



FRESHWATER ZOOPLANKTON DIVERSITY AND CULTURE OF  
SELECTED SPECIES WITH EMPHASIS ON REPRODUCTIVE CAPACITIES  
AND NUTRITIONAL CONTENTS

By

UMI WAHIDAH BINTI AHMAD DINI

Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of  
Philosophy

December 2023

IB 2023 6

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, are copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement for the degree of Doctor of Philosophy

**FRESHWATER ZOOPLANKTON DIVERSITY AND CULTURE OF  
SELECTED SPECIES WITH EMPHASIS ON REPRODUCTIVE CAPACITIES  
AND NUTRITIONAL CONTENTS**

By

**UMI WAHIDAH BINTI AHMAD DINI**

**December 2023**

**Chair : Professor Fatimah binti Md. Yusoff, PhD**  
**Institute : Bioscience**

Zooplankton forms a critical link between the base of the aquatic food web and the top-level consumers; therefore, they are important live-feed organisms in the aquaculture industry. However, the unavailability and unstable production of highly nutritious live feeds are major bottlenecks for the aquaculture industry. Thus, understanding water quality and food requirements for zooplankton is pivotal to the success of sustainable production of zooplankton as live feed in hatcheries. Therefore, the current study aimed to evaluate the distribution and diversity of freshwater zooplankton and identify prospective candidates as live feeds. Subsequently, this study was conducted to ascertain the ideal conditions for zooplankton culture and to assess the effectiveness of microalgae-bacteria encapsulation in enhancing zooplankton quality. A study on zooplankton distribution was carried out at 20 different freshwater water bodies in Malaysia. Physicochemical parameters were measured *in situ*, while water and zooplankton samples were collected for nutrient analyses and for zooplankton identification and enumeration. Selected zooplankton species were molecularly characterised using the 18S rDNA approach. Each of the selected zooplankton species was exposed to several parameters, including temperature (25 °C, 27 °C and 30 °C), pH (pH 5, pH 7, and pH 9), unionized-ammonia, NH<sub>3</sub>-N (0.1 mg L<sup>-1</sup>, 0.3 mg L<sup>-1</sup>, and 0.5 mg L<sup>-1</sup>), and diets (13 diets, including mono and mixed diets). Each treatment was conducted in triplicate. Daily observations were performed for the life cycle, population density, and population growth rate. Proximate analysis was conducted according to standard methods, and fatty acids were analysed using a gas chromatograph–mass spectrometer (GC-MS). A total of 58 zooplankton species, consisting of 12 species of cladocerans, 10 species of copepods, and 36 species of rotifers, were recorded. Environmental parameters associated with eutrophication (high nutrients, chlorophyll a, turbidity, and low water transparency with a Global Test,  $p = 0.508$ ) affect the diversity and composition of zooplankton. Several tolerant species such as *Ceriodaphnia cornuta*, *Brachionus calyciflorus*, *Keratella cochlearis*, *Trichocerca*

*similis*, *Thermocyclops crassus*, and *Mesocyclops thermocyclopoides* can thrive in high eutrophic conditions, indicating that these species could serve as indicators. Among the three freshwater zooplankton groups, Cladocera was selected due to its reproduction mode and nutritional quality. Three species (*C. cornuta*, *Moina micrura* and *Diaphanosoma excisum*) of cladocerans were selected for cultivation based on their abundance and differences in size. The present results pointed out that the optimum water quality requirements for all three cladocerans were 27 °C, pH 7, and 0.1 mg L<sup>-1</sup> NH<sub>3</sub>-N, which supported the fastest maturation, the highest reproductive capacities (production of eggs, offspring, and clutches), and maximized the nutritional contents of all cladocerans species. Furthermore, regarding diets, the combination of the bacterium, *Ochrobactrum haematophilum* with high-quality microalgae, especially the T10 diet (*Cyclotella meneghiniana* + *O. haematophilum*) found to be the ideal diet for all cladocerans, which enhanced the reproductive capacities and nutritional composition. Meanwhile, cyanobacteria diets (*Microcystis aeruginosa* and *Planktothrix agardhii*) were considered as poor diets which hindered the growth of cladocerans. However, the addition of *O. haematophilum* to *M. aeruginosa* and *P. agardhii* diets (T12 and T13) increased the population density and reproductive capacities of all cladocerans but not the nutritional contents. Overall, the selection of optimal culture conditions and microalgaebacteria consortium have important implications for improving the production of high-quality cladocerans for sustainable cultivation, which can be utilized as high-quality live feeds in the aquaculture industry.

Keywords: Cladocera; live food; microalgae; bacteria; nutritional quality

SDG: No poverty; zero hunger; life below water

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KEPELBAGAIAN ZOOPLANKTON AIR TAWAR DAN PENGKULTURAN  
SPESIES TERPILIH DENGAN PENEKANAN KE ATAS KAPASITI  
PEMBIAKAN DAN KANDUNGAN NUTRISI**

Oleh

**UMI WAHIDAH BINTI AHMAD DINI**

**Disember 2023**

Pengerusi : Profesor Fatimah binti Md. Yusoff, PhD  
Institut : Biosains

Zooplankton membentuk hubungan kritikal di antara asas siratan makanan dengan pengguna peringkat atasan; oleh itu, zooplankton adalah organisme makanan hidup yang penting dalam industri akuakultur. Walau bagaimanapun, ketidaksediaan dan ketidakstabilan pengeluaran makanan hidup yang sangat berkhasiat adalah halangan utama bagi industri akuakultur. Memahami kualiti air dan keperluan makanan untuk zooplankton adalah penting untuk kejayaan pengeluaran zooplankton sebagai makanan hidup secara mampan. Justeru, kajian ini bertujuan untuk menilai taburan dan diversiti zooplankton air tawar dan mengenal pasti spesies zooplankton yang sesuai sebagai makanan hidup. Seterusnya, kajian ini dijalankan untuk mengenal pasti kondisi yang ideal untuk pengkulturan zooplankton dan menilai keberkesanan gabungan mikroalga-bakteria dalam meningkatkan kualiti zooplankton. Kajian mengenai taburan zooplankton telah dijalankan di 20 lokasi air tawar yang berbeza di Malaysia. Parameter fizikokimia diukur secara *in situ*, manakala sampel air dan zooplankton dikumpul untuk analisis nutrien dan untuk pengenalpastian dan penghitungan zooplankton. Spesies zooplankton terpilih telah dicirikan secara molekul menggunakan kaedah 18S rDNA. Setiap spesies zooplankton yang dipilih telah didedahkan kepada beberapa parameter, termasuk suhu ( $25^{\circ}\text{C}$ ,  $27^{\circ}\text{C}$  dan  $30^{\circ}\text{C}$ ), pH (pH 5, pH 7, dan pH 9), ammonia,  $\text{NH}_3\text{-N}$  ( $0.1 \text{ mg L}^{-1}$ ,  $0.3 \text{ mg L}^{-1}$ , dan  $0.5 \text{ mg L}^{-1}$ ), dan diet (13 diet, termasuk diet mono dan campuran). Setiap parameter diulang sebanyak tiga kali. Pemerhatian harian dilakukan untuk kitaran hayat, kepadatan populasi, dan kadar pertumbuhan populasi. Analisis proksimat telah dijalankan mengikut kaedah standard, dan asid lemak dianalisis menggunakan spektrometer jisim kromatografi gas (GC-MS). Sebanyak 58 zooplankton spesies, terdiri daripada 12 spesies Kladosera, 10 spesies Copepoda, and 36 spesies Rotifera telah direkodkan. Parameter alam sekitar

yang dikaitkan dengan eutrofikasi (nutrien, kekeruhan, dan klorofil a yang tinggi dengan Ujian global,  $\rho = 0.508$ ) memberi kesan kepada diversiti dan komposisi zooplankton. Beberapa spesies toleran seperti *Ceriodaphnia cornuta*, *Brachionus calyciflorus*, *Keratella cochlearis*, *Trichocerca similis*, *Thermocyclops crassus*, dan *Mesocyclops thermocycloides* mampu bertahan dalam kondisi eutropik yang tinggi; menunjukkan bahawa spesies ini boleh berfungi sebagai indikator. Antara tiga kumpulan zooplankton air tawar, kladosera telah dipilih berdasarkan kaedah pembiakan dan kualiti nutrisi. Tiga spesies (*C. cornuta*, *Moina micrura* and *Diaphanosoma excisum*) kladosera telah dipilih untuk pengkulturan berdasarkan saiz dan kelimpahan yang berbeza. Keputusan menunjukkan bahawa kualiti air yang ideal untuk kesemua kladosera adalah pada suhu  $27^{\circ}\text{C}$ , pH 7, dan  $0.1 \text{ mg L}^{-1}$   $\text{NH}_3\text{-N}$  yang didapati telah mempercepatkan kematangan, meningkatkan hasil pembiakan (pengeluaran telur, anak dan kantung telur), dan memaksimumkan kandungan nutrisi kesemua kladosera. Seterusnya, dari segi diet, campuran bakteria (*Ochrobactrum haematophilum*) dengan mikroalga yang mempunyai kualiti yang tinggi, terutamanya diet T10 (*Cyclotella meneghiniana* + *O. haematophilum*) didapati sebagai diet yang ideal untuk semua kladosera yang telah meningkatkan hasil pembiakan dan kandungan nutrisi. Sementara itu, diet cyanobacteria (*Microcystis aeruginosa* and *Planktothrix agardhii*) dianggap sebagai diet yang tidak elok kerana telah menyekat pertumbuhan kladosera. Walau bagaimanapun, penambahan campuran *O. haematophilum* ke dalam diet *M. aeruginosa* and *P. agardhii* (T12 and T13) telah meningkatkan ketumpatan populasi dan kapasiti pembiakan kladosera, tetapi bukan kandungan nutrisi. Secara keseluruhan, pemilihan keadaan pengkulturan yang optimum dan campuran mikroalga-bakteria sebagai makanan untuk kladosera mempunyai implikasi penting untuk meningkatkan pengeluaran kladosera berkualiti tinggi untuk pengkulturan kladosera secara mampan, yang boleh digunakan sebagai makanan hidup berkualiti tinggi dalam industri akuakultur.

Kata kunci: Kladosera; makanan hidup; microalga; bakteria; kualiti nutrisi  
SDG: Tiada kemiskinan; sifar kelaparan; kehidupan bawah air

## **ACKNOWLEDGEMENTS**

I would like to express my sincere appreciation to Prof. Dr. Fatimah Md. Yusoff for her continuous support as well as her painstaking effort in improving my research work and preparation of the thesis. It was under her tutelage that I developed a focus to complete my thesis. Thanks to the other members of my supervisory committee, Assoc. Prof. Dr. Zetty Norhana Balia Yusof and Dr. Norulhuda Mohamed Ramli for their insightful comments and the assistance they provided at all levels along the way in accomplishing this study. Thanks to all my fellow laboratory mates, especially Siti Balqis, Fareha, Amirah Izyan, Laishatul Aini, and all staff of the AquaHealth, Institute of Bioscience, especially Mr. Mohd Shukri, Mr. Farhan, and Ms. Fadzilah for their assistance and for our exchanges of knowledge and skills during my study period, which helped to enrich my experiences. My appreciation also goes to the Ministry of Higher Education Malaysia through the Fundamental Research Grant Scheme (FRGS) and the SATREPS-COSMOS Project.

Most importantly, I would like to extend my gratitude to my family especially my parents for their unconditional support through my entire life and particularly in my studies. Without encouragement from all of them, I would not be what I am today. I doubt that I will ever be able to convey my appreciation fully, but I owe all of you my eternal gratitude.

Thank you.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Fatimah binti Md. Yusoff, PhD**

Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Chairman)

**Zetty Norhana binti Balia Yusoff, PhD**

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

**Norulhuda binti Mohamed Ramli, PhD**

Senior Lecturer

Faculty of Engineering

Universiti Putra Malaysia

(Member)

---

**ZALILAH BINTI MOHD SHARIFF, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 16 May 2024

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xv
<b>LIST OF FIGURES</b>	xvii
<b>LIST OF ABBREVIATIONS</b>	xxii
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	
1.1 Background of the study	1
1.2 Problem statements	2
1.3 Objectives	3
<b>2 LITERATURE REVIEW</b>	
2.1 Role of zooplankton in aquatic ecosystems	4
2.2 Distribution and diversity of zooplankton in Malaysia and tropical countries	5
2.3 Role of zooplankton as live food in aquaculture industry	14
2.4 Effects of environmental factors on zooplankton growth, reproductive performance, and nutritional composition	15
2.5 Effects of diets on zooplankton growth, reproductive performance, and nutritional contents	22
2.5.1 Microalgae	22
2.5.2 Bacteria/probiotics	23
2.5.3 Combination of microalgae and bacteria	29
<b>3 DISTRIBUTION AND DIVERSITY OF FRESHWATER ZOOPLANKTON IN MALAYSIA AND SELECTION OF POTENTIAL SPECIES FOR LIVE FOOD CULTIVATION</b>	
3.1 Introduction	32
3.2 Materials and methods	33
3.2.1 Study sites	33
3.2.2 In-situ environmental parameter measurement	33
3.2.3 Water and zooplankton sample collection	41
3.2.4 Sample preparation for nutrient and chlorophyll a analyses	41
3.2.4.1 Total nitrogen	41
3.2.4.2 Total phosphorus	41
3.2.4.3 Chlorophyll a	41
3.2.5 Zooplankton identification and enumeration	43
3.2.6 Molecular identification of selected zooplankton species	43

3.2.6.1	Collection, isolation, and culture of selected species	43
3.2.6.2	Degenerate primers design	44
3.2.6.3	Genomic DNA (gDNA) extraction	45
3.2.6.4	Polymerase chain reaction (PCR)	45
3.2.6.5	Cloning and sequencing	46
3.2.7	Statistical Analyses	46
3.3	Results	47
3.3.1	Zooplankton species composition, abundance, and diversity	47
3.3.2	Physicochemical parameters of study sites	57
3.3.3	Relationship between zooplankton and physicochemical parameters	57
3.3.4	Selection of zooplankton species	61
3.3.5	Molecular identification of selected zooplankton species	63
3.4	Discussion	64
3.5	Conclusions	68

#### **4 EFFECTS OF ENVIRONMENTAL FACTORS ON ZOOPLANKTON REPRODUCTIVE CAPACITIES, POPULATION GROWTH, AND NUTRITIONAL COMPOSITIONS**

4.1	Introduction	69
4.2	Materials and methods	70
4.2.1	Zooplankton stock culture	70
4.2.2	Microalgae stock culture	70
4.2.3	Effect of temperature on development and reproductive performance of cladocerans	70
4.2.4	Effect of temperature on population density and growth rate	71
4.2.5	Effect of pH on development and reproductive performance of cladocerans	72
4.2.6	Effect of pH on population density and growth rate	72
4.2.7	Effect of ammonia concentration on development and reproductive performance of cladocerans	73
4.2.8	Effect of ammonia concentration on population density and growth rate	73
4.2.9	Biochemical analyses	74
4.2.9.1	Protein	74
4.2.9.2	Lipid	75
4.2.9.3	Carbohydrate	75
4.2.9.4	Fatty acid	76
4.2.10	Statistical analyses	76
4.3	Results	77
4.3.1	Effect of temperature on development and reproductive capacities	77
4.3.2	Effect of temperature on population density and growth rate	79

4.3.3	Effect of temperature on carbohydrate, protein, and lipid composition	82
4.3.4	Effect of temperature on fatty acid composition	82
4.3.5	Effect of temperature on linoleic acid (LA), arachidonic acid (ARA), eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA) composition	84
4.3.6	Effect of pH on development and reproductive capacities	85
4.3.7	Effect of pH on population density and growth rate	89
4.3.8	Effect of pH on carbohydrate, protein, and lipid composition	91
4.3.9	Effect of pH on fatty acid composition	91
4.3.10	Effect of pH on linoleic acid (LA), arachidonic acid (ARA), eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA) composition	94
4.3.11	Effect of ammonia ( $\text{NH}_3\text{-N}$ ) concentrations on development and reproductive capacities	97
4.3.12	Effect of ammonia ( $\text{NH}_3\text{-N}$ ) concentrations on population density and growth rate	98
4.3.13	Effect of ammonia ( $\text{NH}_3\text{-N}$ ) concentrations on carbohydrate, protein, and lipid composition	99
4.3.14	Effect of ammonia ( $\text{NH}_3\text{-N}$ ) concentrations on fatty acid composition	103
4.3.15	Effect of ammonia ( $\text{NH}_3\text{-N}$ ) concentrations on linoleic acid (LA), arachidonic acid (ARA), eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA) composition	105
4.4	Discussion	108
4.5	Conclusions	111
<b>5</b>	<b>DIETARY EFFECTS OF MICROALGAE AND BACTERIA ON THE REPRODUCTION, GROWTH AND NUTRITIONAL CONTENTS OF ZOOPLANKTON</b>	
5.1	Introduction	112
5.2	Materials and methods	113
5.2.1	Zooplankton stock culture	113
5.2.2	Microalgae stock culture	113
5.2.3	Bacteria culture	114
5.2.4	Effects of diets (mono and mixed microalgae-bacteria diets) on development and reproductive performance of cladocerans	114
5.2.5	Effects of diets (mono and mixed microalgae-bacteria diets) on population density and growth rate	115

