

UNIVERSITI PUTRA MALAYSIA

PHENOTYPIC AND MOLECULAR CHARACTERIZATION OF PASTEURELLA MULTOCIDA OBTAINED FROM POULTRY IN IRAN

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Ву

AHMAD REZA JABBARI,

Thesis Submitted in Fulfilment of the Requirement for the Degree of Doctor of Philosophy in the Faculty of Veterinary Medicine Universiti Putra Malaysia

September 2001



DEDICATED TO MY WIFE, MAHTAB MOZAFFARI, OUR TWO SONS, MOHAMMAD VAHID AND MOHAMMAD MOEIN



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Abstract of thesis submitted to the Senate of Universiti Putra Malaysia in

fulfilment of the requirement for the degree of Doctor of Philosophy

PHENOTYPIC AND MOLECULAR **CHARACTERIZATION**

PASTEURELLA MULTOCIDA OBTAINED FROM POULTRY IN IRAN

By

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September 2001

Chairman: Associate Professor Dr. Abdul Aziz Saharee

Faculty: Veterinary Medicine

A collection of twenty five Pasteurella multocida isolates obtained from avian

pasteurellosis in northern part (endemic area) of Iran were studied for some

of their phenotypic and molecular characteristics. This research is the first

study on conducting of serotyping and molecular characterization of avian

P.multocida in Iran.

Based on the biochemical characteristics, all P.multocida isolates tested

belonged to subspecies (biotype) multocida. Antimicrobial sensitivity test

showed that all the isolates examined were resistant to at least three of the

thirteen antimicrobials tested. Among the antimicrobial

chloramphenicol, combination of sulfametoxazin and trimetoprim and

nitrofurantoin were found to be the most effective (100% sensitivity)

followed by tetracycline (96% sensitivity), penicillin (88% sensitivity) and gentamycin (76% sensitivity). The highest percentage of resistance was found against lincomycin, bacitracin and cloxacillin (100% resistant) followed by furazolidone and colistin (84% and 68% respectively).

Agar gel diffusion precipitation (AGDP) test was used to determine somatic serotypes of the isolates. According to the results of the AGDP test, Serotype 1 was dominant among avian isolates from endemic area. Serotypes 3, 3×4 and 4, found for the first time in the country were also identified among the isolates.

Electrophoresis protein patterns of the isolates were studied by using sodium dodecyl sulphate polyacylamide gel (SDS-PAGE). All strains were similar in the majority of protein bands. The main difference between protein patterns of the isolates was revealed in the position of one of the major band (H Protein) presented in the 34-38 KDa region. According to H protein position, three distinguishable groups were identified. In protein type I, the molecular mass of H protein was about 38 KDa but in protein types II and III it was 36.5 and 34 KDa respectively.

The minimum lethal dose (MLD) of the strains with protein types I, II and III as a virulence determinant was identified in mice. It was revealed that the strain with protein type III had the least virulence and the strain with protein



type I had the greatest virulence in mice. Immunization of mice with strain PMI030 (protein type I) induced a good protection against homologous protein type challenge.

Restriction enzyme analysis (REA) of chromosomal DNA and repetitive extragenic palindromic elements PCR (REP-PCR) were used for determination of genetic diversity among the isolates. DNA fingerprinting by HpaII digestion divided the twenty-five isolates into 7 REA groups, 2 of which contained a single isolate.

DNA fingerprinting with REP-PCR revealed a great genetic diversity among the isolates. According to amplified DNA patterns, a total of 9 REP-PCR groups were determined. REP-PCR produced amplified bands ranging in size from approximately 700 bp to 3.6 Kb with two species-specific bands of 0.8 Kb and 2.3 Kb. REP-PCR was able to differentiate *P.multocida* isolates from different source and geographical areas. Results of this study showed that the use of REP element amplification by polymerase chain reaction is highly reproducible and can be suggested as a suitable epidemiologic tool especially for investigation on the origin of outbreaks and similarity between different avian isolates of *P.multocida*.



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Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai

memenuhi keperluan untuk ijazah Doktor Falsafah

CIRI-CIRI FENOTIPIK DAN MOLEKULAR PASTEURELA MULTOCIDA

YANG DIPEROLEHI DARIPADA TERNAKAN AYAM ITIK DI IRAN

Oleh

AHMAD REZA JABBARI

September 2001

Pengerusi: Profesor Madya Dr. Abdul Aziz Saharee

Fakulti Perubatan Veterinar

Satu kajian tentang sekumpulan 25 Pasteurella multocida yang diperolehi

daripada pasteurelosis avian di bahagian utara (kawasan endemi) Iran telah

dijalankan untuk meneliti ciri-ciri fenotipik dan molekularnya. Kajian ini

merupakan yang pertama mengkaji tentang kelakuan serotip dan ciri-ciri

molekular P.multocida avian di Iran.

Berasaskan ciri-ciri biokimia, kesemua pemencilan *P.multocida* yang diuji

tergolong sebagai subspesies (biotaip) multocida. Ujian sensitiviti antimikrob

menunjukkan bahawa kesemua pemencilan yang diteliti menolak sekurang-

kurangnya 3 daripada 13 antimikrob yang diuji. Di kalangan agen-agen

antimikrob, kloramfenikol, kombinasi sulfametoksazin dan trimetoprim serta

nitrofurantin didapati paling efektif (sensitiviti 100%) diikuti oleh tetrasiklin

(sensitiviti 98%), penisilin (sensitiviti 88%) dan gentamisin (sensitiviti 76%). Peratus tertinggi resistan didapati lebih kepada linkomisin, basitrasin dan kloksasilin (resistan 100%), diikuti oleh furazolidon dan kolistin (masing-masing 84% dan 68%).

