Quantifying myelin in neonates using magnetic resonance imaging: a systematic literature review

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This review aimed to assess the usefulness of various magnetic resonance imaging (MRI) techniques for the quantification of neonatal white matter myelination. The Scopus, PubMed, and Web of Science databases were searched to identify studies following the PRISMA (preferred reporting items for systematic reviews and metaanalyses) statement using quantitative MRI techniques to examine samples collected from neonates to quantify myelin. Twelve studies were ultimately included. The results demonstrated that in validation studies, relaxometry is the most frequently explored approach (83.33%), followed by magnetization transfer imaging (8.33%) and a new automatic segmentation technique (8.33%). Synthetic MRI is recommended for quantifying myelin in neonates because of several advantages that outweigh a few negligible limitations.

Key words: Brain, Magnetic resonance imaging, Myelin, Neonate, Quantitative

Key message

- **Question:** This systematic review attempts to discover the best magnetic resonance imaging (MRI) technique for myelin quantification in neonates by evaluating various MRI parameters and their reproducibility.
- **Finding:** Since the benefits of using synthetic MRI for quantifying myelin in neonates outweigh the very minor drawbacks, it is recommended.
- **Meaning:** The findings suggest the importance of identifying noninvasive MRI techniques available to assess myelin tissue in neonates, which aid in diagnosing neurodevelopmental disorders.

Introduction

White matter (WM) formation and myelination are essen-

tial neurodevelopmental processes. Myelination, the final stage of WM formation, is characterized by the formation of myelin, a segmented neural membrane, along the nerve fibers.¹⁾ These fibers continue to develop throughout childhood and adulthood in a predetermined and directional pattern called spatiotemporal development.²⁾ Myelination enhances the brain's ability to send and receive information in a swift and coordinated manner that is crucial for emotional tasks, movement coordination, decision-making, and other higher-order behavioral and cognitive processes.³⁾ This is made possible by maturation of the myelin sheath. *In vivo* studies of progressive WM development beyond the age of complete somatic growth were reported in 1993, and magnetic resonance imaging (MRI) was first used to study the corpus callosum (CC) in humans.⁴⁾

Myelination peaks after birth and throughout the first year of life, which means that any incident that causes brain injury around the time of delivery or during the perinatal period may impede this process and lead to neurodevelopmental deficits later.⁵⁾ Disorders such as epilepsy,⁶⁾ birth asphyxia,⁵⁾ dyslexia,⁷⁾ autism, attention deficit, hyperactivity, and schizophrenia³⁾ are being investigated for myelination issues. Moreover, injuries to the developing WM and consequent hypomyelination are common out comes of premature birth and other perinatal insults.⁸⁾

Despite a significant need for *in vivo* quantitative evaluation of myelination in a wide range of clinical conditions, including most neurodevelopmental disorders, there is currently no gold standard for myelin assessment.⁹⁾ Standard MRI and diffusion tensor imaging parameters⁹⁾ can be used to assess myelination; however, these approaches provide only indirect and non-specific details of myelin content because they might also represent other tissue features.

While histological investigations were the first to cha-

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Graphical abstract. Quantitative magnetic resonance imaging (MRI) techniques to quantify myelin in neonate.

racterize myelination patterns, the utility of T1- and T2weighted MRI has been established as a preliminary alternative for qualitatively evaluating myelination in utero and shortly after birth.⁸⁾ The myelin sheath is composed of various biochemical compounds; therefore, changes in the properties of brain tissue occur according to the degree of myelination. The main difference between T1and T2-weighted MRI images is the amount of cholesterol, which is assumed to indicate brain maturation, as changes in T1 and T2 relaxation times were linked to age in prior studies.¹⁰ These values were used to quantify myelination disruption in preterm and term infants, which exhibit different relaxation times. One cerebral abnormality seen on MRI of preterm infants at a term-equivalent age related to extremely preterm births is abnormal myelination in the posterior limb of the internal capsule (PLIC), which may indicate developmental delay in the newborn.⁸⁾

A quantitative assessment of myelination in infants was conducted approximately 20 years ago.¹¹ Furthermore, cutting-edge MRI techniques have shown that they can be used to evaluate myelination in the brains of people older than 3 months, including adults. Nonetheless, these approaches have not been frequently applied to study the brains of newborns younger than 3 months of age or critically ill neonates.⁵ The quantification of these myelin changes may increase our understanding of disease severity and the associated prognosis, demonstrate details regarding the spatial location of foci or lesions and the affected associated neural systems, and contribute to the development of a standard measure to assess treatment efficacy.³ However, a quantitative assessment of myelin progression is not possible using T1 and T2 signal intensity alone. A major gap in our understanding of the neurodevelopmental discipline is, therefore, the inability to visualize and quantitatively assess myelination using MRI in human infants or neonates. This systematic review aimed to identify the superior MRI technique for myelin quantification in neonates by evaluating the efficacy and reproducibility of various MRI parameters.

Methodology

1. Review protocol

This systematic literature review was conducted in accordance with the preferred reporting items for systematic reviews and meta-analyses (PRISMA) statement. The PRISMA reporting guidelines are recommended in healthcare journals to improve the transparency and clarity of how reviews are executed while minimizing the possibility of inaccurate reporting.¹²

2. Inclusion and exclusion criteria

The following inclusion criteria were used to select the studies: (1) neonates as study subjects, (2) quantitative studies involving MRI, (3) publication in 2012–2022, (4) publication in English, and (5) peer-reviewed status.

The authors included only studies of full- and preterm neonates. The authors did not exclude studies of neonates who presented with a medical illness or disease. Full-text, peer-reviewed articles published in English were selected to ensure review quality and avoid confusion and misunderstanding. Gray literature was excluded to ensure study reliability since this review focused on published articles only. This also facilitated the eligibility determination and data extraction processes, especially in the methods and discussion sections, which are lacking from the abstracts.

Infants and young children aged 12-121 months were the subjects of a study¹¹⁾ that assessed the myelination of frontal lobe changes published in 2004. In 2011, the first quantitative study¹³⁾ was published on myelination in healthy human neonates aged 3-11 months. However, attempts in newborns have remained limited. Therefore, the publication timeline of 2011-2021 was selected as an inclusion criterion. Early attempts to quantify myelin content in humans experienced a number of limitations, especially long scanning times, which can result in a high specific absorption rate and are not widely available,¹⁴⁾ which is not ideal for implementation in neonatal scanning. Another significant drawback of the early studies was achieving good image resolution, owing to the low myelin content present in the neonatal brain.¹⁵⁾ Furthermore, this review aimed to indirectly assist in determining which methods are available and suitable for addition to the baseline MRI protocol in clinical settings.

3. Systematic searching strategies

We conducted a systematic search of the Scopus, PubMed, and Web of Science databases. The Scopus and Web of Science databases have the potential to be leading databases in a systematic literature review owing to their sophisticated search capabilities, comprehensiveness (indexing more than 5,000 publishers), control over article quality, and multidisciplinary focus, which includes environmental management studies.^{16,17)} In contrast, one significant feature of PubMed that its contents are not replicated by Scopus or Web of Science and is readily updated with literature that has been presented online in an early edition before print publication by a variety of journals in addition to literature that has been printed.¹⁸⁾ The search processes in these 3 databases retrieved a total of 397 open-access articles.

Keywords included Medical Subject Headings terms and text words developed from the authors' discussion, keywords suggested by Scopus, an online thesaurus, and past literature. The searches included all meaningful combinations of the following terms: (1) myelin (WM), (2) MRI, and (3) neonates (newborn or infant). The authors enriched the existing keywords and generated comprehensive search strings (based on the Boolean operator, phrase searching, truncation, wild card, and field code operations) in the 3 databases.

This study screened all 397 selected articles by selecting the criteria for article selection, which were performed automatically based on the sorting function available in the databases. Another 3 studies were identified and manually added to one of the selected articles. The selection criteria were based on the population or patient, intervention, comparator or control, and outcome (PICO) framework or the research question. Since it would be nearly impossible for authors to review all published papers, results from a study by Okoli¹⁹⁾ were used to define the range of time that the authors could review. Because the search process began on May 1, 2022, and ended on July 31, 2022, and the year had not yet ended, the search was limited to 2021. As a result, the period of 2011–2021 was chosen as an inclusion criterion for the search strategy.

During the first screening process, 173 duplicate articles were removed. Ninety articles were excluded after the title screening, while 25 articles were excluded during the abstract screening. After a thorough full-text screening process, 100 articles were excluded for not fitting the PICO framework or research questions. Studies were excluded if nonneonate subjects were involved (e.g., adults, animals, and cadavers), insufficient or nonquantitative techniques involving MRI for the myelin assessment were used, or full-text articles were unavailable. Histology was used to study myelination during infancy; however, postmortem data cannot determine the longitudinal course of WM development.²⁰⁾ Atlas-based analyses, review articles, structural segmentation, and scoring system establishment studies were excluded. The remaining 12 articles were subjected to the eligibility process in which the authors carefully monitored the retrieved literature to ensure that all met the requirements.

4. Quality appraisal

Two authors employed the Quality Assessment of Diagnostic Accuracy Study 2 (QUADAS-2) tool to evaluate the methodological quality of the studies included in this review.²¹⁾ The tool includes 14 questions used to assess the risk of bias and applicability in terms of patient selection, index test, reference standard, and flow and timing domains. The risk of bias for each domain was rated as low, unclear, or high. The total risk of assessment was determined by evaluating all areas and quality of each study. The index test for this review was MRI, and the reference standard was myelin. Bias assessment for reproducibility studies consisted of the same methods used for quantifying myelin except for the reference standard from the QUADAS-2 instrument. If many questions in a study scored positively in that domain, the study was considered to have a low risk of bias in that domain. The risk of bias was generally low across all included articles (Table 1).

Results

1. Study identification and selection

Fig. 1 summarizes the search results. Of the 397 primary

Table 1. Quality assessment of diagnostic accuracy study 2 as	ssessment results
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		Applicability concerns					
Study	Patient selection	Index test	Reference standard	Flow and timing	Patient selection	Index test	Reference standard
Olivieri et al. (2021) ⁵⁾	٢	٢	N/A	0	٢	٢	?
Natu et al. (2021) ²⁶⁾	٢	\odot	N/A	٢	٢	\odot	?
Wang et al. (2019) ⁸⁾	٢	\odot	N/A	٢	٢	\odot	?
Schmidbauer et al. (2019) ²⁷⁾	٢	\odot	N/A	٢	٢	\odot	?
Knight et al. (2018) ²⁸⁾	٢	\odot	N/A	٢	٢	\odot	?
Lee et al. (2018) ²⁴⁾	٢	\odot	N/A	٢	٢	\odot	?
Soun et al. (2017) ¹⁴⁾	٢	٢	N/A	٢	٢	\odot	?
Schneider et al. (2016) ²⁹⁾	٢	\odot	N/A	٢	٢	\odot	?
Melbourne et al. (2016) ³⁰⁾	٢	٢	N/A	٢	٢	\odot	?
Ning et al. (2014) ²²⁾	٢	\odot	N/A	٢	٢	\odot	?
Nossin-Manor et al. (2013) ²⁵⁾	٢	٢	N/A	٢	٢	٢	?
Deoni et al. (2012) ²³⁾	٢	\odot	N/A	٢	3	\odot	?

③, low risk; N/A, not available; ?, unclear risk.



