



Polygonum minus: A tropical medicinal herb with vast applications in food, agricultural, and medicinal fields

Zhongming Yang^a, Xi Deng^a, Zhongguo Yang^b, Mingzhao Han^c, Norsharina Ismail^{a,*}, Kim Wei Chan^a, Ahmad Faizal Abdull Razis^{a,d}, Norhaizan Mohd Esa^{a,e}, Ket Li Ho^f, Md Zuki Abu Bakar^{a,g,**}

^a Natural Medicines and Products Research Laboratory, Institute of Bioscience, Universiti Putra Malaysia, 43400, UPM Serdang, Selangor, Malaysia

^b Potato Institute of Kong Shan, 636799, Tongjiang, Bazhong, China

^c Department of Land Management, Faculty of Agriculture, Universiti Putra Malaysia, 43400, UPM Serdang, Selangor, Malaysia

^d Department of Food Science, Faculty of Food Science and Technology, Universiti Putra Malaysia, 43400, UPM Serdang, Selangor, Malaysia

^e Department of Nutrition, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, 43400, UPM Serdang, Selangor, Malaysia

^f School of Pharmacy, International Medical University, Bukit Jalil, 57000, Kuala Lumpur, Malaysia

^g Department of Veterinary Preclinical Science, Faculty of Veterinary Medicine, Universiti Putra Malaysia, 43400, UPM Serdang, Selangor, Malaysia

ARTICLE INFO

Keywords:

Polygonum minus

Food application

Agricultural application

Medicinal application

ABSTRACT

Polygonum minus (PM) is a tropical medicinal herb that belongs to the family Polygonaceae, widely distributed in Southeast Asia, such as Malaysia, Indonesia, and Thailand. This herb has a sweet and pleasant aroma, commonly used as a flavoring agent. In addition, it is applied as a traditional medicine for multiple diseases, such as fungal infections, poor eyesight, and body aches. Due to its antioxidant and antimicrobial properties, PM is employed as a food additive for delaying or preventing the oxidation reaction of food and as an edible film for protecting food from microbiological contaminants. In livestock, poultry, and aquaculture industries, PM has been proposed as an alternative to traditional growth promoters with anti-parasitic properties. In respect of medicinal applications, PM has been proven to have protective effects on body systems and inhibitory effects on microbes. Moreover, nanotechnology has been applied to improve the efficacy and bioavailability of PM. This review aims to provide insight into the traditional uses, phytochemical properties, and toxicity, as well as applications of PM in food, agricultural, and medicinal fields.

1. Introduction

Polygonum minus (PM) is an annual medicinal herb (Huda-Faujan, Noriham, Norrakiah, & Babji, 2009; Murakami, Ali, Mat-Salleh, Koshimizu, & Ohigashi, 2000) that belongs to the family Polygonaceae (Rahnamaie-Tajadod, Loke, Goh, & Noor, 2017), with sweet and pleasant aroma (Baradaran, HooiLing, ChinChin, & Rahim, 2012). It is a slender, creeping shrub and can reach up to a height of 1.0 m in the lowlands and up to 1.5 m in the highlands (Vikram, Chiruvella, Ripain, & Arifullah, 2014). The stem is a slender twining shrub; its leaves are long and lance-shaped, 5–7 cm long and 0.5–2.0 cm wide. Moreover, the stem is cylindrical, green, and slightly reddish, while the internodes are

short with nodes (P. V. Christopher et al., 2015). The morphology of PM is shown in Fig. 1. Furthermore, PM is widely distributed in Europe (Narasimhulu & Mohamed, 2014) such as Britain, Scandinavia, Spain (Bunawan, Talip, & Noor, 2011), and Southeast Asian (Gattuso, 2001; Udani, George, Musthapa, Pakdaman, & Abas, 2014) such as Malaysia (Qader, Ameen Abdulla, Chua, & Hamdan, 2012), Indonesia (Rusdi, Goh, & Baharum, 2016), Thailand (Vikram et al., 2014; Zakaria, Baba, Ku Bahaudin, & Hamdana, 2015), Vietnam (Rusdi et al., 2016), Laos (Ahmad et al., 2014), and Sri Lanka (Panase & Tiptacho, 2018). Fig. 2 shows the main distribution areas of PM in the world. PM, as synonyms to *Polygonum kawagoeanum*, *Persicaria minor* (Bunawan, Choong, Md-Zain, Baharum, & Noor, 2011), and *Persicaria tenella* (Chia et al.,

* Corresponding author. Natural Medicines and Products Research Laboratory, Institute of Bioscience, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia.

** Corresponding author. Natural Medicines and Products Research Laboratory, Institute of Bioscience, Universiti Putra Malaysia, 43400, UPM Serdang, Selangor, Malaysia.

E-mail addresses: norsharina@upm.edu.my (N. Ismail), zuki@upm.edu.my (M.Z. Abu Bakar).

<https://doi.org/10.1016/j.fbio.2024.104511>

Received 17 March 2024; Received in revised form 3 June 2024; Accepted 4 June 2024

Available online 8 June 2024

2212-4292/© 2024 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

2020), is called as Pygmy smartweed/knotweed in English (Burkill, 1966; Malaysia Biodiversity Information System (MyBIS)), kesum in Malaysia (Aziman, Abdullah, Mohd Noor, Zulkifli, & Wan Kamarudin, 2012; Johnny, Yusuf, & Nulit, 2011; Mohd Ghazali, Al-Naqeb, Krishnan Selvarajan, Hazizul Hasan, & Adam, 2014; Roslan et al., 2012; Shukor, Ismail, Zainal, & Noor, 2013), daun laksa in Singapore (Vimala, Rohana, Rashih, & M, 2011), phakphai in Thailand (Vikram et al., 2014), Kleiner Knöterich in German, Chakhong-macha in Manipuri, and Nghê bé in Vietnamese (P. V. Christopher, Parasuraman, Asmawi, & Murugaiyah, 2017). Additionally, PM has various vernacular names such as kesom (Imelda, Faridah, & Kusumaningrum, 2014), laksa leaves (Abas, Lajis, Israf, Khozirah, & Kalsom, 2006; Noor Hashim, Abas, Shaari, & Lajis, 2012), kusum (Bashir, Aziz, & Noor, 2020), daun laksa or cenohom (Ong & Nordiana, 1999), Jarak Belanda, Kunyit Jawa, Kelima Paya (Qader et al., 2011), cambodian mint, vietnamese mint, water pepper, and marsh pepper (Hamid et al., 2020).

The extensive use of PM, particularly in food and traditional medicinal fields, has attracted the attention of researchers, and various review studies have reported the phytochemical constituents, medicinal properties, or bioactive properties of PM (Asanai, Haron, Camalxaman, & Mohamed, 2020; Nadzirah, Rusop, & Noriham, 2014; Narasimhulu, Reddy, & Mohamed, 2014; Paritalac et al., 2014). However, few researchers have attempted to review the application progress of PM in various fields in recent years. Therefore, the current knowledge about the food, agricultural, and medicinal field applications of PM is reviewed in this article. In addition, we put forward the issues that need to be resolved in future investigations, which is meaningful for broadening PM applications.

