

GROWTH, YIELD AND ISOTOPE COMPOSITION OF CARBON AND NITROGEN IN RICE AS INFLUENCED BY DIFFERENT POTASSIUM FERTILIZER SOURCES IN SELECTED GRANARY AREAS

By

NURAINI SHAFINAZ BINTI MOHD ANUAR

Thesis Submitted to the School of Graduate Studies University Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

June 2023

All materials contained within this thesis, including without limitation of text, logos, icons, photographs, and all other artwork, is copyright material of University Putra Malaysia unless otherwise stated. Use may be made of material contained within the non-commercial thesis purposes from the copyright holder. Commercial use of the material may only be made with the express, prior, written permission of University Putra Malaysia.

Copyright © University Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

GROWTH, YIELD AND ISOTOPE COMPOSITION OF CARBON AND NITROGEN IN RICE AS INFLUENCED BY DIFFERENT POTASSIUM FERTILIZER SOURCES IN SELECTED GRANARY AREAS

By

NURAINI SHAFINAZ BINTI MOHD ANUAR

June 2023

Chair : Adibah Mohd Amin, PhD

Faculty: Agriculture

Rice (*Oryza sativa L.*) plays a vital role in ensuring food security in Malaysia, particularly in the granary areas such as MADA and KADA. These regions serve as significant rice-producing areas, contributing substantially to the nation's rice production. The key role of potassium (K) fertilization is crucial in rice cultivation, impacting various physiological processes and contributing significantly to the quality and yield of rice crops in granary areas. Exploring alternative potassium sources, such as polyhalite, holds potential for improving Malaysian agriculture.

The examination of $\delta^{13}C$ and $\delta^{15}N$ provides significant insights into the acquisition of resources and interactions among plants. This is because the natural abundance of carbon-13 and nitrogen-15 in plants reflects the isotopic composition of their carbon and nitrogen sources and plays a crucial role in advancing our understanding of various ecological and biological processes. However, the extent to which the intake of K affects the isotopic variance in rice has not been well studied.

Therefore, this study aims to comprehensively investigate the effects of potassium (K) fertilizers on rice plant physiological parameters and yield performance. The primary objectives include assessing the impact of K fertilizer sources on stable carbon and nitrogen isotope values (specifically $\delta^{13}C$ and $\delta^{15}N)$ in rice tissues, examining the influence of K fertilizers on rice photosynthesis, and investigating the effects of K fertilizers on rice yield. Field tests in KADA and MADA regions and a rain shelter experiment were conducted to achieve these goals, providing valuable insights into agriculture and crop management.

Rice plants were cultivated in the field and under a rain shelter with five treatments, which are T1 (no-K), T2 (Muriate of Potash), T3 (Sulphate of Potash), T4 (Polyhalite),

and T5 (conventional fertilizer). The fertilizers were applied 3, 15, 55, and 75 days after planting (DAP). Leaf photosynthesis and stomata conductance measurements were taken at 85 DAP and dried and reserved for carbon and nitrogen isotope analyses. Meanwhile, the rice yield was determined at 110 DAP during harvest.

In this study, the rain shelter trials highlighted the positive influence of K fertilizers on rice yield, with Polyhalite consistently providing the highest yields under controlled conditions with 490.4 g/pot for KADA and 489.60 g/pot for MADA soils compared to other treatments. Overall, adding K fertilizer proved beneficial for rice plant health, tiller productivity, and grain yield, emphasizing the importance of potassium fertilization in enhancing overall rice productivity. While soil type does not significantly impact rice yield, MADA soil demonstrates a more conducive environment for photosynthesis and improved efficiency in water and nutrient absorption with a mean of 52.77 µmol CO₂ m⁻² s⁻¹ for rate of photosynthesis and 1.88 mmol m⁻² s⁻¹ for stomata conductance. Polyhalite, a potassium source, significantly boosted photosynthetic rates and stomata conductance. The findings also indicated that K fertilizer application could influence $\delta^{15}N$ in plants, with variations observed in different treatments and environmental conditions. The comprehensive analysis of correlation coefficients highlighted complex interactions between rice yield, rate of photosynthesis, stomata conductance, and isotopic values, providing valuable insights for crop management and sustainable agricultural practices. In conclusion, the study emphasized the importance of K in enhancing rice productivity, influencing plant physiology, and contributing to the intricate dynamics of carbon and nitrogen isotopic signatures in plants.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PERTUMBUHAN, HASIL, DAN KOMPOSISI ISOTOP KARBON DAN NITROGEN DALAM PADI YANG DIPENGARUHI OLEH SUMBER BAJA KALIUM YANG BERBEZA DI KAWASAN JELAPANG PADI YANG TERPILIH

Oleh

NURAINI SHAFINAZ BINTI MOHD ANUAR

Jun 2023

Pengerusi : Adibah Mohd Amin, PhD

Fakulti : Pertanian

Padi (*Oryza sativa L.*) memainkan peranan penting dalam memastikan jaminan makanan di Malaysia, terutamanya di kawasan-kawasan bendang seperti MADA dan KADA. Kawasan-kawasan ini berperanan besar dalam pengeluaran padi negara. Peranan utama baja kalium (K) adalah penting dalam penanaman padi, ia mempengaruhi pelbagai proses fisiologi dan memberi sumbangan yang signifikan kepada kualiti dan hasil tanaman padi di kawasan-kawasan bendang. Penerokaan sumber kalium alternatif, seperti polihalit, memiliki potensi untuk meningkatkan pertanian Malaysia.

Nisbah metabolit Karbon (C) kepada metabolit Nitrogen (N) dalam sel, yang dirujuk sebagai keseimbangan C/N, juga penting untuk pengawalan pertumbuhan dan perkembangan tumbuhan. Oleh itu, pengukuran isotop yang dikaitkan dengan sumber tumbuhan kritikal, seperti karbon dan nitrogen, telah membantu memperdalam pemahaman ekologi tentang pemerolehan sumber tumbuhan dan interaksi tumbuhan. Ini kerana kelimpahan semula jadi karbon-13 dan nitrogen-15 dalam tumbuhan mencerminkan komposisi isotop sumber karbon dan nitrogen serta memainkan peranan penting dalam meningkatkan pemahaman tentang pelbagai proses ekologi dan biologi tumbuhan. Namun, sejauh mana pengambilan kalium (K) mempengaruhi variasi isotop dalam padi masih belum dikaji dengan baik.

Oleh itu, kajian ini bertujuan untuk menyelidik secara komprehensif kesan baja kalium (K) ke atas parameter fisiologi tanaman padi dan prestasi hasil padi. Objektif utama termasuk menilai impak sumber baja K ke atas nilai isotop karbon dan nitrogen yang stabil (khususnya δ^{13} C dan δ^{15} N) dalam tisu padi, mengkaji pengaruh baja K terhadap fotosintesis padi, dan menyiasat kesan baja K terhadap hasil padi. Ujian lapangan di kawasan KADA dan MADA, berserta eksperimen di dalam rumah lindungan hujan,

telah dijalankan untuk mencapai objektif ini untuk memberikan pandangan berharga dalam bidang pertanian dan pengurusan tanaman.

Padi ditanam di lapangan dan di dalam rumah lindungan hujan dengan lima rawatan, iaitu T1 (tanpa K), T2 (Muriate of Potash), T3 (Sulphate of Potash), T4 (Polyhalite), dan T5 (baja konvensional). Baja telah diberikan kepada tanaman pada 3, 15, 55, dan 75 hari selepas tanam (HST). Ukuran fotosintesis daun dan pengukuran konduktiviti stomatal diambil pada 85 (HST) dan kemudiannya dikeringkan serta dikhaskan untuk analisis isotop karbon dan nitrogen. Sementara itu, hasil padi ditentukan pada 110 (HST) semasa menuai.

Dalam kajian ini, eksperimen di dalam rumah lindungan hujan menonjolkan pengaruh positif baja K ke atas hasil padi, dengan Polyhalite secara konsisten memberikan hasil tertinggi di bawah keadaan terkawal dengan 490.4 g/pot untuk tanah KADA dan 489.60 g/pot untuk tanah MADA berbanding rawatan lain. Secara keseluruhan, penambahan baja K terbukti bermanfaat bagi kesihatan tanaman padi, produktiviti anakan padi, dan hasil bijirin, menekankan kepentingan penanaman kalium dalam meningkatkan produktiviti padi secara keseluruhan. Walaupun jenis tanah tidak memberi kesan yang signifikan kepada hasil padi, tanah MADA menunjukkan persekitaran yang lebih kondusif untuk fotosintesis dan penyerapan air serta nutrien yang lebih baik dengan purata 52.77 μmol CO₂ m² s⁻¹ untuk kadar fotosintesis dan 1.88 mmol m² s⁻¹ untuk konduktan stomata. Polyhalite, sebagai sumber kalium, secara signifikan meningkatkan kadar fotosintesis dan konduktan stomata. Penemuan juga menunjukkan bahawa penggunaan baja K boleh mempengaruhi δ¹⁵N dalam tumbuhan, dengan variasi yang diperhatikan dalam rawatan dan keadaan alam sekitar yang berbeza. Analisis menyeluruh bagi pekali korelasi menonjolkan interaksi yang kompleks antara hasil padi, kadar fotosintesis, konduktan stomata, dan nilai isotopik, memberikan pemahaman berharga untuk pengurusan tanaman dan amalan pertanian lestari. Kesimpulannya, kajian ini menekankan kepentingan K dalam meningkatkan produktiviti padi, mempengaruhi fisiologi tanaman, dan menyumbang kepada dinamika rumit nisbah isotopik karbon dan nitrogen dalam tumbuhan.