Fig. 1. PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) flowchart.

data articles, 173 were duplicates. Subsequently, 115 irrelevant articles were removed based on the title and abstract screening. In addition, 100 articles were excluded after the full-text screening because of noncompliance with the research question. Another 3 studies by Olivieri et al.⁵⁾ were deemed compliant with the PICO framework or research question^{14,22,23)} and added manually. Consequently, 12 studies fulfilled the inclusion criteria and were included in the data analysis.^{5,8,14,22–30)} Overall, the selected studies had a low risk of bias.

2. Data extraction

The data extracted from the studies included author names, study location, study period, study design, sample size, sex, average gestational age, gestational age at MRI, MRI modality, MRI metrics, scanning time, and findings. Table 2 shows these variables. Table 3 summarizes the advantages and limitations mentioned in each study as well as the quantitative MRI sequence or technique and software or postprocessing methods used therein. Two authors independently selected the studies to review, extract, and analyze the data and evaluated the quality of each investigation. Overall, all studies provided adequate findings for quantifying myelin in neonates.

The frequency of each region of interest (ROI) mentioned in the studies was calculated (Table 4). These locations represent regional variations in the maturation process of the developing brain. The PLIC is the most frequently mentioned ROI in the literature (83.33% of all relevant studies), followed by the CC (genu and splenium) at 66.67% and optic radiation at 58.33%. Imaging and histological investigations indicate that by term birth, the PLIC is generally highly myelinated compared with other structures; however, the optic radiation possesses low levels of myelin.^{2,31-35}

3. Data synthesis

Olivieri et al.⁵⁾ enrolled neonates with neonatal encephalopathy as well as healthy term neonates as controls. Schmidbauer et al.²⁷⁾ included neonates with hypoxicischemic encephalopathy, intraventricular hemorrhage, or epileptic seizures as the study subjects. Knight et al.²⁸⁾ included groups at an increased risk of developing neurodevelopmental disorders. Meanwhile, the remaining studies ^{8,14,22–26,29,30)} recruited only cerebral low-risk neonates with no history of neurological conditions and excluded those with abnormalities detected on MRI or ultrasound.

The scanning time for the particular quantitative MRI sequences was less than 5 minutes in 3 studies,^{5,22,29} more than 5 minutes in 5 studies^{23-25,29,30} and not mentioned in another 4 studies.^{8,14,28,30} The acquisition time is a concern

Table 2. Data extracted from the literature regarding the main variables of interest

Stud	ly	Place/ country	Study period	Type of study design	Sam- ple	Gender (male: female)	Average gestational age	Age at MRI (wk)	MRI modality	MRI mea- sures	Scann time (qu tative M sequer	ing Janti- VRI NCe)	Findings
					512C		(WK)			(S)	<5 min	≥5 min	
Olivi (20	eri et al. 121) ⁵⁾	Canada, North America	2014- 2018	Prospective longitudinal study	39	10:6 (neonates with NE treated with hypothermia de- veloping brain in- jury) 10:7 (neonates with NE treated with hypothermia not developing brain injury) 4:6 (healthy, term neonates)	 39.33 (neonates with NE treated with hypother- mia developing brain injury) 38.99 (neonates with NE treated with hypother- mia not develop- ing brain injury) 38.73 (healthy, term neonates) 	36–42 (neo- nates with NE 38–40 (heal- thy, term neonates)	Relaxo- metry	T2*	V		T2* values were considerably higher in neonates with NE
Natu (20	u et al. 121) ²⁶⁾	California, United States	Not stated	Prospective longitudinal study	16	9:7 (term neonates)	Not mentioned	>37	Relaxo- metry	T1		V	R1 (and thus myelin) develop- ment is increasingly faster in later visual areas than in early visual areas
Wan (20	ıg et al. 119) ⁸⁾	London, United Kingdom	Not stated	Retrospective study	114	26:32 (preterm neo nates) 30:26 (term neo- nates)	29.4 (preterm neo- nates) 29.7 (term neo- nates)	27–37 (pre- term neo- nates) 37–44 (term neonates)	-	-	Not m	nen- d	The volume of the MLS in the deep brain region seemed to increase in a way that was proportional to the gestational age
Schr et a	nidbauer al. (2019) ²⁷⁾	Vienna, Austria	2017- 2018	Prospective cross-sectio- nal study	25	1:6 (term neonates) 9:9 (preterm neo- nates)	39.0 (term neo- nates)25.0 (preterm neo- nates)	42 (term neo- nates) 38 (preterm neonates)	Relaxo- metry	T1, T2	V		Preterm newborns have con- siderably lower myelination values based on SyMRI than term-born neonates
Knig (20	ht et al. 18) ²⁸⁾	Bristol, England	Not stated	Retrospective study	31	14:6 (late preterm) 6:5 (very preterm)	35.0 (late preterm) 27.0 (very preterm)	38 (late preterm)37 (very preterm)	Relaxo- metry	T2	Not m tioned	nen- d	T2 is longer in extensive regions of WM for extremely preterm neonates than for late pre- term group
Lee (20	et al. 118) ²⁴⁾	South Korea	2015– 2016	Prospective cross-sectio- nalstudy	23	Not specified	Not mentioned	34-43	Relaxo- metry	T1, T2	V		T1 and T2 of the WM areas were both quite high at birth, with a maximum value of roughly 2600 milliseconds for T1 and 280 milliseconds for T2, and declined by more than half in the first year of life
Sour (20	n et al. 117) ¹⁴⁾	New York, United States	Not stated	Retrospective study	10	8:2 (term neonates)	38.7	38-40	Relaxo- metry	T1, T2	Not m	nen- d	The T1/T2 ratio exhibited high intensity values in the PLIC and low intensity values in the optic radiations, which is associated with the progressive brain maturation during the first few months of birth, which re- sults in considerable spatial differences in myelin density
Schr al.	neider et (2016) ²⁹⁾	Lausanne, Switzer- land	2011- 2013	Prospective longitudinal study	39	20:19 (preterm neo- nates)	27.0	1st part: 28- 29 2nd part: 34- 35 3rd part: 37- 40	Relaxo- metry	T1	V		Beginning around 26 weeks, the T1 values in the basal ganglia and thalamus decreased gra- dually due to rapid neuronal densification and continuous myelination
Mell al.	oourne et (2016) ³⁰⁾	London, United Kingdom	Not stated	Prospective longitudinal study	37	10:27 (preterm neo- nates)	26.27	27–58	Relaxo- metry	MWF	Not m	nen- d	In the WM PLIC and ALIC, axonal and myelin density varies, with large myelin volume in the posterior limb and substan- tially lower values in the anterior limb, despite relatively similar intra-axonal volume values
Ning (20	g et al. 014) ²²⁾	Xi'an, China	Not stated	Prospective cross-sectio- nal study	56	36:20 (term neo- nates)	54.0	Not mention- ed	Relaxo- metry	R2*	V		The R2* value of the internal capsule increased with post- menstrual age, indicating rapid maturation in the first year of life
													(Contiuned)

Table 2. Data extracted from the literature regarding the main variables of interest (Contined)

Study	Place/ country	Study period	Type of study design	Sam- ple size	Gender (male: female)	Average gestational age (wk)	Age <i>a</i> t MRI (wk)	MRI modality	MRI mea- sures (s)	Scar time (c tative sequ <5 min	ning quanti- e MRI ence) ≥5 min	Findings
Nossin- Manor et al. (2013) ²⁵⁾	Canada, North America	2008- 2010	Prospective cross-sectio- nal study	54	25:29 (preterm neo- nates)	29.0	26-34	MTI, Relaxo- metry	MTR, T1		V	At approximately 36 weeks gestational age, myelin begins to coat the axons in the PLIC, results in a surge in markers related to macromolecules related with myelination (highest MTR values) as well as signals of significant tissue limitation
Deoni et al. (2012) ²³⁾	Rhode Island, United States	Not stated	Prospective cross-sectio- nal study	13	6:7 (term neonates)	37.0	Not men- tioned	Relaxo- metry	T1, T2, MWF		V	When T1, T2, and MWF measures are com- pared, they show different sensitivity to tissue changes linked with neurode- velopment, with each delivering distinct but complementing insights

MRI, magnetic resonance imaging; NE, neonatal encephalopathy; MLS, myelin-like signal; SyMRI, synthetic magnetic resonance imaging; WM, white matter; PLIC, posterior limb of the internal capsule; MWF, Myelin-water fraction; ALIC, anterior limb of the internal capsule; MTI, magnetization transfer imaging; MTR, magnetization transfer ratio.

Study	Quantitative MRI technique/sequence	Software/ postprocessing	Advantage	Limitation
Olivieri et al. (2021) ⁵⁾	3D multiecho gradient- echo sequence	MRI console appli- cation software, Philips DICOM Viewer software version R3.0	Good signal-to-noise ratio Short scanning time <4 min	Small sample size: unable to do a subanalysis to rule out a potential difference in the myelination of the neonates with NE without injury, compared to the healthy neonates
Natu et al. (2021) ²⁶⁾	Spoiled-gradient echo (SPGR) and inversion recovery-echo planar imaging (IR-EPI)	mrQ software	Quantitative measurements of proton relaxation time (T1, which depends on the physiochemical environment of the tissue) from qMRI allow the amount of brain tissue in a voxel (3D pixel in an MRI image that is 1–2 mm on a side) related to the neuropil and myelin to be measured quantitatively and longitudinally, so, these quantitative metrics are a noninvasive way to find out about changes in the microstructure and separate different theories about how development works, since T1 is lower in tissues with a denser microstructure	Unmyelinated pruned neurites may obscure pruning effects on qMRI Changes in iron due to pruning-associated phagocytosis may
Wang et al. (2019) ⁸⁾	T2	Novel automatic segmentation technique of MLS	The spatiotemporal model shows a steady increase in MLS lends to the theory that they are, in fact, gathering on the formation of myelin T2 remains the most routinely performed MRI image for the neonatal brain	Although the segmentation method given provides a quantitative assessment of the myelinated tissue, it is unable to quantify the fraction of myelin within each voxel
Schmidbauer et al. (2019) ²⁷⁾	Multidynamic multi- echo (MDME)	SyMRI	SyMRI image data are equivalent to T1 and T2 images SyMRI generates quantitative MR maps in seconds SyMRI maps were apparent while measuring myelination of brain stem structures, such as the superior and inferior cerebellar peduncles and the medial lemniscus, which are already myelinated at the usual expected due date	Resolution and slice thickness varies between SyMRI maps and conventional MRI, limiting direct comparison Small sample size
Knight et al. (2018) ²⁸⁾	T2	Not mentioned	T2 mapping is one of the most potent techniques for studying the developing brain, including myelogenesis	T2 measurements are based on a basic mono-expo- nential assumption to a 3-echo turbo-spin echo: lengthy T2 mapping sequence may be required to sample early and late decoherence adequately for high-quality non- exponential or multi-exponential T2 maps
Lee et al. (2018) ²⁴⁾	Multidynamic multi- echo (MDME)	SyMRI	Allows simultaneous quantification of relaxation times, relaxation rate, and PD in 5 to 6 minutes for full head coverage at high resolution	Small sample size: the clinical utility of synthetic sequence quantitative imaging needs a larger research Synthetic sequence correctness and reproducibility are proven at 1.5T but not yet at 3T Only 3T age-related tissue alterations were investigated
Soun et al. (2017) ¹⁴⁾	T1,T2	FSL	 T1/T2 ratio increases contrast-to-noise without increasing scan duration T1/T2 ratio can distinguish between highly and lightly myelinated cortical areas, suggesting it could be used to study myelin development in the neonatal brain Can be used in neonates to identify HII and myelination by comparing T1 signal intensities of distinct brain areas Employs commonly acquired clinical sequences, so it doesn't add scan time 	Possible HII in some subject: studies suggest that severe HII can cause hyperintense T1 signal in the PLIC, while no patient in the cohort had HII, undiscovered subthreshold hypoxia could raise T1 signal intensity in the PLIC and be mistaken as myelin Comparing sequences needed coregistration: T1 and T2 images had modest misregistration abnormalities at cerebrospinal-gray matter interfaces, incorrect contrast increases could be perceived as myelin disease (Contined)

