

UNIVERSITI PUTRA MALAYSIA

CONSTRUCTION OF AN ATTENUATED PASTEURELLA MULTOCIDA B:2 BY MUTATION IN THE GDHA GENE

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FPV 2007 11



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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Partial Fulfillment of the Requirement for the Degree of Master of Science

September 2007



"Things should be made as simple as possible, but not any simpler."
-Albert Einstein-

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

CONSTRUCTION OF AN ATTENUATED PASTEURELLA MULTOCIDA
B:2 BY MUTATION IN THE GDHA GENE

By

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September 2007

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Pasteurella multocida B:2 is a Gram negative bacteria that has been associated with haemorrhagic septicaemia in cattle and buffaloes in Asia. It

has been known to produce endotoxin that leads to haemorrhages and

oedema, causing deaths due to either asphyxiation and dyspnoea or

septicaemia. Vaccination has been used to control the disease but with little

success due to the low vaccination coverage. Therefore, an alternative live

vaccine should be considered.

In preparing an alternative live vaccine, an attenuated P. multocida B:2 is

created by manipulating one of the housekeeping genes of the bacteria. The

selected housekeeping gene, the glutamate dehyrogenase (gdhA) gene, was

successfully isolated via PCR from wild type *P. multocida* B:2. The gene was

then amplified using nested-PCR to determine its functional part. Both PCR

products were cloned into plasmid pCR2.1, producing pSZ1 and pSZ2,

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respectively before being sequenced. The whole sequence of the gene is 1108 bp while the functional part of the gene was 652 bp. The functional part was 99.8% identical to the model sequence, the PM70, which is a model genome sequence of *P. multocida* serotype A.

The pSZ1 was subsequently digested with a unique restriction enzyme, *Munl* before the kanamycin cassette, isolated from plasmid pUC4K via PCR, was inserted at the centre of the housekeeping gene. The recombinant was named pSZ1K. After that, the *gdhA* gene that was disrupted by kanamycin cassette (GK) was isolated from the pSZ1K using restriction enzyme digestion, *EcoRl*. The suicide plasmid, pAKA19 was also digested with the same enzyme to achieve complimentary ligation sites. After ligation, the achieved recombinant plasmid was called pSZ19GK. All cloning products were transformed into *Escherichia coli* DH5a. En route for disruption of the gene in the host genome, both *E. coli* and *P. multocida* B:2 were subjected to spontaneous mutation towards streptomycin. After conveying the pSZ19GK into *P. multocida* B:2 via conjugation, the bacteria was incubated for five days to encourage allelic exchange to occur between disrupted gene and the host chromosome. Subsequently, PCR of the bacteria genome proved that allelic exchange has occurred and the mutant was called *P. multocida* B:2 (GK).

In order to verify the characteristic of the non-pathogenic *P. multocida* B:2 (GK) mutant, *in vitro* stability test and *in vivo* pathogenicity test were done. In *in vitro* stability test, 14 strains out of the 20 survived only up to 15 days of incubation. This proves that the mutants are unable to sustain life without



glutamate supplement and therefore having a short life-span. From there, several strains were picked to be tested *in vivo* using mouse experimental model. Mice infected intraperitoneally or subcutaneously with different concentrations of the mutant survived throughout the 5-day study period. They were compared to the mice that were infected intraperitoneally or subcutaneously with different concentrations of the wild type organism. None of the mice infected with the mutant died but all mice infected with the wild type did not survived and were dead in less than 24 hours. *P. multocida* B:2 were successfully isolated from organs of mice infected with both wild-type and mutant. This confirmed that the mutant, *P. multocida* B:2 (GK) became attenuated by the disruption of the *gdhA* gene and has a good potential to be used as an alternative live vaccine for HS.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PEMBENTUKAN BAKTERIA ATENUAT PASTEURELLA MULTOCIDA B:2
MELALUI PELAKUAN MUTASI DALAM GEN GDHA

Oleh

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Pasteurella multocida B:2 adalah bakteria Gram negatif yang dikaitkan dengan penyakit hawar berdarah di dalam lembu dan kerbau di Asia. Bakteria ini menghasilkan endotoksin yang menyebabkan hemoraj dan edema, kematian diakibatkan sama ada pengasfikasiaan dan dispnea atau septisemia. Vaksinasi telah digunakan untuk mengawal penyakit ini tetapi cara ini kurang efektif disebabkan kawasan liputan vaksinasi adalah rendah. Oleh itu, vaksin hidup alternatif perlu dihasilkan untuk menangani masalah ini.

