

## CAPSULAR SEROTYPE, ANTIBIOTICS SUSCEPTIBILITY AND RESISTANCE GENE PROFILE OF *Streptococcus pneumoniae* FROM NASAL CARRIAGE OF HEALTHY CHILDREN IN SELANGOR, MALAYSIA



By

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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirements for the Degree of Master of Science

November 2022

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

### CAPSULAR SEROTYPE, ANTIBIOTICS SUSCEPTIBILITY AND RESISTANCE GENE PROFILE OF Streptococcus pneumoniae FROM NASAL CARRIAGE OF HEALTHY CHILDREN IN SELANGOR, MALAYSIA

By

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November 2022

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Research on pneumococcal carriage epidemiology are very limited in Malaysia and some other developed countries. This is because most study always focus on symptomatic diseases for prevention and control, meanwhile high level of asymptomatic carriage rate is the starting point of pneumococcal infections. This factors interest us to research on pneumococcal carriage rate, its virulence genes, serotypes distribution, antimicrobial susceptibility pattern and macrolide resistance determinant genes. Three hundred and ten nasal swab samples were collected from healthy children of both  $\leq$ 5 years old and >5 years old with the consent of their parents and guardians. The samples were collected from three selected child care centre; kindergarten, orphanage home and refugee school in Selangor and Kuala Lumpur. Sixty isolates of Streptococcus pneumoniae were recovered from all nasal swabs making the carriage rate of 26.3% for ≤5 years old and 10.4% for >5 years old children respectively. All isolates were carrying the virulence genes i.e 63.3% (38) of ply and lytA gene whereas 36.7% (22) carrying a single gene of either ply or lytA. Among the 22 single gene carrying isolates, we selected 12 lytA gene isolates for 16s rRNA sequence analysis for further confirmation of Streptococcus pneumoniae strain and only 7 isolates were of Streptococcus pneumoniae strain, making the total carriage isolates 17.7% (55). We successfully serotyped the 55 isolates by multiplex PCR techniques with the following descending prevalence sequence result: 7A/7F, 23F, 19F, 11A/D and 15B/15C. The vaccine serotype was accounted for 71% and the non-vaccine serotype was 29%. Antibiotic susceptibility testing showed cefotaxime and ceftriaxone were the most susceptible antibiotics to Streptococcus pneumoniae with 50.9% and 49% respectively, erythromycin susceptible Streptococcus pneumoniae has 38.2%, penicillin on the other hand has 20% susceptibility rate. Despite penicillin having the highest rate of non-susceptibility (80%), erythromycin has the highest rate of resistance with 43.6%. Further study on macrolide resistance determinant genes revealed mefA gene has the higher rate of 67.2% (37) while ermB gene has 25.5% (14) and mefA+ermB has 7.3% (4). This proved that macrolide resistance gene that highly triggers the high resistance level of erythromycin is mefA. Pneumococcal carriage

rate has decreased over time from previous studies. Serotype analysis has shown the importance of vaccinating the children to reduce the carriage rate and infection. Antibiotics of interest on the other hand should always be tested before prescription, so as to avoid misuse, overuse and unsuitable antibiotics. Continuous study is needed to keep track of pneumococcal carriage rate with serotype distribution, and to understand the antibiotics susceptibility pattern. If the resistant rate is higher, it is crucial to analyse the resistance determinant gene with further molecular study for comprehensive knowledge of *Streptococcus pneumoniae* in Malaysia.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

### SEROTYPE CAPSULAR, ANTIBIOTIK KENDERAAN DAN RINTANGAN PROFIL GENE Streptococcus pneumoniae DARIPADA PENGANGKUTAN NASAL KANAK-KANAK SIHAT DI SELANGOR, MALAYSIA

