

### **UNIVERSITI PUTRA MALAYSIA**

THE EFFECT OF IMMUNE ENHANCER ON THE NON-SPECIFIC DEFENSE MECHANISM OF RED TILAPIA HYBRID (OREOCHROMIS NILOTICUS X OREOCHROMIS MOSSAMBICUS) CHALLENGED WITH AEROMONAS HYDROPHILLA

**JOSELITO R. SOMGA** 

FPSS 1995 5



# THE EFFECT OF IMMUNE ENHANCER ON THE NON-SPECIFIC DEFENSE MECHANISM OF RED TILAPIA HYBRID (OREOCHROMIS NILOTICUS X OREOCHROMIS MOSSAMBICUS) CHALLENGED WITH AEROMONAS HYDROPHILA

BY

JOSELITO R. SOMGA

Thesis Submitted in Partial Fulfillment of the Requirement for the Degree of Master of Science in the Faculty of Fisheries and Marine Science Universiti Pertanian Malaysia

December 1995



#### **ACKNOWLEDGEMENTS**

I am grateful to my chairman Prof. Mohammed Shariff Din, for his valuable advice, support, guidance and motivation. My gratitude is extended to my committee members, Dr. Hassan Hj. Mohd Daud and Dr. Abdul Manan Mat Jais for their kind assistance and wise counsel throughout this study.

My appreciation goes to the International Development Research Center (IDRC) Canada for providing the fellowship and research fund for my study.

I express my appreciation and thanks to Dr. Lim of GHF PTE Technologies Ltd. for providing the immune enhancer used in this study. Special thanks is due to P.A.H.L. Jayawardena and Wang Yin Geng for their generous help during the collection of samples. Thanks are also due to the Faculty of Fisheries and Marine Science staff particularly Abdul Gani, Zainan, Rosdi, Zairina Raden Zainal, Mahamud Yusoh and those who have helped me in many ways to accomplish this study.



I am also thankful to Mr. Paul Manalo and family for their kindness and concern during my stay in Malaysia. I also cherish the companionship of John, Ging, Bhebot, Aiza, Ate Da and Ate Dina. Also, I appreciate the moral support extended by my colleague in BFAR. The camaraderie with fellow students at UPM is memorable.

This study is dedicated to my wife Sonia, who patiently assisted me throughout my study, and my family, for their love and support. Above all, to ALMIGHTY GOD for all HIS blessings.



#### TABLE OF CONTENTS

																						P	age
ACKNO	WLE	EDGEM	ENT	3	•	•		•		•	•		•		•	•		•	•		•		ii
LIST	OF	TABL	ES	•		•				•	٠	•	•	•		•	•		•	•	•		vii
LIST	OF	FIGU	RES	•		•				•			•		•	•			•				хi
LIST	OF	PLAT	ES				•						•	•		•		•	•	•	•		xii
LIST	OF	FISH	SP	ECI	ES		•									•				•			xiv
LIST	OF	ABBR	EVI	ATI	ON	S			•			•	•	•	•	•	•		•	•			xvi
ABSTR	RACI	r.				•	•		•					•				•		•			xvii
ABSTF	RAK	•		•	•							•				•			•	•	•		xix
СНАРТ	rer																						
	I	GI	ENER	AL	IN	ТF	ROI	OUC	CT:	IO	N												1
	II	L	TER	ΙΤΑ	JRE	F	SE/	VI)	EW							•							7
		Tì	ne D	efe	ens	e	Мє	ecl	hai	ni	sm						•						7
		No	on-S	pe	cif	ii	: I	De:	fe	ns	е	in	F	is	h								8
				Pro	ote	ct	i	ve	В	ar	ri	er	s										8
				Hui	mor	a]	l I	Fa	ct	or	s							•					9
				Ce	11u	ıla	ar	F	ac	to	rs	•	•		•								14
		A	sses	sm	ent	. (	of	t!	he	N	on	<b>-</b> S	pe	ci	fi	С	Re	sp	on	se			15
			mmun efen							_		Ef											19



