

# UTILIZATION OF *Moringa oleifera* Lam. EXTRACTS AS NATURAL ANTIOXIDANT IN GOAT (*Capra hircus* Linnaeus) MEAT PRODUCTS

By

AL-ZAIDI MOHAMMED AWAD KADHIM

Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

July 2022

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## DEDICATIONS

Assalam for all, first and foremost. This work was purely for the sake of Allah, and after relying on Allah, the implementation of the project was initiated. Therefore, praise be to Allah, it was a simple fruit of success that we put for society and humanity as a whole. The most important element and the active role in the success of that experiment was Professor Awis. For that, all thanks to him. Appreciation and respect for his role in helping me and overcoming all the obstacles that I faced throughout the study period. Thanks are extended to all the respected members of the discussion committee. Dr. Ismail, Dr. Shokri, and Dr. Muhamad Faris for their distinguished role in addition to the friends of the study who helped me a lot and had a vital role and great help to complete the experiment successfully, special thanks Dr. Muideen, Dr. Pavan, Mohammed, Abu Bakar and all Lab Staff in Itafos in UPM. May Allah reward you richly rewarded.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## UTILIZATION OF *Moringa oleifera* Lam. EXTRACTS AS NATURAL ANTIOXIDANT IN GOAT (*Capra hircus* Linnaeus) MEAT PRODUCTS

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July 2022

# Chairman: Awis Qurni Sazili, PhDInstitute: Tropical Agriculture and Food Security

The present study was envisaged to evaluate the use of *Moringa oleifera* leaf extract as natural preservative in goat meat. Aqueous extract of *Moringa oleiferia* mature leaves (60 days) exhibited strong antioxidant activity as determined by radical-scavenging activity of 1,1-diphenyl 2 picrylhydrazyl (DPPH) as IC50 value 18.54 µg/mL, high total phenolic content (48.36 mg of gallic acid equivalent per g). Based on preliminary trial, five treatments were formulated for marination on goat meat as follows: Control (C, without the Moringa oleifera leaves extracts (MOLE) treatment); positive control (PC, treated with 0.1% butylated hydroxytoluene (BHT)); T<sub>0.1</sub> (treated with 0.1% MOLE);  $T_{0.5}$  (treated with 0.5% MOLE) and  $T_{1.0}$  (with 1.0% MOLE). The control and the extracts were treated on the Longissimus dorsi (LD), Infraspinatus (IF), Biceps femoris (BF) and Semimembranous (SM). The samples were marinated under refrigeration in high-density polyethylene (HDPE) bags for 7 days and assessed for various quality attributes (pH, water holding capacity, moisture, colour profile, and radical scavenging activity) on 1, 3 and 7 days, with sensory and texture profile analysis were assessed day 3 in addition to the above parameters. During refrigeration storage, all marinated muscles exhibited increased thiobarbituric acid reactive substances (TBARS) value, lowered water holding capacity (WHC) and increasing moisture content on day 3 followed by marginal decrease in moisture content on day 7. Within muscles, TBARS value of the control was recorded highest followed by T<sub>0.1</sub>,  $T_{0.5}$  and  $T_{1.0}$ . A significant (p<0.05) reduction in redness and increase in yellowness of goat meat upon marination was due to addition of MOLE than in control. MOLE extract when added to meat was found to retard lipid peroxidation. A marked deterioration on sensory attributes was recorded with the advancement of storage period; with control exhibited lowest flavour scores among treatments. Marination with MOLE had significant effect on the mean shear force value of LD muscle. The moisture content of all treatments of all 4 muscles under studies exhibited significantly (p<0.05) lower moisture content on 7<sup>th</sup> day of storage. The pH showed an increasing trend with the advancement of storage days. The results of sensory evaluation showed that all estimation levels are within the scale 8, which lies between moderate like and like very much. The antioxidant activity of MOLE was found to be comparable to BHT. Addition of MOLE did not affect any of the sensory attributes of marinated goat meat. The MOLE at a level of 1% was sufficient to protect goat meat against oxidative rancidity comparable to BHT added samples.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

## PENGGUNAAN Moringa oleifera Lam. EKSTRAK SEBAGAI ANTIOKSIDAN SEMULAJADI DALAM PRODUK DAGING KAMBING (Capra hircus Linnaeus)

Oleh

#### AL-ZAIDI MOHAMMED AWAD KADHIM

Julai 2022

Pengerusi : Awis Qurni Sazili, PhD Institut : Pertanian Tropika Dan Sekuriti Makanan

Kajian ini telah dijalankan untuk menilai kesan penggunaan ekstrak daun Moringa oleifera sebagai pengawet semulajadi dalam daging kambing. Ekstrak akueus daun Moringa oleiferia matang (60 hari) mempamerkan aktiviti antioksidan yang tinggi seperti yang telah dibuktikan melalui aktiviti penghapusan radikal 2,2-diphenyl-1picrylhydrazyl (DPPH) yang menunjukkan nilai IC50 sebanyak 18.54 µg/mL, kandungan fenolik yang tinggi (48.36 mg asid gallik bagi setiap g). Berdasarkan beberapa ujian awal, lima rawatan telah diformulasikan untuk perapan iaitu kawalan (tanpa MOLE dan BHT), Kawalan positif (PC) dengan 0.1% butylated hydroxytoluene (BHT), T<sub>0.1</sub> (dengan 0.1% MOLE) T<sub>0.5</sub> (dengan 0.5% MOLE) dan T1 (dengan 1.0% MOLE) untuk 4 jenis otot iaitu, Longissimus dorsi (LD), Infraspinatus (IF), Biceps femoris (BF) dan semimembranous (SM); membawa kepada jumlah 20 sampel untuk setiap eksperimen. Sampel telah diperap di dalam suhu sejuk di dalam beg HDPE selama 7 hari dan telah dinilai untuk pelbagai ciri-ciri kualiti (pH, kapasiti pegangan air, lembapan, profil warna, bahan tindak balas asid tiobarbiturik TBARS, kiraan plate count (SPC) dan kiraan psikrofilik) pada hari 1, 3 dan 7. Analisis deria (20 panel jurulatih) dan analisis profil tekstur pada hari ke-3 telah dijalankan sebagai tambahan kepada parameter di atas. Semasa penyimpanan penyejukan, semua otot yang diperap menunjukkan peningkatan nilai thiobarbituric acid reactive substances (TBARS), menurunkan kapasiti pegangan air dan meningkatkan kandungan lembapan pada hari ke-3 diikuti dengan penurunan kecil dalam kandungan lembapan pada hari ke-7. Hasil keputusan untuk otot pula menunjukkan, nilai TBARS untuk rawatan kawalan dicatatkan sebagai yang tertinggi diikuti oleh T<sub>0.1</sub>, T<sub>0.5</sub> dan T<sub>1.0</sub>; manakala nilai TBARS T<sub>1.0</sub> adalah setanding dengan sampel kawalan positif (PC). Pengurangan ketara (P <0.05) untuk nilai kemerahan dan peningkatan nilai kekuningan daging kambing semasa perapan adalah disebabkan oleh penambahan MOLE berbanding dalam rawatan kawalan. Apabila ekstrak MOLE ditambah kepada daging, ianya didapati dapat melambatkan peroksidasi lipid. Kemerosotan yang ketara pada ciri-ciri deria telah direkodkan dengan pertambahan tempoh penyimpanan; dengan rawatan kawalan mempamerkan skor perisa yang paling rendah antara rawatan. Aktiviti antioksidan ekstrak MOLE didapati setanding dengan BHT. Penambahan ekstrak MOLE tidak menjejaskan sebarang sifat deria daging kambing yang diperap. Ekstrak MOLE pada tahap 1% adalah mencukupi untuk melindungi daging kambing daripada ketengikan oksidatif yang setanding dengan sampel yang telah ditambah BHT.



