



**NEUROMODULATORY EFFECT OF ZERUMBONE ON ALPHA-_{2A}
ADRENOCEPTOR, NMDA N2B RECEPTOR AND TRPV1 CHANNEL IN LPS-
INDUCED SH-SY5Y CELLS, A NEUROPATHIC PAIN-LIKE *IN VITRO*
MODEL**

By

NOOR AISHAH BINTI MOHAMMED IZHAM

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

July 2021

FPSK (p) 2021 44

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

NEUROMODULATORY EFFECT OF ZERUMBONE ON ALPHA-_{2A} ADRENOCEPTOR, NMDA N2B RECEPTOR AND TRPV1 CHANNEL IN LPS-INDUCED SH-SY5Y CELLS, A NEUROPATHIC PAIN-LIKE *IN VITRO* MODEL

By

NOOR AISHAH BINTI MOHAMMED IZHAM

July 2021

Chair: Associate Professor Enoch Kumar a/l Perimal, PhD
Faculty: Medicine and Health Sciences

Neuropathic pain has been denoted as chronic pain that was prompted by lesion or disease of the somatosensory nervous system. The development and persistence of neuropathic pain involve complex mechanisms intertwined, such as the involvement of neuro-inflammation and neuronal hyperexcitability. The conventional therapies to alleviate neuropathic pain triggered adverse effects as a result of indistinct understanding of the mechanisms. Recent research has proposed that zerumbone, a crystalline sesquiterpene compound derived from *Zingiber zerumbet*, was able to attenuate neuropathic pain symptoms in *in vivo* models. Therefore, the present study was carried out to investigate the neuromodulatory properties of zerumbone via the modulation of alpha-_{2A} adrenoceptor, *N*-methyl-*D*-aspartate (NMDA) subtype N2B receptor and transient receptor potential vanilloid subtype 1 (TRPV1) channel in LPS-induced SH-SY5Y cells, a neuropathic pain-like *in vitro* model. LPS-induced SH-SY5Y cells were employed to allow tight control of physiological environment which could not be established in *in vivo* models, in addition to, reducing the use of animals in the study of neuropathic pain. LPS induction in SH-SY5Y cells enable the observation of the hallmark of neuropathic pain pathophysiology which are the expression of pro-inflammatory mediators and the alteration of receptors and ion channels associated with neuronal hyperexcitability. The optimisation of culture media for SH-SY5Y cells were conducted by observing the effect of growth rate of the cells in different medium and foetal bovine serum (FBS) concentration; Dulbecco's modified Eagle's Medium (DMEM) with 15% FBS, DMEM: Ham's F12 mix (DMEM:F12) with 10% FBS and DMEM:F12 supplemented with 15% FBS. SH-SY5Y cells cultured in DMEM:F12 supplemented with 15% FBS had shown the highest growth rate following 24 hours, hence, the culture medium was used throughout the experiments. SH-SY5Y cells were subjected to neuronal differentiation via 10µM of all *trans* retinoic acid induction and serum deprivation, followed by characterization through immunocytochemistry. Then, 3-(4,5-di methylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay was conducted and confirmed the viability of SH-SY5Y cell to be above 90% following the treatment of 2, 4, 8 and 16 µg/ml of zerumbone for 24 hours. The neuromodulatory effect of zerumbone

was first investigated through the expression level of nitric oxide through Griess' assay, whereby the inhibitory concentration of zerumbone was determined at 8µg/ml. Enzyme-linked immunoassay (ELISA) was then conducted to observe the expression of interleukin-6 (IL-6) and tumour necrosis factor- α (TNF- α), by which only slight downregulation was observed after the treatment of zerumbone. Next, to understand the anti-neuropathic pain effect of zerumbone, molecular docking and Western blot analysis were performed. The inhibitory properties of zerumbone on pain signal transmission was observed to be involved in the modulation of α_{2A} adrenoceptor, NMDA N2B receptor and TRPV1 channel. Molecular docking analysis revealed that the -CDOCKER binding and interaction energy of zerumbone to the respective proteins were higher as compared to the native crystallized ligands. Nevertheless, the amino acid interaction of zerumbone with the proteins indicated its possible anti-neuropathic pain effect by which similar interactions were perceived on NECA (α_{2A} adrenoceptor agonist), ifenprodil (NMDA N2B receptor antagonist) and capsazepine (TRPV1 channel antagonist) with the proteins. Meanwhile Western blot analysis showed that zerumbone increased the expression of α_{2A} adrenoceptor, proposing the inhibitory mechanism of zerumbone through descending modulation, antagonistic towards the down-regulation of the receptor after LPS induction. Treatment of zerumbone down-regulated the expression of NMDA N2B receptor and TRPV1 channel, as opposed to the upregulation of these proteins after LPS induction. Data from each experiments were analysed by using One-way Analysis of Variance (ANOVA) followed by *post hoc* Tukey test, $p < 0.05$. To conclude, the current study proved the neuromodulatory effect of zerumbone on α_{2A} adrenoceptor, NMDA N2B receptor and TRPV1 channel in LPS-induced SH-SY5Y cells, a neuropathic pain-like *in vitro* model.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN NEUROMODULASI OLEH ZERUMBONE TERHADAP
ADRENOSEPTOR ALFA-_{2A}, RESEPTOR NMDA N2B DAN SALURAN ION
TRPV1 DALAM SH-SY5Y SEL TERARUH LIPOPOLISAKARIDA, SEBUAH
MODEL *IN VITRO* BAK SAKIT NEUROPATIK**

Oleh

NOOR AISHAH BINTI MOHAMMED IZHAM

Julai 2021

**Pengerusi: Professor Madya Enoch Kumar a/l Perimal, PhD
Fakulti: Perubatan dan Sains Kesihatan**

Sakit neuropatik ditakrif sebagai sakit kronik yang disebabkan lesion atau penyakit sistem saraf somatosensori. Perkembangan dan perlanjutan sakit neuropatik melibatkan mekanisme kompleks yang saling berkaitan, seperti inflamasi saraf dan keterujaan saraf. Terapi konvensional untuk mengurangkan sakit neuropatik mengakibatkan kesan buruk yang berpunca daripada pemahaman yang tidak jelas berkenaan mekanisme keadaan tersebut. Penyelidikan terkini menunjukkan bahawa zerumbone, sebatian seskuiterpena bak hablur yang berasal daripada *Zingiber zerumbet*, dapat mengatenuasi simptom sakit neuropatik dalam model *in vivo*. Justeru, kajian ini dijalankan untuk menyiasat sifat neuromodulasi zerumbone melalui modulasi adreseptor alfa-_{2a}, reseptor, NMDA N2B dan saluran ion TRPV1 dalam sel SH-SY5Y teraruh lipopolisakarida (LPS), sebuah model *in vitro* bak sakit neuropatik. Sel SH-SY5Y teraruh lipopolisakarida digunakan untuk membolehkan pengawalan ketat terhadap persekitaran fisiologi yang tidak dapat dicapai dalam model *in vivo* serta untuk mengurangkan penggunaan haiwan dalam kajian sakit neuropatik. Aruhan LPS dalam sel SH-SY5Y membolehkan pemerhatian ciri khas patofisiologi sakit neuropatik iaitu ekspresi pengantara pro-inflamasi dan perubahan reseptor dan saluran ion yang berkaitan dengan keterujaan saraf. Pengoptimuman media kultur sel SH-SY5Y dilaksanakan dengan mengkaji kesan kadar pertumbuhan sel dalam setiap jenis kultur media dan serum anak lembu (FBS) dengan kepekatan yang berbeza; Dulbecco's modified Eagle's Medium (DMEM) ditambah dengan 15% FBS, campuran DMEM: Ham's F12 (DMEM:F12) ditambah dengan 10% FBS and DMEM:F12 ditambah dengan 15% FBS. Sel SH-SY5Y dikultur dalam DMEM: F12 ditambah dengan 15% FBS telah menunjukkan kadar pertumbuhan tertinggi berikutan 24 jam, oleh itu, media kultur tersebut digunakan sepanjang eksperimen. Sel SH-SY5Y mengalami pembezaan sel saraf setelah diaruh oleh 10 μ M asid all *trans* retinoik dan deprivasi serum, diikuti dengan pencirian melalui immunositokimia. Kemudian, asai 3-(4,5-di methylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) dilakukan untuk mengesahkan kebolehhidupan sel melebihi 90% selepas rawatan 2, 4, 8 dan 16 μ g/ml zerumbone selepas 24 jam. Kesan neuromodulasi zerumbone diselediki melalui asai Griess' dengan mengkaji tahap ekspresi

nitrik oksida, yang dengannya kepekatan rencatan zerumbone ditentukan pada 8 µg/ml. Seterusnya, asai immunojerapan berpaut enzim (ELISA) dilakukan untuk mengkaji ekspresi interleukin-6 (IL-6) dan faktor nekrosis tumor alfa (TNF- α), dengan pengurangan ekspresi yang kecil setelah rawatan zerumbone. Bagi memahami kesan sakit neuropatik oleh zerumbone, kajian dok molekul dan pedapan Western dilakukan. Berdasarkan pemerhatian terhadap dua analisis tersebut, zerumbone mempunyai sifat rencatan sakit neuropatik yang dimodulasikan melalui adrenoseptor alfa- $_{2a}$, reseptor NMDA N2B dan saluran ion TRPV1. Analisis dok molekul menunjukkan tenaga pengikatan dan tenaga saling tindak - CDCKER zerumbone terhadap protein tersebut, masing-masing lebih tinggi berbanding ligan kristal asli. Namun, interaksi asid amino antara zerumbone dengan protein tersebut menunjukkan persamaan dengan NECA (agonis adrenoseptor alfa- $_{2a}$), ifenprodil (antagonis reseptor NMDA N2B) dan capsazepine (antagonis saluran ion TRPV1), yang membuktikan kemungkinan kesan anti-sakit neuropatik oleh zerumbone. Sementara itu, analisis pedapan Western menunjukkan bahawa zerumbone meningkatkan ungkapan adrenoseptor alfa- $_{2a}$, menyanamkan mekanisme rencatan zermbone melalui modulasi laluan menurun, berantagonis dengan kesan aruhan LPS yang mengurangkan ungkapan reseptor tersebut. Rawatan zerumbone telah mengurangkan ungkapan reseptor NMDA N2B dan saluran ion TRPV1, berlawanan dengan peningkatan ungkapan protein tersebut setelah aruhan LPS. Data daripada setiap kajian dianalisis menggunakan One-Way Analysis of Variance (ANOVA) diikuti dengan ujian *post hoc*, $p < 0.05$. Kesimpulannya, kajian ini membuktikan kesan neuromodulasi oleh zerumbone terhadap adrenoseptor alfa- $_{2a}$, reseptor NMDA N2B dan saluran ion TRPV1 dalam SH-SY5Y sel teraruh lipopolisakarida, sebuah model *in vitro* bak sakit neuropatik.

ACKNOWLEDGEMENTS

All praises to Allah for His blessings and graciousness that have made it possible for me to grow through and complete my PhD journey. The perseverance that I had with me and the knowledge that I attained throughout the study are blessings from Him.

I am eternally grateful for my supervisor, Associate Professor Dr. Enoch Kumar Perimal for his guidance and support. I remembered him telling me, “When things gets hard, it is just another challenge that we need to go through. Whatever happens, even further in life later on, we do not quit. That is why it is important to focus on your strengths, and let the weaknesses improved along the way,” when I was going through one of the low points in my study. He believes in me and make me believe in myself to undertake and persevere throughout my PhD journey.

I would also like to take this opportunity to thank my supervisory committee, Prof. Dr. Mohd Roslan Sulaiman, Prof. Dr. Sharmili Vidyadaran and Dr. Hemabarathy Bharatham for their constructive criticisms and helpful insights that helped in accomplishing my postgraduate study. Thank you to Dr. Say Yee How from the Department of Biomedical Sciences, Universiti Tunku Abdul Rahman, for helping to provide SH-SY5Y cell culture and Dr. Lam Kok Wai from the Faculty of Pharmacy, Universiti Kebangsaan Malaysia for guiding me to perform molecular docking analysis and to conduct the study in the computer laboratory under his supervision.

I am abundantly grateful of my backbone, my father, Mohammed Izham Tajuddin, for his constant encouragement and sacrifices that made it possible for me to further my studies. I am also thankful of my best cheerleader, my mother, Aniza Ahmad for her loving comforts and heartening support. I would also like to thank my only brother, Muhammad Izzat for being thoughtful and understanding.

My sincere thanks for the assistance in providing the study necessities to the staffs of Physiology, Cell Signalling and Immunology Laboratory. To my lab mates, Khalisah, Kavitha and Wawa, thank you for the help, stimulating discussions and staying late in the lab to accompany each other to finish our work. I would like to express my appreciation to Adibah Hanis for being a good listener and sharing motivational playlists, Hanani Amirah' for her trustworthy support and making time for coffee that are always valuable to keep my motivation up to par, and Nurul Farhana for always picking up the phone to listen to my random thoughts.

