



**BROODSTOCK MANAGEMENT, REPRODUCTIVE DEVELOPMENT AND
LARVAL REARING OF THE MARINE ORNAMENTAL SHRIMP,
Lysmata amboinensis (de Man, 1888)**

By

WAN NUR ATIKAH BINTI OMAR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
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Lysmata amboinensis, widely known as "cleaner shrimp," is a favored species in the marine aquarium trade due to its vivid coloration and robust nature. Despite its popularity, attempts to cultivate this species on a large scale have been challenging, primarily due to its extended metamorphosis phase, which can last between 58 and 140 days. This study evaluate the effects of different environmental conditions, specifically salinity and temperature, on major reproductive parameters under controlled laboratory conditions. The study tested the impact of varying salinity levels (30 ppt, 35 ppt, and 40 ppt) on *L. amboinensis* broodstock. The results indicated that while survival rates remained stable across tested salinity, fecundity and egg volume were significantly affected. Broodstock reared at 30 ppt demonstrated the highest values for fecundity, egg volume, and other reproductive parameters ($p < 0.05$). For temperature, the broodstock were divided into four groups (22 °C, 25 °C, 28 °C, and 31 °C). The shrimp showed a wide temperature tolerance range, surviving from 22 °C to 28 °C; however, mortality began at 30 °C, and no shrimp survived beyond 24 hours at 31 °C. While the broodstock could endure temperatures as low as 22 °C, they failed to produce fertile eggs. The optimal temperature for reproduction was 25 °C, yielding nearly twice the fecundity (1420 ± 193) compared to 28 °C (851 ± 60) and producing eggs with greater size ($0.0613 \pm 0.0008 \text{ mm}^3$) and higher biomass ($0.2376 \pm 0.0395 \text{ g}$). The effects of different feeds on broodstock reproduction were also assessed, using four diets: squid (*Loligo* sp.), mussel (*Perna viridis*), polychaetes (*Marphysa moribidii*), and a combination of these. The squid diet led to substantial egg loss throughout the incubation period ($58.67\% \pm 3.67\%$) and lower larval production (97 ± 29) ($p < 0.05$). Conversely, the mussel diet produced the highest larval count (568 ± 23).

The study also explored the life cycle stages of *L. amboinensis*, identifying seven phases of complete embryonic development over approximately 13 days. Histological analysis showed that the gastrula stage, where organ formation begins, was crucial for embryonic

development. Under laboratory conditions, the shrimp achieved ten zoea stages over 55 days, highlighting the lengthy larval development phase. The study further investigated the effect of different diets on larval by feeding larvae six different treatments, including microalgae (*Chaetoceros* sp., *Tetraselmis* sp.), a commercial diet, and combinations thereof. Starved larvae and those fed solely with microalgae or the commercial diet showed poor survival rates, with the highest mortality recorded in starved larvae. The combination diet of *Tetraselmis* sp. and the commercial feed (TSAD) resulted in a 10% higher survival rate ($46.67\% \pm 5.14\%$) demonstrating its superior performance in supporting larval growth. However, even the TSAD diet was insufficient to promote complete metamorphosis to post-larvae, suggesting that current feeding regimens still require optimization. Research showed that the productivity under captive condition can be enhanced by improving the *L. amboinensis* broodstock rearing condition and feeding and understanding the biology of the early life of this species.

Keywords: *Lysmata amboinensis*, Larval development, Reproductive performance

SDG: GOAL 13: Climate action, GOAL 14: Life below water

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGURUSAN INDUK, PERKEMBANGAN PEMBIAKAN DAN PENJAGAAN
LARVA BAGI UDANG HIASAN MARIN,
Lysmata amboinensis (de Man, 1888)**

Oleh

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Lysmata amboinensis yang dikenali sebagai “udang pencuci” merupakan salah satu krustacea yang paling popular dalam industri akuarium marin kerana warna yang menarik dan lasak. Pelbagai percubaan dilakukan untuk membangunkan teknologi perternakan berskala besar, tetapi kejayaannya masih rendah disebabkan fasa metamorfosis yang sangat lama iaitu sekitar 58 hingga 140 hari. Kajian ini menilai kesan keadaan persekitaran yang berbeza (saliniti dan suhu) pada parameter utama pembiakan *L. amboinensis* di dalam keadaan makmal. Untuk eksperimen saliniti, pasangan induk dibahagikan kepada tiga kumpulan (30 ppt, 35 ppt dan 40 ppt). Keputusan menunjukkan bahawa julat saliniti yang besar ini tidak menjelaskan kelangsungan hidup induk. Walau bagaimanapun, saliniti mempengaruhi nilai fekunditi, isipadu telur dan parameter-parameter lain yang diukur di mana induk yang diternak pada saliniti 30 ppt dapat menghasilkan fekunditi dan isipadu telur yang tinggi ($p < 0.05$). Untuk eksperimen suhu, pasangan induk dibahagikan kepada empat kumpulan (22 °C, 25 °C, 28 °C dan 31 °C). Kelangsungan hidup induk bervariasi pada suhu yang berbeza di mana spesies ini mampu bertahan pada suhu serendah 22 °C dan sehingga suhu 28 °C. Kematian bermula apabila suhu mencapai 30 °C. Tiada induk yang hidup selepas 24 jam pada 31°C. Walaupun spesies ini mampu bertahan pada 22 °C, induk tersebut tidak dapat menghasilkan telur yang bersenyawa. Induk di bawah rawatan 25 °C menghasilkan fekunditi (1420 ± 193) iaitu hampir dua kali ganda lebih tinggi berbanding rawatan 28 °C. Telur yang dihasilkan juga lebih besar isipadu dan tinggi nilai biomasnya. Selain itu, kesan makanan yang berlainan terhadap induk *L. amboinensis* juga dijalankan. Empat jenis diet yang terdiri daripada sotong (*Loligo* sp.), kupang (*Perna viridis*), umpsun-umpsun (*Marphysa moribidii*) dan makanan kombinasi diberi kepada induk. Hasil kajian mendapati induk yang diberi diet sotong telah kehilangan sebahagian besar daripada telur yang dihasilkan sepanjang melalui tempoh inkubasi ($58.67\% \pm 3.67\%$), dan diikuti dengan pengeluaran larva yang rendah (97 ± 29) ($p < 0.05$). Sebaliknya, diet kupang menghasilkan jumlah larva tertinggi (568 ± 23).

Kajian ini juga meneroka tahap-tahap kitaran hidup *L. amboinensis* dan mengenal pasti tujuh fasa perkembangan embrio yang lengkap dalam tempoh 13 hari. Analisis histologi menunjukkan bahawa fasa gastrula penting untuk perkembangan embrio kerana pembentukan organ bermula. Dalam kondisi makmal, udang ini mencapai sepuluh tahap zoea dalam tempoh 55 hari. Kajian ini juga menyiasat kesan diet yang berbeza pada larva. Larva diberi enam rawatan diet berbeza, termasuk mikroalga (*Chaetoceros* sp., *Tetraselmis* sp.), diet komersial, dan gabungan diet ini. Larva yang tidak diberi sebarang diet serta larva yang makan samaada mikroalga atau diet komersial secara eksklusif menunjukkan kadar kelangsungan hidup yang rendah. Gabungan diet *Tetraselmis* sp. dan makanan komersial (TSAD) menghasilkan kadar kelangsungan hidup yang 10% lebih tinggi (46.67% ± 5.14%), menunjukkan prestasi yang lebih baik dalam menyokong pertumbuhan larva. Walau bagaimanapun, diet TSAD masih tidak mencukupi bagi menampung pertumbuhan larva untuk bermetamorfosis hingga ke peringkat post larva. Penyelidikan ini menunjukkan bahawa peningkatan produktiviti bagi *L. amboinensis* di bawah keadaan makmal boleh dipertingkatkan melalui penambahbaikan kondisi penjagaan induk dan pemakanan disamping memahami biologi kehidupan awal spesies ini.

Kata Kunci: *Lysmata amboinensis*, Perkembangan larva, Prestasi pembiakan

SDG: MATLAMAT 13: Tindakan iklim, MATLAMAT 14: Kehidupan di dalam air

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LIST OF ABBREVIATIONS

\$	US Dollar sign
€	Euro sign
AA	arachidonic acid
ALA	α -linolenic acid
D	dark
DHA	docosahexaenoic acid
DPA	docosapentaenoic acid
EPA	eicosapentaenoic acid
FP	female-phase
g	gram
h	hour
in	inch
L	light
LOA	linoleic acid
MP	male-phase
MUFA	monounsaturated fatty acid
OA	oleic acid
ppt	parts per thousand
PSH	protandric simultaneous hermaphroditic
PUFA	polyunsaturated fatty acid
RM	Ringgit Malaysia
SA	stearic acid
SFA	saturated fatty acid
SH	simultaneous hermaphrodite
US	United States of America
Z1-Z10	zoea I – zoea X

CHAPTER 1

INTRODUCTION

In the 10th century, aquatic ornamental organism have fascinated humans that began in Asia countries such as China and Japan, and later spread to the western cultural in Turkey, Eastern Europe, and the Roman Empire (Rhyne *et al.*, 2017a). According to Doar (2007), the book, “*The Aquarium: An Unveiling of the Wonders of the Deep Blue Sea*” published in 1854 have influenced people with new hobby of keeping aquariums at that time. Gosse (1854) in his book stated that keeping aquarium make us attached to the unique sea creatures without the need to diving. Nowadays, keeping aquatic organism in household become among top three in the United States of America (US) petting industry. By the year 2017, there are 2.5 million people in the US keeping saltwater aquaria at their home (American Pet Products Association, 2017). According to Ryne *et al.* (2017b), 4.3 million invertebrates representing 545 species being traded in the US alone in the year 2008.

Cleaner shrimp was among the top demanded invertebrates in the marine ornamental industry and have created special attention due to their dazzling coloration and unique ‘personality’. Besides, this high value ornamental shrimp has a major role in maintaining animal health in the reef ecosystem (Fletcher *et al.*, 1995). Due to its behaviour, it is commonly known as cleaner shrimp or scientifically referred as *Lysmata amboinensis*.

