



**EVALUATION OF ANTIMICROBIAL ACTIVITY OF *Aegle marmelos* (L.)
Corrêa AND *Cassia alata* L. AGAINSTS PATHOGENIC MICROORGANISMS**

By

MANIVANNAN A/L VEYAKANDAH

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

December 2022

FS 2022 69

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**EVALUATION OF ANTIMICROBIAL ACTIVITY OF *Aegle marmelos* (L.)
Corrêa AND *Cassia alata* L. AGAINSTS PATHOGENIC MICROORGANISMS**

By

MANIVANNAN A/L VEYAKANDAH

December 2022

Chair : Associate Professor Nur Ain Izzati binti Mohd Zainudin, PhD
Faculty : Science

Traditional medicine makes use of several plant species, including *Aegle marmelos* and *Cassia alata*, to cure a wide range of illnesses. The introduction of novel antibiotic-resistant bacteria and the spread of existing ones are major problems for public health around the world. The primary reason for conducting this research is to determine the efficacy of aqueous and methanol extracts of unripe fruit, *A. marmelos* leaves, and *C. alata* leaves against various microorganisms. Cold methanol and water extraction was used to extract the leaves and unripe fruit of *A. marmelos* and *C. alata*. Pathogenic bacteria (*Streptococcus agalactiae*, *Escherichia coli*, and *Pseudomonas aeruginosa*) and yeasts (*Candida krusei* and *Candida parapsilosis*) were tested for their susceptibility to the plant extracts using disc diffusion and agar well diffusion methods. Microdilution was used to establish the MIC and MFC, or minimal inhibitory and minimum fungicidal concentrations, respectively. Scanning electron microscopy (SEM) was utilised to examine the cellular modification induced by active plant extracts. The antifungal activity of the methanol extract of the fruit leaves of *A. marmelos* and *C. alata* was much higher than that of the water extract, which was ineffective against both *C. parapsilosis* and *C. krusei*. In contrast, the disc diffusion method did not detect any antibacterial activity from the methanol or aqueous extract. The minimum inhibitory concentration (MIC) for both *A. marmelos* leaf and *C. alata* unripe fruit against *C. krusei* and *C. parapsilosis* was found to be 75 mg/ml. However, the MIC for *C. parapsilosis* was reported to be 37.5 mg/ml. The MFC results for *C. krusei* and *C. parapsilosis* on *A. marmelos* leaf and unripe fruit and *C. alata* leaf all indicated 100% mortality at a concentration of 150 mg/ml. Fungistatic activity was demonstrated by the methanolic extract's minimal inhibitory and minimal fungicidal concentrations, respectively. The fungicidal and fungistatic properties of *C. alata* and *A. marmelos* leaves were demonstrated against *C. parapsilosis* and *C. krusei*. Under a scanning electron microscope (SEM), the effects of *A. marmelos* and *C. alata* methanol extract on *C. parapsilosis* and *C. krusei* were seen to cause a wide range of morphologic changes, such as wrinkles, breaks, bumps, holes in the cell wall, ruffles, and a raisin-like appearance. Based on these findings, *A. marmelos* and *C. alata* could serve as promising novel antifungal agent sources.

Keywords: *Aegle marmelos*, *Cassia alata*, MIC, MFC, SEM

SDG: SDG 15- Life on land

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENILAIAN AKTIVITI ANTIMIKROB OLEH *Aegle marmelos* (L.) Corrêa
DAN *Cassia alata* L. TERHADAP MIKROORGANISMA PATOGENIK**

Oleh

MANIVANNAN A/L VEYAKANDAH

Disember 2022

Pengerusi : Profesor Madya Nur Ain Izzati binti Mohd Zainudin, PhD
Fakulti : Sains

Aegle marmelos (L.) Corrêa dan *Cassia alata* Linn adalah spesies tumbuhan yang digunakan dalam perubatan tradisional dalam rawatan pelbagai penyakit. Disebabkan perkembangan pesat rintangan mikrob terhadap antibiotik dan kemunculan strain baru telah membebankan kesihatan komuniti secara global. Oleh itu, objektif utama kajian ini adalah untuk menilai aktiviti antimikrob bagi ekstrak akueus dan metanol daripada buah muda dan daun *A. marmelos*, serta daun *C. alata*. Ekstrak daun dan buah muda *A. marmelos* dan daun *C. alata* telah disediakan melalui pengekstrakan sejuk menggunakan pelarut metanol dan air. Aktiviti antimikrob bagi ekstrak tumbuhan telah diuji ke atas bakteria patogen (*Streptococcus agalactiae*, *Escherichia coli* dan *Pseudomonas aeruginosa*) dan yis (*Candida krusei* dan *Candida parapsilosis*) menggunakan teknik resapan cakera dan telaga agar. Kepekatan perencatan minimum (KPM) dan kepekatan fungisida minimum (KFM) ditentukan dengan kaedah pencairan mikro. Perubahan selular yang disebabkan oleh ekstrak aktif telah diperhatikan dan dinilai menggunakan mikroskop elektron pengimbasan (SEM). Ekstrak metanol daun dan buah muda *A. marmelos* dan daun *C. alata* mempunyai aktiviti antikulat yang ketara terhadap *C. parapsilosis* dan *C. krusei* berbanding akueus yang tidak berkesan terhadap kedua-dua patogen. Sebaliknya, metanol dan ekstrak akueus tidak mendedahkan aktiviti antibakteria dalam kaedah resapan cakera. Nilai kepekatan perencatan minimum (KPM) daun, buah muda *A. marmelos* dan *C. alata* terhadap *C. krusei* dan adalah 75 mg/ml manakala daun *A. marmelos* mempunyai nilai kepekatan perencatan minima 37.5 mg/ml terhadap *C. parapsilosis*. Hasil kepekatan fungisida minimum (KFM) bagi *C. krusei* dan *C. parapsilosis* adalah 150 mg/ml bagi daun, buah muda *A. marmelos* dan daun *C. alata*. KPM dan KFM ekstrak methanol menunjukkan bahawa buah muda *A. marmelos* bertindak sebagai agen fungistatik. Daun *A. marmelos* dan *C. alata* berkesan sebagai agen fungistatik dan fungisida terhadap *C. krusei* dan *C. parapsilosis*. Pemerhatian mikroskop elektron pengimbasan (SEM) ekstrak metanol *A. marmelos* dan *C. alata* pada *C. parapsilosis* dan *C. krusei* menunjukkan pelbagai peringkat perubahan morfologi seperti bentuk tidak sekata dengan kedutan, pecah, bergelombang, kerosakan dinding sel yang berlubang, kecacatan dinding sel, renyuk, dan rupa seperti kismis. Hasil kajian menunjukkan bahawa ekstrak metanol *A. marmelos* dan *C. alata* berpotensi menjadi sumber agen antikulat yang baru.

Kata kunci : *Aegle marmelos*, *Cassia alata*, KPM, KFM, SEM

SDG: SDG 15- Life on land



ACKNOWLEDGEMENTS

First and foremost, praise and thanks to God, the Almighty, for His showers of blessing, throughout my research work to complete this study successfully. A million appreciations to my helpful supervisor, Assoc. Prof. Dr. Nur Ain Izzati Mohd Zainudin for all her guidance, knowledge and support given throughout the journey of research study. My appreciation also goes to both of my co-supervisors, Dr. Meenakshii A/P Nallappan and Dr. Sharina Omar for sparing their time for discussion, encouragement and support every aspect needed for completing my master study. A special thanks to the staff of Bacteriology Laboratory, Faculty of Veterinary Medicine and Mycology Laboratory, Faculty of Science for lending their help during the research work.

I'm highly obliged to Mohd Nor Azlan Bin Abdul Karim, Director of Ministry of Health Malaysia Training Institute (Medical Laboratory Technology) Kuala Lumpur. He is a good Samaritan who has always promoted me for academic excellence. Thank you also to Nur Hanani Binti Salamat from Training Management Division of Malaysian Ministry of Health. It is she who always lend a hand to all tutors in Malaysian Ministry of Health (MOH).

Nobody has been more important to me in pursuit of this project than the family members of my family. My endless gratitude to my parents, Mr. Veyakandah Gounder and Mrs. Kasturi Bai whose love and blessing are with me in whatever I pursue. I'm very grateful to my parents for their prayers, caring and sacrifices educating and preparing me to understand the future of life. They are ultimate role models. Most importantly, I wish to thank my five wonderful children Kowsalya, Rahini and Tivhyan who provide unending inspiration. I'm also thankful to my friends who have directly and indirectly contributed to my research experiences at UPM.

Finally, to my caring, loving and supportive wife, Amutha Valli, my deepest gratitude. Your encouragement when the time is rough are much appreciated and duly noted. It was great comfort and relief to know that you are willing to provide management of our household activities while I completed my work. My heartfelt thanks.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Nur Ain Izzati binti Mohd Zainudin, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Meenakshii a/p Nallappan, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Member)

Sharina binti Omar, PhD

Senior Lecturer
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 13 June 2024

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	ii
ACKNOWLEDGEMENTS	iv
APPROVAL	v
DECLARATION	vii
LIST OF TABLES	xi
LIST OF FIGURES	xii
LIST OF ABBREVIATIONS	xv
 CHAPTER	
 1 INTRODUCTION	
1.1 Background of study	1
1.2 Problem statement and research question	2
1.3 Aims and important of study	3
1.4 Research objective	3
 2 LITERATURE REVIEW	
2.1 Taxonomy and botanical description of <i>Aegle marmelos</i>	4
2.1.2 Traditional uses of <i>Aegle marmelos</i>	5
2.1.3 Bioactive activities of <i>Aegle marmelos</i>	5
2.2 Taxonomy and botanical description of <i>Cassia alata</i>	7
2.2.1 Traditional uses of <i>Cassia alata</i>	8
2.2.2 Bioactive activities of <i>Cassia alata</i>	9
2.2.3 Antibacterial activities of <i>Aegle marmelos</i>	10
2.2.4 Antibacterial activities of <i>Cassia alata</i>	12
2.3 Fungal and bacterial pathogens	16
2.3.1 <i>Candida krusei</i>	16
2.3.2 <i>Candida parapsilosis</i>	17
2.3.3 <i>Escherichia coli</i>	18
2.3.4 <i>Pseudomonas aeruginosa</i>	19
2.3.5 <i>Streptococcus agalactiae</i>	20
2.4 Antimicrobial agent and their mode of action	21
2.5 Antimicrobial resistance	21
 3 MATERIALS AND METHODS	
3.1 Fungal and bacteria cultures	22
3.2 Plant sample	23
3.3 Plant extraction	23
3.3.1 Methanol extraction	23
3.3.2 Aqueous extraction	23
3.3.3 Preparation of stock solution and crude extract	24

3.4	<i>In-vitro</i> evaluating antimicrobial activities	24
3.4.1	Antifungal analysis	24
3.4.2	Antibacterial analysis	24
3.5	Determination of minimum inhibitory concentration (MIC)	25
3.5.1	Minimum fungicidal concentration (MFC)	25
3.6	Determination on effect of active both plant extract on the microorganism through scanning electron microscope.	26
3.7	Statistical analysis	26
4	RESULT	
4.1	Effect of extracting solvent on the extract yields from <i>Aegle marmelos</i> and <i>Cassia alata</i> .	27
4.2	Antimicrobial analysis of methanol extract of <i>Aegle marmelos</i>	27
4.3	Antimicrobial analysis of aqueous extract of <i>Aegle marmelos</i>	31
4.4	Antimicrobial analysis of methanol extract of <i>Cassia alata</i>	33
4.5	Antimicrobial analysis of aqueous extract of <i>Aegle marmelos</i>	35
4.6	Minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) of <i>Aegle marmelos</i> and <i>Cassia alata</i> Linn extract based on microdilution method	37
4.7	Morphological changes of pathogens that treated with plant extract	43
5	DISCUSSION	
5.1	Antimicrobial activities of <i>Aegle marmelos</i> and <i>Cassia alata</i> extract	45
5.2	Minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) of <i>Aegle marmelos</i> and <i>Cassia alata</i> extract.	48
5.3	Morphological changes of pathogen that treated with plant extract.	50
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	
6.1	Conclusion	53
6.2	Recommendation for future study	53
	REFERENCES	55
	BIODATA OF STUDENT	75

