

EFFECTS OF Chlorella vulgaris ENRICHED Artemia sp. AND Moina sp. TO ENHANCE GROWTH PERFORMANCE OF KELAH, Tor tambroides (BLEEKER, 1854) FRY



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

> July 2023 IB 2023 4

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

 \bigcirc



DEDICATION

I would like to dedicate to this dissertation to myself – thank you for hanging on. To my parents, siblings and partner who are always supportive of my ambitions and nurturing me with endless affection and love, you all inspire me to be a better version of myself every day.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFECTS OF Chlorella vulgaris ENRICHED Artemia sp. AND Moina sp. TO ENHANCE GROWTH PERFORMANCE OF KELAH, Tor tambroides (BLEEKER, 1854) FRY

By

WIZILLA JANTI ANAK JOSHUA

July 2023

Chair : Zarirah Mohamed Zulperi, PhD Institute: Bioscience

Tor tambroides is a species valued in Malaysian aquaculture but faces limitations in its potential due to slow growth. This study was designed to investigate the effects of Chlorella vulgaris enriched Artemia sp. and Moina sp. to enhance growth performance of Kelah (Tor tambroides) fry. The objectives of this study were to determine the nutritional profile of *Chlorella* sp. as potential candidate for live food enrichment; to evaluate and compare the composition of unenriched and C. vulgaris enriched Artemia sp. and Moina sp.; and to assess on the effects of C. vulgaris enriched Artemia sp. and Moina sp. in comparison to unenriched counterparts on the growth performance, survival and expression of growth and immune genes of Tor tambroides fry. The identification of C. vulgaris targeting 18S rRNA gene resulted nucleotide sequence of Chlorella sp. with 99.72% to 100% matches with C. vulgaris. The obtained lipid content was 3.29%, with high PUFA content (67.04%). In the second study, Artemia sp. and Moina sp. were enriched with C. vulgaris. The two-way ANOVA revealed that the type of livefood (Artemia and Moina) and unenrichment versus enrichment had significantly influenced the moisture and ash content (p < 0.05). The enriched Artemia had the highest lipid content (25.68%) and it was statistically different as compared to the unenriched and enriched Moina (p<0.05). The linoleic acid was the highest in enriched Moina (12.19%). The total PUFA was the highest in unenriched Artemia (41.61%), followed by enriched Artemia (39.90%) but both treatments were not significant to one another (p>0.05). In the third study, the fries were divided into four diet treatments; unenriched Artemia (UA); enriched Artemia (EA); unenriched Moina (UM); enriched Moina (EM). The growth parameters were significantly higher in fries group fed with enriched Artemia (p<0.05). The variations of the growth genes suggested that there were differences in growth rate in the fry groups, meanwhile the expression of MSTN was proposed to be related to other physiological processes. The immune response genes showed similar pattern expressions in enriched Artemia, suggesting its immunocompetency values. In conclusion, enrichment of Artemia with C. vulgaris enhances the growth performance and expressions of growth and immune response genes, making it a promising diet for slowgrowing species like Tor tambroides.

Keywords: Artemia; Chlorella vulgaris; enrichment; Moina; Tor tambroides



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENGARUH Artemia sp. DAN Moina sp. DIPERKAYAKAN DENGAN Chlorella vulgaris DALAM MENINGKATKAN PRESTASI PERTUMBUHAN BENIH KELAH, Tor tambroides (BLEEKER, 1854)

Oleh

WIZILLA JANTI ANAK JOSHUA



Tor tambroides adalah ikan yang bernilai tinggi dalam akuakultur Malaysia. Spesies ini menghadapi cabaran disebabkan pertumbuhan yang perlahan. Kajian ini fokus pada penggunaan Artemia sp. dan Moina sp. yang diperkaya dengan Chlorella vulgaris untuk meningkatkan prestasi pertumbuhan anak ikan Kelah (Tor tambroides). Objektif kajian merangkumi mengenal pasti profil nutrisi Chlorella sp. sebagai calon untuk pengayaan makanan hidup, membanding komposisi Artemia sp. dan Moina sp. yang diperkaya dan tidak diperkaya dengan C. vulgaris, serta menilai kesan pada pertumbuhan, ketahanan, dan ekspresi gen pertumbuhan dan imun anak ikan Tor tambroides. Pengenalpastian C. vulgaris melibatkan gen 18S rRNA, menunjukkan padanan nukleotida 99.72% hingga 100%, dengan C. vulgaris. Kandungan lipid yang diperoleh adalah 3.29%, dan terdapat 22 asid lemak dengan kandungan PUFA yang tinggi (67.04%). Dalam kajian kedua, Artemia sp. dan Moina sp. masing-masing diperkayakan dengan C. vulgaris selama 6 jam dan 24 jam. Analisis ANOVA dua hala menunjukkan bahawa jenis makanan hidup dan pengayaan mempengaruhi kandungan lembapan dan abu secara signifikan (p<0.05). Artemia yang diperkaya menunjukkan kandungan lipid tertinggi (25.68%), berbeza secara statistik berbanding dengan Moina (p<0.05). Asid linoleik tertinggi terdapat dalam Moina yang diperkaya (12.19%). Kandungan PUFA tertinggi dalam Artemia yang tidak diperkaya (41.61%), diikuti oleh Artemia yang diperkaya (39.90%), tanpa perbezaan signifikan antara keduanya (p>0.05). Dalam kajian ketiga, anak ikan dibahagikan kepada empat rawatan makanan; Artemia yang tidak diperkaya (UA); Artemia yang diperkaya (EA); Moina yang tidak diperkaya (UM); Moina yang diperkaya (EM). Parameter pertumbuhan menunjukkan prestasi yang lebih tinggi secara signifikan dalam kumpulan anak ikan yang diberi Artemia yang diperkaya (p<0.05). Varian gen pertumbuhan menunjukkan perbezaan dalam kadar pertumbuhan di kalangan kumpulan anak ikan, sementara ekspresi MSTN dikemukakan berkaitan dengan proses fisiologi lain. Gen respons imun menunjukkan corak ekspresi yang serupa dalam Artemia yang diperkaya, menunjukkan nilai imunokompeten. Kesimpulannya, pengayaan Artemia dengan C. vulgaris meningkatkan prestasi pertumbuhan dan ekspresi gen pertumbuhan

dan respons imun, menjadikannya diet yang berpotensi untuk spesies yang pertumbuhannya perlahan seperti *Tor tambroides*.



ACKNOWLEDGEMENTS

I would like to sincerely thank my supervisor, Dr. Zarirah Mohamed Zulperi for her guidance, continuous support and motivation throughout this study. I would also like to extend my appreciation to my co-supervisors, Prof. Dr. Mohd Salleh Kamarudin and Assoc. Prof. Dr. Natrah Fatin Mohd Ikhsan for providing advice on their expertise and thoughtfulness during my research period. Their encouragements had inspired me to become a better individual.

Not to mentioned, I would like to reach out my gratitude to the Ministry of Higher Education (MOHE) Malaysia through the Malaysia-Japan SATREPS-COSMOS (JPMJSA 1509) project for funding this study. My deepest appreciation also goes to all the personnel at Aquahealth Laboratory, Institute of Bioscience, Department of Aquaculture, Faculty of Agriculture and Mariculture (AquaLab), International Institute of Aquaculture and Aquatic Sciences (I-AQUAS), UPM for their direct and indirect involvement during my postgraduate journey.

Last but not least, the acknowledgement goes to my parents, siblings and partner who supported me emotionally and believed in my passions and dreams.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Zarirah binti Mohamed Zulperi, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Mohd Salleh bin Kamarudin, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

Natrah Fatin binti Mohd Ikhsan, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 14 March 2024

TABLE OF CONTENTS

n

ABSTRACT ABSTRAK ACKNOWI APPROVA DECLARA LIST OF TA LIST OF FI LIST OF A	LEDGI L TION ABLES IGURE	S ES	i iii v vi viii xiii xvi xviii
CHAPTER			
1	INTR	RODUCTION	
Ĩ	1.1 1.2 1.3 1.4	Background of Study Problem Statement Justification of Study Objectives	1 1 2 3
2	LITE	CRATURE REVIEW	
	2.1	Production of Tor tambroides, Kelah in Malaysia	4
	2.2	Tor tambrodies (Bleeker, 1854)	6
		2.2.1 Nomenclature and Taxonomy	7
		2.2.2 Distributions, Morphology and Biology	7
		2.2.3 Diets of <i>Tor tambroides</i>	8
	2.3	Moina and Artemia as Early Larval and Fry Diets	9
	2.4	Chlorella vulgaris	10
		2.4.1 Nomenclature of <i>Chlorella vulgaris</i>	10
		2.4.2 Potentials of enriched <i>Chlorella vulgaris</i> in	11
	2.5	live food for fish nutrition	12
	2.5 2.6	Growth and immune response in fish fry Enrichment of <i>Moina</i> and <i>Artemia</i> as live food	12
	2.0	Enrichment of <i>Molna</i> and Artemia as live lood	14
3	OF (ERMINATION OF THE NUTRITIONAL PROFILE Chlorella sp. AS POTENTIAL CANDIDATE FOR CFOOD ENRICHMENT	
	3.1	Introduction	16
	3.2	Methodology	17
	3.3	Samples Collection	17
		3.3.1 Molecular identification of <i>Chlorella</i> sp.	17
		3.3.1.1 Total RNA isolation	17
		3.3.1.2 Polymerase Chain Reaction (PCR) Amplification and DNA sequencing	18
	3.4	Phylogenetic Analysis of <i>Chlorella</i> sp.	18
	3.5	Culture Medium Preparation	19
	3.6	Culture of <i>Chlorella</i> sp.	19
		3.6.1 Cell counting	20

	3.6.2 3.6.3	Harvesting of <i>Chlorella</i> sp. Moisture, Ash and Lipid Analysis of <i>Chlorella</i> sp.	20 20
3.7 3.8	3.6.4 Statistica Results	Fatty acids analysis of <i>Chlorella vulgaris</i> I Analysis	21 22 22
	3.8.1	Molecular Identification of Chlorella sp.	22
	3.8.2	Growth curve	25
	3.8.3	Moisture, Ash and Lipid Analysis of <i>Chlorella</i> sp.	25
	3.8.4	Fatty acids analysis of <i>Chlorella vulgaris</i>	26
3.9	Discussio		27
3.10	Conclusi	on	30
		OF Artemia NAUPLII AND Moina sp. AND WITHOUT ENRICHMENT OF	
	ella vulgar		
4.1	Introduct		31
4.2	Methodol		33
	4.2.1	Preparation of livefood	33
	4.2.2	Preparation of Artemia sp.	33
	4.2.3	Preparation of <i>Moina</i> sp.	34
4.3		on and maintenance of Chlorella vulgaris as	35
	enrichme	nt media	
4.4	Enrichme <i>Chlorella</i>	ent of Artemia sp. and Moina sp. using sp.	36
	4.4.1	Enrichment of Artemia sp.	36
	4.4.2	Enrichment of Moina sp.	36
4.5		Ash and Lipid Analysis of enriched and ed Artemia sp. and Moina sp.	37
4.6	Fatty acid sp. and M	d analysis of enriched and unenriched Artemia loina sp.	38
4.7	Statistica	l Analysis	38
4.8	Results		39
	4.8.1	Observation of <i>Artemia</i> and <i>Moina</i> sp. during enrichment	39
	4.8.2	Comparison of Moisture, Ash and Lipid of unenriched and enriched Artemia and Moina	41
	4.8.3	Factors affecting the Moisture, Ash and Lipid of unenriched and enriched <i>Artemia</i> and <i>Moina</i>	44
	4.8.4	Fatty acid analysis of unenriched and enriched Artemia sp. and Moina sp.	46
4.9	Discussio	1 1	48
4 1 0	a 1 ·		C 1

4

6)

4.10 Conclusion 51

	ES OF Tor tamb	proides FRY
5.1	Introduction	
5.2	Methodology	
		nent of Ethics
5.2		les Collection
5.3 5.4	Experimental I Collection of I	
5.4 5.5	Growth perform	
5.5		Weight gained (mg)
		length (mm)
		fic growth rate (% d^{-1})
		ve growth rate (%)
5.6	Survival Rate (
5.7		rowth and Immune Response Genes
		RNA isolation
	5.7.2 Detern	nination of RNA Concentration
	5.7.3 First s	trand of cDNA synthesis
	5.7.4 Polym	
		fication and DNA sequencing
5.8		Growth and Immune Response genes
		titative Real-time PCR
5.9	Statistical Ana	lysis
5.10	Results	
		th performance of fries
	5.10.1	.1 Body Weight gained (mg).2 Final total length (mm)
	5.10.1	
	5.10.1	
		val rate of fries (%)
		ve expression of growth and immune
		nse genes of fries
5.11	Discussions	lie genee er mee
5.12	Conclusion	
	MARY,	CONCLUSION AND
	OMMENDATIO	

