



**SYNTHESIS AND INVESTIGATION OF ANTI-TYROSINASE AND  
ANTIOXIDANT ACTIVITY OF NEW HALOGENATED XANTHONE  
DERIVATIVES**

**By**

**FATIN FARHANA BINTI BAHARUDDIN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Master of Science**

**December 2023**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment  
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**December 2023**

**Chair : Nadiah Mad Nasir, PhD**  
**Faculty : Science**

Excess melanin production in skin cells promotes dermatological diseases such as hyperpigmentation, ageing, and other skin-related diseases. Tyrosinase enzyme catalyses the oxidation of L-tyrosine substrate to melanin, via formation of L-3,4-dihydroxyphenylalanine (L-DOPA). Thus, tyrosinase inhibitors can lower melanin production to achieve whitening effect on the skin. Whitening products such as hydroquinone and kojic acid that are commonly used nowadays, might be harmful to human skin. This research aimed to produce 20 non-toxic halogenated tyrosinase inhibitors synthetically using Grover-Shah and Shah (GSS) method and determine their tyrosinase inhibitory activity using tyrosinase mushroom *Agaricus bisporus* assay with kojic acid as positive control. Six compounds were found to inhibit the enzyme with inhibition percentage of more than 50%. Among these six active compounds, three compounds (**15**, **16** and **17**) showed high potency. Their half-maximal inhibitory concentration ( $IC_{50}$ ), enzyme kinetic analysis and antioxidant properties were determined. The  $IC_{50}$  values for compounds **15** (7.8  $\mu\text{g/ml}$ , 75  $\mu\text{g/ml}$ ), **16** (9  $\mu\text{g/ml}$ ,

118  $\mu\text{g/ml}$ ) and **17** (10  $\mu\text{g/ml}$ , 100  $\mu\text{g/ml}$ ) in L-tyrosine and L-DOPA substrates respectively, demonstrate their strong action in comparison to kojic acid as control ( $\text{IC}_{50}$  4  $\mu\text{g/ml}$ , 7.8  $\mu\text{g/ml}$ ). Enzyme kinetic analysis demonstrated that compounds **15** and **17** as uncompetitive inhibitor and compound **16** as mixed type inhibitor respectively. In antioxidant assays, only compound **16** exhibited a high antioxidant activity ( $\text{IC}_{50}$  18  $\mu\text{g/ml}$ ) in DPPH assay and 2.93 mM/g in FRAP assay with ascorbic acid and quercetin as positive controls. Additionally, compound **16** revealed an  $\text{IC}_{50}$  value of 43.33  $\mu\text{M}$  against the *in vitro* HaCaT skin cell line, indicating a weak toxicity on human skin cells. Compound **16** also revealed a non-toxic property for screening of toxicity test when tested against the *in vitro* brine shrimp toxicity assay at a dose of 1000  $\mu\text{g/ml}$ . From the *in vivo* zebrafish embryo toxicity assay, results have shown that compound **16** was having a non-toxic property based on the parameters such as hatching rate, survival rate, heartbeat rate and obtained a  $\text{LC}_{50}$  value of 197.2  $\mu\text{g/mL}$ . Furthermore, the molecular docking analysis showed their binding interactions between mushroom tyrosinase protein structure (PDB ID: 2Y9X) for compound **16** with a binding affinity of (-7.7 kcal/mol) and the compounds as the ligands through in silico approach. These data suggest that potent halogenated xanthone derivatives have the potential to be developed as new candidates for skin whitening agents with antioxidant and non-toxic properties in cosmetic industries and for pigmentation-related diseases.

**Keywords:** Antioxidant, anti-tyrosinase, synthesis, toxicity, xanthone

**SDG:** GOAL 3: Good Health and Well-being, GOAL 4: Quality Education, GOAL 9: Industry, Innovation, and Infrastructure

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**SINTESIS DAN PENYIASATAN AKTIVITI ANTI-TIROSINASE DAN  
ANTIOKSIDAN BAGI TERBITAN XANTON BERHALOGEN BARU**

Oleh

**FATIN FARHANA BINTI BAHARUDDIN**

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Pengeluaran melanin yang berlebihan dalam sel kulit menggalakkan penyakit dermatologi seperti hiperpigmentasi, penuaan dan penyakit kulit lain yang berkaitan. Enzim tirosinase memungkinkan substrat L-tirosin melalui pengoksidaan, lalu membentuk L-3,4-dihidroksifenilalanin (L-DOPA) dan bertukar kepada melanin. Oleh itu, mendorong perencat tirosinase boleh mengurangkan pengeluaran melanin untuk mendapat sifat pemutihan pada kulit. Pada masa kini, beberapa produk pemutihan seperti hidrokuinon dan asid kojik mungkin berbahaya kepada kulit manusia. Penyelidikan ini bertujuan untuk menghasilkan 20 perencat tirosinase berhalogen tidak toksik secara sintetik dengan menggunakan kaedah Grover-Shah dan Shah (GSS) dan aktiviti perencatan tirosinasenya menggunakan ujian cendawan *Agaricus bisporus* tirosinase dengan asid kojik sebagai kawalan positif. Enam sebatian didapati merencat enzim tyrosinase dengan peratusan perencatan lebih daripada 50%. Di antara enam sebatian aktif ini, tiga sebatian (**15**, **16** dan **17**) menunjukkan potensi

