



**MOLECULAR RESPONSES OF U87-GLIOBLASTOMA MULTIFORM
CANCER STEM CELLS TO HSV-G47Δ ONCOLYTIC VIRUS IN NORMOXIA
AND HYPOXIA TUMOR MICROENVIRONMENTS**

By

REZA VAZIFEHMAND RODPOSHEI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy**

April 2021

FPSK (p) 2021 42

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

**MOLECULAR RESPONSES OF U87-GLIOBLASTOMA MULTIFORM
CANCER STEM CELLS TO HSV-G47 Δ ONCOLYTIC VIRUS IN NORMOXIA
AND HYPOXIA TUMOR MICROENVIRONMENTS**

By

REZA VAZIFEHMAND RODPOSHTEI

April 2021

Chairman : Professor Zamberi bin Sekawi, PhD
Faculty : Medicine and Health Sciences

Among young adults, Glioblastoma (GBM, WHO IV astrocytoma) is the most common and aggressive form of primary brain tumor and it is highly invasive with the potential to spread to the central nervous system. Despite the current standard therapies, which include surgery, chemotherapy and radiation, it is still considered a deadly disease. The mean survival rate of glioblastoma patients is 12-14 months. The GBM cancer stem cell subpopulation within tumor processes play a critical role in tumor initiation, progression, and local recurrence and which are resistant to standard therapies. Different genetic pathways such as *PI3K/AKT*, *RTK/RAS*, *p53*, *RB*, *PKR*, Apoptosis, telomerase, telomere length alterations, autophagy, mitophagy, angiogenesis and multiple functional of noncoding RNAs including microRNAs and Lung non coding RNAs are involved in GBM progression and its invasion. Resistance to chemotherapy treatment can occur in low tumor oxygenation environment (hypoxia). As a novel therapeutic strategy for GBM treatment, live and engineered oncolytic viruses such as HSV-G47 delta (a 3rd generation of HSV-1 with ICP6⁻, γ 34.5⁻, α 47⁻, lac Z⁺) with limited toxicity are applied, and they can specifically target apoptosis-resistance cancer stem cells without cross-resistance with existing therapies; hence, the normal cells are spared.

Although the HSV-G47 Δ oncolytic virus has been applied to treat different kinds of solid tumors such as glioblastoma, its molecular targets in U87-GBM CSCs were in a Curtain of ambiguity. Therefore, *PI3K/AKT*, *RTK/RAS*, *p53*, *RB*, *PKR*, Apoptosis, telomerase, telomere length alterations, autophagy, mitophagy, angiogenesis, and non-coding RNAs were the main objective pathways that were evaluated by HSV-G47 Δ oncolytic virus in normoxia and hypoxia tumor microenvironments. To achieve this purpose, First, GBM-CSCs neurospheres were isolated in DMEM/F12 serum free media and characterized using

monoclonal antibodies (CD133-PE, CD44-FITC and DAPI staining) by immunocytochemistry method. Flow cytometry was conducted for apoptosis and cell cycle distribution. Cell viability assay and CPE effects were performed using a standard protocol. Different genetic pathways (mentioned above) were evaluated at the level of mRNA expression in normoxia and hypoxia niches exposed to HSV-G47Δ oncolytic virus using a custom RT²Profiler™ PCR Array and Q-PCR methods. In-silico pathway analysis was performed using online bioinformatics tool (GENE-MANIA) to detect physical and genetic interactions between dysregulated genes. Findings showed that GBM-CSCs could be specifically targeted by HSV-G47Δ oncolytic virus in both microenvironments and the cells were arrested at early stage of apoptosis and G0/G1 cell cycle. Furthermore, results indicated that HSV-G47Δ is more effective when the glioblastoma cancer stem cells are in hypoxic condition. Out of 169 evaluated genes of different pathways, 51genes were significantly in dysregulated pattern when GBM-CSCs exposed to HSV-G47Δ. One of the impressive results of the study was the effects of HSV-G47Δ virus on telomere length alterations with had increased under normoxic condition while a significant telomere shortening was observed when the U87-CSCs were exposed to HSV-G47Δ virus in hypoxic.

Data showed that out of forty three miRNAs, eight miRNAs including *miR-7-1*, *miR-let-7b*, *miR-130a*, *miR-137*, *miR-200b*, *miR-221*, *miR-222* and *miR-874* were significantly over expressed in normoxic microenvironment. Expression level of LncRNAs including *LEF1-AS1*, *MALAT1*, *LINC00470*, *TUSC7*, *HOTAIR*, *NEAT1* and *XIST* were significantly down regulated in hypoxic microenvironment and *H19* did not have any dysregulated pattern in this niche. In normoxic condition, *LEF1-AS1*, *MALAT1*, *LINC00470*, *H19*, *HOTAIR*, *NEAT1* and *XIST* were under regulated and *TUSC7* was not targeted by HSV-G47Δ. Furthermore, in hypoxic conditions, *PERK*, *ING-G*, *LC3*, *MFN2*, *PINK-1*, and *PARKIN* were significantly downregulated while *INF-G*, *P62*, *LC3*, and *PARKIN* were in the up-regulated pattern. The findings revealed that high-grade glioblastoma cancer stem cells were potentially controlled by HSV-G47Δ oncolytic virus in both autophagy and mitophagy pathways at hypoxic conditions. Our results also showed the expression of the majority of genes in MDR pathway (Exception *DKC1*down regulated) were significantly up in GBM-CSCs when the cells were inoculated with HSV-G47Δ (MOI=1,14h) in normoxic condition while most genes were down regulated under HSV-G47Δ (MOI=1,14h) in hypoxic condition. Pathway analysis showed genetic and physical interactions among dysregulated genes in both microenvironments in the most biological pathways.

In conclusion, HSV-G47Δ has a therapeutic potential to control crucial mechanisms in GBM-CSCs progression and could be considered as a promising strategy in GBM treatment, especially when the cells have a high grade of tumorigenicity in hypoxic niche.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**TINDAKBALAS MOLEKULAR STEM SEL KANSER U87-GLIOBLASTOMA
PELBAGAI TERHADAP VIRUS ONKOLITIK HSV-G47Δ DI
PERSEKITARAN NORMOKSIA DAN HIPOKSIA**

Oleh

REZA VAZIFEHMAND RODPOSHTEI

April 2021

Pengerusi : Profesor Zamberi bin Sekawi, PhD
Fakulti : Perubatan dan Sains Kesihatan

Di kalangan orang muda, Glioblastoma (GBM, WHO IV astrocytoma) adalah sejenis ketumbuhan otak asas yang paling biasa dan agresif. Ia mempunyai potensi untuk merebak dengan cepat sehingga mencapai sistem saraf pusat. Walaupun pembedahan, kemoterapi dan radiasi digunakan sebagai terapi umum, glioblastoma masih dianggap sebagai penyakit yang boleh membawa maut. Purata Kadar kelangsungan hidup pesakit glioblastoma adalah 12-14 bulan. Subpopulasi sel stem kanser GBM dalam proses ketumbuhan memainkan peranan penting dalam permulaan tumor, perkembangan, dan pengulangan tempatan (local recurrence) dan rawatan standard tidak memberi keputusan yang baik. Laluan genetik yang berbeza seperti *PI3K / AKT, RTK / RAS, p53, RB, PKR*, Apoptosis, telomeras, kepanjangan telomer, autopagi, mitopagi, angiogenesis dan pelbagai fungsi RNA bukan pengekodan termasuk microRNAs dan RNA bukan pengcod paru-paru terlibat dalam perkembangan dan perebakan GBM. Rawatan menggunakan kemoterapi sering tidak berjaya dan ini berlaku di persekitaran oksigenasi ketumbuhan kecil (hipoksia). Sebagai strategi terapi baru untuk rawatan GBM, virus onkolik hidup dan rekayasa seperti HSV-G47 delta (generasi ke-3 HSV-1 dengan ICP6-, γ 34.5-, α47-, lac Z +) yang mempunyai toksik rendah digunakan, dan mereka mensasar sel stem barah ketahanan apoptosis tanpa rintangan silang menggunakan terapi yang sediaada; oleh itu, sel normal tidak terjejas.

Walaupun virus onkolitik HSV-G47Δ telah diterapkan untuk merawat pelbagai jenis ketumbuhan padat seperti glioblastoma, sasaran molekulnya dalam CSC U87-GBM tidak begitu jelas. Oleh itu, *PI3K / AKT, RTK / RAS, p53, RB, PKR*, Apoptosis, telomeras, perubahan dalam kepanjangan telome, autopagi, mitopagi, angiogenesis, dan non-coding RNAs adalah jalan masuk (pathway) yang dinilai oleh virus onkolitik= HSV-G47Δ dalam persekitaran ketumbuhan

mikro normoxia dan hipoksia. Untuk mencapai objektif ini, pertama, neurosfera GBM-CSC diasingkan dalam media bebas serum DMEM / F12 dan dicirikan menggunakan antibodi monoklonal (pewarnaan CD133-PE, CD44-FITC dan DAPI) menggunakan kaedah imunokimia. Sitometri aliran digunakan untuk apoptosis dan taburan kitaran sel. Ujian daya maju sel dan kesan CPE dilakukan menggunakan protokol umum. Jalur genetik yang berbeza (seperti di atas) dinilai pada tahap ekspresi mRNA pada persekitaran normoksia dan hipoksia yang terdedah kepada virus onkolistik HSV-G47 Δ menggunakan kaedah RT²Profiler™ PCR Array dan Q-PCR khusus. Analisis jalur ‘in-silico’ dilakukan menggunakan bioinformatik dalam talian (GENE-MANIA) untuk mengesan interaksi fizikal dan genetik antara gen yang tidak teratur. Hasil kajian menunjukkan bahawa GBM-CSC secara khusus disasarkan oleh virus onkolistik HSV-G47 Δ di lingkungan mikro dan sel-sel ditangkap pada tahap awal apoptosis dan kitaran sel G0 / G1. Selanjutnya, hasil menunjukkan bahawa HSV-G47 Δ lebih berkesan apabila sel stem kanser glioblastoma berada dalam keadaan hipoksia. Daripada 169 gen yang dievaluasi dari jalur yang berbeza, 51 gen secara signifikan berada dalam keadaan tidak teratur ketika GBM-CSC terdedah kepada HSV-G47 Δ . Salah satu hasil kajian yang signifikan adalah kesan virus HSV-G47 Δ terhadap kepanjangan telomer dalam persekitaran normoksik dan pemendekan telomer yang ketara diperhatikan bila U87-CSC terdedah kepada virus HSV-G47 Δ dalam persekitaran hipoksia.

Data menunjukkan daripada 43 (empat puluh tiga) miRNA, lapan miRNA termasuk *miR-7-1*, *miR-let-7b*, *miR-130a*, *miR-137*, *miR-200b*, *miR-221*, *miR-222* dan *miR-874* meningkat secara ketara dalam persekitaran normoksik mikro. Tahap ekspresi LncRNA termasuk *LEF1-AS1*, *MALAT1*, *LINC00470*, *TUSC7*, *HOTAIR*, *NEAT1* dan *XIST* di kawal selia dengan ketara dalam lingkungan mikro hipoksia dan H19 tidak mempunyai corak yang berbeza dalam ceruk ini. Di dalam persekitaran normoksik, *LEF1-AS1*, *MALAT1*, *LINC00470*, *H19*, *HOTAIR*, *NEAT1* dan *XIST* diatur dan *TUSC7* tidak disasarkan oleh HSV-G47 Δ . Selanjutnya, dalam keadaan hipoksia, *PERK*, *ING-G*, *LC3*, *MFN2*, *PINK-1*, dan *PARKIN* disaksikan terteratur dengan ketara sementara *INF-G*, *P62*, *LC3*, dan *PARKIN* berada dalam corak yang dikawal selia. Hasil kajian menyaksikan sel stem barah glioblastoma peringkat tinggi mempunyai potensi dikendalikan oleh virus onkolistik HSV-G47 Δ pada kedua-dua jalur autofagus dan mitofagi dalam persekitaran hipoksia. Hasil kajian kami juga menunjukkan ekspresi majoriti gen dalam jalur MDR meningkat secara signifikan dalam GBM-CSCs bila sel-sel dionokulasi dengan HSV-G47 Δ (MOI = 1,14h) dalam keadaan normoksik sementara kebanyakan gen turun dikawal selia di bawah HSV-G47 Δ (MOI = 1,14j) dalam persekitaran hipoksia. Analisis laluan menunjukkan interaksi genetik dan fizikal antara gen yang tidak terkawal (dysregulated genes) di kedua-dua persekitaran mikro di kebanyakan laluan biologi.

Kesimpulannya, HSV-G47 Δ memiliki potensi terapeutik untuk mengawal mekanisme penting dalam perkembangan GBM-CSC dan ia boleh dianggap sebagai strategi baru yang mempunyai potensi dalam rawatan GBM, terutamanya bila sel-sel memiliki tingkat tumorigenitas yang tinggi dalam ceruk hipoksia.

ACKNOWLEDGEMENTS

First and above all, I praise God, the almighty for providing me this opportunity and granting me the capability to proceed successfully.

I would like to express my sincere gratitude to my supervisor Professor Dr Zamberi Sekawi, for the continuous support of my PhD study and related research, for his patience, motivation, and immense knowledge. His guidance helped me in all the time of research and writing of this thesis and manuscripts. I could not have imagined having a better supervisor and mentor for my PhD study.

Besides my supervisor, I would like to thank the rest of my thesis committee: Dr Zulkefely Othman and Dr Chau De Ming, My wife, Dr Dhuha Saeed Ali and Dr Mehdi Shafa from Lonza, USA for their insightful comments and encouragement, but also for the hard question which incited me to widen my research from various perspectives.

Last but not the least; I would like to thank my family, my parents for supporting me spiritually throughout writing this thesis.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Zamberi bin Sekawi, PhD

Professor

Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Zulkefely bin Othman, PhD

Senior Lecturer

Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Chau De Ming, PhD

Senior Lecturer

Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean

School of Graduate Studies
Universiti Putra Malaysia

Date: 10 November 2022

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Professor Dr. Zamperi bin Sekawi

Signature: _____

Name of Member
of Supervisory
Committee:

Dr. Zulkefely bin Othman

Signature: _____

Name of Member
of Supervisory
Committee:

Dr. Chau De Ming

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xviii
 CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 Glioblastoma Multiform (GBM)	4
2.1.1 Epidemiology of GBM	4
2.1.2 Etiology of GBM	4
2.1.3 Pathogenesis of GBM	5
2.1.4 Molecular Pathogenicity of GBM	5
2.1.5 Current Strategy of GBM Treatment and Resistant	6
2.2 Hypoxia in GBM progression	6
2.3 U87 Cell Line and GBM-CSCs	7
2.3.1 U87 Cell Line (ATCC) Characterization	7
2.3.2 Cancer stem cells in GBM	8
2.4 Non Coding RNAs (ncRNAs) in GBM	9
2.4.1 MicroRNAs	9
2.4.2 Long non coding RNAs	14
2.5 Autophagy and mitophagy in GBM	15
2.6 Angiogenesis in GBM	17
2.7 Telomerase and Telomere Length Alterations in GBM	18
2.8 Multi Drug Resistant-Vaults Components in GBM	19
2.9 Oncolytic viruses and oncovir therapy in GBM	20
2.10 HSV-G47Δ oncolytic virus	21
3 MOLECULAR RESPONSES OF U87-GLIOBLASTOMA MULTIFORM CANCER STEM CELLS TO HSV-G47Δ ONCOLYTIC VIRUS, FOCUSING ON PI3K/AKT, RTK/RAS, p53, RB, PKR AND APOPTOSIS SIGNALING PATHWAYS USING ARRAY-PCR	22
3.1 Introduction	22
3.2 Materials and Methods	23
3.2.1 U87-GBMCSCs Cell and G47Δ Oncolytic Virus	23

3.2.2	Isolation and Identification of GBM-U87CSCs	23
3.2.3	HSV-G47Δ Replication and <i>in vitro</i> Cell Viability Assay	23
3.2.4	Cell Cycle and Apoptosis Detection	23
3.2.5	Apoptosis Evaluation	24
3.2.6	Gene Expression Panel Study- PCR Array	24
3.2.7	PCR Array Protocol	26
3.3	Results	26
3.3.1	Isolation and Identification of CSCs	26
3.3.2	<i>In vitro</i> Cell Viability Assay and G47Δ Replication	28
3.3.3	Apoptosis and Cell Cycle Distribution	29
3.3.4	Gene Expression Profiling	31
3.3.5	<i>In-silico</i> Pathways Analysis	35
3.3.6	Statistical Analyses	37
3.4	Discussion	37
3.5	Conclusion and Future Prospects	39
4	EFFECTS OF HSV-G47Δ ONCOLYTIC VIRUS ON TELOMERASE AND TELOMERE LENGTH ALTERATIONS IN BOTH NORMOXIC AND HYPOXIC MICROENVIRONMENTS IN U87-GLIOBLASTOMA MULTIFORM CANCER STEM CELLS	40
4.1	Introduction	40
4.2	Materials and Methods	42
4.2.1	Cell and virus	42
4.2.2	Isolation and Identification of U87GBM-CSCs	43
4.2.3	G47Δ Replication and <i>in vitro</i> Cell Viability Assay	43
4.2.4	Absolute Telomere Length Determination	43
4.2.5	Gene Expression Study	44
4.2.6	Pathway Analysis	44
4.3	Results	45
4.3.1	HSV-G47Δ Effectively Targets U87-CSCs in both Microenvironment Conditions	45
4.3.2	HSV-G47Δ Effects on Absolute Telomere Length in both Microenvironment Conditions	48
4.3.3	HSV-G47Δ Effects on Telomerase Subunits and Telomere Complex	49
4.3.4	Bioinformatics Gene Interactions Analysis	54
4.3.5	Statistical Analyses	55
4.4	Discussion	55
4.5	Conclusion and Future Prospects	59