ASSESSMENT ON THE EFFECTS OF C. vulgaris

REFERENCES	85
APPENDICES	110
BIODATA OF STUDENT	113
LIST OF PUBLICATIONS	114

LIST OF TABLES

Table		Page
1	Production of <i>Tor tambroides</i> (in tonnes) based on the data by the Department of Fisheries Malaysia (2015-2020)	5
2	Summaries of previous studies on enriched Artemia and Moina	15
3	Primers sequences used for PCR amplification	18
4	BLAST similarity percentage of Chlorella sp. UPM	23
5	Percentage of dry weight, moisture, ash and lipid content of C. <i>vulgaris.</i> The data are presented as mean \pm SEM in % dry matter	25
6	Composition of fatty acids (% of total fatty acids) in <i>C. vulgaris</i>	26
7	Percentage of moisture, ash and lipid of the unenriched and enriched <i>Moina</i> and unenriched and enriched <i>Artemia</i> . The data is presented as mean \pm SEM in dry matter basis (%). One-way ANOVA test following post hoc analysis using Tukey's method to determine the statistically significant differences between the unenriched and enriched <i>Artemia</i> and <i>Moina</i> at 95% confidence level	42
8	Comparison of the moisture, ash and lipid content between unenriched and enriched <i>Artemia</i> , unenriched <i>Moina</i> and enriched <i>Moina</i> , based on independent sample t-test.	43
9	Two-way ANOVA results between the livefood (<i>Artemia</i> and <i>Moina</i>), enrichment (unenriched and enriched) and interaction between type of livefood and enrichment on the moisture, ash and lipid composition of the livefood (unenriched <i>Artemia</i> , unenriched <i>Moina</i> , enriched <i>Artemia</i> and enriched <i>Moina</i>)	45
10	Comparisons on the composition of fatty acids (% of total fatty acids) in unenriched and enriched <i>Artemia</i> and <i>Moina</i> . The data is presented in mean \pm SEM of each fatty acid and analysed using non-parametric test following Kruskal-Wallis test at 95% confidence level	47
11	Primers and their sequences for PCR amplification	59
12	Program and cycling conditions	60

13	The body weight gained of the fry groups fed with unenriched <i>Artemia</i> (UA), enriched <i>Artemia</i> (EA), unenriched <i>Moina</i> (UM) and enriched <i>Moina</i> (EM). All data are presented as mean \pm SEM of the body weight gained in fries (Different superscript letter(s) indicate significant different (p<0.05)	62
14	Pairwise comparison in body weight gained (mg) of the fry groups fed with unenriched <i>Artemia</i> (UA), enriched <i>Artemia</i> (EA), unenriched <i>Moina</i> (UM) and enriched <i>Moina</i> (EM)	63
15	Two-way ANOVA results between the livefood (<i>Artemia</i> and <i>Moina</i>), enrichment (unenriched and enriched) and interaction between type of livefood and enrichment on the body weight gained (mg) of the fry groups fed with livefood (unenriched <i>Artemia</i> , unenriched <i>Moina</i> , enriched <i>Artemia</i> and enriched <i>Moina</i>)	64
16	The final total length in fry groups fed with the unenriched <i>Artemia</i> (UA), enriched <i>Artemia</i> (EA), unenriched <i>Moina</i> (UM) and enriched <i>Moina</i> (EM). All data are presented as mean \pm SEM of the final total length in fry. (Different superscript letter(s) indicate significant different (p<0.05)	65
17	Pairwise comparison in final total length (mm) of fry groups fed with unenriched <i>Artemia</i> (UA), enriched <i>Artemia</i> (EA), unenriched <i>Moina</i> (UM) and enriched <i>Moina</i> (EM)	66
18	Two-way ANOVA results between the livefood (<i>Artemia</i> and <i>Moina</i>), enrichment (unenriched and enriched) and interaction between type of livefood and enrichment on the final total length of the fry groups fed with livefood (unenriched <i>Artemia</i> , unenriched <i>Moina</i> , enriched <i>Artemia</i> and enriched <i>Moina</i>)	67
19	The specific growth rate (% d ⁻¹) of the fry groups fed with unenriched <i>Artemia</i> (UA), enriched <i>Artemia</i> (EA), unenriched <i>Moina</i> (UM) and enriched <i>Moina</i> (EM). All data are presented as mean \pm SEM of the specific growth rate in fries (Different superscript letter(s) indicate significant different (p<0.05)	68
20	Pairwise comparison in specific growth rate (%) of the fry groups fed with unenriched <i>Artemia</i> (UA), enriched <i>Artemia</i> (EA), unenriched <i>Moina</i> (UM) and enriched <i>Moina</i> (EM)	69
21	Two-way ANOVA results between the livefood (<i>Artemia</i> and <i>Moina</i>), enrichment (unenriched and enriched) and interaction between type of livefood and enrichment on the specific growth	69

xiv

rate (% d⁻¹) of the fry groups fed with livefood (unenriched *Artemia*, unenriched *Moina*, enriched *Artemia* and enriched *Moina*)

The relative growth rate (%) of the fry groups fed with unenriched Artemia (UA), enriched Artemia (EA), unenriched Moina (UM) and enriched Moina (EM). All data are presented as mean \pm SEM of the relative growth rate in fries (Different superscript letter(s) indicate significant different (p<0.05)

22

23

25

Pairwise comparison in relative growth rate (%) (%) of the fry groups fed with unenriched *Artemia* (UA), enriched *Artemia* (EA), unenriched *Moina* (UM) and enriched *Moina* (EM)

Two-way ANOVA results between the livefood (*Artemia* and *Moina*), enrichment (unenriched and enriched) and interaction between type of livefood and enrichment on the relative growth

24 between type of livefood and enrichment on the relative growth rate (%) of the fry groups fed with livefood (unenriched *Artemia*, unenriched *Moina*, enriched *Artemia* and enriched *Moina*)

> Survival (%) of the fry groups fed with unenriched *Artemia* (UA), enriched *Artemia* (EA), unenriched *Moina* (UM) and enriched *Moina* (EM). All data are presented as mean \pm SEM of the survival (Different superscript letter(s) indicate significant different (p<0.05)

73

70

71

LIST OF FIGURES

Figure		Page	
1	Wholesale and retail values of freshwater aquaculture in Ringgit Malaysia (RM) (DOF, 1991, 2001, 2011, 2021)	4	
2	Wholesale prices of <i>Tor tambroides</i> at major markets from 2015-2020 (RM/kg) (DOF, 2015-2020)	6	
3	Adult <i>Tor tambroides</i> (Empurau) reared in the facilities at Borneo Empurau Farm Sdn. Bhd., Sarawak (Photo Credit: Wizilla Joshua, BEF – 25th May 2018)*	7	
4	<i>Chlorella vulgaris</i> observed under light microscope with the typical cell diameter and size of 2 to 10 μm by Ramaraj et al. (2016). Cultivation of Green Microalga, <i>Chlorella vulgaris</i> for Biogas Purification	11	
5	The culture of <i>Chlorella</i> sp. in Bold's Basal Media. Each bottle contained 5 L culture and it was kept in a room temperature, $25\pm3^{\circ}$ C, 24:0 day/light and continuous aeration	19	
6	The amplified DNA fragments of <i>Chlorella</i> sp. (Lane A – 1kB DNA Ladder Promega, Lane B-D – Sample 1-3)	23	
7	Phylogenetic tree of <i>Chlorella</i> sp. UPM. The bootstrap analysis was performed using the neighbor-joining method and the values are displayed at the fork	24	
8	Comparisons of DNA sequence of <i>Chlorella</i> sp. UPM with the DNA sequences of <i>Chlorella</i> sp. and <i>Chlorella vulgaris</i> obtained on GenBank	24	
9	The growth curve of <i>Chlorella vulgaris</i> until the late logarithmic phase. The graph shows the highest cells density was achieved during Day 11 with cell density of 1.64×10^7 cells/mL	25	
10	The distribution of saturated fatty acid and polyunsaturated fatty acid in <i>Chlorella vulgaris</i> sample	27	
11	The O.S.I. Brand Pro 80 [™] Artemia cysts used in this study	34	
12	Incubation of Artemia cyst which took 15 to 20 hrs to hatch	34	

13	The <i>Moina</i> sp. was starved for 24 hrs before the enrichment	35
14	The images of <i>Artemia</i> sp. observed under stereo microscope (Leica Application Suite EZ) at 0 hr (Instar I), 2 hr, 4 hr and 6 hr of enrichment period. At 0 hr, 2 hr and 4 hr of enrichment, there were no indication of <i>C. vulgaris</i> in the gut of <i>Artemia</i> sp At 6 hr of enrichment, the presence of <i>C. vulgaris</i> in the gut of <i>Artemia</i> sp. was clearly observed as indicated in the figure above	40
15	The images of <i>Moina</i> sp. at 0 hr, 8 hr, 16 hr and 24 hr of enrichment with <i>C. vulgaris</i> . The presence of <i>C. vulgaris</i> was started to be seen in the guts of <i>Moina</i> sp. at 8 hr onwards as indicated in the images. At 16 hr, the <i>C. vulgaris</i> was more visible in the guts and at 24 hr, the gut was clearly loaded	41
16	The fertilized eggs were incubated until hatched at Wet Laboratory at the Faculty of Agriculture, UPM	54
17	Newly hatched larvae before the commencement of the first feeding. During this period the larvae depended on its yolk reserve. Any dead larvae were removed to maintain the water quality and avoid any contamination	55
18	The total length measurement of the fry was taken from the head until the end of tail part	57
19	Comparisons of the body weight gained of the fries group fed with unenriched <i>Artemia</i> , enriched <i>Artemia</i> , unenriched <i>Moina</i> and enriched <i>Moina</i> . The data is presented as mean \pm SEM of body weight gained (mg) of fries	63
20	Comparisons of the final total length of the fry groups fed with unenriched <i>Artemia</i> , enriched <i>Artemia</i> , unenriched <i>Moina</i> and enriched <i>Moina</i> . The data is presented as mean \pm SEM of total length (mm) of fries	65
	Comparisons of the specific growth rate of the fry groups fed with unenriched <i>Artemia</i> , enriched <i>Artemia</i> , unenriched <i>Moina</i> and enriched <i>Moina</i> . The data is presented as mean \pm SEM of specific growth rate	68

Comparisons of the relative growth rate of the fry groups fed with unenriched *Artemia*, enriched *Artemia*, unenriched *Moina* and enriched *Moina*. The data is presented as mean \pm SEM of relative growth rate of fries

22

23

24

25

26

Comparison of GH mRNA relative expression of the fry groups fed with unenriched *Artemia*, enriched *Artemia*, unenriched *Moina* and enriched *Moina*. The data is presented as mean \pm SEM of the GH mRNA relative expression of the fries

Comparison of MSTN mRNA relative expression of the fry groups fed with unenriched *Artemia*, enriched *Artemia*, unenriched *Moina* and enriched *Moina*. The data is presented as mean \pm SEM of the MSTN mRNA relative expression of the fries

Comparison of CC3 mRNA relative of the fry groups fed with unenriched *Artemia*, enriched *Artemia*, unenriched *Moina* and enriched *Moina*. The data is presented as mean \pm SEM of the CC3 mRNA relative of the fries

Comparison of MHCC1a mRNA relative expression of the fry groups fed with unenriched *Artemia*, enriched *Artemia*, unenriched *Moina* and enriched *Moina*. The data is presented as mean \pm SEM of the MHCC1a mRNA relative of the fries

xviii

77

71

74

75

LIST OF ABBREVIATIONS

ALA	α-linolenic acid		
BLAST	Basic Local Alignment Search Tool		
bp	Base pairs		
CC3	Complementary C3		
DAH	Days after hatching		
°C	Degree Celsius		
DOF	Department of Fisheries		
DNA	Deoxyribonucleic acid		
DHA	Docosahexaenoic acid		
EPA	Eicosapentaenoic acid		
FAO	Food and Agriculture Organization		
g	Grams		
GH	Growth hormone		
hr	Hour		
kg	Kilogram		
LNA	Linoleic acid		
L	Liter		
LC- PUFA	Long-chain polyunsaturated fatty acid		
mRNA	Messenger ribonucleic acid		
MHCC1a	Major Histocompatibility Complex Class 1a		
μL	Microliter		
μm	Micrometer		

C)

mg	Milligram
----	-----------

- mL Milliliter
- mm Millimeter
- mm² Millimeter square
- MUFA Monounsaturated fatty acid
- MSTN Myostatin
- nm Nanometer
- % Percentage
- PCR Polymerase chain reaction
- PUFA Polyunsaturated fatty acid
- rpm Revolutions per minute
- RNA Ribonucleic acid
- RM Ringgit Malaysia

G

SFA Saturated fatty acid

CHAPTER 1

INTRODUCTION

1.1 Background of Study

The freshwater aquaculture is defined as the rearing and breeding of aquatic organisms in captivity which include fish, shrimp, shellfish and aquatic plants for commercial aims (Li et al., 2018). Typically, species selected for aquaculture are those with high and fast growth and tolerant to extensive environmental conditions (Julian et al., 2021). Based on the data by FAO (2020), global aquaculture production in 2018 was 48% and out of that, freshwater aquaculture had contributed to 60% of the total production (Keller and Lodge, 2021). Due to such rapid expansion of freshwater aquaculture, numerous of developments in fish nutrition and feed competency have been highlighted with the direction towards sustainability. This is also related to the feeding regimes and suitable feed during the early stage of larviculture, which in turn will support the optimal growth and higher survival of the larvae (Rasdi et al. 2019; Lee, 2003).

