Phenacoccus solenopsis* (Tinsley) (Hemiptera: Pseudococcidae) on cotton. *Proceedings of the national symposium on "Bt Cotton: opportunities and prospects*, CICR, Nagpur. p. 93.
- Rocha, C., San-Blas, F., San-Blas, G. and Vierma, L. (1992). Biosurfactant production by two isolates of *Pseudomonas aeruginosa*. *World Journal of microbiology and biotechnology*, 8(2): 125-128.
- Romeh, A. A. (2013). Phytochemicals from *Ficus sycomorus* L. leaves act as insecticides and acaricides. *African journal of agricultural research*, 8(27): 3571-3579.
- Roonjho, A.R., Gillani, W.A., Rasool, A., Akhtar, N., Mahmood, T., Arsalan, A., Afzal, M., Khan, I., Ranjha, M.A. and Khan, J. (2013). Repellency effects of different plant extracts to cotton mealy bug, *Phenococcus Solenopsis* Tinsley (Hemiptera: Pseudococcidae). *Pakistan Journal of Agricultural Research*, 26(3): 213-219.
- Roper, M. M. (2004). The isolation and characterisation of bacteria with the potential to degrade waxes that cause water repellency in sandy soils. *Soil Research*, 42(4): 427-434.
- Roslan, I., Shamshuddin, J., Fauziah, C.I. and Anuar, A.R. (2011). Fertility and Suitability of the Spodosols Formed on Sandy Beach Ridges Interspersed with Swales in the Kelantan Terengganu Plains of Malaysia for Kenaf Production. *Malaysian Journal of Soil Science*, 15: 1-24.
- Rovesti, L. and Deseö, K. V. (1991). Effectiveness of neem seed kernel extract against *Leucoptera malifoliella* Costa (Lep., Lyonetiidae). *Journal of Applied Entomology*, 111(1-5): 231-236.
- Roy, B., Amin, R., Uddin, M. N., Islam, A. T. M. S., Islam, M. J. and Halder, B. C. (2005). Leaf extracts of Shiyalmutra (*Blumea lacera* Dc.) as botanical pesticides against lesser grain borer and rice weevil. *Journal of Biological Sciences*, 5(2): 201-204.
- Rozendaal, J.A. (1997). *Vector control: methods for use by individuals and communitie*. Geneva, Switzerland: World Health Organization. p 7-177.
- Rugman-Jones, P. F., Morse, J. G. and Stouthamer, R. (2009). Rapid molecular identification of armored scale insects (Hemiptera: Diaspididae) on Mexican 'Hass' avocado. *Journal of Economic Entomology*, 102(5): 1948-1953.

- Rui, L. (2015). Insect pathogenic bacteria in integrated pest management. *Insects*, 6(2): 352-367.
- Ruiz, C. M. and Gomes, J. C. (2000). Effects of ethanol, acetaldehyde, and acetic acid on histamine secretion in guinea pig lung mast cells. *Alcohol*, 20(2): 133-138.
- Rummel D.J. (2005). *Botanical Beauty Book Compendium of Cosmetic Uses*. C and E Publishing, Inc., Quezon City, Philippines, 433 p.
- Rung, A., Scheffer, S. J., Evans, G. and Miller, D. O. U. G. L. A. S. S. (2008). Molecular identification of two closely related species of mealybugs of the genus *Planococcus* (Homoptera: Pseudococcidae). *Annals of the Entomological Society of America*, 101(3): 525-532.
- Ryu, C. M., Farag, M. A., Hu, C. H., Reddy, M. S., Wei, H. X., Paré, P. W. and Kloepper, J. W. (2003). Bacterial volatiles promote growth in *Arabidopsis*. *Proceedings of the National Academy of Sciences*, 100(8): 4927-4932.
- Saeed, S., Ahmad, M., Ahmad, M. and Kwon, Y. J. (2007). Insecticidal control of the mealybug *Phenacoccus gossypiphilus* (Hemiptera: Pseudococcidae), a new pest of cotton in Pakistan. *Entomological Research*, 37(2): 76-80.
- Saeidi, M., Moharramipour, S., Sefidkon, F. and Aghajanzadeh, S. (2011). Insecticidal and repellent activities of *Citrus reticulata*, *Citrus limon* and *Citrus aurantium* essential oils on *Callosobruchus maculatus*. *Integrated Protection of Stored Products IOBC/WPRS Bulletin*, 69: 289-293.
- Sahayaraj, K., Kombiah, P. and Kumar, D. S. (2011). Evaluation of insecticidal activity of fermented plant products on *Spodoptera litura* (Fab.). *Indian Journal of Agricultural Research*, 45(1): 77-82.
- Sahgal, G., Ramanathan, S., Sasidharan, S., Mordi, M. N., Ismail, S. and Mansor, S. M. (2009). In vitro antioxidant and xanthine oxidase inhibitory activities of methanolic *Swietenia mahagoni* seed extracts. *Molecules*, 14(11): 4476-4485.
- Sahito, H. A., Abro, G. H., Mahmood, R. and Malik, A. Q. (2011). Survey of mealybug, *Phenacoccus solenopsis* (Tinsley) and effect of bio-ecological factors on its population in different ecological zones of Sindh. *Pakistan Journal of Agriculture, Agricultural Engineering and Veterinary Sciences*, 27(1): 51-65.
- Sahito, H. A., Soomro, R. B., Talpur, M. A., Memon, S. A. and Dhilloo, K. H. (2012). Biology of mulberry mealybug, *Maconellicoccus hirsutus* (Green) in laboratory conditions. *Basic Research Journal of Agricultural Science and Review*, 1: 11-18.