Dalam percubaan menghasilkan vaksin hidup alternatif, *P. multocida* B:2 yang atenuat, salah satu daripada gen domestik dalam bakteria ini dimanipulasikan. Gen domestik ini, gen glutamate dehidrogenase, telah berjaya diasingkan menggunakan PCR daripada *P. multocida* B:2 jenis liar. Gen itu diamplifikasi sekali lagi menggunakan 'nested-PCR' untuk memperolehi bahagian fungsinya. Kedua-dua produk PCR ini telah diklonkan

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ke dalam plasmid pCR2.1 menghasilkan pSZ1 dan pSZ2 dan kemudian ditentukan jujukannya. Keseluruhan jujukan gen tersebut mempunyai 1108 bp dan bahagian fungsinya mempunyai 652 bp. Bahagian fungsinya telah menghasilkan 99.8% komplimentari terhadap jujukan model, PM70 iaitu model keseluruhan jujukan genom bagi *P. multocida* serotip A.

Dalam pengklonan fasa kedua, pSZ1 telah dibatasi oleh enzim pembatasan yang unik, Munl dan kaset kanamicin yang telah diasingkan daripada plasmid pUC4K melalui PCR dan telah dimasukkan di tengah-tengah gen domestik tersebut (GK). Rekombinan DNA itu dinamakan pSZ1K. Gen gdhA yang telah mutan diasingkan dengan menggunakan enzim pembatasan, EcoRI. Plasmid 'suicide', pAKA19 juga telah dibatasi dengan emzim pembatasan yang sama untuk menghasilkan hujung lekatan yang komplimentari dengan kaset GK. Selepas proses pelekatan, rekombinan pSZ19GK. itu dinamakan Kesemua produk pengklonan plasmid ditransformasi ke dalam Escherichia coli DH5a. Seterusnya untuk menukar gen yang telah dikonstruk pada genom hos, kedua-dua E. coli dan P. multocida B:2 telah melalui mutasi spontan terhadap antibiotik streptomicin. Setelah menghantar pSZ19GK ke dalam P. multocida B:2 melalui proses kojugasi, bakteria itu telah diinkubasi selama lima hari untuk menggalakkan proses pertukaran alel berlaku terhadap kaset GK dengan kromosom perumah. Selepas itu, PCR terhadap genom bakteria membuktikan bahawa pertukaran alel telah berlaku dan mutan yang terhasil dipanggil P. multocida B:2 (GK).



Seterusnya untuk membuktikan mutan P. multocida B:2 (GK) adalah tidak patogenik, ujian kestabilan secara in vitro dan ujian patogenik secara in vivo telah dijalankan. Dalam ujian kestabilan secara in vitro, 14 daripada 20 isolat telah berjaya dihidupkan selama 15 hari secara berterusan. Ini membuktikan adanya mekanisma kawalan dalam penghasilan bakteria yang tidak patogenik dengan jangka hayatnya yang pendek disebabkan oleh kekurangan glutamat. Dari situ, beberapa isolat telah dipilih untuk diuji dalam ujian patogenik secara in vivo menggunakan model eksperimen tikus. Mencit yang telah dijangkiti melalui ruang peritoneum dan dibawah lapisan kulit dengan P. multocida B:2 (GK) hidup yang dihasilkan dengan kepekatan yang berbeza, hidup sepanjang lima hari ujian. Berbanding dengan mencit yang dijangkiti oleh P. multocida B:2 pula, mati dalam masa kurang dari 24 jam. P. multocida B:2 berjaya diisolat dari organ mencit yang dijangkiti oleh keduadua bakteria. Ini membuktikan bakteria mutan itu, P. multocida B:2 (GK) telah atenuat dengan mengubah jujukan pada gen gdhA sekaligus berguna untuk penghasilan vaksin hidup alternatif bagi mengawal penyakit hawar berdarah ini.



