



**EVALUATION OF PANCREATIC CANCER CELL LINES AS A SOURCE OF
CANCER STEM CELLS (CSC) AND THEIR UTILISATION FOR ASSESSING
ANTI-CSC ACTIVITIES OF KRAS BINDERS**

By

TEH YUAN HAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

EVALUATION OF PANCREATIC CANCER CELL LINES AS A SOURCE OF CANCER STEM CELLS (CSC) AND THEIR UTILISATION FOR ASSESSING ANTI-CSC ACTIVITIES OF KRAS BINDERS

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October 2020

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Background: High incidence of recurrence has been recorded at 80% of pancreatic ductal adenocarcinoma (PDAC) cases, despite tumour resection and an adjuvant chemotherapy with gemcitabine are prescribed. Pancreatic cancer stem cells (CSCs) are believed to have initiated recurrent PDAC. Growing evidence indicates that these cells resist gemcitabine as a result of aberrant activation of KRAS, a molecular switch that regulates cancer-associated signalling pathways, including MAPK and PI3K-AKT. Vismodegib is the earliest known anti-pancreatic CSC agent, but its anti-CSC activity has been proven to be not translatable from bench to clinical settings. It is crucial to seek for an alternative to vismodegib. Patient-derived CSC culture is prevalently used for assessing potential anti-CSC agents *in vitro*, but patients' biopsies may not be accessible to some researchers. **Objectives:** In the present study, PDAC cell lines were evaluated as a source of CSCs and they were utilised for assessing the anti-CSC potential of KRAS binders (DCAI and SRJ23) with their mechanisms of action being elucidated. **Methods:** Cell viability assays were performed to evaluate the cytotoxicity of gemcitabine, vismodegib, and KRAS binders in PDAC cell lines (BxPC-3, PANC-1, Capan-2, and MIA PaCa-2). Flow cytometry was employed to analyse the expression of pancreatic CSC surface markers (CD44, CD24, and CD133) on a single-cell basis. While tumoursphere-forming ability was investigated by culturing cells in serum-free and non-adherent conditions *in vitro*, the tumorigenicity of PDAC cells was assessed in immunocompromised mice. Molecular events were delineated by Western blotting and the presence of SOX2-expressing cells in tumour xenografts was evaluated by immunohistochemistry. **Results:** PANC-1 and Capan-2 cell lines showed exceptional resistance to gemcitabine with Capan-2 contains a notable number of putative CSCs co-expressing CD44, CD24, and CD133 (3.7% of the total population). These cells resisted the cytotoxicity of gemcitabine,

but responded to vismodegib. Tumourspheres were found arising from BxPC-3 and Capan-2 cell lines only, but the latter had closer resemblance to CSCs based on their responsiveness to the inhibition induced by foetal bovine serum. The presence of tumoursphere-forming cells is consistent with BxPC-3 and Capan-2 cell lines being highly tumourigenic in immunocompromised mice. DCAI and SRJ23 abrogated the self-renewal of Capan-2 tumourspheres with the effect of SRJ23 being the greatest. DCAI and SRJ23 diminished SOX2 expression and signalling pathways that were essentially activated in Capan-2 tumourspheres. SOX2-expressing cells were found correspondingly present in xenografts of highly tumourigenic BxPC-3 and Capan-2 cell lines, except for PANC-1 xenografts, of which SOX2-expressing cells were present despite poorly tumourigenic. **Conclusion:** Capan-2 cell line contains a small subset of putative CSCs, suggesting that cancer cell line is a valid source of CSCs in the discovery of novel anti-CSC agents. This study provides first evidence of anti-CSC activity in DCAI and SRJ23, which is attained through inhibiting KRAS signalling. It is postulated that SRJ23 induces an additional inhibitory circuitry that involves protein phosphatase 2A, a dephosphorylating enzyme that inactivates NF- κ B and WNT pathways, such that it emerges as a more promising candidate than DCAI in the future development of anti-CSC drug.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
Sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENILAIAN TITISAN SEL KANSER PANKREAS SEBAGAI SUMBER
SEL KANSER STEM DAN PENGGUNAANNYA UNTUK MENGUJI
SIFAT AKTIVITI ANTI-SEL KANSER STEM PERENCAT KRAS**

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Pengenalan: Walaupun tumor dapat dibedah dan gemcitabine dipreskrib dalam kemoterapi, adenokarsinoma duktus pankreas (PDAC) berulang kerap berlaku. Sel kanser stem pankreas telah dikenal pasti sebagai dalang kanser berulang. Pelbagai bukti mengatakan bahawa pengaktifan KRAS, molekul pengawal kepada laluan isyarat yang berhubung kait dengan kanser, yang tidak terkawal menyebabkan sel kanser stem pankreas menentang kesan antikanser gemcitabine. Vismodegib merupakan ejen anti-sel kanser stem yang paling awal ditemui di dalam makmal, namun ia dibuktikan tidak berkesan dalam membasmi kanser berulang. Oleh itu, usaha pencarian gantian vismodegib adalah penting, akan tetapi, tiada hasil yang berjaya setakat ini. **Objektif:** Dalam kajian ini, kehadiran sel kanser stem dalam titisan sel PDAC telah disahkan bahawa ia mampu dijadikan subjek ujikaji yang melibatkan perencat KRAS (DCAI dan SRJ23). Potensi DCAI dan SRJ23 dalam membasmi sel kanser stem dinilai dan mekanismenya turut dijelaskan. **Kaedah:** Aktiviti sitotoksik gemcitabine, vismodegib dan perencat KRAS dalam titisan sel PDAC (BxPC-3, PANC-1, Capan-2, and MIA PaCa-2) dinilai melalui ujian daya maju sel. Sitometri aliran dijalankan untuk menganalisis kehadiran penanda (CD44, CD24 dan CD133) pada permukaan sel kanser stem pancreas tunggal. Kebolehan titisan sel PDAC untuk membentuk sfera tumor dikaji dalam vitro melalui keadaan kultur yang bebas serum dan tidak melekat. Kemampuan ketumbuhan xenograf sel PDAC diuji pada mencit imunokompromi. Perubahan sel pada peringkat molekul dikesan dan digambarkan melalui ujian serap Western dan kehadiran sel xenograf tumor yang mengekspresikan SOX2 dikesan melalui ujian imunohistokimia. **Hasil Ujikaji:** Sel PANC-1 dan Capan-2 menunjukkan kesan rintangan yang luar biasa. Namun, hanya Capan-2 mengandungi sejuzuk sel (3.7% daripada seluruh populasi) yang mengekspresi CD44, CD24 dan CD133. Sel-sel ini menentang sitotoksik gemcitabine, tetapi mereka adalah sensitif

terhadap kesan vismodegib. Walaupun kedua-dua BxPC-3 dan Capan-2 mengandungi sel yang mampu membentuk sfera tumor, sel sfera tumor Capan-2 lebih menyerupai sel kanser stem berdasarkan respons mereka terhadap perencatan pertumbuhan yang disebabkan oleh serum anak lembu. Kehadiran sel-sel yang membentuk sfera tumor selaras dengan keupayaan BxPC-3 dan Capan-2 dalam mencetuskan pertumbuhan tumorigenik pada mencit imunokompromi. DCAI dan SRJ23 merencat kemampuan sel sfera tumor Capan-2 untuk mengswabaharui dan kesan SRJ23 adalah lebih menonjol. Kedua-duanya mengurangkan ekspresi SOX2 dan aktiviti laluan isyarat yang berperanan penting dalam sel sfera tumor Capan-2. Namun begitu, aktiviti SRJ23 dalam merencat isyarat NF- κ B dan WNT adalah lebih kuat daripada DCAI. Kehadiran sel yang mengekspresikan SOX2 dalam xenograf tumor BxPC-3 dan Capan-2 adalah selaras dengan keupayaan mereka dalam mencetuskan pertumbuhan tumor. Walau keupayaan tumorigenik berkurang dalam mencetuskan pertumbuhan tumor, xenograf tumor PANC-1 turut mengandungi sel yang mengekspresikan SOX2. **Kesimpulan:** Titisan sel Capan-2 mengandungi sejumlah kecil sel seiras dengan sel kanser stem yang mengekspresikan CD44, CD24 dan CD133 serta melawan kesan antikanser kemoterapi. Sel-sel ini mungkin terlibat dalam pembentukan sfera tumor dalam vitro dan mencetuskan pertumbuhan xenograf tumor dalam vivo. Sifat anti-sel kanser stem yang dimiliki DCAI dan SRJ23 adalah dilaporkan buat kali pertama dalam kajian ini. Sebagai perencat KRAS, kedua-duanya membantut pertumbuhan sel kanser stem melalui sekatan isyarat KRAS dan laluan isyarat di bawah kawalannya, iaitu MAPK dan AKT, diikuti penyahaktifan isyarat NF- κ B and WNT, dan diakhiri pengurangan ekspresi SOX2. Meskipun ada persamaan antara DCAI dan SRJ23 dalam mekanisme anti-sel kanser stem, adalah dicadangkan bahawa kebolehan SRJ23 melakukan sekatan tambahan terhadap isyarat penting dalam sel kanser stem melalui protein phosphatase 2A merupakan kelebihan yang ada pada SRJ23 apabila dibanding dengan DCAI. SRJ23 berpotensi untuk diusahakan sebagai anti-sel kanser stem.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

7-AAD	7-amino-actinomycin D
ABC	ATP-binding Cassette
ADP	Adenosine Diphosphate
AGP	Andrographolide
ALDH	Aldehyde Dehydrogenase
AML	Acute Myeloid Leukaemia
APC	Allophycocyanin
APL	Acute Promyelocytic Leukaemia
APS	Ammonium Persulfate
ATCC	American Type Culture Collection
α TOS	α -tocopheryl Succinate
ATP	Adenosine Triphosphate
bFGF	Basic Fibroblastic Growth Factor
BSA	Bovine Serum Albumin
BZIM	Benzimidazole
CAI	Carboxyamidotriazole
CI	Confidence Interval
CMYC	Cellular Myelocytomatosis Oncogene
CNS	Central Nervous System
CO	Carbon Monoxide
CSC	Cancer Stem Cell
DAB	3,3'-diaminobenzidine
DCAI	4,6-dichloro-2-methyl-3-aminoethyl-indole
DMEM	Dulbecco's Modified Eagle's Medium