- Saikia, P., Das, D. and Saikia, L. (2000). Evaluation of botanicals and fish oil formulation against bean aphid, *Aphis craccivora* Koch. *Journal of Agricultural Science. North-East, India*, 13: 79-80.
- Saini, R. K. and Ram, P. (2008). Incidence, nature of damage and chemical control of mealybug, *Phenacoccus* sp. on cotton in Haryana. *Proceeding of 2nd Congress on Insect Science*, 21-22.
- Sakthipriya, N., Doble, M. and Sangwai, J. S. (2015). Biosurfactant from *Pseudomonas* species with waxes as carbon source–Their production, modeling and properties. *Journal of Industrial and Engineering Chemistry*, 31: 100-111.
- Saladin, G. and Clément, C. (2005). Physiological side effects of pesticides on non-target plants. *Agriculture and soil pollution: new research*, 53-86.
- Salem, M. Z., Zayed, M. Z., Ali, H. M., El-Kareem, A. and Mamoun, S. M. (2016). Chemical composition, antioxidant and antibacterial activities of extracts from *Schinus molle* wood branch growing in Egypt. *Journal of Wood Science*, 62(6): 548-561.
- Salem, R. E. E. S. (2016). Side effects of certain pesticides on chlorophyll and carotenoids contents in leaves of maize and tomato plants. *Middle East Journal*, 5(4): 566.
- Salunkhe, R. B., Patil, C. D., Salunke, B. K., Rosas-García, N. M. and Patil, S. V. (2013). Effect of wax degrading bacteria on life cycle of the pink hibiscus mealybug, *Maconellicoccus hirsutus* (Green)(Hemiptera: Pseudococcidae). *BioControl*, 58(4): 535-542.
- Samanta, R., Dutta, A. K. and Sen, S. P. (1986). The utilization of leaf wax by N₂-fixing micro-organisms on the leaf surface. *The Journal of Agricultural Science*, 107(3): 681-685.
- Samarasekera, R., Kalhari, K. S. and Weerasinghe, I. S. (2006). Insecticidal activity of essential oils of *Ceylon Cinnamomum* and *Cymbopogon* species against *Musca domestica*. *Journal of essential oil research*, 18(3): 352-354.
- Saminathan, V. R. and Jayaraj, S. (2001). Evaluation of botanical pesticides against the mealybug, *Ferrisia virgata* Cockrell (Homoptera: Pseudococcidae) on cotton. *Madras Agricultural Journal*, 88(7/9): 535-537.
- Samuel, M., Oliver, S. V., Wood, O. R., Coetzee, M. and Brooke, B. D. (2015). Evaluation of the toxicity and repellence of an organic fatty acids mixture (C8910) against insecticide susceptible and resistant strains of the major malaria vector *Anopheles funestus* Giles (Diptera: Culicidae). *Parasites and Vectors*, 8: 321.
- Sánchez-Borzone, M. E., Marin, L. D. and García, D. A. (2017). Effects of insecticidal ketones present in mint plants on GABAA receptor from mammalian neurons. *Pharmacognosy Magazine*, 13(49): 114-117.

- Sanei-Dehkordi, A., Sedaghat, M. M., Vatandoost, H. and Abai, M. R. (2016). Chemical compositions of the peel essential oil of *Citrus aurantium* and its natural larvicidal activity against the malaria vector *Anopheles stephensi* (Diptera: Culicidae) in comparison with *Citrus paradisi*. *Journal of Aarthropod-borne Diseases*, 10(4): 577-585.
- Saramanda, G. and Kaparapu, J. (2017). Antimicrobial activity of fermented citrus fruit peel extract. *International Journal of Engineering Research and Application*, 7: 25-28.
- Sarma, A. K. (2013). Invasion of papaya mealybug, *Paracoccus marginatus* in Assam. *Indian Journal of Entomology*, 75: 355–356.
- Sarma, R., Adhikari, K., Mahanta, S. and Khanikor, B. (2019). Insecticidal activities of *Citrus aurantifolia* essential oil against *Aedes aegypti* (Diptera: Culicidae). *Toxicology Reports*, 6: 1091-1096.
- Sartiami, D., Gillian, Watson, W., Mohamad Roff, M. N. and Idris, A. B. (2016). November. Mealybugs (Hemiptera: Coccoomorpha: Pseudococcidae) attacking *Hibiscus rosasinensis* L. in Malaysia, with two new country records. *AIP Conference. Proceedings.*, 1784: 060007-1–060007-7.
- Sarvananda, L. (2018). Short introduction of DNA barcoding. *International Journal of Research*, 5(4): 673-685.
- SAS (2009). *SAS/IML 9.1 user guide*. SAS Institute. SAS publishing. Cary, North Carolina, USA.
- Sathyaseelan, V. and Bhaskaran, V. (2010). Efficacy of some native botanical extracts on the repellency property against the pink mealybug, *Maconellicoccus hirsutus* (green) in mulberry crop. *Recent Research in Science and Technology*, 2(10): 35-38.
- Satoto, T. B. T., Maniam, S. and Ganesen, K. (2013). Larvicidal effect of ether and chloroform extract of *Kaempferia galanga* against the larvae of *Aedes aegypti* (Diptera: Culicidae). *International Journal of Pharmacognosy and Phytochemical Research*, 5: 96-100.
- Satpute, N.S., Nagane V.V., Barkhade U.P. and Rathod, P.K. (2011). Biology of *Phenacoccus solenopsis* (Tinsley) on different hosts. *Indian Journal of Entomology*, 73 (3):234-236.
- Sayyad, M., Tiang, N., Kumari, Y., Goh, B. H., Jaiswal, Y., Rosli, R. and Shaikh, M. F. (2017). Acute toxicity profiling of the ethyl acetate fraction of *Swietenia macrophylla* seeds and in-vitro neuroprotection studies. *Saudi Pharmaceutical Journal*, 25(2): 196-205.
- Selvakumar, P. and Sivashanmugam, P. (2017). Optimization of lipase production from organic solid waste by anaerobic digestion and its application in biodiesel production. *Fuel Processing Technology*, 165: 1-8.