Thank you to also to whomever I failed to mention here.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Enoch Kumar a/l Perimal, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Mohd Roslan bin Sulaiman, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Sharmili a/p Vidyadaran, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

B. Hemabarathy a/p Bharatham, PhD

Senior Lecturer
Faculty of Health Sciences
Universiti Kebangsaan Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 13 October 2022

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Noor Aishah Binti Mohammed Izham

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xviii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	5
2.1 Pain	5
2.1.1 Pain processing	5
2.1.2 Pain modulation	6
2.1.3 Acute pain	7
2.1.4 Chronic pain	9
2.2 Neuropathic pain	10
2.2.1 Causes and symptoms of neuropathic pain	11
2.2.2 Mechanism of neuropathic pain	11
2.2.2.1 Neuronal sensitization and molecular mechanism	11
2.2.2.2 Neuroinflammation	17
2.3 Current treatments of neuropathic pain	18
2.4 <i>In vitro</i> models of neuropathic pain	19
2.4.1 Type of cells	20
2.4.2 Induction of neuropathy	22
2.5 Zerumbone	23
3 GENERAL METHODOLOGY	25
3.1 Introduction	25
3.2 General methodology	25
3.2.1 SH-SY5Y cell culture	25
3.2.2 Preparation of zerumbone	26
3.2.3 LPS induction and treatment of zerumbone	26
3.2.4 Molecular docking analysis	26
3.2.5 Western blot analysis	27
4 INITIAL CULTURE OF SH-SY5Y CELLS	29
4.1 Introduction	29
4.2 Materials and methods	30
4.2.1 SH-SY5Y cell culture in different media	30

4.2.2	Observation of the morphology of SH-SY5Y cells	33
4.2.3	Determination of viable cells	33
4.2.4	Statistical analysis	33
4.3	Results	34
4.3.1	Effect of different culture media on the cell morphology	34
4.3.2	Effect of different culture media on the number of viable cells	35
4.4	Discussion	35
4.5	Conclusion	37
5	CHARACTERIZATION OF DIFFERENTIATED SH-SY5Y CELLS	38
5.1	Introduction	38
5.2	Materials and methods	38
5.2.1	Induction of SH-SY5Y cells differentiations	38
5.2.1	Immunocytochemistry	39
5.3	Results	40
5.3.1	Induction of SH-SY5Y cells differentiations	40
5.3.2	Characterization of differentiated SH-SY5Y cells	41
5.4	Discussion	43
5.5	Conclusion	46
6	EFFECT OF ZERUMBONE ON THE EXPRESSION OF NO, IL-6 AND TNF-α IN LPS-INDUCED SH-SY5Y CELLS	47
6.1	Introduction	47
6.2	Materials and methods	48
6.2.1	Cell viability assay	48
6.2.2	Statistical analysis	48
6.2.3	Inflammatory cytokine assay	49
6.2.3.1	Measurement of nitric oxide	49
6.2.3.2	Enzyme-linked immunosorbent assay (ELISA)	49
6.2.3.3	Statistical analysis	49
6.3	Results	50
6.3.1	Cell viability assay	50
6.3.2	Nitric oxide assay	51
6.3.3	Enzyme-linked immunosorbent assay (ELISA)	52
6.4	Discussion	53
6.5	Conclusion	59

7	ANTI-NEUROPATHIC PAIN EFFECT OF ZERUMBONE IN LPS-INDUCED SH-SY5Y CELLS VIA THE MODULATION OF ALPHA-2A ADRENOCEPTORS	60
	7.1 Introduction	60
	7.2 Materials and methods	61
	7.3 Results	61
	7.3.1 Validation of docking protocol	61
	7.3.2 CDOCKER binding energy and interaction energy	62
	7.3.3 Binding interaction of zerumbone with target protein	63
	7.3.4 Effect of zerumbone on the protein expression	66
	7.4 Discussion	67
	7.5 Conclusion	69
8	ANTI-NEUROPATHIC PAIN EFFECT OF ZERUMBONE IN LPS-INDUCED SH-SY5Y CELLS VIA THE MODULATION OF NMDA N2B RECEPTORS	70
	8.1 Introduction	70
	8.2 Materials and methods	71
	8.3 Results	71
	8.3.1 Validation of docking protocol	71
	8.3.2 CDOCKER binding energy and interaction energy	72
	8.3.3 Binding interaction of zerumbone with target protein	73
	8.3.4 Effect of zerumbone on the protein expression	76
	8.4 Discussion	77
	8.5 Conclusion	79
9	ANTI-NEUROPATHIC PAIN EFFECT OF ZERUMBONE IN LPS-INDUCED SH-SY5Y CELLS VIA THE MODULATION OF TRPV1 CHANNEL	80
	9.1 Introduction	80
	9.2 Materials and methods	80
	9.3 Results	81
	9.3.1 Validation of docking protocol	82
	9.3.2 CDOCKER binding energy and interaction energy	83
	9.3.3 Binding interaction of zerumbone with target protein	83
	9.3.4 Effect of zerumbone on the protein expression	86
	9.4 Discussion	87
	9.5 Conclusion	89

10	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	90
	REFERENCES	94
	APPENDICES	116
	BIODATA OF STUDENT	121
	LIST OF PUBLICATIONS	122



LIST OF TABLES

		Page
1.1	Summary of the description of neuropathic pain symptoms	11
4.1	Amino acid formulation comparison in DMEM and DMEM:F12 culture media produced by Nacalai Tesque Inc. used in this study	31
4.2	Inorganic salts formulation comparison in DMEM and DMEM:F12 culture media produced by Nacalai Tesque Inc. used in this study	32

LIST OF FIGURES

Figure		Page
2.1	Schematic diagram of the acute pain pathway	9
2.2	Schematic representation of peripheral and central sensitization in chronic pain	10
2.3	Depicted is the illustration of peripheral sensitization mechanism	12
2.4	Depicted is the illustration of central sensitization molecular mechanism	13
2.5	Illustration of the structure of α_{-2A} adrenoceptor	14
2.6	Schematic representation of TRP channels subunits structure organization	15
2.7	Schematic representation of NMDA receptor (subunit GluN1 or N1B and GluN2 or N2B) assembly and modular organization	16
2.8	Illustration of the interaction of neuroinflammation and molecular mechanism in pain signal transmission and transduction	18
2.9	The inflorescences and rhizomes of <i>Zingiber zerumbet</i> and the chemical structure of zerumbone	24
4.1	Morphology of undifferentiated SH-SY5Y cells in different culture media	34
4.2	Quantitative analysis of mean cell count of SH-SY5Y cells in different culture media	35
5.1	Comparison of undifferentiated and differentiated SH-SY5Y cells	40
5.2	Characterization of SH-SY5Y cells differentiation via immunofluorescent staining using Tuj1 antibody	41
5.3	Characterization of SH-SY5Y cells differentiation via immunofluorescent staining using MAP2 antibody	42

5.4	Illustration of the mechanism of retinoic acid signalling	43
5.5	Non-genomic signalling cascade following RAR target gene activation	44
6.1	Effect of zerumbone on the viability of differentiated SH-SY5Y cells	50
6.2	Nitric oxide assay of zerumbone against LPS-induced SH-SY5Y cells	51
6.3	Enzyme-linked immunosorbent assay for IL-6 of zerumbone against LPS-induced SH-SY5Y cells	52
6.4	Enzyme-linked immunosorbent assay for TNF- α of zerumbone against LPS-induced SH-SY5Y cells	53
7.1	The validation of Discovery Studio accuracy	62
7.2	Energy profile of control co-crystallized ligand and zerumbone	63
7.3	The residue of the binding site of α_{-2A} adrenergic receptor interacting with NECA conformation 1 in 2D (right) and 3D (left) perspective	64
7.4	The residue of the binding site of α_{-2A} adrenergic receptor interacting with zerumbone conformation 1 in (a) (b) 3D perspective and (c) 2D perspective	65
7.5	Anti-neuropathic effect of zerumbone on LPS-induced SH-SY5Y cells via the regulation of α_{-2A} adrenoceptor protein expression	66
8.1	The validation of Discovery Studio accuracy	72
8.2	Energy profile of control co-crystallized ligand and zerumbone	73
8.3	The residue of the binding site of NMDA N2B receptor interacting with ifenprodil conformation 1 in 2D (right) and 3D (left) perspective	74
8.4	The residue of the binding site of NMDA N2B receptor interacting with zerumbone conformation 1 in (a) (b) 3D perspective and (c) 2D perspective	75

8.5	Representative western blots of NMDA N2B receptor from LPS-induced SH-SY5Y cells samples of normal, vehicle, LPS only, zerumbone, amitriptyline-treated groups	76
9.1	The validation of Discovery Studio accuracy	82
9.2	Energy profile of control co-crystallized ligand and zerumbone	83
9.3	The residue of the binding site of TRPV1 channel interacting with capsaizpine conformation 1 in 2D (right) and 3D (left) perspective	84
9.4	The residue of the binding site of TRPV1 channel interacting with zerumbone conformation 1 in (a) (b) 3D perspective and (c) 2D perspective	85
9.5	Representative western blots of TRPV1 channel from LPS-induced SH-SY5Y cells samples of normal, vehicle, LPS only, zerumbone, amitriptyline-treated groups	86

LIST OF ABBREVIATIONS

3D	Three-dimensional
5-HT	Serotonin
AEA	Arachidonoyl ethanolamine
AMI	Amitriptyline
AMPA	α -amino-3-hydroxyl-5-methyl-4-isoxazole-propionate
ANOVA	One-way Analysis of Variance
ATP	Adenosine-5-triphosphate
Ca ²⁺	Calcium ion
CAMKIII	Calcium ion calmodulin-dependent protein kinase III
cAMP	Cyclic adenosine monophosphate
CCI	Chronic-constriction injury
COX2	Cyclo-oxygenase-2 enzyme
CPM	Conditioned pain modulation
DAMP	Danger-associated molecular patterns
DMEM	Dulbecco's modified Eagle's medium
DMEM:F12	DMEM: Ham's F12 mix
DMSO	Dimethyl sulfoxide
DNIC	Diffuse noxious inhibitory control
DRG	Dorsal root ganglia
ELISA	Enzyme-linked immunoassay
EPSC	Excitatory post synaptic current
EPSP	Excitatory postsynaptic potentials
ERK	Extracellular signal-regulated kinase
FBS	Foetal bovine serum

hESCs	Human embryonic stem cells
hiFBS	Heat-inactivated foetal bovine serum
HIV	Human immunodeficiency virus
HPLC	High performance liquid chromatography
hPSCs	Human pluripotent stem cells
IASP	International Association for the Study of Pain
IL-1 β	Interleukin-1 β
IL-6	Interleukin-6
iNOS	Inducible nitric oxide synthase
IPSP	Inhibitory postsynaptic potentials
JNK	c-Jun N-terminal kinase
K ⁺	Potassium ion
L-DOPA	L-3, 4-dihydroxyphenylalanine
LPS	Lipopolysaccharides
MAP2	Microtubule-associated protein 2
MAPK	Mitogen-activated protein kinase
MCP-1	Monocyte chemoattractant protein-1
MTT	(3-(4,5-di methylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
N	Normal control group
Na ⁺	Sodium ion
NEAA	Non-essential amino acids
NECA	5'-N-ethylcarboxamidoadenosine
NMDA NR2B	<i>N</i> -methyl- <i>D</i> -aspartate subtype N2B
NO	Nitric oxide

PAG	Periaqueductal gray
PAMP	Pathogen-associated molecular patterns
PBS	Phosphate buffered saline
PDB	Protein Data Bank
PDMS	Polydimethylsiloxane
PI-3K	Phosphoinositide 3-kinases
PKA	Protein kinase A
PKC	Protein kinase C
PVDF	Polyvinylidene difluoride
RA	Retinoic acid
RAR	Retinoic acid receptor
RVM	Rostroventral medulla
RXR	Retinoid X receptor
SDS-PAGE	Sodium dodecyl sulphate polyacrylamide gel electrophoresis
SEM	Standard error mean
TBI	Traumatic brain injury
TNF- α	Tumour necrosis factor- α
TRP	Transient receptor potential
TRPV1	Transient receptor potential cation channel subfamily V member 1
VEH	Vehicle group
ZER	Zerumbone

CHAPTER 1

INTRODUCTION

1.1 Introduction

Neuropathic pain is a condition that could result in the loss of function and reduce the quality of one's life. The International Association for the Study of Pain (IASP) has interpreted neuropathic pain as a pain that is caused by a lesion or disease on the somatosensory system. The prevalence of neuropathic pain has not been ascertained in most countries in Asia including Malaysia, but, worldwide prevalence has been reported to be 7%, with up to 4- to 6-fold of estimated variation among countries (Zghoul *et al.*, 2017; Ye *et al.*, 2018).

The main mechanism that prompts the development and continuance of neuropathic pain involves peripheral and central sensitization (Cohen and Mao, 2014). The neuronal sensitization could be on account of direct lesion due to injury, viral infections, adverse effects of drugs and other metabolic diseases (Khelemsky, Malhotra and Gritsenko, 2019). In spite of the multifariousness of aetiology and vague underlying mechanism, patients with neuropathic pain share common symptoms which are allodynia, pain due to a stimulus that does not normally provoke pain and hyperalgesia, exaggerated response from a normally painful stimulus respectively (He and Kim, 2020). Other symptoms are burning, numbness, tingling, fatigue and sleep disturbance (Goh *et al.*, 2017).

Current treatments of neuropathic pain involve the management of symptoms, with few approaches targeting the mechanism of neuropathic pain. The first-line therapy involves the use of tricyclic antidepressants, such as amitriptyline; calcium ion channel $\alpha_2\delta$ ligands (anticonvulsants), namely gabapentin; and serotonin-noradrenaline reuptake inhibitor (SNRI) namely duloxetine (Fornasari, 2017; Obata, 2017). Earlier studies have exhibited the mechanism of antidepressants in modulating the descending inhibitory pathway, whereby antidepressants managed to inhibit the reuptake of serotonin and noradrenaline at inhibitory neuron synapse (Fornasari, 2017). Moreover, antidepressants have been observed to activate interneurons within the descending pathways which resulted in the release of inhibitory neurotransmitter namely gamma-aminobutyric acid (GABA) (Fornasari, 2017). Besides, treatment of commonly used antidepressants, amitriptyline, was also proposed to modulate NMDA receptor via the calcium ion-dependent desensitization, resulting in the reduced NMDA-activated currents (Stepanenko *et al.*, 2019). Second-line therapy includes the use of lidocaine which blocks voltage-gated sodium channels; capsaicin 8% patches; and opioids which is also used as third-line therapy (Finnerup *et al.*, 2015; Fornasari, 2017). The established mechanism of lidocaine involved the blockage of voltage-gated sodium channels in peripheral nerves (Kurabe, Furue and Kohno, 2016). Moreover, lidocaine has also been investigated to be able to desensitize transient receptor potential (TRP) channels, thus ameliorate neuropathic pain (Hermanns *et al.*, 2019).