2. Phytochemical

2.1. Phytochemical compositions

The diverse chemical constituents present in medicinal plants possess biological attributes that could improve human health through applications in the food and pharmaceutical fields. Additionally, these constituents hold significant values in the perfume, agrochemical, and cosmetic industries (Yi, Cao, Cao, & Xiao, 2019). The functional characteristics of PM are significant due to its diverse range of bioactive compounds, commonly known as secondary metabolites or phytochemicals. A study has demonstrated that PM is abundant in over 70

phytochemicals, including terpenes (monoterpenes and sesquiterpenes), aliphatic compounds, and organic acids (Ahmad et al., 2014). Moreover, to facilitate further research on the single compounds in PM, we summarized the main phytochemicals present in PM as well as their respective activities and/or applications, which are listed in Table 1.

2.2. Factors affecting the phytochemical compositions of PM

The phytochemicals or secondary metabolites of PM are commonly produced to respond to external stimuli including infection, nutrition, or alteration of climatic conditions. These phytochemicals are abundantly found in leaves, stems, and roots, but different compounds are synthesized in specific parts of the PM. Because of this, the phytochemical compositions of PM in different locations and environments are different. The factors affecting the phytochemical compositions of PM are summarized in Fig. 3.

The phytochemical compositions of PM may vary depending on the geographical location, climatic conditions, and varietal differences (Ma, Chai, Hou, Zhao, & Meng, 2022; Punia Bangar, Dunno, Kumar, Mostafa, & Maqsood, 2022; Sabaragamuwa, Perera, & Fedrizzi, 2018; Wong, Li, Li, Razmovski-Naumovski, & Chan, 2011). Notably, PM leaves from Genting Highland in Pahang, Ulu Yam in Selangor, and the lab (a controlled environment lab culturing the cuttings of PM from Genting Highland in Pahang. At 22/16 °C day/night temperatures under 12 h light/dark photoperiod with light intensity of $170 \pm 20 \mu\text{mol m}^{-2} \text{s}^{-1}$ at ~75% RH.) contain different compounds, with a total of 48, 39, and 44 compounds, respectively (Baharum, Bunawan, Ghani, Wan, & Noor, 2010; Rahnamaie-Tajadod, Goh, & Mohd Noor, 2019; Rusdi et al., 2016).

Meanwhile, studies showed that the variation in composition profiles of plant material is influenced by various factors, including the specific plant part utilized, such as barks, leaves, root barks, or buds (Graf & Stappen, 2022; Ribeiro-Santos et al., 2017). Despite the same extraction and identification methods applied to examine the constituents in PM leaves, stems, and roots from the same plant, distinct phytochemicals were obtained (Ahmad, Bunawan, Normah, & Baharum, 2016). In other words, PM leaves, stems, and roots from the same plant contain different compounds. In addition, the extraction techniques also affects the phytochemical composition (Cheng et al., 2023). Ahmad et al. (2014) used Solid Phase Microextraction (SPME) and hydrodistillation coupled with GC-MS to identify the volatile compounds present in the leaves,



Fig. 1. Appearance of *Polygonum minus* (PM). (A) Plant morphology of PM; (B) Whole plant view of PM; (C) Leaves of PM; (D) Stem of PM.

stems, and roots. Specifically, about 300 mg of fresh PM was ground with liquid nitrogen and placed in a 20 mL vial. Then 700 μ L of distilled water was added to the ground leaves and the vial was covered tightly using a hole cap with septum to ensure no volatile could escape during the extraction. The fiber was then exposed to the sample headspace by inserting the fiber through the septum and the vial with the exposed fiber was incubated in a water bath at 45 °C for 15 min. After 15 min, the fiber was thermally desorbed by inserting the fiber into the GC injector at 250 °C for 10 min. On the other hand, essential oil was isolated by the hydrodistillation technique. PM (300 g) were subjected to hydrodistillation with 2 L of distilled water for 8 h using a Clevenger-type apparatus to produce a yellowish essential oil. The essential oils were collected over water, separated, dried over nitrogen gas, and stored. As a result, even though the same plant and the same part were used, different compounds were obtained due to the use of SPME or hydrodistillation respectively, as summarized in Fig. 4.

Besides, the compositions, nutritional values, and functional attributes of phytochemicals are influenced by postharvest interventions, encompassing processing (analytical methods), preservation practices, and storage methodologies (Alwazeer et al., 2023; S. Bakir et al., 2023; Cano-Lamadrid, Martínez-Zamora, Castillejo, & Artés-Hernández, 2022; Cautela, Vella, & Laratta, 2019; Dhimi & Mishra, 2015; Fratianni et al., 2017; Hassan Sakar et al., 2021; Sęczyk, Ahmet Ozdemir, & Kołodziej, 2022). Markom, Hassim, Anuar, and Baharum (2012) found that 70% methanol yielded the highest total extract (33.1%) compared to other co-solvents (water, methanol, ethanol, 50% methanol, 50% ethanol, and 70% ethanol), and exhibited the highest levels of total phenolic content (TPC), total flavonoid content (TFC), ferric-reducing antioxidant power (FRAP), and 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical inhibition. Hassim, Markom, Anuar, and Baharum (2014) corroborated these findings. Conversely, Kartikasari, Rahman, Puspasari, and Ridha (2022) reported that the 50% ethanol extract of PM leaves exhibited the highest TPC, while the methanol extract showed the highest TFC. Moreover, Hassim et al. (2015) observed the ethanol co-solvent system successfully extracted all three aldehydes (decanal, dodecanal, and undecanal). Shevchenko, Muzychikina, Ross, and Korul'kin (2019) concluded that the optimal extraction of the polyphenolic phytocomplex from PM plant raw materials was achieved through repetitive extraction using 50% ethanol or 50% acetone as solvents at their boiling point. In another study, Syaiful, Jayuska, and Harlia (2015) utilized steam distillation techniques to investigate the impact of time variations on the

compounds present in PM leaves. Following that, Azhari, Markom, Ismail, and Anuar (2022) utilized supercritical fluid extraction with carbon dioxide to examine the impact of CO₂ flow rates (2 mL/min, 3 mL/min, and 4 mL/min) on essential oil yield and β -caryophyllene yields from PM roots, as summarized in Fig. 5.

Overall, different processing methods led to variations in the major constituents of the plant extract, and the yield of each principal component was dependent on the specific extraction techniques employed (Ullah, Wilfred, & Shaharun, 2019). Moreover, the type and conditions of the drying treatments influenced the retention of phytochemicals. The findings of Mahanom, Azizah, and Dzulkifly (1999) indicated the herbal preparation produced through oven drying exhibited lower levels of phytochemical content compared to that obtained through freeze-drying. Azhari, Markom, Ismail, and Anuar (2020) also had similar views. Their findings revealed that an increase in drying temperature reduced essential oil yield and resulted in the loss of major chemical compounds in PM roots, and air-drying emerged as the most effective for preserving crucial chemical compounds in PM roots, including β -caryophyllene (1.43%), pentadecane (4.34%), hexadecanoic acid (3.91%), and oleic acid (3.97%). Nonetheless, all types and conditions of drying treatments were able to retain significant amounts of phytochemicals, especially carotenoids, ascorbic acid, niacin, and riboflavin (Mahanom et al., 1999).

In conclusion, the diversity observed in the composition profiles, quantities of individual components, and overall yield of oil or extract from plant materials stems from various factors. These factors encompass the choice of solvent during extraction, the physical characteristics of the sample, the plant species, the specific plant part utilized (such as barks, leaves, root barks, or buds), geographical origin, preservation methods, and encompassing processing (utilized extraction techniques and analytical methods) (Ribeiro-Santos et al., 2017).