Most of commercialized marine ornamental organisms including *L. amboinensis* come directly from the wild ecosystems, mainly from coral reefs (Wabnitz *et al.*, 2003). However, recent studies have shown a decline in the coral reefs of the world. Based on 27 years of data evaluation on reef condition, De’ath *et al.* (2012) found a major decline in hard coral cover from 28.0 % to 13.8% (2,258 surveys of 214 reefs from the year 1985 to 2012). The aquatic organism collection practices also linked to severe destruction of coral reefs through the use of explosions and poisons like cyanide (Corbin, 2001). Moreover, overharvesting of *L. amboinensis* to meets the demand for ornamental industry might also give an ecological impact in the reef area as this species helps to sustained the health of organisms in this ecosystem.

Efforts have been performed to scale down overexploitation on wild marine ornamental sources and to maintain their sustainability (Corbin, 2001; Calado *et al.*, 2003b). Aquaculture or captive culture is recognized as the best alternative to minimize wild collection while sustaining the aquarium industry (Lin, 2004). Aquaculture of ornamental species is a rapid growing sector and often seen as a priority solution in the conservation of reef habitats and relieving collection pressures on wild stocks.

According to Murray *et al.* (2012), the marine ornamental fish that have been successfully bred commercially is still low, approximately 1% to 10%. Presently, the attempts have been made to develop technologies for the mass-scale culture of a few ornamental shrimp including *L. amboinensis*, but with little success (Palmtag and Holt, 2001; Lin *et al.*, 2002). Ornamental shrimp that have been successfully reared in

captivity is only limited to a few species such as *Lysmata debelius*, *Lysmata bogessi* and *Lysmata wurdemanni*.

The major bottlenecks for mass production of *L. amboinensis* are connected to its long metamorphosis phases of larvae on the average of five months (Fletcher *et al.*, 1995) and most of the larvae died during this period. Besides, the larvae ability to delay the metamorphosis worsen the scenario. Consequently, little development has been made in the culture of *L. amboinensis* in the laboratory compared to other species of cleaner shrimps. Due to increasing demand for captive rearing of *L. amboinensis* in the marine ornamental industry as well as the decline of the world's reefs, research is therefore crucial for culturing this ornamental shrimp.

Thus, the problem statement summary for this study are stated below:

1. Currently, all cleaner shrimp, *L. amboinensis* traded in the marine ornamental industry was collected from the wild, mostly from coral reef ecosystems which cause severe damage through the use of poisons such as cyanide or explosions. Therefore, research need to be done to increase the knowledge regarding this species and minimize overexploitation and the reliability on wild specimens in the future.
2. Captive culture of this highly priced ornamental shrimp is regarded as a sustainable alternative. This species has good market value and the culture is highly promising profitable economic activity.
3. Many attempts have been made to develop technologies for the laboratory culture of a few decapod species including *L. amboinensis*, but with only little success. Longer larvae metamorphosis phases up to five months make this species research development slower compared to other species, especially, research focusing on the larvae. Higher larvae survival will increase the mass production potential. So, the information from this study would be vital for the culture production of this species.

Biological information such as broodstock environment condition and feeding, reproductive and mating behaviour, embryonic development, larvae life cycle and larvae feeding in captivity are essential for the development and the realisation of marine ornamental aquaculture technology and future commercial of *L. amboinensis*.

Therefore, the general objectives of this study are to investigate the broodstock environment culturing condition and feeding in order to optimize better output in captivity. The optimum rearing condition are needed to produce continuously reproductive cycle of berried shrimp. During spawning event, biological data and information on berried shrimp and larvae productions are needed for better captive practices. Going through the literature, very little studies were found on the early life of *L. amboinensis*. The description of embryonic development and larvae life cycle is significant for better output in captivity culture of this species.

The specific objectives of this present study are:

1. To examine the effects of salinity and temperature on the major reproductive parameters of *L. amboinensis*, in terms of broodstock survival and fecundity, egg quality and percentage of relative productivity.
2. To evaluate the effect of widely available fresh frozen diets to *L. amboinensis* broodstock based on biochemical compositions, broodstock conditions, reproductive performances and larval quality.
3. To describe the reproductive cycle, the internal and the external embryonic development, and the larval development stages of this species during cultivation.
4. To determine the effect and the suitability of microalgae and commercial diet to *L. amboinensis* larvae in terms of their survival, zoea stages composition and growth development.

REFERENCES

- Adiyodi, K. G. and Adiyodi, R. G. (1994). *Reproductive Biology of Invertebrates, Volume 6, Part B, Asexual Propagation and Reproductive Strategies*. USA: John Wiley.
- Alabi, A. O., Che Cob, Z., Jones, D. A. and Latchford, J. W. (1999). Influence of algal exudates and bacteria on growth and survival of white shrimp larvae fed entirely on microencapsulated diets. *Aquaculture International*, 7: 137–158.
- American Pet Products Association (2017). Pet Industry Market Size: Ownership Statistics. Stamford, CT.
- Amsler, M. O. and George, R. Y. (1984). Seasonal variation in the biochemical composition of the embryos of *Callinectes sapidus* rathbun. *Journal of Crustacean Biology*, 4: 546-553.
- Anger, K. (1989). Growth and exuvial loss during larval and early juvenile development of the hermit crab *Pagurus bernhardus* reared in the laboratory. *Marine Biology*, 103: 503-511.
- Anger, K. (2001). *The biology of decapod crustacean larvae. Crustacean Issues. Vol. 14*. Rotterdam, Netherlands: A. A. Balkema Publishers.
- Anil, M. K., Santhosh, B., Prasad, B.O., Saleela, K. N. and Unnikrishnan, C. (2011). Larval rearing of scarlet skunk cleaner shrimp, *Lysmata amboinensis* and fire shrimp, *Lysmata debelius*. *Renaissance in Fisheries: Outlook and strategies – Book of Abstracts, 9th Indian Fisheries Forum, CMFRI, Kochi and Asian Fisheries Society, Indian Branch, 2011*, Chennai, India. pp. 117.
- Apt, K. E. and Behrens, P. W. (1999). Commercial developments in microalgal biotechnology. *Journal of Phycology*, 35: 215-226.
- Aranyakananda, P. and Lawrence, A. L. (1994). Effects of ingestion rate on dietary protein and energy requirements of *Penaeus vannamei* and the optimal protein to energy ratio. *Memorias 2 Simposio en Nutricion' Acuicola*. Monterrey, Mexico, pp. 1–19.
- Arculeo, M., Payen, G., Cuttitta, A., Galioto, G. and Riggio, S. (1995). A survey of ovarian maturation in a population of *Aristaeus antennatus* (Crustacea: Decapoda). *Animal Biology*, 4: 13-18.
- Association of Official Analytical Chemists (AOAC). (1997). *Official Methods of Analysis of AOAC International*. Washington, USA: AOAC International.
- Aubson, B. and Patlan, D. (1974). Color changes in the ovaries of peneid shrimp as determinant of their maturity. *Marine Fisheries Review*, 36: 23-26.
- Baeza, J. A. (2010). Molecular systematics of peppermint and cleaner shrimps: phylogeny and taxonomy of the genera *lysmta* and *exhippolysmta* (crustacea:

- caridea: hippolytidae). *Zoological Journal of the Linnean Society*, 160(2): 254-265.
- Balina, S., Temperoni, B., Greco, L. S. L and Tropea, C. (2018). Losing reproduction: effect of high temperature on female biochemical composition and egg quality in a freshwater crustacean with direct development, the red cherry shrimp, *Neocaridina davidi* (decapoda, atyidae). *The Biological Bulletin*, 234(3): 139-151.
- Bas, C. and Spivak, E. (2000). Effect of salinity on embryos of two south-western Atlantic estuarine grapsid crab species cultured in vitro. *Journal of Crustacean Biology*, 20: 647-656.
- Bauer, R. T. (2000). Simultaneous hermaphroditism in caridean shrimps: a unique and puzzling sexual system in the decapoda. *Journal of Crustacean Biology*, 20(2): 116-128.
- Bauer, R. T. (2004). *Remarkable Shrimp: Adaptations and Natural History of the Carideans*. Oklahoma: University of Oklahoma Press.
- Bauer, R. T. (2006). Same sexual system but variable sociobiology: evolution of protandric simultaneous hermaphroditism in *Lysmata* shrimps. *Integrative and Comparative Biology*, 46(4): 430-438.
- Bauer, R. T. and Holt, G. J. (1998). Simultaneous hermaphroditism in the marine shrimp *Lysmata wurdemanni* (caridea: hippolytidae): an undescribed sexual system in the decapod. *Crustacea Marine Biology*, 132: 223-235.
- Bolognini, L., Donato, F., Lucchetti, A., Olivotto, I., Truzzi, C., Randazzo, B., Antonucci, M., Illuminati, S. and Grati, F. (2017). A multidisciplinary approach to study the reproductive biology of wild prawns. *Scientific Reports*, 7(1): 1-12.
- Bray, W. A., Lawrence, A. L., and Lester, L. J. (1990). Reproduction of eyestalk-ablated *Penaeus stylirostris* fed various levels of total dietary lipid. *Journal of the World Aquaculture Society*, 21(1): 41-52.
- Brito, R., Rosas, C., Chimal, M. and Gaxiola, G. (2001). Effect of different diets on growth and digestive enzyme activity in *Litopenaeus vannamei* (boone, 1931) early post larvae. *Aquaculture Research*, 32: 257-266.
- Brooks, W. K. and Herrick, F. H. (1891). The embryology and metamorphosis of the Macroura. *Memoirs of the National Academy of Sciences*, 5: 319-576.
- Brown, M. R., Jeffrey, S. W., Volkman, J. K. and Dunstan, G. A. (1997). Nutritional properties of microalgae for mariculture. *Aquaculture*, 151: 315-331.
- Bucca, P. (2008). File: *Stenopus spinosus* 1.jpg. *Wikimedia Commons, the free media repository*. Retrieved July 27, 2018 from https://commons.wikimedia.org/w/index.php?title=File:Stenopus_spinosus_1.jpg&oldid=226143032.