DMEM/F12	Dulbecco's Modified Eagle's Medium: Nutrient Mixture F-12
DMSO	Dimethyl Sulphoxide
EDTA	Ethylenediaminetetraacetic Acid
EGF	Epidermal Growth Factor
EGFR	Epidermal Growth Factor Receptor
ELDA	Extreme Limiting Dilution Analysis
ERK	Extracellular Signal-regulated Kinase
ESA	Epithelial-specific Antigen
ESC	Embryonic Stem Cell
FACS	Fluorescence-activated Cell Sorting
FBS	Foetal Bovine Serum
FGFR1	Fibroblast Growth Factor Receptor 1
FITC	Fluorescein Isothiocyanate
FMO	Fluorescence Minus One
GAP	GTPase-activating Protein
GAPDH	Glyceraldehyde-3-phosphate Dehydrogenase
GDP	Guanosine Diphosphate
GEF	Guanine Nucleotide Exchange Factor
GFP	Green Fluorescent Protein
GTP	Guanosine Triphosphate
GTPase	Guanosine Triphosphatase
GRB2	Growth Factor Receptor-bound Protein 2
GSK3 β	Glycogen Synthase Kinase-3 β
GTPase	Guanosine Triphosphatase
hESCs	Human Embryonic Stem Cells
HSP40	Heatshock Protein 40

HVR	Hypervariable Region	
IHC	Immunohistochemistry	
I κ B α	IkappaB Alpha	
I κ Bs	Inhibitors of NF- κ B	
IKKs	I κ B Kinases	
IL1 β	Interleukin-1 β	
IL1R	Interleukin-1 Receptor	
iPSC	Induced Pluripotent Stem Cell	
KLF4	Kruppel-like Factor 4	
Kobe0065	N-(3-chloro-4-methylphenyl)-2-[2,6-dinitro-4-(trifluoromethyl)phenyl]-hydrazinecarbothioamide	
KRAS	Kirsten rat sarcoma	
LDA	Limiting Dilution Assay	
MACS	Magnetic-activated Cell Sorting	
MAPK	Mitogen-activated Protein Kinase	
MEK	Mitogen-activated Protein Kinase Kinase	
mIBG	Meta-iodobenzylguanidine	
MPTP	1-methyl 4-phenyl 1,2,3,6 Tetrahydropyridine	
mTOR	Mammalian Target of Rapamycin	
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl bromide	tetrazolium
NADH	Nicotinamide Adenine Dinucleotide	
NF- κ B	Nuclear Factor kappa B	
NO	Nitric Oxide	
NSCLC	Non-small Cell Lung Cancer	
OCT4	Octamer-binding Protein 4	
OXPHOS	Oxidative Phosphorylation	

PanIN	Pancreatic Intraductal Neoplasia
PBS	Phosphate-buffered Saline
PDAC	Pancreatic Ductal Adenocarcinoma
PDX1	Duodenal Homeobox Protein 1
PE	R-phycoerythrin
PH	Pleckstrin-homology
PI3K	Phosphoinositide 3-kinase
PIP ₂	Phosphatidylinositol 4,5-bisphosphate
PIP ₃	Phosphatidylinositol 3,4,5-triphosphate
PP2A	Protein Phosphatase 2A
PTCH	Patched
RalGDS	Ral Guanine Nucleotide Dissociation Stimulator
RBD	Ras-binding Domain
RSK-1	Ribosomal S6 Kinase 1
RSK-2	Ribosomal S6 Kinase 2
RTK	Receptor Tyrosine Kinase
SCID	Severe Combined Immune-deficient
SD	Standard Deviation
SDS-PAGE	Sodium Dodecyl Sulphate Polyacrylamide Gel Electrophoresis
SFN	Sulforaphane
SOX2	SRY-box Transcription Factor 2
SP	Side Population
SRJ09	3,19-(2-bromobenzylidene)andrographolide
SRJ23	3,19-(3-chloro-4-fluorobenzylidene)andrographolide
SHH	Sonic Hedgehog

SMO	Smoothened
TBST	Tris-buffered saline/Tween-20
TEMED	Tetramethylethylenediamine
TIC	Tumour-initiating Cell



CHAPTER 1

INTRODUCTION

Background of the Study

With a global incidence of 458,918 cases in 2018, pancreatic ductal adenocarcinoma (PDAC) is ranked 14th in the list of the commonest cancers worldwide (Bray et al., 2018). It is the seventh most lethal cancer with 432,232 deaths were recorded worldwide in 2018 (Bray et al., 2018). It is believed that PDAC arises from the epithelial cells that constitute the lining of pancreatic ducts, through which digestive enzymes produced by acinar cells are channelled into the duodenum (Stark & Eibl, 2015). Even though tumour masses can be removed by resection alone, the prognosis of PDAC patients remains poor (Gbolahan et al., 2019).

PDAC begins with a neoplastic precursor lesion, known as pancreatic intraepithelial neoplasia (PanIN) and its development is described as a progressive accumulation of multiple genetic mutations (Hruban et al., 2000). Among the key proto-oncogenes, Kirsten rat sarcoma (*KRAS*) gene is mutated at the early stage of PDAC development and oncogenic *KRAS* protein has been found in more than 90% of the PDAC cases (Eser et al., 2014), suggesting that it is a promising therapeutic target in the prevention and treatment of PDAC. The pivotal role of *KRAS* in promoting stem cell-like characteristics has also been confirmed in pancreatic cancer cells (Seguin et al., 2014). This finding is consistent with previous studies suggesting that acinar cells (Wei et al., 2016) and β -cells (Pour & Schmied, 1999; Pour et al., 2003) can be transdifferentiated into duct-like cells and function as the precursor of PanIN and PDAC by expressing oncogenic *KRAS* mutant proteins.

Cancer stem cells (CSCs) are identified as the fraction of cells, which show remarkable resistance to the cytotoxic effect of chemotherapeutic drugs, found within the heterogenous population of the bulk tumour. In contrast to their non-CSC counterparts, CSCs express drug-metabolising enzymes and drug efflux transporters (Thomas et al., 2015) as well as demonstrate enhanced ability of DNA repair (Liu et al., 2006). Contemporary chemotherapeutics eliminate fast-dividing non-CSCs effectively, leaving slow-cycling or quiescent CSCs unaffected during chemotherapy (Viale et al., 2009). The residual CSCs give rise to recurrent tumour in few years after the chemotherapy is terminated as a result of undetectable tumour volume.

In a landmark study by Hocker et al. (2013), it was reported that andrographolide (AGP) and its benzylidene derivatives, 3,19-(2-bromobenzylidene)andrographolide (SRJ09) and 3,19-(3-chloro-4-fluorobenzylidene)andrographolide (SRJ23) inhibit both wild-type and oncogenic KRAS proteins. This previous study has paved the way for initiating the present study with the aim of drugging KRAS, which has been known to be undruggable for many decades, in the context of targeting CSCs to eradicate pancreatic cancer recurrence. None of the known KRAS binders, which were found to have antagonistic activity against aberrant KRAS signalling preclinically, have been tested for their anti-pancreatic CSC activities.

Effort of drugging KRAS is still relevant in seeking potential treatment for PDAC patients, despite being challenging. It is believed that KRAS is a prominent therapeutic target in view of its pivotal role in the carcinogenesis and progression of PDAC. In the present study, previously known KRAS binders, namely benzimidazole (BZIM; Maurer et al., 2012), 4,6-dichloro-2-methyl-3-aminoethyl-indole (DCAI; Maurer et al., 2012), N-(3-chloro-4-methylphenyl)-2-[2,6-dinitro-4-(trifluoromethyl)phenyl]-hydrazinecarbothioamide (Kobe0065; Shima et al., 2013), andrographolide (AGP) and its derivatives (SRJ09 and SRJ23), were tested for their anti-pancreatic CSC property in PDAC cell lines with differential KRAS mutational statuses. These cell lines harbour wild-type KRAS (BxPC-3) and mutant KRAS: KRAS^{G12D} (PANC-1), KRAS^{G12V} (Capan-2), KRAS^{G12C} (MIA PaCa-2).

Problem Statement

Treatment options of PDAC remain scarce and they are limited to tumour resection that precedes an adjuvant chemotherapy with the use of either gemcitabine or 5-fluorouracil (Gbolahan et al., 2019). Although the adjuvant gemcitabine therapy was reported to have significantly improved the overall survival and disease-free survival of patients with resectable PDAC (Oettle et al., 2013), 80% of these patients would experience a recurrence that will eventually lead to fatality within two years after diagnosis (Gbolahan et al., 2019).

Pancreatic CSCs are known to have initiated recurrent and metastatic PDAC. Following the isolation and characterisation of pancreatic CSCs based on surface markers (CD44, CD24, and CD133), expression of embryonic SOX2 transcription factor, and an association of these cells with chemoresistance (Hermann et al., 2007; Herreros-Villanueva et al., 2013; Li et al., 2007), a few anti-pancreatic CSC agents were discovered. Vismodegib is the earliest known anti-pancreatic CSC agent that targets Hedgehog signalling (Singh et al., 2011), of which activity against pancreatic CSCs was proven to be promising preclinically. Nevertheless, recent clinical study revealed that combining vismodegib and gemcitabine

did not reduce the CSC content of tumour biopsies and the prognosis of patients with metastatic PDAC was not significantly improved as compared with gemcitabine alone (De Jesus-Acosta et al., 2019). This finding suggested that the anti-pancreatic CSC activity of vismodegib is not translatable from bench to clinical settings. Hence, it is crucial to seek for an alternative to vismodegib.

The existence of CSCs in continuous culture of cancer cell lines remains controversial. Many have questioned the authenticity of these microenvironment-sensitive cells after prolonged culturing in the presence of foetal bovine serum *in vitro*. Although patient-derived CSC culture is preferred for experimentations to study CSC biology and discover novel anti-CSC agents, the inaccessibility of patients' biopsies poses a great hurdle to research activities. Therefore, investigating the presence of CSCs in PDAC cell lines and characterising them molecularly in the context of their self-renewal mechanism are of value to researchers with difficulty in establishing patient-derived CSC culture.

Research Hypothesis

CSCs exist in PDAC cell lines as a rare subset of cells, of which hallmark features include showing resistance to gemcitabine and sensitivity to vismodegib, co-expressing pancreatic CSC surface markers, forming tumourspheres *in vitro*, and being highly tumourigenic *in vivo*. Based on the notion that KRAS signalling plays a determining role in the self-renewal of breast, colorectal, and pancreatic CSCs. It is hypothesised that KRAS binders inhibit the self-renewal of CSCs in PDAC cell lines.

Objectives of the Study

Main Objective

The general objective of the present study was to identify binders of KRAS with anti-pancreatic CSC activity.

Specific Objectives

The specific objectives of the present study were:

1. To characterise CSCs in PDAC cell lines based on resistance to gemcitabine and sensitivity to vismodegib, co-expression of pancreatic CSC surface markers, tumoursphere-forming ability *in vitro*, and tumourigenicity *in vivo*.
2. To establish a test model for assessing the inhibitory effects of KRAS binders on the self-renewal of CSCs in PDAC cell lines.

3. To assess the inhibitory effects of KRAS binders on the self-renewal of CSCs in PDAC cell lines.
4. To elucidate the molecular mechanism of the self-renewal of CSCs in PDAC cell lines and the mechanism of action in abrogating it by KRAS binders.
5. To evaluate the relevance of correlating the presence of SOX2-expressing cells in PDAC tumour xenografts with varying degrees of tumourigenicity *in vivo*.