ACKNOWLEDGEMENTS

First and foremost praises to ALLAH, THE MOST COMPASSIONATE AND MERCIFUL, for granting me strength and courage to complete this thesis.

I would like to express my wholehearted indebtedness especially to my major advisor, Professor Dr. Mohd Zamri Saad for his inestimable and propitious guidance throughout the course of research and in transcription of this thesis. Similarly, my outmost appreciation is extended to my cosupervisors, Dr Zunita Zakaria for her patience, advices and unfailing encouragement that have been indispensable at every stage in designing and executing this thesis and as well as Prof Dr Raha Abdul Rahim, for her guidance, constructive criticism and also for never fail to have faith in me throughout this journey.

I would also like to thank the following people who contributed their efforts in making this thesis a success:

 Prof Madya Dr Abdul Rahman Omar, Dr Md Sabri Mohd Yusoff, Dr Siti Khairani Bejo, Dr Arul Jothi, Mr Mohd Jamil Samad, Mr Kamaruddin and Mr Ariff Ahmad, for their assistance and invaluable time spent.



- Mr Mas Jaffri Masaruddin, Mr Sim Han Koh, Mrs Trang, Mrs Latifah, Ms Ina Salwany and all Histopathology lab member for their help, understanding, encouragement and support.
- Prof Dr Abdul Rani Bahaman, Prof Dr Saleha Abdul Aziz, Dr Zeenathul and Dr Siti Suri for allowing me to use their laboratory facilities.
- To all HS Team, Virology Lab member, Bacteriology Lab member, Molecular Biology Lab Member and Biologic Lab member in FPV and all those who contributed directly or indirectly in sharing their knowledge, skill and assistance throughout the course of my study.

Finally yet notably, I owe all my success to my parents, Tuan Hj Othman Sa'don and Puan Hjh Zahrah Rameli, for their blessing and love, for giving strength and solace and for their unique and special way throughout my upbringing which has led me to where I am now. My heartfelt thanks to my brothers and sister, who make each of my day special and different in its own way. My lovable niece, always my rainbow after the thunderstorm downpour. My dearly loved, Mohd Shahril Myeor, for his companion, patience, guidance, understanding and especially his courage that never fails to amuse me.

Thank you all.

TABLE OF CONTENTS

			Page
ABS ACP APF DEC LIS ¹ LIS ¹	PROVA CLARA T OF T T OF F T OF A APTER	LEDGEMENTS AL ATION ABLES IGURES ABREVIATIONS	ii iii vi x xii xiv xviii xix xxii
2	LITE	RATURE REVIEW	
	2.1 2.2 2.3 2.4 2.5	2.1.1 The Disease 2.1.2 Global Distribution 2.1.3 Epidemiological Cycle 2.1.4 Progression of the disease 2.1.5 Treatment Pasteurella multocida 2.2.1 The Organism 2.2.2 Morphology 2.2.3 Serotyping 2.2.4 Virulent Factors 2.2.5 Metabolic Pathway 2.2.6 Source of Infection 2.2.7 Routes of Infection Vaccination for Prevention and Control 2.3.1 Alum Precipitate Vaccine 2.3.2 Oil Adjuvant Vaccine gdhA Gene Plasmid 2.5.1 Introduction 2.5.2 Suicide Plasmid	2.1 2.2 2.2 2.3 2.4 2.5 2.5 2.6 2.7 2.9 2.10 2.11 2.11 2.12 2.12 2.15 2.17 2.17
	2.6 2.7	Conjugation Allelic Exchange	2.21 2.22
3		ECULAR CLONING AND SEQUENCE ANALYSIS HE gdhA GENE OF PASTEURELLA MULTOCIDA	
	3.1 3.2	Introduction Materials And Methods 3.2.1 Bacterial Strains and Plasmids 3.2.2 Preparation of Stock Cultures 3.2.3 Extraction of Genomic DNA	3.1 3.3 3.3 3.3 3.3