However, due to the complex pathophysiology of neuropathic pain, these treatments were reported to insufficiently provide relief of pain to the patients. Plus, neuropathic pain could also arise from other metabolic diseases, hence making it challenging to decide the accurate treatment. Furthermore, these treatments have been known to have pleiotropic effects which also results in adverse reactions, namely anticholinergic effects, hypertension, seizures, prolongation arrhythmia and nausea (Cavalli *et al.*, 2019).

The foremost mechanism of neuropathic pain are the peripheral and central sensitization which occur because of the alteration of expression of ion channels as well as cellular and molecular changes following the nerve injury and immune response (Khelemsky, Malhotra and Gritsenko, 2019). Monoaminergic pathway, consisting of noradrenergic and serotonergic pathway, could be a prospective target in alleviating neuropathic pain for its role in inhibiting nociceptive signal transmission (Li *et al.*, 2019). α -_{2A} adrenoceptor, one of the key player in modulating the noradrenergic pathway, had been the pharmacological target to alleviate neuropathic pain (Bravo *et al.*, 2019). α -_{2A} adrenoceptor had been reported to be able to modulate other receptor and mediators activation and expression associated to neuropathic pain pathophysiology. As for instances, α -_{2A} adrenoceptor was observed to affect the regulation of glutamate receptors, ion channels and pro-inflammatory mediators, which could synergistically inhibit the nociceptive signal transmission in neuropathic pain (Woo *et al.*, 1997; Liu *et al.*, 2006; Matsushita *et al.*, 2018).

Research have shown that glutamate receptors, specifically *N*-methyl-*D*-aspartate (NMDA) subtype N2B receptors exhibited prominent role in underlying the maintenance of neuropathic pain. In animal models of neuropathic pain, the inhibition or reduction of NMDA N2B receptor expression has been able to attenuate mechanical allodynia and thermal hyperalgesia, the symptoms hallmark of neuropathic pain (Wu *et al.*, 2014; Chia *et al.*, 2020). Alongside NMDA N2B receptor, transient receptor potential subfamily V member 1 (TRPV1) channel has also been proposed to modulate allodynia and hyperalgesia. Similarly to NMDA N2B receptor, the inhibition of expression of TRPV1 channel and the inactivation of the ion channel caused the attenuation of mechanical and thermal allodynia and hyperalgesia (Green *et al.*, 2016; Guo *et al.*, 2019).

The pathophysiology of neuropathic pain also comprises of the role of inflammatory cytokines and chemokines. It is important to take into account that neuronal sensitization and neuroinflammation are not mutually exclusive (Ellis and D.L.H. Bennett, 2013). Commonly identified pro-inflammatory mediators that are suggested to prompt neuropathic pain mechanism are interleukin-6 (IL-6), tumour necrosis factor- α (TNF- α) and nitric oxide (NO) (Perimal *et al.*, 2011a; Colloca, Ludman, Bouhassira, Baron, Anthony H. Dickenson, *et al.*, 2017; Gopalsamy *et al.*, 2017). Upon nerve injury, these inflammatory mediators have been identified as the early markers of neuropathic pain instigation (Ji, Xu and Gao, 2014).

In order to address the current treatment circumstances, research has been exploring the potentials of natural products with pharmacological effectiveness (Quintans *et al.*, 2014).

Herbal medicines have been widely used as ancient therapies to treat various illnesses such as inflammation and pain. One of the moderately large genus of herbs is genus *Zingiber*, with estimated 141 species found mostly within South East Asia and Pacific Islands (Yob *et al.*, 2011). Within the genus *Zingiber*, *Zingiber zerumbet* has been gaining researchers' interest due to its anti-inflammatory and anticancer properties (Gopalsamy *et al.*, 2017; Girisa *et al.*, 2019a). The plant has been traditionally used in the to treat stomach ache, indigestion, fever and other ailments (Fatima *et al.*, 2015). The bioactive sesquiterpene from *Zingiber zerumbet* that is extensively studied is zerumbone. Highly concentrated within the rhizomes of *Zingiber zerumbet*, zerumbone has been observed to possess potentials as anti-proliferative for cancer treatment, antioxidant, antinociceptive and anti-inflammatory effects (Perimal *et al.*, 2011b; Yob *et al.*, 2011; Gopalsamy *et al.*, 2017; Yan *et al.*, 2017).

Although research has provided some information on the anti-allodynic and anti-hyperalgesic effects of zerumbone in *in vivo* models, the mechanism of action of zerumbone in alleviating neuropathic pain is yet to be understood. Plus, animals usage in conducting pain research has been getting ethical concerns, hence there is the need to develop an *in vitro* model to mimic the pathophysiology of neuropathic pain to further evaluate the therapeutic effects of zerumbone.

Several cell lines have been commonly used in neuroscience research, by which in this study, SH-SY5Y cells have been determined to be develop as an *in vitro* model to mimic the pathophysiology of neuropathic pain. This is because SH-SY5Y cells are human neuroblastoma cell line, which eliminates the species variation difference observed in animal cell lines (Namita G Hattangady and Rajadhyaksha, 2009; Kovalevich and Langford, 2013). Furthermore, SH-SY5Y cells were able to be induced to the differentiated state which express important receptors and ion channels involved in nociceptive signal transmission (Encinas *et al.*, 2000; Shipley, Mangold and Szpara, 2017). In order to trigger the development of neuroinflammation and change the expression of receptors and ion channels incorporated in neuropathic pain mechanism, lipopolysaccharides (LPS) induction was opted for to induce the condition in SH-SY5Y cell culture. LPS-induced SH-SY5Y cells have been established to enable the study of oxidative stress, inflammatory mediators, such as interleukins, and proteins associated with pain signal transmission, namely TRP channels and glutamate receptors, attributable to the activation of toll-like receptors (TLR) by LPS (Meseguer *et al.*, 2014; Pandur *et al.*, 2018).

It was hypothesized that the anti-neuropathic pain properties of zerumbone involved the modulation of α_{2A} adrenoceptor, TRPV1 channel, NMDA N2B receptors and pro-inflammatory mediators namely nitric oxide, IL-6 and TNF- α . The findings of this research will help to understand the modulation of the receptors in the pathophysiology of neuropathic pain in *in vitro* model and develop zerumbone as promising therapeutics to attenuate neuropathic pain.

1.2 General Objectives

To study the effect of zerumbone on α_{2A} adrenoceptor, NMDA N2B receptor and TRPV1 channel expression and the pro-inflammatory mediators on *in vitro* model of neuropathic pain-like lipopolysaccharide (LPS)-induced SH-SY5Y cells.

1.3 Specific Objectives

- To determine the effect of different culture medium on the growth of SH-SY5Y cells, to induce differentiation of SH-SY5Y cells and to characterize the differentiated cells via immunostaining of differentiated neuronal markers.
- To determine the effect of zerumbone on the expression of nitric oxide, interleukin-6 and tumour necrosis factor- α in LPS-induced SH-SY5Y cells.
- To study the anti-neuropathic pain effect of zerumbone in LPS-induced SH-SY5Y cells via the modulation of α_{2A} adrenoceptor through molecular docking and Western blot analysis.
- To determine the anti-neuropathic pain effect of zerumbone in LPS-induced SH-SY5Y cells via the modulation of NMDA N2B receptor through molecular docking and Western blot analysis.
- To investigate the anti-neuropathic pain effect of zerumbone in LPS-induced SH-SY5Y cells via the modulation of TRPV1 channel through molecular docking and Western blot analysis.

REFERENCES

- Adams, R. D. *et al.* (2014) 'Electrical and neurotrophin enhancement of neurite outgrowth within a 3D collagen scaffold.', *Annals of biomedical engineering*. NIH Public Access, 42(6), pp. 1282–91. doi: 10.1007/s10439-014-1001-0.
- Afrazi, S. *et al.* (2014) 'Journal of Steroid Biochemistry and Molecular Biology Neurosteroid allopregnanolone attenuates high glucose-induced apoptosis and prevents experimental diabetic neuropathic pain : In vitro and in vivo studies', *Journal of Steroid Biochemistry and Molecular Biology*. Elsevier Ltd, 139, pp. 98–103. doi: 10.1016/j.jsbmb.2013.10.010.
- Aiyer, R. *et al.* (2017) 'A Systematic Review of NMDA Receptor Antagonists for Treatment of Neuropathic Pain in Clinical Practice', *The Clinical Journal of Pain*, 34(5), p. 1. doi: 10.1097/AJP.0000000000000547.
- Akinci, A. *et al.* (2016) 'Predictive factors and clinical biomarkers for treatment in patients with chronic pain caused by osteoarthritis with a central sensitisation component', *International Journal of Clinical Practice*, 70(1), pp. 31–44. doi: 10.1111/ijcp.12749.
- Al-Zubairi, A. S. (2018) 'Anti-Proliferative Activity of Zerumbone Against Tumour Cell Lines 1,2', *OnLine Journal of Biological Sciences Original Research Paper*. doi: 10.3844/ojbsci.2018.123.129.
- Aletta, J. M. (1996) 'Phosphorylation of type III β -tubulin in PC 12 cell neurites during NGF-induced process outgrowth', *Journal of Neurobiology*, 31(4), pp. 461–475. doi: 10.1002/(SICI)1097-4695(199612)31:4<461::AID-NEU6>3.0.CO;2-7.
- Alles, S. R. A. and Smith, P. A. (2018) 'Etiology and Pharmacology of Neuropathic Pain', *Pharmacological Reviews Pharmacol Rev*, 70, pp. 315–347. doi: 10.1124/pr.117.014399.
- Anaeigoudari, A. *et al.* (2016) 'Neuronal nitric oxide synthase has a role in the detrimental effects of lipopolysaccharide on spatial memory and synaptic plasticity in rats', *Pharmacological Reports*, 68(2), pp. 243–249. doi: 10.1016/j.pharep.2015.09.004.
- Arthur, C. P. and Stowell, M. H. B. (2007) 'Structure of synaptophysin: a hexameric MARVEL-domain channel protein.', *Structure (London, England : 1993)*. Structure, 15(6), pp. 707–14. doi: 10.1016/j.str.2007.04.011.
- Bahari, Z. and Meftahi, G. H. (2019) 'Spinal α_2 -adrenoceptors and neuropathic pain modulation; therapeutic target', *British Journal of Pharmacology*. Wiley-Blackwell, 176(14), pp. 2366–2381. doi: 10.1111/bph.14580.
- Bannister, K. and Dickenson, A. H. (2016) 'What do monoamines do in pain modulation?', *Current opinion in supportive and palliative care*. Europe PMC Funders, 10(2), pp. 143–8. doi: 10.1097/SPC.0000000000000207.

- Berta, T. *et al.* (2017) 'Targeting dorsal root ganglia and primary sensory neurons for the treatment of chronic pain.', *Expert opinion on therapeutic targets*. NIH Public Access, 21(7), pp. 695–703. doi: 10.1080/14728222.2017.1328057.
- Billard, J.-M. (2018) 'Changes in Serine Racemase-Dependent Modulation of NMDA Receptor: Impact on Physiological and Pathological Brain Aging', *Frontiers in Molecular Biosciences*. Frontiers, 5, p. 106. doi: 10.3389/fmolb.2018.00106.
- Blackshaw, L. A., Brierley, S. M. and Hughes, P. A. (2010) 'TRP channels: new targets for visceral pain', *Gut*, 59(01), pp. 126–135. doi: 10.1136/gut.2009.179523.
- Bölcskei, K. *et al.* (2005) 'Investigation of the role of TRPV1 receptors in acute and chronic nociceptive processes using gene-deficient mice.', *Pain*. Pain, 117(3), pp. 368–376. doi: 10.1016/j.pain.2005.06.024.
- Bouhassira, D. (2019) 'Neuropathic pain: Definition, assessment and epidemiology', *Revue Neurologique*. Elsevier Masson, 175(1–2), pp. 16–25. doi: 10.1016/J.NEUROL.2018.09.016.
- Brandt, M. R., Beyer, C. E. and Stahl, S. M. (2012) 'TRPV1 Antagonists and Chronic Pain: Beyond Thermal Perception.', *Pharmaceuticals (Basel, Switzerland)*. Multidisciplinary Digital Publishing Institute (MDPI), 5(2), pp. 114–32. doi: 10.3390/ph5020114.
- Bravo, L. *et al.* (2019) 'Monoamines as Drug Targets in Chronic Pain: Focusing on Neuropathic Pain', *Frontiers in Neuroscience*. Frontiers, 13, p. 1268. doi: 10.3389/fnins.2019.01268.
- Brightwell, J. J. and Taylor, B. K. (2009) 'Noradrenergic neurons in the locus coeruleus contribute to neuropathic pain.', *Neuroscience*. NIH Public Access, 160(1), pp. 174–85. doi: 10.1016/j.neuroscience.2009.02.023.
- Bu, F. *et al.* (2015) 'Phosphorylation of NR2B NMDA subunits by protein kinase C in arcuate nucleus contributes to inflammatory pain in rats', *Scientific Reports*. Nature Publishing Group, 5(1), p. 15945. doi: 10.1038/srep15945.
- Carrasco, C. *et al.* (2018) 'Neuropathic Pain: Delving into the Oxidative Origin and the Possible Implication of Transient Receptor Potential Channels.', *Frontiers in physiology*. Frontiers Media SA, 9, p. 95. doi: 10.3389/fphys.2018.00095.
- Cavalli, E. *et al.* (2019) 'The neuropathic pain: An overview of the current treatment and future therapeutic approaches.', *International journal of immunopathology and pharmacology*. SAGE Publications, 33, p. 2058738419838383. doi: 10.1177/2058738419838383.
- Chae, S. (2004) 'Protection of insulin secreting cells from nitric oxide induced cellular damage by crosslinked hemoglobin', *Biomaterials*, 25(5), pp. 843–850. doi: 10.1016/S0142-9612(03)00605-7.
- Chambers, S. M. *et al.* (2013) 'Dual-SMAD Inhibition/WNT Activation-Based Methods to Induce Neural Crest and Derivatives from Human Pluripotent Stem Cells', in. Humana Press, New York, NY, pp. 329–343. doi: 10.1007/7651_2013_59.