5	NON-CODING RNA-BASED TARGETED THERAPY IN U87-GLIOBLASTOMA MULTIFORM CANCER STEM CELLS USING HSV-G47Δ ONCOLYTIC VIRUS	
5.1	Introduction	60
5.2	Materials and methods	62
5.2.1	Isolation and Identification of GBM-U87CSCs in Hypoxic and Normoxic Conditions	62
5.2.2	MicroRNAs Extraction and cDNA Synthesis	63
5.2.3	MicroRNAs Expression Panel Study, miRCURY LNA Custom PCR Array	63
5.2.4	Long non Coding RNAs (lncRNAs) Extraction and cDNA Synthesis	65
5.2.5	LncRNAs Expression Study- qPCR Assay	65
5.2.6	Bioinformatics Interactions Analysis	65
5.3	Results	66
5.3.1	HSV-G47 Δ Oncolytic Virus Effectively Targets miRNAs	66
5.3.2	HSV-G47 Δ oncolytic virus effectively targets lncRNAs in both microenvironments	70
5.3.3	Bioinformatics Analysis of miRNAs-Genes and miRNAs-LncRNAs Interactions	75
5.3.4	Statistical Analyses	76
5.4	Discussion	76
5.5	Conclusion and Future Prospects	78
6	TARGETED THERAPY USING HSV-G47Δ ONCOLYTIC VIRUS TO TREAT GLIOBLASTOMA: EMPHASIS ON AUTOPHAGY AND MITOPHAGY GENE EXPRESSION IN HYPOXIC AND NORMOXIC NICHES	
6.1	Introduction	79
6.2	Material and methods	81
6.2.1	U87-CSCs and HSV-G47 Δ Oncolytic Virus	81
6.2.2	Gene Expression Study	81
6.2.3	<i>In-silico</i> Pathway Analysis	84
6.3	Results	84
6.3.1	HSV-G47 Δ Effectively Targets U87-CSCs	84
6.3.2	Autophagy and Mitophagy Pathways are Selectivity Targeted by HSV-G47 Δ	84
6.3.3	Pathway Analysis	87
6.3.4	Statistical Analyses	89
6.4	Discussion	89
6.5	Conclusion and Future Prospects	90

7	U87-GLIOBLASTOMA MULTIFORM CANCER STEM CELLS MOLECULAR RESPONSES TO HSV-G47Δ ONCOLYTIC VIRUS: EXPERIMENTAL EVIDENCES OF GENE EXPRESSION IN NORMOXIA AND HYPOXIA NICHES	
7.1	Introduction	91
7.2	Materials and Methods	92
7.2.1	U87 GBM-CSCs and HSV-G47Δ Replication Assay	92
7.2.2	Gene expression study	92
7.2.3	Pathway Analysis	95
7.3	Results	95
7.3.1	U87-CSCs molecular response to HSV-G47Δ Oncolytic Virus in Normoxia and Hypoxia Microenvironments, Focusing on Angiogenesis	95
7.3.2	U87-CSCs Molecular Response to HSV-G47Δ Oncolytic Virus in Normoxia and Hypoxia Microenvironments Focusing on Multidrug Resistance Genes	95
7.3.3	U87-CSCs Molecular Response to HSV-G47Δ Oncolytic Virus in Normoxia and Hypoxia Microenvironments Focusing on Epithelial-to-Mesenchymal Transition (EMT)	95
7.3.4	U87-CSCs Molecular Response to HSV-G47Δ Oncolytic Virus in Normoxia and Hypoxia Microenvironments Focusing on Metabolic Genes	96
7.3.5	Genes interaction analysis	99
7.3.6	Statistical Analyses	101
7.4	Discussion	101
7.5	Conclusion and future prospects	102
8	SUMMARY, CONCLUSIONS, AND RECOMMENDATIONS FOR FUTURE RESEARCH	103
REFERENCES		105
APPENDICES		137
BIODATA OF STUDENT		139
LIST OF PUBLICATIONS		140

LIST OF TABLES

Table		Page
2.1	MicroRNAs associated with GBM progression	11
3.1	Genes with altered expression and change in U87-CSCs treated with HSV-G47 Δ oncolytic virus after 6h and 14h infection period respectively	34
4.1	Genes, primers information and altered expression status in hypoxic and normoxic conditions in post infection periodl	50
5.1	miRNAs biological role in glioblastoma pathogenicity, gene targets and expression	67
5.2	LncRNAs biological role in GBM progression, mi-RNAs targets and expression profile	73
5.3	Primers, PCR conditions and fold changes of gene expression profile in U87-CSCs inoculated with HSV-G47 Δ in hypoxic and normoxic	74
6.1	Autophagy and mitophagy genes and qPCR conditions	83
6.2	Gene expression fold changes in both microenvironments. The analysis was performed using THE REST 2009 software. GAPDH served as a housekeeping gene with FC=1.	85
7.1	Genes, Primers and PCR conditions in the current study	93
7.2	Expression pattern of genes involved in angiogenesis, MDR, EMT and metabolism pathways in both normoxia and hypoxia niches	97

LIST OF FIGURES

Figure	Page
2.1 The main signaling pathways affected in high-grade gliomas	6
2.2 Genetic alterations that lead to HIF activation in GBM	7
2.3 A schematic of U87 cell line	8
2.4 GBM cancer stem cells	9
2.5 A schematic of miRNA biogenesis and its function	10
2.6 Well-Characterized lncRNAs in Glioblastoma	14
2.7 A schematic of the gene regulatory network of lncRNAs in GBM	15
2.8 Schematic of autophagosome biogenesis	16
2.9 Schematic diagram of the mitophagy process	17
2.10 Representation of angiogenesis mediators in glioblastoma	18
2.11 A schematic representation of the six subunits of the Shelterin complex on the telomeric region	19
2.12 vRNA and vault particle components	20
2.13 Armed HSV-G47Δ structure	21
3.1 Schematic representation of the Pathways and related experimental genes designed in a 96-well plate of custom PCR-Array using Eppendorf realplex ⁴ real time PCR device	25
3.2 U87-CSCs isolation from U87-GBM cell lines	27
3.3 The q-PCR analysis was performed to confirm U87-CSCs	28
3.4 HSV-G47Δ effectively targets U87-CSCs and killed them in-vitro	29
3.5 Apoptosis distribution and Flowcytometric analysis	30
3.6 Cell cycle distribution of U87-CSCs in mock and HSV-G47Δ treatment	31

3.7	Scatter plot analysis of dysregulated genes in U87-CSCs at 6h (3.7A) and 14h (3.7B) post infection period of HSV- G47 Δ treatment	32
3.8	Dysregulated change in genes	33
3.9	Pathway analysis of genes with altered expression	36
4.1	Schematic illustration of the telomerase structure and shelterin complex	41
4.2	A schematic of loop formation (T-loop and D-loop) in a telomere structure	42
4.3	U87-CSCs isolated from U87 cell lines in a serum free media (DMEM/F12) supplemented with stem cell isolation components	46
4.4	Hif1 α high expression level shows hypoxic induced condition in media	47
4.5	Cell viability assay revealed that HSV-G47 Δ was able to target U87-CSCs in both microenvironment conditions	47
4.6	A schematic diagram shows the effect of HSV-G47 Δ on telomere length in hypoxic and normoxic tumor microenvironment	48
4.7	Regression analysis of absolute telomere length variation under g47 Δ oncolytic virus effects	49
4.8	Relative expression panel of telomerase subunit genes, telomere length control and capping genes in hypoxic tumor microenvironment after 18h post inoculation of HSV- G47 Δ oncolytic virus	52
4.9	Relative expression panel of telomerase subunit genes, telomere length control and capping genes in normoxic tumor microenvironment after 18h post inoculation time of HSV- G47 Δ oncolytic virus	53
4.10	Pathway analysis of telomerase and telomere dependent genes with altered expression	54
4.11	A schematic representation of results that shows the effects of HSV-G47 Δ oncolytic virus on U87-GBMCSCs in both hypoxic and normoxic microenvironments	58

5.1	Biogenesis of microRNAs. They are produced by RNA polymerase II enzyme from nuclear DNA to generate pri-miRNAs.	61
5.2	A schematic presentation of gene expression regulation via LncRNAs	62
5.3	Custom miRNAs, housekeeping genes and miRCURY LNA PCR Array controls in 96 well plate that designed for Eppendorf realplex ⁴ real time PCR device	64
5.4	Scatter plot analysis (SPA) of the relative expression comparison of 43 miRNAs between mock and treated groups in normoxic condition	69
5.5	A diagram of dysregulated relative fold change miRNAs	70
5.6	Relative expression panel of LncRNAs inoculated with HSV-G47Δ	71
5.7	Relative expression panel of LncRNAs inoculated with HSV-G47Δ	72
5.8	<i>In-silico</i> data analysis of dysregulated miRNAs-Genes interactions network	75
5.9	Dysregulated miRNAs- LncRNAs interaction network showing miRNAs target LncRNAs	76
6.1	A schematic illustration of autophagy pathway	80
6.2	A schematic diagram of the mitophagy process	80
6.3	A schematic diagram illustration of autophagy and mitophagy relative gene expression panel in U87CSCs infected with HSV-G47Δ oncolytic virus with MOI=1 during 14hrs in post infection period in both normoxic and hypoxic microenvironments	86
6.4	Pathway analysis of dysregulated genes presented graphically	88
7.1	Relative expression panel of genes involved in angiogenesis, MDR, EMT and metabolism pathways in both normoxia (A) and hypoxia (B) tumor microenvironments	98
7.2	Physical and genetic interactions among dysregulated genes in both normoxia and hypoxia microenvironments	100

LIST OF ABBREVIATIONS

ATCC	American type culture collection
BBB	Blood brain barrier
cDNA	Complementary DNA
CSCs	Cancer stem cells
CPE	Cytopathic effects
D-Loop	Displacement loop
GBM	Glioblastoma Multiform
HGG	High grade glioblastoma
HIF	Hypoxia inducible factor
HSV	Herpes simplex virus
kb	Kilo bases
LGG	Low grade glioblastoma
lncRNAs	Long non coding RNAs
MDR	Multi drug resistance
MIP	Microscopic image processing
miRNAs	MicroRNAs
MOI	Multiplicity of infection
mRNA	Messenger ribonucleic acid
nc-RNAs	Non-coding RNAs
OVs	Oncolytic viruses
PCR	Polymerase chain reaction
SPA	Scatter plot analysis
T-Loop	Telomere loop
TMZ	Temozolomide
WHO	World health organization

CHAPTER 1

INTRODUCTION

In young adults, Glioblastoma (GBM, WHO IV astrocytoma) is the most common and active form of primary brain tumor which grows rapidly targeting the central nervous system (Hanif, Muzaffar, Perveen, Malhi, & Simjee, 2017). It is also highly invasive and heterogeneous influencing therapeutic responses (Neilsen et al., 2019). Exhaustive genetic analysis has shown a variety of deregulated pathways involved in GBM pathogenicity (Martínez, 2012). With the current standard therapies that include surgery, chemotherapy and radiation, the mean survival rate of glioblastoma patients is 12-14 months. (Ohka, Natsume, & Wakabayashi, 2012a). Despite treatment, their prognosis is very poor. The GBM cancer stem cells subpopulation within the tumor processes have the ability for infinite and multi potency thus it is assumed they play a critical role in the tumor initiation, progression, local recurrence of GBM cancer while being resistant to standard therapies (Codrici, Enciu, Popescu, Mihai, & Tanase, 2016). Hypoxia known as low oxygenation (Anoxic) of solid tumors (PO₂ less than 2.5mmHG) is a major concern in glioblastoma cancer treatment since it promotes chemo-radiotherapy resistance (Vaupel & Mayer, 2007).

Telomeres and telomerase play a critical role in abnormal proliferations, metastasis and stemness maintenance (J. Lee et al., 2006). Evidences showed telomerase are expressed in GBM-CSCs and the telomere length are around 3.5kb - three times shorter than the normal human brain cells (Diane E. Handy, Rita Castro, 2011). Telomeres are located at both ends of a chromosome with a nucleoprotein structures composed of a non-coding, repetitive DNA sequence, and they have six kinds of proteins called shelterin complex (Blasco, 2007)(Blasco, 2007) These proteins control the length of the telomere and protect them from degradation(Blackburn, 2010).With the help of the shelterin complex overhanging the single stranded DNA at the very end of all telomeres, two structures known as T-loop (protective cap) and D-loop (displacement loop) are formed (Diotti & Loayza, 2011). De novo repetitive telomeric sequence add to telomeric ends via telomerase enzyme which is a unique transcriptase enzyme (Greider, 2010). Telomerase maintains the length of telomeres (Greider, 2010). It is well-known that different levels of hypoxia regulate telomere length and telomerase activity (Guan, Wei-Ping, Maeda, & Makino, 2011). A study has shown that telomerase activity and short telomere length in glio-astrocytoma tumors are significantly associated with histological potential progression and tumorigenicity (Hiraga et al., 1998a). Despite the existence of many therapies to fight glioblastoma multiform, it is still a deadly disease which has an extremely poor diagnosis (Hanif et al., 2017). Studies have revealed that telomeres and telomerase activity could be an attractive target for cancer therapy (Picariello, 2014).

A large number of studies have pointed to two groups of small non-coding RNAs (ncRNAs) including microRNAs (17-22 nucleotides) and long non-coding RNAs (lncRNAs) >200 nucleotides, deep involvement in glioblastoma global biological process, such as pathogenesis, tumor initiation, progression, proliferation, invasion, angiogenesis, apoptosis, and resistance to chemo-radiotherapy (Rynkeviciene et al., 2019; Stahlhut & Slack, 2013). MicroRNAs(miRNAs) are short, single stranded RNA that post-transcriptionally regulate gene expression (Y. S. Lee & Dutta, 2009; Londina et al., 2015). miRNAs produced in the nucleus are pri-miRNA and they are processed into pre-miRNA that is exported to the cytoplasm by exportin-5 and converted to a mature miRNA by Dicer complex and exert their action by targeting 3' untranslated region (3'-UTR) of mRNA resulting in inhibition of protein synthesis (Bentwich et al., 2005; Berezikov et al., 2005). Figure 1 shows the biogenesis and function process of miRNAs. lncRNAs as non-coding protein exert their biological function at transcriptional, post-transcriptional and epigenetic levels (Mercer, Dinger, & Mattick, 2009). They are also involved in gene expression regulation via multiple mechanisms, such as chromatin modifications, alternative splicing, mRNA stability and interaction with miRNAs (Fernandes, Acuña, Aoki, Floeter-Winter, & Muxel, 2019). Dysregulation of specific lncRNAs play a crucial role in glioblastoma progression and malignancy (X. Zhang et al., 2012). Non-coding RNAs can potentially be applied as therapeutic targets for GBM treatment (Dews et al., 2006; X. Zhang et al., 2012; Qi & Du, 2013).

Under various stimulation factors, an autophagy pathway can be activated as a cytoprotective response to help cells overcome those stressful situations (Singh & Cuervo, 2011). It has been shown that glioblastoma promotes the survival and proliferation of tumor cells (Piao et al., 2012). Autophagy is a lysosome-dependent degradation pathway whereby the cells recycle their damaged cytoplasmic content, such as lipids, proteins and organelles (Vessoni, Muotri, & Okamoto, 2012). There is ample evidence that point to essential role of autophagy in the maintenance and self-renewal of glioblastoma cancer stem cells (Galavotti et al., 2013). Mitophagy on the other hand is a selective autophagy process that promotes cancer cell survival, tumorigenesis and chemo resistance phenotype by removing abnormal mitochondria, and reduction of overall mitochondrial mass in response to certain stress factors such as hypoxia (Chourasia, Boland, & Macleod, 2015; Vara-Perez, Felipe-Abrio, & Agostinis, 2019). Microvascular proliferation, Multi-drug resistance, epithelial-to-mesenchymal transition (EMT), tumor metabolic reprogramming are various mechanisms by which GBM tumor stem cells avoid therapy (Iser, Pereira, Lenz, & Wink, 2017; Pengse Po1, Erin Delaney1, Howard Gamper2, Miklos Szanti-Kis3, Lee Speight3, LiWei Tu1, Andrey Kosolapov1, E. James Petersson3, Ya-Ming Hou2, 2017; Schmalz, Shen, & Park, 2011; W. Zhou & Wahl, 2019).

Despite promising therapies in GBM over the past three decades, there is a risk of relapse phenotype recurrence. Therefore, more effective therapeutic approaches to treat GBM is of critical importance. Among GBM treatment options, oncolytic virotherapy (OV) is promising (Wollmann, Ozduman, & Van Den Pol, 2012a).

Oncolytic viruses (OVs) are either genetically engineered or naturally occurring that selectively replicate in the cancer cells and kill them without harming normal tissues (Fukuhara, Ino, & Todo, 2016). HSV-G47Δ is an engineered oncolytic virus (3rd generation of HSV-1) with three different mutations including ICP6-, γ34.5-,α47- and lac Z+, that have been recently used to treat various tumors, particular brain tumors (Eissa et al., 2018).

