

EFFECT OF TEMPERATURE, CARBON DIOXIDE AND WATER ACTIVITY ON PRODUCTION OF PARENT AND MASKED MYCOTOXINS BY Fusarium spp. AT GERMINATION STAGE

By

MSHELIA LADI PETER

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

December 2020

IPTSM 2020 17

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



EFFECT OF TEMPERATURE, CARBON DIOXIDE AND WATER ACTIVITY ON PRODUCTION OF PARENT AND MASKED MYCOTOXINS BY Fusarium spp. AT GERMINATION STAGE

By

MSHELIA LADI PETER

December 2020

Chairman : Professor Jinap binti Selamat, PhD
Institute : Tropical Agriculture and Food Security

Maize is an important crop cultivated for food and feed in different parts of the world. Like any cereal, maize grain is susceptible to mycotoxin contamination on the farm, during storage and processing. Therefore, the aims of the present work were to (1) evaluate the biodiversity of Fusarium spp. in maize and soil samples collected from different farms, (2) access the effects of climate change factors (aw, CO₂ and temperature) on the production of parent and masked mycotoxins in maize-based growth medium (in vitro), and, (3) determine the effect of climate change factors (aw, CO₂ and temperature) on gene expression and production of parent and masked mycotoxins in germinating and autoclaved maize kernels by F. verticillioides and F. graminearum. A total of 68 Fusarium spp. were isolated from maize and soil samples. Molecular identification by DNA sequencing of TEF-1α gene regions confirmed the isolates. The fungal isolates found were F. verticillioides, F. graminearum, and F. incarnatum. Fusarium spp. contamination was found to be significantly higher in maize (p < 0.05) as compared to the soil samples regardless of the growth medium used. All representative isolates tested using LC-MS/MS were found to be mycotoxigenic. F. verticillioides tested showed Fumonisin production ability, FB₁ (12.0 \pm 0.9 to 15,200 \pm 432 µg/kg), FB₂ (3.39 \pm $0.70 \text{ to } 2510 \pm 692 \text{ } \mu\text{g/kg})$ and FB₃ ($12.0 \pm 0.4 \text{ to } 20.1 \pm 2.1 \text{ } \mu\text{g/kg})$. While for F. graminearum DON (4,760 \pm 580 µg/kg), 3-ADON (3,980 \pm 62 µg/kg) and ZEN $(50 \pm 0.1 \,\mu\text{g/kg})$ was detected and F. incarnatum produced FB₁ $(14.3 \pm 1.2 \text{ to } 4,670 \, \text{m})$ ± 835 μg/kg). F. verticillioides and F. graminearum were further selected because F. incarnatum could not withstand elevated temperature. F. verticillioides was acclimatised at 30 and 35°C while F. graminearum at 30 and 33°C. The effect of aw, CO₂ and temperature were tested on the growth and mycotoxin production maize-based medium. F. verticillioides appears to be more tolerant to elevated temperature, a_w and CO_2 as compared to F. graminearum. However, F. verticillioides produced FB₁ (29.07 \pm 6.70 to 140.49 \pm 1.45 µg/kg) and FB₂ (21.46

 \pm 14.10 to 32.17 \pm 0.54 µg/kg). For the studies on gene expression, the total fungal biomass of the isolate was drastically reduced which was suspected to occur as a result of the effect of climate change factors. This negatively affected the extraction of RNA for further determination of the mycotoxigenic gene expression. However, the results obtained in the present study revealed that mycotoxins were detected in germinating and autoclaved maize kernel by those two acclimatised isolates. The optimum conditions for mycotoxin production were detected at 0.98 aw, 400 ppm CO₂ and 30°C for both isolates in germinating and autoclaved maize kernel. However, masked mycotoxin was not detected in this study. At 30°C, the mycotoxins produced by F. verticillioides were FB₁ and FB₂ while, F. graminearum produced DON, ZEA and \alpha-ZEA. Comparing Fusarium spp. and other species, Aspergillus spp. grow faster and have a high resistance to temperature than Fusarium spp. Therefore, more emphasis must be given to such species. However, it must be noted that the production of mycotoxins other than the targeted ones by these isolates may be possible under the effects of climate change factors. Also, further acclimatisation of the isolates to both temperature and CO₂ might affect the mycotoxigenic fungi due to their flexibility. Therefore, more research is also needed in this area to determine the general types of mycotoxins produced.

KESAN SUHU, KARBON DIOKSIDA DAN AKTIVITI AIR TERHADAP PENGHASILAN MIKOTOKSIN UTAMA DAN TERSEMBUNYI OLEH Fusarium spp. PADA PERINGKAT PECAMBAHAN

Oleh

MSHELIA LADI PETER

Disember 2020

Pengerusi : Profesor Jinap binti Selamat, PhD

Institut : Pertanian Tropika dan Sekuriti Makanan

Jagung adalah tumbuhan yang sangat penting di beberapa kawasan di dunia sebagai makanan bagi manusia dan haiwan. Seperti bijirin yang lain, bijirin jagung juga mudah terdedah kepada pencemaran mikotoksin di ladang, semasa penyimpanan dan pemprosesan. Oleh itu, objektif kajian ini adalah untuk; (1) menilai biodiversiti Fusarium spp. di dalam sampel jagung dan tanah yang diambil dari beberapa ladang jagung, (2) mengakses kesan faktor-faktor perubahan iklim (suhu, karbon dioksida dan aktiviti air) terhadap pengeluaran mikotoksin utama dan mikotoksin tersembunyi di dalam medium pertumbuhan berasaskan jagung (in vitro) dan, (3) mengenalpasti kesan faktor-faktor perubahan iklim (suhu, karbon dioksida dan aktiviti air) terhadap ekspresi gen dan penghasilan mikotoksin utama dan mikotoksin tersembunyi. Sejumlah 68 Fusarium spp. telah diasingkan daripada sampel jagung dan tanah. Pengenalan molekul menggunakan penjujukan DNA TEF-1α kawasan gen telah memastikan isolat tersebut. Tanpa mengambilkira media yang digunakan, jumlah bilangan kulat iaitu F. verticillioides, F. graminearum, and F. incarnatum. Fusarium spp. yang dihasilkan daripada sampel jagung adalah lebih tinggi (p < 0.05) berbanding sampel tanah. Semua isolat diuji menggunakan LC-MS/MS didapati adalah mikotoksigenik. F. verticillioides menunjukkan kebolehan untuk menghasilkan Fumonisin, FB₁ (12.0 ± 0.9 hingga $15,200 \pm 432 \,\mu \text{g/kg}$), FB₂ (3.39 ± 0.70 hingga 2510 ± 692 $\mu \text{g/kg}$) dan FB₃ (12.0 ± 0.4 hingga 20.1 \pm 2.1 μ g/kg). Manakala, untuk *F. graminearum*, DON (4,760 \pm 580 $\mu g/kg$), 3-ADON (3,980 ± 62 $\mu g/kg$) dan ZEN (50 ± 0.1 $\mu g/kg$) telah dikesan dan F. incarnatum menghasilkan FB₁ (14.3 \pm 1.2 hingga 4,670 \pm 835 µg/kg). Hanya F. verticillioides dan F. graminearum telah dipilih selanjutnya kerana F. incarnatum tidak boleh menahan suhu yang tinggi. F. verticillioides telah disesuaikan pada suhu 30 dan 35°C, manakala, F. graminearum pada suhu 30 dan 33°C. Kesan aktiviti air, karbon dioksida dan suhu telah diuji pada pertumbuhan dan pengeluaran mikotoksin pada medium berasaskan jagung. F. verticillioides lebih

bertoleransi terhadap suhu yang tinggi, aktiviti air dan karbon dioksida, jika dibandingkan dengan F. graminearum. Namun begitu, F. verticillioides menghasilkan FB₁ (29.07 \pm 6.70 hingga 140.49 \pm 1.45 µg/kg) dan FB₂ (21.46 \pm 14.10 hingga 32.17 \pm 0.54 µg/kg). Untuk kajian mengenai gen ekspresi, jumlah biomass kulat yang telah diisolat berkurang secara drastik di mana disyaki berlaku akibat faktor perubahan iklim. Ini memberi kesan negatif kepada pengekstrakan RNA untuk penentuan lanjut tentang mikotoksigenik gen ekspresi. Walau bagaimanapun, hasil yang diperoleh dalam kajian ini menunjukkan bahawa mikotoksin berjaya dikesan di dalam biji jagung bercambah dan diautoklaf oleh kedua-dua isolat tersebut. Keadaan optima untuk penghasilan mikotoksin dikesan pada 0.98 aw, 400 ppm CO₂ dan 30°C bagi kedua-dua isolat dalam biji jagung bercambah dan diautoklaf. Walau bagaimanapun, mikotoksin tersembunyi tidak dikesan dalam kajian ini. Pada suhu 30°C, mikotoksin yang dihasilkan oleh F. verticillioides adalah FB₁ dan FB₂, manakala, F. graminearum menghasilkan DON, ZEA dan α-ZEA. Jika dibandingkan Fusarium spp. dengan spesis yang lain, Aspergillus spp. tumbuh lebih cepat dan mempunyai rintangan tinggi terhadap suhu berbanding Fusarium spp. Oleh itu, penekanan lebih tinggi perlu diberikan kepada spesis ini. Walau bagaimanapun, perlu diingatkan bahawa penghasilan mikotoksin selain daripada yang ditarget oleh isolat tersebut adalah berkemungkinan di bawah faktor-faktor perubahan iklim. Selain itu, aklimatisasi selanjutnya bagi isolat terhadap suhu dan karbon dioksida mungkin mempengaruhi kulat mikotoksigenik disebabkan kelenturannya. Oleh itu, penyelidikan yang lebih banyak perlu dijalankan bagi bidang ini bagi menentukan jenis-jenis mikotoksin umum yang dapat dihasilkan.

ACKNOWLEDGEMENTS

I would like to express my sincere gratitude to my supervisor Professor Dr Jinap Selamat who played the most vital role in my PhD study. With her support, motherly care and listening ears to my challenges and high level of patience, tolerances, motivation, and immense knowledge. I would also want to thank my cosupervisors, Dr Nik Iskandar Putra Bin Samsudin, Prof Raffi Mohd Yusop and Assoc. Prof Franz Berthiller for their valuable advice and contribution to my studies.

My heartfelt gratitude goes to my husband, Mr Peter Mshelia who has been by my side throughout my study. I truly want to thank my kids, Lisa, David and Lenah for their patience, and prayers for my success, who brings joy in my life. Special thanks to my parents, Mr and Mrs Hyelsinta Katsalla for their prayers, support and patience throughout my study. I also want to thank my sisters for their moral support. I am also grateful for having great family in-laws for their moral support, prayer and patience.