yang lebih tinggi. Kepekatan perencatan separuh maksimum ( $IC_{50}$ ), analisis kinetik enzim dan sifat antioksidan telah ditentukan. Nilai  $IC_{50}$  untuk sebatian **15** (7.8  $\mu\text{g/ml}$ , 75  $\mu\text{g/ml}$ ), **16** (9  $\mu\text{g/ml}$ , 118  $\mu\text{g/ml}$ ) dan **17** (10  $\mu\text{g/ml}$ , 100  $\mu\text{g/ml}$ ) dalam substrat L-tirosin dan L-DOPA secara masing-masing menunjukkan tindakan penghalangan kuatnya dengan asid kojik sebagai kawalan ( $IC_{50}$  4  $\mu\text{g/ml}$ , 7.8  $\mu\text{g/ml}$ ). Analisis kinetik enzim menunjukkan bahawa sebatian **15** dan **17** sebagai perencat bukan kompetitif dan sebatian **16** sebagai perencat jenis campuran masing-masing. Dalam ujian antioksidan, hanya sebatian **16** yang menunjukkan aktiviti antioksidan yang tinggi ( $IC_{50}$  18  $\mu\text{g/ml}$ ) dalam ujian DPPH dan 2.93 mM/g dalam ujian FRAP dengan asid askorbik dan kuersetin sebagai kawalan positif. Selain itu, sebatian **16** mendedahkan nilai  $IC_{50}$  sebanyak 43.33  $\mu\text{M}$  terhadap ujian *in vitro* MTT sel kulit HaCaT yang menunjukkan ketoksikan yang lemah pada sel kulit manusia. Sebatian **16** juga mendedahkan sifat tidak toksik untuk saringan ujian ketoksikan *in vitro* apabila diuji terhadap ujian ketoksikan udang air garam pada dos 1000  $\mu\text{g/ml}$ . Daripada ujian ketoksikan embrio ikan zebra *in vivo* pula, keputusan telah menunjukkan bahawa sebatian **16** mempunyai sifat tidak toksik berdasarkan parameter-parameter seperti kadar penetasan, kadar kelangsungan hidup, kadar degupan jantung dan memperoleh nilai  $LC_{50}$  iaitu 197.2  $\mu\text{g/mL}$ . Tambahan pula, analisis dok molekul menunjukkan interaksi pengikatannya antara struktur protein cendawan tirosinase (ID PDB: 2Y9X) untuk sebatian **16** dengan nilai pengikatan (-7.7 kcal/mol) dan sebatian ini sebagai ligan melalui pendekatan kaedah secara dalam siliko. Data-data ini menunjukkan bahawa derivatif xanton berhalogen yang kuat berpotensi untuk dibangunkan sebagai calon baharu untuk agen pemutihan kulit dengan sifat antioksidan dan tidak toksik dalam industri kosmetik dan untuk penyakit berkaitan pigmentasi pada waktu akan datang.

**Kata Kunci:** Antioksidan, anti-tirosinas, ketoksikan, sintesis, xanton

**SDG:** GOAL 3: Good Health and Well-being, GOAL 4: Quality Education, GOAL 9: Industry, Innovation, and Infrastructure



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## LIST OF ABBREVIATIONS

$\gamma$	Gamma
$\alpha$	Alpha
$\delta$	Chemical shift in ppm
UV	Ultraviolet
$\mu\text{M}$	Micromolar
$\mu\text{g}$	Microgram
mmol	Milimolar
mL	Mililiter
mg	Miligram
$^{\circ}\text{C}$	degree celcius
%	percentage
$^{13}\text{C}$	carbon-13
$\text{C}_3\text{D}_6\text{O}$	deuterated acetone
AChE	Acetylcholinesterase
$\text{CDCl}_3$	deuterated chloroform
$\text{cm}^{-1}$	per centimeter
d	doublet
dd	doublet of doublet
DI-MS	Direct Injection-Mass Spectrometry
DMSO	dimethyl sulphoxide
DPPH	2,2-diphenylpicrylhydrazyl (DPPH) assay
FRAP	Ferric ion Reducing Antioxidant Power
FT-IR	Fourier Transform-Infrared spectroscopy
GC-MS	Gas Chromatograph-Mass Spectroscopy
$^1\text{H}$	proton

HaCaT	Normal keratinocyte skin cells
hpf	hours post fertilisation
Hz	hertz
IC <sub>50</sub>	Half-maximal inhibitory concentration
m	multiplet
m/z	Mass-to-charge ratio
CD <sub>3</sub> OD	deuterated methanol
t	triplet

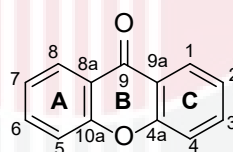


## CHAPTER 1

### INTRODUCTION

#### 1.1 General Introduction

Xanthenes are secondary metabolites found in a variety of plant families, fungi, and lichens (Vieira & Kijjoa, 2005). Xanthenes, with the IUPAC designation 9H-xanthene-9-one, are usually yellowish-colored heterocyclic compounds having the molecular formula  $C_{13}H_8O_2$ . The term “xanthone” is a combination of "xanth" is derived from the Greek word which means "yellow" and “one” which refers to the keto group in the structure. Heterocycle moieties play an important role in the design and development of new physiologically and pharmacologically active molecules. As illustrated in **Figure 1.1**, the xanthone scaffold has a dibenzo- $\gamma$ -pyrone scaffold formed by the coupling of two benzene rings and a pyrone-4-one ring and plays a key role in the development of novel medications with potential biological activities in the pharmaceutical industry.



**Figure 1.1: Xanthone scaffold**

Natural occurring xanthenes include various types of substituents at different positions, resulting in a wide range of pharmacological activities. Based on **Figure 1.1**, a carbonyl group and oxygen bridge linking two aromatic rings, giving the appearance of fused ring system. In terms of molecular symmetry, both C-1, C-8 and C-4, C-5 are acidic sites due to the withdrawing effect of the electronegative oxygen atoms (Odrowaz-Syrbiewski *et al.*, 2009). In contrast to that, the fused ring structure limits



the flexibility and thereby enhances the stability of the xanthone framework. Carbons 1-4 in the xanthone basic skeleton are allocated to acetate-derived ring C, whereas carbons 5-8 are assigned to shikimate-derived ring A. For structural elucidation, the remaining carbon atoms are designated as 4a, 5a, 8a, 9 and 9a (Pedro *et al.*, 2002).

## 1.2 Synthetic Approaches to Xanthone

With varied patterns of substitution, isolated xanthenes have been successfully extracted from famous natural resources such as *Calophyllum inophyllum* and *Garcinia mangostana*, leading to a wide range of compounds with biological significance. The concentration of xanthenes isolated from plants is low in yield, making it challenging to isolate xanthenes from their natural resources. As a result of this, xanthone synthesis has been further studied. Synthetic techniques can be used to create new xanthenes with varied substituent positions and types on the fundamental building block. Synthetic methods can be divided into two methods, via biosynthesis where enzymatic reaction is used to produce numerous xanthone derivatives and chemical synthesis where catalytic reactions are performed in the laboratory.