- Chanchal, S. K. *et al.* (2016) 'In vivo and in vitro protective effects of omeprazole against neuropathic pain', *Scientific Reports*. Nature Publishing Group, 6(1), p. 30007. doi: 10.1038/srep30007.
- Chen, C. *et al.* (2019) 'Develop a 3D neurological disease model of human cortical glutamatergic neurons using micropillar-based scaffolds', *Acta Pharmaceutica Sinica B*. Elsevier, 9(3), pp. 557–564. doi: 10.1016/J.APSB.2019.03.004.
- Chen, J.-Y. *et al.* (2018) 'Valproate reduces neuroinflammation and neuronal death in a rat chronic constriction injury model.', *Scientific reports*. Nature Publishing Group, 8(1), p. 16457. doi: 10.1038/s41598-018-34915-5.
- Chen, T. *et al.* (2017) 'Interactions of Notch1 and TLR4 signaling pathways in DRG neurons of in vivo and in vitro models of diabetic neuropathy', *Scientific Reports*. Nature Publishing Group, 7(1), p. 14923. doi: 10.1038/s41598-017-15053-w.
- Cheung, Y.-T. *et al.* (2009) 'Effects of all-trans-retinoic acid on human SH-SY5Y neuroblastoma as in vitro model in neurotoxicity research', *NeuroToxicology*. Elsevier, 30(1), pp. 127–135. doi: 10.1016/J.NEURO.2008.11.001.
- Chia, J. S. M. *et al.* (2016) 'Zerumbone alleviates chronic constriction injury-induced allodynia and hyperalgesia through serotonin 5-HT receptors', *Biomedicine & Pharmacotherapy*. Elsevier Masson, 83, pp. 1303–1310. doi: 10.1016/J.BIOPHA.2016.08.052.
- Chia, J. S. M. (2018) *Anti-allodynic and antihyperalgesic effects of zerumbone through involvement of monoaminergic pathways in mice model of neuropathic pain*. Available at: http://psasir.upm.edu.my/id/eprint/69665/1/fpsk_2018_33_ir.pdf (Accessed: 9 January 2020).
- Chia, J. S. M. *et al.* (2020) 'Zerumbone Modulates α 2A-Adrenergic, TRPV1, and NMDA NR2B Receptors Plasticity in CCI-Induced Neuropathic Pain In Vivo and LPS-Induced SH-SY5Y Neuroblastoma In Vitro Models', *Frontiers in Pharmacology*. Frontiers, 11, p. 92. doi: 10.3389/fphar.2020.00092.
- Ciccarelli, M. *et al.* (2017) 'Adrenergic Receptors', *Endocrinology of the Heart in Health and Disease*. Academic Press, pp. 285–315. doi: 10.1016/B978-0-12-803111-7.00011-7.
- Cohen, S. P. and Mao, J. (2014) 'Neuropathic pain: mechanisms and their clinical implications', *BMJ*, 348(feb05 6), pp. f7656–f7656. doi: 10.1136/bmj.f7656.
- Colloca, L., Ludman, T., Bouhassira, D., Baron, R., Dickenson, Anthony H., *et al.* (2017) 'Neuropathic pain', *Nature Reviews Disease Primers*. Nature Publishing Group, 3(1), p. 17002. doi: 10.1038/nrdp.2017.2.
- Colloca, L., Ludman, T., Bouhassira, D., Baron, R., Dickenson, Anthony H., *et al.* (2017) 'Neuropathic pain', *Nature Publishing Group*. Macmillan Publishers Limited, 3, pp. 1–20. doi: 10.1038/nrdp.2017.2.
- Costa, A. R. *et al.* (2019) 'Neuropathic Pain Induced Alterations in the Opioidergic Modulation of a Descending Pain Facilitatory Area of the Brain', *Frontiers in Cellular Neuroscience*. Frontiers, 13, p. 287. doi: 10.3389/fncel.2019.00287.

- Costigan, M., Scholz, J. and Woolf, C. J. (2009) 'Neuropathic Pain: A Maladaptive Response of the Nervous System to Damage', *Annual Review of Neuroscience*, 32(1), pp. 1–32. doi: 10.1146/annurev.neuro.051508.135531.
- Courtney, C. A., Fernández-De-Las-Peñas, C. and Bond, S. (2017) 'Mechanisms of chronic pain – key considerations for appropriate physical therapy management', *Journal of Manual & Manipulative therapy*, 25(3), pp. 118–127. doi: 10.1080/10669817.2017.1300397.
- Czapski, G. A. *et al.* (2007) 'Role of nitric oxide in the brain during lipopolysaccharide-evoked systemic inflammation', *Journal of Neuroscience Research*, 85(8), pp. 1694–1703. doi: 10.1002/jnr.21294.
- Damasceno, D. C. *et al.* (2014) 'Streptozotocin-induced diabetes models: Pathophysiological mechanisms and fetal outcomes', *BioMed Research International*, 2014. doi: 10.1155/2014/819065.
- Damien, J. *et al.* (2018) 'Pain Modulation: From Conditioned Pain Modulation to Placebo and Nocebo Effects in Experimental and Clinical Pain.', *International review of neurobiology*. NIH Public Access, 139, pp. 255–296. doi: 10.1016/bs.irm.2018.07.024.
- Das, N. D. *et al.* (2012a) 'Lipopolysaccharide-mediated protein expression profiling on neuronal differentiated SH-SY5Y cells', *Biochip Journal*, 6(2), pp. 165–173. doi: 10.1007/s13206-012-6209-1.
- Das, N. D. *et al.* (2012b) 'Lipopolysaccharide-mediated protein expression profiling on neuronal differentiated SH-SY5Y cells', *BioChip Journal*, 6(2), pp. 165–173. doi: 10.1007/s13206-012-6209-1.
- Das, V. (2015) 'An Introduction to Pain Pathways and Pain “Targets”', in, pp. 1–30. doi: 10.1016/bs.pmbts.2015.01.003.
- Datta, G. *et al.* (2019) 'HIV-1 gp120 Promotes Lysosomal Exocytosis in Human Schwann Cells.', *Frontiers in cellular neuroscience*. Frontiers Media SA, 13, p. 329. doi: 10.3389/fncel.2019.00329.
- Deng, M., Chen, S.-R. and Pan, H.-L. (2019) 'Presynaptic NMDA receptors control nociceptive transmission at the spinal cord level in neuropathic pain', *Cellular and Molecular Life Sciences*, 76(10), pp. 1889–1899. doi: 10.1007/s00018-019-03047-y.
- Dong, C.-J. *et al.* (2008) ' α 2 Adrenergic Modulation of NMDA Receptor Function as a Major Mechanism of RGC Protection in Experimental Glaucoma and Retinal Excitotoxicity', *Investigative Ophthalmology & Visual Science*. The Association for Research in Vision and Ophthalmology, 49(10), p. 4515. doi: 10.1167/iovs.08-2078.
- Du, Q. *et al.* (2019) 'The Role of Transient Receptor Potential Vanilloid 1 in Common Diseases of the Digestive Tract and the Cardiovascular and Respiratory System', *Frontiers in Physiology*. Frontiers, 10, p. 1064. doi: 10.3389/fphys.2019.01064.

- Dureja, G. P. *et al.* (2017) 'Evidence and consensus recommendations for the pharmacological management of pain in India', *Journal of Pain Research*. Dove Press, Volume 10, pp. 709–736. doi: 10.2147/JPR.S128655.
- Eid, E. E. M. *et al.* (2018) 'Zerumbone binding to estrogen receptors: An in-silico investigation', *Journal of Receptors and Signal Transduction*. Taylor & Francis, 38(4), pp. 342–351. doi: 10.1080/10799893.2018.1531886.
- Eldridge, S., Guo, L. and Hamre, J. (2019) 'A Comparative Review of Chemotherapy-Induced Peripheral Neuropathy in In Vivo and In Vitro Models', *Toxicologic Pathology*, 48(1), pp. 190–201. doi: 10.1177/0192623319861937.
- Ellis, A. and Bennett, D.L.H. (2013) 'Neuroinflammation and the generation of neuropathic pain', *British Journal of Anaesthesia*. Elsevier, 111(1), pp. 26–37. doi: 10.1093/bja/aet128.
- Ellis, A. and Bennett, D L H (2013) 'Neuroinflammation and the generation of neuropathic pain', *British Journal of Anaesthesia*, 111(1), pp. 26–37. doi: 10.1093/bja/aet128.
- Elmann, A. *et al.* (2017) 'Glutamate Toxicity to Differentiated Neuroblastoma N2a Cells Is Prevented by the Sesquiterpene Lactone Achillolide A and the Flavonoid 3,5,4'-Trihydroxy-6,7,3'-Trimethoxyflavone from *Achillea fragrantissima*', *Journal of Molecular Neuroscience*, 62(1), pp. 99–105. doi: 10.1007/s12031-017-0916-y.
- Encinas, M. *et al.* (2000) 'Sequential treatment of SH-SY5Y cells with retinoic acid and brain-derived neurotrophic factor gives rise to fully differentiated, neurotrophic factor-dependent, human neuron-like cells.', *Journal of neurochemistry*. J Neurochem, 75(3), pp. 991–1003. doi: 10.1046/j.1471-4159.2000.0750991.x.
- Fang, C.-Y. *et al.* (2017) 'Long-term growth comparison studies of FBS and FBS alternatives in six head and neck cell lines', *PLOS ONE*. Edited by J. J. Cray. Public Library of Science, 12(6), p. e0178960. doi: 10.1371/journal.pone.0178960.
- Fatima, A. *et al.* (2015) 'Binding Mode Analysis of Zerumbone to Key Signal Proteins in the Tumor Necrosis Factor Pathway', pp. 2747–2766. doi: 10.3390/ijms16022747.
- Fatima, A. *et al.* (2018) 'Docking studies reveal zerumbone targets β -catenin of the Wnt- β -catenin pathway in breast cancer', *Journal of the Serbian Chemical Society*, 83(5), pp. 575–591. doi: 10.2298/JSC170313108F.
- Feizerfan, A. and Sheh, G. (2015) 'Transition from acute to chronic pain', *Continuing Education in Anaesthesia Critical Care & Pain*. Narnia, 15(2), pp. 98–102. doi: 10.1093/bjaceaccp/mku044.
- Fernyhough, P., Huang, T.-J. and Verkhatsky, A. (2003) 'Mechanism of mitochondrial dysfunction in diabetic sensory neuropathy', *Journal of the Peripheral Nervous System*, 8(4), pp. 227–235. doi: 10.1111/j.1085-9489.2003.03028.x.

- Finnerup, N. B. *et al.* (2015) 'Pharmacotherapy for neuropathic pain in adults: a systematic review and meta-analysis.', *The Lancet. Neurology*. NIH Public Access, 14(2), pp. 162–73. doi: 10.1016/S1474-4422(14)70251-0.
- Fornasari, D. (2017) 'Pharmacotherapy for Neuropathic Pain: A Review.', *Pain and therapy*. Springer, 6(Suppl 1), pp. 25–33. doi: 10.1007/s40122-017-0091-4.
- Forrest, C. M. *et al.* (2017) 'Quinolinic acid induces neuritogenesis in SH-SY5Y neuroblastoma cells independently of NMDA receptor activation', 45, pp. 700–711. doi: 10.1111/ejn.13499.
- Forster, J. I. *et al.* (2016) 'Characterization of Differentiated SH-SY5Y as Neuronal Screening Model Reveals Increased Oxidative Vulnerability.', *Journal of biomolecular screening*. SAGE Publications, 21(5), pp. 496–509. doi: 10.1177/1087057115625190.
- Förstermann, U. and Sessa, W. C. (2012) 'Nitric oxide synthases: regulation and function.', *European heart journal*. Oxford University Press, 33(7), pp. 829–37, 837a-837d. doi: 10.1093/eurheartj/ehr304.
- Fregnan, F. *et al.* (2012) 'Role of inflammatory cytokines in peripheral nerve injury.', *Neural regeneration research*. Wolters Kluwer -- Medknow Publications, 7(29), pp. 2259–66. doi: 10.3969/j.issn.1673-5374.2012.29.003.
- Freire, M. A. M. *et al.* (2009) 'Pain modulation by nitric oxide in the spinal cord.', *Frontiers in Neuroscience*. Frontiers, 3(2), pp. 175–181. doi: 10.3389/neuro.01.024.2009.
- Fukuoka, T. *et al.* (2002) 'VR1, but not P2X(3), increases in the spared L4 DRG in rats with L5 spinal nerve ligation.', *Pain*. Pain, 99(1–2), pp. 111–20. doi: 10.1016/s0304-3959(02)00067-2.
- Gao, W. *et al.* (2016) 'Quercetin ameliorates paclitaxel-induced neuropathic pain by stabilizing mast cells, and subsequently blocking PKC ϵ -dependent activation of TRPV1.', *Acta pharmacologica Sinica*. Acta Pharmacol Sin, 37(9), pp. 1166–77. doi: 10.1038/aps.2016.58.
- Garland, E. L. (2012) 'Pain processing in the human nervous system: a selective review of nociceptive and biobehavioral pathways.', *Primary care*. NIH Public Access, 39(3), pp. 561–71. doi: 10.1016/j.pop.2012.06.013.
- Gayle, D. A. *et al.* (2002) 'Lipopolysaccharide (LPS)-induced dopamine cell loss in culture: roles of tumor necrosis factor- α , interleukin-1 β , and nitric oxide', *Developmental Brain Research*, 133(1), pp. 27–35. doi: 10.1016/S0165-3806(01)00315-7.
- Gerard, E. *et al.* (2015) 'Chronic constriction injury-induced nociception is relieved by nanomedicine-mediated decrease of rat hippocampal tumor necrosis factor.', *Pain*. Pain, 156(7), pp. 1320–33. doi: 10.1097/j.pain.000000000000181.
- Ghimire, K. *et al.* (2017) 'Nitric oxide: what's new to NO?', *American Journal of Physiology-Cell Physiology*. American Physiological Society Bethesda, MD, 312(3), pp. C254–C262. doi: 10.1152/ajpcell.00315.2016.