3. Toxicity

The main concern in the toxicity of herbal remedies is their capacity to produce lethality and antagonistic impacts when consumed (Azad, Sulaiman, & Kundu, 2022). The prominent uniqueness of herbal plants is the presence of secondary metabolites as a natural defense toward plant growth. They can offer significant benefits but can also be harmful when consumed excessively (van Wyk & Prinsloo, 2020). Therefore, the valuation of toxicity is urgent and vital for medicinal plants to be used in

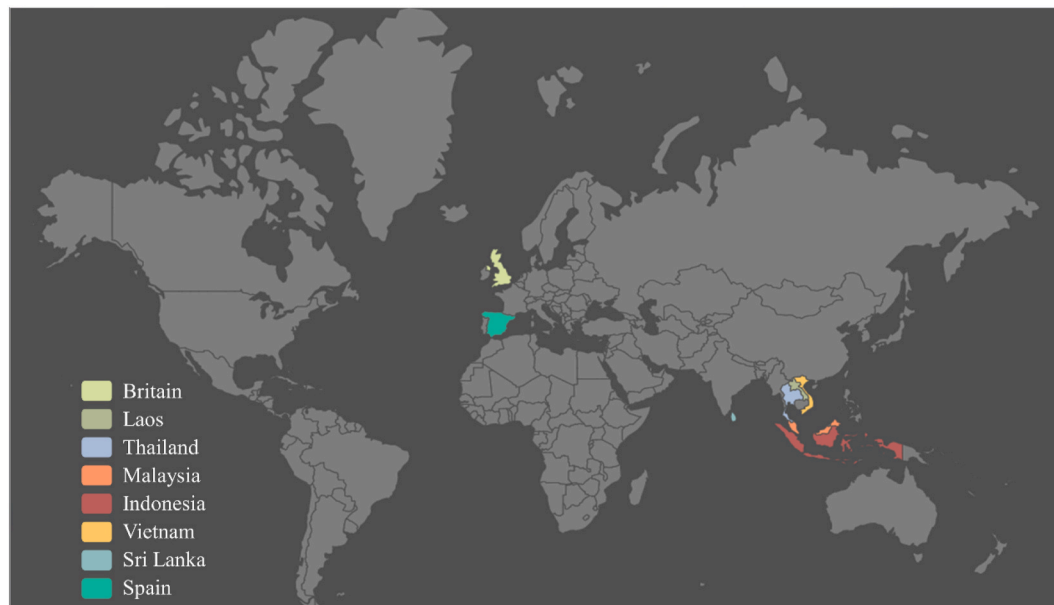


Fig. 2. Main distributions of PM in the world.

Table 1
Phytochemical compositions of PM with their activities and/or applications.

No.	Phytochemical classes	Composition	Chemical formula	Activities/Applications	Reference
1	Terpenes (Monoterpenes)	α -Pinene	C ₁₀ H ₁₆	Anti-microbial; Anti-oxidant; Anti-inflammatory; Chondroprotective	(A. C. R. Da Silva et al., 2012; Rufino et al., 2014)
2		α -Thujene	C ₁₀ H ₁₆	Anti-inflammatory; Anti-oxidant; Anti-microbial	Kazemi (2014)
3		Limonene	C ₁₀ H ₁₆	Anti-Anxiety; Pest Control; Cosmetic; Skincare Formulations; Anti-oxidant; Insect repellent	(Ciriminna, Lomeli Rodriguez, Demma Carà, Lopez Sanchez, & Pagliaro, 2014; Hollingsworth, 2005; Song et al., 2021)
4		β -Ocimene	C ₁₀ H ₁₆	Anti-fungal	Cavaleiro et al. (2015)
5		Myrtenal	C ₁₀ H ₁₄ O	Bronchodilatory; Anti-inflammatory; Anti-aggregative; Anti-hemolytic (<i>in vitro</i>); Anti-bacterial; Anti-tumor; Anti-hyperglycemic, Vasodilating; Heart rate reducing; Hypotensive; Anxiolytic; Anti-oxidant; Neuromodulatory; Sedative; Anti-diabetic; Analgesic; Neuroprotective	(Dragomanova, Tancheva, & Georgieva, 2018, 2019; Henriques et al., 2023)
6		-(Z)-Myrtenol	C ₁₀ H ₁₈ O	Sedative	De Sousa, Raphael, Brocksom, and Brocksom (2007)
7		Borneol	C ₁₀ H ₁₈ O	Analgesia; Putridity elimination; Flesh regeneration; Cell Protection; Anti-thrombotic	(Li, Sun, Zhang, & Wang, 2008; Xiong, Xiao, Xu, Wu, & Jiang, 2013)
8	Terpenes (sesquiterpenes)	(E)-Geranyl acetone	C ₁₃ H ₂₂ O	Anti-bacterial; Anti-fungal	Bonikowski, Świtakowska, and Kula (2015)
9		Germacrene D	C ₁₅ H ₂₄	Mosquito repellency; Anti-aphid; Anti-tick	(J. Liu et al., 2022)
10		Isocaryophyllene	C ₁₅ H ₂₄	Anti-fungi; Anti-cancer	(Legault & Pichette, 2010; Sabulal et al., 2006)
11		Copaene	C ₁₅ H ₂₄	Anti-oxidant; Cytotoxic; Anti-genotoxic	Türkez, Çelik, and Toğar (2014)
12		α -Zingiberene	C ₁₅ H ₂₄	Anticancer; Treatment of chronic wounds; Anti-inflammatory; Angiogenesis suppressing	(Ferreira et al., 2022; Seshadri et al., 2022)
13		δ -Elemene	C ₁₅ H ₂₄	Anti-cancer; Anti-oxidant; Reversal of Drug Resistance	Tan et al. (2021)
14		Aromadendrene	C ₁₅ H ₂₄	Anti-fungal; Anti-bacterial; Anti-viral; Plant growth regulatory; Anti-feedant; Repellent; Cytotoxic; Spasmolytic	(Asakawa, Yoyota, Takemoto, Kubo, & Nakanishi, 1980; Capon & MacLeod, 1988; Gaspar-Marques, Simões, & Rodríguez, 2004; Harada, Sakata, & Ina, 1984; Hubert, Okunade, & Wiemer, 1987; Matsuo, Atsumi, Nakayama, & Hayashi, 1981; Messer et al., 1990; Moreira, Lago, Young, & Roque, 2003; Murata et al., 1990; Pérez-Hernández, Ponce-Monter, Ortiz, Cariño-Cortés, & Joseph-Nathan, 2009; Su et al., 2008; Tada & Yasuda, 1985; Tommasi, Pizza, Conti, Orsi, & Stein, 1990)
15		(E)- α -Bergamotene	C ₁₅ H ₂₄	Insecticidal; Anti-diabetic; Anti-inflammatory; Anti-microbial; Anti-oxidant; Anti-cancer	(Ahmed et al., 2019; Annaz et al., 2023; Bayala et al., 2014; Magalhães et al., 2010; Niu et al., 2021; Urzúa et al., 2010; Ying et al., 2017)
16		β -Guaiene	C ₁₅ H ₂₄	Anti-inflammatory	Sahi (2016)
17		4,11-selinadiene	C ₁₅ H ₂₄	Not reported	\
18		β -Caryophyllene	C ₁₅ H ₂₄	Anti-inflammatory, Anti-diabetic, Anti-tumor, Neuroprotective; Anti-oxidant; Anti-convulsant; Analgesic; Myorelaxant; Sedative; Anti-depressive; Treatment Streptococcus infections, osteoporosis, and steatohepatitis	(Bahi et al., 2014; B. Bakir, Him, Özbek, Düz, & Tütüncü, 2008; Basha & Sankaranarayanan, 2014; Dahham et al., 2015; de Oliveira et al., 2018; Francomano et al., 2019; Legault & Pichette, 2010; Machado et al., 2020)
19		Eremophilene	C ₁₅ H ₂₄	Insecticidal; Anti-bacterial	(Deng et al., 2022; Utegenova et al., 2018)
20		β -farnesene	C ₁₅ H ₂₄	Insecticidal	Sun et al. (2011)
21		Sesquiphellandrene	C ₁₅ H ₂₄	Anti-viral	(Denver, Jackson, Loakes, Ellis, & Young, 1994; Joshi, Sunil Krishnan, & Kaushik, 2020; Zhuang et al., 2012)
22		Alloaromadendrene	C ₁₅ H ₂₄	Anti-oxidant	(Abd-ElGawad, Elshamy, El-Nasser El Gendy, Al-Rowaily, & Assaeed, 2019; Abd-Elgawad et al., 2021)
23		α -Bisabolene	C ₁₅ H ₂₄	Anti-inflammatory; Food flavoring; Anti-cancer	(Jou et al., 2016; C. Wang, Zada, Wei, & Kim, 2017; Y. Zhao et al., 2021)
24		α -Panasinsen	C ₁₅ H ₂₄	Not reported	\
25		α -Cedrene	C ₁₅ H ₂₄	Anti-obesity; Trypanocidal; Anti-leukemic	(T. H. Kim et al., 2015; Nibret & Wink, 2010)
26		Valencene	C ₁₅ H ₂₄	Anti-allergic; Anti-melanogenesis; Anti-septic; Anti-oxidant; Synergistic anti-colon cancer effect with doxorubicin	(Ambroz et al., 2015, 2017; Gallily, Yekhtin, & Hanuš, 2018; Jin, Lee, Kim, & Kim, 2011; K. Liu, Chen, Liu, Zhou, & Wang, 2012; Tsoyi et al., 2011; I. J. Yang, Lee, & Shin, 2016)
27		Nerolidol	C ₁₅ H ₂₆ O	Anti-oxidant; Anti-microbial; Anti-inflammatory; Anti-nociceptive; Anti-parasitic; Anti-ulcer; Skin penetration Enhancement; Insect repellent; Anti-cancer; Anti-biofilm; Insecticidal; Anti-tumor; Anti-nociceptive	(Biazi, Zanetti, Baranoski, Corveloni, & Mantovani, 2017; Chan, Tan, Chan, Lee, & Goh, 2016; Goel, Kaur, & Pahwa, 2016; Javed, Azimullah, Abul Khair et al., 2016; Lee et al., 2007; Nogueira Neto et al., 2013; Russo, 2011; Saito et al., 2016; Tatman & Mo, 2002)
28		α -Himachalene	C ₁₅ H ₂₄	Against coronavirus and HIV-1; Anti-cancer; Anti-inflammatory	(Ajlaoui et al., 2024; Chaudhary et al., 2014; Elias et al., 2019; Faris et al., 2023)
29		Cadinene	C ₁₅ H ₂₄	Anti-oxidant	Kundu, Saha, Wallia, Ahluwalia, and Kaur (2013)