LIST OF TABLES

Table		Page
2.1	Phytochemical in leaves of <i>Cassia alata</i> and their medicinal uses	9
2.2	The comparative of <i>in-vitro</i> antibacterial activity of different parts and extract of <i>C. alata</i> by several studies.	13
3.1	The microorganism used in this study	22
4.1	Effect of different solvent on extraction yields	27
4.2	Inhibition zone of <i>Aegle marmelos</i> methanol extract at various concentration against selected pathogens evaluated by agar well diffusion and disc diffusion method.	29
4.3	Inhibition zone of <i>Aegle marmelos</i> aqueous extract at various concentration against selected pathogens evaluated by agar well diffusion and disc diffusion method	32
4.4	Inhibition zone of <i>Cassia alata</i> methanol extract at various concentration against selected pathogens evaluated by agar well diffusion and disc diffusion method.	34
4.5	Inhibition zone of <i>Cassia alata</i> aqueous extract at various concentration against selected pathogens evaluated by agar well diffusion and disc diffusion method.	36
4.6	The MIC and MFC value of methanol extract of unripe fruit and leaves of <i>Aegle marmelos</i> and leaf of <i>Cassia alata</i> on <i>Candida krusei</i> and <i>Candida parapsilosis</i> (mg/ml).	38

LIST OF FIGURES

Figure		Page
2.1	The leaves of <i>Aegle marmelos</i> (a) and unripe fruits of <i>Aegle marmelos</i> (b)	4
2.2	The structure of bioactive compound isolated from <i>Aegle marmelos</i> : 1-Skimmianine, 2-Aegelin, 3-Lupeol, 4-1,8-Cineole, 5-Citral, 6-Citronellal, 7-4Isopropylbenzaldehyde, 8- Eugenol, 9-Marmesin, 10-Marmelosin, 11-Luvangetin, 12-Aurapten,13-Psoralen.14-Marmelide, 15-Fagarine,16-Marmin	6
2.3	The leaves of <i>Cassia alata</i>	8
4.1	Antimicrobial activity of methanol extract of unripe fruit of <i>A.marmelos</i> by agar well diffusion method against <i>C. krusei</i> (A), <i>C. parapsilosis</i> (B), and methanol extract from leaves of <i>A. marmelos</i> against <i>C. krusei</i> (C), <i>C. parapsilosis</i> (D); Antimicrobial activities of unripe fruit by disc diffusion method against <i>E. coli</i> (E), <i>S. agalactiae</i> (F), <i>P. aeruginosa</i> (G); and methanol extract from leaves of <i>A. marmelos</i> against <i>E. coli</i> (H), <i>S. agalactiae</i> (I), <i>P. aeruginosa</i> (J) at different concentrations [150 mg/ml (1), 75 mg/ml(2), 37.5 mg/ml (3), Itraconazole (30 µg/ml) (4), Gentamicin (10 µg/ml)(5), Gentamicin (120 µg/ml) (6) and Control (7)].	28
4.2	Effect of different concentration of methanol extract from leaf (AML), unripe fruit of <i>Aegle marmelos</i> (AMF) against <i>Candida</i> species. Concentration used for plant extract was mg/ml and for Itraconazole was µg/ml. Samples were incubated at $37 \pm 2^{\circ}\text{C}$ for 48 hours. Fungal plate containing no plant extract or antifungal were considered as negative control. The data are expressed as the mean $\pm$ SE (n=6).	30
4.3	Effect of different concentration of methanol extract from leaf of <i>Cassia alata</i> against <i>Candida</i> species. Concentration used for plant extract was mg/ml and for Itraconazole was µg/ml. Samples were incubated at $37 \pm 2^{\circ}\text{C}$ for 48 hours. Fungal plate containing no plant extract or antifungal were considered as negative control. The data are expressed as the mean $\pm$ SE(n=6).	30
4.4	Antimicrobial activity of aqueous extract of unripe fruit of <i>A. marmelos</i> by agar well diffusion method against <i>C. krusei</i> (A), <i>C. parapsilosis</i> (B), and aqueous extract from leaves of <i>Aegle marmelos</i> against <i>C. krusei</i> (C), <i>C. parapsilosis</i> (D); Antimicrobial activities of unripe fruit by disc diffusion method of aqueous extract against <i>E. coli</i> (E), <i>S. agalactiae</i>	31

(F), *P.aeruginosa* (G); and aqueous extract from leaves of *A. marmelos* against *E.coli* (H), *S. agalactiae* (I), *P. aeruginosa* (J) at different concentrations [150 mg/ml(1), 75 mg/ml (2), 37.5 mg/ml (3), Itraconazole (30 µg/ml) (4), Gentamicin (10 µg/ml) (5), Gentamicin (120 µg/ml) (6) and Control (7)].

- 4.5 Antimicrobial activity of methanol extract from leaf of *C. alata* by agar well diffusion method against *C. krusei* (A), *C. parapsilosis* (B), Antimicrobial activities of leaf *Cassia alata* of methanol extract by disc diffusion method of against *E. coli* (C), *S. agalactiae* (D), *P. aeruginosa* (E) at different concentration[150 mg/ml (1),75 mg/ml (2), 37.5 mg/ml(3), Itraconazole (30 µg/ml) (4), Gentamicin (10 µg/ml)(5), Gentamicin (120 µg/ml) (6) and Control (7)]. 33
- 4.6 Antimicrobial activity of aqueous extract from leaf of *C. alata* by agar well diffusion method against *C. krusei* (A), *C. parapsilosis* (B), Antimicrobial activities of leaf *C. alata* of aqueous extract by disc diffusion method of against *E. coli* (C), *S. agalactiae* (D), *P. aeruginosa* (E) at different concentrations [150 mg/ml (1), 75 mg/ml (2), 37.5 mg/ml(3), Itraconazole (30 µg/ml) (4), Gentamicin (10 µg/ml) (5), Gentamicin (120 µg/ml) (6) and Control (7)]. 35
- 4.7 Broth microdilution method for determination of MIC and MFC value of *Aegle marmelos* leaves using 96 well microtiter plate with MTT assay (A). MIC and MFC ranged from 150–1.17 mg/ml on *C. parapsilosis* ATCC 22019 (Row with letter D-E) and *C. krusei* ATCC 6258 (Row with letter A-E). The purple colouration indicates that the fungal are active; yellow coloration indicate the inhibition of the fungal growth. The leaves of *A. marmelos* completely inhibit the growth of *C. krusei* at concentration of 150 mg/ml (B). *C. parapsilosis* resistance towards methanol extract of *A. marmelos* (C). Fungal growth of *C. krusei* (D) and *C. parapsilosis* (E) observed in control (untreated leaf *A. marmelos*) and no detectable growth is observed at MFC value of leaf of *A. marmelos* (F). 39
- 4.8 Broth microdilution method for determination of MIC and MFC value of extract of *Cassia alata* leaves using 96 well microtiter plate with MTT assay (A) against *C. kruesi* ATCC 6258 (Row A-C) and *C. parapsilosis* ATCC 22019 (RowD-F). MIC and MFC ranged from 150–1.17 mg/ml, respectively. The purple colouration indicates that the fungal are active; yellow coloration indicates the inhibition of the fungal growth. *C. krusei* (ATCC 6258) resistance towards methanol extract of *C.alata* at concentration 150 mg/ml (B). The methanol extract of *C. alata* leaves inhibit the growth of *C. parapsilosis* (ATCC 22019); at concentration 150mg/ml (C). Fungal growth of *C. krusei* (D) and *C. parapsilosis* (E) 40

observed in control (untreated leaf *C. alata*) and no detectable growth is observed at MFC value of leaf of *C. alata* (F).

- 4.9 Broth microdilution method for determination of MIC and MFC value of extract of unripe fruit of *A. marmelos* using 96 well microtiter plate with MTT assay (A) against *C. krusei* (ATCC 6258) (Row A-C) and *C. parapsilosis* (ATCC 22019) (Row D-F). MIC and MFC ranged from 150–1.17 mg/ml, respectively. The purple colouration indicates that the fungal are active; yellow coloration indicates the inhibition of the fungal growth. *C. krusei* (ATCC 6258) shows resistance towards methanol extract of unripe fruit *A. marmelos* (B). MFC of *C. parapsilosis* (ATCC 22019) of methanol extract of unripe fruit *A. marmelos* is 150 mg/ml (C) Fungal growth of *C. krusei* (D) and *C. parapsilosis* (E) observed in control (untreated unripe fruit of *A. marmelos*) and no detectable growth is observed at MFC value of unripe fruit of *A. marmelos* (F). 41
- 4.10 Broth microdilution method for determination of MIC and MFC value of Itraconazole using 96 well microtiter plate with MTT assay (A). MIC and MFC ranged from 30–0.93 µg/mL on *C. parapsilosis* (ATCC 22019) (Row with letter D-E) and *C. krusei* (ATCC 6258) (Row with letter A-C). The Itraconazole completely inhibit the growth of *C. krusei* (ATCC 6258) at concentration of 30 µg/ml (B). *C. parapsilosis* (ATCC 22019) completely inhibit at 30 µg/ml. The MIC value for Itraconazole is 15 µg/ml. 42
- 4.11 Scanning electron micrograph analysis of the effect of *A. marmelos* (leaves and fruits) and *C. alata* (leaves) extract on the morphology of *C. parapsilosis* (A) control cell, morphology alteration in the cell including cell wall deformities and cell wall damaged are indicated by arrow in cell treated with the leaves *A. marmelos* (B), and protuberance are indicated by arrow in the cell treated with *C. alata* leaves (C) and hole are indicated by arrow in the cell treated with unripe fruit of *A. marmelos* (D). Magnification X5000. The screening was performed at least 20 fields. 44
- 4.12 Scanning electron micrograph analysis of the effect of *A. marmelos* (fruit and leaves) and *C. alata* (leaves) extract on the morphology of *C. krusei* (A) control cell, morphology alteration in the cell including hole and cell wall damaged are indicated by arrow in cell treated with the leaves *A. marmelos* (B), wrinkles and protuberance are indicated by arrow in the cell treated with *C. alata* leaves (C) cell wall deformities and raisin like appearance are indicated by arrow in the cell treated with unripe fruit of *A. marmelos* (D). Magnification X5000. The screening was performed at least 20 fields. 44

LIST OF ABBREVIATIONS

ATCC	American Type Culture Collection
ATP	Adenosine triphosphate
MIC	Minimum Inhibitory Concentration
MFC	Minimum Fungicidal Concentration
mg/ml	Miligram per mililitre
µg/mL	Microgram per mililitre
mm	Milimeter
CFU/mL	Colony forming unit per mililitre
MTT	3-bis (2-methoxy-4-nitro- 5-sulphophenyl)-2H-tetrazolium-5-carboxanilide (MTT)
DMSO	Dimethyl Sulfoxide
SEM	Scanning Electron Microscope
SE	Standard Error
Ca ²⁺	Calcium
TLR4	Toll-like receptor 4
SDA	Sabouraud Dextrose Agar
MH	Mueller Hinton Agar
ITZ	Itraconazole
CVC	Central Venous Catheter
ICU	Intensive care unit
MDR	Multidrug Resistant
r.p.m	Revolutions per minute
UPM	Universiti Putra Malaysia

CHAPTER 1

INTRODUCTION

1.1 Background of study

Many cultures have traditional uses for medicinal plants as the era of alternative medicine and holistic health approaches. Furthermore, these natural resources don't have to worry about the negative fallout that can result from man-made synthetic substances. Analysis of the potential antibacterial properties of many plant extracts and therapeutic ointments has found organic effects (Correia et al., 2016). Plant extracts' antimicrobial properties have served as the foundation for a wide variety of uses, including raw and processed food safety, pharmaceuticals, complementary and alternative medicine, and conventional therapies (Mahlo et al., 2016). Eco-friendly and bio-compatible plant-based products have recently received a lot of attention for the prevention and treatment of many human ailments (Sivanathan & Elamaram, 2013). Yuan et al. (2016) found that in both Asia and Africa, around 80% of the population relies on medicinal spices alone or in combination with traditional medicine. Two thousand kinds of medicinal plants are thought to exist in Peninsular Malaysia, with participation from a wide range of ethnic groups (Sabran et al., 2016). Medicinal plant use is based on "common sense," "perception," and "custom practise" derived from socioreligious convictions that have been passed down from generation to generation (Mohamad et al., 2019).