Oleh

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Penyelidikan tentang epidemiologi pembawa pneumokokus sangat terhad di Malaysia dan beberapa negara maju yang lain. Ini kerana kebanyakan kajian memfokuskan kepada penyakit bergejala untuk pencegahan dan kawalan, manakala kadar pembawa tanpa gejala yang tinggi adalah titik permulaan jangkitan pneumokokus. Faktor ini menarik minat kami untuk menyelidik tentang kadar pembawa pneumokokus, gen virulensinya, taburan serotaip, corak kerentanan antimikrob dan gen penentu rintangan makrolid. Tiga ratus sepuluh sampel swab hidung telah dikumpul daripada kanak-kanak yang sihat berumur ≤5 tahun dan >5 tahun dengan persetujuan ibu bapa dan penjaga mereka. Sampel dikumpul dari tiga pusat jagaan kanak-kanak terpilih; tadika, rumah anak yatim dan sekolah pelarian di Selangor dan Kuala Lumpur. Enam puluh isolat S.pneumoniae telah dikenalpasti daripada kesatan hidung kanak2 berdasarkan pengamatan pertumbuhan isolat dan ujian biokimia menjadikan kadar pembawa 26.3% untuk <5tahun dan 10.4% untuk kanak-kanak berumur >5tahun. Semua isolat membawa gen virulensi iaitu 63.3% (38) ply dan gen lytA manakala 36.7% (22) membawa gen tunggal sama ada ply atau lytA. Antara 22 gen tunggal yang membawa isolat, kami memilih 12 isolat gen lytA untuk analisis jujukan rRNA 16s bagi pengesahan lanjut strain Streptococcus pneumoniae. Daripada 12 isolat, hanya 7 disahkan sebagai strain S.pneumoniae, menjadikan kadar pembawa adalah 17.7%(55) di kalangan kanak-kanak. Ujian PCR multipleks terhadap 55 strain S.pneumoniae memberikan hasil jujukan prevalen serotaip secara menurun secara berikut : 7A/7F, 23F, 19F, 11A/D dan 15B/15C. Pecahan jenis serotaip yang terkandung dalam vaksin adalah 71% manakala 29% merupakan serotaip yang tidak terkandung dalam vaksin konjugat. Ujian kerentanan terhadap empat antibiotik menunjukkan cefotaxime dan ceftriaxone adalah antibiotik yang paling rentan kepada Streptococcus pneumoniae dengan 50.9% dan 49% masingmasing, manakala kadar kerentanan erythromycin adalah 38.2% dan penicillin hanya 20%. Walaupun penicillin mempunyai kadar ketidakrentanan (non-susceptible) tertinggi (80%), erythromycin menunjukkan kadar rintangan tertinggi iaitu 43.6%. Kajian lanjut mengenai gen rintangan maklolid menunjukkan gen mefA yang tinggi iaitu 67.2% (37) manakala gen *ermB* adalah 25.5% (14) dan *mefA+ermB* 7.3% (4). Ini menunjukkan *mefA* adalah gen yang mencetuskan tahap rintangan tinggi terhadap erythromycin. Kajian ini menunjukkan kadar pembawa bakteria pneumokokus telah menurun berbanding kajian lepas. Analisis serotaip juga menunjukkan kepentingan vaksin pneumokokus di kalangan kanak-kanak untuk mengurangkan kadar pembawa dan jangkitan penyakit. Antibiotik sebaliknya hendaklah sentiasa diuji sebelum preskripsi, untuk mengelakkan penyalahgunaan, penggunaan berlebihan dan antibiotik yang tidak sesuai. Kajian berterusan diperlukan untuk menjejaki kadar pembawa pneumokokus berserta taburan serotaip, dan memahami taburan kerentanan antibiotik. Jika kadar rintangan lebih tinggi, adalah penting untuk menganalisis gen penentu rintangan seterusnya membuat kajian lanjutan secara terperinci. untuk pengetahuan menyeluruh tentang *Streptococcus pneumoniae* di Malaysia.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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# LIST OF ABBREVIATIONS

	<sup>0</sup> C	Degree Celcius
	%	Percentage
	≤</td <td>Less than</td>	Less than
	>/≥	Greater than
	μg	Microgram
	μΙ	Microlitre
	μm	Micrometre
	AIDS	Acquire Immunodeficiency Syndrome
	ANSORP	Asian Network for Surveillance of Resistant Pathogens
	AMR	Antimicrobial Resistance
	ART	Antiretroviral Therapy
	AST	Antimicrobial Susceptibility Testing
	ATCC	American Type Culture Collection
	BHI	Brain Heart Infusion
	BLAST	Basic Local Alignment Search Tool
E	Вр	Base point
	САР	Community Acquire Pneumonia
	cbpA	Choline binding protein A
	CC	Clonal Complex
	CFU	Colony Forming Unit
	CLSI	Clinical and Laboratory Standards Institutes
	$CO_2$	Carbon dioxide
	CRO	Ceftriaxone
	СТХ	Cefotaxime
	DNA	Deoxyribonucleic acid

