



**EFFECT OF THYMOQUINONE AND THYMOQUINONE-LOADED
NANOSTRUCTURED LIPID CARRIER IN *IN VITRO*
WOUND HEALING MODEL**

By

HENNA ROSHINI ALEXANDER

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Science**

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Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

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Chairman : Sharifah Sakinah Syed Alwi, PhD
Faculty : Medicine and Health Sciences

Wound healing is the body's natural response to wounding. It comprises of four highly integrated phases (haemostasis, inflammation, proliferation and remodelling) resulting in a scar. When there is an impairment to any of the healing components, the healing process gets disrupted and chronic wound ensue. Impaired wound healing is one of the major problems in diabetic patients due to persistence of inflammatory cells and increased apoptosis throughout the healing process. A constant influx of inflammatory cells releases a high level of free radicals such as reactive oxygen species that results in chronic wound. Although the exact mechanism that causes poor wound healing in diabetic patients is unclear, numerous factors have been associated with it, including wound infection, chronic inflammation, sensory neuropathy and hypoxia. Thus, new prospect for therapy to favour speed and optimal healing are emerging. In this study, thymoquinone (TQ), a bioactive compound found in *N. sativa* seed was loaded into a colloidal drug carrier known as a nanostructured lipid carrier (NLC) producing a compound known as thymoquinone-loaded nanostructured lipid carrier (TQ-NLC) (PATENT NO: PI2012001818). The rapidly progressing nanotechnology today set a new alternative carrier to enhance and favour the speed of healing process. This study aimed to determine the effect of TQ and TQ-NLC on cell proliferation and migration, the mode of cell death and the antioxidant levels in normal and diabetic cell models, 3T3 and 3T3-L1. Cytotoxicity of TQ and TQ-NLC was determined by MTT assay. Based on the MTT assay, the IC_{10} obtained for 3T3-L1 treated with TQ and TQ-NLC for 24 hours were 5.3 ± 0.6 and $4.7 \pm 3.3 \mu M$ respectively. As for 3T3 cell, the IC_{10} obtained for TQ and TQ-NLC at 24 hours were 3.9 ± 2.05 and $4.3 \pm 0.17 \mu M$. TQ-NLC was seen to increase the number of healthy cells (89-95%) and gradually decrease early apoptotic cells in time and dose dependant manner compared to TQ in 3T3-L1 cell in the Annexin V analysis. At 72 hours, 3 μM of treatment with TQ-

NLC resulted in the highest number of healthy cells. In 3T3-L1 cells treated with TQ, the apoptotic cells decreased from 16.5% to 11.5 % after 72 hours. 3T3-L1 treated with TQ-NLC showed the lowest number of apoptotic cells and a significant increase in healthy cells compared to control at 48 and 72 hours. In the proliferation and migration assay, 3T3-L1 treated with TQ-NLC showed a higher proliferation and rate of migration ($p < 0.05$) compared to TQ treated cells. In the antioxidant assay, TQ-NLC acted as antioxidant by reducing ROS levels in both the cells after injury at concentration as low as 3 μ M. Both the cells treated with TQ-NLC showed a significantly lower level of ROS ($p < 0.05$) compared to treatment with TQ. Thus, this study demonstrated that TQ-NLC performed better compared to TQ especially on the diabetic mimic cell, 3T3-L1. TQ-NLC accelerated the migration and proliferation of cells while reducing the ROS produced in the wounded cells confirming its ability as a good antidiabetic and antioxidant agent.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra
Malaysia sebagai memenuhi keperluan ijazah Master Sains

**KESAN THYMOQUINONE DAN THYMOQUINONE
BERNANOSTRUKTUR PEMBAWA LIPID DALAM
MODEL *IN VITRO* PENYEMBUHAN LUKA**

Oleh

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Ogos 2019

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Fakulti : Perubatan dan Sains Kesihatan

Penyembuhan luka adalah tindak balas semula jadi tubuh apabila luka berlaku. Ia terdiri daripada empat fasa yang sangat bersepadu (haemostasis, keradangan, proliferasi dan pembentukan semula tisu) yang menghasilkan parut. Apabila terdapat kerosakan pada komponen penyembuhan, proses penyembuhan akan terganggu dan luka kronik berlaku. Penyembuhan luka terjejas adalah salah satu masalah utama pesakit kencing manis akibat ketegangan sel-sel radang dan peningkatan apoptosis sepanjang proses penyembuhan. Kemasukan sel-sel yang menyebabkan inflamasi ini membebaskan paras radikal bebas yang tinggi seperti spesies oksigen reaktif yang mengakibatkan luka kronik. Walaupun mekanisme tepat yang menyebabkan penyembuhan luka yang lemah dalam pesakit diabetes tidak jelas, banyak faktor telah dikaitkan dengannya, termasuk jangkitan luka, keradangan kronik, neuropati deria dan hipoksia. Oleh itu, prospek baru untuk terapi bagi menggalakkan penyembuhan luka yang optimum semakin meningkat. Dalam kajian ini, thymoquinon (TQ), sebatian bioaktif yang terdapat dalam benih *N. sativa* dimasukkan ke dalam pembawa dadah koloid yang dikenali sebagai pembawa lipid nanostruktur (NLC) yang menghasilkan sebatian yang dikenali sebagai pembawa lipid nanostruktur yang dimodikan thymoquinone (TQ-NLC) (PATEN NO: PI2012001818). Nanoteknologi yang berkembang pesat hari ini menetapkan pembawa alternatif baru untuk meningkatkan dan memihak kepada proses penyembuhan yang cepat. Kajian ini bertujuan untuk menentukan kesan TQ dan TQ-NLC terhadap percambahan dan penghijrahan sel, cara kematian sel dalam model sel normal dan diabetik dan tahap antioksidan di dalam sel 3T3 dan 3T3-L1. Sitotoksiti TQ dan TQ-NLC ditentukan oleh ujian MTT. Berdasarkan ujian MTT, IC_{10} yang diperolehi untuk 3T3-L1 yang dirawat dengan TQ dan TQ-NLC selama 24 jam masing-masing adalah 5.3 ± 0.6 dan $4.7 \pm 3.3 \mu M$. Bagi sel 3T3 pula, IC_{10} yang diperolehi untuk TQ dan TQ-NLC pada 24 jam ialah 3.9 ± 2.05 dan $4.3 \pm 0.17 \mu M$. TQ-NLC dilihat meningkatkan jumlah sel yang sihat (89-95%) dan secara beransur-ansur menurunkan sel-sel apoptosis awal dalam masa dan dos berbanding

dengan TQ di dalam sel 3T3-L1 berdasarkan analisis Annexin V. Pada 72 jam, rawatan 3 μ M dengan TQNLC menghasilkan bilangan sel yang paling sihat. Dalam sel 3T3-L1 yang dirawat dengan TQ, sel apoptosis menurun dari 16.5% hingga 11.5% selepas 72 jam. 3T3-L1 yang dirawat dengan TQ-NLC menunjukkan bilangan sel apoptotik yang paling rendah dan peningkatan yang ketara dalam sel-sel sihat berbanding dengan kawalan ($p < 0.05$) pada 48 dan 72 jam. Untuk ujian luka, kepekatan TQ-NLC sebanyak 3 μ M menunjukkan kesan yang ketara pada kedua-dua 3T3 dan 3T3-L1 berbanding kawalan ($p < 0.05$). Sel 3T3-L1 yang dirawat dengan TQ-NLC menunjukkan percambahan dan kadar penghijrahan yang lebih tinggi berbanding sel-sel yang dirawat dengan TQ. Dalam ujian antioksidan, TQ-NLC bertindak sebagai antioksidan dengan mengurangkan tahap ROS dalam kedua-dua sel selepas kecederaan pada kepekatan serendah 3 μ M. Kedua-dua sel yang dirawat dengan TQ-NLC menunjukkan tahap ROS yang lebih rendah berbanding rawatan dengan TQ. Oleh itu, kajian ini menunjukkan bahawa TQ-NLC lebih berkesan berbanding dengan TQ terutama pada sel yang mimik diabetes, 3T3-L1. Rawatan dengan TQ-NLC mempercepat penghijrahan dan pembiakan sel serta mengurangkan ROS yang dihasilkan di sel-sel yang cedera mengesahkan keupayaannya sebagai ejen antidiabetik dan antioksidan yang baik.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

ANOVA	One-Way Analysis of Variance
ATCC	American Type and Culture Collection
CO ₂	Carbon dioxide
DCFH	2',7'-dichlorofluorescein
DCF-DA	2',7'-Dichlorodihydrofluorescein Diacetate
DHTQ	Dihydrothymoquinone
DM	Diabetes mellitus
DMEM	Dulbecco's Modified Eagle's Medium
DMSO	Dimethyl Sulfoxide
DNA	Deoxyribonucleic acid
ECM	Extracellular matrix
EGF	Epidermal growth factor
FBS	Fetal Bovine Serum
FGF	Fibroblast growth factor
FITC	Fluorescein isothiocyanate
G-CSF	Granulocyte colony stimulating factor
GM-CSF	Granulocyte macrophage colony stimulating factor
H ₂ O ₂	Hydrogen peroxide
IC ₁₀	Inhibitory Concentration of 10% Cell Viability
MDA	Malondialdehyde
MMPs	Matrix metalloproteinases
MTT	3-(4,5-dimethylthiazol- 2-yl)-2,5-diphenyl tetrazolium bromide
<i>N. sativa</i>	<i>Nigella sativa</i>
NLC	Nanostructured lipid carrier
O ₂	Oxygen
PBS	Phosphate Buffered Saline

PDGF	Platelet-derived growth factor
PI	Propidium Iodide
PS	Phosphatidylserine
ROS	Reactive oxygen species
SD	Standard Deviation
SEM	Standard error of mean
SLN	Solid lipid nanoparticles
STZ	Streptozotocin
TGF	Transforming growth factor
THQ	Thymohydroquinone
TQ	Thymoquinone
TQ-NLC	Thymoquinone-loaded nanostructured lipid carrier
VECs	Vascular endothelial cells
VEGF	Vascular Endothelial Growth Factor
WHO	World Health Organization

CHAPTER 1

INTRODUCTION

1.1 Background

Wounds are physical insults that result in a break or opening of the skin (Yamini & Gopal, 2016) and are a major cause of physical disability (Nagori & Solanki, 2011). Wounds are generally classified into two types: acute and chronic wound. Acute wounds usually repair themselves in an orderly manner which causes both functional and anatomical restoration (Martin, 1997). Any alterations that interrupt the timely controlled healing processes would prolong tissue damage and the repair process, consequently contributing to chronic wound (Kurahashi & Fujii, 2015) and complications usually entail. Chronic wounds are wounds that display impaired healing. They usually have failed to progress through the normal stages of healing (Guo et al., 2010) and get trapped in a permanent inflammatory stage due to an imperfect or uncoordinated healing process (Briquez et al., 2015; Velnar et al., 2009; García Orue et al., 2016). Wound healing is the body's natural reaction that leads to repair of the injured tissue (Ammar et al., 2015) and is comprised of four highly integrated and overlapping phases (haemostasis, inflammation, proliferation and remodelling of tissue) (Dunill et al., 2015; Shah et al., 2012).