Lastly, I would like to thank my good friend Norlia Mohror for her encouragement and kindness. I also want to appreciate my Lab mates who during my laboratory work we became good friends especially Izzati, Farah, Raehan, Aliah, Sharina, Aida, Fida and Din. I thank the UPM family for their prayers throughout my stay in Malaysia and my life in general. I pray that we shall all live long to enjoy the fruit of this labour. I thank the UPM community and I remain in awe of the mode of organization and proficiency of this great institution.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Jinap binti Selamat, PhD

Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Chairman)

Mohd Rafii bin Yusop, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

Nik Iskandar Putra bin Samsudin, PhD

Senior Lecturer
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Franz Berthiller, PhD

Associate Professor
Institute of Bioanalytics and Agro-Metabolomics
University of Natural Resources and Life Sciences, Vienna
Austria
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 11 May 2023

TABLE OF CONTENTS

				Page
ABSTRAC ABSTRAK ACKNOW APPROVA	LEDG	EMENT	TS .	i iii v vi
DECLARA				viii
LIST OF T				xiv
LIST OF F	IGUR	ES		xvii
LIST OF A	PPEN	DICES		xix
LIST OF A	BBRE	CVIATIO	ONS	XX
СНАРТЕК	2			
1	INTR	ODUCT	TION	1
1	1.1		ound of the study	1
	1.2	_	n statement	2
	1.3		cance of the study	3
	1.4	Objecti		4
		1.4.1	Objective of the study	4
		1.4.2	Specific objectives	4
	1.5	Researc	ch approach	5
2	LITE	RATUR	E REVIEW	6
	2.1	Maize I	Plant	6
		2.1.1	Developmental stages of a maize plant	6
		2.1.2	Nutritional value of maize	7
		2.1.3	Production and consumption of maize for feed	
			in Malaysia	8
		2.1.4	Mycotoxigenic contamination of maize feed by	
			Fusarium spp.	10
	2.2	Fusariu		10
		2.2.1	Overview of the genus Fusarium	10
		2.2.2	Chemical characteristics of Fusarium toxins	11
		2.2.3	Toxicological effects of Fusarium mycotoxins	12
		2.2.4	Morphological and molecular identification of <i>Fusarium</i> spp.	13
	2.3	Mycoto	exins (parent)	15
		2.3.1	Occurrence of mycotoxins in maize and feed	15
		2.3.2	Metabolism of parent mycotoxins	16
		2.3.3	Masked mycotoxins	17
		2.3.4	Types of masked mycotoxins	18
		2.3.5	Metabolism of masked mycotoxins	19
		2.3.6	International regulations of mycotoxins in feed	20
		2.3.7	Malaysian regulatory limit for mycotoxins	21
	2.4	Determ	ination of parents and masked mycotoxins	24

		2.4.1	LC-MS/MS method of determination of parent	24
	2.5	Climate	and masked mycotoxins	
	2.5		e change	28
		2.5.1	8	28
		2.5.2	Effect of climate change on parent and masked mycotoxins in maize	29
	2.6	Biosynt	thetic genes for mycotoxin production by	
		Fusariu	um spp.	30
	2.7		of temperature, CO ₂ and a _w on gene expression arium spp.	31
3	DIVE	RSITY A	AND MYCOTOXIGENIC POTENTIALS OF	
	Fusar	ium SPP	P. FROM MAIZE AND SOIL SAMPLES	32
	3.1	Introdu		32
	3.2		als and methods	35
	3.2	171410110	3.2.1.1 Sampling	35
		3.2.2	Mycotoxin standards and chemicals	35
		3.2.3	Measurement of moisture content in maize and	33
		3.4.3	soil samples	35
		3.2.4	Enumeration, isolation and identification of	33
		3.2.4		35
		2 2 5	fungi Isolation frequency	37
			Molecular identification	
		3.2.6		37
			3.2.6.1 DNA extraction	37
			3.2.6.2 PCR amplification and sequencing of	20
			TEF-1α genes	38
			3.2.6.3 Gel electrophoresis	39
		3.2.7	Determination of toxigenic potential of	
			Fusarium spp.	40
			3.2.7.1 Preparation of growth media	40
			3.2.7.2 Inoculation of <i>Fusarium</i> isolates	40
			3.2.7.3 Extraction of mycotoxins	40
			3.2.7.4 UPLC-MS method	41
		3.2.8	Statistical Analysis	41
	3.3	Results		42
		3.3.1	Moisture content of maize and soil samples	42
		3.3.2	Fungal populations isolated from maize and soil	
			samples	44
		3.3.3	Isolation frequency of Fusarium spp.	46
		3.3.4	pH of the soil samples	48
		3.3.5	Morphological and molecular identification of	
			Fusarium spp.	50
			3.3.5.1 Macro morphological appearance of	
			Fusarium spp.	50
			3.3.5.2 Micro-morphological appearance of	
			Fusarium spp.	54
			3.3.5.3 Molecular identification of <i>Fusarium</i>	Jr
			spp. using PCR amplification	54
			opp. asing I cit ampimication	J 1

		3.3.6	Mycotoxin production potential of Fusarium	56
	3.4	Discuss	spp.	59
	3.5	Conclu		61
	3.3	Conciu	SIOII	01
4	GROY MASI	WTH A	F TEMPERATURE, CO ₂ , AND A _W ON ND THE PRODUCTION OF PARENT AND YCOTOXINS BY F. verticillioides AND F.	62
	4.1	Introdu	ction	62
	4.2		als and methods	63
	1.2	4.2.1	Fungal isolates, mycotoxin standards and	03
		1.2.1	chemicals	63
		4.2.2		64
		4.2.3		64
		4.2.4	Modified atmosphere	65
		4.2.5	Preparation of culture medium and inoculation	
			of fungal strain	66
		4.2.6	Measurement of diametric growth rate	66
		4.2.7	Extraction of mycotoxins	67
		4.2.8	Determination of mycotoxins by LC-MS/MS	67
		4.2.9	Statistical analysis	67
	4.3	Results		67
		4.3.1	Effect of temperature, CO ₂ and a _w on the	
			diametric growth rate of F. verticillioides and	
			F. graminearum	67
		4.3.2	Effect of temperature, CO ₂ and a _w on	
			mycotoxins produced by F. verticillioides and	
		D.	F. graminearum	72
	4.4	Discuss		78
	4.5	Conclu	sion	81
5	FFFF	CT OF	TEMPERATURE, CARBON DIOXIDE AND	
3			TIVITY ON GENE EXPRESSION AND	
		DUCTIO		
		OTOXI		
			ED MAIZE KERNELS BY F. verticillioides	
		F. grami		82
	5.1	Introdu		82
	5.2	Materia	als and methods	83
		5.2.1	Mycotoxin standards and chemicals	83
		5.2.2	Experimental design	83
		5.2.3	Modified atmosphere	83
		5.2.4	Sample preparation	83
		5.2.5	Inoculation and incubation of F. verticillioides	
			and F. graminearum on germinating and	
			autoclaved maize kernels	84
		5.2.6	Extraction of mycotoxins	85
		5.2.7	Determination of mycotoxins using LC-MS/MS	85

		5.2.8	Gene expression	85	
		5.2.9	Extraction of RNA	86	
		5.2.10	Reverse transcription to convert mRNA into cDNA	87	
	5.3	Results	CD1VII	88	
	5.5	5.3.1	Effect of temperature CO_2 , and a_w on gene		
		5.3.2	expression	88 90	
		5.3.3	Expression of RNA gene on gel electrophoresis	90	
		3.3.3	Effect of temperature, CO ₂ and a _w on the production of parent mycotoxins by the		
			acclimatised strain of F. verticillioides in		
			germinating maize kernel	90	
		5.3.4	Effect of temperature, CO ₂ and a _w on the	90	
		3.3.7	production of parent and masked mycotoxins		
			by the acclimatised strain of F. graminearum		
			on germinating maize kernels	93	
		5.3.5	Effect of temperature, CO ₂ and a _w on the		
			production of parent and masked mycotoxins		
			by the acclimatised isolate of F. verticillioides		
			on autoclaved maize kernels	96	
		5.3.6	Effect of temperature, CO ₂ and a _w on the		
			production of parent and masked mycotoxins		
			by the acclimatised strain of F. graminearum		
			on autoclaved maize kernels	99	
	5.4	Discuss		102	
	5.5	Conclus	sion	105	
6	SHMN	MADV	CONCLUSION AND RECOMMENDATION		
U			ER RESEARCH	106	
	6.1	Summa		106	
	6.2		•	108	
	6.3		mendation for future research	108	
REF	ERENC	CES		109	
	APPENDICES				
	BIODATA OF STUDENT				
LIST OF PUBLICATIONS					

LIST OF TABLES

Table		Page
2.1	Proximate composition of maize grain per 100g of the edible portion	8
2.2	Global occurrence of mycotoxins in maize and Finished feed	16
2.3	Types of masked mycotoxins	18
2.4	Maximum levels of mycotoxins in feed materials set by the European Commission (moisture content: 12%)	22
2.5	LCMS/MS determination of mycotoxins in maize, feed and other cereals	25
3.1	50 μL reactions for PCR	38
3.2	GenBank accession number of extype isolates of <i>Fusarium</i> spp. used for the phylogenic analysis	39
3.3	Moisture content (%) of maize and soil samples obtained from different farms	43
3.4	Fungal load (log CFU/g) of maize and soil samples. cultivated on PDA, DG18 and DRBC at 30°C for 7 days	45
3.5	Isolation frequency (%) of <i>Fusarium</i> spp. from sterilized and non-sterilized maize kernels obtained from different locations cultivated on PDA, DG18 and DRBC at 30°C for seven days	47
3.6	pH of soil samples obtained from different farms	49
3.7	Mycotoxins produced by <i>Fusarium</i> spp. (μg/kg) incubated at 30°C on a maize-based medium for 21 days	58
4.1	Experimental design (factorial) on the effect of climate change factors (a _w , temperature and CO ₂) for <i>F. verticillioides</i>	65
4.2	Experimental design (factorial) on the effect of climate change factors (a_w , temperature and CO_2) for $F.\ graminearum$	65
4.3	Effect of temperature, (30, 35°C), CO ₂ levels (400, 800, 1200 ppm), and a _w (0.92, 0.95, 0.98 a _w) on the diametric growth rate of acclimatised strain of <i>F. verticillioides</i> on MMA	68
4.4	Analysis of variance of the effect of temperature, CO_2 and a_w on the diametric growth rate of F . verticillioides	69

4.5	Effect of temperature, (30, 35°C), CO ₂ levels (400, 800, 1200 ppm), and a _w (0.92, 0.95, 0.98 a _w) on the diametric growth rate of acclimatised strain of <i>F. graminearum</i> on MMA	71
4.6	Analysis of variance of the effect of temperature, CO_2 and a_w on the diametric growth rate of F . graminearum	71
4.7	Total concentration of FB ₁ , and FB ₂ produced by F. verticillioides on milled maize agar at various temperatures, a _w and CO ₂ levels after 21 days	74
4.8	Analysis of variance of the effect of temperature, CO2 and aw on the production of FB1 and FB2 by F. verticillioides	75
4.9	Total concentration of DON and ZEA produced by <i>F. graminearum</i> on milled maize agar at various temperatures a_w and CO_2 levels after 21 days	77
4.10	Analysis of variance of the effect of temperature, a_w and CO_2 on the production of DON and ZEA by F. graminearum	78
5.1	Master mix for reverse transcription per reaction	87
5.2	Concentration of RNA extracted from F. verticillioides on autoclaved maize	89
5.3	Concentration of RNA extracted from F. graminearum on autoclaved maize	89
5.4	Fumonisins B ₁ , B ₂ and B ₃ concentration (μg/kg) produced by <i>F. verticillioides</i> on germinating maize kernels at 30°C and various a _w and CO ₂ levels after 21 days	92
5.5	Analysis of variance of the effect of temperature, a_w and CO_2 on the production of FB_1 and FB_2 by the strain of F . verticillioides	93
5.6	Total concentration of DON, 3-ADON, ZEA, and α - ZEA by the acclimatised strain F . graminearum on germinating maize kernels at 30°C at various temperature, a_w and CO_2 levels after 21 days	95
5.7	Analysis of variance of the effect of temperature, a_w and CO_2 on the production of DON, 3-ADON, ZEA and α - ZEA by F . graminearum	96
5.8	Fumonisins B_1 , B_2 and B_3 concentration ($\mu g/kg$) produced by F . verticillioides on autoclaved maize kernels at 30°C at various temperature, a_w and CO_2 levels after 21 days	98
5.9	Analysis of variance of the effect of temperature, a_w and CO_2 on the production of FB_1 and FB_2 by F . verticillioides	99

5.10 Total concentration of DON, ZEA and 3-ADON (μg/kg) produced by the strain *F. graminearum* on autoclaved maize kernels at 30°C at various temperature, a_w and CO₂ levels after 21 days

101

5.11 Analysis of variance of the effect of temperature, a_w and CO_2 on the production of DON, 3-ADON and ZEA by *F. graminearum*