- Giovannitti, J. A. *et al.* (2015) 'Alpha-2 adrenergic receptor agonists: a review of current clinical applications.', *Anesthesia progress*. American Dental Society of Anesthesiology, 62(1), pp. 31–9. doi: 10.2344/0003-3006-62.1.31.
- Girisa, S. *et al.* (2019a) 'Potential of Zerumbone as an Anti-Cancer Agent', *Molecules*. Multidisciplinary Digital Publishing Institute (MDPI), 24(4). doi: 10.3390/MOLECULES24040734.
- Girisa, S. *et al.* (2019b) 'Potential of Zerumbone as an Anti-Cancer Agent', *Molecules*. Multidisciplinary Digital Publishing Institute (MDPI), 24(4). doi: 10.3390/MOLECULES24040734.
- Goh, L.-Y. *et al.* (2017) *The prevalence and associated factors of neuropathic pain symptoms in a cohort of multi-ethnic Malaysian patients with diabetes mellitus, Neurology Asia*. Available at: [https://www.neurology-asia.org/articles/neuroasia-2017-22\(4\)-325.pdf](https://www.neurology-asia.org/articles/neuroasia-2017-22(4)-325.pdf) (Accessed: 28 January 2021).
- Gopalsamy, B. *et al.* (2017) 'Antiallodynic and antihyperalgesic activities of zerumbone via the suppression of IL-1 beta, IL-6, and TNF-alpha in a mouse model of neuropathic pain', *Journal of Pain Research*, 10, pp. 2605–2619. doi: 10.2147/JPR.S143024.
- Gordon, J., Amini, S. and White, M. K. (2013) 'Neuronal Cell Culture', 1078, pp. 1–6. doi: 10.1007/978-1-62703-640-5.
- Greaves, M. and Maley, C. C. (2012) 'Clonal evolution in cancer', *Nature*, 481(7381), pp. 306–313. doi: 10.1038/nature10762.
- Green, D. *et al.* (2016) 'Central activation of TRPV1 and TRPA1 by novel endogenous agonists contributes to mechanical allodynia and thermal hyperalgesia after burn injury.', *Molecular pain*. SAGE Publications, 12. doi: 10.1177/1744806916661725.
- Gregory, N. S. *et al.* (2013) 'An overview of animal models of pain: Disease models and outcome measures', *Journal of Pain*. Elsevier Ltd, 14(11), pp. 1255–1269. doi: 10.1016/j.jpain.2013.06.008.
- Guix, F. X. *et al.* (2005) 'The physiology and pathophysiology of nitric oxide in the brain', *Progress in Neurobiology*, 76(2), pp. 126–152. doi: 10.1016/j.pneurobio.2005.06.001.
- Guo, S.-H. *et al.* (2019) 'Silencing of spinal Trpv1 attenuates neuropathic pain in rats by inhibiting CAMKII expression and ERK2 phosphorylation', *Scientific Reports*. Nature Publishing Group, 9(1), p. 2769. doi: 10.1038/s41598-019-39184-4.
- Gwak, Y. S. and Hulsebosch, C. E. (2011) 'Neuronal hyperexcitability: A substrate for central neuropathic pain after spinal cord injury', *Current Pain and Headache Reports*, 15(3), pp. 215–222. doi: 10.1007/s11916-011-0186-2.
- Haque, A., Jantan, I. and Harikrishnan, H. (2018) 'International Immunopharmacology Zerumbone suppresses the activation of inflammatory mediators in LPS-stimulated U937 macrophages through MyD88-dependent NF- κ B / MAPK / PI3K-Akt signaling pathways', *International Immunopharmacology*. Elsevier, 55(December 2017), pp. 312–322. doi: 10.1016/j.intimp.2018.01.001.

- Hattangady, Namita G and Rajadhyaksha, M. S. (2009) 'A brief review of in vitro models of diabetic neuropathy.', *International journal of diabetes in developing countries*. Springer, 29(4), pp. 143–9. doi: 10.4103/0973-3930.57344.
- Hattangady, Namita G. and Rajadhyaksha, M. S. (2009) 'A brief review of in vitro models of diabetic neuropathy', *International Journal of Diabetes in Developing Countries*, 29(4), pp. 143–149. doi: 10.4103/0973-3930.57344.
- Hattangady, Namita G and Rajadhyaksha, M. (2009) 'A brief review of *in vitro* models of diabetic neuropathy', *International Journal of Diabetes in Developing Countries*, 29(4), p. 143. doi: 10.4103/0973-3930.57344.
- Hayashida, K. and Eisenach, J. C. (2010) 'Spinal α 2-Adrenoceptor-mediated Analgesia in Neuropathic Pain Reflects Brain-derived Nerve Growth Factor and Changes in Spinal Cholinergic Neuronal Function', *Anesthesiology*. American Society of Anesthesiologists, 113(2), pp. 406–412. doi: 10.1097/ALN.0b013e3181de6d2c.
- Hayashida, K. and Obata, H. (2019) 'Strategies to Treat Chronic Pain and Strengthen Impaired Descending Noradrenergic Inhibitory System', *International Journal of Molecular Sciences*. Multidisciplinary Digital Publishing Institute, 20(4), p. 822. doi: 10.3390/ijms20040822.
- He, Y. and Kim, P. Y. (2020) *Allodynia*, StatPearls. StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/30725814> (Accessed: 27 January 2021).
- Heinricher, M. M. (2016) 'Pain Modulation and the Transition from Acute to Chronic Pain', in, pp. 105–115. doi: 10.1007/978-94-017-7537-3_8.
- Heir, R. and Stellwagen, D. (2020) 'TNF-Mediated Homeostatic Synaptic Plasticity: From in vitro to in vivo Models', *Frontiers in Cellular Neuroscience*. Frontiers, 14, p. 297. doi: 10.3389/fncel.2020.565841.
- Hermanns, H. *et al.* (2019) 'Molecular mechanisms of action of systemic lidocaine in acute and chronic pain: a narrative review', *British Journal of Anaesthesia*. Elsevier, 123(3), pp. 335–349. doi: 10.1016/J.BJA.2019.06.014.
- Hosobuchi, Y., Adams, J. and Linchitz, R. (1977) 'Pain relief by electrical stimulation of the central gray matter in humans and its reversal by naloxone', *Science*, 197(4299), pp. 183–186. doi: 10.1126/science.301658.
- Huang, E. P. (1997) 'Synaptic plasticity: A role for nitric oxide in LTP', *Current Biology*. Cell Press, 7(3), pp. R141–R143. doi: 10.1016/S0960-9822(97)70073-3.
- Huang, L.-E. *et al.* (2018) 'N-methyl D-aspartate receptor subtype 2B antagonist, Ro 25-6981, attenuates neuropathic pain by inhibiting postsynaptic density 95 expression', *Scientific Reports*. Nature Publishing Group, 8(1), p. 7848. doi: 10.1038/s41598-018-26209-7.
- Hurtado-Zavala, J. I. *et al.* (2017) 'TRPV1 regulates excitatory innervation of OLM neurons in the hippocampus.', *Nature communications*. Nature Publishing Group, 8, p. 15878. doi: 10.1038/ncomms15878.

- Hutchinson, M. R. *et al.* (2010) 'Evidence that tricyclic small molecules may possess toll-like receptor and myeloid differentiation protein 2 activity.', *Neuroscience*. NIH Public Access, 168(2), pp. 551–63. doi: 10.1016/j.neuroscience.2010.03.067.
- Hutson, P., Backonja, M. and Knurr, H. (2015) 'Intravenous Lidocaine for Neuropathic Pain: A Retrospective Analysis of Tolerability and Efficacy', *Pain Medicine*. Oxford Academic, 16(3), pp. 531–536. doi: 10.1111/pme.12642.
- IASP Terminology - IASP* (2017). Available at: <https://www.iasp-pain.org/Education/Content.aspx?ItemNumber=1698> (Accessed: 2 December 2019).
- Inc., N. T. (2014) *DMEM/Ham's F-12 with L-Gln, Sodium Pyruvate and HEPES Nacalai Tesque*.
- Institute of Medicine (US) Committee on Pain, D. and C. I. B. *et al.* (1987) 'The Anatomy and Physiology of Pain'. National Academies Press (US). Available at: <https://www.ncbi.nlm.nih.gov/books/NBK219252/> (Accessed: 14 October 2020).
- Jämsä, A. *et al.* (2004) 'The retinoic acid and brain-derived neurotrophic factor differentiated SH-SY5Y cell line as a model for Alzheimer's disease-like tau phosphorylation', *Biochemical and Biophysical Research Communications*, 319(3), pp. 993–1000. doi: 10.1016/j.bbrc.2004.05.075.
- Jara-Oseguera, A., Simon, S. A. and Rosenbaum, T. (2008) 'TRPV1: on the road to pain relief.', *Current molecular pharmacology*. NIH Public Access, 1(3), pp. 255–69. doi: 10.2174/1874467210801030255.
- Jardín, I. *et al.* (2017) 'TRPs in Pain Sensation', *Frontiers in Physiology*. Frontiers, 8, p. 392. doi: 10.3389/fphys.2017.00392.
- Jay, G. W. and Barkin, R. L. (2014) 'Neuropathic pain: Etiology, pathophysiology, mechanisms, and evaluations', *Disease-a-Month*. Elsevier, 60(1), pp. 6–47. doi: 10.1016/j.disamonth.2013.12.001.
- Jedema, H. P. *et al.* (2008) 'Chronic cold exposure increases RGS7 expression and decreases alpha(2)-autoreceptor-mediated inhibition of noradrenergic locus coeruleus neurons.', *The European journal of neuroscience*. NIH Public Access, 27(9), pp. 2433–43. doi: 10.1111/j.1460-9568.2008.06208.x.
- Ji, R.-R., Xu, Z.-Z. and Gao, Y.-J. (2014) 'Emerging targets in neuroinflammation-driven chronic pain', *Nature Reviews Drug Discovery*, 13(7), pp. 533–548. doi: 10.1038/nrd4334.
- Jia, D. *et al.* (2019) 'Tempol Attenuates Neuropathic Pain by Inhibiting Nitric Oxide Production', *Analytical Cellular Pathology*, 2019, pp. 1–5. doi: 10.1155/2019/8253850.
- Jones, I. *et al.* (2018) 'Development and validation of an in vitro model system to study peripheral sensory neuron development and injury', *Scientific Reports*. Nature Publishing Group, 8(1), p. 15961. doi: 10.1038/s41598-018-34280-3.

- Jung, J. *et al.* (2004) 'Phosphorylation of Vanilloid Receptor 1 by Ca²⁺/Calmodulin-dependent Kinase II Regulates Its Vanilloid Binding', *Journal of Biological Chemistry*. Elsevier, 279(8), pp. 7048–7054. doi: 10.1074/jbc.M311448200.
- Jung, J. *et al.* (2020) 'Comparison of the use of opioids only and pregabalin add-on for the treatment of neuropathic pain in cervical myelopathy patients: a pilot trial', *Scientific Reports*. Nature Publishing Group, 10(1), p. 8120. doi: 10.1038/s41598-020-65108-8.
- Jurga, A. M. *et al.* (2016) 'Blockade of Toll-Like Receptors (TLR2, TLR4) Attenuates Pain and Potentiates Buprenorphine Analgesia in a Rat Neuropathic Pain Model.', *Neural plasticity*. Hindawi Limited, 2016, p. 5238730. doi: 10.1155/2016/5238730.
- Kaeidi, A. *et al.* (2011) 'Olive (*Olea europaea* L.) leaf extract attenuates early diabetic neuropathic pain through prevention of high glucose-induced apoptosis: In vitro and in vivo studies', *Journal of Ethnopharmacology*. Elsevier Ireland Ltd, 136(1), pp. 188–196. doi: 10.1016/j.jep.2011.04.038.
- Kaswan, N. K. *et al.* (2020) 'Cardamonin inhibits nitric oxide production modulated through NMDA receptor in LPS-Induced SH-SY5Y cell in vitro model', *Life Sciences, Medicine and Biomedicine*, 4(9). doi: 10.28916/lsm.4.9.2020.58.
- Katsetos, C. D. *et al.* (2003) 'Class III β -Tubulin Isotype: A Key Cytoskeletal Protein at the Crossroads of Developmental Neurobiology and Tumor Neuropathology', *Journal of Child Neurology*, 18(12), pp. 851–866. doi: 10.1177/088307380301801205.
- Kaur, G. and Dufour, J. M. (2012) 'Cell lines: Valuable tools or useless artifacts.', *Spermatogenesis*, 2(1), pp. 1–5. doi: 10.4161/spmg.19885.
- Khan, S. U. *et al.* (2020) 'Illustrated step by step protocol to perform molecular docking: Human estrogen receptor complex with 4-hydroxytamoxifen as a case study', *Progress in Drug Discovery & Biomedical Science*, 3(1). doi: 10.36877/pddbs.a0000054.
- Khatib, T. *et al.* (2019) 'Genomic and non-genomic pathways are both crucial for peak induction of neurite outgrowth by retinoids', *Cell Communication and Signaling*. BioMed Central, 17(1), p. 40. doi: 10.1186/s12964-019-0352-4.
- Khelemsky, Y., Malhotra, A. and Gritsenko, K. (eds) (2019) *Academic Pain Medicine*. Cham: Springer International Publishing. doi: 10.1007/978-3-030-18005-8.
- Kiguchi, N. *et al.* (2017) 'Pharmacological Regulation of Neuropathic Pain Driven by Inflammatory Macrophages', *International Journal of Molecular Sciences*. Multidisciplinary Digital Publishing Institute, 18(11), p. 2296. doi: 10.3390/ijms18112296.
- Kim, M.-J. and Yun, J.-M. (2019) 'Molecular Mechanism of the Protective Effect of Zerumbone on Lipopolysaccharide-Induced Inflammation of THP-1 Cell-Derived Macrophages', *Journal of Medicinal Food*, 22(1), pp. 62–73. doi: 10.1089/jmf.2018.4253.