		3.2.4 Quantification of DNA Concentration	3.4
		3.2.5 Amplification of the Gene by Polymerase	3.5
		Chain Reaction and Nested Polymerase	
		Chain Reaction	
		3.2.6 Detection of PCR Products	3.6
		3.2.7 Purification of the PCR Products	3.7
		3.2.8 Cloning Vector	3.8
		3.2.9 TOPO Cloning Ligation Reaction	3.9
		3.2.10 TOP 10 One Shot Chemical Transformation	3.9
		3.2.11 Plasmid Extraction	3.10
		3.2.12Analysis of Positive Clones by Restriction	3.11
		Endonuclease	
		3.2.13 Determination of Insert Orientation in	3.13
		Plasmid pSZ1	
		3.2.14Sequencing the Recombinants	3.14
		3.2.15DNA Sequencing Analysis	3.14
	3.3	Results	3.15
		3.3.1 Molecular Cloning and Sequence Analysis	3.15
		of gdhA Gene of Pasteurella multocida B:2	
		3.3.2 Sequence Analysis of The gdhA Gene	3.20
	3.4	Discussion	3.24
4		JPTION OF THE gdhA GENE TO CREATE AN	
		ILENT MUTANT OF PASTEURELLA MULTOCIDA	
	B:2	1.4	4.4
	4.1	Introduction	4.1
	4.2	Materials and Methods	4.3
		4.2.1 Construction of Disrupted gdhA Gene With	4.3
		Kanamycin Cassette In Suicide Plasmid.	
		· · · · · · · · · · · · · · · · · · ·	4.04
		4.2.2 Development of A Non-Pathogenic	4.24
	4.0	4.2.2 Development of A Non-Pathogenic <i>P. multocida</i> B:2 Via Allelic Exchange.	
	4.3	4.2.2 Development of A Non-PathogenicP. multocida B:2 Via Allelic Exchange.Results	4.30
	4.3	 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With 	
	4.3	 4.2.2 Development of A Non-Pathogenic	4.30 4.30
	4.3	 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With Kanamycin Cassette In Suicide Plasmid 4.3.2 Development Of A Non-Pathogenic 	4.30
		 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With Kanamycin Cassette In Suicide Plasmid 4.3.2 Development Of A Non-Pathogenic P. Multocida B:2 Via Allelic Exchange 	4.30 4.30 4.38
	4.3	 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With Kanamycin Cassette In Suicide Plasmid 4.3.2 Development Of A Non-Pathogenic 	4.30 4.30
5	4.4	 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With Kanamycin Cassette In Suicide Plasmid 4.3.2 Development Of A Non-Pathogenic P. Multocida B:2 Via Allelic Exchange Discussion 	4.30 4.30 4.38
5	4.4 IN VIVO	 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With Kanamycin Cassette In Suicide Plasmid 4.3.2 Development Of A Non-Pathogenic P. Multocida B:2 Via Allelic Exchange Discussion PATHOGENICITY AND IN VITRO STABILITY	4.30 4.30 4.38
5	4.4 IN VIVO TEST O	 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With Kanamycin Cassette In Suicide Plasmid 4.3.2 Development Of A Non-Pathogenic P. Multocida B:2 Via Allelic Exchange Discussion 	4.30 4.30 4.38
5	4.4 IN VIVO TEST O B:2	 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With Kanamycin Cassette In Suicide Plasmid 4.3.2 Development Of A Non-Pathogenic P. Multocida B:2 Via Allelic Exchange Discussion PATHOGENICITY AND IN VITRO STABILITY F THE MUTATED PASTEURELLA MULTOCIDA 	4.30 4.30 4.38 4.42
5	4.4 IN VIVO TEST O B:2 5.1	 4.2.2 Development of A Non-Pathogenic	4.30 4.30 4.38 4.42
5	4.4 IN VIVO TEST O B:2	 4.2.2 Development of A Non-Pathogenic	4.30 4.30 4.38 4.42 5.1 5.2
5	4.4 IN VIVO TEST O B:2 5.1	 4.2.2 Development of A Non-Pathogenic	4.30 4.30 4.38 4.42
5	4.4 IN VIVO TEST O B:2 5.1	 4.2.2 Development of A Non-Pathogenic	4.30 4.30 4.38 4.42 5.1 5.2 5.2
5	4.4 IN VIVO TEST O B:2 5.1	 4.2.2 Development of A Non-Pathogenic	4.30 4.30 4.38 4.42 5.1 5.2
5	4.4 IN VIVO TEST O B:2 5.1 5.2	 4.2.2 Development of A Non-Pathogenic P. multocida B:2 Via Allelic Exchange. Results 4.3.1 Construction of Disrupted gdha Gene With Kanamycin Cassette In Suicide Plasmid 4.3.2 Development Of A Non-Pathogenic P. Multocida B:2 Via Allelic Exchange Discussion PATHOGENICITY AND IN VITRO STABILITY F THE MUTATED PASTEURELLA MULTOCIDA Introduction Materials and Methods 5.2.1 In Vitro Stability of Pasteurella multocida B:2 Mutant 5.2.2 In Vivo Pathogenicity of Pasteurella multocida B:2 Mutant 	4.30 4.30 4.38 4.42 5.1 5.2 5.2 5.2
5	4.4 IN VIVO TEST O B:2 5.1	 4.2.2 Development of A Non-Pathogenic	4.30 4.38 4.42 5.1 5.2 5.2