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# LIST OF ABBREVIATIONS

	ABTS	Measures the relative ability of antioxidants to scavenge
	APA	Anti-peroxide activity
	ANOVA	Analysis of Variance
	AQ	Aqueous solvent
	AOA	Antioxidant activity
	BHA	Butylated hydroxyl anisole
	BHT	Butylated hydroxyl toluene
	CHL	Chlorophyll
	DPPH	2,2-diphenyl-1-picrylhydrazyl
	GAE\G.D.W	Gallic acid equilibrium curve equivalent
	MOLE	Moringa oliefera leaves extract
	MDA	Malondialdehyde salt
	PWFC	Protein water fat coefficient
	PWC	protein water coefficient
	РН	Potential of hydrogen
	РСА	Principal component analysis
	FRAP	Ferric reducing/antioxidant potential
	ТВА	2-thiobarbituric acid
	TCA	Trichloroacetic acid
	TPC	Total Plate Count
	TBARS	Thiobarbituric Acid Reactive Substance
	TFC	Total Flavonoid Content
	TAC	Total Antioxidant capacity
	TPA	Texture profile analysis

### **CHAPTER 1**

### **INTRODUCTION**

## 1.1 Background

Red meat has a vital role in providing a healthy and nutritionally balanced diet, mainly contributed by the sources of animal proteins in the human diet (Ekmekcioglu et al., 2018). Meanwhile, the increase in the global population will require more meat and meat products, mostly for the developing country (FAO, 2011). The growing meat market provides a vast opportunity for livestock rearing and the meat industry in developed nations. Nevertheless, increasing livestock products represent a big challenge (FAO, 2019). Therefore, despite increasing meat products represent a big challenge (FAO, 2019). Therefore, despite increasing meat production, the reduction of meat wastage especially within processing, distribution, and storage, should also be considered in order to meet the growing demand and ensure food security.

World population is projected to increase to reach 9 billion by 2050, most of which is forecasted in developing countries. Meanwhile, the consumption of meat is continuously increasing worldwide because of the increase in the human population. Although the consumption of meat is increasing worldwide significant portion of meat and meat products are spoiled every year (Addis, 2015). However, there is significant portion of meat and meat products are spoiled especially from microbial spoilage and lipid oxidation (Addis, 2015), which leads to the application of antioxidants in the meat and meat products (Islam et al., 2018). Meat preservation is a vital process for production and transporting of meat for a long duration without spoiling texture, colour, and nutritional value after the development and rapid growth of supermarkets (Nychas et al., 2008). The aims of preservation methods are to inhibit microbial spoilage and to minimize oxidation and enzymatic spoilage.

Antioxidants have a major role when added to meat products to prevent lipid oxidation, retard the development of off-flavours, and improve colour stability. In the food industry, they can be divided into natural and synthetic antioxidants. BHA (butylated hydroxyanisole), BHT, PG (propyl gallate), and TBHQ (tert-butylhydroquinone) are examples of synthetic antioxidants; while ingredients obtained from natural sources which exhibit anti-oxidative potential in a food model system are considered as natural antioxidants. These antioxidants play a very important role in the food industry. However, synthetic antioxidants have been identified as toxicological and carcinogenic agents in some studies (Abraham et al., 1986; Ahmad et al., 1995; Sarafian et al. 2002; Faine et al., 2006). Thus, the food industry selected natural products over artificial ones. Consequently, the food market is demanding natural antioxidants, free of synthetic additives and still orientated to diminish the oxidation processes in high-fat meat and meat products. The majority of natural antioxidants are phenolic compounds, and the most important are the tocopherols, flavonoids, and phenolic acids, which may be found widely in plant part, such as grains, fruits, nuts, seeds, leaves, roots, arils, and barks (Santos-Sánchez et al., 2019). Moringa oleifera is a natural antioxidant being focus of research by food technologist recently. *M. oleifera* contains myriad of phytochemicals possess antioxidant potential. Phenolics and flavonoids are the authentic antioxidants found in *Moringa* leaves that have been reported to be safe and bioactive, in which kaempferol and quercetin are recognized as the most effective antioxidants in *Moringa* leaves (Fahey, 2005; Sreelatha and Padma, 2009).

Moringa oleifera is an ornamental plant native to tropical and subtropical. All parts of the *Moringa* plant are edible, with leaves and pods used most frequently. Abundant studies have been done to evaluate the antioxidative capacity of the M. oleifera (Sreelatha and Padma, 2009; Ogbunugafor et al., 2011; Fitriana et al., 2016; Feihrmann et al., 2017). From the abundant research works which showed the antioxidative capacity of the *M. oleifera*, subsequent several studies have been done to evaluate the application of *M. oleifera* as a food additive. The antioxidant potential of *M. oleifera* leaf extract has been shown for the lipid stabilization of sunflower oil (Anwar et al., 2006), butter (Nadeem et al., 2013), and cream cheese (Mohamed et al., 2018). Meanwhile, Feihrmann et al. (2017) showed that M. oleifera leaf extract was effective to inhibit lipid oxidation in beef. This finding was also supported by Islam et al. (2018), who concluded that the effects of *M. oleifera* leaf extract as a natural antioxidant can be used in beef meatballs preparation instead of synthetic antioxidants (BHA) by dosedependent effects. Thus, this showed that M. oleifera leaf extract is a good candidate as a food additive in meat. However, to the best of our knowledge, there is no evidence of *M. oleifera* as preservation on goat meat products (Hocquette et al., 2010).

## 1.2 Problem statement

It is clear that the use of industrial/synthetic antioxidants in preserving meat has a negative impact on consumer health such as food allergy, intolerance, nausea, hyperactivity, and even cancer. Preserving meat from natural plant sources and having a positive effect on the health of meat consumers and increases the effectiveness of its preservation period .Different maturity stages of *Moringa oleifera* leaves could result different levels of phenolic compounds. The effect of maturity stages of *Moringa oleifera* leaves on the extraction process is used as a natural antioxidant incorporated to goat meat causes various concentrations of phenolic compounds. However, different type of muscles of the goat meat could be affected differently by the *Moringa oleifera* extracts. The different types of goat muscle after incorporation with the previously selected *Moringa oleifera* leaves extract may improve the physical and chemical properties and extention of goat meat shelf life.

## 1.3 General objective

The general objective of this study was to determine the effects of *M. oleifera* on the quality of goat meat during storage.

The specific objectives of this study were:

- 1. To determine the phenolic compound of extracts from different *Moringa oleifera* leaves at different maturity levels (30 days and 60 days) using an extraction method.
- 2. To determine the effect of *Moringa oleifera* extracts on different type of muscles of goat meat.

## 1.4 Hypotheses

The total phenolic content may be affected by different age (30-60 days) of *M. oleifera* leaves. The *M. oleifera* leaves extracts in marinade may affect the shelf life and physiochemical properties of different goat muscles.

### 1.5 Significance of study

The current study focuses on finding natural preservatives from plant sources that are available, affordable, easy to use, and of high biological value. The reason for this is because preservatives contain valuable sources and nutrients such as minerals and In addition, they contain highly effective materials such as phenolic vitamins. compounds, carotenoids, vitamin C, and beta-carotene. These materials are used as antioxidants to prevent food spoilage especially red meat and improve the quality and properties of food and meat processing. Consequently, researchers have been working and developing these sources to be used as food additives. The objective of this study was to use Moringa extract as a natural antioxidant as preventive to red meat, especially the various muscles of raw goat meat and to increase their shelf life. In addition, preservatives are good agents to keep the physical and chemical properties intact and, as such, their consumption is healthy and better than the synthetic additives. It is important to mention that the synthetic additives cause several heath problems such as toxicity and cancer. This study is potentially beneficial to consumers' health. This study highlights the severe consequences against food producers to limit the use of the synthetic preservative for the sake of human requirement for health environment.

## 1.6 Gaps of previous studies

Incorporation of natural antioxidants in red goat meat has been experienced in the industries (Kumar et al., 2015). Antioxidants preserve the quantity and quality of meat by scavenging free radicals that cause lipid oxidation which deteriorates the shelf life of meat and meat products (Manessis et al., 2020). Studies have indicated that the use of *Moringa* extract with red goat meat had a positive significant effect. The extract is biologically characterized as active substances against lipid oxidation and meat spoilage; which increases the shelf life of goat meat. Feihrmann et al. (2017) suggested that *M. oleifera* leaf extract substitutes synthetic antioxidant in beef and, as such,

inhibits lipid oxidation of beef. Kenawi and Abd El-Hameed (2018) studied the incorporation of *M. oleifera* leaf extract in buffalo meat product. In the experiment, the pH value of raw and cooked meat patties showed that the pH was not affected by *M. oleifera* and butylated hydroxy toluene (BHT) (Das et al., 2012) and that *M. oleifera* leaf extract is a potential incorporated antioxidant with a privilege of being without adverse effect on the quality. Meanwhile, Rahman et al. (2020) studied the quality and shelf-life of goat meat nuggets incorporated with *M. oleifera* leaf extract at frozen storage. Hence, the gaps in previous studies were in ignoring the effect of maturity age of *Moringa* under a single study. However, these studies did not specify the effect of the *M. oleifera* leaves incorporated with various solvents. The aim of this study was to fill these gaps along with other objectives as listed earlier.