DRSP	Drug Resistance Streptococcus pneumoniae
E-test	Epsilometer test
ENSSP	Erythromycin Non-susceptible Streptococcus pneumoniae
ERSP	Erythromycin Resistance Streptococcus pneumoniae
ermB	Erythromycin Ribosomal Methylase B
ERY	Erythromycin
ESSP	Erythromycin Susceptible Streptococcus pneumoniae
HGT	Horizontal gene transfer
HIV	Human Immunodeficiency Virus
I	Intermediate
ICT	Information Communication Technology
IPD	Invasive Pneumococcal Diseases
JKEUPM	Ethic Committee for Research involving Human Subject
KL	Kuala Lumpur
lytA	Pneumococcal autolysin A
MDR	Multidrug resistant
MDRSP	Multidrug resistant Streptococcus pneumoniae
mefA	Macrolide efflux protein
MIC	Minimum Inhibitory Concentration
MLST	Multi-Locus Sequence Typing
Mm	Millimetre
NC	Negative control
NCBI	National Center for Biotechnology Information
NESp	Non-encapsulated Streptococcus pneumoniae
NIP	National Immunization Programme
NT	Non-typeable
NTP	Non-typeable pneumococcus

NVT	Non-vaccine serotypes
PAF	Platelet-activating factor
PC	Positive control
PCR	Polymerase Chain Reaction
PCV	Pneumococcal conjugate vaccine
PCV7	7-valent conjugate vaccine
PCV10	10-valent conjugate vaccine
PCV13	13-valent conjugate vaccine
PEN	Penicillin
PFGE	Pulsed-Field Gel Electrophoresis
РН	Potential of Hydrogen
PICU	Pediaetric Intensive Care Unit
Ply	Pneumolysin
PPV	Pneumococcal Polysaccharide Vaccine
PRSP	Penicillin resistant Streptococcus pneumoniae
psaA	Pneumococcal surface adhesin A
pspA	Pneumococcal surface protein A
pspC	Pneumococcal surface protein C
R	Resistant
S	Susceptible
S.pneumoniae	Streptococcus pneumoniae
Sp.	Specie
Spp.	Species
ST	Sequence type
UK	United Kingdom
UPM	University Putra Malaysia
URT	Upper Respiratory Tract

UV Ultraviolet radiation

VT Vaccine serotype

 $\bigcirc$ 

WHO World Health Organization



#### **CHAPTER 1**

### INTRODUCTION

### 1.1 Background of the Study

Being a pathogenic bacterium, numerous research has been conducted in developing vaccines and antibiotics for *Streptococcus pneumoniae* prevention and treatment. However, *Streptococcus pneumoniae* is known to develop resistant gene for many of the proposed medications. Due to this, contemporary researches, including this study, are investigating how *Streptococcus pneumoniae* is becoming resistant to antibiotics and what determinant genes of macrolide triggers their prevalence (Schroeder and Stephens, 2016). It is one of the most gravitating infectious disease organism that happens to range from 75.4% (75,400) out of 100,000 hospitalized patients among Ministry of Health hospitals in Malaysia in the year 2010 (Le *et al.*, 2012) as it ranked 25<sup>th</sup> among 99 pathogenic bacteria and pneumonia happens to be the 4<sup>th</sup> leading cause of death (Song, Moon H Nahm and Moseley, 2013). In another study, report analyzed that majority of <2years old hospitalized children and >50 years old adults in Malaysia are infected with *Streptococcus pneumoniae* (Le *et al.*, 2012) and this makes pneumococci serve as one of the major cause of hospitalization in Malaysia.

Also known as pneumococcal, *Streptococcus pneumoniae* is a bacterium that colonizes the human nasopharynx and cause invasive and non-invasive diseases in adults, young adults and children of less than five years of age (Sohail *et al.*, 2018), majorly affected are the infant whose immune system have not be immunized. Nasal cavity is the uppermost part of the respiratory system that is divided into respiratory and olfactory segment and serves as the habitat for microflora and pathogenic organisms that lives on the nasal cavity as colonizers of which isolation frequently takes place as carriage isolates. *Streptococcus pneumoniae* is one the nasal cavity colonizer that brings in disease manifestation; some other related colonizers are *Staphylococcus aureus*, *Haemophilus influenzae*, *Moraxella cattarrhalis* and *Neisseria meningitides*. Pneumococci on its own end enables the colonization by its virulence factor activity that enhance its ability to evade the early components of the human immune response leading to pneumonia, otitis media, sinusitis, meningitis and septicemia diseases (Kadioglu *et al.*, 2008). All of which propagate to high rate of mortality and morbidity (Izurieta *et al.*, 2018).