5.4	Discussion	5.9
6 GEN	NERAL DISCUSSION	6.1
REFER		R.1
APPEN	DICES	A.1
BIODA	TA OF THE AUTHOR	B.1
LIST OF	F PUBLICATIONS	B.3



LIST OF TABLES

Table	Page
4.1 Susceptibility characteristics of <i>P. multocida</i> B:2 and <i>E. coli</i> isolates against different antibiotics.	4.39
5.1 <i>In vitro</i> test results of the 20 mutant isolates of <i>Pasteurella multocida</i> B:2.	5.6
5.2 The virulence of the wild-type and mutant <i>Pasteurella multocida</i> B:2 in mice following intraperitoneal injection with 10 ⁶ cfu/ml.	5.7
5.3 The virulence of wild-type and mutant <i>Pasteurella multocida</i> B:2 at either 10 ⁶ or 10 ⁸ cfu/ml following intraperitoneal and	5.7



LIST OF FIGURES

Figure		Page
3.1	Schematic diagram of the cloning strategy. Directional cloning strategy using two different restriction enzymes for both vectors and PCR product in order to generate complimentary sticky ends. Restriction enzymes were then employed to isolate the desired PCR fragments for further exploitation.	3.12
3.2	Restriction enzyme <i>Narl</i> sites at plasmid pSZ1 with definite fragment size for positive orientation cloned.	3.13
3.3	Agarose gel electrophoresis (AGE) analysis of PCR amplification of the whole sequence of the housekeeping enzyme gene <i>gdhA</i> gene. Lane 1, PCR product of the whole sequence of <i>gdhA</i> gene from PmTB strain, lane 2, PCR product of the whole sequence of <i>gdhA</i> gene from Pm3030 strain, lane 3, GeneRuler TM 100bp DNA ladder (Fermentas, Canada).	3.16
3.4	Agarose gel electrophoresis analysis of PCR amplification of the functional part of the housekeeping enzyme gene <i>gdhA</i> gene with nested-PCR. Lane 1, GeneRuler TM 100bp DNA ladder (Fermentas, Canada), lane 2, Nested-PCR product of the functional part of <i>gdhA</i> gene.	3.17
3.5	Verification of pSZ1 construct by restriction enzymes digestion analysis. Lane 1 & 5, GeneRuler TM 1 kb DNA ladder (Fermentas, Canada), lane 2, plasmid pCR2.1, lane 3, top band is the whole sequence of <i>gdhA</i> gene and two bottom bands were the digested gene with <i>Mun</i> I, lane 4, plasmid pSZ1, lane 6-10, pSZ1 digested with <i>BamHI</i> and <i>XhoI</i> .	3.18
3.6	Determination of orientation of pSZ1 with restriction enzyme digestion, <i>Nar</i> l. Lane 1, GeneRuler TM 1 kb DNA ladder (Fermentas, Canada), lane 2-8, positive pSZ1 isolates from different colonies digested with <i>Nar</i> l.	3.19
3.7	Whole sequence of the <i>gdhA</i> gene, the sequence of the functional part of the gene and also the location of both sets of primer.	3.21
3.8	Sequence comparison of the functional part of the <i>gdhA</i> gene from <i>P. multocida</i> B:2 with <i>P. multocida</i> serotype A (PM70) using BLAST and amino acid sequence of <i>P. multocida</i> B:2.	3.23