102



LIST OF FIGURES

Figure	e	Page
1.1	Flow chart of the study	5
2.1	Developmental stages of maize plant (vegetative and reproductive stages)	7
2.2	Malaysian maize feed importation from 1960 to 2019	9
2.3	Malaysian maize feed consumption from 1960 to 2019	9
2.4	Structures of major important Fusarium spp.	12
2.5	Morphological characteristics of Fusarium spp in situ on CLA	14
2.6	Metabolism of zearalenone in animals	17
2.7	Conversion of DON and ZEA to DON-3-Glc and ZEA-14-Glc	19
2.8	Conversion of ZEA to several metabolites	20
3.1	Macro-morphological appearance of F. verticillioides	51
3.2	Macro-morphological appearance of F. incarnatum	52
3.3	Macro-morphological appearance of F. graminearum	53
3.4	Micromorphology of <i>Fusarium</i> spp. observed on Spezieller Nahrstoffarmer Agar (SNA) in a petri dish under low power microscope 10x	54
3.5	Amplification of the TEF-1α gene (~700 bp) of the representative <i>Fusarium</i> spp. from maize and soil samples. M: 1 kb DNA ladder; C: negative control (without DNA templates)	55
3.6	Maximum likelihood phylogenetic tree of <i>Fusarium</i> spp. based on TEF - α gene sequence. Other isolates of <i>Fusarium</i> spp. from the GenBank were chosen as reference isolates for phylogenetic analysis	56
4.1	The effect of carbon dioxide (400, 800, 1200 ppm), water activity (0.92, 0.95, 0.98 a_w) and temperature (30, 35°C) levels on diametric growth rate (mm/d) of acclimatised strain of F . verticillioides cultivated on milled-maize agar for seven days	70

4.2	The effect of carbon dioxide (400, 800, 1200 ppm), water activity (0.92, 0.95, 0.98 a _w) and temperature (30, 35°C) levels on diametric growth rate (mm/d) of acclimatised strain of <i>F. graminearum</i> cultivated on milled-maize agar for seven days	72
4.3	The effect of carbon dioxide (400, 800, 1200 ppm) and water activity (0.92, 0.95, 0.98 aw) on fumonisins (FB ₁ , FB ₂) production (μ g/kg) by acclimatised strain of <i>F. verticillioides</i> cultivated on milled-maize agar for 21 days at 30°C	73
4.4	The effect of carbon dioxide (400, 800, 1200 ppm) and water activity (0.92, 0.95, 0.98 a_w) on mycotoxin (DON, ZEA) production (μ g/kg) by acclimatised strain of <i>F. graminearum</i> cultivated on milled-maize agar after 21 days at 30°C	76
5.1	Moisture adsorption curve of maize kernels	84
5.2	Flow chart of the gene expression analysis of F. verticillioides and F. graminearum	86
5.3	Growth of F. verticillioides and F. graminearum under the effect of different aw at 30°C and 400 ppm CO ₂ on autoclaved maize kernels	88
5.4	RNA gene of Fusarium spp. on gel electrophoresis.	90
5.5	The effect of CO ₂ (400, 800, 1200 ppm) and water activity (0.92, 0.95, 0.98 a _w) on fumonisins (FB ₁ , FB ₂) production (µg/kg) by acclimatised strain of <i>F. verticillioides</i> cultivated on germinating maize after 21 days at 30°C	91
5.6	The effect of carbon dioxide (400, 800, 1200 ppm) and water activity (0.92, 0.95, 0.98 a_w) on. DON, 3-ADON, ZEN and α -ZEA production ($\mu g/kg$) by acclimatised strain of F . graminearum cultivated on germinating maize after 21 days at 30°C	94
5.7	The effect of CO_2 (400, 800, 1200 ppm) and water activity (0.92, 0.95, 0.98 a_w) on fumonisins (FB ₁ , FB ₂) production (μ g/kg) by acclimatized strain of <i>F. verticillioides</i> cultivated on autoclaved maize after 21 days at 30°C	97
5.8	The effect of carbon dioxide (400, 800, 1200 ppm) and water activity (0.92, 0.95, 0.98 a _w) on DON, ZEA and α -ZEA production (μ g/kg) by acclimatized strain of <i>F. graminearum</i> cultivated on autoclaved maize after 21 days at 30°C	100

LIST OF APPENDICES

Appen	dix	Page
A	Image of maize-based media inoculated with F. verticillioides	131
В	Effect of climate change factors (Temperature, CO_2 , and a_w) on the production of parent and masked mycotoxins inoculated on maize-based media by $F.\ verticillioides$ and $F.\ graminearum$	132
С	Effect of climate change factors (Temperature, CO_2 , and a_w) on the production of parent and masked mycotoxins in germinating maize kernels by $F.$ verticillioides and $F.$ graminearum (in planta)	135
D	Effect of climate change factors(Temperature, CO ₂ , and a _w) on the production of parent and masked mycotoxins in autoclaved maize kernels by <i>F. verticillioides</i> and <i>F. graminearum</i>	138

LIST OF ABBREVIATIONS

a_w Water activity

BLAST Basic Local Alignment Search Tool

CFU Colony Forming Unit

CO₂ Carbon dioxide

DG18 Dichloran 18% glycerol agar

DON deoxynivalenol

3-ADON 3-acetyl-Deoxynivalenol

DRBC Dichloran Rose Bengal Chlorophenerin

EU European Union

FB₁ Fumonisin B₁

FB₂ Fumonisin B₂

FB₃ Fumonisin B₃

FAO Food and Agriculture Organization of the United Union

FAOSTAT FAO Statistical Databases (United Nations)

F. graminearum Fusarium graminearum

F. verticillioides Fusarium verticillioides

GMP Good Manufacturing Practice

HRMS High-resolution mass spectrometry

IARC International Agency for Research on Cancer

IPCC International Panel on Climate Changes

IF Isolation frequency

Kg Kilogram

LC-MS/MS Liquid chromatography-tandem mass spectrophotometry

LOD Limit of detection

LOQ Limit of quantification

MEA Malt Extract Agar

MeOH Methanol

ML Maximum Likelihood

mL Millilitre

MMA Milled maize agar

TEF Translation elongated factor

ULPC Ultra Performance Liquid Chromatography

WHO World Health Organization

α-ZEA Alpha - Zearalenol

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Climate change is described as a global change of weather that may occur as a result of human activities such as clearing of the forest, burning of fossil fuel and others (Paterson and Lima, 2011), causing a drastic increase in carbon dioxide (CO₂) and temperature in the atmosphere resulting to drought, precipitation and change in rainfall (Chakraborty *et al.*, 1998). The climate change scenario has become the center of attention to the scientific community due to the impact it may have on plants, mycotoxigenic fungi, and mycotoxin contamination. Several researchers have predicted that about 1.5 µmol of CO₂ and also 0.03% of temperature is expected to increase yearly (Medina *et al.*, 2017; Magan *et al.*, 2011). However, based on the presently available data, in the next 20-50 years, the concentration of atmospheric CO₂ is expected to double or tripled (from 350 - 400 to 800 - 1200 ppm) (Medina *et al.*, 2015).

Vaughan et al., (2014) reported that in the future, climate change would increase the susceptibility of crops such as maize, rice, and wheat to mycotoxigenic fungal pathogens. Based on a global basis, about 1/3 variations in the yield of crops may be as a result effect of climate change (Ray et al., 2015). In Europe, the European Food Safety (EFSA) has examined the potential impact of climate change and suggested that this could depend on geographical region, which could be detrimental or advantageous (Medina et al., 2015). Similarly, climate change is also expected to have a negative impact in some parts of Asia, Central and South America, they are important producers of various crops such as wheat, maize and soya beans.

Mycotoxigenic fungi can be affected by the impact of climate change factors. Mycotoxins produced by these fungi are toxic chemical substances of secondary metabolites (Andrade *et al.*, 2017; Rahmani *et al.*, 2009). *Aspergillus, Fusarium and Penicillium* are the most important fungal genera that can produce mycotoxins under suitable conditions (Lee and Ryu, 2017; Roseanu *et al.*, 2010). Currently, more than 400 fungal metabolites have been discovered however, not all are toxigenic (Paterson and Lima, 2011). The major fungi associated with maize is *Fusarium* spp. especially *F. verticillioides* and *F. proliferatum* (Cendoya *et al.*, 2014).

Plants are capable of protecting themselves against xenobiotic compounds such as mycotoxins by transforming them into conjugated forms (Berthiller *et al.*, 2005). These conjugated compounds are referred to as masked mycotoxins due to their ability to escape the normal routine analytical methods. Their mechanism of production could be natural or could be formed as a defense against infection by

xenobiotics (Rychlik *et al.*, 2014). Furthermore, these metabolites can be formed as a result of metabolism by plant, animal or alongside during processing of foods from contaminated raw materials.

Several researchers have reported that *Fusarium* spp. are the major species affecting cereal grains during pre and post-harvest under favorable environmental conditions such as temperature, water activity and CO₂ (Smith *et al.*,2016; Medina *et al.*, 2015). The species of *Fusarium* is a natural producer of both parent and masked mycotoxins which are mainly isolated from various food and feed products. Most of the frequently occurring *Fusarium* mycotoxins are DON, 3-ADON, 15-ADON, ZEA, NIV, DAS, HT-2 toxins, T-2 toxins, NEO (Mankevičienč *et al.*, 2011). In cereals, some of the mycotoxins produced accumulated in the kernel as a result of fungal contamination which may lead to their carryover in processed food and feed in toxicologically relevant concentrations (Duan *et al.*, 2016; Rodrigues and Naehrer, 2012). Hence, this has become a global issue.

In Malaysia, maize has been recently gazetted as one of several wealth-creating crops. The interest in maize plantation stemmed from the need to reduce the country's dependence on maize import to feed its livestock. Basically, nearly 100% maize for feed is imported from other countries which caused a heavy economic burden. Therefore, the Malaysian government under the Department of Agriculture with co-operation from Green World Genetics (GWG), has taken an initiative to cultivate Malaysian first commercialized maize. The pilot project of maize cultivation was carried out using GWG 888 (maize seed).

Therefore, there is a need to carry out extensive research on the prevalence of *Fusarium* spp., and the effect of climate change on the prevalent species at different stages in maize and their mycotoxigenic potentials.

1.2 Problem statement

Mycotoxins are toxic chemical substances contaminating a large variety of food and feed in a relevant toxicological quantity. This occurs due to suitable environmental conditions such as temperature, humidity, water activity and CO₂. Indeed, many researchers in Malaysia have reported contamination of animal feed. Malaysia is a tropical country with high humidity, temperature and rainfall therefore a high risk of mycotoxin contamination is expected to occur (Bhat *et al.*, 2010). However, *Fusarium* spp. is among the mycotoxigenic fungi that invade many varieties of agricultural products and can produce both parent and masked mycotoxins that are harmful to both human and animals.

Masked mycotoxin is of great threat to both human and animal health due to its ability to hydrolysed into their parent compound in the animal and human body, it is reabsorbed into the blood thereby increasing its exposure and toxicological properties (Berthiller *et al.*, 2009). Although, various research has been conducted on the co-occurrence of parent and masked mycotoxins in different varieties of food and feed, unfortunately, classical risk analysis does not include the presence of masked mycotoxin into account as the total mycotoxins and current regulatory limits for food and feed are based on parent mycotoxins only.

Besides, contamination of maize in the field cannot be totally overcome due to the ubiquitous nature of fungi. Several issues of *Fusarium* contamination in different product has been reported in Malaysia. Therefore, due to these reasons, extensive research focusing on the diversity, identification, toxigenic potentials and the influence of climate change on *Fusarium* spp. is needed in order to ensure the safety of both humans and animal and also to recommend a relevant way to tackle the issue of contamination by *Fusarium* spp. as a result of future climate change.

Furthermore, the gene expression by the *Fusarium* spp. must also be investigated based on the effect of climate change to understand the impact on the clustered gene of *Fusarium* spp. in maize since there are no reported kinds of literature on such study. This information is important in order to understand the behavior of *Fusarium* spp. in maize under certain climatic conditions, which in turn could be used as an actual dataset based on the previous prediction about the effect of climate change factors on mycotoxigenic fungi.

1.3 Significance of the study

The data obtained from this study will contribute to better awareness of the prevalent *Fusarium* spp. in maize. The data and information will also be useful to the Policy Makers (National Policy on Climate Change) about the current occurrence and the feature perception of *Fusarium* based on the influence of climate change, the target for intervention due to the present occurrence of *Fusarium* spp. in maize.