Diabetes mellitus (DM) is a group of metabolic disease characterized by high glucose level in the blood. This can be a result of insulin resistance, absence of insulin or inadequate levels of insulin released due to the destruction of β -cells that are responsible for insulin production (Reece, 2010). Patients with DM often suffer from various metabolic abnormalities including delayed wound healing that usually results in leg amputations. (Cakirca et al., 2014; Asmat et al., 2016). The cause of chronic ulcers in DM patients has been attributed to a variety of abnormal biological mechanisms: ischemia, neuropathy, infection, prolonged inflammatory response, cytokine and growth factors deficits (Falanga, 2005; Medina et al., 2005; Hu & Lan, 2016).

Since ancient times, preparations from plants were used to speed up the process of wound healing (Schmidt et al., 2009; Fronza et al., 2009). Usage of medicinal plants dates from the earliest years of men's evolution (Dattner, 2003). They have been extensively used to prepare herbal medicines as they are known to be safer than most modern medicines (Ahmad et al., 2013). Examples of medicinal plants used for wound healing are *Morinda citrifolia* (Nayak et al., 2007), *Radix paeoniae* (Malviya et al., 2009), *Lawsonia alba* (Mandawgade & Patil., 2003) and *Ginko biloba* (Bairya & Rao, 2001). The use of natural products to replace man-made drugs has risen significantly in the last decade as there are concerns about side effects of conventional medicine (Darakshan et al., 2015).

Nigella sativa Linn., (*N. sativa*) is an annual flowering plant belonging to Ranunculaceae family (Khan et al., 2011). It is an amazing herb with a rich historical background (Ng et al., 2011). It is native to North Africa, Southern Europe and Southwest Asia (Ahmad et al., 2013). *N. sativa* is also known as Habbatus Sauda, black seed, roman coriander and kalonji (Hussain and Hussain, 2016). Black seed has been extensively used as spice, condiment (Yarnell & Abascal, 2011) and was one of the earliest cultivated plants in human history (Desai et al., 2015). It also has been utilized as herbal medicine by various cultures to treat various ailments (Butt & Sultan, 2010; Khan et al., 2015) including high blood pressure, headache, fever, dizziness and influenza (Baharetha et al., 2013; Agbaria et al., 2015). The principle active ingredient isolated from the essential oil of *N. sativa* is thymoquinone (TQ) (Mahfouz & El-Dakhkhany, 1960; Hajhashemi et al., 2004). TQ is responsible for the therapeutic properties of this plant (Ahmad et al., 2013). The pharmacological investigation of the seed extracts reveals a variety of activities including antihypertensive (El Tahir et al., 1993), anti-inflammatory (Ali et al., 2003; Houghton et al., 1995), antihistaminic (Mahfouz et al., 1965), antimicrobial (El-Alfy et al., 1975; Kouidhi et al., 2011), antioxidant (Staneik & Gille, 2010) and antidiabetic (Al-Hader et al., 1993; Abdelmeguid et al., 2010).

Many studies on *N. sativa* and TQ have been previously recorded to find a solution for diabetes mellitus yet none have been carried out to observe the effect of TQ on wounds in diabetic patients. Although TQ is known for its varied functions, because its bioavailability is poor, its clinical use is limited (Pathan et al., 2011). Therefore, to overcome this problem, together with the rapidly progressing nanotechnology today, it is indeed important to look at a new alternative can effectively reach beneath the skin to speed the healing process.

Nanostructured lipid carrier (NLC), a colloidal drug carrier is tagged to a compound or extract that has trouble with bioavailability or absorption in the body. As Ng et al. (2015) previously described, thymoquinone-loaded nanostructured lipid carrier (TQ-NLC) (PATENT NO: PI201200181) was produced through a hot high-pressure homogenization method and yielded remarkable physicochemical properties. With high encapsulation efficiency, and a particle size of less than 50 nm in diameter, TQ-NLC has a stability of up to two years (Ng et al., 2015). It presents a larger surface area for reaction with its target component and also minimizes the probability of it being phagocytosed by macrophages (zur Mühlen et al., 1998; Ong et al., 2016). TQ-NLC has a huge potential to be used for wounds of diabetic patients. With all the attributes of NLC and the effectiveness of TQ as an anti-oxidant, anti-inflammation and antidiabetic agent, TQ-NLC may be an effective agent for diabetic wound healing.

1.2 Problem statement

Diabetic patients develop wounds that are prone to infection and display impaired wound healing that leads to many other complications such as chronic ulceration, and resultant limb amputation (Bowling et al., 2015). A great deal of effort has gone into developing treatments for diabetic wounds. Still, current treatment for diabetic wound comes with adverse side effects that lead to further problems such as damage to the large blood vessels of the heart and kidney complication (Chaudhury et al., 2017). Therefore, it is crucial to discover an alternative treatment to accelerate the healing of diabetic wound with lesser side effects. One target has been the reduction of chronic inflammation by restoring levels of endogenous antioxidants and reducing the level of reactive oxygen species (Griffith et al., 2009). TQ was reported to significantly reduce the levels of reactive oxygen species and act as a good antioxidant (Badary, 2003). It was also found to be a good anti-diabetic agent through numerous studies (Al-Trad et al., 2016; Al Wafai et al., 2013). Hence, with the integration of the new advance nanotechnology together with the anti-diabetic and anti-oxidant property of TQ, we decided to look at the beneficial effect of TQ-nanostructured lipid carrier to enhance healing process, not only in normal condition but also in prolonged tissue damage of diabetic patients. There are no previous reports on the *in vitro* wound healing properties of TQ and TQ-NLC.

1.3 Objectives

1.3.1 General objective

To elucidate the healing effect of TQ and TQ-NLC in normal and diabetic cell models.

1.3.2 Specific Objectives

1. To measure cell viability upon treatment with TQ and TQ-NLC using MTT assay
2. To examine the mode of cell death in both the cell lines using Annexin V assay
3. To evaluate the effect of TQ and TQ-NLC on cell migration in normal and diabetic models using scratch assay.
4. To evaluate the ROS level of both compounds toward human skin fibroblast 3T3 and 3T3-L1 cells.

1.4 Hypothesis

It is hypothesized that TQ-NLC is a better candidate and have better healing properties by enhancing the proliferation and migration process as well as reducing the ROS levels compared to TQ.



REFERENCES

- Abdelwahab, S. I., Sheikh, B. Y., Taha, M. M., How, C. W., Abdullah, R., Yagoub, U., ... Eid, E. E. (2013). Thymoquinone-loaded nanostructured lipid carriers: preparation, gastroprotection, *in vitro* toxicity, and pharmacokinetic properties after extravascular administration. *International Journal of Nanomedicine*, 8:2163–2172.
- Abd El-Ghany RM, Sharaf NM, Kassem LA, Mahran LG, Heikal OA. (2009). Thymoquinone triggers anti-apoptotic signaling targeting death ligand and apoptotic regulators in a model of hepatic ischemia reperfusion injury. *Drug Discov Ther*, 3:296–306.
- Abdelmeguid, N. E., Fakhoury, R., Kamal, S. M., & Al Wafai, R. J. A. L. (2010). Effects of *Nigella sativa* and thymoquinone on biochemical and subcellular changes in pancreatic β -cells of streptozotocin-induced diabetic rats. *Journal of Diabetes*, 2(4):256–266.
- Abdul Latif, M., Zulasyraf Mohd Zaki, M., May Leng, T., Hidayah Abdul Rahman, N., Aisyah Arshad, S., & Hamid, A. (2015). *Alocasia denudata* Engler treatment enhance open wound healing activities in Wistar rat's skin. *Journal of Ethnopharmacology*, 176:258–267.
- Abdulla, M., Fard, A., Sabaratnam, V., Kah Hui, W., Kuppusamy, U. R., Abdullah, N., & Ismail, S. (2011). Potential Activity of Aqueous Extract of Culinary-Medicinal Lion's Mane Mushroom, *Hericium erinaceus* (Bull.: Fr.) Pers. (Aphyllphoromycetideae) in Accelerating Wound Healing in Rats. *International journal of medicinal mushrooms*, 13:33–39.
- Abukhader, M. M. (2013). Thymoquinone in the clinical treatment of cancer: Fact or fiction? *Pharmacognosy reviews*, 7(14):117–120.
- Agbaria, R., Gabarin, A., Dahan, A., & Ben-Shabat, S. (2015). Anticancer activity of *Nigella sativa* (black seed) and its relationship with the thermal processing and quinone composition of the seed. *Drug design, development and therapy*, 9:3119–3124.
- Ahmad, A., Husain, A., Mujeeb, M., Khan, S. A., Najmi, A. K., Siddique, N. A., Damanhour, Z.A., Anwar F. (2013). A review on therapeutic potential of *Nigella sativa*: A miracle herb. *Asian Pacific journal of tropical biomedicine*, 3(5):337–352.
- Al-Gubory, K. H., Fowler, P. A., & Garrel, C. (2010). The roles of cellular reactive oxygen species, oxidative stress and antioxidants in pregnancy outcomes. *The International Journal of Biochemistry & Cell Biology*, 42(10):1634–1650.