Plants also contain bioactive combinations that have been shown to be antimicrobial whether used singly or in combination with other antimicrobial drugs (Hu et al., 2020). There are less negative effects on animals, humans, and the environment when these mixes are used in traditional medicine (Ncube et al., 2012; Saha & Dasgupta, 2005), hence they are widely used as a crude extract for natural remedies. *Cassia alata* Linn and *Aegle marmelos* (L) Corrêa were used in this research as potential options for combating fungal and bacterial infections.

In addition to its more common names, the *Aegle marmelos* tree is also known as sripal or bilwa in Sanskrit, vilvam in Tamil, and stone apple or bael in English or Pokok Maja Batu in Malay. In addition to its native range in India, *A. marmelos* can also be found in a number of other countries and territories, including China, Nepal, Ceylon, Myanmar, Pakistan, Bangladesh, Nepal, Vietnam, Laos, Cambodia, Thailand, Indonesia, Malaysia, Tibet, Java, the Philippines, and Fiji (Yadav et al., 2015). The tree thrives in well-drained soil and can reach a height of 12-15 m in spite of the harsh and arid climate. The leaflets of its leaves range in size from three to five ovals and are pointy and shallowly serrated. The flowers can be seen in clusters all along the young branches, and they have a pleasant scent (Baliga et al., 2011). The tree is also said to have produced its first fruit in 1941 in Northern Malaysia (Sharma et al., 2007).

The *A. marmelos* plant, including its stem, bark, root, leaves, and fruits, has been used as a natural medicine in India's Ayurvedic, Unani, and Siddha medical systems (Raja et al., 2011). Aegelin, skimmianine, lupeol, cineol, citral, marmesinin, marmelide, cuminaldehyde, eugenol, -sitosterol, flavone, glycoside, and marmeline are just few of the bioactive chemicals isolated from *A. marmelos* leaf, fruits, bark, and root (Monika et al., 2023). Therapeutic qualities such as anti-diabetic, anti-ulcer, anti-oxidant, anti-malarial, anti-inflammatory, anti-cancer, anti-hyperlipidaemic, and anti-bacterial and anti-viral activities have been attributed to their extracts and used in traditional medicine (Somu et al., 2019).

Common names for the bush *Cassia alata* [synonym: *Senna alata* (L.) Roxb.] include Candle, Dadmardan, and Semaigathi in India; the Roman Candle tree in Fiji; Gelenggan in Malaysia; and Ketepeng Badak in Indonesia (Alioes & Kartika, 2019). For optimal results, a small amount of lime is sometimes combined with the concentrate when the plant is used to treat ringworm infection and parasitic skin disease in Malaysia and Nigeria (Owoyale et al., 2006; Ibrahim & Osman, 1995). It is used to treat a wide variety of illnesses in India and South America, including skin disorders, asthma, snake bites, fever, digestive problems, and even heart diseases (Yeong et al., 2017).

According to Somchit et al. (2003), *Cassia alata* found in these regions Southeast Asia, Fiji, Northern Australia, Africa, and Latin America. The anti-inflammatory, analgesic, antifungal, hypoglycemic, diuretic, and wound-healing properties of a *C. alata* leaf extract have been well documented (Mohideen et al., 2005). There are some flavonoids and anthraquinones in the ethanol extract of *C. alata* leaves (Kumar et al., 2016). While chrysophanol, emodin, and rhein all have antibacterial properties (Wuthi-Udomler et al., 2010), rhein is the most potent.

1.2 Problem statement and research questions

Plants have long been a go-to source for restorative drugs, playing an important role in healthcare and providing a wealth of potential novel antimicrobial drugs for use against bacteria and other drug-resistant microbes. It is possible that the new antibacterial agents will be cloaked in natural ointments and extracts from medicinal plants (Sabo & Knezevic, 2019). However, antimicrobial drug supplies are limited, and widespread use of these drugs for treatment has contributed to the rise of drug-resistant microorganisms. This research aimed to answer the following analytical questions about the antimicrobial and antifungal efficacy of *A. marmelos* and *C. alata* extracts against infectious microorganisms: i. two, how effective are *A. marmelos* and *C. alata* extracts against bacteria and fungus, as measured by their minimal inhibitory, bactericidal, and fungicidal concentrations? Extracts of *A. marmelos* and *C. alata* are hypothesised to be effective against *Candida* spp., *Escherichia coli*, *Pseudomonas aeruginosa*, and *Streptococcus agalactiae*, and to be useful in the treatment of related disorders. This research will help determine whether or not plant extracts may be used as an alternative to readily available conventional medication for treating microbial infections.

1.3 Aims and importance of study

Pathogenic microorganisms, such as fungus and bacteria, are becoming increasingly resistant to antibacterial treatments in the coming years (Raoufi & Shafaghat, 2018). Presently, there is a serious threat to the clinical application of anti-infection drugs and the effective treatment of bacterial infections due to the proliferation of bacteria that have developed resistance to numerous types of antibiotics (Mandrone et al., 2019). Eighty percent of people in non-industrialized countries rely on plant medicine as their primary source of healthcare (Dulger, 2009). Half of all newly synthesised particles derived from natural sources have demonstrated their value in the development of drugs for the treatment of infectious disorders (Baliga et al., 2011).

Traditional plant medicines for bacterial and infectious diseases may provide inspiration for the creation of new, effective natural antibiotics (Murugan & Kamaraj, 2018). Tannins, terpenoids, alkaloids, flavonoids, and glycosides are just some of the many optional metabolites found in plants that have been shown to exhibit antibacterial activities when tested in vitro (Dubey et al., 2019). In addition, they are effective against bacteria, fungi, viruses, and protozoa, making them useful for treating a wide variety of skin conditions, including dermatitis, burns, diarrhoea, fever, wounds, cuts, sores, hacking, and localised skin swellings (Pesewu et al., 2008). Because of the plant's origin, these factors compelled botanists and pharmaceutical companies to go elsewhere for treatments (Al Farraj et al., 2020). These explanations suggest that traditional medicine has merit when compared to modern medicine (Mahlo et al., 2016).

1.4 Research objectives

The objectives of this study are:

- i. To evaluate the antimicrobial activities of *A. marmelos* and *C. alata* on *Candida krusei* (ATCC 6258), *Candida parapsilosis* (ATCC 22019), *Escherichia coli*, *Pseudomonas aeruginosa* and *Streptococcus agalactiae*.
- ii. To examine the morphological changes of selected pathogens treated with plant extract under the scanning electron microscope (SEM).

REFERENCES

- Abirami, S. K. G., Vivekanandhan, K., Hemanthkumar, R., Prasanth, S., & Kumar, J. R. (2014). Study of antimicrobial potential of *Aegle marmelos*. *Journal of Medicinal Plants Studies*, 2, 113–116.
- Abubacker, M. N., & Ramanathan, R. (2005). Antibacterial activity of flower extract of *Cassia alata*, *Journal Tropical Medicine Plants*, 6(2), 183-185.
- Abubacker, M. N., Ramanathan, R., & Senthil Kumar, T. (2008). *In vitro* antifungal activity of *Cassia alata* flower extract. *Indian Journal of Natural Products and Resources*, 7(1), 6–9.
- Agyare, C., Bempah, S. B., Boakye, Y. D., Ayande, P. G., Adarkwa-Yiadom, M. & Mensah, K. B. (2013). Evaluation of antimicrobial and wound healing potential of *Justicia flava* and *Lannea welwitschii*. *Evidence-Based Complementary and Alternative Medicine*, vol 2013, 10 pages.
- Ahmad, A., Kaleem, M., Ahmed, Z., & Shafiq, H. (2015). Therapeutic potential of flavonoids and their mechanism of action against microbial and viral infections- A review. *Food Research International*, 77, 221–235.
- Ahmad, S., & Akram, M. (2019). Antifungal activity in the methanolic, aqueous and hexane extracts of *Calligonum polygonoides*. *International Journal of Immunopathology and Pharmacology*, 33, 1-5.
- Akharaiyi, F. C., & Dina, A. O. (2019). Antimicrobial activities, antioxidant and phytochemical analysis of methanol and aqueous leaf extract of *Phyllanthus muellerianus* (KUNTZ) Exell. *International Journal of Pharmacy Research and Technology*, 8(1), 29–38.
- Al Farraj, D. A., Ragab Abdel Gawwad, M., Mehmood, A., Alsalmeh, A., Darwish, N. M., Al-Zaqri, N., & Warad, I. (2020). *In vitro* antimicrobial activities of organic solvent extracts obtained from *Dipcadi viride* (L.) Moench. *Journal of King Saud University - Science*, 32(3), 1965–1968.
- Alalor, C. A., Igwilo, C. I., & Jeroh, E. (2012). Evaluation of the antibacterial properties of aqueous and methanol extracts of *Cassia alata*. *Journal of Pharmacy and Allied Health Sciences*, 2(2), 40-46.
- Aldholmi, M., Marchand, P., Ourliac-garnier, I., Pape, P. Le, & Ganesan, A. (2019). A decade of antifungal leads from natural products: 2010 - 2019. *Pharmaceuticals*, 12(4), 182, 1-21.
- Alioes, Y., & Kartika, A. (2019). Uji potensi antijamur *Candida albicans* ekstrak daun gelinggang (*Cassia alata* L.) dibandingkan dengan sediaan daun sirih yang beredar di pasaran secara *in vitro*. *Jurnal Kimia Riset*, 3(2), 108.