4.0	Schematic diagram of the overall work-flow showing ligation and cloning process in First Route and Second Route.	4.4
4.1	Schematic diagram of the cloning strategy. Ligation of suicide plasmid, pAKA19 and <i>gdhA</i> gene using <i>BamHI</i> and <i>XhoI</i> enzymes. <i>MunI</i> , a unique restriction enzyme situated in the middle of the gene will soon be used for insertion and also detection of the disrupted gene.	4.9
4.2	Directional cloning strategy using <i>Mun</i> I for pSZ19 and kanamycin cassette.	4.14
4.3	Ligation of pSZ1 and kanamycin cassette in order to generate GK (<i>gdhA</i> ~ kanamycin cassette) using a unique restriction enzyme.	4.18
4.4	Directional cloning strategy using restriction enzymes for pAKA19 and GK (<i>gdhA</i> and kanamycin construct) in order to generate complimentary sticky ends.	4.21
4.5	Verification of pSZ19 construct by restriction enzymes analysis. Lane 1, GeneRuler TM 1 kb DNA ladder (Fermentas, Canada), lane 2 and 3, digested pSZ19 with BamHI and XhoI.	4.31
4.6	PCR product from the amplification of plasmid pUC4K with pUC4K (2) primers. Lane 1, GeneRuler TM 1 kb DNA ladder (Fermentas, Canada), lane 2, digested PCR product, kanamycin cassette with Mun I.	4.32
4.7	Verification of pCR2.1 with kanamycin cassette construct by restriction enzymes analysis. Lane 1, GeneRuler TM 1 kb DNA ladder (Fermentas, Canada), lane 2, digested pCR2.1 with kanamycin cassette with <i>Mun</i> I.	4.33
4.8	Authentication of pSZ1K with <i>gdhA</i> gene and kanamycin cassette construct (GK) by restriction enzymes analysis. Lane 1, digested plasmid pCR2.1 TOPO with <i>Bam</i> HI and <i>Xho</i> I, lane 2, digested pSZ1 with first <i>Mun</i> I followed by <i>Bam</i> HI and <i>Xho</i> I prior to the analysis, lane 3&4, digested pSZ1K with <i>Bam</i> HI and <i>Xho</i> I, lane 5, GeneRuler TM 1 kb DNA ladder (Fermentas, Canada).	4.35
4.9	Isolation of GK from pSZ1K with restriction enzyme. Lane 1, plasmid pCR2.1 TOPO, lane 2, digested pSZ1K with <i>EcoRI</i> , lane 3&4, GeneRuler TM 1 kb DNA ladder (Fermentas), lane 5, purified GK from digestion of pSZ1K with FooRI	4.36