*Streptococcus pneumoniae* are gram positive bacteria that belongs to *Firmicutes phylum* with a cell wall that plays an important key roles in it shape, growth, cell division and interactions with the component of human host. Pneumococci has it means of being differentiated from other *Streptococcus spp* with it alpha hemolysis pattern on sheep blood agar. It is also an encapsulated and non-capsulated diplococcus bacterial with close to 98 serotypes which serves as it main virulence factor in their capsular polysaccharide (Arushothy *et al.*, 2019). All these serotypes does not cause severe pneumococcal diseases but differ in invasiveness and non-invasiveness, age, geographical distribution,

antimicrobial resistance, and time. With its over 90 serotypes varying geographically and increase in resistance (Arushothy *et al.*, 2019), research and investigation on serotypes distribution severity and antimicrobial resistance of pneumococcus in Malaysia keep increasing rapidly (Le *et al.*, 2011). Moreover, Pneumococcal surface protein A(PspA), Pneumococcal surface protein C (PspC), Pneumococcal surface adhesin A(PsaA), Choline binding protein A(CbpA), pneumolysin (ply), autolysin (LytA), hyaluronidase, neuraminidase are the pneumococcal proteins and enzymes that precipitate the prevalence of pneumococcal diseases as virulence factors (Song, Moon H. Nahm and Moseley, 2013).

### 1.2 Problem Statement

Regardless of the availability of antimicrobial therapy and vaccination program, pneumococcal infection keeps prevailing (Cornick and Bentley, 2012) and based on clinical failure, macrolide are associated with the treatment of pneumococcal infections (Schroeder and Stephens, 2016). Pneumococcal serotypes also differ in geographic areas, age, time and period of distribution (Henriques-normark and Tuomanen, 2013). Thus, this increases the rate of macrolide resistance as the bacteriostatics antibiotics became widely use and variation in serotype which have impact on the vaccination. We approach this research for proper and further studies on the serotype distributions and trends of antibiotics susceptibility pattern as well as distribution of the resistance gene on macrolide antibiotics of *Streptococcus pneumoniae*.

## 1.3 Research Hypothesis

- H1. The carriage differs according to age, time and location.
- H2. *Streptococcus pneumoniae* possess the virulence properties of autolysin (*lytA*) and pneumolysin (*ply*).
- H3. The capsular serotype differs in antibiotics susceptibility and resistance gene profile.
- H4. Antibiotics susceptibility pattern and macrolide resistance genes of *Streptococcus pneumoniae* differs in rate.

#### 1.4 Objectives

## **1.4.1 General Objective**

To characterize the capsular serotype, antibiotics susceptibility pattern and the genetic analysis of macrolide resistance to *Streptococcus pneumoniae* in children carriage.

## 1.4.2 Specific Objectives

- a) To identify the proportion of pneumococcal nasal carriage among health children from some selected nursery homes in Malaysia.
- b) To determine the difference in serotype distribution of the nasal carriage isolates of *Streptococcus pneumoniae* by multiplex PCR
- c) To determine the minimum inhibitory concentration (MIC) profile of antibiotics against *Streptococcus pneumoniae* isolates using E-test method.
- d) To determine the macrolide-resistance determinant genes of *Streptococcus pneumoniae* isolates by PCR.
- e) To relate macrolide-resistance genes and erythromycin susceptibility profile of *Streptococcus pneumoniae* isolates.

## 1.5 Thesis Organization

This thesis is organized as follows:

- **Chapter 1** is an introductory into the domain of the research where the aim, objectives, hypothesis and problem statements are presented.
- Chapter 2 provides a comprehensive review on the previous and current state of *Streptococcus pneumoniae* regarding its carriage rate, serotype distribution, antibiotics susceptibility and macrolide resistance genes.
- **Chapter 3** presents the methodology of the research defining suitable techniques of microbiological experimentation and molecular methods.
- Chapter 4 describes the resulting outcome of the method used on the research.
- **Chapter 5** discusses the out of the research methodology and results in relation with previously related research.
- **Chapter 6** concludes the research with the summarization of the research achievements, conclusions, potential enhancements and future recommended investigation.

#### REFERENCES

- Abdullah, N. *et al.* (2014) 'The development of a 16S rRNA gene based PCR for the identification of Streptococcus pneumoniae and comparison with four other species specific PCR assays', (April 2010). doi: 10.1186/1471-2334-10-104.
- Al-tawfiq, J. A. *et al.* (2016) 'Prevention of pneumococcal infections during mass gathering Prevention of pneumococcal infections during mass gathering', 5515. doi: 10.1080/21645515.2015.1058456.
- Al-Tawfiq, J. A. and Memish, Z. A. (2016) 'Prevention of pneumococcal infections during mass gathering', *Human Vaccines and Immunotherapeutics*, 12(2), pp. 326–330. doi: 10.1080/21645515.2015.1058456.
- Ammar et al. (2017) 'crossm Capsule Type and Amount Affect Streptococcus pneumoniae', 8(4), pp. 1–12.
- Anderson, R. *et al.* (2015) 'Key virulence factors of Streptococcus pneumoniae and nontypeable Haemophilus infuenzae : roles in host defence and immunisation Key virulence factors of Streptococcus pneumoniae and roles in host defence and immunisation', 8782. doi: 10.1080/10158782.2011.11441412.
- Arushothy, R. et al. (2019) 'International Journal of Infectious Diseases Pneumococcal serotype distribution and antibiotic susceptibility in Malaysia: A four-year study (2014 – 2017) on invasive paediatric isolates', *International Journal of Infectious Diseases*. International Society for Infectious Diseases, 80, pp. 129– 133. doi: 10.1016/j.ijid.2018.12.009.
- Arushothy, R. et al. (2020) 'International Journal of Infectious Diseases Multidrugresistant Streptococcus pneumoniae causing invasive pneumococcal disease isolated from a paediatric patient', *International Journal of Infectious Diseases*. International Society for Infectious Diseases, 90, pp. 219–222. doi: 10.1016/j.ijid.2019.10.037.
- Bao, Y. et al. (2019) 'The changing phenotypes and genotypes of invasive pneumococcal isolates from children in Shenzhen during 2013 – 2017', Vaccine. Elsevier Ltd, 37(49), pp. 7248–7255. doi: 10.1016/j.vaccine.2019.09.069.