ACKNOWLEDGEMENTS

I am grateful to God Almighty for His protection, guidance, wisdom, and strength throughout my studies. He has brought me to a successful end, just as he promised. Therefore, all praise and thanks are due to Allah.

I want to express my sincere gratitude to my esteemed supervisors, Dr. Adibah Mohd Amin and Dr. Muhammad Firdaus Sulaiman. Their invaluable support and guidance during the course of my thesis and presentation have been instrumental in shaping my academic career. Their timely feedback and insightful comments have been immensely helpful in refining the quality of my work. I am genuinely thankful for their unwavering support and encouragement throughout this journey.

I want to express my sincere appreciation for the collaborative efforts of KADA, MADA, and the Department of Chemistry Malaysia. Expressly, the Department of Land Management, Faculty of Agriculture, Universiti Putra Malaysia (UPM), in partnership with the Quality Management and Research Center of the Department of Chemistry Malaysia, has provided consumables according to the number of tree tissue samples required for the analysis of carbon 13 and nitrogen 15 isotope levels in tissue samples of paddy. This collaboration has been invaluable in our research efforts, and we are grateful for the support and expertise of all involved.

I thank Dr. Zulkarami Berahim, a teacher at the Institute of Tropical Agriculture and Food Security (ITAFoS), for assisting me with my project on leaf physiology. He has given me valuable advice and input. In particular, his direction in operating the open-top portable photosynthesis system (Li-6400XT, LI-COR, Lincoln, Nebraska, USA) has been indispensable to the research.

Words cannot express my sincere appreciation to the research assistants who played a significant role in this project's success. Their unwavering guidance, contributions, and expertise, particularly in statistical analysis and lab experiment management, have been invaluable. I would also like to acknowledge their constant support, motivation, and encouragement during my low points, which gave me the strength to persevere. I am equally grateful to my classmates and friends whose constructive recommendations and comments have significantly enhanced the quality of my thesis. I am confident that this project's outcome will be of immense value, and I pray that God blesses everyone who played a part in its success.

I want to take a moment to express my sincere gratitude to my family for their incredible patience, sacrifice, support, and encouragement during my research. I am so lucky to have such kind and gentle parents who have always been there for me, believing in me and supporting me no matter what. Finally, to all those who have provided me with their encouragement and support throughout my studies, I want to say a heartfelt thank you. Your kindness and generosity have meant more to me than words can express.

This thesis was submitted to the Senate of University Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master OF Science. The members of the Supervisory Committee were as follows:

Adibah binti Mohd Amin, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Muhammad Firdaus Sulaiman, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies University Putra Malaysia

Date: 14 March 2024

TABLE OF CONTENTS

APPROV DECLAR LIST OF LIST OF	K WLEDE VAL RATION TABLE FIGURI		Page i iii v vi viii xiii xv
СНАРТН	ER		
1	1.2 1.3 1.4 1.5	Introduction 1.1.1 The World Rice Situation 1.1.2 Malaysia Rice Production 1.1.3 Potassium (K) Fertilizer 1.1.4 Natural Abundance of Isotopic Ratios Problem Statement Research Objectives Significance of the Study Organization of the Thesis	1 1 1 2 6 6 7 8 8 9
2	LITE 2.1	Botanical of Rice 2.1.1 Physical Properties of Rice 2.1.2 Mechanical Properties of Rice	10 10 10 11
	2.2	Soil Properties of Paddy Field in KADA and MADA Soil Properties of Paddy Field in KADA and MADA	11 12
		2.3.1 Source and Types of Potassium Fertilizer The Comparison Between Types of K Fertilizer	12 13
	2.4	Roles of Potassium (K) in Plants 2.4.1 Role of K in Photosynthesis 2.4.2 Role of K in Stomatal Conductance 2.4.3 Role of K in the Metabolism of Plants 2.4.4 Role of K in Cell Elongation and Cell Membrane Activity 2.4.5 Role of K in Protein Synthesis of Rice	14 15 16 16 17
	2.5 2.6 2.7 2.8 2.9	2.4.5 Role of K in Protein Synthesis of Rice 2.4.6 Role of K in Total Soluble Sugar of Rice Plant Nutrient Interaction Synergistic and Antagonistic Nutrient Interaction Uses of Carbon Isotope in Plants Carbon Isotope Discrimination Uses of Nitrogen Isotope in Plants Nitrogen Isotope Discrimination	17 17 18 18 19 19 20

3	METH	IODOLOGY	23
	3.1	Planting Material	23
	3.2	Study Area	23
	3.3	Treatments	25
	3.4	Experimental Design	26
		3.4.1 Determination of Land Suitability	27
		3.4.2 Cultivation	29
		3.4.3 Fertilizer and Water Management	30
	3.5	Measurement of Parameters	35
		3.5.1 Soil Analysis	35
		3.5.2 Harvesting and Yield Component	36
		Measurements	
		3.5.3 Plant Analysis	36
		3.5.4 Leaf Physiology	37
		3.5.5 Stable Isotopes Ratio Determination	38
	3.6	Data Statistical Analysis	38
4		LTS AND DISCUSSION	40
	4.1	Effect of K on Soil Chemical Characteristics	40
		4.1.1 KADA: Paddy Field Study	41
		4.1.2 MADA: Paddy Field Study	43
		4.1.3 KADA: Rain Shelter Experiment	46
	//	4.1.4 MADA Rain Shelter Experiment	49
	4.2	Effects of K Fertilizer on Rice Yield	50
	4.3	Rice Yield Parameter	52
		4.3.1 KADA: Paddy Field Study	52
		4.3.2 MADA: Paddy Field Study	53
		4.3.3 KADA: Rain Shelter Experiment	54
		4.3.4 MADA: Rain Shelter Experiment	56
	4.4	Effect of K on Nutrient Content of Rice Plant	57
		Tissue	
		4.4.1 KADA: Paddy Field Study	57
		4.4.2 MADA: Paddy Field Study	58
		4.4.3 KADA: Rain Shelter Experiment	59
	4.5	4.4.4 MADA: Rain Shelter Experiment	60
	4.5	Correlation Between Yield, Rate of Photosynthesis,	61
	4.6	Stomata Conductance, δ13C and δ15N Effects of K Fertilizer on Photosynthesis Rate	62
	4.0	Effects of K Fertilizer on Stomatal Conductance	64
	4.7	δ13C Value in Rice Plant Tissue at Different	04
	4.0	Fertilizer Applications	65
	4.9	Total C in Rice Plant Tissue at Different Fertilizer	
	4.7	Applications	68
	4.10	δ15N in Rice Plant Tissue at Different Fertilizer	
	4.10	Applications	69
	4.11	Total N in Rice Plant Tissue at Different Fertilizer	
	7.11	Applications	72
	4.12	Rain Shelter Experiment of KADA and MADA	
	1.12	Soils	73
		50110	

5 SUI	MMARY, CONCLUSION AND	77
RE	COMMENDATIONS FOR FUTURE RESEARCH	77
5.1	Summary of the Findings	77
5.2	Recommendations	79
5.3	Limitations of the Study	80
5.4	Suggestions for Future Study	81
5.5	Conclusion	81
REFERENCES		83
APPENDICES		93
BIODATA OF S	STUDENT	121
LIST OF PUBL	ICATIONS	122

LIST OF TABLES

Table		Page
1.1	Total population estimates, by region, 1950 – 2100 (billion)	4
1.2	Distribution Map of Rice Production Areas in Peninsular Malaysia	5
2.1	Physico-Chemical Properties of the Soil used for each Paddy Field Trial	12
2.2	The comparison between K fertilizer Sources in the Form of SOP, MOP, and Polyhalite	14
3.1	Treatments in the Paddy Field and Rain Shelter Experiment	25
3.2	Experimental Layout for the Field Trials	27
3.3	Lime Requirement according to Soil pH	27
3.4	Cultivation Method and Seed Rate	29
3.5	Recommended Fertilizer Application for Rice Production	31
3.6	Water Depth According to Rice Growth Stage	32
4.1	Fertility classes for Exchangeable Cations	40
4.2	Soil Chemical Characteristics before Rice Planting in KADA Paddy Field.	41
4.3	Soil Chemical Characteristics at Harvest in KADA Paddy Field	42
4.4	Soil Chemical Characteristics before Planting of MADA Paddy Field.	43
4.5	Soil Chemical Characteristics at Harvest in MADA Paddy Field	45
4.6	Soil Chemical Characteristics before Planting of KADA Rain Shelter Experiment	46
4.7	Soil Chemical Characteristics at Harvest of KADA Rain Shelter Experiment	47
4.8	Soil Chemical Characteristics before Planting of MADA Rain Shelter Experiment	48
4.9	Chemical Characteristics at Harvest of MADA Rain Shelter Experiment	49