#### REFERENCES

- Abad, A., & Shahidi, F. (2020). Fatty acid, triacylglycerol and minor component profiles affect oxidative stability of camelina and sophia seed oils. Food Bioscience, 100849.
- Abdulsalam, S., Yahaya, M. S., & Yakasai, M. A. (2015). Performance of Broiler Chickens Fed on *Moringa Oleifera* leaf meal supplemented poultry feed. Nigeria Agricultural Journal. 46(1), 139–146.
- Abraham, J. A., Whang, J. L., Tumolo, A., Mergia, A., Friedman, J., Gospodarowicz, D., & Fiddes, J. C. (1986). Human basic fibroblast growth factor: nucleotide sequence and genomic organization. The EMBO Journal, 5(10), 2523-2528.
- Addis, M. (2015). Major Causes of Meat Spoilage and Preservation Techniques: A. changes, 41. ISSN 2224-6088 101-114.
- Adeyemi, K. D., El-Imam, A. M. A., Olorunsanya, A. O., Sola-ojo, F. E., Olatunji, O. S., Okukpe, K. M., & Fatai, I. A. (2014). Lipid peroxidation in smoke-dried African catfish treated with *Moringa oleifera* marinade, salt or butylated hydroxyl anisole. Croatian Journal of Fisheries, 72, 36-40.
- Afable, E. A., de la Cruz, R. J., Paras, A. T., & Segui, M. E. (2020). Diagonalizability with respect to perplectic and pseudo-unitary similarity transformations. Linear Algebra and its Applications, 591, 61-71.
- Agamou, A. J. A., Fombang, E. N., & Mbofung, C. M. (2015). Particular benefits can be attributed to *Moringa oleifera* Lam leaves based on origin and stage of maturity. Journal of Experimental Biology and Agricultural Sciences, 3(6), 541-555.
- Ahmad, R. S., Imran, A., & Hussain, M. B. (2018). Nutritional composition of meat. Meat Science and Nutrition, 4..7704,
- Ahmad, S. (1995). Oxidative stress from environmental pollutants. Archives of Insect Biochemistry and Physiology, 29(2), 135-157.
- Ahmadinejad, F., Geir Moller, S., Hashemzadeh-Chaleshtori, M., Bidkhori, G., & Jami, M.-S. (2017). Molecular mechanisms behind free radical scavengers function against oxidative stress. Antioxidants, 6(3), 51.
- Akerib, D. S., Akerlof, C. W., Alsum, S. K., Araújo, H. M., Arthurs, M., Bai, X., & Bailey A. J. (2020). Projected WIMP sensitivity of the LUX-ZEPLIN dark matter experiment. Physical Review, 1015, 052002.
- Akhtar, S., Khan, M.I., & Faiz, F. (2013). Effect of thawing on frozen meat quality: A comprehensive review. Pakistan Journal of Food Sciences, 23(4), 198-211.

- Akula, R., & Ravishankar, G.A. (2011). Influence of abiotic stress signals on secondary metabolites in plants. Plant Signal Behaviour, 6, 1720–1731.
- Alabi, O., Malik, A., NgAmbi, J., Obaje, P., & Ojo, B. (2017). Effect of aqueous *Moringa Oleifera* (Lam) leaf extracts on growth performance and carcass characteristics of Hubbard broiler chicken. Revista Brasileira De Ciência Avícola, 19(2), 273-280.
- Alamgir, A. N. M. (2018). Secondary metabolites: secondary metabolic products consisting of C and H; C, H, and O; N, S, and P elements; and O/N heterocycles. In Therapeutic Use of Medicinal Plants and their Extracts: Volume 2 (pp. 165-309). Springer, Cham.
- Al-Asmari, A.K., Albalawi, S.M., Athar, M.T., Khan, A.Q., Al-Shahrani, H. & Islam, M. (2015). *Moringa oleifera* as an anti-cancer agent against breast and colorectal cancer cell lines. PloS one 10(8), e0135814.
- Al-Malki, A.L. & El Rabey, H.A., 2015. The antidiabetic effect of low doses of *Moringa oleifera* Lam. seeds on streptozotocin induced diabetes and diabetic nephropathy in male rats. BioMed Research International, 2015, 381040.
- Al-Wabel, M. I., Al-Omran, A., El-Naggar, A. H., Nadeem, M., & Usman, A. R. (2013). Pyrolysis temperature induced changes in characteristics and chemical composition of biochar produced from conocarpus wastes. Bioresource Technology, 131, 374-379.
- Aminzare, M., Hashemi, M., Ansarian, E., Bimkar, M., Azar, H. H., Mehrasbi, M. R., & Afshari, A. (2019). Using natural antioxidants in meat and meat products as preservatives: a review. Advances in Animal and Veterinary Sciences, 7(5), 417-426.
- Amiri, E., Aminzare, M., Azar, H. H., & Mehrasbi, M. R. (2019). Combined antioxidant and sensory effects of corn starch films with nanoemulsion of Zataria multiflora essential oil fortified with cinnamaldehyde on fresh ground beef patties. Meat Science, 153, 66-74.
- Amit, S. K., Uddin, M. M., Rahman, R., Islam, S. R., & Khan, M. S. (2017). A review on mechanisms and commercial aspects of food preservation and processing. Agriculture and Food Security, 6(1), 1-22.
- Anaeto, M., Adeyeye, J. A., Chioma, G. O., Olarinmoye, A. O., & Tayo, G. O. (2010). Goat products: Meeting the challenges of human health and nutrition. Agriculture and Biology Journal of North America, 1(6), 1231-1236.
- Anand, S. P., & Sati, N. (2013). Artificial preservatives and their harmful effects: looking toward nature for safer alternatives. International Journal of Pharmaceutical Sciences and Research, 4(7), 2496.

- Anwar, F., Latif, S., Ashraf, M. & Gilani, A.H., 2007. Moringa oleifera: a food plant with multiple medicinal uses. Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives, 21(1),17-25.
- Arain, M. A., Khaskheli, M., Rajput, I., Rao, S., Faraz, S., Fazlani, S., & Umer, M. (2010). Examination of physical properties of goat meat. Pakistan Journal of Nutrition, 9(5), 422-425.
- Asiedu-Gyekye, I. J., Frimpong-Manso, S., Awortwe, C., Antwi, D. A., & Nyarko, A. K. (2014). Micro- and macroelemental composition and safety evaluation of the nutraceutical *Moringa oleifera* leaves. Journal of Toxicology, 2014, 1-13.
- Aslam, M., Anwar, F., Nadeem, R., Rashid, U., Kazi, T., G. & Nadeem, M. (2005). Mineral composition of *Moringa oleifera* leaves and pods from different regions of Punjab, Pakistan. Asian Journal of Plant Science, 4, 417-421.
- Awad, A.M., Kumar, P., Ismail-Fitry, M.R., Jusoh, S., Ab Aziz, M.F., & Sazili, A.Q. (2021). Green extraction of bioactive compounds from plant biomass and their application in meat as natural antioxidant. Antioxidants (Basel), 10(9),1465.
- Awadh, J. A., Jian, L., De-sheng, Z., & De-fang, F. (2001). Determination of multiple pyrethroid insecticides in chrysanthemum flower. Chinese Jornal of Pesticide Research, 3(4), 81-85.
- Awodele, O., Oreagba, I. A., Odoma, S., Silva, J. A., & Osunkalu, V. O. (2012). Toxicological evaluation of the aqueous leaf extract of *Moringa oleifera* Lam. (Moringaceae). Journal of Ethnopharmacology, 139(2), 330-336.
- Aykan, N. F. (2015). Red meat and colorectal cancer. Oncology Reviews, 9(1), 288.
- Bajovic, B., Bolumar, T., & Heinz, V. (2012). Quality considerations with high pressure processing of fresh and value added meat products. Meat Science, 92(3), 280-289.
- Banerjee, R., & Maheswarappa, N. B. (2019). Superchilling of muscle foods: Potential alternative for chilling and freezing. Critical Reviews in Food Science and Nutrition, 59(8), 1256-1263.
- Bartley, G. E., & Scolnik, P. A. (1995). Plant carotenoids: pigments for photoprotection, visual attraction, and human health. The Plant Cell, 7(7), 1027.
- Bellostas, N., Soslash, J. C., Nikiema, A., Soslash, H., Pasternak, D., & Kumar, S. (2010). Glucosinolates in leaves of Moringa species grown and disseminated in Niger. African Journal of Agricultural Research, 5(11), 1338-1340.
- Bendall, J. R. (1975). Cold- contracture and atp- turnover in the red and white musculature of the pig, post mortem. Journal of the Science of Food and Agriculture, 26(1), 55-71.