- Cahu, C. L., Cuzon, G. and Quazuguel, P. (1995). Effect of highly unsaturated fatty acids, α -tocopherol and ascorbic acid in broodstock diet on egg composition and development of *Penaeus indicus*. *Comparative Biochemistry and Physiology*, 112: 417-424.
- Cahu, C. L., Guillaume, J., Stephan, G. and Chim, L. (1994). Influence of phospholipid and highly unsaturated fatty acids on spawning rate and egg and tissue composition in *Penaeus vannamei* fed semi purified diets. *Aquaculture*, 126: 159-170.
- Cahu, C. L., Zambonino, I. J. L., Peres, A., Quazuguel, P. and Le Gall, M. M. (1998). Algal addition in sea bass (*Dicentrarchus labrax*) larvae rearing: effect on digestive enzymes. *Aquaculture*, 161: 479-489.
- Calado, R. (2008). *Marine Ornamental Shrimp. Biology, Aquaculture and Conservation*. U. K: Wiley-Blackwell.
- Calado, R. and Narciso, L. (2003). Seasonal variation in embryo production and brood loss in the Monaco shrimp *Lysmata seticaudata* (decapoda: hippolytidae). *Journal of the Marine Biological Association of the United Kingdom*, 83: 959–62.
- Calado, R., Dinisio, G., Nunes, C. and Dinia, M. T. (2007b). Facultative secondary lecithotrophy in the megalopa of the shrimp *Lysmata seticaudata* (risso, 1816) (decapoda: hippolytidae) under laboratory conditions. *Journal of Plankton Research*, 29(7): 599-603.
- Calado, R., Dionisio, G. and Dinis, M. T. (2007a). Starvation resistance of early zoeal stages of marine ornamental shrimps *Lysmata* spp. (decapoda: hippolytidae) from different habitats. *Journal of Experimental Marine Biology and Ecology* 351: 226-233.
- Calado, R., Dionisio, G., Bartilotti, C., Nunes, C., Santos, A. D. and Dinis, M. T. (2008). Importance of light and larval morphology in starvation resistance and feeding ability of newly hatched marine ornamental shrimps *Lysmata* spp. (decapoda: hippolytidae). *Aquaculture*. 283: 56-63.
- Calado, R., Lin, J., Rhyne, A. L., Araújo, R., and Narciso, L. (2003a). Marine ornamental decapods-popular, pricey, and poorly studied. *Journal of Crustacean Biology*, 23(4): 963–973.
- Calado, R., Martins, C., Santos, O. and Narciso, L. (2001). Larval development of the Mediterranean cleaner shrimp *Lysmata seticaudata* (risso, 1816) (caridea; hippolytidae) fed on different diets: costs and benefits of mark-time molting. *European Aquaculture Society, Special Publication*, 30: 96-99.
- Calado, R., Narciso, L., Araujo, R. and Lin, J. (2003b). Overview of Marine Ornamental Shrimp Aquaculture. In: Cato, J. C., Brown, C. L. (Eds.), *Marine Ornamental Species: Collection, Culture & Conservation* (pp. 221-230). Iowa State Press.
- Calado, R., Olivotto, I., Oliver, M. P. and Holt, J. (2017). *Marine Ornamental Species Aquaculture*. U. K: Wiley-Blackwell.

- Calado, R., Pimentel, T., Cleary, D. F. R., Dionisio, G., Nunes, C., Silva, T. L. D., Dinis, M. T. and Reis, A. (2010a). Providing a common diet to different marine decapods does not standardize the fatty acid profiles of their larvae: a warning sign for experimentation using invertebrate larvae produced in captivity. *Marine Biology*, 157: 2427–2434.
- Calado, R., Pimentel, T., Pochelon, P., Olaguer-Feliú, A. O. and Queiroga, H. (2010b). Effect of food deprivation in late larval development and early benthic life of temperate marine coastal and estuarine caridean shrimp. *Journal of Experimental Marine Biology and Ecology*. 384: 107-112.
- Calado, R., Vitorino, A., Reis, A., Lopes Da Silva, T., and Dinis, M. T. (2009). Effect of different diets on larval production, quality and fatty acid profile of the marine ornamental shrimp *Lysmata amboinensis* (de Man, 1888), using wild larvae as a standard. *Aquaculture Nutrition*, 15(5): 484-491.
- Callan, C. K., Rietfors, M. D., Laidley, C. W. and Moss, S. M. (2011). *Spawning, Larval Rearing, and Feeding of the Harlequin Shrimp, Hymenocera picta*. Paper presented at the Crustacean Society Summer Meeting, Hawaii.
- Calvo, N. S., Roldán-Luna, M., Argáez-Sosa, J. A., Martínez-Moreno, G. L., Mascaró, M. and Simões, N. (2016). Reflected-light influences the coloration of the peppermint shrimp, *Lysmata bogessi* (Decapoda: Caridea). *Journal of the World Aquaculture Society*, 47(5): 701-711.
- Carmona-Osalde, C., Rodriguez-Serna, M., Olvera-Novoa, M. A. and Gutierrez-Yurrita, P. J. (2004). Gonadal development, spawning, growth and survival of the crayfish *Procambarus llamasii* at three different water temperatures. *Aquaculture*, 232: 305-316.
- Cavalli R. O., Tamtin, M., Lavens, P. and Sorgeloos, P. (2001). Variation in lipid classes and fatty acid content in tissues of wild *Macrobrachium rosenbergii* (de Man) females during maturation. *Aquaculture*, 193: 311-324.
- Cavalli, R. O., Lavens, P. and Sorgeloos P. (1999). Performance of *Macrobrachium rosenbergii* broodstock fed diets with different fatty acid composition. *Aquaculture*, 179: 387-402.
- Cavalli, R. O., Scardua, M. P. and Wasielesky, W. J. (1997). Reproductive performance of different sized wild and pond-reared *Penaeus paulensis* females. *Journal of the World Aquaculture Society*, 28: 260-267.
- Chace, F. A. Jr. (1997). The caridean shrimps (crustacea: decapoda) of the Albatross Philippine expedition, 1907–1910, part 7: families Atyidae, Eugonatonotidae, Rhynchocinetidae, Bathypalaemonellidae, Processidae, and Hippolytidae. *Smithsonian Contributions to Zoology*, 587: 1–106.
- Charmantier, G., Charmantier-Daures, M. and Towle, D. (2009). Osmotic and Ionic Regulation in Aquatic Arthropods. In D. H. Evans (Ed.), *Osmotic and Ionic Regulation. Cells and Animals* (pp. 165-230). Boca Raton: CRC Press.

- Childs, S. (2006a). File: *Hymenocera picta*.jpg. *Wikimedia Commons, the free media repository*. Retrieved July 27, 2018 from https://commons.wikimedia.org/w/index.php?title=File:Hymenocera_picta.jpg&oldid=297455454.
- Childs, S. (2006b). File: *Thor amboinensis* - Popcorn Shrimp (234709333).jpg. *Wikimedia Commons, the free media repository*. Retrieved July 27, 2018 from [https://commons.wikimedia.org/w/index.php?title=File:Thor_amboinensis_-_Popcorn_Shrimp_\(234709333\).jpg&oldid=270476720](https://commons.wikimedia.org/w/index.php?title=File:Thor_amboinensis_-_Popcorn_Shrimp_(234709333).jpg&oldid=270476720).
- Clarke, A. (1993). Reproductive trade-offs in caridean shrimps. *Functional Ecology*, 7(4): 411-419.
- Clarke, A. and Gore, D. J. (1993). Egg Size and Composition in *Ceratoserolis* (Crustacea: Isopoda) from the Weddell Sea. In Hempel, G. (Ed.), *Weddell Sea Ecology* (pp. 129-134). Berlin: Springer-Verlag.
- Clement, S. E. (2003). Cultured Marine Ornamentals - Retail Consumer Perspectives. In: Cato, J. C., Brown, C. L. (Eds.), *Marine Ornamental Species: Collection, Culture & Conservation* (pp. 343-349). Iowa State Press.
- Cobo, V. J. and Okamori, C. M. (2008). Fecundity of the spider crab *Mithraculus forceps* (decapoda, mithracidae) from the northeastern coast of the state of São Paulo, Brazil. *Iheringia, Séries Zoologia, Porto Alegre*, 98(1), 84-87.
- Corbin, J. S. (2001). Marine ornamentals '99: conference highlights and priority recommendations. *Aquarium Sciences and Conservation*, 3: 3-11.
- Correa, C. and Thiel, M. (2003). Mating systems in caridean shrimp (decapoda: caridea) and their evolutionary consequences for sexual dimorphism and reproductive biology. *Revista Chilena de Historia Natural*, 76: 187-203.
- Cowgill, U. M., Keating, K. I. and Takahashi, I. T. (1985). Fecundity and longevity of *Ceriodaphnia dubia/affinis* in relation to diet at two different temperatures. *Journal of Crustacean Biology*, 5: 420-429.
- Crisp, J. A., Partridge, G. J., D'Souza, F. M. L., Tweedley, J. R. and Moheimani, N. R. (2017). Effects of temperature and salinity on larval survival and development of the western school prawn *Metapenaeus dalli*. *International Aquatic Research*, 9(1): 1-10.
- Cunha, L., Mascaro, M., Chiapa, X., Costa, A. and Simoes, N. (2008). Experimental studies on the effect of food in early larvae of the cleaner shrimp *Lysmata amboinensis* (de man, 1888) (decapoda: caridea: hippolytidae). *Aquaculture*, 277: 117-123.
- Cuvin-Aralar, M. L. A. (2014). Embryonic development of the caridean prawn *Macrobrachium mammillodactylus* (crustacea: decapoda: palaemonidae). *Invertebrate Reproduction and Development*, 58(4): 306-313.