- Alishir, A., Yu, J. S., Park, M., Kim, J. C., Pang, C., Kim, J. K., Kim, K. H. (2021). Ulmusakidian, a new coumarin glycoside and antifungal phenolic compounds from the root bark of *Ulmus davidiana* var. *japonica*. *Bioorganic and Medicinal Chemistry Letters*, 36(Mar), 127828.
- Alyousef, A. A. (2021). Antifungal activity and mechanism of action of different parts of *Myrtus communis* growing in Saudi Arabia against *Candida* species. *Journal of Nanomaterials*, 2021, 1-10.
- Amaral-Lopes, S., & Moura, A. (2012). Neonatal fungal sepsis by *Candida krusei*: A report of three cases and a literature review. *Medical Mycology Case Reports*, 1(1), 24–26.
- Andleeb, S., Alsalmeh, A., Al-zaqri, N., Warad, I., & Alkahtani, J. (2020). *In-vitro* antibacterial and antifungal properties of the organic solvent extract of *Argemone mexicana* L. *Journal of King Saud University – Science*, 32(3), 2053-2058.
- Andrews, R. E., Parks, L. W., & Spence, K. D. (1980). Some effects of douglas fir terpenes on certain microorganisms. *Applied and Environmental Microbiology*, 40(2), 301–304.
- Angel Ezhilarasi, A., Judith Vijaya, J., Kaviyarasu, K., John Kennedy, L., Ramalingam, R. J., & Al-Lohedan, H. A. (2018). Green synthesis of NiO nanoparticles using *Aegle marmelos* leaf extract for the evaluation of *in-vitro* cytotoxicity, antibacterial and photocatalytic properties. *Journal of Photochemistry and Photobiology B: Biology*, 180(mar), 39-50.
- Appiah, T., Boakye, Y. D., & Agyare, C. (2017). Antimicrobial activities and time-kill kinetics of extracts of selected Ghanaian mushrooms. *Evidence-Based Complementary and Alternative Medicine*, 2017, 1-15
- Appleyard R., K. (1954). Segregation of new lysogenic types during growth of a doubly lysogenic strain derived from *Escherichia coli* K12. *Genetics*, 39(4), 440-452
- Arendrup, M. C., Cuenca-Estrella, M., Lass-Flörl, C., Hope, W., Arendrup, M. C., Hope, W. W., Verweij, P. (2012). EUCAST technical note on *Aspergillus* and amphotericin B, itraconazole, and posaconazole. *Clinical Microbiology and Infection*, 18(7), E248-E250.
- Ariharan, V. N., & Nagendra Prasad, P. (2014). Anti-bacterial activity of three morphological traits of *Aegle Marmelos* (Linn.) Corr.-‘Vilvam.’ *Rasayan Journal of Chemistry*, 7(3), 260–263.
- Azam M., W., & Khan, U., A. (2019). Updates on the pathogenicity status of *Pseudomonas aeruginosa*. *Drug Discocery Today*, 24(1), 350-359.
- Bac, N. D., Anh, L. T., Quang, L. B., Luc, N. K., Thi, T., & Nga, T. (2019). Prevalence of *Candida* bloodstream isolates from patients in two hospitals in Vietnam. *Iranian Journal of Microbiology*, 11(2), 108–113.

- Bahri, S., Tamrin, M., Tukur, A. M., D. M., Nasir, M., & Siti, K. (2016). Prevalence of *Escherichia Coli* O157 : H7 and *Enterobacteriaceae* on hands of workers in halal cattle abattoirs in Peninsular Malaysia. *The Malaysian Journal of Medical Science*, 23(5), 65–71.
- Balakumar, S., Rajan, S., Thirunalasundari, T., & Jeeva, S. (2011). Antifungal activity of *Aegle marmelos* (L.) *Corrêa* (Rutaceae) leaf extract on dermatophytes. *Asian Pacific Journal of Tropical Biomedicine*, 1(4), 309–312.
- Balashanmugam, P., Balakumaran, M. D., Murugan, R., Dhanapal, K., & Kalaichelvan, P. T. (2016). Phytogetic synthesis of silver nanoparticles, optimization and evaluation of *in vitro* antifungal activity against human and plant pathogens. *Microbiological Research*, 192, 52–64.
- Baliga, M. S., Bhat, H. P., Joseph, N., & Fazal, F. (2011). Phytochemistry and medicinal uses of the bael fruit (*Aegle marmelos* Correa): A concise review. *Food Research International*, 44(7), 1768–1775.
- Baraboutis, I. G., Doris, K., Papanikolaou, K., Tsagalou, E. P., Chatsiou, K., Papathanasiou, E., Papastamopoulos, V. (2010). An outbreak of hemodialysis catheter-related bacteremia with sepsis caused by *Streptococcus agalactiae* in a hemodialysis unit. *International Journal of Infectious Diseases*, 14(5), 418–422.
- Behera, P., Vennel Raj, V., & Basavaraju, R. (2014). Phytochemical and antimicrobial activity of fruit pulp of *Aegle marmelos*. *Journal of Chemical and Pharmaceutical Research*, 6(8), 319–326.
- Bélanger, L., Garenaux, A., Harel, J., Boulianne, M., Nadeau¹ E., & Dozois, C.M. (2011). *Escherichia coli* from animal reservoirs as a potential source of human extraintestinal pathogenic *E. coli*. *FEMS Immunology and Medical Microbiology*, 62 (1),1-10.
- Berkow, E.L., Lockhart, S.R., & Luis, O.Z. (2020). Antifungal susceptibility testing: Current approaches. *Clinical Microbiology Review*, 33 (3),1-30.
- Bhanot, M. (2018). Development and sensory evaluation of unripe and ripe Bael fruit powder (*Aegle marmelos*), *Journal of Pharmacognosy and Phytochemistry*, 7(4), 2894–2897.
- Bhuyan, D. J., Vuong, Q. V., Chalmers, A. C., van Altena, I. A., Bowyer, M. C., & Scarlett, C. J. (2017). Phytochemical, antibacterial and antifungal properties of an aqueous extract of *Eucalyptus microcorys* leaves. *South African Journal of Botany*, 112, 180–185.
- Bilkis, Z., Jamal, M. A. H. M., Alam, M. K., & Islam, M. R. (2017). Assessment of the antimicrobial activities of wood apple (*Aegle marmelos* L.) seed extracts against selected fungal and bacterial strains. *Anti-Infective Agents*, 15(1), 33-37.

- Bismelah, N.A. Rohana, A., Zety, H.M.K., Nor Hadiani, I., & Nurul fazlina E.R. (2022) The antibacterial effect of *Plectranthus scutellarioides* (L.) R.BR. leaves extract against bacteria associated with peri-implantitis. *Journal of Traditional and Complementary Medicine*, 12(6), 556–566.
- Bonfietti, L. X., Martins, M. dos A., Szeszs, M. W., Pukiskas, S. B. S., Purisco, S. U., Pimentel, F. C., Pereira, G.H., Silva, D.C., Olivera, L., & Melhem, M. S. C. (2012). Prevalence, distribution and antifungal susceptibility profiles of *Candida parapsilosis*, *Candida orthopsilosis* and *Candida metapsilosis* bloodstream isolates. *Journal of Medical Microbiology*, 61(Pt7), 1003–1008.
- Boregowda, R. S., Murali, N., Udayashankar, A. C., Niranjana, S. R., Lund, O. S., & Prakash, H. S. (2019). Antifungal activity of *Eclipta alba* metabolites against sorghum pathogens. *Plants*, 8(3),72.
- Brindhadevi, K., LewisOscar, F., Mylonakis, E., Shanmugam, S., Verma, T. N., & Pugazhendhi, A. (2020). Biofilm and quorum sensing mediated pathogenicity in *Pseudomonas aeruginosa*. *Process Biochemistry*, 96, 49–57.
- Chakthong, S., Weaaryee, P., Puangphet, P., Mahabusarakam, W., Plodpai, P., Voravuthikunchai, S. P., & Kanjana-Opas, A. (2012). Alkaloid and coumarins from the green fruits of *Aegle marmelos*. *Phytochemistry*, 75, 108–113.
- Channa, U., Shah, A. M., Bhatti, S., Memon, A. A., Ghanghro, A. B., & Memon, M. N. (2020). Phytochemical analysis and antibacterial properties of *Cassia Senna alata*. *Rawal Medical Journal*, 45(1), 223–226.
- Chase, Mark W., Morton C. M. & Kallunki J. A. (1999) Phylogenetic relationships of rutaceae: A cladistic analysis of the subfamilies using evidence from RBC and ATP sequence variation, *American Journal of Botany*, 86 (8),1191-1199.
- Chinchansure, A., Shamnani, N., Arkile - Shingare, M., Sarkar, D., & Joshi, S. (2015). Antimycobacterium activity of coumarin from fruit of *Aegle marmelos* (L) Correa. *International Journal of Basic and Applied Chemical Sciences*, 5, 2277–207339.
- Choudhary, P. (2017). Evaluation of *Aegle marmelos* extracts for antibacterial, antifungal, antioxidant and cytotoxic properties. *Current Trends in Biomedical Engineering and Biosciences*, 9(4), 7–10.
- Choukri, F., Benderdouche, M., & Sednaoui, P. (2014). *In vitro* susceptibility profile of 200 recent clinical isolates of *Candida* species to topical antifungal treatments of vulvovaginal candidiasis, the imidazoles and nystatin agents. *Journal de Mycologie Medicale*, 24(4), 303–307.
- Clinical and Laboratory Standards Institute (CLSI), 2009. Method for antifungal disk diffusion susceptibility testing of yeasts; approved guideline, 2nd ed., M44-A2 Clinical and Laboratory Standards Institute, Wayne, PA.

- CLSI. (2012). Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; twenty-second informational supplement. CLSI document M100-S22. Clinical and Laboratory Standard Institute, Wayne.
- Correia, A. F., Silveira, D., Fonseca-Bazzo, Y. M., Magalhães, P. O., Fagg, C. W., da Silva, E. C., de Medeiros Nóbrega, Y. K. (2016). Activity of crude extracts from Brazilian cerrado plants against clinically relevant *Candida* species. *BMC Complementary and Alternative Medicine*, 16(1), 1-9.
- Cox, S. D., Mann, C. M., Markham, J. L., Bell, H. C., Gustafson, J. E., Warmington, J. R., & Wyllie, S. G. (2000). The mode of antimicrobial action of the essential oil of *Melaleuca alternifolia* (Tea tree oil). *Journal of Applied Microbiology*, 88(1), 170–175.
- Dahiya, R., Tomar, R. S., & Shrivastava, V. (2015). Evaluation of antimicrobial potential of *Aegle marmelos* fruit extract against selected microorganisms. *Journal of Pharmaceutical Sciences and Research*, 7(9), 681–684.
- Davidson, P. M., & Brannen, A. L. (2005). Food antimicrobials: an introduction. In P. M. Davidson, J. N. Sofos, & A. L. Brannen (Eds.). *Antimicrobial in food* (pp. 1–10). (3rd ed.). New York, NY: CRC Press.
- De Oliveira Santos, G. C., Vasconcelos, C. C., Lopes, A. J. O., de Sousa Cartágenes, M. do S., Filho, A. K. D. B., do Nascimento, F. R. F., Ramos RM., Pires ERRB, de Andrade Monteiro, C. (2018). *Candida* infections and therapeutic strategies: mechanisms of action for traditional and alternative agents. *Frontiers in Microbiology*, 9, 1351.
- Dhankhar, S., Ruhil, S., Balhara, M., Dhankhar, S., & Chhillar, A. K. (2011). *Aegle marmelos* (Linn.) Correa: A potential source of Phytomedicine. *Journal of Medicinal Plants Research*, 5(9), 1497–1507.
- Duan, C. *et al.* (2023) Gram-negative bacterial infection causes aggravated innate immune response in sepsis: Studies from clinical samples and cellular models. *Biochemical and Biophysical Research Communications*, 650, 137–144.
- Dubey, S., Kumar Dixit, A., Dehariya, R., & Kumar Dixit, A. (2019). Preliminary phytochemical screening of *Carthamus tinctorius* L. *Research Journal of Life Sciences, Bioinformation, Pharmaceutical and Chemical Sciences*, 5(3), 330.
- Dulger, B. (2009). Antifungal activity of *Lamium tenuiflorum* against some medical yeast *Candida* and *Cryptococcus* species. *Pharmaceutical Biology*, 47(5), 467–470.
- Duraipandiyan, V., & Ignacimuthu, S. (2007). Antibacterial and antifungal activity of *Cassia fistula* L.: an ethnomedicinal plant. *Journal of Ethnopharmacology*, 112, 590–594.