In Malaysia, *Tor tambroides* is one of the fish species with aquaculture potential. It is a riverine cyprinid with high value in Southeast Asia (Lau et al., 2021). It is treasured and highly regard as an important ornamental and sport fish with stable and high market value (Azfar-Ismail et al., 2020). According to Department of Fisheries (DOF), the wholesale value of this species could reach up to RM 233.37 per kilogram and had the production of 105,904.01 tonnes in 2021. Thus, making it a great potential in aquaculture industry (Ingram et al., 2005). Various efforts have been taken to further understand this species in different aspects, including its breeding performance (Ingram et al., 2007; Ingram et al., 2005), molecular systematics (Esa et al., 2008), morphometric analysis (Pollar et al., 2007), ontogenic development (Ramezani-Fard et al., 2011) and its environmental water parameters in the farms (Soon et al., 2014).

1.2 Problem statement

Unlike other fast-growing species in aquaculture, *T. tambroides* is a slow grower fish as compared to other cyprinids (Asaduzzaman et al., 2018). It may take a year to achieve 500 g to 600 g body weight in its natural environment (Ng, 2004). Several researches had emerged in order to enhance the knowledge on the nutrition requirements of this fish species at various life stages. Among the studies focusing on its dietary requirements during the fingerling stage were the effects of the dietary protein requirement and the lack of protein-sparing action by dietary lipid on its growth performance based on semi-purified diets and different levels of proteins and lipids (Ng et al., 2008), effects of commercial poultry offal meal replacing fishmeal on its growth, body composition and feed intake efficiency (Ishak et al., 2021). Meanwhile, the nutritional studies during the juvenile stage were highlighting on the effects of different dietary lipid percentage over time of feeding on its muscle fatty acid compositions

(Ramezani-Fard et al., 2012) and the effects of crude illipe (*Shorea macrophylla*) oil on its growth performance, body composition and fatty acid profile (Kamarudin et al., 2018). The earliest study on the larvae stage of *T. tambroides* were done by Asaduzzaman et al. (2016) through the feeding with live and formulated feeds (65% fishmeal, 2% squid meal and 2% shrimp meal) and their impacts on the biochemical composition and growth performance of the larvae, whereby the formulated feed had demonstrated better results than live foods. Although the formulated feed was tested in T. *tambroides* larvae (Asaduzzaman et al., 2016), it might not be cost effective and sustainable for early larval feeding as it includes high inclusion of fishmeal (65%).

1.3 Justification of study

The development of larviculture and hatchery are deemed as the critical parts in aquaculture. It is known as a bottleneck in seed production for aquaculture (Herath and Atapaththu, 2013). In regard to hatchery management and operation is the use of live food as the diets of fry. Generally, live foods are more desirable as compared to artificial feeds during the early fish and crustacean larval stage (Das et al., 2007). The dependency on live food in the hatcheries are expected to be continuously applied in the future (Dhont et al., 2013). The digestive enzymes are found in abundance in live foods and it stimulates larval appetite (Zeng et al., 2018). Among the commonly used live food sources in the hatcheries for the culture of both marine and freshwater larvae and fry are *Artemia* and *Moina*. These live foods are known as the filter feeders. They have the ability to filter any compound available in the water column (Sorgeloos et al., 2001; Manklinniam et al., 2018). Such characteristics in *Artemia* and *Moina* are utilized to deliver certain nutritional contents through enrichment technique to target species.

Microalgae possess high nutritional values such as long chain polyunsaturated fatty acid (LC-PUFA) which constitute of phospholipids (Li et al, 2018; Li and Olsen, 2015). The green microalgae, *Chlorella vulgaris* is a good candidate for enrichment of live food. It is known as the super food with various nutrients such as fats, protein, vitamins, bioactive compounds and carbohydrates (Andrade et al., 2018). It contains immunostimulant properties which is beneficial in enhancing the lifespan of fish (Gouveia et al., 2002) and boost the growth of several fish species including Olive flounder (*Paralichthys olivaceus*) (Rahimnejad et al., 2017), koi carp (*Cyprinus carpio*) (Khani et al., 2017) and African catfish (*Clarias gariepinus*).

Besides, the administration of enriched live food, particularly *Artemia* sp. and *Moina* sp. enriched with *C. vulgaris* is yet to be discovered in *T. tambroides* fry focusing on the growth performance, survival and expression of growth and immune response genes. The long culture period of *T. tambroides* in captive condition is contributed by the fact that it is a slow-growing fish (Chowdhury et al., 2016; Soon et al., 2014). This drawback could impact the seeds availability and the period of this species to reach the marketable size which indirectly hinder its potential in aquaculture industry. The growth and immune systems are related to the interaction and metabolism of nutrients (Martin and Król, 2017; Moriyama et al., 2001; Peter and Marchant, 1995). Studies supported that *Artemia* and *Moina* enriched with various enrichment media had resulted in positive effects in terms of growth performance and survival as compared to the

unenriched counterparts. For instance, *Artemia* enriched with *Chlorella* sp. had resulted in higher survival of juvenile long snout seahorse, *Hippocampus guttulatus* (Palma et al., 2011), whereas *Chlorella* sp. enriched *Moina* had contributed to the high specific growth rate and survival in Siamese fighting fish (*Betta splendens*) juvenile (Rasdi et al., 2020b). These promising effects could be based on species specific as different larval species may be affected differently to the enrichment diets and it does not always contribute to positive effects. For example, *Artemia* enriched with vitamin A did not give any significant effects to the growth performance and survival of strip trumpeter (*Latris lineata*) (Negm et al., 2014). The *C. vulgaris* enriched *Artemia* and *Moina* could be potential candidates for *T. tambroides* fry during weaning period before artificial and formulated feed are being offered as the sole diet and this could possibly enhance its growth performance and survival during early stage of lifecycle.

1.4 Objectives

Hence, the objectives of this study were:

- 1. To screen and analyze of the nutritional profile of *Chlorella* sp. as potential candidate for live food enrichment
- 2. To evaluate and compare the composition of *Artemia* sp. and *Moina* sp. enriched and without enrichment of *C. vulgaris*
- 3. To assess on the effects of *C. vulgaris* enriched *Artemia* sp. and *Moina* sp. in comparison to unenriched counterparts on the growth performance, survival and expression of growth and immune genes of *Tor tambroides* fry.

Hypothesis:

- H0: The C. *vulgaris* enriched *Artemia* sp. and *Moina* sp. does not affect the growth performance, survival and expression of immune and growth genes of *Tor tambroides* fry
- H1: The C. *vulgaris* enriched *Artemia* sp. and *Moina* sp. does affect the growth performance, survival and expression of immune and growth genes of *Tor tambroides* fry

REFERENCES

- Abdolahnejad, Z., Pourkazemi, M., Khoshkholgh, M. R., & Yarmohammadi, M. (2015). Expression of growth hormone gene during early development of Siberian sturgeon (*Acipenser baerii*). *Molecular Biology Research Communications*, 4(4), 181.
- Abdollahi, R., Heidari, B., & Aghamaali, M. (2016). Evaluation of lysozyme, complement C3, and total protein in different developmental stages of Caspian kutum (*Rutilus frisii kutum* K.). Fisheries & Aquatic Life, 24(1), 15-22.
- Abduh, M. Y., Koh, I. C. C., Abol-Munafi, A. B., Norazmi-Lokman, N. H., & Mat Noordin, N. (2021). Effects of dietary fish oil and corn oil on gonadosomatic and hepatosomatic index, gonadal histology, 17β-oestradiol level and fatty acids profile of mahseer (*Tor tambroides*) broodstock in captivity. *Aquaculture Nutrition*, 27(5), 1448-1459.
- Abedi, E., & Sahari, M. A. (2014). Long-chain polyunsaturated fatty acid sources and evaluation of their nutritional and functional properties. *Food science & Nutrition*, 2(5), 443-463.
- Abou-Shanab, R. A., Matter, I. A., Kim, S. N., Oh, Y. K., Choi, J., & Jeon, B. H. (2011). Characterization and identification of lipid-producing microalgae species isolated from a freshwater lake. *Biomass and Bioenergy*, 35(7), 3079-3085.
- Abu-Serie, M. M., Habashy, N. H., & Attia, W. E. (2018). In vitro evaluation of the synergistic antioxidant and anti-inflammatory activities of the combined extracts from Malaysian *Ganoderma lucidum* and Egyptian *Chlorella vulgaris*. BMC Complementary and Alternative Medicine, 18(1), 1-13.
- Agh, N., & Sorgeloos, P. (2005). Handbook of protocols and guidelines for culture and enrichment of live food for use in larviculture. Urmia-Iran: Ediciones Artemia & Aquatic Animals Research Center, 60.
- Ahlgren, G., Gustafsson, I. B. & Boberg, M. (1992). Fatty acid content and chemical composition of freshwater microalgae. *Journal of Phycology*, 28, 37-50.
- Ahmad, M. T., Shariff, M., Md. Yusoff, F., Goh, Y. M., & Banerjee, S. (2020). Applications of microalga *Chlorella vulgaris* in aquaculture. *Reviews in Aquaculture*, 12(1), 328-346.
- Akbary, P., Hosseini, S. A., & Imanpoor, M. R. (2011). Enrichment of Artemia nauplii with essential fatty acids and vitamin C: effect on rainbow trout (Oncorhynchus mykiss) larvae performance. Iranian Journal of Fisheries Sciences, 10, 557-569.

- Aluma, M. O. (2020). Degradation of *Highly Unsaturated Fatty Acids in Artemia sp.* (Doctoral dissertation, Ghent University).
- Ammar, S. H. (2016). Cultivation of Microalgae Chlorella vulgaris in Airlift photobioreactor for Biomass Production using commercial NPK Nutrients. Al-Khwarizmi Engineering Journal, 12(1), 90-99.
- An, B. K., Kim, K. E., Jeon, J. Y., & Lee, K. W. (2016). Effect of dried *Chlorella vulgaris* and Chlorella growth factor on growth performance, meat qualities and humoral immune responses in broiler chickens. *SpringerPlus*, 5, 718.
- Andersen, R. A. (Ed.). (2005). Algal culturing techniques. Elsevier.
- Andrade, L. M., Andrade, C. J., Dias, M., Nascimento, C., & Mendes, M. A. (2018). Chlorella and Spirulina microalgae as sources of functional foods. Nutraceuticals, and Food Supplements, 6(1), 45-58.
- Anthony, J., Sivashankarasubbiah, K. T., Thonthula, S., Rangamaran, V. R., Gopal, D., & Ramalingam, K. (2018). An efficient method for the sequential production of lipid and carotenoids from the Chlorella Growth Factor-extracted biomass of *Chlorella vulgaris. Journal of Applied Phycology*, 30, 2325-2335.
- AOAC (1995). Official methods of analysis 16th Ed. Association of official analytical chemists. Washington DC, USA. Sci. Educ.
- AOAC (1999). Official methods of analysis. Washington DC, USA. Sci. Educ.
- Araújo, F. G., & Rosa, P. V. (2016). Docosahexaenoic acid (C22: 6n-3) alters cortisol response after air exposure in *Prochilodus lineatus* (Valenciennes) larvae fed on enriched Artemia. Aquaculture Nutrition, 23(6), 1216-1224.
- Asaduzzaman, M., Kader, M. A., Bulbul, M., Abol-Munafi, A. B., Abd Ghaffer, M., & Verdegem, M. (2016). Biochemical composition and growth performances of Malaysian Mahseer *Tor tambroides* larvae fed with live and formulated feeds in indoor nursery rearing system. *Aquaculture Reports*, 4, 156-163.
- Asaduzzaman, M., Sofia, E., Shakil, A., Haque, N. F., Khan, M. N. A., Ikeda, D., ... & Abol-Munafi, A. B. (2018). Host gut-derived probiotic bacteria promote hypertrophic muscle progression and upregulate growth-related gene expression of slow-growing Malaysian Mahseer *Tor tambroides. Aquaculture Reports*, 9, 37-45.
- Ashfaq, H., Soliman, H., Saleh, M., & El-Matbouli, M. (2019). CD4: a vital player in the teleost fish immune system. *Veterinary Research*, 50, 1-11.
- Aya, F. A., Nillasca, V. S. N., & Garcia, L. M. B. (2021). Improved survival and growth of silver therapon *Leiopotherapon plumbeus* early juveniles through co-feeding

with *Artemia* and commercial feeds. *Journal of Applied Ichthyology*, *37*(6), 925-931.