HYBR	NON-SPECIFIC RESPONSE OF RED TILAPIA RID FED WITH ENCAP AND CHALLENGED WITH OMONAS HYDROPHILA
Intr	oduction
Mate	erials and Methods
	Maintenance of Stock Fish
	Experimental Fish
	Culture and Maintenance of Aeromonas hydrophila
	Preparation of the Standard Curve of Aeromonas hydrophila
	Determination of Median Lethal Dose (LD <sub>50</sub> ) of <i>A. hydrophila</i> in Red Tilapia Hybrid
	Preparation of Formalin Killed Aeromonas hydrophila
	Incorporation of Immune Enhancer to the Feed
	Experiment Proper
	ermination of the Non-Specific ponse
	Hematocrit
	Total White Blood Cell (WBC) Counts .
	Nitroblue Tetrazolium (NBT) Assay - Potential Killing Activity of Neutrophils, Monocytes and Macrophages
	Lysozyme Assay
	Total Plasma Protein
Sta	tistical Analysis
	4



	DIBCUBBION	43
IV	SURVIVABILITY OF RED TILAPIA HYBRID FED WITH ENCAP AND CHALLENGED WITH AEROMONAS HYDROPHILA	50
		30
	Introduction	50
	Materials and Methods	51
	Statistical Analysis	52
	Results	52
	Discussion	54
v	HISTOPATHOLOGY OF RED TILAPIA HYBRID	
	FED WITH ENCAP AND CHALLENGED WITH AEROMONAS HYDROPHILA	58
	Introduction	58
	Materials and Methods	60
	Results	60
	Discussion	70
VI	GENERAL DISCUSSION, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE STUDY	74
REFERENCE	s	79
APPENDIX		92
BIOGDADHI	CAI SKETCH	105



# LIST OF TABLES

Table		Pag	ge
1	Hematocrit Values, White Blood Cell Counts and NBT/Potential Killing Activity in Tilapia Hybrid Fed with Different Concentrations of ENCAP and Challenged with A. hydrophila		92
2	Lysozyme Activity and Total Plasma Protein in Tilapia Hybrid Fed with Different Concentrations of ENCAP and Challenged with A. hydrophila		93
3	Summary of Water Quality Parameters Taken in the Duration of the Experiment	•	94
4	LD <sub>50</sub> of <i>Aeromonas hydrophila</i> to Red Tilapia Hybrid		94
5	Median Lethal Concentration (LD <sub>50</sub> ) of A. hydrophila in Red Tilapia Hybrid Analyzed Using Spearman-Karber		95
6	ANOVA for Hematocrit Values Between Treatment Groups in Red Tilapia Hybrid Before Injection of A. hydrophila	<b>10</b>	96
7	ANOVA for Hematocrit Values Between Treatment Groups in Red Tilapia Hybrid at Day One Post Injection of A. hydrophila		96
8	ANOVA for Hematocrit Values Between Treatment Groups in Red Tilapia Hybrid at Day Two Post Injection of A. hydrophila	•	96
9	ANOVA for Hematocrit Values Between Treatments in Red Tilapia Hybrid at Day		97



10	Treatment Groups in Red Tilapia Hybrid at Day Seven Post Injection of A. hydrophila	7
11	ANOVA for WBC Counts Between Treatment Groups of Red Tilapia Hybrid Before Injection of A. hydrophila 9	7
12	ANOVA for WBC Counts Between Treatment Groups in Red Tilapia Hybrid At Day One Post Injection of A. hydrophila 9	8
13	ANOVA for WBC Counts Between Treatment Groups in Red Tilapia Hybrid at Day Two Post Injection of A. hydrophila 9	8
14	ANOVA for WBC Counts Between Treatment Groups in Red Tilapia Hybrid at Day Four Post Injection of A. hydrophila 9	8
15	ANOVA for WBC Counts Between Treatment Groups in Red Tilapia Hybrid at Day Seven Post injection of A. hydrophila	9
16	ANOVA for NBT/Potential Killing Activity Between Treatment Groups in Red Tilapia Hybrid Before Injection of A. hydrophila	9
17	ANOVA for NBT/Potential Killing Activity Between Treatment Groups in Red Tilapia Hybrid at Day One Post Injection of A. hydrophila	9
18	ANOVA for NBT/Potential Killing Activity Between Treatment Groups in Red Tilapia Hybrid at Day Two Post Injection of A. hydrophila	00
19	ANOVA for NBT/Potential Killing Activity Between Treatment Groups in Red Tilapia Hybrid at Day Four Post Injection of A. hydrophila	00
20	ANOVA for NBT/Potential Killing Activity Between Treatment Groups in Red Tilapia Hybrid at Day Seven Post Injection of A. hydrophila	00