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Study	Quantitative MRI technique/sequence	Software/ postprocessing	Advantage	Limitation
Schneider et al. (2016) ²⁹⁾	3D magnetization prepared dual rapid acquisition of gradient echo (MP2RAGE)	Not mentioned	By creating a purely T1-weighted picture, MP2RAGE may generate whole-brain T1 tissue relaxation time maps for quantitative tissue characterization T1 relaxometry's descriptive features are of special relevance in the preterm population because they convey structural information about tissue, such as water content and lipid and macromolecule compo- sitions, and depict myelin's chronologic maturation T1 relaxation time gives information regarding myelin synthesis, cholesterol, and macromolecules (galac tocerebrosides), making it an ideal marker of brain maturity	Values from moderate or severe brain lesions cannot be compared to those from low-risk patients since they were very few No healthy control fetuses or term neonates were available for comparison
Melbourne et al. (2016) ³⁰⁾	2D Gradient and Spin Echo (GraSE)	Not mentioned	Single component relaxometry is a non-specific predictor of myelin and myelination but a MWF derived from multicomponent T2 relaxometry correlates with histological staining	Small sample size for longitudinal data With 2 longitudinal imaging time points, it's difficult to support an alternative model, and it's unclear how cross-sectional models should be used longitudinally
Soun et al. (2017) ¹⁴⁾	T1, T2	FSL	 T1/T2 ratio increases contrast-to-noise without increasing scan duration T1/T2 ratio can distinguish between highly and lightly myelinated cortical areas, suggesting it could be used to study myelin development in the neonatal brain Can be used in neonates to identify HII and myelination by comparing T1 signal intensities of distinct brain areas Employs commonly acquired clinical sequences, so it doesn't add scan time 	Possible HII in some subject: studies suggest that severe HII can cause hyperintense T1 signal in the PLIC, while no patient in the cohort had HII, undiscovered subthreshold hypoxia could raise T1 signal intensity in the PLIC and be mistaken as myelin Comparing sequences needed coregistration: T1 and T2 images had modest misregistration abnormalities at cerebrospinal-gray matter interfaces, incorrect contrast increases could be perceived as myelin disease
Schneider et al. (2016) ²⁹⁾	3D magnetization prepared dual rapid acquisition of gradient echo (MP2RAGE)	Not mentioned	By creating a purely T1-weighted picture, MP2RAGE may generate whole-brain T1 tissue relaxation time maps for quantitative tissue characterization T1 relaxometry's descriptive features are of special relevance in the preterm population because they convey structural information about tissue, such as water content and lipid and macromolecule compo- sitions, and depict myelin's chronologic maturation T1 relaxation time gives information regarding myelin synthesis, cholesterol, and macromolecules (galac tocerebrosides), making it an ideal marker of brain maturity	Values from moderate or severe brain lesions cannot be compared to those from low-risk patients since they were very few No healthy control fetuses or term neonates were available for comparison
Melbourne et al. (2016) ³⁰⁾	2D Gradient and Spin Echo (GraSE)	Not mentioned	Single component relaxometry is a non-specific predictor of myelin and myelination but a MWF derived from multicomponent T2 relaxometry correlates with histological staining	Small sample size for longitudinal data With 2 longitudinal imaging time points, it's difficult to support an alternative model, and it's unclear how cross-sectional models should be used longitudinally
Ning et al. (2014) ²²⁾	3D enhanced T2 star weighted angiography (ESWAN)	ADW4.3 workstation	R2* is more sensitive than R2 for iron deposition and white matter maturation in the brain	No phase and R2* templates: neonates' increased brain water content and lower iron concentration make it difficult to automatically separate and register many brain areas, they have more brain water than adults and it decreases with age Reduced water fractions may have affected the R2* value by age Considering the potential influence of sedation on R2* and phase values, some correction should be made using systemic oxygenation parameters such as oxygen or carbon dioxide pressure and oxygen saturation
Nossin-Manor et al. (2013) ²⁵⁾	PDW 3D spoiled gradient recalled (SPGR) and T1 3D spoiled gradient recalled (SPGR)	Not mentioned	MTR and T1 values are sensitive (but not specific) to water content in tissue, the development of tissue organization, and early myelination events in the developing brain	MTR could be more sensitive to inflammatory diseases than myelination and demyelination
Deoni et al. (2012) ²³⁾	Spoiled gradient echo (SPGR, spoiled FLASH), fully- balanced steady-state free precession (bSSFP) and inversion-prepared (IR)- SPGR	Not mentioned	Excellent resolution compared to other quantitative approaches	Long scanning time: difficult to add to the baseline MRI protocol in a population of critically ill neonates Not widely used to study neonatal brains under 3 months of age or the brains of critically ill neonates Concerns regarding the accuracy of the technique

MRI, magnetic resonance imaging; 3D, 3-dimensional; 2D, 2-dimensional; MLS, myelin-like signal; SyMRI, synthetic magnetic resonance imaging; qMRI, quantitative MRI; PD, Proton density; HII, hypoxic-ischemic injury; MWF, Myelin-water fraction; MTR, magnetization transfer ratio; PLIC, posterior limb of the internal capsule.

Table 4. List of regions of interests (ROIs) from the literature and how often they were mentioned