- Selvasundaram, R. and Muraleedharan, N. (1999). Neem formulations for control of pink mite in tea. *Biopesticides in Insect Pest Management*, Phoenix Publishing House, New Delhi, India pp, 33-7.
- Seni, A. and Chongtham, S. (2013). Papaya mealybug *Paracoccus marginatus* williams and granara de willink (Hemiptera: pseudococcidae), acurrent threat to agriculture-A review. *Agricultural Reviews*, 34(3): 216-222.
- Seni, A. and Naik, B. S. 2017. Bio-efficacy of some insecticides against mustard aphid, *Lipaphis erysimi* (Kalt.) (Hemiptera: Aphididae). *Journal of Entomology and Zoology Studies*, 5(6): 541-543.
- Seni, A. and Sahoo, A. K. (2014). Biology of the papaya mealybug, *paracoccus marginatus* williams and granara de willink (Hemiptera: pseudococcidae). *International Journal of Agriculture, Environment and Biotechnology*, 7(4): 875-881.
- Seo, K. I., Ha, K. J., Bae, Y. I., Jang, J. K. and Shim, K. H. (2000). Antimicrobial activities of oak smoke flavoring. *Korean Journal of Food Preservation*, 7(3): 337-341.
- Seyoum, A., Pålsson, K., Kung'a, S., Kabiru, E. W., Lwande, W., Killeen, G. F. and Knots, B. G. J. (2002). Traditional use of mosquito-repellent plants in western Kenya and their evaluation in semi-field experimental huts against *Anopheles gambiae*: ethnobotanical studies and application by thermal expulsion and direct burning. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 96(3): 225-231.
- Shaalán, E. A., Canyon, D. V., Younes, M. W., Abdel-Wahab, H. and Mansour, A. H. (2006). Efficacy of eight larvicidal botanical extracts from *Khaya senegalensis* and *Daucus carota* against *Culex annulirostris*. *Journal of the American Mosquito Control Association*, 22(3): 433-436.
- Shahid, M. R., Arif, M., Suhail, A., Khan, M.A. and Ali, A. (2013). Seasonal activity, host plant susceptibility and carryover of mealy bug *Phenacoccus solenopsis* (Tinsley) in Pakistan. *Journal of Agri-Food and Applied Sciences*, 1(3): 71-74.
- Shaji, S. M., Shahana, J., Thomas, A., Jiju, V. and Elesy, A. (2017). Herbal Insecticide and Pesticide-Save the Life and Future. *Journal of Pharmaceutical Sciences and Pharmacology*, 4(3): 34-40.
- Shanmugarajan, T. S., Arunsundar, M., Somasundaram, I., Krishnakumar, E., Sivaraman, D. and Ravichandiran, V. (2008). Cardioprotective effect of *Ficus hispida* Linn. on cyclophosphamide provoked oxidative myocardial injury in a rat model. *International Journal of Pharmacology*, 1: 1-10.
- Sharma, A. K., Sharma, V. and Saxena, J. (2016). A review paper on properties of fungal lipases. *International Journal of Current Microbiology and Applied Sciences*, 5(12), 123-130.