4.10	Yield of each Research Plot	51
4.11	Rice Yield Parameters upon Harvest at 110 DAP at KADA Paddy Field	53
4.12	Rice Yield Parameters upon Harvest at 110 DAP at MADA Paddy Field	54
4.13	Rice yield parameters upon harvest at 110 DAP at KADA Rain Shelter Experiment	55
4.14	Rice yield parameters upon harvest at 110 DAP at MADA Rain Shelter Experiment	56
4.15	Nutrient content of rice plant upon harvest at 110 DAP at KADA Paddy Field	57
4.16	Nutrient Content of Rice Plant upon Harvest at 110 DAP at MADA Paddy Field	58
4.17	Nutrient Content of Rice Plant upon Harvest at 110 DAP at KADA Rain Shelter Experiment	59
4.18	Nutrient Content of Rice Plant upon Harvest at 110 DAP at MADA Rain Shelter Experiment	60
4.19	Correlation Between Yield, Rate of Photosynthesis, Stomata Conductance, $\delta^{13}C$ and $\delta^{15}N$ derived from Pearson correlation coefficients in SAS	61
4.20	Photosynthesis Rate of Each Research Plot	63
4.21	Stomatal Conductance of Each Research Plot	64
4.22	δ ¹³ C Value of Each Research Plot	65
4.23	Total C of Each Research Plot	69
4.24	$\delta^{15}N$ Value of Each Research Plot	70
4.25	Total N of Each Research Plot	72
4.26	Rice Yield, Rate of Photosynthesis, Stomata Conductance, $\delta^{13}C$ and $\delta^{15}N$ between treatment of KADA and MADA Soils in Rain Shelter Experiment	74
4.27	Rice Yield, Rate of Photosynthesis, Stomata Conductance, $\delta^{13}C$ and $\delta^{15}N$ between Soil Types of KADA and MADA Soils in Rain Shelter Experiment	75

LIST OF FIGURES

Figure		Page
1.1	Total population estimates, by region, 1950 – 2100 (billion)	2
1.2	Distribution Map of Rice Production Areas in Peninsular Malaysia	3
2.1	General Parts of Rice showing the Panicles, Stems, Leaves and Roots	10
2.2	Rice Crop Stages	11
3.1	Soil Sampling before Planting and Land Preparations at the Research Plot	28
3.2	Soil sampling preparations before planting at the experimental plot in the rain shelter	28
3.3	Planting of rice seeds by broadcast method at the research plot	30
3.4	Planting of rice seedling by transplanting method under rain shelter	30
3.5	Rice plant at 15 DAP for second fertilizer application according to the standard fertilization programme	32
3.6	Rice plant at 55 DAP for forth fertilizer application according to the standard fertilization programme	33
3.7	Rice plants at 110 DAP in field during harvest	33
3.8	Rice plant at 15 DAP for second fertilizer application in rain shelter	33
3.9	Rice plant at 35 DAP for third fertilizer application in rain shelter	34
3.10	Rice plant at 55 DAP for forth fertilizer application in rain shelter	34
3.11	Rice plant at 75 DAP for the last fertilizer application in rain shelter	34
3.12	Rice plants at 110 DAP in rain shelter during harvest	35
3.13	A Li-6400XT (LI-COR) was used for measurements of net CO2 rate of photosynthesis and stomatal conductance	37
3.14	Isotope Ratio Mass Spectrometer (IRMS) Machine in Jabatan Kimia Malaysia	38

LIST OF ABBREVIATIONS

UPM Universiti Putra Malaysia

KRI Khazanah Research Institute

DOA Department of Agriculture

KADA Kemubu Agricultural Development Authority

MADA Muda Agricultural Development Authority

IADA Integrated Agricultural Development Area

BLS Barat Laut Selangor

K Potassium

C Carbon

N Nitrogen

P Phosphorus

Mg Magnesium

Ca Calcium

Al Aluminium

Fe Iron

CO₂ Carbon Dioxide

O₂ Oxygen

H⁺ Hydrogen

NH⁴⁺ Ammonium

NO³- Nitrate

K₂O Potassium Oxide

K₂SO₄ Potassium Sulphate

KCl Potassium Chloride

MgO Magnesium Oxide

CaO Calcium Oxide

SOP Sulphate of Potash

MOP Muriate of Potash

ROS Reactive Oxygen Species

NADPH Nicotinamide Adenine Dinucleotide Phosphate

WUE Water Use Efficiency

Days After Planting

HST Hari Selepas Tanam

Δ13C Carbon 13 Discrimination

Δ15N Nitrogen 15 Discrimination

Stomata Conductance

T1 Treatment 1

T2 Treatment 2

T3 Treatment 3

T4 Treatment 4

T5 Treatment 5

AAS Atomic Absorption Spectrometry

AES Atomic Emission Spectrometry

ICP Inductively Coupled Plasma

LI-COR Portable Photosynthesis System

IRMS Isotope Ratio Mass Spectrometry

SAS Statistical Analysis Software

CHAPTER 1

INTRODUCTION

1.1 Research Background

Rice (Oryza sativa L.), a staple crop globally, is crucial for food security and agricultural sustainability. Malaysia, a leading global importer of rice, faces the challenge of meeting the yearly domestic rice demand exceeding 1000 metric tonnes. Potassium (K) is an essential macronutrient that influences plant well-being, strength, and ability to withstand biological and environmental challenges. It plays a significant role in controlling stomatal conductance, increasing photosynthesis, osmotic adjustment, enzymatic processes, protein synthesis, and preserving ionic balance in plant cells.

Stable isotope analysis, specifically targeting $\delta^{13}C$ (carbon isotopes) and $\delta^{15}N$ (nitrogen isotopes), has become an effective methodology for understanding plant resource acquisition and interactions. This study aims to fill a gap in current knowledge by examining the impact of various potassium fertilizer sources on rice growth, yield, and isotopic ratio. The experimental design involves rice plants treated with five different treatments, including T1 (absence of K), T2 (Muriate of Potash), T3 (Sulphate of Potash), T4 (Polyhalite), and T5 (conventional fertilizer).

The experiments were conducted in the field and under a rain shelter. The MR219 were planted in Malaysia's two largest granary areas (KADA and MADA). These areas are significant in the rice industry and contribute to the country's overall production. This study also explains the selection of these regions for the field trial sites due to their strategic importance in agricultural research and their valuable insights into optimizing rice production and nutrient management practices. Additionally, including a rain shelter experiment addresses the challenges of conducting field trials and creates a controlled environment for studying the effects of potassium fertilizers on rice growth. The rain shelter provided a consistent and controlled space where important environmental factors could be managed, leading to more accurate and reliable data collection.

1.1.1 The World Rice Situation

In 2017, the United Nations estimated the worldwide population to be 7.5 billion, with Asia contributing the largest share; regionally, Asia has the world's greatest population (Figure 1.1). While rice is a staple in the majority of Asian nations, the region uses more than 80% of the global supply. It is anticipated that rice demand will increase in the future. Rice consumption is already high, and this tendency is anticipated to continue as

the population increases. Most of the world's rice production and consumption is concerted in Asia.

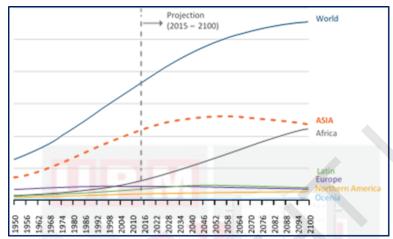


Figure 1.1: Total Population estimates by Region, the Year 1950 – 2100 (Billion) (Source: Khazanah Research Institute, 2019)

1.1.2 Malaysia Rice Production

Located in Southeast Asia, Malaysia encounters an equatorial environment distinguished by elevated temperatures (averaging from 21 to 32 °C) and humidity (with an annual precipitation of approximately 2500 mm) during the entirety of the year (Tan et al., 2020; Suhaila et al., 2012). In the significant granary areas, rice farmers typically engage in two planting cycles annually: the off-season cycle, which spans from March to July, and the main season cycle, which occurs from August to February. The primary season aligns with the northeast monsoon, leading to substantial precipitation and elevated atmospheric moisture levels. On the other hand, it has been observed that during the off-season, there is a decrease in air humidity and a reduction in rainfall (Firdaus et al., 2014).

Rice is Malaysia's third most widely planted crop, after oil palm and rubber. In 2020, 644,859 ha of its land was planted with rice. In 2021, the area was 647,859 ha, of which 525.984 ha were in Peninsular Malaysia, with the remaining in Sabah and Sarawak (Khazanah Research Institute, 2019).