## 1.3 Biological Activities of Xanthone

Xanthone plays an important role in biological active heterocycles which have wide potential in pharmacological field in recent years. Hence, many researchers tend to focus in synthesizing various xanthone derivatives for the development of new lead drug.

Xanthone derivatives are mainly shown in producing as many types of biological activities reported whereby their derivatives can interact with several different

medicinal targets such as antioxidant, anticancer and antibacterial (Li *et al.*, 2011). Researchers reported that xanthenes which bearing aromatic benzene rings in the structure can produce bioactivities due to the aromatic rings are able imitate the behavior of most inhibitors on the respective bioactivities.

Studies have been conducted showing that the xanthone derivatives possesses biological activities such as antityrosinase (Rosa *et al.*, 2021), anticancer (Pedro *et al.*, 2002), antioxidant (Wairata *et al.*, 2021), antitumor (Luo *et al.*, 2013) and anti-melanogenesis activities (Hosseinpour *et al.*, 2021) and more.

Alzheimer's disease is commonly referred to as an illness with multiple factors due to genetic, environmental, and endogenous aspects. Cholinergic neuronal transmission loss and oxidative stress have been recognised as important markers of this Alzheimer's disease. In 2017, it was stated that xanthone derivatives have the potential to be used as dual agents in the treatment of Alzheimer's disease, alongside acetylcholinesterase inhibitors and antioxidants (Cidade *et al.*, 2017).

Melanin is the most abundant natural biological polymer pigment in skin and hair (Lam *et al.*, 2010). However, excessive formation of melanin can result in dermatological conditions such as freckles, melasma, and lentigo (Kim *et al.*, 2008). Although the mechanism of melanogenesis is complex, it has been widely reported that the tyrosinase enzyme may regulate the biosynthetic pathway for the formation of melanin (Gartner *et al.*, 2012).

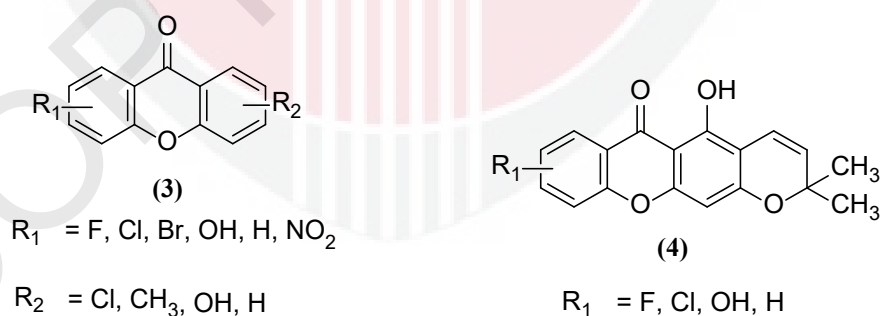
In recent years, Rosa and her colleagues managed to discover that xanthone derivatives are able to inhibit tyrosinase enzyme with a potency in their discovery as a whitening agent (Rosa *et al.*, 2021). They also reported that most of their synthesized xanthone derivatives are showing a result as tyrosinase inhibitors with the best results that beats approximately six times lower than the IC<sub>50</sub> value from the positive control, kojic acid.

Previous non-halogenated xanthone derivatives are lacking to be reported in having anti-tyrosinase properties and recently, a study done by Rosa and colleagues in 2021 managed to discover that a halogenated xanthone possesses an antityrosinase activity which can be potential as a skin whitening agent in future. Also, based on our knowledge, prenylated xanthenes are yet to be reported to possess an antityrosinase activity too. Therefore, the purpose of this work was to synthesize xanthone scaffolds with halogens, then inducing a cyclic ring into the scaffold, and subsequently with the evaluation of their biological activities, on tyrosinase enzyme in two pathways: monophenolase (L-tyrosine) and diphenolase (L-3,4-dihydroxyphenylalanine; L-DOPA).

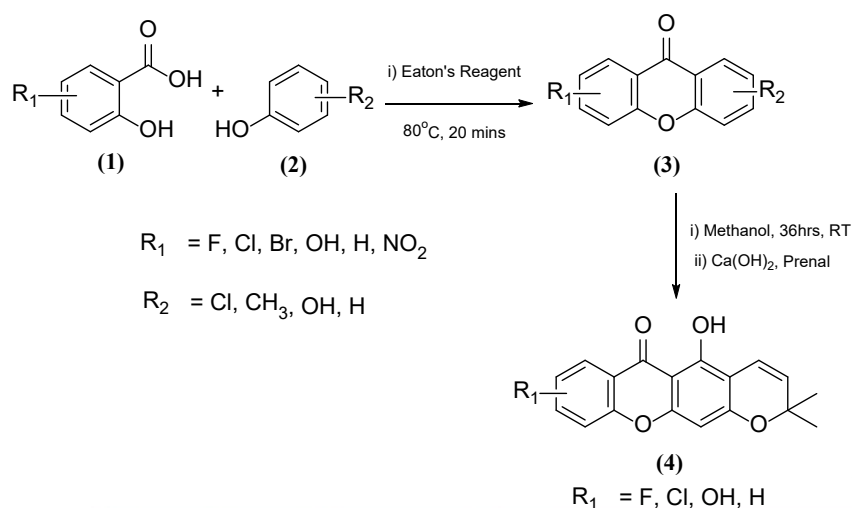
Halogenation of compounds are a common approach in drug discovery and a significant number of compounds in clinical development are halogenated (Lu *et al.*, 2010; Hernandez *et al.*, 2010). Halogen atoms are involved in protein-ligand noncovalent interactions, such as formation of hydrogen bonds and halogen bonds in the enzyme activity (Hernandez *et al.*, 2010). Introducing halogen atoms may also contribute to steric effects, through the ability of these bulky atoms to occupy the binding site of molecular targets (Hernandez *et al.*, 2010).

Most skin whitening agents in the cosmetic industry nowadays are found to be using toxic ingredients such as hydroquinone and mercury. Although these ingredients can exhibit a high whitening effects, they produce an adverse effect such as inflammation, irritation and blisters towards the users (Owolabi *et al.*, 2020). Thus, in our study, we would like to develop a new skin whitening agent with non-toxic properties that are beneficial for the cosmetic industries.