BC Center for Disease Control (2012) 'Pneumococcal Infection', pp. 11-13.

Bradshaw, J. L. and Mcdaniel, L. S. (2019) 'Selective pressure: Rise of the nonencapsulated pneumococcus', pp. 1–6.

Carbajal, I. et al. (2018) 'Streptococcus pneumoniae', pp. 3-6.

Carvalho, G. *et al.* (2010) 'Revisiting Pneumococcal Carriage by Use of Broth Enrichment and PCR Techniques for Enhanced Detection of Carriage and Serotypes □', 48(5), pp. 1611–1618. doi: 10.1128/JCM.02243-09.

- Chaguza, C. *et al.* (2016) 'Recombination in Streptococcus pneumoniae Lineages Increase with Carriage Duration and Size of the Polysaccharide Capsule', 7(5). doi: 10.1128/mBio.01053-16.Editor.
- Chaguza, C., Cornick, J. E. and Everett, D. B. (2015) 'Mechanisms and impact of genetic recombination in the evolution of Streptococcus pneumoniae', *CSBJ*. Elsevier B.V., 13, pp. 241–247. doi: 10.1016/j.csbj.2015.03.007.
- Chen, H., Hsu, M. and Wu, T. (2018) 'ScienceDirect Non-typeable Streptococcus pneumoniae infection in a medical center in Taiwan after wide use of pneumococcal conjugate vaccine', *Journal of Microbiology, Immunology and Infection.* Elsevier Taiwan LLC, pp. 1–5. doi: 10.1016/j.jmii.2018.04.001.
- Cornick, J. E. and Bentley, S. D. (2012) 'Streptococcus pneumoniae: The evolution of antimicrobial resistance to beta-lactams, fluoroquinolones and macrolides', *Microbes and Infection*. Elsevier Masson SAS, 14(7–8), pp. 573–583. doi: 10.1016/j.micinf.2012.01.012.
- Diana, N. *et al.* (2020) 'International Journal of Medical Microbiology Pilus islets and the clonal spread of piliated Streptococcus pneumoniae: A review', *International Journal of Medical Microbiology*. Elsevier GmbH, 310(7), p. 151449. doi: 10.1016/j.ijmm.2020.151449.
- Dilokthornsakul, P. *et al.* (2019) 'An updated cost-effectiveness analysis of pneumococcal conjugate vaccine among children in Thailand', *Vaccine*. Elsevier Ltd, 37(32), pp. 4551–4560. doi: 10.1016/j.vaccine.2019.06.015.
- Dube, F. S. *et al.* (2013) 'Detection of Streptococcus pneumoniae from Different Types of Nasopharyngeal Swabs in Children', 8(6), pp. 4–9. doi: 10.1371/journal.pone.0068097.
- et al., D. (no date) 'Pneumococcal Conjugate Vaccine for Children Below Five Years Old Executive Summary'.
- Gloria, et al (2014) 'Table 1 : List of oligonucleotide primers used in 41 conventional multiplex \* PCR assays for pneumococcal serotype deduction of 70 serotypes'.
- Harimurti, K. *et al.* (2016) 'Nasopharyngeal carriage of Streptococcus pneumoniae in adults infected with human immunodeficiency virus in Jakarta , Indonesia', *Journal of Infection and Public Health.* King Saud Bin Abdulaziz University for Health Sciences, 9(5), pp. 633–638. doi: 10.1016/j.jiph.2016.01.004.
- Henriques-normark, B. and Tuomanen, E. I. (2013) 'The Pneumococcus : Epidemiology , Microbiology , and Pathogenesis', pp. 1–16.
- Henriques-normark, B. and Tuomanen, E. I. (2018) 'The Pneumococcus : Epidemiology , Microbiology , and Pathogenesis', pp. 1–16. doi: 10.1101/cshperspect.a010215.