- Al-Hader, A., Aqel, M., & Hasan, Z. (1993). Hypoglycemic Effects of the Volatile Oil of *Nigella sativa* Seeds. *International Journal of Pharmacognosy*, 31(2):96–100.
- Ali, B. H. and Blunden, G. (2003), Pharmacological and toxicological properties of *Nigella sativa*. *Phytotherapy Research*, 17:299-305.
- Alkharfy K. M., Ahmad A., Khan R. M., Al-Shagha W. M. (2015). Pharmacokinetic plasma behaviors of intravenous and oral bioavailability of thymoquinone in a rabbit model. *Eur. J. Drug Metab. Pharmacokinet*, 40:319–323.
- Al-Malki, A. L., & Sayed, A. A. R. (2014). Thymoquinone attenuates cisplatin-induced hepatotoxicity via nuclear factor kappa- β . *BMC Complementary and Alternative Medicine*, 14(1):282.
- Al-Qubaisi, M., Rozita, R., Yeap, S. K., Omar, A. R., Ali, A. M., & Alitheen, N. B. (2011). Selective cytotoxicity of goniiothalamine against hepatoblastoma HepG2 cells. *Molecules*, 16(4):2944–2959.
- AI-Trad B, AI-Batayneh K, EI-Metwally S, Alhazimi A, Ginawi I, Alaraj M, et al. *Nigella sativa* oil and thymoquinone ameliorate albuminuria and renal extracellular matrix accumulation in the experimental diabetic rats. *Eur Rev Med Pharmacol Sci* 2016; 20(12): 2680-2688
- Al Wafai RJ. *Nigella sativa* and thymoquinone suppress cyclooxygenase-2 and oxidative stress in pancreatic tissue of streptozotocin-induced diabetic rats. *Pancreas* 2013; 42(5): 841-849.
- Amin, M. N., Banik, S., Ibrahim, M., Moghal, M. M. R., Majumder, M. S., Siddika, R., Alam, M.K., Rahat Maruf, K.M., Anonna, S. N. (2015). A study on *Ardisia solanacea* for evaluation of phytochemical and pharmacological properties. *International Journal of Pharmacognosy and Phytochemical Research*, 7(1):8–15.
- Ammar, I., Bardaa, S., Mzid, M., Sahnoun, Z., Rebaii, T., Attia, H., & Ennouri, M. (2015). Antioxidant, antibacterial and *in vivo* dermal wound healing effects of *Opuntia* flower extracts. *International Journal of Biological Macromolecules*, 81:483–490.
- Armania, N., Saiful Yazan, L., Noorhidayah Musa, S., Ismail, I. S., Foo, J. B., Chan, K. W., Noreen, H., Hisyam, A.H., Zulfahmi, S., Ismail, M. (2013). *Dillenia suffruticosa* exhibited antioxidant and cytotoxic activity through induction of apoptosis and G2/M cell cycle arrest. *Journal of Ethnopharmacology*, 146(2):525-535.

- Asaduzzaman Khan, M., Tania, M., Fu, S., & Fu, J. (2017). Thymoquinone, as an anticancer molecule: from basic research to clinical investigation. *Oncotarget*, 8(31):51907–51919.
- Asmat, U., Abad, K., & Ismail, K. (2016). Diabetes mellitus and oxidative stress—A concise review. *Saudi Pharmaceutical Journal*, 24(5):547–553.
- Ayyanar, M., and Ignacimuthu, S. (2009). Herbal medicines for wound healing among tribal people in Southern India: ethnobotanical and scientific evidences. *International Journal of Applied Research in Natural Products*, 2(3):29–42.
- Badary O. A. (1999). Thymoquinone attenuates ifosfamide-induced Fanconi syndrome in rats and enhances its antitumor activity in mice. *J Ethnopharmacology*, 67:135–42.
- Badary, O. A., Taha, R. A., Gamal El-Din, A. M., & Abdel-Wahab, M. H. (2003). Thymoquinone Is a Potent Superoxide Anion Scavenger. *Drug and Chemical Toxicology*, 26(2):87–98.
- Baharetha, H. M., Nassar, Z. D., Aisha, A. F., Ahamed, M. B., Al-Suede, F. S., Abd Kadir, M. O., Ismail, Z., Majid, A. M. (2013). Proapoptotic and antimetastatic properties of supercritical CO₂ extract of *Nigella sativa* Linn. against breast cancer cells. *Journal of medicinal food*, 16(12):1121–1130.
- Bainbridge. P. (2013). Wound healing and the role of fibroblasts. *Journal of Wound Care*, 22(8):407–412.
- Bairy, K., & Rao, C. (2001). Wound Healing Profiles of *Ginkgo biloba*. *Journal Of Natural Remedies*, 1(1), 25-27.
- Balekar, N., Pasi, A. K., Parihar, G., & Jain, D. K. (2013). Antiarthritic activity of hydroalcoholic seed extract of *Cassia tora* Linn. *Topclass Journal of Herbal Medicine*, 2(11):254-262.
- Baltzis, D., Eleftheriadou, I., Veves, A. (2014). Pathogenesis and treatment of impaired wound healing in diabetes mellitus: new insights. *Advances in Therapy*, 31(8):817-836.
- Banerjee S, Kaseb AO, Wang Z, Kong D, Mohammad M, et al. (2009). Antitumor activity of gemcitabine and oxaliplatin is augmented by thymoquinone in pancreatic cancer. *Cancer Res*, 69:5575– 5583.
- Barreto, S. R., Albuquerque-Júnior, R. L., Araújo, A. A., Almeida, J. R., Santos, M. R., Barreto, A. S., DeSantana J. M., Siqueira-Lima P. S., Quintans, J. S., Quintans-Júnior, J. L. (2014). A Systematic Review of the Wound-Healing Effects of Monoterpenes and Iridoid Derivatives. *Molecules*. 19(1):846-862.

- Barrientos, S., Stojadinovic, O., Golinko, M. S., Brem, H. and Tomic- Canic, M. (2008). PERSPECTIVE ARTICLE: Growth factors and cytokines in wound healing. *Wound Repair and Regeneration*, 16(5):585-601.
- Beckmann, K. H., Meyer-Hamme, G., & Schröder, S. (2014). Low level laser therapy for the treatment of diabetic foot ulcers: a critical survey. *Evidence-based complementary and alternative medicine: eCAM*, 2014:1-9.
- Beloqui, A., Solinís, M. Á., Rodríguez-Gascón, A., Almeida, A. J., & Prést, V. (2016). Nanostructured lipid carriers: Promising drug delivery systems for future clinics. *Nanomedicine: Nanotechnology, Biology and Medicine*, 12(1):143–161.
- Bennett SP, Griffiths GD, Schor AM, Leese GP, Schor SL. Growth factors in the treatment of diabetic foot ulcers. *Br J Surg* 2003; 90: 133–46
- Berlanga-Acosta, J., Schultz, G. S., López-Mola, E., Guillen-Nieto, G., García-Siverio, M., & Herrera-Martínez, L. (2013). Glucose toxic effects on granulation tissue productive cells: the diabetics' impaired healing. *BioMed Research International*, 2013:1–15.
- Berrios-Torres, S. I., Umscheid, C. A., Bratzler, D. W., Leas, B., Stone, E. C., Kelz, R. R., et al. (2017). Centers for disease control and prevention guideline for the prevention of surgical site infection. *JAMA Surgery*, 152(8):784–791.
- Bhaskar A, Nithya V. (2012). Evaluation of the wound-healing activity of *Hibiscus rosa sinensis* L (Malvaceae) in Wistar albino rats. *Indian Journal of Pharmacology*, 44(6):450–457.
- Bowling FL, Rashid ST, Boulton AJ. Preventing and treating foot complications associated with diabetes mellitus. *Nat Rev Endocrinol* 2015;11:606-16.
- Briquez, P. S., Hubbell, J. A., & Martino, M. M. (2015). Extracellular Matrix-Inspired Growth Factor Delivery Systems for Skin Wound Healing. *Advances in wound care*, 4(8):479–489.
- Butt, M. S., & Sultan, M. T. (2010). *Nigella sativa*: Reduces the Risk of Various Maladies. *Critical Reviews in Food Science and Nutrition*, 50(7):654–665.
- Cakirca M, Karatoprak C, Zorlu M, et al. Effect of vildagliptin add-on treatment to metformin on plasma asymmetric dimethylarginine in type 2 diabetes mellitus patients. *Drug Des Devel Ther*. 2014;8:239–243.
- Cheeseman, K. H. (1993). Tissue Injury by Free Radicals. *Toxicology and Industrial Health*, 9(1-2):39–51.