5.2.6	Biochemical analyses	117
5.2.6.1	Protein	117
5.2.6.2	Lipid	117
5.2.6.3	Carbohydrate	117
5.2.6.4	Fatty acid	117
5.2.7	Statistical analyses	117
5.3	Results	118
5.3.1	Population growth density and specific growth rate of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> and <i>Diaphanosoma excisum</i> fed with different diets	118
5.3.2	Reproductive performance of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> and <i>Diaphanosoma excisum</i> fed with different diets	121
5.3.2.1	Maturation	121
5.3.2.2	Number of eggs	121
5.3.2.3	Number of offsprings	123
5.3.2.4	Number of clutches	125
5.3.3	Life span and body size of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> and <i>Diaphanosoma excisum</i> fed with different diets	127
5.3.3.1	Life span	127
5.3.3.2	Body size	129
5.3.4	Carbohydrate, protein, and lipid contents of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> and <i>Diaphanosoma excisum</i> fed with different diets	129
5.3.4.1	Carbohydrate	129
5.3.4.2	Protein	132
5.3.4.3	Lipid	135
5.3.5	Fatty acid composition of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> and <i>Diaphanosoma excisum</i> fed with different diets	135
5.3.5.1	Saturated fatty acid (SAFA)	135
5.3.5.2	Monounsaturated fatty acid (MUFA)	137
5.3.5.3	Polyunsaturated fatty acid (PUFA)	140
5.3.6	Linoleic acid (LA), arachidonic acid (ARA), eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA) composition of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> and <i>Diaphanosoma excisum</i> fed with Different Diets	140
5.3.6.1	Linoleic acid (LA)	140
5.3.6.2	Arachidonic acid (ARA)	142
5.3.6.3	Eicosapentaenoic acid (EPA)	142
5.3.6.4	Docosahexaenoic acid (DHA)	145
5.3.7	Interaction between diets and biochemical composition of cladocerans	148
5.3.7.1	<i>Ceriodaphnia cornuta</i>	148
5.3.7.2	<i>Moina micrura</i>	149

5.3.7.3	<i>Diaphanosoma excisum</i>	151
4.4	Discussion	152
4.5	Conclusions	156
<b>6</b>	<b>GENERAL DISCUSSION AND RECOMMENDATIONS</b>	
6.1	General discussion	157
6.2	Recommendations	158
<b>REFERENCES</b>		160
<b>APPENDICES</b>		192
<b>BIODATA OF STUDENT</b>		195
<b>LIST OF PUBLICATIONS</b>		196

## LIST OF TABLES

Table	Page
2.1 Zooplankton composition and diversity in Malaysia and other tropical countries	8
2.2 Biochemical composition (%) of different zooplankton species	16
2.3 Fatty acid profile (%) of different zooplankton species	17
2.4 Growth and reproductive capacities of zooplankton at various temperature, pH, and ammonia levels	19
2.5 Peak population density and specific growth rate of zooplankton species fed with different microalgae	24
2.6 Reproductive capacities of zooplankton species fed with different microalgae	25
2.7 Body size and life span of zooplankton species fed with different microalgae	26
2.8 Nutritional composition of zooplankton species fed with different microalgae	27
2.9 Growth, reproductive capacities, and nutritional contents of zooplankton fed with mixed microalgae-bacteria and probiotics	31
3.1 Description of the sampling sites	35
3.2 Utilisation of available gene sequences from GenBank of the closely related species for molecular identification primers design	44
3.3 Mean densities (ind. L <sup>-1</sup> ) and percentages (%) of zooplankton species in natural and artificial lakes	50
3.4 Percentage contribution (%) of major zooplankton species (> 70 %) for each lake based on similarity percentage (SIMPER) analysis	49
3.5 Spatial of environmental parameters and nutrients (Mean ± SE) at the study sites. *na: data not available	58
3.6 Overall correlation metrics (non-parametric Spearman's rank order correlation) for environmental and zooplankton species at the study sites	59

3.7	Combination of variables that explain the zooplankton community in the study area, showing spearman rank correlation ( $\rho$ ) obtained by BIO-ENV analysis	60
3.8	Identification of zooplankton species based on 18S rRNA partial sequence analysis	63
4.1	Development and reproductive capacity of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> cultured at different temperature levels. Means with different letters are significantly different at $P < 0.05$	78
4.2	Development and reproductive capacity of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> cultured at different pH levels. Means with different letters are significantly different at $P < 0.05$	88
4.3	Development and reproductive capacity of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> cultured at different ammonia ( $\text{NH}_3\text{-N}$ ). Means with different letters are significantly different at $P < 0.05$	100
5.1	Feeding treatments on the reproductive performance, growth, and nutritional of composition of cladocerans	116
5.2	Eigenvector, eigenvalue and % variations derived from the principal component analysis (PCA) for determination of biochemical composition of <i>Ceriodaphnia cornuta</i> in different diets	148
5.3	Eigenvector, eigenvalue and % variations derived from the principal component analysis (PCA) for determination of biochemical composition of <i>Moina micrura</i> in different diets	150
5.4	Eigenvector, eigenvalue and % variations derived from the principal component analysis (PCA) for determination of biochemical composition of <i>Diaphanosoma excisum</i> in different diets	151

## LIST OF FIGURES

Figure		Page
3.1	Map of the sampling sites including eight natural lakes and 12 artificial lakes	34
3.2	Composition of zooplankton in natural and artificial lakes	47
3.3	Composition of zooplankton group in natural and artificial lakes	48
3.4	nMDS of zooplankton densities ( $\text{ind. L}^{-1}$ ) and diversity ( $H'$ ) illustrating that zooplankton densities and diversity were clustered in different groups corresponding to the lake types	54
3.5	Shade plot of zooplankton species densities ( $\text{ind. L}^{-1}$ ) in the sampling sites. White spaces denote absence of the species in that specific site; depth of colour scale is linearly proportional to a fourth-root transformation of density	55
3.6	Non-metric multidimensional scaling (nMDS) of major zooplankton species distribution in natural and artificial lakes. The size of the ball denotes the density in $\text{ind. L}^{-1}$	56
3.7	Bi-plots of the canonical correspondence analysis (CCA) for zooplankton species and physical and chemical parameters showing the distribution of zooplankton species in relation to environmental conditions in different sites. DO = dissolved oxygen, transparency = water transparency, TN = total nitrogen, TP = total phosphorus, Chl. a = chlorophyll a	60
3.8	Overall composition of cladoceran species	61
3.9	Composition of cladoceran species each lake	61
3.10	Three cladocerans species that were selected as the potential live food	62
3.11	Gradient PCR amplification of 18s rRNA from <i>Moina</i> and <i>Ceriodaphnia</i> cultures genomic DNA. Lane 1 – 4 represents amplification of 18s rRNA from <i>Moina</i> sp. DNA at 63°C, 61°C, 58°C and 56°C. Lane 5 – 8 represents amplification of 18s rRNA from <i>Ceriodaphnia</i> sp. DNA at 63°C, 61°C, 58°C and 56°C	63

4.1	Population growth density of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> and c) <i>Diaphanosoma excisum</i> at different temperature levels	80
4.2	Population growth rates of a) <i>Ceriodaphnia cornuta</i> , a) <i>Moina micrura</i> and c) <i>Diaphanosoma excisum</i> cultured at different temperature levels. Means with different letters are significantly different at $P < 0.05$	81
4.3	a) Carbohydrate, b) protein, and c) lipid composition of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphasonoma excisum</i> cultured at different temperature levels. Means with different letters are significantly different at $P < 0.05$	83
4.4	Composition of a) saturated fatty acid (SAFA), b) monounsaturated fatty acid (MUFA), and c) polyunsaturated fatty acid (PUFA) <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphasonoma excisum</i> cultured at different temperature levels. Means with different letters are significantly different at $P < 0.05$	86
4.5	Composition of a) linoleic acid (LA), b) arachidonic acid (ARA), c) eicosapentaenoic acid (EPA), and d) docosahexaenoic acid (DHA) (% of total fatty acids) of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphanosoma excisum</i> cultured at different temperature levels. Means with different letters are significantly different at $P < 0.05$	87
4.6	Population growth density of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> and c) <i>Diaphanosoma excisum</i> at different pH levels	90
4.7	Population growth rate of a) <i>Ceriodaphnia cornuta</i> , a) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> cultured at different pH. Means with different letters are significantly different at $P < 0.05$	92
4.8	Figure 4.8: a) Carbohydrate, b) protein, and c) lipid composition of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphasonoma excisum</i> cultured at different pH levels. Means with different letters are significantly different at $P < 0.05$	93
4.9	Composition of a) saturated fatty acid (SAFA), b) monounsaturated fatty acid (MUFA), and c) polyunsaturated fatty acid (PUFA) of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphanosoma excisum</i> cultured at different pH levels. Means with different letters are significantly different at $P < 0.05$	95

4.10	Composition of a) linoleic acid (LA), b) arachidonic acid (ARA), c) eicosapentaenoic acid (EPA), and d) docosahexaenoic acid (DHA) of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphanosoma excisum</i> cultured at different pH levels. Means with different letters are significantly different at $P < 0.05$	96
4.11	Population growth density of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> and c) <i>Diaphanosoma excisum</i> at different ammonia ( $\text{NH}_3\text{-N}$ ) concentrations	101
4.12	Population growth rate of a) <i>Ceriodaphnia cornuta</i> , a) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> cultured at different ammonia ( $\text{NH}_3\text{-N}$ ) concentrations. Means with different letters are significantly different at $P < 0.05$	102
4.13	a) Carbohydrate, b) protein, and c) lipid composition (%) of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphasonoma excisum</i> cultured at different ammonia ( $\text{NH}_3\text{-N}$ ) concentrations. Means with different letters are significantly different at $P < 0.05$	104
4.14	Composition of a) saturated fatty acid (SAFA), b) monounsaturated fatty acid (MUFA), and c) polyunsaturated fatty acid (PUFA) of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphasonoma excisum</i> cultured at different ammonia ( $\text{NH}_3\text{-N}$ ) concentrations. Means with different letters are significantly different at $P < 0.05$	106
4.15	Composition of a) linoleic acid (LA), b) arachidonic acid (ARA), c) eicosapentaenoic acid (EPA), and d) docosahexaenoic acid (DHA) of <i>Ceriodaphnia cornuta</i> , <i>Moina micrura</i> , and <i>Diaphanosoma excisum</i> cultured at different ammonia ( $\text{NH}_3\text{-N}$ ) concentrations. Means with different letters are significantly different at $P < 0.05$	107
5.1	Population growth density of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> and c) <i>Diaphanosoma excisum</i> in different diets	119
5.2	Population growth rate of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> and c) <i>Diaphanosoma excisum</i> in different diets	120
5.3	Maturation of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	122

5.4	Number of eggs of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	124
5.5	Number of offspring of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	126
5.6	Number of clutches of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	128
5.7	Life span of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	130
5.8	Body size of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	131
5.9	Carbohydrate content (%) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	133
5.10	Protein content (%) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	134
5.11	Lipid content (%) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	136
5.12	Saturated fatty acids (SAFA) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	138
5.13	Monounsaturated fatty acids (MUFA) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	139

5.14	Polyunsaturated fatty acids (PUFA) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	141
5.15	Composition of inoleic acid (LA) (% of total fatty acids) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	143
5.16	Composition of arachidonic acid (ARA) (% of total fatty acids) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$ . ND: Not detected	144
5.17	Composition of eicosapentaenoic acid (EPA) (% of total fatty acids) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$	146
5.18	Composition of docosahexaenoic acid (DHA) (% of total fatty acids) of a) <i>Ceriodaphnia cornuta</i> , b) <i>Moina micrura</i> , and c) <i>Diaphanosoma excisum</i> fed with different diets. Means with different letters are significantly different at $P < 0.05$ . ND: Not detected	147
5.19	Principal Component Analysis (PCA) ordination of nutritional composition parameters that best described the <i>Ceriodaphnia cornuta</i> fed with different diets	149
5.20	Principal Component Analysis (PCA) ordination of nutritional composition parameters that best described the <i>Moina micrura</i> fed with different diets	150
5.21	Principal Component Analysis (PCA) ordination of nutritional composition parameters that best described the <i>Diaphanosoma excisum</i> fed with different diets	152

## LIST OF ABBREVIATIONS

%	Percentage
(NH <sub>4</sub> ) <sub>6</sub> Mo <sub>7</sub> O <sub>24</sub>	Ammonium molybdate
<	Less than
>	More than
µg L <sup>-1</sup>	Microgram per liter
µL	Microliter
µm	Micrometer
ALA	Linolenic acid
ANOVA	Analysis of variance
AOAC	Association of Official Analytical Chemists
APHA	American Public Health Association
ARA	Arachidonic acid
BBM	Bold's Basal Medium
BIO-ENV	Biotic-environmental analysis
bp	Base pair
°C	Degree Celsius
C <sub>4</sub> H <sub>4</sub> O <sub>7</sub> SbK	Antimony potassium tartarate
CCA	Canonical correspondence analysis
CFU	Colony forming unit
Chl a	Chlorophyll a
CTSI	Carlson Trophic Status Index
CuSO <sub>4</sub> 5H <sub>2</sub> O	Copper sulfate pentahydrate
DHA	Docosahexaenoic acid
DNA	Deoxyribonucleic acid

DO	Dissolved oxygen
EPA	Eicosapentaenoic acid
g	Gram
h	Hour
H'	Shannon–Wiener diversity index
H <sub>2</sub> SO <sub>4</sub>	Sulfuric acid
HABs	Harmful algal blooms
HCl	Hydrochloric acid
ind. L	Individual per liter
K <sub>2</sub> S <sub>2</sub> O <sub>8</sub>	Potassium peroxobisulfate
kg	Kilogram
KH <sub>2</sub> PO <sub>4</sub>	Potassium dihydrogen phosphate
KNO <sub>3</sub>	Potassium nitrate
L	Liter
LA	Linoleic acid
m	Meter
mg	Milligram
mg L <sup>-1</sup>	Milligram per liter
mL	Milliliter
mm	Millimeter
MUFA	Monounsaturated fatty acid
N <sub>2</sub> H <sub>6</sub> SO <sub>4</sub>	Hydrazine sulfate
NaOH	Sodium hydroxide
NED	N-(1-naphthyl) ethylenediamine dihydrochloride
NH <sub>2</sub> C <sub>6</sub> H <sub>4</sub> SO <sub>2</sub>	Sulfanilamide
NH <sub>3</sub> -N	Ammonia nitrogen

NH <sub>4</sub> Cl	Ammonium chloride
nm	Nanometer
nMDS	Non-metric multidimensional scaling
NTU	Nephelometric turbidity unit
OD	Optical density
PCA	Principle component analysis
PCR	polymerase chain reaction
PRIMER	Plymouth Routines in Multivariate Ecological Research
PUFA	polyunsaturated fatty acid
<i>r</i>	Correlation
rpm	Revolutions per minute
rRNA	Ribosomal ribonucleic acid
SAFA	Saturated fatty acid
SD	Standard division
SGR	Specific growth rate
SIMPER	Similarity percentage analysis
Sp.	Species
TAN	Total ammonia nitrogen
TN	Total nitrogen
TP	Total phosphorus
TSB	Tryptic soy broth
UPM	Universiti Putra Malaysia
v/v	Volume/volume
ZnSO <sub>4</sub>	Zinc sulfate heptahydrate

# CHAPTER 1

## INTRODUCTION

### 1.1 Background of the study

Zooplankton are microscopic aquatic organisms which respond quickly to many environmental stressors, such as hydrological changes and anthropogenic activity-induced water pollution. Zooplankton abundance and diversity are affected by abiotic and biotic factors. They represent the channel of transmission of energy flux and organic matter from the primary producers to the top consumers in aquatic ecosystems (Bakhtiyar et al., 2020). In addition, zooplankton are suitable for culturing as live food because they can reach a high population size rapidly which can assure a sustainable supply for aquaculture (Rasdi et al., 2020a; Chakraborty and Mallick, 2023). Zooplankton is also recognized as a superior live food based on its nutritional composition which may match the dietary requirements of aquaculture animals, particularly fish larvae, to foster their growth and reproduction (Samat et al., 2020b).