- Kim, S.-M. *et al.* (2019) 'Retinoic acid-stimulated ERK1/2 pathway regulates meiotic initiation in cultured fetal germ cells', *PLOS ONE*. Edited by S. V. Pizzo. Public Library of Science, 14(11), p. e0224628. doi: 10.1371/journal.pone.0224628.
- Kobayashi, M., Kurihara, K. and Matsuoka, I. (1994) 'Retinoic acid induces BDNF responsiveness of sympathetic neurons by alteration of Trk neurotrophin receptor expression', *FEBS Letters*, 356(1), pp. 60–65. doi: 10.1016/0014-5793(94)01238-5.
- Kong, J.-T. *et al.* (2013) 'Understanding central mechanisms of acupuncture analgesia using dynamic quantitative sensory testing: a review.', *Evidence-based complementary and alternative medicine: eCAM*. Hindawi Limited, 2013, p. 187182. doi: 10.1155/2013/187182.
- Kong, W.-L., Peng, Y.-Y. and Peng, B.-W. (2017) 'Modulation of neuroinflammation: Role and therapeutic potential of TRPV1 in the neuro-immune axis', *Brain, Behavior, and Immunity*. Academic Press, 64, pp. 354–366. doi: 10.1016/J.BBI.2017.03.007.
- Kopincová, J., Púzszerová, A. and Bernátová, I. (2012) 'L-NAME in the cardiovascular system - nitric oxide synthase activator?', *Pharmacological reports: PR*. Pharmacol Rep, 64(3), pp. 511–20. doi: 10.1016/s1734-1140(12)70846-0.
- Korzhevskii, D. E., Karpenko, M. N. and Kirik, O. V. (2012) 'Microtubule-Associated Proteins as Indicators of Differentiation and the Functional State of Nerve Cells', *Neuroscience and Behavioral Physiology*, 42(3), pp. 215–222. doi: 10.1007/s11055-012-9556-4.
- Kovalevich, J. and Langford, D. (2013) 'Considerations for the use of SH-SY5Y neuroblastoma cells in neurobiology.', *Methods in molecular biology (Clifton, N.J.)*. NIH Public Access, 1078, pp. 9–21. doi: 10.1007/978-1-62703-640-5_2.
- Krames, E. S. (2014) 'The Role of the Dorsal Root Ganglion in the Development of Neuropathic Pain', *Pain Medicine*. Narnia, 15(10), pp. 1669–1685. doi: 10.1111/pme.12413.
- Kuhar, M. J., Couceyro, P. R. and Lambert, P. D. (1999) ' α - and β -Adrenergic Receptors'. Lippincott-Raven. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK28138/> (Accessed: 9 January 2020).
- Kumar, A. *et al.* (2014) 'Clonidine for management of chronic pain: A brief review of the current evidences.', *Saudi journal of anaesthesia*. Wolters Kluwer -- Medknow Publications, 8(1), pp. 92–6. doi: 10.4103/1658-354X.125955.
- Kurabe, M., Furue, H. and Kohno, T. (2016) 'Intravenous administration of lidocaine directly acts on spinal dorsal horn and produces analgesic effect: An in vivo patch-clamp analysis.', *Scientific reports*. Nature Publishing Group, 6, p. 26253. doi: 10.1038/srep26253.
- Laumet, G., Chen, S.-R. and Pan, H.-L. (2017) 'NMDA Receptors and Signaling in Chronic Neuropathic Pain', in *The NMDA Receptors*. Cham: Springer International Publishing, pp. 103–119. doi: 10.1007/978-3-319-49795-2_6.

- Leung, L. and Cahill, C. (2010) 'TNF-alpha and neuropathic pain - a review', *Journal of Neuroinflammation*, 7(1), p. 27. doi: 10.1186/1742-2094-7-27.
- Levy, D. and Zochodne, D. W. (2004) 'NO Pain: Potential Roles of Nitric Oxide in Neuropathic Pain', *Pain Practice*, 4(1), pp. 11–18. doi: 10.1111/j.1533-2500.2004.04002.x.
- Li, C. *et al.* (2019) 'Role of Descending Dopaminergic Pathways in Pain Modulation.', *Current neuropharmacology*. Bentham Science Publishers, 17(12), pp. 1176–1182. doi: 10.2174/1570159X17666190430102531.
- Lin, H.-C. *et al.* (2014) '5-Lipoxygenase Inhibitors Attenuate TNF- α -Induced Inflammation in Human Synovial Fibroblasts', *PLoS ONE*. Edited by C.-M. Yang, 9(9), p. e107890. doi: 10.1371/journal.pone.0107890.
- Liu, R. *et al.* (2017) 'Zerumbone reduced the inflammatory response of acute lung injury in endotoxin-treated mice via Akt-NF κ B pathway Chemico-Biological Interactions Zerumbone reduced the inflammatory response of acute lung injury in endotoxin-treated mice via Akt-NF κ B p', *Chemico-Biological Interactions*. Elsevier Ltd, 271(July 2018), pp. 9–14. doi: 10.1016/j.cbi.2017.04.017.
- Liu, T. *et al.* (2017) 'NF- κ B signaling in inflammation', *Signal Transduction and Targeted Therapy*. Nature Publishing Group, 2(1), p. 17023. doi: 10.1038/sigtrans.2017.23.
- Liu, W. *et al.* (2006) *Adrenergic modulation of NMDA receptors in prefrontal cortex is differentially regulated by RGS proteins and spinophilin*. Available at: www.pnas.org/cgi/doi/10.1073/pnas.0604560103 (Accessed: 9 January 2020).
- Liu, W. Y., Tzeng, T. and Liu, I. (2016) 'Zerumbone, a Bioactive Sesquiterpene, Ameliorates Diabetes-Induced Retinal Microvascular Damage through Inhibition of Phospho-p38 Mitogen-Activated Protein Kinase and Nuclear Factor- κ B Pathways'. doi: 10.3390/molecules21121708.
- Lloyd, D. R. and Murphy, A. Z. (2009) 'The Role of the Periaqueductal Gray in the Modulation of Pain in Males and Females: Are the Anatomy and Physiology Really that Different?', *Neural Plasticity*, 2009, pp. 1–12. doi: 10.1155/2009/462879.
- Luccini, E. *et al.* (2007) 'Functional interactions between presynaptic NMDA receptors and metabotropic glutamate receptors co-expressed on rat and human noradrenergic terminals.', *British journal of pharmacology*. Br J Pharmacol, 151(7), pp. 1087–94. doi: 10.1038/sj.bjp.0707280.
- Lundberg, J. O., Weitzberg, E. and Gladwin, M. T. (2008) 'The nitrate–nitrite–nitric oxide pathway in physiology and therapeutics', *Nature Reviews Drug Discovery*, 7(2), pp. 156–167. doi: 10.1038/nrd2466.
- Malek, N. *et al.* (2015) 'The importance of TRPV1-sensitisation factors for the development of neuropathic pain', *Molecular and Cellular Neuroscience*. Academic Press, 65, pp. 1–10. doi: 10.1016/J.MCN.2015.02.001.

- Marrone, M. C. *et al.* (2017) 'TRPV1 channels are critical brain inflammation detectors and neuropathic pain biomarkers in mice', *Nature Communications*. Nature Publishing Group, 8(1), p. 15292. doi: 10.1038/ncomms15292.
- Matsushita, Y. *et al.* (2018) 'Adrenergic receptors inhibit TRPV1 activity in the dorsal root ganglion neurons of rats', *PLOS ONE*. Edited by M. Minami. Public Library of Science, 13(1), p. e0191032. doi: 10.1371/journal.pone.0191032.
- McEntire, D. M. *et al.* (2016) 'Pain transduction: a pharmacologic perspective.', *Expert review of clinical pharmacology*. NIH Public Access, 9(8), pp. 1069–80. doi: 10.1080/17512433.2016.1183481.
- Meacham, K. *et al.* (2017) 'Neuropathic Pain: Central vs. Peripheral Mechanisms'. doi: 10.1007/s11916-017-0629-5.
- Mengke, N. A. S. *et al.* (2016) 'Rapamycin inhibits lipopolysaccharide-induced neuroinflammation in vitro and in vivo', pp. 4957–4966. doi: 10.3892/mmr.2016.5883.
- Meseguer, V. *et al.* (2014) 'TRPA1 channels mediate acute neurogenic inflammation and pain produced by bacterial endotoxins', *Nature Communications*. Nature Publishing Group, 5(1), p. 3125. doi: 10.1038/ncomms4125.
- Messeguer, A., Planells-Cases, R. and Ferrer-Montiel, A. (2006) 'Physiology and pharmacology of the vanilloid receptor.', *Current neuropharmacology*. Bentham Science Publishers, 4(1), pp. 1–15. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/18615132> (Accessed: 3 February 2019).
- Meyr, A. J. and Steinberg, J. S. (2008) 'The Physiology of the Acute Pain Pathway', *Clinics in Podiatric Medicine and Surgery*, 25(3), pp. 305–326. doi: 10.1016/j.cpm.2008.02.012.
- Mogil, J. S. (2009) 'Animal models of pain: Progress and challenges', *Nature Reviews Neuroscience*, 10(4), pp. 283–294. doi: 10.1038/nrn2606.
- Mohammed Izham, A. *et al.* (2018) 'The Effect of DMEM and DMEM:F12 Culture Media on the Growth of SH-SY5Y Cells', *Life Sciences, Medicine and Biomedicine*, 2(3), pp. 3–6. doi: 10.28916/lsm.2.3.2018.23.
- Murakami, A., Miyamoto, M. and Ohigashi, H. (2004) 'phytochemical , induces expression of proinflammatory cytokine genes in human colon adenocarcinoma cell lines', 21, pp. 95–101.
- Murillo, J. R. *et al.* (2017) 'Quantitative proteomic analysis identifies proteins and pathways related to neuronal development in differentiated SH-SY5Y neuroblastoma cells', *EuPA Open Proteomics*. Elsevier, 16, pp. 1–11. doi: 10.1016/J.EUPROT.2017.06.001.
- Nacalai Product Information* (2021). Available at: <https://www.nacalai.co.jp/ss/ec/EC-srchdetl.cfm?HP=1&l=EN&lc=1&syohin=2354721&syubetsu=3&catalog=&SiireC=&MakerC=&yoro=> (Accessed: 24 February 2021).

- Ng, Y.-W. and Say, Y.-H. (2018) 'Palmitic acid induces neurotoxicity and gliotoxicity in SH-SY5Y human neuroblastoma and T98G human glioblastoma cells', *PeerJ*. doi: 10.7717/peerj.4696.
- Nikulina, E. M. *et al.* (2014) 'Neurotrophins in the ventral tegmental area: Role in social stress, mood disorders and drug abuse', *Neuroscience*. Pergamon, 282, pp. 122–138. doi: 10.1016/J.Neuroscience.2014.05.028.
- Nopparat, C. *et al.* (2017) 'The anti-inflammatory effect of melatonin in SH-SY5Y neuroblastoma cells exposed to sublethal dose of hydrogen peroxide', *Mechanisms of Ageing and Development*. Elsevier, 164(November 2016), pp. 49–60. doi: 10.1016/j.mad.2017.04.001.
- O'Neill, E. *et al.* (2016) 'Amitriptyline protects against TNF- α -induced atrophy and reduction in synaptic markers via a Trk-dependent mechanism.', *Pharmacology research & perspectives*. Wiley-Blackwell, 4(2), p. e00195. doi: 10.1002/prp2.195.
- Obata, H. (2017) 'Analgesic Mechanisms of Antidepressants for Neuropathic Pain.', *International journal of molecular sciences*. Multidisciplinary Digital Publishing Institute (MDPI), 18(11). doi: 10.3390/ijms18112483.
- Obuchowicz, E. *et al.* (2006) 'Amitriptyline and nortriptyline inhibit interleukin-1 release by rat mixed glial and microglial cell cultures.', *The international journal of neuropsychopharmacology*. Int J Neuropsychopharmacol, 9(1), pp. 27–35. doi: 10.1017/S146114570500547X.
- Ogawa, N. *et al.* (2014) 'Gene Therapy for Neuropathic Pain by Silencing of TNF- α Expression with Lentiviral Vectors Targeting the Dorsal Root Ganglion in Mice', *PLoS ONE*. Edited by S. Di Giovanni. Public Library of Science, 9(3), p. e92073. doi: 10.1371/journal.pone.0092073.
- Olmos, G. and Lladó, J. (2014) 'Tumor Necrosis Factor Alpha: A Link between Neuroinflammation and Excitotoxicity', 2014.
- Pandur, E. *et al.* (2018) 'Effect of Inflammatory Mediators Lipopolysaccharide and Lipoteichoic Acid on Iron Metabolism of Differentiated SH-SY5Y Cells Alters in the Presence of BV-2 Microglia.', *International journal of molecular sciences*. Multidisciplinary Digital Publishing Institute (MDPI), 20(1). doi: 10.3390/ijms20010017.
- Park, C.-K. *et al.* (2011) 'Resolving TRPV1- and TNF- α -mediated spinal cord synaptic plasticity and inflammatory pain with neuroprotectin D1.', *The Journal of neuroscience: the official journal of the Society for Neuroscience*. Society for Neuroscience, 31(42), pp. 15072–85. doi: 10.1523/JNEUROSCI.2443-11.2011.
- Peppin, J. F. and Pappagallo, M. (2014) 'Capsaicinoids in the treatment of neuropathic pain: a review.', *Therapeutic advances in neurological disorders*. SAGE Publications, 7(1), pp. 22–32. doi: 10.1177/1756285613501576.
- Perimal, E. K. *et al.* (2011a) 'Zerubone-Induced Antinociception: Involvement of the l-Arginine-Nitric Oxide-cGMP -PKC-K+ATP Channel Pathways', *Basic & Clinical Pharmacology & Toxicology*. John Wiley & Sons, Ltd, 108(3), pp. 155–162. doi: 10.1111/j.1742-7843.2010.00635.x.