- Chen, Q., Jin, M., Yang, F., Zhu, J., Xiao, Q., & Zhang, L. (2013). Matrix metalloproteinases: inflammatory regulators of cell behaviours in vascular formation and remodelling. *Mediators of inflammation*, 2013:1-14
- Chaudhury, A., Duvoor, C., Reddy Dendi, V. S., Kraleti, S., Chada, A., Ravilla, R., ... Mirza, W. (2017). Clinical Review of Antidiabetic Drugs: Implications for Type 2 Diabetes Mellitus Management. *Frontiers in endocrinology*, 8, 6. doi:10.3389/fendo.2017.00006
- Chung A. S., Ferrara M. D. (2010). The Extracellular Matrix and Angiogenesis: The Role of the Extracellular Matrix in Developing Vessels and Tumor Angiogenesis. *Pathways*, 11:1-5.
- Chushri S., Settharaksa S., Chokpaisarn J., Limsuwan S., Voravuthikunchai S. P. (2013). Thai herbal formulas used for wound treatment: a study of their antibacterial potency, anti-inflammatory, antioxidant, and cytotoxicity effects. *Journal of Alternative and Complement Medicine*, 19(7):671-676.
- Cordeiro MF, Reichel MB, Gay JA, D'Esposito F, Alexander RA, Khaw PT. Transforming growth factor-beta1, -beta2, and -beta3 in vivo: effects on normal and mitomycin C-modulated conjunctival scarring. *Invest Ophthalmol Vis Sci* 1999; 40: 1975–82.
- Cork, M. J., Danby, S. G., Vasilopoulos, Y., Hadgraft, J., Lane, M. E., Moustafa, M., Guy, R.H., Macgowan, A.L., Tazi-Ahnini, R., Ward, S. J. (2009). Epidermal Barrier Dysfunction in Atopic Dermatitis. *Journal of Investigative Dermatology*, 129(8): 1892–1908.
- Daniel, C., Weigmann, B., Bronson, R., & Boehmer, H. Von. (2011). Prevention of type 1 diabetes in mice by tolerogenic vaccination with a strong agonist insulin mimotope. *The Journal of Experimental Medicine*, 208(7):1501-1511.
- Darakhshan, S., Bidmeshki Pour, A., Hosseinzadeh Colagar, A., & Sisakhtnezhad, S. (2015). Thymoquinone and its therapeutic potentials. *Pharmacological Research*, 95–96:138–158.
- Dartsch DC, Schaefer A, Boldt S, Kolch W, Marquardt H. Comparison of anthracycline-induced death of human leukemia cells: programmed cell death versus necrosis. *Apoptosis*. 2002;7:537–48.
- Dattner, A. M. (2003). From medical herbalism to phytotherapy in dermatology: back to the future. *Dermatologic Therapy*, 16(2):106–113.
- Desai, S. D., Saheb, S., Das, K., & Haseena, S. (2015). Phytochemical analysis of *Nigella sativa* and its antidiabetic effect. *Journal of Pharmaceutical Sciences and Research*, 7(8):527-532.

- Dewangan H., Bais M., Jaiswal V., Verma VK. (2012). Potential wound healing activity of the ethanolic extract of *Solanum xanthocarpum* Schrad and wendl leaves. *Pakistan Journal of Pharmaceutical Science*, 25:189-194.
- Dhall, S., Do, D. C., Garcia, M., Kim, J., Mirebrahim, S. H., Lyubovitsky, J., ... Martins-Green, M. (2014). Generating and Reversing Chronic Wounds in Diabetic Mice by Manipulating Wound Redox Parameters. *Journal of Diabetes Research*, 2014:1–18.
- Dhivya, S., Padma, V. V., & Santhini, E. (2015). Wound dressings - a review. *BioMedicine*, 5(4):22.
- Diegelmann, R. F., & Evans, M. C. (2004). Wound healing: an overview of acute, fibrotic and delayed healing. *Frontiers in Bioscience*, 9:283-289.
- Dong, H., Wang, N., Zhao, L., & Lu, F. (2012). Berberine in the Treatment of Type 2 Diabetes Mellitus: A Systemic Review and Meta-Analysis. *Evidence-Based Complementary and Alternative Medicine*, 2012:1–12.
- Dubey, A., Prabhu, P., & Kamath, J. V. (2012). Nano Structured lipid carriers: A Novel Topical drug delivery system, *International Journal of PharmTech Research*, 4(2):705–714.
- Dubinsky, J., Kristal, B., & Elizondo-Fournier, M. (1995). An obligate role for oxygen in the early stages of glutamate-induced, delayed neuronal death. *The Journal of Neuroscience*, 15(11):7071–7078.
- Dunnill, C., Patton, T., Brennan, J., *et al.* (2015). Reactive oxygen species (ROS) and wound healing: the functional role of ROS and emerging ROS modulating technologies for augmentation of the healing process. *International Wound Journal*, 14 (1):89–96.
- Durmşkahya C. & Öztürk M. (2013). Ethnobotanical Survey of Medicinal Plants Used for the Treatment of Diabetes in Manisa, Turkey. *Sains Malaysiana*, 42(10):1431–1438.
- Eckes, B., Moinszadeh, P., Sengle, G., Hunzelmann, N., Krieg, T. (2014) Molecular and cellular basis of scleroderma. *Journal of Molecular Medicine (Berlin, Germany)*, 92(9):913-924.
- El Alfy, S. T., M. El Fatatry, H., & Toama, M. A. (1975). Isolation and Structure Assignment of an Antimicrobial Principle from the Volatile Oil of *Nigella sativa* L. Seeds. *Pharmazie*, 30(2):109-111.
- El-Dakhakhny M. (1963). Studies on the chemical constituents of Egyptian *Nigella sativa* L. seeds. The essential oil. *Planta Medica*, 11(4): 465–470.

- El-Mesallamy, H. O., Diab, M. R., Hamdy, N. M., & Dardir, S. M. (2014). Cell-Based Regenerative Strategies for Treatment of Diabetic Skin Wounds, a Comparative Study between Human Umbilical Cord Blood-Mononuclear Cells and Calves' Blood Haemodialysate. *PLoS ONE*, 9(3):e89853
- El Tahir, E. H. K., Ashour, M., & M. Al-Harbi, M. (1993). The respiratory effects of the volatile oil of the black seed (*Nigella sativa*) in guinea-pigs: Elucidation of the mechanism(s) of action. *General pharmacology*, 24(5):1115-1122.
- S. Y. Latifah, W. K. Ng, G. Al-Naqeeb, and I. Maznah, "Cytotoxicity of thymoquinone (TQ) from *Nigella sativa* towards human cervical carcinoma cell (HeLa)," *Journal of Pharmacy Research*, vol. 2, no. 4, pp. 585–589, 2009.
- W. K. Ng, L. S. Yazan, and M. Ismail, "Thymoquinone from *Nigella sativa* was more potent than cisplatin in eliminating of SiHa cells via apoptosis with down-regulation of Bcl-2 protein," *Toxicology In Vitro*, vol. 25, no. 7, pp. 1392–1398, 2011.
- El-Zawahrawy, B., Fatma Al-Zahraa, A. H. (1998). Effect of *Nigella sativa* on blood level and structure of liver and pancreas in adult male albino rats. *Al-Azhar Medical Journal*, 27:479–483.
- Falanga V. Wound healing and its impairment in the diabetic foot. *Lancet*. 2005;366(9498):1736–1743.
- Famili, A., Ammar, D. A., & Kahook, M. Y. (2013). Ethyl pyruvate treatment mitigates oxidative stress damage in cultured trabecular meshwork cells. *Molecular vision*, 19, 1304–1309.
- Fang, C., Al-Suwayeh, S., & Fang, J.-Y. (2012). Nanostructured Lipid Carriers (NLCs) for Drug Delivery and Targeting. *Recent patents on nanotechnology* Vol. 7(1):41-55.
- Fanjul-Fernández, M., Folgueras, A. R., Cabrera, S., & López-Otín, C. (2010). Matrix metalloproteinases: Evolution, gene regulation and functional analysis in mouse models. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*, 1803(1):3–19.
- Farah, K. M., Atoji, Y., Shimizu, Y., Shiina, T., Nikami, H., Takewaki, T. (2004). Mechanisms of the hypoglycaemic and immunopotentiating effects of *Nigella sativa* L. oil in streptozotocin-induced diabetic hamsters. *Research in Veterinary Science*, 77(2):23-29.
- Fitzmaurice, S. D., Sivamani, R. K., Isseroff, R. R. (2011). Antioxidant therapies for wound healing: A clinical guide to currently commercially available products. *Skin Pharmacology Physiology*, 24:113–26.

- Fronza, M., Heinzmann, B., Hamburger, M., Laufer, S., & Merfort, I. (2009). Determination of the wound healing effect of Calendula extracts using the scratch assay with 3T3 fibroblasts. *Journal of Ethnopharmacology*, 126(3): 463–467.
- Frykberg, R. G., & Banks, J. (2015). Challenges in the Treatment of Chronic Wounds. *Advances in Wound Care*, 4(9):560–582.
- Gainza, G., Villullas, S., Pedraz, J. L., Hernandez, R. M., & Igartua, M. (2015). Advances in drug delivery systems (DDSs) to release growth factors for wound healing and skin regeneration. *Nanomedicine: Nanotechnology, Biology and Medicine*, 11(6):1551–1573.
- García Orue, I., Gainza, G., Gutierrez, F. B., Aguirre Anda, J. J., Evora, C., Luis Pedraz, J., ... Igartua, M. (2016). Novel nanofibrous dressings containing rhEGF and Aloe vera for wound healing applications. *International Journal of Pharmaceutics*, 523(2): 556–566.
- Ghosheh, O. A., Houdi, A. A., & Crooks, P. A. (1999). High performance liquid chromatographic analysis of the pharmacologically active quinones and related compounds in the oil of the black seed (*Nigella sativa* L.). *Journal of Pharmaceutical and Biomedical Analysis*, 19(5):757–762.
- Gill, S. E., & Parks, W. C. (2008). Metalloproteinases and their inhibitors: regulators of wound healing. *The International Journal of Biochemistry & Cell Biology*, 40(6-7):1334–1347.
- Goletz, S., Zillikens, D., & Schmidt, E. (2017). Structural proteins of the dermal-epidermal junction targeted by autoantibodies in pemphigoid diseases. *Experimental Dermatology*, 26(12):1154–1162.
- Gonçalves R. V., Novaes R. D., Matta S. L. P., Benevides G. P., Faria F. R., Pinto M. V. M. (2010). Comparative study of the effects of gallium-aluminum-arsenide laser photobiomodulation and healing oil on skin wounds in wistar rats: a histomorphometric study. *Photomedicine and Laser Surgery*, 28(5):597–602.
- Gonzalez, A. C., Costa, T. F., Andrade, Z. A., & Medrado, A. R. (2016). Wound healing - A literature review. *Anais brasileiros de dermatologia*, 91(5):614–620.
- Greenish H. S. (1880). Contribution to the Chemistry of *Nigella sativa*. *Pharmac J Trans.*, 10: 909-911.
- Green, H., & Meuth, M. (1974). An established pre-adipose cell line and its differentiation in culture. *Cell*, 3(2):127–133.
- Griffith B, Pendyala S, Hecker L, Lee PJ, Natarajan V, and Thannickal VJ. NOX enzymes and pulmonary disease. *Antioxid Redox Signal* 11: 2505–2516.

