



**PARP INHIBITOR COMBINATIONS WITH OLAPARIB FOR SYNERGISTIC
EFFICACY IN OLAPARIB-RESISTANT TRIPLE-NEGATIVE BREAST
CANCER CELLS**

By

NUR AININIE BINTI YUSOH

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Chair : Haslina Ahmad, PhD
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Triple-negative breast cancer (TNBC) is associated with aggressive and heterogenous tumour phenotype, early tumour relapse and poor prognosis than other types of invasive breast cancer. TNBC does not respond to endocrine therapy due to the lack of oestrogen receptor (ER), progesterone receptor (PR) and human epidermal growth factor receptor type 2 (HER2) expression. Although poly(ADP-ribose) polymerase (PARP) inhibitors (PARPi) have been successfully developed to treat breast cancer; however, these PARPi as monotherapies are limited to TNBC patients with defective gene where BRCA-deficient TNBC accounts for a small subset (< 30%) of TNBC patients. With few exceptions, monotherapy remains ineffective due to insufficient tumour suppression, requires high dose of drug, dose-limiting toxicity and acquired drug resistance is common. Thus, combining PARPi with other therapeutics to increase efficacy, reduce drug dosage, and overcome drug resistance is a promising strategy to treat TNBC. Moreover, drug combination screening of a PARPi is a relatively new concept and having a compound library with diverse chemical space to be screened alongside PARPi holds potential to new drug combination discovery. Therefore, this study aims to discover new combinations of Olaparib, the most successful PARPi to date, alongside other therapeutic in MDA-MB-231 TNBC cells without BRCA mutation. The library used in this screen comprises of a total of > 100 molecules including DNA-binding ruthenium compounds and commercially-available drugs. In this study, Olaparib-resistant (OlaR) MDA-MB-231R cells were developed from the parental MDA-MB-231 cells after prolonged treatment (~8 months) with Olaparib. Olaparib resistance was assessed by clonogenic survival assay in which 28-fold level of Olaparib resistance was observed in MDA-MB-231R cells in comparison to the parental cells. The acquisition of Olaparib resistance is associated with upregulation of drug efflux pumps of P-glycoprotein (P-gp), loss of poly(ADP-ribose) glycohydrolase (PARG) and the activation of ATR/Chk1 signalling pathway. Synergistic drug combinations were identified using Chou and Talalay

combination index (CI) method in which $CI < 0.9$ demonstrated synergy. This study identified four hits including a natural compound, curcumin; two ruthenium(II)-rhenium(I) metallomacrocycles, Ru(bpy)Re and Ru(dppz)Re; and FDA-approved Exemestane ($CI < 0.9$ across the tested concentrations). Synergy was confirmed using long-term clonogenic survival assay in which single agents showed survival fractions (S.F.) of $> 45\%$, and upon co-treatments, significant ($P < 0.01$) loss in clonogenic potential of cells was observed. Moreover, synergy was retained in the acquired OlaR MDA-MB-231R cells. Mechanistic studies indicated synergy was achieved via significant ($P < 0.05$) enhancement of DNA damage and the resultant apoptosis. Most importantly, all identified combinations showed low cytotoxicity in MCF10A normal breast cells with substantial cancer versus normal cell selectivity (selectivity index (SI) > 50). These identified combinations were also able to inhibit the growth of 3D breast cancer spheroids. Thus, this study supports the concept that PARPi Olaparib combination strategy represents a promising approach towards aggressive strains of BRCA-proficient TNBC and can overcome acquired PARPi resistance. Additionally, co-treatment with the synergistic pairing of Olaparib and Ru-PIP, a ruthenium compound, in non-small cell lung cancer (NSCLC) cells showed enhanced ($P < 0.05$) DNA damages and reactive oxygen species (ROS) levels, and was able to inhibit the lung cancer spheroids growth, providing further affirmation of expanding the benefits of these identified combinations to other cancer types beyond TNBC. Moreover, exposure of zebrafish embryos to these identified synergistic pairings over 96 h did not lead to any noticeable signs of toxicity, making these newly identified combinations as suitable candidates for future *in vivo* investigations.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**RAWATAN GABUNGAN PERENCAT PARP OLAPARIB YANG MEMBERI
KESAN SINERGI TERHADAP SEL KANSER PAYUDARA *TRIPLE-NEGATIF*
YANG MEMPUNYAI RINTANGAN TERHADAP OLAPARIB**

Oleh

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Kanser payudara *triple-negatif* (TNBC) telah dikaitkan dengan fenotip tumor yang agresif, kepelbagaian tumor, kanser kambuh semula dan mempunyai prognosis yang buruk berbanding kanser payudara bersifat invasif yang lain. TNBC tidak memberi tindak balas terhadap terapi endokrin kerana kanser ini tidak mempunyai ekspresi reseptor progesterone, reseptor estrogen dan reseptor faktor pertumbuhan epidermis manusia jenis 2 (HER2). Walaupun perencat polimerase poli(ADP-ribosa) (PARP) telah berjaya dibangunkan untuk merawat kanser payudara, tetapi keberkesanan aktiviti PARPi sebagai agen tunggal adalah terhad kepada pesakit kanser yang mempunyai mutasi pada gen BRCA dimana bilangan pesakit yang mempunyai mutasi pada gen BRCA adalah sangat kecil (< 30% daripada pesakit TNBC). Tambahan pula, rawatan agen tunggal masih tidak berkesan untuk merawat kanser disebabkan tidak dapat membunuh sel-sel kanser dengan sepenuhnya, memerlukan dos ubat yang tinggi, ketoksikan ubat, dan terdapat risiko berlakunya rintangan terhadap agen tunggal. Oleh itu, strategi menggunakan rawatan gabungan PARPi yang dapat memperluas kesan keberkesanan, mengurangkan dos ubat dan dapat mengatasi masalah rintangan ubat merupakan strategi yang berpotensi untuk merawat TNBC. Di samping itu, rawatan gabungan PARPi adalah satu konsep baharu dan dengan penggunaan agen yang pelbagai menjadikan strategi ini berpotensi tinggi untuk menemukan rawatan gabungan yang baharu. Oleh itu, kajian ini bertujuan untuk menemukan rawatan gabungan Olaparib, salah satu PARPi yang paling berpotensi buat masa sekarang, bersama agen yang lain terhadap sel kanser MDA-MB-231 yang tidak mempunyai mutasi pada gen BRCA. Agen yang digunakan dalam kajian ini berjumlah sebanyak > 100 agen, yang terdiri daripada agen antikanser yang telah berada di pasaran, dan kompleks rutenium yang mengikat dengan DNA. Dalam kajian ini, sel MDA-MB-231 yang menunjukkan rintangan terhadap Olaparib telah berjaya dibangunkan daripada sel MDA-MB-231 di mana sel tersebut telah dirawat secara berpanjangan dengan ubat Olaparib (~8 bulan). Berbanding sel MDA-MB-231,

sel MDA-MB-231R menunjukkan tahap rintangan sebanyak 28 kali ganda terhadap agen Olaparib apabila menggunakan ujian klonogenik. Rintangan terhadap agen Olaparib juga dikaitkan dengan peningkatan jumlah pam efluks P-glikoprotein (P-gp), pengurangan aktiviti *glycohydrolase* poli(ADP-ribosa) (PARG) dan peningkatan aktiviti ATR/Chk1. Gabungan ubat-ubatan yang menghasilkan kesan sinergistik dinilai dengan menggunakan kaedah Chou dan Talalay kombinasi indeks (CI) di mana $CI < 0.9$ menunjukkan kesan sinergistik. Hasil dapatan kajian ini telah dapat mengenal pasti empat *hits* yang terdiri daripada bahan aktif daripada kunyit yang dikenali sebagai kurkumin; dua *macrocytes* yang berasaskan logam rutenium(II)-renium(I), Ru(bpy)Re dan Ru(dppz)Re; dan ubat yang telah mendapat kelulusan FDA iaitu Exemestane. Seterusnya, kesan sinergistik rawatan gabungan baharu ini juga dikaji dengan menggunakan ujian klonogenik. Agen tunggal menunjukkan kelangsungan hidup sel (S.F.) $> 45\%$, dan rawatan gabungan menunjukkan pengurangan kelangsungan hidup sel yang ketara ($P < 0.01$). Di samping itu, rawatan gabungan ini juga menunjukkan kesan sinergistik terhadap sel MDA-MB-231R. Kajian mekanistik menunjukkan kesan sinergistik telah diperolehi melalui peningkatan kerosakan DNA yang ketara ($P < 0.05$) yang menyebabkan kematian sel melalui mekanisme *apoptosis*. Pertama sekali, semua rawatan gabungan baharu yang dikenal pasti menunjukkan kesan sitotoksik yang rendah terhadap sel payudara normal MCF10A dimana rawatan ini menunjukkan tahap selektif yang tinggi terhadap sel kanser berbanding sel normal (nilai indeks selektif (SI) > 50). Rawatan gabungan baharu yang dikenal pasti ini juga turut berjaya menghalang pertumbuhan sferoid 3D kanser payudara. Oleh itu, kajian ini menyokong konsep dimana rawatan gabungan PARPi Olaparib berpotensi untuk merawat kanser TNBC yang agresif termasuklah kanser yang tidak mempunyai mutasi pada gen BRCA, dan rawatan ini dapat mengatasi masalah rintangan ubat PARPi. Tambahan lagi, rawatan gabungan Olaparib dan kompleks rutenium Ru-PIP yang menunjukkan kesan sinergistik terhadap sel kanser paru-paru (NSCLC) menunjukkan peningkatan kerosakan DNA yang ketara ($P < 0.05$) dan peningkatan spesies oksigen reaktif (ROS), dan juga dapat menghalang pertumbuhan sferoid 3D kanser paru-paru. Ini memberi justifikasi yang kukuh untuk memperluas manfaat rawatan gabungan yang telah dikenal pasti ini untuk digunakan terhadap jenis kanser yang lain selain TNBC. Selain itu, melalui ujian ketoksikan embrio ikan Zebra dimana walaupun rawatan gabungan baharu ini telah diberikan kepada embrio ikan Zebra selama 96 jam, tiada tanda kesan ketoksikan yang ketara yang ditunjukkan oleh rawatan tersebut. Ini menjadikan rawatan gabungan baharu ini sebagai rawatan yang sesuai untuk dikaji dalam kajian lanjutan secara *in vivo*.

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LIST OF ABBREVIATIONS

°C	degree celsius
3-AB	3-aminobenzamide
ABCB1	ATP-binding cassette sub-family B member 1
AI	Aromatase inhibitor
ALC1	Amplified in Liver Cancer 1
ANOVA	One-way analysis of variance
ATCC	American Type Culture Collection
ATM	Ataxia-telangiectasia mutated
ATR	Ataxia-telangiectasia-mutated-and-Rad3-related kinase
ATRi	ATR inhibitor
BCA	Bicinchoninic acid
BER	Base excision repair
bpy	2,2'-bipyrene
BRCA	Breast cancer susceptibility genes
BSA	Bovine serum albumin
Chk1	Checkpoint kinase 1
CI	Combination index
CO ₂	Carbon dioxide
COVID-19	Coronavirus disease 2019
CUR	Curcumin
DCF	2',7'-dichlorofluorescein
DCFDA	2',7'-dichlorofluorescein diacetate
DDB2	DNA damage-binding protein 2
DDR	DNA damage response

dmb	4,4'-dimethyl-2,2'-bipyridine
DMEM	Dulbecco's modified Eagle's medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
dppz	dipyrido[3,2-a:2',3'-c]phenazine
DSB	Double-strand breaks
EGFR	Epidermal growth factor receptor
ER	Estrogen receptor
ER β	Oestrogen receptor beta
EXE	Exemestane
FBS	Fetal bovine serum
FDA	U.S. Food and Drug Administration
HDACi	Histone deacetylase inhibitors
hEGF	Recombinant human EGF
HER2	Human epidermal growth factor receptor type 2
hpf	Hours of post-fertilisation
HR	Homologous recombination
HRP	Horseradish peroxidase
IACUC	Institutional Animal Care and Use Committee
IC ₅₀	Half maximal inhibitory concentration
Im	Imidazole
Kb	Binding constant
KP1019	indazolium trans-tetrachlorobis(1H-indazole)ruthenate(III)
LC ₅₀	Half maximal lethal concentration
LIG3	DNA ligase III

LL'	2-((2',2'':5'',2'''-terthiophene)-imidazo[4,5-f][1,10]phenanthroline)
log P	Octanol/water partition coefficient
MCE	MedChemExpress
MDA-MB-231R	Olaparib-resistant MDA-MB-231 cells
MDR1	Multi drug resistance protein 1
mTOR	Mammalian target of rapamycin
MTT	Thiazolyl blue tetrazolium bromide
NAMI-A	ImH][trans-RuCl ₄ (DMSO)(Im)
NCI	National Cancer Institute
NER	Nucleotide excision repair
NHEJ	Non-homologous end joining
NSCLC	Non-small cell lung cancer cells
OLAP	Olaparib
OlaR	Olaparib-resistant
ORR	Objective response rates
P/S	Penicillin/streptomycin
PAIN	Pan-assay interference
PARG	Poly(ADP-ribose) glycohydrolase
PARP	Poly(ADP-ribose) polymerase
PARPi	PARP inhibitor
p-ATM	ATM phosphorylated at Ser1981
p-ATR	ATR phosphorylated at Thr1989
PBS	Phosphate buffered saline
p-Chk1	Chk1 phosphorylated at Ser345
PDT	Photodynamic therapy

PDX	Patient-derived tumour xenograft
PFA	Paraformaldehyde
PFS	Progression-free survival
P-gp	p-glycoprotein
phen	1,10-phenanthroline
p-HPIP	2-(4-hydroxyphenyl)imidazo[4,5-f][1,10]phenanthroline
PI	Propidium iodide
PIP	2-(phenyl)imidazo[4,5-f][1,10]phenanthroline
Polβ	DNA polymerase β
PR	Progesterone receptor
qtpy	2,2':4,4''4',4''' quaterpyridyl
RI	Resistance index
RIPA	Radioimmunoprecipitation assay
ROS	Reactive oxygen species
RPC	Ruthenium polypyridyl complex
RT	Room temperature
BR	Ru(bpy)Re
DR	Ru(dppz)Re
Ru-PIP	[Ru(dppz) ₂ (PIP)] ²⁺
S.F.	Survival fraction
SCLC	Small cell lung cancer
SD	Standard deviation
SDS-PAGE	Sodium dodecyl sulphate-polyacrylamide gel
SERM	Selective ER modulator
SI	Selectivity Index

siRNA	Small-interfering RNA
SSB	Single-strand breaks
T2	Cell doubling time
TBS	Tris buffered saline
TBS-T	0.1% Tween 20 in 1X TBS
TI	Therapeutic index
TLD1433	[Ru(dmb) ₂ (LL')] ²⁺
TNBC	Triple-negative breast cancer
UPM	Universiti Putra Malaysia
UV	Ultraviolet light
VEGF	Vascular endothelial growth factor
vs.	versus
XRCC1	X-ray repair cross complementing protein 1
γH2AX	H2AX phosphorylated at Ser139

CHAPTER 1

INTRODUCTION

1.1 Research background

Triple-negative breast cancer (TNBC) is a subtype of breast cancer which is characterised by the lack of oestrogen receptor (ER), progesterone receptor (PR) and human epidermal growth factor receptor type 2 (HER2) expression (Almansour, 2022; Zagami & Carey, 2022). TNBC accounts for approximately 15-20% of all breast cancer cases and nearly 12-18% of Malaysian breast cancer patients are TNBC (Abdul Aziz et al., 2020; Almansour, 2022). Due to the heterogenous nature of TNBC together with high metastatic potential and high invasiveness, TNBC is associated with early tumour relapse leading to poor prognosis compared to non-TNBC subtypes (Bianchini et al., 2022). Despite successful development of advanced therapies for breast cancers, limited treatment options are available for TNBC patients. In contrast to non-TNBC patients with ER, PR and/or HER2 expression that are more susceptible to therapies targeting these receptors (endocrine therapies), these therapies are not effective for TNBC patients which lack these ER, PR and HER2 receptors (Yin et al., 2020).

Nowadays, with greater understanding and insights on the hallmarks of cancer including genomic instability have led to some important advances and breakthroughs in treating TNBC (Huang & Zhou, 2021; Groelly et al., 2023). Cellular DNA is continuously being subjected to various endogenous damage such as reactive oxygen species (ROS) generated from cellular metabolism, and exogenous damages such as ultraviolet light (UV), ionising radiation or DNA damaging chemicals, that can be detrimental to cells. Thus, in response to the accumulation of DNA lesion, cancer cells have evolved numerous complicated and entangled network of DNA damage response (DDR) signalling pathways to repair DNA lesion to ensure cell survival (Alhmoud et al., 2020; Huang & Zhou, 2021; Nikfarjam & Singh, 2023). Based on this understanding, the development of targeted therapeutics in particular, the inhibitors to DDR, have resulted in a paradigm shift in small molecule-based cancer therapy (Huang & Zhou, 2021; Huang et al., 2022a). This is perhaps best exemplified by the development of poly(ADP-ribose) polymerase (PARP) inhibitors (PARPi) which have high selectivity towards cancers harbouring mutations in breast cancer susceptibility (BRCA) genes, achieved by the concept of synthetic lethality (Mateo et al., 2019; Curtin & Szabo, 2020). This cancer selectivity is achieved through the exploitation of cancer cells with impaired DNA repair characteristics together with targeting the compensatory DNA repair pathways (Bryant et al., 2005; Lord & Ashworth, 2017). Briefly, PARP enzyme is crucial for DNA single-strand breaks (SSBs) repair; meanwhile, BRCA1/2 genes are required for DNA double-strand breaks (DSBs) repair by homologous recombination (HR), and the simultaneous impairment of both repair mechanisms leads to cancer selective cell death

through synthetic lethality. Several PARPi have gained U.S. Food and Drug Administration (FDA) approval for treating BRCA-deficient cancers, with prominent examples include Olaparib, niraparib, rucaparib and talazoparib (Curtin & Szabo, 2020). However, BRCA-deficient cancers account for a relatively small subset (< 30%) of TNBC patients which substantially limits the population of TNBC patients that could benefit from PARPi-based therapy (Chen et al., 2018). Besides, PARPi monotherapies may not be effective for BRCA-proficient TNBC.

Moreover, with very few exceptions, anticancer monotherapies including PARPi single agents are clinically limited due to insufficient tumour suppression, intolerable side effects and the prevalence of acquired PARPi resistance, in which any of these occurrences will result in treatment failure (Mokhtari et al., 2017; Toss et al., 2022). Nowadays, drug resistance remains a major therapeutic challenge to successful cancer therapy. Besides, TNBC is heterogenous in nature and exhibits inherently aggressive clinical behaviour with significantly higher risk of relapse compared to non-TNBC subtypes. Based on this background, adopting the therapeutic approach to combine PARPi with other therapeutics for synergy to enhance drug efficacy and overcome acquired PARPi resistance is a promising approach in treating the aggressive and heterogenous cancer BRCA-proficient TNBC (Wang et al., 2022). Moreover, reduced drug dosage as a result of synergistic response is predicted to represent a lower risk in a clinical setting by minimising adverse effects (Mokhtari et al., 2017). Newly identified combinations also need to have a high cancer selectivity, as an indication of margin of safety for future clinical investigations. Accordingly, drug combination will result in manageable toxicity profile, higher survival rate and improve the cost-effectiveness of the treatment over monotherapies.