- Azfar-Ismail, M., Kamarudin, M. S., Syukri, F., & Latif, K. (2020). Larval development of a new hybrid Malaysian mahseer (*Barbonymus gonionotus*♀× *Tor tambroides*♂). Aquaculture Reports, 18, 100416.
- Azuadi, N. M., Siraj, S. S., Daud, S. K., Christianus, A., Harmin, S. A., Sungan, S., & Britin, R. (2011). Enhancing ovulation of Malaysian mahseer (*Tor tambroides*) in captivity by removal of dopaminergic inhibition. *Journal of Fisheries and Aquatic Science*, 6(7), 740.
- Bai, S. C., Katya, K., & Yun, H. (2015). Additives in aquafeed: An overview. Feed and Feeding Practices in Aquaculture, 171-202.
- Bami, M. L., Kamarudin, M. S., Saad, C. R., Arshad, A., & Ebrahimi, M. (2017). Effects of palm oil products on growth performance, body composition and fatty acid profile of juvenile Malaysian mahseer (*Tor tambroides*). *Journal of Oil Palm Research*, 29(3), 387-400.
- Becker, B., & Marin, B. (2009). Streptophyte algae and the origin of embryophytes. *Annals of Botany*, 103(7), 999-1004.
- Beijerinck, M. W. (1890). Culture experiments with zoochlorella, Lichenengonidia and other lower algae. *Botanisches Centralblatt*, 47, 725-739, 741-754, 757-768, 781-785.
- Bengtson, D. A. (2003). Status of marine aquaculture in relation to live prey: past, present and future. *Live feeds in Marine Aquaculture*, 1-16.
- Bertoldi, F. C., Sant'Anna, E., da Costa Braga, M. V., & Oliveira, J. L. B. (2006). Lipids, fatty acids composition and carotenoids of *Chlorella vulgaris* cultivated in hydroponic wastewater. *Grasas y Aceites*, 57(3), 270-274.
- Bertucci, J. I., Blanco, A. M., Sundarrajan, L., Rajeswari, J. J., Velasco, C., & Unniappan, S. (2019). Nutrient regulation of endocrine factors influencing feeding and growth in fish. *Frontiers in Endocrinology*, 10, 83.
- Bleeker, P. (1854). Overview of the ichthyological fauna of Sumatra, with a description of some new species naturally. *Timesheet Dutch East Indies*, 7, 49-108.
- Bleeker, P. (1863). Atlas ichthyologique des Indes orientales néêrlandaises: publié sous les auspices du gouvernement colonial néêrlandais (Vol. 3). F. Muller.
- Boackle, S. A. (2003). Complement and autoimmunity. *Biomedicine & Pharmacotherapy*, 57(7), 269-273.

- Brauner, C. J., & Richards, J. G. (2020). Physiological performance in aquaculture: Using physiology to help define optimal conditions for growth and environmental tolerance. *Fish Physiology*, *38*, 83-121.
- Cahu, C., & Infante, J. Z. (2001). Substitution of live food by formulated diets in marine fish larvae. *Aquaculture*, 200(1-2), 161-180.
- Campbell, M. N. (2008). Biodiesel: Algae as a renewable source for liquid fuel. *Guelph Engineering Journal*, *1*(1), 2-7.
- Cao, Z., He, M., Chen, X., Wang, S., Cai, Y., Xie, Z., ... & Zhou, Y. (2017). Identification, polymorphism and expression of MHC class Ia in golden pompano, *Trachinotus ovatus*. Fish & shellfish immunology, 67, 55-65.
- Carter, C. G. (2015). Feeding in hatcheries. In D. Allen-Davis (Ed.), Feed and Feeding Practices in Aquaculture (pp. 317-348). Woodhead Publishing.
- Catalán, V., Frühbeck, G., & Gómez-Ambrosi, J. (2018). *Chapter 8—Inflammatory and* oxidative stress markers in skeletal muscle of obese subjects. Obesity; Del Moral, AM; Aguilera García.
- Cha, S. T., Chen, J. W., Goh, E. G., Aziz, A., & Loh, S. H. (2011). Differential regulation of fatty acid biosynthesis in two *Chlorella* species in response to nitrate treatments and the potential of binary blending microalgae oils for biodiesel application. *Bioresource Technology*, 102(22), 10633-10640.
- Chacón-Lee, T. L., & González-Mariño, G. E. (2010). Microalgae for "healthy" foods possibilities and challenges. *Comprehensive Reviews in Food Science and Food Safety*, 9(6), 655-675.
- Chang, W. S. (2000). Features of Indigenous Fish Species Having Potentials for Aquaculture (pp. 2-7). Inland Fisheries Division of Department of Agriculture, Sarawak.
- Chen, W., Luo, L., Han, D., Long, F., Chi, Q., & Hu, Q. (2021). Effect of dietary supplementation with *Chlorella sorokiniana* meal on the growth performance, antioxidant status, and immune response of rainbow trout (*Oncorhynchus mykiss*). *Journal of Applied Phycology*, *33*, 3113-3122.
- Chin, P. K. (1990). The freshwater fishes of North Borneo. Supplementary chapter. *The Freshwater Fishes of North Borneo.*
- Chowdhury, A. J. K., Zakaria, N. H., Abidin, Z. A. Z., & Rahman, M. M. (2016). Phototrophic purple bacteria as feed supplement on the growth, feed utilization and body compositions of Malaysian Mahseer, *Tor tambroides* juveniles. *Sains Malaysiana*, 45(1), 135-140.

- Clarke, E. V., Weist, B. M., Walsh, C. M., & Tenner, A. J. (2015). Complement protein C1q bound to apoptotic cells suppresses human macrophage and dendritic cellmediated Th17 and Th1 T cell subset proliferation. *Journal of Leucocyte Biology*, 97(1), 147-160.
- Clay, T. A., Suchy, M. D., Ferrara, A. M., Fontenot, Q. C., & Lorio, W. (2011). Early growth and survival of larval Alligator Gar, *Atractosteus spatula*, reared on artificial floating feed with or without a live *Artemia* spp. supplement. *Journal of the World Aquaculture Society*, 42(3), 412-416.
- Conceição, L. E., Yúfera, M., Makridis, P., Morais, S., & Dinis, M. T. (2010). Live feeds for early stages of fish rearing. *Aquaculture Research*, *41*(5), 613-640.
- Converti, A., Casazza, A. A., Ortiz, E. Y., Perego, P., & Del Borghi, M. (2009). Effect of temperature and nitrogen concentration on the growth and lipid content of *Nannochloropsis oculata* and *Chlorella vulgaris* for biodiesel production. *Chemical Engineering and Processing: Process Intensification*, 48(6), 1146-1151.
- Dang, Y., & Liu, C. (2018). Real-time PCR array to study the effects of chemicals on the growth hormone/insulin-like growth factors (GH/IGFs) axis of zebrafish embryos/larvae. *Chemosphere*, 207, 365-376.
- Das, P., Mandal, S. C., Bhagabati, S. K., Akhtar, M. S., & Singh, S. K. (2012). Important live food organisms and their role in aquaculture. *Frontiers in Aquaculture*, 5(4), 69-86.
- Das, S. K., Tiwari, V. K., Venkateshwarlu, G., Reddy, A. K., Parhi, J., Sharma, P., & Chettri, J. K. (2007). Growth, survival and fatty acid composition of *Macrobrachium rosenbergii* (de Man, 1879) post larvae fed HUFA-enriched *Moina micrura. Aquaculture*, 269(1-4), 464-475.
- Davis, D. A., Ii, T. J. D., & Head, M. E. (2018). Culture of Small Zooplankton for the Feeding of Larval Fish. SRAC Publication, 701, 1-6.
- Day, F. (1876). On some of the fishes of the Deccan. *Zoological Journal of the Linnean Society*, *12*(64), 565-578.
- Defoirdt, T., Verstraete, W., & Bossier, P. (2008). Luminescence, virulence and quorum sensing signal production by pathogenic Vibrio campbellii and Vibrio harveyi isolates. Journal of Applied Microbiology, 104(5), 1480-1487.
- Deng, B., Zhang, F., Wen, J., Ye, S., Wang, L., Yang, Y., ... & Jiang, S. (2017). The function of myostatin in the regulation of fat mass in mammals. *Nutrition & Metabolism*, 14(1), 1-6.

- Department of Fisheries (DOF). (1991, 2001, 2011-2015, 2020-2021). Annual Fisheries Statistics. Fisheries Data Collection Branch. Ministry of Agriculture and Agro-Based Industry Malaysia.
- Devi, G., Balasundaram, C., & Harikrishnan, R. (2020). Effect of madecassic acid on innate-adaptive immune response and cytokine gene expression in *Labeo rohita* against *Argulus siamensis*. *Recent Trends Biotechnology*, 8, 1-11.
- Devlin, R. H., Leggatt, R. A., & Benfey, T. J. (2020). Genetic modification of growth in fish species used in aquaculture: phenotypic and physiological responses. *Fish Physiology*, 38, 237-272.
- Dey, T., Ghosh, P. K., Nandi, S. K., Chowdhury, G., Mian, S., & Uddin, M. S. (2022). A Review on n-3 HUFA and Live Food Organism for Marine Fish Larvae Nutrition. American Journal of Agricultural Science, Engineering, and Technology, 6(3), 88-102.
- Dhont, J., Dierckens, K., Støttrup, J., Van Stappen, G., Wille, M., & Sorgeloos, P. (2013). Rotifers, Artemia and copepods as live feeds for fish larvae in aquaculture. Advances in Aquaculture Hatchery Technology (pp. 157-202). Woodhead Publishing.
- Dinesh, K., & Nandeesha, M. C. (2007). Status of Mahseers, the King of Freshwater systems in India-A review. Proceeding of the International Symposium on the Mahseer Kuala Lumpur: Malaysian Fish. Soc (pp. 3-35).
- Dineshbabu, G., Goswami, G., Kumar, R., Sinha, A., & Das, D. (2019). Microalgae– nutritious, sustainable aqua-and animal feed source. *Journal of Functional Foods*, 62, 103545.
- Dorigo, U., Bérard, A., & Humbert, J. F. (2002). Comparison of eukaryotic phytobenthic community composition in a polluted river by partial 18S rRNA gene cloning and sequencing. *Microbial Ecology*, 44, 372-380.
- Eddy, S. P. T. (1997, November). Angling for Kelah. Rod and Line Magazine, 68-74.
- Esa, Y. B., Siraj, S. S., Daud, S. K., Ryan, J. J. R., Rahim, K. A. A., & Tan, S. G. (2008). Molecular systematics of mahseers (Cyprinidae) in Malaysia inferred from sequencing of a mitochondrial Cytochrome C Oxidase I (COI) gene. *Pertanika Journal of Tropical Agricultural Science*, 31(2), 263-269.
- Eschmeyer, W. N., Fricke, R., & Van der Laan, R. (2015). *Catalog of fishes: Genera, Species, References*. California Academy of Sciences, San Francisco.
- Estévez A., McEvoy L.A., Bell J.G. & Sargent, J.R (1998). Effects of temperature and starvation time on the pattern and rate of loss of essential fatty acids in *Artemia*

nauplii previously enriched using arachidonic acid and eicosapentaenoic acid-rich emulsions. *Aquaculture*, 165, 295-311.

- FAO. (2020). *The State of World Fisheries and Aquaculture 2020*. Sustainability in Action. Rome: Food and Agriculture Organization of the United Nations.
- Ferraresso, S., Bonaldo, A., Parma, L., Buonocore, F., Scapigliati, G., Gatta, P. P., & Bargelloni, L. (2016). Ontogenetic onset of immune-relevant genes in the common sole (*Solea solea*). Fish & Shellfish Immunology, 57, 278-292.
- Ferreira, G. F., Pinto, L. R., Maciel Filho, R., & Fregolente, L. V. (2019). A review on lipid production from microalgae: Association between cultivation using waste streams and fatty acid profiles. *Renewable and Sustainable Energy Reviews*, 109, 448-466.
- Ferrer-Álvarez, Y. I., Ortega-Clemente, L. A., Pérez-Legaspi, I. A., Hernández-Vergara, M. P., Robledo-Narváez, P. N., Ríos-Leal, E., & Poggi-Varaldo, H. M. (2015). Growth of *Chlorella vulgaris* and *Nannochloris oculata* in effluents of Tilapia farming for the production of fatty acids with potential in biofuels. *African Journal of Biotechnology*, *14*(20), 1710-1717.
- Firdaus-Nawi, M., & Zamri-Saad, M. (2016). Major components of fish immunity: a review. Pertanika Journal of Tropical Agricultural Science, 39(4).
- Fischer, U., Dijkstra, J. M., Köllner, B., Kiryu, I., Koppang, E. O., Hordvik, I., ... & Ototake, M. (2005). The ontogeny of MHC class I expression in rainbow trout (Oncorhynchus mykiss). Fish & Shellfish Immunology, 18(1), 49-60.
- Folch, J., Lees, M., & Sloane Stanley, G. H. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*, 226(1), 497-509.
- Franz, G. P., Tönißen, K., Rebl, A., Lutze, P., & Grunow, B. (2022). The expression of myogenic gene markers during the embryo-larval-transition in Pikeperch (*Sander lucioperca*). Aquaculture Research, 53(13), 4767-4781.
- Froese, R., & Pauly, D. (2015). FishBase. World Wide Web electronic publication version (11/2014). <u>http://www.fishbase.org</u>.
- Fuentes, E. N., Pino, K., Navarro, C., Delgado, I., Valdés, J. A., & Molina, A. (2013). Transient inactivation of myostatin induces muscle hypertrophy and overcompensatory growth in zebrafish via inactivation of the SMAD signalling pathway. *Journal of Biotechnology*, 168(4), 295-302.
- Furuita, H., Takeuchi, T., Toyota, M., & Watanabe, T. (1996). EPA and DHA requirements in early juvenile red sea bream using HUFA enriched Artemia nauplii. Fisheries Science, 62(2), 246-251.