Posterior limb of the internal capsule (PLC) Obtion et al. (2017) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Nossin Manor et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Desoni et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ming et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ning et al. (2019) ⁴⁵ , Ning et et al. (2019) ⁴⁵ , Schmeidzner et al. (2019) ⁴⁵ , Ning et al. (2019) ⁴⁵ , Ning et al. (2019) ⁴⁵ , Schmeidzner et a	ROI(s)	Mentioned in study	No. of studies
Corpus callosum (genu) Obiesn' et al. (2021) ⁹⁷ , Kright et al. (2019) ⁹⁸ , Issain Manor et al. (2019) ⁹⁷ , Issain Man	Posterior limb of the internal capsule (PLIC)	Olivieri et al. (2021) ⁵), Wang et al. (2019) ⁸), Schmidbauer et al. (2019) ²⁷), Knight et al. (2018) ²⁸), Soun et al. (2017) ¹⁴), Schneider et al. (2016) ²⁹), Melbourne et al. (2016) ³⁰), Ning et al. (2014) ²²), Nossin-Manor et al. (2013) ²⁵). Deoni et al. (2012) ²³	10
Corpus callosum (splenium)Olivien et al. (2019) ¹⁶ , Ning et al. (2019) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2019) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2017) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2017) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2017) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2016) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2016) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2016) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2016) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2016) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Schneider et al. (2016) ²⁷ , Nossin-Manor et al. (2013) ²⁷ , Doon et al. (2	Corpus callosum (genu)	Olivieri et al. (2021) ⁵⁾ , Knight et al. (2018) ²⁸⁾ , Lee et al. (2018) ²⁴⁾ , Schneider et al. (2016) ²⁹⁾ , Melbourne et al. (2016) ³⁰⁾ , Ning et al. (2014) ²²⁾ , Nossin-Manor et al. (2013) ²⁵⁾ , Deoni et al. (2012) ²³⁾	8
Optic radiationsObject et al. (2019) ¹⁷ , Schmidbauer et al. (2019) ²⁷ , Icen et al. (2018) ¹⁷ , Icen	Corpus callosum (splenium)	Olivieri et al. (2021) ⁵⁾ , Knight et al. (2018) ²⁸⁾ , Lee et al. (2018) ²⁴⁾ , Schneider et al. (2016) ²⁹⁾ , Melbourne et al. (2016) ³⁰⁾ , Ning et al. (2014) ²²⁾ , Nossin-Manor et al. (2013) ²⁵⁾ , Deoni et al. (2012) ²³⁾	8
The lamosObjerent ed. 12021% Schmidbauer et al. (2016)% et al. (2013% et al. (2013%)% schmidbauer et al. (2016)% schmidbauer et al. (2016)% schmidbauer et al. (2016)% 	Optic radiations	Olivieri et al. (2021) ⁵⁾ , Schmidbauer et al. (2019) ²⁷⁾ , Knight et al. (2018) ²⁸⁾ , Lee et al. (2018) ²⁴⁾ , Soun et al. (2017) ¹⁴⁾ , Schneider et al. (2016) ²⁹⁾ , Deoni et al. (2012) ²³⁾	7
Frontal region (white matter)Olivair et al. (2021) ⁹ , Schmidbauer et al. (2016) ²⁹ , Icae et al. (2016) ²⁹ , Isosin-Manor et al. (2012) ²¹ 6Parietal region (white matter)Lee et al. (2016) ²⁹ , Schmidber et al. (2016) ²⁹ , Nossin-Manor et al. (2012) ²¹ 3Anterior imb of the internal capsule (ALLC)Melbourne et al. (2016) ²⁹ , Nossin-Manor et al. (2016) ²¹ , Isosin-Manor et al. (2012) ²¹ 3Corpus callosum (body)Knighte et al. (2018) ⁴⁹ , Schneider et al. (2016) ²⁷ 2Corpus callosum (body)Knighte et al. (2018) ⁴⁹ , Schneider et al. (2016) ²⁷ 2Central region (white matter)Schmidbauer et al. (2016) ²⁷ , Schneider et al. (2016) ²⁷ 2Central region (gray matter)Lee et al. (2018) ⁴⁷ , Schneider et al. (2016) ²⁷ 2Parieal region (gray matter)Lee et al. (2018) ⁴⁷ , Schneider et al. (2016) ²⁷ 2Caudate nucleusLee et al. (2018) ⁴⁷ , Nossin-Manor et al. (2016) ²⁷⁰ 2PutamenNing et al. (2014) ²⁷¹ , Nossin-Manor et al. (2016) ²⁷⁰ 2Caudate nucleusLee et al. (2018) ⁴⁷¹ , Nossin-Manor et al. (2013) ²⁷⁰ 2Somatosensory (S1)Natu et al. (221) ⁴⁷¹ , Nossin-Manor et al. (2013) ²⁷⁰ 2Somatosensory (S1)Natu et al. (221) ⁴⁷¹ , Nossin-Manor et al. (2013) ²⁷⁰ 2Somatosensory (S1)Natu et al. (221) ⁴⁷¹ , Nossin-Manor et al. (2013) ²⁷⁰ 2Somatosensory (S1)Natu et al. (221) ⁴⁷¹ , Nossin-Manor et al. (2013) ²⁷⁰ 1Subatalinic nucleusWang et al. (2019) ⁴⁷⁰ 1Subatalinic nucleusWang et al. (2019) ⁴⁷⁰ 1Subatalinic nucleusWang et al. (2	Thalamus	Olivieri et al. (2021) ⁵⁾ , Schmidbauer et al. (2019) ²⁷⁾ , Lee et al. (2018) ²⁴⁾ , Schneider et al. (2016) ²⁹⁾ , Ning et al. (2014) ²²⁾ , Nossin-Manor et al. (2013) ²⁵⁾	6
Parietal region (white mater)Lee et al. (2016) ²⁶ , Schneider et al. (2014) ²⁷). Deoni et al. (2012) ²³ Schneider et al. (2016) ²⁷ , Deoni et al. (2012) ²³ Corpus callosum (body)Kinght et al. (2018) ²⁶ , Deoni et al. (2012) ²⁷³ 2Corpus callosum (body)Kinght et al. (2018) ²⁷⁰ , Deoni et al. (2012) ²⁷³ 2Corpus callosum (body)Kinght et al. (2018) ²⁷⁰ , Deoni et al. (2015) ²⁷⁶ 2CingulumKinght et al. (2018) ²⁷⁶ , Schneider et al. (2016) ²⁷⁶ 2Central region (white mater)Schneidbare et al. (2016) ²⁷⁶ , Schneider et al. (2016) ²⁷⁶ 2Perirolandic region (gray mater)Lee et al. (2018) ²⁷⁶ , Schneider et al. (2016) ²⁷⁶ 2Caddate nucleusLee et al. (2018) ²⁷⁶ , Schneider et al. (2016) ²⁷⁶ 2PutamenNing et al. (2014) ²⁷¹ , Noscin-Manor et al. (2013) ²⁷⁶ 2Caddate nucleusLee et al. (2014) ²⁷¹ , Noscin-Manor et al. (2013) ²⁷⁶ 2PutamenNing et al. (2014) ²⁷¹ , Noscin-Manor et al. (2013) ²⁷⁶ 2PutamenNau et al. (2014) ²⁷¹ , Noscin-Manor et al. (2013) ²⁷⁶ 2Motor (M1)Nau et al. (2014) ²⁷¹ , Noscin-Manor et al. (2013) ²⁷⁶ 2Motor (M1)Nau et al. (2014) ²⁷¹ , Noscin-Manor et al. (2013) ²⁷⁶ 2Motor (M1)Nau et al. (2014) ²⁷¹ , Noscin-Manor et al. (2013) ²⁷⁶ 2Motor (M1)Nau et al. (2014) ²⁷¹ , Noscin-Manor et al. (2013) ²⁷⁶ 2Motor (M1)Nau et al. (2014) ²⁷¹ , Ninght et al. (2016) ²⁷⁶ 2Motor (M1)Nau et al. (2014) ²⁷¹ , Ninght et al. (2016) ²⁷⁶ 2Subtalanic nucleusWang et al. (2018) ²⁷⁶ </td <td>Frontal region (white matter)</td> <td>Olivieri et al. (2021)⁵⁾, Schmidbauer et al. (2019)²⁷⁾, Lee et al. (2018)²⁴⁾, Schneider et al. (2016)²⁹⁾, Nossin-Manor et al. (2013)²⁵⁾, Deoni et al. (2012)²³⁾</td> <td>6</td>	Frontal region (white matter)	Olivieri et al. (2021) ⁵⁾ , Schmidbauer et al. (2019) ²⁷⁾ , Lee et al. (2018) ²⁴⁾ , Schneider et al. (2016) ²⁹⁾ , Nossin-Manor et al. (2013) ²⁵⁾ , Deoni et al. (2012) ²³⁾	6
Anterior limb of the internal capsule (ALIC)Melbourne et al. (2016) ¹⁶ , Ning et al. (2017) ¹⁶ , Noneid er et al. (2016) ¹⁷⁰ 2Lentform nucleusOlivier et al. (2021) ¹⁶ , Noneid er et al. (2013) ³⁷³ 2Corpus callossing (body)Knight et al. (2019) ¹⁶ , Dooni et al. (2013) ³⁷³ 2CingulumKnight et al. (2018) ⁴⁶ , Dooni et al. (2016) ⁵⁷⁰ 2Corpus callossing (hoth)Schnidbauer et al. (2016) ⁵⁷⁰ 2Fontal region (gray matter)Lee et al. (2018) ⁴⁶ , Schneidber et al. (2016) ⁵⁷⁰ 2Perirolandic region (gray matter)Lee et al. (2018) ⁴⁷ , Schneidber et al. (2016) ⁵⁷⁰ 2Caduate nucleusLee et al. (2018) ⁴⁷ , Schneidber et al. (2016) ⁵⁷⁰ 2Caduate nucleusLee et al. (2018) ⁴⁷ , Schneidber et al. (2016) ⁵⁷⁰ 2Caduate nucleusLee et al. (2018) ⁴⁷ , Nossin-Manor et al. (2013) ⁵⁷⁰ 2Ciobus pallidusNing et al. (2014) ²⁷⁰ , Nossin-Manor et al. (2013) ⁵⁷⁰ 2Visual (V1)Natu et al. (2021) ⁵⁷⁰ , Knight et al. (2018) ⁵⁸¹ 2Somatosensory (S1)Natu et al. (2021) ⁵⁷⁰ , Knight et al. (2018) ⁵⁸¹ 2Auditory (A1)Natu et al. (2021) ⁵⁷⁰ , Knight et al. (2018) ⁵⁸¹ 1Motor (M1)Natu et al. (2021) ⁵⁷⁰ , Knight et al. (2018) ⁵⁸¹ 1Subchainei nucleusWang et al. (2019) ⁵⁷⁰ 1Motor (M1)Natu et al. (2018) ⁵⁷⁰ 1Inferior oriclusWang et al. (2019) ⁵⁷⁰ 1Motor (M1)Natu et al. (2018) ⁵⁷⁰ 1Inferior oriclusWang et al. (2019) ⁵⁷⁰ 1Medial longitudinal fasciculus <td>Parietal region (white matter)</td> <td>Lee et al. (2018)²⁴⁾, Schneider et al. (2016)²⁹⁾, Nossin-Manor et al. (2013)²⁵⁾, Deoni et al. (2012)²³⁾</td> <td>4</td>	Parietal region (white matter)	Lee et al. (2018) ²⁴⁾ , Schneider et al. (2016) ²⁹⁾ , Nossin-Manor et al. (2013) ²⁵⁾ , Deoni et al. (2012) ²³⁾	4
Lentiform nucleusOlivieri et al. (2019) ¹⁹ , Schneider et al. (2016) ¹⁹ 2Corpus callosum (body)Knight et al. (2018) ¹⁹ , Deoni et al. (2012) ¹³ 2CingulumKnight et al. (2018) ¹⁹ , Deoni et al. (2012) ¹³ 2Cingulur (ava matter)Schnidbauer et al. (2016) ¹⁹ , Schneider et al. (2016) ¹⁹ 2Perirolandic region (gray matter)Lee et al. (2018) ¹⁴ , Schneider et al. (2016) ¹⁹ 2Parietal region (gray matter)Lee et al. (2018) ¹⁴ , Schneider et al. (2016) ¹⁹ 2Parietal region (gray matter)Lee et al. (2018) ¹⁴ , Schneider et al. (2016) ¹⁹ 2PutamenNing et al. (2014) ¹⁷ , Nossin-Manor et al. (2013) ¹⁵ 2PutamenNing et al. (2014) ¹⁷ , Nossin-Manor et al. (2013) ¹⁵ 2Primary sensory-motro cortices:7Visual (V1)Natu et al. (2021) ¹⁹ , Knight et al. (2018) ¹⁹ 2Auditory (A1)Natu et al. (2021) ¹⁰ , Knight et al. (2018) ¹⁹ 2Auditory (A1)Natu et al. (2021) ¹⁰ , Knight et al. (2018) ¹⁹ 2Auditory (A1)Natu et al. (2021) ¹⁰ , Knight et al. (2018) ¹⁰ 1Superior cerebellar peduncleWang et al. (2018) ¹⁰ 1Superior cerebellar peduncleWang et al. (2018) ¹⁰ 1Decussation of the superior cerebellar peduncleKnight et al. (2018) ¹⁰ 1Inferior colliculusWang et al. (2019) ¹⁰ 1Inferior colliculusWang et al. (2019) ¹⁰ 1Medial longitudinal fasciculusKnight et al. (2018) ¹⁰ 1Medial longitudinal fasciculusKnight et al. (2018) ¹⁰ 1 <td>Anterior limb of the internal capsule (ALIC)</td> <td>Melbourne et al. (2016)³⁰⁾, Ning et al. (2014)²²⁾, Deoni et al. (2012)²³⁾</td> <td>3</td>	Anterior limb of the internal capsule (ALIC)	Melbourne et al. (2016) ³⁰⁾ , Ning et al. (2014) ²²⁾ , Deoni et al. (2012) ²³⁾	3
Corpus callosum (body)Knight et al. (2019) ⁷⁹ , Deoni et al. (2012) ²⁷¹ 2Ventrolateral nucleusWang et al. (2019) ⁷⁹ , Nosin-Manor et al. (2012) ²⁷¹ 2Central region (knite matter)Schmidbauer et al. (2019) ⁷⁷ , Schneider et al. (2016) ⁷⁹¹ 2Penricoland: cengoin (gray matter)Lee et al. (2018) ⁴⁷ , Schneider et al. (2016) ⁷⁹¹ 2Parietal region (gray matter)Lee et al. (2018) ⁴⁷ , Schneider et al. (2016) ⁷⁹¹ 2Parietal region (gray matter)Lee et al. (2018) ⁴⁷ , Schneider et al. (2016) ⁷⁹¹ 2Caudate nucleusLee et al. (2018) ⁴⁷ , Schneider et al. (2016) ⁷⁹¹ 2Parietal region (gray matter)Lee et al. (2018) ⁴⁷ , Schneider et al. (2016) ⁷⁹¹ 2Caudate nucleusLee et al. (2014) ⁴⁷² , Nossin-Manor et al. (2013) ⁵⁷³ 2Pitmary sensory-motor cortics:77Visual (V1)Natu et al. (2021) ⁵⁷⁰ , Knight et al. (2018) ⁵⁷⁰ 2Motor (M1)Natu et al. (2021) ⁵⁷⁰ , Knight et al. (2018) ⁷⁸⁰ 2Auditory (A1)Natu et al. (2021) ⁵⁷⁰ , Knight et al. (2018) ⁷⁸⁰ 1Subhalamic nucleusWang et al. (2018) ⁷⁸⁰ 1Subhalamic nucleusWang et al. (2018) ⁷⁸⁰ 1Decussation of the superior cerebellar peduncleKnight et al. (2018) ⁷⁸⁰ 1Inferior officulusWang et al. (2018) ⁷⁸⁰ 1Inferior officulusWang et al. (2018) ⁷⁸⁰ 1Midde et al. Mag et al. (2018) ⁷⁸⁰ 11Midde et al. Mag et al. (2018) ⁷⁸⁰ 11Medial longitudinal fasciculusKnight et al. (2018) ⁷⁸⁰ <t< td=""><td>Lentiform nucleus</td><td>Olivieri et al. (2021)⁵⁾, Schneider et al. (2016)²⁹⁾</td><td>2</td></t<>	Lentiform nucleus	Olivieri et al. (2021) ⁵⁾ , Schneider et al. (2016) ²⁹⁾	2
Ventrolateral nucleusWang et al. (2019) ⁸¹ , Nossin-Manor et al. (2013) ⁷³⁰ 2CingulumKnight et al. (2018) ⁸¹ , Schneider et al. (2016) ⁷³⁰ 2Fontal region (yray matter)Lee et al. (2018) ⁸¹ , Schneider et al. (2016) ⁷³⁰ 2Perirolandic region (gray matter)Lee et al. (2018) ⁸¹ , Schneider et al. (2016) ⁷³⁰ 2Parietal region (gray matter)Lee et al. (2018) ⁸¹ , Schneider et al. (2016) ⁷³⁰ 2Caudate nucleusLee et al. (2018) ⁸¹ , Schneider et al. (2016) ⁷³⁰ 2PutamenNing et al. (2014) ⁷²⁰ , Nossin-Manor et al. (2013) ⁷³⁰ 2Pitmary sensory-motor cortices:*********************************	Corpus callosum (body)	Knight et al. (2018) ²⁸⁾ , Deoni et al. (2012) ²³⁾	2
CingulumKnight et al. (2019) ²⁰ , Schneider et al. (2016) ²⁰ 2Central region (white matter)Schmidbauer et al. (2016) ²⁰ , Schneider et al. (2016) ²⁰ 2Princland region (gray matter)Lee et al. (2018) ²⁰ , Schneider et al. (2016) ²⁰ 2Parietal region (gray matter)Lee et al. (2018) ²⁰ , Schneider et al. (2016) ²⁰ 2Caudate nucleusLee et al. (2018) ²⁰ , Schneider et al. (2016) ²⁰ 2Caudate nucleusLee et al. (2018) ²⁰ , Schneider et al. (2016) ²⁰ 2Gibbus pallidusNing et al. (2014) ²² , Nossin-Manor et al. (2013) ²⁰¹ 2Visual (V1)Ning et al. (2014) ²² , Nossin-Manor et al. (2013) ²⁰¹ 2Somatosensory (S1)Natu et al. (2021) ²⁰ , Knight et al. (2018) ²⁰¹ 2Motor (M1)Natu et al. (2021) ²⁰ , Knight et al. (2018) ²⁰¹ 2Auditory (A1)Natu et al. (2021) ²⁰¹ , Knight et al. (2018) ²⁰¹ 1Subthalmic nucleusWang et al. (2019) ²⁰ 1Superior cerebellar peduncleKnight et al. (2018) ²⁰¹ 1Decusation of the superior cerebellar peduncleKnight et al. (2018) ²⁰¹ 1Medial longitudinal fasciculusWang et al. (2019) ⁸⁰ 1Medial longitudinal fasciculusKnight et al. (2018) ²⁰¹ 1Medial longitudinal fasciculus <t< td=""><td>Ventrolateral nucleus</td><td>Wang et al. (2019)⁸⁾, Nossin-Manor et al. (2013)²⁵⁾</td><td>2</td></t<>	Ventrolateral nucleus	Wang et al. (2019) ⁸⁾ , Nossin-Manor et al. (2013) ²⁵⁾	2
Central region (white matter) Schnidbauer et al. (2019) ³⁷⁾ , Schneider et al. (2016) ³⁷⁰ 2 Frontal region (gray matter) Lee et al. (2018) ³⁴⁾ , Schneider et al. (2016) ³⁷⁰ 2 Perirolandic region (gray matter) Lee et al. (2018) ³⁴⁾ , Schneider et al. (2016) ³⁷⁰ 2 Parietal region (gray matter) Lee et al. (2018) ³⁴ , Schneider et al. (2016) ³⁷⁰ 2 Caudate nucleus Lee et al. (2018) ³⁴ , Schneider et al. (2013) ³⁷⁰ 2 Odobus palidus Ning et al. (2014) ³² , Nossin-Manor et al. (2013) ³⁷⁰ 2 Odobus palidus Ning et al. (2014) ³² , Nossin-Manor et al. (2013) ³⁷⁰ 2 Somatosensory (S1) Natu et al. (2021) ³⁶ , Knight et al. (2018) ³⁸¹ 2 Motor (M1) Natu et al. (2021) ³⁶¹ , Knight et al. (2018) ³⁸¹ 2 Auditory (A1) Natu et al. (2021) ³⁶¹ , Knight et al. (2018) ³⁸¹ 2 Subthalamic nucleus Wang et al. (2019) ³⁶¹ 1 Subthalamic nucleus Wang et al. (2018) ³⁸¹ 1 Subthalamic nucleus Wang et al. (2018) ³⁶¹ 1 Subthalamic nucleus Wang et al. (2018) ³⁶¹ 1 Subthalamic nucleus Wang et al. (2018) ³⁶¹ 1 </td <td>Cinqulum</td> <td>Knight et al. (2018)²⁸⁾. Deoni et al. (2012)²³⁾</td> <td>2</td>	Cinqulum	Knight et al. (2018) ²⁸⁾ . Deoni et al. (2012) ²³⁾	2
constances constan	Central region (white matter)	Schmidhauer et al. $(2019)^{27}$ Schneider et al. $(2016)^{29}$	2
Numery Springery (network) Construction Construction <th< td=""><td>Frontal region (grav matter)</td><td>Lee et al. $(2018)^{24}$ Schneider et al. $(2016)^{29}$</td><td>2</td></th<>	Frontal region (grav matter)	Lee et al. $(2018)^{24}$ Schneider et al. $(2016)^{29}$	2
Canadate region (gap matter) Lee et al. (2018) ⁴⁶ , Ninej et al. (2016) ⁵⁶ , 2 Caudate nucleus Lee et al. (2018) ⁴⁶ , Ninej et al. (2014) ²² , 2 Putamen Ning et al. (2014) ²² , Nossin-Manor et al. (2013) ²⁵) 2 Primary sensory-motor cortices: * * Visual (V1) Natu et al. (2021) ⁵⁶ , Knight et al. (2018) ³⁶ , Sosin-Manor et al. (2018) ³⁷⁰ 2 Somatosensory (S1) Natu et al. (2021) ⁵⁶ , Knight et al. (2018) ³⁷⁰ 2 Auditory (A1) Natu et al. (2021) ⁵⁶ , Knight et al. (2018) ³⁷⁰ 2 Auditory (A1) Natu et al. (2021) ⁵⁶ , Knight et al. (2018) ³⁷⁰ 2 Superior cerebellar peduncle Wang et al. (2019) ⁵⁷⁰ , Knight et al. (2018) ³⁷⁰ 1 Superior cerebellar peduncle Wang et al. (2019) ⁵⁷⁰ , Knight et al. (2018) ³⁷⁰ 1 Superior cerebellar peduncle Wang et al. (2019) ⁵⁷⁰ 1 Inferior colliculus Wang et al. (2019) ⁵⁷⁰ 1 Leteral lemniscus Wang et al. (2019) ⁵⁷⁰ 1 Medial longituinal fasciculus Knight et al. (2018) ⁵⁷⁰ 1 Inferior fonto-occipital fasciculus Knight et al. (2019) ⁵⁷⁰ 1 <	Perirolandic region (gray matter)	Lee et al. (2018) ²⁴⁾ Schneider et al. (2016) ²⁹⁾	2
Intergenergies Exect al. (2019) ²⁸ , Ning et al. (2014) ²² , 2 Quaden nucleus Lee et al. (2014) ²² , Nossin-Manor et al. (2013) ²⁵) 2 Putamen Ning et al. (2014) ²² , Nossin-Manor et al. (2013) ²⁵) 2 Primary sensory-motor cortices: ************************************	Parietal region (gray matter)	Lee et al. (2018) ²⁴⁾ Schneider et al. (2016) ²⁹⁾	2
Catadac Indicas Execution (Conf) ^{27,1} Norsin-Manor et al. (2013) ^{25,1} 2 Globus pallidus Ning et al. (2014) ^{22,1} , Nossin-Manor et al. (2013) ^{25,1} 2 Primary sensory-motor cortices: ************************************	Caudate pucleus	Lee et al. $(2018)^{24}$ Ning et al. $(2014)^{22}$	2
Tutation Ning et al. (2014) ^{2,10} , Nossin-Manor et al. (2013) ²⁵¹ 2 Primary sensory-motor cortices: ************************************	Putamen	Ning et al. $(2014)^{22}$ Nossin Manor et al. $(2013)^{25}$	2
King et al. (2019) King et	Globus pallidus	Ning et al. $(2014)^{22}$ Nossin-Manor et al. $(2013)^{25}$	2
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Corticospinal tract Knight et al. (2018) ²⁸⁾ 1 Uncus Knight et al. (2018) ²⁸⁾ 1	Cerebellar region (white matter)	Deoni et al. (2012) ²³⁾	1
Uncus Knight et al. (2018) ²⁸⁾ 1	Corticospinal tract	Knight et al. (2018) ²⁸⁾	1
	Uncus	Knight et al. (2018) ²⁸⁾	1