Most of the irrigated rice areas in Peninsular Malaysia are in the ten designated granary areas (Figure 1.2), totaling approximately 416,415 ha. Granary areas refer to major irrigation schemes (areas greater than 4,000 ha) and are recognized by the government in the National Agricultural Policy as the main rice-producing areas. There are rice planting area of 10 granary areas in Peninsular Malaysia, namely MADA, KADA, IADA

KERIAN, IADA BLS, IADA Pulau Pinang, IADA Seberang Perak, IADA KETARA and IADA Kemasin-Semerak, IADA Pekan, and IADA Rompin since 2017 until 2021 is shown in Table 1.1.

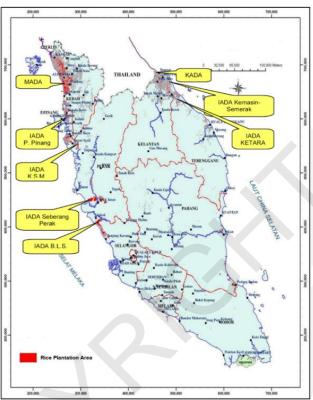


Figure 1.2: Distribution Map of Rice Production Areas in Peninsular Malaysia (Source: Khazanah Research Institute, 2019)

Domestic rice production in Malaysia relies heavily on ten important granary locations. The country produced 2.7 million metric tonnes of rice in 2017, where granary lands produced a total of 2.0 million MT or 74.1% of the total paddy yield for that year. Muda Agricultural Development Authority (MADA) in Malaysia's Northern Peninsular produced approximately 38.8% of the country's total paddy production and is known as the "Rice Bowl," followed by Kemubu Agricultural Development Authority (KADA) at 9.1% and Integrated Agricultural Development Area (IADA) Barat Laut Selangor (BLS) at 8.1% (DOA, 2022). These granary regions have varied levels of agricultural output due to differences in their locations (various environmental conditions) and soil management.

Table 1.1: The Rice Planting Area for Granary Areas in Malaysia

Tuble	1.1: The Rice Planting Area for Granary Areas in Malaysia						
	Year	2017	2018	2019	2020	2021 ^e	
No	Locations	Planted Area (ha)					
1	MADA	201,259	210,324	201,338	201,306	210,347	
2	KADA	54,067	53,710	50.348	52,164	52,589	
3	IADA, Kerian	41.898	41,898	41,898	38,578	36,994	
4	IADA Barat, LautSelangor	36,708	36,868	36,602	36,004	35,885	
5	IADA P.Pinang	25,564	25,564	25.564	24,210	24,210	
6	IADA SeberangPerak	27,735	27,735	27,334	26,296	26,296	
7	IADA KETARA	9,752	9.752	9,752	9,752	9,752	
8	IADA Kemasin, Semerak	7,129	6.902	7,564	8,129	8,401	
9	IADA Pekan	6,832	6,429	6,634	4,764	7,446	
10	IADA Rompin	5,101	5,071	5,108	5,158	5,272	

(Source: DOA, 2022)

Table 1.2 shows that the national average yield is approximately 4000 kg/ha, with yields surpassing 5000 kg/ha in regions such as IADA Pulau Pinang, IADA Ketara, and MADA. Conversely, granaries such as IADA Pekan and Batang Lupar are low-yield producers, with yields of less than 3000 kg/ha (DOA, 2022).

Table 1.2: The Average Yield of Rice for Granary Areas in Malaysia

Table 1	ole 1.2: The Average Yield of Rice for Granary Areas in Malaysia							
	Year	2017	2018	2019	2020	2021 ^e		
No	Locations	Average Yield (kg/ha)						
1	MADA	4,841	5,111	4,933	4,833	4,192		
2	KADA	4,448	4,695	4,032	4,621	4,874		
3	IADA, Kerian	4,087	3,957	3,584	3,223	3,830		
4	IADA Barat, Laut Selangor	4,510	4,731	4,756	4,431	4,337		
5	IADA P.Pinang	5,737	5,228	5,012	5,022	5,655		
6	IADA Seberang Perak	3,180	3,417	2,923	2,774	2,613		
7	IADA KETARA	5,172	5,349	5,162	5,407	5,218		
8	IADA Kemasin, Semerak	3,779	4,079	3,733	3,666	3,656		
9	IADA Pekan	1,506	2,673	2,637	2,707	1,860		
10	IADA Rompin	3,338	2,910	2,373	4,156	4,610		
11	IADA Kota Belud	2,511	3,112	2,908	2,914	3,540		
12	IADA Batang Lupar	2,009	2,492	2,754	2,599	2,847		

(Source: DOA, 2022)

1.1.3 Potassium (K) Fertilizer

Potassium (K) is an essential nutrient for plants. Plants absorb K+, which is found in soils and fertilizers like potassium chloride (KCl), potassium nitrate (KNO₃), potassium sulfate (K₂SO₄), and potassium carbonate (K₂CO₃). With a few exceptions, KCl, or muriate of potash, is the most affordable and widely utilized form for agronomic crops (Kafkafi et al., 2001). Although more expensive, other forms, such as K₂SO₄ and KNO₃, are utilized for some crops sensitive to chloride (Cl).

In addition, Jin et al. (2011) found that plants absorb more K than any other mineral element (except for nitrogen), and K is the nutrient that most frequently inhibits plant growth and crop yields. Additionally, increased K application has been demonstrated to increase photosynthetic rate, plant development, yield, and drought resistance in diverse crops (Egilla et al., 2005; Pervez et al., 2001).

Potassium (K) is the most numerous inorganic cation essential for plant growth (White and Karley, 2010). It is vital for expanding yield and improving quality (Oosterhuis et al., 2014). Potassium activates enzymes involved in, amongst other processes, protein synthesis, sugar transport, N and C metabolism, and photosynthesis. Potassium is also required for cell division, an essential aspect of plant development and function (Hepler et al., 2001).

Many researches on the impact of K on plant development have been undertaken. Through these mechanisms, potassium governs stomatal opening and shutting, cell elongation, and other vital physiological functions. Due to its high mobility, K is essential for controlling cell osmotic pressure and balancing cations and anions in the cytoplasm of plants (Hu et al., 2016). Insufficient potassium levels can impair photosynthetic CO_2 fixation and the transportation and utilization of assimilates (Hasanuzzaman et al., 2018).

Other than that, the availability of K+ from soil or fertilizers depends on soil texture, moisture, pH, and other factors (Hasanuzzaman et al., 2018). As a macronutrient, K is usually used as a soil base. In addition to the preceding considerations, plant species' K+ uptake regulates K release from soil minerals or fertilizers. Barley (*Hordeum vulgare L.*), rice (*Oryza sativa L.*), and capsicum (*Capsicum annuum L.*) contain low-affinity and high-affinity transporters identified using molecular methods (Nieves et al., 2014).

1.1.4 Natural Abundance of Isotopic Ratios

The term "natural abundance" relates to the relative ratios of various isotopes of an element that are present in the environment. The natural occurrence of isotopic abundance in plants is attributed to isotopic fractionation processes that occur during various biological and environmental interactions.

Carbon is an essential element found in plants, serving as the foundational component of organic compounds. Carbon is naturally present in two stable isotopes, namely carbon-12 (12C) and carbon-13 (13C). Plants actively uptake carbon dioxide (CO2) from the surrounding atmosphere during photosynthesis. Plants exhibit a preference for the lighter 12C isotope over the slightly heavier 13C isotope when engaging in the process of photosynthesis. The discrimination occurs due to the variances in kinetic reactions involving the two isotopes. Consequently, the carbon in plant tissues undergoes enrichment at 12C compared to atmospheric CO_2 , resulting in a decreased $\delta^{13}C$ value in plant biomass.

Nitrogen is crucial for promoting optimal plant growth as it plays a fundamental role in forming proteins, nucleic acids, and other essential molecules. Nitrogen is present in two isotopic forms, namely nitrogen-14 (14N) and the comparatively heavier nitrogen-15 (15N) isotopes. Plants exhibit a preference for incorporating the lighter 14N isotope when they uptake nitrogen from different sources. The degree of this preference may vary depending on the source of nitrogen. For example, using organic fertilizers or leguminous plants, which can fix atmospheric nitrogen, typically leads to plant tissues exhibiting lower $\delta^{15}N$ values compared to those primarily reliant on inorganic nitrogen fertilizers.

In a nutshell, the natural occurrence of isotopic ratios in plants can be attributed to the processes of isotopic fractionation that take place during photosynthesis and nitrogen uptake. This phenomenon plays a critical role in comprehending plant physiology, nutrient sources, and ecological interactions. Through analyzing δ^{13} C and δ^{15} N values in plant tissues, researchers can obtain valuable information about nutrient cycling, environmental conditions, and agricultural practices.

1.2 Problem Statement

The research problem discussed in this thesis arises from the vital role that potassium (K) plays in the growth and productivity of rice, which is a crucial crop for ensuring food security worldwide. Previous research has shown that potassium significantly impacts photosynthetic rates, plant growth, and rice production. When plants do not have enough potassium, it causes them to experience physiological stress (Wang et al., 2013). This stress affects how their stomata open and close, impacting their exchange of gases and their metabolic processes. This situation highlights the importance of investigating rice's isotopic ratio of carbon and nitrogen. By doing so, we can gain insights into how the availability of potassium affects these essential physiological processes.