Logical interaction between two or more genes can affects the phenotype of cells that can identified using various types of bioinformatics tools. The ultimate goals of gene-gene interactions are to recognize gene functions, identify pathways and discover potential drug targets(Koo, Liew, Mohamad, Hakim, & Salleh, 2013). GeneMANIA is a flexible, user-friendly web interface for generating hypotheses about gene function, analyzing gene lists and prioritizing genes for functional assays(Warde-farley, Donaldson, Comes, Zuberi, Badrawi, et al., 2010).The mechanism of molecular targeting of HSV-G47Δ is still in a grey zone and therefore, this study was undertaken to evaluate the expression pattern of genes involved in GBM progression pathways. Furthermore, Gene-gene interactions will be evaluated using GENEMANIA bioinformatics tool.

The objectives of the current research study were as follows:

1. To detect molecular responses of U87-GBM-CSCs to HSV-G47Δ oncolytic virus focusing on apoptosis, cell cycle distribution and PI3K /AKT, RTK/RAS, p53, RB, PKR signaling pathways using Flowcytometry and Array-PCR method.
2. To detect effects of HSV-G47Δ oncolytic virus on telomerase and telomere length alterations using Q-PCR method.
3. To detect noncoding RNA responses to HSV-G47Δ on GBM-CSCs in normoxia and hypoxia tumor environments using QPCR method.
4. To detect HSV-G47Δ effects on GBM-CSCs in autophagy and mitophagy pathways in normoxia and hypoxia microenvironments using Q-PCR method.
5. To detect molecular responses of GBM-CSCs to HSV-G47Δ oncolytic virus in normoxia and hypoxia tumor microenvironments, focusing on angiogenesis, multiple drug resistance, Epithelial-to-mesenchymal transition and metabolic pathways using Q-PCR method

Hypothesis

HSV-G47Δ oncolytic virus significantly targets U87-GBMCSCs in different GBM biological pathways and has ability to control GBM progression.

REFERENCES

- Adamson, C., Kanu, O. O., Mehta, A. I., Di, C., Lin, N., Mattox, A. K., & Bigner, D. D. (2009). Glioblastoma multiforme: A review of where we have been and where we are going. *Expert Opinion on Investigational Drugs*, 18(8), 1061–1083. <https://doi.org/10.1517/13543780903052764>
- Aghi, M. K., Liu, T. C., Rabkin, S., & Martuza, R. L. (2009). Hypoxia enhances the replication of oncolytic herpes simplex virus. *Molecular Therapy*, 17(1), 51–56. <https://doi.org/10.1038/mt.2008.232>
- Aghi, M., & Martuza, R. L. (2005). Oncolytic viral therapies - The clinical experience. *Oncogene*, 24(52), 7802–7816. <https://doi.org/10.1038/sj.onc.1209037>
- Agnihotri, S., Burrell, K. E., Wolf, A., Jalali, S., Hawkins, C., Rutka, J. T., & Zadeh, G. (2013). Glioblastoma, a brief review of history, molecular genetics, animal models and novel therapeutic strategies. *Archivum Immunologiae et Therapiae Experimentalis*, 61(1), 25–41. <https://doi.org/10.1007/s00005-012-0203-0>
- Agnihotri, S., Golbourn, B., Huang, X., Remke, M., Younger, S., Cairns, R. A., ... Rutka, J. T. (2016). PINK1 is a negative regulator of growth and the warburg effect in glioblastoma. *Cancer Research*, 76(16), 4708–4719. <https://doi.org/10.1158/0008-5472.CAN-15-3079>
- Ahir, B. K., Engelhard, H. H., & Lakka, S. S. (2020). Tumor Development and Angiogenesis in Adult Brain Tumor: Glioblastoma. *Molecular Neurobiology*, 57(5), 2461–2478. <https://doi.org/10.1007/s12035-020-01892-8>
- Aoki, H., Kondo, Y., Aldape, K., Yamamoto, A., Yokoyama, T., Hollingsworth, E. F., ... Kondo, S. (2008). *io s ci en ce o r u*. 8627. <https://doi.org/10.4161/auto.5668>
- Auffinger, B., Tobias, A. L., Han, Y., Lee, G., Guo, D., Dey, M., ... Ahmed, A. U. (2014). Conversion of differentiated cancer cells into cancer stem-like cells in a glioblastoma model after primary chemotherapy. *Cell Death and Differentiation*, 21(7), 1119–1131. <https://doi.org/10.1038/cdd.2014.31>
- Avivar-Valderas, A., Salas, E., Bobrovnikova-Marjon, E., Diehl, J. A., Nagi, C., Debnath, J., & Aguirre-Ghiso, J. A. (2011). PERK Integrates Autophagy and Oxidative Stress Responses To Promote Survival during Extracellular Matrix Detachment. *Molecular and Cellular Biology*, 31(17), 3616–3629. <https://doi.org/10.1128/mcb.05164-11>
- Babae, N., Bourajjaj, M., Liu, Y., Van Beijnum, J. R., Cerisoli, F., Scaria, P. V., ... Schiffelers, R. M. (2014). Systemic miRNA-7 delivery inhibits tumor angiogenesis and growth in murine xenograft glioblastoma. *Oncotarget*, 5(16), 6687–6700. <https://doi.org/10.18632/oncotarget.2235>

- Bai, Y., Lathia, J. D., Zhang, P., Flavahan, W., Rich, J. N., & Mattson, M. P. (2014). Molecular targeting of TRF2 suppresses the growth and tumorigenesis of glioblastoma stem cells. *Glia*, 62(10), 1687–1698. <https://doi.org/10.1002/glia.22708>
- Banelli, B., Forlani, A., Allemani, G., Morabito, A., Pistillo, M. P., & Romani, M. (2017). MicroRNA in glioblastoma: An overview. *International Journal of Genomics*, 2017. <https://doi.org/10.1155/2017/7639084>
- Bar, E. E. (2011). Glioblastoma, cancer stem cells and hypoxia. *Brain Pathology*, 21(2), 119–129. <https://doi.org/10.1111/j.1750-3639.2010.00460.x>
- Bar, E. E., Lin, A., Mahairaki, V., Matsui, W., & Eberhart, C. G. (2010). Hypoxia increases the expression of stem-cell markers and promotes clonogenicity in glioblastoma neurospheres. *American Journal of Pathology*, 177(3), 1491–1502. <https://doi.org/10.2353/ajpath.2010.091021>
- Beck, S., Jin, X., Sohn, Y. W., Kim, J. K., Kim, S. H., Yin, J., ... Kim, H. (2011). Telomerase activity-independent function of TERT allows glioma cells to attain cancer stem cell characteristics by inducing EGFR expression. *Molecules and Cells*, 31(1), 9–15. <https://doi.org/10.1007/s10059-011-0008-8>
- Beier, D., Schriefer, B., Brawanski, K., Hau, P., Weis, J., Schulz, J. B., & Beier, C. P. (2012). Efficacy of clinically relevant temozolamide dosing schemes in glioblastoma cancer stem cell lines. *Journal of Neuro-Oncology*, 109(1), 45–52. <https://doi.org/10.1007/s11060-012-0878-4>
- Bentwich, I., Avniel, A., Karov, Y., Aharonov, R., Gilad, S., Barad, O., ... Bentwich, Z. (2005). Identification of hundreds of conserved and nonconserved human microRNAs. *Nature Genetics*, 37(7), 766–770. <https://doi.org/10.1038/ng1590>
- Berezikov, E., Guryev, V., Van De Belt, J., Wienholds, E., Plasterk, R. H. A., & Cuppen, E. (2005). Phylogenetic shadowing and computational identification of human microRNA genes. *Cell*, 120(1), 21–24. <https://doi.org/10.1016/j.cell.2004.12.031>
- Berger, W., Steiner, E., Grusch, M., Elbling, L., & Micksche, M. (2009). Vaults and the major vault protein: Novel roles in signal pathway regulation and immunity. *Cellular and Molecular Life Sciences*, 66(1), 43–61. <https://doi.org/10.1007/s00018-008-8364-z>
- Berger, Walter, Spiegl-Kreinecker, S., Buchroithner, J., Elbling, L., Pirker, C., Fischer, J., & Micksche, M. (2001). Overexpression of the human major vault protein in astrocytic brain tumor cells. *International Journal of Cancer*, 94(3), 377–382. <https://doi.org/10.1002/ijc.1486>
- Bhattacharyya, S., Sandy, A., & Groden, J. (2010). Unwinding protein complexes in ALternative telomere maintenance. *Journal of Cellular Biochemistry*, 109(1), 7–15. <https://doi.org/10.1002/jcb.22388>

- Bian, L., Meng, Y., Zhang, M., & Li, D. (2019). MRE11-RAD50-NBS1 complex alterations and DNA damage response: Implications for cancer treatment. *Molecular Cancer*, 18(1), 1–14. <https://doi.org/10.1186/s12943-019-1100-5>
- Bier, A., Giladi, N., Kronfeld, N., Lee, H. K., Cazacu, S., Finniss, S., ... Brodie, C. (2013). MicroRNA-137 is downregulated in glioblastoma and inhibits the stemness of glioma stem cells by targeting RTVP-1. *Oncotarget*, 4(5), 665–676. <https://doi.org/10.18632/oncotarget.928>
- Birney, E., Stamatoyannopoulos, J. A., Dutta, A., Guigó, R., Gingeras, T. R., Margulies, E. H., ... De Jong, P. J. (2007). Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project. *Nature*, 447(7146), 799–816. <https://doi.org/10.1038/nature05874>
- Blackburn, E. H. (2010). Telomeres and telomerase: The means to the end (Nobel lecture). *Angewandte Chemie - International Edition*, 49(41), 7405–7421. <https://doi.org/10.1002/anie.201002387>
- Blasco, M. A. (2007). Telomere length, stem cells and aging. *Nature Chemical Biology*, 3(10), 640–649. <https://doi.org/10.1038/nchembio.2007.38>
- Boado, R. J., Black, K. L., & Pardridge, W. M. (1994). Gene expression of GLUT3 and GLUT1 glucose transporters in human brain tumors. *Molecular Brain Research*, 27(1), 51–57. [https://doi.org/10.1016/0169-328X\(94\)90183-X](https://doi.org/10.1016/0169-328X(94)90183-X)
- Bondy, M. L., Davis, F. G., II, D., Kruchko, C., McCarthy, J., Rajaraman, P., & Schwartzbaum, J. A. (2008). Brain Tumor Epidemiology: Consensus from the Brain Tumor Epidemiology Consortium (BTEC). *Cancer*, 113, 1953–1968. <https://doi.org/10.1002/cncr.23741>
- Bossy-Wetzel, E., Bakiri, L., & Yaniv, M. (1997). Induction of apoptosis by the transcription factor c-Jun. *EMBO Journal*, 16(7), 1695–1709. <https://doi.org/10.1093/emboj/16.7.1695>
- Brat, D. J., & Van Meir, E. G. (2001). Glomeruloid microvascular proliferation orchestrated by VPF/VEGF: A new world of angiogenesis research. *American Journal of Pathology*, 158(3), 789–796. [https://doi.org/10.1016/S0002-9440\(10\)64025-4](https://doi.org/10.1016/S0002-9440(10)64025-4)
- Brech, A., Ahlquist, T., Lothe, R. A., & Stenmark, H. (2009). Autophagy in tumour suppression and promotion. *Molecular Oncology*, 3(4), 366–375. <https://doi.org/10.1016/j.molonc.2009.05.007>
- Bredel, M. (2001). Anticancer drug resistance in primary human brain tumors. *Brain Research Reviews*, 35(2), 161–204. [https://doi.org/10.1016/S0165-0173\(01\)00045-5](https://doi.org/10.1016/S0165-0173(01)00045-5)

- Brown, D. V., Filiz, G., Daniel, P. M., Hollande, F., Dworkin, S., Amiridis, S., ... Mantamadiotis, T. (2017). Expression of CD133 and CD44 in glioblastoma stem cells correlates with cell proliferation, phenotype stability and intratumor heterogeneity. *PLoS ONE*, 12(2), 1–17. <https://doi.org/10.1371/journal.pone.0172791>
- Cai, G., Qiao, S., & Chen, K. (2015). Suppression of miR-221 inhibits glioma cells proliferation and invasion via targeting SEMA3B. *Biological Research*, 48, 1–8. <https://doi.org/10.1186/s40659-015-0030-y>
- Cao, S., Wang, Y., Li, J., Lv, M., Niu, H., & Tian, Y. (2016). Tumor-suppressive function of long noncoding RNA MALAT1 in glioma cells by suppressing miR-155 expression and activating FBXW7 function. *American Journal of Cancer Research*, 6(11), 2561–2574.
- Carmeliet, P., & Jain, R. K. (2011). Principles and mechanisms of vessel normalization for cancer and other angiogenic diseases. *Nature Reviews Drug Discovery*, 10(6), 417–427. <https://doi.org/10.1038/nrd3455>
- Carninci, P., Kasukawa, T., Katayama, S., Gough, J., Frith, M. C., Maeda, N., ... Hayashizaki, Y. (2005). Molecular biology: The transcriptional landscape of the mammalian genome. *Science*, 309(5740), 1559–1563. <https://doi.org/10.1126/science.1112014>
- Cheema, T. A., Wakimoto, H., Fecci, P. E., Ning, J., Kuroda, T., Jeyaretna, D. S., ... Rabkin, S. D. (2013). Multifaceted oncolytic virus therapy for glioblastoma in an immunocompetent cancer stem cell model. *Proceedings of the National Academy of Sciences of the United States of America*, 110(29), 12006–12011. <https://doi.org/10.1073/pnas.1307935110>
- Chen, H., Li, X., Li, W., & Zheng, H. (2015). miR-130a can predict response to temozolamide in patients with glioblastoma multiforme, independently of O6-methylguanine-DNA methyltransferase. *Journal of Translational Medicine*, 13(1), 1. <https://doi.org/10.1186/s12967-015-0435-y>
- Chen, J., Li, Y., Yu, T.-S., McKay, R. M., Burns, D., Kernie, S. G., & Parada, L. F. (2012a). Following Chemotherapy. *Nature*, 488(7412), 522–526. <https://doi.org/10.1038/nature11287.A>
- Chen, J., Li, Y., Yu, T. S., McKay, R. M., Burns, D. K., Kernie, S. G., & Parada, L. F. (2012b). A restricted cell population propagates glioblastoma growth after chemotherapy. *Nature*, 488(7412), 522–526. <https://doi.org/10.1038/nature11287>
- Chen, W., Xu, X. K., Li, J. L., Kong, K. K., Li, H., Chen, C., ... Li, F. C. (2017). MALAT1 is a prognostic factor in glioblastoma multiforme and induces chemoresistance to temozolamide through suppressing miR-203 and promoting thymidylate synthase expression. *Oncotarget*, 8(14), 22783–22799. <https://doi.org/10.18632/oncotarget.15199>

- Cheng, Z., Li, Z., Ma, K., Li, X., Tian, N., Duan, J., ... Wang, Y. (2017). Long non-coding RNA XIST promotes glioma tumorigenicity and angiogenesis by acting as a molecular sponge of miR-429. *Journal of Cancer*, 8(19), 4106–4116. <https://doi.org/10.7150/jca.21024>
- Chien, C.-H., Hsueh, W.-T., Chuang, J.-Y., & Chang, K.-Y. (2019). Role of autophagy in therapeutic resistance of glioblastoma. *Journal of Cancer Metastasis and Treatment*, 2019. <https://doi.org/10.20517/2394-4722.2019.016>
- Chittenden, T. W., Chittenden, T. W., Howe, E. a., Howe, E. a., Culhane, a. C., Culhane, a. C., ... Quackenbush, J. (2008). Functional classification analysis of somatically mutated genes in human breast and colorectal cancers. *Genomics*, 455(7216), 1061–1068. <https://doi.org/10.1038/nature07385>.Comprehensive
- Choi, K. H., Farrell, A. S., Lakamp, A. S., & Ouellette, M. M. (2011). Characterization of the DNA binding specificity of Shelterin complexes. *Nucleic Acids Research*, 39(21), 9206–9223. <https://doi.org/10.1093/nar/gkr665>
- Chourasia, A. H., Boland, M. L., & Macleod, K. F. (2015). Mitophagy and cancer. *Cancer and Metabolism*, 3(1), 1–11. <https://doi.org/10.1186/s40170-015-0130-8>
- Ciafrè, S. A., Galardi, S., Mangiola, A., Ferracin, M., Liu, C. G., Sabatino, G., ... Farace, M. G. (2005). Extensive modulation of a set of microRNAs in primary glioblastoma. *Biochemical and Biophysical Research Communications*, 334(4), 1351–1358. <https://doi.org/10.1016/j.bbrc.2005.07.030>
- Cj, P., Hv, E., Vijayakurup, V., R Menon, G., Nair, S., & Gopala, S. (2019). High LC3/Beclin Expression Correlates with Poor Survival in Glioma: a Definitive Role for Autophagy as Evidenced by In Vitro Autophagic Flux. *Pathology and Oncology Research*, 25(1), 137–148. <https://doi.org/10.1007/s12253-017-0310-7>
- Clarke, K., Smith, K., Gullick, W. J., & Harris, A. L. (2001). Mutant epidermal growth factor receptor enhances induction of vascular endothelial growth factor by hypoxia and insulin-like growth factor-1 via a PI3 kinase dependent pathway. *British Journal of Cancer*, 84(10), 1322–1329. <https://doi.org/10.1054/bjoc.2001.1805>
- Cloughesy, T. F., Cavenee, W. K., & Mischel, P. S. (2014). Glioblastoma: From molecular pathology to targeted treatment. *Annual Review of Pathology: Mechanisms of Disease*, 9, 1–25. <https://doi.org/10.1146/annurev-pathol-011110-130324>