Typically, a rational combination strategy is the most common method to establish new effective drug combination in which complimentary small molecules were selected based on their mechanisms of action (Falchi et al., 2017; Kim et al., 2020a). However, unbiased (hypothesis-free) drug combination screen may allow for serendipity in drug discovery (Kummar et al., 2010; Al-Lazikani et al., 2012). Moreover, screening a compound library with diverse chemical space aided by the inclusion of the DNA-binding ruthenium compounds holds the potential to new drug combination discovery with different chemical reactivities. The interest in metal-based anticancer drug is based on the discovery of cis-diamminedichloroplatinum (II), or cisplatin, by the Rosenberg group in 1960 (Rosenberg et al., 1969), where cisplatin becomes the first FDA-approved metallodrug in 1978 (Kelland, 2007). Since then, extensive efforts were made in designing complexes based on other alternative transition metal centres including ruthenium to replace or as an alternative to cisplatin (Zeng et al., 2017; Lee et al., 2020). Several ruthenium compounds have successfully entered clinical trials including NAMI-A, KP1019, NKP1339 and until now, the most successful clinical ruthenium compound is TLD1433 which has entered phase II trial to treat bladder cancer patients (Trondl et al., 2014; Monro et al., 2019). Interestingly, some ruthenium compounds target DNA through intercalation (reversible), a unique and different binding mode compared to the covalent binding showed by cisplatin (Zeng et al., 2017; Rilak Simović et al.,

2019). Briefly, these ruthenium compounds intercalate DNA inducing structural changes to the DNA strand by lengthening of the DNA strand or twisting of the base pairs resulting to its functional arrest. For example, our group have developed the ruthenium(II) polypyridyl complex (RPC) of $[\text{Ru}(\text{dppz})_2(\text{PIP})]^{2+}$ (dppz = dipyrdo[3,2-a:2',3'-c]phenazine and PIP = 2-(phenyl)imidazo[4,5-f][1,10]phenanthroline) (or Ru-PIP) which intercalates DNA with high affinity (Gill et al., 2016). The addition of Ru-PIP led to the stalling of DNA replication fork progression resulting in the activation of DDR and G1 cell cycle arrest (Gill et al., 2016).

Previously, our group described the potential of combining Ru-PIP with PARPi Olaparib in breast cancer cells where strong synergy was observed as a result of complementary mechanisms of action, with low cytotoxicity in normal healthy cells (Yusoh et al., 2020b). For this reason, ruthenium compounds have appeared as suitable and attractive candidates for exploration in combination therapy for cancer. Moreover, the combination of ruthenium compounds with DDR-targeting agents for cancer is still relatively a new concept. Therefore, having these emerging DNA-binding ruthenium compounds in the library provides chemical novelty to this drug combination screen as this library uses much diverse chemical space than the small molecules that are typically used in PARPi combination strategy. In this study, the most successful PARPi to date, Olaparib, will be screened alongside a compound library comprising of commercially available drugs and ruthenium compounds in BRCA-proficient TNBC. This approach may expand the applications of PARPi in BRCA-proficient cancers or other cancers with different sources of genomic instability and overcome acquired drug resistance.

1.2 Problem statement

Although PARPi have successfully been developed for BRCA-deficient TNBC cancers based on synthetic lethality concept; however, BRCA-deficient cancers accounts for a small subset (< 30%) of TNBC patients (Chen et al., 2018). Besides, PARPi monotherapies may not be effective for BRCA-proficient TNBC. Moreover, with very few exceptions, monotherapy was not able to prolong patient survival as single drug therapies may lead to insufficient tumour suppression and high dose of drug required for single therapeutic may cause intolerable side effects (Mokhtari et al., 2017). Furthermore, constant or prolonged exposure to single drug may lead to rapid emergence of acquired resistance which continues to be a challenge. For example, upon prolonged oral administration of PARPi single agents, more than 40% BRCA1/2-deficient patients did not respond to PARPi treatment indicating that they have acquired PARPi resistance (Li et al., 2020). Relapse is also frequent in a significant number of TNBC patients (occurs mostly in the first 5 years following diagnosis) which resulted in failure of the currently available treatments (Toss et al., 2022). Stewart et al. (2019) described that 40% of patients with stage I to III TNBC will suffer cancer recurrence after standard neoadjuvant chemotherapy and surgery (Stewart et al., 2019). Although PARPi combinations have been examined, these strategies are mainly effective in BRCA-deficient cancers (Lee et al., 2016; Fasching et al., 2021;

Drewett et al., 2022). This justifies the needs to find other effective PARPi combinations for BRCA-proficient cancers including for the aggressive cancer of TNBC and in the acquired PARPi-resistant tumours.

1.3 Aims and research objectives

This study aims to discover new synergistic combinations of PARPi Olaparib alongside other therapeutics from a compound library comprising of commercially available anticancer agents and ruthenium anticancer compounds in BRCA-proficient MDA-MB-231 TNBC cells including in Olaparib-resistant cells, and expand this approach to other cancer cell line beyond TNBC such as non-small cell lung cancer (NSCLC) cells.

- 1) To establish Olaparib-resistant (OlaR) MDA-MB-231R TNBC cells and determine the Olaparib resistance mechanisms.
- 2) To evaluate new synergistic combinations of Olaparib with a mixed organic/ruthenium compounds library in MDA-MB-231 cells, determine the mechanisms of synergy, evaluate their cytotoxic effects in OlaR MDA-MB-231R cells and in breast cancer spheroids and assess their acute toxicity in zebrafish embryos.
- 3) To evaluate new synergistic combinations of Olaparib with FDA-approved anticancer drugs library in MDA-MB-231 cells, determine the mechanisms of synergy and evaluate their cytotoxic effects in OlaR MDA-MB-231R cells.
- 4) To evaluate the previously identified synergistic combination of Olaparib with Ru-PIP, a ruthenium metalloc compound, in lung cancer cells, determine the mechanisms of synergy, evaluate their cytotoxic effects in lung cancer spheroids and assess their acute toxicity in zebrafish embryos.

REFERENCES

- Abdou, Y., Goudarzi, A., Yu, J. X., Upadhaya, S., Vincent, B., & Carey, L. A. (2022). Immunotherapy in triple negative breast cancer: beyond checkpoint inhibitors. *npj Breast Cancer*, 8(1), 121.
- Abdul Aziz, A. A., Md Salleh, M. S., & Ankathil, R. (2020). Clinicopathological and Prognostic Characteristics of Malaysian Triple Negative Breast Cancer Patients Undergoing TAC Chemotherapy Regimen. *Int J Breast Cancer*, 2020, 8424365.
- Ahel, D., Horejsí, Z., Wiechens, N., Polo, S. E., Garcia-Wilson, E., Ahel, I., Flynn, H., Skehel, M., West, S. C., Jackson, S. P., Owen-Hughes, T., & Boulton, S. J. (2009). Poly(ADP-ribose)-dependent regulation of DNA repair by the chromatin remodeling enzyme ALC1. *Science*, 325(5945), 1240-1243.
- Ahmad, H., Ghosh, D., & Thomas, J. A. (2014). Using ancillary ligands to tune the DNA binding properties of self-assembled luminescent metallomacrocycles. *Chemical Communications*, 50(29), 3859-3861.
- Al-Lazikani, B., Banerji, U., & Workman, P. (2012). Combinatorial drug therapy for cancer in the post-genomic era. *Nature Biotechnology*, 30(7), 679-692.
- Alcindor, T., & Beauger, N. (2011). Oxaliplatin: a review in the era of molecularly targeted therapy. *Curr Oncol*, 18(1), 18-25.
- Alessio, E., & Messori, L. (2019). NAMI-A and KP1019/1339, Two Iconic Ruthenium Anticancer Drug Candidates Face-to-Face: A Case Story in Medicinal Inorganic Chemistry. *Molecules*, 24(10).
- Alexandrova, E., Giurato, G., Saggese, P., Pecoraro, G., Lamberti, J., Ravo, M., Rizzo, F., Rocco, D., Tarallo, R., Nyman, T. A., Collina, F., Cantile, M., Di Bonito, M., Botti, G., Nassa, G., & Weisz, A. (2020). Interaction Proteomics Identifies ERbeta Association with Chromatin Repressive Complexes to Inhibit Cholesterol Biosynthesis and Exert An Oncosuppressive Role in Triple-negative Breast Cancer. *Mol Cell Proteomics*, 19(2), 245-260.
- Alhmoud, J. F., Woolley, J. F., Al Moustafa, A. E., & Malki, M. I. (2020). DNA Damage/Repair Management in Cancers. *Cancers (Basel)*, 12(4).
- Almansour, N. M. (2022). Triple-Negative Breast Cancer: A Brief Review About Epidemiology, Risk Factors, Signaling Pathways, Treatment and Role of Artificial Intelligence. *Front Mol Biosci*, 9, 836417.

- Alvarado-Miranda, A., Lara-Medina, F. U., Muñoz-Montaño, W. R., Zinser-Sierra, J. W., Galeana, P. A. C., Garza, C. V., Sanchez Benitez, D., Limón Rodríguez, J. A., Arce Salinas, C. H., Guijosa, A., & Arrieta, O. (2023). Capecitabine Plus Aromatase Inhibitor as First Line Therapy for Hormone Receptor Positive, HER2 Negative Metastatic Breast Cancer. *Curr Oncol*, 30(7), 6097-6110.
- Amaral, C., Augusto, T. V., Almada, M., Cunha, S. C., Correia-da-Silva, G., & Teixeira, N. (2020). The potential clinical benefit of targeting androgen receptor (AR) in estrogen-receptor positive breast cancer cells treated with Exemestane. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, 1866(5), 165661.
- Angelini, M., Teglia, F., Astolfi, L., Casolari, G., & Boffetta, P. (2023). Decrease of cancer diagnosis during COVID-19 pandemic: a systematic review and meta-analysis. *Eur J Epidemiol*, 38(1), 31-38.
- Ataollahi, M. R., Sharifi, J., Paknahad, M. R., & Paknahad, A. (2015). Breast cancer and associated factors: a review. *J Med Life*, 8(Spec Iss 4), 6-11.
- Badisa, R. B., Darling-Reed, S. F., Joseph, P., Cooperwood, J. S., Latinwo, L. M., & Goodman, C. B. (2009). Selective cytotoxic activities of two novel synthetic drugs on human breast carcinoma MCF-7 cells. *Anticancer Res*, 29(8), 2993-2996.
- Bagnolini, G., Milano, D., Manerba, M., Schipani, F., Ortega, J. A., Gioia, D., Falchi, F., Balboni, A., Farabegoli, F., De Franco, F., Robertson, J., Pellicciari, R., Pallavicini, I., Peri, S., Minucci, S., Girotto, S., Di Stefano, G., Roberti, M., & Cavalli, A. (2020). Synthetic Lethality in Pancreatic Cancer: Discovery of a New RAD51-BRCA2 Small Molecule Disruptor That Inhibits Homologous Recombination and Synergizes with Olaparib. *Journal of Medicinal Chemistry*, 63(5), 2588-2619.
- Balmaña, J., Tung, N. M., Isakoff, S. J., Graña, B., Ryan, P. D., Saura, C., Lowe, E. S., Frewer, P., Winer, E., Baselga, J., & Garber, J. E. (2014). Phase I trial of olaparib in combination with cisplatin for the treatment of patients with advanced breast, ovarian and other solid tumors. *Annals of Oncology*, 25(8), 1656-1663.
- Banerjee, S., Stewart, J., Porta, N., Toms, C., Leary, A., Lheureux, S., Khaliq, S., Tai, J., Attygalle, A., Vroobel, K., Lord, C. J., Natrajan, R., & Bliss, J. (2021). ATARI trial: ATR inhibitor in combination with olaparib in gynecological cancers with ARID1A loss or no loss (ENGOT/GYN1/NCRI). *Int J Gynecol Cancer*, 31(11), 1471-1475.
- Barchiesi, G., Roberto, M., Verrico, M., Vici, P., Tomao, S., & Tomao, F. (2021). Emerging Role of PARP Inhibitors in Metastatic Triple Negative

Breast Cancer. Current Scenario and Future Perspectives. *Front Oncol*, 11, 769280.

Bardia, A., Hurvitz, S. A., DeMichele, A., Clark, A. S., Zelnak, A., Yardley, D. A., Karuturi, M., Sanft, T., Blau, S., Hart, L., Ma, C., Rugo, H. S., Purkayastha, D., & Moulder, S. (2021). Phase I/II Trial of Exemestane, Ribociclib, and Everolimus in Women with HR(+)/HER2(-) Advanced Breast Cancer after Progression on CDK4/6 Inhibitors (TRINITI-1). *Clin Cancer Res*, 27(15), 4177-4185.

Baselga, J., Campone, M., Piccart, M., Burris, H. A., 3rd, Rugo, H. S., Sahmoud, T., Noguchi, S., Gnant, M., Pritchard, K. I., Lebrun, F., Beck, J. T., Ito, Y., Yardley, D., Deleu, I., Perez, A., Bachelot, T., Vittori, L., Xu, Z., Mukhopadhyay, P., Lebwohl, D., & Hortobagyi, G. N. (2012). Everolimus in postmenopausal hormone-receptor-positive advanced breast cancer. *N Engl J Med*, 366(6), 520-529.

Basourakos, S. P., Li, L., Aparicio, A. M., Corn, P. G., Kim, J., & Thompson, T. C. (2017). Combination Platinum-based and DNA Damage Response-targeting Cancer Therapy: Evolution and Future Directions. *Curr Med Chem*, 24(15), 1586-1606.

Bendell, J., O'Reilly, E. M., Middleton, M. R., Chau, I., Hochster, H., Fielding, A., Burke, W., & Burris, H., 3rd. (2015). Phase I study of olaparib plus gemcitabine in patients with advanced solid tumours and comparison with gemcitabine alone in patients with locally advanced/metastatic pancreatic cancer. *Ann Oncol*, 26(4), 804-811.

Benton, G., DeGray, G., Kleinman, H. K., George, J., & Arnaoutova, I. (2015). In vitro microtumors provide a physiologically predictive tool for breast cancer therapeutic screening. *PLoS One*, 10(4), e0123312.

Berenbaum, M. C. (1977). Synergy, additivism and antagonism in immunosuppression. A critical review. *Clin Exp Immunol*, 28(1), 1-18.

Berenbaum, M. C. (1978). A Method for Testing for Synergy with Any Number of Agents. *The Journal of Infectious Diseases*, 137(2), 122-130.

Bergamo, A., Riedel, T., Dyson, P. J., & Sava, G. (2015). Preclinical combination therapy of the investigational drug NAMI-A(+) with doxorubicin for mammary cancer. *Invest New Drugs*, 33(1), 53-63.

Bernardi, R., & Gianni, L. (2014). Hallmarks of triple negative breast cancer emerging at last? *Cell Research*, 24(8), 904-905.

Berndsen, R. H., Weiss, A., Abdul, U. K., Wong, T. J., Meraldi, P., Griffioen, A. W., Dyson, P. J., & Nowak-Sliwinska, P. (2017). Combination of ruthenium(II)-arene complex [Ru(η 6-p-cymene)Cl₂(pta)] (RAPTA-C) and the epidermal growth factor receptor inhibitor erlotinib results in

efficient angiostatic and antitumor activity. *Scientific Reports*, 7(1), 43005.

- Bertin, P., Rector-Brooks, J., Sharma, D., Gaudelet, T., Anighoro, A., Gross, T., Martínez-Peña, F., Tang, E. L., Suraj, M. S., Regep, C., Hayter, J. B. R., Korablyov, M., Valiante, N., van der Sloot, A., Tyers, M., Roberts, C. E. S., Bronstein, M. M., Lairson, L. L., Taylor-King, J. P., & Bengio, Y. (2023). RECOVER identifies synergistic drug combinations in vitro through sequential model optimization. *Cell Rep Methods*, 3(10), 100599.
- Bhamidipati, D., Haro-Silerio, J. I., Yap, T. A., & Ngoi, N. (2023). PARP inhibitors: enhancing efficacy through rational combinations. *British Journal of Cancer*.
- Bianchi, J. J., Zhao, X., Mays, J. C., & Davoli, T. (2020). Not all cancers are created equal: Tissue specificity in cancer genes and pathways. *Current Opinion in Cell Biology*, 63, 135-143.
- Bianchini, G., Balko, J. M., Mayer, I. A., Sanders, M. E., & Gianni, L. (2016). Triple-negative breast cancer: challenges and opportunities of a heterogeneous disease. *Nat Rev Clin Oncol*, 13(11), 674-690.
- Bianchini, G., De Angelis, C., Licata, L., & Gianni, L. (2022). Treatment landscape of triple-negative breast cancer — expanded options, evolving needs. *Nature Reviews Clinical Oncology*, 19(2), 91-113.
- Biegała, Ł., Gajek, A., Marczak, A., & Rogalska, A. (2023). Olaparib-Resistant BRCA2(MUT) Ovarian Cancer Cells with Restored BRCA2 Abrogate Olaparib-Induced DNA Damage and G2/M Arrest Controlled by the ATR/CHK1 Pathway for Survival. *Cells*, 12(7).
- Blackadar, C. B. (2016). Historical review of the causes of cancer. *World J Clin Oncol*, 7(1), 54-86.
- Blackford, A. N., & Jackson, S. P. (2017). ATM, ATR, and DNA-PK: The Trinity at the Heart of the DNA Damage Response. *Molecular Cell*, 66(6), 801-817.
- Borisy, A. A., Elliott, P. J., Hurst, N. W., Lee, M. S., Lehar, J., Price, E. R., Serbedzija, G., Zimmermann, G. R., Foley, M. A., Stockwell, B. R., & Keith, C. T. (2003). Systematic discovery of multicomponent therapeutics. *Proc Natl Acad Sci U S A*, 100(13), 7977-7982.
- Bowman, K. J., Newell, D. R., Calvert, A. H., & Curtin, N. J. (2001). Differential effects of the poly (ADP-ribose) polymerase (PARP) inhibitor NU1025 on topoisomerase I and II inhibitor cytotoxicity in L1210 cells in vitro. *Br J Cancer*, 84(1), 106-112.