Hocknell, R. E. et al. (2019) 'Serotype distribution of disease-causing Streptococcus

pneumoniae in Thailand : A systematic review', *Vaccine*. Elsevier Ltd, 37(24), pp. 3159–3166. doi: 10.1016/j.vaccine.2019.04.085.

- Hung, I. F. et al. (2013) 'International Journal of Infectious Diseases Regional epidemiology of invasive pneumococcal disease in Asian adults : epidemiology , disease burden , serotype distribution , and antimicrobial resistance patterns and prevention', *International Journal of Infectious Diseases*. International Society for Infectious Diseases, 17(6), pp. e364–e373. doi: 10.1016/j.ijid.2013.01.004.
- Hyun, S. *et al.* (2019) 'Changes in serotype distribution and antimicrobial resistance of Streptococcus pneumoniae isolates from adult patients in Asia : Emergence of drug-resistant non-vaccine serotypes', *Vaccine*. Elsevier Ltd, (xxxx), pp. 1–9. doi: 10.1016/j.vaccine.2019.09.065.
- Ismail, N. (2017) 'Streptococcus pneumoniae acquisition among vaccinated Malaysian hajj pilgrims and its associated factors', *Journal of Microbiology and Infectious Diseases*, 7(July 2016), pp. 56–56. doi: 10.5799/jmid.vi.328784.
- Isturiz, R. *et al.* (2018) 'Response to Mungall et al. letter to the editor on Streptococcus pneumoniae serotype 19A: worldwide epidemiology. Expert review of vaccines 2017;16(10):1007–27', *Expert Review of Vaccines*. Taylor & Francis, 17(8), pp. 669–671. doi: 10.1080/14760584.2018.1506207.
- Izurieta, P. *et al.* (2018) 'Public health impact of pneumococcal conjugate vaccine infant immunization programs: assessment of invasive pneumococcal disease burden and serotype distribution', *Expert Review of Vaccines*. Taylor & Francis, 17(6), pp. 479–493. doi: 10.1080/14760584.2018.1413354.
- Jindal, H. M. *et al.* (2018) 'Comparative genomic analysis of ten clinical Streptococcus pneumoniae collected from a Malaysian hospital reveal 31 new unique drugresistant SNPs using whole genome sequencing', *Journal of Biomedical Science*. Journal of Biomedical Science, 25(1), pp. 1–14. doi: 10.1186/s12929-018-0414-8.
- Kadioglu, A. *et al.* (2008) 'The role of Streptococcus pneumoniae virulence factors in host respiratory colonization and disease', (May). doi: 10.1038/nrmicro1871.
- Keller, L. E., Robinson, D. A. and Mcdaniel, L. S. (2016) 'Nonencapsulated Streptococcus pneumoniae: Emergence and', 7(2), pp. 1–12. doi: 10.1128/mBio.01792-15.Invited.
- Kilian, M. et al. (2008) 'Evolution of Streptococcus pneumoniae and Its Close Commensal Relatives', 3(7). doi: 10.1371/journal.pone.0002683.
- Kim, K. H. et al. (2011) 'Nasopharyngeal pneumococcal carriage of children attending day care centers in Korea: Comparison between children immunized with 7valent pneumococcal conjugate vaccine and non-immunized', Journal of Korean Medical Science, 26(2), pp. 184–190. doi: 10.3346/jkms.2011.26.2.184.