- Codrici, E., Enciu, A. M., Popescu, I. D., Mihai, S., & Tanase, C. (2016). Glioma Stem Cells and Their Microenvironments: Providers of Challenging Therapeutic Targets. *Stem Cells International*, 2016. <https://doi.org/10.1155/2016/5728438>
- Coleman, J. F. (2010). Robbins and Cotran's Pathologic Basis of Disease, 8th Edition. In *The American Journal of Surgical Pathology* (Vol. 34). <https://doi.org/10.1097/pas.0b013e3181bc5f0f>
- Cong, Y.-S., Wright, W. E., & Shay, J. W. (2002). Human Telomerase and Its Regulation. *Microbiology and Molecular Biology Reviews*, 66(3), 407–425. <https://doi.org/10.1128/mmbr.66.3.407-425.2002>
- Counter, C. M., Hahn, W. C., Wei, W., Caddle, S. D., Beijersbergen, R. L., Lansdorp, P. M., ... Weinberg, R. A. (1998). Dissociation among in vitro telomerase activity, telomere maintenance, and cellular immortalization. *Proceedings of the National Academy of Sciences of the United States of America*, 95(25), 14723–14728. <https://doi.org/10.1073/pnas.95.25.14723>
- D 'adda, F., Fagagna, D., Reaper, P. M., Clay-Farrace, L., Fiegler, H., Carr, P., ... Jackson, S. P. (2003). A DNA damage checkpoint response in telomere-initiated senescence. *Nature*, 426(November), 194–198.
- Dadey, D. Y. A., Kapoor, V., Khudanyan, A., Thotala, D., & Hallahan, D. E. (2018). PERK regulates glioblastoma sensitivity to ER stress although promoting radiation resistance. *Molecular Cancer Research*, 16(10), 1447–1453. <https://doi.org/10.1158/1541-7786.MCR-18-0224>
- de Biase, D., Visani, M., Morandi, L., Marucci, G., Taccioli, C., Cerasoli, S., ... Nobile, C. (2012). miRNAs expression analysis in paired fresh/Frozen and dissected formalin fixed and paraffin embedded glioblastoma using real-time PCR. *PLoS ONE*, 7(4), 1–7. <https://doi.org/10.1371/journal.pone.0035596>
- de Lange T. (2009). How Telomeres Solve the End-Protection Problem. *Science*, 326(5955), 948. <https://doi.org/10.1126/science.1170633>.How
- Denchi, E. L., & De Lange, T. (2007). Protection of telomeres through independent control of ATM and ATR by TRF2 and POT1. *Nature*, 448(7157), 1068–1071. <https://doi.org/10.1038/nature06065>
- Deng, Z., Kim, E. T., Vladimirova, O., Dheekollu, J., Wang, Z., Liu, D., ... Paul, M. (2015). *HHS Public Access*. 9(6), 2263–2278. <https://doi.org/10.1016/j.celrep.2014.11.019.HSV-1>
- Derrien, T., Johnson, R., Bussotti, G., Tanzer, A., Djebali, S., Tilgner, H., ... Guigó, R. (2012). The GENCODE v7 catalog of human long noncoding RNAs: Analysis of their gene structure, evolution, and expression. *Genome Research*, 22(9), 1775–1789. <https://doi.org/10.1101/gr.132159.111>

- Dews, M., Homayouni, A., Yu, D., Murphy, D., Sevignani, C., Wentzel, E., ... Thomas-Tikhonenko, A. (2006). Augmentation of tumor angiogenesis by a Myc-activated microRNA cluster. *Nature Genetics*, 38(9), 1060–1065. <https://doi.org/10.1038/ng1855>
- Di, H., Li, C., Wang, X., Du, C., Sun, S., Li, Y., ... Wang, J. (2017). Overexpression of miR-874 enhances chemosensitivity of glioma cells to temozolomide by the oncogenic STAT3 pathway. *International Journal of Clinical and Experimental Pathology*, 10(3), 2852–2860.
- Diane E. Handy, Rita Castro, J. L. (2011). 基因的改变NIH Public Access. *Bone*, 23(1), 1–7. <https://doi.org/10.1161/CIRCULATIONAHA.110.956839>
- Dimov, I., Tasić-Dimov, D., Conić, I., & Stefanovic, V. (2011). Glioblastoma multiiforme stem cells. *TheScientificWorldJournal*, 11(June 2014), 930–958. <https://doi.org/10.1100/tsw.2011.42>
- Dinger, M. E., Amaral, P. P., Mercer, T. R., & Mattick, J. S. (2009). Pervasive transcription of the eukaryotic genome: Functional indices and conceptual implications. *Briefings in Functional Genomics and Proteomics*, 8(6), 407–423. <https://doi.org/10.1093/bfgp/elp038>
- Diotti, R., & Loayza, D. (2011). Shelterin complex and associated factors at human telomeres. *Nucleus*, 2(2). <https://doi.org/10.4161/nucl.2.2.15135>
- Du, H., Cui, S., Li, Y., Yang, G., Wang, P., Fikrig, E., & You, F. (2018). MiR-221 negatively regulates innate anti-viral response. *PLoS ONE*, 13(8), 1–13. <https://doi.org/10.1371/journal.pone.0200385>
- Du, P., Zhao, H., Peng, R., Liu, Q., Yuan, J., Peng, G., & Liao, Y. (2017). LncRNA-XIST interacts with miR-29c to modulate the chemoresistance of glioma cell to TMZ through DNA mismatch repair pathway. *Bioscience Reports*, 37(5), 1–12. <https://doi.org/10.1042/BSR20170696>
- Duncan, C. G., Killela, P. J., Payne, C. A., Lampson, B., Chen, W. C., Liu, J., ... Yan, H. (2010). Integrated genomic analyses identify ERRFI1 and TACC3 as glioblastoma-targeted genes. *Oncotarget*, 1(4), 265–277. <https://doi.org/10.18632/oncotarget.137>
- Dunn, A. M., Hofmann, O. S., Waters, B., & Witchel, E. (2011). Cloaking malware with the trusted platform module. *Proceedings of the 20th USENIX Security Symposium*, pp. 395–410.
- Durant, S. T. (2012). Telomerase-independent paths to immortality in predictable cancer sub-types. *Journal of Cancer*, 3(1), 67–82. <https://doi.org/10.7150/jca.3965>
- Eissa, I. R., Bustos-Villalobos, I., Ichinose, T., Matsumura, S., Naoe, Y., Miyajima, N., ... Kasuya, H. (2018). The current status and future prospects of oncolytic viruses in clinical trials against melanoma, glioma, pancreatic, and breast cancers. *Cancers*, 10(10). <https://doi.org/10.3390/cancers10100356>

- Elias, M. C., Tozer, K. R., Silber, J. R., Mikheeva, S., Deng, M., Morrison, R. S., ... Rostomily, R. C. (2005). TWIST is expressed in human gliomas and promotes invasion. *Neoplasia*, 7(9), 824–837.
<https://doi.org/10.1593/neo.04352>
- Fairall, L., Chapman, L., Moss, H., De Lange, T., & Rhodes, D. (2001). Structure of the TRFH dimerization domain of the human telomeric proteins TRF1 and TRF2. *Molecular Cell*, 8(2), 351–361. [https://doi.org/10.1016/S1097-2765\(01\)00321-5](https://doi.org/10.1016/S1097-2765(01)00321-5)
- Family, H. D. (2009). *Radiation-induced gliomas*. 9(10), 1511–1517.
- Fernandes, J. C. R., Acuña, S. M., Aoki, J. I., Floeter-Winter, L. M., & Muxel, S. M. (2019). Long non-coding RNAs in the regulation of gene expression: Physiology and disease. *Non-Coding RNA*, 5(1).
<https://doi.org/10.3390/ncrna5010017>
- Fischer, P. M. (2004). The Use of CDK Inhibitors in Oncology and beyond. *Biochemical and Biophysical Research Communications*, 327(3), 742–746. [https://doi.org/10.1016/j.bbrc.2004.09.077 \[pii\]](https://doi.org/10.1016/j.bbrc.2004.09.077)
- Fisher, J. L., Schwartzbaum, J. A., Wrensch, M., & Wiemels, J. L. (2007). Epidemiology of Brain Tumors. *Neurologic Clinics*, 25(4), 867–890. <https://doi.org/10.1016/j.ncl.2007.07.002>
- Frederick, L., Wang, X. Y., Eley, G., & James, C. D. (2000). Diversity and frequency of epidermal growth factor receptor mutations in human glioblastomas. *Cancer Research*, 60(5), 1383–1387.
- Fu, Z., Luo, W., Wang, J., Peng, T., Sun, G., Shi, J., ... Zhang, B. (2017). Malat1 activates autophagy and promotes cell proliferation by sponging miR-101 and upregulating STMN1, RAB5A and ATG4D expression in glioma. *Biochemical and Biophysical Research Communications*, 492(3), 480–486. <https://doi.org/10.1016/j.bbrc.2017.08.070>
- Fukuhara, H., Ino, Y., & Todo, T. (2016). Oncolytic virus therapy: A new era of cancer treatment at dawn. *Cancer Science*, 107(10), 1373–1379. <https://doi.org/10.1111/cas.13027>
- Furnari, F. B., Fenton, T., Bachoo, R. M., Mukasa, A., Stommel, J. M., Stegh, A., ... Cavenee, W. K. (2007). Malignant astrocytic glioma: Genetics, biology, and paths to treatment. *Genes and Development*, 21(21), 2683–2710. <https://doi.org/10.1101/gad.1596707>
- Furuno, M., Pang, K. C., Ninomiya, N., Fukuda, S., Frith, M. C., Bult, C., ... Suzuki, H. (2006). Clusters of internally primed transcripts reveal novel long noncoding RNAs. *PLoS Genetics*, 2(4), 537–553. <https://doi.org/10.1371/journal.pgen.0020037>

- Galavotti, S., Bartesaghi, S., Faccenda, D., Shaked-Rabi, M., Sanzone, S., McEvoy, A., ... Salomoni, P. (2013). The autophagy-associated factors DRAM1 and p62 regulate cell migration and invasion in glioblastoma stem cells. *Oncogene*, 32(6), 699–712. <https://doi.org/10.1038/onc.2012.111>
- Galli, R., Binda, E., Orfanelli, U., Cipelletti, B., Gritti, A., De Vitis, S., ... Vescovi, A. (2004). Erratum: Isolation and characterization of tumorigenic, stem-like neural precursors from human glioblastoma (Cancer Research (October 2004) 64 (7011-7021). *Cancer Research*, 64(21), 8130.
- Garofalo, M., Quintavalle, C., Romano, G., M. Croce, C., & Condorelli, G. (2011). miR221/222 in Cancer: Their Role in Tumor Progression and Response to Therapy. *Current Molecular Medicine*, 12(1), 27–33. <https://doi.org/10.2174/156652412798376170>
- Gatei, M., Jakob, B., Chen, P., Kijas, A. W., Becherel, O. J., Gueven, N., ... Lavin, M. F. (2011). ATM protein-dependent phosphorylation of Rad50 protein Regulates DNA repair and cell cycle control. *Journal of Biological Chemistry*, 286(36), 31542–31556. <https://doi.org/10.1074/jbc.M111.258152>
- GmbH, M. B. (n.d.). *Annexin V-FITC Kit*. (130), 1–2.
- Goker Bagca, B., & Biray Avci, C. (2018). *Cancer Stem Cells and Autophagy: Present Knowledge and Future Perspectives*. (January), 163–177. https://doi.org/10.1007/978-3-319-98146-8_11
- Gopinath, S. C. B., Wadhwa, R., & Kumar, P. K. R. (2010). Expression of noncoding vault RNA in human malignant cells and its importance in mitoxantrone resistance. *Molecular Cancer Research*, 8(11), 1536–1546. <https://doi.org/10.1158/1541-7786.MCR-10-0242>
- Grant, R., Kolb, L., & Moliterno, J. (2014). Molecular and genetic pathways in gliomas: the future of personalized therapeutics. *CNS Oncology*, 3(2), 123–136. <https://doi.org/10.2217/cns.14.7>
- Greider, C. W. (2010). Telomerase discovery: The excitement of putting together pieces of the puzzle (Nobel lecture). *Angewandte Chemie - International Edition*, 49(41), 7422–7439. <https://doi.org/10.1002/anie.201002408>
- Greider, C. W., & Blackburn, E. H. (1985). Identification of a specific telomere terminal transferase activity in tetrahymena extracts. *Cell*, 43(2 PART 1), 405–413. [https://doi.org/10.1016/0092-8674\(85\)90170-9](https://doi.org/10.1016/0092-8674(85)90170-9)
- Grigorenko, E. V., Ortenberg, E., Hurley, J., Bond, A., & Munnelly, K. (2011). MicroRNA and Cancer - miRNA Profiling on High-Throughput OpenArray™ System. *BOOK: MicroRNA and Cancer*, 676, 101–110. <https://doi.org/10.1007/978-1-60761-863-8>
- Guan, J.-Z., Wei-Ping, Maeda, T., & Makino, N. (2011). Differential effect of hypoxia regulating telomere length and telomerase activity. *Aging Clinical and Experimental Research*, 24(3), 3–7.

- Guo, J., Cai, H., Zheng, J., Liu, X., Liu, Y., Ma, J., ... Xue, Y. (2017). Long non-coding RNA NEAT1 regulates permeability of the blood-tumor barrier via miR-181d-5p-mediated expression changes in ZO-1, occludin, and claudin-5. *Biochimica et Biophysica Acta - Molecular Basis of Disease*, 1863(9), 2240–2254. <https://doi.org/10.1016/j.bbadi.2017.02.005>
- Guo, Y., Yan, K., Fang, J., Qu, Q., Zhou, M., & Chen, F. (2013). Let-7b expression determines response to chemotherapy through the regulation of Cyclin D1 in Glioblastoma. *Journal of Experimental and Clinical Cancer Research*, 32(1). <https://doi.org/10.1186/1756-9966-32-41>
- Guo, Z. S., Liu, Z., Kowalsky, S., Feist, M., Kalinski, P., Lu, B., ... Bartlett, D. L. (2017). Oncolytic immunotherapy: Conceptual evolution, current strategies, and future perspectives. *Frontiers in Immunology*, 8(MAY). <https://doi.org/10.3389/fimmu.2017.00555>
- Haas-kogan, D. A., Francisco, S., James, C. D., Francisco, S., Debnath, J., Francisco, S., ... Francisco, S. (2010). NIH Public Access. (November). <https://doi.org/10.1126/scisignal.2001017>
- Hamasaki, M., Furuta, N., Matsuda, A., Nezu, A., Yamamoto, A., Fujita, N., ... Yoshimori, T. (2013). Autophagosomes form at ER-mitochondria contact sites. *Nature*, 495(7441), 389–393. <https://doi.org/10.1038/nature11910>
- Han, Y., Wu, Z., Wu, T., Huang, Y., Cheng, Z., Li, X., ... Du, Z. (2016). Tumor-suppressive function of long noncoding RNA MALAT1 in glioma cells by downregulation of MMP2 and inactivation of ERK/MAPK signaling. *Cell Death & Disease*, 7, e2123. <https://doi.org/10.1038/cddis.2015.407>
- Hanahan, D., & Folkman, J. (1996). Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. *Cell*, 86(3), 353–364. [https://doi.org/10.1016/S0092-8674\(00\)80108-7](https://doi.org/10.1016/S0092-8674(00)80108-7)
- Hanif, F., Muzaffar, K., Perveen, K., Malhi, S. M., & Simjee, S. U. (2017). Glioblastoma multiforme: A review of its epidemiology and pathogenesis through clinical presentation and treatment. *Asian Pacific Journal of Cancer Prevention*, 18(1), 3–9. <https://doi.org/10.22034/APJCP.2017.18.1.3>
- Hao, J., Zhang, C., Zhang, A., Wang, K., Zhifan, J. A., Wang, G., ... Pu, P. (2012). miR-221/222 is the regulator of Cx43 expression in human glioblastoma cells. *Oncology Reports*, 27(5), 1504–1510. <https://doi.org/10.3892/or.2012.1652>
- Harley, C. B. (1991). Telomere loss: mitotic clock or genetic time bomb? *Mutation Research DNAging*, 256(2–6), 271–282. [https://doi.org/10.1016/0921-8734\(91\)90018-7](https://doi.org/10.1016/0921-8734(91)90018-7)
- Harrington, L., Mcphail, T., Mar, V., Zhou, W., Oulton, R., Program, A. E. S. T., ... Robinson, M. (1997). A Mammalian Telomerase-Associated Protein. 275(February), 973–977.