- D'Abramo, L. R. (1997). Triacylglycerol and fatty acids. In E. Halver (Ed.), *Crustacean Nutrition*, Vol. 6 (pp. 71–84). Baton Rouge, Los Angeles: World Aquaculture Society.
- D'Abramo, L. R. (1998). Nutritional requirements of the freshwater prawn *Macrobrachium rosenbergii*: comparisons with species of penaeid shrimp. *Reviews in Fisheries Science*, 6: 153–163.
- D'Abramo, L. R. (2002). Challenges in developing successful formulated feed for culture of larval fish and crustaceans. In L. E. Cruz-Suárez, D. Ricque-Marie, M. Tapia-Salazar, M.G. Gaxiola-Cortés and N. Simões (Eds.) (pp. 143-151). Cancún, México: *Avances en Nutrición Acuícola VI. Memorias del VI Simposium Internacional de Nutrición Acuícola*.
- D'souza, F. M. L. and Kelly G. J. (2000). Effects of a diet of a nitrogen-limited alga (*Tetraselmis suecica*) on growth, survival and biochemical composition of tiger prawn (*Penaeus semisulcatus*) larvae. *Aquaculture*, 181: 311–329.
- D'Souza, F. M. L. and Loneragan, N. R. (1999). Effects of monospecific and mixed-algae diets on survival, development and fatty acid composition of penaeid prawn (*Penaeus* sp.) larvae. *Marine Biology*, 133: 621–633.
- Dall, W., Smith, D. M. and Moore, L. E. (1995). Carotenoids in the tiger prawn *Penaeus esculentus* during ovarian maturation. *Marine Biology*, 123: 435-441.
- Das, T., Pal, A., Chakraborty, S. K., Manush, S. M., Dalvi, R. S., Sarma, K. and Mukherjee, S. C. (2006). Thermal dependence of embryonic development and hatching rate in *Labeo rohita* (Hamilton, 1822). *Aquaculture*. 255: 536–541.
- Davis, C. C. (1965). A study of the hatching process in aquatic invertebrates: XX. the blue crabs, *Callinectes sapidus*, rathbun. *Chesapeake Science*, 6(4): 201-208.
- De'ath, G., Fabricius, K. E., Sweatman, H. and Puotinen, M. (2012). The 27-year decline of coral cover on the Great Barrier Reef and its causes. *PNAS Early Edition*, 1-5.
- Debelius, H. (2001). *Crustacea Guide of the World: Atlantic Ocean, Indian Ocean, Pacific Ocean*. Unterwasserarchive, Frankfurt: IKAN.
- Diaz, H. and Costlow, J. D. (1972). Larval development of *Ocypode quadrata* (brachyura: crustacea) under laboratory conditions. *Marine Biology*, 15: 120-131.
- Dinakaran, G. K., Soundarapandian, P. and Varadharajan, D. (2013). Embryonic development of the palaemonid prawn *Macrobrachium idella idella* (Hilgendorf, 1898). *Cell and Developmental Biology*, 2(1): 1-6.
- Doar, K. H. (2007). *Case studies in aquarium history: trends discovered in studying the history of three regional aquariums*. (Master thesis). The Florida State University College of Arts and Sciences, United Stated.

- Donelson, J. M., Munday, P. L., McCormick, M. I. and Nilsson, G. E. (2011). Acclimation to predicted ocean warming through developmental plasticity in a tropical reef fish. *Global Change Biology*, 17: 1712-1719.
- Enright, C. T., Newkirk, G. F., Craigie, J. S. and Castell, J. D. (1986). Evaluation of phytoplankton as diets for juvenile *Ostrea edulis* L. *Journal of Experimental Marine Biology and Ecology*, 96: 1-13.
- Fabregas, J., Abalde, J. and Herrero, C. (1984). Growth of the marine microalga *Tetraselmis suecica* in batch cultures with different salinities and nutrient concentrations. *Aquaculture*, 42: 207-215.
- Felder, D. L., Martin, J. W. and Goy, J. W. (1985). Patterns in early post-larval development of decapods. In A. M. Wenner (Ed.), *Larval growth, Crustacean Issues 2* (pp. 163-226). Rotterdam, Netherlands: Balkema Press.
- Fiedler, G. C. (1998). Functional, simultaneous hermaphroditism in female-phase *Lysmata amboinensis* (decapoda: caridea: hippolytidae). *Pacific Science*, 52: 161-169.
- Figueiredo, J., Penha-Lopes, G., Anto, J., Narciso, L. and Lin, J. (2008). Fecundity, brood loss and egg development through embryogenesis of *Armases cinereum* (decapoda: grapsidae). *Marine Biology*, 154(2): 287-294.
- Fletcher, D. J., Kotter, I., Wunsch, M. and Yasir, I. (1995). Preliminary observations on the reproductive biology of ornamental cleaner prawns. *International Zoo Yearbook*, 34: 73-77.
- Folch, A. J., Less, M., and Sloan-Stanley, G. H. (1957). A simple method for isolation and purification of total lipids from animal tissues. *The Journal of Biological Chemistry*, 226: 497-509.
- Food and Agriculture Organization (FAO). (2007). *Aquaculture management and conservation service. Improving Penaeus monodon hatchery practices. Manual based on experience in India*. Fisheries Technical Paper. Retrieved June 7, 2018, from <http://www.fao.org/documents/card/en/c/0e6b8d11-81ed-5afb-b374-06605dbe01d1/>.
- Freire, C. A., Amado, E. M., Souza, L. R., Veiga, M. P. T., Vitule, J. R. S., Souza, M. M. and Prodocimo, V. (2008). Muscle water control in crustaceans and fishes as a function of habitat, osmoregulatory capacity, and degree of euryhalinity. *Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology*. 149(4): 435-446.
- García-Guerrero, M. U. and Hendrickx, M. E. (2005). Embryology of decapod crustaceans, II: gross embryonic development of *Petrolisthes robsonae* (Glassell, 1945) and *Petrolisthes armatus* (gibbes, 1850) (decapoda, anomura, porcellanidae). *Crustaceana*, 78(9): 1089-1097.
- García-Guerrerol, M. and Hendrickx, M. E. (2006). Embryology of decapod crustaceans III: embryonic development of *Eurypanopeus canalensis* Abele & Kim, 1989,

- and *Panopeus chilensis* H. Milne Edwards & Lucas, 1844 (decapoda, brachyura, panopeidae). *Belgian Journal of Zoology*, 136 (2): 249-253.
- Gebauer, P., Paschke, K. and Anger, K. (1999). Costs of delayed metamorphosis: reduced growth and survival of early juveniles of an estuarine grapsid crab, *Chasmagnathus granulata*. *Journal of Experimental Marine Biology and Ecology*, 238: 271-281.
- Gebauer, P., Paschke, K. and Anger, K. (2003). Delayed metamorphosis in decapod crustaceans: evidence and consequences. *Revista Chilena de Historia Natural*, 76: 169-175.
- Gimenez, L. and Anger, K. (2001). Relationships among salinity, egg size, embryonic development, and larval biomass in the estuarine crab *Chasmagnathus granulata dana*, 1851. *Journal of Experimental Marine Biology and Ecology*, 260(2): 241-257.
- Gimenez, L. and Anger, K. (2005). Effects of temporary food limitation on survival and development of brachyuran crab larvae. *Journal of Plankton Research*, 27: 485-494.
- Glencross, B. D., Booth, M. and Allan, G. L. (2007). A feed is only as good as its ingredients - a review of ingredient evaluation strategies for aquaculture feeds. *Aquaculture Nutrition*, 13: 17-34.
- Gonza'lez, R. A., Diaz, F., Licea, A., Re, A. D., Sa'nchez, L. N. and Garcia-Esquivel, Z. (2010). Thermal preference, tolerance and oxygen consumption of adult white shrimp *Litopenaeus vannamei* (Boone) exposed to different acclimation temperatures. *Journal of Thermal Biology*, 35: 218-224.
- Gonzalez-Baro, M. R. and Pollero, R. J. (1993). Palmitic acid metabolism in hepatopancreas of the freshwater crustacean *Macrobrachium borellii* during an annual cycle. *Comparative Biochemistry and Physiology*, 106B (1): 71-75.
- Gore, R. H. (1985). Molting and Growth in Decapod Larvae. In A. M. Wenner (Ed.), *Crustacean Issues, Vol. 2 (Larval Growth)* (pp. 1-65). Rotterdam, Netherlands: A. A. Balkema.
- Gosse, P. H. (1854). *The Aquarium: An Unveiling of the Wonders of the Deep Blue Sea*. London: Paternoster Row.
- Goy, J. W. (1990). *Components of Reproductive Effort and Delay of Larval Metamorphosis in Tropical Marine Shrimp (Crustacea: Decapoda: Caridea and Stenopodidea)*. (Ph.D. Dissertation). A & M University, Texas.
- Greenley, G. (2013). Kings of the Arthropods: An Incredible Myriad of Shrimp. *Reef Hobbyist Magazine*, 7, 26-33.
- Gregrati, R. A., Fransozo, V., López-Greco, L. S. and Negreiros-Fransozo, M. L. (2010). Reproductive cycle and ovarian development of the marine ornamental shrimp *Stenopus hispidus* in captivity, *Aquaculture*. 1-6.