1.4 Objective

1.4.1 Objective of the study

Generally, this study aimed to evaluate the prevalent *Fusarium* spp. in maize and determine the effect of climate change on the production parent and masked mycotoxins by the *Fusarium* spp. in maize at different developmental stages.

1.4.2 Specific objectives

- 1. To evaluate the biodiversity of *Fusarium* spp. in maize and soil samples collected from different maize farms.
- 2. To access the effects of climate change factors (CO₂, temperature and a_w) on the production of parent mycotoxins in maize-based growth medium (*in vitro*).
- 3. To determine the effect of climate change factors (CO₂, temperature and a_w) on gene expression and production of parent and masked mycotoxins in germinating and autoclaved maize kernels by *F. verticillioides* and *F. graminearum*

1.5 Research approach

Determine the biodiversity and mycotoxigenic potentials of *Fusarium* spp. in maize and soil samples

Obj 1

Isolation and identification of *Fusarium* spp.

Morphological and molecular Identification of Fusarium spp.

Mycotoxigenic potentials of Fusarium spp.

Acclimatisation of the prevalent strain (F. verticillioides and F. graminearum by sub culturing to the 10th generation)

Obj 2

Determine the effect of climate change factors on the production of parent and masked mycotoxins produced by *F. verticillioides* and *F. graminearum in vitro*

Obj 3

Determine the effect of climate change factors on the production of parent and masked mycotoxins in germinating maize kernels and gene expression (FUM 1, TRI 5 and PKS4) by the acclimatised isolate of F. verticillioides and F. graminearum

Figure 1.1: Flow chart of the study

REFERENCES

- Afzal, I., Bakhtavar, M. A., Ishfaq, M., Sagheer, M., & Baributsa, D. (2017). Maintaining dryness during storage contributes to higher maize seed quality. *Journal of Stored Products Research*, 72, 49–53.
- Ahangarkani, F., Rouhi, S., & Azizi, I. (2014). A review on incidence and toxicity of fumonisins. *Toxin Reviews*, 33, 95–100.
- Akbar, A., Medina, A., & Magan, N. (2016). Impact of interacting climate change factors on growth and ochratoxin A production by *Aspergillus* section Circumdati and Nigri species on coffee. *World Mycotoxin Journal*, 9(5), 863-874.
- Al-Hindi, R. R., Aly, S. E., Hathout, A. S., Alharbi, M. G., Al-Masaudi, S., Al-Jaouni, S. K., & Harakeh, S. M. (2018). Isolation and molecular characterization of mycotoxigenic fungi in agarwood. *Saudi Journal of Biological Sciences*, 25, 1781–1787.
- Andrade, P. D., Dantas, R. R., de Moura, T. L. D. S., & Caldas, E. D. (2017). Determination of multi-mycotoxins in cereals and of total fumonisins in maize products using isotope-labelled internal standard and liquid chromatography/tandem mass spectrometry with positive ionization. *Journal of Chromatography A*, 1490, 138–147.
- Anfossi, L., Giovannoli, C., & Baggiani, C. (2016). Mycotoxin detection. *Current Opinion in Biotechnology*, 37, 120–126.
- Arroyo-Manzanares, N., Huertas-Pérez, J. F., García-Campaña, A. M., & Gámiz-Gracia, L. (2014). Mycotoxin analysis: New proposals for sample treatment. *Advances in Chemistry*, 2014, 1–12
- Bankole, S., Schollenberger, M., & Drochner, W. (2006). Mycotoxins in food systems in Sub Saharan Africa: A review. *Mycotoxin Research*, 22, 163–169.
- Battilani, P. & Leggieri, M. C. (2015). Predictive modelling of aflatoxin contamination to support maize chain management. *World Mycotoxin Journal*, 8, 161–170.
- Belizán, M. M., Gomez, A. D. L. A., Baptista, Z. P. T., Jimenez, C. M., Matías, M. D. H. S., Catalán, C. A., & Sampietro, D. A. (2019). Influence of water activity and temperature on growth and production of trichothecenes by *Fusarium* graminearum sensu stricto and related species in maize grains. *International Journal of Food Microbiology*, 305, 108242.
- Bernáldez, V., Rodríguez, A., Rodríguez, M., Sánchez-Montero, L., & Córdoba, J. J. (2017). Evaluation of different RNA extraction methods of filamentous fungi in various food matrices. *LWT*, 78, 47–53.

- Berthiller, F., Dall'Asta, C., Corradini, R., Marchelli, R., Sulyok, M., Krska, R., ... & Schuhmacher, R. (2009). Occurrence of deoxynivalenol and its 3-β-D-glucoside in wheat and maize. *Food Additives and Contaminants*, 26, 507–511.
- Berthiller, F, Brera, C., Iha, M. H., Krska, R., Lattanzio, V. M. T., Macdonald, S., ... & Maragos, C. (2017). Developments in mycotoxin analysis: An update for 2015-2016. *World Mycotoxin Journal*, 10, 5–29.
- Berthiller, F, Lemmens, M., Werner, U., Krska, R., Hauser, M., & Adam, G. (2007). Short review: Metabolism of the *Fusarium* mycotoxins deoxynivalenol and zearalenone in plants. *Mycotoxin Research*. 23, 68–72.
- Berthiller, F., Crews, C., Dall'Asta, C., De Saeger, S., Haesaert, G., Karlovsky, P., ... & Stroka, J. (2013). Masked mycotoxins: A review. *Molecular Nutrition and Food Research*, 57, 165–186.
- Berthiller, F., Dall'Asta, C., Schuhmacher, R., Lemmens, M., Adam, G., & Krska, A. R. (2005). Masked mycotoxins: Determination of a deoxynivalenol glucoside in artificially and naturally contaminated wheat by liquid chromatography-tandem mass spectrometry. *Journal of Agricultural and Food Chemistry*, 53, 3421–3425.
- Berthiller, F., Maragos, C. M., & Dall'Asta, C. (2015). Introduction to Masked Mycotoxins, in *Masked Mycotoxins in Food: Formation, Occurrence and Toxicological Relevance* (pp1-13) Royal society chemistry, USA.
- Berthiller, F., Schuhmacher, R., Adam, G., & Krska, R. (2009). Formation, determination and significance of masked and other conjugated mycotoxins. *Analytical and Bioanalytical Chemistry*, 395, 1243–1252.
- Berthiller, F., Schuhmacher, R., Buttinger, G., & Krska, R. (2005). Rapid simultaneous determination of major type A- and B-trichothecenes as well as zearalenone in maize by high-performance liquid chromatographytandem mass spectrometry. *Journal of Chromatography A*, 1062, 209–216.
- Berthiller, F., Sulyok, M., Krska, R., & Schuhmacher, R. (2007). Chromatographic methods for the simultaneous determination of mycotoxins and their conjugates in cereals. *International Journal of Food Microbiology*, 119, 33–37.
- Bhat, R., Rai, R. V., & Karim, A. A. (2010). Mycotoxins in food and feed: Present status and future concerns. *Comprehensive Reviews in Food Science and Food Safety*, 9, 57–81.
- Breidbach, A. (2017). A greener, quick and comprehensive extraction approach for LC-MS of multiple mycotoxins. *Toxins*, *9*, 91.

- Bryła, M., Waśkiewicz, A., Ksieniewicz-Woźniak, E., Szymczyk, K., & Jędrzejczak, R. (2018). Modified *Fusarium* mycotoxins in cereals and their products. Metabolism, occurrence, and toxicity: An updated review. *Molecules*, 23, 963–972.
- Cairns Fuller, V., Aldred, D., & Magan, N. (2005). Water, temperature and gas composition interactions affect growth and ochratoxin A production by isolates of *Penicillium verrucosum* on wheat grain. *Journal of Applied Microbiology*, 99, 1215–1221.
- Campo, S., Carrascal, M., Coca, M., Abián, J., & San Segundo, B. (2004). The defence response of germinating maize embryos against fungal infection: A proteomics approach. *Proteomics*, *4*, 383–396.
- Cendoya, E., Monge, M.P., Palacios, S.A., Chiacchiera, S.M., Torres, A.M., Farnochi, M.C., Ramirez, M.L. (2014). Fumonisin occurrence in naturally contaminated wheat grain harvested in Argentina. *Food Control* 37:56–61
- Chakraborty, S., Murray, G. M., Magarey, P. A., Yonow, T., O'Brien, R. G., Croft, B. J., ... & Sutherst, R. W. (1998). Potential impact of climate change on plant diseases of economic significance to Australia. *Australasian Plant Pathology*, 27, 15–35.
- Chulze, S. N. (2010) Strategies to reduce mycotoxin levels in maize during storage: a review. Food Addit Contam Part A Chem Anal Control Expo Risk Assess 27(5):651–657.
- Covarelli, L., Stifano, S., Beccari, G., Raggi, L., Lattanzio, V. M. T., & Albertini, E. (2012). Characterization of *Fusarium verticillioides* isolates isolated from maize in Italy: Fumonisin production, pathogenicity and genetic variability. *Food Microbiology*, 31, 17–24.
- Cuevas-Rodríguez, E. O., Verdugo-Montoya, N. M., Angulo-Bejarano, P. I., Milán-Carrillo, J., Mora-Escobedo, R., Bello-Pérez, L. A., ...& Reyes-Moreno, C. (2006). Nutritional properties of tempeh flour from quality protein maize (*Zea mays L.*). *LWT Food Science and Technology*, 39, 1072–1079.
- Da Luz, S. R., Pazdiora, P. C., Dallagnol, L. J., Dors, G. C., & Chaves, F. C. (2017). Mycotoxin and fungicide residues in wheat grains from fungicide-treated plants measured by a validated LC-MS method. *Food Chemistry*, 220, 510–516.
- Dagnac, T., Latorre, A., Fernández Lorenzo, B., & Llompart, M. (2016). Validation and application of a liquid chromatography-tandem mass spectrometry-based method for the assessment of the co-occurrence of mycotoxins in maize silages from dairy farms in NW Spain. *Food Additives & Contaminants: Part A*, 33, 1850–1863.

- De Boevre, M., Di Mavungu, J. D., Maene, P., Audenaert, K., Deforce, D., Haesaert, G., ... & De Saeger, S. (2012). Development and validation of an LC-MS/MS method for the simultaneous determination of deoxynivalenol, zearalenone, T-2-toxin and some masked metabolites in different cereals and cereal-derived food. Food Additives & Contaminants: Part A, 29, 819–835.
- De Boevre, D. M., & Saeger, D. S. (2015). Metabolism of modified mycotoxins studied through in vitro and in vivo models: An overview. *Toxicology Letters*, 233, 24–28.
- De Boevre, M., Jacxsens, L., Lachat, C., Eeckhout, M., Di Mavungu, J. D., Audenaert, K., ... De Saeger, S. (2013). Human exposure to mycotoxins and their masked forms through cereal-based foods in Belgium. *Toxicology Letters*, 218, 281–292.
- Dellafiora, L., Perotti, A., Galaverna, G., Buschini, A., & Dall'Asta, C. (2016). On the masked mycotoxin zearalenone-14-glucoside. Does the mask truly hide?. *Toxicon*, 111, 139-142.
- De Oliveira R., L., Reis, G. M., Da Silva, V. N., Braghini, R., Teixeira, M. M. G., & Corrêa, B. (2011). Molecular characterization and fumonisin production by *Fusarium verticillioides* isolated from corn grains of different geographic origins in Brazil. *International Journal of Food Microbiology*, 145, 9–21.
- Deepa, N., & Sreenivasa, M. Y. (2019). Food bioscience molecular methods and key genes targeted for the detection of fumonisin producing *Fusarium* verticillioides An updated review. *Food Bioscience*, 32, 100473.
- Desjardins, A. E. (2006). Fusarium mycotoxins: chemistry, genetics, and biology. American Phytopathological Society (APS Press).
- Duan, C., Qin, Z., Yang, Z., Li, W., Sun, S., Zhu, Z., & Wang, X. (2016). Identification of pathogenic *Fusarium* spp. causing maize ear rot and potential mycotoxin production in China. *Toxins*, 8, 186.
- Eckard, S., Wettstein, F. E., Forrer, H. R., & Vogelgsang, S. (2011). Incidence of *Fusarium* species and mycotoxins in silage maize. *Toxins*, 3, 949–967.
- EFSA Panel on Contaminants in the Food Chain (CONTAM). (2013). Scientific Opinion on the risk for public and animal health related to the presence of sterigmatocystin in food and feed. *EFSA Journal*, 11, 3254.
- Engelhardt, G., Ruhland, M., & Wallnöfer, P. R. (1999). Metabolism of mycotoxins in plants. *Advance in Food Science*, 21, 71–78.
- Engelhardt, G., Zill, G., Wohner, B., & Wallnöfer, P. R. (1988). Transformation of the *Fusarium* mycotoxin zearalenone in maize cell suspension cultures. *Naturwissenschaften*, 75, 309–310.