in neonatal imaging because of the risk of heat and motion artifacts.³⁶⁾

A few selected studies excluded some of the data owing to low image quality due to motion artifacts and significantly devastated brain structures,²⁷⁾ and because of extensive lesions on brain MRI, death, withdrawal of consent, or abnormal neurologic examination findings at term-equivalent age.²⁹⁾ Some subjects failed to produce usable data²⁶⁾ because they were unable to remain asleep after the MRI sequences commenced. Meanwhile, one study excluded a few subjects because some of them had abnormal cerebral ultrasonography findings and patient movement rendered the diffusion or relaxometry acquisitions unsatisfactory.³⁰⁾ Another study eliminated some images with pronounced motion artifacts from the final analysis.²⁴⁾

Discussion

The findings show that relaxometry (83.33%) is by far the most frequently investigated approach in validation studies, followed by magnetization transfer imaging (MTI) (8.33%) and the novel automatic segmentation technique (8.33%). A brief description of the general concepts of relaxometry, MTI, and automated segmentation techniques is provided in Table 5.

1. Relaxometry

Standard MRI images are qualitative, and the signal intensity depends on the number of complementary contrast mechanisms altered by the MRI hardware and software. Without a quantitative metric for the absolute analysis of pixel signal intensities, independent of scanner hardware and sequences, comparing MRI images from various individuals or from different parts of the same individual's body is difficult. Quantitative relaxometry separates the effects of each MRI contrast mechanism (T1, T2, and T2*), generates maps that are unaffected by the MRI protocol, and has a physical meaning that is often expressed in absolute units. Quantitative relaxometry not only provides an unbiased way to compare MRI scans but also uses the relationship between MRI maps and physiology to offer a noninvasive alternative to histology and biopsy.