(continued on next page)

Table 1 (continued)

No.	Phytochemical classes	Composition	Chemical formula	Activities/Applications	Reference
30		Gurjunene	C ₁₅ H ₂₄	Anti-oxidant; Anti-bacterial	Namata Abba, Ilagouma, Amadou, and Romane (2021)
31		Caryophyllene oxide	C ₁₅ H ₂₄ O	Anti-fungal; Anti-inflammatory; Analgesic; Anti-cancer; Insecticidal; Anti-feedant; Anti-platelet aggregation; Preservative in food, drugs, and cosmetics	(Bettarini et al., 1993; Fidy, Fiedorowicz, Strzadala, & Szumny, 2016; Gyrdymova & Rubtsova, 2022; Javed, Azimullah, Haque, & Ojha, 2016; Langenheim, 1994; Lin et al., 2003; Russo, 2011; Sain et al., 2014; D. Yang, Michel, Chaumont, & Millet-Clerc, 2000)
32		Humulene epoxide	C ₁₅ H ₂₄ O	Anti-oxidant	(P. Sharma & Shah, 2015)
33		Seychellene	C ₁₅ H ₂₄	Anti-inflammatory	(Luo et al., 2019; L. Zhang et al., 2021)
34		α-Curcumene	C ₁₅ H ₂₂	Anti-Tumor; Anti-breast cancer	(Al-Amin, Rahman, Khairuddean, & Salhimi, 2023; Y. Shin & Lee, 2013)
35		Cubenol	C ₁₅ H ₂₆ O	Anti-bacterial; Anti-oxidant; Anti-microbial; Anti-fungal; Insecticidal	(Basile et al., 2022; Bueno, Escobar, Martínez, Leal, & Stashenko, 2011; Djeddi et al., 2012; González et al., 2012; Mancini, De Martino, Malova, & De Feo, 2013; Moiteiro et al., 2013; Takao et al., 2012)
36		Thujopsene	C ₁₅ H ₂₄	Anti-fungal	Syahmina and Usuki (2020)
37		Longipinocarvone	C ₁₅ H ₂₂ O	Not reported	\
38		Aristolene	C ₁₅ H ₂₄	Not reported	\
39		dehydro- Cyclolongifolene oxide	C ₁₅ H ₂₂ O	Anti-oxidant	Abd-ElGawad et al. (2023)
40		α-Cadinol	C ₁₅ H ₂₆ O	Anti-mite; Anti-bacterial; Anti-fungal; Anti-oxidant	(Chang, Chen, Wang, & Wu, 2001; González et al., 2012)
41		β-Bisobolol	C ₁₅ H ₂₆ O	Not reported	\
42		α-Eudesmol	C ₁₅ H ₂₆ O	Anti-microbial; Anti-tumor	(Britto et al., 2012; Ho, Wang, Hsu, Lee, & Su, 2009)
43		Drimenol	C ₁₅ H ₂₆ O	Anti-feedant; Anti-bacterial; Anti-fungal; Cytotoxic; Insecticidal; Anti-allergic; Piscicidal; Molluscicidal; Regulating the plant growth	(T. de M. Silva et al., 2013; Vlad, 2006)
44		Isolongifolol	C ₁₅ H ₂₆ O	Not reported	\
45		Drimenin	C ₁₅ H ₂₂ O ₂	Anti-fungal; Anti-bacterial; Anti-fungal; Anti-feedant; Plant-growth regulatory; Cytotoxic; Phytotoxic; Piscicidal; Molluscicidal	(Jansen & De Groot, 2004; Paz et al., 2020)
46	Aliphatic compounds	Undecane	C ₁₁ H ₂₄	Anti-inflammatory; Anti-allergic	(D. Choi, Kang, & Park, 2020)
47		Decanal	C ₁₀ H ₂₀ O	Anti-oxidant; Flavor and fragrance applications; Inhibitory and bactericidal; Cytotoxic	(K. Liu et al., 2012)
48		Decane, 4-methyl	C ₁₁ H ₂₄	Not reported	\
49		Decanol	C ₁₀ H ₂₂ O	Not reported	\
50		Undecanal	C ₁₁ H ₂₂ O	Anti-bacterial	(Basu & Banik, 2020; Faudale, Viladomat, Bastida, Poli, & Codina, 2008)
51		Cyclohexane, 1-ethenyl-1-methyl-2,4-bis(1-methylethene)	C ₁₅ H ₂₄	Not reported	\
52		Dodecanal	C ₁₂ H ₂₄ O	Anti-microbial	Chanprapai, Kubo, and Chavasiri (2018)
53		Pentadecane	C ₁₅ H ₃₂	Anti-microbial; Cytotoxic; Anti-parasitic	Bruno et al. (2015)
54		1-Dodecanol	C ₁₂ H ₂₆ O	Anti-fungal; Anti-microbial	Chanprapai et al. (2018)
55		Pentadecanal	C ₁₅ H ₃₀ O	Anti-biofilm	Casillo et al. (2017)
56	Heptadecane	C ₁₇ H ₃₆	Anti-inflammatory; Anti-proliferative; Anti-aging	(D. H. Kim et al., 2013; L. C. Wu, Ho, Shieh, & Lu, 2005)	
57	Octadecane	C ₁₈ H ₃₈	Not reported	\	
58	1-Hexadecanol	C ₁₆ H ₃₄	Not reported	\	
59	Eicosane	C ₂₀ H ₄₂	Anti-fungal	Ahsan, Chen, Zhao, Irfan, and Wu (2017)	
60	Tetracosane	C ₂₄ H ₅₀	Not reported	\	
61	Bicyclo[5.3.0]decane, 2-methylene-5-(1-methylvinyl)-8-methyl-	C ₁₅ H ₂₄	Anti-microbial; Anti-inflammatory; Anti-oxidant; Anti-proliferative; Anti-cancer; Anti-tumors; Anaesthetic	(Shaheena, Chintagunta, Dirisala, & Sampath Kumar, 2019; Zahin, Ahmad, & Aqil, 2017)	
62	Bicyclo[5.2.0]nonane, 2-methylene-4,8,8-trimethyl-4-vinyl-	C ₁₅ H ₂₄	Not reported	\	
63	Naphthalene, 1,2,3,4,4a,5,6,8a-octahydro-4a,8-dimethyl-2-(1-methylethenyl)-,	C ₁₅ H ₂₄	Anti-inflammatory; Anti-microbial; Anti-cancer	(M. H. Huang et al., 2003; Ibrahim & Mohamed, 2016; Osmaniye et al., 2023)	
64	Phytane	C ₂₀ H ₄₂	Not reported	\	
65	Perhydrofarnesyl acetone	C ₁₈ H ₃₀ O	Not reported	\	
66	3,7,11,15-Tetramethyl-2-hexadecen-1-ol	C ₂₀ H ₄₀ O	Fragrance ingredient	McGinty, Letizia, and Api (2010)	
67	Phytol	C ₂₀ H ₄₀ O	Anxiolytic; Metabolism-modulating; Cytotoxic; Anti-oxidant; Autophagy and apoptosis inducing; Anti-nociceptive; Anti-inflammatory; Anti-microbial; Immune-modulating; Anti-tumor; Anti-mutagenic; Anti-atherogenic; Anti-diabetic; Lipid-lowering; Anti-spasmodic; Anti-epileptic; Anti-nociceptive; Anti-depressant	(Alencar et al., 2018; Islam et al., 2018)	