- Erenstein, O. (2010). The evolving maize sector in Asia: Challenges and opportunities. *Journal of New Seeds*, 11, 1–15.
- Etcheverry, M., Torres, A., Ramirez, M. L., Chulze, S., & Magan, N. (2001). *In vitro* control of growth and fumonisin production by *Fusarium verticillioides* and *F. proliferatum* using antioxidants under different water availability and temperature regimes. *Journal of Applied Microbiology*, 92, 624–632.
- Faltusová, Z., Chrpová, J., Salačová, L., Džuman, Z., Pavel, J., Zachariášová, M., ... & Ovesná, J. (2015). Effect of *Fusarium culmorum* Tri gene transcription on deoxynivalenol and D3G levels in two different barley cultivars. *Journal of Phytopathology*, *163*, 593–603.
- Foroud, N. A., & Eudes, F. (2009). Trichothecenes in cereal grains. *International Journal of Molecular Sciences*, 10, 147–173.
- Frandsen, T. P., Lok, F., Mirgorodskaya, E., Roepstorff, P., & Svensson, B. (2000). Purification, enzymatic characterization, and nucleotide sequence of a high- isoelectric-point alpha-glucosidase from barley malt. *Plant Physiology*, 123, 275–286.
- Galaverna, G., Dallsta, C., Mangia, M. A., Dossena, A., & Marchelli, R. (2009). Masked mycotoxins: An emerging issue for food safety. *Czech Journal of Food Sciences*, 27, 89–92.
- Gao, Z., Feng, H. Y., Liang, X. G., Zhang, L., Lin, S., Zhao, X., ... & Zhou, S. L. (2018). Limits to maize productivity in the North China Plain: A comparison analysis for spring and summer maize. *Field Crops Research*, 228, 39–47.
- Gareis, M., Bauer, J., Thiem, J., Plank, G., Grabley, S., & Gedek, B. (1990). Cleavage of zearalenone-glycoside, a "masked" mycotoxin, during digestion in swine. *Journal of Veterinary Medicine, Series B*, *37*, 236–240.
- Geary, P. A., Chen, G., Kimanya, M. E., Shirima, C. P., Oplatowska-Stachowiak, M., Elliott, C. T., ...& Yun, Y. (2016). Determination of multi-mycotoxin occurrence in maize-based porridges from selected regions of Tanzania by liquid chromatography-tandem mass spectrometry (LC-MS / MS), a longitudinal study. *Food Control*, 68, 337–343.
- Geiser, D. M., del Mar Jiménez-Gasco, M., Kang, S., Makalowska, I., Veeraraghavan, N., Ward, T. J., ... & O'Donnell, K. (2004). FUSARIUM-ID v. 1.0: a DNA sequence database for identifying *Fusarium*. *European Journal of Plant Pathology*, 110, 473–479.
- Gil-Serna, J., Vázquez, C., & Patiño, B. (2019). Genetic regulation of aflatoxin, ochratoxin A, trichothecene, and fumonisin biosynthesis: A review. *International Microbiology*, 1, 8–16

- Gong, F., Wu, X., Zhang, H., Chen, Y., & Wang, W. (2015). Making better maize plants for sustainable grain production in a changing climate. *Frontiers in Plant Science*, 6, 1–6.
- Goswami, R. S., & Kistler, H. C. (2005). Pathogenicity and *in planta* mycotoxin accumulation among members of the *Fusarium graminearum* species complex on wheat and rice. *Phytopathology*, 95, 1397–1404.
- Gratz, S. W. (2017). Do plant-bound masked mycotoxins contribute to toxicity? *Toxins*, 9, 1–10.
- Gruber-Dorninger, C., Jenkins, T., & Schatzmayr, G. (2019). Global mycotoxin occurrence in feed: A ten-year survey. *Toxins*, 11(7), 375.
- Gupta, R. K. (2017). Foodborne infectious diseases. In *Food Safety in the 21st Century* (pp. 13–28). Academic Press.
- Hansen, F. T., Sørensen, J. L., Giese, H., Sondergaard, T. E., & Frandsen, R. J. (2012). Quick guide to polyketide synthase and nonribosomal synthetase genes in *Fusarium*. *International Journal of Food Microbiology*, 155, 128– 136.
- Haschek, W. M., Gumprecht, L. A., Smith, G., Tumbleson, M. E., & Constable, P. D. (2001). Fumonisin toxicosis in swine: An overview of porcine pulmonary oedema and current perspectives. *Environmental health perspectives*, 109, 251–257.
- Hassan, Z. U., Al-Thani, R. F., Migheli, Q., & Jaoua, S. (2018). Detection of toxigenic mycobiota and mycotoxins in cereal feed market. *Food Control*, 84, 389–394.
- He, J., Zhou, T., Young, J. C., Boland, G. J., & Scott, P. M. (2010). Chemical and biological transformations for detoxification of trichothecene mycotoxins in human and animal food chains: A review. *Trends in Food Science and Technology*, 21, 67–76.
- Hope, R., Aldred, D., & Magan, N. (2005). Comparison of environmental profiles for growth and deoxynivalenol production by *Fusarium culmorum* and *F. graminearum* on wheat grain. *Letters of Applied Microbiology*, 40, 295–300.
- Hsuan, H. M., Salleh, B., & Zakaria, L. (2011). Molecular identification of *Fusarium* species in *Gibberella fujikuroi* species complex from rice, sugarcane and maize from Peninsular Malaysia. *International Journal of Molecular Sciences*, 12, 6722–6732.
- Hu, X., Hu, R., Zhang, Z., Li, P., Zhang, Q., & Wang, M. (2016). Development of a multiple immunoaffinity column for simultaneous determination of multiple mycotoxins in feeds using UPLC–MS/MS. *Analytical and Bioanalytical Chemistry*, 408, 6027–6036.

- IPCC., A. R. (2007). Intergovernmental panel on climate change. *Climate change* 2007: *Synthesis report*. Geneva, Switzerland
- IPCC, C. C. (2014). Mitigation of climate change. Contribution of working group III to the fifth assessment report of the intergovernmental panel on climate change.
- Jackson, L., & Jablonski, J. (2004). Fumonisins. In *Mycotoxins in food*, (pp36–405). Woodhead Publishing.
- Jacobs, A., Mojela, L., Summerell, B., & Venter, E. (2018). Characterisation of members of the *Fusarium incarnatum-equiseti* species complex from undisturbed soils in South Africa. *Antonie van Leeuwenhoek*, 111, 1999– 2008.
- James, A., & Zikankuba, V. L. (2018). Mycotoxins contamination in maize alarms food safety in sub-Sahara Africa. *Food Control*, 90, 372–381.
- Jestoi, M., Ritieni, A., & Rizzo, A. (2004). Analysis of the *Fusarium* mycotoxins fusaproliferin and trichothecenes in grains using gas chromatography-mass spectrometry. *Journal of Agriculture and Food Chemistry*, 52, 1464–1469.
- Johnson, E. T., Proctor, R. H., Dunlap, C. A., & Busman, M. (2018). Reducing production of fumonisin mycotoxins in *Fusarium verticillioides* by RNA interference. *Mycotoxin Research*, 34, 29-37.
- Jones, T. J. (2009). Maize tissue culture and transformation: the first 20 years. In *Molecular Genetic Approaches to Maize Improvement* (pp. 7–27). Springer, Berlin, Heidelberg.
- Junior, R., Ferreira, O., Dalein, M. S., Nascimento, V. L., Haesbaert, F. M., Ferreira, T. P. D. S., ... & Santos, G. R. D. (2019). Fumonisin production by *Fusarium verticillioides* in maize genotypes cultivated in different environments. *Toxins*, 11, 215–220
- Kafouris, D., Christofidou, M., Christodoulou, M., Christou, E., & Ioannou-Kakouri, E. (2017). A validated UPLC-MS/MS multi-mycotoxin method for nuts and cereals: Results of the official control in Cyprus within the EU requirements. *Food and Agricultural Immunology*, 28, 90–108.
- Kamala, A., Ortiz, J., Kimanya, M., Haesaert, G., Donoso, S., Tiisekwa, B., & De Meulenaer, B. (2015). Multiple mycotoxin co-occurrence in maize grown in three agro-ecological zones of Tanzania. *Food Control*, *54*, 208–215.
- Keller, N. P., Butchko, R.A.E., Sarr, B & Phillips, T. D. (1994). A visual patterns of mycotoxin production in maize kernels by Aspergillus spp. *Phytopathology* 84: 483–488
- Knutsen, H. K., Alexander, J., Barregård, L., Bignami, M., Brüschweiler, B., ... & Grasl-Kraupp, B. (2017). Risks for animal health related to the presence of zearalenone and its modified forms in feed. *EFSA Journal*, *15*(7), e04851.

- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution*, *33*, 1870–1874.
- Lahouar, A., Crespo-Sempere, A., Marín, S., Saïd, S., & Sanchis, V. (2015). Toxigenic moulds in Tunisian and Egyptian sorghum for human consumption. *Journal of Stored Products Research*, 63, 57–62.
- Lahouar, A., Marin, S., Crespo-Sempere, A., Saïd, S., & Sanchis, V. (2016). Effects of temperature, water activity and incubation time on fungal growth and aflatoxin B₁ production by toxinogenic *Aspergillus flavus* isolates on sorghum seeds. *Revista Argentina de Microbiologia*, 48, 78–85.
- Lahouar, A., Marin, S., Crespo-Sempere, A., Saïd, S., & Sanchis, V. (2017). Influence of temperature, water activity and incubation time on fungal growth and production of ochratoxin A and zearalenone by toxigenic Aspergillus tubingensis and Fusarium incarnatum isolates in sorghum seeds. International Journal of Food Microbiology, 242, 53–60.
- Lancova, K., Hajslova, J., Poustka, J., Krplova, A., Zachariasova, M., Dostálek, P., & Sachambula, L. (2008). Transfer of *Fusarium* mycotoxins and 'masked' deoxynivalenol (deoxynivalenol-3-glucoside) from field barley through malt to beer. *Food Additives and Contaminants*, 25, 732–744.
- Latiffah, Z., Padzilah, M. I., Baharuddin, S., & Maziah, Z. (2009). Fusarium species in forest soil of Bird Valley. Malaysian Journal of Microbiology, 5, 132–133.
- Lattanzio, V. M. T., Pascale, M., & Visconti, A. (2009). Current analytical methods for trichothecene mycotoxins in cereals. *Trends in Analytical Chemistry*, 28, 758–768.
- Lauber, C. L., Strickland, M. S., Bradford, M. A., & Fierer, N. (2008). The influence of soil properties on the structure of bacterial and fungal communities across land-use types. Soil Biology & Biochemistry 40, 2407— 2415.
- Lauer, J. (1997). Healthy corn growth and development in Wisconsin. *Field Crops*, 28, 1–16.
- Lazzaro, I., Susca, A., Mulè, G., Ritieni, A., Ferracane, R., Marocco, A., & Battilani, P. (2012). Effects of temperature and water activity on *FUM2* and *FUM21* gene expression and fumonisin B production in *Fusarium* verticillioides. European Journal of Plant Pathology, 134, 685–695.
- Lee, H. J., & Ryu, D. (2017). Worldwide occurrence of mycotoxins in cereals and cereal-derived food products: Public health perspectives of their co-occurrence. *Journal of Agricultural and Food Chemistry*, 65, 7034–7051.