Zooplankton distribution, growth, reproductive capacities, and nutritional values are affected by environmental parameters and food sources. Temperature, pH, dissolved oxygen (DO), and ammonia are among the important parameters that have a substantial impact on zooplankton (Liang et al., 2020a; Qin et al., 2021; Xue et al., 2022). In general, warmer temperatures increase the metabolic rate of zooplankton, leading to higher growth, reproduction, and feeding rates (Garzke et al., 2020). However, decreased in monounsaturated fatty acid (MUFA) and polyunsaturated fatty acid (PUFA) was observed when *Brachionus calyciflorus* exposed to increasing temperature (30 °C). Water pH affects the regular physiological processes of aquatic animals, such as the exchange of ions with water and respiration (Tresguerres et al., 2023). Acidic pH exposure, whether chronic or acute, can affect zooplankton metabolic activity, egg production, population growth, and survival (Lee et al., 2020; Gao et al., 2022). Ammonia is the primary nitrogenous product of the protein catabolism. Total ammonia-nitrogen consists of toxic un-ionized NH<sub>3</sub>-N and nontoxic ionized NH<sub>4</sub><sup>+</sup>-N, and their equilibrium is mainly dependent on temperature and pH. High levels of NH<sub>3</sub>-N can disrupt the normal metabolism of aquatic organisms, resulting in growth retardation and inhibition of zooplankton metamorphosis (Han et al., 2022).

Of all the aforementioned factors, the food source is the most critical factor. Differences in fatty acid content among zooplankton taxa could affect their demands for essential fatty acids from their diet. Fatty acids especially polyunsaturated fatty acids (PUFAs) composition is important for the growth of zooplankton as they function as precursors for hormones involved in gonad development, reproduction, and immune responses (Ilić et al., 2019). Since Omega-3 and Omega-6 fatty acids cannot be synthesized de novo by

zooplankton, they must be included in the diet, either as eicosapentaenoic acid (EPA), docosahexaenoic acid (DHA) and arachidonic acid (ARA) or as their precursors, such as  $\alpha$ -linolenic acid (ALA: precursor of EPA and DHA) and linoleic acid (LIN: precursor of ARA). Therefore, these essential fatty acids must be supplied from diets to support zooplankton for the need for zooplankton growth. In nature, zooplankton are fed on a wide range of microalgal species and bacteria. Being the primary producers, microalgae are the zooplankton's main food source, which ultimately helps to enrich zooplanktonic organisms by giving protein, energy, vitamins, minerals, and amino acids (Altaff and Vijayaraj, 2021). At the same time, microalgae and bacteria also develop interactions over time. Several investigations have found that heterotrophic bacteria are important for microalgal growth and survival (Samo et al., 2018; Sandhya and Vijayan, 2019). Indirectly, these symbiotic microalgae–bacteria consortia with valuable chemical and energy compounds could be utilized to enrich the zooplankton or optimizing zooplankton growth and quality. In recent years, the enrichment of zooplankton as live feeds with high-quality microalgae and potential bacteria either by mono diets or mixed diets has become a popular approach to boost zooplankton production (Wenzel et al., 2021; Samat et al., 2020a).

Among freshwater zooplankton, cladocerans such as *Ceriodaphnia*, *Moina*, and *Diaphanosoma* have been explored as potential live feed candidates (Chilmawati et al., 2019; Neri et al., 2020; Suhaimi et al., 2022). Cladocerans reproduce by cyclic parthenogenesis, which involves alternation between the asexual and sexual phases as part of their reproduction. Females exhibit parthenogenesis during the asexual phase, in which they can self-replicate without fertilization, reproduce offspring that are all females, and reach a high population in a short period. In addition, cladoceran's ability to produce ephippia (cysts/resting eggs) during the sexual phase is also one of the reasons to utilize them as live feed candidates. For culturing purposes, cladocerans display a high potential for large-scale production because of their high growth rate. Apart from elucidating the capabilities of cladocerans in high-density cultures, a better understanding of the factors influencing the production of high-quality zooplankton must be acquired improve the productivity of live feed in aquaculture.

## 1.2 Problem statements

Aquaculture production in Malaysia including freshwater, brackish, and marine aquaculture contributing significantly to the country's food security and economic growth. However, in the aquaculture industry, artificial feed is not always sufficient to fulfill the growth requirements of larvae compared to live food. Mass rearing of healthy fish larvae is a key step in the aquaculture production processes. The rearing of more sensitive freshwater and marine fish larvae depends heavily on live food, particularly zooplankton. During the hatchery and nursery phases, fish larvae undergo rapid growth and development in the early stages of their life cycle. During this time, their mouthparts (mouth sizes) undergo changes in size and structure. Larvae with smaller mouths may not be able to consume larger zooplankton, leading to poor nutrition and growth. Conversely,

larvae with larger mouths may struggle to capture and ingest smaller zooplankton efficiently. They can only ingest specific live feed with a specific size. For example, the ideal size of live feed is 50–90 µm for silver carp larvae, 90–150 µm for grass carp larvae and 150–270 µm for bighead carp larvae (Dabrowski and Bardega, 1984). Matching the size of the live food to the developmental stage of the larvae helps ensure efficient feeding, optimal growth, and overall success in raising healthy fish fry for stocking or further grow-out (Lahnsteiner et al., 2023). At the moment, *Artemia* is the most preferred and reliable live food, however, the size of *Artemia* not suitable for all fish larvae developmental stages. Hence, different species of zooplankton with different size should be utilized. Nowadays, the mass production of high-quality zooplankton as live feed is a bottleneck for sustainable aquaculture production. A poor understanding of their optimum requirements renders large-scale zooplankton production. Previous studies have revealed that the successful mass culture production of zooplankton is immensely related to environmental conditions and food quality, which are prerequisites for growth and reproduction (Ogello et al., 2019; Hegab et al., 2020; Ramlee et al., 2021). In addition, to obtain high-quality live food, they should be fed with high nutritional food sources that influence the supply of various nutritional contents to zooplankton. The production of zooplankton depends on the biochemical composition of the microalgae and bacteria fed to them. However, mono-specific diets may cause nutritional deficiencies because of the inadequate content of one or more essential nutrients. To reduce this risk, several authors have suggested the use of mixed diets because their combined nutrient contents are more likely to meet the nutritional requirements of the target species (Rasdi et al., 2020c; Contreras-Tapia et al., 2020; Fouzi et al., 2021; Azani et al., 2022; Yuslan et al., 2022). From this study, potential zooplankton species were identified as candidates for live feed. Additionally, the optimal environmental conditions and diet necessary for their growth and reproduction were established. The findings from this study can be used to develop an economical and practical technique for high-quality live feed, which would be valuable to the aquaculture industry.

### 1.3 Objectives

The utilization of freshwater zooplankton as live feed is worth considering to meet the demand of the aquaculture sector. Hence, optimum conditions, especially environmental variables and food sources are crucial for high-quality zooplankton production. Therefore, the objectives of this study were:

1. To assess freshwater zooplankton community structure (species composition, distribution, and diversity) in natural and artificial lakes with different levels of eutrophication and to identify prospective candidates for live food production.
2. To investigate the effect of environmental factors (temperature, pH, and ammonia) on the reproductive performance, growth, and nutritional contents of the selected zooplankton species
3. To evaluate the dietary effect (mono-diets and microalgae-bacterial combined diets) on zooplankton growth performance and nutritional contents.

## REFERENCES

- A Rahman, A. R. A., Sinang, S. C. & Nayan, N. (2022). Response of algal biomass and macrophyte communities to internal or external nutrient loading. *Environmental Monitoring and Assessment*, 194(7), 491.
- Abbas, M., Dia, S., Deutsch, E. S. & Alameddine, I. (2023). Analyzing eutrophication and harmful algal bloom dynamics in a deep Mediterranean hypereutrophic reservoir. *Environmental Science and Pollution Research*, 30(13), 37607-37621.
- Abbasi, Y., Ahmadifard, N. & Tukmechi, A. (2019). Effect of probiotic *Pediococcus acidilactici* on growth, reproductive and bacterial count of marine rotifer *Brachionus plicatilis*. *International Journal of Aquatic Biology*, 7(1), 27-34.
- Abd. Razak, S.B. & Sharip, Z (2019). Spatio-temporal variation of zooplankton community structure in tropical urban waterbodies along trophic and urban gradients. *Ecological Process*, 8, 44.
- Abou-Shanab, R. A., Singh, M., Rivera-Cruz, A., Power, G., Bagby-Moon, T. & Das, K. (2016). Effect of *Brachionus rubens* on the growth characteristics of various species of microalgae. *Electronic Journal of Biotechnology*, 22, 68-74.
- Alekseev, V. R., Haffner, D. G., Vaillant, J. J. and Yusoff, F. M. (2013). Cyclopoid and calanoid copepod biodiversity in Indonesia. *Journal of Limnology*, 72(2s), 12.
- Shakri, A., Yusoff F. M, Ismail, I. S. & Toda, T. (2020). Reduced reproductive capacity in *Moina micrura* Kurz, 1875 exposed to toxic *Microcystis* spp. *Asian Fisheries Science*, 33(1), 42 - 49.
- Aditya, L., Mahlia, T. I., Nguyen, L. N., Vu, H. P. & Nghiem, L. D. (2022). Microalgae-bacteria consortium for wastewater treatment and biomass production. *Science of the total environment*, 838, 155871.
- Agha, R., Saebelfeld, M., Manthey, C., Rohrlack, T. & Wolinska, J. (2016). Chytrid parasitism facilitates trophic transfer between bloom-forming cyanobacteria and zooplankton (*Daphnia*). *Scientific Reports*, 6(1), 35039.
- Ahmadifard, N., Rezaei Aminlooi, V., Tukmechi, A. & Agh, N. (2019). Evaluation of the impacts of long-term enriched *Artemia* with *Bacillus subtilis* on growth performance, reproduction, intestinal microflora, and resistance to *Aeromonas hydrophila* of ornamental fish *Poecilia latipinna*. *Probiotics and antimicrobial Proteins*, 11, 957-965.

- Aiman, W. M., Yusoff, F. M., Arshad, A., Kamal, A. H. M., Ismail, J., Idris, M. H., Karim N. U. & Al Asif, A. (2020). Distribution of zooplankton community in Toli shad (*Tenualoosa toli*) habitats, Sarawak, Malaysia. *Biodiversitas Journal of Biological Diversity*, 21(9), 4022 – 4033.
- Ajepe, R. G., Hammed, A. M., Amosu, A. O. & Fashina-Bombata, H. A. (2014). Comparative study of *Artemia* (Brine Shrimp) and *Ceriodaphnia* (Zooplankton) as foods for catfish larvae. *American Journal of Experimental Agriculture*, 4(7), 857 – 865.
- Almeida, R., Formigo, N. E., Sousa-Pinto, I. & Antunes, S. C. (2020). Contribution of zooplankton as a biological element in the assessment of reservoir water quality. *Limnetica*, 39(1), 245-261.
- Altaff, K. & Janakiraman, A. (2015). Effect of temperature on mass culture of three species of zooplankton, *Brachionus plicatilis*, *Ceriodaphnia reticulata* and *Apocyclops dengizicus*. *International Journal of Fisheries and Aquatic Studies*, 2(4), 49-53.
- Altaff, K. & Vijayaraj, R. (2021). Micro-algal diet for copepod culture with reference to their nutritive value—A Review. *International Journal of Current Research and Review*, 13(7), 86 – 96.
- Amenyogbe, E. (2023). Application of probiotics for sustainable and environment-friendly aquaculture management-A review. *Cogent Food & Agriculture*, 9(1), 2226425.
- Amorim, C. A. & do Nascimento Moura, A. (2021). Ecological impacts of freshwater algal blooms on water quality, plankton biodiversity, structure, and ecosystem functioning. *Science of the Total Environment*, 758, 143605.
- Aravind, R., Anand, P. S., Vinay, T. N., Biju, I. F., Sandeep, K. P., Raymond, J. A. J. & Vijayan, K. K. (2021). Population growth and mass production of brackish water cladoceran *Eury cercus beringi* sp. nov. under different diet and salinity regime, and its role in *P. indicus* larval rearing. *Regional Studies in Marine Science*, 44, 101777.
- Aquino, M. R. Y., Cho, C. D., Cruz, M. A. S., Saguiguit, M. A. G., & Papa, R. D. S. (2008). Zooplankton composition and diversity in Paoay Lake, Luzon Is., Philippines. *Philippine Journal of Science*, 137, 169-177.
- Arias, M. J., Vaschetto, P. A., Marchese, M., Regaldo, L. & Gagneten, A. M. (2022). Benthic macroinvertebrates and zooplankton communities as ecological indicators in urban wetlands of Argentina. *Sustainability*, 14(7), 4045.
- Atmaja, D. M., Budiastuti, M. S. & Setyono, P. (2021). The distributional pattern of zooplankton community in Beratan Lake, Bali. *IOP Conference Series: Earth and Environmental Science*, 683, 012002.

- Ayele, H. S. & Atlabachew, M. (2021). Review of characterization, factors, impacts, and solutions of lake eutrophication: Lesson for lake Tana, Ethiopia. *Environmental Science and Pollution Research*, 28(12), 14233-14252.
- Ayón, P. & Hirche, H. J. (2021). Zooplankton of the low-oxygen waters of Bahia Callao (Central Peru)-with special reference to the reproductive activity of the copepod *Acartia tonsa*. *Estuarine, Coastal and Shelf Science*, 262, 107572.
- Azani, N., Ambok Bolong, A. M., Liew, H. J., Kamarudin, S., Arshad, A., Hassan, M. M. & Rasdi, N. W. (2022). Different dietary effects on growth and reproduction of freshwater zooplankton *Ceriodaphnia cornuta* (Sars, 1885) and its potential use in *Pangasius nasutus* larval rearing. *International Aquatic Research*.
- Bakhtiyar, Y., Arafat, M. Y., Andrabí, S. & Tak, H. I. (2020). Zooplankton: The significant ecosystem service provider in aquatic environment. *Bioremediation and Biotechnology, Vol 3: Persistent and Recalcitrant Toxic Substances*, 3, 227-244.
- Bakr, A. R., Fu, G. Y. & Hedeen, D. (2020). Water quality impacts of bridge stormwater runoff from scupper drains on receiving waters: A review. *Science of the Total Environment*, 726, 138068.
- Balachandar, S. & Rajaram, R. (2019). Influence of different diets on the growth, survival, fecundity and proximate composition of brine shrimp *Artemia franciscana* (Kellog, 1906). *Aquaculture research*, 50(2), 376-389.
- Ballesteros-Redondo, L., Palm, H. W., Bährs, H., Wacker, A. & Bischoff, A. A. (2023) Pikeperch larviculture (*Sander lucioperca* [L., 1758]) with *Brachionus plicatilis* (Mueller, 1786) (Rotifera) and *Apocyclops panamensis* (Marsh, 1913) (Copepoda). *Journal of the World Aquaculture Society*, 54 (4), 1026-1039.
- Benayache, N. Y., Nguyen-Quang, T., Hushchyna, K., McLellan, K., Afri-Mehennaoui, F. Z. & Bouaïcha, N. (2019). An overview of cyanobacteria harmful algal bloom (CyanoHAB) issues in freshwater ecosystems. *Limnology: Some new aspects of inland water ecology*, 13-37.
- Beyene, G., Kifle, D. & Fetahi, T. (2022). Spatial distribution of zooplankton in relation to some selected physicochemical water quality parameters of Lake Hawassa, Ethiopia. *African Journal of Aquatic Science*, 47(2), 163-172.
- Bhatia, S. K., Ahuja, V., Chandel, N., Mehariya, S., Kumar, P., Vinayak, V. & Yang, Y. H. (2022). An overview on microalgal-bacterial granular consortia for resource recovery and wastewater treatment. *Bioresource technology*, 351, 127028.