In addition, temperature and precipitation are essential factors that significantly influence the variation of plant carbon and nitrogen isotopes. In 2000, Robinson and colleagues suggested that by measuring the levels of carbon-13 (13C) and nitrogen-15 (15N) isotopes in plants, we can gain valuable information about how they react to environmental stressors such as nutrient deficiency. This insight applies to rice and other crops such as barley (Ellis et al., 2002) and quinoa (Hussain et al., 2018). Isotopic analysis has been used to study how these crops respond to stressful conditions (Trandel

et al., 2018). It is interesting that much research has been conducted on the variability of isotope in plants. However, there is a significant gap in our knowledge regarding understanding how the availability of nutrients, specifically potassium, affects this variation in isotopes.

Furthermore, researchers in the past have conducted studies on carbon isotope discrimination in crops such as cotton, specifically concerning potassium deficiencies (Bednarz et al., 1998). These studies have emphasized the significance of potassium in the process of carbon metabolism. However, there is currently a lack of studies that quantitatively evaluate the degree of isotopic variation caused by potassium absorption in rice.

The main issue that this thesis aims to tackle is the necessity of studying how carbon $(\delta^{13}C)$ and nitrogen $(\delta^{15}N)$ isotopic responses, as well as the total carbon (C) and nitrogen (N) content, photosynthesis rates, stomatal conductance, and rice yield, are affected by various potassium fertilizer treatments. This research aims to bridge the existing knowledge gap by examining how varying potassium sources affect the isotopic composition of rice, providing insights into its physiological responses and ultimately contributing to the enhancement of rice production in granary areas.

1.3 Research Objectives

Generally, this study aimed to determine the effect of potassium fertilizer on stable carbon and nitrogen isotopic ratios in rice. The specific objectives of the study are as follows:

- 1. To determine the influences of different potassium fertilizer sources on the stable carbon and nitrogen isotope value in rice.
- 2. To investigate the effect of different types of potassium fertilizer on the rate of photosynthesis in rice.
- 3. To examine the effect of different types of potassium fertilizer on yield performance in rice.

1.4 Significance of the Study

First and foremost, it is important to acknowledge that rice is a crucial crop that serves as a staple food for over half of the world's population. This makes it essential to maintain food security. The study aims to understand how various K fertilizer sources affect the growth and yield of rice. This research is valuable in our efforts to improve rice production. This is especially important when the world population is growing and the need for rice is increasing.

In addition, this research holds the potential to improve the use of fertilizers, which is a crucial part of sustainable agriculture. Potassium is an essential nutrient that plants need

for their growth. By studying how different sources of potassium fertilizer affect the growth of rice, we can find ways to use fertilizers more effectively and in a more targeted manner. This has the potential to increase crop yields and decrease the negative impact on the environment caused by excessive fertilizer use, all while benefiting the financial situation of farmers.

The study examines how different K fertilizer treatments affect the carbon (δ^{13} C) and nitrogen (δ^{15} N) isotopic ratio in rice. This research could potentially lead to the development of rice varieties that are more resistant to environmental challenges such as nutrient deficiency. Ensuring that crops can thrive even in adverse conditions is crucial for maintaining agricultural sustainability.

Moreover, this study is important because it helps us understand how potassium uptake affects the variation in isotopes in rice. It contributes to our knowledge of plant physiology and how nutrients interact with each other. The knowledge acquired can be applied not only to rice but also to other types of crops and ecosystems, resulting in additional benefits.

The findings of this research are extremely valuable for farmers, agronomists, and policymakers. By analyzing the data, we can provide recommendations on which K fertilizer sources would be the best choice, ultimately resulting in better crop yields and improved food security.

1.5 Organization of the Thesis

This thesis is organized into five (5) chapters. The first (1) chapter begins with the introduction that includes the introduction of Malaysia's rice production, types of fertilizer, problem statement, objectives, and significance of the study. The second (2) chapter consists of the literature reviews of past studies and information relevant to the study. The third (3) chapter includes an explanation of the research methodology, including sampling techniques, data collection methods, study area descriptions, and statistical data analysis. The fourth (4) chapter provides results of the descriptive analysis, analysis of variance (oneway ANOVA) and in-depth discussion of the findings is also discussed in this chapter. The last chapter, chapter five (5), consists of a summary of the study, recommendations, study limitations, suggestions for further study, and the overall conclusion of the study.

REFERENCES

- Anderson, K. A., & Smith, B. W. (2006). Effect of season and variety on the differentiation of geographic growing origin of pistachios by stable isotope profiling. *Journal of Agricultural and Food Chemistry*, 54(5), 1747–1752. https://doi.org/10.1021/jf052928m
- Ashraf, M., Ahmad, A., & McNeilly, T. (2001). Growth and photosynthetic characteristics in pearl millet under water stress and different potassium supply. *Photosynthetica*, *39*(3), 389–394. https://doi.org/10.1023/a:1015182310754
- Aulakh, M. S., & Malhi, S. S. (2005). Interactions of Nitrogen with Other Nutrients and Water: Effect on Crop Yield and Quality, Nutrient Use Efficiency, Carbon Sequestration, and Environmental Pollution. *Advances in Agronomy*, 341–409. https://doi.org/10.1016/s0065-2113(05)86007-9
- Azman, E. A., Ismail, R., & Jusop, S. (2022). Effects of Various Liming Materials on the Growth of Rice under Rainshelter Condition. AgroTech Food Science, Technology and Environment, 1(2), 1–13. https://doi.org/10.53797/agrotech.v1i2.1.2022
- Bednarz, C. W., Oosterhuis, D. M., & Evans, R. D. (1998). Leaf photosynthesis and carbon isotope discrimination of cotton in response to potassium deficiency. *Environmental and Experimental Botany*, 39(2), 131–139. https://doi.org/10.1016/s0098-8472(97)00039-7
- Berahim, Z., Dorairaj, D., Omar, M. H., Saud, H. M., & Ismail, M. R. (2021). Spermine mediated improvements on stomatal features, growth, grain filling and yield of rice under differing water availability. *Scientific Reports*, 11(1). https://doi.org/10.1038/s41598-021-89812-1
- Bouman, B. A. M., & Tuong, T. P. (2001). Field water management to save water and increase its productivity in irrigated lowland rice. *Agricultural Water Management*, 49(1), 11–30. https://doi.org/10.1016/s0378-3774(00)00128-1
- Busch, F. A., Holloway-Phillips, M., Stuart-Williams, H., & Farquhar, G. D. (2020). Revisiting carbon isotope discrimination in C3 plants shows respiration rules when photosynthesis is low. *Nature Plants*, 6(3), 245–258. https://doi.org/10.1038/s41477-020-0606-6
- Caemmerer, S. V., Ghannoum, O., Pengelly, J. J. L., & Cousins, A. B. (2014). Carbon isotope discrimination as a tool to explore C4 photosynthesis. *Journal of Experimental Botany*, 65(13), 3459–3470. https://doi.org/10.1093/jxb/eru127
- Cakmak, I. (2005). The role of potassium in alleviating detrimental effects of abiotic stresses in plants. *Journal of Plant Nutrition and Soil Science*, *168*(4), 521–530. https://doi.org/10.1002/jpln.200420485

- Cakmak. I. (2000). Tansley Review no. 111. *New Phytologist*, *146*(2), 185–205. https://doi.org/10.1046/j.1469-8137.2000.00630.x
- Cecchetti, A. R., Sytsma, A., Stiegler, A. N., Dawson, T. E., & Sedlak, D. L. (2020). Use of stable nitrogen isotopes to track plant uptake of nitrogen in a nature-based treatment system. *Water Research X*, 9, 100070. https://doi.org/10.1016/j.wroa.2020.100070
- Chalk, P. M., Inácio, C. T., & Chen, D. (2019). An overview of contemporary advances in the usage of 15N natural abundance (δ¹⁵N) as a tracer of agro-ecosystem N cycle processes that impact the environment. Agriculture, *Ecosystems & Environment*, 283, 106570. https://doi.org/10.1016/j.agee.2019.106570
- Comstock, J. (2001). Steady-state isotopic fractionation in branched pathways using plant uptake of NO3 as an example. *Planta*, 214(2), 220–234. https://doi.org/10.1007/s004250100602
- Condon, A. G., Richards, R. A., Rebetzke, G. J., & Farquhar, G. D. (2002). Improving intrinsic water-use efficiency and crop yield. *Crop Science*, 42(1), 122–131. https://doi.org/10.2135/cropsci2002.1220
- Coskun, D., Britto, D. T., & Kronzucker, H. J. (2016). The nitrogen-potassium intersection: Membranes, metabolism, and mechanism. *Plant, Cell & Environment*, 40(10), 2029–2041. https://doi.org/10.1111/pce.12671
- Craig, H. (1953). The geochemistry of the stable carbon isotopes. *Geochimica Et Cosmochimica Acta*, 3(2-3), 53–92. https://doi.org/10.1016/0016-7037(53)90001-5
- Craig, H. (1954). Carbon 13 in plants and the relationships between carbon 13 and carbon 14 variations in nature. *The Journal of Geology*, 62(2), 115–149. https://doi.org/10.1086/626141
- Craine, J. M., Elmore, A. J., Aidar, M. P., Bustamante, M., Dawson, T. E., Hobbie, E. A., Kahmen, A., Mack, M. C., McLauchlan, K. K., Michelsen, A., Nardoto, G. B., Pardo, L. H., Peñuelas, J., Reich, P. B., Schuur, E. A., Stock, W. D., Templer, P. H., Virginia, R. A., Welker, J. M., & Wright, I. J. (2009). Global patterns of foliar nitrogen isotopes and their relationships with climate, mycorrhizal fungi, foliar nutrient concentrations, and nitrogen availability. *New Phytologist*, *183*(4), 980–992. https://doi.org/10.1111/j.1469-8137.2009.02917.x
- Dash, A. K., Singh, H. K., Mahakud, T., Pradhan, K. C., & Jena, D. (2015). Interaction effect of nitrogen, phosphorus, potassium with sulphur, boron and zinc on yield and nutrient uptake by rice under rice-rice cropping system in inceptisol of coastal Odisha. *International Research Journal of Agricultural Science and Soil Science*, 5(1), 14-21
- Dat, J., Vandenabeele, S., Vranová, E., Van Montagu, M., Inzé*, D., & Van Breusegem, F. (2000). Dual action of the active oxygen species during plant stress responses. Cellular and Molecular Life Sciences (CMLS), 57(5), 779–795. https://doi.org/10.1007/s000180050041