- Bennett, R. N., Mellon, F. A., Foidl, N., Pratt, J. H., Dupont, M. S., Perkins, L., & Kroon, P. A. (2003). Profiling glucosinolates and phenolics in vegetative and reproductive tissues of the multi-purpose trees *Moringa oleifera* L. (Horseradish Tree) and *Moringa stenopetala* L. Journal of Agricultural and Food Chemistry, 51(12), 3546-3553.
- Böttcher, S., Steinhäuser, U., & Drusch, S. (2015). Off-flavour masking of secondary lipid oxidation products by pea dextrin. Food Chemistry, 169, 492-498.
- Brilhante, R. S. N., Sales, J. A., Pereira, V. S., Castelo-Branco, D. C. M., Cordeiro, R. A., Sampaio, C. M. S., Paiva, M. A. N., Santos, J. B. F., Sidrim, J. J. C., & Rocha, M. F. G. (2017). Research advances on the multiple uses of *Moringa oleifera*: A sustainable alternative for socially neglected population. Asian Pacific Journal of Tropical Medicine, 10(7), 621–630.
- Bukar, A., Uba, A., & Oyeyi, T. (2010). Antimicrobial profile of *Moringa oleifera* Lam. extracts against some food-borne microorganisms. Bayero Journal of Pure and Applied Sciences, 3(1), 58706.
- Burneiko, R. C., Diniz, Y. S., Galhardi, C. M., Rodrigues, H. G., Ebaid, G. M., Faine, L. A., & Novelli, E. L. (2006). Interaction of hypercaloric diet and physical exercise on lipid profile, oxidative stress and antioxidant defenses. Food and Chemical Toxicology, 44(7), 1167-1172.
- Campêlo, M. C. S., Medeiros, J. M. S., & Silva, J. B. A. (2019) Natural products in food preservation. International Food Research Journal 26(1), 41-46.
- Cao, J., Gargano, Ho, Y. S., Magnenat, J. L., Bronson, R. T., M., Sugawara, M., & Funk, C. D. (1997). Mice deficient in cellular glutathione peroxidase develop normally and show no increased sensitivity to hyperoxia. Journal of Biological Chemistry, 272(26), 16644-16651.
- Casey, N. H., & Webb, E. C. (2010). Managing goat production for meat quality. Small Ruminant Research, 89(2-3), 218-224.
- Celada, P., Sánchez-Muniz, F. J., Delgado-Pando, G., Bastida, S., Rodilla, M. E., Jiménez-Colmenero, F., & Olmedilla-Alonso, B. (2016). Effects of improved fat meat products consumption on emergent cardiovascular disease markers of male volunteers at cardiovascular risk. Journal of Physiology and Biochemistry, 72(4), 669-678.
- Chellaiah, R., Shanmugasundaram, M. & Kizhekkedath, J., (2020). Advances in meat preservation and safety. International Journal of Science and Research (IJSR), 9(3),1499-1502.
- Choudhary, M. K., Bodakhe, S. H., & Gupta, S. K. (2013). Assessment of the antiulcer potential of *Moringa oleifera* root-bark extract in rats. Journal of Acupuncture and Meridian Studies, 6(4), 214-220.

- Chuang, P.H., Lee, C.W., Chou, J.Y., Murugan, M., Shieh, B.J. & Chen, H.M., (2007). Anti-fungal activity of crude extracts and essential oil of *Moringa oleifera* Lam. Bioresource Technology, 98(1), 232-236.
- Collie, J. T., Greaves, R. F., Jones, O. A., Eastwood, G., & Bellomo, R. (2020). Vitamin C measurement in critical illness: challenges, methodologies and quality improvements. Clinical Chemistry and Laboratory Medicine (CCLM), 58(4), 460-470.
- Contini, C., Álvarez, R., O'sullivan, M., Dowling, D. P., Gargan, S. Ó., & Monahan, F. J. (2014). Effect of an active packaging with citrus extract on lipid oxidation and sensory quality of cooked turkey meat. Meat Science, 96(3), 1171-1176.
- Das, A. K., Rajkumar, V., Verma, A. K., & Swarup, D. (2012). Moringa oleiferia leaves extract: A natural antioxidant for retarding lipid peroxidation in cooked goat meat patties. International Journal of Food Science and Technology, 47(3), 585-591.
- Dave, D., & Ghaly, A. E. (2011). Meat spoilage mechanisms and preservation techniques: a critical review. American Journal of Agricultural and Biological Sciences, 6(4), 486-510.
- Devatkal, S. K., Narsaiah, K., & Borah, A. (2010). Anti-oxidant effect of extracts of kinnow rind, pomegranate rind and seed powders in cooked goat meat patties. Meat Science, 85(1), 155-159.
- Devendra, C. (2011). Integrated tree crops-ruminants systems in South East Asia: Advances in productivity enhancement and environmental sustainability. Asian-Australasian Journal of Animal Sciences, 24(5), 587-602.
- Dhakar, R., Pooniya, B., Gupta, M., Maurya, S., Bairwa, N., & Sanwarmal. (2011). Moringa: The herbal gold to combat malnutrition. Chronicles of Young Scientists, 2(3), 119.
- Djeridane, A., Yousfi, M., Nadjemi, B., Boutassouna, D., Stocker, P., & Vidal, N. (2006). Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. Food chemistry, 97(4), 654-660.
- Dolojan, F. M. (2015). Standardization and commercialization of chevon products. Journal of Agricultural Technology, 11(8), 2099-2109.
- Duthie, G. G., Duthie, S. J. and Kyle, J. A. M. (2000). Plant polyphenols in cancer and heart disease: Implications as nutritional antioxidants. Nutrition Research Review, 13, 79-106,
- Ebrahimzadeh, M. A., Nabavi, S. F., Nabavi, S. M., & Eslami, B. (2010). Antihemolytic and antioxidant activities of *Allium paradoxum*. Central European Journal of Biology, 5(3), 338-345.

- Ekmekcioglu, C., Wallner, P., Kundi, M., Weisz, U., Haas, W., & Hutter, H. P. (2018). Red meat, diseases, and healthy alternatives: A critical review. Critical Reviews in Food Science and Nutrition, 58(2), 247-261.
- El Dib, R., Gameiro, O.L., Ogata, M.S., Modolo, N.S., Braz, L.G., Jorge, E.C., do Nascimento Junior, P. & Beletate, V., (2015). Zinc supplementation for the prevention of type 2 diabetes mellitus in adults with insulin resistance. Cochrane Database of Systematic Reviews, (5).CD005525.
- El-Deep, M. H., Dawood, M. A. O., Assar, M. H., Ijiri, D., & Ohtsuka, A. (2019). Dietary *Moringa oleifera* improves growth performance, oxidative status, and immune related gene expression in broilers under normal and high temperature conditions. Journal of Thermal Biology, 82, 157–163.
- Escarpa, A., & Gonzalez, M. C. (2001). An overview of analytical chemistry of phenolic compounds in foods. Critical Reviews in Analytical Chemistry, 318, 57-139.
- Fahey, J. W. (2005). Moringa oleifera: a review of the medical evidence for its nutritional, therapeutic, and prophylactic properties. Part 1. Trees for life Journal, 1(5), 1-15.
- Falowo, A. B., Fayemi, P. O., & Muchenje, V. (2014). Natural antioxidants against lipid–protein oxidative deterioration in meat and meat products: A review. Food Research International, 64, 171-181.
- FAO (2011-2019). Study: Global Food Losses and Food Waste-extent, causes and prevention"-FAO.
- Faustman, C., & Suman, S. P. (2017). The Eating Quality of Meat. Lawrie's Meat Science, 329-356.
- Feihrmann, A. C., Nascimento, M. G., Belluco, C. Z., Fioroto, P., Cardozo-Filho, L., & Tonon, L. C. (2017). Evaluation of the effect of antioxidant Moringa Oleifera. Chemical Engineering Transactions, 57, 1993-1998.
- Firuzi, M. R., Niakousari, M., Eskandari, M. H., Keramat, M., Gahruie, H. H., & Khaneghah, A. M. (2019). Incorporation of pomegranate juice concentrate and pomegranate rind powder extract to improve the oxidative stability of frankfurter during refrigerated storage. LWT, 102, 237-245.
- Fitriana, W.D., Ersam, T., Shimizu, K. & Fatmawati, S., (2016). Antioxidant activity of *Moringa oleifera* extracts. Indonesian Journal of Chemistry, 16(3), 297-301.
- Font-i-Furnols, M., & Guerrero, L. (2014). Consumer preference, behavior and perception about meat and meat products: An overview. Meat Science, 98(3), 361-371.