- Effendy, Laydiana. (2013), Potensi antijamur kombinasi ekstrak etanol daun sirih merah (*Piper crocatum* Ruiz & Pav.) dan kelopak bunga rosella (*Hibiscus sabdariffa* Linn.) terhadap *Candida albicans*. *Calyptra*, 2(1), 1-10.
- Elisha, I. L., Botha, F. S., McGaw, L. J., & Eloff, J. N. (2017). The antibacterial activity of extracts of nine plant species with good activity against *Escherichia coli* against five other bacteria and cytotoxicity of extracts. *BMC Complementary and Alternative Medicine*, 17(1), 133.
- El-Mahmood A. M, Doughari J. H. (2008). Phytochemical screening and antibacterial evaluation of the leaf root extracts of *Cassia* or Linn. *African Journal Pharmacy Pharmacology*, 2 (7), 124- 129.
- Esimone, C. O., Nworu, C. S., Ekong, U. S., & Okereke, B. (2007). Evaluation of the antiseptic properties of *Cassia alata*-based soap. *The Internet Journal of Alternative Medicine*, 6(1), 1–5.
- Evans, J. J., Klesius, P. H., & Shoemaker, C. A. (2004). Efficacy of *Streptococcus agalactiae* (group B) vaccine in tilapia (*Oreochromis niloticus*) by intraperitoneal and bath immersion administration. *Vaccine*, 22,(27-28), 3769-3773.
- Fatmawati, S., Yuliana, Purnomo, A. S., & Abu Bakar, M. F. (2020). Chemical constituents, usage and pharmacological activity of *Cassia alata*. *Heliyon*, 6(7), 04396.
- Fernández-Torres, B., Carrillo-Muñoz, A., Inza, I. and Guarro, J. (2006). Effect of culture medium on the disk diffusion method for determining antifungal susceptibilities of dermatophytes. *Antimicrobial Agents and Chemotherapy*, 50(6), 2222–2224.
- Ganapathy, S., & Karpagam, S. (2016). *In vitro* antibacterial and phytochemical potential of *Aegle marmelos* against multiple drug resistant (MDR) *Escherichia coli*. *Journal of Pharmacognosy and Phytochemistry*, 5(1), 253–255.
- Garcia-rubio, R., Oliveira, H. C. De, Rivera, J., Niño-vega, G. A., & Hall, R. A. (2020). The fungal cell wall *Candida*, *Cryptococcus*, and *Aspergillus* species. *Frontiers in Microbiology*, 10, 2993.
- Gómez-Gaviria, M., & Mora-Montes, H. M. (2020). Current aspects in the biology, pathogeny, and treatment of *Candida krusei*, a neglected fungal pathogen. *Infection and Drug Resistance*, 13, 1673–1689.
- Griffith, M., Postelnick, M., Scheetz, M. (2012).Antimicrobial stewardship programs: methods of operation and suggested outcomes. *Expert Review Anti-Infective*,10, 63–73.

- Gutiérrez JA, Caballero S, Díaz LA, Guerrero MA, Ruiz J, Ortiz CC. (2017). High antifungal activity against candida species of monometallic and bimetallic nanoparticles synthesized in nanoreactors. *ACS Biomaterial Science Engineering*, 4(2):647–653.
- Halim-Lim, S., Ramli, N. ., Fadzil, F. A., & Abd Rahim, M. H. A. (2020). The antimicrobial and antioxidant properties of *Cassia alata* extraction under different temperature profiles. *Food Research*, 1-6.
- Hammouche-Mokrane, N., León-González, A. J., Navarro, I., Boulila, F., Benallaoua, S., & Martín-Cordero, C. (2017). Phytochemical profile and antibacterial activity of *Retama raetam* and *R. sphaerocarpa cladodes* from Algeria. *Natural Product Communications*, 12(12), 1857–1860.
- Hasanthi, K. B., Gamage, R. N. N., & Kumari, K. (2018). Anti-microbial activity of different solvent extracts of dried flowers of *Aegle marmelos*. *Journal of Pharmacognosy and Phytochemistry*, 7(1), 1827–1830.
- Hennebelle, T., Weniger, B., Joseph, H., Sahpaz, S., & Bailleul, F. (2009). *Senna alata*. *Fitoterapia*, 80(7), 385–393.
- Ho, S. E., Subramaniam, G., Palasubramaniam, S., & Navaratnam, P. (2002). Carbapenem-resistant *Pseudomonas aeruginosa* in Malaysia Producing IMP-7 Lactamase. *Antimicrobial Agents and Chemotherapy*, 46(10), 3286–3287.
- Hossan, M. S., Hanif, A., Khan, M., Bari, S., Jahan, R., & Rahmatullah, M. (2009). Ethnobotanical survey of the Tripura tribe of Bangladesh. *American-Eurasian Journal of Sustainable Agriculture*, 3(2), 253–261.
- Hugh R & Leifson E. (1964). The proposed neotype strains of *Pseudomonas aeruginosa*. (Schroeter 1872) Migula 1900. *International Journal of Systematic Bacteriology*, 20(1), 15-16.
- Hurley R., & Stanley V.C. (1969). Cytopathic effects of pathogenic and non-pathogenic species of *Candida* on cultured mouse epithelial cells: relation to the growth rate and morphology of the fungi. *Journal of Medical Microbiology*, 2, 63-74.
- Ibrahim, D., & Osman, H. (1995). Antimicrobial activity of *Cassia alata* from Malaysia. *Journal of Ethnopharmacology*, 45(3), 151–156.
- Ibrahim, N. A., El-Sakhawy, F. M., Mohammed, M. M. D., Farid, M. A., Abdel-Wahed, N. A. M., & Deabes, D. A. H. (2015). Chemical composition, antimicrobial and antifungal activities of essential oils of the leaves of *Aegle marmelos* (L.) Correa growing in Egypt. *Journal of Applied Pharmaceutical Science*, 5(2), 1–5.
- Iraqi, P., Chakraborty, T., Das, M. K., & Yadav, R. N. (2019). Herbal antimicrobial gel with leaf extract of *Cassia alata* L. *Journal of Drug Delivery and Therapeutics*, 9(3), 82–94.

- Jeeva, G., Jeeva, S., & Kingston, C. (2007). Traditional treatment of skin diseases in South Travancore, southern peninsular India. *Indian Journal of Traditional Knowledge (IJTK)*, 06(3), 498–501.
- Jenson, B. N., Ibrahim, H., Danmalam, H. U., Olorunmola, J., Aliyu, I. I., & Jenson, B. N. (2020). Phytochemical and antimicrobial studies on *Senna alata* leaf extracts and fractions. *Journal of Pharmaceutical Development and Industrial Pharmacy*, 2(2), 1–10.
- Jud, P., Valentin, T., Regauer, S., Gary, T., Hackl, G., Rief, P., Brodmann, M., Hafner, F. (2017). Invasive *Candida krusei* infection and *Candida vasculitis* of a leg ulcer in an immunocompetent patient: A case report. *International Journal of Infectious Diseases*, 55, 96–98.
- Kadosh, D., & Mundodi, V. (2020). A re-evaluation of the relationship between morphology and pathogenicity in *Candida* species. *Journal of Fungi*, 6(1), 16–18
- Kaper, J. B., Nataro, J. P., & Mobley, H. L. T. (2004). Pathogenic *Escherichia coli*. *Nature Reviews Microbiology*, 2(2), 123–140
- Karumaran, S., Nethaji, S., & Rajakumar, R. (2016). Antimicrobial and antioxidant activity of leaf extracts of *Aegle marmelos*. *Advances in Applied Science Research*, 7(3), 205–208.
- Kenconoajati, H., Ulkhaq, M. F., Azhar, M. H., & Budi, D. S. (2019). Evaluation of antibacterial activity of different solvent extract from *Coffea canephora* leaves against *Edwardsiella tarda* and *Streptococcus agalactiae*. *AACL Bioflux*, 12(6), 2371–2377.
- Khan, M. R., Kihara, M., Omoloso, A. D. (2001). Antimicrobial activity of *Cassia alata*. *Fitoterapia*, 75(2), 561-564.
- Khan, S., Imran, M., Imran, M., & Pindari, N. (2017). Antimicrobial activity of various ethanolic plant extracts against pathogenic multi drug resistant *Candida* spp. *Bioinformation*, 13(03), 67–72.
- Kibret M., & Abera, B. (2011). Antimicrobial susceptibiltiy pattern of *E.coli* from clinical source `in noreast Ethiopia. *African Health Sciences*, 11(1),40-45.
- Kim, J. H., Choi, W. G., Moon, J. Y., Lee, J. Y., Lee, S., & Lee, H. S. (2018). Metabolomics-assisted metabolite profiling of itraconazole in human liver preparations. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 1083(March), 68–74.
- Kirmusaoğ lu, S. (Ed). (2019). Antimicrobials, antibiotic resistance, antibiofilm strategies and activity methods. IntechOpen. 78751.

- Kline, K. A., & Lewis, A. L. (2016.). Gram-positive uropathogens, polymicrobial urinary tract infection, and the emerging microbiota of the urinary tract. *Microbial Spectrum*, 4(2),1-31
- Koluman, A., Akar, N., & Haznedaroğlu, İ. C. (2017). Antibacterial activity of Ankaferd Hemostat (ABS) on shiga toxin producing *Escherichia coli* and other pathogens significant in foodborne disease. *Turkish Journal of Hematology*, 34(1), 93–98.
- Kothari, S., Mishra, V., Bharat, S., & Tonpay, S. D. (2011). Antimicrobial activity and phytochemical screening of serial extracts from leaves of *Aegle marmelos* (Linn.). *Acta Poloniae Pharmaceutica - Drug Research*, 68(5),687-692.
- Kronvall G., & Karlsson I. (2001). Fluconazole and voriconazole multidisk testing of *Candida* species for disk test calibration and mic estimation. *Journal of Clinical Microbiology*,39(4):1422-8
- Kub, T. N. T., Manaf, N. A. A. & Ibrahim, A. S. C. (2021). Comparison of well diffusion, disc diffusion and broth dilution methods for antimicrobial activity of andrographis paniculata herbal gel against acne-associated pathogen. *Malaysian Journal of Microbiology*, 17(1), 1-7.
- Kumar Sekar, D., Kumar, G., Karthik, L., & Rao, K. V. B. (2011). A review on pharmacological and phytochemical properties of *Aegle marmelos* (L.) Corr. Serr. (Rutaceae). *Asian Journal of Plant Science and Research*, 1(2), 8–17.
- Kumar, A., Zarychanski, R., Pisipati, A., Kumar, A., Kethireddy, S. & Bow, E.J. (2018) Fungicidal versusfungistatic therapy of invasive *Candida* infection in non-neutropenic adults: a meta-analysis. *Mycology*, 9, 116–128
- Kumar, R., Anburaj, G., & Vasantha, S. (2016). Phytochemical investigation of ethanol, methanol, hydro-alcoholic and aqueous flower extracts of *Cassia* species.*Asian Journal of Innovative Research*, 32-35.
- Kyle, A.A. and Dahl, M.V. (2004) Topical therapy for fungalinfections. *Amrican Journal of Clinical Dermatology*, 5, 443–451.
- Lallitto, F., Prigitano A., Mangione F., Piralla A., Tamarozzi F., Marone P., Cavanna C. (2018). Presence of L701 M mutation in the FKS1 gene of echinocandin-susceptible *Candida krusei* isolates. *Diagnostic Microbiol Infection Disease*. 92(4), 311-314.
- Lamia, S. S., Shimo, M. S., Sakib, S., Rashed, B., Prima, A. A., & Mony, A. T. (2018). Phytochemistry and pharmacological properties of *Aegle marmelos* l (rutaceae) : A review. *International Journal of Research in Pharmacy and Pharmaceutical Sciences*, 3(3), 45–54.
- Laphookhieo, S., Phungpanya, C., Tantapakul, C., Techa, S., Tha-in, S., & Narmdorkmai, W. (2011). Chemical constituents from *Aegle marmelos*. *Journal of Brazilian Chemical Society*, 22(1), 176–178.