- Guedes, A. C. and Malcata, M. X. (2012). Nutritional Value and Uses of Microalgae in Aquaculture. In M. Zainal (Ed.), *Aquaculture* (pp. 59–78). Rijeka: InTech.
- Habashy, M. M., Sharshar, K. M. and Hassan, M. M. S. (2012). Morphological and histological studies on the embryonic development of the freshwater prawn, *Macrobrachium rosenbergii* (crustacea, decapoda). *The Journal of Basic & Applied Zoology*, 65(3): 157-165.
- Haesman, M. P. and Fielder, D. E. (1983). Laboratory spawning and mass rearing of the mangrove crab *Scylla serrata* (forskal), from first zoea to first crab stage. *Aquaculture*, 34: 303-316.
- Haplochromis (2007a). File: *Lysmata debelius*.jpg. *Wikimedia Commons, the free media repository*. Retrieved July 26, 2018 from https://commons.wikimedia.org/w/index.php?title=File:Lysmata_debelius.JPG&oldid=268102076.
- Haplochromis (2007b). File: Gobie and Shrimp.jpg. *Wikimedia Commons, the free media repository*. Retrieved July 27, 2018 from https://commons.wikimedia.org/w/index.php?title=File:Gobie_and_Shrimp.JPG&oldid=124072430.
- Harrison, K. E. (1990). The role of nutrition in maturation, reproduction and embryonic development of decapod crustaceans: a review. *Journal of Shellfish Research*. 9: 1–28.
- Harrison, K. E. (1997). Broodstock Nutrition and Maturation Diets. In L. R. D'Abramo (Ed.), *Crustacean Nutrition, Advances in World Aquaculture, Volume 6* (pp. 390-401). Louisiana, USA: World Aquaculture Society.
- Hayashi, K. I. (1975). *Hippolysmata grabhami* Gordon, a synonym of *Lysmata amboinensis* (De Man) (decapoda, caridea, hippolytidae). *Publications of the Seto Marine Biological Laboratory*, 19(5): 285-296.
- Helm, M. M., N. Bourne, and A. Lovatelli. (2004). *Hatchery Culture of Bivalves: A Practical Manual*. FAO Fisheries Technical Paper Number 471. Rome, Italy: FAO Fisheries.
- Hertrampf, J. W. and Piedad-Pascual, F. (2000) Mollusc Products. In J. W., Hertrampf and F. Piedad-Pascual (Eds.), *Handbook on ingredients for aquaculture feeds* (pp 314-321). Dordrecht, Netherlands: Springer.
- Hettiarachchi, H. A. S. U. and Edirisinghe, U. (2016). Captive breeding of fire shrimp (*Lysmata debelius*) under Sri Lankan conditions. *Tropical Agricultural Research*, 28(1): 88-99.
- Hewitt, D. R. and Duncan, P. F. (2001). Effect of high water temperature on the survival, molting and food consumption of *Penaeus (Marsupenaeus) japonicus* (Bate, 1888). *Aquaculture Research*, 32: 305-313.
- Hines, A. H. (1986). Larval pattern in the life histories of brachyuran crabs (crustacea, decapoda, brachyura). *Bulletin of Marine Science*, 39: 444-466.

- Hinz, S., Sulkin, S. D., Strom, S. and Testermann, J. (2001). Discrimination in ingestion of protistan prey by larval crabs. *Marine Ecology Progress Series*, 222: 155-162.
- Hirche, H. J., Meyer, U. and Niehoff, B. (1997). Egg production of *Calanus finmarchicus*: effect of temperature, food and season. *Marine Biology*, 127: 609-620.
- Holland, D. (1978). Lipid Reserves and Energy Metabolism in the Larvae of Benthic Marine Invertebrates. In D. C. Malins (Ed.), *Biochemical and Biophysical Perspectives in Marine Biology* (pp. 85-123). Seattle: Academic Press.
- Houser, R. and Akiyama, D. (1997). Feed Formulation Principles. In L. D'Abramo, D. Conklin and D. Akiyama (Eds.), *Advances in World Aquaculture, Vol. 6 (Crustacean Nutrition)* (pp. 493-520). Louisiana, USA: The World Aquaculture Society.
- Huffman, L. (2009). File: *Lysmata amboinensis* shrimp.jpg. *Wikimedia Commons, the free media repository*. Retrieved July 26, 2018 from https://commons.wikimedia.org/w/index.php?title=File:Lysmata_amboinensis_Shrimp.jpg&oldid=142879170.
- IUCN (2018a). (2018, July 24). *The IUCN red list of threatened species version 2018-1*. Retrieved from <http://www.iucnredlist.org>.
- IUCN (2018b). (2018, July 24). *The IUCN red list of threatened species version 2018-1*. Retrieved from <http://www.iucnredlist.org/news/saving-nemo>.
- Jensen, T. G. (2001). Arabian sea and bay of bengal exchange of salt and tracers in an ocean model. *Geophysical Research Letters*, 28(20): 3967-3970.
- Johnson, V. R. Jr. (1977). Individual recognition in the banded shrimp *Stenopus hispidus* (olivier). *Animal Behaviour*, 25: 418-428.
- Johnston, M. D., Johnston, D. J., Knott, B. and Jones, C. (2005). Mouthpart and foregut ontogeny in phyllosomata of *Panulirus ornatus* and their implications for development of a formulated larval diet. *European Aquaculture Society, Special Publication*, 36: 223-226.
- Jones, D. A., Kumlu, M., LeVay, L. and Fletcher, D. J. (1997). The digestive physiology of herbivorous, omnivorous and carnivorous crustacean larvae: a review. *Aquaculture*, 155: 285-295.
- Jun-jie, Y., Yun-long, Z., Qun, W., Zhong-liang, Z., Xian-cheng, H., Xiao-wei, D. and Chuan-guang, A. (2006). Biochemical compositions and digestive enzyme activities during the embryonic development of prawn, *Macrobrachium rosenbergii*. *Aquaculture*, 253: 573-582.
- Kraul, S. (1999). Commercial culture of the harlequin shrimp *Hymenocera picta* and other ornamental marine shrimp. *Marine Ornamentals '99, Book of Abstracts. Hawaii, USA*. pp. 50.

- Krogh, A. (1966). *Osmotic Regulation in Aquatic Animals*. New York: Dover Publications.
- Kumlu, M. and Jones, D. A. (1995). The effect of live and artificial diets on growth, survival, and trypsin activity in larvae of *Penaeus indicus*. *Journal of World Aquaculture Society*, 26: 406-415.
- Kuris, A. M. (1991). A Review of Patterns and Causes of Crustacean Brood Mortality. In A. Wenner and A. Kuris (Eds.), *Crustacean Egg Production Vol.7*. (pp. 117-141) Rotterdam, Netherlands: *Crustacean Issues*.
- Lange, R. and Mostad, A. (1967). Cell volume regulation in osmotically adjusting marine animals. *Journal of Experimental Marine Biology and Ecology*, 1(2): 209-219.
- Le Moullac, G., Wormhoudt, A. V. and AQUACOP. (1994). Adaptation of digestive enzymes to dietary protein, carbohydrate and fibre levels and influence of protein and carbohydrate quality in *Penaeus vannamei* larvae (crustacea, decapoda). *Aquatic Living Resources*, 7: 203-210.
- Le Vay, L., Jones, D., Puello-Cruz, A., Sangha, R. and Ngamphongsai, C. (2001). Digestion in relation to feeding strategies exhibited by crustacean larvae. *Comparative Biochemistry and Physiology*. 128: 623–630.
- Liao, I. C., Su, H. M. and Lin, J. H. (1993). Larval Foods for Penaeid Prawns. In J. P. McVey (Ed.), *Handbook of mariculture, crustacean aquaculture, vol 1* (pp. 43–69). Boca Raton: CRC Press.
- Lin, J. (2004). Aquaculture of ornamental marine shrimp provides alternative to wild collection. *Global Aquaculture Advocate*, 55–56.
- Lin, J. and Shi, P. (2002). Effect of broodstock diet on reproductive performance of the golden banded coral shrimp *Stenopus scutellatus*. *Journal of the World Aquaculture Society*, 33(3): 383–386. .
- Lin, J. and Zhang, D. (2001a). Effects of broodstock diet on reproductive performance of the peppermint shrimp *Lysmata wurdemanni*. *Journal of Shellfish Research*, 20: 361–363.
- Lin, J. and Zhang, D. (2001b). Reproduction in simultaneous hermaphroditic shrimp *Lysmata wurdemanni*: any two will do?. *Marine Biology*, 139: 919-922.
- Lin, J., Zhang, D., and Rhyne, A. (2002). Broodstock and Larval Nutrition of Marine Ornamental Shrimp. In L. E. Cruz-Suaréz, D. Ricque-Marie, M. Tapia-Salazar, M. G. Graxiola-Cortés, and N., Simoes, (Eds.), *Avances em Nutrición Acuícola VI. Memorias del VI*. (pp. 277-280). Cancun, Mexico: *Símposium Internacional de Nutrición Acuícola*.
- Lin, Q., Lu, J., Gao, Y., Shen, L., Cai, J. and Luo, J. (2006). The effect of temperature on gonad, embryonic development and survival rate of juvenile seahorses, *Hippocampus kuda* Bleeker. *Aquaculture*, 254: 701–713.

- Liñán-Cabello, M. A., Paniagua, M. and Hopkins, P. M. (2002). Bioactive roles of carotenoids and retinoids in crustaceans. *Aquaculture Nutrition*, 8: 299-309.
- Ling, R. (2006). File: *Stenopus hispidus* 1.jpg. *Wikimedia Commons, the free media repository*. Retrieved July 26, 2018 from https://commons.wikimedia.org/w/index.php?title=File:Stenopus_hispidus_1.jpg&oldid=297457146.
- Llodra, R. E. (2002). Fecundity and life-history strategies in marine invertebrates. *Advances in Marine Biology*, 43: 87-170.
- Lopez-Elias, J. A., Voltolina, D., Chavira-Ortega, C. O., Rodríguez, B. B., Saenz-Gaxiola, L. M., Esquivel, B. C., Nieves, M. (2003). Mass production of microalgae in six commercial shrimp hatcheries of the Mexican northwest. *Aquacultural Engineering*. 29: 155-164.
- Makridis, P., Costa, R. A. and Dinis, M. T. (2006). Microbial conditions and antimicrobial activity in cultures of *Chlorella minutissima* and effect on bacterial load of enriched artemia metanauplii. *Aquaculture*, 255: 76-81.
- Manning, R. B. and Chace, Jr. F. A. (1990). Decapod and stomatopod crustacea from Ascension Island, South Atlantic Ocean. *Smithsonian Contributions to Zoology*, 503: 1-91.
- Mantelatto, F. L. M. and Fransozo, A. (1997). Fecundity of the crab *Callinectes ornatus Ordway*, 1863 (decapoda, brachyura, portunidae) from the Ubatuba region, São Paulo, Brazil. *Crustaceana*, 70: 214-226.
- Marichamy, R. and Rajapackiam, S. (1992). Experiment on larval rearing and seed production of the mud crab *Sylla serrata* (Forskal), in: Angell, C. A. (ed.). Report of the seminar on the mud crab culture and trade, The Mud Crab. Madras, Bay of Bengal Program. Pp. 135-141.
- Martin, R. X., Turco, E. B. R., and Bazan, N. G. (1994). Developmental maturation of hepatic n-3 polyunsaturated fatty acid metabolism: supply of docosahexaenoic acid to retina and brain. *The Journal of Nutritional Biochemistry*, 5: 151-160.
- McClanahan, T. R., Sheppard, C. R. C and Obura, D. O. (2000). *Coral Reefs of the Indian Ocean: Their Ecology and Conservation*. New York: Oxford University Press.
- Mcedward, L. R. and Morgan, K. H. (2001). Interspecific relationships between egg size and the level of parental investment per offspring in echinoderms. *The Biological Bulletin*. 200(1): 33-50.
- McGaw, I. J. and Curtis, D. L. (2012). A review of gastric processing in decapod crustaceans. *Journal of Comparative Physiology B*, 183(4): 443-465.
- McLaren, I. A. (1965). Some relationships between temperature and egg size, body size, development rate, and fecundity of the copepod *Pseudocalanus*. *Limnology and Oceanography*, 10: 528-538.