Thesis Outline

The present study has two main aims:

1. Identifying and characterising pancreatic CSCs to establish a test model for evaluating the anti-pancreatic CSC property of KRAS binders.
2. Evaluating the anti-pancreatic CSC property of KRAS binders.

It is presented in a total of 7 chapters. A brief preview of each chapter is as follows:

Chapter 1

An introductory chapter includes the background, significance, problem statements, hypotheses, and objectives of the present study to help the readers in appreciating the rationale behind the initiation of the present study.

Chapter 2

This chapter notes fundamental knowledge for the conception, design, and execution of the present study. In addition, the knowledge is helpful to the readers in apprehending and appreciating the work done in the present study and the content of this thesis.

Chapter 3

This chapter explores the existence of CSCs in PDAC cell lines based on the chemoresistant nature of CSCs and co-expression of pancreatic CSC surface markers (CD44, CD24, and CD133). The CD44⁺CD24⁺CD133⁺ cells of Capan-2 cell line were found to be resistant to gemcitabine, a chemotherapeutic drug, and sensitive to vismodegib, an anti-pancreatic CSC agent.

Chapter 4

This chapter addresses the question of whether CSCs exist in PDAC cell lines based on two hallmark characteristics of CSCs, namely tumoursphere formation *in vitro* and tumourigenicity *in vivo*.

Chapter 5

This chapter reports the cytotoxic activities of KRAS binders, namely benzimidazole (BZIM), DCAI, Kobe0065, AGP, SRJ09, and SRJ23 in PDAC cell lines. DCAI and SRJ23 were selected to be further tested for their activities in abrogating the self-renewal of Capan-2 tumourspheres, which is pertaining to anti-pancreatic CSC property.

Chapter 6

This chapter delineates the molecular mechanism of the formation of Capan-2 tumourspheres in serum-free and non-adherent culture conditions. In addition, the modulating effects of gemcitabine, vismodegib, DCAI, and SRJ23 on the biology of Capan-2 tumourspheres were revealed. The relevance of relating the presence of SOX2-expressing cells, which are known to be pancreatic CSCs, in PDAC tumour xenografts to varying degrees of their tumorigenicity *in vivo* was investigated and reported.

Chapter 7

This chapter summarises the discussion and conclusion of each chapter. Future direction of the present study is suggested in this chapter.

REFERENCES

- Adjei, A. A. (2001). Blocking oncogenic Ras signaling for cancer therapy. *Journal of the National Cancer Institute*, 93(14): 1062-1074.
- Akita, H., Marquardt, J. U., Durkin, M. E., Kitade, M., Seo, D., Conner, E. A.,...Thorgeirsson, S. S. (2014). MYC activates stem-like cell potential in hepatocarcinoma by a p53-dependent mechanism. *Cancer Research*, 74(20): 5903-5913.
- Al-Hajj, M., Wicha, M. S., Benito-Hernandez, A., Morrison, S. J., & Clarke, M. F. (2003). Prospective identification of tumorigenic breast cancer cells. *Proceedings of the National Academy of Sciences of the United States of America*, 100(7): 3983-3988.
- Arlt, A., Gehrz, A., Muerköster, S., Vorndamm, J., Kruse, M. L., Fölsch, U. R., & Schäfer, H. (2003). Role of NF-kappaB and Akt/PI3K in the resistance of pancreatic carcinoma cell lines against gemcitabine-induced cell death. *Oncogene*, 22(21): 3243-3251.
- Arlt, A., Vorndamm, J., Breitenbroich, M., Fölsch, U. R., Kalthoff, H., Schmidt, W. E., & Schäfer, H. (2001). Inhibition of NF-kappaB sensitizes human pancreatic carcinoma cells to apoptosis induced by etoposide (VP16) or doxorubicin. *Oncogene*, 20(7): 859-868.
- Armstrong, L., Hughes, O., Yung, S., Hyslop, L., Stewart, R., Wappler, I.,...Lako, M. (2006). The role of PI3K/AKT, MAPK/ERK and NFkappabeta signalling in the maintenance of human embryonic stem cell pluripotency and viability highlighted by transcriptional profiling and functional analysis. *Human Molecular Genetics*, 15(11): 1894-1913.
- Ashton, T. M., McKenna, W. G., Kunz-Schughart, L. A., & Higgins, G. S. (2018). Oxidative Phosphorylation as an emerging target in cancer therapy. *Clinical Cancer Research*, 24(11): 2482-2490.
- Atwood, S. X., Sarin, K. Y., Whitson, R. J., Li, J. R., Kim, G., Rezaee, M.,...Tang, J. Y. (2015). Smoothed variants explain the majority of drug resistance in basal cell carcinoma. *Cancer Cell*, 27(3): 342-353.
- Bai, D., Ueno, L., & Vogt, P. K. (2009). Akt-mediated regulation of NFkappaB and the essentialness of NFkappaB for the oncogenicity of PI3K and Akt. *International Journal of Cancer*, 125(12): 2863-2870.
- Balic, A., Sørensen, M. D., Trabulo, S. M., Sainz, B. Jr., Cioffi, M., Vieira, C. R.,...Heeschen, C. (2014). Chloroquine targets pancreatic cancer stem cells via inhibition of CXCR4 and hedgehog signalling. *Molecular Cancer Therapeutics*, 13(7): 1758-1771.

- Bardeesy, N. & DePinho, R. A. (2002). Pancreatic cancer biology and genetics. *Nature Reviews Cancer*, 2(12): 897-909.
- Bass, A. J., Watanabe, H., Mermel, C. H., Yu, S., Perner, S., Verhaak, R. G.,... Meyerson, M. (2009). SOX2 is an amplified lineage-survival oncogene in lung and esophageal squamous cell carcinomas. *Nature Genetics*, 41(11): 1238-1242.
- Bell, R. H. Jr. & Pour, P. M. (1987). Pancreatic carcinogenicity of N-nitrosobis(2-oxopropyl)-amine in diabetic and non-diabetic Chinese hamsters. *Cancer Letters*, 34(2): 221-230.
- Berman, D. M., Karhadkar, S. S., Maitra, A., Montes De Oca, R., Gerstenblith, M. R., Briggs, K.,... Beachy, P. A. (2003). Widespread requirement for hedgehog ligand stimulation in growth of digestive tract tumours. *Nature*, 425(6960): 846-851.
- Bernards, A. & Settleman, J. (2004). GAP control: regulating the regulators of small GTPases. *Trends in Cell Biology*, 14(7): 377-385.
- Bettiol, E., Sartiani, L., Chicha, L., Krause, K. H., Cerbai, E., & Jaconi, M. E. (2007). Fetal bovine serum enables cardiac differentiation of human embryonic stem cells. *Differentiation*, 75(8): 669-681.
- Birecree, E., King, L. E. Jr., & Nanney, L. B. (1991). Epidermal growth factor and its receptor in the developing human nervous system. *Developmental Brain Research*, 60(2): 145-154.
- Bodine, D., Seidel, N. E., & Orlic, D. (1996). Bone marrow collected 14 days after *in vivo* administration of granulocyte colony-stimulating factor and stem cell factor to mice has 10-fold more repopulating ability than untreated bone marrow. *Blood*, 88(1): 89-97.
- Bohusiav, J., Chen, L. F., Kwon, H., Mu, Y., & Greene, W. C. (2004). p53 induces NK-kappaB activation by an IkappaB kinase-independent mechanism involving phosphorylation of p65 by ribosomal S6 kinase 1. *Journal of Biological Chemistry*, 279(25): 26115-26125.
- Bonner-Weir, S. & Sharma, A. (2002). Pancreatic stem cells. *Journal of Pathology*, 197(4): 519-526.
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., & Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians*, 68(6): 394-424.
- Brewer, J. R., Mazot, P., & Soriano, P. (2016). Genetic insights into the mechanisms of Fgf signaling. *Genes & Development*, 30(7): 751-771.

- Cammareri, P., Scopelliti, A., Todaro, M., Eterno, V., Francescangeli, F., Moyer, M. P.,...Stassi, G. (2010). Aurora-a is essential for the tumorigenic capacity and chemoresistance of colorectal cancer stem cells. *Cancer Research*, 70(11): 4655-4665.
- Cantley, L. C. (2002). The phosphoinositide 3-kinase pathway. *Science*, 296(5573): 1655-1657.
- Cao, Y., Luo, J. L., & Karin, M. (2007). I κ B kinase α kinase activity is required for self-renewal of ErbB2/Her2-transformed mammary tumor-initiating cells. *Proceedings of the National Academy of Sciences of the United States of America*, 104(40): 15852-15857.
- Castillo-Azofeifa, D., Seidel, K., Gross, L., Golden, E. J., Jacquez, B., Klein, O. D., & Barlow, L. A. (2018). SOX2 regulation by hedgehog signalling controls adult lingual epithelium homeostasis. *Development*, 145(14). pii: dev164889.
- Catenacci, D. V., Junttila, M. R., Karrison, T., Bahary, N., Horiba, M. N., Nattam, S. R.,...Kindler, H. L. (2015). Randomized phase Ib/II study of gemcitabine plus placebo or vismodegib, a Hedgehog pathway inhibitor, in patients with metastatic pancreatic cancer. *Journal of Clinical Oncology*, 33(36): 4284-4292.
- Chandrakesan, P., Ahmed, I., Anwar, T., Wang, Y., Sarkar, S., Singh, P.,...Umar, S. (2010). Novel changes in NF- κ B activity during progression and regression phases of hyperplasia: role of MEK, ERK, and p38. *Journal of Biological Chemistry*, 285(43): 33485-33498.
- Chen, J., Li, Y., Yu, T. S., McKay, R. M., Burns, D. K., Kernie, S. G., & Parada, L. F. (2012). A restricted cell population propagates glioblastoma growth following chemotherapy. *Nature*, 488(7412): 522-526.
- Chen, L. F., Williams, S. A., Mu, Y., akano, H., Duerr, J. M., Buckbinder, L., & Greene, W. C. (2005). NF- κ B RelA phosphorylation regulates RelA acetylation. *Molecular and Cellular Biology*, 25(18): 7966-7975.
- Chenn, A. & McConnell, S. K. (1995). Cleavage orientation and the asymmetric inheritance of Notch1 immunoreactivity in mammalian neurogenesis. *Cell*, 82(4): 631-641.
- Chow, E. K., Fan, L. L., Chen, X., & Bishop, J. M. (2012). Oncogene-specific formation of chemoresistant murine hepatic cancer stem cells. *Hepatology*, 56(4): 1331-1341.
- Clarke, M. F., Dick, J. E., Dirks, P. B., Eaves, C. J., Jamieson, C. H., Jones, D. L.,...Wahl, G. M. (2006). Cancer stem cells-perspectives

on current status and future directions: AACR workshop on cancer stem cells. *Cancer Research*, 66(19): 9339-9344.