21	ANOVA for Lysozyme Activity Between Treatment Groups in Red Tilapia Hybrid Before Injection of A. hydrophila 101
22	ANOVA for Lysozyme Activity Between Treatment Groups in Red Tilapia Hybrid at Day One Post Injection of A. hydrophila
23	ANOVA for Lysozyme Activity Between Treatment Groups in Red Tilapia Hybrid at Day Two Post Injection of A. hydrophila
24	ANOVA for Lysozyme Activity Between Treatment Groups in Red Tilapia Hybrid at Day Four Post Injection of A. hydrophila
25	ANOVA for Lysozyme Activity Between Treatment Groups in Red Tilapia Hybrid at Day Seven Post Injection of A. hydrophila
26	ANOVA for Total Plasma Protein Between Treatment Groups in Red Tilapia Hybrid Before Injection of A. hydrophila 102
27	ANOVA for Total Plasma Protein Between Treatment Groups in Red Tilapia Hybrid at Day One Post Injection of A. hydrophila
28	ANOVA for Total Plasma Protein Between Treatment Groups in Red Tilapia Hybrid at Day Two Post Injection of A. hydrophila
29	ANOVA for Total Plasma Protein Between Treatment Groups in Red Tilapia Hybrid at Day Four Post Injection of A. hydrophila
30	ANOVA for Total Plasma Protein Between Treatment Groups in Red Tilapia Hybrid at Day Seven Post Injection of A.



31	for 21 Days Post Bacterial Challenged										
	Between Treatment Groups of Red Tilapia Hybrid	104									
32	ANOVA for The Survivability Between Treatment Groups Post Bacterial Challenged	104									



## LIST OF FIGURES

Figure		Pā	age
1	Hematocrit Value in Red Tilapia Hybrid Fed with ENCAP and Challenged with A. hydrophila		36
2	Total WBC Count in Red Tilapia Hybrid Fed with ENCAP and Challenged with A. hydrophila		37
3	NBT/ Potential Killing Activities of Neutrophils and Other Phagocytic Cells in Red Tilapia Hybrid Fed with ENCAP and Challenged with A. hydrophila	•	39
4	Lysozyme Activity in Red Tilapia Hybrid Fed with ENCAP and Challenged with A. hydrophila	•	41
5	Total Plasma Protein in Red Tilapia Hybrid Fed with ENCAP and Challenged with A. hydrophila	•	42
6	Percent Survivability of Red Tilapia Hybrid Fed with ENCAP and Challenged with A. hydrophila		53



## LIST OF PLATES

Plate			Pa	ge
1	Red Tilapia Hybrid Before Challenge of A. hydrophila		•	61
2	Red Tilapia Hybrid with Exposed Epidermal Surface and Necrotic Fins After 2-3 Days Post Bacterial Challenged (PBC)		•	61
3	Tissue Section Showing the Sloughed Epidermal Layer After 7 Days PBC		•	62
4	Congestion of RBC's in Blood Vessels and Surrounding Muscle Tissue After 7 Days PBC		•	62
5	Tissue Section of the Liver Showing Vacoulation After 2 Days PBC			64
6	Bacteria in the Liver Section Undergoing Necrosis After 7 Days PBC		•	64
7	Liver Section Showing Severe Necrosis After 7 Days PBC. The Structural Integrity was Lost	•		65
8	Intrahepatic Pancreatic Tissue Undergoing Disintegration of the Acinar Cells and Marked Proliferation of Mononuclear Cells After 2 Days PBC	•	•	65
9	Congestion of the Portal Vessel After 2 Days PBC. Pancreatic Tissue Undergoing Necrosis			66
10	Tissue Section of the Spleen Showing Vacuolated Areas, Enlarged Melanomacrophages Centers (MMC) and Scattered Melanin Pigments After 2			6.6