- Leggieri, C. M., Giorni, P., Pietri, A., & Battilani, P. (2019). *Aspergillus flavus* and *Fusarium verticillioides* interaction: modelling the impact on mycotoxin production. *Frontiers in Microbiology*, *10*, 2653.
- Leong, P., Yong, Y.C., Shin, K.H., How, S.P., Lim, H.C., & Lim, B. K. (2014). An update of mycotoxins contamination in feed ingredients in Malaysia from 2012 to 2013. *Veterinary Research.*, *5*, 122–126.
- Leong, Y. H., Rosma, A., Latiff, A. A., & Ahmad, N. I. (2011). Exposure assessment and risk characterization of aflatoxin B₁ in Malaysia. *Mycotoxin Research*, 27, 207–214.
- Leslie, J. F., & Summerell, B. A. (2006). *The Fusarium laboratory manual* (First edition). Ames, Iowa, Avenue, USA.: Blackwell Publishing.
- Li, X., Liu, B., Wang, F., Ma, X., Li, Z., Guo, D., ...& Zhang, S. (2017). Determination of 16 mycotoxins in maize by ultrahigh-performance liquid chromatography-tandem mass spectrometry. *Analytical Letters*, 51, 702–716
- Liao, C. D., Wong, J. W., Zhang, K., Hayward, D. G., Lee, N. S., & Trucksess, M. W. (2013). Multi-mycotoxin analysis of finished grain and nut products using high-performance liquid chromatography-triple-quadrupole mass spectrometry. *Journal of Agricultural and Food Chemistry*, 61, 4771–4782.
- Lippolis, V., Porricelli, A.C., Mancini, E., Ciasca, B., Lattanzio, V.M., De Girolamo, A.,& Pascale M. (2019). Fluorescence polarization immunoassay for the determination of T-2 and HT-2 toxins and their glucosides in wheat. *Toxins.*, 11, 380–388
- Logrieco, A., Moretti, A., Perrone, G., & Mulè, G. (2007). Biodiversity of complexes of mycotoxigenic fungal species associated with *Fusarium* ear rot of maize and *Aspergillus* rot of grape. *International Journal of Food Microbiology*, 119, 11–16.
- López-errasquín, E., Vázquez, C., Jiménez, M., & González-jaén, M. T. (2007). Real-Time RT-PCR assay to quantify the expression of *FUM1* and *FUM19* genes from the fumonisin-producing *Fusarium verticillioides*. *Journal of Microbiological Methods*, 68, 312–317.
- Lu, X., Liu, J., Ren, W., Yang, Q., Chai, Z., Chen, R., ...& Zhang, C. (2018). Gene-indexed mutations in maize. *Molecular Plant*, 11, 496–504.
- Luck, J., Spackman, M., Freeman, A., Tre bicki, P., Griffiths, W., Finlay, K., & Chakraborty, S. (2011). Climate change and diseases of food crops. *Plant Pathology.*, 60, 113–121.
- Madania, A., Altawil, M., Naffaa, W., Volker, P. H., & Hawat, M. (2013). Morphological and molecular characterization of *Fusarium* isolated from maize in Syria. *Journal of Phytopathology*, *161*, 452–458.

- Madgwick, J. W., West, J. S., White, R. P., Semenov, M. A., Townsend, J. A., Turner, J. A., & Fitt, B. D. (2011). Impacts of climate change on wheat anthesis and *Fusarium* ear blight in the UK. *European Journal of Plant Pathology*, 130, 117–131.
- Magan, N., & Aldred, D. (2007). Post-harvest control strategies: minimizing mycotoxins in the food chain. *International journal of food microbiology*, 119(1-2), 131-139
- Magan, N., Aldred, D., Mylona, K., & Lambert, R. J. (2010). Limiting mycotoxins in stored wheat. *Food Additives and Contaminants*, 27, 644–650.
- Magan, N., Hope, R., Cairns, V., & Aldred, D. (2003). Post-harvest fungal ecology: Impact of fungal growth and mycotoxin accumulation in stored grain. *European Journal of Plant Pathology*, 109, 723–730.
- Magan, N., Medina, A., & Aldred, D. (2011). Possible climate change effects on mycotoxin contamination of food crops pre-and postharvest. *Plant Pathology*, 60, 150–163.
- Majeed, S., De Boevre, M., De Saeger, S., Rauf, W., Tawab, A., Fazal-e-Habib, ... & Iqbal, M. (2018). Multiple mycotoxins in rice: Occurrence and health risk assessment in children and adults of Punjab, Pakistan. *Toxins*, 10, 77.
- Malekinejad, H., Maas-Bakker, R., & Fink-Gremmels, J. (2006). Species differences in the hepatic biotransformation of zearalenone. *Veterinary Journal*, 172, 96–102.
- Manizan, A. L., Oplatowska-Stachowiak, M., Piro-Metayer, I., Campbell, K., Koffi-Nevry, R., Elliott, C.,& Brabet, C. (2018). Multi-mycotoxin determination in rice, maize and peanut products most consumed in Côte d'Ivoire by UHPLC-MS/MS. *Food Control*, 87, 22–30.
- Mankevičienč, A., Butkutč, B., Gaurilčikienč, I., Dabkevičius, Z., & Supronienč, S. (2011). Risk assessment of *Fusarium* mycotoxins in Lithuanian small cereal grains. *Food Control*, 22, 970–976.
- Manstretta, V., & Rossi, V. (2016). Effects of temperature and moisture on development of *Fusarium graminearum* perithecia in maize stalk residues. *Applied and Environmental Microbiology*, 82:184–191
- Maragos, C. M. (2004). Emerging technologies for mycotoxin detection. *Journal of Toxicology: Toxin Reviews*, 23, 317–344.
- Marasas, W.F., Rheeder, J.P., Lamprecht, S.C., Zeller, K.A., & Leslie, F. J. (2001). Fusarium andiyazi spp. nov., a new species from sorghum. Mycologia, 93, 1203–1210.

- Marín, P., Jurado, M., Magan, N., Vázquez, C., & González-Jaén, M. T. (2010). Effect of solute stress and temperature on growth rate and TRI5 gene expression using real time RT-PCR in *Fusarium graminearum* from Spanish wheat. *International Journal of Food Microbiology*, 140, 169–174.
- Marín, P., Magan, N., Vázquez, C., & González-Jaén, M. T. (2010). Differential effect of environmental conditions on the growth and regulation of the fumonisin biosynthetic gene *FUM1* in the maize pathogens and fumonisin producers *Fusarium verticillioides* and *Fusarium proliferatum*. *FEMS Microbiology Ecology*, 73, 303–311.
- Marin, S., Magan, N., Belli, N., Ramos, A. J., Canela, R., & Sanchis, V. (1999). Two-dimensional profiles of fumonisin B1 production by *Fusarium moniliforme* and *Fusarium proliferatum* in relation to environmental factors and potential for modelling toxin formation in maize grain. *International Journal of Food Microbiology*, 51, 159–167.
- Marin, S., Ramos, A. J., Cano-Sancho, G., & Sanchis, V. (2013). Mycotoxins: Occurrence, toxicology, and exposure assessment. *Food and Chemical Toxicology*, 60, 218–237.
- Marin, Sonia, Sanchis, V., & Magan, N. (1995). Water activity, temperature, and pH effects on growth of *Fusarium moniliforme* and *Fusarium proliferatum* isolates from maize. *Canadian Journal of Microbiology*, 41, 1063–1070.
- Marroquín-Cardona, A. G., Johnson, N. M., Phillips, T. D., & Hayes, A. W. (2014). Mycotoxins in a changing global environment-A review. *Food and Chemical Toxicology*, 69, 220–230.
- Maul, R., Müller, C., Rieß, S., Koch, M., Methner, F., & Irene, N. (2012). Germination induces the glucosylation of the *Fusarium* mycotoxin deoxynivalenol in various grains. *Food Chemistry*, 131, 274–279.
- Medema, M. H., Cimermancic, P., Sali, A., Takano, E.,& Fischbach, M. A. (2014). A systematic computational analysis of biosynthetic gene cluster evolution: Lessons for engineering biosynthesis. *PLoS Computational Biology*, 10, e1004016.
- Medina, A., Akbar, A., Baazeem, A., Rodriguez, A., & Magan, N. (2017). Climate change, food security and mycotoxins: Do we know enough? *Fungal Biology Reviews*, 31, 143–154.
- Medina, A., Gilbert, M. K., Mack, B. M., OBrian, G. R., Rodríguez, A., Bhatnagar,& Magan, N. (2017). Interactions between water activity and temperature on the *Aspergillus flavus* transcriptome and aflatoxin B₁ production. *International Journal of Food Microbiology*, 256, 36–44.
- Medina, Á., González-Jartín, J. M., & Sainz, M. J. (2017). Impact of global warming on mycotoxins. *Current Opinion in Food Science*, 18, 76–81.

- Medina, A., Rodriguez, A., & Magan, N. (2014). Effect of climate change on Aspergillus flavus and aflatoxin B1 production. *Frontiers in Microbiology*, 5, 348.
- Medina, A., Rodríguez, A., Sultan, Y., & Magan, N. (2015). Climate change factors and *Aspergillus flavus*: Effects on gene expression, growth and aflatoxin production. *World Mycotoxin Journal*, 8, 171–179.
- Medina, A., Rodríguez, A., & Magan, N. (2015). Climate change and mycotoxigenic fungi: Impacts on mycotoxin production. *Current Opinion in Food Science*, *5*, 99–104.
- Medina, A., Schmidt-Heydt, M., Cárdenas-Chávez, D. L., Parra, R., Geisen, R., & Magan, N. (2013). Integrating toxin gene expression, growth and fumonisin B₁ and B₂ production by a strain of *Fusarium verticillioides* under different environmental factors. *Journal of the Royal Society Interface*, 10, 20130320
- Miller, J. D., Young, J. C., & Trenholm, H. L. (1983). *Fusarium* toxins in field corn. I. Time course of fungal growth and production of deoxynivalenol and other mycotoxins. *Canadian Journal of Botany*, 61, 3080–3087.
- Minervini, F., & Dell'Aquila, M. E. (2008). Zearalenone and reproductive function in farm animals. *International Journal of Molecular Sciences*, 9, 2570–2584.
- Miraglia, M., Marvin, H. J. P., Kleter, G. A., Battilani, P., Brera, C., Coni, E., ... & Filippi, L. (2009). Climate change and food safety: an emerging issue with special focus on Europe. *Food and Chemical Toxicology*, 47, 1009–1021.
- Moretti, A. N. (2009). Taxonomy of *Fusarium* genus, a continuous fight between lumpers and splitters. *Zbornik Matice srpske za prirodne nauke*,, 117, 7–13.
- Nahle, S., El Khoury, A., & Atoui, A. (2020). Current status on the molecular biology of zearalenone: its biosynthesis and molecular detection of zearalenone producing *Fusarium* species. *European Journal of Plant Pathology*, 1-12.
- Nathanail, A. V., Varga, E., Meng-Reiterer, J., Bueschl, C., Michlmayr, H., Malachova, A.,& Berthiller, F. (2015). Metabolism of the *Fusarium* mycotoxins T-2 toxin and HT-2 toxin in Wheat. *Journal of Agricultural* and Food Chemistry, 63, 7862–7872.
- Nicholson, P., Chandler, E., Draeger, R. C., Gosman, N. E., Simpson, D. R., Thomsett, M. (2003). Molecular tools to study epidemiology and toxicology of *Fusarium* head blight of cereals. *European Journal of Plant Pathology*, 109, 691–703.
- Nicolaisen, M., Kærgaard, L., Lazzaro, I., Henrik, N., & Fejer, A. (2009). Realtime PCR for quantification of eleven individual *Fusarium* species in cereals. *Journal of Microbiological Methods*, 76, 234–240.