Cox, A. D. & Der, C. J. (2010). Ras history: The saga continues. *Small GTPases*, 1(1): 2-27.

Cubilla, A. L. & Fitzgerald, P. J. (1976). Morphological lesions associated with human primary invasive nonendocrine pancreas cancer. *Cancer Research*, 36(7 PT 2): 2690-2698.

Cunningham, J. J., Ulbright, T. M., Pera, M. F., & Looijenga, L. H. (2012). Lessons from human teratomas to guide development of safe stem cell therapies. *Nature Biotechnology*, 30(9): 849-857.

De Jesus-Acosta, A., Sugar, E. A., O'Dwyer, P. J., Ramanathan, R. K., Von Hoff, D. D., Rasheed, Z.,...Laheru, D. A. (2019). Phase 2 study of vismodegib, a hedgehog inhibitor, combined with gemcitabine and nab-paclitaxel in patients with untreated metastatic pancreatic adenocarcinoma. *British Journal of Cancer*, 122(4): 498-505.

De Sousa Cavalcante, L. & Monteiro, G. (2014). Gemcitabine: metabolism and molecular mechanisms of action, sensitivity and chemoresistance in pancreatic cancer. *European Journal of Pharmacology*, 741: 8-16.

Dierks, C., Beigi, R., Guo G. R., Zirlik, K., Stegert, M. R., Manley, P.,...Warmuth, M. (2008). Expansion of Bcr-Abl-positive leukemic stem cells is dependent on Hedgehog pathway activation. *Cancer Cell*, 14(3): 238-249.

Dylla, S. J., Beviglia, L., Park, I. K., Chartier, C., Raval, J., Ngan, L.,...Smith-Berdan, S. (2008). Colorectal cancer stem cells are enriched in xenogeneic tumors following chemotherapy. *PLoS One*, 3(6): e2428.

Eramo, A., Lotti, F., Sette, G., Piloizzi, E., Biffoni, M., Di Virgilio, A.,...De Maria, R. (2008). Identification and expansion of tumorigenic lung cancer stem cell population. *Cell Death & Differentiation*, 15(3): 504-514.

Eser, S., Schnieke, A., Schneider, G., & Saur, D. (2014). Oncogenic KRAS signalling in pancreatic cancer. *British Journal of Cancer*, 111(5): 817-822.

Fallon, J. H., Seroogy, K. B., Loughlin, S. E., Morrison, R. S., Bradshaw, R. A., Knaver, D. J., & Cunningham, D. D. (1984). Epidermal growth factor immunoreactive material in the central nervous system: location and development. *Science*, 224(4653):1107-1109.

- Fang, X., Yu, S. X., Lu, Y., Bast, R. C. Jr., Woodgett, J. R., & Mills, G. B. (2000). Phosphorylation and inactivation of glycogen synthase kinase 3 by protein kinase A. *Proceedings of the National Academy of Sciences of the United States of America*, 97(22): 11960-11965.
- Farnie, G., Clarke, R. B., Spence, K., Pinnock, N., Brennan, K., Anderson, N. G., & Bundred, N. J. (2007). Novel cell culture technique for primary ductal carcinoma in situ: role of Notch and epidermal growth factor receptor signalling pathways. *Journal of the National Cancer Institute*, 99(8): 616-627.
- Feldmann, G., Dhara, S., Fendrich, V., Bedja, D., Beaty, R., Mullendore, M.,...Maitra, A. (2007). Blockade of Hedgehog signalling inhibits pancreatic cancer invasion and metastases: a new paradigm for combination therapy in solid cancers. *Cancer Research*, 67(5): 2187-2196.
- Ferreira, R. M. M., Sancho, R., Messal, H. A., Nye, E., Spencer-Dene, B., Stone, R. K.,...Behrens, A. (2017). Duct- and acinar-derived pancreatic ductal adenocarcinomas show distinct tumor progression and marker expression. *Cell Reports*, 21(4): 966-978.
- Franke, J., Abs, V., Zizzadoro, C., & Abraham, G. (2014). Comparative study of the effects of fetal bovine serum versus horse serum on growth and differentiation of primary equine bronchial fibroblasts. *BMC Veterinary Research*, 10: 119.
- Fu, D., Calvo, J. A., Samson, L. D. (2012). Balancing repair and tolerance of DNA damage caused by alkylating agents. *Nature Reviews Cancer*, 12(2): 104-120.
- Fu, J., Rodova, M., Roy, S. K., Sharma, J., Singh, K. P., Srivastava, R. K., & Shankar, S. (2013). GANT-61 inhibits pancreatic cancer stem cell growth in vitro and in NOD/SCID/IL2R gamma null mice xenograft. *Cancer Letters*, 330(1): 22-32.
- Furukawa, T., Chiba, R., Kobari, M., Matsuno, S., Nagura, H., & Takahashi, T. (1994). Varying grades of epithelial atypia in the pancreatic ducts of humans. Classification based on morphometry and multivariate analysis and correlated with positive reactions of carcinoembryonic antigen. *Archives of Pathology & Laboratory Medicine*, 118(3): 227-234.
- Gage, F. H., Ray, J., & Fisher, L. J. (1995). Isolation, characterization, and use of stem cells from the CNS. *Annual Review of Neuroscience*, 18: 159-192.
- Gamer, J. M., Fan, M., Yang, C. H., Du, Z., Sims, M., Davidoff, A. M., & Pfeffer, L. M. (2013). Constitutive activation of signal transducer and activator of transcription 3 (STAT3) and nuclear factor κB

signalling in glioblastoma cancer stem cells regulates the Notch pathway. *Journal of Biological Chemistry*, 288(36): 26167-26176.

Gbolahan, O. B., Tong, Y., Sehdev, A., O'Neil, B., & Shahda, S. (2019). Overall survival of patients with recurrent pancreatic cancer treated with systemic therapy: a retrospective study. *BMC Cancer*, 19(1): 468.

Guzman, M. L., Neering, S. J., Upchurch, D., Grimes, B., Howard, D. S., Rizzieri, D. A., ... Jordan, C. T. (2001). Nuclear factor-kappa B is constitutively activated in primitive human acute myelogenous leukemia cells. *Blood*, 98(8): 2301-2307.

Hariharan, D., Saied, A., & Kocher, H. M. (2008). Analysis of mortality rates for pancreatic cancer across the world. *HPB (Oxford)*, 10(1): 58-62.

Hermann, P. C., Huber, S. L., Herrler, T., Aicher, A., Ellwart, J. W., Guba, M., ... Heeschen, C. (2007). Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer. *Cell Stem Cell*, 1(3): 313-323.

Hermann, P. C. & Sainz, B. Jr. (2018). Pancreatic cancer stem cells: A state or an entity? *Seminars in Cancer Biology*, 53: 223-231.

Herrmann, C. (2003). Ras-effector interactions: after one decade. *Current Opinion in Structural Biology*, 13(1): 122-129.

Herreros-Villanueva, M., Zhang, J. S., Koenig, A., Abel, E. V., Smyrk, T. C., Bamlet, W. R., ... Billadeau, D. D. (2013). SOX2 promotes dedifferentiation and imparts stem cell-like features to pancreatic cancer cells. *Oncogenesis*, 2: e61.

Hocker, H. J., Cho, K. J., Chen, C. Y., Rambahal, N., Sagineedu, S. R., Shaari, K., ... Gorfe, A. A. (2013). Andrographolide derivatives inhibit guanine nucleotide exchange and abrogate oncogenic Ras function. *Proceedings of the National Academy of Sciences of the United States of America*, 110(25): 10201-10206.

Hruban, R. H., Goggins, M., Parsons, J., & Kern, S. E. (2000). Progression model for pancreatic cancer. *Clinical Cancer Research*, 6(8): 2969-2972.

Hruban, R. H., van Mansfeld, A. D., Offerhaus, G. J., van Weering, D. H., Allison, D. C., Goodman, S. N., ... Bos, J. L. (1993). K-ras oncogene activation in adenocarcinoma of the human pancreas. A study of 82 carcinomas using a combination of mutant-enriched polymerase chain reaction analysis and allele-specific oligonucleotide hybridization. *American Journal of Pathology*, 143(2): 545-554.

- Hsieh, C. Y., Hsu, M. J., Hsiao, G., Wang, Y. H., Huang, C. W., Chen, S. W.,...Sheu, J. R. (2011). Andrographolide enhances nuclear factor-kappa B subunit p65 Ser536 dephosphorylation through activation of protein phosphatase 2A in vascular smooth muscle cells. *Journal of Biological Chemistry*, 286(8): 5942-5955.
- Hu, L., McArthur, C., & Jaffe, R. B. (2010). Ovarian cancer stem-like side-population cells are tumourigenic and chemoresistant. *British Journal of Cancer*, 102(8): 1276-1283.
- Hu, Y. & Smyth, G. K. (2009). ELDA: Extreme limiting dilution analysis for comparing depleted and enriched populations in stem cell and other assays. *Journal of Immunological Methods*, 347: 70-78.
- Huang, H. L., Hsing, H. W., Lai, T. C., Chen, Y. W., Lee, T. R., Chan, H. T.,...Chan, H. L. (2010). Trypsin-induced proteome alteration during cell subculture in mammalian cells. *Journal of Biomedical Science*, 17: 36.
- Huang, P., Zheng, S., Wierbowski, B. M., Kim, Y., Nedelcu, D., Aravena, L.,...Salic, A. (2018). Structural basis of Smoothened activation in Hedgehog signaling. *Cell*, 175(1): 295-297.
- Huang, F. T., Zhuan-Sun, Y. X., Zhuang, Y. Y., Wei, S. L., Tang, J., Chen, W. B.,...Zhang, S. N. (2012). Inhibition of hedgehog signaling depresses self-renewal of pancreatic cancer stem cells and reverses chemoresistance. *International Journal of Oncology*, 41(5): 1707-1714.
- Ilmer, M., Boiles, A. R., Regel, I., Yokoi, K., Michalski, C. W., Wistuba, I. I.,...Vykroukal, J. (2015). RSPO2 enhances canonical Wnt signaling to confer stemness-associated traits to susceptible pancreatic cancer cells. *Cancer Research*, 75(9): 1883-1896.
- Ischenko, I., Petrenko, O., & Hayman, M. J. (2014). Analysis of the tumor-initiating and metastatic capacity of PDX1-positive cells from the adult pancreas. *Proceedings of the National Academy of Sciences of the United States of America*, 111(9): 3466-3471.
- Ishikawa, F., Yoshida, S., Saito, Y., Hijikata, A., Kitamura, H., Tanaka, S.,...Shultz, L. D. (2007). Chemotherapy-resistant human AML stem cells home to and engraft within the bone-marrow endosteal region. *Nature Biotechnology*, 25(11): 1315-1321.
- Jaafar Marican, N. H., Cruz-Migoni, S. B., & Borycki, A. G. (2016). Asymmetric distribution of primary cilia allocates satellite cells for self-renewal. *Stem Cell Reports*, 6(6): 798-805.
- Jada, S. R., Matthews, C., Saad, M. S., Hamzah, A. S., Lajis, N. H., Stevens, M. F., & Stanslas, J. (2008). Benzylidene derivatives of andrographolide inhibit growth of breast and colon cancer cells in

vitro by inducing G(1) arrest and apoptosis. *British Journal of Pharmacology*, 155(5): 641-654.