11	Tissue Section of the Spleen Showing Marked Reduction of RBC's and Vacuolated Areas After 7 Days PBC	67
12	Tissue Section of the Kidney Showing Massive Infiltration of RBC's and other Inflammatory Cells in the Intertubular Spaces After 7 Days PBC. Melanomacrophage Centers (MMC)	68
13	Kidney Section Showing Extensive Tubular and Glomerular Necrosis After 7 Days PBC	69
14	Higher Magnification Showing Mononuclear and Polymorphonuclear Cells in the Intertubular Spaces	69



#### LIST OF FISH SPECIES

African catfish Clarias gariepinus
American eel
Atlantic salmon Salmo salar
Ayu Plecoglossus altevelis
Brook trout Salvelinus fontinalis
Channel catfish Ictalurus punctatus
Chinook salmon Onchorynchus tshawystscha
Coho salmon Onchorynchus kisutch
Common carp
Dabs Limanda limanda
Eel
European eel Anguilla anguilla
Fathead minnow Pimephales promelas
Flatfish Paralicthys olivaceus
Goldfish Carassius auratus
Largemouth bass Micropterus salmoides
Plaice Pleuronectes platessa
Porgy Pagrus major
Rainbow trout Onchorynchus mykiss
Rainbow trout Salmo gairdneri
Red seabream Pagrus major
Sea bass Dicentrarchus labrax
Steelhead trout Salmo gairdneri



Tilapia	•	•	•	٠	•	٠	•	•	•	•	•	Tilapia ni	ilotica
Turbot	•			•		٠	•			•		Scophthalmus	maximus
Walleye		•					٠			•		Stizostedion vitreum	vitreum
Winter	flo	our	nde	er		•						Psedopleuronectes ame	ricanus
Yellowta	ai.	l I	_									Seriola quinque	radiata



#### LIST OF ABBREVIATIONS

CFU Colony Forming Unit

CP Crude Protein

DMSO Dimethyl sulfoxide

EDTA Ethylene Diamine Tetraacetic Acid

EUS Epizootic Ulcerative Syndrome

LD<sub>50</sub> Median Lethal Dose

MMC Melanomacrophages Center

MS-222 Tricane Methanesulfonate

NBT Nitroblue Tetrazolium

PBC Post Bacterial Challenge

PBS Physiological Buffered Saline

RBC Red Blood Cells

RS Reimlers Schott

SD Standard Deviation

TSA Trypticase Soy Agar

WBC White Blood Cells



Abstract of the thesis submitted to the Senate of the Universiti Pertanian Malaysia in partial fulfillment of the requirement for the degree of Master of Science

# THE EFFECT OF IMMUNE ENHANCER ON THE NON-SPECIFIC DEFENSE MECHANISM OF RED TILAPIA HYBRID (OREOCHROMIS NILOTICUS X OREOCHROMIS MOSSAMBICUS) CHALLENGED WITH AEROMONAS HYDROPHILA

by

Joselito R. Somga

December 1995

Chairman: Prof. Mohd. Shariff Mohd. Din, Ph.D

Faculty: Fisheries and Marine Science

Immunomodulation of ENCAP in red tilapia hybrid against Aeromonas hydrophila was studied. Different concentrations of ENCAP (0, 500, 750 and 1000 mg/kg of feed) were fed to different groups of fish and later challenged by intraperitoneal injection of 8 x 10<sup>8</sup> CFU/ml A. hydrophila. The non-specific immune response was determined after one, two, four and seven days post bacterial challenge using haematological and serological assays such as haematocrit, WBC counts, potential killing activity of neutrophils and other phagocytic cells by NBT, lysozyme activity and total plasma protein. Different concentrations of ENCAP showed different levels of immunopotentiation.