- Bischoff, A. A., Kubitz, M., Wranik, C. M., Ballesteros-Redondo, L., Fink, P. & Palm, H. W. (2022). The effect of *Brachionus calyciflorus* (Rotifera) on larviculture and fatty acid composition of Pikeperch (*Sander lucioperca* (L.)) cultured under pseudo-green water conditions. *Sustainability*, 14(11), 6607.
- Bizani, M., Bornman, T. G., Campbell, E. E., Perissinotto, R. & Deyzel, S. H. P. (2023). Mesozooplankton community responses to a large-scale harmful algal bloom induced by the non-indigenous dinoflagellate *Lingulodinium polyedra*. *Science of The Total Environment*, 860, 161030.
- Bomfim, F. F., Braghin, L. S., Bonecker, C. C. & Lansac-Tôha, F. A. (2018). High food availability linked to dominance of small zooplankton in a subtropical floodplain. *International Review of Hydrobiology*, 103(1-2), 26-34.
- Bomfim, F. F., Melao, M. G., Gebara, R. C. & Lansac-Tôha, F. A. (2022). Warming alters the metabolic rates and life-history parameters of *Ceriodaphnia silvestrii* (Cladocera). *Anais da Academia Brasileira de Ciências*, 94(2), 1-14.
- Bonecker, C. C., Diniz, L. P., Braghin, L. D. S. M., Mantovano, T., da Silva, J. V. F., de Fátima Bomfim, F. & Lansac-Tôha, F. A. (2020). Synergistic effects of natural and anthropogenic impacts on zooplankton diversity in a subtropical floodplain: a long-term study. *Oecologia Australis*, 24(2), 524-537.
- Boonmak, P. & Sanoamuang, L. (2022). Diversity of freshwater calanoid copepods (Crustacea: Copepoda: Calanoida) in southern Vietnam with an updated checklist for the country. *Diversity*, 14(7), 523.
- Brito, M. T. D. S., Heino, J., Pozzobom, U. M. & Landeiro, V. L. (2020). Ecological uniqueness and species richness of zooplankton in subtropical floodplain lakes. *Aquatic Sciences*, 82(2), 43.
- Burbano, M. F., Torres, G. A., Prieto, M. J., Gamboa, J. H. & Chapman, F. A. (2020). Increased survival of larval spotted rose snapper *Lutjanus guttatus* (Steindachner, 1869) when fed with the copepod *Cyclopina* sp. and *Artemia* nauplii. *Aquaculture*, 519, 734912.
- Butt, U. D., Lin, N., Akhter, N., Siddiqui, T., Li, S. & Wu, B. (2021). Overview of the latest developments in the role of probiotics, prebiotics and synbiotics in shrimp aquaculture. *Fish & Shellfish Immunology*, 114, 263-281.
- Cabral, C. R., Diniz, L. P., da Silva, A. J., Fonseca, G., Carneiro, L. S., de Melo Júnior, M. & Caliman, A. (2020). Zooplankton species distribution, richness and composition across tropical shallow lakes: A large scale assessment by biome, lake origin, and lake habitat. *Annales de Limnologie-International Journal of Limnology*, 56 (25), 1-22.

- Camus, T., Rolla, L., Jiang, J. & Zeng, C. (2021). Effects of microalgal food quantity on several productivity-related parameters of the calanoid copepod *Bestiolina similis* (Calanoida: Paracalanidae). *Frontiers in Marine Science*, 8, 812240.
- Casas-Rodriguez, A., Cameán, A. M. & Jos, A. (2022). Potential endocrine disruption of cyanobacterial toxins, microcystins and cylindrospermopsin: A review. *Toxins*, 14(12), 882.
- Castro, J. M., Castro, G. M., Dávila, F. S. & Castro, A. E. C. (2017). Density comparison of *Moina macrocopa* (Straus, 1820) cultured at different temperature conditions (19, 23 and 25 C) fed with bacteria obtained from Biofloc system. *Journal of Entomology and Zoology Studies*, 5(6), 2433-2437.
- Celewicz-Gołdyn, S. & Kuczyńska-Kippen, N. (2017). Ecological value of macrophyte cover in creating habitat for microalgae (diatoms) and zooplankton (rotifers and crustaceans) in small field and forest water bodies. *PLoS One*, 12(5), e0177317.
- Chai, W. S., Chew, C. H., Munawaroh, H. S. H., Ashokkumar, V., Cheng, C. K., Park, Y. K. & Show, P. L. (2021). Microalgae and ammonia: a review on inter-relationship. *Fuel*, 303, 121303.
- Chakraborty, S. & Mallick, P. H. (2023). Cladocera as a substitute for Artemia as live feed in aquaculture practices: A review. *Sustainability, Agri, Food and Environmental Research*, 11.
- Chang, K. C., Wen, J. D. & Yang, L. W. (2015). Functional importance of mobile ribosomal proteins. *BioMed research international*, 2015. 539238
- Chen, R., Xu, N., Zhao, F., Wu, Y., Huang, Y. & Yang, Z. (2015). Temperature-dependent effect of food size on the reproductive performances of the small-sized cladoceran *Moina micrura*. *Biochemical Systematics and Ecology*, 59, 297-301.
- Chilmawati, D., Susilowati, T. & Adhinugroho, I. (2019). The effects of microalgal diet with enrichment of fermented organic matters (tofu waste, rice bran and fish meal) on growth and reproduction of *Diaphanosoma brachyurum*. *IOP Conference Series: Earth and Environmental Science*, 246(1), 012036.
- Chizhayeva, A., Amangeldi, A., Oleinikova, Y., Alybaeva, A. & Sadanov, A. (2022). Lactic acid bacteria as probiotics in sustainable development of aquaculture. *Aquatic Living Resources*, 35, 10.
- Choi, J. Y. & Kim, S. K. (2020). Responses of rotifer community to microhabitat changes caused by summer-concentrated rainfall in a shallow reservoir, South Korea. *Diversity*, 12(3), 113.

- Choueri, R. B., Melão, M. D. G. G., Lombardi, A. T. & Vieira, A. A. (2007). Effects of cyanobacterium exopolysaccharides on life-history of *Ceriodaphnia cornuta* SARS. *Journal of plankton research*, 29(4), 339-345.
- Coelho, P. N. & Henry, R. (2021). Functional groups of microcrustaceans along a horizontal gradient in a neotropical lake colonized by macrophytes. *Aquatic Sciences*, 83, 1-13.
- Contreras-Tapia, R. A., Castellanos-Páez, M. E., Nandini, S., Castillo-Rivera, M., Benítez-Díaz-Mirón, M. I. & Garza-Mouriño, G. (2020). Effect of commercial probiotic on the population dynamics of selected rotifers (*Brachionus angularis*, *Platynus patulus*) and cladocerans (*Moina cf. macrocopa* and *Simocephalus mixtus*). *Aquaculture Research*, 51(11), 4482-4494.
- Coone, M., Vanoverberghe, I., Houwenhuyse, S., Verslype, C. & Decaestecker, E. (2023). The effect of hypoxia on *Daphnia magna* performance and its associated microbial and bacterioplankton community: A scope for phenotypic plasticity and microbiome community interactions upon environmental stress?. *Frontiers in Ecology and Evolution*, 11, 1131203.
- Dabrowski, K., & Bardega, R. (1984). Mouth size and predicted food size preferences of larvae of three cyprinid fish species. *Aquaculture*, 40, 41-46.
- Damayanti, K. Y., Mubarak, A. S. & Sari, L. A. (2020). The effect of giving fermented rice bran suspension on fecundity and production of *Moina macrocopa* offspring per parent. *IOP Conference Series: Earth and Environmental Science*, 441(1), 012156.
- das Candeias, D. A., Moi, D. A., Simões, N. R., Azevedo, F., Meerhoff, M. & Bonecker, C. C. (2022). High temperature, predation, nutrient, and food quality drive dominance of small-sized zooplankton in Neotropical lakes. *Aquatic Sciences*, 84(4), 49.
- Dayras, P., Bialais, C., Sadovskaya, I., Lee, M. C., Lee, J. S. & Souissi, S. (2021). Microalgal diet influences the nutritive quality and reproductive investment of the cyclopoid copepod paracyclops nana. *Frontiers in Marine Science*, 8, 697561.
- de Alkmin, G. D., Nunes, B., Soares, A. M., Bellot, M., Gómez-Canela, C. & Barata, C. (2020). *Daphnia magna* responses to fish kairomone and chlorpromazine exposures. *Chemico-Biological Interactions*, 325, 109123.
- de Aquino Santos, A. S., Vilar, M. C. P., Amorim, C. A., Molica, R. J. R. & do Nascimento Moura, A. (2022). Exposure to toxic *Microcystis* via intact cell ingestion and cell crude extract differently affects small-bodied cladocerans. *Environmental Science and Pollution Research*, 29(16), 23194-23205.

- Deng, W., Sun, J., Chang, Z. G., Gou, N. N., Wu, W. Y., Luo, X. L. & Ji, H. (2020). Energy response and fatty acid metabolism in *Onychostoma macrolepis* exposed to low-temperature stress. *Journal of Thermal Biology*, 94, 102725.
- Derevenskaia, O. I., Borisova, N. I. & Unkovskaia, E. N. (2021). Zooplankton indices in the evaluation of the ecological state of the eutrophic lake (Case study: Karasikha Lake, Russia). *Caspian Journal of Environmental Sciences*, 19(4), 701-708.
- Derevenskaya, O. Y., Prytkova, E. S. & Unkovskaya, E. N. (2019). Zooplankton as water quality indicator in shallow lakes. *Journal of Computational and Theoretical Nanoscience*, 16(11), 4486-4490.
- Deubel, A., Gransee, A. & Merbach, W. (2000). Transformation of organic rhizodepositions by rhizosphere bacteria and its influence on the availability of tertiary calcium phosphate. *Journal of Plant Nutrition and Soil Science*, 163(4), 387-392.
- Dillon, R. A., Conroy, J. D., Lang, K. J., Pangle, K. L. & Ludsin, S. A. (2021). Bottom hypoxia alters the spatial distribution of pelagic intermediate consumers and their prey. *Canadian Journal of Fisheries and Aquatic Sciences*, 78(5), 522-538.
- Domingues, C. D., da Silva, L. H. S., Rangel, L. M., de Magalhães, L., de Melo Rocha, A., Lobão, L. M. & Sarmento, H. (2017). Microbial food-web drivers in tropical reservoirs. *Microbial Ecology*, 73, 505-520.
- Doubek, J. P., Campbell, K. L., Doubek, K. M., Hamre, K. D., Lofton, M. E., McClure, R. P. & Carey, C. C. (2018). The effects of hypolimnetic anoxia on the diel vertical migration of freshwater crustacean zooplankton. *Ecosphere*, 9(7), e02332.
- Dubois, M., Gilles, K. A., Hamilton, J. K., Rebers, P. A. & Smith, F. (1956). Colorimetric method for determination of sugars and related substances. *Analytical Chemistry*, 28(3), 350–356.
- Dyall, S. C., Balas, L., Bazan, N. G., Brenna, J. T., Chiang, N., da Costa Souza, F. & Taha, A. Y. (2022). Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures, and functions. *Progress in lipid research*, 86, 101165.
- Edeh, I. C., Nsofor, C. I., Ikeogu, C. F., Amobi, M. I., Ikechukwu, C. C., Ogbonnaya, H. F. & Avwernoya, F. (2021). Comparative study on the growth and survival of *Heterocladarias* fry fed on *Artemia* nauplii and *Moina Micrura*. *The Bioscientist Journal*, 9(1), 1-8.
- Eiras, B. J. C. F., de Sousa, L. M., Magalhães, A. & da Costa, R. M. (2023a). Development of *Moina minuta* (Hansen, 1899) (Cladocera: Anomopoda: Moinidae) under different food sources. *Observatório De La Economía Latinoamericana*, 21(6), 4232-4245.

- Eiras, B. J. C. F., Campelo, D. A. V., de Moura, L. B., de Oliveira, L. C. C., de Sousa, L. M. & da Costa, R. M. (2023b). Replacement of *Artemia franciscana* Nauplii by *Moina minuta* Neonates as live food on the larviculture of Angelfish (*Pterophyllum scalare*- Schultze, 1823) and Severum (*Heros severus*-Heckel, 1840). *Aquaculture Research*, 2023, 6118659.
- Ejsmont-Karabin, J. & Karabin, A. (2013). The suitability of zooplankton as lake ecosystem indicators: crustacean trophic state index. *Polish Journal of Ecology*, 61(3), 561-573.
- Eryalcin, K. (2018). Effects of different commercial feeds and enrichments on biochemical composition and fatty acid profile of rotifer (*Brachionus Plicatilis*, Muller 1786) and *Artemia Franciscana*. *Turkish Journal of Fisheries and Aquatic Sciences*, 18, 81-90.
- Espinosa-Rodríguez, C. A., Sarma, S. S. S. & Nandini, S. (2021). Zooplankton community changes in relation to different macrophyte species: Effects of *Egeria densa* removal. *Ecohydrology & Hydrobiologia*, 21(1), 153-163.
- Falfushynska, H., Kasianchuk, N., Siemens, E., Henao, E. & Rzymski, P. (2023). A review of common cyanotoxins and their effects on fish. *Toxics*, 11(2), 118.
- Fajardo, L.J.; Lebeng, R.S.; Morales, M.L.; Reyes, A.T. (2022). Plankton abundance and diversity in Pantabangan Reservoir, Panta-bangan, Nueva Ecija, Philippines. *AACL Bioflux*, 15, 1541-1552.
- Farhadian, O., Khanjani, M. H., Keivany, Y. & Ebrahimi, E. (2012). Culture experiments with a freshwater cladoceran, *Ceriodaphnia quadrangula* (OF Müller, 1785), as suitable live food for Mayan cichlid (*Cichlasoma urophthalmus*) larvae. *Brazilian Journal of Aquatic Science and Technology*, 16(2), 1-11.
- Fatma, C. A. F., Özdemir, N. Ş. & Gökçe, Z. (2020). Biochemical content (fatty acids, sterols, total protein, lipophilic vitamins) and antioxidant activity (GSH, MDA) of cladocerans (*Daphnia* sp., *Diaphasoma branchyrum*). *Mediterranean Fisheries and Aquaculture Research*, 3(1), 34-46.
- Fernandez, R., Oseguera, L. A. & Alcocer, J. (2020). Zooplankton biodiversity in tropical karst lakes of southeast Mexico, Chiapas. *Revista Mexicana de Biodiversidad*, 91, e913184.
- Fernando, C. H. and Ponyi, J. E. (1981). The freelifing freshwater cyclopoid Copepoda (Crustacea) of Malaysia and Singapore. *Hydrobiologia*, 78(2), 113-123.
- Ferrão-Filho, A. D. S. & da Silva, D. A. C. (2020). Saxitoxin-producing *Raphidiopsis raciborskii* (cyanobacteria) inhibits swimming and

- physiological parameters in *Daphnia similis*. *Science of the Total Environment*, 706, 135751.
- Fouzi, M. N. M., Surakshima, H. A. B. & Withanage, P. M. (2021). Influence of fish meal, yeast and maize on the growth and survival of freshwater zooplankton *Daphnia magna*. *Sri Langkan Journal of Technology*. 2021, 46-52.
- Freese, H. M. & Martin-Creuzburg, D. (2013). Food quality of mixed bacteria-algae diets for *Daphnia magna*. *Hydrobiologia*, 715, 63-76.
- Fujii, M., Takayama, Y., Imaizumi, Y., Yusoff, F. M., Yago, K., Nagao, N. & Toda, T. (2023). Energy-saving in fucoxanthin production by *Chaetoceros gracilis* in a flat-bag photobioreactor with intermittent mixing. *Biocatalysis and Agricultural Biotechnology*, 50, 102693.
- Gama Flores, J. L., Sarma, S. S. S., López Rocha, A. N. & Nandini, S. (2019). Effects of cladoceran-conditioned medium on the demography of brachionid rotifers (Rotifera: Brachionidae). *Hydrobiologia*, 844, 21-30.
- Gama-Flores, J. L., Huidobro-Salas, M. E., Sarma, S. S. S. & Nandini, S. (2020). Population responses and fatty acid profiles of *Brachionus calyciflorus* (Rotifera) in relation to different thermal regimes. *Journal of Thermal Biology*, 94, 102752.
- Gama-Flores, J. L., Huidobro-Salas, M. E., Sarma, S. S. S., Nandini, S., Zepeda-Mejia, R. & Gulati, R. D. (2015). Temperature and age affect the life history characteristics and fatty acid profiles of *Moina macrocopa* (Cladocera). *Journal of thermal biology*, 53, 135-142.
- Gao, H., Liu, X. & Ban, S. (2022). Effect of acute acidic stress on survival and metabolic activity of zooplankton from Lake Biwa, Japan. *Inland Waters*, 12(4), 488-498.
- García-Chicote, J., Armengol, X. & Rojo, C. (2019). Zooplankton species as indicators of trophic state in reservoirs from Mediterranean river basins. *Inland Waters*, 9(1), 113-123.
- García-García, G., Jiménez-Contreras, J., Vargas-Hernández, A. A., Nandini, S. & Sarma, S. S. S. (2017). Is aluminum innocuous to zooplankton at pH below 6?. *Bulletin of environmental contamination and toxicology*, 98, 489-495.
- Garzke, J., Sommer, U. & Ismar-Rebitz, S. M. (2020). Zooplankton growth and survival differentially respond to interactive warming and acidification effects. *Journal of Plankton Research*, 42(2), 189-202.
- Gasim, M. B., Toriman, M. E., Muftah, S., Bargig, A., Aziz, N. A. A., Azaman, F. & Muhamad, H. (2015). Water quality degradation of Cempaka Lake, Bangi, Selangor, Malaysia as an impact of excessive *E. coli* and nutrient concentrations. *Malaysian Journal of Analytical Sciences*, 19(6), 1391-1404.