- Gerelt, B., Ikeuchi, Y., Nishiumi, T., & Suzuki A. (2002). Meat tenderization by calcium chloride after osmotic dehydration. Meat Science, 60(3), 237-44.
- Giannakourou, M., & Giannou, V. (2014). Chilling and freezing. Food Engineering Handbook: Food Process Engineering, CRC Press, Boca Raton, FL, 319-370.
- Gómez, I., Janardhanan, R., Ibañez, F. C., & Beriain, M. J. (2020). The effects of processing and preservation technologies on meat quality: Sensory and nutritional aspects. Foods, 9(10), 1416.
- Gopalakrishnan, L., Doriya, K. & Kumar, D.S., (2016). Moringa oleifera: A review on nutritive importance and its medicinal application. Food Science and Human Wellness, 5(2), 49-56.
- Green, M. A., & Fry, S. C. (2005). Vitamin C degradation in plant cells via enzymatic hydrolysis of 4-O-oxalyl-L-threonate. Nature, 433(7021), 83-87.
- Hannan, M. A., Kang, J., Mohibbullah, M., Hong, Y., Lee, H., Choi, J., Moon, I. S. (2014). *Moringa oleifera* with promising neuronal survival and neurite outgrowth promoting potentials. Journal of Ethnopharmacology, 152(1), 142-150.
- Hasanuzzaman, M., Nahar, K., Alam, M., Roychowdhury, R., & Fujita, M. (2013). Physiological, biochemical, and molecular mechanisms of heat stress tolerance in plants. International Journal of Molecular Sciences, 14(5), 9643-9684.
- Heerthana, V. R., & Preetha, R. (2019). Biosensors: a potential tool for quality assurance and food safety pertaining to biogenic amines/volatile amines formation in aquaculture systems/products. Reviews in Aquaculture, 11(1), 220-233.
- Heijnen, C., Boogaard, B. V. D., & Maas van Berkel, B. (2004). Preservation of fish and meat. Agrodok. Editor: Marja de Goffau-Markusse, Translation: Joost Guijt, Catharina de Kat-Reynen (editing), Printed by: Digigrafi, Wageningen, the Netherlands
- Hocquette, J. F., Gondret, F., Baéza, E., Médale, F., Jurie, C., & Pethick, D. W. (2010). Intramuscular fat content in meat-producing animals: development, genetic and nutritional control, and identification of putative markers. Animal, 4(2), 303-319.
- Huang, C., Wang, Y., Li, X., Ren, L., Zhao, J., Hu, Y., & Cao, B. (2020). Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. The lancet, 395(10223), 497-506.
- Hunter, R.S.; Harold, R.W. (1987) The Measurement of Appearance; Hunter Associates Laboratory. Inc.: Reston, VA, USA.

- Hussein, R.A. and El-Anssary, A.A., (2019). Plants secondary metabolites: the key drivers of the pharmacological actions of medicinal plants. Herbal Medicine, 1(3),76193.
- Islam, M. J., Sayeed, M. A., Akhtar, S., Hossain, M. S., & Liza, A. A. (2018). Consumers profile analysis towards chicken, beef, mutton, fish and egg consumption in Bangladesh. British Food Journal, 120, 2818-2831.
- Jakopič, J., Štampar, F. & Veberič, R., (2010). Influence of hail net and reflective foil on cyanidin glycosides and quercetin glycosides in 'Fuji'apple skin. HortScience, 45(10),1447-1452.
- Jamal, M., Ahmad, W., Andleeb, S., Jalil, F., Imran, M., Nawaz, M. A., & Kamil, M. A. (2018). Bacterial biofilm and associated infections. Journal of the Chinese Medical Association, 81(1), 7-11.
- Jeffery, E. H., Brown, A. F., Kurilich, A. C., Keck, A. S., Matusheski, N., Klein, B. P., & Juvik, J. A. (2003). Variation in content of bioactive components in broccoli. Journal of Food Composition and Analysis, 16(3), 323-330.
- Jiang, J., & Xiong, Y. L. (2016). Natural antioxidants as food and feed additives to promote health benefits and quality of meat products: A review. Meat Science, 120, 107-117.
- Jomova, K., & Valko, M. (2011). Advances in metal-induced oxidative stress and human disease. Toxicology, 283(2-3), 65–87.
- Jung, S., Choe, J. H., Kim, B., Yun, H., Kruk, Z. A., & Jo, C. (2010). Effect of dietary mixture of gallic acid and linoleic acid on antioxidative potential and quality of breast meat from broilers. Meat Science, 86(2), 520-526.
- Kang, J., Senanayake, G., Sohn, J., & Shin, S. M. (2010). Recovery of cobalt sulfate from spent lithium ion batteries by reductive leaching and solvent extraction with Cyanex 272. Hydrometallurgy, 100(3-4), 168-171.
- Karre, L., Lopez, K., & Getty, K. J. (2013). Natural antioxidants in meat and poultry products. Meat Science, 94(2), 220-227.
- Kasolo, J. N., Bimenya, G. S., Ojok, L., Ochieng, J. & Ogwal-Okeng, J. W. (2010). Phytochemicals and uses of *Moringa oleifera* leaves in Ugandan rural communities. Journal of Medicinal Plants Research. 4(9), 753-757.
- Kenawi, M, A., & Sanaa M. Abd El-Hameed. (2018). Effect of *Moringa oleifera* leaves extract on quality of buffalo meat product. Analytical methods, 1, 204867994.
- Keren-Shaul, H., Spinrad, A., Weiner, A., Matcovitch-Natan, O., Dvir-Szternfeld, R., Ulland, T. K., & Amit, I. (2017). A unique microglia type associated with restricting development of Alzheimer's disease. Cell, 169(7), 1276-1290.

- Khan, I., Zaneb, H., Masood, S., Yousaf, M. S., Rehman, H. F., & Rehman, H. (2017). Effect of *Moringa oleifera* leaf powder supplementation on growth performance and intestinal morphology in broiler chickens. Journal of Animal Physiology and Animal Nutrition, 101, 114-121.
- Kholif, A. E., Gouda, G. A., Anele, U. Y., & Galyean, M. L. (2018). Extract of *Moringa oleifera* leaves improves feed utilization of lactating Nubian goats. Small Ruminant Research, 158, 69-75.
- Kim, Y. H. B., Warner, R. D., & Rosenvold, K. (2014). Influence of high pre-rigor temperature and fast pH fall on muscle proteins and meat quality: a review. Animal Production Science, 54(4), 375-395.
- Kim, Y. H., Huff-Lonergan, E., Sebranek, J. G., & Lonergan, S. M. (2010). Highoxygen modified atmosphere packaging system induces lipid and myoglobin oxidation and protein polymerization. Meat Science, 85(4), 759-767.
- Kirisattayakul, W., Wattanathorn, J., Tong-Un, T., Muchimapura, S., Wannanon, P., & Jittiwat, J. (2013). Cerebroprotective effect of *Moringa oleifera* against focal ischemic stroke induced by middle cerebral artery occlusion. Oxidative Medicine and Cellular Longevity, 2013, 1-10.
- Koutchma, T., Popović, V., Ros- Polski, V., & Popielarz, A. (2016). Effects of ultraviolet light and high- pressure processing on quality and health- related constituents of fresh juice products. Comprehensive Reviews in Food Science and Food Safety, 15(5), 844-867.
- Kudi, A. C., Umoh, J. U., Eduvie, L. O., & Gefu, J. (1999). Screening of some Nigerian medicinal plants for antibacterial activity. Journal of Ethnopharmacology, 67(2), 225-228.
- Kumar, P., Sharma, R., Ray, S., Mehariya, S., Patel, S. K., Lee, J. K., & Kalia, V. C. (2015). Dark fermentative bioconversion of glycerol to hydrogen by Bacillus thuringiensis. Bioresource Technology, 182, 383-388.
- Landrum, M. A. (2018). Microbial treatments to reduce or eliminate campylobacter from broiler carcasses, parts and livers (Doctoral dissertation, University of Georgia).