Hematocrit levels and WBC counts decreased in all the groups due to migration of erythrocytes and leukocytes to the infected areas. Probably, toxins released by the bacteria also contributed to these lowered levels. Neutrophils and other phagocytic cells demonstrated an increase in the potential killing activity. Lysozyme activity also increased in fish fed with ENCAP, while total plasma protein decreased brought about by the abnormal function of the liver to synthesize protein. Based on these cellular and humoral factors, fish fed with 750 mg/kg ENCAP had a consistently higher immune response. Fish fed with 500 mg/kg and 1000 mg/kg showed a lower immune response which suggests slight immunopotentiation mild immunosuppression, respectively.

Histopathology showed that both the control and fish fed with different concentrations of ENCAP exhibited varying lesions in the spleen, liver, pancreatic tissue and kidney. However fish fed with ENCAP showed a significantly higher survivability. Results of this study indicated that ENCAP caused immunomodulation. The enhanced non-specific response contributed to the increased survivability.



Abstrak tesis dikemukakan kepada Senat Universiti Pertanian Malaysia sebagai memenuhi sebahagian syarat untuk mendapat Ijazah Master Sains

# KESAN PERANGSANG IMMUN KE ATAS MEKANISME PERTAHANAN TIDAK SPECIFIK DALAM HIBRID TILAPIA MERAH (OREOCHROMIS NILOTICUS X OREOCHROMIS MOSSAMBICUS) YANG DICABAR DENGAN AEROMONAS HYDROPHILA

oleh

Joselito R. Somga

Disember 1995

Pengerusi: Prof. Mohd. Shariff Mohd. Din, Ph.D

Faculti : Perikanan dan Sains Samudera

Immunomodulasi oleh ENCAP pada hibrid tilapia merah terhadap Aeromonas hydrophila telah dikaji. Kepekatan ENCAP yang berbeza (0, 500, 750 dan 1000 mg/kg makanan) telah diberi kepada beberapa kumpulan ikan dan kemudian dicabar suntikan intraperitoneal 8 x 108 CFU/ml hydrophila. Tindakbalas tidak specifik imun telah ditentukan pada hari pertama, kedua, keempat dan ketujuh selepas suntikan bakteria dengan menggunakan hematologikal dan serologikal asei seperti hematokrit, pengiraan sel darah putih, potensi aktiviti membunuh neutrofil dan selsel fagositik lain secara NBT, aktiviti lisozim dan jumlah protein plasma. Kepekatan ENCAP yang berbeza menunjukkan tahap immunopotensasi yang berbeza.



Paras hematokrit dan jumlah WBC berkurangan dalam kumpulan disebabkan oleh migrasi eritosit leukosit ke kawasan yang dijangkiti. Mungkin, toksin yang dilepaskan oleh bakteria menyumbang kepada tahap yang rendah tersebut. Neutrofil dan sel-sel fagositik lain menunjukkan peningkatan potensi aktiviti membunuh. Aktiviti lisozim telah juga ditingkatkan pada ikan yang diberi makan ENCAP. Sementara jumlah protein plasma berkurangan akibat mengsintesiskan fungsi abnormal hati protein. Berdasarkan faktor-faktor sellular dan humoral, ikan yang diberi makan 750 mg/kg ENCAP mempunyai tindakbalas immunisasi tinggi yang berpanjangan. Ikan yang diberi makan 500 mg/kg menunjukkan tindakbalas immunisasi rendah yang mencadangkan sedikit immunopotensasi dan manakala pada 1000 mg/kg menunjukkan immunosupresi.

Histopatologi menunjukkan kedua-dua kawalan dan ikan yang diberi makan dengan kepekatan ENCAP yang berlainan mempamirkan lesi yang berbeza dalam limpa, hati, tisu pankreatik dan ginjal. Walaubagaimanapun, ikan yang diberi makan dengan ENCAP menunjukkan kemandirian yang jelas tinggi. Keputusan kajian ini menunjukkan bahawa ENCAP menyebabkan immunomodulasi. Tindakbalas tidak specifik yang diransang menyumbang kepada peningkatan kemandirian.