- Sharma, A., Kumar, V., Thukral, A. K. and Bhardwaj, R. (2019). Responses of plants to pesticide toxicity: An overview. *Planta Daninha*, 37(6): e019184291.
- Shawer, R., Donati, I., Cellini, A., Spinelli, F. and Mori, N. (2018). Insecticidal Activity of *Photorhabdus luminescens* against *Drosophila suzukii*. *Insects*, 9(4), 148.
- Shetu, H. J., Trisha, K. T., Sikta, S. A., Anwar, R., Rashed, S. S. B. and Dash, P. R. (2018). Pharmacological importance of *Kaempferia galanga* (Zingiberaceae): a mini review. *International Journal of Research in Pharmaceutical Sciences*, 3: 32-39.
- Shibulal, B., Al-Bahry, S. N., Al-Wahaibi, Y. M., Elshafie, A. E., Al-Bemani, A. S. and Joshi, S. J. (2017). The potential of indigenous *Paenibacillus ehimensis* BS1 for recovering heavy crude oil by biotransformation to light fractions. *PloS one*, 12(2): e0171432.
- Showler, A. T. (2017). Botanically based repellent and insecticidal effects against horn flies and stable flies (Diptera: Muscidae). *Journal of Integrated Pest Management*, 8,1(15): 1–11.
- Silva, C. A. D. D. (2012). Occurrence of new species of mealybug on cotton fields in the states of Bahia and Paraíba, Brazil. *Bragantia*, 71: 467-470.
- Simon, C., Frati, F., Beckenbach, A., Crespi, B., Liu, H. and Flook, P. (1994). Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Annals of the entomological Society of America*, 87(6): 651-701.
- Simpson, M. G. (2010). Diversity and classification of flowering plants: eudicots. *Plant Systematics (Second Edition)*, Academic Press, San Diego, 275-448.
- Singer, M. V. and Finnerty, W. R. (1990). Physiology of biosurfactant synthesis by *Rhodococcus* species H13-A. *Canadian journal of microbiology*, 36(11): 741-745.
- Singh, B.B. and Singh, R. (2014). Major rice insect pests in northeastern UP. *International Journal Life Science. Bt and Pharm.* 3(1): 124-143.
- Singh, F. and Khoshoo, T. N. (1989). Cytogenetic basis of evolution in garden Hibiscus. *Nucleus (Calcutta)*, 32(1-2), 62-67.
- Singh, M. J. and Sedhuraman, P. (2015). Biosurfactant, polythene, plastic, and diesel biodegradation activity of endophytic *Nocardiopsis* sp. mrinalini9 isolated from *Hibiscus rosa sinensis* leaves. *Bioresources and Bioprocessing*, 2: 1-7.
- Singh, V., Sood, A. K., Verma, K. S., Mehta, P. K. and Sood, S. (2016). Record of host plants and natural enemies of *Solenopsis* mealybug, *Phenacoccus solenopsis* Tinsley: A potential pest under protected cultivation in Himachal Pradesh. *Bioscan*, 11(1): 251-254.

- Sirisena, U. G. A. I., Watson, G. W., Hemachandra, K. S., Sage, O. and Wijayagunasekara, H. N. P. (2015). Scanning electron microscopy of six selected mealybugs (Hemiptera: Pseudococcidae) species of Sri Lanka. *Tropical Agricultural Research*, 26(2): 237–247.
- Sirisha, K., Kumar, C. G., Ramakrishna, K. V. S. and Gunda, S. K. (2017). Phenazine-1-carboxamide, an Extrolite Produced by *Pseudomonas aeruginosa* Strain (CGK-KS-1) Isolated from Ladakh and India, and its Evaluation Against Various *Xanthomonas* spp. *Microbiology and Biotechnology Letters*, 45(3): 209-217.
- Smith, K. and Dalton, S. (2015). 24, 48 and 72 hours contact toxicity test of peppermint (*Mentha piperita*), garlic (*Allium sativum* L.) and lemon (*Citrus limon* L.) essential oil on the long-tailed mealybug (*Pseudococcus longispinus*). *URSCA Proceedings*, 1.
- Soe, T. N., Ngampongsai, A. and Sittichaya, W. (2020). Bioactivity of some plant essential oils for seed treatment against pulse beetle, *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae) on mung bean. *Bulgarian Journal of Agricultural Science*, 26(1): 141-147.
- Soh, W. I. (2010). Evaluation of the mosquito repellent action of Cuban oregano (*Plectranthus amboinicus*) essential oil. *School of Bioprocess Engineering (FYP) [272]*. Universiti Malaysia Perlis.
- Solangi, G. S., Lohar, M. K., Abro, G. H. and Buriro, A. S. (2012). Biology and release of exotic predator *Cryptolaemus montrouzieri* Mulsant on mealybug *Phenacoccus solenopsis* Tinsley at Tandojam. *Sarhad Journal of Agriculture*, 28(3): 429-435.
- Soliman, A. A. A. (2015). *Effect of Neem, Jimson weed and propolis Powder Against bug (Phenacoccus solenopsis) (Hemiptera: Pseudococcidae) infesting okra fruits* (Doctoral dissertation, Sudan University of Science and Technology).
- Sood, N. and Lal, B. (2008). Isolation and characterization of a potential paraffin-wax degrading thermophilic bacterial strain *Geobacillus kaustophilus* TERI NSM for application in oil wells with paraffin deposition problems. *Chemosphere*, 70(8): 1445-1451.
- Sorkhoh, N. A., Ibrahim, A. S., Ghannoum, M. A. and Radwan, S. S. (1993). High-temperature hydrocarbon degradation by *Bacillus stearothermophilus* from oil-polluted Kuwaiti desert. *Applied Microbiology and Biotechnology*, 39(1): 123-126.
- Sparks Jr, A. N., Norman Jr, J. W. and Wolfenbarger, D. A. (1996). Efficacy of selected insecticides against the beet armyworm, *Spodoptera exigua*-field and laboratory evaluations. In *Beltwide Cotton Conferences*. National Cotton Council, Memphis, TN, 844-846.