Furthermore, the structure-activity relationships (SARs) on selected xanthenes that demonstrates higher activity *in vitro* test will be investigated. Our hypothesis is to create a series of xanthone derivatives by modifying the basic scaffold which are predicted to be a promising lead as whitening agents for pharmaceuticals purposes based on the reported study which shows the activity in helping skin related diseases (Rosa *et al.*, 2021). **Figure 1.2** and **Scheme 1.1** illustrates the reaction scheme as well as the general structures of the targeted compounds. Meanwhile, **Table 1.1** lists the structures of all twenty compounds, including ten new compounds.



**Figure 1.2: General structure of targeted compounds**



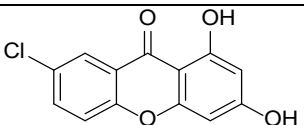
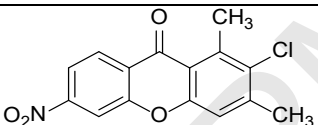
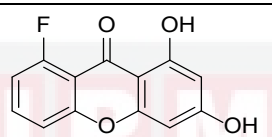
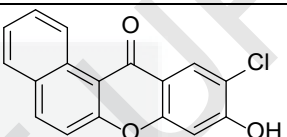
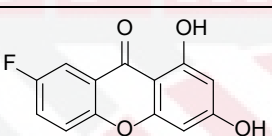
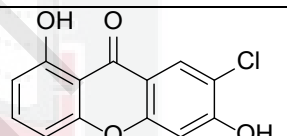
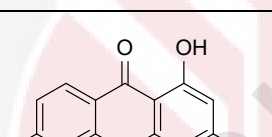
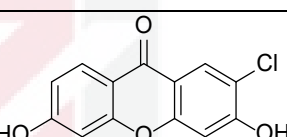
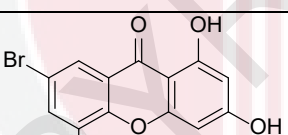
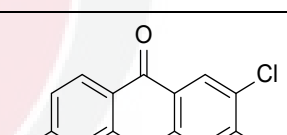
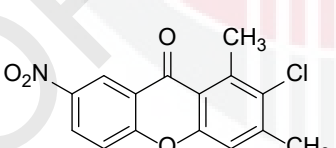
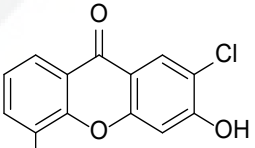
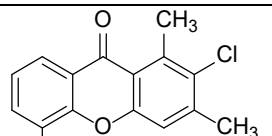
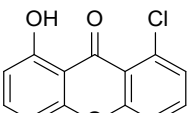
**Scheme 1.1: General scheme for synthesis of halogenated xanthenes (3) and prenylated xanthenes (4)**

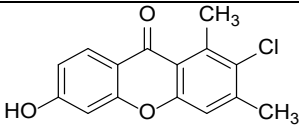
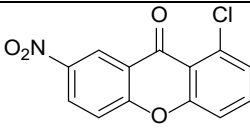
In this study, we chose halogenated xanthone such as fluorine, chlorine and bromine as different substituents and prenylated xanthone derivatives to discover their potential in inhibiting the tyrosinase enzyme as previous studies also have supported that halogenated xanthone produces this inhibitory effect. For the prenylated derivatives, studies are yet to be reported in this tyrosinase inhibitory, thus, we would like to initiate in discovering their results which can be added into the xanthonoic derivatives of library among the other researchers for future.

Most previous studies reported that compounds that bears an aromatic ring moiety shows a potency in most biological inhibitory properties (Rosa *et al.*, 2021; Zhu *et al.*, 2013; Haldys *et al.*, 2020). Moreover, aromatic rings structurally mimic the tyrosinase substrates which are L-tyrosine and L-3,4-dihydroxyphenylalanine (L-DOPA) that brings an interaction with the hydrophobic tyrosinase enzyme site via van der Waals interactions (Zhu *et al.*, 2013). Therefore, xanthone scaffold possesses a great potential

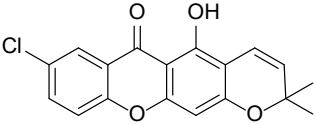
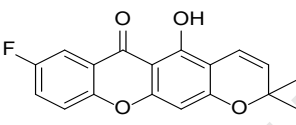
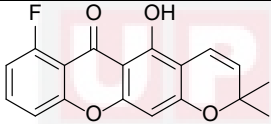
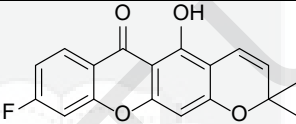
in holding as a tyrosinase inhibitor that are potential for skin whitening agent due to the presence of aromatic ring.

**Table 1.1: Structure of synthesized xanthone derivatives**

Halogenated xanthenes			
5		13*	
6		14*	
7		15*	
8		16*	
9		17*	
10*		18*	
11*		19	

12*		20*	
-----	---	-----	--

**Table 1.1: Continued**

Prenylated xanthones			
21		23	
22		24	

\*Indicates new compounds

## 1.4 Problem Statement

Nowadays, in the modern world, a lot of UV radiations are emitted from environments such as natural sunlight and man-made UV rays from the screen of technologies such as smartphones, television and more. The main types of UV rays that are able to affect the skin are UVA rays and UVB rays. People who tend to be more exposed towards the UV rays are at greater risk for skin cancer, melasma, freckles, and age spots which can accelerate skin aging by several years in difference. In addition, continuous exposure of UV rays may speed up the formation of melanin in the skin, DNA damage, gene mutation, impairment of immune system, hyperpigmentation and photoaging (Kim *et al.*, 2021; cancer.org, 2023). To achieve a skin whitening effect, tyrosinase inhibitors are introduced in any cosmetic products. Unfortunately, due to the high toxicity of the most known tyrosinase inhibitor such as hydroquinone, kojic acid and

mercury (Burnett *et al.*, 2010; Owolabi *et al.*, 2020), there is a need in discovering new tyrosinase inhibitors which are safe and non-toxic for the users.

Most isolated xanthenes from natural products will have a low yield with a limited type of substituents in the structures, therefore, a synthetic approach in producing xanthenes with various positions of substituents leads to new possibilities in discovering new areas of their biological activities with diverse of patterns in substituents.