- Njumbe E, E., Van Poucke, C., & De Saeger, S. (2015). A multi-analyte LC-MS/MS method for the analysis of 23 mycotoxins in different Sorghum varieties: The forgotten sample matrix. *Food Chemistry*, 177, 397–404.
- Nuss, E.T., & Tanumihardjo, S. A. (2010). Maize: A paramount staple crop in the context of global nutrition. *Comprehensive Reviews in Food Science and Food Safety*, 9, 417–436.
- O'Donnell, K., Cigelnik, E., & Nirenberg, H. I. (1998). Molecular systematics and phylogeography of the *Gibberella fujikuroi* species complex. *Mycologia*, 90, 465–493.
- Omar, N. H., Mohd, M., Nor, N. M. I. M., & Zakaria, L. (2018). Characterization and pathogenicity of *Fusarium* species associated with leaf spot of mango (*Mangifera indica* L.). *Microbial pathogenesis*, 114, 362-368.
- Orsi, R. B., Corrêa, B., Possi, C. R., Schammass, E. A., Nogueira, J. R., Dias, S. M., & Malozzi, M. A. (2000). Mycoflora and occurrence of fumonisins in freshly harvested and stored hybrid maize. *Journal of Stored Products Research*, 36, 75–87.
- Ort, D.R., & Long, S. (2014). Limits on yields in the corn belt. *Science*, 344, 484–485.
- Panwar, V., Aggarwal, A., Paul, S., Singh, V., Singh, P. K., Sharma, D., & Saharan, M. S. (2016) Effect of temperature and pH on the growth of *Fusarium* spp. causing *Fusarium* head blight (FHB) in wheat. *South Asian Journal of Experimental Biology*, 6:186–193
- Pardo, E., Marín, S., Solsona, A., Sanchis, V., & Ramos, A. J. (2004). Modelling of germination and growth of ochratoxigenic isolates of *Aspergillus ochraceus* as affected by water activity and temperature on a barley-based medium. *Food Microbiology*, 21, 267–274.
- Pascari, X., Ramos, A. J., Marín, S., & Sanchís, V. (2018). Mycotoxins and beer. Impact of beer production process on mycotoxin contamination. A review. *Food Research International*, 103, 121–129.
- Pasquali, M., Beyer, M., Logrieco, A., Audenaert, K., Balmas, V., Basler, R., Boutigny, A.L. (2016). A European database of *Fusarium graminearum* and *F. culmorum* trichothecene genotypes. *Frontiers in microbiology*, 7:article no. 406.
- Paterson, R. R.M., & Lima, N. (2011). Further mycotoxin effects from climate change. *Food Research International*, 44, 2555–2566.
- Paterson, R. R. M., & Lima, N. (2010). How will climate change affect mycotoxins in food? *Food Research International*, 43, 1902–1914.

- Pazderů, K., Vepříková, Z., Capouchová, I., Konvalina, P., & Prokinová, E. (2016). Changes in the content of various *Fusarium* mycotoxins forms in germinating winter wheat and spring barley kernels. *Plant Soil Environment*, 62, 42–46.
- Pena, G. A., Cavaglieri, L. R., & Chulze, S. N. (2019). Fusarium species and moniliformin occurrence in sorghum grains used as ingredient for animal feed in Argentina. Journal of the Science of Food and Agriculture, 99(1), 47-54
- Peng, W. X., Marchal, J. L. M., & Van der Poel, A. F. B. (2018). Strategies to prevent and reduce mycotoxins for compound feed manufacturing. *Animal Feed Science and Technology*, 237, 129-153.
- Pereyra, C. M., Ren, L., Chiacchiera, S. M., & Mar, A. (2010). Fungi and mycotoxins in feed intended for sows at different reproductive stages in Argentina. *Veterinary Medicine International*, 2010, 7.
- Pesek, J. J., Matyska, M. T., Hoffmann, J. F., Madruga, N. A., Crizel, R. L., Elias, M. C.,& Chaves, F. C. (2017). Liquid chromatography with mass spectrometry analysis of mycotoxins in food samples using silica hydride based stationary phases. *Journal of Separation Science*, 40, 1953–1959.
- Pfeiffer, E., Kommer, A., Dempe, J. S., Hildebrand, A. A., & Metzler, M. (2011). Absorption and metabolism of the mycotoxin zearalenone and the growth promotor zeranol in Caco-2 cells *in vitro*. *Molecular Nutrition and Food Research*, 55, 560–567.
- Picot, A., Hourcade-Marcolla, D., Barreau, C., Pinson-Gadais, L., Caron, D., & Richard-Forget, F. L. C. (2012). Interactions between *Fusarium verticillioides* and *Fusarium graminearum* in maize ears and consequences for fungal development and mycotoxin accumulation. *Plant Pathology.*, 61, 140-51.
- Pinotti, L., Ottoboni, M., Giromini, C., Orto, V.D., & Cheli, F. (2016). Mycotoxin contamination in the EU feed supply chain: A focus on cereal by-products. *Toxins*, 8, 45.
- Pinton, P., & Oswald, I. P. (2014). Effect of deoxynivalenol and other type B trichothecenes on the intestine: A review. *Toxins*, 6, 1615–1643.
- Popovski, S., & Celar, F. A. (2013). The impact of environmental factors on the infection of cereals with *Fusarium* species and mycotoxin production: A review. *Acta Agriculturae Slovenica 101*, 105–116.
- Prasanna, B. M., Pixley, K., Warburton, M. L., & Xie, C. X. (2010). Molecular marker-assisted breeding options for maize improvement in Asia. *Molecular Breeding*, 26, 339-356.

- Prassana, B. M., Vasal, S. K., Kassahun, B., & Singh, N. N. (2001). Quality protein maize. *Current Science*, 81, 1308–1319.
- Proctor, R.H., Brown, D.W., Plattner, R.D., & Desjardins, A. E. (2003). Co-expression of 15 contiguous genes delineates a fumonisin biosynthetic gene cluster in *Gibberella moniliformis*. Fungal Genetics and Biology, 38, 237–249.
- Proctor, R. H., Busman, M., Seo, J. A., Lee, Y. W., & Plattner, R. D. (2008). A fumonisin biosynthetic gene cluster in *Fusarium oxysporum* strain O-1890 and the genetic basis for B versus C fumonisin production. *Fungal Genetics and Biology*, 45, 1016–1026.
- Proctor, R. H., Plattner, R. D., Brown, D. W., Seo, E. O. J., & Yin-Won, L. E. E. (2004). Discontinuous distribution of fumonisin biosynthetic genes in the *Gibberella fujikuroi* species complex. *Mycological Research*, 1108, 815–822.
- Rahmani, A., Jinap, S., & Soleimany, F. (2009). Qualitative and quantitative analysis of mycotoxins. *Comprehensive Reviews in Food Science and Food Safety*, 8, 202–251.
- Rai, A., Das, M., & Tripathi, A. (2019). Occurrence and toxicity of a Fusarium mycotoxin, zearalenone. Critical Reviews in Food Science and Nutrition., 23, 1–20.
- Ramdial, H., Hosein, F., & Rampersad, S. (2016). First report of *Fusarium incarnatum* associated with fruit disease of bell peppers in Trinidad. *Plant Disease*, 100, 526–529
- Ray, D. K., Gerber, J. S., Macdonald, G. K., & West, P. C. (2015). Climate variation explains a third of global crop yield variability. *Nature Communications*, 6, 1–9.
- Reddy, K. R. N., & Salleh, B. (2011). Co-occurrence of moulds and mycotoxins in corn grains used for animal feeds in Malaysia. *Journal of Animal and Veterinary Advances*, 10, 668–673.
- Ren, G., Hu, Y., Zhang, J., Zou, L., & Zhao, G. (2018). Determination of multiclass mycotoxins in Tartary buckwheat by ultra-fast liquid chromatography coupled with triple quadrupole mass spectrometry. *Toxins*, 10, 28.
- Rheeder, J. P., Marasas, W. F., & Vismer, H. F. (2002). Production of fumonisin analogues by *Fusarium* species. *Applied and Environmental Microbiology*, 68, 2101–2105.
- Rodrigues, I., & Naehrer, K. (2012). A Three-year survey on the worldwide occurrence of mycotoxins in feedstuffs and feed. *Toxins*, 4, 663–675.

- Romera, D., Mateo, E. M., Mateo-Castro, R., Gómez, J. V., Gimeno-Adelantado, J. V., & Jiménez, M. (2018). Determination of multiple mycotoxins in feedstuffs by combined use of UPLC-MS/MS and UPLC-QTOF-MS. Food Chemistry, 267, 140–148.
- Roseanu, A., Jecu, L., Badea, M., & Evans, R. W. (2010). Mycotoxins: An overview of their quantification methods. *Romanian Journal of Biochemistry*, 47, 79–86.
- Rousk, J., Bååth, E., Brookes, P. C., Lauber, C. L., Lozupone, C., Caporaso, J. G., ... & Fierer, N. (2010). Soil bacterial and fungal communities across a pH gradient in an arable soil. *The ISME Journal*, *4*, 1340–1351.
- Rubert, J., Dzuman, Z., Vaclavikova, M., Zachariasova, M., Soler, C., & Hajslova, J. (2012). Analysis of mycotoxins in barley using ultra-high liquid chromatography high-resolution mass spectrometry: comparison of efficiency and efficacy of different extraction procedures. *Talanta*, *99*, 712–719.
- Ruhland, M., Engelhardt, G., Schäfer, W., & Wallnöfer, P. R. (1996). Transformation of the mycotoxin ochratoxin A in plants: Isolation and identification of metabolites formed in cell suspension cultures of wheat and maize. *Natural Toxins*, 4, 254–260.
- Rychlik, M., Humpf, H. U., Marko, D., Dänicke, S., Mally, A., Berthiller, F., ... & Lorenz, N. (2014). Proposal for a comprehensive definition of modified and other forms of mycotoxins including "masked" mycotoxins. *Mycotoxin Research*, 30, 197–205.
- Saleemi, M. K., Khan, M. Z., Khan, A., Hameed, M. R., Khatoon, A., Abadin, Z. U., & Hassan, Z. U. (2017). Study of fungi and their toxigenic potential isolated from wheat and wheat bran. *Toxin Reviews*, 36, 80–88.
- Samapundo, S., De Meulenaer, B., Atukwase, A., Debevere, J., & Devlieghere, F. (2007). The influence of modified atmospheres and their interaction with water activity on the radial growth and fumonisin B₁ production of Fusarium verticillioides and F. proliferatum on corn. Part I: The effect of initial headspace carbon dioxide concentration. International Journal of Food Microbiology, 114, 160–167.
- Samapundo, S., Devlieghere, F., De Meulenaer, B., Geeraerd, A. H., Van Impe, J. F., & Debevere, J. M. (2005). Predictive modelling of the individual and combined effect of water activity and temperature on the radial growth of *Fusarium verticillioides* and *F. proliferatum* on corn. *International Journal of Food Microbiology*, 105, 35–52.
- Samsudin, N. I. P., & Magan, N. (2016). Efficacy of potential biocontrol agents for control of *Fusarium verticillioides* and fumonisin B₁ under different environmental conditions. *World Mycotoxin Journal*, 9, 205–213.