- Bowman, K. J., White, A., Golding, B. T., Griffin, R. J., & Curtin, N. J. (1998). Potentiation of anti-cancer agent cytotoxicity by the potent poly(ADP-ribose) polymerase inhibitors NU1025 and NU1064. *Br J Cancer*, 78(10), 1269-1277.
- Brown, J. S., O'Carrigan, B., Jackson, S. P., & Yap, T. A. (2017). Targeting DNA Repair in Cancer: Beyond PARP Inhibitors. *Cancer Discov*, 7(1), 20-37.
- Bryant, H. E., Petermann, E., Schultz, N., Jemth, A. S., Loseva, O., Issaeva, N., Johansson, F., Fernandez, S., McGlynn, P., & Helleday, T. (2009). PARP is activated at stalled forks to mediate Mre11-dependent replication restart and recombination. *Embo j*, 28(17), 2601-2615.
- Bryant, H. E., Schultz, N., Thomas, H. D., Parker, K. M., Flower, D., Lopez, E., Kyle, S., Meuth, M., Curtin, N. J., & Helleday, T. (2005). Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase. *Nature*, 434(7035), 913-917.
- Buzdar, A. U., Robertson, J. F., Eiermann, W., & Nabholz, J. M. (2002). An overview of the pharmacology and pharmacokinetics of the newer generation aromatase inhibitors anastrozole, letrozole, and exemestane. *Cancer*, 95(9), 2006-2016.
- Cahuzac, M., Péant, B., Mes-Masson, A.-M., & Saad, F. (2022). Development of Olaparib-Resistance Prostate Cancer Cell Lines to Identify Mechanisms Associated with Acquired Resistance. *Cancers*, 14(16), 3877.
- Cai, L., Xu, X., & Chen, W. (2022). The Current State of the Art in PARP Inhibitor-Based Delivery Nanosystems. *Pharmaceutics*, 14(8).
- Calvert, A. H., Harland, S. J., Newell, D. R., Siddik, Z. H., Jones, A. C., McElwain, T. J., Raju, S., Wiltshaw, E., Smith, I. E., Baker, J. M., Peckham, M. J., & Harrap, K. R. (1982). Early clinical studies with cis-diammine-1,1-cyclobutane dicarboxylate platinum II. *Cancer Chemother Pharmacol*, 9(3), 140-147.
- Carey, J. P. W., Karakas, C., Bui, T., Chen, X., Vijayaraghavan, S., Zhao, Y., Wang, J., Mikule, K., Litton, J. K., Hunt, K. K., & Keyomarsi, K. (2018). Synthetic Lethality of PARP Inhibitors in Combination with MYC Blockade Is Independent of BRCA Status in Triple-Negative Breast Cancer. *Cancer Res*, 78(3), 742-757.
- Carlsson, J., & Yuhas, J. M. (1984). Liquid-Overlay Culture of Cellular Spheroids. In H. Acker, J. Carlsson, R. Durand, & R. M. Sutherland (Eds.), *Spheroids in Cancer Research: Methods and Perspectives* (pp. 1-23). Berlin, Heidelberg: Springer Berlin Heidelberg.

- Cassar, S., Adatto, I., Freeman, J. L., Gamse, J. T., Iturria, I., Lawrence, C., Muriana, A., Peterson, R. T., Van Cruchten, S., & Zon, L. I. (2020). Use of Zebrafish in Drug Discovery Toxicology. *Chemical Research in Toxicology*, 33(1), 95-118.
- Chakraborty, C., Sharma, A. R., Sharma, G., & Lee, S.-S. (2016). Zebrafish: A complete animal model to enumerate the nanoparticle toxicity. *Journal of Nanobiotechnology*, 14(1), 65.
- Chaudhuri, A. R., Callen, E., Ding, X., Gogola, E., Duarte, A. A., Lee, J. E., Wong, N., Lafarga, V., Calvo, J. A., Panzarino, N. J., John, S., Day, A., Crespo, A. V., Shen, B., Starnes, L. M., de Ruiter, J. R., Daniel, J. A., Konstantinopoulos, P. A., Cortez, D., Cantor, S. B., Fernandez-Capetillo, O., Ge, K., Jonkers, J., Rottenberg, S., Sharan, S. K., & Nussenzweig, A. (2016). Replication fork stability confers chemoresistance in BRCA-deficient cells. *Nature*, 535(7612), 382-387.
- Chaudhuri, A. R., & Nussenzweig, A. (2017). The multifaceted roles of PARP1 in DNA repair and chromatin remodelling. *Nat Rev Mol Cell Biol*, 18(10), 610-621.
- Chen, D., Guo, S., Tang, X., Rong, Y., Bo, H., Shen, H., Zhao, Z., Qiao, A., Shen, J., & Wang, J. (2022a). Combination of ruthenium (II) polypyridyl complex Δ -Ru1 and Taxol enhances the anti-cancer effect on Taxol-resistant cancer cells through Caspase-1/GSDMD-mediated pyroptosis. *Journal of Inorganic Biochemistry*, 230, 111749.
- Chen, H., Wu, J., Zhang, Z., Tang, Y., Li, X., Liu, S., Cao, S., & Li, X. (2018). Association Between BRCA Status and Triple-Negative Breast Cancer: A Meta-Analysis. *Front Pharmacol*, 9, 909.
- Chen, H. D., Guo, N., Song, S. S., Chen, C. H., Miao, Z. H., & He, J. X. (2020). Novel mutations in BRCA2 intron 11 and overexpression of COX-2 and BIRC3 mediate cellular resistance to PARP inhibitors. *Am J Cancer Res*, 10(9), 2813-2831.
- Chen, I.-C., Lin, C.-H., Chang, D.-Y., Chen, T. W.-W., Wang, M.-Y., Ma, W.-L., Lin, Y.-T., Huang, S.-M., & Lu, Y.-S. (2021a). Abstract CT028: A pilot study of pembrolizumab and exemestane/leuprolide in premenopausal hormone receptor positive/HER2 negative locally advanced or metastatic breast cancer (PEER). *Cancer Research*, 81(13_Supplement), CT028-CT028.
- Chen, I. C., Lin, C.-H., Chang, D.-Y., Chen, T. W.-W., Wang, M.-Y., Ma, W.-L., Lin, Y.-T., Huang, S.-M., & Lu, Y.-S. (2021b). Abstract CT028: A pilot study of pembrolizumab and exemestane/leuprolide in premenopausal hormone receptor positive/HER2 negative locally advanced or metastatic breast cancer (PEER). *Cancer Research*, 81(13_Supplement), CT028-CT028.

- Chen, J. Q., & Russo, J. (2009). ERalpha-negative and triple negative breast cancer: molecular features and potential therapeutic approaches. *Biochim Biophys Acta*, 1796(2), 162-175.
- Chen, T., Tongpeng, S., Lu, Z., Topatana, W., Juengpanich, S., Li, S., Hu, J., Cao, J., Lee, C., Tian, Y., Chen, M., & Cai, X. (2022b). DNA damage response inhibition-based combination therapies in cancer treatment: Recent advances and future directions. *Aging and Cancer*, 3(1), 44-67.
- Cheng, H., Zhang, Z., Borczuk, A., Powell, C. A., Balajee, A. S., Lieberman, H. B., & Halmos, B. (2013). PARP inhibition selectively increases sensitivity to cisplatin in ERCC1-low non-small cell lung cancer cells. *Carcinogenesis*, 34(4), 739-749.
- Chou, T.-C., & Talalay, P. (1983). Analysis of combined drug effects: a new look at a very old problem. *Trends in Pharmacological Sciences*, 4, 450-454.
- Chou, T.-C., & Talalay, P. (1984). Quantitative analysis of dose-effect relationships: the combined effects of multiple drugs or enzyme inhibitors. *Advances in Enzyme Regulation*, 22, 27-55.
- Chou, T., & Martin, N. (2005). CompuSyn for drug combinations: PC software and user's guide: a computer program for quantitation of synergism and antagonism in drug combinations, and the determination of IC50 and ED50 and LD50 values. . Retrieved from <https://www.combosyn.com>.
- Chou, T. C. (2006). Theoretical basis, experimental design, and computerized simulation of synergism and antagonism in drug combination studies. *Pharmacol Rev*, 58(3), 621-681.
- Chou, T. C. (2010). Drug combination studies and their synergy quantification using the Chou-Talalay method. *Cancer Res*, 70(2), 440-446.
- Chun, H. G., Leyland-Jones, B. R., Caryk, S. M., & Hoth, D. F. (1986). Central nervous system toxicity of fludarabine phosphate. *Cancer Treat Rep*, 70(10), 1225-1228.
- Cimprich, K. A., & Cortez, D. (2008). ATR: an essential regulator of genome integrity. *Nat Rev Mol Cell Biol*, 9(8), 616-627.
- Clarke, N. W., Armstrong, A. J., Thiery-Vuillemin, A., Oya, M., Shore, N., Lored, E., Procopio, G., Menezes, J. d., Giroto, G., Arslan, C., Mehra, N., Parnis, F., Brown, E., Schlürmann, F., Joung, J. Y., Sugimoto, M., Virizuela, J. A., Emmenegger, U., Navratil, J., Buchschacher, G. L., Poehlein, C., Harrington, E. A., Desai, C., Kang, J., & Saad, F. (2022). Abiraterone and Olaparib for Metastatic Castration-Resistant Prostate Cancer. *NEJM Evidence*, 1(9), EVIDoA2200043.

- Cokol, M., Chua, H. N., Tasan, M., Mutlu, B., Weinstein, Z. B., Suzuki, Y., Nergiz, M. E., Costanzo, M., Baryshnikova, A., Giaever, G., Nislow, C., Myers, C. L., Andrews, B. J., Boone, C., & Roth, F. P. (2011). Systematic exploration of synergistic drug pairs. *Mol Syst Biol*, 7, 544.
- Cole, S. P. (1986). Rapid chemosensitivity testing of human lung tumor cells using the MTT assay. *Cancer Chemother Pharmacol*, 17(3), 259-263.
- Coleman, R. L., Oza, A. M., Lorusso, D., Aghajanian, C., Oaknin, A., Dean, A., Colombo, N., Weberpals, J. I., Clamp, A., Scambia, G., Leary, A., Holloway, R. W., Gancedo, M. A., Fong, P. C., Goh, J. C., O'Malley, D. M., Armstrong, D. K., Garcia-Donas, J., Swisher, E. M., Floquet, A., Konecny, G. E., McNeish, I. A., Scott, C. L., Cameron, T., Maloney, L., Isaacson, J., Goble, S., Grace, C., Harding, T. C., Raponi, M., Sun, J., Lin, K. K., Giordano, H., & Ledermann, J. A. (2017). Rucaparib maintenance treatment for recurrent ovarian carcinoma after response to platinum therapy (ARIEL3): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet*, 390(10106), 1949-1961.
- Costa, E. C., Moreira, A. F., de Melo-Diogo, D., Gaspar, V. M., Carvalho, M. P., & Correia, I. J. (2016). 3D tumor spheroids: an overview on the tools and techniques used for their analysis. *Biotechnol Adv*, 34(8), 1427-1441.
- Costăchel, O., Fadei, L., & Badea, E. (1969). Tumor cell suspension culture on non adhesive substratum. *Z Krebsforsch*, 72(1), 24-31.
- Curtin, N. J., & Szabo, C. (2020). Poly(ADP-ribose) polymerase inhibition: past, present and future. *Nature Reviews Drug Discovery*, 19(10), 711-736.
- D'Amours, D., Desnoyers, S., D'Silva, I., & Poirier, G. G. (1999). Poly(ADP-ribosyl)ation reactions in the regulation of nuclear functions. *Biochem J*, 342 (Pt 2)(Pt 2), 249-268.
- Dasari, S., & Bernard Tchounwou, P. (2014). Cisplatin in cancer therapy: Molecular mechanisms of action. *European Journal of Pharmacology*, 740, 364-378.
- Däster, S., Amatruda, N., Calabrese, D., Ivanek, R., Turrini, E., Drieser, R. A., Zajac, P., Fimognari, C., Spagnoli, G. C., Iezzi, G., Mele, V., & Muraro, M. G. (2017). Induction of hypoxia and necrosis in multicellular tumor spheroids is associated with resistance to chemotherapy treatment. *Oncotarget*, 8(1), 1725-1736.
- de Bono, J., Mateo, J., Fizazi, K., Saad, F., Shore, N., Sandhu, S., Chi, K. N., Sartor, O., Agarwal, N., Olmos, D., Thiery-Vuillemin, A., Twardowski, P., Mehra, N., Goessl, C., Kang, J., Burgents, J., Wu, W., Kohlmann, A.,

- Adelman, C. A., & Hussain, M. (2020). Olaparib for Metastatic Castration-Resistant Prostate Cancer. *New England Journal of Medicine*, 382(22), 2091-2102.
- de Murcia, J. M., Niedergang, C., Trucco, C., Ricoul, M., Dutrillaux, B., Mark, M., Oliver, F. J., Masson, M., Dierich, A., LeMeur, M., Walztinger, C., Chambon, P., & de Murcia, G. (1997). Requirement of poly(ADP-ribose) polymerase in recovery from DNA damage in mice and in cells. *Proceedings of the National Academy of Sciences*, 94(14), 7303-7307.
- Delgado, J. L., Hsieh, C. M., Chan, N. L., & Hiasa, H. (2018). Topoisomerases as anticancer targets. *Biochem J*, 475(2), 373-398.
- Demény, M. A., & Virág, L. (2021). The PARP Enzyme Family and the Hallmarks of Cancer Part 1. Cell Intrinsic Hallmarks. *Cancers (Basel)*, 13(9).
- Dent, R. A., Lindeman, G. J., Clemons, M., Wildiers, H., Chan, A., McCarthy, N. J., Singer, C. F., Lowe, E. S., Watkins, C. L., & Carmichael, J. (2013). Phase I trial of the oral PARP inhibitor olaparib in combination with paclitaxel for first- or second-line treatment of patients with metastatic triple-negative breast cancer. *Breast Cancer Res*, 15(5), R88.
- Derakhshan, F., & Reis-Filho, J. S. (2022). Pathogenesis of Triple-Negative Breast Cancer. *Annu Rev Pathol*, 17, 181-204.
- Derby, S. J., Chalmers, A. J., & Carruthers, R. D. (2022). Radiotherapy-Poly(ADP-ribose) Polymerase Inhibitor Combinations: Progress to Date. *Seminars in Radiation Oncology*, 32(1), 15-28.
- Do, K. T., Kochupurakkal, B., Kelland, S., de Jonge, A., Hedglin, J., Powers, A., Quinn, N., Gannon, C., Vuong, L., Parmar, K., Lazaro, J. B., D'Andrea, A. D., & Shapiro, G. I. (2021). Phase 1 Combination Study of the CHK1 Inhibitor Prexasertib and the PARP Inhibitor Olaparib in High-grade Serous Ovarian Cancer and Other Solid Tumors. *Clin Cancer Res*, 27(17), 4710-4716.
- Dobzhansky, T. (1946). Genetics of natural populations; recombination and variability in populations of *Drosophila pseudoobscura*. *Genetics*, 31(3), 269-290.
- Donson, A. M., Amani, V., Warner, E. A., Griesinger, A. M., Witt, D. A., Levy, J. M. M., Hoffman, L. M., Hankinson, T. C., Handler, M. H., Vibhakkar, R., Dorris, K., & Foreman, N. K. (2018). Identification of FDA-Approved Oncology Drugs with Selective Potency in High-Risk Childhood Ependymoma. *Molecular Cancer Therapeutics*, 17(9), 1984-1994.
- Drew, Y., Ledermann, J., Hall, G., Rea, D., Glasspool, R., Highley, M., Jayson, G., Sludden, J., Murray, J., Jamieson, D., Halford, S., Acton, G.,

- Backholer, Z., Mangano, R., Boddy, A., Curtin, N., & Plummer, R. (2016). Phase 2 multicentre trial investigating intermittent and continuous dosing schedules of the poly(ADP-ribose) polymerase inhibitor rucaparib in germline BRCA mutation carriers with advanced ovarian and breast cancer. *Br J Cancer*, 114(7), 723-730.
- Drewett, L., Lucey, R., Pinilla, K. A., Grybowicz, L., Wulff, J., Dayimu, A., Demir, N., Vallier, A.-L., Qian, W., Machin, A., McAdam, K., Roylance, R., Copson, E., Armstrong, A. C., Levitt, N., Provenzano, E., Tischkowitz, M. D., McMurtry, E., Earl, H. M., Abraham, J., Team, P. R., & Team, P. R. (2022). PARTNER: A randomized, phase II/III trial to evaluate the safety and efficacy of the addition of olaparib to platinum-based neoadjuvant chemotherapy in patients with triple-negative and/or germline BRCA-mutated breast cancer. *Journal of Clinical Oncology*, 40(16_suppl), TPS619-TPS619.
- Duijf, P. H. G., Nanayakkara, D., Nones, K., Srihari, S., Kalimutho, M., & Khanna, K. K. (2019). Mechanisms of Genomic Instability in Breast Cancer. *Trends Mol Med*, 25(7), 595-611.
- Duma, N., Santana-Davila, R., & Molina, J. R. (2019). Non-Small Cell Lung Cancer: Epidemiology, Screening, Diagnosis, and Treatment. *Mayo Clinic Proceedings*, 94(8), 1623-1640.
- Edwards, S. L., Brough, R., Lord, C. J., Natrajan, R., Vatcheva, R., Levine, D. A., Boyd, J., Reis-Filho, J. S., & Ashworth, A. (2008). Resistance to therapy caused by intragenic deletion in BRCA2. *Nature*, 451(7182), 1111-1115.
- Etezadi, S., Evans, J. C., Shen, Y. T., De Souza, R., Piquette-Miller, M., & Allen, C. (2018). Ratio-Dependent Synergism of a Doxorubicin and Olaparib Combination in 2D and Spheroid Models of Ovarian Cancer. *Mol Pharm*, 15(2), 472-485.
- El-Khamisy, S. F., Masutani, M., Suzuki, H., & Caldecott, K. W. (2003). A requirement for PARP-1 for the assembly or stability of XRCC1 nuclear foci at sites of oxidative DNA damage. *Nucleic Acids Res*, 31(19), 5526-5533.
- Eleutherakis-Papaïakovou, E., Kanellias, N., Kastritis, E., Gavriatopoulou, M., Terpos, E., & Dimopoulos, M. A. (2020). Efficacy of Panobinostat for the Treatment of Multiple Myeloma. *J Oncol*, 2020, 7131802.
- Elgar, C. E., Yusoh, N. A., Tiley, P. R., Kolozsvári, N., Bennett, L. G., Gamble, A., Péan, E. V., Davies, M. L., Staples, C. J., Ahmad, H., & Gill, M. R. (2023). Ruthenium(II) Polypyridyl Complexes as FRET Donors: Structure- and Sequence-Selective DNA-Binding and Anticancer Properties. *Journal of the American Chemical Society*, 145(2), 1236-1246.