- Geletu, T. T. (2023). Lake eutrophication: Control of phytoplankton overgrowth and invasive aquatic weeds. *Lakes & Reservoirs: Research & Management*, 28(1), e12425.
- Geng, Y., Li, M., Yu, R., Sun, H., Zhang, L., Sun, L. & Xu, J. (2022). Response of planktonic diversity and stability to environmental drivers in a shallow eutrophic lake. *Ecological Indicators*, 144, 109560.
- Gerasimova, T. N., Pogozhev, P. I. & Sadchikov, A. P. (2019). Suppression of Cyanobacterial Blooms by Zooplankton: Experiments in Natural Water Bodies with the Use of Flow-Through Ecosystems. *Russian Journal of General Chemistry*, 89, 2840-2844.
- Gilbert, J. J. (2022). Food niches of planktonic rotifers: Diversification and implications. *Limnology and Oceanography*, 67(10), 2218-2251.
- González, C. & del Carmen, A. (2022). *Unchaining the biomarker potential of aerobic methanotrophic bacteria* (Doctoral dissertation, Staats-und Universitätsbibliothek Hamburg Carl von Ossietzky).
- Gorokhova, E., El-Shehawy, R., Lehtiniemi, M. & Garbaras, A. (2021). How copepods can eat toxins without getting sick: Gut bacteria help zooplankton to feed in cyanobacteria blooms. *Frontiers in Microbiology*, 11, 589816.
- Gu, L., Qin, S., Zhu, S., Lu, N., Sun, Y., Zhang, L. & Yang, Z. (2020). *Microcystis aeruginosa* affects the inducible anti-predator responses of *Ceriodaphnia cornuta*. *Environmental Pollution*, 259, 113952.
- Gutierrez, M. F., Molina, F. R., Teixeira-de-Mello, F., Frau, D. & Antoniazzi, C. (2021). Influence of fish predation on the dynamic of zooplankton and macroinvertebrates in floodplain lakes under different turbidity conditions: an experimental study. *Aquatic Sciences*, 83(3), 48.
- Halsband, C., Dix, M. F., Sperre, K. H. & Reinardy, H. C. (2021). Reduced pH increases mortality and genotoxicity in an Arctic coastal copepod, *Acartia longiremis*. *Aquatic Toxicology*, 239, 105961.
- Hamil, S., Bouchelouche, D., Arab, S., Alili, M., Baha, M. & Arab, A. (2021). The relationship between zooplankton community and environmental factors of Ghrib Dam in Algeria. *Environmental Science and Pollution Research*, 28, 46592-46602.
- Han, C., Shimotsu, K., Kim, H. J., Sakakura, Y., Lee, J. S., Souissi, S. & Hagiwara, A. (2022). Comparison of ammonia thresholds for survival and reproduction between two copepods: The planktonic calanoid *Eurytemora affinis* and the benthic harpacticoid *Tigriopus japonicus*. *Aquaculture*, 560, 738534.

- Han, T., Kong, M., Tang, C., Xu, X., Zhu, Y., Gao, Y. & Li, W. (2021). Influence of algal blooms on the efficacy of La/Al-based phoslock in the control of phosphorus release from sediment in shallow lakes: a microcosm study. *Journal of Soils and Sediments*, 21(10), 3405-3414.
- Hashim, S. I. N. S., Talib, S. H. A. & Abustan, M. S (2020). Nutrient correlation analysis between sediment and water at Sembrong Dam, Johor. *International Journal of Advanced Research in Engineering and Technology*. 11. 986-993.
- Mourov, S. M., Mirzayeva, D. A., Khujamshukurov, N. A., Shakirov, Z. S., Kuchkarova, D. K. & Abdinazarov, K. K. (2020). The role of microalgae in the cultivation of zooplankton (*Daphnia*). *International Journal of Current Microbiology and Applied Sciences I*, 9(5), 3356-3365.
- He, H., Jeppesen, E., Bruhn, D., Yde, M., Hansen, J. K., Spanggaard, L. & Lauridsen, T. L. (2020). Decadal changes in zooplankton biomass, composition, and body mass in four shallow brackish lakes in Denmark subjected to varying degrees of eutrophication. *Inland Waters*, 10(2), 186-196.
- Hegab, M. H., Abdelhameed, M., Nasr, H. & Abd El Mola, H. (2020). Applying a new feeding protocol for enhancing mass culture and nutritional value of the rotifer *Brachionus plicatilis* Müller, 1786. *Aquaculture Studies*, 20(2), 81-89.
- Herath, S. S. & Satoh, S. (2022). Environmental impacts of nitrogen and phosphorus from aquaculture. In *Feed and Feeding Practices in Aquaculture* (pp. 427-444). Woodhead Publishing.
- Hernández-León, S., Calles, S. & de Puelles, M. L. F. (2019). The estimation of metabolism in the mesopelagic zone: disentangling deep-sea zooplankton respiration. *Progress in Oceanography*, 178, 102163.
- Hiltunen, M., Vehniäinen, E. R. & Kukkonen, J. V. (2021). Interacting effects of simulated eutrophication, temperature increase, and microplastic exposure on *Daphnia*. *Environmental Research*, 192, 110304.
- Hou, L., Yang, Y., Sun, B., Jing, Y. & Deng, W. (2021). Dietary fiber, gut microbiota, short-chain fatty acids, and host metabolism. *American Journal of Life Sciences*, 9, 162-172.
- Howarth, R. W., Chan, F., Swaney, D. P., Marino, R. M. & Hayn, M. (2021). Role of external inputs of nutrients to aquatic ecosystems in determining prevalence of nitrogen vs. phosphorus limitation of net primary productivity. *Biogeochemistry*, 154(2), 293-306.
- Huanacuni, J. I., Pepe-Victoriano, R., Lora-Vilchis, M. C., Merino, G. E., Torres-Taipe, F. G. & Espinoza-Ramos, L. A. (2021). Influence of microalgae diets on the biological and growth parameters of *Oithona nana* (Copepoda: Cyclopoida). *Animals*, 11, 3544.

- Huang, Y., Liang, Y., Li, Y., Cai, M., Jiang, Q., Li, D. & Zhao, Y. (2018). The effect of acute ammonia exposure on selected immunological parameters of *Daphnia similoides* (Cladocera). *Crustaceana*, 91(4), 403-411.
- Idris, B. A. G. (1983). Freshwater zooplankton of Malaysia (Crustacea: cladocera). Universiti Pertanian Malaysia Press. Malaysia.
- Ilić, M., Werner, C. & Fink, P. (2019). Equal relevance of omega-3 and omega-6 polyunsaturated fatty acids for the fitness of *Daphnia* spp. *Limnology and Oceanography*, 64(6), 2512-2525.
- Imran, A., Saadalla, M. J. A., Khan, S. U., Mirza, M. S., Malik, K. A. & Hafeez, F. Y. (2014). *Ochrobactrum* sp. Pv2Z2 exhibits multiple traits of plant growth promotion, biodegradation and N-acyl-homoserine-lactone quorum sensing. *Annals of microbiology*, 64, 1797-1806.
- Ishibashi, Y., Goda, H., Hamaguchi, R., Sakaguchi, K., Sekiguchi, T., Ishiwata, Y. & Ito, M. (2021). PUFA synthase-independent DHA synthesis pathway in *Parietichytrium* sp. and its modification to produce EPA and n-3DPA. *Communications Biology*, 4(1), 1378.
- Ismail, A. H. & Adnan, A. A. M. (2016). Zooplankton composition and abundance as indicators of eutrophication in two small man-made lakes. *Tropical life sciences research*, 27(suppl.1), 31-38.
- Ismail, A. H., Aziz, N. S., Zainal, Z. F., Abdullahi, Z. H., Haziqah, M. T. F. & Maznah, W. W. (2021a). Zooplankton study in Bukit Merah Reservoir, Malaysia a preliminary biodiversity assessment. *IOP Conference Series: Earth and Environmental Science*, 711 (1), 012003.
- Ismail, A., Haziqah, M. F., & Alawiyah, J. S. (2021b). The zooplankton community as an ecological indicator in a reservoir at Ulu Muda Forest Reserve, Kedah, Peninsular Malaysia. *Malayan Nature Journal*, 73(3).
- Ismail, A. H., Lim, C. C. & Wan Omar, W. M. (2019). Evaluation of spatial and temporal variations in zooplankton community structure with reference to water quality in Teluk Bahang Reservoir, Malaysia. *Tropical Ecology*, 60, 186-198.
- Ismail, H. N., Dini, U. W. A. & Tay, C. C. (2014). Biological responses of tropical cladoceran, *Ceriodaphnia cornuta* to different algae diets. *Journal of Life Sciences and Technology*, 2, 48-54.
- Jawan, A. & Sumin, V. (2012). The effect of land used on the water quality of oxbow lakes in Sabah. *The Malaysian Journal of Analytical Sciences*, 16(3), 273-276.
- Jeffries, D. S., Dieken, F. P. and Jones, D. E. (1979). Performance of the autoclave digestion method for total phosphorus analysis. *Water Research* 13(3), 275-279.

- Jithlang, I., & Wongrat, L. (2006). Composition and distribution of zooplankton in the Pasak Jolasid reservoir, Lop Buri province. *Journal of Fisheries and Environment*, 30, 1-18.
- Jiang, W., Chen, R., Zhao, L., Duan, Y., Wang, H., Yan, Z. & Mao, Z. (2023). Isolation of phloridzin-degrading, IAA-producing bacterium *Ochrobactrum haematophilum* and its effects on the apple replant soil environment. *Horticultural Plant Journal*, 9(2), 199-208.
- Kagali, R. N., Kim, H. J., Koga, T., Sakakura, Y. & Hagiwara, A. (2019). Effect of two commercial probiotic products on population growth of rotifer *Brachionus rotundiformis* Tschugunoff. *Hydrobiologia*, 844, 173-182.
- Kamilya, D. & Devi, W. M. (2022). *Bacillus* probiotics and bioremediation: An aquaculture perspective. In *Bacilli in Agrobiotechnology: Plant Stress Tolerance, Bioremediation, and Bioprospecting* (pp. 335-347). Cham: Springer International Publishing.
- Kandasamy, S., Palanichamy, M. & Mani, S. R. (2020). Mass culture of Cladocerans, *Diaphanasoma sarsi* and *Ceriodaphnia cornuta* using chicken manure. *Algae*, 10, 104.
- Kandathil Radhakrishnan, D., AkbarAli, I., Schmidt, B. V., John, E. M., Sivanpillai, S. & Thazhakot Vasunambesan, S. (2020). Improvement of nutritional quality of live feed for aquaculture: An overview. *Aquaculture Research*, 51(1), 1-17.
- Karpowicz, M., Ejsmont-Karabin, J., Kozlowska, J., Feniova, I. & Dzialowski, A. R. (2020). Zooplankton community responses to oxygen stress. *Water*, 12(3), 706.
- Kaviyarasan, M., Santhanam, P., Ananth, S., Kumar, S. D., Raju, P. & Kandan, S. (2020). Population growth, nauplii production and post-embryonic development of *Pseudodiaptomus annandalei* (Sewell, 1919) in response to temperature, light intensity, pH, salinity and diets. *Indian Journal of Geo Marine Sciences*, 49(6), 1000-1009.
- Kazamia, E., Czesnick, H., Nguyen, T. T. V., Croft, M. T., Sherwood, E., Sasso, S. & Smith, A. G. (2012). Mutualistic interactions between vitamin B12-dependent algae and heterotrophic bacteria exhibit regulation. *Environmental microbiology*, 14(6), 1466-1476.
- Kazmi, S. S. U. H., Wang, Y. Y. L., Cai, Y. E. & Wang, Z. (2022). Temperature effects in single or combined with chemicals to the aquatic organisms: An overview of thermo-chemical stress. *Ecological Indicators*, 143, 109354.
- Keister, J. E., Winans, A. K. & Herrmann, B. (2020). Zooplankton community response to seasonal hypoxia: A test of three hypotheses. *Diversity*, 12(1), 21.

- Khalifa, U., Ebenezer, V. & Pierson, J. J. (2022). Elevated temperature and low pH affect the development, reproduction, and feeding preference of the tropical cyclopoid copepod *Oithona rigida*. *International Journal of Environmental Studies*, 1-17.
- Khaw, Y. S., Yusoff, F. M., Tan, H. T., Noor Mazli, N. A. I., Nazarudin, M. F., Shaharuddin, N. A. & Takahashi, K. (2022). Fucoxanthin production of microalgae under different culture factors: A systematic review. *Marine Drugs*, 20(10), 592.
- Khaw, Y. S., Yusoff, F. M., Tan, H. T., Noor Mazli, N. A. I., Nazarudin, M. F., Shaharuddin, N. A. & Omar, A. R. (2021). The critical studies of fucoxanthin research trends from 1928 to June 2021: A bibliometric review. *Marine Drugs*, 19(11), 606.
- Khoo, K. S., Ooi, C. W., Chew, K. W., Chia, S. R., Foo, S. C., Ng, H. S. & Show, P. L. (2022). Extraction of Fucoxanthin from *Chaetoceros calcitrans* by electropermeabilization-assisted liquid biphasic flotation system. *Journal of Chromatography A*, 1668, 462915.
- Khoo, K. S., Ooi, C. W., Chew, K. W., Foo, S. C. & Show, P. L. (2021) Bioprocessing of *Chaetoceros calcitrans* for the recovery of fucoxanthin using CO<sub>2</sub>-based alkyl carbamate ionic liquids. *Bioresource Technology*, 322, 124520.
- Kim, S. K., Yun, J. H., Joo, G. J. & Choi, J. Y. (2022). Hydrological characteristics and trophic status as dominant drivers of rotifer community composition in artificially created riverine wetlands. *Animals*, 12(4), 461.
- Kitamura, H., Ishitani, H., Kuge, Y. & Nakamoto, M. (1982). Determination of nitrate in freshwater and seawater by a hydrazine reduction method. *Japan Journal of Water Pollution Research* 5, 35-42.
- Kolarova, N. & Napiórkowski, P. (2023). Are rotifer indices suitable for assessing the trophic status in slow-flowing waters of canals?. *Hydrobiologia*, 1-11.
- Kouzuma, A. & Watanabe, K. (2015). Exploring the potential of algae/bacteria interactions. *Current Opinion in Biotechnology*, 33, 125-129.
- Kriech, A. J. & Osborn, L. V. (2022). Review of the impact of stormwater and leaching from pavements on the environment. *Journal of Environmental Management*, 319, 115687.
- Krupa, E., Romanova, S., Berkinbaev, G., Yakovleva, N. & Sadvakasov, E. (2020). Zooplankton as indicator of the ecological state of protected aquatic ecosystems (Lake Borovoe, Burabay National Nature Park, Northern Kazakhstan). *Water*, 12(9), 2580.
- Krztoń, W., Kosiba, J. & Wilk-Woźniak, E. (2022). Features that matter: studying how phytoplankton drives zooplankton community functional traits. *Hydrobiologia*, 849(12), 2647-2662.

- Kudnar, P. S. (2022). Comparative study of nutritional potential of cultured and wild zooplankton species *Moina macrocופה* (Straus, 1820). *GORTERIA Journal*, 35 (3), 1-10.
- Kumar, S., Radhakrishnan, D. K., Akbar Ali, I. & Nareshkumar, A. (2022). Probiotics and its application in aquaculture. In *Aquaculture Science and Engineering* (pp. 379-400). Singapore: Springer Nature Singapore.
- Kumari, N., Mishra, S., Häder, D. P. & Sinha, R. P. (2022). Cyanobacterial blooms and their implications in the changing environment. *Advances in Environmental and Engineering Research*, 3(1), 1-41.
- Kumari, S., Lianthuamluai, L., Sarkar, U. K., Puthiyottil, M., Karnatak, G., Meena, D. K. & Das, B. K. (2023). Environmental characterization of two ecologically distinct Gangetic oxbow lakes using zooplankton taxonomic indices through comparative approach for wetland monitoring. *Wetlands*, 43(1), 14.
- Lahnsteiner, F., Lahnsteiner, E. & Duenser, A. (2023). Suitability of different live feed for first feeding of freshwater fish larvae. *Aquaculture Journal*, 3(2), 107-120.
- Lai, H. C. and Fernando, C. H. (1978). The freshwater calanoida (crustacea: copepoda) of Singapore and Peninsular Malaysia. *Hydrobiologia*, 61(2), 113-127.
- Lal, J., Shatrupa, T. & Kashyap, N. (2022). Enriched live food and it's important on larval rearing of fish. JUST Agriculture. 2
- Latib, N. L., Yusoff, F., Nagao, N. & Nizar, H. (2020). Growth of tropical cladocerans *Ceriodaphnia cornuta* GO Sars, 1885 and *Moina micrura* Kurz, 1875 fed with different diets. *Journal of Environmental Biology*, 41, 1224-1229.
- Lee, A. K., Wei, J. H. & Welander, P. V. (2022a). De novo cholesterol biosynthesis in the bacterial domain. *bioRxiv*, 2022-10.
- Lee, A. K., Wei, J. H. & Welander, P. V. (2023). De novo cholesterol biosynthesis in bacteria. *Nature Communications*, 14(1), 2904.
- Lee, E. H., Choi, S. Y., Seo, M. H., Lee, S. J. & Soh, H. Y. (2020). Effects of temperature and pH on the egg production and hatching success of a common Korean copepod. *Diversity*, 12(10), 372.
- Lee, Y., Kim, M. S., Park, J. J. C., Lee, Y. H. & Lee, J. S. (2022b). Oxidative stress-mediated synergistic deleterious effects of nano-and microplastics in the hypoxia-conditioned marine rotifer *Brachionus plicatilis*. *Marine Pollution Bulletin*, 181, 113933.
- Li, S. N., Zhang, C., Li, F., Ren, N. Q. & Ho, S. H. (2023). Recent advances of algae-bacteria consortia in aquatic remediation. *Critical Reviews in Environmental Science and Technology*, 53(3), 315-339.