A previous study done by Rosa and her colleagues in 2021 managed to discover that xanthone derivatives are found to be exhibiting an antityrosinase effect based on the mushroom tyrosinase enzyme study. However, up to these recent years, a small number of studies were reported on halogenated xanthone derivatives as tyrosinase inhibitors. Thus, our study decided in producing varieties of halogenated xanthone derivatives to investigate their tyrosinase inhibitory effect which can also be beneficial among other researchers in tyrosinase inhibitors studies. For prenylated derivatives, our group managed to discover its anticancer bioactivity which were tested on a breast cancer cells and very lacking studies were done for prenylated derivatives on tyrosinase inhibition study. Therefore, our group took this new approach in discovering the tyrosinase inhibitory effects by prenylated derivatives.

Thus, we are aiming in producing a variety of halogenated xanthenes and prenylated xanthone derivatives with different designated functional groups on the xanthone scaffold were developed and synthesized to promote their bioactivity in inhibition of tyrosinase enzyme with their antioxidant characteristics. Later, the toxicity of the



potent compounds will be examined, and their SARs will be clarified in order to understand the mechanisms of xanthone interaction with the mushroom tyrosinase enzyme. (Rosa *et al.*, 2021; Zhou *et al.*, 2019).

### 1.5 Objectives of Study

- To synthesize and characterize a series of halogenated and prenylated xanthenes via spectroscopic methods such as Nuclear Magnetic Resonance (NMR), Fourier Transform Infrared (FTIR) and Gas Chromatography Mass Spectrometry (GCMS).
- To evaluate the *in vitro* inhibitory activity on mushroom tyrosinase enzyme and antioxidant activity of the synthesized compounds.
- To determine the binding mechanisms of the potent compounds (bioactive) via enzyme kinetic study analysis supported by *in silico* molecular docking analysis.
- To determine the cytotoxicity properties of the potent compounds on normal keratinocyte skin (HaCaT) cell line, brine shrimp assay and zebrafish embryo toxicity test.

## REFERENCES

- Abu, N., Zamberi, N. R., Yeap, S. K., Nordin, N., Mohamad, N. E., Romli, M. F., Rasol, N. E., Subramani, T., Ismail, N. H., Alitheen, N. B. (2018). Subchronic toxicity, immunoregulation and anti-breast tumor effect of Nordamnacantal, an anthraquinone extracted from the stems of *Morinda citrifolia* L. *BMC complementary and alternative medicine*, 18: 1-10.
- Alasalvar, C., Al-Farsi, M., Quantick, P. C., Shahidi, F., Wiktorowicz, R. (2005). Effect of chill storage and modified atmosphere packaging (MAP) on antioxidant activity, anthocyanins, carotenoids, phenolics and sensory quality of ready-to-eat shredded orange and purple carrots. *Food Chemistry*, 89: 69-76.
- Athipornchai, A., Niyomtham, N., Pabuprapap, W., Ajavakom, V., Duca, M., Azoulay, S., Suksamrarn, A. (2021). Potent tyrosinase inhibitory activity of curcuminoid analogues and inhibition kinetics studies. *Cosmetics*, 8: 35.
- Bagherzadeh, K., Shirgahi Talari, F., Sharifi, A., Ganjali, M. R., Saboury, A. A., Amanlou, M. (2015). A new insight into mushroom tyrosinase inhibitors: docking, pharmacophore-based virtual screening and molecular modeling studies, *Journal of Biomolecular Structure and Dynamics*, 33: 487-501.
- Baharuddin, F. F., Nasir, N. M., Tejo, B. A., Koh, S. P., Ramakrishnan, S., Ahmad Nordin, N. Q. A., Adzahar, A. N., Devakrishnan, P., Mohd Razib, S. (2023). Potent halogenated xanthone derivatives: synthesis, molecular docking, and study on antityrosinase activity, *Journal of Asian Natural Products Research*.
- Bakar, S. A., Ali, A. M., Ahmad, N. H. (2019). Differential antiproliferative activity of Goniothalamine against selected human cancer cell lines. *Malaysian Journal of Medicine and Health Sciences*, 15: 66-73.
- Burnett, C. L., Bergfeld, W. F., Belsito, D. V., Hill, R. A., Klaassen, C. D., Liebler, D. C., Marks, J. G., Shank, R. C., Slaga, T. J., Snyder, P. W. (2010). Final report of the safety assessment of kojic acid as used in cosmetics. *International journal of toxicology*, 29(6): 244S-273S.
- Chantarasriwong, O., Milcarek, A. T., Morales, T. H., Settle, A. L., Rezende, C. O., Althufairi, B.D., Theodoraki, M. A., Alpaugh, M. L., & Theodorakis, E. A. (2019). Synthesis, structure-activity relationship and in vitro pharmacodynamics of A-ring modified caged xanthenes in a preclinical model of inflammatory breast cancer. *European Journal of Medicinal Chemistry*, 168: 405–413.
- Chao, W. W., Su, C. C., Peng, H. Y., Chou, S. T. (2017). *Melaleuca quinquenervia* essential oil inhibits  $\alpha$ -melanocyte-stimulating hormone-induced melanin production and oxidative stress in B16 melanoma cells. *Phytomedicine*, 34: 191-201.
- Cidade, H., Cruz, I., Puthongking, P., Cravo, S., Palmeira, A., Pinto, M., Sousa, E. (2017). Xanthone and flavone derivatives as dual agents with acetylcholinesterase

inhibition and antioxidant activity as potential anti-alzheimer agents. *Journal of Chemistry*.

Cidade, H., Rocha, V., Palmeira, A., Marques, C., Tiritan, M. E., Ferreira, H., Lobo, J. S., Almeida, I. F. Sousa, M. E., Pinto, M. (2020). In silico and in vitro antioxidant and cytotoxicity evaluation of oxygenated xanthone derivatives. *Arabian Journal of Chemistry*, 13: 17-26.

Decker, H., Tuczec, F. (2000). Tyrosinase/catecholoxidase activity of hemocyanins: structural basis and molecular mechanism. *Trends in biochemical sciences*, 25: 392-397.