- Emran, T. B., Shahriar, A., Mahmud, A. R., Rahman, T., Abir, M. H., Siddiquee, M. F.-R., Ahmed, H., Rahman, N., Nainu, F., Wahyudin, E., Mitra, S., Dhama, K., Habiballah, M. M., Haque, S., Islam, A., & Hassan, M. M. (2022a). Multidrug Resistance in Cancer: Understanding Molecular Mechanisms, Immunoprevention and Therapeutic Approaches. *Frontiers in Oncology*, 12.
- Emran, T. B., Shahriar, A., Mahmud, A. R., Rahman, T., Abir, M. H., Siddiquee, M. F. R., Ahmed, H., Rahman, N., Nainu, F., Wahyudin, E., Mitra, S., Dhama, K., Habiballah, M. M., Haque, S., Islam, A., & Hassan, M. M. (2022b). Multidrug Resistance in Cancer: Understanding Molecular Mechanisms, Immunoprevention and Therapeutic Approaches. *Frontiers in Oncology*, 12.
- Engbrecht, M., & Mangerich, A. (2020). The Nucleolus and PARP1 in Cancer Biology. *Cancers (Basel)*, 12(7).
- Evers, B., Schut, E., van der Burg, E., Braumuller, T. M., Egan, D. A., Holstege, H., Edser, P., Adams, D. J., Wade-Martins, R., Bouwman, P., & Jonkers, J. (2010). A high-throughput pharmaceutical screen identifies compounds with specific toxicity against BRCA2-deficient tumors. *Clin Cancer Res*, 16(1), 99-108.
- Falchi, F., Giacomini, E., Masini, T., Boutard, N., Di Ianni, L., Manerba, M., Farabegoli, F., Rossini, L., Robertson, J., Minucci, S., Pallavicini, I., Di Stefano, G., Roberti, M., Pellicciari, R., & Cavalli, A. (2017). Synthetic Lethality Triggered by Combining Olaparib with BRCA2–Rad51 Disruptors. *ACS Chemical Biology*, 12(10), 2491-2497.
- Farago, A. F., Yeap, B. Y., Stanzione, M., Hung, Y. P., Heist, R. S., Marcoux, J. P., Zhong, J., Rangachari, D., Barbie, D. A., Phat, S., Myers, D. T., Morris, R., Kem, M., Dubash, T. D., Kennedy, E. A., Digumarthy, S. R., Sequist, L. V., Hata, A. N., Maheswaran, S., Haber, D. A., Lawrence, M. S., Shaw, A. T., Mino-Kenudson, M., Dyson, N. J., & Drapkin, B. J. (2019). Combination Olaparib and Temozolomide in Relapsed Small-Cell Lung Cancer. *Cancer Discov*, 9(10), 1372-1387.
- Farmer, H., McCabe, N., Lord, C. J., Tutt, A. N. J., Johnson, D. A., Richardson, T. B., Santarosa, M., Dillon, K. J., Hickson, I., Knights, C., Martin, N. M. B., Jackson, S. P., Smith, G. C. M., & Ashworth, A. (2005). Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature*, 434(7035), 917-921.
- Fasching, P. A., Link, T., Hauke, J., Seither, F., Jackisch, C., Klare, P., Schmatloch, S., Hanusch, C., Huober, J., Stefek, A., Seiler, S., Schmitt, W. D., Uleer, C., Doering, G., Rhiem, K., Schneeweiss, A., Engels, K., Denkert, C., Schmutzler, R. K., Hahnen, E., Untch, M., Burchardi, N., Blohmer, J. U., & Loibl, S. (2021). Neoadjuvant paclitaxel/olaparib in

comparison to paclitaxel/carboplatinum in patients with HER2-negative breast cancer and homologous recombination deficiency (GeparOLA study). *Ann Oncol*, 32(1), 49-57.

Ferlay, J., Colombet, M., Soerjomataram, I., Mathers, C., Parkin, D. M., Piñeros, M., Znaor, A., & Bray, F. (2019). Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. *International Journal of Cancer*, 144(8), 1941-1953.

Ferlay, J., Colombet, M., Soerjomataram, I., Parkin, D. M., Piñeros, M., Znaor, A., & Bray, F. (2021). Cancer statistics for the year 2020: An overview. *International Journal of Cancer*, 149(4), 778-789.

Ferlay, J., Ervik, M., Lam, F., Colombet, M., Mery, L., Piñeros, M., Znaor, A., Soerjomataram, I., & Bray, F. (2020). Global Cancer Observatory: Cancer Today. Retrieved from <https://gco.iarc.fr/today>

Finkel, T. (2011). Signal transduction by reactive oxygen species. *J Cell Biol*, 194(1), 7-15.

Flobak, Å., Niederdorfer, B., Nakstad, V. T., Thommesen, L., Klinkenberg, G., & Lægreid, A. (2019). A high-throughput drug combination screen of targeted small molecule inhibitors in cancer cell lines. *Scientific Data*, 6(1), 237.

Flocke, L. S., Trondl, R., Jakupec, M. A., & Keppler, B. K. (2016). Molecular mode of action of NKP-1339 - a clinically investigated ruthenium-based drug - involves ER- and ROS-related effects in colon carcinoma cell lines. *Invest New Drugs*, 34(3), 261-268.

Florea, A. M., & Büsselberg, D. (2011). Cisplatin as an anti-tumor drug: cellular mechanisms of activity, drug resistance and induced side effects. *Cancers (Basel)*, 3(1), 1351-1371.

Fojo, A. T., Ueda, K., Slamon, D. J., Poplack, D. G., Gottesman, M. M., & Pastan, I. (1987). Expression of a multidrug-resistance gene in human tumors and tissues. *Proc Natl Acad Sci U S A*, 84(1), 265-269.

Fong, P. C., Boss, D. S., Yap, T. A., Tutt, A., Wu, P., Mergui-Roelvink, M., Mortimer, P., Swaisland, H., Lau, A., O'Connor, M. J., Ashworth, A., Carmichael, J., Kaye, S. B., Schellens, J. H., & de Bono, J. S. (2009). Inhibition of poly(ADP-ribose) polymerase in tumors from BRCA mutation carriers. *N Engl J Med*, 361(2), 123-134.

Foty, R. (2011). A simple hanging drop cell culture protocol for generation of 3D spheroids. *J Vis Exp*(51).

- Foucquier, J., & Guedj, M. (2015). Analysis of drug combinations: current methodological landscape. *Pharmacology Research & Perspectives*, 3(3), e00149.
- Franken, N. A. P., Rodermond, H. M., Stap, J., Haveman, J., & van Bree, C. (2006). Clonogenic assay of cells in vitro. *Nature Protocols*, 1(5), 2315-2319.
- Fu, D., Calvo, J. A., & Samson, L. D. (2012). Balancing repair and tolerance of DNA damage caused by alkylating agents. *Nature Reviews Cancer*, 12(2), 104-120.
- Gajan, A., Sarma, A., Kim, S., Gurdziel, K., Wu, G. S., & Shekhar, M. P. (2021). Analysis of Adaptive Olaparib Resistance Effects on Cisplatin Sensitivity in Triple Negative Breast Cancer Cells. *Frontiers in Oncology*, 11.
- Galluzzi, L., Senovilla, L., Vitale, I., Michels, J., Martins, I., Kepp, O., Castedo, M., & Kroemer, G. (2012). Molecular mechanisms of cisplatin resistance. *Oncogene*, 31(15), 1869-1883.
- García-Becerra, R., Santos, N., Díaz, L., & Camacho, J. (2012). Mechanisms of resistance to endocrine therapy in breast cancer: focus on signaling pathways, miRNAs and genetically based resistance. *Int J Mol Sci*, 14(1), 108-145.
- Garcia Jimenez, D., Poongavanam, V., & Kihlberg, J. (2023). Macrocycles in Drug Discovery—Learning from the Past for the Future. *Journal of Medicinal Chemistry*, 66(8), 5377-5396.
- Gasser, G., & Metzler-Nolte, N. (2012). The potential of organometallic complexes in medicinal chemistry. *Curr Opin Chem Biol*, 16(1-2), 84-91.
- Ge, X., Yost, S. E., Lee, J. S., Frankel, P. H., Ruel, C., Cui, Y., Murga, M., Tang, A., Martinez, N., Chung, S., Yeon, C., Stewart, D., Li, D., Rajurkar, S., Somlo, G., Mortimer, J., Waisman, J., & Yuan, Y. (2022). Phase II Study Combining Pembrolizumab with Aromatase Inhibitor in Patients with Metastatic Hormone Receptor Positive Breast Cancer. *Cancers (Basel)*, 14(17).
- Geldmacher, Y., Oleszak, M., & Sheldrick, W. S. (2012). Rhodium(III) and iridium(III) complexes as anticancer agents. *Inorganica Chimica Acta*, 393, 84-102.
- Gelmon, K. A., Tischkowitz, M., Mackay, H., Swenerton, K., Robidoux, A., Tonkin, K., Hirte, H., Huntsman, D., Clemons, M., Gilks, B., Yerushalmi, R., Macpherson, E., Carmichael, J., & Oza, A. (2011). Olaparib in patients with recurrent high-grade serous or poorly differentiated ovarian

- carcinoma or triple-negative breast cancer: a phase 2, multicentre, open-label, non-randomised study. *Lancet Oncol*, 12(9), 852-861.
- Ghosh, D., Ahmad, H., & A. Thomas, J. (2009). Kinetically locked luminescent metallomacrocycles as duplex DNA binding substrates. *Chemical Communications*(20), 2947-2949.
- Gill, M. R., Garcia-Lara, J., Foster, S. J., Smythe, C., Battaglia, G., & Thomas, J. A. (2009). A ruthenium(II) polypyridyl complex for direct imaging of DNA structure in living cells. *Nature Chemistry*, 1(8), 662-667.
- Gill, M. R., Harun, S. N., Halder, S., Boghoozian, R. A., Ramadan, K., Ahmad, H., & Vallis, K. A. (2016). A ruthenium polypyridyl intercalator stalls DNA replication forks, radiosensitizes human cancer cells and is enhanced by Chk1 inhibition. *Scientific Reports*, 6(1), 31973.
- Gill, M. R., & Thomas, J. A. (2012). Ruthenium(ii) polypyridyl complexes and DNA—from structural probes to cellular imaging and therapeutics. *Chemical Society Reviews*, 41(8), 3179-3192.
- Gill, M. R., & Vallis, K. A. (2019). Transition metal compounds as cancer radiosensitizers. *Chemical Society Reviews*, 48(2), 540-557.
- Giudice, E., Gentile, M., Salutari, V., Ricci, C., Musacchio, L., Carbone, M. V., Ghizzoni, V., Camarda, F., Tronconi, F., Nero, C., Ciccarone, F., Scambia, G., & Lorusso, D. (2022). PARP Inhibitors Resistance: Mechanisms and Perspectives. *Cancers*, 14(6), 1420.
- Gogola, E., Duarte, A. A., de Ruiter, J. R., Wiegant, W. W., Schmid, J. A., de Bruijn, R., James, D. I., Guerrero Llobet, S., Vis, D. J., Annunziato, S., van den Broek, B., Barazas, M., Kersbergen, A., van de Ven, M., Tarsounas, M., Ogilvie, D. J., van Vugt, M., Wessels, L. F. A., Bartkova, J., Gromova, I., Andújar-Sánchez, M., Bartek, J., Lopes, M., van Attikum, H., Borst, P., Jonkers, J., & Rottenberg, S. (2018). Selective Loss of PARG Restores PARylation and Counteracts PARP Inhibitor-Mediated Synthetic Lethality. *Cancer Cell*, 33(6), 1078-1093.e1012.
- Golan, T., Hammel, P., Reni, M., Van Cutsem, E., Macarulla, T., Hall, M. J., Park, J. O., Hochhauser, D., Arnold, D., Oh, D. Y., Reinacher-Schick, A., Tortora, G., Algül, H., O'Reilly, E. M., McGuinness, D., Cui, K. Y., Schlienger, K., Locker, G. Y., & Kindler, H. L. (2019). Maintenance Olaparib for Germline BRCA-Mutated Metastatic Pancreatic Cancer. *N Engl J Med*, 381(4), 317-327.
- Golbaghi, G., Haghdoust, M. M., Yancu, D., López de los Santos, Y., Doucet, N., Patten, S. A., Sanderson, J. T., & Castonguay, A. (2019). Organoruthenium(II) Complexes Bearing an Aromatase Inhibitor: Synthesis, Characterization, in Vitro Biological Activity and in Vivo Toxicity in Zebrafish Embryos. *Organometallics*, 38(3), 702-711.

- Gomez, M. K., Illuzzi, G., Colomer, C., Churchman, M., Hollis, R. L., O'Connor, M. J., Gourley, C., Leo, E., & Melton, D. W. (2020). Identifying and Overcoming Mechanisms of PARP Inhibitor Resistance in Homologous Recombination Repair-Deficient and Repair-Proficient High Grade Serous Ovarian Cancer Cells. *Cancers (Basel)*, 12(6).
- Gonzalez-Angulo, A. M., Timms, K. M., Liu, S., Chen, H., Litton, J. K., Potter, J., Lanchbury, J. S., Stemke-Hale, K., Hennessy, B. T., Arun, B. K., Hortobagyi, G. N., Do, K. A., Mills, G. B., & Meric-Bernstam, F. (2011). Incidence and outcome of BRCA mutations in unselected patients with triple receptor-negative breast cancer. *Clin Cancer Res*, 17(5), 1082-1089.
- Griffin, R. J., Pemberton, L. C., Rhodes, D., Bleasdale, C., Bowman, K., Calvert, A. H., Curtin, N. J., Durkacz, B. W., Newell, D. R., Porteous, J. K., & et al. (1995). Novel potent inhibitors of the DNA repair enzyme poly(ADP-ribose)polymerase (PARP). *Anticancer Drug Des*, 10(6), 507-514.
- Griffin, R. J., Srinivasan, S., White, A. W., Bowman, K., Calvert, A. H., Curtin, N. J., Newell, D. R., & Golding, B. T. (1996). Novel Benzimidazole and Quinazolinone Inhibitors of the DNA Repair Enzyme Poly(ADP-ribose)polymerase*. *Pharmacy and Pharmacology Communications*, 2(1), 43-47.
- Groelly, F. J., Fawkes, M., Dagg, R. A., Blackford, A. N., & Tarsounas, M. (2023). Targeting DNA damage response pathways in cancer. *Nature Reviews Cancer*, 23(2), 78-94.
- Gyori, B. M., Venkatachalam, G., Thiagarajan, P. S., Hsu, D., & Clement, M.-V. (2014). OpenComet: An automated tool for comet assay image analysis. *Redox Biology*, 2, 457-465.
- Haji-Karim, M., & Carlsson, J. (1978). Proliferation and viability in cellular spheroids of human origin. *Cancer Res*, 38(5), 1457-1464.
- Hamburger, A. W., & Salmon, S. E. (1977). Primary bioassay of human tumor stem cells. *Science*, 197(4302), 461-463.
- Han, S. J., Kwon, S., & Kim, K. S. (2021). Challenges of applying multicellular tumor spheroids in preclinical phase. *Cancer Cell International*, 21(1), 152.
- Hanahan, D. (2022). Hallmarks of Cancer: New Dimensions. *Cancer Discov*, 12(1), 31-46.
- Hanif, M., & Hartinger, C. G. (2018). Anticancer metallodrugs: where is the next cisplatin? *Future Med Chem*, 10(6), 615-617.

- Hanna, C., Kurian, K. M., Williams, K., Watts, C., Jackson, A., Carruthers, R., Strathdee, K., Cruickshank, G., Dunn, L., Erridge, S., Godfrey, L., Jefferies, S., McBain, C., Sleigh, R., McCormick, A., Pittman, M., Halford, S., & Chalmers, A. J. (2020). Pharmacokinetics, safety, and tolerability of olaparib and temozolomide for recurrent glioblastoma: results of the phase I OPARATIC trial. *Neuro Oncol*, 22(12), 1840-1850.
- Hartinger, C. G., Jakupec, M. A., Zorbas-Seifried, S., Groessl, M., Egger, A., Berger, W., Zorbas, H., Dyson, P. J., & Keppler, B. K. (2008). KP1019, a new redox-active anticancer agent--preclinical development and results of a clinical phase I study in tumor patients. *Chem Biodivers*, 5(10), 2140-2155.
- Hartinger, C. G., Zorbas-Seifried, S., Jakupec, M. A., Kynast, B., Zorbas, H., & Keppler, B. K. (2006). From bench to bedside – preclinical and early clinical development of the anticancer agent indazolium trans-[tetrachlorobis(1H-indazole)ruthenate(III)] (KP1019 or FFC14A). *Journal of Inorganic Biochemistry*, 100(5), 891-904.
- He, L., Kulesskiy, E., Saarela, J., Turunen, L., Wennerberg, K., Aittokallio, T., & Tang, J. (2018). Methods for High-throughput Drug Combination Screening and Synergy Scoring. *Methods Mol Biol*, 1711, 351-398.
- Heffeter, P., Atil, B., Kryeziu, K., Groza, D., Koellensperger, G., Körner, W., Jungwirth, U., Mohr, T., Keppler, B. K., & Berger, W. (2013). The ruthenium compound KP1339 potentiates the anticancer activity of sorafenib in vitro and in vivo. *Eur J Cancer*, 49(15), 3366-3375.
- Hernandez, J. J., Pryszlak, M., Smith, L., Yanchus, C., Kurji, N., Shahani, V. M., & Molinski, S. V. (2017). Giving Drugs a Second Chance: Overcoming Regulatory and Financial Hurdles in Repurposing Approved Drugs As Cancer Therapeutics. *Front Oncol*, 7, 273.
- Hirschhaeuser, F., Menne, H., Dittfeld, C., West, J., Mueller-Klieser, W., & Kunz-Schughart, L. A. (2010). Multicellular tumor spheroids: an underestimated tool is catching up again. *J Biotechnol*, 148(1), 3-15.
- Hodges, L. M., Markova, S. M., Chinn, L. W., Gow, J. M., Kroetz, D. L., Klein, T. E., & Altman, R. B. (2011). Very important pharmacogene summary: ABCB1 (MDR1, P-glycoprotein). *Pharmacogenet Genomics*, 21(3), 152-161.
- Holbeck, S. L., Camalier, R., Crowell, J. A., Govindharajulu, J. P., Hollingshead, M., Anderson, L. W., Polley, E., Rubinstein, L., Srivastava, A., Wilsker, D., Collins, J. M., & Doroshow, J. H. (2017). The National Cancer Institute ALMANAC: A Comprehensive Screening Resource for the Detection of Anticancer Drug Pairs with Enhanced Therapeutic Activity. *Cancer Res*, 77(13), 3564-3576.

- Holbeck, S. L., Collins, J. M., & Doroshow, J. H. (2010). Analysis of Food and Drug Administration-approved anticancer agents in the NCI60 panel of human tumor cell lines. *Mol Cancer Ther*, 9(5), 1451-1460.
- Holford, N. H. G., & Sheiner, L. B. (1981). Understanding the Dose-Effect Relationship. *Clinical Pharmacokinetics*, 6(6), 429-453.
- Holliday, D. L., & Speirs, V. (2011). Choosing the right cell line for breast cancer research. *Breast Cancer Res*, 13(4), 215.
- Hong, R., & Xu, B. (2022). Breast cancer: an up-to-date review and future perspectives. *Cancer Communications*, 42(10), 913-936.
- Hornberg, J. J., Bruggeman, F. J., Westerhoff, H. V., & Lankelma, J. (2006). Cancer: a Systems Biology disease. *Biosystems*, 83(2-3), 81-90.
- Hou, D., Liu, Z., Xu, X., Liu, Q., Zhang, X., Kong, B., Wei, J. J., Gong, Y., & Shao, C. (2018). Increased oxidative stress mediates the antitumor effect of PARP inhibition in ovarian cancer. *Redox Biol*, 17, 99-111.
- Hovest, M. G., Brüggelolte, N., Hosseini, K. S., Krieg, T., & Herrmann, G. (2006). Senescence of Human Fibroblasts after Psoralen Photoactivation Is Mediated by ATR Kinase and Persistent DNA Damage Foci at Telomeres. *Molecular Biology of the Cell*, 17(4), 1758-1767.
- Hsu, J.-Y., Chang, C.-J., & Cheng, J.-S. (2022). Survival, treatment regimens and medical costs of women newly diagnosed with metastatic triple-negative breast cancer. *Scientific Reports*, 12(1), 729.
- Huang, A., Garraway, L. A., Ashworth, A., & Weber, B. (2020). Synthetic lethality as an engine for cancer drug target discovery. *Nature Reviews Drug Discovery*, 19(1), 23-38.
- Huang, J. L., Chang, Y. T., Hong, Z. Y., & Lin, C. S. (2022a). Targeting DNA Damage Response and Immune Checkpoint for Anticancer Therapy. *Int J Mol Sci*, 23(6).
- Huang, M., Haiderali, A., Fox, G. E., Frederickson, A., Cortes, J., Fasching, P. A., & O'Shaughnessy, J. (2022b). Economic and Humanistic Burden of Triple-Negative Breast Cancer: A Systematic Literature Review. *Pharmacoeconomics*, 40(5), 519-558.
- Huang, R.-X., & Zhou, P.-K. (2020). DNA damage response signaling pathways and targets for radiotherapy sensitization in cancer. *Signal Transduction and Targeted Therapy*, 5(1), 60.