- Le, P. T. Q. (2019). Phytochemical screening and antimicrobial activity of extracts of *Cassia alata* L. leaves and seeds. *Bulgarian Chemical Communications*, 51(3), 378–383.
- Lei, J., Xu, J., & Wang, T. (2018). *In vitro* susceptibility of *Candida* species to fluconazole, itraconazole and voriconazole and the correlation between triazoles susceptibility: Results from a five-year study. *Journal of Medical Mycology*, 28(2), 310–313.
- Limsuwan, S., & Voravuthikunchai, S. (2013). Anti *Streptococcus pyogenes* activity of selected medicinal plant extracts used in thai traditional medicine. *Tropical Journal of Pharmaceutical Research*, 12(4), 535–540.
- Lino A, & Deogracious O. (2006). The *in-vitro* antibacterial activity of *Annona senegalensis*, *Securidacca longipendiculata* and *Steganotaenia araliacea* – Ugandan medicinal plants. *African Health Sciences*, 6, 31–35.
- Liu, Y., Kang, M., Ye, H., Zong, Z., & Lv, X. (2018). Analysis on clinical characteristics and drug resistance of *Candida parapsilosis* bloodstream infections in West China Hospital from 2012 to 2015. *Journal of Mycology Medical*, 28(1), 222–226.
- Lotfali E, Kordbacheh P, Mirhendi H, Zaini, F., Gharaji, A., Mohammadi, R., Noorbakhsh, F., Moazeni, M., Fallahi, A., Rezaie, S. (2016). Antifungal susceptibility analysis of clinical isolates of *Candida parapsilosis* in Iran. *Iran Journal of Public Health*, 45(3), 322–328.
- Lv, Q., Yan, L., & Jiang, Y. (2016). The synthesis, regulation and functions of sterols in *Candida albicans*: Well-known but still lots to learn. *Virulence*, 7(6), 649–659.
- Mabhiza, D., Chitemerere, T., & Mukanganyama, S. (2016). Antibacterial properties of alkaloid extracts from *Callistemon citrinus* and *Vernonia adoensis* against *Staphylococcus aureus* and *Pseudomonas aeruginosa*. *International Journal of Medicinal Chemistry*, 1–7, 6304163.
- Maccallum, D. M. (2012). Hosting infection : Experimental models to assay *Candida* virulence. *International Journal of Microbiology*, 363764.
- Mahlo, S. M., Chauke, H. R., McGaw, L., & Eloff, J. (2016). Antioxidant and antifungal activity of selected medicinal plant extracts against phytopathogenic fungi. *African Journal of Traditional, Complementary and Alternative Medicines*, 13(4), 216–222.
- Mailafia, S., Nagappan, D., & Bamaïyi, P. H. (2017). Phenotypic and molecular characterization of *Escherichia coli* isolates from Japanese quails (*Coturnixcoturnix japonica*) in Kelantan, Malaysia. *IOSI Journal of Microbiology, Biotechnology and Food Science*, 3(1), 1–8.
- Maity, P., Hansda, D., Bandyopadhyay, U., & Mishra, D. K. (2009). Biological activities of crude extracts and chemical constituents of bael, *Aegle marmelos* (L.) Corr.

- Mallikarjun, S., Rao, A., Rajesh, G., Shenoy, R., & Pai, M. (2016). Antimicrobial efficacy of Tulsi leaf (*Ocimum sanctum*) extract on periodontal pathogens: An *in vitro* study. *Journal of Indian Society of Periodontology*, 20(2), 145–150.
- Manandhar S., Luitel S., & Dahal R., K. (2019). *In vitro* antimicrobial activity of some medicinal plants against human pathogenic bacteria. *Journal of Tropical Medicine*, 2013,1-5.
- Mandrone, M., Bonvicini, F., Lianza, M., Sanna, C., Maxia, A., Gentilomi, G. A., & Poli, F. (2019). Sardinian plants with antimicrobial potential. Biological screening with multivariate data treatment of thirty-six extracts. *Industrial Crops and Products*, 137(5), 557–565.
- Manish, K. (2022). Ecological informatics medicinal plants in peril due to climate change in the Himalaya. *Ecological Informatics*, 68(11), 101546.
- Marak, M. B., & Dhanashree, B. (2018). Antifungal susceptibility and biofilm production of *Candida* spp. isolated from clinical samples. *International Journal of Microbiology*, 1-5.
- Martinez de Tejada, G., Sanchez-Gomez, S., Razquin-Olazarán, I., Kowalski, I., Kaconis, Y., Heinbockel, L., Brandenburg, K. (2012). Bacterial cell wall compounds as promising targets of antimicrobial agents I. Antimicrobial peptides and lipopolyamines. *Current Drug Targets*, 13(9), 1121–1130.
- Mazzariol, A., Bazaj, A., & Cornaglia, G. (2017). Multi-drug-resistant Gram-negative bacteria causing urinary tract infections: a review. *Journal of Chemotherapy*, 29, 2–9.
- Mishra, B. B., Kishore, N., Tiwari, V. K., Singh, D. D., & Tripathi, V. (2010). A novel antifungal anthraquinone from seeds of *Aegle marmelos* Correa (family Rutaceae). *Fitoterapia*, 81(2), 104–107.
- Mohamad, T. A. S. T., Islahudin, F., Jasamai, M., & Jamal, J. A. (2019). Preference, perception and predictors of herbal medicine use among Malay women in Malaysia. *Patient Preference and Adherence*, 13, 1829–1837.
- Mohammed, M. M. D., Ibrahim, N. A., El-Sakhawy, F. S., Mohamed, K. M., & Deabes, D. A.-H. (2016). Two new cytotoxic furoquinoline alkaloids isolated from *Aegle marmelos* (Linn.) Correa. *Natural Product Research*, 30(22), 2559–2566.
- Mohanasundaram, S., Doss, V. A., Haripriya, G., Varsha, M., Daniya, S., & Madhankumar. (2017). GC-MS analysis of bioactive compounds and comparative antibacterial potentials of aqueous, ethanolic and hydroethanolic extracts of *Senna alata* L. against enteric pathogens. *International Journal of Research in Pharmaceutical Sciences*, 8(1), 22–27.
- Mohideen, S., Sasikala, E., & Aruhaj, P. (2005). Pharmacognosy of *Cassia alata* Linn -

- leaves. *Ancient Science of Life*, 24(4), 192-198.
- Monika, S., Thirumal, M., Kumar, P., R.(2023). Phytochemical and biological review of *Aegle marmelos* Linn. *Future Science OA*, 9(3),3-14.
- Moradali, M. F, Ghods, S. & Rehm, BHA. (2017) *Pseudomonas aeruginosa* lifestyle: A paradigm for adaptation, survival, and persistence. *Frontiers Cellular Infection Microbiology*. 7(39),1-29.
- Mordi, R. M., Osula, A. N., Igeleke, C. L., Odjadjare, E. E., Oboh, F. O. J., & Uwadiae, E. O. (2016). The evaluation of the antimicrobial property of *Cassia alata* leaves in Benin city, Nigeria. *Biomedicine and Nursing*, 2(3), 14–23.
- More, Y. D., Gade, R. M., & Shitole, A. V. (2017). Evaluation of antifungal activities of extracts of *Aegle marmelos*, *Syzygium cumini* and *Pongamia pinnata* against *Pythium debaryanum*. *Indian Journal of Pharmaceutical Sciences*, 79(3), 377–384.
- Muniyan, A. S., & Anandhan, A. S. (2015). Evaluation of antifungal activity of crude leaf extracts of indian sacred trees. *Journal of Pharmaceutical, Chemical and Biological Sciences*, 3(8), 240–246.
- Murugan, M., & Kamaraj, M. (2018). Phytochemical screening and antimicrobial activity of medicinal plant-*hybunthus enneaspermus* L. F. muell. *Journal of Drug Delivery and Therapeutics*, 8(4), 323–330.
- Nanasombat, S., Kuncharoen, N., Ritcharoon, B., & Sukcharoen, P. (2018). Antibacterial activity of thai medicinal plant extracts against oral and gastrointestinal pathogenic bacteria and prebiotic effect on the growth of *Lactobacillus acidophilus*. *Chiang Mai Journal of Science*, 45(1), 33–44.
- Nayak, B. K., Mukilarasi, V., & Nanda, A. (2015). Antibacterial activity of leaf extract of *Cassia alata* separated by soxhlet extraction method. *Der Pharmacia Lettre*, 7(4), 254-257.
- Ncube B., Finnie J.F., & Van Staden J. (2012). *In vitro* antimicrobial synergism within plant extract combinations from three South African medicinal bulbs. *Journal of Ethnopharmacology*, 139(1),81-89.
- Neji, S., Hadrich, I., Trabelsi, H., Abbes, S., Cheikhrouhou, F., Sellami, H., Ayadi, A. (2017). Virulence factors, antifungal susceptibility and molecular mechanisms of azole resistance among *Candida parapsilosis* complex isolates recovered from clinical specimens. *Journal of Biomedical Science*, 24(1), 1–16.
- Nemkul, C. M., Bajracharya, G. B., & Shrestha, I. (2018). Phytochemical , antibacterial and DPPH free radical scavenging evaluations of the barks of *Aegle marmelos* (L.) Correa. *Journal of Pharmacognosy and Phytochemistry*, 7(4), 1637–1641.
- Nikaein, D., Sharifzadeh, A., & Khosravi, A. R. (2018). Fungicidal versus Fungistatic activity of five Iranian essences against fluconazole resistant *Candida* species.

- Oladeji, O. S., Adelowo, F. E., Oluyori, A. P., & Bankole, D. T. (2020). Ethnobotanical description and biological activities of *Senna alata*. *Evidence-Based Complementary and Alternative Medicine*, 1-12.
- Otto, R., B., D., Ameso, S., & Onegi, B. (2014) Assessment of antibacterial activity of crude leaf and roots of *Cassia alata* against *Neisseria gonorrhea*. *Africans Health Science*.14(4),840-848.
- Owoyale, J., Olatunji, G., & Oguntoye, S. (2006). Antifungal and antibacterial activities of an alcoholic extract of *Senna alata* leaves. *Journal of Applied Sciences and Environmental Management*, 9(3),105-107.
- Parihar N., & Kumar S. (2013). Study of antifungal potential of *Aegle marmelos*: a medicinal plant. *International Journal of Plant, Animal and Environmental Science*, 3(1),126-129.
- Paterson, G. K., Mitchell, T. J. (2004). The biology of gram positive sortase enzyme. *Trends Microbiology*, 12(2),89-95
- Patil, R. H., Bhushanb, C., & Sailaxmic, S. (2009). Antifungal and antiaflatoxicogenic activity of *Aegle marmelos* L. *Pharmacognosy Journal*, 1(4), 298–301.
- Patil, S., & Muthusamy, P. (2020). A bio-inspired approach of formulation and evaluation of *Aegle marmelos* fruit extract mediated silver nanoparticle gel and comparison of its antibacterial activity with antiseptic cream. *European Journal of Integrative Medicine*, (33), 101025.
- Patil, S., Venckatesh, R., & Seenivasan, R. (2015). Green synthesis of silver nanoparticle from leaf extract of *Aegle marmelos* and evaluation of its antibacterial activity. *International Journal of Pharmacy and Pharmaceutical Sciences*, 7(6), 169–173.
- Paul, B., Mitra, P., Ghosh, T., Salhan, R. N., Chakrabarti, A., Singh, T. A., & Mitra, P. K. (2012). Effect of leaves of *Cassia alata* Linnaeus (*Caesalpinaceae*) on the growth of *Staphylococcus aureus* Rosenbach. *Pleione*, 6(2), 341–347.
- Perumal, A., Krishna, S., & Madhusree, M. (2018). GC-MS analysis, antioxidant and antibacterial activities of ethanol extract of leaves of *Aegle Marmelos* (L.) Correa. *Journal of Drug Delivery and Therapeutics*, 8(4), 247–255.
- Pesewu, G. A., Cutler, R. R., & Humber, D. P. (2008). Antibacterial activity of plants used in traditional medicines of Ghana with particular reference to MRSA. *Journal of Ethnopharmacology*, 116(1), 102–111.
- Pfaller, M. A., Messer, S. A., Boyken, L., Rice, C., Tendolkar, S., Hollis, R. J., Diekema, D. J. (2003). Caspofungin activity against clinical isolates of fluconazole-resistance *Candida*. *Journal of Clinical Microbiology*, 41(12), 5729–5731.