- Spiders, J. D., Davies, T., He, C., Heinz, K. M., Bogran, C. E. and Starman, T. W. (2008). Do insecticide affect plant growth and development. *Greenh. Grow*, 2: 1-3.
- Sreerag, R. S. and Jayaprakas, C. A. (2014). Management of two major sucking pests using neem oil formulation. *Journal of Biofertilizers and Biopesticides*, 6(1): 1.
- Sreerag, R. S., Nishanth Kumar, S. and Jayaprakas, C. A. (2016). Characterization of mealy substance secreted by the tuber crops pest mealy bug, *Rhizoecus amorphophalli*. *Archives of Phytopathology and Plant Protection*, 49(7-8): 149-156.
- Srisukh, V., Tribuddharat, C., Nukoolkarn, V., Bunyaphrathatsara, N., Chokephaibulkit, K., Phoomniyom, S., Chuanphung, S. and Srifuengfung, S. (2012). Antibacterial activity of essential oils from *Citrus hystrix* (makrut lime) against respiratory tract pathogens. *Science Asia*, 38(2): 212-217.
- Srivastava, N., Singh, S., Gupta, A. C., Shanker, K., Bawankule, D. U. and Luqman, S. (2019). Aromatic ginger (*Kaempferia galanga* L.) extracts with ameliorative and protective potential as a functional food, beyond its flavor and nutritional benefits. *Toxicology Reports*, 6: 521-528.
- Steinkraus, K. (2018). Indigenous fermented foods involving an alkaline fermentation. In *Handbook of Indigenous Fermented Foods, revised and expanded* (pp. 365-378). CRC Press.
- Stevenson, P. C., Anderson, J. C., Blaney, W. M. and Simmonds, M. S. J. (1993). Developmental inhibition of *Spodoptera litura* (Fab.) larvae by a novel caffeoylquinic acid from the wild groundnut, *Arachis paraguariensis* (Chod et Hassl.). *Journal of Chemical Ecology*, 19(12): 2917-2933.
- Straub, T. M., Pepper, I. L. and Gerba, C. P. (1993). Hazards from pathogenic microorganisms in land-disposed sewage sludge. *Reviews of environmental contamination and toxicology*, 55-91.
- Su, W. C., Ge, Y. H., Wu, R. H., Xu, H. L., Xue, F. and Lu, C. T. (2016). Effects of bensulfuron-methyl residue on photosynthesis traits and chlorophyll fluorescence of corn seedlings. *Journal of Maize Science*, 24: 67-74.
- Subash, N. and Raju, G. (2014). Insecticidal activity of certain fermented plant extract against *Spodoptera litura* (Fab.) (Lepidoptera: Noctuidae) *International Journal of Institutional Pharmacy and Life Sciences* 4(5), September-October 2014.
- Suganthi, M., Janaki, I. and Sakthivel, P. (2012). Biology of mealy bugs, *Paracoccus marginatus* (Williams and Granara de Willink) and *Phenacoccus solenopsis* (Tinsley) on sunflower under greenhouse and laboratory. *Madras Agricultural Journal*, 99(4/6): 371-373.

- Sujana, P., Sridhar, T. M., Josthna, P. and Naidu, C. V. (2013). Antibacterial activity and phytochemical analysis of *Mentha piperita* L. (Peppermint)-An important multipurpose medicinal plant. *American Journal of Plant Sciences*, 4(1): 77–83.
- Sukari, M. A., Rashid, N. Y., Neoh, B. K., Bakar, N. A. and Riyanto, S. (2010). Larvicidal activity of some *Curcuma* and *Kaempferia* rhizome extracts against dengue fever mosquito *Aedes aegypti* Linnaeus (Diptera: Culicidae). *Asian Journal of Chemistry*, 22(10): 7915-7919.
- Sun, R., Zheng, X., Wang, X., Dang, L. Z., Yang, Z. S., Luo, S. L. and Li, G. H. (2011). Two new benzofuran derivatives from the fungus *Stereum* sp. YMF1. 1684. *Phytochemistry Letters*, 4(3): 320-322.
- Suresh, S., Jothimani, R., Sivasubramanian, P., Karuppuchamy, P., Samiyappan, R. and Jonathan, E. I. (2010). Invasive mealybugs of Tamil Nadu and their management. *Karnataka Journal of Agricultural Sciences*, 23(1): 6-9.
- Swofford, D. L., Thorne, J. L., Felsenstein, J. and Wiegmann, B. M. (1996). The topology-dependent permutation test for monophyly does not test for monophyly. *Systematic Biology*, 45(4): 575-579.
- Tamokou, J. D. D., Mbaveng, A. T. and Kuete, V. (2017). Antimicrobial activities of African medicinal spices and vegetables. In *Medicinal spices and vegetables from Africa* (pp. 207-237). Academic Press.
- Tamura, K. and Nei, M. (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular biology and evolution*, 10(3): 512-526.
- Tang, F. E. and Tong, C. W. (2011). A study of the garbage enzyme's effects in domestic wastewater. *International Journal of Environmental and Ecological Engineering*, 5(12): 887-892.
- Tankeo, S. B., Lacmata, S. T., Noumedem, J. A., Dzoyem, J. P., Kuate, J. R. and Kuete, V. (2014). Antibacterial and antibiotic-potential activities of some Cameroonian food plants against multi-drug resistant Gram-negative bacteria. *Chinese Journal of Integrative Medicine*, 20(7): 546-554.
- Tanwar, R. K., Jeyakumar, P. and Monga, D. (2007). Mealybugs and their management Technical Bulletin 19, September, 2007. *National Centre for Integrated Pest Management LBS Building, Pusa Campus, New Delhi*, 110(12): 1-10.
- Tanwar, R. K., Jeyakumar, P., Singh, A., Jafri, A. A. and Bambawale, O. M. (2011). Survey for cotton mealybug, *Phenacoccus solenopsis* (Tinsley) and its natural enemies. *Journal of Environmental Biology*, 32(3): 381.
- Tavares, W., Akhtar, Y., Gonçalves, G. L. P., Zanuncio, J. C. and Isman, M. B. (2016). Turmeric powder and its derivatives from *Curcuma longa* rhizomes: insecticidal effects on cabbage looper and the role of synergists. *Scientific reports*, 6: 34093.