Lattanzio, V. (2013). Phenolic compounds: introduction. natural products, 1543–1580.

Le, N. T. T., Bach, L. G., Nguyen, D. C., Le, T. H. X., Pham, K. H., Nguyen, D. H., & Hoang Thi, T. T. (2019). Evaluation of factors affecting antimicrobial activity of bacteriocin from Lactobacillus plantarum microencapsulated in alginate-gelatin capsules and its application on pork meat as a bio-preservative. International Journal of Environmental Research and Public Health, 16(6), 1017.

- Lemon, K.P., Higgins, D.E., & Kolter, R. (2007). Flagellar motility is critical for *Listeria monocytogenes* biofilm formation. Journal of Bacteriology, 189, 4418-4424.
- Leygonie, C., Britz, T. J., & Hoffman, L. C. (2012). Impact of freezing and thawing on the quality of meat. Meat Science, 91(2), 93-98.
- Li, H. B., Cheng, K. W., Wong, C. C., Fan, K. W., Chen, F., & Jiang, Y. (2007). Evaluation of antioxidant capacity and total phenolic content of different fractions of selected microalgae. Food Chemistry, 102(3), 771-776.
- Li, T. T., Jalbani, Y. M., Zhang, G. L., Zhao, Z. Y., Wang, Z. Y., Zhao, X. Y., & Chen, A. L. (2019). Detection of goat meat adulteration by real-time PCR based on a reference primer. Food Chemistry, 277, 554-557.
- Li, Y., Zhang, Q., Zhang, J., Wu, L., Qi, Y., & Zhou, J. M. (2010). Identification of microRNAs involved in pathogen-associated molecular pattern-triggered plant innate immunity. Plant Physiology, 152(4), 2222-2231.
- Liu, R. H. (2004). Potential synergy of phytochemicals in cancer prevention: Mechanism of action, Journal of Nutrition, 134, 3479-3485.
- Maina, J. W. (2018). Analysis of the factors that determine food acceptability. The Pharma Innovation, 7(5, Part D), 253-257
- Malekmohammad, K., Sewell, R. D., & Rafieian-Kopaei, M. (2019). Antioxidants and atherosclerosis: mechanistic aspects. Biomolecules, 9(8), 301.
- Manessis, G., Kalogianni, A. I., Lazou, T., Moschovas, M., Bossis, I., & Gelasakis, A. I. (2020). Plant-derived natural antioxidants in meat and meat products. Antioxidants, 9(12), 1215.
- Manguro, L.O.A., & Lemmen. P. (2007). Phenolics of *Moringa oleifera* leaves. Natural Product Research, 21 56-68.
- Maqsood, S., Benjakul, S., Abushelaibi, A., & Alam, A. (2014). Phenolic compounds and plant phenolic extracts as natural antioxidants in prevention of lipid oxidation in seafood: A detailed review. Comprehensive Reviews in Food Science and Food Safety, 13(6), 1125-1140.
- Mashau, M.E., Ramatsetse, K.E., & Ramashia, S.E. (2021). Effects of adding moringa oleifera leaves powder on the nutritional properties, lipid oxidation and microbial growth in ground beef during cold storage. Applied Science, 11, 2944.
- Matic, I., Guidi, A., Kenzo, M., Mattei, M. & Galgani, A., (2018). Investigation of medicinal plants traditionally used as dietary supplements: A review on *Moringa oleifera*. Journal of Public Health in Africa, 9(3), 841.

- Matshediso, P. G., Cukrowska, E., & Chimuka, L. (2015). Development of pressurised hot water extraction (PHWE) for essential compounds from Moringa oleifera leaf extracts. Food Chemistry, 172, 423-427.
- Mazhangara, I. R., Chivandi, E., Mupangwa, J. F., & Muchenje, V. (2019). The potential of goat meat in the red meat industry. Sustainability, 11(13), 3671.
- Mbikay, M. (2012). Therapeutic potential of *Moringa oleifera* leaves in chronic hyperglycemia and dyslipidemia: a review. Frontiers in Pharmacology, 3, 24.
- McKeen, D., Pospelov, M. & Ritz, A., (2012). Modified Higgs branching ratios versus C P and lepton flavor violation. Physical Review D, 86(11), 113004.
- Medini, F., Fellah, H., Ksouri, R., & Abdelly, C. (2014). Total phenolic, flavonoid and tannin contents and antioxidant and antimicrobial activities of organic extracts of shoots of the plant *Limonium delicatulum*. Journal of Taibah University for Science, 8(3), 216-224.
- Mitterer-Daltoé, M., Bordim, J., Lise, C., Breda, L., Casagrande, M., & Lima, V. (2020). Consumer awareness of food antioxidants. synthetic vs. natural. Food Science and Technology, 41, 208-212.
- Mohamed, F. A., Heba H. Salama, S. M. El-Sayed, Hoda S. El-Sayed, & H. A. Zahran. (2018). Utilization of natural antimicrobial and antioxidant of *Moringa oleifera* leaves extract in manufacture of cream cheese. Journal of Biological Sciences, 18, 92-106.
- Moyo, B., Masika, P.J., Hugo, A. & Muchenje, V., 2011. Nutritional characterization of Moringa (*Moringa oleifera* Lam.) leaves. African Journal of Biotechnology, 10(60), 12925-12933.
- Muthukumar, M., Naveena, B.M., Vaithiyanathan, S., Sen, A.R., & Sureshkumar, K. (2014). Effect of incorporation of *Moringa oleifera* leaves extract on quality of ground pork patties. Journal of Food Science and Technology, 51(11), 3172-3180.
- Nafee, A. M., Pasha, H. F., Abd El Aal, S. M., & Mostafa, N. A. (2012). Clinical significance of serum clusterin as a biomarker for evaluating diagnosis and metastasis potential of viral-related hepatocellular carcinoma. Clinical Biochemistry, 45(13-14), 1070-1074.
- Nakyinsige, K., Fatimah, A. B., Aghwan, Z. A., Zulkifli, I., Goh, Y. M., and Sazili, A. Q. (2014). Bleeding efficiency and meat oxidative stability and microbiological quality of New Zealand White rabbits subjected to halal slaughter without stunning and gas stun-killing. Asian-Australasian Journal of Animal Sciences, 27, 406-413.

- Neveu, V., Perez-Jiménez, J., Vos, F., Crespy, V., du Chaffaut, L., Mennen, L., Knox, C., Eisner, R., Cruz, J., Wishart, D. & Scalbert, A., (2010). Phenol-Explorer: an online comprehensive database on polyphenol contents in foods. Database, 2010, bap204.
- Nielsen, H. B., Sonne, A. M., Grunert, K. G., Banati, D., Pollák-Tóth, A., Lakner, Z., & Peterman, M. (2009). Consumer perception of the use of high-pressure processing and pulsed electric field technologies in food production. Appetite, 52(1), 115-126.
- Nobossé, P., Fombang, E. N., & Mbofung, C. M. (2018). Effects of age and extraction solvent on phytochemical content and antioxidant activity of fresh *Moringa oleifera* L. leaves. Food Science and Nutrition, *6*(8), 2188-2198.
- Nychas, G. J. E., Skandamis, P. N., Tassou, C. C., & Koutsoumanis, K. P. (2008). Meat spoilage during distribution. Meat Science, 78(1-2), 77-89.
- Ogbe, A. O., & John P. Affiku. (2011). Proximate study, mineral and anti-nutrient composition of *Moringa oleifera* leaves harvested from Lafia, Nigeria: potential benefits in poultry nutrition and health. Journal of Microbiology, Biotechnology and Food Sciences, 1, 296-308.
- Ogbunugafor, H. A., Eneh, F. U., Ozumba, A. N., Igwo-Ezikpe, M. N., Okpuzor, J., Igwilo, I. O., & Onyekwelu, O. A. (2011). Physico-chemical and antioxidant properties of *Moringa oleifera* seed oil. Pakistan Journal of Nutrition, 10(5), 409-414.
- Okumu, M. O. (2016). Prophylactic Efficacy of *Moringa oleifera* leaf extracts against liver injury induced by artesunate-amodiaquine antimalarial combination (Doctoral dissertation, University of Nairobi).
- Onsare, J. G., & Arora, D. S. (2015). Antibiofilm potential of flavonoids extracted from *Moringa oleifera* seed coat against *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Candida albicans*. Journal of Applied Microbiology, 118, 313-325.
- Oyeyinka, A. T., & Oyeyinka, S. A. (2018). *Moringa oleifera* as a food fortificant: Recent trends and prospects. Journal of the Saudi Society of Agricultural Sciences, 17(2), 127-136.
- Pal, M., Devrani, M., & Pinto, S. (2018). Significance of hygienic processing of milk and dairy products. Madridge Journal of Food Technology, 3, 133-137.
- Pandjaitan, N., Howard, L. R., Morelock, T., & Gil, M. I. (2005). Antioxidant capacity and phenolic content of spinach as affected by genetics and maturation. Journal of Agricultural and Food Chemistry, 53(22), 8618-8623.
- Pathare, P. B., & Roskilly, A. P. (2016). Quality and energy evaluation in meat cooking. Food Engineering Reviews, 8(4), 435-447.