- Kim, S. H. *et al.* (2012) 'Changing Trends in Antimicrobial Resistance and Serotypes of Streptococcus pneumoniae Isolates in Asian Countries : an Asian Network for Surveillance of Resistant Pathogens (ANSORP) Study', pp. 1418–1426. doi: 10.1128/AAC.05658-11.
- Klugman, K. P. and Feldman, C. (2011) 'Pneumococcal Infections Guerrant, Richard L.', pp. 199–202. doi: https://doi.org/10.1016/B978-0-7020-3935-5.00029-X.
- Langereis, J. D. and de Jonge, M. I. (2017) 'Non-encapsulated Streptococcus pneumoniae, vaccination as a measure to interfere with horizontal gene transfer', *Virulence*. Taylor & Francis, 8(6), pp. 1–3. doi: 10.1080/21505594.2017.1309492.
- Le, C. *et al.* (2011) 'Capsular Serotype and Antibiotic Resistance of Streptococcus pneumoniae Isolates in Malaysia', 6(5), pp. 1–8. doi: 10.1371/journal.pone.0019547.
- Le, C. et al. (2012) 'The epidemiology of pneumococcal carriage and infections in Malaysia', pp. 707–719.
- Lee, N. Y. *et al.* (2001) 'Carriage of Antibiotic-Resistant Pneumococci among Asian Children : A Multinational Surveillance by the Asian Network for Surveillance of Resistant Pathogens (ANSORP)', pp. 1463–1469.
- M. T., N. et al. (2012) 'Community acquired pneumonia in Malaysia: Is Streptococcus pneumoniae an important pathogen?', African Journal of Microbiology Research, 6(3), pp. 512–519. doi: 10.5897/AJMR11.804.
- Madhi, S. A. and Nunes, M. C. (2016) 'The potential impact of pneumococcal conjugate vaccine in Africa: Considerations and early lessons learned from the South African experience', *Human Vaccines and Immunotherapeutics*, 12(2), pp. 314–325. doi: 10.1080/21645515.2015.1084450.
- Malik, S. *et al.* (1998) 'Susceptibility Pattern of Streptococcus pneumoniae Among Preschool Children in Kota Bharu, Malaysia', 44(no.5, p. 107–108.).
- McNeil, H. C. and Clarke, S. C. (2016) 'Serotype prevalence of Streptococcus pneumoniae in Malaysia – The need for carriage studies', *Medical Journal of Malaysia*, 71(3), pp. 134–138.
- Mohd, M. et al. (2013) 'Determination of phenotypes and pneumococcal surface protein A family types of Streptococcus pneumoniae from Malaysian healthy children', *Journal of Microbiology, Immunology and Infection*. Elsevier Taiwan LLC, 46(3), pp. 180–186. doi: 10.1016/j.jmii.2012.04.004.
- Na, X. I. E. *et al.* (2018) 'Letter to the Editor A Cross-sectional Survey Assessing Carriage of Streptococcus pneumoniae in a Healthy Population in Xinjiang Uygur Autonomous Region of China \*', 31(3), pp. 233–237. doi: 10.3967/bes2018.029.

- Nathan, J. J. *et al.* (2014) 'Prevalence of macrolide resistance and in vitro activities of six antimicrobial agents against clinical isolates of Streptococcus pneumoniae from a multi-center surveillance in Malaysia', (June).
- Neves, F. P. G. *et al.* (2013) 'Nasopharyngeal carriage, serotype distribution and antimicrobial resistance of Streptococcus pneumoniae among children from Brazil before the introduction of the 10-valent conjugate vaccine', *BMC Infectious Diseases*, 13(1), pp. 1–7. doi: 10.1186/1471-2334-13-318.
- Noorul, N. and Mohamed, S. (2014) 'Evaluation of PCR-based approach for serotype determination of Streptococcus pneumoniae Evaluation of PCR-based approach for serotype determination of Streptococcus pneumoniae', (pp. 5-7,).
- Nuermberger, E. L. and Bishai, W. R. (2004) 'Antibiotic Resistance in Streptococcus pneumoniae : What Does the Future Hold ?', 38(Suppl 4), pp. 363–371.
- Pai, R., Gertz, R. E. and Beall, B. (2006) 'Sequential Multiplex PCR Approach for Determining Capsular Serotypes of Streptococcus pneumoniae Isolates', 44(1), pp. 124–131. doi: 10.1128/JCM.44.1.124.
- Pelton, S. I. (2018) 'crossm Deconstructing Pneumococcal Progression from Colonization to Disease', pp. 17–19.
- Pichichero, M. E. *et al.* (2012) 'Antibody response to Streptococcus pneumoniae proteins PhtD, LytB, PcpA, PhtE and Ply after nasopharyngeal colonization and acute otitis media in children', *Human Vaccines and Immunotherapeutics*, 8(6), pp. 799–805. doi: 10.4161/hv.19820.
- Prasanna Subramaniam et al. (2018) 'Serotypes & penicillin susceptibility of Streptococcus pneumoniae isolated from children admitted to a tertiary teaching hospital in Malaysia', *Journal of Medical Microbiology*, 76(11), pp. 1532– 1539. doi: 10.4103/ijmr.IJMR.
- Rohani, M. Y. *et al.* (1999) 'Epidemiology of Streptococcus pneumoniae infection in Malaysia', *Epidemiology and Infection*, 122(1), pp. 77–82. doi: 10.1017/S0950268898001605.
- Scholz, C. F. P., Poulsen, K. and Kilian, M. (2012) 'Novel Molecular Method for Identification of Streptococcus pneumoniae Applicable to Clinical Microbiology and 16S rRNA Sequence-Based Microbiome Studies', 50(6), pp. 1968–1973. doi: 10.1128/JCM.00365-12.
- Schroeder, M. R. and Stephens, D. S. (2016) 'Macrolide Resistance in Streptococcus pneumoniae', 6(September), pp. 1–9. doi: 10.3389/fcimb.2016.00098.
- Shi, W. et al. (2018) 'Antimicrobial susceptibility and fluctuations in clonal complexes of serogroup 6 Streptococcus pneumoniae isolates collected from children in Beijing, China, between 1997 and 2016', *Brazilian Journal of Microbiology*. Sociedade Brasileira de Microbiologia, 49(4), pp. 891–899. doi: 10.1016/j.bjm.2018.02.004.