- Hastie, N. D., Dempster, M., Dunlop, M. G., Thompson, A. M., Green, D. K., & Allshire, R. C. (1990). Telomere reduction in human colorectal carcinoma and with ageing. *Nature*, 346(6287), 866–868.
<https://doi.org/10.1038/346866a0>
- Hausott, B., Park, J. W., Valovka, T., Offterdinger, M., Hess, M. W., Geley, S., & Klimaschewski, L. (2019). Subcellular localization of sprouty2 in human glioma cells. *Frontiers in Molecular Neuroscience*, 12(March), 1–11.
<https://doi.org/10.3389/fnmol.2019.00073>
- Hayashi-Nishino, M., Fujita, N., Noda, T., Yamaguchi, A., Yoshimori, T., & Yamamoto, A. (2009). A subdomain of the endoplasmic reticulum forms a cradle for autophagosome formation. *Nature Cell Biology*, 11(12), 1433–1437. <https://doi.org/10.1038/ncb1991>
- He, C., Jiang, B., Ma, J., & Li, Q. (2016). Aberrant NEAT1 expression is associated with clinical outcome in high grade glioma patients. *Apmis*, 124(3), 169–174. <https://doi.org/10.1111/apm.12480>
- He, H., & Qin, M. (2020). Long non-coding RNA LEF1-AS1 is involved in the progression of retinoblastoma through regulating the Wnt/β-catenin pathway. *Clinical and Experimental Pharmacology and Physiology*, (December 2019), 1–6. <https://doi.org/10.1111/1440-1681.13263>
- Heddleston, J. M., Li, Z., Lathia, J. D., Bao, S., Hjelmeland, A. B., & Rich, J. N. (2010). Hypoxia inducible factors in cancer stem cells. *British Journal of Cancer*, 102(5), 789–795. <https://doi.org/10.1038/sj.bjc.6605551>
- Hemmati, H. D., Nakano, I., Lazareff, J. A., Masterman-Smith, M., Geschwind, D. H., Bronner-Fraser, M., & Kornblum, H. I. (2003). Cancerous stem cells can arise from pediatric brain tumors. *Proceedings of the National Academy of Sciences of the United States of America*, 100(25), 15178–15183. <https://doi.org/10.1073/pnas.2036535100>
- Hermann, B., Dietrich, R., & Volker, A. (2009). Cancer Epidemiology: Vol 2, Modifiable Factors. *Methods in Molecular Biology*, 472, 467–475. <https://doi.org/10.1007/978-1-60327-492-0>
- Herrmann, M., Pusceddu, I., März, W., & Herrmann, W. (2018). Telomere biology and age-related diseases. *Clinical Chemistry and Laboratory Medicine*, 56(8), 1210–1222. <https://doi.org/10.1515/cclm-2017-0870>
- Hiraga, S., Ohnishi, T., Izumoto, S., Miyahara, E., Kanemura, Y., Matsumura, H., & Arita, N. (1998a). Telomerase activity and alterations in telomere length in human brain tumors. *Cancer Research*, 58(10), 2117–2125.
- Hiraga, S., Ohnishi, T., Izumoto, S., Miyahara, E., Kanemura, Y., Matsumura, H., & Arita, N. (1998b). Telomerase activity and alterations in telomere length in human brain tumors. *Cancer Research*, 58(10), 2117–2125.

- Hjelmeland, A. B., Wu, Q., Heddleston, J. M., Choudhary, G. S., MacSwords, J., Lathia, J. D., ... Rich, J. N. (2011). Acidic stress promotes a glioma stem cell phenotype. *Cell Death and Differentiation*, 18(5), 829–840. <https://doi.org/10.1038/cdd.2010.150>
- Holland, E. C. (2000). Glioblastoma multiforme: The terminator. *Proceedings of the National Academy of Sciences of the United States of America*, 97(12), 6242–6244. <https://doi.org/10.1073/pnas.97.12.6242>
- Hurst, D., & Cody, J. (2015). Promising oncolytic agents for metastatic breast cancer treatment. *Oncolytic Virotherapy*, 63. <https://doi.org/10.2147/ov.s63045>
- Iacob, G., & Dinca, E. B. (2009). Current data and strategy in glioblastoma multiforme. *Journal of Medicine and Life*, 2(4), 386–393.
- Inskip, P. D., Tarone, R. E., Hatch, E. E., Wilcosky, T. C., Shapiro, W. R., Selker, R. G., ... Linet, M. S. (2001). Cellular-telephone use and brain tumors. *New England Journal of Medicine*, 344(2), 79–86. <https://doi.org/10.1056/NEJM200101113440201>
- Iser, I. C., Pereira, M. B., Lenz, G., & Wink, M. R. (2017). The Epithelial-to-Mesenchymal Transition-Like Process in Glioblastoma: An Updated Systematic Review and In Silico Investigation. *Medicinal Research Reviews*, 37(2), 271–313. <https://doi.org/10.1002/med.21408>
- Jiang, L., Mao, P., Song, L., Wu, J., Huang, J., Lin, C., ... Li, J. (2010). miR-182 as a prognostic marker for glioma progression and patient survival. *American Journal of Pathology*, 177(1), 29–38. <https://doi.org/10.2353/ajpath.2010.090812>
- Jiang, P., Wang, P., Sun, X., Yuan, Z., Zhan, R., Ma, X., & Li, W. (2016). Knockdown of long noncoding RNARNARNA H19 sensitizes human glioma cells to temozolomide therapy. *OncoTargets and Therapy*, 9, 3501–3509. <https://doi.org/10.2147/OTT.S96278>
- Jiang, X., Yan, Y., Hu, M., Chen, X., Wang, Y., Dai, Y., ... Xia, H. (2016). Increased level of H19 long noncoding RNA promotes invasion, angiogenesis, and stemness of glioblastoma cells. *Journal of Neurosurgery*, 124(1), 129–136. <https://doi.org/10.3171/2014.12.JNS1426>
- Johnson, S. M., Grosshans, H., Shingara, J., Byrom, M., Jarvis, R., Cheng, A., ... Slack, F. J. (2005). RAS is regulated by the let-7 microRNA family. *Cell*, 120(5), 635–647. <https://doi.org/10.1016/j.cell.2005.01.014>
- JOVČEVSKA, I., KOČEVAR, N., & KOMEL, R. (2013). Glioma and glioblastoma - how much do we (not) know? *Molecular and Clinical Oncology*, 1(6), 935–941. <https://doi.org/10.3892/mco.2013.172>

- Kabat, G. C., Etgen, A. M., & Rohan, T. E. (2010). Do steroid hormones play a role in the etiology of glioma? *Cancer Epidemiology Biomarkers and Prevention*, 19(10), 2421–2427. <https://doi.org/10.1158/1055-9965.EPI-10-0658>
- Kaur, B., Khwaja, F. W., Severson, E. A., Matheny, S. L., Brat, D. J., & Van Meir, E. G. (2005). Hypoxia and the hypoxia-inducible-factor pathway in glioma growth and angiogenesis. *Neuro-Oncology*, 7(2), 134–153. <https://doi.org/10.1215/S1152851704001115>
- Kawamata, T., Yoda, M., & Tomari, Y. (2011). Multilayer checkpoints for microRNA authenticity during RISC assembly. *EMBO Reports*, 12(9), 944–949. <https://doi.org/10.1038/embor.2011.128>
- Kaza, N., Kohli, L., & Roth, K. A. (2012). Autophagy in brain tumors: A new target for therapeutic intervention. *Brain Pathology*, 22(1), 89–98. <https://doi.org/10.1111/j.1750-3639.2011.00544.x>
- Ke, J., Yao, Y. L., Zheng, J., Wang, P., Liu, Y. H., Ma, J., ... Xue, Y. X. (2015). Knockdown of long non-coding RNA HOTAIR inhibits malignant biological behaviors of human glioma cells via modulation of miR-326. *Oncotarget*, 6(26), 21934–21949. <https://doi.org/10.18632/oncotarget.4290>
- Kefas, B., Godlewski, J., Comeau, L., Li, Y., Abounader, R., Hawkinson, M., ... Purow, B. (2008). microRNA-7 inhibits the epidermal growth factor receptor and the akt pathway and is down-regulated in glioblastoma. *Cancer Research*, 68(10), 3566–3572. <https://doi.org/10.1158/0008-5472.CAN-07-6639>
- Khalil, A. M., Guttman, M., Huarte, M., Garber, M., Raj, A., Morales, D. R., ... Rinn, J. L. (2009). Many human large intergenic noncoding RNAs associate with chromatin-modifying complexes and affect gene expression. *Proceedings of the National Academy of Sciences of the United States of America*, 106(28), 11667–11672. <https://doi.org/10.1073/pnas.0904715106>
- Kickhoefer, V. A., Rajavel, K. S., Scheffer, G. L., Dalton, W. S., Schepers, R. J., & Rome, L. H. (1998). Vaults are up-regulated in multidrug-resistant cancer cell lines. *Journal of Biological Chemistry*, 273(15), 8971–8974. <https://doi.org/10.1074/jbc.273.15.8971>
- Kickhoefer, V. A., Siva, A. C., Kedersha, N. L., Inman, E. M., Ruland, C., Streuli, M., & Rome, L. H. (1999). The 193-kD vault protein, VPARP, is a novel poly(ADP-ribose) polymerase. *Journal of Cell Biology*, 146(5), 917–928. <https://doi.org/10.1083/jcb.146.5.917>
- Kickhoefer, V. A., Vasu, S. K., & Rome, L. H. (1996). Vaults are the answer, what is the question? *Trends in Cell Biology*, 6(5), 174–178. [https://doi.org/10.1016/0962-8924\(96\)10014-3](https://doi.org/10.1016/0962-8924(96)10014-3)

- Kieran, M. W. (2004). Anti-angiogenic chemotherapy in central nervous system tumors. *Cancer Treatment and Research*, 117, 337–349. https://doi.org/10.1007/978-1-4419-8871-3_19
- Kim, Y. D., Jang, S. J., Lim, E. J., Ha, J. S., Shivakumar, S. B., Jeong, G. J., ... Jeon, B. G. (2017). Induction of telomere shortening and cellular apoptosis by sodium meta-arsenite in human cancer cell lines. *Animal Cells and Systems*, 21(4), 241–254. <https://doi.org/10.1080/19768354.2017.1342691>
- Kirn, D., Martuza, R. L., & Zwiebel, J. (2001). Replication-selective virotherapy for cancer: Biological principles, risk management and future directions. *Nature Medicine*, 7(7), 781–787. <https://doi.org/10.1038/89901>
- Koga, K., Todaka, T., Morioka, M., Hamada, J. ichiro, Kai, Y., Yano, S., ... Suda, T. (2001). Expression of angiopoietin-2 in human glioma cells and its role for angiogenesis. *Cancer Research*, 61(16), 6248–6254.
- Kondo, T., Setoguchi, T., & Taga, T. (2004). Persistence of a small subpopulation of cancer stem-like cells in the C6 glioma cell line. *Proceedings of the National Academy of Sciences of the United States of America*, 101(3), 781–786. <https://doi.org/10.1073/pnas.0307618100>
- Konopaske GT, Dorph-Petersen KA, Sweet RA, Pierri JN, Zhang W, Sampson AR, L. DA. (2008). 基因的改变NIH Public Access. *Bone*, 23(1), 1–7. <https://doi.org/10.1038/jid.2014.371>
- Koo, C. L., Liew, M. J., Mohamad, M. S., Hakim, A., & Salleh, M. (2013). A Review for Detecting Gene-Gene Interactions Using Machine Learning Methods in Genetic Epidemiology. 2013.
- Kreitman, M. &. (1991). © 19 9 1 Nature Publishing Group 그라첼꺼. *Nature*, 353, 412–414.
- Kuroda, S., Fujiwara, T., Shirakawa, Y., Yamasaki, Y., Yano, S., Uno, F., ... Fujiwara, T. (2010). Telomerase-dependent oncolytic adenovirus sensitizes human cancer cells to ionizing radiation via inhibition of DNA repair machinery. *Cancer Research*, 70(22), 9339–9348. <https://doi.org/10.1158/0008-5472.CAN-10-2333>
- Kurth, I., Peitzsch, C., Baumann, M., & Dubrovska, A. (2014). The Role of Cancer Stem Cells in Tumor Radioresistance. *Cancer Stem Cells*, 9781118356(December), 473–491. <https://doi.org/10.1002/9781118356203.ch35>
- La, D., Raffa, G., Tomasello, C., Aguennouz, Mh., & Germano, A. (2013). Telomeres and Brain Tumors. *Clinical Management and Evolving Novel Therapeutic Strategies for Patients with Brain Tumors*, (April). <https://doi.org/10.5772/52426>

- Lakomy, R., Sana, J., Hankeova, S., Fadrus, P., Kren, L., Lzicarova, E., ... Slaby, O. (2011). MiR-195, miR-196b, miR-181c, miR-21 expression levels and O-6-methylguanine-DNA methyltransferase methylation status are associated with clinical outcome in glioblastoma patients. *Cancer Science*, 102(12), 2186–2190. <https://doi.org/10.1111/j.1349-7006.2011.02092.x>
- Lang, M. F., Yang, S., Zhao, C., Sun, G., Murai, K., Wu, X., ... Shi, Y. (2012). Genome-wide profiling identified a set of miRNAs that are differentially expressed in glioblastoma stem cells and normal neural stem cells. *PLoS ONE*, 7(4). <https://doi.org/10.1371/journal.pone.0036248>
- Lathia, J. D., Gallagher, J., Myers, J. T., Li, M., Vasanji, A., McLendon, R. E., ... Rich, J. N. (2011). Direct In Vivo Evidence for Tumor Propagation by Glioblastoma Cancer Stem Cells. 6(9), 1–9. <https://doi.org/10.1371/journal.pone.0024807>
- Leão, R., Apolónio, J. D., Lee, D., Figueiredo, A., Tabori, U., & Castelo-Branco, P. (2018). Mechanisms of human telomerase reverse transcriptase (hTERT) regulation: Clinical impacts in cancer. *Journal of Biomedical Science*, 25(1), 1–12. <https://doi.org/10.1186/s12929-018-0422-8>
- Lee, H. K., Lund, J. M., Ramanathan, B., Mizushima, N., & Iwasaki, A. (2007). Autophagy-Dependent Viral Recognition by Plasmacytoid Dendritic Cells Autophagy-Dependent Viral Recognition by Plasmacytoid Dendritic Cells. (April). <https://doi.org/10.1126/science.1136880>
- Lee, H. Y., Zhou, K., Smith, A. M., Noland, C. L., & Doudna, J. A. (2013). Differential roles of human Dicer-binding proteins TRBP and PACT in small RNA processing. *Nucleic Acids Research*, 41(13), 6568–6576. <https://doi.org/10.1093/nar/gkt361>
- Lee, J., Kotliarov, S., Kotliarov, Y., Li, A., Su, Q., Donin, N. M., ... Fine, H. A. (2006). Tumor stem cells derived from glioblastomas cultured in bFGF and EGF more closely mirror the phenotype and genotype of primary tumors than do serum-cultured cell lines. *Cancer Cell*, 9(5), 391–403. <https://doi.org/10.1016/j.ccr.2006.03.030>
- Lee, S. T., Chu, K., Oh, H. J., Im, W. S., Lim, J. Y., Kim, S. K., ... Roh, J. K. (2011). Let-7 microRNA inhibits the proliferation of human glioblastoma cells. *Journal of Neuro-Oncology*, 102(1), 19–24. <https://doi.org/10.1007/s11060-010-0286-6>
- Lee, Y., Ahn, C., Han, J., Choi, H., Kim, J., Yim, J., ... Kim, V. N. (2003). The nuclear RNase III Drosha initiates microRNA processing. *Nature*, 425(6956), 415–419. <https://doi.org/10.1038/nature01957>
- Lee, Y., Kim, M., Han, J., Yeom, K. H., Lee, S., Baek, S. H., & Kim, V. N. (2004). MicroRNA genes are transcribed by RNA polymerase II. *EMBO Journal*, 23(20), 4051–4060. <https://doi.org/10.1038/sj.emboj.7600385>

- Lee, Y. S., & Dutta, A. (2009). MicroRNAs in Cancer Contents: *Annu Rev Pathol*, 4, 199–227.
[https://doi.org/10.1146/annurev.pathol.4.110807.092222.MicroRNAs](https://doi.org/10.1146/annurev.pathol.4.110807.092222)
- Lei, M., Podell, E. R., & Cech, T. R. (2004). Structure of human POT1 bound to telomeric single-stranded DNA provides a model for chromosome end-protection. *Nature Structural and Molecular Biology*, 11(12), 1223–1229.
<https://doi.org/10.1038/nsmb867>
- Levine, B., & Kroemer, G. (2008). Autophagy in the Pathogenesis of Disease. *Cell*, 132(1), 27–42. <https://doi.org/10.1016/j.cell.2007.12.018>
- Li, C., Lei, B., Huang, S., Zheng, M., Liu, Z., Li, Z., & Deng, Y. (2015). H19 derived microRNA-675 regulates cell proliferation and migration through CDK6 in glioma. *American Journal of Translational Research*, 7(10), 1747–1764.
- Li, H. Y., Li, Y. M., Li, Y., Shi, X. W., & Chen, H. (2016). Circulating microRNA-137 is a potential biomarker for human glioblastoma. *European Review for Medical and Pharmacological Sciences*, 20(17), 3599–3604.
- Li, J., Zhu, Y., Wang, H., & Ji, X. (2018). Targeting Long Noncoding RNA in Glioma: A Pathway Perspective. *Molecular Therapy - Nucleic Acids*, 13(December), 431–441. <https://doi.org/10.1016/j.omtn.2018.09.023>
- Li, Z., Bao, S., Wu, Q., Wang, H., Eyler, C., Sathornsumetee, S., ... Rich, J. N. (2009). Hypoxia-Inducible Factors Regulate Tumorigenic Capacity of Glioma Stem Cells cancer stem cell specific molecules involved in neoangiogenesis, including HIF2α and its regulated factors. *Cancer Cell*, 15(6), 501–513. <https://doi.org/10.1016/j.ccr.2009.03.018>
- Lin, Q., & Yun, Z. (2010). Impact of the hypoxic tumor microenvironment on the regulation of cancer stem cell characteristics. *Cancer Biology and Therapy*, 9(12), 949–956. <https://doi.org/10.4161/cbt.9.12.12347>
- Lipovich, L., Johnson, R., & Lin, C. Y. (2010). MacroRNA underdogs in a microRNA world: Evolutionary, regulatory, and biomedical significance of mammalian long non-protein-coding RNA. *Biochimica et Biophysica Acta - Gene Regulatory Mechanisms*, 1799(9), 597–615.
<https://doi.org/10.1016/j.bbagr.2010.10.001>
- Liu, A., Yu, Q., Peng, Z., Huang, Y., Diao, S., Cheng, J., ... Hong, M. (2017). miR-200b inhibits CD133+ glioma cells by targeting the AKT pathway. *Oncology Letters*, 13(6), 4701–4707. <https://doi.org/10.3892/ol.2017.6055>
- Liu, C., Zhang, Y., She, X., Fan, L., Li, P., Feng, J., ... Wu, M. (2018). A cytoplasmic long noncoding RNA LINC00470 as a new AKT activator to mediate glioblastoma cell autophagy. *Journal of Hematology and Oncology*, 11(1). <https://doi.org/10.1186/s13045-018-0619-z>