- Tawatsin, A., Wratten, S. D., Scott, R. R., Thavara, U. and Techadamrongsin, Y. (2001). Repellency of volatile oils from plants against three mosquito vectors. *Journal of Vector Ecology*, 26: 76-82.
- Taylor, L. D. and Magdalita, P. M. (2021). Incidence of insect pests on *Hibiscus rosa sinensis* L. germplasm in the plant nursery. *Philippine Science Letters*, 14(01).
- Tevini, M. and Teramura, A. H. (1989). UV-B effects on terrestrial plants. *Photochemistry and Photobiology*, 50(4): 479-487.
- Thangamalar, A., Subramanian, S. and Mahalingam, C. A. (2010). Bionomics of papaya mealybug, *Paracoccus marginatus* and its predator *Spalgus epius* in mulberry ecosystem. *Karnataka Journal of Agricultural Sciences*, 23(1): 39-41.
- Thavara, U., Tawatsin, A., Bhakdeenuan, P., Wongsinkongman, P., Boonruad, T., Bansiddhi, J. and Mulla, M. S. (2007). Repellent activity of essential oils against cockroaches (Dictyoptera: Blattidae, Blattellidae, and Blaberidae) in Thailand. *The Southeast Asian Journal of Tropical Medicine and Public Health*, 38(4): 663-673.
- Thinnaluri, M., Bhaskar, R., Mahesh, N. and Narayanaswamy, T. K. (2014). Evaluation of botanical extracts on the repellency property against the Pink Mealybug, *Maconellicoccus hirsutus* (Green) in mulberry. *International Journal of Development Research*, 4(8): 1504-1507.
- Thomas, A. and Ramamurthy, V. V. (2008). On the problems in diagnostics of cotton mealybug *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae). *Entomon*, 33: 277-282.
- Throne-Holst, M., Wentzel, A., Ellingsen, T. E., Kotlar, H. K. and Zotchev, S. B. (2007). Identification of novel genes involved in long-chain n-alkane degradation by *Acinetobacter* sp. strain DSM 17874. *Applied and environmental microbiology*, 73(10): 3327-3332.
- Townsend, M. L., Oetting, R. D. and Chong, J. H. (2000). Management of the mealybug *Phenacoccus madeirensis*. In *Proceeding of Southern Nursing Association and Research Conference*, 45: 162-166.
- Tworowski, T. (2002). Herbicide effects of essential oils. *Weed science*, 50(4): 425-431.
- Ugborogho, R. E. and Shofoyeke, B. A. (1983). Observations on the morphology, reproductive biology and economic importance of *Hibiscus esculentus* L. and *H. sabdariffa* L. (Malvaceae) in Nigeria. *Nigerian Field*, 48: 79-80.
- Umar, M. I., Asmawi, M. Z. B., Sadikun, A., Altaf, R. and Iqbal, M. A. (2011). Phytochemistry and medicinal properties of *Kaempferia galanga* L. (Zingiberaceae) extracts. *Afr. J. Pharm. Pharmacol*, 5(14): 1638-1647.