- Meseck, L. S. T., Jennifer, H. A. and Wikfors, H. G. (2005). Photoperiod and light intensity effects on growth and utilization of nutrients by the aquaculture feed microalgae, *Tetraselmis chuii*. *Aquaculture*, 246: 393–404.
- Militz T. A. and Hutson, K. S. (2015). Beyond symbiosis: cleaner shrimp clean up in culture. *PLoS ONE*, 10(2): 1-11.
- Monroig, O., Navarro, J.C., Amat, F., González, P. and Hontoria, F. (2006). Effects of nauplii density, product concentration and product dosage on the survival of the nauplii and EFA incorporation during artemia enrichment with liposomes. *Aquaculture*, 261: 659-669.
- Moran, A. L. and McAlister, J. S. (2009). Egg size as a life history character of marine invertebrates: is it all it's cracked up to be?. *The Biological Bulletin*, 216: 226-242.
- Mourente, G. and Rodriguez, A. (1991). Variation in the lipid content of wild-caught females of marine shrimp *Penaeus kerathurus* during sexual maturation. *Marine Biology*, 110(1): 21-28.
- Mourente, G. and Tocher, D. R. (1992). Effects of weaning onto a pelleted diet on docosahexaenoic acid (22:6n-3) levels in brain of developing turbot (*Scophthalmus maximus*). *Aquaculture*, 105: 363-377.
- Muller, Y. M. R., Nazari, E. M. and Simões-Costa, M. S. (2003). Embryonic stages of the freshwater prawn *Macrobrachium olfersi* (decapoda, palaemonidae). *Journal of Crustacean Biology*, 23(4): 869–875.
- Munday, P. L., Kingsford, M. J., O'callaghan, M. and Donelson, J. M. (2008). Elevated temperature restricts growth potential of the coral reef fish *Acanthochromis polyacanthus*. *Coral Reefs*, 27: 927–931.
- Murray, J. M., Watson, G. J., Giangrande, A., Licciano, M. and Bentley, M. G. (2012). Managing the marine aquarium trade: revealing the data gaps using ornamental polychaetes. *PLoS ONE*, 7(1): 1-8.
- Naessens, E., Lavens, P., Gomez, L., Browdy, C., McGovern-Hopkins, K., Spencer, A., Kawahigashi, D. and Sorgeloos, P. (1997). Maturation performance of *Penaeus vannamei* co-fed Artemia biomass preparations. *Aquaculture*, 155: 87-101.
- Nagao, J., Munehara, H. and Shimazaki K. (1999). Embryonic development of the hair crab *Erimacrus isenbeckii*. *Journal of Crustacean Biology*, 19: 77-83.
- Nair, K. B. (1948). The embryology of *Caridina laevis* heller. *Proceedings of the National Academy of Sciences India (B)*, 29: 211-288.
- Narciso, L., and Morais, S. (2001). Fatty acid profile of *Palaemon serratus* (palaemonidae) eggs and larvae during embryonic and larval development using different live diets. *Journal of Crustacean Biology*, 21(3): 566-574.
- Nazari, E. M., Simões-Costa, M. S., Müller, Y. M. R., Ammar, D. and Dias, M. (2003). Comparisons of fecundity, egg size, and egg mass volume of the freshwater

- prawns *Macrobrachium potiuna* and *Macrobrachium olfersi* (decapoda, palaemonidae). *Journal of Crustacean Biology*, 23(4): 862-868.
- Nilsson, G. E., Crawley, N., Lunde, I. G. and Munday, P. L. (2009). Elevated temperature reduces the respiratory scope of coral reef fishes. *Global Change Biology*, 15: 1405-1412.
- Oh, C. and Hartnoll, R. G. (1999). Brood loss during incubation in *Philocheras trispinosus* (decapoda) in port erin bay, isle of man. *Journal of Crustacean Biology*, 19: 467-476.
- Orrell T. (custodian) (2018). ITIS Regional: The Integrated Taxonomic Information System (version Dec 2014). In: Species 2000 & ITIS Catalogue of Life, 2015 Annual Checklist (Roskov Y., Abucay L., Orrell T., Nicolson D., Kunze T., Flann C., Bailly N., Kirk P., Bourgoin T., DeWalt R. E., Decock W., De Wever A., eds). Digital resource at http://www.itis.gov/servlet/SingleRpt/SingleRpt?search_topic=TSN&search_value=659974,Netherlands. ISSN 2405-8858.
- Palacios E., Pérez-Rostro C. I., Ramírez J. L., Ibarra A. M. and Racotta I. S. (1999). Reproductive exhaustion in shrimp (*Penaeus vannamei*) reflected in larval biochemical composition, survival, and growth. *Aquaculture*, 171: 209-221.
- Palacios, E. Racotta, I. S., Heras, H., Marty, Y., Marty, Y., Moal, J. and Samain, F. (2001). Relation between lipid and fatty acid composition of eggs and larval survival in white pacific shrimp (*Penaeus vannamei*, boone, 1931). *Aquaculture International*, 9(6): 531-543.
- Palacious, E., Ibarra, A. M. and Racotta, I. S. (2000). Tissue biochemical composition in relation to multiple spawning in wild and pond-reared *Penaeus vannamei* broodstock. *Aquaculture*, 185: 353-371.
- Palanisamy, V., Latif, F. A. and Resat, R. M. (1991). A Guide on the Production of Algal Culture for Use in Shrimp Hatcheries. Kuala Lumpur: Department of fisheries, Ministry of Agriculture, Malaysia.
- Palmtag, M. R. and G. J. Holt. (2007). Experimental studies to evaluate larval survival of the fire shrimp, *Lysmata debelius*, to the juvenile stage. *Journal of the World Aquaculture Society*, 38: 102-113.
- Palmtag, M. R. and Holt, G. J. (2001). Captive rearing of fire shrimp (*Lysmata debelius*). Texas Sea Grant College Program Research Report, pp. 1-4.
- Pechenik, J. A. (1999). On the advantages and disadvantages of larval stages in benthic marine invertebrate life cycles. *Marine Ecology Progress Series*, 177: 269-297.
- Penisson, S. (2016). Boxing Shrimps In The Ring. *Reef Hobbyist Magazine*, 10, 34-41.
- Perez, M. F. and Sulkin, S. D. (2005). The palatability of autotrophic dinoflagellates in larval crabs. *Marine Biology*. 146: 771-780.

- Perez-Velazquez, M., Gonzalez-Felix, M. L., Lawrence, A. L., Bray, W. A. and Gatlin, D. M. (2003). Dietary effects on sperm quality of *Litopenaeus vannamei* broodstock. *Journal of the World Aquaculture Society*, 34(1): 92-98.
- Pinheiro, M. A. A. and Hattori, G. Y. (2003). Embryology of the mangrove crab *Ucides cordatus* (brachyura: ocypodidae). *Journal of Crustacean Biology*, 23: 729-737.
- Poulin, E., Boletzky, S. V. and Feral, J. P. (2001). Combined ecological factors permit classification of developmental patterns in benthic marine invertebrates: a discussion note. *Journal of Experimental Marine Biology and Ecology*, 257(1): 109-115.
- Prakash, S., Kumar, T. T. A and Subramoniam, T. (2010). New records of marine ornamental shrimps (decapoda: stenopodidea and caridea) from the Gulf of Mannar, Tamil Nadu, India. *Check List*, 12(6): 1-6.
- Provenzano, A. J. (1985). Commercial culture of decapod crustaceans. In A. J. Provenzano, Jr. (Ed.), *Economic Aspects: Fisheries and Culture, The Biology of Crustacea. Vol. 10* (pp. 269–314). New York: Academic Press.
- Querijero, B.V.L., Teshima, S., Koshio, S. and Ishikawa, M. (1997). Utilization of monounsaturated fatty acid (18:1n-9, oleic acid) by freshwater prawn *Macrobrachium rosenbergii* (de man) juveniles. *Aquaculture Nutrition*, 3: 127-139.
- Quintero, M. E. S. and Gracia, A. (1998). Stages of gonadal development in the spotted pink shrimp *Penaeus brasiliensis*. *Journal of Crustacean Biology*, 18(4): 680-685.
- Rabalais, N. N. and Gore, R. H. (1985). Abbreviated developments in decapods. *Crustacean Issues*, 2: 67-126.
- Racotta, I. S., Palacios, E. and Ibarra, A. M. (2003). Shrimp larval quality in relation to broodstock condition. *Aquaculture*, 227: 107-130.
- Rajion, M. A., McLean, J. G. and Cahill, R. N. (1985). Essential fatty acids in the fetal and newborn lamb. *Australian Journal of Biological Sciences*, 38(1): 33-40.
- Reiber, C. L. and McMahon, B. R. (1998). Progressive hypoxia's effects on the crustacean cardiovascular system: a comparison of the freshwater crayfish (*Procambarus clarkii*) and the lobster (*Homarus americanus*). *Journal of Comparative Physiology*, 168(3): 168–176.
- Rhyne, A. J., Lin, J., Calado, R. and Turingan, R. (2001). Improvements in marine ornamentals shrimp culture: high speed video analysis of feeding kinematics in dietary study. *Book of Abstracts Marine Ornamentals 2001. 'Collection, Culture and Conservation'*. Orlando, Florida, USA, pp. 29-30.
- Rhyne, A. L. and Lin, J. (2004). Effects of different diets on larval development in a peppermint shrimp (*Lysmata* sp. (Risso)). *Aquaculture Research*, 35: 1179-1185.