- Liu, G., Yuan, X., Zeng, Z., Tunici, P., Ng, H., Abdulkadir, I. R., ... Yu, J. S. (2006). Analysis of gene expression and chemoresistance of CD133+ cancer stem cells in glioblastoma. *Molecular Cancer*, 5(February). <https://doi.org/10.1186/1476-4598-5-67>
- Liu, K. W., Hu, B., & Cheng, S. Y. (2011). Platelet-derived growth factor receptor alpha in Glioma: A bed seed. *Chinese Journal of Cancer*, 30(9), 590–602. <https://doi.org/10.5732/cjc.011.10236>
- Liu, Q., Tang, H., Liu, X., Liao, Y., Li, H., Zhao, Z., ... Jiang, W. (2014). MiR-200b as a prognostic factor targets multiple members of RAB family in glioma. *Medical Oncology*, 31(3). <https://doi.org/10.1007/s12032-014-0859-x>
- Liu, R., Martuza, R. L., & Rabkin, S. D. (2005). Intracarotid delivery of oncolytic HSV vector G47Δ to metastatic breast cancer in the brain. *Gene Therapy*, 12(8), 647–654. <https://doi.org/10.1038/sj.gt.3302445>
- Liu, Renbin, Varghese, S., & Rabkin, S. D. (2005). Oncolytic herpes simplex virus vector therapy of breast cancer in C3(1)/SV40 T-antigen transgenic mice. *Cancer Research*, 65(4), 1532–1540. <https://doi.org/10.1158/0008-5472.CAN-04-3353>
- Liu, T. C., Zhang, T., Fukuhara, H., Kuroda, T., Todo, T., Martuza, R. L., ... Kurtz, A. (2006). Oncolytic HSV Armed with Platelet Factor 4, an Antiangiogenic Agent, Shows Enhanced Efficacy. *Molecular Therapy*, 14(6), 789–797. <https://doi.org/10.1016/j.ymthe.2006.07.011>
- Liu, Y., Snow, B. E., Hande, M. P., Baerlocher, G., Kickhoefer, V. A., Yeung, D., ... Harrington, L. (2000). Telomerase-Associated Protein TEP1 Is Not Essential for Telomerase Activity or Telomere Length Maintenance In Vivo. *Molecular and Cellular Biology*, 20(21), 8178–8184. <https://doi.org/10.1128/mcb.20.21.8178-8184.2000>
- Liu, Zhenlin, Jiang, Z., Huang, J., Huang, S., Li, Y., Yu, S., ... Liu, X. (2014). miR-7 inhibits glioblastoma growth by simultaneously interfering with the PI3K/ATK and Raf/MEK/ERK pathways. *International Journal of Oncology*, 44(5), 1571–1580. <https://doi.org/10.3892/ijo.2014.2322>
- Liu, Zhiguo, Liu, Y., Li, L., Xu, Z., Bi, B., Wang, Y., & Li, J. Y. (2014). MiR-7-5p is frequently downregulated in glioblastoma microvasculature and inhibits vascular endothelial cell proliferation by targeting RAF1. *Tumor Biology*, 35(10), 10177–10184. <https://doi.org/10.1007/s13277-014-2318-x>
- Londina, E., Lohera, P., Telonis, A. G., Quann, K., Clark, P., Jinga, Y., ... Rigoutsos, I. (2015). Analysis of 13 cell types reveals evidence for the expression of numerous novel primate- And tissue-specific microRNAs. *Proceedings of the National Academy of Sciences of the United States of America*, 112(10), E1106–E1115. <https://doi.org/10.1073/pnas.1420955112>

- Lötsch, D., Steiner, E., Holzmann, K., Spiegl-Kreinecker, S., Pirker, C., Hlavaty, J., ... Berger, W. (2013). Major vault protein supports glioblastoma survival and migration by upregulating the EGFR/PI3K signalling axis. *Oncotarget*, 4(11), 1904–1918. <https://doi.org/10.18632/oncotarget.1264>
- Louis, D. N., Ohgaki, H., Wiestler, O. D., Cavenee, W. K., Burger, P. C., Jouvet, A., ... Kleihues, P. (2007). The 2007 WHO classification of tumours of the central nervous system. *Acta Neuropathologica*, 114(2), 97–109. <https://doi.org/10.1007/s00401-007-0243-4>
- Louis, D. N., Perry, A., Reifenberger, G., von Deimling, A., Figarella-Branger, D., Cavenee, W. K., ... Ellison, D. W. (2016). The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. *Acta Neuropathologica*, 131(6), 803–820. <https://doi.org/10.1007/s00401-016-1545-1>
- Ma, J., Wang, P., Yao, Y., Liu, Y., Li, Z., Liu, X., ... Xue, Y. (2016). Knockdown of long non-coding RNA MALAT1 increases the blood-tumor barrier permeability by up-regulating miR-140. *Biochimica et Biophysica Acta - Gene Regulatory Mechanisms*, 1859(2), 324–338. <https://doi.org/10.1016/j.bbagr.2015.11.008>
- Ma, X., Li, Z., Li, T., Zhu, L., Li, Z., & Tian, N. (2017). Long non-coding RNA HOTAIR enhances angiogenesis by induction of vegfa expression in glioma cells and transmission to endothelial cells via glioma cell derived-extracellular vesicles. *American Journal of Translational Research*, 9(11), 5012–5021.
- Machida, K., McNamara, G., Cheng, K. T.-H., Huang, J., Wang, C.-H., Comai, L., ... Lai, M. M. C. (2010). Hepatitis C Virus Inhibits DNA Damage Repair through Reactive Oxygen and Nitrogen Species and by Interfering with the ATM-NBS1/Mre11/Rad50 DNA Repair Pathway in Monocytes and Hepatocytes. *The Journal of Immunology*, 185(11), 6985–6998. <https://doi.org/10.4049/jimmunol.1000618>
- Madan, E., Dikshit, B., Gowda, S. H., Srivastava, C., Sarkar, C., Chattopadhyay, P., ... Chosdol, K. (2016). FAT1 is a novel upstream regulator of HIF1α and invasion of high grade glioma. *International Journal of Cancer*, 139(11), 2570–2582. <https://doi.org/10.1002/ijc.30386>
- Maher, E. A., Furnari, F. B., Bachoo, R. M., Rowitch, D. H., Louis, D. N., Cavenee, W. K., & DePinho, R. A. (2001). Malignant glioma: Genetics and biology of a grave matter. *Genes and Development*, 15(11), 1311–1333. <https://doi.org/10.1101/gad.891601>
- Majmundar, A. J., Wong, W. J., & Simon, M. C. (2010). Hypoxia-Inducible Factors and the Response to Hypoxic Stress. *Molecular Cell*, 40(2), 294–309. <https://doi.org/10.1016/j.molcel.2010.09.022>

- Martínez, R. (2012). Beyond Genetics in Glioma Pathways: The Ever-Increasing Crosstalk between Epigenomic and Genomic Events. *Journal of Signal Transduction*, 2012, 1–9. <https://doi.org/10.1155/2012/519807>
- Mattick, J. S., & Gagen, M. J. (2001). The evolution of controlled multitasked gene networks: The role of introns and other noncoding RNAs in the development of complex organisms. *Molecular Biology and Evolution*, 18(9), 1611–1630. <https://doi.org/10.1093/oxfordjournals.molbev.a003951>
- Medina, R. A., & Owen, G. I. (2002). Glucose transporters: Expression, regulation and cancer. *Biological Research*, 35(1), 9–26. <https://doi.org/10.4067/S0716-97602002000100004>
- Mei-Yee Kiang, K., Zhang, X. Q., & Leung, G. K. K. (2015). Long non-coding RNAs: The key players in glioma pathogenesis. *Cancers*, 7(3), 1406–1424. <https://doi.org/10.3390/cancers7030843>
- Mercer, T. R., Dinger, M. E., & Mattick, J. S. (2009). Long non-coding RNAs: Insights into functions. *Nature Reviews Genetics*, 10(3), 155–159. <https://doi.org/10.1038/nrg2521>
- Miao, F. an, Chu, K., Chen, H. rong, Zhang, M., Shi, P. cong, Bai, J., & You, Y. ping. (2019a). Increased DKC1 expression in glioma and its significance in tumor cell proliferation, migration and invasion. *Investigational New Drugs*, 37(6), 1177–1186. <https://doi.org/10.1007/s10637-019-00748-w>
- Miao, F. an, Chu, K., Chen, H. rong, Zhang, M., Shi, P. cong, Bai, J., & You, Y. ping. (2019b). Increased DKC1 expression in glioma and its significance in tumor cell proliferation, migration and invasion. *Investigational New Drugs*, 37(6), 1177–1186. <https://doi.org/10.1007/s10637-019-00748-w>
- Miyatake, S., Iyer, A., Martuza, R. L., & Rabkin, S. D. (1997). Transcriptional targeting of herpes simplex virus for cell-specific replication. *Journal of Virology*, 71(7), 5124–5132. <https://doi.org/10.1128/jvi.71.7.5124-5132.1997>
- Mizushima, N., Yoshimori, T., & Ohsumi, Y. (2011). The role of atg proteins in autophagosome formation. *Annual Review of Cell and Developmental Biology*, 27, 107–132. <https://doi.org/10.1146/annurev-cellbio-092910-154005>
- Møller, H. G., Rasmussen, A. P., Andersen, H. H., Johnsen, K. B., Henriksen, M., & Duroux, M. (2013). A systematic review of MicroRNA in glioblastoma multiforme: Micro-modulators in the mesenchymal mode of migration and invasion. *Molecular Neurobiology*, 47(1), 131–144. <https://doi.org/10.1007/s12035-012-8349-7>
- Monteiro, A., Hill, R., Pilkington, G., & Madureira, P. (2017a). The Role of Hypoxia in Glioblastoma Invasion. *Cells*, 6(4), 45. <https://doi.org/10.3390/cells6040045>

- Monteiro, A., Hill, R., Pilkington, G., & Madureira, P. (2017b). The Role of Hypoxia in Glioblastoma Invasion. *Cells*, 6(4), 45. <https://doi.org/10.3390/cells6040045>
- Mueller, W. C., & Deimling, A. Von. (2009). by Methylation. *Recent Results in Cancer Research*. <https://doi.org/10.1007/978-3-540-31206-2>
- Murnyák, B., Kouhsari, M. C., Hershkovich, R., Kálmán, B., Marko-Varga, G., Klekner, Á., & Hortobágyi, T. (2017). PARP1 expression and its correlation with survival is tumour molecular subtype dependent in glioblastoma. *Oncotarget*, 8(28), 46348–46362. <https://doi.org/10.18632/oncotarget.18013>
- Myung, J. K., Choi, S. A., Kim, S. K., Wang, K. C., & Park, S. H. (2014). Snail plays an oncogenic role in glioblastoma by promoting epithelial mesenchymal transition. *International Journal of Clinical and Experimental Pathology*, 7(5), 1977–1987.
- Nakada, M., Kita, D., Watanabe, T., Hayashi, Y., Teng, L., Pyko, I. V., & Hamada, J. I. (2011). Aberrant signaling pathways in Glioma. *Cancers*, 3(3), 3242–3278. <https://doi.org/10.3390/cancers3033242>
- Nakai, E., Park, K., Yawata, T., Chihara, T., Kumazawa, A., Nakabayashi, H., & Shimizu, K. (2009). Enhanced mdr1 expression and chemoresistance of cancer stem cells derived from glioblastoma. *Cancer Investigation*, 27(9), 901–908. <https://doi.org/10.3109/07357900801946679>
- Nazio, F., Bordi, M., Cianfanelli, V., Locatelli, F., & Cecconi, F. (2019). Autophagy and cancer stem cells: molecular mechanisms and therapeutic applications. *Cell Death and Differentiation*, 26(4), 690–702. <https://doi.org/10.1038/s41418-019-0292-y>
- nd Kelsey C. MartinMhatre V. Ho, Ji-Ann Lee, A. (2012). 基因的改变NIH Public Access. *Bone*, 23(1), 1–7. <https://doi.org/10.1038/jid.2014.371>
- Neilsen, B. K., Sleighholm, R., McComb, R., Ramkissoon, S. H., Ross, J. S., Corona, R. J., ... Aizenberg, M. R. (2019). Comprehensive genetic alteration profiling in primary and recurrent glioblastoma. *Journal of Neuro-Oncology*, 142(1), 111–118. <https://doi.org/10.1007/s11060-018-03070-2>
- Nickel, G. C., Barnholtz-Sloan, J., Gould, M. P., McMahon, S., Cohen, A., Adams, M. D., ... LaFramboise, T. (2012). Characterizing mutational heterogeneity in a glioblastoma patient with double recurrence. *PLoS ONE*, 7(4), 1–8. <https://doi.org/10.1371/journal.pone.0035262>
- Noonan, J., Zarrer, J., & Murphy, B. M. (2016). Targeting autophagy in glioblastoma. *Critical Reviews in Oncogenesis*, 21(3–4), 241–252. <https://doi.org/10.1615/CritRevOncog.2016017008>
- Ohgaki, H., & Kleihues, P. (2005). Epidemiology and etiology of gliomas. *Acta Neuropathologica*, 109(1), 93–108. <https://doi.org/10.1007/s00401-005-0991-y>

- Ohka, F., Natsume, A., & Wakabayashi, T. (2012a). Current trends in targeted therapies for glioblastoma multiforme. *Neurology Research International*, 2012. <https://doi.org/10.1155/2012/878425>
- Ohka, F., Natsume, A., & Wakabayashi, T. (2012b). Current trends in targeted therapies for glioblastoma multiforme. *Neurology Research International*, 2012(March). <https://doi.org/10.1155/2012/878425>
- Ojha, R., Bhattacharyya, S., & Singh, S. K. (2015). *Autophagy in Cancer Stem Cells : A Potential Link Between Chemosensitivity , Recurrence , and Metastasis.* 4, 97–108. <https://doi.org/10.1089/biores.2014.0035>
- Orisme, W., Punchihewa, C., Parker, M., Qaddoumi, I., Chen, X., Hedlund, E., ... Tang, C. (2013). *HHS Public Access.* 45(6), 602–612. <https://doi.org/10.1038/ng.2611.Whole-genome>
- Osada, H., Tokunaga, T., Hatanaka, H., Kawakami, T., Tsuchida, T., Abe, Y., ... Nakamura, M. (2001). Gene expression of angiogenesis related factors in glioma. *International Journal of Oncology*, 18(2), 305–309. <https://doi.org/10.3892/ijo.18.2.305>
- Ostrom, Q. T., Gittleman, H., Farah, P., Ondracek, A., Chen, Y., Wolinsky, Y., ... Barnholtz-Sloan, J. S. (2013). CBTRUS statistical report: Primary brain and central nervous system tumors diagnosed in the United States in 2006–2010. *Neuro-Oncology*, 15(SUPPL.2). <https://doi.org/10.1093/neuonc/not151>
- Ostrom, Q. T., Gittleman, H., Fulop, J., Liu, M., Blanda, R., Kromer, C., ... Barnholtz-Sloan, J. S. (2015). CBTRUS statistical Report: primary brain and central nervous system tumors diagnosed in the United States in 2008–2012. *Neuro-Oncology*, 17, iv1–iv62. <https://doi.org/10.1093/neuonc/nov189>
- Palm, W., & de Lange, T. (2008). How Shelterin Protects Mammalian Telomeres. *Annual Review of Genetics*, 42(1), 301–334. <https://doi.org/10.1146/annurev.genet.41.110306.130350>
- Park, J. W., Wollmann, G., Urbiola, C., Fogli, B., Florio, T., Geley, S., & Klimaszewski, L. (2018). Sprouty2 enhances the tumorigenic potential of glioblastoma cells. *Neuro-Oncology*, 20(8), 1044–1054. <https://doi.org/10.1093/neuonc/noy028>
- Pastori, C., Kapranov, P., Penas, C., Peschansky, V., Volmar, C. H., Sarkaria, J. N., ... Roberts, T. C. (2015). The bromodomain protein BRD4 controls HOTAIR, a long noncoding RNA essential for glioblastoma proliferation. *Proceedings of the National Academy of Sciences of the United States of America*, 112(27), 8326–8331. <https://doi.org/10.1073/pnas.1424220112>
- Peng, L., Fu, J., & Ming, Y. (2018). The miR-200 family: Multiple effects on gliomas. *Cancer Management and Research*, 10, 1987–1992. <https://doi.org/10.2147/CMAR.S160945>