- 4.10 Screening of pSZ19GK. Lane 1,2,3&5, positive selection of transformants confirmed with the existence of the ~8kb band, lane 4, negative transformant with only self-ligation of the plasmid band (upper band) and self-ligation of the insert band (lower band), lane 6&9, GeneRulerTM 1 kb DNA ladder (Fermentas, Canada) (From top to bottom, lane 6: 8.0 kb, 4.0 kb, 3.5 kb & 2.5 kb and lane 9: 10 kb, 6.0 kb, 3.5 kb, 3.0 kb & 2.0 kb), lane 7, ligation mixture of vector:vector, pAKA19, lane 8, plasmid pSZ19 and lane 10,11&12, PCR product of GK amplified with gdhA(1) set primers from pSZ19GK.
- 4.11 Plasmid purification of *P. multocida* B:2 (GK) from allelic 4.40 exchange incubation period. Lane 1, GeneRulerTM 1 kb DNA ladder (Fermentas, Canada) (0.25 to 10 kb), lane 2-5, selected *P. multocida* B:2 (GK) isolate taken from incubated plates.
- 4.12 Indication of allelic exchange in *P. multocida* B:2 (GK) via Direct Colony PCR. Lane 1, GK cassette (~2.4kb), lane 2, GeneRulerTM 1 kb DNA ladder (Fermentas, Canada) (0.25 to 10 kb) and lane 3, *gdhA* gene from parent strain, *P. multocida* B:2 (~1.108KB).



LIST OF ABBREVIATIONS

A₂₆₀ absorbance at 260 nm

A_{450 nm} absorbance at 450 nm

aa amino acid

AcmA *N*- acetylmuramidase

BLAST basic local alignment search tool

bp base pair

BSA bovine serum albumin

CaCl₂ calcium chloride

cDNA complementary DNA

cfu colony forming unit

CO₂ carbon dioxide

C-terminal the carboxyl-terminal (-COOH) of a

polypeptide

dH₂O distilled water

DNA deoxyribonucleic acid

dNTPs deoxyribonucleic acid

EDTA ethylenediamine tetraacetate

EtBr ethidium bromide

GMO genetically modified organisms

h hour

HCI hydrochloric acid

His histidine



i.p intraperitoneally

i.v intravenously

in an experimental situation outside the

organism

in vivo in a living cell or organism

IPTG isopropyl-D-galactopyranoside

kb kilo base pair

kDa kilo Daltons

LB Luria Bertani

M molarity

MCS multiple cloning site

MgCl₂ magnesium chloride

min minute

ml millilitre

mM milliMolar

NaCl sodium chloride

NAG *N*-acetylglucosamine

NAM *N*-acetylmuramic acid

ng nanogram

N-terminal the amino-terminal (NH₂) of a polypeptide

°C degrees centigrade

OD optical density

ORF open reading frame

PBS phosphate buffered saline

PCR polymerase chain reaction



PEG polyethylene glycol

pH isoelectric point

POD peroxidase

RE restriction enzyme

RNase ribonuclease

rpm revolution per minute

RT room temperature

s second

TBE tris-boric-EDTA

Taq Thermus aquaticus

TCA tri-carboxylic acid

TE tris EDTA

U unit

UV ultraviolet

v/v volume per volume

w/v weight per volume

% percent/ pencentage

 μ I microlitre

 μg microgram



CHAPTER 1

INTRODUCTION

Haemorrhagic septicaemia (HS) is a contagious bacterial disease caused by two serotypes of *Pasteurella multocida*; B2 and E2. It affects cattle (*Bos taurus* and *B. indicus*) and water buffaloes (*Bubalus bubalis*) with a high mortality rate in infected animals. It is irrefutably regarded as endemic in most parts of tropical Asia, Africa and India. Moreover, it is also considered to be the most economically important disease of livestock or large ruminants in South East Asia and causes significant economic losses in India and Africa. In Malaysia, the disease is enzootic in Kelantan, Terengganu, Kedah, Perak, Pahang and Negeri Sembilan due to the high population of cattle and buffaloes (Chadrasekeran 1981).

Outbreaks of haemorrhagic septicaemia have been observed during stressful conditions such as radical changes in weather, including the advent of monsoons, rainy seasons, debility caused by seasonal levels of low nutrition or changes in diet and the pressure of work (draft animals), such as increased activity during paddy cultivation. It is believed that the disease is spread by direct and indirect contact (fomites). The patterns of outbreaks revealed that carrier animals were subjected to stressful condition and start to shed the organisms and subsequently infect other susceptible animals.