Dodean, R. A., Kelly, J. X., Peyton, D., Gard, G. L., Riscoe, M. K., & Winter, R. W. (2008). Synthesis and heme-binding correlation with antimalarial activity of 3,6-bis-( $\omega$ -N, N-diethylaminoamyoxy)-4,5-difluoroxanthone. *Bioorganic and Medicinal Chemistry*, 16(3): 1174–1183.

Dubinska-Magiera, M., Daczewska, M., Lewicka, A., Migocka-Patrzalek, M., Niedbalska-Tarnowska, J., Jagla., K. (2016). Zebrafish: a model for the study of toxicants affecting muscle development and function. *International Journal of Molecular Sciences*, 17: 1941.

Eaton, P. E., Carlson, G. R., Lee, J. T. (1973). Phosphorus pentoxide-methanesulfonic acid. Convenient alternative to polyphosphoric acid. *The Journal of Organic Chemistry*, 38: 4071-4073.

Flieger, J. (2000). ALKALOIDS | Thin-Layer (Planar) Chromatography. *Encyclopedia of Separation Science*.

Friedman M. (1996). Food browning and its prevention: an overview. *Journal of Agricultural and Food Chemistry*, 44: 631-653.

Gartner L. P., Hiatt, J. L. (2012). *Color atlas and text of histology*, Lippincott Williams & Wilkins.

Gohari, A. R., Hajimehdipoor, H., Saeidnia, S., Ajani, Y., Hadjiakhoondi, A. (2011). Antioxidant activity of some medicinal species using FRAP assay. *Journal of Medicinal Plants*, 0: 54-60.

Grover, P. K., Shah, G. D., & Shah, R. C. (1955). Xanthones. Part IV. A new synthesis of hydroxyxanthones and hydroxybenzophenones. *Journal of Chemistry Society*, (0): 3982–3985.

Haldys, K., Goldeman, W., Jewginski, M., Wolinska, E., Anger-Gora, N., Rossowska, J., Latajka, R. (2020). Halogenated aromatic thiosemicarbazones as potent inhibitors of tyrosinase and melanogenesis. *Bioorganic Chemistry*, 94: 103419.

Hashim, N. M., Rahmani, M., Ee, G. C. L., Sukari, M. A., Yahayu, M., Amin, M. A. M., Ali, A. M., Go, R. (2012). Antioxidant, antimicrobial and tyrosinase inhibitory

activities of xanthenes isolated from *Artocarpus obtusus* FM Jarrett. *Molecules*, 17: 6071-6082.

Hedayati, A., Shaluei, F., Jahanbakhshi, A., Kolangi, H., Alizadeh, M. (2017). Detection of LC50, NOEC, and LOEC of some heavy metals (mercury, plumb and zinc) in freshwater fish Roach (*Rutilus rutilus*). *Toxicology & Industrial Health*, 33: NP4-NP10.

Hernandes, M. Z., Cavalcanti, S. M. T., Moreira, D. R. M., de-Azevedo Junior, W. F., Leite, A. C. L. (2010). Halogen atoms in the modern medicinal chemistry: hints for the drug design. *Current drug targets*, 11(3): 303-314.

Hosseinpour, H., Farid, S. M., Iraj, A., Asgari, M. S., Edraki, N., Hosseini, S., Jamshidzadeh, A., Larijani, B., Attarroshan, M., Pirhadi, S. (2021). Anti-melanogenesis and anti-tyrosinase properties of aryl-substituted acetamides of phenoxy methyl triazole conjugated with thiosemicarbazide: Design, synthesis and biological evaluations. *Bioorganic Chemistry*, 114: 104979.

Hruschka, S., Yoshida, S., Kirk, K. L., Haufe, G. (2008). Fluorinated phenylcyclopropylamines: Part 6. Effects of electron withdrawing or donating aryl substituents on the inhibition of tyramine oxidase from *Arthrobacter* sp. by diastereomeric 2-aryl-2-fluoro-cyclopropylamines. *Journal of Fluorine Chemistry*, 129: 875-800.

<https://www.cancer.org/cancer/types/skin-cancer.html>

Hu, Y. H., Chen, Q. X., Cui, Y., Gao, H. J., Xu, L., Yu, X. Y., Wang, Y., Yang, C. L., Wang, Q. (2016). 4-Hydroxy cinnamic acid as mushroom preservation: anti-tyrosinase activity kinetics and application. *International Journal of biological macromolecules*, 86: 489-495.

Hyun, S. K., Lee, W. H., Jeong, D. M., Kim, Y., Choi, J. S. (2008). Inhibitory effects of kurarinol, kuraridinol, and trifolirhizin from *Sophora flavescens* on tyrosinase and melanin synthesis. *Biological and Pharmaceutical Bulletin*, 31: 154-158.

Jimenez-Atienzar M., Escribano, J., Cabanes, J., Gandia-Herrero, F., Garcia-Carmona, F. (2005). Oxidation of the flavonoid eriodictyol by tyrosinase. *Plant Physiology and Biochemistry*, 43: 866-873.

Khammee, T., Athipornchai, A., Upamai, W., Jaisin, Y., & Suksamrarn, S. (2014). Synthesis of Hydroxyxanthenes and Evaluations for their Acetylcholinesterase Inhibitory and Neurotoxicity Activities. *KKU Science Journal*, 42: 212–220.

Kim, H. M., Oh, S., Yang, J. Y., Sun, H. J., Jang, M., Kang, D., Son, K. K., Byun, K. (2021). Evaluating whether radiofrequency irradiation attenuated UV-B-induced skin pigmentation by increasing melanosomal autophagy and decreasing melanin synthesis. *International Journal of Molecular Sciences*, 22: 10724.