- Jeong, S. J., Pise-Masison, C. A., Radonovich, M. F., Park, H. U., & Brady, J. N. (2005). Activated AKT regulates NF-kappaB activation, p53 inhibition and cell survival in HTLV-1-transformed cells. *Oncogene*, 24(44): 6719-6728.
- Ji, Z., Mei, F. C., Xie, J., & Cheng, X. (2007). Oncogenic KRAS activates hedgehog signaling pathway in pancreatic cancer cells. *Journal of Biological Chemistry*, 282(19): 14048-14055.
- Kang, Y. W., Lee, J. E., Jung, K. H., Son, M. K., Shin, S. M., Kim, S. J.,...Hong, S. S. (2018). KRAS targeting antibody synergizes anti-cancer activity of gemcitabine against pancreatic cancer. *Cancer Letters*, 438: 174-186.
- Kapoor, A., Yao, W., Ying, H., Hua, S., Liewen, A., Wang, Q.,...DePinho, R. A. (2014). Yap1 activation enables bypass of oncogenic Kras addiction in pancreatic cancer. *Cell*, 158(1): 185-197.
- Kern, S., Hruban, R., Hollingsworth, M. A., Brand, R., Adrian, T. E., Jaffee, E., & Tempero, M. A. (2001). A white paper: the product of a pancreas cancer think tank. *Cancer Research*, 61(12): 4923-4932.
- Kim, M. P. & Gallick, G. E. (2008). Gemcitabine resistance in pancreatic cancer: Picking the key players. *Clinical Cancer Research*, 14(5): 1284-1285.
- Kim, E. J., Sahai, V., Abel, E. V., Griffith, K. A., Greenson, J. K., Takebe, N.,...Simeone, D. M. (2014). Pilot clinical trial of hedgehog pathway inhibitor GDC-0449 (vismodegib) in combination with gemcitabine in patients with metastatic pancreatic adenocarcinoma. *Clinical Cancer Research*, 20(23): 5937-5945.
- Kimura, W., Morikane, K., Esaki, Y., Chan, W. C., & Pour, P. M. (1998). Histologic and biologic patterns of microscopic pancreatic ductal adenocarcinomas detected incidentally at autopsy. *Cancer*, 82(10): 1839-1849.
- Konstantinopoulos, P. A., Karamouzis, M. V., & Papavassiliou, A. G. (2007). Post-translational modifications and regulation of the RAS superfamily of GTPases as anticancer targets. *Nature Reviews Drug Discovery*, 6(7): 541-555.
- Koo, B. S., Lee, S. H., Kim, J. M., Huang, S., Kim, S. H., Rho, Y. S.,...Lim, Y. C. (2015). Oct4 is a critical regulator of stemness in head and neck squamous carcinoma cells. *Oncogene*, 34(18): 2317-2324.

- Kozuka, S., Sassa, R., Taki, T., Masamoto, K., Nagasawa, S., Saga, S.,...Takeuchi, M. (1979). Relation of pancreatic duct hyperplasia to carcinoma. *Cancer*, 43(4): 1418-1428.
- Lapidot, T., Sirard, C., Vormoor, J., Murdoch, B., Hoang, T., Caceres-Cortes, J.,...Dick, J. E. (1994). A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature*, 367(6464): 645-648.
- Le Rolle, A. F., Chiu, T. K., Zeng, Z., Shia, J., Weiser, M. R., Paty, P. B., & Chiu, V. K. (2016). Oncogenic KRAS activates an embryonic stem cell-like program in human colon cancer initiation. *Oncotarget*, 7(3): 2159-2174.
- Lechler, T. & Fuchs, E. (2005). Asymmetric cell divisions promote stratification and differentiation of mammalian skin. *Nature*, 437(7056): 275-280.
- Lee, C. H., Yu, C. C., Wang, B. Y., & Chang, W. W. (2016). Tumorsphere as an effective in vitro platform for screening anti-cancer stem cell drugs. *Oncotarget*, 7(2): 1215-1226.
- Lee, J., Kotliarova, S., Kotliarov, Y., Li, A., Su, Q., Donin, N. M., & Fine, H. A. (2006). Tumor stem cells derived from glioblastomas cultured in bFGF and EGF more closely mirror the phenotype and genotype of primary tumors than do serum-cultured cell lines. *Cancer Cell*, 9(5): 391-403.
- Lennerz, J. K. & Stenzinger, A. (2015). Allelic ratio of KRAS mutations in pancreatic cancer. *Oncologist*, 20(4): e8-9.
- Li, S. H., Fu, J., Watkins, D. N., Srivastava, R. K., & Shankar, S. (2013). Sulforaphane regulates self-renewal of pancreatic cancer stem cells through the modulation of Sonic Hedgehog-Gli pathway. *Molecular and Cellular Biochemistry*, 373(1-2): 217-227.
- Li, C., Heidt, D. G., Dalerba, P., Burant, C. F., Zhang, L., Adsay, V.,...Simeone, D. M. (2007). Identification of pancreatic cancer stem cells. *Cancer Research*, 67(3): 1030-1037.
- Li, X., Lewis, M. T., Huang, J., Gutierrez, C., Osborne, C. K., Wu, M. F.,...Chang, J. C. (2008). Intrinsic resistance of tumorigenic breast cancer cells to chemotherapy. *Journal of National Cancer Institute*, 100(9): 672-679.
- Li, Y., Xi, Z, Chen, X., Cai, S., Liang, C., Wang, Z.,...Xu, H. (2018). Natural compound Oblongifolin C confers gemcitabine resistance in pancreatic cancer by downregulating Src/MAPK/ERK pathways. *Cell Death & Disease*, 9(5): 538.

- Li, Q., Yang, G., Feng, M., Zheng, S., Cao, Z., Qiu, J.,...Zhao, Y. (2018). NF- κ B in pancreatic cancer: Its key role in chemoresistance. *Cancer Letters*, 421: 127-134.
- Liang, C., Shi, S., Meng, Q., Liang, D., Ji, S., Zhang, B.,...Yu, X. (2017). Complex roles of the stroma in the intrinsic resistance to gemcitabine in pancreatic cancer: where we are and where we are going. *Experimental & Molecular Medicine*, 49: e406.
- Lim, J. C., Chan, T. K., Ng, D. S., Sagineedu, S. R., Stanslas, J., & Wong, W. S. (2012). Andrographolide and its analogues: versatile bioactive molecules for combating inflammation and cancer. *Clinical and Experimental Pharmacology and Physiology*, 39(3): 300-310.
- Lin, W. C., Rajbhandari, N., Liu, C., Sakamoto, K., Zhang, Q., Triplett, A. A.,...Wagner, K. U. (2013). Dormant cancer cells contribute to residual disease in a model of reversible pancreatic cancer. *Cancer Research*, 73(6): 1821-1830.
- Liston, D. R. & Davis, M. (2017). Clinically relevant concentrations of anticancer drugs: A guide for nonclinical studies. *Clinical Cancer Research*, 23(14): 3489-3498.
- Liu, S., Dontu, G., Mantle, I. D., Patel, S., Ahn, N. S., Jackson, K. W.,...Wicha, M. S. (2006). Hedgehog signalling and Bmi-1 regulate self-renewal of normal and malignant human mammary stem cells. *Cancer Research*, 66(12): 6063-6071.
- Liu, M., Sakamaki, T., Casimiro, M. C., Willmarth, N. E., Quong, A. A., Ju, X.,...Pestell, R. G. (2010). The canonical NF- κ B pathway governs mammary tumorigenesis in transgenic mice and tumor stem cell expansion. *Cancer Research*, 70(24): 10464-10473.
- Liu, T., Xu, F., Du, X., Lai, D., Liu, T., Zhao, Y.,...Liu, Z. (2010). Establishment and characterization of multi-drug resistant, prostate carcinoma-initiating stem-like cells from human prostate cancer cell lines 22RV1. *Molecular and Cellular Biochemistry*, 340(1-2): 265-273.
- Liu, G., Yuan, X., Zeng, Z., Tunici, P., Ng, H., Abdulkadir, I. R.,...Yu, J. S. (2006). Analysis of gene expression and chemoresistance of CD133+ cancer stem cells in glioblastoma. *Molecular Cancer*, 5: 67.
- Lombardo, Y., de Giorgio, A., Coombes, C. R., Stebbing, J., & Castellano, L. (2015). Mammosphere formation assay from human breast cancer tissues and cell lines. *Journal of Visualized Experiments*, (97): 52671.

- Lonardo, E., Cioffi, M., Sancho, P., Crusz, S., & Heeschen, C. (2015). Studying pancreatic cancer stem cell characteristics for developing new treatment strategies. *Journal of Visualized Experiments*, (100): e52801.
- Lonardo, E., Cioffi, M., Sancho, P., Sanchez-Ripoll, Y., Trabulo, S. M., Dorado, J.,...Heeschen, C. (2013). Metformin targets the metabolic achilles heel of human pancreatic cancer stem cells. *PLoS One*, 8(10): e76518.
- Lonardo, E., Hermann, P. C., Heeschen, C. (2010). Pancreatic cancer stem cells-update and future perspectives. *Molecular Oncology*, 4(5): 431-442.
- Lonardo, E., Hermann, P. C., Mueller, M. T., Huber, S., Balic, A., Miranda-Lorenzo, I.,...Heeschen, C. (2011). Nodal/Activin signaling drives self-renewal and tumorigenicity of pancreatic cancer stem cells and provides a target for combined drug therapy. *Cell Stem Cell*, 9(5): 433-446.
- Lu, Y., Liu, Y., Xu, Z., Li, H., Liu, H., & Zhu, W. (2012). Halogen bonding for rational drug design and new drug discovery. *Expert Opinion on Drug Discovery*, 7(5): 375-383.
- Lu, C., Yang, D., Sabbatini, M. E., Colby, A. H., Grinstaff, M. W., Oberlies, N. H.,...Liu, K. (2018). Contrasting roles of H3K4me3 and H3K9me3 in regulation of apoptosis and gemcitabine resistance in human pancreatic cancer cells. *BMC Cancer*, 18(1): 149.
- Lu, Y., Zhu, H., Shan, H., Lu, J., Chang, X., Li, X.,...Wang, Z. (2013). Knockdown of Oct4 and Nanog expression inhibits the stemness of pancreatic cancer cells. *Cancer Letters*, 340(1): 113-123.
- Ma, Y., Yu, W., Shrivastava, A., Srivastava, R. K., & Shankar, S. (2019). Inhibition of pancreatic cancer stem cell characteristics by α -Mangostin: Molecular mechanisms involving Sonic hedgehog and Nanog. *Journal of Cellular and Molecular Medicine*, 23(4): 2719-2730.
- Manikam, S. D. & Stanslas, J. (2009). Andrographolide inhibits growth of acute promyelocytic leukaemia cells by inducing retinoic acid receptor-independent cell differentiation and apoptosis. *Journal of Pharmacy and Pharmacology*, 61(1): 69-78.
- Matsubara, S., Ding, Q., Miyazaki, Y., Kuwahata, T., Tsukasa, K., & Takao, S. (2013). mTOR plays critical roles in pancreatic cancer stem cells through specific and stemness-related functions. *Scientific Reports*, 3: 3230.
- Maurer, T., Garrenton, L. S., Oh, A., Pitts, K., Anderson, D. J., Skelton N. J.,...Fang, G. (2012). Small-molecule ligands bind to a distinct

pocket in Ras and inhibit SOS-mediated nucleotide exchange activity. *Proceedings of the National Academy of Sciences of the United States of America*, 109(14): 5299-5304.