- Huang, R., & Zhou, P.-K. (2021). DNA damage repair: historical perspectives, mechanistic pathways and clinical translation for targeted cancer therapy. *Signal Transduction and Targeted Therapy*, 6(1), 254.
- Hughes, J. P., Rees, S., Kalindjian, S. B., & Philpott, K. L. (2011). Principles of early drug discovery. *Br J Pharmacol*, 162(6), 1239-1249.
- Humphrey, R. W., Brockway-Lunardi, L. M., Bonk, D. T., Dohoney, K. M., Doroshow, J. H., Meech, S. J., Ratain, M. J., Topalian, S. L., & Pardoll, D. M. (2011). Opportunities and challenges in the development of experimental drug combinations for cancer. *J Natl Cancer Inst*, 103(16), 1222-1226.
- Iida, J., Bell-Loncella, E. T., Purazo, M. L., Lu, Y., Dorchak, J., Clancy, R., Slavik, J., Cutler, M. L., & Shriver, C. D. (2016). Inhibition of cancer cell growth by ruthenium complexes. *J Transl Med*, 14, 48.
- Ivascu, A., & Kubbies, M. (2006). Rapid Generation of Single-Tumor Spheroids for High-Throughput Cell Function and Toxicity Analysis. *SLAS Discovery*, 11(8), 922-932.
- Iyoda, M., Yamakawa, J., & Rahman, M. J. (2011). Conjugated Macrocycles: Concepts and Applications. *Angewandte Chemie International Edition*, 50(45), 10522-10553.
- Jackson, D. A., & Pombo, A. (1998). Replicon clusters are stable units of chromosome structure: evidence that nuclear organization contributes to the efficient activation and propagation of S phase in human cells. *J Cell Biol*, 140(6), 1285-1295.
- Jackson, L. M., & Moldovan, G.-L. (2022). Mechanisms of PARP1 inhibitor resistance and their implications for cancer treatment. *NAR Cancer*, 4(4).
- Jerusalem, G., de Boer, R. H., Hurvitz, S., Yardley, D. A., Kovalenko, E., Ejlertsen, B., Blau, S., Özgüroglu, M., Landherr, L., Ewertz, M., Taran, T., Fan, J., Noel-Baron, F., Louveau, A. L., & Burris, H. (2018). Everolimus Plus Exemestane vs Everolimus or Capecitabine Monotherapy for Estrogen Receptor-Positive, HER2-Negative Advanced Breast Cancer: The BOLERO-6 Randomized Clinical Trial. *JAMA Oncol*, 4(10), 1367-1374.
- Ji, L., Zhang, Q., & Liu, J. (2001). DNA structure, binding mechanism and biology functions of polypyridyl complexes in biomedicine. *Science in China Series B: Chemistry*, 44(3), 246-259.
- Jiang, Y., Dai, H., Li, Y., Yin, J., Guo, S., Lin, S. Y., & McGrail, D. J. (2019). PARP inhibitors synergize with gemcitabine by potentiating DNA damage in non-small-cell lung cancer. *Int J Cancer*, 144(5), 1092-1103.

- Jin, H., Wang, L., & Bernards, R. (2023). Rational combinations of targeted cancer therapies: background, advances and challenges. *Nature Reviews Drug Discovery*, 22(3), 213-234.
- Jorgensen, P., & Tyers, M. (2004). How Cells Coordinate Growth and Division. *Current Biology*, 14(23), R1014-R1027.
- Jourdan, J.-P., Bureau, R., Rochais, C., & Dallemagne, P. (2020). Drug repositioning: a brief overview. *Journal of Pharmacy and Pharmacology*, 72(9), 1145-1151.
- Juarez-Salinas, H., Sims, J. L., & Jacobson, M. K. (1979). Poly(ADP-ribose) levels in carcinogen-treated cells. *Nature*, 282(5740), 740-741.
- Jubin, T., Kadam, A., Jariwala, M., Bhatt, S., Sutariya, S., Gani, A. R., Gautam, S., & Begum, R. (2016). The PARP family: insights into functional aspects of poly (ADP-ribose) polymerase-1 in cell growth and survival. *Cell Prolif*, 49(4), 421-437.
- Kajstura, M., Halicka, H. D., Pryjma, J., & Darzynkiewicz, Z. (2007). Discontinuous fragmentation of nuclear DNA during apoptosis revealed by discrete "sub-G1" peaks on DNA content histograms. *Cytometry A*, 71(3), 125-131.
- Kapdi, A. R., & Fairlamb, I. J. S. (2014). Anti-cancer palladium complexes: a focus on PdX₂L₂, palladacycles and related complexes. *Chemical Society Reviews*, 43(13), 4751-4777.
- Karges, J., Heinemann, F., Jakubaszek, M., Maschietto, F., Subecz, C., Dotou, M., Vinck, R., Blacque, O., Tharaud, M., Goud, B., Viñuelas Zahinos, E., Spingler, B., Ciofini, I., & Gasser, G. (2020a). Rationally Designed Long-Wavelength Absorbing Ru(II) Polypyridyl Complexes as Photosensitizers for Photodynamic Therapy. *Journal of the American Chemical Society*, 142(14), 6578-6587.
- Karges, J., Kuang, S., Maschietto, F., Blacque, O., Ciofini, I., Chao, H., & Gasser, G. (2020b). Rationally designed ruthenium complexes for 1- and 2-photon photodynamic therapy. *Nature Communications*, 11(1), 3262.
- Katt, M. E., Placone, A. L., Wong, A. D., Xu, Z. S., & Searson, P. C. (2016). In Vitro Tumor Models: Advantages, Disadvantages, Variables, and Selecting the Right Platform. *Frontiers in Bioengineering and Biotechnology*, 4.
- Kelland, L. (2007). The resurgence of platinum-based cancer chemotherapy. *Nature Reviews Cancer*, 7(8), 573-584.

- Keung, M. Y., Wu, Y., Badar, F., & Vadgama, J. V. (2020). Response of Breast Cancer Cells to PARP Inhibitors Is Independent of BRCA Status. *Journal of Clinical Medicine*, 9(4), 940.
- Kim, D.-S., Camacho, C. V., & Kraus, W. L. (2021). Alternate therapeutic pathways for PARP inhibitors and potential mechanisms of resistance. *Experimental & Molecular Medicine*, 53(1), 42-51.
- Kim, H., George, E., Ragland, R., Rafail, S., Zhang, R., Krepler, C., Morgan, M., Herlyn, M., Brown, E., & Simpkins, F. (2017). Targeting the ATR/CHK1 Axis with PARP Inhibition Results in Tumor Regression in BRCA-Mutant Ovarian Cancer Models. *Clin Cancer Res*, 23(12), 3097-3108.
- Kim, H., Xu, H., George, E., Hallberg, D., Kumar, S., Jagannathan, V., Medvedev, S., Kinose, Y., Devins, K., Verma, P., Ly, K., Wang, Y., Greenberg, R. A., Schwartz, L., Johnson, N., Scharpf, R. B., Mills, G. B., Zhang, R., Velculescu, V. E., Brown, E. J., & Simpkins, F. (2020a). Combining PARP with ATR inhibition overcomes PARP inhibitor and platinum resistance in ovarian cancer models. *Nature Communications*, 11(1), 3726.
- Kim, J. W., Min, A., Im, S. A., Jang, H., Kim, Y. J., Kim, H. J., Lee, K. H., Kim, T. Y., Lee, K. W., Oh, D. Y., Kim, J. H., & Bang, Y. J. (2020b). TDP1 and TOP1 Modulation in Olaparib-Resistant Cancer Determines the Efficacy of Subsequent Chemotherapy. *Cancers (Basel)*, 12(2).
- Kinner, A., Wu, W., Staudt, C., & Iliakis, G. (2008). Gamma-H2AX in recognition and signaling of DNA double-strand breaks in the context of chromatin. *Nucleic Acids Res*, 36(17), 5678-5694.
- Knelson, E. H., Patel, S. A., & Sands, J. M. (2021). PARP Inhibitors in Small-Cell Lung Cancer: Rational Combinations to Improve Responses. *Cancers (Basel)*, 13(4).
- Koh, D. W., Lawler, A. M., Poitras, M. F., Sasaki, M., Wattler, S., Nehls, M. C., Stöger, T., Poirier, G. G., Dawson, V. L., & Dawson, T. M. (2004). Failure to degrade poly(ADP-ribose) causes increased sensitivity to cytotoxicity and early embryonic lethality. *Proc Natl Acad Sci U S A*, 101(51), 17699-17704.
- Kong, W., Midena, G., Chen, Y., Athanasiadis, P., Wang, T., Rousu, J., He, L., & Aittokallio, T. (2022). Systematic review of computational methods for drug combination prediction. *Computational and Structural Biotechnology Journal*, 20, 2807-2814.
- Korzyńska, A., & Zychowicz, M. (2008). A Method of Estimation of the Cell Doubling Time on Basis of the Cell Culture Monitoring Data. *Biocybernetics and Biomedical Engineering*, 28, 75-82.

- Koster, K. L., Huober, J., & Joerger, M. (2022). New antibody-drug conjugates (ADCs) in breast cancer-an overview of ADCs recently approved and in later stages of development. *Explor Target Antitumor Ther*, 3(1), 27-36.
- Kovács, R., Bakos, K., Urbányi, B., Kövesi, J., Gazsi, G., Csepeli, A., Appl Á, J., Bencsik, D., Csenki, Z., & Horváth, Á. (2016). Acute and sub-chronic toxicity of four cytostatic drugs in zebrafish. *Environ Sci Pollut Res Int*, 23(15), 14718-14729.
- Kreeger, P. K., & Lauffenburger, D. A. (2009). Cancer systems biology: a network modeling perspective. *Carcinogenesis*, 31(1), 2-8.
- Kristeleit, R., Shapiro, G. I., Burris, H. A., Oza, A. M., LoRusso, P., Patel, M. R., Domchek, S. M., Balmaña, J., Drew, Y., Chen, L. M., Safrá, T., Montes, A., Giordano, H., Maloney, L., Goble, S., Isaacson, J., Xiao, J., Borrow, J., Rolfe, L., & Shapira-Frommer, R. (2017). A Phase I-II Study of the Oral PARP Inhibitor Rucaparib in Patients with Germline BRCA1/2-Mutated Ovarian Carcinoma or Other Solid Tumors. *Clin Cancer Res*, 23(15), 4095-4106.
- Kulkarni, G. S., Lilge, L., Nesbitt, M., Dumoulin-White, R. J., Mandel, A., & Jewett, M. A. S. (2022). A Phase 1b Clinical Study of Intravesical Photodynamic Therapy in Patients with Bacillus Calmette-Guérin-unresponsive Non-muscle-invasive Bladder Cancer. *Eur Urol Open Sci*, 41, 105-111.
- Kummar, S., Chen, H. X., Wright, J., Holbeck, S., Millin, M. D., Tomaszewski, J., Zweibel, J., Collins, J., & Doroshov, J. H. (2010). Utilizing targeted cancer therapeutic agents in combination: novel approaches and urgent requirements. *Nature Reviews Drug Discovery*, 9(11), 843-856.
- Langelier, M.-F., Eisemann, T., Riccio, A. A., & Pascal, J. M. (2018). PARP family enzymes: regulation and catalysis of the poly(ADP-ribose) posttranslational modification. *Current Opinion in Structural Biology*, 53, 187-198.
- Ledermann, J., Harter, P., Gourley, C., Friedlander, M., Vergote, I., Rustin, G., Scott, C. L., Meier, W., Shapira-Frommer, R., Safrá, T., Matei, D., Fielding, A., Spencer, S., Dougherty, B., Orr, M., Hodgson, D., Barrett, J. C., & Matulonis, U. (2014). Olaparib maintenance therapy in patients with platinum-sensitive relapsed serous ovarian cancer: a preplanned retrospective analysis of outcomes by BRCA status in a randomised phase 2 trial. *Lancet Oncol*, 15(8), 852-861.
- Ledermann, J. A., Oza, A. M., Lorusso, D., Aghajanian, C., Oaknin, A., Dean, A., Colombo, N., Weberpals, J. I., Clamp, A. R., Scambia, G., Leary, A., Holloway, R. W., Gancedo, M. A., Fong, P. C., Goh, J. C., O'Malley, D.

- M., Armstrong, D. K., Banerjee, S., García-Donas, J., Swisher, E. M., Cameron, T., Maloney, L., Goble, S., & Coleman, R. L. (2020). Rucaparib for patients with platinum-sensitive, recurrent ovarian carcinoma (ARIEL3): post-progression outcomes and updated safety results from a randomised, placebo-controlled, phase 3 trial. *The Lancet Oncology*, 21(5), 710-722.
- Lee, J.-m., Zimmer, A. D. S., Lipkowitz, S., Annunziata, C. M., Ho, T. W., Chiou, V. L., Minasian, L. M., Houston, N. D., Ekwede, I., & Kohn, E. C. (2016). Phase I study of the PD-L1 inhibitor, durvalumab (MEDI4736; D) in combination with a PARP inhibitor, olaparib (O) or a VEGFR inhibitor, cediranib (C) in women's cancers (NCT02484404). *Journal of Clinical Oncology*, 34(15_suppl), 3015-3015.
- Lee, J. M., Peer, C. J., Yu, M., Amable, L., Gordon, N., Annunziata, C. M., Houston, N., Goey, A. K., Sissung, T. M., Parker, B., Minasian, L., Chiou, V. L., Murphy, R. F., Widemann, B. C., Figg, W. D., & Kohn, E. C. (2017). Sequence-Specific Pharmacokinetic and Pharmacodynamic Phase I/II Study of Olaparib Tablets and Carboplatin in Women's Cancer. *Clin Cancer Res*, 23(6), 1397-1406.
- Lee, S. Y., Kim, C. Y., & Nam, T. G. (2020). Ruthenium Complexes as Anticancer Agents: A Brief History and Perspectives. *Drug Des Devel Ther*, 14, 5375-5392.
- Leijen, S., Burgers, S. A., Baas, P., Pluim, D., Tibben, M., van Werkhoven, E., Alessio, E., Sava, G., Beijnen, J. H., & Schellens, J. H. (2015). Phase I/II study with ruthenium compound NAMI-A and gemcitabine in patients with non-small cell lung cancer after first line therapy. *Invest New Drugs*, 33(1), 201-214.
- Li, H., Liu, Z.-Y., Wu, N., Chen, Y.-C., Cheng, Q., & Wang, J. (2020). PARP inhibitor resistance: the underlying mechanisms and clinical implications. *Molecular Cancer*, 19(1), 107.
- Li, K., Zong, D., Sun, J., Chen, D., Ma, M., & Jia, L. (2022a). Rewiring of the Endocrine Network in Triple-Negative Breast Cancer. *Frontiers in Oncology*, 12.
- Li, Y., Zhang, H., Merkher, Y., Chen, L., Liu, N., Leonov, S., & Chen, Y. (2022b). Recent advances in therapeutic strategies for triple-negative breast cancer. *Journal of Hematology & Oncology*, 15(1), 121.
- Liao, H., Ji, F., Helleday, T., & Ying, S. (2018). Mechanisms for stalled replication fork stabilization: new targets for synthetic lethality strategies in cancer treatments. *EMBO Rep*, 19(9).
- Lin, K., Rong, Y., Chen, D., Zhao, Z., Bo, H., Qiao, A., Hao, X., & Wang, J. (2020). Combination of Ruthenium Complex and Doxorubicin

Synergistically Inhibits Cancer Cell Growth by Down-Regulating PI3K/AKT Signaling Pathway. *Front Oncol*, 10, 141.

Lin, K., Zhao, Z.-Z., Bo, H.-B., Hao, X.-J., & Wang, J.-Q. (2018). Applications of Ruthenium Complex in Tumor Diagnosis and Therapy. *Frontiers in Pharmacology*, 9.

Lindoy, L. F., Park, K.-M., & Lee, S. S. (2013). Metals, macrocycles and molecular assemblies – macrocyclic complexes in metallo-supramolecular chemistry. *Chemical Society Reviews*, 42(4), 1713-1727.

Litton, J. K., Rugo, H. S., Ettl, J., Hurvitz, S. A., Gonçalves, A., Lee, K.-H., Fehrenbacher, L., Yerushalmi, R., Mina, L. A., Martin, M., Roché, H., Im, Y.-H., Quek, R. G. W., Markova, D., Tudor, I. C., Hannah, A. L., Eiermann, W., & Blum, J. L. (2018). Talazoparib in Patients with Advanced Breast Cancer and a Germline BRCA Mutation. *New England Journal of Medicine*, 379(8), 753-763.

Liu, J., Zheng, W., Shi, S., Tan, C., Chen, J., Zheng, K., & Ji, L. (2008). Synthesis, antitumor activity and structure-activity relationships of a series of Ru(II) complexes. *J Inorg Biochem*, 102(2), 193-202.

Liu, S., Shiotani, B., Lahiri, M., Maréchal, A., Tse, A., Leung, C. C., Glover, J. N., Yang, X. H., & Zou, L. (2011). ATR autophosphorylation as a molecular switch for checkpoint activation. *Mol Cell*, 43(2), 192-202.

Lombard, A. P., Armstrong, C. M., D'Abronzio, L. S., Ning, S., Leslie, A. R., Sharifi, M., Lou, W., Evans, C. P., Dall'Era, M., Chen, H. W., Chen, X., & Gao, A. C. (2022). Olaparib-Induced Senescence Is Bypassed through G2-M Checkpoint Override in Olaparib-Resistant Prostate Cancer. *Mol Cancer Ther*, 21(4), 677-685.

Lombardi, P. (2002). Exemestane, a new steroidal aromatase inhibitor of clinical relevance. *Biochim Biophys Acta*, 1587(2-3), 326-337.

Lord, C. J., & Ashworth, A. (2017). PARP inhibitors: Synthetic lethality in the clinic. *Science*, 355(6330), 1152-1158.

Lui, G. Y. L., Shaw, R., Schaub, F. X., Stork, I. N., Gurley, K. E., Bridgwater, C., Diaz, R. L., Rosati, R., Swan, H. A., Ince, T. A., Harding, T. C., Gadi, V. K., Goff, B. A., Kemp, C. J., Swisher, E. M., & Grandori, C. (2020). BET, SRC, and BCL2 family inhibitors are synergistic drug combinations with PARP inhibitors in ovarian cancer. *EBioMedicine*, 60, 102988.

Łukasiewicz, S., Czezelewski, M., Forma, A., Baj, J., Sitarz, R., & Stanisławek, A. (2021). Breast Cancer-Epidemiology, Risk Factors, Classification, Prognostic Markers, and Current Treatment Strategies-An Updated Review. *Cancers (Basel)*, 13(17).