- Gulati, V., Gulati, P., Harding, I. H., & Palombo, E. A. (2015). Exploring the anti-diabetic potential of Australian Aboriginal and Indian Ayurvedic plant extracts using cell-based assays. *BMC Complementary and Alternative Medicine*, 15(1):8.
- Guo, S., & DiPietro, L. A. (2010). Factors Affecting Wound Healing. *Journal of Dental Research*, 89(3): 219–229.
- Guo, J. J., Yang, H., Qian, H., Huang, L., Guo, Z., & Tang, T. (2010). The Effects of Different Nutritional Measurements on Delayed Wound Healing After Hip Fracture in the Elderly. *Journal of Surgical Research*, 159(1): 503–508.
- Guo, T., Zhang, Y., Zhao, J., Zhu, C., & Feng, N. (2015). Nanostructured lipid carriers for percutaneous administration of alkaloids isolated from *Aconitum sinomontanum*. *Journal of Nanobiotechnology*, 13(1):47.
- Gupta, A., Upadhyay, N. K., Sawhney, R. and Kumar, R. (2008). A poly- herbal formulation accelerates normal and impaired diabetic wound healing. *Wound Repair and Regeneration*, 16: 784-790.
- Gurtner, G. C., Werner, S., Barrandon, Y., & Longaker, M. T. (2008). Wound repair and regeneration. *Nature*, 453(7193):314–32.
- Hajhashemi, V., Ghannadi, A. and Jafarabadi, H. (2004), Black cumin seed essential oil, as a potent analgesic and anti-inflammatory drug. *Phytotherapy Research*, 18: 195-199.
- Hajiaghaalipour, F., Kanthimathi, M. S., Abdulla, M. A., & Sanusi, J. (2013). The Effect of *Camellia sinensis* on Wound Healing Potential in an Animal Model. *Evidence-Based Complementary and Alternative Medicine: eCAM*, 2013:1–7.
- Hantash, B., M. (2008). Adult and fetal wound healing. *Frontiers in Bioscience*, 13(13):51.
- Haron, A. S., Syed Alwi, S. S., Saiful Yazan, L., Abd Razak, R., Ong, Y. S., Zakarial Ansar, F. H., & Roshini Alexander, H. (2018). Cytotoxic Effect of Thymoquinone-Loaded Nanostructured Lipid Carrier (TQ-NLC) on Liver Cancer Cell Integrated with Hepatitis B Genome, Hep3B. *Evidence-Based Complementary and Alternative Medicine*, 2018:1–13.
- Heldin CH, Westermark B. Mechanism of action and in vivo role of platelet-derived growth factor. *Physiol Rev* 1999; 79: 1283–316.
- Horton JW. Free radicals and lipid peroxidation mediated injury in burn trauma: the role of antioxidant therapy. *Toxicology* 2003;189:75–88.

- Hosseinkhani, A., Falahatzadeh, M., Raoofi, E., & Zarshenas, M. M. (2016). An Evidence-Based Review on Wound Healing Herbal Remedies from Reports of Traditional Persian Medicine. *Journal of evidence-based complementary & alternative medicine*, 22(2):334–343.
- Houghton, P. J., Zarka, R., Delasheras, B., Hoult, J.R. (1995). Fixed oil of *Nigella sativa* and derived thymoquinone inhibit eicosanoid generation in leukocytes and membrane lipid peroxidation. *Planta Medica*, 61: 33–36
- Houghton, P. J., Hylands, P. J., Mensah, A. Y., et al. (2005). *In vitro* tests and ethnopharmacological investigations: Wound healing as an example. *Journal of Ethnopharmacology*, 100:100–7.
- Hsu, C. C., Lee, C. H., Wahlqvist, M. L., Huang, H. L., Chang, H. Y., Chen, L., ... Cheng, J. S. (2012). Poverty increases type 2 diabetes incidence and inequality of care despite universal health coverage. *Diabetes care*, 35(11):2286–2292.
- Hsu, Y.-C., Li, L., & Fuchs, E. (2014). Emerging interactions between skin stem cells and their niches. *Nature Medicine*, 20(8):847–856.
- Hu, S. C.-S., & Lan, C.-C. E. (2016). High-glucose environment disturbs the physiologic functions of keratinocytes: Focusing on diabetic wound healing. *Journal of Dermatological Science*, 84(2): 121–127.
- Hussain, D. A., & Hussain, M. M. (2016). *Nigella sativa* (black seed) is an effective herbal remedy for every disease except death – a Prophetic statement which modern scientists confirm unanimously: A review. *Advancement in Medicinal Plant Research*, 4(2): 27–57.
- Ilango, K., & Chitra, V. (2010). Wound Healing and Anti-oxidant Activities of the Fruit Pulp of *Limonia acidissima* Linn (Rutaceae) in Rats. *Tropical Journal of Pharmaceutical Research*, 9(3):223–230.
- Jores, K., Mehnert, W., & Mäder, K. (2003). Physicochemical Investigations on Solid Lipid Nanoparticles and on Oil-Loaded Solid Lipid Nanoparticles: A Nuclear Magnetic Resonance and Electron Spin Resonance Study. *Pharmaceutical research*, 20(8): 1274–1283.
- Kaseb AO, Chinnakannu K, Chen D, Sivanandam A, Tejwari S, et al. Androgen receptor and E2F-1 targeted thymoquinone therapy for hormone-refractory prostate cancer. *Cancer Res.* 2007; 67:7782– 7788. [PubMed: 17699783]
- Khader M., Bresgen N., Eckl P. (2009). *In vitro* toxicological properties of thymoquinone. *Food Chem. Toxicol*, 47(1):129–33.

- Khan, M. A., Chen, H. C., Tania, M., & Zhang, D. Z. (2011). Anticancer activities of *Nigella sativa* (black cumin). *African journal of traditional, complementary, and alternative medicines: AJTCAM*, 8(5 Suppl):226–232.
- Khan, M. A., Tania, M., Wei, C., Mei, Z., Fu, S., Cheng, J., ... Fu, J. (2015). Thymoquinone inhibits cancer metastasis by downregulating TWIST1 expression to reduce epithelial to mesenchymal transition. *Oncotarget*, 6(23):19580–19591.
- Kido, D. K. Mizutani, K. Takeda, R. Mikami, T. Matsuura, K. Iwasaki, et al. (2017). Impact of diabetes on gingival wound healing via oxidative stress *PLoS One*, 12(12): e0189601
- Kmiecik, B., Skotny, A., Wawrzaszek, R., & Rybak, Z. (2013). Influence of oxidative stress on tissue regeneration. *Polimers in Medicine*, 43(3):191–197.
- Kokane, D., More, Y., Kale, M., Nehete, M., Mehendale, P., & Gadgoli, C. (2009). Evaluation of wound healing activity of root of *Mimosa pudica*. *Journal of Ethnopharmacology*, 124(2):311-315.
- Koh, T. J., & DiPietro, L. A. (2011). Inflammation and wound healing: the role of the macrophage. *Expert reviews in molecular medicine*, 13:e23.
- Kouidhi, B., Zmantar, T., Jrah, H., Souiden, Y., Chaieb, K., Mahdouani, K., & Bakhrouf, A. (2011). Antibacterial and resistance-modifying activities of thymoquinone against oral pathogens. *Annals of Clinical Microbiology and Antimicrobials*, 10(1):29.
- Krishnaswamy, V.R., Mintz, D., Sagi I. (2017). Matrix metalloproteinases: the sculptors of chronic cutaneous wounds. *Biochim Biophys Acta*, 1864(11 Pt B):2220-2227.
- Kunkemoeller, B., & Kyriakides, T. R. (2017). Redox Signaling in Diabetic Wound Healing Regulates Extracellular Matrix Deposition. *Antioxidants & redox signaling*, 27(12):823–838.
- Kurahashi, T., & Fujii, J. (2015). Roles of Antioxidative Enzymes in Wound Healing. *Journal of Developmental Biology*, 3(2): 57–70.
- Lauffenburger, D. A, & Horwitz, A. F. (1996). Cell migration: a physically integrated molecular process. *Cell.*, 9;84(3):359-69.
- Lee HS, Kooshesh F, Sauder DN, Kondo S. Modulation of TGF-beta 1 production from human keratinocytes by UVB. *Exp Dermatol* 1997; 6: 105–10.