(continued on next page)

Table 1 (continued)

No.	Phytochemical classes	Composition	Chemical formula	Activities/Applications	Reference
68	Organic acids	Dodecanoic acid	C ₁₂ H ₂₄ O ₂	Anti-bacterial; Anti-oxidant; Anti-apoptotic	Renugadevi, Valli Nachiyar, and Zaveri (2021) Iguchi et al. (2001)
69		Myristoleic acid	C ₁₄ H ₂₆ O ₂	Cytotoxic	
70		Tetradecanoic acid	C ₁₄ H ₂₈ O ₂	Anti-microbial	
71		Pentadecanoic acid	C ₁₅ H ₃₀ O ₂	Anti-bacterial; Anti-fungal; Anti-inflammatory; Anti-fibrotic; Red blood cell stabilizing; Mitochondrial repairing; Essential fatty acid; Regulation of stemness in breast cancer cells; Anti-proliferative; Cytotoxic; Anti-cancer; Anti-microbial	(To, Nguyen, Moon, Ediriweera, & Cho, 2020; S. Venn-Watson, Lumpkin, & Dennis, 2020; S. K. Venn-Watson & Butterworth, 2022)
72		Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	Anti-cancer; Anti-inflammatory; Anti-bacterial; Anti-oxidant; Anti-microbial	(Aparna et al., 2012; Shaaban, Ghaly, & Fahmi, 2021; Sianipar, Purnamaningsih, & Rosaria, 2016; Siswadi & Saragih, 2021)
73		Oleic acid	C ₁₈ H ₃₄ O ₂	Anti-fungi; Immunomodulatory; Blood pressure reducing; Regulating body weight	(Alonso-Torre, Carrillo, & Cavia, 2012; Owolabi, L, & B, 2018; Terés et al., 2008; Tutunchi, Ostadrahimi, & Saghafi-Asl, 2020)
74		Octadecanoic acid	C ₁₈ H ₃₆ O ₂	Anti-oxidants; Anti-microbial; Anti-inflammatory; Anti-cancer; Anti-measles virus	(Entigu, Linton, Lihan, & Ahmad, 2013; Mujeeb et al., 2014; Othman, Abdullah, Ahmad, Ismail, & Zakaria, 2015; Reza et al., 2021; Siswadi & Saragih, 2021)
75		Hexanedioic acid	C ₆ H ₁₀ O ₄	Anti-bacterial; Anti-microbial	(W. H. Choi & Jiang, 2014; Iornumbe, Yiase, & Ato, 2016; Sianipar et al., 2016; Siswadi & Saragih, 2021)
76		2-Propenoic acid	C ₃ H ₄ O ₂	Not reported	\
77		1,2-Benzenedicarboxylic acid	C ₈ H ₆ O ₄	Cytotoxicity; Anti-microbial; Anti-cancer; Anti-bacterial	(Albratty et al., 2021; Krishnan, Mani, & Jasmine, 2014; Shoge, Garba, & Labaran, 2017)

Table 2

PM-induced toxic effects.

Category	Plant part	Solvent used in plant solvent extraction	Model	Highest dose	Results	Reference
Acute toxicity	Leaf	Water	Rats	5 g/kg bw	Non-toxic	(S. Q. Wasman et al., 2010)
	Stem and leaf	Water		2000 mg/kg bw		Ming et al. (2013)
	Leaf	Water		2000 mg/kg bw		(N. Muhammad et al., 2013)
	Leaf	Methanol		2000 mg/kg bw		(P. V. Christopher et al., 2017)
Sub-acute toxicity	Stem and leaf	Water	Rats	1000 mg/kg bw	Non-toxic	Ming et al. (2013)
	Leaf	Water	Rats	2000 mg/kg bw	Large dose whereby it may lead to electrolyte imbalance, anemia, and hypocalcemia	(N. Muhammad et al., 2013)
	Leaf	Methanol	Rats	2000 mg/kg bw	Non-toxic	(P. V. Christopher et al., 2017)
Genotoxicity	Leaf	Water	Human lymphocytes	\	No genotoxic	Wan-Ibrahim, Sidik, and Kuppusamy (2010)
	Stem and leaf	Methanol	Zebrafish		No teratogenic	Omar, Kue, Dianita, and Yu (2020)

medicine and pharmacology (Pham et al., 2023). Table 2 and 3 summarize the toxicity of PM in detail, including acute toxicity, subacute toxicity, genotoxicity, and cytotoxicity.