- Perimal, E. K. *et al.* (2011b) ‘Zerumbone-Induced Antinociception: Involvement of the l-Arginine-Nitric Oxide-cGMP -PKC-K+ATP Channel Pathways’, *Basic and Clinical Pharmacology and Toxicology*, 108(3), pp. 155–162. doi: 10.1111/j.1742-7843.2010.00635.x.
- Perimal, E. K. *et al.* (2011c) ‘Zerumbone-Induced Antinociception: Involvement of the l-Arginine-Nitric Oxide-cGMP -PKC-K+ATP Channel Pathways’, *Basic and Clinical Pharmacology and Toxicology*, 108(3), pp. 155–162. doi: 10.1111/j.1742-7843.2010.00635.x.
- Perimal, E. K. *et al.* (2011d) ‘Zerumbone-Induced Antinociception: Involvement of the l-Arginine-Nitric Oxide-cGMP -PKC-K+ATP Channel Pathways’, *Basic and Clinical Pharmacology and Toxicology*, 108(3), pp. 155–162. doi: 10.1111/j.1742-7843.2010.00635.x.
- Pertovaara, A. (2013) ‘The noradrenergic pain regulation system: A potential target for pain therapy’, *European Journal of Pharmacology*, 716(1–3), pp. 2–7. doi: 10.1016/j.ejphar.2013.01.067.
- Pfeiffer, S. *et al.* (1996) *Inhibition of nitric oxide synthesis by NG^{*}nitro-L-arginine methyl ester (L-NAME): requirement for bioactivation to the free acid, NG-nitro-L-arginine*. Stockton Press All rights reserved. Available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1909689/pdf/brjpharm00085-0114.pdf> (Accessed: 26 October 2020).
- Podratz, J. L. *et al.* (2016) ‘Neurotoxicity to DRG neurons varies between rodent strains treated with cisplatin and bortezomib.’, *Journal of the neurological sciences*. NIH Public Access, 362, pp. 131–5. doi: 10.1016/j.jns.2015.12.038.
- Popiolek-Barczyk, K. *et al.* (2017) ‘Biphalin, a Dimeric Enkephalin, Alleviates LPS-Induced Activation in Rat Primary Microglial Cultures in Opioid Receptor-Dependent and Receptor-Independent Manners’, *Neural Plasticity*. Hindawi, 2017. doi: 10.1155/2017/3829472.
- Pousinha, P. A. *et al.* (2017) ‘Physiological and pathophysiological control of synaptic GluN2B-NMDA receptors by the C-terminal domain of amyloid precursor protein.’, *eLife*. eLife Sciences Publications, Ltd, 6. doi: 10.7554/eLife.25659.
- Pribrag, H. and Stellwagen, D. (2013) ‘TNF- α Downregulates Inhibitory Neurotransmission through Protein Phosphatase 1-Dependent Trafficking of GABA_A Receptors’, 33(40), pp. 15879–15893. doi: 10.1523/JNEUROSCI.0530-13.2013.
- Puopolo, M. *et al.* (2013) ‘Permeation and block of TRPV1 channels by the cationic lidocaine derivative QX-314’, *Journal of Neurophysiology*. American Physiological Society Bethesda, MD, 109(7), pp. 1704–1712. doi: 10.1152/jn.00012.2013.
- Puopolo, M. (2019) ‘The hypothalamic-spinal dopaminergic system: a target for pain modulation.’, *Neural regeneration research*. Wolters Kluwer -- Medknow Publications, 14(6), pp. 925–930. doi: 10.4103/1673-5374.250567.
- Purves, D. *et al.* (2001) ‘Glutamate Receptors’. Sinauer Associates. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK10802/> (Accessed: 9 January 2020).

- Qiao, J. *et al.* (2012) 'PI3K/AKT and ERK regulate retinoic acid-induced neuroblastoma cellular differentiation.', *Biochemical and biophysical research communications*. NIH Public Access, 424(3), pp. 421–6. doi: 10.1016/j.bbrc.2012.06.125.
- Qu, X.-X. *et al.* (2009) 'Role of the spinal cord NR2B-containing NMDA receptors in the development of neuropathic pain', *Experimental Neurology*. Academic Press, 215(2), pp. 298–307. doi: 10.1016/J.EXPNEUROL.2008.10.018.
- Quintans, J. S. S. *et al.* (2014) 'Natural Products Evaluated in Neuropathic Pain Models - A Systematic Review', *Basic & Clinical Pharmacology & Toxicology*. John Wiley & Sons, Ltd, 114(6), pp. 442–450. doi: 10.1111/bcpt.12178.
- Rahman, H. S. *et al.* (2014) 'Zerumbone Induces G2/M Cell Cycle Arrest and Apoptosis via Mitochondrial Pathway in Jurkat cell Line', *Natural Product Communications*. SAGE PublicationsSage CA: Los Angeles, CA, 9(9), p. 1934578X1400900. doi: 10.1177/1934578X1400900904.
- Raju, K. *et al.* (2015) 'Regulation of brain glutamate metabolism by nitric oxide and S-nitrosylation.', *Science signaling*. NIH Public Access, 8(384), p. ra68. doi: 10.1126/scisignal.aaa4312.
- Rampogu, S. and Rampogu Lemuel, M. (2016) 'Network Based Approach in the Establishment of the Relationship between Type 2 Diabetes Mellitus and Its Complications at the Molecular Level Coupled with Molecular Docking Mechanism', *BioMed Research International*, 2016, pp. 1–6. doi: 10.1155/2016/6068437.
- Riediger, C. *et al.* (2017) 'Adverse Effects of Antidepressants for Chronic Pain: A Systematic Review and Meta-analysis', *Frontiers in Neurology*. Frontiers, 8, p. 307. doi: 10.3389/fneur.2017.00307.
- Rohm, B. *et al.* (2018) 'Nonivamide , a capsaicin analog , increases dopamine and serotonin release in SH-SY5Y cells via a TRPV1-independent pathway', pp. 2008–2018. doi: 10.1002/mnfr.201200846.
- Rosenbaum, T. and Simon, S. A. (2007) *TRPV1 Receptors and Signal Transduction, TRP Ion Channel Function in Sensory Transduction and Cellular Signaling Cascades*. CRC Press/Taylor & Francis. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/21204507> (Accessed: 9 January 2020).
- Ruan, Y. *et al.* (2020) 'The Role of Adrenoceptors in the Retina', *Cells*. Multidisciplinary Digital Publishing Institute, 9(12), p. 2594. doi: 10.3390/cells9122594.
- Sakagami, H. *et al.* (2017) 'Re-evaluation of Culture Condition of PC12 and SH-SY5Y Cells Based on Growth Rate and Amino Acid Consumption', 1095, pp. 1089–1095. doi: 10.21873/invivo.11174.
- Samanta, A. *et al.* (2018) 'Transient Receptor Potential (TRP) Channels HHS Public Access Author manuscript', *Subcell Biochem*, 87, pp. 141–165. doi: 10.1007/978-981-10-7757-9_6.

- Sambasevam, Y. *et al.* (2017) 'Cardamonin attenuates hyperalgesia and allodynia in a mouse model of chronic constriction injury-induced neuropathic pain: Possible involvement of the opioid system', *European Journal of Pharmacology*. Elsevier, 796, pp. 32–38. doi: 10.1016/j.ejphar.2016.12.020.
- Sanchez-Perez, A. *et al.* (2005) 'Modulation of NMDA receptors in the cerebellum. II. Signaling pathways and physiological modulators regulating NMDA receptor function', *The Cerebellum*. Springer, 4(3), pp. 162–170. doi: 10.1080/14734220510008003.
- Sango, K. *et al.* (2011) 'Immortalized adult rodent Schwann cells as in vitro models to study diabetic neuropathy.', *Experimental diabetes research*. Hindawi Limited, 2011, p. 374943. doi: 10.1155/2011/374943.
- Sansone, R. A. and Sansone, L. A. (2008) 'Pain, pain, go away: antidepressants and pain management.', *Psychiatry (Edgmont (Pa.: Township))*. Matrix Medical Communications, 5(12), pp. 16–9. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/19724772> (Accessed: 12 October 2021).
- Schäfers, M. *et al.* (2003) 'Increased sensitivity of injured and adjacent uninjured rat primary sensory neurons to exogenous tumor necrosis factor-alpha after spinal nerve ligation.', *The Journal of neuroscience : the official journal of the Society for Neuroscience*. J Neurosci, 23(7), pp. 3028–38. doi: 10.1523/JNeurosci.23-07-03028.2003.
- Scheinin, M. *et al.* (1994) 'Distribution of alpha 2-adrenergic receptor subtype gene expression in rat brain.', *Brain research. Molecular brain research*. Brain Res Mol Brain Res, 21(1–2), pp. 133–49. doi: 10.1016/0169-328x(94)90386-7.
- Schmidt-Nielsen, K. (1997) *Animal Physiology: Adaptation and Environment - Knut Schmidt-Nielsen - Google Books*. Fifth. New York: Cambridge University Press. Available at: https://books.google.com.my/books?hl=en&lr=&id=Af7IwQWJoCMC&oi=fnd&pg=PP9&ots=T0j_Xs-rHf&sig=E80dL4ZZKc0lSkMIJ7SEsgcooqc&redir_esc=y#v=onepage&q&f=false.
- Schug, S. A. and Stannard, K. J. (2011) *Treatment of Neuropathic Pain, Mechanisms of Vascular Disease: A Reference Book for Vascular Specialists*. University of Adelaide Press. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/30485013>.
- Shen, H. *et al.* (2019) 'An integrated cell isolation and purification method for rat dorsal root ganglion neurons.', *The Journal of international medical research*. SAGE Publications, 47(7), pp. 3253–3260. doi: 10.1177/0300060519855585.
- Shiple, M. M. *et al.* (2017) 'Differentiated Human SH-SY5Y Cells Provide a Reductionist Model of Herpes Simplex Virus 1 Neurotropism.', *Journal of virology*. American Society for Microbiology (ASM), 91(23). doi: 10.1128/JVI.00958-17.
- Shiple, M. M., Mangold, C. A. and Szpara, M. L. (2017) 'HHS Public Access', (108). doi: 10.3791/53193.Differentiation.

- Shubayev, V. I. and Myers, R. R. (2002) 'Anterograde TNF alpha transport from rat dorsal root ganglion to spinal cord and injured sciatic nerve.', *Neuroscience letters*. *Neurosci Lett*, 320(1–2), pp. 99–101. doi: 10.1016/s0304-3940(02)00010-1.
- Siddall, P. J. *et al.* (2000) 'The efficacy of intrathecal morphine and clonidine in the treatment of pain after spinal cord injury.', *Anesthesia and analgesia*. *Anesth Analg*, 91(6), pp. 1493–8. doi: 10.1097/00000539-200012000-00037.
- Sikand, P. and Premkumar, L. S. (2007) 'Potentiation of glutamatergic synaptic transmission by protein kinase C-mediated sensitization of TRPV1 at the first sensory synapse.', *The Journal of physiology*. *J Physiol*, 581(Pt 2), pp. 631–47. doi: 10.1113/jphysiol.2006.118620.
- Silva, M. *et al.* (2016) 'Endovanilloid control of pain modulation by the rostroventromedial medulla in an animal model of diabetic neuropathy.', *Neuropharmacology*. *Neuropharmacology*, 107, pp. 49–57. doi: 10.1016/j.neuropharm.2016.03.007.
- Skaper, S. (2008) 'The Biology of Neurotrophins, Signalling Pathways, and Functional Peptide Mimetics of Neurotrophins and their Receptors', *CNS & Neurological Disorders - Drug Targets*, 7(1), pp. 46–62. doi: 10.2174/187152708783885174.
- Smerker, H., Advisor, F. and Ewing, S. (2014) 'Weaning and Adaptation of SH-SY5Y Cells to Low Glucose Media for Manganese Exposure Studies', pp. 7–9.
- Sousa, A. M. *et al.* (2016) 'Experimental models for the study of neuropathic pain', *Revista Dor*. *Sociedade Brasileira para o Estudo da Dor*, 17, pp. 27–30. doi: 10.5935/1806-0013.20160043.
- Srinivasan, A. and Toh, Y.-C. (2019) 'Human Pluripotent Stem Cell-Derived Neural Crest Cells for Tissue Regeneration and Disease Modeling', *Frontiers in Molecular Neuroscience*. *Frontiers*, 12, p. 39. doi: 10.3389/fnmol.2019.00039.
- Stacey, G. (2006) 'Primary Cell Cultures and Immortal Cell Lines', in *Encyclopedia of Life Sciences*. Chichester, UK: John Wiley & Sons, Ltd. doi: 10.1038/npg.els.0003960.
- Stepanenko, Y. D. *et al.* (2019) 'Dual action of amitriptyline on NMDA receptors: enhancement of Ca-dependent desensitization and trapping channel block', *Scientific Reports*. *Nature Publishing Group*, 9(1), p. 19454. doi: 10.1038/s41598-019-56072-z.
- Stevison, F. *et al.* (2017) 'Inhibition of the *all-trans* Retinoic Acid (*at* RA) Hydroxylases CYP26A1 and CYP26B1 Results in Dynamic, Tissue-Specific Changes in Endogenous *at* RA Signaling', *Drug Metabolism and Disposition*, 45(7), pp. 846–854. doi: 10.1124/dmd.117.075341.
- Storozhuk, M. V., Moroz, O. F. and Zholos, A. V. (2019) 'Multifunctional TRPV1 Ion Channels in Physiology and Pathology with Focus on the Brain, Vasculature, and Some Visceral Systems', *BioMed Research International*. *Hindawi*, 2019, pp. 1–12. doi: 10.1155/2019/5806321.