- Senghor, A., Dioh, R. M., Müller, C., & Youm, I. (2017). Cereal crops for biogas production: A review of possible impact of elevated CO₂. *Renewable and Sustainable Energy Reviews*, 1, 548–54.
- Sewald, N., Von Gleissenthall, J. L., Schuster, M., Müller, G., & Aplin, R. T. (1992). Structure elucidation of a plant metabolite of 4-deoxynivalenol. *Tetrahedron: Asymmetry*, *3*, 953–960.
- Shah, T. R., Prasad, K., & Kumar, P. (2016). Maize-A potential source of human nutrition and health: A review. *Cogent Food & Agriculture*, 2(1), 1166995.
- Sharma, A., Khan, R., Catanante, G., Sherazi, T. A., Bhand, S., Hayat, A., & Marty, J. L. (2018). Designed strategies for fluorescence-based biosensors for the detection of mycotoxins. *Toxins*, *10*, 197.
- Shi, W., Tan, Y., Wang, S., Gardiner, D. M., De Saeger, S., Liao, Y.,& Wu, A. (2017). Mycotoxigenic potentials of *Fusarium* species in various culture matrices revealed by mycotoxin profiling. *Toxins*, 9, 6.
- Shwab, E. K., & Keller, N. P. (2008). Regulation of secondary metabolite production in filamentous ascomycetes. *Mycological Research*, 112, 225–230.
- Smith, D. S., & Eremin, S. A. (2008). Fluorescence polarization immunoassays and related methods for simple, high-throughput screening of small molecules. *Analytical and Bioanalytical Chemistry*, 391, 1499–1507.
- Smith, M. C., Madec, S., Coton, E., & Hymery, N. (2016). Natural co-occurrence of mycotoxins in foods and feeds and their *in vitro* combined toxicological effects. *Toxins*, 8, 94.
- Soleimany, F., Jinap, S., & Abas, F. (2012). Determination of mycotoxins in cereals by liquid chromatography-tandem mass spectrometry. *Food Chemistry*, 130, 1055–1060.
- Soleimany, F., Jinap, S., Rahmani, A., & Khatib, A. (2011). Simultaneous detection of 12 mycotoxins in cereals using RP-HPLC-PDA-FLD with PHRED and a post-column derivatization system. *Food Additives and Contaminants*, 28, 494–501.
- Stoev, S. D. (2015). Foodborne mycotoxicoses, risk assessment and underestimated hazard of masked mycotoxins and joint mycotoxin effects or interaction. *Environmental Toxicology and Pharmacology*, *39*, 794–809.
- Streit, E., Schwab, C., Sulyok, M., Naehrer, K., Krska, R., & Schatzmayr, G. (2013). Multi-mycotoxin screening reveals the occurrence of 139 different secondary metabolites in feed and feed ingredients. *Toxins*, 5, 504–523.
- Suhr, K. I., & Nielsen, V. (2005). Inhibition of fungal growth on wheat and rye bread by modified atmosphere packaging and active packaging using volatile mustard essential oil. *Journal of Food Science*, 70, M37–M44.

- Suleiman, R. A., Rosentrater, K. A., & Bern, C. J. (2013). Effects of deterioration parameters on storage of maize: A review. *Journal of Natural Sciences Research*, *3*, 147–165
- Sultana, S., Kitajima, M., Kobayashi, H., Nakagawa, H., Shimizu, M., Kageyama, K., & Suga, H. (2019). A natural variation of fumonisin gene cluster associated with fumonisin production difference in *Fusarium fujikuroi*. *Toxins*, 11(4), 200.
- Summerell, B. A., & Leslie, J. F. (2011). Fifty years of *Fusarium*: how could nine species have ever been enough? *Fungal Diversity*, 50, 135.
- Tanaka, H., Takino, M., Sugita-konishi, Y., & Tanaka, T. (2006). Development of a liquid chromatography/time-of-flight mass spectrometric method for the simultaneous determination of trichothecenes, zearalenone and aflatoxins in foodstuffs. *Rapid Communications in Mass Spectrometry*, 20, 1422–1428.
- Tangni, E. K., Debongnie, P., Huybrechts, B., Van Hove, F., & Callebaut, A. (2017). Towards the development of innovative multi-mycotoxin reference materials as promising metrological tool for emerging and regulated mycotoxin analyses. *Mycotoxin Research*, 33, 15–24.
- Taylor, J. W., Jacobson, D. J., Kroken, S., Kasuga, T., Geiser, D. M., Hibbett, D. & Fisher, M. C. (2000). Phylogenetic species recognition and species concepts in fungi. Fungal Genetics and Biology, 31, 21–32.
- Tola, M., & Kebede, B. (2016). Occurrence, importance and control of mycotoxins: A review. *Cogent Food & Agriculture*, 2, 1191103.
- Torres, M. R., Ramos, A. J., Soler, J., Sanchis, V., & Marín, S. (2003). SEM study of water activity and temperature effects on the initial growth of *Aspergillus ochraceus, Alternaria alternata* and *Fusarium verticillioides* on maize grain. *International Journal of Food Microbiology*, 81, 185–193.
- Vaclavik, L., Zachariasova, M., Hrbek, V., & Hajslova, J. (2010). Analysis of multiple mycotoxins in cereals under ambient conditions using direct analysis in real-time (DART) ionization coupled to high-resolution mass spectrometry. *Talanta*, 82, 1950–1957.
- van Diepeningen, A. D., & De Hoog, G. S. (2016). Challenges in *Fusarium*, a trans-kingdom pathogen. *Mycopathologia*, 181, 161–163
- Van Egmond, H. P., Schothorst, R. C., & Jonker, M. A. (2007). Regulations relating to mycotoxins in food: perspectives in a global and European context. *Analytical and Bioanalytical Chemistry*, 389, 147–157.
- Váry, Z., Mullins, E., Mcelwain, J. C., & Doohan, F. M. (2015). The severity of wheat diseases increases when plants and pathogens are acclimatised to elevated carbon dioxide. *Global Change Biology*, 21, 2661–2669.

- Vaughan, M. M., Huffaker, A., Schmelz, E. A., Dafoe, N. J., Christensen, S., Sims, J., ... Teal, P. E. A. (2014). Effects of elevated [CO₂] on maize defence against mycotoxigenic *Fusarium verticillioides*. *Plant, Cell and Environment*, 37, 2691–2706.
- Velluti, A., Marin, S., Bbettucci, L., Ramos, A. J., & Sanchis, V. (2000). The effect of fungal competition on colonization of maize grain by *Fusarium moniliforme*, F. proliferatum and F. graminearum and on fumonisin B₁ and zearalenone formation. International Journal of Food Microbiology, 59, 59–66.
- Vendl, O., Berthiller, F., Crews, C., & Krska, R. (2009). Simultaneous determination of deoxynivalenol, zearalenone, and their major masked metabolites in cereal-based food by LC-MS-MS. *Analytical and Bioanalytical Chemistry*, 395, 1347–1354.
- Vermeulen, J., De Preter, K., Lefever, S., Nuytens, J., De Vloed, F., Derveaux, S., ... & Vandesompele, J. (2011). Measurable impact of RNA quality on gene expression results from quantitative PCR. *Nucleic Acids Research*, 39, e63-e63
- Vesonder, R. F., Ellis, J. J., Kwolek, W. F., & DeMarini, D. J. (1982). Production of vomitoxin on corn by *Fusarium graminearum* NRRL 5883 and *F. roseum* NRRL 6101. *Applied and environmental microbiology*, 43(4), 967-970.
- Villani, A., Moretti, A., De Saeger, S., Han, Z., Di Mavungu, J. D., Soares, C. M., ...& Paciolla, C. (2016). A polyphasic approach for characterization of a collection of cereal isolates of the *Fusarium incarnatum-equiseti* species complex. *International Journal of Food Microbiology*, 234, 24–35.
- Villani, A., Proctor, R. H., Kim, H. S., Brown, D. W., Logrieco, A. F., Amatulli, M. T., ... & Susca, A. (2019). Variation in secondary metabolite production potential in the *Fusarium incarnatum-equiseti* species complex revealed by comparative analysis of 13 genomes. *BMC Genomics*, 20, 314.
- Villafana, R. T., Ramdass, A. C., & Rampersad, S. N. (2019). Selection of *Fusarium* trichothecene toxin genes for molecular detection depends on TRI gene cluster organization and gene function. *Toxins*, 11(1), 36.
- Waśkiewicz, A. & Stępień, L. (2012). Mycotoxins Biosynthesized by Plant-Derived Fusarium Isolates. Archives of Industrial Hygiene and Toxicology, 63, 437–446.
- Wielogórska, E., Elliott, C. T., Danaher, M., Chevallier, O., & Connolly, L. (2015). Validation of an ultra-high-performance liquid chromatography-tandem mass spectrometry method for detection and quantitation of 19 endocrine disruptors in milk. *Food Control*, 48, 48–55.

- Williams, W. V., Rosenbaum, H., & Weiner, D. B. (1992). Effect of RNA concentration on cDNA synthesis for DNA amplification. *Genome Research*, 2, 86–88.
- Wong. (1992). Morfologi dan variety-varieti jagung. In Zaharah, H. (Eds.), Penanaman Jagung. Kuala Lumpur, Malaysia: Institut Penyelidikan Dan Kemajuan Pertanian, 7–12.
- Xing, F., Liu, X., Wang, L., Selvaraj, J. N., Jin, N., Wang, Y., ... & Liu, Y. (2017).
 Distribution and variation of fungi and major mycotoxins in pre-and post-nature drying maize in North China Plain. Food Control, 80, 244–251.
- Yan, H., Zhou, Z., & Shim, W. B. (2021). Two Regulators of G-protein signaling (RGS) proteins FlbA1 and FlbA2 differentially regulate fumonisin B₁ biosynthesis in *Fusarium verticillioides*. *Current Genetics*, 1-11.
- Yazid, S. N. E., Thanggavelu, H., Mahror, N., Selamat, J., & Samsudin, N. I. P. (2018). Formulation of maize- and peanut-based semi-synthetic growth media for the ecophysiological studies of aflatoxigenic *Aspergillus flavus* in maize and peanut agro-ecosystems. *International Journal of Food Microbiology*, 282, 57–65.
- Zachariasova, M., Vaclavikova, M., Lacina, O., Vaclavik, L., & Hajslova, J. (2012). Deoxynivalenol oligoglycosides: New "masked" Fusarium toxins occurring in malt, beer, and breadstuff. Journal of Agricultural and Food Chemistry, 60, 9280–9291.
- Zahra, N., Saeed, M. K., Sheikh, A., Kalim, I., Ahmad, S. R., & Jamil, N. (2019). A review of mycotoxin types, occurrence, toxicity, detection methods and control. *Biological Sciences-PJSIR*, 62, 206–218.
- Zain, M. E. (2011). Impact of mycotoxins on humans and animals. *Journal of Saudi Chemical Society*, 15, 129–144.
- Zhang, K., Schaab, M. R., Southwood, G., Tor, E. R., Aston, L. S., Song, W., ... & Tran, K. (2017). A collaborative study: Determination of mycotoxins in corn, peanut butter, and wheat flour using stable isotope dilution assay (SIDA) and liquid chromatography-tandem mass spectrometry (LC-MS/MS). Journal of Agricultural and Food Chemistry, 65, 7138–7152.
- Zhang, K., Wong, J. W., Krynitsky, A. J., & Trucksess, M. W. (2016). Perspective on advancing FDA regulatory monitoring for mycotoxins in foods using liquid chromatography and mass spectrometry. *Journal of AOAC International*, 99, 890–894
- Zhao, X., Tong, C., Pang, X., Wang, Z., Guo, Y., Du, F., & Wu, R. (2012). Functional mapping of ontogeny in flowering plants. *Briefings in bioinformatics*, 13(3), 317-328.

- Zhang, Z., Hu, X., Zhang, Q., & Li, P. (2016). Determination for multiple mycotoxins in agricultural products using HPLC-MS/MS via a multiple antibody immunoaffinity column. *Journal of Chromatography B*, 1021, 145–152.
- Zheng, M. Z., Richard, J. L., & Binder, J. (2006). A review of rapid methods for the analysis of mycotoxins. *Mycopathologia*, 161, 261–273.
- Zielonka, L., Gajęcka, M., Zmudzki, J., & Gajęcki, M. (2015). The effect of selected environmental *Fusarium* mycotoxins on the ovaries in the female wild boar (Sus scrofa). *Polish Journal of Veterinary Sciences*, 18, 391–399.



Websites

Malaysian Department of Meteorology. Available online: http://www.met.gov.my/ (accessed on 24 March 2019)

Merriam-Webster Inc. 2006. Cereal processing, Corn: layers and structures of corn kernel. Art, from Encyclopedia Britannica. Online Academic Edition. Inc., 2012. Available at: http://www.britannica.com/EBchecked/topic/103350/cereal-processing.

(Accessed 03 October 2019)

Index Mundi (2019). Malaysian maize feed importation and consumption from 1960 to 2019. (http://www.indexmundi.com.) (Accessed on 27 August 2019)

USDA, accessed in 2017. USDA Food Composition Databases, Agricultural Research Service. Available at: United States Department of Agriculture (USDA).https://ndb.nal.usda.gov/ndb/