- Maajani, K., Jalali, A., Alipour, S., Khodadost, M., Tohidinik, H. R., & Yazdani, K. (2019). The Global and Regional Survival Rate of Women With Breast Cancer: A Systematic Review and Meta-analysis. *Clinical Breast Cancer*, 19(3), 165-177.
- Mahdi, H., Hafez, N., Doroshov, D., Sohal, D., Keedy, V., Do, K. T., LoRusso, P., Jürgensmeier, J., Avedissian, M., Sklar, J., Glover, C., Felicetti, B., Dean, E., Mortimer, P., Shapiro, G. I., & Eder, J. P. (2021). Ceralasertib-Mediated ATR Inhibition Combined With Olaparib in Advanced Cancers Harboring DNA Damage Response and Repair Alterations (Olaparib Combinations). *JCO Precis Oncol*, 5.
- Mahmood, T., & Yang, P. C. (2012). Western blot: technique, theory, and trouble shooting. *N Am J Med Sci*, 4(9), 429-434.
- Makhlin, I., McAndrew, N. P., Wileyto, E. P., Clark, A. S., Holmes, R., Bottalico, L. N., Mesaros, C., Blair, I. A., Jeschke, G. R., Fox, K. R., Domchek, S. M., Matro, J. M., Bradbury, A. R., Feldman, M. D., Hexner, E. O., Bromberg, J. F., & DeMichele, A. (2022). Ruxolitinib and exemestane for estrogen receptor positive, aromatase inhibitor resistant advanced breast cancer. *npj Breast Cancer*, 8(1), 122.
- Makovec, T. (2019). Cisplatin and beyond: molecular mechanisms of action and drug resistance development in cancer chemotherapy. *Radiol Oncol*, 53(2), 148-158.
- Mandapati, A., & Lukong, K. E. (2023). Triple negative breast cancer: approved treatment options and their mechanisms of action. *J Cancer Res Clin Oncol*, 149(7), 3701-3719.
- Maréchal, A., & Zou, L. (2013). DNA damage sensing by the ATM and ATR kinases. *Cold Spring Harb Perspect Biol*, 5(9).
- Marsault, E., & Peterson, M. L. (2011). Macrocycles Are Great Cycles: Applications, Opportunities, and Challenges of Synthetic Macrocycles in Drug Discovery. *Journal of Medicinal Chemistry*, 54(7), 1961-2004.
- Marzio, A., Puccini, J., Kwon, Y., Maverakis, N. K., Arbin, A., Sung, P., Bar-Sagi, D., & Pagano, M. (2019). The F-Box Domain-Dependent Activity of EMI1 Regulates PARPi Sensitivity in Triple-Negative Breast Cancers. *Mol Cell*, 73(2), 224-237.e226.
- Mateo, J., Lord, C. J., Serra, V., Tutt, A., Balmaña, J., Castroviejo-Bermejo, M., Cruz, C., Oaknin, A., Kaye, S. B., & de Bono, J. S. (2019). A decade of clinical development of PARP inhibitors in perspective. *Ann Oncol*, 30(9), 1437-1447.

- Mathews Griner, L. A., Guha, R., Shinn, P., Young, R. M., Keller, J. M., Liu, D., Goldlust, I. S., Yasgar, A., McKnight, C., Boxer, M. B., Duveau, D. Y., Jiang, J. K., Michael, S., Mierzwa, T., Huang, W., Walsh, M. J., Mott, B. T., Patel, P., Leister, W., Maloney, D. J., Leclair, C. A., Rai, G., Jadhav, A., Peyser, B. D., Austin, C. P., Martin, S. E., Simeonov, A., Ferrer, M., Staudt, L. M., & Thomas, C. J. (2014). High-throughput combinatorial screening identifies drugs that cooperate with ibrutinib to kill activated B-cell-like diffuse large B-cell lymphoma cells. *Proc Natl Acad Sci U S A*, 111(6), 2349-2354.
- McCartan, D., Bolger, J. C., Fagan, A., Byrne, C., Hao, Y., Qin, L., McIlroy, M., Xu, J., Hill, A. D., Gaora, P., & Young, L. S. (2012). Global characterization of the SRC-1 transcriptome identifies ADAM22 as an ER-independent mediator of endocrine-resistant breast cancer. *Cancer Res*, 72(1), 220-229.
- Mehta, G., Hsiao, A. Y., Ingram, M., Luker, G. D., & Takayama, S. (2012). Opportunities and challenges for use of tumor spheroids as models to test drug delivery and efficacy. *J Control Release*, 164(2), 192-204.
- Mello-Andrade, F., Cardoso, C. G., Silva, C. R. e., Chen-Chen, L., Melo-Reis, P. R. d., Lima, A. P. d., Oliveira, R., Ferraz, I. B. M., Grisolia, C. K., Almeida, M. A. P., Batista, A. A., & Silveira-Lacerda, E. d. P. (2018). Acute toxic effects of ruthenium (II)/amino acid/diphosphine complexes on Swiss mice and zebrafish embryos. *Biomedicine & Pharmacotherapy*, 107, 1082-1092.
- Michalet, X., Ekong, R., Fougereousse, F., Rousseaux, S., Schurra, C., Hornigold, N., van Slegtenhorst, M., Wolfe, J., Povey, S., Beckmann, J. S., & Bensimon, A. (1997). Dynamic molecular combing: stretching the whole human genome for high-resolution studies. *Science*, 277(5331), 1518-1523.
- Miller, E. G. (1975). Stimulation of nuclear poly (adenosine diphosphate-ribose) polymerase activity from HeLa cells by endonucleases. *Biochim Biophys Acta*, 395(2), 191-200.
- Miller, M. L., Molinelli, E. J., Nair, J. S., Sheikh, T., Samy, R., Jing, X., He, Q., Korkut, A., Crago, A. M., Singer, S., Schwartz, G. K., & Sander, C. (2013). Drug synergy screen and network modeling in dedifferentiated liposarcoma identifies CDK4 and IGF1R as synergistic drug targets. *Sci Signal*, 6(294), ra85.
- Mirza, M. R., Monk, B. J., Herrstedt, J., Oza, A. M., Mahner, S., Redondo, A., Fabbro, M., Ledermann, J. A., Lorusso, D., Vergote, I., Ben-Baruch, N. E., Marth, C., Mądry, R., Christensen, R. D., Berek, J. S., Dørum, A., Tinker, A. V., du Bois, A., González-Martín, A., Follana, P., Benigno, B., Rosenberg, P., Gilbert, L., Rimel, B. J., Buscema, J., Balser, J. P., Agarwal, S., & Matulonis, U. A. (2016). Niraparib Maintenance Therapy

in Platinum-Sensitive, Recurrent Ovarian Cancer. *N Engl J Med*, 375(22), 2154-2164.

Mishra, A. K., Abrahamsson, A., & Dabrosin, C. (2016). Fulvestrant inhibits growth of triple negative breast cancer and synergizes with tamoxifen in ER α positive breast cancer by up-regulation of ER β . *Oncotarget*, 7(35), 56876-56888.

Miwa, M., & Sugimura, T. (1971). Splitting of the ribose-ribose linkage of poly(adenosine diphosphate-ribose) by a calf thymus extract. *J Biol Chem*, 246(20), 6362-6364.

Miwa, M., Tanaka, M., Matsushima, T., & Sugimura, T. (1974). Purification and Properties of a Glycohydrolase from Calf Thymus Splitting Ribose-Ribose Linkages of Poly(Adenosine Diphosphate Ribose). *Journal of Biological Chemistry*, 249(11), 3475-3482.

Mokhtari, R. B., Homayouni, T. S., Baluch, N., Morgatskaya, E., Kumar, S., Das, B., & Yeger, H. (2017). Combination therapy in combating cancer. *Oncotarget*, 8(23), 38022-38043.

Molla, S., Chatterjee, S., Sethy, C., Sinha, S., & Kundu, C. N. (2021). Olaparib enhances curcumin-mediated apoptosis in oral cancer cells by inducing PARP trapping through modulation of BER and chromatin assembly. *DNA Repair (Amst)*, 105, 103157.

Molla, S., Hembram, K. C., Chatterjee, S., Nayak, D., Sethy, C., Pradhan, R., & Kundu, C. N. (2020). PARP inhibitor Olaparib Enhances the Apoptotic Potentiality of Curcumin by Increasing the DNA Damage in Oral Cancer Cells through Inhibition of BER Cascade. *Pathol Oncol Res*, 26(4), 2091-2103.

Monro, S., Colón, K. L., Yin, H., Roque, J., III, Konda, P., Gujar, S., Thummel, R. P., Lilge, L., Cameron, C. G., & McFarland, S. A. (2019). Transition Metal Complexes and Photodynamic Therapy from a Tumor-Centered Approach: Challenges, Opportunities, and Highlights from the Development of TLD1433. *Chemical Reviews*, 119(2), 797-828.

Moreno-Alcántar, G., Picchetti, P., & Casini, A. (2023). Gold Complexes in Anticancer Therapy: From New Design Principles to Particle-Based Delivery Systems. *Angewandte Chemie International Edition*, 62(22), e202218000.

Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods*, 65(1-2), 55-63.

- Murai, J., Huang, S. Y., Das, B. B., Renaud, A., Zhang, Y., Doroshow, J. H., Ji, J., Takeda, S., & Pommier, Y. (2012). Trapping of PARP1 and PARP2 by Clinical PARP Inhibitors. *Cancer Res*, 72(21), 5588-5599.
- Nagayama, A., Vidula, N., Ellisen, L., & Bardia, A. (2020). Novel antibody-drug conjugates for triple negative breast cancer. *Ther Adv Med Oncol*, 12, 1758835920915980.
- Nelson, K. M., Dahlin, J. L., Bisson, J., Graham, J., Pauli, G. F., & Walters, M. A. (2017). The Essential Medicinal Chemistry of Curcumin. *Journal of Medicinal Chemistry*, 60(5), 1620-1637.
- Nickoloff, J. A., Sharma, N., & Taylor, L. (2020). Clustered DNA Double-Strand Breaks: Biological Effects and Relevance to Cancer Radiotherapy. *Genes (Basel)*, 11(1).
- Nikfarjam, S., & Singh, K. K. (2023). DNA damage response signaling: A common link between cancer and cardiovascular diseases. *Cancer Medicine*, 12(4), 4380-4404.
- Nishizuka, Y., Ueda, K., Nakazawa, K., & Hayaishi, O. (1967). Studies on the polymer of adenosine diphosphate ribose. I. Enzymic formation from nicotinamide adenine dinucleotide in mammalian nuclei. *J Biol Chem*, 242(13), 3164-3171.
- Noordermeer, S. M., Adam, S., Setiawati, D., Barazas, M., Pettitt, S. J., Ling, A. K., Olivieri, M., Álvarez-Quilón, A., Moatti, N., Zimmermann, M., Annunziato, S., Krastev, D. B., Song, F., Brandsma, I., Frankum, J., Brough, R., Sherker, A., Landry, S., Szilard, R. K., Munro, M. M., McEwan, A., Gouillet de Rugy, T., Lin, Z. Y., Hart, T., Moffat, J., Gingras, A. C., Martin, A., van Attikum, H., Jonkers, J., Lord, C. J., Rottenberg, S., & Durocher, D. (2018). The shieldin complex mediates 53BP1-dependent DNA repair. *Nature*, 560(7716), 117-121.
- Nosengo, N. (2016). Can you teach old drugs new tricks? *Nature*, 534(7607), 314-316.
- Novohradsky, V., Zajac, J., Vrana, O., Kasparkova, J., & Brabec, V. (2018). Simultaneous delivery of olaparib and carboplatin in PEGylated liposomes imparts this drug combination hypersensitivity and selectivity for breast tumor cells. *Oncotarget*, 9(47), 28456-28473.
- Nuvoli, B., Camera, E., Mastrofrancesco, A., Briganti, S., & Galati, R. (2018). Modulation of reactive oxygen species via ERK and STAT3 dependent signalling are involved in the response of mesothelioma cells to exemestane. *Free Radic Biol Med*, 115, 266-277.
- O'Neil, N. J., Bailey, M. L., & Hieter, P. (2017). Synthetic lethality and cancer. *Nature Reviews Genetics*, 18(10), 613-623.

- OECD. (2013). Guideline for Testing of Chemicals, 236. Fish Embryo Acute Toxicity (FET) Test. In: OECD Paris.
- Ogiwara, H., Ui, A., Shiotani, B., Zou, L., Yasui, A., & Kohno, T. (2013). Curcumin suppresses multiple DNA damage response pathways and has potency as a sensitizer to PARP inhibitor. *Carcinogenesis*, 34(11), 2486-2497.
- Olive, P. L., & Banáth, J. P. (2006). The comet assay: a method to measure DNA damage in individual cells. *Nature Protocols*, 1(1), 23-29.
- Ostling, O., & Johanson, K. J. (1984). Microelectrophoretic study of radiation-induced DNA damages in individual mammalian cells. *Biochem Biophys Res Commun*, 123(1), 291-298.
- Oza, A. M., Cibula, D., Benzaquen, A. O., Poole, C., Mathijssen, R. H., Sonke, G. S., Colombo, N., Špaček, J., Vuylsteke, P., Hirte, H., Mahner, S., Plante, M., Schmalfeldt, B., Mackay, H., Rowbottom, J., Lowe, E. S., Dougherty, B., Barrett, J. C., & Friedlander, M. (2015). Olaparib combined with chemotherapy for recurrent platinum-sensitive ovarian cancer: a randomised phase 2 trial. *Lancet Oncol*, 16(1), 87-97.
- Oza, A. M., Tinker, A. V., Oaknin, A., Shapira-Frommer, R., McNeish, I. A., Swisher, E. M., Ray-Coquard, I., Bell-McGuinn, K., Coleman, R. L., O'Malley, D. M., Leary, A., Chen, L. M., Provencher, D., Ma, L., Brenton, J. D., Konecny, G. E., Castro, C. M., Giordano, H., Maloney, L., Goble, S., Lin, K. K., Sun, J., Raponi, M., Rolfe, L., & Kristeleit, R. S. (2017). Antitumor activity and safety of the PARP inhibitor rucaparib in patients with high-grade ovarian carcinoma and a germline or somatic BRCA1 or BRCA2 mutation: Integrated analysis of data from Study 10 and ARIEL2. *Gynecol Oncol*, 147(2), 267-275.
- Pal, R., & Seleem, M. N. (2020). Screening of Natural Products and Approved Oncology Drug Libraries for Activity against *Clostridioides difficile*. *Scientific Reports*, 10(1), 5966.
- Palmer, R. G., & Denman, A. M. (1984). Malignancies induced by chlorambucil. *Cancer Treat Rev*, 11(2), 121-129.
- Pascal, J. M. (2018). The comings and goings of PARP-1 in response to DNA damage. *DNA Repair (Amst)*, 71, 177-182.
- Patton, E. E., Zon, L. I., & Langenau, D. M. (2021). Zebrafish disease models in drug discovery: from preclinical modelling to clinical trials. *Nat Rev Drug Discov*, 20(8), 611-628.
- Pavese, F., Capoluongo, E. D., Muratore, M., Minucci, A., Santonocito, C., Fuso, P., Concolino, P., Di Stasio, E., Carbognin, L., Tiberi, G.,

- Garganese, G., Corrado, G., Di Leone, A., Generali, D., Fragomeni, S. M., D'Angelo, T., Franceschini, G., Masetti, R., Fabi, A., Mulè, A., Santoro, A., Belli, P., Tortora, G., Scambia, G., & Paris, I. (2022). BRCA Mutation Status in Triple-Negative Breast Cancer Patients Treated with Neoadjuvant Chemotherapy: A Pivotal Role for Treatment Decision-Making. *Cancers (Basel)*, *14*(19).
- Perillo, B., Di Donato, M., Pezone, A., Di Zazzo, E., Giovannelli, P., Galasso, G., Castoria, G., & Migliaccio, A. (2020). ROS in cancer therapy: the bright side of the moon. *Experimental & Molecular Medicine*, *52*(2), 192-203.
- Pilié, P. G., Gay, C. M., Byers, L. A., O'Connor, M. J., & Yap, T. A. (2019). PARP Inhibitors: Extending Benefit Beyond BRCA-Mutant Cancers. *Clin Cancer Res*, *25*(13), 3759-3771.
- Pines, A., Vrouwe, M. G., Martijn, J. A., Typas, D., Luijsterburg, M. S., Cansoy, M., Hensbergen, P., Deelder, A., de Groot, A., Matsumoto, S., Sugawara, K., Thoma, N., Vermeulen, W., Vrieling, H., & Mullenders, L. (2012). PARP1 promotes nucleotide excision repair through DDB2 stabilization and recruitment of ALC1. *J Cell Biol*, *199*(2), 235-249.
- Plesca, D., Mazumder, S., & Almasan, A. (2008). DNA damage response and apoptosis. *Methods Enzymol*, *446*, 107-122.
- Plummer, R., Jones, C., Middleton, M., Wilson, R., Evans, J., Olsen, A., Curtin, N., Boddy, A., McHugh, P., Newell, D., Harris, A., Johnson, P., Steinfeldt, H., Dewji, R., Wang, D., Robson, L., & Calvert, H. (2008). Phase I Study of the Poly(ADP-Ribose) Polymerase Inhibitor, AG014699, in Combination with Temozolomide in Patients with Advanced Solid Tumors. *Clinical Cancer Research*, *14*(23), 7917-7923.
- Plummer, R., Verheul, H. M., De Vos, F., Leunen, K., Molife, L. R., Rolfo, C., Grundtvig-Sørensen, P., De Grève, J., Rottey, S., Jerusalem, G., Italiano, A., Spicer, J., Dirix, L., Goessl, C., Birkett, J., Spencer, S., Learoyd, M., Bailey, C., & Dean, E. (2018). Pharmacokinetic Effects and Safety of Olaparib Administered with Endocrine Therapy: A Phase I Study in Patients with Advanced Solid Tumours. *Adv Ther*, *35*(11), 1945-1964.
- Podhorecka, M., Skladanowski, A., & Bozko, P. (2010). H2AX Phosphorylation: Its Role in DNA Damage Response and Cancer Therapy. *J Nucleic Acids*, *2010*.
- Poveda, A., Floquet, A., Ledermann, J. A., Asher, R., Penson, R. T., Oza, A. M., Korach, J., Huzarski, T., Pignata, S., Friedlander, M., Baldoni, A., Park-Simon, T.-W., Sonke, G. S., Lisyanskaya, A. S., Kim, J.-H., Filho, E. A., Vergote, I., Rowe, P., & Pujade-Lauraine, E. (2020). Final overall survival (OS) results from SOLO2/ENGOT-ov21: A phase III trial

assessing maintenance olaparib in patients (pts) with platinum-sensitive, relapsed ovarian cancer and a BRCA mutation. *Journal of Clinical Oncology*, 38(15_suppl), 6002-6002.