- Untergasser, A., Cutcutache, I., Koressaar, T., Ye, J., Faircloth, B. C., Remm, M. and Rozen, S. G. (2012). Primer3—new capabilities and interfaces. *Nucleic acids research*, 40(15): e115-e115.
- Untiedt, R. and Blanke, M. M. (2004). Effects of fungicide and insecticide mixtures on apple tree canopy photosynthesis, dark respiration and carbon economy. *Crop Protection*, 23(10): 1001-1006.
- Uses of Garbage Enzyme. (2010). WWW.o3enzyme.com.<http://www.o3enzyme.com> (accessed June 5, 2010).
- Utsumi, Y., Utsumi, C., Tanaka, M., Ha, C. V., Takahashi, S., Matsui, A. and Seki, M. (2019). Acetic acid treatment enhances drought avoidance in cassava (*Manihot esculenta* Crantz). *Frontiers in Plant Science*, 10: 521.
- Van Lenteren, J. E. and Woets, J. V. (1988). Biological and integrated pest control in greenhouses. *Annual review of Entomology*, 33(1): 239-269.
- Varjani, S. J. and Upasani, V. N. (2016). Biodegradation of petroleum hydrocarbons by oleophilic strain of *Pseudomonas aeruginosa* NCIM 5514. *Bioresource technology*, 222: 195-201.
- Veerapagu, M., Jeya, K. R., Kalaivani, R., Jeyanthi, K. A. and Geethanjali, S. (2019). Screening of hydrocarbon degrading bacteria isolated from oil contaminated soil. *The Pharma Innovation Journal*, 8(6): 69-72.
- Vennila, S., Deshmukh, A. J., Pinjarkar, D., Agarwal, M., Ramamurthy, V. V., Joshi, S. and Bambawale, O. M. (2010). Biology of the mealybug, *Phenacoccus solenopsis* on cotton in the laboratory. *Journal of Insect Science*, 10(1): 115.
- Vennila, S., Prasad, Y. G., Prabhakar, M., Kumar, R., Nagrare, V. and Amutha, M. (2011). Spatio-temporal distribution of host plants of cotton mealybug *Phenacoccus solenopsis* Tinsley in India, NCIPM. *Technical Bulletin No. 26*, National Centre for Integrated Pest Management, New Delhi. P 50.
- Verger, P. J. and Boobis, A. R. (2013). Reevaluate pesticides for food security and safety. *Science*, 341(6147): 717-718.
- Verma, S. (2017). International Journal of Research in *Hibiscus rosa-sinensis* L. (Malvaceae): A Multipurpose Ornamental Plant, 6: 61-64.
- Voet, V. (2012). "Ways to Save Energy", [Online], Available at <http://http://www.Ways to save energy.net> [8] Nov [2012].
- Vokou, D., Douvli, P., Blionis, G. J. and Halley, J. M. (2003). Effects of monoterpenoids, acting alone or in pairs, on seed germination and subsequent seedling growth. *Journal of chemical ecology*, 29(10): 2281-2301.
- Vuković, S. M., Indić, D. V. and Gvozdenac, S. M. (2014). Phytotoxic effects of fungicides, insecticides and nonpesticidal components on pepper depending on

water quality. *Pesticides and Phytomedicine/Pesticidi i fitomedicina*, 29(2): 145-153.

Wakeil, N. E. (2013). Botanical pesticides and their mode of actions. *Review article Gesunde Pflanzen*, 65(4): 125-149.

Walia, S., Saha, S. and Rana, V. S. (2014). Phytochemical pesticides. In *Advances in Plant Biopesticides* (pp.295-322). Springer, New Delhi.

Walker, A., Hoy, M. and Meyerdirk, D. (2006). Papaya mealybug (*Paracoccus marginatus* Williams and Granara de Willink (Insecta: Hemiptera: Pseudococcidae)). *Featured Creatures. Entomology and Nematology Department, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences (IFAS), University of Florida, Gainesville, FL.*

Wand, M. E., McCowen, J. W., Nugent, P. G. and Sutton, J. M. (2013). Complex interactions of *Klebsiella pneumoniae* with the host immune system in a *Galleria mellonella* infection model. *Journal of medical microbiology*, 62(12): 1790-1798.

Wang, W. and Shao, Z. (2013). Enzymes and genes involved in aerobic alkane degradation. *Frontiers in microbiology*, 4: 116.

Wang, X. G., Wei, X. Y., Huang, X. Y., Shen, L. T., Tian, Y. Q. and Xu, H. H. (2011). Insecticidal constructure and bioactivities of compounds from *Ficus sarmentosa* var. henryi. *Agricultural Sciences in China*, 10(9): 1402-1409.

Wang, Y., Wu, S. and Zhang, R. (2009). Pest risk analysis of a new invasive pest, *Phenacoccus solenopsis*, to China. *Chinese Bulletin of Entomology*, 46(1): 101-106.

Wanna, R. and Kwang-Ngoen, P. (2019). Efficiency of Indian borage essential oil against cowpea bruchids. *International Journal*, 16(56): 129-134.

Warabi, E., Usui, K., Tanaka, Y. and Matsumoto, H. (2001). Resistance of a soybean cell line to oxyfluorfen by overproduction of mitochondrial protoporphyrinogen oxidase. *Pest Management Science: formerly Pesticide Science*, 57(8): 743-748.

Waterfield, N., Dowling, A., Sharma, S., Daborn, P. J., Potter, U. and Ffrench-Constant, R. H. (2001). Oral toxicity of *Photographus luminescens* W14 toxin complexes in *Escherichia coli*. *Applied and Environmental Microbiology*, 67(11): 5017-5024.

Watson, G. W. and Kubiriba, J. (2005). Identification of mealybugs (Hemiptera: Pseudococcidae) on banana and plantain in Africa. *African Entomology*, 13(1): 35-47.

Williams, D. J. (1996). A brief account of the hibiscus mealybug *Maconellicoccus hirsutus* (Hemiptera: Pseudococcidae), a pest of agriculture and horticulture,