- Denton, T. M., Schmidt, S., Critchley, C., & Stewart, G. R. (2001). Natural abundance of stable carbon and nitrogen isotopes in cannabis sativa reflects growth conditions. *Functional Plant Biology*, 28(10), 1005. https://doi.org/10.1071/pp01066
- Diao, H., Wang, A., Yuan, F., Guan, D., Dai, G., & Wu, J. (2020). Environmental effects on carbon isotope discrimination from assimilation to respiration in a coniferous and broad-leaved mixed forest of Northeast China. *Forests*, 11(11), 1156. https://doi.org/10.3390/f11111156
- Ding, Y., Luo, W., & Xu, G. (2006). Characterisation of magnesium nutrition and interaction of magnesium and potassium in rice. *Annals of Applied Biology*, 149(2), 111–123. https://doi.org/10.1111/j.1744-7348.2006.00080.x
- DOA. (2022). RICE CHECK: PADI. Retrieved October 11, 2023, from https://www.doa.gov.my/doa/resources/aktiviti_sumber/sumber_awam/penerbit_an/pakej_teknologi/padi/rice_check_padi_2022.pdf
- Dobermann, A., Witt, C., Dawe, D., Abdulrachman, S., Gines, H., Nagarajan, R., Satawathananont, S., Son, T., Tan, P., Wang, G., Chien, N., Thoa, V., Phung, C., Stalin, P., Muthukrishnan, P., Ravi, V., Babu, M., Chatuporn, S., Sookthongsa, J., Adviento, M. (2002). Site-specific nutrient management for intensive rice cropping systems in Asia. *Field Crops Research*, 74(1), 37–66. https://doi.org/10.1016/s0378-4290(01)00197-6
- Egilla, J. N., Davies, F. T., & Boutton, T. W. (2005). Drought stress influences leaf water content, photosynthesis, and water-use efficiency of Hibiscus rosa-sinensis at three potassium concentrations. *Photosynthetica*, 43(1), 135–140. https://doi.org/10.1007/s11099-005-5140-2
- Ellis, R. P., Forster, B. P., Gordon, D. C., Handley, L. L., Keith, R. P., Lawrence, P., Meyer, R., Powell, W., Robinson, D., Scrimgeour, C. M., Young, G., & Thomas, W. T. B. (2002). Phenotype/genotype associations for yield and salt tolerance in a barley mapping population segregating for two dwarfing genes. *Journal of Experimental Botany*, 53(371), 1163–1176. https://doi.org/10.1093/jexbot/53.371.1163
- Engels, C., & Kirkby, E. A. (2001). Cycling of nitrogen and potassium between shoot and roots in maize as affected by shoot and root growth. *Journal of Plant Nutrition and Soil Science*, 164(2), 183–191. <a href="https://doi.org/10.1002/1522-2624(200104)164:2<183::aid-jpln183>3.0.co;2-b">https://doi.org/10.1002/1522-2624(200104)164:2<183::aid-jpln183>3.0.co;2-b
- Evans, R. D. (2001). Physiological mechanisms influencing plant nitrogen isotope composition. *Trends in Plant Science*, 6(3), 121–126. https://doi.org/10.1016/s1360-1385(01)01889-1
- Farooq, M., Wahid, A., Kobayashi, N., Fujita, D., & Basra, S. M. (2009). Plant drought stress: Effects, mechanisms and management. *Agronomy for Sustainable Development*, 29(1), 185–212. https://doi.org/10.1051/agro:2008021

- Farquhar, G. D., Ehleringer, J. R., & Hubick, K. T. (1989). Carbon isotope discrimination and photosynthesis. *Annual Review of Plant Physiology and Plant Molecular Biology*, 40(1), 503–537. https://doi.org/10.1146/annurev.pp.40.060189.002443
- Farquhar, G. D., O'Leary, M. H., & Berry, J. A. (1982). On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Functional Plant Biology*, *9*(2), 121–137. https://doi.org/10.1071/pp9820121
- Gibson, S. I. (2005). Control of plant development and gene expression by sugar signaling. *Current Opinion in Plant Biology*, 8(1), 93–102. https://doi.org/10.1016/j.pbi.2004.11.003
- Handley, L. L., & Raven, J. A. (1992). The use of natural abundance of nitrogen isotopes in plant physiology and ecology. *Plant, Cell and Environment*, *15*(9), 965–985. https://doi.org/10.1111/j.1365-3040.1992.tb01650.x
- Hasanuzzaman, M., Bhuyan, M., Nahar, K., Hossain, M., Mahmud, J., Hossen, M., Masud, A., Moumita, & Fujita, M. (2018). Potassium: A Vital Regulator of Plant Responses and Tolerance to Abiotic Stresses. *Agronomy*, 8(3), 31. https://doi.org/10.3390/agronomy8030031
- Hepler, P. K., Vidali, L., & Cheung, A. Y. (2001). Polarized cell growth in higher plants. Annual Review of Cell and Developmental Biology, 17(1), 159–187. https://doi.org/10.1146/annurev.cellbio.17.1.159
- Holthusen, D., Peth, S., & Horn, R. (2010). Impact of potassium concentration and matric potential on soil stability derived from rheological parameters. *Soil and Tillage Research*, 111(1), 75–85. https://doi.org/10.1016/j.still.2010.08.002
- Hou, E., Chen, C., Wen, D., Kuang, Y., & Sun, F. (2015). Plant and soil Δ13C and Δ15N are linked to community biomass, litter production, and litter turnover rate in mature subtropical forests. *Plant Ecology*, 216(6), 859–872. https://doi.org/10.1007/s11258-015-0473-9
- Hou, W., Yan, J., Jákli, B., Lu, J., Ren, T., Cong, R., & Li, X. (2018). Synergistic Effects of Nitrogen and Potassium on Quantitative Limitations to Photosynthesis in Rice (Oryza sativa L.). *Journal of Agricultural and Food Chemistry*, 66(20), 5125–5132. https://doi.org/10.1021/acs.jafc.8b01135
- Hsing, Y. I. C. (2016). Rice in Taiwan. Encyclopaedia of the History of Science, Technology, and Medicine in Non-Western Cultures, 3769–3772. https://doi.org/10.1007/978-94-007-7747-7_10245
- Hu, W., Coomer, T. D., Loka, D. A., Oosterhuis, D. M., & Zhou, Z. (2017). Potassium deficiency affects the carbon-nitrogen balance in cotton leaves. *Plant Physiology and Biochemistry*, 115, 408–417. https://doi.org/10.1016/j.plaphy.2017.04.005
- Hu, W., Jiang, N., Yang, J., Meng, Y., Wang, Y., Chen, B., Zhao, W., Oosterhuis, D. M., & Zhou, Z. (2016). Potassium (K) supply affects K accumulation and photosynthetic physiology in two cotton (Gossypium hirsutum L.) cultivars with