- Peixoto, C., Cruz, T., & Peixoto, M.F. (2011). Análise quantitativa do crescimento de plantas: conceitos e prática. Enciclopédia Biosfera, 7,13.
- Pinelo, M., Del Fabbro, P., Manzocco, L., Nuñez, M. J., & Nicoli, M. C. (2005). Optimization of continuous phenol extraction from Vitis vinifera byproducts. Food Chemistry, 92(1), 109-117.
- Polacek, N., & Ivanov, P. (2020). The regulatory world of tRNA fragments beyond canonical tRNA biology. RNA Biology, 17, 1057-1059.
- Prüss-Ustün, A., Vickers, C., Haefliger, P., & Bertollini, R. (2011). Knowns and unknowns on burden of disease due to chemicals: a systematic review. Environmental Health, 10(1), 1-15.
- Rahman, M. H., Alam, M. S., Monir, M. M., & Rahman, S. M. E. (2020). Effect of Moringa oleifera leaf extract and synthetic antioxidant on quality and shelf-life of goat meat nuggets at frozen storage. International Journal of Food Research, 7, 34-45.
- Ramanathan, R., Hunt, M.C., Mancini, R.A., Nair, M.N., Denzer, M.L., Suman, S.P. & Mafi, G.G., (2020). Recent updates in meat color research: Integrating traditional and high-throughput approaches. Meat and Muscle Biology, 4(2), 9598.
- Resconi, V., Escudero, A., & Campo, M. (2013). The development of aromas in ruminant meat. Molecules, 18(6), 6748–6781.
- Richter, N., Siddhuraju, P., & Becker, K. (2003). Evaluation of nutritional quality of Moringa (*Moringa oleifera* Lam.) leaves as an alternative protein source for Nile tilapia (*Oreochromis niloticus* L.). Aquaculture, 217(1), 599-611.
- Rockwood, J.L., Anderson, B.G., Casamatta, D.A. (2013). Potential uses of Moringa oleifera and an examination of antibiotic efficacy conferred by *M. oleifera* seed and leaf extracts using crude extraction techniques available to underserved indigenous populations, International Journal of Phytotherapy Research, 3 61–71.
- Ross, A. C., Taylor, C. L., Yaktine, A. L., & Del Valle, H. B. (2011). Overview of calcium. in dietary reference intakes for calcium and vitamin D. National Academies Press (US).
- Saeed, M., Alagawany, M., Fazlani, S. A., Kalhoro, S. A., Naveed, M., Ali, N., & Chao, S. (2019). Health promoting and pharmaceutical potential of ferulic acid for the poultry industry. World's Poultry Science Journal, 75(1), 83-92.
- Saini, R.K., Sivanesan, I. & Keum, Y.S., (2016). Phytochemicals of *Moringa oleifera*: a review of their nutritional, therapeutic and industrial significance. 3 Biotech, 6(2),1-14.

- Sajjadi, B., Chen, W. Y., Raman, A. A. A., & Ibrahim, S. (2018). Microalgae lipid and biomass for biofuel production: A comprehensive review on lipid enhancement strategies and their effects on fatty acid composition. Renewable and Sustainable Energy Reviews, 97, 200-232.
- Santos, A. F., Argolo, A. C., Paiva, P. M., & Coelho, L. C. (2012). Antioxidant activity of *Moringa oleifera* tissue extracts. Phytotherapy Research, 26(9), 1366-1370.
- Santos-Sánchez, N. F., Salas-Coronado, R., Villanueva-Cañongo, C., & Hernández-Carlos, B. (2019). Antioxidant compounds and their antioxidant mechanism (pp. 1-28). London, UK: Intech Open.
- Sapperstein, S. K., Walter, D. M., Grosvenor, A. R., Heuser, J. E., & Waters, M. G. (1995). p115 is a general vesicular transport factor related to the yeast endoplasmic reticulum to Golgi transport factor Uso1p. Proceedings of the National Academy of Sciences, 92(2), 522-526.
- Shah, M. A., Bosco, S. J. D., & Mir, S. A. (2014). Plant extracts as natural antioxidants in meat and meat products. Meat Science, 98(1), 21-33.
- Shahidi, F., Liyana-Pathirana, C.M. & Wall, D.S., (2006). Antioxidant activity of white and black sesame seeds and their hull fractions. Food Chemistry, 99(3),478-483.
- Sheridan, R., Hoffman, L. C., & Ferreira A. V. (2003). Meat quality of boer goat kids and mutton merino lambs: Commercial yields and chemical composition. Animal Science, 76, 63-71.
- Sherwin, H. W., & Farrant, J. M. (1998). Protection mechanisms against excess light in the resurrection plants Craterostigma wilmsii and Xerophyta viscosa. Plant Growth Regulation, 24(3), 203-210.
- Siddhuraju, P., & Becker, K. (2003). Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera* Lam.) leaves. Journal of Agricultural and Food Chemistry, 51(8), 2144-2155.
- Silva, E. M., Rogez, H., & Larondelle, Y. (2007). Optimization of extraction of phenolics from *Inga edulis* leaves using response surface methodology. Separation and Purification Technology, 55(3), 381-387.
- Singh, B.N., Singh, B.R., Singh, R.L., Prakash, D., Dhakarey, R., Upadhyay, G. and Singh, H.B., 2009. Oxidative DNA damage protective activity, antioxidant and anti-quorum sensing potentials of *Moringa oleifera*. Food and Chemical Toxicology, 47(6),1109-1116.
- Singh, T.P., Singh, P., & Kumar, P. (2015). Drumstick (*Moringa oleifera*) as a food additive in livestock products. Nutrition and Food Science, 45, 423-432.

- Smet, C., Noriega, E., Rosier, F., Walsh, J. L., Valdramidis, V. P., & Van Impe, J. F. (2016). Influence of food intrinsic factors on the inactivation efficacy of cold atmospheric plasma: Impact of osmotic stress, suboptimal pH and food structure. Innovative Food Science and Emerging Technologies, 38, 393-406.
- Sreelatha, S., & Padma, P. R. (2009). Antioxidant activity and total phenolic content of *Moringa oleifera* leaves in two stages of maturity. Plant Foods for Human Nutrition, 64(4), 303-311.
- Sreelatha, S., & Padma, P.R. (2010). Protective mechanisms of *Moringa oleifera* against CCl4-induced oxidative stress in precision-cut liver slices. Forsch Komplementmed,17(4):189-194.
- Sultana, B., Anwar, F., Asi, M. R., & Chatha, S. A. S. (2008). Antioxidant potential of extracts from different agro wastes: Stabilization of corn oil. Grasas y Aceites, 59, 205-217.
- Sun, Q., Zhao, X., Zhang, C., Xia, X., Sun, F., & Kong, B. (2019). Ultrasound-assisted immersion freezing accelerates the freezing process and improves the quality of common carp (*Cyprinus carpio*) at different power levels. LWT, 108, 106-112.
- Sun, X. D., & Holley, R. A. (2010). High hydrostatic pressure effects on the texture of meat and meat products. Journal of Food Science, 75(1), R17-R23.
- Suphachai, C. (2014). Antioxidant and anticancer activities of Moringa oleifera leaves. Journal of Medicinal Plants Research, 8(7), 318-325.
- Sutalangka, C., Wattanathorn, J., Muchimapura, S., & Thukham-mee, W. (2013). Moringa oleifera mitigates memory impairment and neurodegeneration in animal model of age-related dementia. Oxidative Medicine and Cellular Longevity, 23, 695936.
- Tangkham, W. (2018). Evaluation of 4-methyloctanoic acid compound in goat meat. Annals of Short Reports, 1, 1021.
- Tapia, M. S., Alzamora, S. M., & Chirife, J. (2020). Effects of water activity (a<sub>w</sub>) on microbial stability as a hurdle in food preservation. Water Activity in Foods:Fundamentals and Applications, 323-355.
- Tashkin, D. P., Baldwin, G. C., Sarafian, T., Dubinett, S., & Roth, M. D. (2002). Respiratory and immunologic consequences of marijuana smoking. The Journal of Clinical Pharmacology, 42(S1), 71S-81S.
- Thamban, C., Mathew, A. C., & Jaganathan, D. (2017). Drip irrigation for sustainable coconut farming-institutional and technology perspectives. Indian Coconut Journal, 59(11), 5-9.