McKay, R. (1997). Stem cells in the central nervous system. *Science*, 276(5309): 66-71.

Messina, E., De Angelis, L., Frati, G., Morrone, S., Chimenti, S., Fiordaliso, F.,...Giacomello, A. (2004). Isolation and expansion of adult cardiac stem cells from human and murine heart. *Circulation Research*, 95(9): 911-921.

Michaud, D. S. (2004). Epidemiology of pancreatic cancer. *Minerva Chirurgica*, 59(2): 99-111.

Mitra, A., Menezes, M. E., Pannell, L. K., Mulekar, M. S., Honkanen, R. E., Shevde, L. A., & Samant, R. S. (2012). DNAJB6 chaperones PP2A mediated dephosphorylation of GSK3 β to downregulate β -catenin transcription target, osteopontin. *Oncogene*, 31(41): 4472-4483.

Modi, S., Kir, D., Banerjee, S., & Saluja, A. (2016). Control of apoptosis in treatment and biology of pancreatic cancer. *Journal of Cellular Biochemistry*, 117(2): 279-288.

Moon, B. S., Jeong, W. J., Park, J., Kim, T. I., Min do, S., & Choi, K. Y. (2014). Role of oncogenic K-Ras in cancer stem cell activation by aberrant Wnt/ β -catenin signaling. *Journal of the National Cancer Institute*, 106(2): djt373.

Moreira, D., Zhang, Q., Hossain, D. M., Nechaev, S., Li, H., Kowolik, C. M.,...Kortylewski, M. (2015). TLR9 signaling through NF- κ B/RELA and STAT3 promotes tumor-propagating potential of prostate cancer cells. *Oncotarget*, 6(19): 17302-17313.

Morrison, S. J. & Kimble, J. (2006). Asymmetric and symmetric stem-cell divisions in development and cancer. *Nature*, 441(7097): 1068-1074.

Morrison, S. J., Wright, D. E., & Weissman, I. L. (1997). Cyclophosphamide/granulocyte colony-stimulating factor induces hematopoietic stem cells to proliferate prior to mobilization. *Proceedings of the National Academy of Sciences of the United States of America*, 94(5): 1908-1913.

Morshead, C. M., Craig, C. G., & van der Kooy, D. (1998). *In vivo* clonal analyses reveal the properties of endogenous neural stem cell proliferation in the adult mammalian forebrain. *Development*, 125(12): 2251-2261.

- Moskaluk, C. A., Hruban, R. H., & Kern, S. E. (1997). p16 and K-ras gene mutations in the intraductal precursors of human pancreatic adenocarcinoma. *Cancer Research*, 57(11): 2140-2143.
- Mueller, M. T., Hermann, P. C., Witthauer, J., Rubio-Viguira, B., Leicht, S. F., Huber, S.,...Heeschen, C. (2009). Combined targeted treatment to eliminate tumorigenic cancer stem cells in human pancreatic cancer. *Gastroenterology*, 137(3): 1102-1113.
- Müller, M., Hermann, P. C., Liebau, S., Weidgang, C., Seufferlein, T., Kleger, A., & Perkhofer, L. (2016). The role of pluripotency factors to drive stemness in gastrointestinal cancer. *Stem Cell Research*, 16(2): 349-357.
- Nakanishi, C. & Toi, M. (2005). Nuclear factor-kappaB inhibitors as sensitizers to anticancer drugs. *Nature Reviews Cancer*, 5(4): 297-309.
- Nassar, N., Horn, G., Herrmann, C., Scherer, A., McCormick, F., & Wittinghofer, A. (1995). The 2.2 Å crystal structure of the Ras-binding domain of the serine/threonine kinase c-Raf1 in complex with Rap1A and a GTP analogue. *Nature*, 375(6532): 554-560.
- Niess, H., Camaj, P., Renner, A., Ischenko, I., Zhao, Y., Krebs, S.,...Bruns, C. J. (2015). Side population cells of pancreatic cancer show characteristics of cancer stem cells responsible for resistance and metastasis. *Targeted Oncology*, 10(2): 215-227.
- Noctor, S. C., Martínez-Cerdeño, V., Ivic, L., Kriegstein, A. R. (2004). Cortical neurons arise in symmetric and asymmetric division zones and migrate through specific phases. *Nature Neuroscience*, 7(2): 136-144.
- Nolan-Stevaux, O., Lau, J., Truitt, M. L., Chu, G. C., Hebrok, M., Fernández-Zapico, M. E., & Hanahan, D. (2009). GLI1 is regulated through Smoothened-independent mechanisms in neoplastic pancreatic ducts and mediates PDAC cell survival and transformation. *Genes & Development*, 23(1): 24-36.
- Nomura, A., Gupta, V. K., Dauer, P., Sharma, N. S., Dudeja, V., Merchant, N.,...Banerjee, S. (2018). NFκB-mediated invasiveness in CD133⁺ pancreatic TICs is regulated by autocrine and paracrine activation of IL1 signaling. *Molecular Cancer Research*, 16(1): 162-172.
- Nomura, A., McGinn, O., Dudeja, V., Sangwan, V., Saluja, A. K., & Banerjee, S. (2015). Minnelide effectively eliminates CD133(+) side population in pancreatic cancer. *Molecular Cancer*, 14: 200.
- O'Brien, C. A., Kreso, A., & Jamieson, C. H. (2010). Cancer stem cells and self-renewal. *Clinical Cancer Research*, 16(12): 3113-3120.

- Oda, K., Matsuoka, Y., Funahashi, A., & Kitano, H. (2005). A comprehensive pathway map of epidermal growth factor receptor signaling. *Molecular Systems Biology*, 1: 2005.0010.
- Oettle, H., Neuhaus, P., Hochhaus, A., Hartmann, J. T., Gellert, K., Ridwelski, K.,...Riess, H. (2013). Adjuvant chemotherapy with gemcitabine and long-term outcomes among patients with resected pancreatic cancer: The CONKO-001 randomized trial. *The Journal of the American Medical Association*, 310(14): 1473-1481.
- Ornitz, D. M. & Itoh, N. (2015). The fibroblast growth factor signaling pathway. *Wiley Interdisciplinary Reviews Developmental Biology*, 4(3): 215-266.
- Ouyang, W., Li, J., Ma, Q., & Huang, C. (2006). Essential roles of PI3K/Akt/IKKbeta/NFkappaB pathway in cyclin D1 induction by arsenite in JB6 Cl41 cells. *Carcinogenesis*, 27(4):864-873.
- Pacold, M. E., Suire, S., Perisic, O., Lara-Gonzalez, S., Davis, C. T., Walker, E. H.,...Williams, R. L. (2000). Crystal structure and functional analysis of Ras binding to its effector phosphoinositide 3-kinase gamma. *Cell*, 103(6): 931-943.
- Pardal, R., Clarke, M. F., & Morrison, S. J. (2003). Applying the principles of stem-cell biology to cancer. *Nature Reviews Cancer*, 3(12): 895-902.
- Pastrana, E., Silva-Vargas, V., & Doetsch, F. (2011). Eyes wide open: a critical review of sphere-formation as an assay for stem cells. *Cell Stem Cell*, 8(5): 486-498.
- Patrawala, L., Calhoun, T., Schneider-Broussard, R., Li, H., Bhatia, B., Tang, S.,...Tang, D. G. (2006). Highly purified CD44⁺ prostate cancer cells from xenograft human tumors are enriched in tumorigenic and metastatic progenitor cells. *Oncogene*, 25(12): 1696-1708.
- Polakis, P. (2000). Wnt signaling and cancer. *Genes & Development*, 14(15): 1837-1851.
- Ponti, D., Costa, A., Zaffaroni, N., Pratesi, G., Petrangolini, G., Coradini, D.,...Daidone, M. G. (2005). Isolation and in vitro propagation of tumorigenic breast cancer cells with stem/progenitor cell properties. *Cancer Research*, 65(13): 5506-5511.
- Pour, P. M. & Kazakoff, K. (1996). Stimulation of islet cell proliferation enhances pancreatic ductal carcinogenesis in the hamster model. *American Journal of Pathology*, 149(3): 1017-1025.

- Pour, P. M., Kazakoff, K., & Carlson, K. (1990). Inhibition of streptozotocin-induced islet cell tumors and N-nitrosobis(2-oxopropyl)amine-induced pancreatic exocrine tumors in Syrian hamsters by exogenous insulin. *Cancer Research*, 50(5): 1634-1639.
- Pour, P. M., Pandey, K. K., & Batra, S. K. (2003). What is the origin of pancreatic adenocarcinoma? *Molecular Cancer*, 2: 13.
- Pour, P. M. & Schmied, B. M. (1999). One thousand faces of Langerhans islets. *International Journal of Pancreatology*, 25(3): 181-193.
- Pour, P. M., Schmied, B. M., Ulrich, A. B., Friess, H., Andren-Sandberg, A., & Buchler, M. W. (2001). Abnormal differentiation of islet cells in pancreatic cancer. *Pancreatology*, 1(2): 110-116.
- Pramanik, K. C., Makena, M. R., Bhowmick, K., & Pandey, M. K. (2018). Advancement of NF- κ B signaling pathway: A novel target in pancreatic cancer. *International Journal of Molecular Sciences*, 19(12). pii: E3890.
- Prior, I. A., Lewis, P. D., & Mattos, C. (2012). A comprehensive survey of Ras mutations in cancer. *Cancer Research*, 72(10): 2457-2467.
- Qin, L., Dong, Z., & Zhang, J. T. (2014). Reversible epigenetic regulation of 14-3-3 σ expression in acquired gemcitabine resistance by uhrf1 and DNA methyltransferase 1. *Molecular Pharmacology*, 86(5): 561-569.
- Quint, K., Tonigold, M., Di Fazio, P., Montalbano, R., Lingelbach, S., Rückert, F.,...Neureiter, D. (2012). Pancreatic cancer cells surviving gemcitabine treatment express markers of stem cell differentiation and epithelial-mesenchymal transition. *International Journal of Oncology*, 41(6): 2093-2102.
- Regan, J. L., Schumacher, D., Staudte, S., Steffen, A., Haybaeck, J., Keilholz, U.,...Lange, M. (2017). Non-canonical hedgehog signalling is a positive regulator of the WNT pathway and is required for the survival of colon cancer stem cells. *Cell Reports*, 21(10): 2813-2828.
- Reid, C. B., Tavazoie, S. F., & Walsh, C. A. (1997). Clonal dispersion and evidence for asymmetric cell division in ferret cortex. *Development*, 124(12): 2441-2450.
- Réjiba, S., Wack, S., Aprahamian, M., & Hajri, A. (2007). K-ras oncogene silencing strategy reduces tumor growth and enhances gemcitabine chemotherapy efficacy for pancreatic cancer treatment. *Cancer Science*, 98(7): 1128-1136.