- Rhyne, A. L. and Lin, J. (2006). A western Atlantic peppermint shrimp complex: redescription of *Lysmata wurdemanni*, description of four new species, and remarks on *Lysmata rathbunae* (Crustacea: Decapoda: Hippolytidae). *Bulletin of Marine Science*. 79: 165–204.
- Rhyne, A. L., Tlusty, M. F. and Szczebak, J. T. (2017a). Early culture trials and an overview on U.S. In Calado, R., Olivotto, I., Oliver, M. P. and Holt, G. J. (Eds.), *Marine ornamental species trade marine ornamental species aquaculture* (pp. 51-70). John Wiley and Sons Ltd.
- Rhyne, A. L., Tlusty, M. F., Holmberg, R. J. and Szczebak J. T. (2015). (2018, June 23). Marine aquarium biodiversity and trade flow. Retrieved from Aquariumtradedata.org.
- Rhyne, A. L., Tlusty, M. F., Szczebak J. T. and Holmberg, R. J. (2017b). Expanding our understanding of the trade in marine aquarium animals. *PeerJ*. 1-36.
- Ricker, W. E. (1979). Growth rates and models. In W. S. Hoar, D. J. Randall and J. R. Brett. (Eds.), *Fish physiology* (pp. 677-743). New York: Academic Press.
- Robertson, J. D. (1960). Ionic regulation in the crab *Carcinus maenas* (L.) in relation to the moulting cycle. *Comparative Biochemistry and Physiology*, 1: 183-212.
- Rocha, A. P. P. (2007). *Experimental and commercial production of Lysmata seticaudata* (Master thesis), University of Lisbon, Portugal.
- Rodriguez, A., Vay, L. L., Mourente, G. and Jones, D. A (1994) Biochemical composition and digestive enzyme activity in larvae and postlarvae of *Penaeus japonicus* during herbivorous and carnivorous feeding. *Marine Biology*, 118: 45–51.
- Romano, N. and Zeng, C. (2012). Osmoregulation in decapod crustaceans: Implications to aquaculture productivity, methods for potential improvement and interactions with elevated ammonia exposure. *Aquaculture*, 334-337: 12-23.
- Rosa, R., Calado, R., Andrade, A. M., Narciso, L. and Nunes, M. L. (2005). Changes in amino acids and lipids during embryogenesis of european lobster, *Homarus gammarus* (Crustacea: Decapoda). *Comparative Biochemistry and Physiology B*, 140: 241-249.
- Rosa, R., Calado, R., Narciso, L. and Nunes, M. L. (2007). Embryogenesis of decapod crustaceans with different life history traits, feeding ecologies and habitats: a fatty acid approach. *Marine Biology*, 151: 935-947.
- Rosa, R., Lopes, A. R., Pimentel, M., Faleiro, F., Baptista, M., Trubenbachi, K. J., Narciso, L., Dionisio, G., Pegado, M. R. Repolho, T., Calado, R. and Diniz, M. (2014). Ocean cleaning stations under a changing climate : biological responses of tropical and temperate fish-cleaner shrimp to global warming, 20: 3068–3079.
- Rosa, R., Morais, S., Calado, R., Narciso, L. and Nunes, M. L. (2003). Biochemical changes during the embryonic development of norway lobster, *Nephrops norvegicus* (crustacea: decapoda). *Aquaculture*, 221: 507-522.

- Roughley, T. C. (1936). *Wonders of the Great Barrier Reef*. Sydney and London: Angus & Robertson Limited.
- Rufino, M. M. and Jones, D. A. (2001a). Binary individual recognition in *Lysmata debelius* (decapoda: hippolytidae) under laboratory conditions. *Journal of Crustacean Biology*, 21(2): 388-392.
- Rufino, M. M. and Jones, D. A. (2001b). Observations on the function of the fifth pereiopod in late stage larvae of *Lysmata debelius* (decapoda, hippolytidae). *Crustaceana*, 74: 977-990.
- Sampaio, L. A. and Bianchini, A. (2002). Salinity effects on osmoregulation and growth of the euryhaline flounder *Paralichthys orbignyanus*. *Journal of Experimental Marine Biology and Ecology*, 269(2): 187-196.
- Sandeman, R. and Sandeman, R. (1991). Periods in the development of the embryo of the freshwater crayfish *Cherax destructor*. *Roux's Archives of Developmental Biology*, 200: 27-37.
- Sang, H. M. and Fotedar, R. (2004). Growth, survival, haemolymph osmolality and organosomatic indices of the western king prawn (*Penaeus latisulcatus* kishinouye, 1896) reared at different salinities. *Aquaculture*, 234: 601-614.
- Santos, A. D., Calado, R., Bartolotti, C. and Narciso, L. (2004). The larval development of the partner shrimp *Periclimenes sagittifer* (norman, 1861) (decapoda: caridea: palaemonidae: pontoniinae) described from laboratory-reared material, with a note on chemical settlement cues. *Helgoland Marine Research*, 58: 129-139.
- Sargent, J. R. (1995). Origins and functions of egg lipids: nutritional implications. In N. R. Bromage and R. J. Roberts (Eds.) *Broodstock management and egg and larval quality* (pp. 353-372). Oxford, United Kingdom: Blackwell Science.
- Saritha, K., Mary, D. and Patterson, J. (2015). Nutritional status of green mussel *Perna viridis* at Tamil Nadu, Southwest Coast of India. *Journal of Nutrition & Food Sciences*, S14: 1-4.
- Sastray, A. N. (1983). Ecological Aspects of Reproduction. In F. J. Vernberg and W. B. Vernberg (Eds.), *The biology of Crustacea. Environmental Adaptations* (pp. 179-270). New York: Academic Press.
- Saxena, A. (2005). *Textbook of Crustacea*. New Delhi: Discovery Publishing House.
- Schwamborn, R., Ekau, W., Silva, A. P., Schwamborn, S. H. L., Silva, T. A., Neumann-Leitão, S. and Saint-Paul, U. (2006). Ingestion of large centric diatoms, mangrove detritus, and zooplankton by zoeae of *Aratus pisonii* (crustacea: brachyura: grapsidae). *Hydrobiologia*, 560: 1-13.
- Seotaro, (2007). File: *Rhynchocinetes uritai.jpg*. *Wikimedia Commons, the free media repository*. Retrieved July 26, 2018 from [https://commons.wikimedia.org/w/index.php?title=File:Rhynchocinetes_uritai\(Shimoda,Shizuoka,Japan,2007\).jpg&oldid=155555978](https://commons.wikimedia.org/w/index.php?title=File:Rhynchocinetes_uritai(Shimoda,Shizuoka,Japan,2007).jpg&oldid=155555978).

- Smith, D. J. B., Diele, K. and Abrunhosa F. A. (2013). Carry-over effects of delayed larval metamorphosis on early juvenile performance in the mangrove crab *Ucidès cordatus* (ucididae). *Journal of Experimental Marine Biology and Ecology*, 440: 61-68.
- Simoes, F., Ribeiro, F. and Jones, D. A. (1998). The effect of diet on the reproductive performance of marine cleaner shrimps *L. debelius* (Bruce, 1983) and *Lysmata amboinensis* (de Man, 1888) (Caridea, Hippolytidae) in captivity. *Aquaculture 98 Book of Abstracts*, Las Vegas, Nevada, USA, pp. 497.
- Simoes, F., Ribeiro, F. and Jones, D. A. (2002). Feeding early larval stages of fire shrimp *Lysmata debelius* (caridea, hippolytidae). *Aquaculture International*, 10: 349-360.
- Sin, N. N. N., Kian, A. Y. S. and Shapawi, R. (2016). Effects of different protein sources in the broodstock diet on reproductive performance of giant freshwater prawn (*Macrobrachium rosenbergii*). *International Journal of Aquatic Science*, 7(2): 87-94.
- Singh, I. S. B. and Philip, R. (1995). A simple device for the separation of weak larvae of *Macrobrachium rosenbergii* (de man). *Aquaculture Research*, 26: 225-227.
- Smith, L. L., Beidenbach, J. M. and Lawrence, A. L. (1992). Penaeid larviculture: Galveston method. In Fast, A. W. and Lester, L. J. (Eds.), *Marine shrimp culture: principles and practises. Development in aquaculture and fisheries science* (pp. 171-193). Amsterdam, Holland: Elsevier.
- Soundarapandian, P., Dinakaran, G. K. and Varadharajan, D. (2014). Effect of temperatures on the embryonic development, morphometrics and survival of *Macrobrachium Idella* (hilgendorf, 1898). *Journal of Aquaculture Research and Development*, 5(7): 1-6.
- Steele, D. H. and Steele, V. J. (1991). Effects of salinity on the survival, growth rate and reproductive output of *Gammarus lawrencianus* (crustacea, amphipoda). *Marine Ecology Progress Series*, 78: 49-56.
- Subramoniam, T. (1991). Yolk utilization and esterase activity in the mole crab *Emerita asiatica*. *Crustacean Issues*, 7: 19-30.
- Suprayudi, M. A., Takeuchi, T. and Hamasaki, K. (2004) Essential fatty acids for larval mud crab *Scylla serrata*: implications of lack of the ability to bioconvert C18 unsaturated fatty acids to highly unsaturated fatty acids. *Aquaculture*, 231: 403-416.
- Sverdrup, H. N., Johnson, M. W. and Fleming, R. H. (1942). *The oceans*. New Jersey: Prentice Hall.
- Takeuchi, T., Satoh, N., Sekiya, S., Shimizu, T. and Watanabe, T. (1999). The effect of dietary EPA and DHA on molting rate of larval swimming crab *Portunus trituberculatus*. *Nippon Suisan Gakkaishi*, 65(6): 988-1004.