- Shiri, T. et al. (2019) 'Pneumococcal Disease: A Systematic Review of Health Utilities, Resource Use, Costs, and Economic Evaluations of Interventions', Value in Health. Elsevier Inc, 22(11), pp. 1329–1344. doi: 10.1016/j.jval.2019.06.011.
- Sigurdsson, S. *et al.* (2017) 'Pneumococcal vaccination : Direct and herd effect on carriage of vaccine types and antibiotic resistance in Icelandic children', 35, pp. 5242–5248. doi: 10.1016/j.vaccine.2017.08.020.
- Sirekbasan, L. et al. (2015) 'Phenotypes and Genotypes of Macrolide-Resistant Streptococcus Pneumoniae', pp. 84–88. doi: 10.5152/balkanmedj.2015.15169.
- Sleeman, K. L. et al. (2006) 'Capsular Serotype–Specific Attack Rates and Duration of Carriage of Streptococcus pneumoniae in a Population of Children', *The Journal of Infectious Diseases*, 194(5), pp. 682–688. doi: 10.1086/505710.
- Sohail, I. et al. (2018) 'Role of inflammatory risk factors in the pathogenesis of streptococcus pneumoniae', *Frontiers in Immunology*, 9(OCT), pp. 1–8. doi: 10.3389/fimmu.2018.02275.
- Song, J. *et al.* (2004) 'Macrolide resistance and genotypic characterization of Streptococcus pneumoniae in Asian countries : a study of the Asian Network for Surveillance of Resistant Pathogens (ANSORP)', 53(3), pp. 457–463. doi: 10.1093/jac/dkh118.
- Song, J. Y., Nahm, M. H. and Moseley, M. A. (2013) 'Clinical Implications of Pneumococcal Serotypes : Invasive Disease Potential, Clinical Presentations, and Antibiotic Resistance', pp. 4–15.
- Song, J. Y., Nahm, M. H. and Moseley, M. A. (2013) 'Clinical implications of pneumococcal serotypes: Invasive disease potential, clinical presentations, and antibiotic resistance', *Journal of Korean Medical Science*, 28(1), pp. 4–15. doi: 10.3346/jkms.2013.28.1.4.
- Song, K. S. K. and J.-H. (2004) 'Evolution of Erythromycin-Resistant Streptococcus pneumoniae from Asian Countries That Contains erm (B) and mef (A) Genes', 190.
- Souli, M. et al. (2006) 'Characterisation of macrolide-non-susceptible Streptococcus pneumoniae colonising children attending day-care centres in Athens, Greece during 2000 and 2003', *Clinical Microbiology and Infection*. European Society of Clinical Infectious Diseases, 13(1), pp. 70–77. doi: 10.1111/j.1469-0691.2006.01555.x.

Subhash Chandra Parija (2012) Microbiology and Immunology.

- Watson, D. A. et al. (1993) 'A Brief History of the Pneumococcus in Biomedical Research: A Panoply of Scientific Discovery', Clinical Infectious Diseases.
- Weil-Olivier, C. et al. (2012) 'Prevention of pneumococcal diseases in the post-seven valent vaccine era: A European perspective', BMC Infectious Diseases. BMC

Infectious Diseases, 12(1), p. 1. doi: 10.1186/1471-2334-12-207.

- Wessels, E. *et al.* (2012) 'Evaluation of Several Biochemical and Molecular Techniques for Identification of Streptococcus pneumoniae and Streptococcus pseudopneumoniae and Their Detection in Respiratory Samples', pp. 1171– 1177. doi: 10.1128/JCM.06609-11.
- Wouters, I. *et al.* (2019) 'Follow-up of serotype distribution and antimicrobial susceptibility of Streptococcus pneumoniae in child carriage after a PCV13-to-PCV10 vaccine switch in Belgium', *Vaccine*. The Authors, 37(8), pp. 1080– 1086. doi: 10.1016/j.vaccine.2018.12.068.
- Zivich, P. N. *et al.* (2018) 'Streptococcus pneumoniae outbreaks and implications for transmission and control: a systematic review', *Pneumonia*. Pneumonia, 10(1), p. 11. doi: 10.1186/s41479-018-0055-4.