- Gama-Flores, J. L., Huidobro-Salas, M. E., Sarma, S. S. S., Nandini, S., Zepeda-Mejia, R., & Gulati, R. D. (2015). Temperature and age affect the life history characteristics and fatty acid profiles of *Moina macrocopa* (Cladocera). *Journal* of Thermal Biology, 53, 135-142.
- Ghosh, S., & Love, N. G. (2011). Application of rbcL based molecular diversity analysis to algae in wastewater treatment plants. *Bioresource Technology*, 102(3), 3619-3622.
- Gilannejad, N., de Las Heras, V., Martos-Sitcha, J. A., Moyano, F. J., Yúfera, M., & Martínez-Rodríguez, G. (2020). Ontogeny of Expression and Activity of Digestive Enzymes and Establishment of gh/igf1 Axis in the Omnivorous Fish *Chelon labrosus. Animals*, 10(5), 874.
- Glencross, B. D. (2009). Exploring the nutritional demand for essential fatty acids by aquaculture species. *Reviews in Aquaculture*, 1(2), 71-124.
- Gogoi, B., Safi, V., & Das, D. N. (2016). The Cladoceran as live feed in fish culture: A brief review. *Research Journal of Animal, Veterinary and Fishery Sciences*, 4(3), 7-12.
- Gouveia, L., Choubert, G., Gomes, E., Pereira, N., Santinha, J., & Empis, J. (2002). Colouringation of gilthead seabream, *Sparus aurata* (Lin 1875), using *Chlorella* vulgaris microalga. Aquaculture Research, 33(12), 987-993.

Grimholt, U. (2016). MHC and Evolution in Teleosts. Biology, 5(1), 6.

- Grimholt, U., Drabløs, F., Jørgensen, S., Høyheim, B., & Stet, R. J. (2002). The major histocompatibility class I locus in Atlantic salmon (*Salmo salar* L.): Polymorphism, linkage analysis and protein modelling. *Immunogenetics*, 54, 570-581.
- Guedes, A. C., Sousa-Pinto, I., & Malcata, F. X. (2015). *Application of microalgae protein to aquafeed*. In Handbook of Marine Microalgae (pp. 93-125). Academic Press.
- Guillard, R. R., & Sieracki, M. S. (2005). Counting cells in cultures with the light microscope. *Algal Culturing Techniques*, 239-252.
- Gupta, S. K., Jha, A. K., Pal, A. K., & Venkateshwarlu, G. (2007). Use of natural carotenoids for pigmentation in fishes. *Reviews in Fish Biology and Fisheries*, 17(3), 441-455.
- Habib, M. A. B., Yusoff, F. M., Phang, S. M., & Mohamed, S. (2003). Growth and nutritional values of *Moina micrura* fed on *Chlorella vulgaris* grown in digested palm oil mill effluent. *Asian Fisheries Science*, 16(1-2), 107-120.

- Hanaee, J., Agh, N., Hanaee, M., Delazar, A., & Sarker, S. D. (2005). Studies on the enrichment of *Artemia urmiana* cysts for improving fish food value. *Animal Feed Science and Technology*, 120(1-2), 107-112.
- Haryono, H., & Tjakrawidjaja, A. H. (2006). Morphological study for identification improvement of *Tambra* fish (*Tor* spp.: Cyprinidae) from Indonesia. *Biodiversitas Journal of Biological Diversity*, 7(1), 1-4.
- Hashimoto, K., Nakanishi, T., & Kurosawa, Y. (1990). Isolation of carp genes encoding major histocompatibility complex antigens. *Proceedings of the National Academy of Sciences*, 87(17), 6863-6867.
- He, Z. H., Qin, J. G., Wang, Y., Jiang, H., & Wen, Z. (2001). Biology of *Moina mongolica* (Moinidae, Cladocera) and perspective as live food for marine fish larvae. *Hydrobiologia*, 457, 25-37.
- Herath, S. S., & Atapaththu, K. S. S. (2013). Sudden weaning of angel fish *Pterophyllum* scalare (Lichtenstein) (Pisces; Cichlidae) larvae from brine shrimp (*Artemia* sp.) nauplii to formulated larval feed. *SpringerPlus*, *2*, 1-7.
- Hewitt, E. W. (2003). The MHC class I antigen presentation pathway: Strategies for viral immune evasion. *Immunology*, 110(2), 163-169.
- Holt, G. J. (2011). Larval fish nutrition. John Wiley & Sons.
- Hopkins, K. D. (1992). Reporting fish growth: A review of the basics 1. Journal of the World Aquaculture Society, 23(3), 173-179.
- Idenyi, J. N., Ebenyi, L. N., Ogah, O., Nwali, B. U., & Ogbanshi, M. E. (2016). Effect of different growth media on the cell densities of freshwater microalgae isolates. *IOSR-JPBS*, 11(3), 24-28.
- Ilavarasi, A., Mubarakali, D., Praveenkumar, R., Baldev, E., & Thajuddin, N. (2011). Optimization of various growth media to freshwater microalgae for biomass production. *Biotechnology*, 10(6), 540-545.
- Ingram, B., Sungan, S., Gooley, G., Sim, S. Y., Tinggi, D., & De Silva, S. S. (2005). Induced spawning, larval development and rearing of two indigenous Malaysian mahseer, *Tor tambroides* and *T. douronensis*. *Aquaculture Research*, 36(10), 983-995.
- Ingram, B., Sungan, S., Tinggi, D., Sim, S. Y., & De Silva, S. S. (2007). Breeding performance of Malaysian mahseer, *Tor tambroides* and *T. douronensis* broodfish in captivity. *Aquaculture Research*, 38(8), 809-818.
- Ishak, S. D. B. (2018). *Dietary carbohydrate utilization by the Malaysian mahseer, Tor tambroides (Bleeker, 1854)* (Master's dissertation). Universiti Putra Malaysia.

- Ishak, S. D., Yusof, Y. A., Abol-Munafi, A. B., & Kamarudin, M. S. (2021). Different starch sources in extruded diets for the Malaysian mahseer (*Tor tambroides*) effects on growth, feed utilisation and tissue histology. *Journal of Sustainability Science and Management*, 16, 94-108.
- Islam, M. R., Hassan, M. R., Begum, M., Punom, N. J., Begum, M. K., Sultana, N., & Rahman, M. S. (2017). Effects of feeding zooplankton, *Moina macrocopa* (Straus, 1820) on the growth of Nile tilapia *Oreochromis niloticus* L. *Bangladesh Journal of Scientific and Industrial Research*, 52(2), 81-88.
- Ismail, S., Kam, M. S., & Ramezani-F, E. (2013). Performance of commercial poultry offal meal as fishmeal replacement in the diet of juvenile Malaysian mahseer, *Tor tambroides*. *Asian Journal of Animal and Veterinary Advances*, 8(2), 284-292.
- Izquierdo, M. S., Socorro, J., Arantzamendi, L., & Hernández-Cruz, C. M. (2000). Recent advances in lipid nutrition in fish larvae. *Fish Physiology and Biochemistry*, 22(2), 97.
- Jaafar, F., Na-Nakorn, U., Srisapoome, P., Amornsakun, T., Duong, T. Y., Gonzales-Plasus, M. M., ... & Parhar, I. S. (2021). A current update on the distribution, morphological features, and genetic identity of the Southeast Asian mahseers, *Tor* species. *Biology*, 10(4), 286.
- Jardine, T. D., Galloway, A. W., & Kainz, M. J. (2020). Unlocking the power of fatty acids as dietary tracers and metabolic signals in fishes and aquatic invertebrates. *Philosophical Transactions of the Royal Society B*, 375(1804), 20190639.
- Jo, B. H., Lee, C. S., Song, H. R., Lee, H. G., & Oh, H. M. (2014). Development of novel microsatellite markers for strain-specific identification of *Chlorella* vulgaris. Journal of Microbiology and Biotechnology, 24(9), 1189-1195.
- Jumatli, A., & Ismail, M. S. (2021). Promotion of sustainable aquaculture in Malaysia.
 Proceedings of the International Workshop on the Promotion of Sustainable
 Aquaculture, Aquatic Animal Health, and Resource Enhancement in Southeast
 Asia (pp. 31-40). Aquaculture Department, Southeast Asian Fisheries
 Development Center.
- Kamarudin, M. S., Bami, M. L., Arshad, A., Saad, C. R., & Ebrahimi, M. (2018). Preliminary study of the performance of crude illipe oil (*Shorea macrophylla*) as a dietary lipid source for riverine cyprinid *Tor tambroides*. *Fisheries Science*, 84, 385-397.
- Kamarudin, M. S., Otoi, S., & Saad, C. R. (2011). Changes in growth, survival and digestive enzyme activities of Asian redtail catfish, *Mystus nemurus*, larvae fed on different diets. *African Journal of Biotechnology*, 10(21), 4484-4493.

- Kamaszewski, M., Ostaszewska, T., Prusińska, M., Kolman, R., Chojnacki, M., Zabytyvskij, J., ... & Kasprzak, R. (2014). Effects of *Artemia* sp. enrichment with essential fatty acids on functional and morphological aspects of the digestive system in *Acipenser gueldenstaedtii* larvae. *Turkish Journal of Fisheries and Aquatic Sciences*, 14(4), 929-938.
- Kandathil R., D., Velayudhannair, K., & Schmidt, B. V. (2020). Effects of bioflocculated algae on the growth, digestive enzyme activity and microflora of freshwater fish *Catla catla* (Hamilton 1922). *Aquaculture Research*, 51(11), 4533-4540.
- Kandathil Radhakrishnan, D., AkbarAli, I., Schmidt, B. V., John, E. M., Sivanpillai, S.,
 & Thazhakot Vasunambesan, S. (2019). Improvement of nutritional quality of live feed for aquaculture: An overview. *Aquaculture Research*, *51*(1), 1-17.
- Kandathil Radhakrishnan, D., AkbarAli, I., Schmidt, B. V., John, E. M., Sivanpillai, S.,
 & Thazhakot Vasunambesan, S. (2020). Improvement of nutritional quality of live feed for aquaculture: An overview. *Aquaculture Research*, *51*(1), 1-17.
- Kaneko, G., Furukawa, S., Kurosu, Y., Yamada, T., Takeshima, H., Nishida, M., ... & Watabe, S. (2011). Correlation with larval body size of mRNA levels of growth hormone, growth hormone receptor I and insulin-like growth factor I in larval torafugu *Takifugu rubripes. Journal of Fish Biology*, 79(4), 854-874.
- Kang, H. K., Salim, H. M., Akter, N., Kim, D. W., Kim, J. H., Bang, H. T., ... & Suh, O. S. (2013). Effect of various forms of dietary *Chlorella* supplementation on growth performance, immune characteristics, and intestinal microflora population of broiler chickens. *Journal of Applied Poultry Research*, 22(1), 100-108.
- Kawaroe, M., Prartono, T., Hwangbo, J., Sunuddin, A., Augustine, D., & Gustina, A. S. (2015). Effect of ethyl methane sulfonate (EMS) on cell size, fatty acid content, growth rate, and antioxidant activities of microalgae *Dunaliella* sp. *Aquaculture*, *Aquarium*, *Conservation and Legislation*, 8(6), 924-932.
- Keller, R. P., & Lodge, D. M. (2021). Invasive species. Oxford University Press.
- Khalil, N. A., & Mousa, M. A. (2013). Experimental study on the activation of growth hormone-secreting cells during larval development of Nile tilapia, *Oreochromis niloticus*. *The Egyptian Journal of Aquatic Research*, *39*(1), 67-74.
- Khani, M., Soltani, M., Mehrjan, M. S., Foroudi, F., & Ghaeni, M. (2017). The effect of *Chlorella vulgaris* (Chlorophyta, Volvocales) microalga on some hematological and immune system parameters of Koi carp (*Cyprinus carpio*). *Iranian Journal* of Ichthyology, 4(1), 62-68.