- different K sensitivities. *Field Crops Research*, 196, 51–63. https://doi.org/10.1016/j.fcr.2016.06.005
- Hu, W., Yang, J., Meng, Y., Wang, Y., Chen, B., Zhao, W., Oosterhuis, D. M., & Zhou, Z. (2015). Potassium application affects carbohydrate metabolism in the leaf subtending gossypiumn (gossypium hirsutum L.) boll and its relationship with boll biomass. Field Crops Research, 179, 120–131. https://doi.org/10.1016/j.fcr.2015.04.017
- Huang, C. L., & Schulte, E. E. (1985). Digestion of plant tissue for analysis by ICP emission spectroscopy. *Communications in Soil Science and Plant Analysis*, 16(9), 943–958. https://doi.org/10.1080/00103628509367657
- Hussain, M. I., Al- Dakheel, A. J., & Reigosa, M. J. (2018). Genotypic differences in agro-physiological, biochemical and isotopic responses to salinity stress in quinoa (Chenopodium Quinoa Willd.) plants: Prospects for salinity tolerance and yield stability. *Plant Physiology and Biochemistry*, 129, 411–420. https://doi.org/10.1016/j.plaphy.2018.06.023
- Igamberdiev, A. U., Mikkelsen, T. N., Ambus, P., Bauwe, H., Lea, P. J., & Gardeström, P. (2004). Photorespiration Contributes to Stomatal Regulation and Carbon Isotope Fractionation: A Study with Barley, Potato and Arabidopsis Plants Deficient in Glycine Decarboxylase. *Photosynthesis Research*, 81(2), 139–152. https://doi.org/10.1023/b:pres.0000035026.05237.ec
- Jamaludin, A. F. (2016). Alleviating soil acidity, aluminium and iron toxicity in an acid sulfate soil using lime and bio-organic fertilizer to increase rice yield. http://psasir.upm.edu.my/id/eprint/67513/
- Jang, S., Lee, Y., Lee, G., Seo, J., Lee, D., Yu, Y., Chin, J. H., & Koh, H. J. (2018). Association between sequence variants in panicle development genes and the number of spikelets per panicle in rice. *BMC Genetics*, 19(1). https://doi.org/10.1186/s12863-017-0591-6
- Kalcsits, L. A., Buschhaus, H. A., & Guy, R. D. (2014). Nitrogen isotope discrimination as an integrated measure of nitrogen fluxes, assimilation and allocation in plants. *Physiologia Plantarum*, *151*(3), 293–304. https://doi.org/10.1111/ppl.12167
- Ketterings, Q. M., Reid, S., & Rao, R. (2007). Agronomy Fact Sheet Series, Fact Sheet 22, Cornell University Cooperative Extension. (Cornell University, Ithaca.) http://nmsp.cals.cornell.edu/guidelines/factsheets.html
- Khan, S. A., Mulvaney, R. L., & Ellsworth, T. R. (2013). The potassium paradox: Implications for soil fertility, crop production and human health. *Renewable Agriculture and Food Systems*, 29(1), 3–27. https://doi.org/10.1017/s1742170513000318
- Khazanah Research Institute. (2019). The Status of the Paddy and Rice Industry in Malaysia. Kuala Lumpur: Khazanah Research Institute. https://www.krinstitute.org/assets/contentMS/img/template/editor/20190409 Rice Report_Full% 20Report_Final.pdf

- Kohn, M. (2016). Carbon isotope discrimination in C3 land plants is independent of natural variations in pCO₂. *Geochemical Perspectives Letters*, 35–43. https://doi.org/10.7185/geochemlet.1604
- Kumar, A., Sen, A., Upadhyay, P. K., & Singh, R. K. (2017). Effect of Zinc, Iron and Manganese Levels on Quality, Micro and Macro Nutrients Content of Rice and Their Relationship with Yield. *Communications in Soil Science and Plant Analysis*, 48(13), 1539–1551. https://doi.org/10.1080/00103624.2017.1373799
- Kundu, C. A., Ishii, M., Sato, K., Masunaga, T., Wanjogu, R. K., Njagi, R. E., Yamauchi, A., & Makihara, D. (2016). Evaluation of Soil Chemical Properties under Paddy Production System in Central Kenya: Soil Exchangeable Cations. *Journal of Agricultural Science*, 8(8), 136. https://doi.org/10.5539/jas.v8n8p136
- Kurdali, F., & Al-Shamma'a, M. (2010). Natural abundances of 15nitrogen and 13carbon indicative of growth and n2 fixation in potassium fed lentil grown under water stress. *Journal of Plant Nutrition*, 33(2), 157–174. https://doi.org/10.1080/01904160903434238
- Kusumi, K., Hirotsuka, S., Kumamaru, T., & Iba, K. (2012). Increased leaf photosynthesis caused by elevated stomatal conductance in a rice mutant deficient in slac1, a guard cell anion channel protein. *Journal of Experimental Botany*, 63(15), 5635–5644. https://doi.org/10.1093/jxb/ers216
- Lacombe, R. J. S., & Bazinet, R. P. (2020). Natural abundance carbon isotope ratio analysis and its application in the study of diet and metabolism. *Nutrition Reviews*, 79(8), 869–888. https://doi.org/10.1093/nutrit/nuaa109
- Lewis, T. D., Hallett, P. D., Paton, G. I., & Harrold, L. (2019). Retention and release of nutrients from polyhalite to soil. *Soil Use and Management*, 36(1), 117–122. https://doi.org/10.1111/sum.12548
- Liang, Y., Hu, F., Yang, M., & Yu, J. (2003). Antioxidative defenses and water deficit-induced oxidative damage in rice (Oryza sativa L.) growing on non-flooded paddy soils with ground mulching. *Plant and Soil*, 257(2), 407–416. https://doi.org/10.1023/a:1027313902195
- Liu, S. H., Chen, G. X., Yin, J. J., & Lu, C. G. (2011). Response of the flag leaves of a super-hybrid rice variety to drought stress during grain filling period. *Journal of Agronomy and Crop Science*, 197(4), 322–328. https://doi.org/10.1111/j.1439-037x.2010.00458.x
- Ma, S., He, F., Tian, D., Zou, D., Yan, Z., Yang, Y., Zhou, T., Huang, K., Shen, H., & Fang, J. (2018). Variations and determinants of carbon content in plants: a global synthesis. *Biogeosciences*, *15*(3), 693–702. https://doi.org/10.5194/bg-15-693-2018
- Malvi, U. R. (2011). Interaction of micronutrients with major nutrients with special reference to potassium. *Karnataka Journal of Agricultural Sciences*, 24, 106-109.

- Martin, E., & Hine, R. (2015). *A Dictionary of Biology*. Oxford University Press, USA, page 320. http://books.google.ie/books?id=gMf9CAAAQBAJ&printsec=frontcover&dq=9 78-0-19-871437-8&hl=&cd=1&source=gbs_api
- Mohd Zain, N. A., & Ismail, M. R. (2016). Effects of potassium rates and types on growth, leaf gas exchange and biochemical changes in rice (Oryza sativa) planted under cyclic water stress. *Agricultural Water Management*, *164*, 83–90. https://doi.org/10.1016/j.agwat.2015.09.022
- Morris, K. A., Saetre, P., Norton, U., & Stark, J. M. (2022). Plant community effects on soil moisture and nitrogen cycling in a semi-arid ecosystem. *Biogeochemistry*, 159(2), 215–232. https://doi.org/10.1007/s10533-022-00922-y
- Musa, A. (2020). Crop Yield Estimation (Biological yield and economic yield). Basic Agricultural Study. https://agriculturistmusa.com/crop-yield-estimation/
- Nieves-Cordones, M., Alemán, F., Martínez, V., & Rubio, F. (2014). K+ uptake in plant roots. The systems involved, their regulation and parallels in other organisms. Journal of Plant Physiology, 171(9), 688–695. https://doi.org/10.1016/j.jplph.2013.09.021
- Nuraini, S. M. A., Adibah, M. A., & Sulaiman, M. F. (2022). Influence of Potassium Fertilizers on Stable Carbon Isotopic Ratio in Rice. *Malaysian Journal of Soil Science*, 26.
- O'Leary, M. H. (1988). Carbon isotopes in photosynthesis. *BioScience*, 38(5), 328–336. https://doi.org/10.2307/'310735
- O'Leary, M. H. (1993). Biochemical basis of carbon isotope fractionation. Stable Isotopes and Plant Carbon-Water Relations, 19–28. https://doi.org/10.1016/b978-0-08-091801-3.50009-x
- Oosterhuis, D. M., Loka, D. A., Kawakami, E. M., & Pettigrew, W. T. (2014). The physiology of potassium in crop production. *Advances in Agronomy*, *126*, 203–233. https://doi.org/10.1016/b978-0-12-800132-5.00003-1
- Pervez, H., Ashraf, M., & Makhdum, M. I. (2004). Influence of potassium nutrition on gas exchange characteristics and water relationsossypiumn (gossypium hirsutum L.). *Photosynthetica*, 42(2), 251–255. https://doi.org/10.1023/b:phot.0000040597.62743.5b
- Policies and Economic Development of Rice Production in Malaysia. (2015, March 17).