- Pengse Po1, Erin Delaney1, Howard Gamper2, Miklos Szanti-Kis3, Lee Speight3, LiWei Tu1, Andrey Kosolapov1, E. James Petersson3, Ya-Ming Hou2, and C. D. (2017). 乳鼠心肌提取 HHS Public Access. *Physiology & Behavior*, 176(12), 139–148.
<https://doi.org/10.1016/j.physbeh.2017.03.040>
- Piao, Y., Liang, J., Holmes, L., Zurita, A. J., Henry, V., Heymach, J. V., & De Groot, J. F. (2012). Glioblastoma resistance to anti-VEGF therapy is associated with myeloid cell infiltration, stem cell accumulation, and a mesenchymal phenotype. *Neuro-Oncology*, 14(11), 1379–1392.
<https://doi.org/10.1093/neuonc/nos158>
- Picariello, L. (2014). Telomerase activity: An attractive target for cancer therapeutics. *World Journal of Pharmacology*, 3(4), 86.
<https://doi.org/10.5497/wjp.v3.i4.86>
- Piccirillo, S. G. M., Binda, E., Fiocco, R., Vescovi, A. L., & Shah, K. (2009). Brain cancer stem cells. *Journal of Molecular Medicine*, 87(11), 1087–1095.
<https://doi.org/10.1007/s00109-009-0535-3>
- Plate, K. H., & Risau, W. (1995). Angiogenesis in malignant gliomas. *Glia*, 15(3), 339–347. <https://doi.org/10.1002/glia.440150313>
- Qi, P., & Du, X. (2013). The long non-coding RNAs, a new cancer diagnostic and therapeutic gold mine. *Modern Pathology*, 26(2), 155–165.
<https://doi.org/10.1038/modpathol.2012.160>
- Qiu, S., Lin, S., Hu, D., Feng, Y., Tan, Y., & Peng, Y. (2013). Interactions of miR-323/miR-326/miR-329 and miR-130a/miR-155/miR-210 as prognostic indicators for clinical outcome of glioblastoma patients. *Journal of Translational Medicine*, 11(1), 1–11. <https://doi.org/10.1186/1479-5876-11-10>
- Quintavalle, C., Garofalo, M., Zanca, C., Romano, G., Iaboni, M., Del Basso De Caro, M., ... Condorelli, G. (2012). MiR-221/222 overexpression in human glioblastoma increases invasiveness by targeting the protein phosphate PTP. *Oncogene*, 31(7), 858–868. <https://doi.org/10.1038/onc.2011.280>
- Ravi, R., Mookerjee, B., Bhujwalla, Z. M., Sutter, C. H., Artemov, D., Zeng, Q., ... Bedi, A. (2000). Regulation of tumor angiogenesis by p53-induced degradation of hypoxia- inducible factor 1α. *Genes and Development*, 14(1), 34–44. <https://doi.org/10.1101/gad.14.1.34>
- Regazzo, G., Terrenato, I., Spagnuolo, M., Carosi, M., Cognetti, G., Cicchillitti, L., ... Rizzo, M. G. (2016). A restricted signature of serum miRNAs distinguishes glioblastoma from lower grade gliomas. *Journal of Experimental and Clinical Cancer Research*, 35(1), 1–11.
<https://doi.org/10.1186/s13046-016-0393-0>

- Remondelli, P., & Renna, M. (2017). *The Endoplasmic Reticulum Unfolded Protein Response in Neurodegenerative Disorders and Its Potential Therapeutic Significance*. 10(June), 1–16.
<https://doi.org/10.3389/fnmol.2017.00187>
- Roberto Zoncu, David M. Sabatini, and A. E. (2011). NIH Public Access Nat Rev Mol Cell Biol mTOR: from growth signal integration to cancer, diabetes and Nat Rev Mol Cell Biol Author Manuscript Biol mTOR: from growth signal integration to cancer, diabetes and Nat Rev Mol Cell Biol Author Manuscript . Author. *Nature Reviews. Molecular Cell Biology*, 12(1), 21–35. <https://doi.org/10.1038/nrm3025.mTOR>
- Rousseau, A., Mokhtari, K., & Duyckaerts, C. (2008). The 2007 WHO classification of tumors of the central nervous system - What has changed? *Current Opinion in Neurology*, 21(6), 720–727.
<https://doi.org/10.1097/WCO.0b013e328312c3a7>
- Rynkeviciene, R., Simiene, J., Strainiene, E., Stankevicius, V., Usinskiene, J., Kaubriene, E. M., ... Suziedelis, K. (2019). Non-coding RNAs in glioma. *Cancers*, 11(1), 1–35. <https://doi.org/10.3390/cancers11010017>
- Ryskalin, L., Gaglione, A., Limanaqi, F., Biagioli, F., Familiari, P., Frati, A., ... Fornai, F. (2019). The autophagy status of cancer stem cells in glioblastoma multiforme: From cancer promotion to therapeutic strategies. *International Journal of Molecular Sciences*, 20(15). <https://doi.org/10.3390/ijms20153824>
- Sa, L., Li, Y., Zhao, L., Liu, Y., Wang, P., Liu, L., ... Xue, Y. (2017). The role of HOTAIR/miR-148b-3p/USF1 on regulating the permeability of BTB. *Frontiers in Molecular Neuroscience*, 10(June), 1–17. <https://doi.org/10.3389/fnmol.2017.00194>
- Sampson, V. B., Rong, N. H., Han, J., Yang, Q., Aris, V., Soteropoulos, P., ... Krueger, L. J. (2007). MicroRNA let-7a down-regulates MYC and reverts MYC-induced growth in Burkitt lymphoma cells. *Cancer Research*, 67(20), 9762–9770. <https://doi.org/10.1158/0008-5472.CAN-07-2462>
- Sana, J., Faltejskova, P., Svoboda, M., & Slaby, O. (2012). Novel classes of non-coding RNAs and cancer. *Journal of Translational Medicine*, 10(1), 1–21. <https://doi.org/10.1186/1479-5876-10-103>
- Sarafian, V. S., Koev, I., Mehterov, N., Kazakova, M., & Dangalov, K. (2018). LAMP-1 gene is overexpressed in high grade glioma. *Apmis*, 126(8), 657–662. <https://doi.org/10.1111/apm.12856>
- Schmalz, P. G. R., Shen, M. J., & Park, J. K. (2011). Treatment resistance mechanisms of malignant glioma tumor stem cells. *Cancers*, 3(1), 621–635. <https://doi.org/10.3390/cancers3010621>
- Schmeisser, H., Bekisz, J., & Zoon, K. C. (2014). New function of Type I IFN: Induction of autophagy. *Journal of Interferon and Cytokine Research*, 34(2), 71–78. <https://doi.org/10.1089/jir.2013.0128>

- Schraivogel, D., Weinmann, L., Beier, D., Tabatabai, G., Eichner, A., Zhu, J. Y., ... Meister, G. (2011). CAMTA1 is a novel tumour suppressor regulated by miR-9/9 * in glioblastoma stem cells. *EMBO Journal*, 30(20), 4309–4322. <https://doi.org/10.1038/emboj.2011.301>
- Schwartzbaum, J. A., Fisher, J. L., Aldape, K. D., & Wrensch, M. (2006). Epidemiology and molecular pathology of glioma. *Nature Clinical Practice Neurology*, 2(9), 494–503. <https://doi.org/10.1038/ncpneuro0289>
- Senft, D., & Ronai, Z. A. (2016). Regulators of mitochondrial dynamics in cancer. *Current Opinion in Cell Biology*, 39, 43–52. <https://doi.org/10.1016/j.ceb.2016.02.001>
- Sgubin, D., Wakimoto, H., Kanai, R., Rabkin, S. D., & Martuza, R. L. (2012). Oncolytic Herpes Simplex Virus Counteracts the Hypoxia-Induced Modulation of Glioblastoma Stem-Like Cells. *STEM CELLS TRANSLATIONAL MEDICINE*, 1(4), 322–332. <https://doi.org/10.5966/sctm.2011-0035>
- Shang, C., Guo, Y., Hong, Y., & Xue, Y. X. (2016). Long non-coding RNA TUSC7, a target of miR-23b, plays tumor-suppressing roles in human gliomas. *Frontiers in Cellular Neuroscience*, 10(OCT2016), 1–9. <https://doi.org/10.3389/fncel.2016.00235>
- Shea, A., Harish, V., Afzal, Z., Chijioke, J., Kedir, H., Dusmatova, S., ... Kumar, D. (2016a). MicroRNAs in glioblastoma multiforme pathogenesis and therapeutics. *Cancer Medicine*, 5(8), 1917–1946. <https://doi.org/10.1002/cam4.775>
- Shea, A., Harish, V., Afzal, Z., Chijioke, J., Kedir, H., Dusmatova, S., ... Kumar, D. (2016b). MicroRNAs in glioblastoma multiforme pathogenesis and therapeutics. *Cancer Medicine*, 5(8), 1917–1946. <https://doi.org/10.1002/cam4.775>
- Shervington, A., Patel, R., Lu, C., Cruickshanks, N., Lea, R., Roberts, G., ... Shervington, L. (2007). Telomerase subunits expression variation between biopsy samples and cell lines derived from malignant glioma. *Brain Research*, 1134(1), 45–52. <https://doi.org/10.1016/j.brainres.2006.11.093>
- Silber, J., Lim, D. A., Petritsch, C., Persson, A. I., Maunakea, A. K., Yu, M., ... Hodgson, J. G. (2008). miR-124 and miR-137 inhibit proliferation of glioblastoma multiforme cells and induce differentiation of brain tumor stem cells. *BMC Medicine*, 6, 1–17. <https://doi.org/10.1186/1741-7015-6-14>
- Simon, M. C. (2010). Abstract SY34-02: The impact of O₂ availability on human cancer. 8(12), SY34-02-SY34-02. <https://doi.org/10.1158/1538-7445.am10-sy34-02>
- Singh, R., & Cuervo, A. M. (2011). Autophagy in the cellular energetic balance. *Cell Metabolism*, 13(5), 495–504. <https://doi.org/10.1016/j.cmet.2011.04.004>

- Skalsky, R. L., & Cullen, B. R. (2011). Reduced expression of brain-enriched microRNAs in glioblastomas permits targeted regulation of a cell death gene. *PLoS ONE*, 6(9). <https://doi.org/10.1371/journal.pone.0024248>
- Smith, C., & Ironside, J. W. (2007). Diagnosis and pathogenesis of gliomas. *Current Diagnostic Pathology*, 13(3), 180–192. <https://doi.org/10.1016/j.cdip.2007.04.002>
- Smogorzewska, A., & De Lange, T. (2002). Different telomere damage signaling pathways in human and mouse cells. *EMBO Journal*, 21(16), 4338–4348. <https://doi.org/10.1093/emboj/cdf433>
- Song, J., Ouyang, Y., Che, J., Li, X., Zhao, Y., Yang, K., ... Yuan, W. (2017). Potential value of miR-221/222 as diagnostic, prognostic, and therapeutic biomarkers for diseases. *Frontiers in Immunology*, 8(FEB), 1–9. <https://doi.org/10.3389/fimmu.2017.00056>
- Srinivasan, S., Patric, I. R. P., & Somasundaram, K. (2011). A Ten-microRNA expression signature predicts survival in Glioblastoma. *PLoS ONE*, 6(3). <https://doi.org/10.1371/journal.pone.0017438>
- Stahlhut, C., & Slack, F. J. (2013). MicroRNAs and the cancer phenotype: profiling, signatures and clinical implications. *Genome Medicine*, 5(12), 111. <https://doi.org/10.1186/gm516>
- Steiner, E., Holzmann, K., Elbling, L., Micksche, M., & Berger, W. (2006). Cellular Functions of Vaults and their Involvement in Multidrug Resistance. *Current Drug Targets*, 7(8), 923–934. <https://doi.org/10.2174/138945006778019345>
- Stopschinski, B. E., Beier, C. P., & Beier, D. (2013). Glioblastoma cancer stem cells - From concept to clinical application. *Cancer Letters*, 338(1), 32–40. <https://doi.org/10.1016/j.canlet.2012.05.033>
- Stucki, M., Clapperton, J. A., Mohammad, D., Yaffe, M. B., Smerdon, S. J., Jackson, S. P., ... Carpenter, P. B. (2016). Structure of Human DROSHA. *Cell*, 164(1), 81–90. <https://doi.org/10.1101/gad.1262504.mic>
- Sun, C., Wang, Z., Song, W., Chen, B., Zhang, J., Dai, X., ... Dong, J. (2015). Alteration of DNA damage signaling pathway profile in radiation-treated glioblastoma stem-like cells. *Oncology Letters*, 10(3), 1769–1774. <https://doi.org/10.3892/ol.2015.3411>
- Sun, G., Cao, Y., Shi, L., Sun, L., Wang, Y., Chen, C., ... You, Y. (2013). Overexpressed miRNA-137 inhibits human glioma cells growth by targeting Rac1. *Cancer Biotherapy and Radiopharmaceuticals*, 28(4), 327–334. <https://doi.org/10.1089/cbr.2012.1380>
- Suvà, M. L., Rheinbay, E., Gillespie, S. M., Patel, A. P., Wakimoto, H., Rabkin, S. D., ... Bernstein, B. E. (2014). Reconstructing and reprogramming the tumor-propagating potential of glioblastoma stem-like cells. *Cell*, 157(3),

580–594. <https://doi.org/10.1016/j.cell.2014.02.030>

Suzuki, K., Kawataki, T., Endo, K., Miyazawa, K., Kinouchi, H., & Saitoh, M. (2018). Expression of zebs in gliomas is associated with invasive properties and histopathological grade. *Oncology Letters*, 16(2), 1758–1764. <https://doi.org/10.3892/ol.2018.8852>

Szabo, E., Schneider, H., Seystahl, K., Rushing, E. J., Herting, F., Weidner, K. M., & Weller, M. (2016). Autocrine VEGFR1 and VEGFR2 signaling promotes survival in human glioblastoma models in vitro and in vivo. *Neuro-Oncology*, 18(9), 1242–1252.
<https://doi.org/10.1093/neuonc/now043>

Tabori, U., Vukovic, B., Zielenska, M., Hawkins, C., Braude, I., Rutka, J., ... Malkin, D. (2006). The role of telomere maintenance in the spontaneous growth arrest of pediatric low-grade gliomas. *Neoplasia*, 8(2), 136–142. <https://doi.org/10.1593/neo.05715>

Tamim, S., Vo, D. T., Uren, P. J., Qiao, M., Bindewald, E., Kasprzak, W. K., ... Penalva, L. O. F. (2014). Genomic analyses reveal broad impact of miR-137 on genes associated with malignant transformation and neuronal differentiation in glioblastoma cells. *PLoS ONE*, 9(1). <https://doi.org/10.1371/journal.pone.0085591>

Targets, T. (2015). *Glioblastoma: Molecular Pathways, Stem Cells and Therapeutic Targets*. 538–555. <https://doi.org/10.3390/cancers7020538>

Tate, M. C., & Aghi, M. K. (2009). Biology of Angiogenesis and Invasion in Glioma. *Neurotherapeutics*, 6(3), 447–457.
<https://doi.org/10.1016/j.nurt.2009.04.001>

Teodorczyk, M., & Martin-Villalba, A. (2010). Sensing invasion: Cell surface receptors driving spreading of glioblastoma. *Journal of Cellular Physiology*, 222(1), 1–10. <https://doi.org/10.1002/jcp.21901>

Thakkar, J. P., Dolecek, T. A., Horbinski, C., Ostrom, Q. T., Lightner, D. D., Barnholtz-Sloan, J. S., & Villano, J. L. (2014). Epidemiologic and molecular prognostic review of glioblastoma. *Cancer Epidemiology Biomarkers and Prevention*, 23(10), 1985–1996. <https://doi.org/10.1158/1055-9965.EPI-14-0275>

Thornton, B., & Basu, C. (2011). Real-time PCR (qPCR) primer design using free online software. *Biochemistry and Molecular Biology Education*, 39(2), 145–154. <https://doi.org/10.1002/bmb.20461>

Todo, T., Martuza, R. L., Rabkin, S. D., & Johnson, P. A. (2001). Oncolytic herpes simplex virus vector with enhanced MHC class I presentation and tumor cell killing. *Proceedings of the National Academy of Sciences of the United States of America*, 98(11), 6396–6401. <https://doi.org/10.1073/pnas.101136398>