- Phongpaichit, S., Pujenjob, N., Rukachaisirikul, V., & Ongsakul M. (2004). Antifungal activity from leaf extracts of *Cassia alata* L., *Cassia fistula* L. and *Cassia tora* L. *Songklanakarin Journal of Science*, 26(5),741-8.
- Poonkothai, M., & Saravanan, M. (2008). Antibacterial activity of *Aegle marmelos* against leaf, bark and fruit extracts. *Ancient Science of Life*, 27(3), 15–158.
- Poowanawittayakom, N., Dutta, A., Stock, S., Touray, S., Ellison, R. T., & Levitz, S. M. (2018). Reemergence of intravenous drug use as risk factor for candidemia, Massachusetts, USA. *Emerging Infectious Diseases*, 24(4), 631–637.
- Prateeksha, Yusuf, M.A., Singh, B.N., Sudheer, S., Kharwar, R. N., Siddiqui, S., Abdel-Azeem, A.M., Fernandes F.L., Dashora, K., Gupta, V.K. (2019). Chrysophanol: A natural anthraquinone with multifaceted biotherapeutic potential. *Biomolecules*, 9(2), 68,2-24.
- Pryszcz L. P., Németh, T., Gácsér, A., Gabaldón, T. (2013). Unexpected genomic variability in clinical and environmental strains of the pathogenic yeast *Candida parapsilosis*. *Genome Biology Evolution*, 5(12), 2382-92.
- Qadri, H., Haseeb Shah, A., Mudasir Ahmad, S., Alshehri, B., Almilaibary, A., Ahmad Mir, M. (2022) “Natural products and their semi-synthetic derivatives against antimicrobial-resistant human pathogenic bacteria and fungi,” *Saudi Journal of Biological Sciences*, 29(9), 103-376.
- Raabe, V. N & Shane, A. L. (2019). Group B *Streptococcus* (*Streptococcus agalactiae*). *Microbiology Spectrum*, 7(2), 1-22.
- Rahmatullah, M., Jahan, R., Khatun, M. A., Jahan, F. I., Azad, A. K., Bashar., A.B.M., Miajee Z.U.M., Ahsan. S., Nahar. N., Ahmad. I., & Chowdhury, M.H. (2010). A pharmacological evaluation of medicinal plants used by folk medicinal practitioners of Station Purbo Para Village of Jamalpur Sadar Upazila in Jamalpur district, Bangladesh. *American-Eurasian Journal Sustain Agriculture*, 4, 170-195.
- Rahmawati, F., Prihantini, N. N., & Hady, B. C. (2022). *In vitro* bioactivity test of *Senna alata* (L.) roxb leaves extract. *International Journal of Health Sciences and Research*, 12(2), 304–317.
- Raja, S. B., Murali, M. R., Kumar, N. K., & Devaraj, S. N. (2011). Isolation and partial characterisation of a novel lectin from *Aegle marmelos* fruit and its effect on adherence and invasion of *Shigellae* to HT29 cells. *PLoS ONE*, 6(1), 1–9.
- Rajendran, R., Mowat, E., Jones, B., William, C., & Ramage, G. (2015) Prior *in vitro* exposure to voriconazole confers resistance to amphotericin B in *Aspergillus fumigatus* biofilms. *International Journal of Antimicrobiol Agents*, 46:342–345.

- Raji, A. A., Jummai, K. Z., Busu, M. S., Ya'aba, Y., & Kolo, I. (2019). *In-vitro* antimicrobial susceptibility and phytochemical constituents of methanol leaf extract of *Prosopis africana* against some selected microorganisms. *Journal of Advances in Microbiology*, 18(1), 1–8.
- Ramdani, D, Marjuki, M. & Chuzaemi, S. (2017). Pengaruh Perbedaan jenis pelarut dalam proses ekstraksi buah Mengkudu (*Morinda citrifolia* L.) pada pakan terhadap viabilitas protozoa dan produksi gas *in-vitro*, *Jurnal Ilmu-Ilmu Peternakan*, 27(2), 54-62.
- Rana, B. K.; Singh, U. P. & Taneja, V. 1997. Antifungal activity of kinetics of inhibition by essential oil solated rom leaves of *Aegle marmelos*, *Journal of Ethnopharmacology*, 57, 29 – 34.
- Rautela, R., Das, G. K., Khan, F. A., Prasad, S., Kumar, A., Prasad, J. K., Srivastava, S. K. (2018). Antibacterial, anti-inflammatory and antioxidant effects of *Aegle marmelos* and *Murraya koenigii* in dairy cows with endometritis. *Livestock Science*, 214 (2), 142-148.
- Razmavar, S., Abdulla, M. A., Ismail, S. B., & Hassandarvish, P. (2014). Antibacterial activity of leaf extracts of *Baeckea frutescens* against methicillin-resistant *Staphylococcus aureus*. *BioMed Research International*, 1-5.
- Reda, F. M., El-Zawahry, Y., & Omar, A. (2017). Synergistic effect of combined antibiotic and methanol extract of *Eucalyptus camaldulensis* leaf against *Staphylococcus aureus* and *Pseudomonas aeruginosa*. *International Journal of Applied Sciences and Biotechnology*, 5(4), 486–497.
- Rizwana, H., Fatimah, A., Alharbi, R. I., Albasher, G., Nadine, M. S., & Alqusumi, R. (2021). Morphology and ultrastructure of some pathogenic fungi altered by lead extract of senna alexandrina mill. *Pakistan Journal of Argiculture Science*, 58(1), 389-408.
- Kupczynski, R., Szumny, A., Bednarski, M., Piasecki, T., Spitalniak-Bajerska, K., & Roman, A. (2019). Application of *Potentilla anserine*, *Polygonum aviculare* and *Rumex crispus* mixture extracts in a rabbit model with rxperimentally induced *E. coli* infection. *Animals*, 9(774), 2-5
- Sabran, S. F., Mohamed, M., & Abu Bakar, M. F. (2016). Ethnomedical knowledge of plants used for the treatment of tuberculosis in Johor, Malaysia. *Evidence-Based Complementary and Alternative Medicine*, 1-12.
- Saito, S. T., Trentin, D. D. S., MacEdo, A. J., Pungartnik, C., Gosmann, G., Silveira, J. D. D., & Brendel, M. (2012). Bioguided fractionation shows *Cassia alata* extract to inhibit *Staphylococcus epidermidis* and *Pseudomonas aeruginosa* growth and biofilm formation. *Evidence-Based Complementary and Alternative Medicine*, 1-13.

- Samaranayake, Y. H., & Samaranayake, L. P. (1994). *Candida krusei*: Biology, epidemiology, pathogenicity and clinical manifestations of an emerging pathogen. *Journal of Medical Microbiology*, 41(5), 295–310.
- Sankar Ganesh, P., & Ravishankar Rai, V. (2018). Attenuation of quorum-sensing-dependent virulence factors and biofilm formation by medicinal plants against antibiotic resistant *Pseudomonas aeruginosa*. *Journal of Traditional and Complementary Medicine*, 8(1), 170-177.
- Santhanam, J., Yahaya, N., & Aziz, M. N. (2013). Species distribution and antifungal susceptibility patterns of *Candida* species: Is low susceptibility to itraconazole a trend in Malaysia. *Medical Journal of Malaysia*, 68(4), 343–347.
- Saravanasingh, K., Frdrik, P. G., & Ramamurthy, M. (2016). A study on antibacterial and antifungal activities of extracts of medicinal plant *Aegle marmelos*. *International Journal of Advanced Research in Biological Sciences*, 3(2), 321–328.
- Sarkar, & Solanki. (2011). Isolation, characterization and antibacterial activity of leaves extract of bael (*Aegle marmelos*). *International Journal of Pharmacy and Life Sciences*, 2(12), 1303–1305.
- Sedighi, N. S., Salari, S., & Izadi, A. R. (2017). Evaluation of antifungal effect of iron-oxide nanoparticles against different *Candida* species. *The Institute of Engineering and Technology Nanobiotechnology*, 11(7), 883–888.
- Selvi, V., Isaivani, I., & Karpagam, S. (2012). Studies on antimicrobial activities from flower extract of *Cassia alata* Linn. *International Journal of Current Science*, 2, 299–303.
- Sen, A., & Batra, A. (2012). Determination of antimicrobial potentialities of different solvent extracts of the medicinal plant: *Phyllanthus amarus* Schum. and Thonn. *International Journal of Green Pharmacy*, 6(1), 50–56.
- Senthilkumar, R. P., Malayaman, V., & Sindhuja, S. (2013). Phytochemical screening and antibacterial evaluation of the leaf, flower and seed coat extracts of *Cassia alata* L. *Journal of Chemical & Pharmaceutical Research*, 5(11), 740–744.
- Seyfarth, F., Schliemann, S., Elsner, P., & Hipler, U. C. (2008). Antifungal effect of high- and low-molecular-weight chitosan hydrochloride, carboxymethyl chitosan, chitosan oligosaccharide and N-acetyl-d-glucosamine against *Candida albicans*, *Candida krusei* and *Candida glabrata*. *International Journal of Pharmaceutics*, 353(1–2), 139–148.
- Seyedmousavi, S., Bosco, S. M. G, de Hoog, S., Ebel, F., Elad, D., Gomes, R. R., Jacobsen, I. D., Jensen, H. E., Martel, A., Mignon, B., Pasmans, F., Piecková, E., Rodrigues, A. M., Singh, K., Vicente, V.A., Wibbelt, G., Wiederhold, N. P., Guillot, J. (2018). Fungal infections in animals: a patchwork of different situations. *Medical Mycology*, 1(56), 165-187.