- Li, S., Chu, Y., Xie, P., Xie, Y., Chang, H. & Ho, S. H. (2022a). Insights into the microalgae-bacteria consortia treating swine wastewater: symbiotic mechanism and resistance genes analysis. *Bioresource Technology*, 349, 126892.
- Li, Y. & Chen, F. (2020). Are zooplankton useful indicators of water quality in subtropical lakes with high human impacts?. *Ecological Indicators*, 113, 106167.
- Li, Y., Geng, M., Yu, J., Du, Y., Xu, M., Zhang, W. & Chen, F. (2022b). Eutrophication decrease compositional dissimilarity in freshwater plankton communities. *Science of The Total Environment*, 821, 153434.
- Liang, D., Wang, Q., Wei, N., Tang, C., Sun, X. & Yang, Y. (2020a). Biological indicators of ecological quality in typical urban river-lake ecosystems: The planktonic rotifer community and its response to environmental factors. *Ecological Indicators*, 112, 106127.
- Liang, Y., Gao, T., Shao, L., Min, Y. & Yang, J. (2020b). Effects of microcystin-LR and nitrite on the lifespan, reproduction, and heat shock responses of rotifer *Brachionus calyciflorus* at different temperatures. *Aquatic Sciences*, 82, 1-14.
- Liang, Y., Lu, X., Min, Y., Liu, L. & Yang, J. (2018). Interactive effects of microcystin and ammonia on the reproductive performance and phenotypic traits of the rotifer *Brachionus calyciflorus*. *Ecotoxicology and environmental safety*, 147, 413-422.
- Liguori, A. (2022). Multigenerational life-history Responses to pH in distinct populations of the copepod *Tigriopus californicus*. *The Biological Bulletin*, 242(2), 97-117.
- Lin, S. S., Shen, S. L., Zhou, A. & Lyu, H. M. (2021). Assessment and management of lake eutrophication: A case study in Lake Erhai, China. *Science of the Total Environment*, 751, 141618.
- Liu, C., Dong, S., Zhou, Y., Shi, K., Pan, Z., Sun, D. & Gao, Q. (2019). Temperature-dependent fatty acid composition change of phospholipid in steelhead trout (*Oncorhynchus mykiss*) tissues. *Journal of Ocean University of China*, 18, 519-527.
- Liu, P., Pan, J., Lin, J., Wen, Z., Huang, Q., Pajk, F. & Han, B. P. (2020b). Temperature niche difference and interspecific competition determine the parapatric distribution of two congeneric species in *Diaphanosoma*. *Research square*, 1-15.
- Liu, X., Dur, G., Ban, S., Sakai, Y., Ohmae, S. & Morita, T. (2020a). Planktivorous fish predation masks anthropogenic disturbances on decadal trends in zooplankton biomass and body size structure in Lake Biwa, Japan. *Limnology and Oceanography*, 65(3), 667-682.

- Liu, Y., Yao, S., Xu, P., Cao, Y., Li, J., Wang, J. & Cheng, C. (2014). Composition and diversity of endophytic bacterial communities in Noni (*Morinda citrifolia* L.) seeds. *International Journal of Agricultural Policy and Research*, 2(3), 98-104.
- Locke, A. & Sprules, W. G. (2000). Effects of acidic pH and phytoplankton on survival and condition of *Bosmina longirostris* and *Daphnia pulex*. *Hydrobiologia*, 437, 187-196.
- Lorenzen, C. J. (1967). Determination of chlorophyll and pheo-pigments: spectrophotometric equations. *Limnology and Oceanography* 12(2), 343-346.
- Lowery, C. M., Bown, P. R., Fraass, A. J. & Hull, P. M. (2020). Ecological response of plankton to environmental change: Thresholds for extinction. *Annual Review of Earth and Planetary Sciences*, 48, 403-429.
- Lucchesi, D. O., Chipps, S. R. & Schumann, D. A. (2022). Effects of seasonal hypoxia on macroinvertebrate communities in a small reservoir. *Lakes & Reservoirs: Research & Management*, 27(1), e12395.
- Lučić, D., Hure, M., Bobanović-Čolić, S., Njire, J., Vidjak, O., Onofri, I. & Batistić, M. (2019). The effect of temperature change and oxygen reduction on zooplankton composition and vertical distribution in a semi-enclosed marine system. *Marine Biology Research*, 15(4-6), 325-342.
- Lyu, K., Cao, H., Chen, R., Wang, Q. & Yang, Z. (2013). Combined effects of hypoxia and ammonia to *Daphnia similis* estimated with life-history traits. *Environmental Science and Pollution Research*, 20, 5379-5387.
- Lyu, K., Meng, Q., Zhu, X., Dai, D., Zhang, L., Huang, Y. & Yang, Z. (2016). Changes in iTRAQ-based proteomic profiling of the cladoceran *Daphnia magna* exposed to microcystin-producing and microcystin-free *Microcystis aeruginosa*. *Environmental Science & Technology*, 50(9), 4798-4807.
- Macedo, R. L., Franco, A. C. S., Klippe, G., Oliveira, E. F., Silva, L. H. S., dos Santos, L. N. & Branco, C. W. (2020). Small in size but rather pervasive: the spread of the North American rotifer *Kellicottia bostoniensis* (Rousselet, 1908) through Neotropical basins. *BioInvasions Rec*, 9(2), 287-302.
- Macke, E., Callens, M., De Meester, L. & Decaestecker, E. (2017). Host-genotype dependent gut microbiota drives zooplankton tolerance to toxic cyanobacteria. *Nature Communications*, 8(1), 1608.
- Magouz, F. I., Essa, M. A., Matter, M., Tageldein Mansour, A., Alkafafy, M. & Ashour, M. (2021). Population dynamics, fecundity and fatty acid

- composition of *Oithona nana* (Cyclopoida, Copepoda), fed on different diets. *Animals*, 11(5), 1188.
- Makwinja, R., Mengistou, S., Kaunda, E. & Alamirew, T. (2021). Spatial distribution of zooplankton in response to ecological dynamics in tropical shallow lake: Insight from Lake Malombe, Malawi. *Journal of Freshwater Ecology*, 36(1), 127-148.
- Manzi, F., Agha, R., Mühlenhaupt, M. & Wolinska, J. (2022). Prior exposure of a fungal parasite to cyanobacterial extracts does not impair infection of its *Daphnia* host. *Hydrobiologia*, 849(12), 2731-2744.
- Mao, Z., Cao, Y., Gu, X., Zeng, Q., Chen, H. & Jeppesen, E. (2023). Response of zooplankton to nutrient reduction and enhanced fish predation in a shallow eutrophic lake. *Ecological Applications*, 33(1), e2750.
- Mao, Z., Gu, X., Cao, Y., Zhang, M., Zeng, Q., Chen, H. & Jeppesen, E. (2020). The role of top-down and bottom-up control for phytoplankton in a subtropical shallow eutrophic lake: evidence based on long-term monitoring and modeling. *Ecosystems*, 23, 1449-1463.
- Marileo, L., Acuña, J., Rilling, J., Díaz, P., Langellotti, A. L., Russo, G. L. & Viscardi, S. (2023). Protist-lactic acid bacteria co-culture as a strategy to bioaccumulate polyunsaturated fatty acids in the protist *Aurantiochytrium* sp. T66. *Marine Drugs*, 21(3), 142.
- Martin-Creuzburg, D., Beck, B. & Freese, H. M. (2011). Food quality of heterotrophic bacteria for *Daphnia magna*: evidence for a limitation by sterols. *FEMS Microbiology Ecology*, 76(3), 592-601.
- Martin-Creuzburg, D., Sperfeld, E. & Wacker, A. (2009). Colimitation of a freshwater herbivore by sterols and polyunsaturated fatty acids. *Proceedings of the Royal Society B: Biological Sciences*, 276(1663), 1805-1814.
- Martínez-Córdova, L. R., Robles-Porcha, G. R., Vargas-Albores, F., Porcha-Cornejo, M. A., & Martínez-Porcha, M. (2022). Microbial bioremediation of aquaculture effluents. In *Microbial Biodegradation and Bioremediation* (pp. 409-417). Elsevier.
- Mathieu, F., Guo, F. & Kainz, M. J. (2022). Tracking dietary fatty acids in triacylglycerols and phospholipids of zooplankton. *Freshwater Biology*, 67(11), 1949-1959.
- McDonald, K., DesRochers, N., Renaud, J. B., Sumarah, M. W. & McMullin, D. R. (2023). Metabolomics reveals strain-specific cyanopeptide profiles and their production dynamics in *Microcystis aeruginosa* and *M. flos-aquae*. *Toxins*, 15(4), 254.
- Mejias, C., Riquelme, C., Sayes, C., Plaza, J. & Silva-Aciar, F. (2018). Production of the rotifer *Brachionus plicatilis* (Müller 1786) in closed outdoor systems fed with the microalgae *Nannochloropsis gaditana* and

- supplemented with probiotic bacteria *Pseudoalteromonas* sp. (SLP1). *Aquaculture International*, 26, 869-884.
- Mejri, S. C., Tremblay, R., Audet, C., Wills, P. S. & Riche, M. (2021). Essential fatty acid requirements in tropical and cold-water marine fish larvae and juveniles. *Frontiers in Marine Science*, 8, 680003.
- Meremo, W.T., Reuben, O., Wamalwa, Y.A. & Ndegwa, D.M. (2022) Changes in water quality parameters and their effect on zooplankton distribution in a shallow bay of Lake Victoria, Kenya. *International Journal of Fisheries and Aquatic Studies*, 10(4), 206–212.
- Min, C., Johansson, L. S., Søndergaard, M., Lauridsen, T. L., Chen, F., Sh, T. & Jeppesen, E. (2021). Copepods as environmental indicator in lakes: Special focus on changes in the proportion of calanoids along nutrient and pH gradients. *Aquatic Ecology*, 55, 1241-1252.
- Mohd-Asharuddin, S., Zayadi, N., Rasit, W. & Othman, N. (2016). Water quality characteristics of Sembrong Dam Reservoir, Johor, Malaysia. *IOP Conference Series: Materials Science and Engineering*, 136 (1), 012058.
- Moody, E. K. & Wilkinson, G. M. (2019). Functional shifts in lake zooplankton communities with hypereutrophication. *Freshwater Biology*, 64(3), 608-616.
- Moser, G. A. O., Barrera-Alba, J. J., Ortega, M. J., Alves-de-Souza, C. & Bartual, A. (2022). Comparative characterization of three *Tetraselmis chui* (Chlorophyta) strains as sources of nutraceuticals. *Journal of Applied Phycology*, 34, 821-835.
- Moustaka-Gouni, M & Sommer, U. (2020). Effects of harmful blooms of large-sized and colonial cyanobacteria on aquatic food webs. *Water*, 12(6), 1587.
- Mubarak, A.S., Jusadi, D, Junior, M.Z. & Suprayudi, M.A. (2017). The population growth and the nutritional status of *Moina macrocopa* feed with rice bran and cassava bran suspensions. *J. Akuak. Indon.*, 16, 223-233.
- Munoz-Colmenares, M. E., Soria, J. M. & Vicente, E. (2021). Can zooplankton species be used as indicators of trophic status and ecological potential of reservoirs?. *Aquatic Ecology*, 55(4), 1143-1156.
- Muttharasi, C., Gayathri, V., Muralisankar, T., Mohan, K., Uthayakumar, V., Radhakrishnan, S. & Palanisamy, M. (2021). Growth performance, digestive enzymes and antioxidants activities in the shrimp *Litopenaeus vannamei* fed with *Amphiroa fragilissima* crude polysaccharides encapsulated *Artemia* nauplii. *Aquaculture*, 545, 737263.
- Naman, N., Kassim, Z. & Rasdi, N. W. (2021). The effect of copepod enriched-vegetable based diet on Giant Tiger Prawn (*Penaeus monodon*) post-

- larvae. *IOP Conference Series: Earth and Environmental Science*, 674 (1), 012081.
- Nandini, S. & Sarma, S. S. S. (2023). Experimental studies on zooplankton-toxic cyanobacteria interactions: A Review. *Toxics*, 11(2), 176.
- Nandini, S., Araiza-Vázquez, D. A. & Sarma, S. S. S. (2021a). *Moina macrocopa* demographic response to harmful cyanobacteria. *Ecohydrology & Hydrobiology*, 21(2), 333-340.
- Nandini, S., Miracle, M. R., Vicente, E. & Sarma, S. S. S. (2021b). Strain-related differences in bacterivory and demography of *Diaphanosoma mongolianum* (Cladocera) in relation to diet and previous exposure to cyanobacteria in nature. *Aquatic Ecology*, 55(4), 1225-1239.
- Napiórkowski, P. & Napiórkowska, T. (2017). Limnophase versus potamophase: how hydrological connectivity affects the zooplankton community in an oxbow lake (Vistula River, Poland). In *Annales de Limnologie-International Journal of Limnology*, 53, 143-151.
- Napiórkowski, P., Bałkowska, M., Mrozińska, N., Szymańska, M., Kolarova, N. & Obolewski, K. (2019). The effect of hydrological connectivity on the zooplankton structure in floodplain lakes of a regulated large river (the Lower Vistula, Poland). *Water*, 11(9), 1924.
- Neri, T. A., Rohmah, Z., Ticar, B. F. & Choi, B. D. (2020). Effect of different culture conditions on nutritional value of *Moina macrocopa* as a live feed for fish fry production. *농업생명과학연구*, 54(6), 91-98.
- Nevejan, N., De Schryver, P., Wille, M., Dierckens, K., Baruah, K. & Van Stappen, G. (2018). Bacteria as food in aquaculture: do they make a difference?. *Reviews in Aquaculture*, 10(1), 180-212.
- Nieman, C. L. & Gray, S. M. (2019). Visual performance impaired by elevated sedimentary and algal turbidity in walleye *Sander vitreus* and emerald shiner *Notropis atherinoides*. *Journal of Fish Biology*, 95(1), 186-199.
- Noor Mazli, N. A. I., Yusoff, F. M., Karim, M (2022). Improvement of microalgae growth rate and nutritional contents through symbiotic bacterial-microalgae interaction. Master Thesis. UPM.
- Nugroho, T. S. A., Ekasari, J., Jusadi, D. & Setiawati, M. (2021). Productivity and nutritional quality of *Moina* sp. cultivated in various culture media. *Jurnal Akuakultur Indonesia*, 20(2), 148-162.
- Ogello, E. O., Wullur, S. & Hagiwara, A. (2019). Blending fishwastes and chicken manure extract as low-cost and stable diet for mass culture of freshwater zooplankton, optimized for aquaculture. *IOP Conference Series: Materials Science and Engineering*, 567 (1), 012022).