- Tu, Y., & Liu, N. (2015). Systematic Review of MicroRNAs and its Therapeutic Potential in Glioma. *Cancer Translational Medicine*, 1(2), 50. <https://doi.org/10.4103/2395-3977.155924>
- Tunici, P., Bissola, L., Lualdi, E., Pollo, B., Cajola, L., Broggi, G., ... Finocchiaro, G. (2004). Genetic alterations and in vivo tumorigenicity of neurospheres derived from an adult glioblastoma. *Molecular Cancer*, 3, 1–5. <https://doi.org/10.1186/1476-4598-3-25>
- Valente, V., Teixeira, S. A., Neder, L., Okamoto, O. K., Oba-Shinjo, S. M., Marie, S. K. N., ... Carlotti, C. G. (2009). Selection of suitable housekeeping genes for expression analysis in glioblastoma using quantitative RT-PCR. *BMC Molecular Biology*, 10, 1–11. <https://doi.org/10.1186/1471-2199-10-17>
- Van Den Bent, M. J., Snijders, T. J., & Bromberg, J. E. C. (2012). Current treatment of low grade gliomas. *Memo - Magazine of European Medical Oncology*, 5(3), 223–227. <https://doi.org/10.1007/s12254-012-0014-3>
- Vara-Perez, M., Felipe-Abrio, B., & Agostinis, P. (2019). Mitophagy in Cancer: A Tale of Adaptation. *Cells*, 8(5), 493. <https://doi.org/10.3390/cells8050493>
- Varghese, S., Rabkin, S. D., Liu, R., Nielsen, P. G., Ipe, T., & Martuza, R. L. (2006). Enhanced therapeutic efficacy of IL-12, but not GM-CSF, expressing oncolytic herpes simplex virus for transgenic mouse derived prostate cancers. *Cancer Gene Therapy*, 13(3), 253–265. <https://doi.org/10.1038/sj.cgt.7700900>
- Vaupel, P., & Mayer, A. (2007). Hypoxia in cancer: Significance and impact on clinical outcome. *Cancer and Metastasis Reviews*, 26(2), 225–239. <https://doi.org/10.1007/s10555-007-9055-1>
- Vella, M. C., Choi, E. Y., Lin, S. Y., Reinert, K., & Slack, F. J. (2004). The *C. elegans* microRNA let-7 binds to imperfect let-7 complementary sites from the lin-41 3'UTR. *Genes and Development*, 18(2), 132–137. <https://doi.org/10.1101/gad.1165404>
- Verheul, H. M. W., & Pinedo, H. M. (2003). Vascular endothelial growth factor and its inhibitors. *Drugs of Today*, 39(SUPPL. C), 81–93.
- Vescovi, A. L., Galli, R., & Reynolds, B. A. (2006). Brain tumour stem cells. *Nature Reviews Cancer*, 6(6), 425–436. <https://doi.org/10.1038/nrc1889>
- Vessoni, A. T., Muotri, A. R., & Okamoto, O. K. (2012). Autophagy in stem cell maintenance and differentiation. *Stem Cells and Development*, 21(4), 513–520. <https://doi.org/10.1089/scd.2011.0526>
- Visani, M., de Biase, D., Marucci, G., Cerasoli, S., Nigrisoli, E., Bacchi Reggiani, M. L., ... Nobile, C. (2014). Expression of 19 microRNAs in glioblastoma and comparison with other brain neoplasia of grades I-III. *Molecular Oncology*, 8(2), 417–430. <https://doi.org/10.1016/j.molonc.2013.12.010>

- Visani, M., de Biase, D., Marucci, G., Taccioli, C., Baruzzi, A., Pessione, A., ... Nobile, C. (2013). Definition of miRNAs Expression Profile in Glioblastoma Samples: The Relevance of Non-Neoplastic Brain Reference. *PLoS ONE*, 8(1), 3–8. <https://doi.org/10.1371/journal.pone.0055314>
- Vlahopoulos, S. A., Logothetis, S., Mikas, D., Giarika, A., Gorgoulis, V., & Zoumpourlis, V. (2008). The role of ATF-2 in oncogenesis. *BioEssays*, 30(4), 314–327. <https://doi.org/10.1002/bies.20734>
- Wan, Y. Y., Zhang, J. F., Yang, Z. J., Jiang, L. P., Wei, Y. F., Lai, Q. N., ... Han, X. J. (2014). Involvement of Drp1 in hypoxia-induced migration of human glioblastoma U251 cells. *Oncology Reports*, 32(2), 619–626. <https://doi.org/10.3892/or.2014.3235>
- Wang, B., Li, M., Wu, Z., Li, X., Li, Y., Shi, X., & Cheng, W. (2015). Associations between SOX2 and miR-200b expression with the clinicopathological characteristics and prognosis of patients with glioma. *Experimental and Therapeutic Medicine*, 10(1), 88–96. <https://doi.org/10.3892/etm.2015.2488>
- Wang, G., Li, Z., Tian, N., Han, L., Fu, Y., Guo, Z., & Tian, Y. (2016). MiR-148b-3p inhibits malignant biological behaviors of human glioma cells induced by high HOTAIR expression. *Oncology Letters*, 12(2), 879–886. <https://doi.org/10.3892/ol.2016.4743>
- Wang, J. N., Xu, L. H., Zeng, W. G., Hu, P., Rabkin, S. D., & Liu, R. R. (2015). Treatment of human thyroid carcinoma cells with the G47delta oncolytic herpes simplex virus. *Asian Pacific Journal of Cancer Prevention*, 16(3), 1241–1245. <https://doi.org/10.7314/APJCP.2015.16.3.1241>
- Wang, J., Xu, L., Zeng, W., Hu, P., Zeng, M., Rabkin, S. D., & Liu, R. (2014). Treatment of human hepatocellular carcinoma by the oncolytic herpes simplex virus G47delta. *Cancer Cell International*, 14(1), 1–9. <https://doi.org/10.1186/s12935-014-0083-y>
- Wang, X. R., Luo, H., Li, H. L., Cao, L., Wang, X. F., Yan, W., ... You, Y. P. (2013). Overexpressed let-7a inhibits glioma cell malignancy by directly targeting k-ras, independently of pten. *Neuro-Oncology*, 15(11), 1491–1501. <https://doi.org/10.1093/neuonc/not107>
- Warde-farley, D., Donaldson, S. L., Comes, O., Zuberi, K., Badrawi, R., Chao, P., ... Morris, Q. (2010). *The GeneMANIA prediction server: biological network integration for gene prioritization and predicting gene function*. 38, 214–220. <https://doi.org/10.1093/nar/gkq537>
- Warde-farley, D., Donaldson, S. L., Comes, O., Zuberi, K., Lopes, C. T., Maitland, A., ... Montojo, J. (2010). *The GeneMANIA prediction server: biological network integration for gene prioritization and predicting gene function*. (May 2014). <https://doi.org/10.1093/nar/gkq537>
- Watson, G., Ronai, Z., & Lau, E. (2018). *Factors in Biology and Disease*. (813), 347–357. <https://doi.org/10.1016/j.phrs.2017.02.004.ATF2>

- Wen, P. Y., & Kesari, S. (2008). Malignant gliomas in adults. *New England Journal of Medicine*, 359(5), 492–507.
<https://doi.org/10.1056/NEJMra0708126>
- Winter, J., Jung, S., Keller, S., Gregory, R. I., & Diederichs, S. (2009). Many roads to maturity: MicroRNA biogenesis pathways and their regulation. *Nature Cell Biology*, 11(3), 228–234. <https://doi.org/10.1038/ncb0309-228>
- Wojtowicz, K., Januchowski, R., Nowicki, M., & Zabel, M. (2017). VPARP adjusts MVP expression in drug-resistant cell lines in conjunction with MDR proteins. *Anticancer Research*, 37(6), 3015–3023.
<https://doi.org/10.21873/anticanres.11656>
- Wollmann, G., Ozduman, K., & Van Den Pol, A. N. (2012a). Oncolytic virus therapy for glioblastoma multiforme: Concepts and candidates. *Cancer Journal*, 18(1), 69–81. <https://doi.org/10.1097/PPO.0b013e31824671c9>
- Wollmann, G., Ozduman, K., & Van Den Pol, A. N. (2012b). Oncolytic virus therapy for glioblastoma multiforme: Concepts and candidates. *Cancer Journal*, 18(1), 69–81. <https://doi.org/10.1097/PPO.0b013e31824671c9>
- Wright, W. E., & Shay, J. W. (1995). Time, telomeres and tumours: is cellular senescence more than an anticancer mechanism? *Trends in Cell Biology*, 5(8), 293–297. [https://doi.org/10.1016/S0962-8924\(00\)89044-3](https://doi.org/10.1016/S0962-8924(00)89044-3)
- Wright, W. E., & Shay, J. W. (2005). Telomere biology in aging and cancer. *Journal of the American Geriatrics Society*, 53(9 SUPPL.), 292–294.
<https://doi.org/10.1111/j.1532-5415.2005.53492.x>
- Wu, D. G., Wang, Y. Y., Fan, L. G., Luo, H., Han, B., Sun, L. H., ... Liu, N. (2011). MicroRNA-7 regulates glioblastoma cell invasion via targeting focal adhesion kinase expression. *Chinese Medical Journal*, 124(17), 2616–2621. <https://doi.org/10.3760/cma.j.issn.0366-6999.2011.17.010>
- Xiang, J., Guo, S., Jiang, S., Xu, Y., Li, J., Li, L., & Xiang, J. (2016). Silencing of long non-coding RNA MALAT1 promotes apoptosis of glioma cells. *Journal of Korean Medical Science*, 31(5), 688–694.
<https://doi.org/10.3346/jkms.2016.31.5.688>
- Xie, Q., Yan, Y., Huang, Z., Zhong, X., & Huang, L. (2014). MicroRNA-221 targeting PI3-K/Akt signaling axis induces cell proliferation and BCNU resistance in human glioblastoma. *Neuropathology*, 34(5), 455–464.
<https://doi.org/10.1111/neup.12129>
- Yan, C., & Li, T. S. (2018). Dual role of mitophagy in cancer drug resistance. *Anticancer Research*, 38(2), 617–621.
<https://doi.org/10.21873/anticanres.12266>
- Yang, B., Wei, Z. Y., Wang, B. Q., Yang, H. C., Wang, J. Y., & Bu, X. Y. (2018). Down-regulation of the long noncoding RNA-HOX transcript antisense intergenic RNA inhibits the occurrence and progression of glioma. *Journal of Cellular Biochemistry*, 119(2), 2278–2287.

<https://doi.org/10.1002/jcb.26390>

- Yang, H. W., Menon, L. G., Black, P. M., Carroll, R. S., & Johnson, M. D. (2010). SNAI2/Slug promotes growth and invasion in human gliomas. *BMC Cancer*, 10. <https://doi.org/10.1186/1471-2407-10-301>
- Yang, X., Xiao, Z., Du, X., Huang, L., & Du, G. (2017). Silencing of the long non-coding RNA NEAT1 suppresses glioma stem-like properties through modulation of the miR-107/CDK6 pathway. *Oncology Reports*, 37(1), 555–562. <https://doi.org/10.3892/or.2016.5266>
- Yang, Z., & Klionsky, D. J. (2010). Eaten alive: A history of macroautophagy. *Nature Cell Biology*, 12(9), 814–822. <https://doi.org/10.1038/ncb0910-814>
- Yao, N., Wang, C., Hu, N., Li, Y., Liu, M., Lei, Y., ... Zhang, D. (2019). Inhibition of PINK1/Parkin-dependent mitophagy sensitizes multidrug-resistant cancer cells to B5G1, a new betulinic acid analog. *Cell Death and Disease*, 10(3). <https://doi.org/10.1038/s41419-019-1470-z>
- Yao, Y., Ma, J., Xue, Y., Wang, P., Li, Z., Liu, J., ... Liu, Y. (2015). Knockdown of long non-coding RNA XIST exerts tumor-suppressive functions in human glioblastoma stem cells by up-regulating miR-152. *Cancer Letters*, 359(1), 75–86. <https://doi.org/10.1016/j.canlet.2014.12.051>
- Ye, J. Z. S., Donigian, J. R., Van Overbeek, M., Loayza, D., Luo, Y., Krutchinsky, A. N., ... De Lange, T. (2004). TIN2 binds TRF1 and TRF2 simultaneously and stabilizes the TRF2 complex on telomeres. *Journal of Biological Chemistry*, 279(45), 47264–47271. <https://doi.org/10.1074/jbc.M409047200>
- Yi, R., Qin, Y., Macara, I. G., & Cullen, B. R. (2003). Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. *Genes and Development*, 17(24), 3011–3016. <https://doi.org/10.1101/gad.1158803>
- Yin, C. Y., Kong, W. E. I., Jiang, J., Xu, H. A. O., & Zhao, W. E. I. (2019). miR-7-5p inhibits cell migration and invasion in glioblastoma through targeting SATB1. *Oncology Letters*, 17(2), 1819–1825. <https://doi.org/10.3892/ol.2018.9777>
- Youle, R. J., & Narendra, D. P. (2011). Mechanisms of mitophagy. *Nature Reviews Molecular Cell Biology*, 12(1), 9–14. <https://doi.org/10.1038/nrm3028>
- Yu, H., Xue, Y., Wang, P., Liu, X., Ma, J., Zheng, J., ... Liu, Y. (2017). Knockdown of long non-coding RNA XIST increases blood-tumor barrier permeability and inhibits glioma angiogenesis by targeting miR-137. *Oncogenesis*, 6(3). <https://doi.org/10.1038/oncsis.2017.7>
- Yuan, X., Curtin, J., Xiong, Y., Liu, G., Waschsmann-Hogiu, S., Farkas, D. L., ... Yu, J. S. (2004). Isolation of cancer stem cells from adult glioblastoma multiforme. *Oncogene*, 23(58), 9392–9400. <https://doi.org/10.1038/sj.onc.1208311>

- Zeng, W., Hu, P., Wu, J., Wang, J., Li, J., Lei, L., & Liu, R. (2013). The oncolytic herpes simplex virus vector G47Δ effectively targets breast cancer stem cells. *Oncology Reports*, 29(3), 1108–1114. <https://doi.org/10.3892/or.2012.2211>
- Zhang, C. Z., Zhang, J. X., Zhang, A. L., Shi, Z. D., Han, L., Jia, Z. F., ... Kang, C. S. (2010). MiR-221 and miR-222 target PUMA to induce cell survival in glioblastoma. *Molecular Cancer*, 9, 1–9. <https://doi.org/10.1186/1476-4598-9-229>
- Zhang, C., Zhang, J., Hao, J., Shi, Z., Wang, Y., Han, L., ... Kang, C. (2012). High level of miR-221/222 confers increased cell invasion and poor prognosis in glioma. *Journal of Translational Medicine*, 10(1). <https://doi.org/10.1186/1479-5876-10-119>
- Zhang, T., Wang, Y.-R., Zeng, F., Cao, H.-Y., Zhou, H.-D., & Wang, Y.-J. (2016). *LncRNA H19 is overexpressed in glioma tissue, is negatively associated with patient survival.* 4891–4897. Retrieved from <https://www.europeanreview.org/wp/wp-content/uploads/4891-4897-LncRNA-H19-is-overexpressed-in-glioma-tissue-is-negatively-associated-with-patient-survival.pdf>
- Zhang, X., Kiang, K. M., Zhang, G. P., & Leung, G. K. (2015). Long non-coding RNAs dysregulation and function in glioblastoma stem cells. *Non-Coding RNA*, 1(1), 69–86. <https://doi.org/10.3390/ncrna1010069>
- Zhang, X. Q., Sun, S., Lam, K. F., Kiang, K. M. Y., Pu, J. K. S., Ho, A. S. W., ... Leung, G. K. K. (2013). A long non-coding RNA signature in glioblastoma multiforme predicts survival. *Neurobiology of Disease*, 58, 123–131. <https://doi.org/10.1016/j.nbd.2013.05.011>
- Zhang, X., Sun, S., Pu, J. K. S., Tsang, A. C. O., Lee, D., Man, V. O. Y., ... Leung, G. K. K. (2012). Long non-coding RNA expression profiles predict clinical phenotypes in glioma. *Neurobiology of Disease*, 48(1), 1–8. <https://doi.org/10.1016/j.nbd.2012.06.004>
- Zhen, L., Yun-hui, L., Hong-yu, D., Jun, M., & Yi-long, Y. (2016). Long noncoding RNA NEAT1 promotes glioma pathogenesis by regulating miR-449b-5p/c-Met axis. *Tumor Biology*, 37(1), 673–683. <https://doi.org/10.1007/s13277-015-3843-y>
- Zhong, H., Chiles, K., Feldser, D., Laughner, E., Hanrahan, C., Georgescu, M. M., ... Semenza, G. L. (2000). Modulation of hypoxia-inducible factor 1α expression by the epidermal growth factor/phosphatidylinositol 3-kinase/PTEN/AKT/FRAP pathway in human prostate cancer cells: Implications for tumor angiogenesis and therapeutics. *Cancer Research*, 60(6), 1541–1545.
- Zhou, J., Xiang, W., Li, S., Hu, Q., Peng, T., Chen, L., & Ming, Y. (2018). Association between long non-coding RNAs expression and pathogenesis and progression of gliomas (review). *Oncology Letters*, 15(4), 4070–4078.

<https://doi.org/10.3892/ol.2018.7875>

Zhou, W., & Wahl, D. R. (2019). Metabolic abnormalities in glioblastoma and metabolic strategies to overcome treatment resistance. *Cancers*, 11(9). <https://doi.org/10.3390/cancers11091231>

Zong, M. Z., Feng, W. T., Du, N., Yu, X. J., & Yu, W. Y. (2019). Upregulation of long noncoding RNA LEF1-AS1 predicts a poor prognosis in patients with esophageal squamous cell carcinoma. *European Review for Medical and Pharmacological Sciences*, 23(18), 7929–7934. https://doi.org/10.26355/eurrev_201909_19007