- Li, B., & Wang, J. H. (2011). Fibroblasts and myofibroblasts in wound healing: force generation and measurement. *Journal of tissue viability*, 20(4):108–120.
- Linjaw, S. A., Khalil, W. K., Hassanane, M. M., & Ahmed, E. S. (2015). Evaluation of the protective effect of *Nigella sativa* extract and its primary active component thymoquinone against DMBA-induced breast cancer in female rats. *Archives of medical science: AMS*, 11(1), 220–229. doi:10.5114/aoms.2013.33329
- Lima, M. H. M., Caricilli, A. M., de Abreu, L. L., Araújo, E. P., Pelegrinelli, F. F., et al. (2012). Topical Insulin Accelerates Wound Healing in Diabetes by Enhancing the AKT and ERK Pathways: A Double-Blind Placebo-Controlled Clinical Trial. *PLOS ONE*, 7(5): e36974
- Lipsky, B. A., & Hoey, C. (2009) Topical antimicrobial therapy for treating chronic wounds. *Clinical Infectious Disease*, 49(10):1541–9.
- Mabrouk, A., & Cheikh, H. B. (2015). Thymoquinone supplementation reverses lead-induced oxidative stress in adult rat testes. *General Physiology and Biophysics*, 34(01):65–72.
- Mahfouz, M., & El-Dakhakhny, M. (1960). The isolation of a crystalline active principle from *Nigella sativa* L. seeds. *J Pharm Sci UAR*, 1:9-19.
- Mahfouz, M., Abdel-Maguid, R., El-Dakhakhny, M. (1965). The effect of “nigellone therapy” on the histaminopexic power of the blood sera in asthmatic patients. *Arzneim Forsch (Drug Res.)*, 15:1230–1234.
- Maiga, A., Diallo, D., Fane, S., Sanogo, R., Paulsen, B. S. & Cisse, B. (2005). “A survey of toxic plants on the market in the district of Bamako, Mali: traditional knowledge compared with a literature search of modern pharmacology and toxicology,” *Journal of Ethnopharmacology*, vol. 96, no. 1-2, pp. 183–193,
- Mandawgade, S. D., & Patil, K. S. (2003). Wound healing potential of some active principles of *Lawsonia alba* Lam. leaves. *Indian Journal of Pharmaceutical Sciences*, 65, 390–394.
- Malviya, N., Malviya, N., & Jain, S. (2009). Wound healing activity of aqueous extract of *Radix paeoniae* root. *Acta Poloniae Pharmaceutica*, 66, 543–547.
- Majdalawieh, A. F., & Fayyad, M. W. (2015). Immunomodulatory and anti-inflammatory action of *Nigella sativa* and thymoquinone: A comprehensive review. *International Immunopharmacology*, 28(1):295–304.

- Mandavia, C. H., Aroor, A. R., Demarco, V. G., & Sowers, J. R. (2013). Molecular and metabolic mechanisms of cardiac dysfunction in diabetes. *Life sciences*, 92(11):601–608.
- Manjunath, K., Reddy, J. S., & Venkateswarlu, V. (2005). Solid lipid nanoparticles as drug delivery systems. *Methods and Findings in Experimental and Clinical Pharmacology*, 27(2):127.
- Mannello, F. (2011). What does matrix metalloproteinase-1 expression in patients with breast cancer really tell us? *BMC Medicine*, 9(1):95.
- Mansour, M. A., Nagi, M. N., El- Khatib, A. S. and Al- Bekairi, A. M. (2002). Effects of thymoquinone on antioxidant enzyme activities, lipid peroxidation and DT- diaphorase in different tissues of mice: a possible mechanism of action. *Cell Biochem. Funct*, 20:143-151.
- Martin, P. (1997). Wound healing-aiming for perfect skin regeneration. *Science*, 276(5309): 75–81.
- Matsusaki, M., Fujimoto, K., Shirakata, Y., Hirakawa, S., Hashimoto, K., & Akashi, M. (2015). Development of full-thickness human skin equivalents with blood and lymph-like capillary networks by cell coating technology. *J. Biomed. Mater. Res. A*, 103(10):3386-3396.
- McGuire S. (2014) Centers for Disease Control and Prevention. State indicator report on Physical Activity, 2014. Atlanta, GA: U.S. *Department of Health and Human Services Adv Nutr*, 5(2014): 762-763.
- McLafferty E, Hendry C, Alistair F. (2012). The integumentary system: anatomy, physiology and function of skin. *Nurs Stand*, 27(3):35-42.
- McLennan, S.V., Min, D., & Dk, Yue. (2008). Matrix metalloproteinases and their roles in poor wound healing in diabetes. *Wound practice and research*, 16(3):116-121.
- Medina A, Scott PG, Ghahary A, Tredget EE. Pathophysiology of chronic nonhealing wounds. *J Burn Care Rehabil*. 2005;26(4):306–319.
- Meral I, Yener Z, Kahraman T, Mert N. (2001). Effect of *Nigella sativa* on glucose concentration, lipid peroxidation, anti-oxidant defence system and liver damage in experimentally-induced diabetic rabbits. *J Vet Med A Physiol Pathol Clin Med*, 48(10):593–599.
- Mehnert, W., & Mäder, K. (2001). Solid lipid nanoparticles: Production, characterization and applications. *Advanced Drug Delivery Reviews*, 47(2):165–196.
- Mishra, M., Kumar, H., & Tripathi, K. (2008). Diabetic delayed wound healing and the role of silver nanoparticles. *Digest Journal of Nanomaterials and Biostructures*, 3(2):49-54.

- Miyoshi, Y., Ogawa, O., & Oyama, Y. (2016). Nivolumab, an Anti-Programmed Cell Death-1 Antibody, Induces Fulminant Type 1 Diabetes. *The Tohoku Journal of Experimental Medicine*, 239(2):155–158.
- Mognetti B., Barberis A., Marino S., et al. (2012). *In vitro* enhancement of anticancer activity of paclitaxel by a Cremophor free cyclodextrin-based nanosponge formulation. *Journal of Inclusion Phenomena and Macrocyclic Chemistry*, 74(1–4):201–210.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *J Immunol Methods*, 65:55–63.
- Mosser, D. M., & Edwards, J. P. (2008). Exploring the full spectrum of macrophage activation. *Nature reviews. Immunology*, 8(12):958–969.
- Morton, L. M., & Phillips, T. J. (2016). Wound healing and treating wounds: Differential diagnosis and evaluation of chronic wounds. *Journal of the American Academy of Dermatology*, 74(4):589–605.
- Müller, R. H., Petersen, R. D., Hommoss, A., & Pardeike, J. (2007). Nanostructured lipid carriers (NLC) in cosmetic dermal products. *Advanced Drug Delivery Reviews*, 59(6), 522–530.
- Murphy, P. S., & Evans, G. R. (2012). Advances in wound healing: a review of current wound healing products. *Plastic surgery international*, 2012:1–8.
- Nagi, M. N., & Mansour, M. A. (2000). Protective effect of thymoquinone against doxorubicin-induced cardiotoxicity in rats: A possible mechanism of protection. *Pharmacological Research*, 41(3):283–289.
- Nagori B. P., & Solanki R. (2011). Role of medicinal plants in wound healing. *Res J Med Plant*; 5(4): 392-405.
- Namiki A, Brogi E, Kearney M, Kim EA, Wu T, Couffinhal T, Varticovski L, Isner JM. Hypoxia induces vascular endothelial growth factor in cultured human endothelial cells. *J Biol Chem* 1995; 270: 31189–95
- National diabetes fact sheet: national estimates and general information on diabetes and prediabetes in the United States (2011). National Diabetes Fact Sheet, 2011 [PDF file]. https://www.cdc.gov/diabetes/pubs/pdf/ndfs_2011.pdf. Retrieved 20th February, 2019.
- Nayak, B. S., Isitor, G. N., Maxwell, A., Bhogadi, V., & Ramdath, D. D. (2007). Wound-healing activity of *Morinda citrifolia* fruit juice on diabetes-induced rats. *Journal of Wound Care*, 16(2), 83–86.

- Nazaruk, J., & Galicka, A. (2014). The Influence of Selected Flavonoids from the Leaves of *Cirsium palustre* (L.) Scop. on Collagen Expression in Human Skin Fibroblasts. *Phytotherapy Research*, 28(9):1399–1405.
- Ng, W. K., Yazan, L. S., & Ismail, M. (2011). Thymoquinone from *Nigella sativa* was more potent than cisplatin in eliminating of SiHa cells via apoptosis with down-regulation of Bcl-2 protein. *Toxicology in Vitro*, 25(7): 1392–1398.
- Ng, W. K., Saiful Yazan, L., Yap, L. H., Wan Nor Hafiza, W. A. G., How, C. W., & Abdullah, R. (2015). Thymoquinone-Loaded Nanostructured Lipid Carrier Exhibited Cytotoxicity towards Breast Cancer Cell Lines (MDA-MB-231 and MCF-7) and Cervical Cancer Cell Lines (HeLa and SiHa). *BioMed Research International*, 2015:1–10.
- Nikam A.P., Mukesh P., R., C. S. P. (2014). Nanoparticles – an overview. *International Journal of Research and Development in Pharmacy & Life Sciences*, 3(5):1121–1127.
- Novak, I., Kirkin, V., McEwan, D. G., Zhang, J., Wild, P., Rozenknop, A., ... Dikic, I. (2010). Nix is a selective autophagy receptor for mitochondrial clearance. *EMBO Reports*, 11(1):45–51.
- Okonkwo, U.A., & DiPietro L.A. (2017). Diabetes and wound angiogenesis. *International Journal of Molecular Science*, 18(7):1419.
- Ong, Y. S., Saiful Yazan, L., Ng, W. K., Noordin, M. M., Sapuan, S., Foo, J. B., & Tor, Y. S. (2016). Acute and subacute toxicity profiles of thymoquinone-loaded nanostructured lipid carrier in BALB/c mice. *International Journal of Nanomedicine*, 11:5905–5915.
- Pardeike, J., Hommoss, A., & Müller, R. H. (2009). Lipid nanoparticles (SLN, NLC) in cosmetic and pharmaceutical dermal products. *International Journal of Pharmaceutics*, 366(1-2):170–184.
- Paradis, V., Mathurin, P., Kollinger, M., Imbert-Bismut, F., Charlotte, F., Piton, A., Opolon, P., Holstege, A., Poynard, T. & Bedossa, P. (1997). In situ detection of lipid peroxidation in chronic hepatitis C: correlation with pathological features. *Journal of Clinical Pathology*, 50(5):401–406.
- Pari, L., & Sankaranarayanan, C. (2009). Beneficial effects of thymoquinone on hepatic key enzymes in streptozotocin–nicotinamide induced diabetic rats. *Life Sciences*, 85(23):830–834.
- Park, K.-S., & Park, D.-H. (2018). The effect of Korean red ginseng on full-thickness skin wound healing in rats. *Journal of Ginseng Research*, 43(2):226–235.