- Khaw, Y. S., Khong, N. M. H., Shaharuddin, N. A., & Yusoff, F. M. (2020). A simple 18S rDNA approach for the identification of cultured eukaryotic microalgae with an emphasis on primers. *Journal of Microbiological Methods*, 172, 105890.
- Kolkovski, S., Czesny, S., Yackey, C., Moreau, R., Cihla, F., Mahan, D., & Dabrowski, K. (2000). The effect of vitamins C and E in (n-3) highly unsaturated fatty acidsenriched *Artemia* nauplii on growth, survival, and stress resistance of fresh water walleye *Stizostedion vitreum* larvae. *Aquaculture Nutrition*, 6(3), 199.
- Kotani, T., Imari, H., Miyashima, A., & Fushimi, H. (2016). Effects of feeding with frozen freshwater cladoceran *Moina macrocopa* on the performance of red sea bream *Pagrus major* larviculture. *Aquaculture International*, 24, 183-197.
- Kottelat, M. (1998). Fishes of the Nam Theum and Xe Bangai basins, Laos, with diagnosis of twenty-two new species (Teleostei: Cyprinidae, Balitoridae, Cobitidae, Coiidae and Odontobutidae). *Ichthyological Exploration of Freshwaters*, 9, 1-128.
- Kottelat, M. (2000). Notes on taxonomy, nomenclature and distribution of some fishes of Laos. *Journal of South Asian Natural History*, 5(1), 83-90.
- Kottelat, M. (2011). Fishes of the Xe Kong drainage in Laos, especially from the Xe Kaman. Co-Management of freshwater biodiversity in the Sekong Basin (pp. 1-29). WWF & Critical Ecosystem Partnership Fund.
- Kottelat, M. (2013). The fishes of the inland waters of Southeast Asia: A catalogue and core bibliography of the fishes known to occur in freshwaters, mangroves and estuaries. *Raffles Bulletin of Zoology*.
- Kottelat, M., Bairdz, I. G., Kullanderg, S. O., Ng, H. H., Parenti, L. R., Rainboth, W. J.,
 & Vidthayanon, C. (2012). The status and distribution of. *The status and distribution of freshwater biodiversity in Indo-Burma*, 38.
- Kottelat, M., Pinder, A. C., & Harrison, A. (2018). Tor tambra. *The IUCN Red List of Threatened Species*.
- Kottelat, M., Whitten, A. J., Kartikasari, S. N., & Wirjoatmodjo, S. (1993). Freshwater fishes of Western Indonesia and Sulawesi. Periplus Editions.
- Krienitz, L., Hegewald, E. H., Hepperle, D., Huss, V. A., Rohr, T., & Wolf, M. (2004).
 Phylogenetic relationship of *Chlorella* and *Parachlorella* gen. nov. (Chlorophyta, Trebouxiophyceae). *Phycologia*, 43(5), 529-542.
- Kumar, R., Sahoo, P. K., & Barat, A. (2017). Transcriptome profiling and expression analysis of immune responsive genes in the liver of Golden mahseer (*Tor putitora*) challenged with *Aeromonas hydrophila*. *Fish & Shellfish Immunology*, 67, 655-666.

- Kumaran, M., Palanisamy, K. M., Bhuyar, P., Maniam, G. P., Rahim, M. H. A., & Govindan, N. (2023). Agriculture of microalgae *Chlorella vulgaris* for polyunsaturated fatty acids (PUFAs) production employing palm oil mill effluents (POME) for future food, wastewater, and energy nexus. *Energy Nexus*, 9, 100169.
- Kumari, P., Varma, A. K., Shankar, R., Thakur, L. S., & Mondal, P. (2021). Phycoremediation of wastewater by *Chlorella pyrenoidosa* and utilization of its biomass for biogas production. *Journal of Environmental Chemical Engineering*, 9(1), 104974.
- Lange, S., Bambir, S. H., Dodds, A. W., Bowden, T., Bricknell, I., Espelid, S., & Magnadóttir, B. (2006). Complement component C3 transcription in Atlantic halibut (*Hippoglossus hippoglossus* L.) larvae. Fish & Shellfish Immunology, 20(3), 285-294.
- Lange, S., Dodds, A. W., Gudmundsdóttir, S., Bambir, S. H., & Magnadóttir, B. (2005). The ontogenic transcription of complement component C3 and Apolipoprotein AI tRNA in Atlantic cod (*Gadus morhua* L.)—A role in development and homeostasis?. *Developmental and Comparative Immunology*, 29(12), 1065-1077.
- Lau, M. M. L., Lim, L. W. K., Ishak, S. D., Abol-Munafi, A. B., & Chung, H. H. (2021). A review on the emerging asian aquaculture fish, the Malaysian Mahseer (*Tor tambroides*): Current status and the way forward. *Proceedings of the Zoological Society* (Vol. 74, No. 2, pp. 227-237). New Delhi: Springer India.
- Le, T. H., Hoa, N. V., Sorgeloos, P., & Van Stappen, G. (2019). Artemia feeds: a review of brine shrimp production in the Mekong Delta, Vietnam. Reviews in Aquaculture, 11(4), 1169-1175.
- Lee, J. W., Lee, Y. M., Lee, J. H., Noh, J. K., Kim, H. C., Park, C. J., ... & Kim, S. Y. (2013). The expression analysis of complement component C3 during early developmental stages in Olive Flounder (*Paralichthys olivaceus*). *Development* and Reproduction, 17(4), 311.
- Li, K. & Olsen, Y. (2015). Effect of enrichment time and dietary DHA and non-highly unsaturated fatty acid composition on the efficiency of DHA enrichment in phospholipid of rotifer (*Brachionus Cayman*). Aquaculture, 446, 310-317.
- Li, K., Olsen, R. E., Jin, Y., & Olsen, Y. (2018). Phospholipids in marine larval rearing. *Emerging Issues in Fish Larvae Research*, 131-158.
- Li, M. F., & Zhang, H. Q. (2022). An overview of complement systems in teleosts. *Developmental & Comparative Immunology*, 104520.

- Limtipsuntorn, U., Rungsin, W., Thongprajukaew, K., Boonyung, W., & Rangsin, W. (2018). Ontogenic development of enzymatic activity and digestive system in Jullien's golden carp (*Probarbus jullieni* Sauvage, 1880). Aquaculture Research, 49(10), 3362-3373
- Liu, L., Li, Y. L., Xu, S. D., Wang, K. Z., Wu, P., Chu, W. Y., & Wang, X. Q. (2016). Molecular characterization of the myosatin gene and the effect of fasting on its expression in Chinese perch (*Siniperca chuatsi*). *Genetic and Molecular Research*, 15(2), 10-4238.
- Liu, L., Yu, X., & Tong, J. (2012). Molecular characterization of myostatin (MSTN) gene and association analysis with growth traits in the bighead carp (*Aristichthys nobilis*). *Molecular Biology Reports*, 39, 9211-9221.
- Liu, X., Zeng, S., Liu, S., Wang, G., Lai, H., Zhao, X., ... & Li, G. (2020). Identifying the related genes of muscle growth and exploring the functions by compensatory growth in mandarin fish (*Siniperca chuatsi*). *Frontiers in Physiology*, 11, 553563.
- Loh, J. Y., Ong, H. K. A., Hii, Y. S., Smith, T. J., Lock, M. W., & Khoo, G. (2012). Highly unsaturated fatty acid (HUFA) retention in the freshwater cladoceran, *Moina macrocopa*, enriched with lipid emulsions. *Aquaculture*, 354-355, 30-36.
- Lopalco, P., Lobasso, S., Lopes-dos-Santos, R. M. A., Van Stappen, G., & Corcelli, A. (2019). Lipid profile changes during the development of *Artemia franciscana*, from cysts to the first two naupliar stages. *Frontiers in Physiology*, *9*, 1872.
- Løvoll, M., Dalmo, R. A., & Bøgwald, J. (2007). Extrahepatic synthesis of complement components in the rainbow trout (Oncorhynchus mykiss). Fish and Shellfish Immunology, 23(4), 721-731.
- Løvoll, M., Kilvik, T., Boshra, H., Bøgwald, J., Sunyer, J. O., & Dalmo, R. A. (2006). Maternal transfer of complement components C3-1, C3-3, C3-4, C4, C5, C7, Bf, and Df to offspring in rainbow trout (Oncorhynchus mykiss). Immunogenetics, 58, 168-179.
- Lum, K. K., Kim, J., & Lei, X. G. (2013). Dual potential of microalgae as a sustainable biofuel feedstock and animal feed. *Journal of Animal Science and Biotechnology*, 4, 1-7.
- Magnadóttir, B., Lange, S., Gudmundsdottir, S., Bøgwald, J., & Dalmo, R. A. (2005). Ontogeny of humoral immune parameters in fish. *Fish & Shellfish Immunology*, 19(5), 429-439.
- Manklinniam, P., Chittapun, S., & Maiphae, S. (2018). Growth and nutritional value of Moina macrocopa (Straus, 1820) fed with Saccharomyces cerevisiae and Phaffia rhodozyma. Crustaceana, 91(8), 897-912.

- Markou, G., & Nerantzis, E. (2013). Microalgae for high-value compounds and biofuels production: A review with focus on cultivation under stress conditions. *Biotechnology Advances*, 31(8), 1532-1542.
- Marshall, J. S., Warrington, R., Watson, W., & Lim, H. L. (2018). An introduction to immunology and immunopathology. *Allergy, Asthma & Clinical Immunology*, 14(1), 49.
- Martin, S. A., & Król, E. (2017). Nutrigenomics and immune function in fish: New insights from omics technologies. *Developmental & Comparative Immunology*, 75, 86-98.
- Mathimani, T., & Nair, B. B. (2016). Evaluation of microalga for biodiesel using lipid and fatty acid as a marker-a central composite design approach. *Journal of the Energy Institute*, 89(3), 436-446.
- McKinnon, A. D., Duggan, S., Nichols, P. D., Rimmer, M. A., Semmens, G., & Robino,
 B. (2003). The potential of tropical paracalanid copepods as livefeeds in aquaculture. *Aquaculture*, 223, 89-106.
- McPherron, A. C., Lawler, A. M., & Lee, S. J. (1997). Regulation of skeletal muscle mass in mice by a new TGF-p superfamily member. *Nature*, 387(6628), 83-90.
- Méndez-Martínez, Y., García-Guerrero, M. U., Lora-Vilchis, M. C., Martínez-Córdova, L. R., Arcos-Ortega, F. G., Alpuche, J. J., & Cortés-Jacinto, E. (2018). Nutritional effect of Artemia nauplii enriched with Tetraselmis suecica and Chaetoceros calcitrans microalgae on growth and survival on the river prawn Macrobrachium americanum larvae. Aquaculture International, 26, 1001-1015.
- Miah, M. F., Roy, S., Jinnat, E., & Khan, Z. K. (2013). Assessment of Daphnia, Moina and Cylops in freshwater ecosystems and the evaluation of mixed culture in laboratory. American International Journal of Research in Formal, Applied & Natural Sciences, 4(1), 1-7.
- Mian, S., Kader, M. A., & Abol-Munafi, A. B. (2017). Effects of dietary phospholipid levels and sources on growth performance, fatty acid composition and oxidative responsiveness of juvenile Malaysian mahseer, *Tor tambroides. Aquaculture, Aquarium, Conservation and Legislation, 10*(5), 1127-1139.
- Miandare, H. K., Farahmand, H., Akbarzadeh, A., Ramezanpour, S., Kaiya, H., Miyazato, M., ... & Nikinmaa, M. (2013). Developmental transcription of genes putatively associated with growth in two sturgeon species of different growth rate. *General and Comparative Endocrinology*, 182, 41-47.

- Misieng, J. D., Kamarudin, M. S., & Musa, M. (2011). Optimum dietary protein requirement of Malaysian mahseer (*Tor tambroides*) fingerling. *Pakistan Journal* of Biological Sciences, 14(3), 232-235.
- Mitreva, M. (2017). The Microbiome in infectious diseases. In *Infectious Diseases, 2-Volume Set* (pp. 68-74). Elsevier.
- Mohapatra, B. C., Sahoo, S. K., Das Gupta, S., & Gupta, S. D. (2017). Biology of Mahanadi Mahseer, *Tor Mosal Mahanadicus* (David) Reared in Freshwater Pond Culture System. *Current Agriculture Research Journal*, 5(2).
- Mohsin, A. M., & Ambak, M. A. (1983). *Freshwater fishes of Peninsular Malaysia*. Penerbit Universiti Pertanian Malaysia.
- Mokhtar, D. M., Zaccone, G., Alesci, A., Kuciel, M., Hussein, M. T., & Sayed, R. K. (2023). Main components of fish immunity: An overview of the fish immune system. *Fishes*, 8(2), 93.
- Mona, M. H., El-Gamal, M. M., Razek, F. A., & Eldeen, M. N. (2017). Utilization of Daphnia longispina as supplementary food for rearing Marsupenaeus japonicus post larvae. Journal of the Marine Biological Association of India, 59(2), 74.
- Moradi-Kheibari, N., Ahmadzadeh, H., & Lyon, S. R. (2022). Correlation of total lipid content of *Chlorella vulgaris* with the dynamics of individual fatty acid growth rates. *Frontiers in Marine Science*, *9*, 837067.
- Morais, S., Conceição, L. E. C., Rønnestad, I., Koven, W., Cahu, C., Infante, J. Z., & Dinis, M. T. (2007). Dietary neutral lipid level and source in marine fish larvae: effects on digestive physiology and food intake. *Aquaculture*, 268(1-4), 106-122.
- Moriyama, S., Ayson, F. G., & Kawauchi, H. (2000). Growth regulation by insulin-like growth factor-I in fish. *Bioscience*, *Biotechnology*, and *Biochemistry*, 64(8), 1553-1562.
- Mousavi-Sabet, H., Eagderi, S., Moshayedi, F., & Jalili, P. (2015). The effects of supplemental ascorbic acid and unsaturated fatty acids in enriched *Artemia* on growth performance and stress resistance of sailfin molly fry, *Poecilia latipinna*. *Poecilia Research*, 5(1), 31-38.
- Munirasu, S., Ramasubramanian, V., Uthayakumar, V., & Muthukumar, S. (2013). Bioenrichment of live feed *Daphnia magna* for the survival and growth of freshwater fish *Catla catla*. *International Journal of Current Research and Review*, 5(8), 20.
- Naceur, H. B., Romdhan, M. S., & Stappen, G. V. (2020). Potential Use of fatty acid profile for Artemia sp. discrimination. Inland Water Biology, 13, 434-444.