- Olmos, J., Acosta, M., Mendoza, G. & Pitones, V. (2020). *Bacillus subtilis*, an ideal probiotic bacterium to shrimp and fish aquaculture that increase feed digestibility, prevent microbial diseases, and avoid water pollution. *Archives of microbiology*, 202, 427-435.
- Ovie, S. I. & Ovie, S. O. (2006). Moisture, protein, and amino acid contents of three freshwater zooplankton used as feed for aquacultured larvae and postlarvae. *The Israeli Journal of Aquaculture – Bamidgeh* 58(1), 2006, 29-33.
- Pan, Y. J., Dahms, H. U., Hwang, J. S. & Souissi, S. (2022). Recent trends in live feeds for marine larviculture: a mini review. *Frontiers in Marine Science*, 9, 864165.
- Papa, R. D. S., & Zafaralla, M. T. (2011). The composition, diversity and community dynamics of limnetic zooplankton in a tropical caldera lake (Lake Taal, Philippines). *Raffles Bulletin of Zoology*, 59.
- Park, J. C. & Park, H. G. (2010). Growth of the brackish water flea, *Diaphanosoma celebensis*, on different foods and food concentrations. *Korean Journal of Fisheries and Aquatic Sciences*, 43(2), 131-138.
- Parsons, T. R., Maita, Y. and Lalli, C. M. (1984). A manual of chemical and biological methods for seawater analysis. (pp. 173). England, Pergamon Press Ltd.
- Pawlak-Skowrońska, B. & Bownik, A. (2021). Cyanobacterial anabaenopeptin-B, microcystins and their mixture cause toxic effects on the behavior of the freshwater crustacean *Daphnia magna* (Cladocera). *Toxicon*, 198, 1-11.
- Pawlak-Skowrońska, B., Toporowska, M. & Mazur-Marzec, H. (2019). Effects of secondary metabolites produced by different cyanobacterial populations on the freshwater zooplankters *Brachionus calyciflorus* and *Daphnia pulex*. *Environmental Science and Pollution Research*, 26, 11793-11804.
- Peerakietkhajorn, S., Tsukada, K., Kato, Y., Matsuura, T. & Watanabe, H. (2015). Symbiotic bacteria contribute to increasing the population size of a freshwater crustacean, *Daphnia magna*. *Environmental Microbiology Reports*, 7(2), 364-372.
- Peerakietkhajorn, S., Kato, Y., Kasalický, V., Matsuura, T. & Watanabe, H. (2016). Betaproteobacteria *Limnohabitans* strains increase fecundity in the crustacean *Daphnia magna*: Symbiotic relationship between major bacterioplankton and zooplankton in freshwater ecosystem. *Environmental microbiology*, 18(8), 2366-2374.
- Pereira, W. A., Mendonça, C. M. N., Urquiza, A. V., Marteinsson, V. P., LeBlanc, J. G., Cotter, P. D. & Oliveira, R. P. (2022). Use of probiotic bacteria and

- bacteriocins as an alternative to antibiotics in aquaculture. *Microorganisms*, 10(9), 1705.
- Pérez-Morales, A., Sarma, S. S. S., Nandini, S., Espinosa-Rodríguez, C. A. & Rivera-De la Parra, L. (2020). Demographic responses of selected rotifers (Rotifera) and cladocerans (Cladocera) fed toxic *Microcystis aeruginosa* (Cyanobacteria). *Fundamental and Applied Limnology*, 193, 261-274.
- Phan, D. D., Nguyen V. K., Le, T. N., Dang Ngoc, T. and Ho T. H. (2015). Identification Handbook of Freshwater Zooplankton of the Mekong River and its Tributaries, Mekong River Commission, Vientiane. 207pp.
- Phan, N. T., Duong, Q. H., Tran-Nguyen, Q. A. & Trinh-Dang, M. (2021). The species diversity of tropical freshwater rotifers (Rotifera: Monogononta) in relation to environmental factors. *Water*, 13(9), 1156.
- Picapedra, P. H. S., Fernandes, C., Baumgartner, G. & Sanches, P. V. (2020). Zooplankton communities and their relationship with water quality in eight reservoirs from the midwestern and southeastern regions of Brazil. *Brazilian Journal of Biology*, 81, 701-713.
- Pinto, I., Nogueira, S., Rodrigues, S., Formigo, N. & Antunes, S. C. (2023). Can zooplankton add value to monitoring water quality? A case study of a meso/eutrophic Portuguese Reservoir. *Water*, 15(9), 1678.
- Pontin R. (1978). A Key to the Freshwater Planktonic and Semi-Planktonic Rotifera of the British Isles. Freshwater Biological Association, Ambleside. 178pp.
- Prasertphon, R., Chaichana, R., & Jitchum, P. (2023). Seasonal variation of zooplankton assemblages and their responses to water chemistry and microcystin content in shallow lakes in Thailand. *Archives of Biological Sciences*, 75, 369-378.
- Qin, H., Cao, X., Cui, L., Lv, Q. & Chen, T. (2020). The influence of human interference on zooplankton and fungal diversity in Poyang Lake watershed in China. *Diversity*, 12(8), 296.
- Qin, S., Ma, L., Li, D., Huang, J., Zhang, L., Sun, Y. & Yang, Z. (2021). Rising temperature accelerates the responses of inducible anti-predator morphological defenses of *Ceriodaphnia cornuta* but decreases the responsive intensity. *Ecological Indicators*, 120, 106919.
- Qin, S., Yang, T., Yu, B., Zhang, L., Gu, L., Sun, Y. & Yang, Z. (2022). The stress effect of atrazine on the inducible defense traits of *Daphnia pulex* in response to fish predation risk: Evidences from morphology, life history traits, and expression of the defense-related genes. *Environmental Pollution*, 311, 119965.

- Quirino, B. A., Teixeira de Mello, F., Deosti, S., Bonecker, C. C., Cardozo, A. L. P., Yofukuji, K. Y. & Fugi, R. (2021). Interactions between a planktivorous fish and planktonic microcrustaceans mediated by the biomass of aquatic macrophytes. *Journal of Plankton Research*, 43(1), 46-60.
- Rahmati, R., Esmaeili Fereidouni, A., Rouhi, A. & Agh, N. (2020). Effects of different diets on population growth and fatty acids composition in cyclopoid copepod, *Acanthocyclops trajani* (Mirabdullayev and Defaye, 2002): A potential supplementary live food for freshwater fish larvae. *Iranian Journal of Fisheries Sciences*, 19(3), 1447-1462.
- Ramaekers, L., Vanschoenwinkel, B., Brendonck, L. & Pinceel, T. (2023). Elevated dissolved carbon dioxide and associated acidification delays maturation and decreases calcification and survival in the freshwater crustacean *Daphnia magna*. *Limnology and Oceanography*, 68(7), 1624-1635.
- Ramlee, A., Chembaruthy, M., Gunaseelan, H., Yatim, S. R. M., Taufek, H., & Rasdi, N. W. (2021). Enhancement of nutritional value on zooplankton by alteration of algal media composition: A review. *IOP conference series: earth and environmental science*, 869 (1), 012006).
- Ramlee, A., Suhami, H. And Rasdi, N. W. (2022). Diversity and abundance of plankton in different habitat zonation of Papan River, Lake Kenyir, Malaysia. *Biodiversitas Journal of Biological Diversity*, 23(1).
- Ramón, A., Esteves, A., Villadóniga, C., Chalar, C. & Castro-Sowinski, S. (2023). A general overview of the multifactorial adaptation to cold: biochemical mechanisms and strategies. *Brazilian Journal of Microbiology*, 1-29.
- Rasdi, N. W., Arshad, A., Ikhwanuddin, M., Hagiwara, A., Yusoff, F. M. & Azani, N. (2020a). A review on the improvement of cladocera (*Moina*) nutrition as live food for aquaculture: Using valuable plankton fisheries resources. *Journal of Environmental Biology*, 41, 1239-1248.
- Rasdi, N. W., Ikhwannuddin, M., Azani, N., Ramlee, A., Yuslan, A., Suhami, H. & Arshad, A. (2020c). The effect of different feeds on the growth, survival and reproduction of rotifer, *Brachionus plicatilis*. *Journal of Environmental Biology*, 41, 1275-1280.
- Rasdi, N. W., Ikhwannuddin, M., Syafika, C. A., Azani, N. & Ramli, A. (2021b). Effects of using enriched copepod with microalgae on growth, survival, and proximate composition of giant freshwater prawn (*Macrobrachium rosenbergii*). *Iranian Journal of Fisheries Sciences*, 20(4), 986-1003.
- Rasdi, N. W., Qin, J. G., Naseer, N. M., Ikhwanuddin, M., Yik Sung, Y., Md Yusoff, F. & Hagiwara, A. (2021a). The impact of feeding algae and canola oil on the growth, survival and reproduction of *Moina* sp. *Songklaakarin Journal of Science & Technology*, 43(3).

- Rasdi, N. W., Ramlee, A., Abol-Munafi, A. B., Ikhwanuddin, M., Azani, N., Yuslan, A. & Arshad, A. (2020b). The effect of enriched Cladocera on growth, survivability and body coloration of Siamese fighting fish. *Journal of Environmental Biology*, 41, 1257-1263.
- Rashid, H. & Prakash, M. M. (2022). Zooplanktons as bioindicators of water pollution from Vikram Tearth Sarovar Ujjain (MP). *Journal of Pharmaceutical Negative Results*, 888-895.
- Rizo, E. Z. C., Liu, P., Niu, H., Yang, Y., Lin, Q., Papa, R. D. S. & Han, B. P. (2020). Zooplankton in a continuous waterscape: environmental and spatial factors shaping spring zooplankton community structure in a large canyon reservoir at the tropic of cancer. *Hydrobiologia*, 847, 3621-3635.
- Rodmongkoldee, M. & Taparhudee, W. (2020a). Life table responses of *Moina micrura* fed with different food concentrations. *Burapha Science Journal* (วารสาร วิทยาศาสตร์บูรพา), 25(3), 1136-1146.
- Rodmongkoldee, M., Taparhudee, W. & Saengphan, N. (2020b). Laboratory study on life history of three water flea species (Cladocera: Moinidae) in Thailand. *Burapha Science Journal* (วารสาร วิทยาศาสตร์บูรพา), 25(1), 129-140.
- Roman, M. R., Brandt, S. B., Houde, E. D. & Pierson, J. J. (2019). Interactive effects of hypoxia and temperature on coastal pelagic zooplankton and fish. *Frontiers in Marine Science*, 6, 139.
- Ross, A. J. & Arnott, S. E. (2022). Similar zooplankton responses to low pH and calcium may impair long-term recovery from acidification. *Ecological Applications*, 32(3), e2512.
- Ruthena, Y., Anggoro, S., & Soeprobawati, T. R. (2023). Diversity of Plankton Bio-indicators on Water Quality of Lutan Lake, Palangka Raya, Central Kalimantan, Indonesia. *E3S Web of Conferences*, 448, 03068.
- Sa-Ardrit, P., Pholpunthin, P. and Segers, H. (2013). A checklist of the freshwater rotifer fauna of Thailand (Rotifera, Monogononta, Bdelloidea). *Journal of Limnology* 72(s2), 18.
- Saha, H., Wisdom, K. S., Devi, A. L., Pde, D. U. R., Devi, S. T., Kamei, M. & Pal, P. (2017). Effects of water pH on life history parameters of a new bosminiid Cladocera: *Bosmina (Bosmina) Tripurae* (Korinek, Saha and Bhattacharya, 1999) in laboratory condition. *Bulletin of Environmental Contamination and Toxicology*, 99, 23-26.
- Sahandi, J., Sorgeloos, P., Xiao, H., Wang, H., Qi, Z., Zheng, Y. & Tang, X. (2020). Possible effects of probiotic strains on suppression of Vibrio and enhancement of growth in rotifer, *Brachionus plicatilis*. *Iranian Journal of Fisheries Sciences*, 19(6), 3018-3033.

- Sahandi, J., Sorgeloos, P. & Zhang, W. (2022). Culture of *Artemia franciscana* nauplii with selected microbes suppressed Vibrio loading and enhanced survival, population stability, enzyme activity, and chemical composition. *Aquaculture International*, 30(5), 2279-2293.
- Sunardi, S., Yoshimatsu, T., Junianto, N., Istiqamah, N., & Deweber, T. (2016). Long-term variability of zooplankton community under climate warming in tropical eutrophic man-made lake. *Biodiversitas Journal of Biological Diversity*, 17.
- Samat, N. A., Yusoff, F. M., Chong, C. M. & Karim, M. (2020a). Enrichment of freshwater zooplankton *Moina micrura* with probiotics isolated from microalgae. *Journal of Environmental Biology*, 41, 1215-1223.
- Samat, N. A., Yusoff, F. M., Lim, K. C., Rasdi, N. W., Syukri, F. & Karim, M. (2022). Effects of temperature, pH, and photoperiod on the performance of a freshwater cladoceran *Moina micrura* culture enriched with *Lysinibacillus fusiformis* and *Bacillus pocheonensis*. *Latin american journal of aquatic research*, 50(5), 681-691.
- Samat, N. A., Yusoff, F. M., Rasdi, N. W. & Karim, M. (2020b). Enhancement of live food nutritional status with essential nutrients for improving aquatic animal health: A review. *Animals*, 10(12), 2457.
- Samat, N. A., Yusoff, F. M., Rasdi, N. W. & Karim, M. (2021). The efficacy of *Moina micrura* enriched with probiotic *Bacillus pocheonensis* in enhancing survival and disease resistance of red hybrid tilapia (*Oreochromis* spp.) larvae. *Antibiotics*, 10(8), 989.
- Samo, T.J., Kimbrel, J.A., Nilson, D.J., Pett-Ridge, J., Weber, P.K. & Mayali, X. (2018). Attachment between heterotrophic bacteria and microalgae influences symbiotic microscale interactions. *Environmental Microbiology*, 20, 4385–4400.
- Sánchez-Colón, Y. M. & Schaffner, F. C. (2021). Identifying nonpoint sources of phosphorus and nitrogen: A case study of pollution that enters a freshwater wetland (Laguna Cartagena, Puerto Rico). *Journal of Water resource and Protection*, 13(8), 588-604.
- Sandhya, S. V. & Vijayan, K. K. (2019). Symbiotic association among marine microalgae and bacterial flora: a study with special reference to commercially important *Isochrysis galbana* culture. *Journal of Applied Phycology*, 31, 2259-2266.
- Sashidhar B. & Podile A. R. (2010) Mineral phosphate solubilization by rhizosphere bacteria and scope for manipulation of the direct oxidation pathway involving glucose dehydrogenase. *Journal of Applied Microbiology*, 109, 1-12

- Schenone, L., Modenutti, B., Martyniuk, N., Bastidas Navarro, M., Laspoumaderes, C. & Balseiro, E. (2021). Modelling key variables for understanding the effects of grazing and nutrient recycling by zooplankton on the freshwater microbial loop. *Freshwater Biology*, 66(12), 2322-2337.
- Segers, H. (2008). Global diversity of rotifers (Rotifera) in freshwater. *Hydrobiologia*, 595(1), 49-59.
- Shah, A. S. R. M., Ismail, J., Latief, D., & Omar, W. M. W. (2012). The spatial structure of zooplankton communities of Pedu Reservoir, Malaysia. *Wetland Science*, 10, 423-428.
- Sharip Z. & Mohamad M. F. (2019) Microbial contamination in urban tropical lentic waterbodies and ponds along urbanisation gradient. *Journal of Tropical Agricultural Sciences* 42, 165–184.
- Sharip, Z. (2019). Spatio-temporal variation of zooplankton community structure in tropical urban waterbodies along trophic and urban gradients. *Ecological processes*, 8(1), 1-12.
- Sharip, Z. (2021). Changes in phytoplankton and zooplankton abundance and diversity in macrophyte-dominated and open pelagic ecosystem of shallow reservoirs. *Inland Water Biology*, 14(4), 427-437.
- Shen, J., Qin, G., Yu, R., Zhao, Y., Yang, J., An, S. & Wan, Y. (2021). Urbanization has changed the distribution pattern of zooplankton species diversity and the structure of functional groups. *Ecological Indicators*, 120, 106944.
- Shen, Q., Zhan, Y., Jia, X., Li, B., Zhu, X. & Gao, T. (2022). Combined effects of the pesticide spinetoram and the cyanobacterium *Microcystis* on the water flea *Daphnia pulex*. *Environmental Science and Pollution Research*, 29(31), 47148-47158.
- Shield, R. J. (1995). A guide to identification of rotifers, cladocerans and copepods from Australian inland waters. Co-operative Research Centre for Freshwater Ecology. Albury, New South Wales, Australia.
- Shrivastava, J., Ndugwa, M., Caneos, W. & De Boeck, G. (2019). Physiological trade-offs, acid-base balance and ion-osmoregulatory plasticity in European sea bass (*Dicentrarchus labrax*) juveniles under complex scenarios of salinity variation, ocean acidification and high ammonia challenge. *Aquatic Toxicology*, 212, 54-69.
- Sinev, A. Y. and Yusoff, F. M. (2015). Cladocera (Crustacea: Branchiopoda) of Sabah state in Borneo Island, Malaysia. *Zootaxa*, 4000(5), 581-591.
- Singh, K., Munilkumar, S., Sahu, N. P., Das, A. & Devi, G. A. (2019). Feeding HUFA and vitamin C-enriched *Moina micrura* enhances growth and