1) T1

T1, the longitudinal relaxation time, is a time constant that represents the time required for magnetization to return to its initial state after being shifted.³⁷⁾ This recovery is affected by the change in the magnetic fields of the atoms around it at the proton resonance frequency. Because of its proximity, the proton next to the water molecule is typically the main source of fluctuations. The random spinning of the water molecule, which is described by the rotational correlation time, changes the angle between the protons

Table 5. Overview of relaxometry, magnetization transfer imaging, and automated segmentation technique

MRI modality	Definition	How does it/they work in MRI to detect/quantify myelin?
Relaxometry	The term "relaxometry" is employed in MRI to denote the process of quantifying the decay of signals (known as relaxation) subsequent to the excitation of protons through RF pulses. This process involves extracting precise values for the relaxation times, which represent the inherent characteristics of the tissues being investigated, from the aforementioned measurements. The estimation of the metrics (e.g., T1, T2, and T2*) can be achieved through the utilization of the suitable pulse sequence and corresponding parameters.	Multicomponent relaxation-based methods represent tissue water compartments by analyzing relaxation times. Water trapped between myelin bilayers and water inside or outside axons have different T1 and T2 relaxation times, therefore, for example, the MWF can be determined by measuring their relative signal contributions. Derived technique/sequence: MWF (derived from T2) R2* (derived from T2*, reciprocal of T2*) T1/T2 ratio (derived from T1 and T2) SyMRI (derived from T1, T2, and PD)
Magnetization transfer imaging	MRI technique that applies the contrast between tissues contain- ing hydrogen protons in 3 distinct states: those bound to macromolecules, those in free water, and those in the hydration layer between macromolecules and free water.	MT is an observable occurrence in which the spins of protons, when stimulated by an RF pulse; transmit a portion of their energy to adjacent mobile proton spins that are bound to macromolecules. MTI is a technique used to estimate the size of the macromolecular proton pool. This estimation is achieved by utilizing the ultra-short T2 relaxation and transferring the magnetization to the observable mobile water pool. The MTR has been widely employed in accordance with this theoretical framework and has demonstrated a strong association with histological myelin content.
Automatic segmentation	Segmentation refers to the computational process of separating an image into distinct regions that exhibit comparable attributes, such as gray level, color, texture, brightness, and contrast. Automated segmentation in the radiology field typically relies on fundamental image processing algorithms that analyze pixel intensities and/or textural characteristics, such as inter-pixel relationships. The concept is comparable to that of natural image segmentation, wherein the objective is to extract the target organ or region of interest from a medical image (2D or 3D).	The strategy for segmenting myelin has been built upon a range of computational methodologies and algorithms, necessitating the application of engineering principles in the domains of deep learning and artificial intelligence. Numerous ongoing studies primarily focus on achieving instant, rapid, sensitive, and reliable outcomes. Ongoing research is being carried out on this subject matter.

MRI, magnetic resonance imaging; RF, radiofrequency; MWF, Myelin-water fraction; SyMRI, synthetic magnetic resonance imaging; MT, magnetization transfer; MTI, magnetization transfer ratio; Magnetization transfer ratio; 3D, 3-dimensional; 2D, 2-dimensional.

over time, which causes magnetic fluctuations. The T1 relaxation due to this interaction occurs most quickly when the rotational correlation time is equal to the inverse of the proton resonance frequency. Similarly, protons in lipids, especially triglycerides, are affected by protons close to them. Because the protons in lipids do not move as much, there is usually more energy at the proton resonance frequency. This shortens the T1 of the lipids. Therefore, T1 shows how mobile the molecules are (mostly water protons) and how tightly they are bound to macromolecules. T1 is mostly used to measure the macromolecular content, water binding, and water content in a number of diseases, such as inflammation and changes in the amount of myelin in the brain.

Schneider et al.²⁹⁾ conducted a serial imaging investigation of preterm infants with low cerebral risk, aimed to establish reference values for T1 relaxation time, and evaluated the hypothesis that its evolution is equivalent to that of the apparent diffusion coefficient (ADC) and fractional anisotropy (FA) values, thus providing more accurate information about tissue structure. Magnetization prepared dual rapid acquisition of gradient echo, which was used in their study as a recently developed technology obtains a purely T1-weighted image and enables the generation of wholebrain T1 tissue relaxation time maps for quantitative tissue characterization.³⁸⁾ The descriptive properties of T1 relaxometry are of particular interest in the preterm group because they provide structural information about tissue, such as water content and lipid and macromolecule composition, as well as predict the chronological maturation of myelin and can be regarded as one of the best markers of brain maturation. In the WM fiber tracts (PLIC, optic radiation, and corona radiata), the linear drop in ADC and T1 represented a reduction in water content, fiber packaging, and early myelination processes, particularly in the PLIC after 36 weeks. The T1 relaxation levels (milliseconds) measured in the PLIC matured rapidly compared to those in each other ROI.

In another study, Natu et al.²⁶⁾ quantitatively examined T1 in 4 key sensorimotor regions. An inversion recoveryecho planar imaging (IR-EPI) sequence was used to measure the relaxation time (R1) at every voxel, an IR-EPI sequence was utilized. Spoiled gradient echo images were used in conjunction with EPI sequences to produce synthetic T1-weighted whole-brain images. R1 (R1=1/T1) was estimated for each voxel using the IR-EPI data. Contrary to the assumption that visual areas are already myelinated at birth,³⁹⁾ their investigation of R1 indicated that while the primary visual cortex (V1) is more mature than other visual areas at birth, it continues to develop and myelinate extensively over the first 6 months of life. Second, contrary to the belief that primary sensory-motor regions (such as V1) also myelinate the swiftest postnatally,³⁹⁾ an evaluation of the rate of R1 development during the first 6 months of life demonstrates that V1 does not myelinate the fastest in the human visual system. Despite having lower values at birth, R1 across several visual areas catches up to R1 in V1 by 6 months of age. The third finding was that microstructural development differed throughout the visual hierarchy. Thus, at birth, the higher visual areas of the ventral and dorsal streams had lower R1 values and were presumably less myelinated than the early retinotopic areas. In contrast, the development of R1 (and subsequently myelin) was increasingly rapid in later visual areas than in earlier areas.

2) T1/T2 ratio

Soun et al.¹⁴⁾ evaluated the ratio intensity values in 10 term neonates with no injury and hypothesized that the values may be sensitive to hyperintensities such as those observed in neonates with encephalopathy who experience injury. The T1/T2 ratio approach emphasizes the natural contrast of myelin.⁴⁰ The superimposition of opposing features of T1 and T2 weighting was used in this ratio method. The T1/ T2 ratio has the advantage of enhancing the WM contrastto-noise ratio without increasing scan time. Furthermore, the T1/T2 ratio may differentiate between highly and lightly myelinated cortical areas, implying that this technique can be utilized to investigate spatial variation in myelin formation in the neonatal brain. While no patient in their study population was known to have hypoxic-ischemic injury, the undiscovered subthreshold effects of hypoxia may have increased the T1 signal intensity in the PLIC and been mistaken as myelin tissue.

Owing to the rapid development of the brain during the first few months after birth, significant regional differences occur in myelin density. Their findings corroborate prior research indicating that the PLIC is highly myelinated by term birth, whereas the optic radiation is not fully myelinated.²⁾ According to these studies, despite variations in tissue characteristics between neonates and adults, the T1/T2 ratio indicates myelin density.

Coregistration is necessary for sequence comparison. The T1 and T2 images exhibited moderate misregistration abnormalities, particularly at the cerebrospinal-gray matter interfaces. These abnormalities may lead to incorrect contrast increases that could be mischaracterized as a myelin-related disorder. The T1/T2 ratio, if proven successful, could enhance the ability to detect myelin impairment in the developing brain. Additionally, because this approach uses routinely acquired diagnostic sequences, it does not require any extra scan time, which is critical in the neonatal population, in which movement is a major contributing factor to image quality.

3) T2* and R2*

Susceptibility-weighted imaging detects phase shifts and improves the visibility of iron, calcifications, veins, and blood byproducts based on paramagnetic or diamagnetic characteristics.⁴¹⁻⁴³

Relaxation time T2* is susceptible to deoxyhemoglobin, water content, and iron levels, with an even higher sensitivity than T2. It has been utilized to analyze brain function (due to changes in deoxyhemoglobin levels), myocardial oxygenation, tumor hypoxia, hemorrhage, cardiac calcification (low water content), and liver and cardiac iron.³⁷⁾ Several studies²²⁾ have found a correlation between T2* values and myelin content, which reflects nuclear interactions and field inhomogeneity due to the presence of paraor diamagnetic substances in iron and myelin. However, the neonatal brain is less likely to have high concentrations of iron and calcium because myelin is most likely the primary contributor to T2* values in the neonatal brain. The differences in susceptibility between myelin and most other water-based tissues result in signal intensity degradation in T2*-weighted imaging, which causes brain regions with higher myelin content to appear darker.⁴¹⁾

Although susceptibility- and T2*-based methods are primarily used to detect calcium deposition, measure iron content, and image hemorrhagic infarcts in adults,⁴⁴⁾ numerous studies have reported that myelin content is the primary contributor to T2* decay,²²⁾ particularly in neonates who are less likely to have high concentrations of iron and calcium. Considering the multifactorial nature of T2*, several studies suggested that myelin content may be the most significant contributor to this signal.²²⁾

Olivieri et al.⁵⁾ employed T2* mapping to study brain myelination in healthy neonates and those with NE throughout the first month of life. Their findings support previous histopathological studies reporting that T2* values were significantly higher in neonates with NE who developed injury in the PLIC, thalami, and lentiform nuclei, which are actively myelinated around the time of term birth and in the month thereafter.⁴⁵⁾ Higher T2* values indicate a lower myelin content, while lower T2* values indicate a higher myelin content. In addition, this approach has a high signal-to-noise ratio (SNR) and a short scanning period of less than 4 min, allowing it to be readily implemented in baseline procedures for imaging healthy and critically ill neonates.

Ning et al. $(2014)^{22}$ used a variant of this technique called enhanced T2*-weighted angiography, which focused on R2* values (R2*=1/T2*) to analyze the myelin content in termborn healthy infants who were approximately 3 months old at the time of MRI. They found that R2* values in the gray nuclei and WM were positively associated with postmenstrual age. The phase values of the PLIC and splenium of the CC were substantially greater than those of the anterior limb of the internal capsule (ALIC) and genu of the CC in neonates. However, they mentioned that, owing to infants' greater brain water content and significantly lower iron concentration, the boundaries of many brain regions are insufficiently distinct for automated segmentation and registration. In addition, they have more brain water than adults, which diminishes with age. Reduced water fractions may have influenced the change in R2* value with age.⁴⁵⁾

4) Myelin-water fraction

The T2 relaxation time is susceptible to tissue composition and structure as well as iron and water content. It has been used to evaluate myelin content in the brain; inflammation, collagen composition, and structure of the cartilage and heart muscle; and edema, hemorrhage, and iron content in the heart and liver.³⁷⁾ A multicomponent analysis of T1 and T2 relaxation, known as multicomponent relaxometry, may offer a more specific indicator of myelin maturation.⁴⁶⁾ A T2 decay analysis of the brain parenchyma revealed at least 2 discrete microanatomical water domains: a rapidrelaxing water pool normally related to water bound in the lipid bilayers of the myelin sheath and a slower-relaxing water pool pertaining to water within and outside of the myelinated axon.⁴⁶⁾

The myelin-water fraction (MWF) is a quantitative T2 measurement that enhances the pathological specificity of myelin breakdown.⁴⁷⁾ The MWF is estimated using a multi-exponential T2 fit, with a short T2 component (10–50 msec) attributed to water trapped between the myelin sheaths. Histopathology has established an association between MWF and myelin loss.⁴⁸⁾ Myelin content can be measured indirectly by measuring the myelin-bound water signal or MWF, which correlates well with the results of histological evaluations.⁴⁸⁾

Multicompartment T2 relaxometry has been used³⁰⁾ to assess myelin by quantifying the increase in MWF in the thalamus and posterior WM (PWM) areas; the increase in the thalamus is significantly greater. In the thalamus, the overall change in T2 values was attributable to an increase in myelination, whereas in the WM, the change was associated with a decrease in free water content and an increase in tissue volume. In the PWM, the amount of myelin measured was rather insignificant, and the main change was tentatively attributed to (unmyelinated) axonal and glial proliferation confirmed by T2 relaxometry. Results from the WM PLIC and ALIC demonstrated axonal and myelin density differentiation, with high MWF values in the PLIC and significantly lower values in the ALIC.