- Shantaram, B. P., Ramachandra, B. S., & Eknath, G. A. (2016). Preliminary phytochemical analysis of seeds and leaves of *Aegle marmelos* extracts and in-vitro assessment of their antibacterial activity. *International Journal of Pharma Research and Health Sciences*, 4(4), 1315–1319.
- Sharma, J., & Kumar, P. (2017). Comparative study of antimicrobial activity and phytochemical screening of serial extracts from leaves and fruit of *Aegle marmelos* and *Carica papaya*. *International Journal of Pharmacy & Pharmaceutical Sciences*, 9(12), 119.
- Sharma, K., & Chauhan, E. S. (2017). Nutritional and phytochemical evaluation of fruit pulp powder of *Aegle marmelos* (Bael). *Journal of Chemical and Pharmaceutical Sciences*, 10(2), 809–814.
- Sharma, N., & Dubey. (2013). History and Taxonomy of *Aegle marmelos*: A Review. *International Journal of Pure and Applied Bioscience*, 1(6), 7-13.
- Sharmin, M., Nur, I.T., Acharjee, M., Munshe, S. K & Noor, R. (2014). Microbiological profiling and the demonstration of *in vitro* antibacterial traits of the major oral herbal medicines used in DhakaMetropolis. *Springer Plus*, 3, 739.
- Shin, M. G., Suh, S. P. A. L., & Ryang, D. W. (2005). Differences in biofilm production by three genotypes of *Candida parapsilosis* from clinical sources. *Medical Mycology*, 43(7), 657–661.
- Shiv, G., Gaherwal, S., Shrivastava, C. S., Wast, N., Holkar, G., College, S., & Indore, M. P. (2015a). Antifungal activity of *Aegle marmelos*, *Calotropis procera* and *Solanum xanthocarpum* extract against *Aspergillus niger*, *Candida albicans* and *Phenerochaete chrysosporium*. *European Journal of Experimental Biology*, 5(2), 81–89.
- Siddiqui, Z.N., Farooq, F., Musthafa, T.N.M., Ahmad, A., Khan, A. U. (2013). Synthesis, characterization and antimicrobial evaluation of novel halopyrazole derivatives. *Journal of Saudi Chemical Society*, 17(2), 237–43.
- Silva, S., Negri, M., Henriques, M., Oliveira, R., Williams, D. W., Azeredo, J. (2011). Adherence and biofilm formation of non-*Candida albicans* *Candida* species. *Trends Microbiology*, 19, 241–247.
- Singh, S., Sobel, J. D., Bhargava, P., Boikov, D., & Vazquez, J. A. (2002). Vaginitis due to *Candida krusei* : *Epidemiology, Clinical Aspects, and Therapy*, 48201, 1066–1070.
- Sivananthan, M., Elamaram, M., Tras, J., Kemacahaya, J., & Kemacahaya, T. (2013). *In vitro* evaluation of antibacterial activity of chloroform extract *Andrographis paniculata* leaves and roots, *Durio zibethinus* wood bark and *Psidium guajava* leaves against selected bacterial strains Department of Biomedical Science, Faculty of Biomedicine. *International Journal of Biomolecules and Biomedicine*, 3(1), 12–19.

- Sivaraj, R., Balakrishnan A., Thenmozhi, M., Venckatesh, R., (2011). Antimicrobial activity of *Aegle marmelos*, *Ruta graveolens*, *Opuntia delini*, *Euphorbia royleana* and *Euphorbia antiquorum*, *Journal of Pharmacy Research* 4(5), 1507–1508.
- Somchit, M. N., Reezal, I., Nur, I. E., & Mutalib, A. R. (2003). *In vitro* antimicrobial activity of ethanol and water extracts of *Cassia alata*, *Journal of Ethnopharmacology*, 84, 1–4.
- Somu, C., Karuppiyah, H., & Sundaram, J. (2019). Antiviral activity of seselin from *Aegle marmelos* against nuclear polyhedrosis virus infection in the larvae of silkworm, *Bombyx mori*. *Journal of Ethnopharmacology*, 245(5), 112155.
- Su, P. W., Yang, C. H., Yang, J. F., Su, P. Y., & Chuang, L. Y. (2015). Antibacterial activities and antibacterial mechanism of *Polygonum cuspidatum* extracts against nosocomial drug-resistant pathogens. *Molecules*, 20(6), 11119–11130.
- Su, Y., Feng, J., Liu, C., Li, W., Xie, Y., & Li, A. (2017). Dynamic bacterial colonization and microscopic lesions in multiple organs of tilapia infected with low and high pathogenic *Streptococcus agalactiae* strains. *Aquaculture*, 471, 190–203.
- Suja, K. P., Jose, L., Lakshmanan, Di., Vidya, A. G., Reshmi Nair, R. J., & Kumar, R. A. (2017). Isolation and characterization of antimycobacterial compounds from fruits of *Aegle marmelos* (L.) correa. *Journal of Communicable Diseases*, 49(4), 32–38.
- Sujatha, J., & Asokan, S. (2017). Antidermatophytic and antioxidant activity of hexane extracts of *Cassia alata* leaves and its phytochemical screening. Antidermatophytic and antioxidant activity. *International Research Journal of Pharmaceutical and Biosciences*, 4(5), 25–35.
- Sujatha, J., Rajesh Kumar, S., & Lakshmi, T. (2019). Antidermatophytic, the anticancer and antioxidant activity of *Cassia alata* ethanolic extract and its phytochemical analysis. *International Journal of Research in Pharmaceutical Sciences*, 10(2), 838–845.
- Sukenda, Achmad S., Muhammad Zairin, Jr., Angela, M.L, Sri Nuryati., & Dendi Hidayatullah (2021). Virulence gene profiling and pathogenicity of *Streptococcus agalactiae* isolated from tilapia, *Oreochromis niloticus* farms in Indonesia. *Journal of Indonesian Aquaculture*, 16(2), 119–125.
- Sulaiman, L., Abidin, Z. H. Z., Yunus, S. A. M., Mazni, N. A., Hadi, A. N., Arof, A. K., & Simarani, K. (2019). Dual functions of Cu (NO³)(2) as antifungal and colour stabilizer for coating paint film consisting of chlorophyll. *Pigment and Resin Technology*, 50(2), 177–182.
- Sule, W. F., Okonko, I. O., Omo-Ogun, S., Nwanze, J. C., Ojezele, M. O., Ojezele, O. J., Alli J. A., Soyemi, E. T., & Olaonipekun, T. O. (2011). Phytochemical properties and *in-vitro* antifungal activity of *Senna alata* Linn. crude stem bark extract. *Journal of Medicinal Plants Research*, 5(2), 176–183.

- Summer, K, Browne, J, Hollanders, M., & Benkendorff, K. (2022). Out of control: The need for standardised solvent approaches and data reporting in antibiofilm assays incorporating dimethyl-sulfoxide (DMSO). *Biofilm*, 17(4),1-12.
- Sun, Y., Deng, Y., Kwapong, W. R., & Ma, L. (2019). Clinical distribution, drug resistance and risk factor analysis of *Pseudomonas aeruginosa* infections. *International Journal of Clinical and Experimental Medicine*, 12(10), 12322–12330.
- Tacconelli, E. (2009). Antimicrobial use: risk driver of multidrug resistant microorganisms in healthcare settings. *Current Opinion Infectious Diseases*, 22, 352–358.
- Tamba, M., Rocca, R., Prosperi, A., Pupillo, G., Bassi, P., Galletti, G., Martini, E., Santi, A., Casadei, G., & Arrigoni N (2022) Evaluation of control program against *Streptococcus agalactiae* infection in dairy herds during 2019–2021 in Emilia-Romagna region, Northern Italy. *Frontier in. Veterinary. Science*, 9,1-10.
- Timothy, S.Y., Lamu, F. W., Rhoda, A. S., Adati, R. G., Maspalma, I. D., Askira, M. (2012). Acute toxicity, phytochemistry and antibacterial activity of aqueous and ethanolic leaf extracts of *Cassia alata* linn. *International Research Journal of Pharmacy*, 3(6), 73-76.
- Trofa, D., Gácsér, A., & Nosanchuk, J. D. (2008). *Candida parapsilosis*, an emerging fungal pathogen. *Clinical Microbiology Reviews*, 21(4), 606–625.
- Vaou, N., Stavropoulou, E., Voidarou, C., Tsigalou, C., Bezirtzoglou, E. (2021). Towards advances in medicinal plant antimicrobial activity: A review study on challenges and future perspectives. *Microorganisms*, 9(10), 2-28.
- Veena, H., Navya, N., Sandesh, G. K., & Thippeswamy, N. B. (2019). Antibacterial and antidiarrheal activity of *Simarouba amara* (Aubl.) bark. *Journal of Applied Pharmaceutical Science*, 9(5), 88–96.
- Veljic, M., Tarbuk, M., Marin, P. D., Ciric, A., Sokovic, M., & Marin, M. (2008). Antimicrobial activity of methanol extracts of mosses from Serbia. *Pharmaceutical Biology*, 46(12), 871–875.
- Vijayakumar, A., & Sinthiya, A. (2015). Biosynthesis of phytochemicals coated silver nanoparticles using aqueous extract of leaves of *Cassia alata* – characterization, antibacterial and antioxidant activities. *International Journal of Pharmaceutical and Clinical Research*, 10(5), 138–149.
- Villasenor, I. M., Canlas, A. P., Pascua, M. P. I., Sabando, M. N., & Soliven, L. A. P. (2002). Bioactivity studies on *Cassia alata* L. leaf extracts. *Phytotherapy Research*, 16(S1), 93–96.
- Vincent, J., Marshall, J., Anzueto, A., Martin, C. D., & Gomersall, C. (2009). International study of the prevalence and outcomes of infection in intensive care units. *The Journal of American Medical Association*, 302(21), 2323–2329.

- Wahab, A., Rahman, N. B. A., Yahaya, M. L. Bin, Baharuddin, N. S. B., & Jamil, (2017). Antimicrobial effects of *Quercus infectoria* gall extract on the cell morphology of *Streptococcus agalactiae*, *Proteus vulgaris* and *Candida albicans*. *Malaysian Journal of Microbiology*, 13(1), 13–19.
- Waris, M. A. A., Farid, A. M., & Misra. (2021). Antifungal activity of *Cassia alata* L. ethyl acetate and n-hexane leaves extract against *Candida albicans*. *Journal of Public Health and Pharmacy*, 1(2), 27–29.
- Webster, D., Taschereau, P., Belland, R. J., Sand, C., & Rennie, R. P. (2008). Antifungal activity of medicinal plant extracts; preliminary screening studies. *Journal of Ethnopharmacology*, 115(1), 140–146.
- Whaley, S. G., Berkow, E. L., Rybak, J. M., Nishimoto, A. T., Barker, K. S., & Rogers, P. D. (2017). Azole antifungal resistance in *Candida albicans* and emerging non-albicans *Candida* species. *Frontiers in Microbiology*, 7(21), 1–12.
- Wuthi-Udomlert, M., Kupittayanant, P., & Gritsanapan, W. (2010). *In vitro* evaluation of antifungal activity of anthraquinone derivatives of *Senna alata*. *Journal of Health Research*, 24(3), 117–122.
- Yadav, N. P., & Chanotia, C. S. (2009). Phytochemical and pharmacological profile of leaves of *Aegle marmelos*. *The Pharmacology Review*, 144–150.
- Yadav, S. S., Dahiya, K., Ganie, S. A., & Gulia, S. K. (2015). Antibacterial activity of *Aegle marmelos* (L) Correa. *International Journal of Pharmacy and Pharmaceutical Sciences*, 7(3), 462–464.
- Yin, H., Deng, Y., Wang, H., Liu, W., Zhuang, X., & Chu, W. (2015). Tea polyphenols as an antivirulence compound disrupt quorum-Sensing regulated pathogenicity of *Pseudomonas aeruginosa*. *Scientific Reports*, 5(11), 1–11.
- Yuan, H., Ma, Q., Ye, L., & Piao, G. (2016). The traditional medicine and modern medicine from natural products. *Molecules*, 21(5), 559.
- Yunus Reni & Anita Rosanty. (2018.). Inhibition of sabandara leaves (*Cassia alata*) against malassezia fungi furfur. *Proceeding 1st International Conference Health Polytechnic of Kupang*, 582-591.