- Poveda, A., Floquet, A., Ledermann, J. A., Asher, R., Penson, R. T., Oza, A. M., Korach, J., Huzarski, T., Pignata, S., Friedlander, M., Baldoni, A., Park-Simon, T. W., Tamura, K., Sonke, G. S., Lisyanskaya, A., Kim, J. H., Filho, E. A., Milenkova, T., Lowe, E. S., Rowe, P., Vergote, I., & Pujade-Lauraine, E. (2021). Olaparib tablets as maintenance therapy in patients with platinum-sensitive relapsed ovarian cancer and a BRCA1/2 mutation (SOLO2/ENGOT-Ov21): a final analysis of a double-blind, randomised, placebo-controlled, phase 3 trial. *Lancet Oncol*, 22(5), 620-631.
- Poveda, A., Lheureux, S., Colombo, N., Cibula, D., Lindemann, K., Weberpals, J., Bjurberg, M., Oaknin, A., Sikorska, M., González-Martín, A., Madry, R., Pérez, M. J. R., Ledermann, J., Davidson, R., Blakeley, C., Bennett, J., Barnicle, A., & Škof, E. (2022). Olaparib maintenance monotherapy in platinum-sensitive relapsed ovarian cancer patients without a germline BRCA1/BRCA2 mutation: OPINION primary analysis. *Gynecologic Oncology*, 164(3), 498-504.
- Poynton, F. E., Bright, S. A., Blasco, S., Williams, D. C., Kelly, J. M., & Gunnlaugsson, T. (2017). The development of ruthenium(ii) polypyridyl complexes and conjugates for in vitro cellular and in vivo applications. *Chemical Society Reviews*, 46(24), 7706-7756.
- Prasad, C. B., Prasad, S. B., Yadav, S. S., Pandey, L. K., Singh, S., Pradhan, S., & Narayan, G. (2017). Olaparib modulates DNA repair efficiency, sensitizes cervical cancer cells to cisplatin and exhibits anti-metastatic property. *Scientific Reports*, 7(1), 12876.
- Psyrris, A., Economopoulou, P., Kotsantis, I., Koutsodontis, G., Cheila, M., Papaxoinis, G., Pectasides, D. G., Fountzilas, G., Krikoni, L., & Souliotis, V. (2018). A phase II window of opportunity study of preoperative olaparib (O) with cisplatin (C) or durvalumab (D) or olaparib alone in patients with operable squamous cell head and neck carcinoma (HNSCC) (OPHELIA). *Annals of Oncology*, 29, viii372.
- Puckett, C. A., & Barton, J. K. (2007). Methods to Explore Cellular Uptake of Ruthenium Complexes. *Journal of the American Chemical Society*, 129(1), 46-47.
- Puckett, C. A., Ernst, R. J., & Barton, J. K. (2010). Exploring the cellular accumulation of metal complexes. *Dalton Trans*, 39(5), 1159-1170.
- Purnell, M. R., & Whish, W. J. (1980). Novel inhibitors of poly(ADP-ribose) synthetase. *Biochem J*, 185(3), 775-777.

- Pushpakom, S., Iorio, F., Eyers, P. A., Escott, K. J., Hopper, S., Wells, A., Doig, A., Williams, T., Latimer, J., McNamee, C., Norris, A., Sanseau, P., Cavalla, D., & Pirmohamed, M. (2019). Drug repurposing: progress, challenges and recommendations. *Nature Reviews Drug Discovery*, 18(1), 41-58.
- Qiu, Z., Oleinick, N. L., & Zhang, J. (2018). ATR/CHK1 inhibitors and cancer therapy. *Radiother Oncol*, 126(3), 450-464.
- Rademaker-Lakhai, J. M., van den Bongard, D., Pluim, D., Beijnen, J. H., & Schellens, J. H. (2004). A Phase I and pharmacological study with imidazolium-trans-DMSO-imidazole-tetrachlororuthenate, a novel ruthenium anticancer agent. *Clin Cancer Res*, 10(11), 3717-3727.
- Rani, A., Stebbing, J., Giamas, G., & Murphy, J. (2019). Endocrine Resistance in Hormone Receptor Positive Breast Cancer—From Mechanism to Therapy. *Frontiers in Endocrinology*, 10.
- Ransy, C., Vaz, C., Lombès, A., & Bouillaud, F. (2020). Use of H₂O₂ to Cause Oxidative Stress, the Catalase Issue. *Int J Mol Sci*, 21(23).
- Rasouli, R., & Tabrizian, M. (2021). Rapid Formation of Multicellular Spheroids in Boundary-Driven Acoustic Microstreams. *Small*, 17(39), 2101931.
- Raynal, N. J., Da Costa, E. M., Lee, J. T., Gharibyan, V., Ahmed, S., Zhang, H., Sato, T., Malouf, G. G., & Issa, J. J. (2017). Repositioning FDA-Approved Drugs in Combination with Epigenetic Drugs to Reprogram Colon Cancer Epigenome. *Mol Cancer Ther*, 16(2), 397-407.
- Redza-Dutordoir, M., & Averill-Bates, D. A. (2016). Activation of apoptosis signalling pathways by reactive oxygen species. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*, 1863(12), 2977-2992.
- Remon, J., Besse, B., Leary, A., Bièche, I., Job, B., Lacroix, L., Auguste, A., Mauduit, M., Audigier-Valette, C., Raimbourg, J., Madroszyk, A., Michels, S., Bayar, M. A., Jimenez, M., Soria, J.-C., Rouleau, E., & Barlesi, F. (2020). Somatic and Germline BRCA 1 and 2 Mutations in Advanced NSCLC From the SAFIR02-Lung Trial. *JTO Clinical and Research Reports*, 1(3), 100068.
- Riccardi, C., & Nicoletti, I. (2006). Analysis of apoptosis by propidium iodide staining and flow cytometry. *Nature Protocols*, 1(3), 1458-1461.
- Richards, M., Anderson, M., Carter, P., Ebert, B. L., & Mossialos, E. (2020). The impact of the COVID-19 pandemic on cancer care. *Nature Cancer*, 1(6), 565-567.

- Rilak Simović, A., Masnikosa, R., Bratsos, I., & Alessio, E. (2019). Chemistry and reactivity of ruthenium(II) complexes: DNA/protein binding mode and anticancer activity are related to the complex structure. *Coordination Chemistry Reviews*, 398, 113011.
- Robson, M. E., Im, S. A., Senkus, E., Xu, B., Domchek, S. M., Masuda, N., Delaloge, S., Li, W., Tung, N., Armstrong, A., Wu, W., Goessl, C., Runswick, S., & Conte, P. (2017). Olaparib for Metastatic Breast Cancer in Patients with a Germline BRCA Mutation. *N Engl J Med*, 377(6), 523-533.
- Robson, M. E., Tung, N., Conte, P., Im, S. A., Senkus, E., Xu, B., Masuda, N., Delaloge, S., Li, W., Armstrong, A., Wu, W., Goessl, C., Runswick, S., & Domchek, S. M. (2019). OlympiAD final overall survival and tolerability results: Olaparib versus chemotherapy treatment of physician's choice in patients with a germline BRCA mutation and HER2-negative metastatic breast cancer. *Ann Oncol*, 30(4), 558-566.
- Rodea-Palomares, I., González-Pleiter, M., Martín-Betancor, K., Rosal, R., & Fernández-Piñas, F. (2015). Additivity and Interactions in Ecotoxicity of Pollutant Mixtures: Some Patterns, Conclusions, and Open Questions. *Toxics*, 3(4), 342-369.
- Roell, K. R., Reif, D. M., & Motsinger-Reif, A. A. (2017). An Introduction to Terminology and Methodology of Chemical Synergy—Perspectives from Across Disciplines. *Frontiers in Pharmacology*, 8.
- Roller, D. G., Axelrod, M., Capaldo, B. J., Jensen, K., Mackey, A., Weber, M. J., & Gioeli, D. (2012). Synthetic lethal screening with small-molecule inhibitors provides a pathway to rational combination therapies for melanoma. *Mol Cancer Ther*, 11(11), 2505-2515.
- Roos, W. P., & Kaina, B. (2006). DNA damage-induced cell death by apoptosis. *Trends Mol Med*, 12(9), 440-450.
- Rose, M., Burgess, J. T., O'Byrne, K., Richard, D. J., & Bolderson, E. (2020). PARP Inhibitors: Clinical Relevance, Mechanisms of Action and Tumor Resistance. *Front Cell Dev Biol*, 8, 564601.
- Rosenberg, B., Vancamp, L., Trosko, J. E., & Mansour, V. H. (1969). Platinum Compounds: a New Class of Potent Antitumour Agents. *Nature*, 222(5191), 385-386.
- Rothenberg, M. L. (1997). Topoisomerase I inhibitors: Review and update. *Annals of Oncology*, 8(9), 837-855.
- Rottenberg, S., Jaspers, J. E., Kersbergen, A., van der Burg, E., Nygren, A. O. H., Zander, S. A. L., Derksen, P. W. B., de Bruin, M., Zevenhoven, J., Lau, A., Boulter, R., Cranston, A., O'Connor, M. J., Martin, N. M. B.,

- Borst, P., & Jonkers, J. (2008). High sensitivity of BRCA1-deficient mammary tumors to the PARP inhibitor AZD2281 alone and in combination with platinum drugs. *Proceedings of the National Academy of Sciences*, 105(44), 17079-17084.
- Rouleau, M., Patel, A., Hendzel, M. J., Kaufmann, S. H., & Poirier, G. G. (2010). PARP inhibition: PARP1 and beyond. *Nat Rev Cancer*, 10(4), 293-301.
- Sabnis, G. J., Goloubeva, O., Chumsri, S., Nguyen, N., Sukumar, S., & Brodie, A. M. (2011). Functional activation of the estrogen receptor- α and aromatase by the HDAC inhibitor entinostat sensitizes ER-negative tumors to letrozole. *Cancer Res*, 71(5), 1893-1903.
- Sadoughi, F., Hallajzadeh, J., Asemi, Z., Mansournia, M. A., Alemi, F., & Yousefi, B. (2021). Signaling pathways involved in cell cycle arrest during the DNA breaks. *DNA Repair*, 98, 103047.
- Sakai, W., Swisher, E. M., Karlan, B. Y., Agarwal, M. K., Higgins, J., Friedman, C., Villegas, E., Jacquemont, C., Farrugia, D. J., Couch, F. J., Urban, N., & Taniguchi, T. (2008). Secondary mutations as a mechanism of cisplatin resistance in BRCA2-mutated cancers. *Nature*, 451(7182), 1116-1120.
- Saldivar, J. C., Cortez, D., & Cimprich, K. A. (2017). The essential kinase ATR: ensuring faithful duplication of a challenging genome. *Nat Rev Mol Cell Biol*, 18(10), 622-636.
- Sarin, N., Engel, F., Kalayda, G. V., Mannewitz, M., Cinatl, J., Jr., Rothweiler, F., Michaelis, M., Saafan, H., Ritter, C. A., Jaehde, U., & Frötschl, R. (2017). Cisplatin resistance in non-small cell lung cancer cells is associated with an abrogation of cisplatin-induced G2/M cell cycle arrest. *PLoS One*, 12(7), e0181081.
- Sartorius, C. A., Hanna, C. T., Gril, B., Cruz, H., Serkova, N. J., Huber, K. M., Kabos, P., Schedin, T. B., Borges, V. F., Steeg, P. S., & Cittelly, D. M. (2016). Estrogen promotes the brain metastatic colonization of triple negative breast cancer cells via an astrocyte-mediated paracrine mechanism. *Oncogene*, 35(22), 2881-2892.
- Sato, M. S., & Lindahl, T. (1992). Role of poly(ADP-ribose) formation in DNA repair. *Nature*, 356(6367), 356-358.
- Scandlyn, M. J., Stuart, E. C., Somers-Edgar, T. J., Menzies, A. R., & Rosengren, R. J. (2008). A new role for tamoxifen in oestrogen receptor-negative breast cancer when it is combined with epigallocatechin gallate. *Br J Cancer*, 99(7), 1056-1063.

- Scattolin, T., Voloshkin, V. A., Visentin, F., & Nolan, S. P. (2021). A critical review of palladium organometallic anticancer agents. *Cell Reports Physical Science*, 2(6), 100446.
- Schaefer, M. H., & Serrano, L. (2016). Cell type-specific properties and environment shape tissue specificity of cancer genes. *Scientific Reports*, 6(1), 20707.
- Schmidt, L., Kling, T., Monsefi, N., Olsson, M., Hansson, C., Baskaran, S., Lundgren, B., Martens, U., Häggblad, M., Westermarck, B., Forsberg Nilsson, K., Uhrbom, L., Karlsson-Lindahl, L., Gerlee, P., & Nelander, S. (2013). Comparative drug pair screening across multiple glioblastoma cell lines reveals novel drug-drug interactions. *Neuro Oncol*, 15(11), 1469-1478.
- Senga, S. S., & Grose, R. P. (2021). Hallmarks of cancer-the new testament. *Open Biol*, 11(1), 200358.
- Sgouros, G., Bodei, L., McDevitt, M. R., & Nedrow, J. R. (2020). Radiopharmaceutical therapy in cancer: clinical advances and challenges. *Nature Reviews Drug Discovery*, 19(9), 589-608.
- Shah, P. D., Wethington, S. L., Pagan, C., Latif, N., Tanyi, J., Martin, L. P., Morgan, M., Burger, R. A., Haggerty, A., Zarrin, H., Rodriguez, D., Domchek, S., Drapkin, R., Shih, I. M., Smith, S. A., Dean, E., Gaillard, S., Armstrong, D., Torigian, D. A., Hwang, W. T., Giuntoli, R., & Simpkins, F. (2021). Combination ATR and PARP Inhibitor (CAPRI): A phase 2 study of ceralasertib plus olaparib in patients with recurrent, platinum-resistant epithelial ovarian cancer. *Gynecol Oncol*, 163(2), 246-253.
- Shanle, E. K., Onitilo, A. A., Huang, W., Kim, K., Zang, C., Engel, J. M., Xu, W., & Wisinski, K. B. (2015). Prognostic significance of full-length estrogen receptor beta expression in stage I-III triple negative breast cancer. *Am J Transl Res*, 7(7), 1246-1259.
- Sharma, D., Rasool, F., Bharti, M., Vyas, K. M., & Magani, S. K. J. (2023). Regorafenib and Ruthenium Complex Combination Inhibit Cancer Cell Growth by Targeting PI3K/AKT/ERK Signalling in Colorectal Cancer Cells. *International Journal of Molecular Sciences*, 24(1), 686.
- Shen, D. W., Fojo, A., Chin, J. E., Roninson, I. B., Richert, N., Pastan, I., & Gottesman, M. M. (1986). Human multidrug-resistant cell lines: increased *mdr1* expression can precede gene amplification. *Science*, 232(4750), 643-645.
- Shen, Y., Aoyagi-Scharber, M., & Wang, B. (2015). Trapping Poly(ADP-Ribose) Polymerase. *J Pharmacol Exp Ther*, 353(3), 446-457.

- Shen, Y. T., Evans, J. C., Zafarana, G., Allen, C., & Piquette-Miller, M. (2018). BRCA Status Does Not Predict Synergism of a Carboplatin and Olaparib Combination in High-Grade Serous Ovarian Cancer Cell Lines. *Mol Pharm*, 15(7), 2742-2753.
- Shum, J., Leung, P. K.-K., & Lo, K. K.-W. (2019). Luminescent Ruthenium(II) Polypyridine Complexes for a Wide Variety of Biomolecular and Cellular Applications. *Inorganic Chemistry*, 58(4), 2231-2247.
- Siegel, R. L., Miller, K. D., Wagle, N. S., & Jemal, A. (2023). Cancer statistics, 2023. *CA: A Cancer Journal for Clinicians*, 73(1), 17-48.
- Singh, D. D., Parveen, A., & Yadav, D. K. (2021). Role of PARP in TNBC: Mechanism of Inhibition, Clinical Applications, and Resistance. *Biomedicines*, 9(11).
- Singh, N. P., McCoy, M. T., Tice, R. R., & Schneider, E. L. (1988). A simple technique for quantitation of low levels of DNA damage in individual cells. *Exp Cell Res*, 175(1), 184-191.
- Skidmore, C. J., Davies, M. I., Goodwin, P. M., Halldorsson, H., Lewis, P. J., Shall, S., & Zia'ee, A. A. (1979). The involvement of poly(ADP-ribose) polymerase in the degradation of NAD caused by gamma-radiation and N-methyl-N-nitrosourea. *Eur J Biochem*, 101(1), 135-142.
- Skok, Ž., Zidar, N., Kikelj, D., & Ilaš, J. (2020). Dual Inhibitors of Human DNA Topoisomerase II and Other Cancer-Related Targets. *Journal of Medicinal Chemistry*, 63(3), 884-904.
- Slootbeek, P. H. J., Kloots, I. S. H., van Oort, I. M., Kroeze, L. I., Schalken, J. A., Bloemendal, H. J., & Mehra, N. (2023). Cross-Resistance between Platinum-Based Chemotherapy and PARP Inhibitors in Castration-Resistant Prostate Cancer. *Cancers (Basel)*, 15(10).
- Spigel, D. R. (2012). PARP inhibitors in lung cancer. *J Thorac Oncol*, 7(16 Suppl 5), S392-393.
- Stewart, R. L., Updike, K. L., Factor, R. E., Henry, N. L., Boucher, K. M., Bernard, P. S., & Varley, K. E. (2019). A Multigene Assay Determines Risk of Recurrence in Patients with Triple-Negative Breast Cancer. *Cancer Res*, 79(13), 3466-3478.
- Strähle, U., Scholz, S., Geisler, R., Greiner, P., Hollert, H., Rastegar, S., Schumacher, A., Selderslaghs, I., Weiss, C., Witters, H., & Braunbeck, T. (2012). Zebrafish embryos as an alternative to animal experiments--a commentary on the definition of the onset of protected life stages in animal welfare regulations. *Reprod Toxicol*, 33(2), 128-132.

- Sun, D., Gao, W., Hu, H., & Zhou, S. (2022). Why 90% of clinical drug development fails and how to improve it? *Acta Pharm Sin B*, 12(7), 3049-3062.
- Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., & Bray, F. (2021). Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer Journal for Clinicians*, 71(3), 209-249.
- Swisher, E. M., Kwan, T. T., Oza, A. M., Tinker, A. V., Ray-Coquard, I., Oaknin, A., Coleman, R. L., Aghajanian, C., Konecny, G. E., O'Malley, D. M., Leary, A., Provencher, D., Welch, S., Chen, L.-m., Wahner Hendrickson, A. E., Ma, L., Ghatage, P., Kristeleit, R. S., Dorigo, O., Muser, A., Kaufmann, S. H., Elvin, J. A., Lin, D. I., Chambers, S. K., Dominy, E., Vo, L.-T., Goble, S., Maloney, L., Giordano, H., Harding, T., Dobrovic, A., Scott, C. L., Lin, K. K., & McNeish, I. A. (2021). Molecular and clinical determinants of response and resistance to rucaparib for recurrent ovarian cancer treatment in ARIEL2 (Parts 1 and 2). *Nature Communications*, 12(1), 2487.
- Swisher, E. M., Lin, K. K., Oza, A. M., Scott, C. L., Giordano, H., Sun, J., Konecny, G. E., Coleman, R. L., Tinker, A. V., O'Malley, D. M., Kristeleit, R. S., Ma, L., Bell-McGuinn, K. M., Brenton, J. D., Cragun, J. M., Oaknin, A., Ray-Coquard, I., Harrell, M. I., Mann, E., Kaufmann, S. H., Floquet, A., Leary, A., Harding, T. C., Goble, S., Maloney, L., Isaacson, J., Allen, A. R., Rolfe, L., Yelensky, R., Raponi, M., & McNeish, I. A. (2017). Rucaparib in relapsed, platinum-sensitive high-grade ovarian carcinoma (ARIEL2 Part 1): an international, multicentre, open-label, phase 2 trial. *Lancet Oncol*, 18(1), 75-87.
- Szostakowska, M., Trębińska-Stryjewska, A., Grzybowska, E. A., & Fabisiowicz, A. (2019). Resistance to endocrine therapy in breast cancer: molecular mechanisms and future goals. *Breast Cancer Research and Treatment*, 173(3), 489-497.
- Tagliatela, A., Alvarez, S., Leuzzi, G., Sannino, V., Ranjha, L., Huang, J. W., Madubata, C., Anand, R., Levy, B., Rabadan, R., Cejka, P., Costanzo, V., & Ciccio, A. (2017). Restoration of Replication Fork Stability in BRCA1- and BRCA2-Deficient Cells by Inactivation of SNF2-Family Fork Remodelers. *Mol Cell*, 68(2), 414-430.e418.
- Tallarida, R. J. (2011). Quantitative methods for assessing drug synergism. *Genes Cancer*, 2(11), 1003-1008.
- Tang, C., Livingston, M. J., Safirstein, R., & Dong, Z. (2023). Cisplatin nephrotoxicity: new insights and therapeutic implications. *Nature Reviews Nephrology*, 19(1), 53-72.