- Reynolds, B. A., Tetziaff, W., & Weiss, S. (1992). A multipotent EGF-responsive striatal embryonic progenitor cell produces neurons and astrocytes. *Journal of Neuroscience*, 12(11): 4565-4574.
- Reynolds, B. A. & Weiss, S. (1992). Generation of neurons and astrocytes from isolated cells of the adult mammalian central nervous system. *Science*, 255: 1707-1710.
- Ricci-Vitiani, L., Lombardi, D. G., Piloizzi, E., Biffoni, M., Todaro, M., Peschle, C., & De Maria, R. (2007). Identification and expansion of human colon-cancer-initiating cells. *Nature*, 445(7123): 111-115.
- Roux, P. P. & Blenis, J. (2004). ERK and p38 MAPK-activated protein kinases: a family of protein kinases with diverse biological functions. *Microbiology and Molecular Biology Reviews*, 68(2): 320-344.
- Rovira, M., Scott, S. G., Liss, A. S., Jensen, J., Thayer, S. P., & Leach, S. D. (2010). Isolation and characterization of centroacinar/terminal ductal progenitor cells in adult mouse pancreas. *Proceedings of the National Academy of Sciences of the United States of America*, 107(1): 75-80.
- Sainz, B. Jr., Alcala, S., Garcia, E., Sanchez-Ripoll, Y., Azevedo, M. M., Cioffi, M.,...Heeschen, C. (2015). Microenvironmental hCAP-18/LL-37 promotes pancreatic ductal adenocarcinoma by activating its cancer stem cell compartment. *Gut*, 64(12): 1921-1935.
- Saito, Y., Uchida, N., Tanaka, S., Suzuki, N., Tomizawa-Murasawa, M., Sone, A.,...Ishikawa, F. (2010). Induction of cell cycle entry eliminates human leukemia stem cells in a mouse model of AML. *Nature Biotechnology*, 28(3): 275-280.
- Sakurai, H., Chiba, H., Miyoshi, H., Sugita, T., & Toriumi, W. (1999). IkkappaB kinases phosphorylate NF-kappaB p65 subunit on serine 536 in the transactivation domain. *Journal of Biological Chemistry*, 274(43): 30353-30356.
- Sancho, P., Burgos-Ramos, E., Tavera, A., Bou Kheir, T., Jagust, P., Schoenhals, M.,...Heeschen, C. (2015). MYC/PGC-1 α balance determines the metabolic phenotype and plasticity of pancreatic cancer stem cells. *Cell Metabolism*, 22(4): 590-605.
- Sarkar, A. & Hochedlinger, K. (2013). The sox family of transcription factors: versatile regulators of stem and progenitor cell fate. *Cell Stem Cell*, 12(1): 15-30.
- Sasaki, C. Y., Barberi, T. J., Ghosh, P., & Longo, D. L. (2005). Phosphorylation of RelA/p65 on serine 536 defines an

- IkappaB α -independent NF- κ B pathway. *Journal of Biological Chemistry*, 280(41): 34538-34547.
- Sato, S., Rancourt, A., Sato, Y., & Satoh, M. S. (2016). Single-cell lineage tracking analysis reveals that an established cell line comprises putative cancer stem cells and their heterogeneous progeny. *Scientific Reports*, 6: 23328.
- Scaltriti, M. & Baselga, J. (2006). The epidermal growth factor receptor pathway: a model for targeted therapy. *Clinical Cancer Research*, 12(18): 5268-5272.
- Scheffzek, K., Ahmadian, M. R., Kabsch, W., Wiesmuller, L., Lautwen, A., Schmitz, & Wittinghofer, A. (1997). The Ras-RasGAP complex: structural basis for GTPase activation and its loss in oncogenic Ras mutants. *Science*, 277(5324): 333-338.
- Schmidt, A. & Hall, A. (2002). Guanine nucleotide exchange factors for Rho GTPases: turning on the switch. *Genes and Development*, 16(13): 1587-1609.
- Schmied, B. M., Ulrich, A. B., Matsuzaki, H., Li, C., Friess, H., Buchler, M. W.,...Pour, P. M. (2000). Alteration of the Langerhans islet in pancreatic cancer patients. *International Journal of Pancreatology*, 28(3): 187-197.
- Seery, J. P. & Watt, F. M. (2000). Asymmetric stem-cell divisions define the architecture of human oesophageal epithelium. *Current Biology*, 10(22): 1447-1450.
- Seguin, L., Kato, S., Franovic, A., Camargo, M. F., Lesperance, J., Elliott, K. C.,...Cheresh, D. A. (2014). An Integrin β_3 -KRAS-RalB complex drives tumour stemness and resistance to EGFR inhibition. *Nature Cell Biology*, 16(5): 457-468.
- Seino, T., Kawasaki, S., Shimokawa, M., Tamagawa, H., Toshimitsu, K., Fujii, M.,...Sato, T. (2018). Human pancreatic tumor organoids reveal loss of stem cell niche factor dependence during disease progression. *Cell Stem Cell*, 22(3): 454-467.
- Sergeant, G., Vankelecom, H., Gremeaux, L., & Topal, B. (2009). Role of cancer stem cells in pancreatic ductal adenocarcinoma. *Nature Reviews Clinical Oncology*, 6(10): 580-586.
- Shackleton, M. (2010). Normal stem cells and cancer stem cells: similar and different. *Seminars in Cancer Biology*, 20(2): 85-92.
- Shackleton, M., Quintana, E., Fearon, E. R., & Morrison, S. J. (2009). Heterogeneity in cancer: cancer stem cells versus clonal evolution. *Cell*, 138(5): 822-829.

- Shafee, N., Smith, C. R., Wei, S., Kim, Y., Mills, G. B., Hortobagyi, G. N.,...Lee, E. Y. (2008). Cancer stem cells contribute to cisplatin resistance in Brca1/p53-mediated mouse mammary tumors. *Cancer Research*, 68(9): 3243-3250.
- Shao, D. D., Xue, W., Krall, E. B., Bhutkar, A., Piccioni, F., Wang, X.,...Hahn, W. C. (2014). KRAS and YAP1 converge to regulate EMT ad tumor survival. *Cell*, 158(1): 171-184.
- Sharpe, H. J., Pau, G., Dijkgraaf, G. J., Basset-Seguín, N., Modrusan, Z., Januario, T.,...de Sauvage, F. J. (2015). Genomic analysis of smoothened inhibitor resistance in basal cell carcinoma. *Cancer Cell*, 27(3): 327-341.
- Shima, F., Yoshikawa, Y., Ye, M., Araki, M., Matsumoto, S., Liao, J.,...Kataoka, T. (2013). In silico discovery of small-molecule Ras inhibitors that disolay antitumor activity by blocking the Ras-effector interaction. *Proceedings of the National Academy of Sciences of the United States of America*, 110(20): 8182-8187.
- Simeone, D. M. (2008). Pancreatic cancer stem cells: implications for the treatment of pancreatic cancer. *Clinical Cancer Research*, 14(18):5646-5648.
- Singh, S. K., Clarke, I. D., Terasaki, M., Bonn, V. E., Hawkins, C., Squire, J., & Dirks, P. B. (2003). Identification of a cancer stem cell in human brain tumors. *Cancer Research*, 63: 5821-5828.
- Singh, J. K., Farnie, G., Bundred, N. J., Simões, B. M., Shergill, A., Landberg, G.,...Clarke, R. B. (2013). Targeting CXCR1/2 significantly reduces breast cancer stem cell activity and increases the efficacy of inhibiting HER2 via HER2-dependent and -independent mechanisms. *Clinical Cancer Research*, 19(3): 643-656.
- Singh, B. N., Fu, J., Srivastava, R. K., & Shankar, S. (2011). Hedgehog signaling antagonist GDC-0449 (Vismodegib) inhibits pancreatic cancer stem cell characteristics: molecular mechanisms. *PLoS One*, 6(11): e27306.
- Singh, S. K., Hawkins, C., Clarke, I. D., Squire, J., Bayani, J., Hide, T.,...Dirks, P. B. (2004). Identification of human brain tumour initiating cells. *Nature*, 432(7015): 396-401.
- Singh, S., Trevino, J., Bora-Singhal, N., Coppola, D., Haura, E., Altiock, S., & Chellappan, S. P. (2012). EGFR/Src/Akt signaling modulates SOX2 expression and self-renewal of stem-like side-population cells in non-small cell lung cancer. *Molecular Cancer*, 11: 73.