- Nadjar-Boger, E., & Funkenstein, B. (2011). Myostatin-2 gene structure and polymorphism of the promoter and first intron in the marine fish *Sparus aurata*: evidence for DNA duplications and/or translocations. *BMC Genetics*, 12, 1-19.
- Najafpour, B., Cardoso, J. C., Canário, A. V., & Power, D. M. (2020). Specific evolution and gene family expansion of complement 3 and regulatory factor H in fish. *Frontiers in Immunology*, 11, 568631.
- Nanton, D. A. & Castell, J. D. (1998) The effects of dietary fatty acids on the fatty acid composition of the harpacticoid copepod, *Tisbesp.*, for use as a live food for marine fish larvae. *Aquaculture*, 163, 251-261.
- Navarro, J. C., Henderson, R. J., McEvoy, L. A., Bell, M. V., & Amat, F. (1999). Lipid conversions during enrichment of *Artemia*. Aquaculture, 174(1), 155-166.
- Nayak, S., Al Ashhab, A., Zilberg, D., & Khozin-Goldberg, I. (2020). Dietary supplementation with omega-6 LC-PUFA-rich microalgae regulates mucosal immune response and promotes microbial diversity in the zebrafish gut. *Biology*, 9(6), 119.
- Negm, R. K., Cobcroft, J. M., Brown, M. R., Nowak, B. F., & Battaglene, S. C. (2014). Performance and skeletal abnormality of striped trumpeter *Latris lineata* larvae and post larvae fed vitamin A enriched *Artemia. Aquaculture*, 422, 115-123.
- Neri, T. A., Rohmah, Z., Ticar, B. F., & Choi, B. D. (2020). Effect of different culture conditions on nutritional value of *Moina macrocopa* as a live feed for fish fry production. *Journal of Agriculture & Life Sciences Research*, 54(6), 91-98.
- Neustupa, J., Němcová, Y., Eliáš, M., & Škaloud, P. (2009). Kalinella bambusicola gen. et sp. nov.(Trebouxiophyceae, Chlorophyta), a novel coccoid Chlorella-like subaerial alga from Southeast Asia. Phycological Research, 57(3), 159-169.
- Ng, C. K. (2004). *Kings of the Rivers: Mahseer in Malaysia and the Region*. Inter Sea Fishery.
- Ng, J. Y., Chua, M. L., Zhang, C., Hong, S., Kumar, Y., Gokhale, R., & Ee, P. L. R. (2020). *Chlorella vulgaris* extract as a serum replacement that enhances mammalian cell growth and protein expression. *Frontiers in Bioengineering and Biotechnology*, 8, 564667.
- Ng, W. K., & Andin, V. C. (2011). The Malaysian mahseer, *Tor tambroides* (Bleeker), requires low dietary lipid levels with a preference for lipid sources with high omega-6 and low omega-3 polyunsaturated fatty acids. *Aquaculture*, 322, 82-90.
- Ng, W. K., Abdullah, N., & De Silva, S. S. (2008). The dietary protein requirement of the Malaysian mahseer, *Tor tambroides* (Bleeker), and the lack of protein-sparing action by dietary lipid. *Aquaculture*, 284(1-4), 201-206.

- Nguyen, T. T., Na-Nakorn, U., Sukmanomon, S., & ZiMing, C. (2008). A study on phylogeny and biogeography of mahseer species (Pisces: Cyprinidae) using sequences of three mitochondrial DNA gene regions. *Molecular Phylogenetics* and Evolution, 48(3), 1223-1231.
- Nichols, H. W., & Bold, H. C. (1965). Trichosarcina polymorpha gen. et sp. nov. Journal of Phycology, 1(1), 34-38.
- Nick, G. L. (2003). Addressing human exposure to environmental toxins with Chlorella pyrenoidosa (Medicinal Properties in Whole Foods). Townsend Letter for Doctors and Patients, (237), 28-33.
- Ohs, C. L., Cassiano, E. J., & Rhodes, A. (2010). Choosing an Appropriate Live feed for Larviculture of Marine Fish: FA167/FA167, 12/2009. *EDIS*, 2010(2).
- Øie, G., Makridis, P., Reitan, K. I., & Olsen, Y. (1997). Protein and carbon utilization of rotifers (*Brachionus plicatilis*) in first feeding of turbot larvae (*Scophthalmus maximus* L.). Aquaculture, 153(1-2), 103-122.
- Opazo, R., Valladares, L., & Romero, J. (2017). Comparison of gene expression patterns of key growth genes between different rate growths in zebrafish (*Danio rerio*) siblings. *Latin American Journal of Aquatic Research*, 45(4), 766-775.
- Palma, J., Bureau, D. P., & Andrade, J. P. (2011). Effect of different Artemia enrichments and feeding protocol for rearing juvenile long snout seahorse, *Hippocampus* guttulatus. Aquaculture, 318, 439-443.
- Patruno, M., Sivieri, S., Poltronieri, C., Sacchetto, R., Maccatrozzo, L., Martinello, T., ... & Radaelli, G. (2008). Real-time polymerase chain reaction, in situ hybridization and immunohistochemical localization of insulin-like growth factor-I and myostatin during development of *Dicentrarchus labrax* (Pisces: Osteichthyes). *Cell and Tissue Research*, 331, 643-658.
- Pepin, P. (2023). Feeding by larval fish: how taxonomy, body length, mouth size, and behaviour contribute to differences among individuals and species from a coastal ecosystem. *ICES Journal of Marine Science*, *80*(1), 91-106.
- Peter, R. E., & Marchant, T. A. (1995). The endocrinology of growth in carp and related species. *Aquaculture*, *129*(1-4), 299-321
- Philia, J., & Wibisono, J. (2018). Cultivation of microalgae *Chlorella* sp. on fresh water and waste water of tofu industry. In *E3S Web of Conferences* (Vol. 31, p. 04009). EDP Sciences.
- Pina-Pérez, M. C., Brück, W. M., Brück, T., & Beyrer, M. (2019). Microalgae as healthy ingredients for functional foods. In *The role of alternative and innovative food ingredients and products in consumer wellness* (pp. 103-137). Academic Press.

- Pinder, A. C., Britton, J. R., Harrison, A. J., Nautiyal, P., Bower, S. D., Cooke, S. J., ... & Raghavan, R. (2019). Mahseer (*Tor* spp.) fishes of the world: status, challenges and opportunities for conservation. *Reviews in Fish Biology and Fisheries*, 29, 417-452.
- Pittman, K., Yúfera, M., Pavlidis, M., Geffen, A. J., Koven, W., Ribeiro, L., ... & Tandler, A. (2013). Fantastically plastic: fish larvae equipped for a new world. *Reviews in Aquaculture*, 5, S224-S267.
- Poh, K. R. B. (2006). Morphological and Genetic Identification, and Population Structure of Kelah (Tor tambroides) (Doctoral dissertation). Universiti Putra Malaysia.
- Pollar, M., Jaroensutasinee, M., & Jaroensutasinee, K. (2007). Morphometric analysis of *Tor tambroides* by stepwise discriminant and neural network analysis. *International Journal of Bioengineering and Life Sciences*, 1(9), 106-110.
- Praveen, K., Abinandan, S., Natarajan, R., & Kavitha, M. S. (2018). Biochemical responses from biomass of isolated *Chlorella* sp., under different cultivation modes: non-linear modelling of growth kinetics. *Brazilian Journal of Chemical Engineering*, 35, 489-496.
- Priyadarshani I, Rath B (2012) Commercial and industrial applications of microalgae a review. *Journal of Algal Biomass Utilisation*, *3*, 89-100.
- Radha, S., Fathima, A. A., Iyappan, S., & Ramya, M. (2013). Direct colony PCR for rapid identification of varied microalgae from freshwater environment. *Journal* of Applied Phycology, 25, 609-613.
- Radhakrishnan, S., Bhavan, P. S., Scenivasan, C., Shanthi, R., & Muralisankar, T. (2014). Replacement of fishmeal with *Spirulina platensis, Chlorella vulgaris* and *Azolla pinnata* on non-enzymatic and enzymatic antioxidant activities of *Macrobrachium rosenbergii. The Journal of Basic & Applied Zoology*, 67(2), 25-33.
- Rahimnejad, S., Lee, S. M., Park, H. G., & Choi, J. (2017). Effects of dietary inclusion of *Chlorella vulgaris* on growth, blood biochemical parameters, and antioxidant enzyme activity in olive flounder, *Paralichthys olivaceus*. *Journal of the World Aquaculture Society*, 48(1), 103-112.
- Rahman, M. M., & Sorgeloos, P. (2022). A training manual on *Artemia* cyst hatching and decapsulation. Retrieved from <u>https://hdl.handle.net/20.500.12348/5375</u>
- Raji, A. A., Jimoh, W. A., Bakar, N. A., Taufek, N. M., Muin, H., Alias, Z., ... & Razak, S. A. (2020). Dietary use of *Spirulina (Arthrospira)* and *Chlorella* instead of fish

meal on growth and digestibility of nutrients, amino acids and fatty acids by African catfish. *Journal of Applied Phycology*, *32*, 1763-1770.

- Ramezani-Fard, E., Kamarudin, M. S., Harmin, S. A., Saad, C. R., Abd Satar, M. K., & Daud, S. K. (2011). Ontogenic development of the mouth and digestive tract in larval Malaysian mahseer, *Tor tambroides* Bleeker. *Journal of Applied Ichthyology*, 27(3), 920-927.
- Ramezani-Fard, E., Kamarudin, M. S., Saad, C. R., Harmin, S. A., & Meng, G. Y. (2012). Dietary lipid levels affect growth and fatty acid profiles of Malaysian mahseer, *Tor tambroides. North American Journal of Aquaculture*, 74(4), 530-536.
- Ramírez-López, C., Chairez, I., & Fernández-Linares, L. (2016). A novel culture medium designed for the simultaneous enhancement of biomass and lipid production by *Chlorella vulgaris* UTEX 26. *Bioresource Technology*, 212, 207-216.
- Rasdi, N. W., Abdullah, M. I., Azman, S., Karim, M., Syukri, F. & Hagiwara, A. (2021). The effects of enriched *Moina* on the growth, survival, and proximate analysis of marine shrimp (*Penaeus monodon*). *Journal of Sustainability Science and Management*, 16, 56-70.
- Rasdi, N. W., Arshad, A., Ikhwanuddin, M., Hagiwara, A., Yusoff, F. M., & Azani, N. (2020a). A review on the improvement of cladocera (*Moina*) nutrition as live food for aquaculture: Using valuable plankton fisheries resources. *Journal of Environmental Biology*, 41, 1239-1248.
- Rasdi, N. W., Ramlee, A., Abol-Munafi, A. B., Ikhwanuddin, M., Azani, N., Yuslan, A.,
 ... & Arshad, A. (2020b). The effect of enriched Cladocera on growth, survivability and body coloration of Siamese fighting fish. *Journal of Environmental Biology*, 41, 1257-1263.
- Rasdi, N. W., Suhaimi, H., Hagiwara, A., Ikhwanuddin, M., Ghaffar, M. A., Yuslan, A., & Najuwa, S. (2019). Effect of different salinities gradient on fatty acid composition, growth, survival and reproductive performance of *Moina* macrocopa (Straus 1820) (Crustacea, Cladocera). Journal of Sustainability Science and Management, 16(4), 56-70
- Ricker, W. E. (1979). Growth rates and models. In: *Hoar, W. S., Randall, D. J., & Brett, J. R. (Eds.), Fish Physiology, Volume VIII. Bioenergetics and Growth* (pp. 679-743). New York. Academic Press.
- Roberts, T. R. (1999). Fishes of the cyprinid genus *Tor* in the Nam Theun watershed (Mekong basin) of Laos, with description of a new species. *Raffles Bulletin of Zoology*, 47, 225-236.