- with descriptions of two related species from southern Asia. *Bulletin of Entomological Research*, 86(5): 617-628.
- Williams, D. J. (2004). *Mealybugs of southern Asia* (pp. 1-896). Natural History Museum/Southdene Sdn Bhd, Kuala Lumpur, Malaysia.
- Williams, D. J. and Granara De Willink (1992). *Mealybugs of Central and South America*. CAB International. Wallingford, England, 635 p.
- Wilson, F. D. and Menzel, M. Y. (1964). Kenaf (*Hibiscus cannabinus*), roselle (*Hibiscus sabdariffa*). *Economic Botany*, 18(1): 80-91.
- Wink, M. (1993). Production and application of phytochemicals from an agricultural perspective. *Phytochemistry and agriculture*, 34: 171-213.
- Wink, M. (2006). Importance of plant secondary metabolites for protection against insects and microbial infections. *Advances in Phytomedicine*, 3: 251-268.
- Wong, S. K., Lim, Y. Y. and Chan, E. W. C. (2009). Antioxidant properties of Hibiscus: Species variation, altitudinal change, coastal influence and floral colour change. *Journal of Tropical Forest Science*, 21(4): 307-315.
- Wood, B. W. and Payne, J. A. (1986). Net photosynthesis of orchard grown pecan leaves reduced by insecticide sprays. *HortScience*, 21(1): 112-113.
- Wood, M. T., Miles, R. and Tabora, P. (1997). EM fermented plant extract and EM5 for controlling pickleworm (*Diaphania nitidalis*) in organic cucumber. *School of Natural Resources, University of Missouri, USA and EARTH College, Limon, Costa Rica*.
- Wu, F., Liu, Z., Shen, H., Yu, F., Ma, J., Hu, X. and Zeng, L. (2014). Morphological and molecular identification of *Paracoccus marginatus* (Hemiptera: Pseudococcidae) in Yunnan, China. *Florida Entomologist*, 97(4): 1469-1473.
- Wu, S. and Zhang, R. (2009). A new invasive pest, *Phenacoccus solenopsis*, threatening seriously to cotton production. *Chinese bulletin of entomology*, 46(1): 159-162.
- Xia, X. J., Huang, Y. Y., Wang, L., Huang, L. F., Yu, Y. L., Zhou, Y. H. and Yu, J. Q. (2006). Pesticides-induced depression of photosynthesis was alleviated by 24-epibrassinolide pretreatment in *Cucumis sativus* L. *Pesticide Biochemistry and Physiology*, 86(1): 42-48.
- Xu, J., Zhang, Y., Hung, T. and Deng, H. X. (2013). Growth characteristics of seven hydrocarbon-degrading active bacteria isolated from oil contaminated soil. *Nature Environment and Pollution Technology*, 12(1): 17.
- Yaghoobi-Ershadi, M.R., Akhavan, A. A., Jahanifard, E., Vatandoust, H., Amin, G. H., Mousavi, L. and Arandian, M. H. (2006). Repellency effect of Myrtle essential oil and DEET against *Phlebotomus papatasi*, under laboratory conditions.

- Yakubu, B. L. and Nda, A. J. (2012). Evaluation of (*Khaya senegallensis* for the control of *Tribolium confusum* Duval (Coleoptera: Tenebrionidae) on Stored Pearl Millet, *Advances in life Science and Technology*, 4: 58-63.
- Yasmin, M. S., Bachchu, M. A. A. and Hossain, M. A. (2017). Toxic and repellent effects of three botanical oils against adult *Aphis craccivora* Koch (Homoptera: Aphididae) under laboratory conditions. *University Journal of Zoology Rajshahi University*, 36: 39-48.
- Youngman, R. R., Leigh, T. F., Kerby, T. A., Toscano, N. C. and Jackson, C. E. (1990). Pesticides and cotton: effect on photosynthesis, growth, and fruiting. *Journal of economic entomology*, 83(4): 1549-1557.
- Yunus, A. and Ho, T. H. (1980). List of economic pests, host plants, parasites and predators in West Malaysia (1920-1978. *Bulletin Ministry of Agriculture, Malaysia*, (153).
- Zhang Y., Smith, P., Maximova, S. N. and Guiltinan, M. J., (2015). Application of glycerol as a foliar spray activates the defense response and enhances disease resistance of *Theobroma cacao*. *Molecular Plant Pathology*, 16(1): 27–37.
- Zhang, M., Zeiss, M. R. and Geng, S. (2015). Agricultural pesticide use and food safety: California's model. *Journal of Integrative Agriculture*, 14(11): 2340-2357.
- Zhang, Z. Shao, M. Qi, S. Guo, X. Ma, H. and Ren, H. (1988). Study on wax secretion capacity of the white wax insect (*Ericeruspela Chavannes*) in China. *Zoology Research*, 9(01): 93-97.
- Zhou, Y. Q., Liu, H., He, M. X., Wang, R., Zeng, Q. Q., Wang, Y., Ye, W. C. and Zhang, Q. W. (2018). Chapter 11-A Review of the Botany, Phytochemical, and Pharmacological Properties of Galangal. In *Natural and Artificial Flavoring Agents and Food Dyes*; Grumezescu A. M.; Holban A. M., Eds.; Academic Press, pp 351–396.
- Zibae, I. (2015). Synergistic effect of some essential oils on toxicity and knockdown effects, against mosquitos, cockroaches and housefly. *Arthropods*, 4(4): 107.
- Zulfikar, A. W. and Sitepu, F. Y. (2019). The effect of lemongrass (*Cymbopogon nardus*) extract as insecticide against *Aedes aegypti*. *Int J Mosq Res*, 6(1): 101-103.
- Zurovcova, M., Havelka, J. A. N., Starý, P. E. T. R., Vechtova, P., Chundelova, D., Jarosova, A. and Kucerova, L. (2010). "DNA barcoding" is of limited value for identifying adelgids (Hemiptera: Adelgidae) but supports traditional morphological taxonomy. *European Journal of Entomology*, 107(2): 147.