- survival of *Anabas testudineus* (Bloch, 1792) larvae. *Aquaculture*, 500, 378-384.
- Sipaúba-Tavares, L. H., Truzzi, B. S., & Berchielli-Morais, F. D. A. (2014). Growth and development time of subtropical Cladocera *Diaphanosoma birgei* Korinek, 1981 fed with different microalgal diets. *Brazilian Journal of Biology*, 74, 464-471.
- Siqwepu, O., Richoux, N. B. & Vine, N. G. (2017). The effect of different dietary microalgae on the fatty acid profile, fecundity and population development of the calanoid copepod *Pseudodiaptomus hessei* (Copepoda: Calanoida). *Aquaculture*, 468, 162-168.
- Sobko, E. I., Shirokova, L. S., Klimov, S. I., Chupakov, A. V., Zabelina, S. A., Shorina, N. V. & Vorobieva, T. Y. (2023). Environmental factors controlling zooplankton communities in Thermokarst Lakes of the Bolshezemelskaya Tundra Permafrost peatlands (NE Europe). *Water*, 15(3), 511.
- Stamou, G., Katsiapi, M., Moustaka-Gouni, M. (2019). Trophic state assessment based on zooplankton communities in Mediterranean lakes. *Hydrobiologia* 844, 83–103.
- Suhaimi, H., Yuslan, A., Ikhwanuddin, M., Yusoff, F. M., Mazlan, A. G., Habib, A. & Rasdi, N. W. (2022). Effect of diet on productivity and body composition of *Moina macrocopa* (Straus, 1820) (Branchiopoda, Cladocera, Anomopoda). *Crustaceana*, 95(1), 1-28.
- Sultana, S., Awal, S., Shaika, N. A. & Khan, S. (2022). Cyanobacterial blooms in earthen aquaculture ponds and their impact on fisheries and human health in Bangladesh. *Aquaculture Research*, 53(15), 5129-5141.
- Suminto, Chilmawati, D., Susilowati, T. & Adhinugroho, I. (2019). The effects of microalgal diet with enrichment of fermented organic matters (tofu waste, rice bran and fish meal) on growth and reproduction of *Diaphanosoma brachyurum*. *IOP Conference Series: Earth and Environmental Science*, 246 (1), 012036.
- Sun, C., Wang, S., Wang, H., Hu, X., Yang, F., Tang, M. & Zhong, J. (2022). Internal nitrogen and phosphorus loading in a seasonally stratified reservoir: Implications for eutrophication management of deep-water ecosystems. *Journal of Environmental Management*, 319, 115681.
- Syukri, F., Hassan, N. H. & Ani, A. S. N. (2022). Performances of FS Feed, *Artemia* Nauplii and Commercial. *Journal of Sustainability Science and Management*, 17(2), 35-45.
- Tachihana, S., Nagao, N., Katayama, T., Yusoff, F. M., Banerjee, S., Shariff, M. & Furuya, K. (2023). High productivity of fucoxanthin and eicosapentaenoic acid in a marine diatom *Chaetoceros gracilis* by perfusion culture under high irradiance. *Algal Research*, 72, 103123.

- Tadeo, A. J. D. & Veracruz, E. M. (2018). Larval rearing of giant gourami, *Osphronemus goramy* Lacépède 1801 fed with different live food organisms. *Asian Fisheries Science*, 31(2), 113-126.
- Taib, M., Zaki S., Azani N., Kama, A. H. M., Manan T. S. & Rasdi N. W. (2022). The effect of enriched *Artemia* sp. on growth, nutritional composition, and survival performance of *Macrobrachium rosenbergii* (giant freshwater prawn). *Egyptian Journal of Aquatic Biology and Fisheries*, 26(3), 625-635.
- Taipale, S. J., Brett, M. T., Pulkkinen, K. & Kainz, M. J. (2012). The influence of bacteria-dominated diets on *Daphnia magna* somatic growth, reproduction, and lipid composition. *FEMS Microbiology Ecology*, 82(1), 50-62.
- Taipale, S. J., Ventelä, A. M., Litmanen, J. & Anttila, L. (2022). Poor nutritional quality of primary producers and zooplankton driven by eutrophication is mitigated at upper trophic levels. *Ecology and Evolution*, 12(3), e8687.
- Tang, C., Yi, Y., Yang, Z., Zhou, Y., Zerizghi, T., Wang, X. & Duan, P. (2019). Planktonic indicators of trophic states for a shallow lake (Baiyangdian Lake, China). *Limnologica*, 78, 125712.
- Tang, Y., Su, L., Xu, R., Wang, S., Su, Y., Liu, Z. & Jeppesen, E. (2023). Response of zooplankton to inputs of terrestrial dissolved organic matter: Food quality constraints induced by microbes. *Limnology and Oceanography*, 68(3), 709-722.
- Tanvir, R. U., Hu, Z., Zhang, Y. & Lu, J. (2021). Cyanobacterial community succession and associated cyanotoxin production in hypereutrophic and eutrophic freshwaters. *Environmental Pollution*, 290, 118056.
- Tellenbach, C., Tardent, N., Pomati, F., Keller, B., Hairston Jr, N. G., Wolinska, J. & Spaak, P. (2016). Cyanobacteria facilitate parasite epidemics in *Daphnia*. *Ecology*, 97(12), 3422-3432.
- Thor, P., Vermandele, F., Bailey, A., Guscelli, E., Loubet-Sartrou, L., Dupont, S. & Calosi, P. (2022). Ocean acidification causes fundamental changes in the cellular metabolism of the Arctic copepod *Calanus glacialis* as detected by metabolomic analysis. *Scientific Reports*, 12(1), 22223.
- Titocci, J. & Fink, P. (2022). Food quality impacts on reproductive traits, development and fatty acid composition of the freshwater calanoid copepod *Eudiaptomus* sp. *Journal of Plankton Research*, 44(4), 528-541.
- Toruan, R. L. (2021). Zooplankton diversity in Lake Tondano, Indonesia. *IOP Conference Series: Earth and Environmental Science*, 744 (1), 012092.

- Tresguerres, M., Kwan, G. T. & Weinrauch, A. (2023). Evolving views of ionic, osmotic, and acid–base regulation in aquatic animals. *Journal of Experimental Biology*, 226(14).
- Tudi, M., Daniel Ruan, H., Wang, L., Lyu, J., Sadler, R., Connell, D. & Phung, D. T. (2021). Agriculture development, pesticide application and its impact on the environment. *International journal of environmental research and public health*, 18(3), 1112.
- Turcihan, G., Isinibilir, M., Zeybek, Y. G. & Eryalçın, K. M. (2022). Effect of different feeds on reproduction performance, nutritional components, and fatty acid composition of cladocera water flea (*Daphnia magna*). *Aquaculture Research*, 53(6), 2420-2430.
- Tyrell, A. S., Fisher, N. S. & Fields, D. M. (2020). Separating thermal and viscous effects of temperature on copepod respiration and energy budget. *The Biological Bulletin*, 239(1), 62-71.
- Umi, W. A. D., Yusoff, F. M., Aris, A. Z. & Sharip, Z. (2018). Rotifer community structure in tropical lakes with different environmental characteristics related to ecosystem health. *Journal of Environmental Biology*, 39(5), 795-807.
- Umi, W. A. D., Yusoff, F. M., Aris, A. Z., Sharip, Z. & Sinev, A. Y. (2020). Planktonic microcrustacean community structure varies with trophic status and environmental variables in tropical shallow lakes in Malaysia. *Diversity*, 12(9), 322.
- Valencia-Vargas, M. A., Nandini, S. & Sarma, S. S. S. (2020). Demographic characteristics of two freshwater cyclopoid copepods in Mexico, fed a plankton diet: the native *Mesocyclops longisetus* Thiébaud and the invasive *Mesocyclops pehpeinsis* Hu. *Inland Waters*, 10(1), 128-136.
- Vidya, P. V., Rajathy, S., Kumar, R., Maneesh Kumar, S. K., Akhilesh, K. B., Mohan, R., & Sudha, A. (2023). Geochemical partitioning of sediment-bound phosphorous in Thrissur Kole Wetlands, Southwest India: Status of bioavailability and trophic state. *International Journal of Environmental Science and Technology*, 1-16.
- Vilar, M. C. P., da Costa Pena Rodrigues, T. F., da Silva Ferrão-Filho, A. & de Oliveira e Azevedo, S. M. F. (2021). Grazer-induced chemical defense in a microcystin-producing *Microcystis aeruginosa* (Cyanobacteria) exposed to *Daphnia gessneri* infochemicals. *Journal of Chemical Ecology*, 47, 847-858.
- Villa, J. A., Ray, E. E. & Barney, B. M. (2014). *Azotobacter vinelandii* siderophore can provide nitrogen to support the culture of the green algae *Neochloris oleoabundans* and *Scenedesmus* sp. BA032. *FEMS Microbiology Letters*, 351(1), 70-77.

- Walczyska, A. & Sobczyk, M. (2022a). Body size response to hypoxia in experimental evolution and the post-evolution plastic body size response to different temperature in *Lecane inermis* rotifer.
- Walczyska, A. & Sobczyk, M. (2022b). Experimental evolution shows body size decrease in response to hypoxia, with a complex effect on plastic size response to temperature. *The Biological Bulletin*, 243(2), 272-281.
- Wang, H., Huo, T., Du, X., Wang, L., Song, D., Huang, X. & Zhao, C. (2022a). Zooplankton community and its environmental driving factors in Ulungur Lake, China. *Journal of Freshwater Ecology*, 37(1), 387-403.
- Wang, H., Molinos, J. G., Heino, J., Zhang, H., Zhang, P. & Xu, J. (2021). Eutrophication causes invertebrate biodiversity loss and decreases cross-taxon congruence across anthropogenically-disturbed lakes. *Environment International*, 153, 106494.
- Wang, J., Lu, J., Zhang, Z., Han, X., Zhang, C. & Chen, X. (2022b). Agricultural non-point sources and their effects on chlorophyll *a* in a eutrophic lake over three decades (1985–2020). *Environmental Science and Pollution Research*, 29(31), 46634-46648.
- Wang, Y., Ning, W., Han, M., Gao, C., Guo, W., Chang, J. S. & Ho, S. H. (2023). Algae-mediated bioremediation of ciprofloxacin through a symbiotic microalgae-bacteria consortium. *Algal Research*, 71, 103062.
- Weber, A. K., & Pirow, R. (2009). Physiological responses of *Daphnia pulex* to acid stress. *BMC physiology*, 9, 1-25.
- Weinstock, J. B., Vargas, L. & Collin, R. (2022). Zooplankton abundance reflects oxygen concentration and dissolved organic matter in a seasonally hypoxic estuary. *Journal of Marine Science and Engineering*, 10(3), 427.
- Wenzel, A., Vrede, T., Jansson, M. & Bergström, A. K. (2021). *Daphnia* performance on diets containing different combinations of high-quality algae, heterotrophic bacteria, and allochthonous particulate organic matter. *Freshwater Biology*, 66(1), 157-168.
- Xiao, J., Li, Q. Y., Tu, J. P., Chen, X. L., Chen, X. H., Liu, Q. Y. & Wang, H. L. (2019). Stress response and tolerance mechanisms of ammonia exposure based on transcriptomics and metabolomics in *Litopenaeus vannamei*. *Ecotoxicology and environmental safety*, 180, 491-500.
- Xiong, W., Ni, P., Chen, Y., Gao, Y., Li, S. & Zhan, A. (2019). Biological consequences of environmental pollution in running water ecosystems: A case study in zooplankton. *Environmental Pollution*, 252, 1483-1490.
- Xu, S., Jiang, Y., Liu, Y. & Zhang, J. (2021). Antibiotic-accelerated cyanobacterial growth and aquatic community succession towards the formation of cyanobacterial bloom in eutrophic lake water. *Environmental Pollution*, 290, 118057.

- Xue, W., Jin, J., Zhang, F., Chen, H., Yang, D., Zhang, Y. & Wei, W. (2022). *Bosmina fatalis* adapting to ammonia through oxidative stress and ribosome increase. *Journal of Freshwater Ecology*, 37(1), 117-129.
- Yang, Z., Lü, K., Chen, Y. & Montagnes, D. J. (2012). The interactive effects of ammonia and microcystin on life-history traits of the cladoceran *Daphnia magna*: Synergistic or antagonistic?. *PLoS One*, 7(3), e32285.
- Yin, X. W. & Niu, C. J. (2008). Effect of pH on survival, reproduction, egg viability and growth rate of five closely related rotifer species. *Aquatic Ecology*, 42, 607-616.
- Yousuf, S., Tyagi, A. & Singh, R. (2022). Probiotic supplementation as an emerging alternative to chemical therapeutics in finfish aquaculture: A Review. *Probiotics and Antimicrobial Proteins*, 1-18.
- Yu, B., Lyu, K., Li, J., Yang, Z. & Sun, Y. (2022). Combined toxic effects of nitrite and ammonia on life history traits of *Daphnia pulex*. *Frontiers in Environmental Science*, 10, 1019483.
- Yufera, M. & Darias, M.J. (2007). The onset of exogenous feeding in marine fish larvae. *Aquaculture* 268:53–63.
- Yunandar, Y., Effendi, H., Widiatmaka, W., & Setiawan, Y. (2021). Spatial distribution and structure of plankton in Paminggir Swamp of the Hulu Sungai Utara Regency, South Kalimantan Indonesia. *IOP Conference Series: Earth and Environmental Science*, 869, 012026
- Yuslan, A., Suhaimi, H., Taufek, H. M. & Rasdi, N. W. (2022). Effect of bio-organic fertilizer and agro-industrial residue on the growth and reproduction of cyclopoid copepod, *Oithona rigida* (Giesbrecht, 1896). *International Journal of Aquatic Biology*, 10(2), 151-168.
- Yusof Hanan, M., Amatul-Samahah, M. A., Jaapar, M. Z., Ramli, N. S. F., & Mohamad, S. N. (2023). *Moina* sp. as Artemia replacement in the larval rearing of river catfish, *Pangasius nasutus* (Bleeker, 1863). *Journal of Applied Aquaculture*, 1-17.
- Zahoor, I. & Mushtaq, A. (2023). Water pollution from agricultural activities: A critical global review. *International Journal of Chemical and Biochemical Science*, 23, 164-176.
- Zeis, B., Buchen, I., Wacker, A. & Martin-Creuzburg, D. (2019). Temperature-induced changes in body lipid composition affect vulnerability to oxidative stress in *Daphnia magna*. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 232, 101-107.
- Zhang, H., Hollander, J. & Hansson, L. A. (2017). Bi-directional plasticity: rotifer prey adjust spine length to different predator regimes. *Scientific Reports*, 7(1), 10254.

- Zhang, L., Fang, W., Li, X., Lu, W. & Li, J. (2020). Strong linkages between dissolved organic matter and the aquatic bacterial community in an urban river. *Water Research*, 184, 116089.
- Zhang, S., Lu, W., Zhou, Z. & Chen, W. (2022). Spatial differences in zooplankton community structure between two fluvial lakes in the middle and lower reaches of the Yangtze River: Effects of land use patterns and physicochemical factors. *Diversity*, 14(11), 908.
- Zhang, Y., Feng, S., Zhu, L., Li, M. & Xiang, X. (2023a). Population dynamics of *Brachionus calyciflorus* driven by the associated natural bacterioplankton. *Frontiers in Microbiology*, 13, 1076620.
- Zhang, Y., Whalen, J. K., Cai, C., Shan, K. & Zhou, H. (2023b). Harmful cyanobacteria-diatom/dinoflagellate blooms and their cyanotoxins in freshwaters: A nonnegligible chronic health and ecological hazard. *Water Research*, 119807.
- Zhang, Y., Zhang, H., Zhang, Z., Liu, C., Sun, C., Zhang, W. & Marhaba, T. (2018). pH effect on heavy metal release from a polluted sediment. *Journal of Chemistry*, 2018, 1-7.
- Zhao, K., Wang, L., You, Q., Pan, Y., Liu, T., Zhou, Y. & Wang, Q. (2021). Influence of cyanobacterial blooms and environmental variation on zooplankton and eukaryotic phytoplankton in a large, shallow, eutrophic lake in China. *Science of The Total Environment*, 773, 145421.
- Zhao, L., Xu, Y. & Lai, X. (2018). Antagonistic endophytic bacteria associated with nodules of soybean (*Glycine max L.*) and plant growth-promoting properties. *Brazilian Journal of Microbiology*, 49, 269-278.
- Zhu, X., Wang, Q., Zhang, L., Liu, J., Zhu, C. & Yang, Z. (2015). Offspring performance of *Daphnia magna* after short-term maternal exposure to mixtures of microcystin and ammonia. *Environmental Science and Pollution Research*, 22, 2800-280