- Thurber, M. D., & Fahey, J. W. (2009). Adoption of Moringa oleifera to combat undernutrition viewed through the lens of the "Diffusion of Innovations" theory. Ecology of Food and Nutrition, 48(3), 212-225.
- Truong, D. H., Nguyen, D. H., Ta, N. T. A., Bui, A. V., Do, T. H., & Nguyen, H. C. (2019). Evaluation of the use of different solvents for phytochemical constituents, antioxidants, and in vitro anti-inflammatory activities of Severinia buxifolia. Journal of Food Quality, 3, 8178294.
- Twinomuhwezi, H., Awuchi, C. G., & Rachael, M. (2020). Comparative study of the proximate composition and functional properties of composite flours of amaranth, rice, millet, and soybean. American Journal of of Food Science and Nutrition, 6, 6-19.
- Uçar, A., Yilmaz, M. V., & Cakiroglu, F. P. (2016). Food safety-problems and solutions. Significance, Prevention and Control of Food Related Diseases, 1-26.
- Udechukwu, M. C., Abbey, L., Nwodo, U., & Udenigwe, C. C. (2018). Potential of *Moringa oleifera* seeds and leaves as functional food ingredients for human health promotion. Journal of Food and Nutrition Research, 57(1), 1-14.
- Valdez-Solana, M. A., Mejia-Garcia, V. Y., Tellez-Valencia, A., Garcia-Arenas, G., Salas-Pacheco, J., Alba-Romero, J. J., & Sierra-Campos, E. (2015). Nutritional content and elemental and phytochemical analyses of *Moringa oleifera* grown in Mexico. Journal of Chemistry, 2015, 860381
- Vázquez-León, L. A., Páramo-Calderón, D. E., Robles-Olvera, V. J., Valdés-Rodríguez, O. A., Pérez-Vázquez, A., García-Alvarado, M. A., & Rodríguez-Jimenes, G. C. (2017). Variation in bioactive compounds and antiradical activity of *Moringa oleifera* leaves: influence of climatic factors, tree age, and soil parameters. European Food Research and Technology, 243(9), 1593-1608.
- Verbeke, W., Marcu, A., Rutsaert, P., Gaspar, R., Seibt, B., Fletcher, D., & Barnett, J. (2015). Would you eat cultured meat?': Consumers' reactions and attitude formation in Belgium, Portugal and the United Kingdom. Meat Science, 102, 49-58.
- Verma, A.K., Chatli, M.K., Kumar, D., Kumar, P., & Mehta N. (2015). Efficacy of sweet potato powder and added water as fat replacer on the quality attributes of low-fat pork patties. Asian-Australasian Journal of Animal Science, 28(2), 252-259.
- Vigne, J. D. (2011). The origins of animal domestication and husbandry: a major change in the history of humanity and the biosphere. Comptes Rendus Biologies, 334(3), 171-181.

- Vongsak, B., Sithisarn, P., Mangmool, S., Thongpraditchote, S., Wongkrajang, Y., & Gritsanapan, W. (2013). Maximizing total phenolics, total flavonoids contents and antioxidant activity of *Moringa oleifera* leaf extract by the appropriate extraction method. Industrial crops and products, 44, 566-571.
- Wan, Y. J., Zhu, P. L., Yu, S. H., Sun, R., Wong, C. P., & Liao, W. H. (2018). Anticorrosive, ultralight, and flexible carbon- wrapped metallic nanowire hybrid sponges for highly efficient electromagnetic interference shielding. Small, 14(27), 1800534.
- Wang, Q. Q., Gonell, S., Leenders, S. H., Dürr, M., Ivanović-Burmazović, I., & Reek, J. N. (2016). Self-assembled nanospheres with multiple endohedral binding sites pre-organize catalysts and substrates for highly efficient reactions. Nature Chemistry, 8(3), 225.
- Warner, R. D. (2017). The eating quality of meat—IV water-holding capacity and juiciness: Lawrie's meat science (pp. 419–459). Kidlington, England: Woodhead Publishing.
- Webb, E. C. (2014). Goat meat production, composition, and quality. Animal Frontiers, 4(4), 33–37.
- Webb, E., Casey, N., & Simela, L. (2005). Goat meat quality. Small Ruminant Research, 60(1-2), 153-166.
- Weston, A. R., Rogers, R. W., & Althen, T. G. (2002). The role of collagen in meat tenderness. The Professional Animal Scientist, 18(2), 107-111.
- Whiting, R. C., & Jenkins, R. K. (1981). Comparison of rabbit, beef, and chicken meats for functional properties and frankfurter processing. Journal of Food Science, 46, 1693-1696.
- Whitnall, T., & Pitts, N. (2019). Global trends in meat consumption. Agricultural Commodities, 9(1), 96.
- Xing, T., Gao, F., Tume, R. K., Zhou, G., & Xu, X. (2019). Stress effects on meat quality: a mechanistic perspective. Comprehensive Reviews in Food Science and Food Safety, 18(2), 380-401.
- Yahia, E. M., Maldonado Celis, M. E., & Svendsen, M. (2017). The contribution of fruit and vegetable consumption to human health. Fruit and Vegetable Phytochemicals: Chemistry and Human Health, 2nd Edition, 1-52.USA.
- Yang, R. Y., Chang, L. C., Hsu, J. C., Weng, B. B., Palada, M. C., Chadha, M. L., & Levasseur, V. (2006). Nutritional and functional properties of Moringa leaves– From germplasm, to plant, to food, to health. Moringa leaves: Strategies, standards and markets for a better impact on nutrition in Africa. Moringanews, CDE, CTA, GFU. Paris.

- Yuan Y.V., Bone D.E., Carrington M.F. (2005) Antioxidant activity of dulse (*Palmaria palmata*) extract evaluated in vitro. Food Chemistry. 91: 485-494.
- Zaku, S. G., Emmanuel, S., Tukur, A. A., & Kabir, A. (2015). *Moringa oleífera*: An underutilized tree in Nigeria with amazing versatility: A Review. African Journal of Food Science, 9(9), 456-461.
- Zhang, W., Naveena, B. M., Jo, C., Sakata, R., Zhou, G., Banerjee, R., & Nishiumi, T. (2017). Technological demands of meat processing–An Asian perspective. Meat Science, 132, 35-44.
- Zhao, S., & Zhang, D. (2013). Supercritical fluid extraction and characterisation of essential oil from *Moringa oleifera* leaves. Separation and Purification Technology, 118, 497–502.
- Zhou, W., Liu, Q., Zhu, Z., & Zhang, J. (2010). Preparation and properties of vanadium-doped TiO2 photocatalysts. Journal of Physics D: Applied Physics, 43(3), 035301.
- Zimmermann, P., & Zentgraf, U. (2005) The correlation between oxidative stress and leaf senescence during plant development. Cell and Molecular Biology Letters, 10, 515–534.
- Zupan, A., Mikulic-Petkovsek, M., Slatnar, A., Stampar, F., & Veberic, R. (2014). Individual phenolic response and peroxidase activity in peel of differently sunexposed apples in the period favorable for sunburn occurrence. Journal of plant Physiology, 171(18), 1706-1712.