- Spiegel, J., Cromm, P. M., Zimmermann, G., Grossmann, T. N., & Waldmann, H. (2014). Small-molecule modulation of Ras signaling. *Nature Chemical Biology*, 10(8): 613-622.
- Stambolic, V. & Woodgett, J. R. (1994). Mitogen inactivation of glycogen synthase kinase-3 beta in intact cells via serine 9 phosphorylation. *The Biochemical Journal*, 303 (Pt 3): 701-704.
- Stark, A. & Eibl, G. (2015). Pancreatic Ductal Adenocarcinoma. *Pancreapedia: Exocrine Pancreas Knowledge Base*, DOI: 10.3998/panc.2015.14.
- Steg, A. D., Bevis, K. S., Katre, A. A., Ziebarth, A., Dobbin, Z. C., Alvarez, R. D.,...Landen, C. N. (2011). Stem cell pathways contribute to clinical chemoresistance in ovarian cancer. *Clinical Cancer Research*, 18(3): 869-881.
- Sun, Y., Kong, W., Falk, A., Hu, J., Zhou, L., Pollard, S., & Smith, A. (2009). CD133 (Prominin) negative human neural stem cells are clonogenic and tripotent. *PLoS ONE*, 4: e5498.
- Sun, L., Mathews, L. A., Cabarcas, S. M., Zhang, X., Yang, A., Zhang, Y.,...Farrar, W. L. (2013). Epigenetic regulation of SOX9 by the NF- κ B signaling pathway in pancreatic cancer stem cells. *Stem Cells*, 31(8): 1454-1466.
- Sutherland, C., Leighton, I. A., Cohen, P. (1993). Inactivation of glycogen synthase kinase-3 beta by phosphorylation: new kinase connections in insulin and growth-factor signalling. *The Biochemical Journal*, 296 (Pt 1): 15-19.
- Taipale, J. & Beachy, P. A. (2001). The Hedgehog and Wnt signalling pathways in cancer. *Nature*, 411(6835): 349-354.
- Takahashi, K., Tanabe, K., Ohnuki, M., Narita, M., Ichisaka, T., Tomoda, K., & Yamanaka, S. (2007). Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*, 131(5): 861-872.
- Takahashi, K. & Yamanaka, S. (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*, 126(4): 663-676.
- Takebe, N., Miele, L., Harris, P. J., Jeong, W., Bando, H., Kahn, M.,...Ivy, S. P. (2015). Targeting Notch, Hedgehog, and Wnt pathways in cancer stem cells: clinical update. *Nature Reviews Clinical Oncology*, 12(8): 445-464.
- Tam, W. L. & Ng, H. H. (2014). Sox2: Masterminding the root of cancer. *Cancer Cell*, 26(1): 3-5.

- Tan, B. T., Park, C. Y., Ailles, L. E., & Weissman, I. L. (2006). The cancer stem cell hypothesis: a work in progress. *Laboratory Investigation*, 86(12): 1203-1207.
- Thayer, S. P., di Magliano, M. P., Heiser, P. W., Nielsen, C. M., Roberts, D. J., Lauwers, G. Y.,...Hebrok, M. (2003). Hedgehog is an early and late mediator of pancreatic cancer tumorigenesis. *Nature*, 425(6960): 851-856.
- Thomas, M. L., Coyle, K. M., Sultan, M., & Marcato, P. (2015). Cancer stem cells and chemoresistance: Strategies to overcome therapeutic resistance. In: Babashah, S. (eds) *Cancer stem cells: Emerging concepts and future perspectives in translational oncology*. Springer, Cham.
- Ulrich, A. B., Standop, J., Schmied, B. M., Schneider, M. B. Lawson, T. A., & Pour, P. M. (2002). Species differences in the distribution of drug-metabolizing enzymes in the pancreas. *Toxicologic Pathology*, 30(2): 247-253.
- Vandermoere, F., El Yazidi-Belkoura, I., Adriaenssens, E., Lemoine, J., & Hondermarck, H. (2005). The antiapoptotic effect of fibroblast growth factor-2 is mediated through nuclear factor-kappaB activation induced via interaction between Akt and IkappaB kinase-beta in breast cancer cells. *Oncogene*, 24(35): 5482-5491.
- Vasiliou, V. & Nebert, D. W. (2005). Analysis and update of the human aldehyde dehydrogenase (ALDH) gene family. *Human Genomics*, 2(2): 138-143.
- Verheyen, E. M. & Gottardi, C. J. (2010). Regulation of Wnt/beta-catenin signaling by protein kinases. *Developmental Dynamics*, 239(1): 34-44.
- Vetter, I. R. & Wittinghofer, A. (2001). The guanine nucleotide-binding switch in three dimensions. *Science*, 294(5545): 1299-1304.
- Viale, A., De Franco, F., Orleth, A., Cambiaghi, V., Giuliani, V., Bossi, D.,...Pelicci, P. G. (2009). Cell-cycle restriction limits DNA damage and maintains self-renewal of leukaemia stem cells. *Nature*, 457(7225): 51-56.
- Wang, V. M., Ferreira, R. M. M., Almagro, J., Evan, T., Legrave, N., Zaw Thin, M.,...Behrens, A. (2019). CD9 identifies pancreatic cancer stem cells and modulates glutamine metabolism to fuel tumour growth. *Nature Cell Biology*, 21(11): 1425-1435.
- Wang, K., Ji, W., Yu, Y., Li, Z., Niu, X., Xia, W., & Lu, S. (2018). FGFR1-ERK1/2-SOX2 axis promotes cell proliferation, epithelial-mesenchymal transition, and metastasis in FGFR1-amplified lung cancer. *Oncogene*, 37(39): 5340-5354.

- Wang, F., Ma, L., Zhang, Z., Liu, X., Gao, H., Zhuang, Y.,...Yang, Y. (2016). Hedgehog signaling regulates epithelial-mesenchymal transition in pancreatic cancer stem-like cells. *Journal of Cancer*, 7(4): 408-417.
- Wang, Y. K., Zhu, Y. L., Qiu, F. M., Zhang, T., Chen, Z. G., Zheng, S., & Huang, J. (2010). Activation of Akt and MAPK pathways enhances the tumorigenicity of CD133+ primary colon cancer cells. *Carcinogenesis*, 31(8): 1376-1380.
- Wei, D., Wang, L., Yan, Y., Jia, Z., Gagea, M., Li, Z.,...Xie, K. (2016). KLF4 is essential for induction of cellular identity change and acinar-to ductal reprogramming during early pancreatic carcinogenesis. *Cancer Cell*, 29(3): 324-338.
- Wen, J., Park, J. Y., Park, K. H., Chung, H. W., Bang, S., Park, S. W., & Song, S. Y. (2010). Oct4 and Nanog expression is associated with early stages of pancreatic carcinogenesis. *Pancreas*, 39(5): 622-626.
- Wennerberg, K., Rossman, K. L., & Der, C. J. (2005). The Ras superfamily at a glance. *Journal of Cell Science*, 118(Pt 5): 843-846.
- Wright, D. E., Cheshier, S. H., Wagers, A. J., Randall, T. D., Christensen, J. L., & Weissman, I. L. (2001). Cyclophosphamide/granulocyte colony-stimulating factor causes selective mobilization of bone marrow hematopoietic stem cells into the blood after M phase of the cell cycle. *Blood*, 97(8): 2278-2285.
- Wu, D. & Pan, W. (2010). GSK3: a multifaceted kinase in WNT signaling. *Trends in Biochemical Sciences*, 35(3): 161-168.
- Xia, Y. F., Ye, B. Q., Li, Y. D., Wang, J. G., He, X. J., Lin, X.,...Geng, J. G. (2004). Andrographolide attenuates inflammation by inhibition of NF-kappa B activation through covalent modification of reduced cysteine 62 of p50. *Journal of Immunology*, 173(6): 4207-4217.
- Xiang, T., Long, H., He, L., Han, X., Lin, K., Liang, Z.,...Zhu, B. (2015). Interleukin-17 produced by tumor microenvironment promotes self-renewal of CD133+ cancer stem-like cells in ovarian cancer. *Oncogene*, 34(2): 165-176.
- Xu, C., Sun, X., Qin, S., Wang, H., Zheng, Z., Xu, S.,...Ren, H. (2015). Let-7a regulates mammosphere formation capacity through Ras/NF- κ B and Ras/MAPK/ERK pathway in breast cancer stem cells. *Cell Cycle*, 14(11): 1686-1697.
- Yamano, M., Fujii, H., Takagaki, T., Kadowaki, N., Watanabe, H., & Shirai, T. (2000). Genetic progression and divergence in pancreatic carcinoma. *American Journal of Pathology*, 156(6): 2123-2133.

- Yan, Y., Li, Z., Kong, X., Jia, Z., Zuo, X., Gagea, M.,...Xie, K. (2016). KLF4-mediated suppression of CD44 signaling negatively impacts pancreatic cancer stemness and metastasis. *Cancer Research*, 76(8): 2419-2431.
- Yang, M. C., Wang, H. C., Hou, Y. C., Tung, H. L., Chiu, T. J., & Shan, Y. S. (2015). Blockade of autophagy reduces pancreatic cancer stem cell activity and potentiates the tumoricidal effect of gemcitabine. *Molecular Cancer*, 14: 179.
- Yauch, R. L., Dijkgraaf, G. J., Alicka, B., Januario, T., Ahn, C. P., Holcomb, T.,...de Sauvage, F. J. (2009). Smoothened mutation confers resistance to a Hedgehog pathway inhibitor in medulloblastoma. *Science*, 326(5952): 572-574.
- Yost, C., Torres, M., Miller, J. R., Huang, E., Kimelman, D., & Moon, R. T. (1996). The axis-inducing activity, stability, and subcellular distribution of beta-catenin is regulated in *Xenopus* embryos by glycogen synthase kinase 3. *Genes & Development*, 10(12): 1443-1454.
- Yu, F., Li, J., Chen, H., Fu, J., Ray, S., Huang, S.,... Ai, W. (2011). Kruppel-like factor 4 (KLF4) is required for maintenance of breast cancer stem cells and for cell migration and invasion. *Oncogene*, 30(18): 2161-2172.
- Zeng, G., Germinaro, M., Micsenyi, A., Monga, N. K., Bell, A., Sood, A.,...Monga, S. P. (2006). Abberant Wnt/ β -catenin signalling in pancreatic adenocarcinoma. *Neoplasia*, 8(4): 279-289.
- Zhang, Z., Duan, Q., Zhao, H., Liu, T., Wu, H., Shen, Q.,...Yin, T. (2016). Gemcitabine treatment promotes pancreatic cancer stemness through the Nox/ROS/NF- κ B/STAT3 signaling cascade. *Cancer Letters*, 382(1): 53-63.
- Zhang, Y., Morris, JP 4th, Yan, W., Schofield, H. K., Gurney, A., Simeone, D. M.,...Pasca di Magliano, M. (2013). Canonical wnt signaling is required for pancreatic carcinogenesis. *Cancer Research*, 73(15): 4909-4922.
- Zhang, W., Tan, W., Wu, X., Poustovoitov, M., Strasner, A., Li, W.,...Karin, M. (2013). A NIK-IKK α module expands ErbB2-induced tumor-initiating cells by stimulating nuclear export of p27/Kip1. *Cancer Cell*, 23(5): 647-659.
- Zhao, C., Chen, A., Jamieson, C. H., Fereshteh, M., Abrahamsson, A., Blum, J.,...Reya, T. (2009). Hedgehog signalling is essential for maintenance of cancer stem cells in myeloid leukaemia. *Nature*, 458(7239): 776-779.

Zhao, D., Pan, C., Sun, J., Gilbert, C., Drews-Elger, K., Azzam, D. J.,...Slingerland, J. M. (2015). VEGF drives cancer-initiating stem cells through VEGFR-2/Stat3 signaling to upregulate Myc and Sox2. *Oncogene*, 34(24): 3107-3119.