- Su, W.-F. *et al.* (2019) 'Overexpression of P2X4 receptor in Schwann cells promotes motor and sensory functional recovery and remyelination via BDNF secretion after nerve injury', *Glia*. John Wiley & Sons, Ltd, 67(1), pp. 78–90. doi: 10.1002/glia.23527.
- Sudo, R. T. *et al.* (2017a) 'Antinociception induced by a novel α 2A adrenergic receptor agonist in rodents acute and chronic pain models', *European Journal of Pharmacology*. Elsevier, 815, pp. 210–218. doi: 10.1016/J.EJPHAR.2017.09.018.
- Sudo, R. T. *et al.* (2017b) 'Antinociception induced by a novel α 2A adrenergic receptor agonist in rodents acute and chronic pain models', *European Journal of Pharmacology*. Elsevier, 815, pp. 210–218. doi: 10.1016/J.EJPHAR.2017.09.018.
- Sulaiman, M. R. *et al.* (2009) 'Preliminary analysis of the antinociceptive activity of zerumbone', *Fitoterapia*, 80(4), pp. 230–232. doi: 10.1016/j.fitote.2009.02.002.
- Sun, W.-H., Su, Y.-S. and Chen, C.-C. (2019) 'The Transition from Acute to Chronic Pain of a single chapter of a title in Oxford Handbooks Online for personal use (for details see Privacy Policy and Legal Notice Subject: Neuroscience, Sensory and Motor Systems Online Publication The Transition from Acute to Chronic Pain The Oxford Handbook of the Neurobiology of Pain'. doi: 10.1093/oxfordhb/9780190860509.013.28.
- Tanabe, K. *et al.* (2010) 'Mechanisms of tumor necrosis factor- α -induced interleukin-6 synthesis in glioma cells', *Journal of Neuroinflammation*. BioMed Central, 7(1), p. 16. doi: 10.1186/1742-2094-7-16.
- Tauben, D. and Stacey, B. R. (2021) *Pharmacologic management of chronic non-cancer pain in adults - UpToDate*. Available at: <https://www.uptodate.com/contents/pharmacologic-management-of-chronic-non-cancer-pain-in-adults> (Accessed: 13 October 2021).
- Teppola, H., Tuula, J. S. and Linne, O. J. M. (2016) 'Morphological Differentiation Towards Neuronal Phenotype of SH-SY5Y Neuroblastoma Cells by Estradiol, Retinoic Acid and Cholesterol', *Neurochemical Research*. Springer US, 41(4), pp. 731–747. doi: 10.1007/s11064-015-1743-6.
- Thomson, A. C. *et al.* (2021) 'The Effects of Serum Removal on Gene Expression and Morphological Plasticity Markers in Differentiated SH-SY5Y Cells', *Cellular and Molecular Neurobiology*. Springer, pp. 1–11. doi: 10.1007/s10571-021-01062-x.
- Torp, K. D., Metheny, E. and Simon, L. V. (2021) *Lidocaine Toxicity*, *StatPearls*. StatPearls Publishing. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/29494086> (Accessed: 13 October 2021).
- Tremblay, R. G. *et al.* (2010) 'Differentiation of mouse Neuro 2A cells into dopamine neurons', *Journal of Neuroscience Methods*. Elsevier, 186(1), pp. 60–67. doi: 10.1016/J.JNEUMETH.2009.11.004.

- Tsarouchas, T. M. *et al.* (2018) 'Dynamic control of proinflammatory cytokines IL-1 β and TNF- α by macrophages in zebrafish spinal cord regeneration', *Nature Communications*. Nature Publishing Group, 9(1), p. 4670. doi: 10.1038/s41467-018-07036-w.
- Tsukahara, R. and Ueda, H. (2016) 'Myelin-related gene silencing mediated by LPA1 – Rho/ROCK signaling is correlated to acetylation of NF κ B in S16 Schwann cells', *Journal of Pharmacological Sciences*. Elsevier, 132(2), pp. 162–165. doi: 10.1016/J.JPHS.2016.07.010.
- Uys, M. M., Shahid, M. and Harvey, B. H. (2017) 'Therapeutic Potential of Selectively Targeting the α 2C-Adrenoceptor in Cognition, Depression, and Schizophrenia—New Developments and Future Perspective', *Frontiers in Psychiatry*. Frontiers, 8, p. 144. doi: 10.3389/fpsy.2017.00144.
- Vardeh, D. and Naranjo, J. F. (2017) 'Peripheral and Central Sensitization', in *Pain Medicine*. Cham: Springer International Publishing, pp. 15–17. doi: 10.1007/978-3-319-43133-8_4.
- Vincent, A. M., Brownlee, M. and Russell, J. W. (2002) 'Oxidative Stress and Programmed Cell Death in Diabetic Neuropathy', *Annals of the New York Academy of Sciences*. John Wiley & Sons, Ltd (10.1111), 959(1), pp. 368–383. doi: 10.1111/j.1749-6632.2002.tb02108.x.
- Waataja, J. J. *et al.* (2019) 'Agmatine preferentially antagonizes GluN2B-containing N-methyl- d -aspartate receptors in spinal cord', *Journal of Neurophysiology*. American Physiological Society Bethesda, MD , 121(2), pp. 662–671. doi: 10.1152/jn.00172.2018.
- Wang, B. *et al.* (2017) 'Effects of α 2A Adrenoceptors on Norepinephrine Secretion from the Locus Coeruleus during Chronic Stress-Induced Depression', *Frontiers in Neuroscience*. Frontiers, 11, p. 243. doi: 10.3389/fnins.2017.00243.
- Wang, C.-C. *et al.* (2011) 'Article A Critical Role for GluN2B-Containing NMDA Receptors in Cortical Development and Function', *Neuron*, 72, pp. 789–805. doi: 10.1016/j.neuron.2011.09.023.
- Wang, L. X. and Wang, Z. J. (2003) 'Animal and cellular models of chronic pain', *Advanced Drug Delivery Reviews*, 55(8), pp. 949–965. doi: 10.1016/S0169-409X(03)00098-X.
- Wang, Y. *et al.* (2018) 'Sensitization of TRPV1 receptors by TNF- α orchestrates the development of vincristine-induced pain.', *Oncology letters*. Spandidos Publications, 15(4), pp. 5013–5019. doi: 10.3892/ol.2018.7986.
- Wei, Z. *et al.* (2019) 'Emerging Role of Schwann Cells in Neuropathic Pain: Receptors, Glial Mediators and Myelination', *Frontiers in Cellular Neuroscience*. Frontiers, 13, p. 116. doi: 10.3389/fncel.2019.00116.
- Wong-Riley, M. T. T. (2012) 'Bigenomic regulation of cytochrome c oxidase in neurons and the tight coupling between neuronal activity and energy metabolism.', *Advances in experimental medicine and biology*. NIH Public Access, 748, pp. 283–304. doi: 10.1007/978-1-4614-3573-0_12.

- Woo, K. S. *et al.* (1997) 'Chinese adults are less susceptible than whites to age-related endothelial dysfunction.', *Journal of the American College of Cardiology*, 30(1), pp. 113–8. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/9207630> (Accessed: 2 December 2016).
- Wu, F. *et al.* (2014) 'Lentivirus mediated siRNA against GluN2B subunit of NMDA receptor reduces nociception in a rat model of neuropathic pain.', *BioMed research international*. Hindawi, 2014, p. 871637. doi: 10.1155/2014/871637.
- Wu, L.-J. and Zhuo, M. (2009) 'Targeting the NMDA Receptor Subunit NR2B for the Treatment of Neuropathic Pain'. Available at: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5084290/pdf/13311_2011_Article_60400693.pdf.
- Xicoy, H., Wieringa, B. and Martens, G. J. M. (2017) 'The SH-SY5Y cell line in Parkinson's disease research: a systematic review.', *Molecular neurodegeneration*. BioMed Central, 12(1), p. 10. doi: 10.1186/s13024-017-0149-0.
- Yam, M. F. *et al.* (2018) 'General pathways of pain sensation and the major neurotransmitters involved in pain regulation', *International Journal of Molecular Sciences*, 19(8). doi: 10.3390/ijms19082164.
- Yan, H. *et al.* (2017) 'Zerumbone inhibits melanoma cell proliferation and migration by altering mitochondrial functions', *Oncology Letters*, 13(4), pp. 2397–2402. doi: 10.3892/ol.2017.5742.
- Yan, X. *et al.* (2013) 'Endogenous activation of presynaptic NMDA receptors enhances glutamate release from the primary afferents in the spinal dorsal horn in a rat model of neuropathic pain.', *The Journal of physiology*. Wiley-Blackwell, 591(7), pp. 2001–19. doi: 10.1113/jphysiol.2012.250522.
- Yang, M. H. *et al.* (2019) 'Pleiotropic Pharmacological Actions of Capsazepine, a Synthetic Analogue of Capsaicin, against Various Cancers and Inflammatory Diseases.', *Molecules (Basel, Switzerland)*. Multidisciplinary Digital Publishing Institute (MDPI), 24(5). doi: 10.3390/molecules24050995.
- Yao, S. Y. *et al.* (2010) 'In vitro and in vivo induction and activation of nNOS by LPS in oligodendrocytes.', *Journal of neuroimmunology*. J Neuroimmunol, 229(1–2), pp. 146–56. doi: 10.1016/j.jneuroim.2010.07.023.
- Ye, J. *et al.* (2018) 'The publication trend of neuropathic pain in the world and China: a 20–years bibliometric analysis', *The Journal of Headache and Pain*. BioMed Central, 19(1), p. 110. doi: 10.1186/s10194-018-0941-4.
- Yin, K., Baillie, G. J. and Vetter, I. (2016) 'Neuronal cell lines as model dorsal root ganglion neurons: A transcriptomic comparison.', *Molecular pain*. SAGE Publications, 12. doi: 10.1177/1744806916646111.
- Yob, N. J. *et al.* (2011) 'Zingiber zerumbet (L.) Smith: A Review of Its Ethnomedicinal, Chemical, and Pharmacological Uses.', *Evidence-based complementary and alternative medicine: eCAM*. Hindawi, 2011, p. 543216. doi: 10.1155/2011/543216.

- Yu, Y.-M., Han, P.-L. and Lee, J.-K. (2003) 'JNK pathway is required for retinoic acid-induced neurite outgrowth of human neuroblastoma, SH-SY5Y', *NeuroReport*, 14(7), pp. 941–945. doi: 10.1097/01.wnr.0000074341.81633.b8.
- Yuan, S. *et al.* (2014) 'Gp120 in the pathogenesis of human immunodeficiency virus-associated pain', *Ann Neurol*, 75(6), pp. 837–850. doi: 10.1002/ana.24139.Gp120.
- Zghoul, N. *et al.* (2017) 'Prevalence of chronic pain with neuropathic characteristics: a randomized telephone survey among medical center patients in Kuwait.', *Journal of pain research*. Dove Press, 10, pp. 679–687. doi: 10.2147/JPR.S123966.
- Zhang, J.-M. and An, J. (2007) 'Cytokines, inflammation, and pain.', *International anesthesiology clinics*. NIH Public Access, 45(2), pp. 27–37. doi: 10.1097/AIA.0b013e318034194e.
- Zhao, H. *et al.* (2017) 'The role of microglia in the pathobiology of neuropathic pain development: what do we know?', *British Journal of Anaesthesia*. Narnia, 118(4), pp. 504–516. doi: 10.1093/bja/aex006.
- Zhou, X. L. *et al.* (2019) 'ROR2 modulates neuropathic pain via phosphorylation of NMDA receptor subunit GluN2B in rats'. doi: 10.1016/j.bja.2018.08.025.
- Zhu, B. *et al.* (2019) 'Intra-Venous Lidocaine to Relieve Neuropathic Pain: A Systematic Review and Meta-Analysis', *Frontiers in Neurology*. Frontiers, 10, p. 954. doi: 10.3389/fneur.2019.00954.
- Zhuo, M., Wu, G. and Wu, L.-J. (2011) 'Neuronal and microglial mechanisms of neuropathic pain', *Molecular Brain*, 4(1), p. 31. doi: 10.1186/1756-6606-4-31.