3.1. Acute toxicity

An acute toxicity study examines potential harmful effects that arise when organisms are exposed to single or multiple doses of a test substance within 24 h through various routes like oral, dermal, or inhalation (Saganuwan, 2017). The information gathered from the acute toxicity study serves as a guide for determining appropriate dosage levels in long-term toxicity studies and other animal research (Erhirhie, Ihekwereme, & Ilodigwe, 2018).

PM has well-established traditional applications in folk medicine and, in general, is believed to be safe. Even so, it is essential to experiment with its safety level for long-term usage. The acute toxicity assessment of PM aqueous leaf extract appears promising, as no adverse effects were observed in experimental animals during a 14-day observation period. It is reported that, in the acute toxicity test, oral

administration of 2000 mg/kg of the PM aqueous extract produced neither mortality nor changes in behavior or any other physiological activities (Ming, Zulkawi, Cy, & Choudhary, 2013; N. Muhammad et al., 2013). The highest administered dose of 5 g/kg body weight (bw) did not induce any toxicity, as examination of the liver and kidney in treated rats revealed no significant changes compared to the control group. Hematology and clinical biochemistry values fell within the range of those observed in control animals. This indicates that the PM aqueous leaf extract is relatively non-toxic (S. Q. Wasman, Mahmood, Salehuddin, Zahra, & Salmah, 2010). Another study found that the methanol extract of PM at 2000 mg/kg did not exhibit any signs of acute toxicity. Thus, the LD₅₀ is determined to be greater than 2000 mg/kg (P. V. Christopher et al., 2017). According to Sharma et al. (2020), all the test animals stayed healthy and exhibited normal behavior throughout the experiment, so this herb could be considered safe. Therefore, PM could be considered a tool for developing medications to address contemporary health issues.

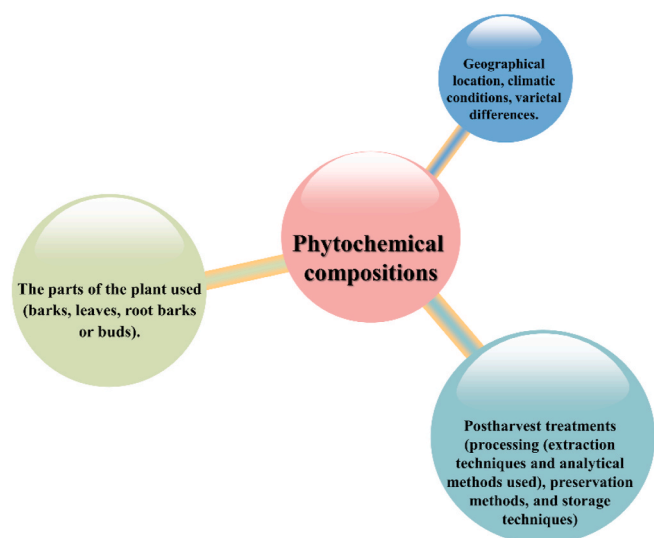


Fig. 3. Factors affecting the phytochemical compositions of PM.

3.2. Sub-acute toxicity

The sub-acute toxicity study is essential for the assessment of the safety of an agent when it is administered repeatedly over a period of time (P. V. Christopher et al., 2017) because subacute toxicity determines the long-term adverse effects of the repeated daily dose administration (Nakakaawa, Gbala, Cheseto, Bargul, & Wesonga, 2023).

In a subacute 28-day repeated dose oral toxicity study, no mortality or toxic signs were observed when the three doses of 125, 500, and 1000 mg/kg of PM aqueous extract were administered orally for a period of 28 days (Ming et al., 2013). In the subacute toxicity study led by Muhammad and colleagues, the results indicated an absence of toxicity, behavioral changes, or fatalities during the study. However, when the extract was given at a dose of 2000 mg/kg bw for 14 days, there was a significant reduction in hemoglobin, calcium, and sodium levels, along with an increase in potassium levels. In summary, oral administration of PM leaf extract did not induce subacute toxicity in male rats, except at an extremely high dose, which could lead to electrolyte imbalance, anemia, and hypocalcemia (N. Muhammad et al., 2013). The study by Christopher et al. revealed that the methanol extract of PM leaves does

not possess any neurotoxicity as the animals were normal. The locomotor activity of the treated group of animals was compared and found no reduction or stimulation. Thus, the result indicates the absence of any inhibitory or stimulatory activity on the mental activity. In the present study, chronic treatment of PM methanol extracts up to 2000 mg/kg did not produce any mortality (P. V. Christopher et al., 2017).

Thus far, sub-chronic and chronic toxicological studies have not been carried out on various parts and solvent extracts of PM. Hence, a thorough toxicological evaluation of the safety level of the extracts is required for consideration of PM as an innocuous herbal drug.

3.3. Genotoxicity

Studies have revealed that some plants frequently used in folk medicine are potentially genotoxic (Ananthi, Chandra, Santhiya, & Ramesh, 2010; Marques, De Medeiros, Da Silva Dias, Barbosa-Filho, & Agnez-Lima, 2003; Melo-Reis, Bezerra, Vale, Canhêta, & Chen-Chen, 2011; Regner et al., 2011; I. S. Shin et al., 2011). Wan-Ibrahim et al. have proved that aqueous extract of PM has no genotoxic effect on human lymphocytes (Wan-Ibrahim et al., 2010). Omar and colleagues investigated the teratogenic potential of PM using a zebrafish model. The results showed that the tested methanol extract of PM did not interfere with the development of zebrafish embryogenesis (Omar et al., 2020).

3.4. Cytotoxicity

Over the past decade, a number of *in vitro* methods have been evaluated with the aim of replacing the mouse bioassay for toxicity testing. Cell culture-based toxicity tests are of interest, having the potential to detect more general cytotoxicity endpoints (Dzoyem, Kuete, McGaw, & Eloff, 2014).

For cancer cells, the PM leaf 50%, 70%, and 100% ethanol extracts showed significant suppression of viability of HCT116, HT29, and CT26 cell lines (Rohin et al., 2020), and meanwhile PM leaf 100% ethanol extract showed antiproliferative activity on HeLa cells (Ali et al., 1996; Rohin et al., 2020). However, PM leaf acetone extract displayed no toxicity against the HCT116, HT29, and CT26 cell lines (Rohin et al., 2020). This indicates that the function of the extracts can be influenced by the choice of various extraction solvents. In addition, the ethyl acetate extract of PM leaf had potent cytotoxicity against HCT116, HT29, CT26, HeLa, and HepG2 cell lines (Abdullah et al., 2017; Mohd Ghazali