Meanwhile, Knight et al.²⁸⁾ found that T2 is longer in extremely preterm newborns than in late preterm infants across many WM regions, and these effects are particularly pronounced in WM regions that myelinate earlier and rapidly. Deoni et al.⁴⁹⁾ further established the use of multicomponent relaxometry in the investigation of infant brain development, demonstrating qualitative agreement between MWF trends acquired from magnetic resonance (MR) and spatiotemporal myelination patterns found by histology. Deoni et al.²³⁾ assessed myelin content in healthy infants aged 3 months to 7.5 years using a new multicomponent relaxation approach called multicomponent driven equilibrium single-pulse observation of T1 and T2. Despite its superior resolution, this method is not ideal for neonatal patients owing to the lengthier scan periods,¹⁴⁾ making it more challenging to incorporate into the baseline MRI procedure in a critically ill neonate population. Concerns have recently been raised regarding its accuracy.⁵⁰⁾

However, a significant disadvantage of this technique is its relatively poor SNR,¹⁵⁾ which makes imaging more difficult because of the low myelin concentration in the neonatal brain. Until recently, obtaining MWF maps in clinical practice was challenging because acquisition was limited to a single slice and the scanning time was unduly long.³⁷⁾

5) Synthetic MRI

Specific parameters, such as the T1-relaxation constants, T2-relaxation constants, and proton density (PD) of the studied tissue, can be measured using the quantitative multidynamic multiecho (MDME) sequence which acquires all the required parameters for image postprocessing in less than 6 minutes.⁵¹⁾ Since the repetition time, echo time, and inversion time are all extrinsic scan parameters that can be established and adjusted postprocessing, they are not predefined in this method.⁵²⁾ T1-weighted, T2-weighted, PD-weighted, and inversion recovery contrasts can all be produced from SyMRI in less than 1 min after intrinsic tissue parameters have been collected and extrinsic scan parameters have been specified.⁵¹⁾

Lee et al.²⁴⁾ and Schmidbauer et al.²⁷⁾ used a vendor-provided tool (SyMRI 8.0; SyntheticMR, Linkoping, Sweden) to construct quantification maps (T1, T2, and PD maps) simultaneously from raw data acquired by the MDME sequence. Using the MDME pulse sequence, the partial volume of myelin was calculated using R1, R2, and PD.⁵³⁾

The SyMRI software generates maps relying on T1- and T2-relaxation constants and the PD, which vary among different tissues.⁴⁶⁾ This permits the imaging of various tissue types and the quantification of myelin, resulting in a more accurate evaluation of brain development and demyelinating disorders.⁵⁴⁾ Using quantitative maps, myelin could be better differentiated from non-myelinated gray and WM than on conventional images.

The SyMRI software automatically calculated the total

estimated myelin partial volume (Msum) and the ratio of Msum to the total brain parenchyma.²⁴⁾ T1 and T2 of the WM areas were extremely high at birth, with maximum values of approximately 2,600 msec and 280 msec, respectively; however, these values reduced by more than half in the first year of life.

Time is a crucial factor in clinical settings. Even using current techniques, conventional approaches for MR mapping would take 15-30 minutes.⁵¹⁾ Beyond the neonatal brain imaging technique, SyMRI is a quantitative imaging technique that detects myelination-related changes on an individual level. In addition, it typically reduces the examination time while offering several MR contrasts such as T1, T2, PD, and inversion recovery. Thus, SyMRI enables the generation of quantitative maps and different MR contrasts in one-third of the time required by typical quantitative mapping methods. Recent evidence indicates that SvMRI imaging data are equivalent to those of conventional T1 and T2 images.⁵²⁾ In addition to standard MR contrasts, SyMRI enables the rapid generation of quantitative MR maps.⁵¹⁾ This suggests new avenues in diagnostic neonatal brain imaging.

2. Magnetization transfer imaging

MTI is an MRI technique that was developed to investigate the properties of nonwater tissue components. The MTI theory considers the influence of additional parameters, namely those reflecting the exchange of protons between water molecules and molecules of more solid structural components. Consequently, MTI can reveal tissue structures and structural components that are typically not discernible using standard MRI. This theory assumes that the spin magnetization of tissue macromolecular components can be indirectly observed by observing the normally visible spins of tissue water. The magnetization transfer ratio (MTR) is an adjusted index derived from 2 MR images that reflect the degree of magnetization transfer. The MTR is a semiquantitative marker of myelination in the brain in its most basic form.⁵⁵)

The PLIC, which is not as densely packed as the genu and splenium of the CC⁵⁶⁾ and is in the premyelinating stage in the preterm period, has a significantly lower macro-molecular density (lower MTR values) than the ventrolateral thalamic nucleus (VLN) and CC structures and higher water content (higher T1 values) than myelinated gray matter. However, the VLN and the posterior part of the PLIC had greater MTR values than the nonmyelinated CC at term-equivalent age while remaining in the same order of FA and axial diffusivity values and exhibiting the lowest mean diffusivity and radial diffusivity values, together with the pons. These alterations indicate myelination.

During axonal myelination, the ratio of free and bound

water molecules is assumed to represent the myelin content.⁵⁷⁾ However, some studies have highlighted the contentious results achieved by applying MTR to measure myelination. Myelination was found to be inaccurately described MTR values in an experimental mouse model of autoimmune encephalomyelitis, which is characterized by lymphocyte-mediated inflammation followed by demyelination, axonal degeneration, and neuronal loss and did not correlate with the histopathological myelin content according to one such study.⁵⁵⁾ Based on these and other findings,⁵⁵⁾ the MTR approach may be more sensitive to inflammatory diseases than myelination and demyelination.

3. Automatic segmentation technique of myelin-like signals

Numerous advancements in radiology have been attributed to interdisciplinary collaborations in the field of engineering. A significant example is the application of segmentation techniques to analyze different anatomical structures within medical imaging data, thereby enhancing the diagnostic potential of such images. Using T2-weighted MRI, Wang et al.⁸⁾ presented quantitative measures of myelination patterns in preterm infants with a gestational age of 29–44 weeks. Based on an innovative automatic segmentation method for myelin-like signals (MLSs) on T2-weighted MRI, Wang et al.⁸⁾ developed a quantitative marker for myelination to assess preterm neonatal brain development. On T2-weighted neonatal brain MR images, they referred to the tissue that was likely to consist of myelin as MLS.

To create quantifiable markers of myelination, the suggested segmentation method was applied to T2-weighted scans of 114 preterm newborns. First, they performed a volumetric examination of the progression of myelination and demonstrated that myelination increases in regions of the deep brain while remaining unchanged in the brainstem. Second, they constructed a spatiotemporal model of myelination progression and compared it to qualitative studies of myelination.⁵⁸⁾ In conclusion, they demonstrated that the spatiotemporal atlas of progressive myelination may accurately predict gestational age at the time of the scan and, thus, make a possible quantitative diagnosis of developmental delay.

Developing an automatic approach for segmenting MLS, which can then be used in volumetric analyses and spatiotemporal modeling of progressive myelination, is a strategy for developing quantitative indicators of myelination. Automatic MLS segmentation is difficult and there are currently no dedicated techniques for segmenting MLS throughout the preterm and neonatal periods. In the NeoBrainS12 challenge,⁵⁹⁾ recently developed approaches for neonatal brain segmentation showed great potential for segmenting a variety of brain structures on neonatal brain MR images. However, none of these methods perform well in terms of myelination segmentation. Most neonatal brain segmentation algorithms use a probabilistic atlas or manual annotations to collect previous knowledge of the estimated tissue placement as developed for adults. However, myelin is not included in any available neonatal brain atlases or manual annotation databases, nor was it considered in the development of the Human Connectome Project⁶⁰ segmentation technique. Another issue that causes correct segmentation of the MLS in the prenatal stage is its low volume compared to MRI resolution. Therefore, the partial volume effect (i.e., mixing 2 or more tissues in a single voxel) must be considered. Many approaches for partial volume modeling have been proposed; however, they all require prior knowledge of the site of tissue mixing, which is currently unavailable for MLS.

Although the segmentation method⁸⁾ provides a quantitative assessment of the volume of myelinated tissue, it cannot quantify the myelin fraction in each voxel. Quantitative approaches such as relaxometry and MTI can generate more precise measurements of myelin maturation if substantial databases in this age range are obtained.

Conclusion

A comprehensive review of research that validates the quantification of myelin using MRI to consolidate the existing knowledge on neonatal subjects has been provided. From the authors' viewpoint, SyMRI holds great potential in pediatrics for visualizing the developing brain because contrast images can be optimized post-scanning.54) In addition, the maps generated using this technique are coregistered because of the simultaneous quantification of R1 and R2 relaxation rates and PD. A significant advancement over conventional quantification approaches that require separate scans of each dataset has been observed. SyMRI offers the potential to reduce the overall MRI scan time, particularly when multiple contrast-weighted images are required, which is common in clinical practice. It has the potential benefit of a shortened scan time, especially for pediatric or uncooperative patients. SyMRI is not widely used in clinical settings because of its postprocessing time. However, using the SyMRI software, postprocessing required less than 1 min. A fast processing time is a clear benefit for clinical application. A quick scanning time is essential for neonatal imaging and reduces the need for sedation. Thus, considering all of these factors, researchers favor SyMRI as the most ideal and versatile approach for quantifying myelin in neonates.

Footnote

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References

- 1. Raine CS. Morphology of myelin and myelination. In: Morell P, editor. Myelin. Boston (MA): Springer, 1984:1-50.
- 2. Barkovich AJ. Concepts of myelin and myelination in neuroradiology. AJNR Am J Neuroradiol 2000;21:1099-109.
- 3. Spader HS, Ellermeier A, O'Muircheartaigh J, Dean DC 3rd, Dirks H, Boxerman JL, et al. Advances in myelin imaging with potential clinical application to pediatric imaging. Neurosurg Focus 2013;34:E9.
- 4. Pujol J, Vendrell P, Junque C, Martia-Vilalta JL, Capdevila A. When does human brain development end? Evidence of corpus callosum growth up to adulthood. Ann Neurol 1993;34:71-5.
- Olivieri B, Rampakakis E, Gilbert G, Fezoua A, Wintermark P. Myelination may be impaired in neonates following birth asphyxia. NeuroImage Clin 2021:31:102678.
- 6. Widjaja E, Kis A, Go C, Raybaud C, Snead OC, Smith ML. Abnormal white matter on diffusion tensor imaging in children with new-onset seizures. Epilepsy Res 2013;104:105-11.
- Beaulieu C, Yip E, Low PB, M\u00e4dler B, Lebel CA, Siegel L, et al. Myelin water imaging demonstrates lower brain myelination in children and adolescents with poor reading ability. Front Hum Neurosci 2020;14:568395.
- 8. Wang S, Ledig C, Hajnal JV, Counsell SJ, Schnabel JA, Deprez M. Quantitative assessment of myelination patterns in preterm neonates using T2-weighted MRI. Sci Rep 2019;9:12938.
- 9. Kulikova S, Hertz-Pannier L, Dehaene-Lambertz G, Poupon C, Dubois J. A new strategy for fast MRI-based quantification of the myelin water fraction: application to brain imaging in infants. PLoS One 2016;11:e0163143.
- Yu N, Kim JY, Han D, Kim SY, Lee HM, Kim DH, et al. Threedimensional magnetic resonance fingerprinting in neonates. Invest Radiol 2022;57:44-51.
- 11. Carmody DP, Dunn SM, Boddie-Willis AS, DeMarco JK, Lewis M. A quantitative measure of myelination development in

infants, using MR images. Neuroradiology 2004;46:781-6.

- Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gøtzsche PC, Ioannidis JPA, et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. J Clin Epidemiol 2009;62:e1-34.
- 13. Deoni SCL, Mercure E, Blasi A, Gasston D, Thomson A, Johnson M, et al. Mapping infant brain myelination with magnetic resonance imaging. J Neurosci 2011;31:784-91.
- 14. Soun JE, Liu MZ, Cauley KA, Grinband J. Evaluation of neonatal brain myelination using the T1- and T2-weighted MRI ratio. J Magn Reson Imaging 2017;46:690-6.
- 15. Weber AM, Zhang Y, Kames C, Rauscher A. Myelin water imaging and R2* mapping in neonates: Investigating R2* dependence on myelin and fibre orientation in whole brain white matter. NMR Biomed 2020;33:e4222.
- Martín-martín A, Orduna-malea E, López-cózar ED, Martínmartín A. Google Scholar, Web of Science, and Scopus: a systematic comparison of citations in 252 subject categories. Scientometrics 2021;126:871-906.
- 17. Gusenbauer M, Haddaway NR. Which academic search systems are suitable for systematic reviews or meta-analyses? Evaluating retrieval qualities of Google Scholar, PubMed, and 26 other resources. Res Synth Methods 2020;11:181-217.
- Falagas ME, Pitsouni EI, Malietzis GA, Pappas G. Comparison of PubMed, Scopus, Web of Science, and Google Scholar: strengths and weaknesses. FASEB J 2008;22:338-42.
- 19. Okoli C. A guide to conducting a standalone systematic literature review. Commun Assoc Inf Syst 2015;37:879-910.
- 20. Grotheer M, Rosenke M, Wu H, Kular H, Querdasi FR, Natu VS, et al. White matter myelination during early infancy is linked to spatial gradients and myelin content at birth. Nat Commun 2022;13:997.
- 21. Whiting PF, Rutjes AWS, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med 2011;155:529-36.
- 22. Ning N, Zhang L, Gao J, Zhang Y, Ren Z, Niu G, et al. Assessment of iron deposition and white matter maturation in infant brains by using enhanced T2 star weighted angiography (ESWAN): R2* versus phase values. PLoS One 2014;9:e89888.
- 23. Deoni SCL, Dean DC, O'Muircheartaigh J, Dirks H, Jerskey BA. Investigating white matter development in infancy and early childhood using myelin water faction and relaxation time mapping. Neuroimage 2012;63:1038-53.
- 24. Lee SM, Choi YH, You SK, Lee WK, Kim WH, Kim HJ, et al. Age-related changes in tissue value properties in children: simultaneous quantification of relaxation times and proton density using synthetic magnetic resonance imaging. Invest Radiol 2018;53:236-45.
- 25. Nossin-Manor R, Card D, Morris D, Noormohamed S, Shroff MM, Whyte HE, et al. Quantitative MRI in the very preterm brain: Assessing tissue organization and myelination using magnetization transfer, diffusion tensor and T1 imaging. Neuroimage 2013;64:505-16.
- 26. Natu VS, Rosenke M, Wu H, Querdasi FR, Kular H, Lopez-Alvarez N, et al. Infants' cortex undergoes microstructural growth coupled with myelination during development. Commun Biol 2021;4:1191.
- 27. Schmidbauer V, Geisl G, Diogo M, Weber M, Goeral K, Klebermass-Schrehof K, et al. SyMRI detects delayed myelination in preterm neonates. Eur Radiol 2019;29:7063-72.
- 28. Knight MJ, Smith-Collins A, Newell S, Denbow M, Kauppinen

RA. Cerebral white matter maturation patterns in preterm infants: an mri t2 relaxation anisotropy and diffusion tensor imaging study. J Neuroimaging 2018;28:86-94.

- 29. Schneider J, Kober T, Bickle Graz M, Meuli R, Hüppi PS, Hagmann P, et al. Evolution of T1 relaxation, ADC, and fractional anisotropy during early brain maturation: a serial imaging study on preterm infants. Am J Neuroradiol 2016;37:155-62.
- 30. Melbourne A, Eaton-Rosen Z, Orasanu E, Price D, Bainbridge A, Cardoso MJ, et al. Longitudinal development in the preterm thalamus and posterior white matter: MRI correlations between diffusion weighted imaging and T2 relaxometry. Hum Brain Mapp 2016;37:2479-92.
- Dubois J, Dehaene-Lambertz G, Kulikova S, Poupon C, Hüppi PS, Hertz-Pannier L. The early development of brain white matter: a review of imaging studies in fetuses, newborns and infants. Neuroscience 2014;276:48-71.
- 32. Dietrich RB, Bradley WG, Zaragoza EJ 4th, Otto RJ, Taira RK, Wilson GH, et al. MR evaluation of early myelination patterns in normal and developmentally delayed infants. AJR Am J Roentgenol 1988;150:889-96.
- 33. Paus T, Collins DL, Evans AC, Leonard G, Pike B, Zijdenbos A. Maturation of white matter in the human brain: a review of magnetic resonance studies. Brain Res Bull 2001;54:255-66.
- Brody BA, Kinney HC, Kloman AS, Gilles FH. Sequence of central nervous system myelination in human infancy. I. An autopsy study of myelination. J Neuropathol Exp Neurol 1987; 46:283-301.
- 35. Kinney HC, Brody BA, Kloman AS, Gilles FH. Sequence of central nervous system myelination in human infancy. II. Patterns of myelination in autopsied infants. J Neuropathol Exp Neurol 1988;47:217-34.
- Dubois J, Alison M, Counsell SJ, Hertz-Pannier L, Hüppi PS, Benders MJNL. MRI of the neonatal brain: a review of methodological challenges and neuroscientific advances. J Magn Reson Imaging 2021;53:1318-43.
- Margaret Cheng HL, Stikov N, Ghugre NR, Wright GA. Practical medical applications of quantitative MR relaxometry. J Magn Reson Imaging 2012;36:805-24.
- Marques JP, Kober T, Krueger G, van der Zwaag W, Van de Moortele PF, Gruetter R. MP2RAGE, a self bias-field corrected sequence for improved segmentation and T1-mapping at high field. Neuroimage 2010;49:1271-81.
- Yakovlev PI, Lecours AR. The myelogenetic cycles of regional maturation of the brain. In: Minkowski A, editor. Regional development of brain in early life. Oxford: Blackwell, 1967:3-70.
- Glasser MF, van Essen DC. Mapping human cortical areas in vivo based on myelin content as revealed by T1- and T2weighted MRI. J Neurosci 2011;31:11597-616.
- Chavhan GB, Babyn PS, Thomas B, Shroff MM, Mark Haacke E. Principles, techniques, and applications of T2*-based MR imaging and its special applications. Radiographics 2009;29: 1433-49.
- Haacke EM, Xu Y, Cheng YCN, Reichenbach JR. Susceptibility weighted imaging (SWI). Magn Reson Med 2004;52:612-8.
- Sehgal V, Delproposto Z, Haacke EM, Tong KA, Wycliffe N, Kido DK, et al. Clinical applications of neuroimaging with susceptibility-weighted imaging. J Magn Reson Imaging 2005; 22:439-50.
- Wu G, Xi G, Hua Y, Sagher O. T2* Magnetic resonance imaging sequences reflect brain tissue iron deposition following intracerebral hemorrhage gang. Transl Stroke Res 2010;1:31-4.
- 45. Mukherjee P, Miller JH, Shimony JS, Conturo TE, Lee BCP,

Almli CR, et al. Normal brain maturation during childhood: Developmental trends characterized with diffusion-tensor MR imaging. Radiology 2001;221:349-58.

- Whittall KP, MacKay AL, Graeb DA, Nugent RA, Li DKB, Paty DW. In vivo measurement of T2 distributions and water contents in normal human brain. Magn Reson Med 1997;37:34-43.
- Mackay A, Kenneth W, Julian A, David L, Donald P, Douglas G. In vivo visualization of myelin water in brain by magnetic resonance. Magn Reson Med 1994;31:673-7.
- Laule C, Leung E, Li DKB, Traboulsee AL, Paty DW, MacKay AL, et al. Myelin water imaging in multiple sclerosis: Quantitative correlations with histopathology. Mult Scler 2006;12:747-53.
- 49. Deoni SCL. Correction of main and transmit magnetic field (B0 and B 1) inhomogeneity effects in multicomponent-driven equilibrium single-pulse observation of T1 and T2. Magn Reson Med 2011;65:1021-35.
- West DJ, Teixeira RPAG, Wood TC, Hajnal JV, Tournier JD, Malik SJ. Inherent and unpredictable bias in multi-component DESPOT myelin water fraction estimation. Neuroimage 2019; 195:78-88.
- 51. Hagiwara A, Warntjes M, Hori M, Andica C, Nakazawa M, Kumamaru KK, et al. SyMRI of the brain: rapid quantification of relaxation rates and proton density, with synthetic MRI, automatic brain segmentation, and myelin measurement. Invest Radiol 2017;52:647-57.
- 52. Trial M, Tanenbaum XLN, Tsiouris XAJ, Johnson XAN, Naidich XTP, Delano XMC, et al. Synthetic MRI for clinical neuroimaging: results of the magnetic resonance image compilation (MAGiC) prospective, multicenter, multireader trial. Am J Neuroradiol 2017;38:1103-10.
- 53. Warntjes M, Engström M, Tisell A, Lundberg P. Modeling the presence of myelin and edema in the brain based on multi-parametric quantitative MRI. Front Neurol 2016:7:16.
- 54. West H, Leach JL, Jones BV, Care M, Radhakrishnan R, Merrow AC, et al. Clinical validation of synthetic brain MRI in children: initial experience. Neuroradiology 2017;59:43-50.
- Fjær S, Bø L, Myhr KM, Torkildsen O, Wergeland S. Magnetization transfer ratio does not correlate to myelin content in the brain in the MOG-EAE mouse model. Neurochem Int 2015;83-84:28-40.
- 56. Aboitiz F, Scheibel AB, Fisher RS, Zaidel E. Fiber composition of the human corpus callosum. Brain Res 1992;598:143-53.
- Kucharczyk W, Macdonald PM, Stanisz GJ, Henkelman RM. Relaxivity and magnetization transfer of white matter lipids at MR imaging: Importance of cerebrosides and pH. Radiology 1994;192:521-9.
- Barkovich AJ. Magnetic resonance techniques in the assessment of myelin and myelination. J Inherit Metab Dis 2005;28:311-43.
- 59. Wang S, Kuklisova-Murgasova M, Schnabel JA. An atlas-based method for neonatal MR brain tissue segmentation. MICCAI NeoBrainS12 2012;1-8.
- 60. Makropoulos A, Robinson EC, Schuh A, Wright R, Fitzgibbon S, Bozek J, et al. The developing human connectome project: a minimal processing pipeline for neonatal cortical surface reconstruction. Neuroimage 2018;173:88-112.

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