- Patel, P., Gupta, P., Burns, A., Mohamed, A. A., Cole, R., Lane, D., ... Khunti, K. (2019). Biochemical Urine Testing of Adherence to Cardiovascular Medications Reveals High Rates of Nonadherence in People Attending Their Annual Review for Type 2 Diabetes. *Diabetes Care*, dc181453.
- Pathan, S. A., Jain, G. K., Zaidi, S. M. A., Akhter, S., Vohora, D., Chander, P., ... Khar, R. K. (2010). Stability-indicating ultra-performance liquid chromatography method for the estimation of thymoquinone and its application in biopharmaceutical studies. *Biomedical Chromatography*, 25(5):613–620.
- Pawar, R. S., & Toppo, F. A. (2012). Plants that heal wounds. A review. *Kerba Polonica*, 58(1), 47–65.
- Petreaca, M. L., Do, D., Dhall, S., McLelland, D., Serafino, A., et al. (2012). Deletion of a tumor necrosis superfamily gene in mice leads to impaired healing that mimics chronic wounds in humans. *Wound Repair Regen*, 20:353–366.
- Pitz, H. da S., Pereira, A., Blasius, M. B., Voytena, A. P. L., Affonso, R. C. L., Fanan, S., ... Maraschin, M. (2016). *In vitro* Evaluation of the Antioxidant Activity and Wound Healing Properties of Jaboticaba (*Plinia peruviana*) Fruit Peel Hydroalcoholic Extract. *Oxidative Medicine and Cellular Longevity*, 2016:1–6.
- Plikus, M. V., Guerrero-Juarez, C. F., Ito, M. Li., Y. R., Dedhia, P. H., Zheng, Y., et al. Regeneration of fat cells from myofibroblasts during wound healing. *Science*, 355(6326):748-752.
- Polat, R., Satil, F., & Çakılcıoğlu, U. (2011). Medicinal plants and their use properties of sold in herbal market in Bingöl (Turkey) district. *Biological Diversity and Conservation*, 4(3):25-35.
- Ponnusamy, L., Mahalingaiah, P. S., & Singh, K. P. (2015). Chronic oxidative stress increases resistance in renal carcinoma cells to doxorubicin-induced cytotoxicity potentially through epigenetic mechanism. *Cancer Research*, 75(15): 4571–4571.
- Powers CJ, McLeskey SW, Wellstein A. Fibroblast growth factors, their receptors and signaling. *Endocr Relat Cancer* 2000; 7: 165–97
- Profyris, C., Tziotziou, C., & Do Vale, I. (2012). Cutaneous scarring: Pathophysiology, molecular mechanisms, and scar reduction therapeutics: Part I. The molecular basis of scar formation. *Journal of the American Academy of Dermatology*, 66(1):1–10.
- Puri, D., Bhandari, A., Sharma, P., & Choudhary, D. (2010). Lipid nanoparticles (SLN, NLC): A novel approach for cosmetic and dermal pharmaceutical. *Journal of Global Pharma Technology*, 2(9):1-15.

- Qing, C. (2017). The molecular biology in wound healing & non-healing wound. *Chinese Journal of Traumatology*, 20(4): 189-193.
- Qi, J., Lu, Y., & Wu, W. (2012). Absorption, Disposition and Pharmacokinetics of Solid Lipid Nanoparticles, *Current Drug Metabolism*, 13(4):418-428.
- Rabeta, M. S., & Nur Faraniza, R. (2013). Total phenolic content and ferric reducing antioxidant power of the leaves and fruits of *garcinia atrovirdis* and *cynometra cauliflora*. *International Food Research Journal*, 20(4):1691–1696.
- Raher, E. (1998). Collagen and the Phases of Wound Healing. *Wound Caring* 5(2).
- Raja, Sivamani K, Garcia MS, Isseroff RR. Wound reepithelialization: modulating keratinocyte migration in wound healing. *Front Biosci* 2007; 12: 2849–68.
- Randhawa M. A. & Alghamdi M. S. (2002). Review of the Pharmaco-Therapeutic Effects of *Nigella sativa*. *Pakistan Journal of Medical Research*, 41(2):77-83.
- Raposo, S. C., Simões, S. D., Almeida, A. J., & Ribeiro, H. M. (2013). Advanced systems for glucocorticoids' dermal delivery. *Expert Opinion on Drug Delivery*, 10(6):857–877.
- Reece E.A. (2010). The fetal and maternal consequences of gestational diabetes mellitus. *The Journal of Maternal-Fetal & Neonatal Medicine*, 23(3): 199 203.
- Richard, J.-L., Lavigne, J.-P., Got, I., Hartemann, A., Malgrange, D., Tsirtsikolou, D., ... Senneville, E. (2011). Management of patients hospitalized for diabetic foot infection: Results of the French OPIDIA study. *Diabetes & Metabolism*, 37(3):208–215.
- Rigoulet, M., Yoboue, E. D., & Devin, A. (2011). Mitochondrial ROS Generation and Its Regulation: Mechanisms Involved in H₂O₂ Signalling. *Antioxidants & Redox Signaling*, 14(3):459–468.
- Roepke M, Diestel A, Bajbouj K, Walluscheek D, Schonfeld P, et al. Lack of p53 augments thymoquinone-induced apoptosis and caspase activation in human osteosarcoma cells. *Cancer Biol Ther*. 2007; 6:160–169. [PubMed: 17218778]
- Rodrigues da Silva, G. H., Ribeiro, L. N. M., Mitsutake, H., Guilherme, V. A., Castro, S. R., Poppi, R. J., ... de Paula, E. (2017). Optimised NLC: a nanotechnological approach to improve the anaesthetic effect of bupivacaine. *International Journal of Pharmaceutics*, 529(1):253–263.

- Sahak, M. K. A., Kabir, N., Abbas, G., Draman, S., Hashim, N. H., & Hasan Adli, D. S. (2016). The Role of *Nigella sativa* and Its Active Constituents in Learning and Memory. *Evidence-Based Complementary and Alternative Medicine*, 2016:1–6.
- Sánchez-Campillo, M., Larqué, E., González-Silvera, D., Martínez-Tomás, R., García-Fernández, M., Avilés, F., ... Pérez-Llamas, F. (2012). Changes in the carotenoid concentration in human postprandial chylomicron and antioxidant effect in HepG2 caused by differently processed fruit and vegetable soups. *Food Chemistry*, 133(1): 38–44.
- Sawant, K., & Dodiya, S. (2008). Recent Advances and Patents on Solid Lipid Nanoparticles. *Recent Patents on Drug Delivery & Formulation*, 2(2):120–135.
- Schäfer-Korting, M., Mehnert, W., & Korting, H.-C. (2007). Lipid nanoparticles for improved topical application of drugs for skin diseases. *Advanced Drug Delivery Reviews*, 59(6):427–443.
- Schäfer, M., & Werner, S. (2008). Oxidative stress in normal and impaired wound repair. *Pharmacological Research*, 58(2):165–171.
- Schmidt, C., Fronza, M., Goettert, F., Geller, F., Luik, S., Flores, E.M.M., Bittencourt, C.F., Heinzmann, B.M., Laufer, S., Merfort, I. (2009). Biological studies on Brazilian plants used in wound healing. *Journal of Ethnopharmacology*, 122: 523–532.
- Schreml, S., Szeimies, R., Prantl, L., Karrer, S., Landthaler, M., & Babilas, P. (2010). Oxygen in acute and chronic wound healing. *The British Journal of Dermatology*, 163(2):257–68.
- Schultz G, Rotatori DS, Clark W. EGF and TGF-alpha in wound healing and repair. *J Cell Biochem* 1991; 45: 346–52.
- Schwarz, C., Mehnert, W., Lucks, J. S., & Müller, R. H. (1994). Solid lipid nanoparticles (SLN) for controlled drug delivery. I. Production, characterization and sterilization. *Journal of Controlled Release*, 30(1):83–96.
- Selçuk CT, Durgun M, Tekin R, Yolbas L, Bozkurt M, Akçay C, et al. Evaluation of the effect of thymoquinone treatment on wound healing in a rat burn model. *J Burn Care Res* 2013;34:e274–81
- Severino, P., Pinho, S. C., Souto, E. B., & Santana, M. H. A. (2011). Polymorphism, crystallinity and hydrophilic–lipophilic balance of stearic acid and stearic acid–capric/caprylic triglyceride matrices for production of stable nanoparticles. *Colloids and Surfaces B: Biointerfaces*, 86(1):125–130.