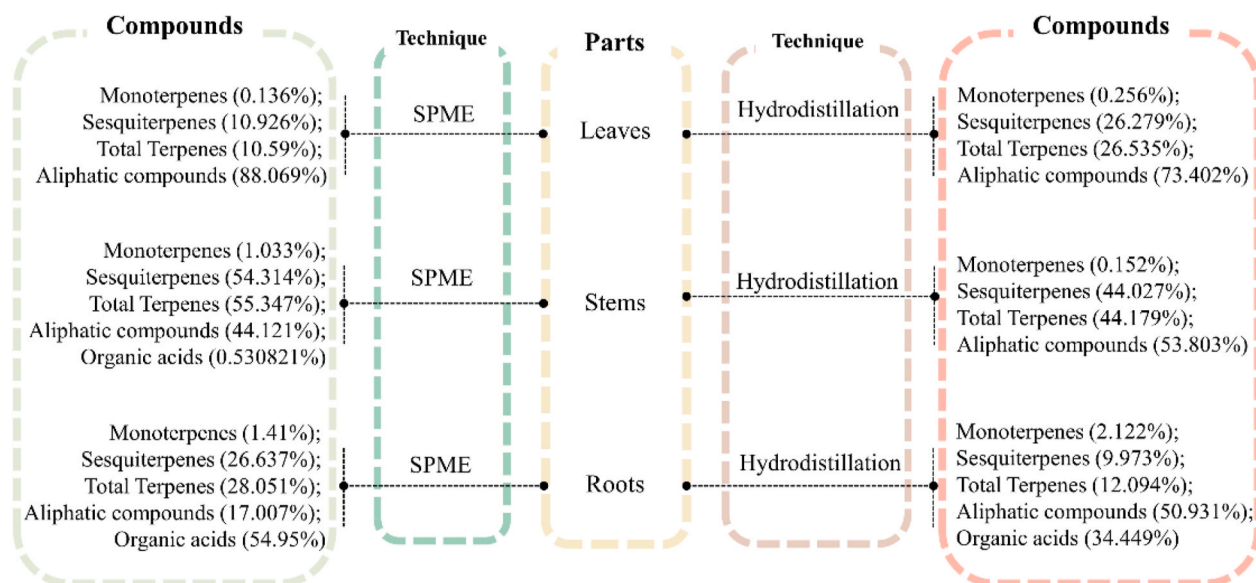


Fig. 4. Plant parts and extraction techniques affect phytochemical composition.

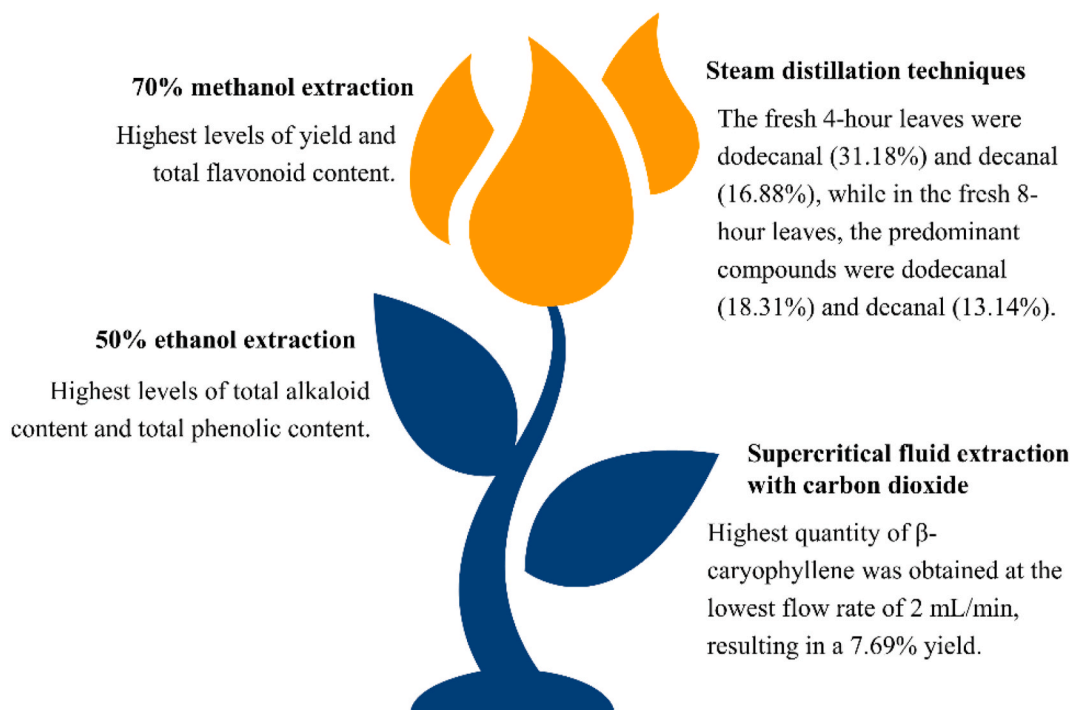


Fig. 5. Extraction affects the phytochemical composition.

et al., 2014; Rohin et al., 2020). The PM leaf ethyl acetate extract showed cytotoxicity toward MCF7 cells, but $IC_{50} > 250 \mu\text{g/mL}$ (Mohd Ghazali et al., 2014). The hexane extract of PM leaf exhibited promising cytotoxicity against the HCT116 cells with an IC_{50} value of $40 \mu\text{g/mL}$ (Abdullah et al., 2017). Another study showed that PM leaf methanol extract had antiproliferative effects against HCT116, HT29, CT26, and HeLa cell lines, and cytotoxicity toward HepG2 and MCF7 cells, but $IC_{50} > 250 \mu\text{g/mL}$ (P. V. Christopher, Parasuraman, et al., 2016; Mohd Ghazali et al., 2014; Rohin et al., 2020). Research findings showed that petroleum ether extract from PM leaf was able to suppress the growth of HCT116, HeLa, HepG2, and MCF7 cells (Mohd Ghazali et al., 2014). Besides, another *in vitro* experiment showed that water extract of PM possessed antiproliferative effects against a panel of cancer cells, including HCT116, HT29, CT26, HeLa, HepG2, and MCF7 cells (P. V. Christopher, Parasuraman, et al., 2016; Mohd Ghazali et al., 2014; Rohin et al., 2020).

For normal cells, the water, ethanol, ethyl acetate, methanol, and petroleum ether extracts of PM leaf showed no activity against the Hs888Lu and Vero cells and gave an $IC_{50} > 250 \mu\text{g/mL}$ for CCD841, Chang liver, and WRL-68 cells (Abdullah et al., 2017; Mohd Ghazali et al., 2014; Qader et al., 2011; Vimala et al., 2011; Wahab et al., 2015). In addition, on the CCD841 and EA.hy926 cells, PM leaf hexane, methanol, and water extracts demonstrated cytotoxic action, while on the Vero cell, PM stems methanol extract demonstrated cytotoxic action (Abdullah et al., 2017; P. V. Christopher, Parasuraman, et al., 2016; Wahab et al., 2015).

Consequently, some PM extracts have been verified to treat cancers effectively due to their lower toxicity and minimal side effects on normal cells. This high selectivity makes it a safe option for managing and treating cancer.

According to Table 3 and it is evident that researchers using the same solvent extract obtained varying results on identical cells. Some demonstrated robust cytotoxic activity effects, while others exhibited weaker ones. This variability may be attributed to differences in the plant phytochemical compositions, influenced by geographical location, climate, and variations in plant varieties (as summarized in part 2).

4. Traditional usages

The whole parts of this plant (aerial, root, root bark, stem, stem bark, flower, and seeds) are used for various purposes (P. V. Christopher et al., 2017). PM is traditionally used as a food and flavoring agent (Abdullah et al., 2017; Han et al., 2024). In Japan, China, and Europe, PM has long been applied as a hot-tasting spice (Ahmad et al., 2014). In Malaysia, PM is frequently added to vegetable salad and many other dishes as a flavoring enhancer (Abdullah et al., 2017), such as laksa (spicy noodle dish), kerabu (fried herbal rice), asam pedas (spicy tamarind curry), and tom yam (spicy tangy soup) (Rahim et al., 2022; Vimala, Adenan, Ahmad, & Shahdan, 2003). The Malaysian Government has listed this plant in the National Agro-Food Policy to ensure sufficient food supply (Embong, 2007). In Indonesia, particularly in West Kalimantan, padas porridge made using PM is commonly known as the unique cuisine of West Kalimantan (Alves, Ribeiro, Kloos, & Zani, 2001; Phatik, Das, & Boruah, 2014). Moreover, PM is also an important traditional medicine for relieving physical discomforts, including rid of fungal infections, treating poor eyesight, and alleviating body aches as well as others.