 Policies and Economic Development of Rice Production in Malaysia | FFTC Agricultural Policy Platform (FFTC-AP). https://ap.fftc.org.tw/article/826
- Robinson, D. (2001). Δ15N as an integrator of the nitrogen cycle. *Trends in Ecology & Evolution*, 16(3), 153–162. https://doi.org/10.1016/s0169-5347(00)02098-x
- Robinson, D., Handley, L. L., Scrimgeour, C. M., Gordon, D. C., Forster, B. P., & Ellis, R. P. (2000). Using stable isotope natural abundances (Δ15N and Δ13C) to

- integrate the stress responses of wild barley (Hordeum spontaneum C. Koch.) Genotypes. *Journal of Experimental Botany*, 51(342), 41–50. https://doi.org/10.1093/jexbot/51.342.41
- Ruiz, J. M., & Romero, L. (2002). Relationship between potassium fertilization and nitrate assimilation in leaves and fruits of cucumber (Cucumis sativus) plants. Annals of Applied Biology, 140(3), 241–245. https://doi.org/10.1111/j.1744-7348.2002.tb00177.x
- Römheld, V., & Kirkby, E. A. (2010). Research on potassium in agriculture: Needs and prospects. *Plant and Soil*, 335(1-2), 155–180. https://doi.org/10.1007/s11104-010-0520-1
- Salehi-Lisar, S. Y., & Bakhshayeshan-Agdam, H. (2016). Drought stress in plants: Causes, consequences, and tolerance. *Drought Stress Tolerance in Plants*, *Vol 1*, 1–16. https://doi.org/10.1007/978-3-319-28899-4_1
- Schleuss, P. M., Widdig, M., Heintz-Buschart, A., Kirkman, K., & Spohn, M. (2020, March 17). Interactions of nitrogen and phosphorus cycling promote P acquisition and explain synergistic plant-growth responses. *Ecology*, 101(5). https://doi.org/10.1002/ecy.3003
- Shrestha, J., Kandel, M., Subedi, S., & Shah, K. K. (2020). Role of nutrients in rice (Oryza sativa L.): A review. *Agrica*, 9(1), 53. https://doi.org/10.5958/2394-448x.2020.00008.5
- Sirius Minerals, 2019 POLY4 brochure. Version March 2019. http://www.siriusminerals.com
- Sohrabi, M., Rafii, M. Y., Hanafi, M. M., Siti Nor Akmar, A., & Latif, M. A. (2012). Genetic diversity of Upland Rice Germplasm in Malaysia based on quantitative traits. *The Scientific World Journal*, 2012, 1–9. https://doi.org/10.1100/2012/416291
- Swap, R. J., Aranibar, J. N., Dowty, P. R., Gilhooly, W. P., & Macko, S. A. (2003). Natural abundance of 13C and 15N in C3 and C4 vegetation of southern Africa: patterns and implications. *Global Change Biology*, *10*(3), 350–358. https://doi.org/10.1111/j.1365-2486.2003.00702.x
- Tan, H., Cui, Y., Liu, C., Zeng, F., Han, C., Zhang, H., Fan, X., Mitchell, R., Yan, D., & Zhang, D. (2022). Toward the replacement of conventional fertilizer with polyhalite in eastern China to improve peanut growth and soil quality. *Chemical and Biological Technologies in Agriculture*, 9(1). https://doi.org/10.1186/s40538-022-00363-7
- Tan, Z. X., Lal, R., & Wiebe, K. D. (2005). Global Soil Nutrient Depletion and Yield Reduction. *Journal of Sustainable Agriculture*, 26(1), 123–146. https://doi.org/10.1300/j064v26n01_10
- Taub, D. R., & Wang, X. (2008). Why are nitrogen concentrations in plant tissues lower under elevated CO₂? A critical examination of the hypotheses. *Journal of*

- Integrative Plant Biology, 50(11), 1365–1374. https://doi.org/10.1111/j.1744-7909.2008.00754.x
- Tränkner, M., Tavakol, E., & Jákli, B. (2018). Functioning of potassium and magnesium in photosynthesis, photosynthate translocation and photoprotection. *Physiologia Plantarum*, *163*(3), 414–431. https://doi.org/10.1111/ppl.12747
- Trandel, M. A., Vigardt, A., Walters, S. A., Lefticariu, M., & Kinsel, M. (2018). Nitrogen isotope composition, nitrogen amount, and fruit yield of tomato plants affected by the soil–fertilizer types. *ACS Omega*, *3*(6), 6419–6426. https://doi.org/10.1021/acsomega.8b00296
- Ubierna, N., Holloway-Phillips, M. M., & Farquhar, G. D. (2018). Using Stable Carbon Isotopes to Study C3 and C4 Photosynthesis: Models and Calculations. *Methods in Molecular Biology*, 155–196. https://doi.org/10.1007/978-1-4939-7786-4_10
- Unkovich, M. (2013). Isotope discrimination provides new insight into biological nitrogen fixation. *New Phytologist*, 198(3), 643–646. https://doi.org/10.1111/nph.12227
- Vale, F., & Sério, D. R. (2017). Introducing polyhalite to Brazil: first steps of a new fertilizer. International Potash Institute (IPI) *e-ifc*, 48, 3-11
- Van Aken, O., Giraud, E., Clifton, R., & Whelan, J. (2009). Alternative oxidase: A target and regulator of stress responses. *Physiologia Plantarum*, 137(4), 354–361. https://doi.org/10.1111/j.1399-3054.2009.01240.x
- Wang, M., Zheng, Q., Shen, Q., & Guo, S. (2013). The critical role of potassium in plant stress response. *International Journal of Molecular Sciences*, 14(4), 7370–7390. https://doi.org/10.3390/ijms14047370
- Wang, N., Hua, H., Egrinya Eneji, A., Li, Z., Duan, L., & Tian, X. (2012). Genotypic variations in photosynthetic and physiological adjustment to potassium deficiency in cotton (Gossypium hirsutum). *Journal of Photochemistry and Photobiology B: Biology*, 110, 1–8. https://doi.org/10.1016/j.jphotobiol.2012.02.002
- Waraich, E., Ahmad, R., Halim, A., & Aziz, T. (2012). Alleviation of temperature stress by nutrient management in crop plants: a review. *Journal of Soil Science and Plant Nutrition*, 12(2), 221–244. https://doi.org/10.4067/s0718-95162012000200003
- White, P. J., & Karley, A. J. (2010). Potassium. *Plant Cell Monographs*, 199–224. https://doi.org/10.1007/978-3-642-10613-2_9
- Watzka, M., Buchgraber, K., & Wanek, W. (2006, July). Natural 15N abundance of plants and soils under different management practices in a montane grassland. *Soil Biology and Biochemistry*, 38(7), 1564–1576. https://doi.org/10.1016/j.soilbio.2005.11.007
- Xu, X., Du, X., Wang, F., Sha, J., Chen, Q., Tian, G., Zhu, Z., Ge, S., & Jiang, Y. (2020). Effects of Potassium Levels on Plant Growth, Accumulation and Distribution of

- Carbon, and Nitrate Metabolism in Apple Dwarf Rootstock Seedlings. *Frontiers in Plant Science*, 11. https://doi.org/10.3389/fpls.2020.00904
- Kafkafi, U., Xu, G., Imas, P., Magen, H., Tarchitzky, J., & Johnston, A. E. (2001). Potassium and chloride in crops and soils: the role of potassium chloride fertilizer in crop nutrition (pp. 101-103). Horgen, Switzerlands: International Potash Institute.
- Yin, X., & Struik, P. C. (2017). Can increased leaf photosynthesis be converted into higher crop mass production? A simulation study for rice using the Crop Model Gecros. *Journal of Experimental Botany*, 68(9), 2345–2360. https://doi.org/10.1093/jxb/erx085
- Yousfi, S., Serret, M. D., Márquez, A. J., Voltas, J., & Araus, J. L. (2012). Combined use of δ 13 C, δ 18 O and δ 15 n tracks nitrogen metabolism and genotypic adaptation of durum wheat to salinity and water deficit. *New Phytologist*, 194(1), 230–244. https://doi.org/10.1111/j.1469-8137.2011.04036.x
- Yuzugullu, O., Marelli, S., Erten, E., Sudret, B., & Hajnsek, I. (2017). Determining rice growth stage with X-band sar: A Metamodel based inversion. *Remote Sensing*, 9(5), 460. https://doi.org/10.3390/rs9050460
- Zain, N. A. B. M. (2015). Agro-Morphological, Physiological and Biochemical Characteristics of Rice Subjected to Cyclic Water Stress and Potassium Fertilizers.
- Zhang, H. Y., Hartmann, H., Gleixner, G., Thoma, M., & Schwab, V. F. (2019). Carbon isotope fractionation including photosynthetic and post-photosynthetic processes in C3 plants: Low [CO₂] matters. Geochimica Et Cosmochimica Acta, 245, 1–15. https://doi.org/10.1016/j.gca.2018.09.035
- Zhao, D., Oosterhuis, D., & Bednarz, C. (2001). Influence of Potassium Deficiency on Photosynthesis, Chlorophyll Content, and Chloroplast Ultrastructure of Cotton Plants. *Photosynthetica*, 39(1), 103–109. https://doi.org/10.1023/a:1012404204910
- Zheng, X., Chen, B., Lu, G., & Han, B. (2009). Overexpression of a NAC transcription factor enhances rice drought and salt tolerance. *Biochemical and Biophysical Research Communications*, 379(4), 985–989. https://doi.org/10.1016/j.bbrc.2008.12.163