- Shah, J. M., Omar, E., Pai, D. R., & Sood, S. (2012). Cellular events and biomarkers of wound healing. *Indian journal of plastic surgery: official publication of the Association of Plastic Surgeons of India*, 45(2): 220–228.
- Sharath, R., Harish, B., Krishna, V., Sathyanarayana, B. and Swamy, H. K. (2010). Wound healing and protease inhibition activity of Bacoside- A, isolated from *Bacopa monnieri* wettest. *Phytotherapy Research*, 24:1217-1222.
- Sharma Y, Jeyabalan J, Singh R. (2013) Potential wound healing agents from medicinal plants: a review. *Pharmacologia*, 4:1–17.
- Shaw, T. J., & Martin, P. (2009). Wound repair at a glance. *Journal of Cell Science*, 122(18):3209–3213.
- Shiraha H, Glading A, Gupta K, Wells A. IP-10 inhibits epidermal growth factor-induced motility by decreasing epidermal growth factor receptor-mediated calpain activity. *J Cell Biol* 1999; 146: 243–54. 45.
- Solomonov, I., Zoharai, E., Talmi-Frank, D., Wolf, S.G., Shainskaya, A., Zhuravlev, A., Kartvelishvily, E., Visse, R., Levin, L., Kampf, N, Jaitin, D. A., David, E., Amit, I., Nagase, H., Sagi, I. (2016). Distinct biological events generated by ECM proteolysis by two homologous collagenases. *Proc. Natl. Acad. Sci. U. S. A*, 113(39):10884-10889.
- Sorrell, J. M., & Caplan, A. I. (2004). Fibroblast heterogeneity: more than skin deep. *Journal of Cell Science*, 117(5):667–675.
- Staniek, K., & Gille, L. (2010). Is thymoquinone an antioxidant? *BMC Pharmacology*, 10(1):A9.
- Stojadinovic, A., Carlson, J. W., Schultz, G. S., Davis, T. A., & Elster, E. A. (2008). Topical advances in wound care. *Gynecologic Oncology*, 111(2):70–80.
- Sudsai T., Wattanapiromsakul C., Tewtrakul S. (2016). Wound healing property of isolated compounds from *Boesenbergia kingii* rhizomes. *Journal of Ethnopharmacology*, 184:42-48.
- Sumaria, N., Roediger, B., Ng, L. G., Qin, J., Pinto, R., Cavanagh, L. L., ... Weninger, W. (2011). Cutaneous immunosurveillance by self-renewing dermal $\gamma\delta$ T cells. *The Journal of Experimental Medicine*, 208(3):505-518.
- Tam, J. C., Lau, K. M., Liu, C. L., To, M. H., Kwok, H. F., Lai, K. K., Lau, C. P., Ko, C. H., Leung, P. C., Fung, K. P., Lau, C. B. S. (2011). The *in vivo* and *in vitro* diabetic wound healing effects of a 2-herb formula and its mechanisms of action. *Journal of Ethnopharmacology*, 134(3):831-838.

- Tang A, Gilchrist BA. Regulation of keratinocyte growth factor gene expression in human skin fibroblasts. *J Dermatol Sci* 1996; 11: 41–50.
- Teller, P., & White, T. K. (2009) The physiology of wound healing: injury through maturation. *Surg Clin North Am*, 89(3):599-610.
- Tembhurne, S., S, F., More B., H., & D, M. S. (2014). A review on therapeutic potential of *Nigella sativa* (Kalonji) seeds. *Journal of Medicinal Plants Research*, 8(3):167-177.
- Thakare, V. M., Chaudhari, R. Y., & Patil, V. R. (2011). Wound Healing Evaluation of Some Herbal Formulations Containing Curcuma Longa and Cynodon Dactylon Extract. *International Journal of Phytomedicine*, 3(2011):325-332.
- Thakur, R., Jain, N., Pathak, R., & Sandhu, S. S. (2011). Practices in Wound Healing Studies of Plants. *Evidence-Based Complementary and Alternative Medicine*, 2011:1–17.
- Thestrup- Pedersen, K. (2000). Clinical aspects of atopic dermatitis. *Clinical and Experimental Dermatology*, 25(7):535-543.
- Thummuri, D., Jeengar, M. K., Shrivastava, S., Nemani, H., Ramavat, R. N., Chaudhari, P., & Naidu, V. G. M. (2015). Thymoquinone prevents RANKL-induced osteoclastogenesis activation and osteolysis in an in vivo model of inflammation by suppressing NF-KB and MAPK Signalling. *Pharmacological Research*, 99:63–73.
- Tiwari, R., & Pathak, K. (2011). Nanostructured lipid carrier versus solid lipid nanoparticles of simvastatin: Comparative analysis of characteristics, pharmacokinetics and tissue uptake. *International Journal of Pharmaceutics*, 415(1):232–243.
- Tokumaru S, Higashiyama S, Endo T, Nakagawa T, Miyagawa JI, Yamamori K, Hanakawa Y, Ohmoto H, Yoshino K, Shirakata Y, Matsuzawa Y, Hashimoto K, Taniguchi N. Ectodomain shedding of epidermal growth factor receptor ligands is required for keratinocyte migration in cutaneous wound healing. *J Cell Biol* 2000; 151: 209– 20. 38.
- Tubessa, Z., Abu Bakar, M. Z., & Ismail, M. (2013). Characterization and Stability Evaluation of Thymoquinone Nanoemulsions Prepared by High-Pressure Homogenization. *Journal of Nanomaterials*, 2013:1-16.
- Tyrone JW, Marcus JR, Bonomo SR, Mogford JE, Xia Y, Mustoe TA. Transforming growth factor beta3 promotes fascial wound healing in a new animal model. *Arch Surg* 2000; 135: 1154–9.
- Uutela M, Wirzenius M, Paavonen K, Rajantie I, He Y, Karpanen T, Lohela M, Wiig H, Salven P, Pajusola K, Eriksson U, Alitalo K. PDGF-D induces

- macrophage recruitment, increased interstitial pressure, and blood vessel maturation during angiogenesis. *Blood* 2004; 104: 3198–204.
- Velnar, T., Bailey, T., & Smrkolj, V. (2009). The Wound Healing Process: An Overview of the Cellular and Molecular Mechanisms. *Journal of International Medical Research*, 37(5):1528–1542.
- Venus, M., Waterman, J., & McNab, I. (2011). Basic physiology of the skin. *Surgery*, 29(10):471-474.
- Vermes, I., Haanen, C., Steffens-Nakken, H., & Reutellingsperger, C. (1995). A novel assay for apoptosis Flow cytometric detection of phosphatidylserine expression on early apoptotic cells using fluorescein labelled Annexin V. *Journal of Immunological Methods*, 184(1):39–51.
- Vinardell, M. P., Sordé, A., Díaz, J., Baccarin, T., & Mitjans, M. (2015). Comparative effects of macro-sized aluminum oxide and aluminum oxide nanoparticles on erythrocyte hemolysis: influence of cell source, temperature, and size. *Journal of Nanoparticle Research*, 17(2):80-90.
- Vollmar, B., El-Gibaly, A., Scheuer, C., W Strik, M., Bruch, H.-P., & Menger, M. (2002). Acceleration of Cutaneous Wound Healing by Transient p53 Inhibition. *Laboratory investigation; a journal of technical methods and pathology*, 82(8):1063-71.
- Walter, C., Klein, M. O., Pabst, A., Al-Nawas, B., Duschner, H., & Ziebart, T. (2010). Influence of bisphosphonates on endothelial cells, fibroblasts, and osteogenic cells. *Clinical Oral Investigations*, 14(1):35–41.
- Wang, L., Xu, M. L., Rasmussen, S. K., and Wang, M. H. (2011). Vomifoliol 9-O- α -arabinofuranosyl (1 $\rightarrow$ 6)- β -D-glucopyranoside from the leaves of *Diospyros Kaki* stimulates the glucose uptake in HepG2 and 3T3-L1 cells. *Carbohydrate Research Journal*, 4:21.
- Weiss, E., Zohary, D., & Hopf, M. (2012). Domestication of Plants in the Old World - The Origin and Spread of Domesticated Plants in South-west Asia, Europe, and the Mediterranean Basin. 4th Oxford University Press.
- Wong, H. L., Rauth, A. M., Bendayan, R., Manias, J. L., Ramaswamy, M., Liu, Z., ... Wu, X. Y. (2006). A New Polymer–Lipid Hybrid Nanoparticle System Increases Cytotoxicity of Doxorubicin Against Multidrug-Resistant Human Breast Cancer Cells. *Pharmaceutical Research*, 23(7):1574–1585.
- Yaman, İ., & Balıkcı, E. (2010). Protective effects of *Nigella sativa* against gentamicin-induced nephrotoxicity in rats. *Experimental and Toxicologic Pathology*, 62(2):183–190.

- Yamini, K., & Gopal, V. (2016). Herbal Challenge for Wound Healing – A Traditional Review. *Journal of Pharmacognosy and Phytochemistry*, 4(3): 5–12.
- Yarnell, E., & Abascal, K. (2011). *Nigella sativa*: Holy Herb of the Middle East. *Alternative and Complementary Therapies*, 17(2): 99–105.
- Yebra M, Parry GC, Stromblad S, Mackman N, Rosenberg S, Mueller BM, Cheresh DA. Requirement of receptorbound urokinase-type plasminogen activator for integrin alphavbeta5-directed cell migration. *J Biol Chem* 1996; 271: 29393–9.
- Zhang, Y., Lin, Y., Bowles, C., & Wang, F. (2004). Direct Cell Cycle Regulation by the Fibroblast Growth Factor Receptor (FGFR) Kinase through Phosphorylation-dependent Release of Cks1 from FGFR Substrate 2. *Journal of Biological Chemistry*, 279(53):55348–55354.
- Zubair H, Khan HY, Sohail A, Azim S, Ullah MF, Ahmad A, Sarkar FH, Hadi SM. Redox cycling of endogenous copper by thymoquinone leads to ROS-mediated DNA breakage and consequent cell death: putative anticancer mechanism of antioxidants. *Cell Death Dis.* 2013;4:e660.
- zur Mühlen, A., Schwarz, C., & Mehnert, W. (1998). Solid lipid nanoparticles (SLN) for controlled drug delivery – Drug release and release mechanism. *European Journal of Pharmaceutics and Biopharmaceutics*, 45(2):149–155.