#### CHAPTER I

#### GENERAL INTRODUCTION

Aquaculture plays a vital role in the production of fish and other fishery products. Aquaculture has expanded around the world due to the increasing demand of protein from the growing human population and the decline of available natural aquatic resources. However, the rapid expansion and intensification of fish farming lead to the occurrence of various economically important diseases. Consideration on the intimate relationship between the fish, pathogen and environment seems to be neglected. The unwise increase of stocking density together with the deterioration of the aquatic environment can cause stress to the cultured fish. Stress lowers the resistance of the fish thereby giving chance to opportunistic pathogens to invasive. become Thus fish in this scenario, will inevitably succumb to diseases cause by either viruses, bacteria, parasites and fungi.

To overcome such problems, fish culturists became more dependent on the use of chemotherapeutic agents. But with the limitation of approved chemotherapeutic products, overused or misused of antibiotics generate the risk of



bacterial resistant pathogens and the problems of drug residues in the environment and fish products (Ellis, 1988; Ghittino et al., 1984; Anderson, 1992; Baticados and Paclibare, 1992; Nikl et al., 1993). Rijkers et al. (1981) reported that prolonged used of oxytetracycline cause depression of the humoral and cellular immunity in common carp. Some chemicals such as malachite green, a known parasiticide and pyridylmercuric acetate, an effective fungicide cause cancer and mercury accumulation in tissues respectively (Anderson et al., 1984).

The use of vaccine to stimulate the production of antibody against specific pathogen has been studied. The first experimental vaccination in fish was reported by Duff 1942 against furunculosis using killed Aeromonas salmonicida given orally. But since then only few vaccines have been proven to be effective on commercial scale. Other vaccinations that have successfully been done experimentally were not reliably reproduced even using other techniques of administration and antigen preparation (Ellis, 1988). Although vaccination is a valuable approach for disease prevention (Alderman and Michel, 1991) its usefulness is limited by their specificity, lack availability and high cost to produce commercially (Ellis, 1988; Yoshida et al., 1993).



The constraints on the use of chemotherapeutic agents and vaccines in fish farming further the development of more effective ways and means to protect the fish from various disease causing organisms. The use and application of immunostimulants for protecting the fish against diseases has been attempted. Immunostimulant elevates the non-specific defense mechanism or the specific immune response (Anderson, 1992). This may be administered alone or in combination with vaccine to activate the non-specific defense mechanism as well as heightening the specific immune response.

The non-specific defense mechanism is the first line of defense which constitutes the protective barriers such as skin and scales, humoral factors in mucus and sera such lysozymes, C-reactive protein, transferrin interferon, and the cellular factors such as phagocytic cells, neutrophils and macrophages (Fletcher, Roberts, 1989; Robertsen et al., 1990; Kaige et al., 1990; Anderson, 1992). On the otherhand, the specific defense mechanism is responsible for initiating and mediating the humoral, cell mediated immunity (CMI) and the memory. The humoral immunity refers to the production of soluble antibody, whereas the CMI refers to responses which are mediated by lymphocytes and macrophages and the memory constitutes an adaptive change in the lymphoid cells causing an enhanced magnitude with subsequent challenge by the same antigen (Roberts, 1989).



The use of immunostimulants is being intensified in the areas of cancer and AIDS (Acquired immunodeficiency syndrome) research (Fudenberg and Whitten, 1984; Azuma and Jolles, 1987; WHO, 1990 as cited by Anderson, 1992). It activates macrophages, T- and B-lymphocytes, and natural killer cells that increase the body's ability to destroy tumour cells (Raa et al., 1992). Immunostimulants were also used for activating early protection against diseases in domestic animals (Kehrli et al., 1990).

Immunostimulants can be obtained from a very diverse natural sources and a large number have been made by chemical synthesis with natural products as structural models (Raa et al., 1992). Different substances have been tested to stimulate immune response in fish. Glucans, a long-chain polysaccharides extracted from yeast given parenterally or orally were evaluated in fish for their ability to enhance protection against different bacterial pathogens (Yano et al., 1989; Robertsen et al., 1990; Raa et al., 1992; Chen and Ainsworth, 1992; Nikl et al., 1993; Jeney and Anderson, 1993).

Some drugs such as levamisole, quaternary ammonium compound (QAC) and short chain polypeptide (ISK) affect the non-specific defense mechanism activities (Jeney and Anderson, 1993). Immunoactive peptide FK 565 (Kitao and