- Tan-Fermin, J. D. (1991). Effects of unilateral eyestalk ablation on ovarian histology and oocyte size frequency of wild and pond-reared *Penaeus monodon* (Fabricius) broodstock. *Aquaculture*, 93: 77-86.
- Teshima, S. (1998). Nutrition of *Penaeus japonicas*. *Reviews in Fisheries Science*, 6(2): 97-111.
- Teshima, S. I., Koshio, S., Ishikawa, M., Alam, M. S., and Hernandez, L. H. (2006). Protein requirements of the freshwater prawn *Macrobrachium rosenbergii* evaluated by the factorial method. *Journal of the World Aquaculture Society*, 37(2): 145–153.
- Tewksbury, J. J., Huey, R. B. and Deutsch, C. A. (2008). Putting the heat on tropical animals. *Science*, 320: 1296-1297.
- Thompson, P. A., Guo, M. and Harrison, P. J. (1993). The influence of irradiance on the biochemical composition of three phytoplankton species and their nutritional value for larvae of the Pacific oyster (*Crassostrea gigas*). *Marine Biology*, 117: 259–268.
- Thoney, D. A., Warmolts, D. I. and Andrews, C. (2003). Acquisition of fishes and aquatic invertebrates for zoological collections. Is there a future?. *Zoo Biology*, 22: 519–527.
- Tompkins, J. (1995). Culture Collection of Algae and Protozoa. In M. M., Day and M. F. Turner (Eds.), *Catalogue of Strains: Natural Environment Research Council* (pp. 144-173). United Kingdom: Titus Wilson and Sons Ltd, Kendal.
- Tropea, C., Stumpf, L. and López-Greco, L. S. (2015). Effect of temperature on biochemical composition, growth and reproduction of the ornamental red cherry shrimp *Neocaridina heteropoda heteropoda* (decapoda, caridea). *PLoS ONE*, 10(3): 1-14.
- Tsuji, H., Zeng, C., Romano, N., Southgate, P. C. and Ye, H. (2015). Development of larval culture methods for blue-legged gold coral banded shrimp *Stenopus cyanoscelis*: effects of prey type and prey density. *Fish Science*, 81: 731–736.
- Tziouveli, V. (2011). *Broodstock conditioning and larval rearing of the marine ornamental white-striped cleaner shrimp, Lysmata amboinensis (de Man, 1888)*. (PhD thesis). James Cook University, Australia.
- Tziouveli, V. and Smith, G. (2009). Sexual maturity and environmental sex determination in the white-striped cleaner shrimp *Lysmata amboinensis*. *Invertebrate Reproduction and Development*, 53(3): 155-163.
- Tziouveli, V. and Smith, G. G. (2012). A comparison of the fatty acid profiles of adult tissues, and newly hatched, fed and starved *Lysmata amboinensis* larvae. *Aquaculture Research*, 43(4): 577-587.
- Tziouveli, V., Hall, M. and Smith, G. (2011). The effect of maturation diets on the reproductive output of the white-striped cleaner shrimp, *Lysmata amboinensis*.

- Tziouveli, V., Hall, M., and Smith, G. G. (2012). Evaluation of lipid-enriched artemia on the reproductive performance of the white-striped cleaner shrimp, *Lysmata amboinensis*. *Aquaculture International*, 20(2): 201-211.
- Utinomi, H. (1956). *Coloured Illustrations of Sea Shore Animals of Japan*. Hoikusha, Osaka: Hoikusha Publication Corporation.
- Venkataramiah, A., Lakshmi, G. J. and Gunter, G. (1974). *Studies on the effects of salinity and temperature on the commercial shrimp, Penaeus aztecus Ives: With special regard to survival limits growth, oxygen consumption and ionic regulation*. Mississippi: Gulf Coast Research Laboratory (Ocean Springs).
- Vos, A. D., Pattiarchi, C. B. and Wijeratne, E. M. S. (2014). Surface circulation and upwelling patterns around Sri Lanka. *Biogeosciences*, 11: 5909-5930.
- Wabnitz, C., Taylor, M., Green, E., Razak, T. (2003). *From ocean to aquarium*. Cambridge: UNEP-WCMC.
- Waloszek, D. and Maas, A. (2005). The evolutionary history of crustacean segmentation: a fossil-based perspective. *Evolution & Development*, 7: 515-527.
- Wear, R. G. (1974). Incubation in British decapod crustacean, and the effects of temperature on the rate and success of embryonic development. *Journal of the Marine Biological Association of the United Kingdom*, 54: 745-762.
- Wen, X. B., Chen, L. Q., Zhou, Z. L., Ai, C. X. and Deng, G. Y. (2002). Reproduction response of chinese mittenhanded crab (*Eriocheir sinensis*) fed different source of dietary lipid. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 131(3): 675-681.
- Williams, K. C. (2007). Nutritional requirements and feeds development for post-larval spiny lobster: a review. *Aquaculture*, 263: 1-14.
- Williamson, D. I. (1982). Larval morphology and diversity. In L.G. Abele (Ed.), *The Biology of Crustacea, Vol. 2 (Embryology, Morphology and Genetics)* (pp. 43-110). New York: Academic Press.
- Wirtz, P. (1997). Crustacean symbiots of the sea anemone *Telmatostylis cricoides* at madeira and the canary island. *Journal of Zoology (London)*, 242: 799-811.
- Wong, J. W., and Michiels, N. K. (2011). Control of social monogamy through aggression in a hermaphroditic shrimp. *Frontiers in Zoology*, 8(30): 1-7.
- Wouters, R., Gomez, L., Lavens, P. and Calderon, J. (1999). Feeding enriched artemia biomass to *Penaeus vannamei* broodstock: its effect on reproductive performance and larval quality. *Journal of Shellfish Research*, 18(2), 651-659.
- Wouters, R., Lavens, P., Nieto, J. and Sorgeloos, P. (2001). Penaeid shrimp broodstock nutrition: an updated review on research and development. *Aquaculture*, 202: 1-21.

- Wu, X. (2013). *Lipid nutrition of early life history of two commercially important tropical crustaceans, the blue swimmer crab (*Portunus pelagicus*) and the ornate rock lobster (*Panulirus ornatus*), with emphasis on highly unsaturated fatty acids* (Doctoral thesis), James Cook University, Australia.
- Xu, X. L., Ji, W. J., Castell, J. D., and O'Dor, R. K. (1994). Essential fatty acid requirement of the chinese prawn *Penaeus chinensis*. *Aquaculture*, 127: 29-40.
- Xue, S., Fang, J., Zhang, J., Jiang, Z., Mao, Y. and Zhao, F. (2013). Effects of temperature and salinity on the development of the amphipod crustacean *Eogammarus sinensis*. *Chinese Journal of Oceanology and Limnology*. 31(5), 1010-1017.
- Yajing, Z., Y., Shengli, Y. and Hong, L. (2012). Morphological observation of embryonic development of marine ornamental shrimp (*Lysmata amboinensis*). *Journal of Fishery Sciences of China*, 19(6): 923-929.
- Yamaguchi, T. (2001). Incubation of eggs and embryonic development of the fiddler crab, *Uca lactea* (decapoda, brachyura, ocypodidae). *Crustaceana*, 74: 449-458.
- Yang, H. J. and Kim, W. (2006). A new record of *Lysmata amboinensis* (de man) (decapoda: hippolytidae) from Jeju-do Island, Korea. *Journal of Fisheries Science and Technology*, 9(3): 118-122.
- Yao, J., Zhao, Y. L., Wang, Q., Zhou, Z. L., Hu, X. C., Duan, X. W. and An-Chuan, G. (2006). Biochemical compositions and digestive enzyme activities during the embryonic development of prawn, *M. rosenbergii*. *Aquaculture*, 253: 573-582.
- Zaki, M. L. and Saad, H. (2010). Comparative study on growth and survival of larval and juvenile *Dicentrarchus labrax* rearing on rotifer and artemia enriched with four different microalgae species. *African Journal of Biotechnology*, 9: 3576-3588.
- Zhang D, Rhyne L. A and Lin, J. (2007). Density-dependent effect on reproductive behaviour of *Lysmata amboinensis* and *L. boggessi* (Decapoda: Caridea: Hippolytidae). *Journal of the Marine Biological Association of the United Kingdom*, 87: 517-522.
- Zhang, D. and Lin, J. (2005). Comparative mating success of smaller male-phase and larger male-role euhermaphrodite phase shrimp, *Lysmata wurdemani* (caridea: hippolytidae). *Marine Biology*, 147: 1387-1392.
- Zhang, D., Lin, J. and Creswell, R. (1997). Larviculture and effect of food on larval survival and development in golden coral shrimp *Stenopus scutellatus*. *Journal of Shellfish Research*, 16(2): 367-369.
- Zhang, D., Lin, J. and Creswell, R. L. (1998a). Effects of food and temperature on survival and development in the peppermint shrimp *Lysmata wurdemani*, *Journal of the World Aquaculture Society*. 29(4): 471-476.

- Zhang, D., Lin, J. and Creswell, R. L. (1998b). Mating behavior and spawning of the banded coral shrimp *Stenopus hispidus* in the laboratory. *Journal of Crustacean Biology*, 18: 511–518.
- Zhang, D., Lin, J. and Creswell, R. L. (1998c). Ingestion rate and feeding behavior of the peppermint shrimp *Lysmata wurdemanni* on artemia nauplii. *Journal of the World Aquaculture Society*, 29, 97–103.
- Zhang, D., Rhyne, A. L. and Lin, J. (2007). Density-dependent effect on reproductive behavior of *Lysmata amboinensis* and *L. bogessi* (decapoda: caridean: hippolytidae). *Journal of the Marine Biological Association of the United Kingdom*, 87: 517-522.
- Zhang, P., Zhang, X., Li, J. and Huang, G. (2006). The effects of body weight, temperature, salinity, pH, light intensity and feeding condition on lethal DO levels of whiteleg shrimp, *Litopenaeus vannamei* (Boone, 1931). *Aquaculture*, 256: 579–587.
- Zhao, Y., Zhao, Y. and Zeng, C. (2007). Morphogenesis and variations in biochemical composition of the eggs of *Macrobrachium nipponense* (de Haan, 1849) (decapoda, caridea, palaemonidae) during embryonic development. *Crustaceana*, 80(9): 1057-1070.