- Kim, Y. J., Kang, K. S., Yokozawa, T. (2008). The anti-melanogenic effect of pycnogenol by its anti-oxidative actions. *Food and chemical toxicology*, 46: 2466-2471.
- Kou, X., Song, L., Wang, Y., Yu, Q., Ju, H., Yang, A., & Shen, R. (2019). Design, synthesis and anti-Alzheimer's disease activity study of xanthone derivatives based on multi-target strategy. *Bioorganic & Medicinal Chemistry Letters*, 126927.
- Lam, R. Y. Y., Lin, Z. X., Sviderskaya, E., Cheng, C. H. K. (2010). Application of a combined sulphorhodamine B and melanin assay to the evaluation of Chinese medicines and their constituent compounds for hyperpigmentation treatment. *Journal of Ethnopharmacology*, 132: 274-279.
- Li, G. L., He, J. Y., Zhang, A., Wan, Y., Wang, B., Chen, W. H. (2011). Toward potent  $\alpha$ -glucosidase inhibitors based on xanthenes: A closer look into the structure–activity correlations. *European Journal of Medicinal Chemistry*, 46(9): 4050-4055.
- Liang, X., Huang, Z. H., Ma, X., Zheng, Z. H., Lu, X. H., Qi, S. H. (2021). Mycotoxins as inhibitors of protein tyrosine phosphatases from deep-sea-derived fungus *Aspergillus puniceus* SCSIOz21. *Bioorganic Chemistry*, 107: 104571.
- Luo, L., Qin, J. K., Dai, Z. K., & Gao, S. H. (2013). Synthesis and biological evaluation of novel benzo[b]xanthone derivatives as potential antitumor agents. *Journal of the Serbian Chemical Society*, 78(9): 1301–1308.
- Mahabusarakam, W., Nuangnaowarat, W., Taylor, W.C. (2006). Xanthone derivatives from *Cratoxylum cochinchinense* roots. *Phytochemistry*, 67: 470-74.
- Matoba, Y., Kumagai, T., Yamamoto, A., Yoshitsu, H., Sugiyama, M. (2006). Crystallographic evidence that the dinuclear copper center of tyrosinase is flexible during catalysis. *Journal of Biological Chemistry*, 281: 8981-8990.
- Menéndez, C. A., Biscussi, B., Accordino, S., Paula Murray, A., Gerbino, D. C., Appignanesi, G. A. (2017). Design, synthesis and biological evaluation of 1,3-dihydroxyxanthone derivatives: Effective agents against acetylcholinesterase. *Bioorganic Chemistry*, 75: 201–209.
- Merza, J., Aumond, M-C., Rondeau, D., Dumontet, V., Ray, A. M. L., Seraphin, D., Richomme, P. (2004). Prenylated xanthenes and tocotrienols from *Garcinia virgata*. *Phytochemistry*, 65: 2915-20.
- Meyer, B. N., Ferrigni, N. R., Putnam, J. E., Jacobsen, L. B., Nichols, D. E. J., McLaughlin, J. L. (1982). Brine shrimp: a convenient general bioassay for active plant constituents. *Planta medica*. 45: 31-34.
- Miguel, C., Azevedo, G., Manuel, C., Afonso, M., Maria, M., & Pinto, M. (2012). Routes to Xanthenes: An Update on the Synthetic Approaches. *Current Organic Chemistry*, 16(23): 2818–2867.



Mine, M., Mizuguchi, H., Takayanagi, T. (2022). Kinetic analyses of two-steps oxidation from l-tyrosine to l-dopaquinone with tyrosinase by capillary electrophoresis/dynamic frontal analysis. *Analytical Biochemistry*, 655: 114856.

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Mondal, M., Puranik, V. G., & Argade, N. P. (2006). Facile Synthesis of 1,3,7-Trihydroxyxanthone and Its Regioselective Coupling Reactions with Prenal: Simple and Efficient Access to Osajaxanthone and Nigrolineaxanthone F. *Journal of Organic Chemistry*, 71(2): 4992–4995.

Moreau, S., Varache-Lembège, M., Larroure, S., Fall, D., Neveu, A., Deffieux, G., Nuhrich, A. (2002). (2-Arylhyaazonomethyl)-substituted xanthenes as antimycotics: Synthesis and fungistatic activity against *Candida* species. *European Journal of Medicinal Chemistry*, 37: 237–253.

Odrowaz-Sypniewski, M. R., Tsoungas, P. G., Varvounis, G., & Cordopatis, P. (2009). Xanthone In Synthesis: A Reactivity Profile Via Directed Lithiation of its Dimethyl Ketal. *Tetrahedron Letters*, 50(44): 5981–5983.

Osman, M. S., Shantier, S. W., Awad, T. A., Garelnabi, E. A. E. (2022). In vitro evaluation, in silico studies and toxicological assay of some xanthenes as potential *Leishmania donovani* inhibitors. *Informatics in Medicine Unlocked*. 30: 100954.

Owolabi, J. O., Fabiyi, O. S., Adelakin, L. A., Ekwerike, M. C. (2020). Effects of skin lightening cream agents–hydroquinone and kojic acid, on the skin of adult female experimental rats. *Clinical, Cosmetic and Investigational Dermatology*, (0): 283–289.

Panda, S. S., Chand, M., Sakhuja, R., Jain, S. C. (2013). Xanthenes as potential antioxidants. *Current Medicinal Chemistry*, 20: 4491–4507.

Pedro, M., Cerqueira, F., Sousa, M. E., Nascimento, M. S. J., & Pinto, M. (2002). Xanthenes as inhibitors of growth of human cancer cell lines and their effects on the proliferation of human lymphocytes in vitro. *Bioorganic and Medicinal Chemistry*, 10(12): 3725–3730.

Pillai, K. M., Naiksatam, P., Johnson, F., & Rajagopalan, R. (1986). Thermorubin 11: 1,3-Dihydroxy-9H-xanthenes and 1,3-Dihydroxy-9H-xanthenes. *New Methods of Synthesis*, 14: 717–723.

Pinto, M. M. M., Palmeira, A., Fernandes, C., Resende, D. I. S. P., Sousa, E., Cidade, H., Tiritan, M. E., Correia-da-Silva, M., Cravo, S. (2021). From natural products to new synthetic small molecules: A journey through the world of xanthenes. *Molecules*, 26: (2).

Quillinan, A. J., & Scheinmann, F. (1973). Studies in the xanthone series. Part XII. A general synthesis of polyoxygenated xanthenes from benzophenone precursors. *Journal of Chemistry Society Perkin Transactions 1*, (0): 1329–1337.