- Roberts, T. R., & Khaironizam, M. Z. (2008). Trophic polymorphism in the Malaysian fish *Neolissochilus soroides* and other old-world barbs (Teleostei, Cyprinidae). *Natural History Bulletin of the Siam Society*.
- Rocha, J. M., Garcia, J. E., & Henriques, M. H. (2003). Growth aspects of the marine microalga Nannochloropsis gaditana. Biomolecular Engineering, 20(4-6), 237-242.
- Roslina, K. (2018). Contribution of Brackish and Freshwater Aquaculture to Livelihood of Small-Scale Rural Aquaculture Farmers in Kedah, Malaysia. *Pertanika Journal of Social Sciences & Humanities*, 26(3).
- Roy, S. S., & Pal, R. (2015). Microalgae in aquaculture: a review with special references to nutritional value and fish dietetics. *Proceedings of the Zoological Society* (Vol. 68, pp. 1-8). Springer India.
- Roy, U., Sawant, P. B., Debroy, S., & Haque, R. (2022). Live Feed Enrichment–A Possible Step Towards a Sustainable Blue Revolution. *International Journal of Research in Engineering, Science and Management*, 5(10), 40-42.
- Ru, I. T. K., Sung, Y. Y., Jusoh, M., Wahid, M. E. A., & Nagappan, T. (2020). Chlorella vulgaris: A perspective on its potential for combining high biomass with high value bioproducts. Applied Phycology, 1(1), 2-11.
- Rushan, N. H., Mat Yasin, N. H., & Said, F. M. (2021). The effect of culture medium on the oil yield and fatty acid methyl ester of freshwater microalgae *Chlorella* vulgaris. Chemical Engineering Communications, 208(4), 592-600.
- Safi, C., Zebib, B., Merah, O., Pontalier, P. Y., & Vaca-Garcia, C. (2014). Morphology, composition, production, processing and applications of *Chlorella vulgaris*: A review. *Renewable and Sustainable Energy Reviews*, 35, 265–278.
- Sam, K. K., Merosha, P., Janaranjani, M., Athirah, I., & Shu-Chien, A. C. (2021). The Malaysian Mahseer, *Tor tambroides* possess all required biosynthesis enzymes for the conversion of C18 polyunsaturated fatty acids to long-chain polyunsaturated fatty acids. *Aquaculture*, 543, 736942
- Samat, N. A., Yusoff, F. M., Rasdi, N. W., & Karim, M. (2020). Enhancement of live food nutritional status with essential nutrients for improving aquatic animal health: A review. *Animals*, 10(12), 2457.
- Schoch, C. L., Seifert, K. A., Huhndorf, S., Robert, V., Spouge, J. L., & Levesque, C. A. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences of the United States of America*, 109, 6241-6246.

- Secombes, C. J., & Wang, T. (2012). The innate and adaptive immune system of fish. In *Infectious disease in aquaculture*, 3-68. Woodhead Publishing.
- Segev-Hadar, A., Alupo, G., Tal, K., Nitzan, T., & Biran, J. (2020). Identification and characterization of a non-muscular myostatin in the Nile tilapia. *Frontiers in Endocrinology*, 11, 94.
- Shalaby, E. A., & Dubey, N. K. (2018). Polysaccharides from cyanobacteria: response to biotic and abiotic stress and their antiviral activity. *Indian Journal of Geo Marine Sciences*, 47(1), 21-33.
- Sharma, P., Gujjala, L. K. S., Varjani, S., & Kumar, S. (2022). Emerging microalgaebased technologies in biorefinery and risk assessment issues: Bioeconomy for sustainable development. *Science of The Total Environment*, 813, 152417.
- Shen, T., Lei, M., Wang, J., He, X., Li, X., & Li, J. (2014). Molecular cloning, organization, expression and 3D structural analysis of the MHC class 1a gene in the whitespotted bamboo shark (*Chiloscyllium plagiosum*). Veterinary Immunology and Immunopathology, 157(1-2), 111-118.
- Sheng, Y., Sun, Y., Zhang, X., Wan, H., Yao, C., Liang, K., ... & Wang, Y. (2020). Characterization of two myostatin genes in pufferfish *Takifugu bimaculatus*: Sequence, genomic structure, and expression. *PeerJ*, 8, e9655.
- Shepard, A. K. (2015). Responses of heterotrophic and autotrophic pico-and nanoplankton to nutrient availability and enrichment across marine systems in the Northern Gulf of Mexico (Doctoral dissertation). Louisiana State University.
- Shi, X., Luo, Z., Chen, F., Wei, C. C., Wu, K., Zhu, X. M., & Liu, X. (2017). Effect of fish meal replacement by *Chlorella* meal with dietary cellulase addition on growth performance, digestive enzymatic activities, histology and myogenic genes' expression for crucian carp *Carassius auratus*. *Aquaculture Research*, 48(6), 3244-3256.
- Singh, K., Munilkumar, S., Sahu, N. P., Das, A., & Devi, G. A. (2019). Feeding HUFA and vitamin C-enriched *Moina micrura* enhances growth and survival of *Anabas testudineus* (Bloch, 1792) larvae. *Aquaculture*, 500, 378-384.
- Smith, N. C., Rise, M. L., & Christian, S. L. (2019). A comparison of the innate and adaptive immune systems in cartilaginous fish, ray-finned fish, and lobe-finned fish. *Frontiers in Immunology*, 10, 2292.
- Soeta, N., Terashima, M., Gotoh, M., Mori, S., Nishiyama, K., Ishioka, K., ... & Suzutani, T. (2009). An improved rapid quantitative detection and identification method for a wide range of fungi. *Journal of Medical Microbiology*, 58(8), 1037-1044.

- Soon, L. K., Lihan, S., Dasthagir, F. F. G., Mikal, K. M., Collick, F., & Hua, N. K. (2014). Microbiological and physicochemical analysis of water from Empurau fish (*Tor tambroides*) farm in Kuching, Sarawak, Malaysian Borneo. *International Journal of Scientific & Technology Research*, 3(6), 285-292.
- Sorgeloos, P., Dhert, P., & Candreva, P. (2001). Use of the brine shrimp, Artemia spp., in marine fish larviculture. Aquaculture, 200(1-2), 147-159.
- Sorgeloos, P., Lavens, P., Leger, P., & Tackaert, W. (1993). The use of Artemia in marine fish larviculture. In TML Conference Proceedings, 3, 73-86.
- Sprecher, H. (2000). Metabolism of highly unsaturated n-3 and n-6 fatty acids. *Biochimica et Biophysica Acta*, 1486, 219-231.
- Srichanun, M., Tantikitti, C., Vatanakul, V., & Musikarune, P. (2012). Digestive enzyme activity during ontogenetic development and effect of live feed in green catfish larvae (*Mystus nemurus* Cuv. & Val. Songklanakarin Journal of Science & Technology, 34(3).
- Steinbacher, P., Haslett, J. R., Six, M., Gollmann, H. P., Sänger, A. M., & Stoiber, W. (2010). Phases of myogenic cell activation and possible role of dermomyotome cells in teleost muscle formation. *Developmental dynamics*, 235(11), 3132-3143.
- Støttrup, J., & McEvoy, L. (Eds.). (2008). Live feeds in marine aquaculture. John Wiley & Sons.
- Strandberg, U., Taipale, S. J., Kainz, M. J., & Brett, M. T. (2014). Retroconversion of docosapentaenoic acid (n-6): an alternative pathway for biosynthesis of arachidonic acid in *Daphnia magna*. *Lipids*, 49, 591-595.
- Sun, Z., Tan, X., Liu, Q., Ye, H., Zou, C., Xu, M., ... & Ye, C. (2019). Physiological, immune responses and liver lipid metabolism of orange-spotted grouper (*Epinephelus coioides*) under cold stress. *Aquaculture*, 498, 545-555.
- Teng, T., Zhao, X., Li, C., Guo, J., Wang, Y., Pan, C., ... & Ling, Q. (2020). Cloning and expression of IGF-I, IGF-II, and GHR genes and the role of their single-nucleotide polymorphisms in the growth of pikeperch (*Sander lucioperca*). Aquaculture International, 28, 1547-1561.

Thomas, H. S. (1873). The Rod in India... Naval & Military Press.

Ticiani, D., Delariva, R. L., Iquematsu, M. S., & Bialetzki, A. (2022). Larval development of *Characidium orientale* (Actinopterygii: Crenuchidae) a small Neotropical fish. *Iheringia. Série Zoologia*, 112, e2022003.

- Tomaselli, L. (2004). The microalgal cell. *Handbook of microalgal culture: Biotechnology and Applied Phycology*, 1, 3-19.
- Treece, G. D. (2000). *Artemia production for marine larval fish culture*, 702. Stoneville, Mississippi: Southern Regional Aquaculture Center.
- Triantaphyllopoulos, K. A., Cartas, D., & Miliou, H. (2020). Factors influencing GH and IGF-I gene expression on growth in teleost fish: how can aquaculture industry benefit?. *Reviews in Aquaculture*, 12(3), 1637-1662.
- Valente, L. M., Moutou, K. A., Conceição, L. E., Engrola, S., Fernandes, J. M., & Johnston, I. A. (2013). What determines growth potential and juvenile quality of farmed fish species?. *Reviews in Aquaculture*, 5, S168-S193.
- Van Wychen, S., & Laurens, L. M. L. (2013). Determination of total lipids as fatty acid methyl esters (FAME) by in situ transesterification. *Contract*, 303, 275-3000.
- Vangansbeke, D., Nguyen, D. T., Audenaert, J., Gobin, B., Tirry, L., & De Clercq, P. (2016). Establishment of *Amblyseius swirskii* in greenhouse crops using food supplements. *Systematic and Applied Acarology*, 21(9), 1174-1184.
- Venegas-Calerón, M., Sayanova, O., & Napier, J. A. (2010). An alternative to fish oils: metabolic engineering of oil-seed crops to produce omega-3 long chain polyunsaturated fatty acids. *Progress in Lipid Research*, 49(2), 108-119.
- Volkman, J. K., Jeffrey, S. W., Nichols, P. D., Rogers, G. I., & Garland, C. D. (1989). Fatty acid and lipid composition of 10 species of microalgae used in mariculture. *Journal of Experimental Marine Biology and Ecology*, *128*(3), 219-240.
- Vrijenhoek, R. C. (1998). Conservation genetics of freshwater fish. Journal of fish Biology, 53, 394-412.
- Walock, C. D., Kittilson, J. D. & Sheridan, M. A. (2014) Characterization of a novel growth hormone receptor-encoding cDNA in rainbow trout and regulation of its expression by nutritional state. *Gene*, 533, 286-294.
- Walton, S. E., Gan, H. M., Raghavan, R., Pinder, A. C., & Ahmad, A. (2016).
 Disentangling the taxonomy of the mahseers (*Tor spp.*) of Malaysia: An integrated approach using morphology, genetics and historical records. *Reviews in Fisheries Science & Aquaculture*, 25(3), 171-183.
- Wang, C., Perera, T. V., Ford, H. L., & Dascher, C. C. (2003). Characterization of a divergent non-classical MHC class I gene in sharks. *Immunogenetics*, 55, 57-61.
- Wilson, A. B. (2017). MHC and adaptive immunity in teleost fishes. *Immunogenetics*, 69(8-9), 521-528.

- Wiwattanapatapee, R., Padoongsombat, N., Choochom, T., Tang, S., & Chaimongkol, A. (2002). Water flea *Moina macrocopa* as a novel biocarrier of norfloxacin in aquaculture. *Journal of Controlled Release*, 83(1), 23-28.
- Wong, Y., Ho, Y. H., Ho, K. C., Leung, H. M., & Yung, K. K. L. (2017). Growth medium screening for *Chlorella vulgaris* growth and lipid production. *Journal of Aquaculture and Marine Biology*, 6(1), 00143.
- Wu, X. M., Hu, Y. W., Xue, N. N., Ren, S. S., Chen, S. N., Nie, P., & Chang, M. X. (2017). Role of zebrafish NLRC5 in antiviral response and transcriptional regulation of MHC related genes. *Developmental & Comparative Immunology*, 68, 58-68.
- Xie, S., Aiguo Z., Yongyong F., Zhenlu W., Lanfen F., Yue Z., ... & Jixing Zou. (2019). Effects of fasting and re-feeding on mstn and mstnb genes expressions in *Cranoglanis bouderius. Gene*, 682, 1-12.
- Yan, K., Guo, F., Kainz, M. J., Li, F., Gao, W., Bunn, S. E., & Zhang, Y. (2023). The importance of omega-3 polyunsaturated fatty acids as high-quality food in freshwater ecosystems with implications of global change. *Biological Reviews*.
- Zaleha, K., & Busra, I. (2012). Culture of harpacticoid copepods: understanding the reproduction and effect of environmental factors. *Aquaculture*, 343-360.
- Zapata, A., Diez, B., Cejalvo, T., Gutierrez-de Frias, C., & Cortés, A. (2006). Ontogeny of the immune system of fish. *Fish & Shellfish Immunology*, 20(2), 126-136.
- Zeng, C., Shao, L., Ricketts, A., & Moorhead, J. (2018). The importance of copepods as live feed for larval rearing of the green mandarin fish *Synchiropus splendidus*. *Aquaculture*, 491, 65-71.
- Zhang, S., Li, Y., Shao, J., Liu, H., Wang, J., Wang, M., ... & Bian, W. (2020). Functional identification and characterization of IpMSTNa, a novel orthologous myostatin (MSTN) gene in channel catfish *Ictalurus punctatus*. *International Journal of Biological Macromolecules*, 152, 1-10.
- Zhou, W., & Cui, G. H. (1996). A review of *Tor* species from the Lancangjiang River (Upper Mekong River), China (Teleostei: Cyprinidae). *Ichthyological Exploration of Freshwaters*, 7, 131-142.
- Zhu, K., H. Wang, H. Wang, Y. Gul, M. Yang, C. Zeng, and W. Wang. (2014). Characterization of muscle morphology and satellite cells, and expression of muscle-related genes in skeletal muscle of juvenile and adult *Megalobrama amblycephala*. *Micron* 64: 66-75.