Ujian Agar Gel Diffusion Precipitation (AGDP) telah digunakan untuk menentukan pemencilan somatik serotip. Berdasarkan keputusan ujian AGDP, Serotip 1 didapati dominan berbanding avian yang dipencilkan daripada kawasan endemik. Serotip 3, 3 x 4 dan 4 yang pertama kali ditemui di negara Iran juga dikenal pasti di kalangan pemencilan-pemencilan lain.

Corak protein elektroforisis bagi pemencilan ini telah dikaji dengan menggunakan gel sodium dodisil sulfat poliklamid (SDS-PAGE). Kesemua strain adalah sama dalam kebanyakan jalur-jalur protein. Perbezaan utama antara corak-corak protein pemencilan ini begitu ketara pada kedudukan salah satu daripada jalur utama (H Protein) yang dibentangkan dalam kawasan 34-38 Kda. Merujuk kepada kedudukan protein H, tiga kumpulan yang berbeza telah dikenal pasti. Pada protein jenis I, jisim molekular protein H adalah lebih kurang 38 Kda tetapi di dalam protein jenis II dan jenis III, masing-masing 36.5 dan 34 Kda.



Dos lethal minimum (MLD) strain berprotein jenis I, II dan III sebagai penentu virulen telah dikenal pasti pada tikus. Ia menunjukkan bahawa strain berprotein jenis III mempunyai virulen paling sedikit dan strain berprotein jenis I mempunyai virulen paling banyak terdapat pada tikus.

Pembatasan analisis enzim (REA) kromosomal DNA dan REP-PCR digunakan untuk menentukan kepelbagaian genetik di kalangan pemencilan. Cap jari DNA oleh pencernaan HpaII membahagikan 25 pemencilan ini kepada 7 kumpulan REA, yang mana dua daripadanya mengandungi pemencilan tunggal.

Cap jari DNA beserta REP-PCR menunjukkan kepelbagaian genetik yang amat ketara di kalangan pemencilan. Berdasarkan corak DNA yang luas, sejumlah 9 REP-PCR telah ditentukan. REP-PCR menghasilkan jalur-jalur yang kuat, selari mengikut saiz daripada kira-kira 700 bp hingga 3.6 Kb beserta dua jalur spesifik-spesies 0.8 Kb dan 2.3 Kb. REP-PCR berupaya untuk membezakan pemencilan *P.multocida* di kalangan sumber-sumber dan kawasan-kawasan geografi yang berbeza. Kesimpulan daripada kajian ini menunjukkan bahawa penggunaan amplifikasi elemen-elemen REP oleh tindak balas berantai polimeras sangat produktif dan boleh dicadangkan sebagai alat epidemiologi yang sesuai untuk penyelidikan terhadap kerebakan asal dan keserupaan antara pemencilan avian *P.multocida* yang berlainan.



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LIST OF ABBREVIATIONS

AGDP Agar Gel Diffusion Precipitin Test

AP-PCR
APS
Arbitrary Primed PCR
APS
Ammonium Persulfate
BHI
Brain Heart Infusion
°C
Degree Celcius

CCA Crude Capsule Antigens **CCF** Cell-Free Culture Filtrate **CFU** Colony Forming Unit CP Capsular Polysaccharide **CPFs Cross-Protection Factors** CU strain Clemson University Strain **DATP** Deoxyadenosine Triphosphate **DCTP** Deoxycytidine Triphosphate **DGTP** Deoxyguanosine Triphosphate **DTTP** Deoxythymidine Triphosphate **DNTP** Deoxynucleutide Triphosphate DIE Descriptive Identification Epithet **EDTA** Ethylendiamine Tetra Acetate

ELISA Enzyme Linked Immunosorbent Assay

FC Fowl Cholera

HS Haemorrhagic Septicaemia

HSA Heat Stable Antigen HSPs Heat Shock Proteins

IHA Indirect Haemaglutination Test IVO Iranian Veterinary Organization

KDa Kilo Daltons

KSCN Potassium Thiocyanate Extracts
LD 100 Lethal Dose of 100 Percent

LPS Lipopolysaccharide MAb Monoclonal Antibody

Mda Mega Daltons

MHA Muller-Hinton Agar MLD Minimum Lethal Dose

MOMPs Major Outer Membrane Proteins
OMPs Outer Memberane Proteins

P.multocida Pasteurella multocida

PAGE Polyacrylamide Gel Electrophoresis

PCR Polymerase Chain Reaction
PMSF Phenylmethyl Sulfonyl Fluoride
PMT Pasteurella multocida Toxin



RAPD Random Amplified Polymorphic DNA

RD Complex
REA
Respiratory Disease Complex
Restriction Endonuclease Analysis
REP
Repetitive Extragenic Palindrome

rRNA Ribosomal RNA

RVSRI Razi Vaccine and Serum Research Institute

TaqTermophilus aquaticusSDSSodium Dodecyl sulphateSPFSpecific Pathogen Free

Subspecies Subspecies

TAE Tris Acetate EDTA
TEMED Tetrametylendiamine

Tox Toxin

TSI Three Sugar Iron Agar

U Uni

VRI Veterinary Research Institute

WC Whole Cell