- Ter Brugge, P., Kristel, P., van der Burg, E., Boon, U., de Maaker, M., Lips, E., Mulder, L., de Ruiter, J., Moutinho, C., Gevensleben, H., Marangoni, E., Majewski, I., Józwiak, K., Kloosterman, W., van Roosmalen, M., Duran, K., Hogervorst, F., Turner, N., Esteller, M., Cuppen, E., Wesseling, J., & Jonkers, J. (2016). Mechanisms of Therapy Resistance in Patient-Derived Xenograft Models of BRCA1-Deficient Breast Cancer. *J Natl Cancer Inst*, 108(11).
- Thomas, R., Al-Khadairi, G., & Decock, J. (2021). Immune Checkpoint Inhibitors in Triple Negative Breast Cancer Treatment: Promising Future Prospects. *Frontiers in Oncology*, 10.
- Tomasik, B., Bieńkowski, M., & Jassem, J. (2021). PARP inhibitors beyond BRCA -mutated cancers: precision medicine at the crossroads. *Precision Cancer Medicine*, 4.
- Toss, A., Venturelli, M., Civallero, M., Piombino, C., Domati, F., Ficarra, G., Combi, F., Cabitza, E., Caggia, F., Barbieri, E., Barbolini, M., Moschetti, L., Omarini, C., Piacentini, F., Tazzioli, G., Dominici, M., & Cortesi, L. (2022). Predictive factors for relapse in triple-negative breast cancer patients without pathological complete response after neoadjuvant chemotherapy. *Front Oncol*, 12, 1016295.
- Trondl, R., Heffeter, P., Kowol, C. R., Jakupiec, M. A., Berger, W., & Keppler, B. K. (2014). NKP-1339, the first ruthenium-based anticancer drug on the edge to clinical application. *Chem. Sci.*, 5(8), 2925-2932.
- Trucco, C., Javier Oliver, F., de Murcia, G., & Ménissier-de Murcia, J. (1998). DNA repair defect in poly(ADP-ribose) polymerase-deficient cell lines. *Nucleic Acids Research*, 26(11), 2644-2649.
- Tubbs, A., & Nussenzweig, A. (2017). Endogenous DNA Damage as a Source of Genomic Instability in Cancer. *Cell*, 168(4), 644-656.
- Ueda, K., Cardarelli, C., Gottesman, M. M., & Pastan, I. (1987). Expression of a full-length cDNA for the human "MDR1" gene confers resistance to colchicine, doxorubicin, and vinblastine. *Proc Natl Acad Sci U S A*, 84(9), 3004-3008.
- Vaidyanathan, A., Sawers, L., Gannon, A. L., Chakravarty, P., Scott, A. L., Bray, S. E., Ferguson, M. J., & Smith, G. (2016). ABCB1 (MDR1) induction defines a common resistance mechanism in paclitaxel- and olaparib-resistant ovarian cancer cells. *Br J Cancer*, 115(4), 431-441.
- Van Asten, K., Neven, P., Lintermans, A., Wildiers, H., & Paridaens, R. (2014). Aromatase inhibitors in the breast cancer clinic: focus on exemestane. *Endocrine-Related Cancer*, 21(1), R31-R49.

- Varbanov, H. P., Kuttler, F., Banfi, D., Turcatti, G., & Dyson, P. J. (2017). Repositioning approved drugs for the treatment of problematic cancers using a screening approach. *PLoS One*, 12(2), e0171052.
- Vinod, S. K., & Hau, E. (2020). Radiotherapy treatment for lung cancer: Current status and future directions. *Respirology*, 25(S2), 61-71.
- Visconti, R., Della Monica, R., & Grieco, D. (2016). Cell cycle checkpoint in cancer: a therapeutically targetable double-edged sword. *Journal of Experimental & Clinical Cancer Research*, 35(1), 153.
- Walker, M. G., Jarman, P. J., Gill, M. R., Tian, X., Ahmad, H., Reddy, P. A. N., McKenzie, L., Weinstein, J. A., Meijer, A. J. H. M., Battaglia, G., Smythe, C. G. W., & Thomas, J. A. (2016). A Self-Assembled Metallomacrocyclic Singlet Oxygen Sensitizer for Photodynamic Therapy. *Chemistry – A European Journal*, 22(17), 5996-6000.
- Wang, J., Aroumougame, A., Loblrich, M., Li, Y., Chen, D., Chen, J., & Gong, Z. (2014). PTIP associates with Artemis to dictate DNA repair pathway choice. *Genes Dev*, 28(24), 2693-2698.
- Wang, J. Y. J. (2001). DNA damage and apoptosis. *Cell Death & Differentiation*, 8(11), 1047-1048.
- Wang, L.-M., Wang, P., Chen, X.-M., Yang, H., Song, S.-S., Song, Z., Jia, L., Chen, H.-D., Bao, X.-B., Guo, N., Huan, X.-J., Xi, Y., Shen, Y.-Y., Yang, X.-Y., Su, Y., Sun, Y.-M., Gao, Y.-L., Chen, Y., Ding, J., Lang, J.-Y., Miao, Z.-H., Zhang, A., & He, J.-X. (2023). Thioparib inhibits homologous recombination repair, activates the type I IFN response, and overcomes olaparib resistance. *EMBO Molecular Medicine*, 15(3), e16235.
- Wang, N., Yang, Y., Jin, D., Zhang, Z., Shen, K., Yang, J., Chen, H., Zhao, X., Yang, L., & Lu, H. (2022). PARP inhibitor resistance in breast and gynecological cancer: Resistance mechanisms and combination therapy strategies. *Frontiers in Pharmacology*, 13.
- Wang, Q., Cui, K., Espin-Garcia, O., Cheng, D., Qiu, X., Chen, Z., Moore, M., Bristow, R. G., Xu, W., Der, S., & Liu, G. (2013a). Resistance to bleomycin in cancer cell lines is characterized by prolonged doubling time, reduced DNA damage and evasion of G2/M arrest and apoptosis. *PLoS One*, 8(12), e82363.
- Wang, X., Pan, L., Mao, N., Sun, L., Qin, X., & Yin, J. (2013b). Cell-cycle synchronization reverses Taxol resistance of human ovarian cancer cell lines. *Cancer Cell International*, 13(1), 77.
- Wang, X., Zhang, H., & Chen, X. (2019). Drug resistance and combating drug resistance in cancer. *Cancer Drug Resist*, 2(2), 141-160.

- Wang, Y. T., Yuan, B., Chen, H. D., Xu, L., Tian, Y. N., Zhang, A., He, J. X., & Miao, Z. H. (2018). Acquired resistance of phosphatase and tensin homolog-deficient cells to poly(ADP-ribose) polymerase inhibitor and Ara-C mediated by 53BP1 loss and SAMHD1 overexpression. *Cancer Sci*, 109(3), 821-831.
- Weiss, A., Bonvin, D., Berndsen, R. H., Scherrer, E., Wong, T. J., Dyson, P. J., Griffioen, A. W., & Nowak-Sliwinska, P. (2015). Angiostatic treatment prior to chemo- or photodynamic therapy improves anti-tumor efficacy. *Scientific Reports*, 5(1), 8990.
- Westin, S. N., Coleman, R. L., Fellman, B. M., Yuan, Y., Sood, A. K., Soliman, P. T., Wright, A. A., Horowitz, N. S., Campos, S. M., Konstantinopoulos, P. A., Levenback, C. F., Gershenson, D. M., Lu, K. H., Bayer, V., Tuki, S., Rabbit, A., Ottesen, L., Godin, R., Mills, G. B., & Liu, J. F. (2021). EFFORT: EFFicacy Of adavosertib in parp ResisTance: A randomized two-arm non-comparative phase II study of adavosertib with or without olaparib in women with PARP-resistant ovarian cancer. *Journal of Clinical Oncology*, 39(15_suppl), 5505-5505.
- Wilson, J. J., & Lippard, S. J. (2012). In Vitro Anticancer Activity of cis-Diammineplatinum(II) Complexes with β -Diketone Leaving Group Ligands. *Journal of Medicinal Chemistry*, 55(11), 5326-5336.
- Wu, W., Warner, M., Wang, L., He, W.-W., Zhao, R., Guan, X., Botero, C., Huang, B., Ion, C., Coombes, C., & Gustafsson, J.-A. (2021). Drivers and suppressors of triple-negative breast cancer. *Proceedings of the National Academy of Sciences*, 118(33), e2104162118.
- Xu, G., Chapman, J. R., Brandsma, I., Yuan, J., Mistrik, M., Bouwman, P., Bartkova, J., Gogola, E., Warmerdam, D., Barazas, M., Jaspers, J. E., Watanabe, K., Pieterse, M., Kersbergen, A., Sol, W., Celie, P. H. N., Schouten, P. C., van den Broek, B., Salman, A., Nieuwland, M., de Rink, I., de Ronde, J., Jalink, K., Boulton, S. J., Chen, J., van Gent, D. C., Bartek, J., Jonkers, J., Borst, P., & Rottenberg, S. (2015). REV7 counteracts DNA double-strand break resection and affects PARP inhibition. *Nature*, 521(7553), 541-544.
- Xu, J., Keenan, T. E., Overmoyer, B., Tung, N. M., Gelman, R. S., Habin, K., Garber, J. E., Ellisen, L. W., Winer, E. P., Goss, P. E., Yeap, B. Y., Chabner, B. A., & Isakoff, S. J. (2021a). Phase II trial of veliparib and temozolomide in metastatic breast cancer patients with and without BRCA1/2 mutations. *Breast Cancer Res Treat*, 189(3), 641-651.
- Xu, J., Wang, J., Ye, J., Jiao, J., Liu, Z., Zhao, C., Li, B., & Fu, Y. (2021b). Metal-Coordinated Supramolecular Self-Assemblies for Cancer Theranostics. *Advanced Science*, 8(16), 2101101.

- Yabroff, K. R., Wu, X. C., Negoita, S., Stevens, J., Coyle, L., Zhao, J., Mumphy, B. J., Jemal, A., & Ward, K. C. (2022). Association of the COVID-19 Pandemic With Patterns of Statewide Cancer Services. *J Natl Cancer Inst*, 114(6), 907-909.
- Yao, H., He, G., Yan, S., Chen, C., Song, L., Rosol, T. J., & Deng, X. (2017). Triple-negative breast cancer: is there a treatment on the horizon? *Oncotarget*, 8(1), 1913-1924.
- Yap, T. A., Ngoi, N., Dumbrava, E. E., Karp, D. D., Rodon Ahnert, J., Fu, S., Hong, D. S., Naing, A., Pant, S., Piha-Paul, S. A., Subbiah, V., Tsimberidou, A. M., Dufner, D., Rhudy, J., Gore, S., Ivy, S. P., Yuan, Y., Westin, S. N., Mills, G. B., & Meric-Bernstam, F. (2022). NC10329: Phase Ib Sequential Trial of Agents against DNA Repair (STAR) Study to investigate the sequential combination of the Poly (ADP-Ribose) Polymerase inhibitor (PARPi) olaparib (ola) and WEE1 inhibitor (WEE1i) adavosertib (ada) in patients (pts) with DNA Damage Response (DDR)-aberrant advanced tumors, enriched for BRCA1/2 mutated and CCNE1 amplified cancers. *European Journal of Cancer*, 174, S7.
- Yazinski, S. A., Comaills, V., Buisson, R., Genois, M. M., Nguyen, H. D., Ho, C. K., Todorova Kwan, T., Morris, R., Lauffer, S., Nussenzweig, A., Ramaswamy, S., Benes, C. H., Haber, D. A., Maheswaran, S., Birrer, M. J., & Zou, L. (2017). ATR inhibition disrupts rewired homologous recombination and fork protection pathways in PARP inhibitor-resistant BRCA-deficient cancer cells. *Genes Dev*, 31(3), 318-332.
- Yeo, C. I., Ooi, K. K., & Tiekink, E. R. T. (2018). Gold-Based Medicine: A Paradigm Shift in Anti-Cancer Therapy? *Molecules*, 23(6).
- Yi, J., Liu, C., Tao, Z., Wang, M., Jia, Y., Sang, X., Shen, L., Xue, Y., Jiang, K., Luo, F., Liu, P., & Cheng, H. (2019). MYC status as a determinant of synergistic response to Olaparib and Palbociclib in ovarian cancer. *EBioMedicine*, 43, 225-237.
- Yin, L., Duan, J.-J., Bian, X.-W., & Yu, S.-c. (2020). Triple-negative breast cancer molecular subtyping and treatment progress. *Breast Cancer Research*, 22(1), 61.
- Yoshioka, K. I., Kusumoto-Matsuo, R., Matsuno, Y., & Ishiai, M. (2021). Genomic Instability and Cancer Risk Associated with Erroneous DNA Repair. *Int J Mol Sci*, 22(22).
- Young, P. A., Márquez-Garbán, D. C., Noor, Z. S., Moatamed, N., Elashoff, D., Grogan, T., Romero, T., Sasano, H., Saito, R., Rausch, R., Hamilton, N., Dubinett, S. M., Garon, E. B., & Pietras, R. J. (2021). Investigation of Combination Treatment With an Aromatase Inhibitor Exemestane and Carboplatin-Based Therapy for Postmenopausal Women With Advanced NSCLC. *JTO Clin Res Rep*, 2(4), 100150.

- Yu, J., Lu, R., Nedrow, J. R., & Sgouros, G. (2020). Response of breast cancer carcinoma spheroids to combination therapy with radiation and DNA-PK inhibitor: growth arrest without a change in α/β ratio. *International Journal of Radiation Biology*, 96(12), 1534-1540.
- Yu, X., & Sun, D. (2013). Macrocyclic Drugs and Synthetic Methodologies toward Macrocycles. *Molecules*, 18(6), 6230-6268.
- Yudin, A. K. (2015). Macrocycles: lessons from the distant past, recent developments, and future directions. *Chemical Science*, 6(1), 30-49.
- Yue, S., Luo, M., Liu, H., & Wei, S. (2020). Recent Advances of Gold Compounds in Anticancer Immunity. *Front Chem*, 8, 543.
- Yuhas, J. M., Li, A. P., Martinez, A. O., & Ladman, A. J. (1977). A simplified method for production and growth of multicellular tumor spheroids. *Cancer Res*, 37(10), 3639-3643.
- Yusoh, N. A. (2020). *Effect of combination therapy of ruthenium polypyridyl complex, [Ru(dppz)2(PIP)]2+ and PARP inhibitors against several cancer cell lines*. (Master's thesis), Universiti Putra Malaysia.
- Yusoh, N. A., Ahmad, H., & Gill, M. R. (2020a). Combining PARP Inhibition with Platinum, Ruthenium or Gold Complexes for Cancer Therapy. *ChemMedChem*, 15(22), 2121-2135.
- Yusoh, N. A., Leong, S. W., Chia, S. L., Harun, S. N., Rahman, M. B. A., Vallis, K. A., Gill, M. R., & Ahmad, H. (2020b). Metallointercalator [Ru(dppz)2(PIP)]2+ Renders BRCA Wild-Type Triple-Negative Breast Cancer Cells Hypersensitive to PARP Inhibition. *ACS Chemical Biology*, 15(2), 378-387.
- Zagami, P., & Carey, L. A. (2022). Triple negative breast cancer: Pitfalls and progress. *npj Breast Cancer*, 8(1), 95.
- Zanoni, M., Piccinini, F., Arienti, C., Zamagni, A., Santi, S., Polico, R., Bevilacqua, A., & Tesei, A. (2016). 3D tumor spheroid models for in vitro therapeutic screening: a systematic approach to enhance the biological relevance of data obtained. *Sci Rep*, 6, 19103.
- Zappa, C., & Mousa, S. A. (2016). Non-small cell lung cancer: current treatment and future advances. *Transl Lung Cancer Res*, 5(3), 288-300.
- Zeman, M. K., & Cimprich, K. A. (2014). Causes and consequences of replication stress. *Nat Cell Biol*, 16(1), 2-9.
- Zeng, L., Gupta, P., Chen, Y., Wang, E., Ji, L., Chao, H., & Chen, Z.-S. (2017). The development of anticancer ruthenium(II) complexes: from

single molecule compounds to nanomaterials. *Chemical Society Reviews*, 46(19), 5771-5804.

- Zhang, Z., Zhang, Y., Liu, C., Shao, J., Chen, Y., Zhu, Y., Zhang, L., Qin, B., Kong, Z., Wang, X., Wang, Y., Huang, D., Liu, L., Zhou, Y., Tao, R., Yang, Z., Liu, M., & Zhao, W. (2023). A real-world study of immune checkpoint inhibitors in advanced triple-negative breast cancer. *Cancer Innovation*, 2(3), 172-180.
- Zhao, H., Yang, Q., Hu, Y., & Zhang, J. (2018). Antitumor effects and mechanisms of olaparib in combination with carboplatin and BKM120 on human triple-negative breast cancer cells. *Oncol Rep*, 40(6), 3223-3234.
- Zhong, L., Yin, R., & Song, L. (2023). Resistance to PARP Inhibitors After First-Line Platinum-Based Chemotherapy in a Patient with Advanced Ovarian Cancer with a Pathogenic Somatic BRCA1 Mutation. *Pharmgenomics Pers Med*, 16, 195-200.
- Zhou, Z., Song, J., Nie, L., & Chen, X. (2016). Reactive oxygen species generating systems meeting challenges of photodynamic cancer therapy. *Chemical Society Reviews*, 45(23), 6597-6626.
- Zhu, L., Liu, J., Chen, J., & Zhou, Q. (2021a). The developing landscape of combinatorial therapies of immune checkpoint blockade with DNA damage repair inhibitors for the treatment of breast and ovarian cancers. *Journal of Hematology & Oncology*, 14(1), 206.
- Zhu, X., Chen, L., Huang, B., Li, X., Yang, L., Hu, X., Jiang, Y., Shao, Z., & Wang, Z. (2021b). Efficacy and mechanism of the combination of PARP and CDK4/6 inhibitors in the treatment of triple-negative breast cancer. *Journal of Experimental & Clinical Cancer Research*, 40(1), 122.
- Živković, M. D., Kljun, J., Ilic-Tomic, T., Pavic, A., Veselinović, A., Manojlović, D. D., Nikodinovic-Runic, J., & Turel, I. (2018). A new class of platinum(ii) complexes with the phosphine ligand pta which show potent anticancer activity. *Inorganic Chemistry Frontiers*, 5(1), 39-53.
- Zou, L., & Elledge, S. J. (2003). Sensing DNA damage through ATRIP recognition of RPA-ssDNA complexes. *Science*, 300(5625), 1542-1548.