- Ramakrishnan, S., Nasir, N. M., Stanslas, J., Hamdi, A. I. F., Latif, M. A. M., Baharuddin, F. F. (2023). One-pot two-component synthesis of halogenated xanthone, 3-o substituted xanthone, and prenylated xanthone derivatives as aromatase inhibitors. *Results in Chemistry*, 5: 100789.
- Reutrakul, V., Anantachoke, N., Pohmakotr, M., Jaipetch, T., Sophasan, S., Yoosook, C., Kasisit, J., Napaswat, C., Santisuk, T., Tuchinda, P. (2007). Cytotoxic and anti-HIV-1 caged xanthenes from the resin and fruits of *Garcinia hanburyi*. *Planta Medicine*, 73: 33-40.
- Roberts, J. C. (1961). Naturally Occurring Xanthenes. *Chemical Reviews*, 6: 591–605.
- Rosa, G.P., Palmeira, A., Almeida, I. F., Kane-Pagés, A., Barreto, M. C., Sousa, E., Pinto, M. M. M. (2021). Xanthenes for melanogenesis inhibition: Molecular docking and QSAR studies to understand their anti-tyrosinase activity, *Bioorganic & Medicinal Chemistry*, 29: 115873.
- Saeed, A., Mahesar, P. A., Channar, P. A., Abbas, Q., Larik, F. A., Hassan, M., Raza, H., Seo, S. Y. (2017). Synthesis, molecular docking studies of coumarinyl-pyrazolinyl substituted thiazoles as non-competitive inhibitors of mushroom tyrosinase. *Bioorganic Chemistry*, 74: 187-196.
- Sanchez-Ferrer, A., Rodriguez-Lopez, J. N., Garcia-Canovas, F., Garcia-Carmona, F. (1995). Tyrosinase: A comprehensive review of its mechanism. *Biochimica et Biophysica Acta (BBA)-Protein Structure and Molecular Enzymology*, 1247: 1-11.
- Sheng, Z., Ge, S., Xu, P., Li, C., Zhao, D., Tang, X. (2018). Design, synthesis and evaluation of cinnamic acid ester derivatives as mushroom tyrosinase inhibitors. *Medicinal Chemistry Communications*, 9: 853-861.
- Shimizu, K., Kondo, R., Sakai, K. (2000). Inhibition of tyrosinase by flavonoids, stilbenes and related 4-substituted resorcinols: structure-activity investigations. *Planta medica*. 66: 11-15.
- Sousa, M. E., & Pinto, M. M. M. (2005). Synthesis of Xanthenes: An Overview. In *Current Medicinal Chemistry*, 12: 2517-2538.
- Sztanke, M., Rzymowska, J., Sztanke, K. (2021). Anticancer active trifluoromethylated fused triazinones are safe for early-life stages of zebrafish (*Danio rerio*) and reveal a proapoptotic action. *Journal of Enzyme Inhibition and Medicinal Chemistry*. 36: 336-344.
- Thiagarajan, S. K., Rama-Krishnan, K., Husna Shafie, N., Arapoc, D. J., Bahari, H. (2019). Evaluation of the effect of aqueous *Momordica charantia* Linn. extract on zebrafish embryo model through acute toxicity assay assessment. *Evidence-based complementary and alternative medicine*.
- Varache-Lembège, M., Moreau, S., Larroure, S., Montaudon, D., Robert, J., & Nuhrich, A. (2008). Synthesis and antiproliferative activity of aryl- and

heteroaryl-hydrazones derived from xanthone carbaldehydes. *European Journal of Medicinal Chemistry*, 43(6): 1336–1343.

- Veshalini, K., Arapoc, D. J., Adam, Z., Razali, R., Suliman, N. A., Bakar, N. A. A. (2022). Phytochemical Screening, In vitro Antioxidant Activities and Zebrafish Embryotoxicity of *Abelmoschus esculentus* Extracts. *Pharmacognosy Journal*. 14: 3.
- Vieira, L. M. M., & Kijjoa, A. (2005). Naturally-Occurring Xanthenes: Recent Developments. *Current Medicinal Chemistry*, 12: 2413–2446.
- Wairata, J., Sukandar, E. R., Fadlan, A., Purnomo, A. S., Taher, M., Ersam, T. (2021). Evaluation of the Antioxidant, Antidiabetic, and Antiplatelet Activities of Xanthenes Isolated from *Garcinia forbesii* and their in-silico studies. *Biomedicines*, 9: 1380.
- Wu, J., Dai, J., Huang, L., Ding, H., Mao, J., & Yu, S. (2019). Synthesis of Novel Xanthone Analogues and Their Growth Inhibitory Activity Against Human Lung Cancer A549 Cells. 4239–4246.
- Xie, W., Zhang, H., He, J., Zhang, J., Yu, Q., Luo, C., Li, S. (2017). Synthesis and biological evaluation of novel hydroxybenzaldehyde-based kojic acid analogues as inhibitors of mushroom tyrosinase. *Bioorganic & Medicinal Chemistry Letters*, 27: 530-532.
- Yi, W., Cao, R., Peng, W., Wen, H., Yan, Q., Zhou, B., Ma, L., Song, H. (2010). Synthesis and biological evaluation of novel 4-hydroxybenzaldehyde derivatives as tyrosinase inhibitors. *European Journal of Medicinal Chemistry*, 45: 639-646.
- Yimdjo, M.C., Azebaze, A.G., Nkengfack, A.E., Meyer, A.M., Bodo, B., Fomum, Z. T. (2004). Antimicrobial and cytotoxic agents from *Calophyllum inophyllum*. *Phytochemistry*, 65: 2789-95.
- Zhao, Z., Liu, G., Meng Y., Tian, J., Chen, X., Shen, M., Li, Y., Li, B., Gao, C., Wu S. (2019). Synthesis and anti-tyrosinase mechanism of the substituted vanillyl cinnamate analogues. *Bioorganic Chemistry*, 93: 103316.
- Zhou, B., Wang, X., Weng, Z., Huang, B., Ma, Z., Yu, B., Ruan, L., & Hu, D. (2019). Synthesis and antitumor, antityrosinase, and antiplatelet aggregation activities of xanthone. *Journal of Chinese Pharmaceutical Sciences*, 28(4): 247–256.
- Zhu, T. H., Yu, Y. Y., Cao, S.W. (2013). Tyrosinase inhibitory effects and antioxidant properties of paeonol and its analogues. *Food Science and Technology Research*, 19(4): 609-615.