Given various biological activities besides enhancing the flavor of food dishes (Imelda et al., 2014), traditionally, PM leaves are used as medicine for the treatment of different ailments (Hassan, Noor, & Kader, 2015, pp. 41–48). It has been used to treat digestive disorders (George, Ng, O'Callaghan, Jensen, & Wong, 2014; Han et al., 2024), reduce dandruff (Z. Muhammad & Mustafa, 2015), treat poor eyesight (Jarukamjorn & Nemoto, 2008), warm the body up (Embong, 2007), improve blood circulation (Vikram et al., 2014), get rid of fungal infections (Ong & Nordiana, 1999), alleviate body aches (Amiruddin, Wahab, & Mohsin, 2020; Wiart, 2006), and be a beverage after child-birth (P. Christopher, Vikneswaran, et al., 2015). In India, PM has been applied as diuretics, central nervous system stimulants, diaphoretics, and stypitics (Ahmad et al., 2016). Moreover, most Chinese Buddhists have taken the leaves primarily for the fact that PM can reduce sexual desire; thus, the monks usually grow PM as their garden plant and consume it as a supportive stride in their celibate life (Abubakar et al., 2015). Fig. 6 displays the traditional application of PM in ethnopharmacology.

Table 3
Detailed information on cytotoxicity of PM.

Plant part	Solvent used in plant solvent extraction	Cells	IC ₅₀	Reference
Leaf	Ethanol	HeLa	0.1 mg/mL	Ali et al. (1996)
Leaf	Water	Hs888Lu	No	Qader et al. (2011)
Leaf	Ethanol		cytotoxicity	
Leaf	Water	WRL-68	No	Vimala et al. (2011)
		Vero	cytotoxicity	
Leaf	Petroleum ether	HepG2	>250 µg/mL	Mohd Ghazali et al. (2014)
		WRL-68	56.23 µg/mL	
		HeLa	127.21 µg/mL	
		HCT116	145.32 µg/mL	
		MCF7	>250 µg/mL	
		Chang liver	>250 µg/mL	
	Methanol	HepG2	>250 µg/mL	
		WRL-68	>250 µg/mL	
		HeLa	205 µg/mL	
		HCT116	86.3 µg/mL	
		MCF7	>250 µg/mL	
		Chang liver	>250 µg/mL	
	Ethyl acetate	HepG2	32.25 µg/mL	
		WRL-68	122.38 µg/mL	
		HeLa	63.09 µg/mL	
		HCT116	199.52 µg/mL	
		MCF7	>250 µg/mL	
		Chang liver	285.72 µg/mL	
	Water	HepG2	>250 µg/mL	
		WRL-68	>250 µg/mL	
		HeLa	>250 µg/mL	
		HCT116	>250 µg/mL	
		MCF7	>250 µg/mL	
		Chang liver	>250 µg/mL	
Leaf Stem	Methanol	Vero	875 mg/L	Wahab, Bunawan, and Ibrahim (2015)
Leaf	Water	HCT116	>200 µg/mL	(P. V. Christopher, Parasuraman, et al., 2016)
		HT29	>200 µg/mL	
		HeLa	>200 µg/mL	
		EA. hy926	92.2 µg/mL	
Leaf	Methanol	HCT116	>200 µg/mL	
		HT29	>200 µg/mL	
		HeLa	>200 µg/mL	
		EA. hy926	214 µg/mL	
Leaf	Hexane	HCT116	40 µg/mL	Abdullah, Mohd Ali, Abolmaesoomi,
		CCD841	190.62 µg/mL	

Table 3 (continued)

Plant part	Solvent used in plant solvent extraction	Cells	IC ₅₀	Reference
	Ethyl acetate	HCT116	43.18 µg/mL	Abdul-Rahman, and Hashim (2017)
		CCD841	89.2 µg/mL	
	Methanol	HCT116	>333 µg/mL	
		CCD841	>333 µg/mL	
	Water	HCT116	>333 µg/mL	
		CCD841	>333 µg/mL	
Leaf	Water	HT29	75 µg/mL	Rohin, Hadi, and Ridzwan (2020)
		HCT116	33 µg/mL	
		CT26	29 µg/mL	
	Methanol	HT29	78 µg/mL	
		HCT116	30 µg/mL	
		CT26	20 µg/mL	
	Ethanol	HT29	73 µg/mL	
		HCT116	31 µg/mL	
		CT26	26 µg/mL	
	70% Ethanol	HT29	34 µg/mL	
		HCT116	13 µg/mL	
		CT26	10 µg/mL	
	50% Ethanol	HT29	24 µg/mL	
		HCT116	20 µg/mL	
		CT26	23 µg/mL	
	Acetone	HT29	No	
		HCT116	cytotoxicity	
		CT26		
	Ethyl acetate	HT29	33 µg/mL	
		HCT116	7 µg/mL	
		CT26	7 µg/mL	

5. Food application

Recent studies found that PM had different applications in the food field such as food formulation and encapsulation. Therefore, PM and their bioactive properties hold significant potential for utilization in diverse food industry contexts as illustrated in Fig. 7.

5.1. Food additive

In recent decades, particular attention has been emphasis on the importance of natural products in safeguarding meat products against spoilage (Konfo et al., 2023). Antioxidants, functioning as hydrogen atom donors, are utilized in meat products to retard or prevent oxidative reactions (Hadidi et al., 2022). The incorporation of plant extracts as food additives serves to diminish the reliance on synthetic additives while addressing the demand for natural food options (Santos et al., 2022). This situation also poses a new challenge for food scientists who are actively seeking novel natural sources of antioxidant molecules for potential use as food additives (Felipe Alzate-Arbeláez, Dorta, López-Alarcón, Cortés, & Rojano, 2019). On the other hand, to ensure the microbial safety of food products, the food industry employs various preservatives (Premanath, James, Karunasagar, Vaňková, & Scholtz, 2022). Natural plant extracts are recognized as effective natural antimicrobial agents and thus gaining importance as food additives (Friedlein, Dorn-In, & Schwaiger, 2021; N. Sharma & Gulati, 2023).

PM exhibits antioxidant attributes, as evidenced by spent hen meat burgers containing PM water extracts, which displayed significantly lower peroxide value (PV) and thiobarbituric acid (TBA) values in comparison to the control group over a three-month frozen (−18 °C) storage period (Huda-Fujan, Noriham, Norrakiah, & Babji, 2006). Similarly, in a separate study, the PM extract-treated duck meatballs demonstrated superior antioxidant effects, texture, and microbial potency compared to the control group Abdul Azziz et al. (2015) during

Table 4
Summary of the inhibition effects of PM on the bacterium.

Plant part	Extraction method	Solvent used	Method	Inhibited bacterium	Reference
Leaf	Solvent extraction	n-Hexane	Disk diffusion method	<i>Staphylococcus aureus</i> , <i>Staphylococcus epidermidis</i> , and <i>Streptococcus pyogenes</i>	(P.M. Ridzuan et al., 2013)
		Dichloromethane			
		Methanol			
		Water			
Leaf	The first stage covers extraction, which was done by steam distillation to obtain essential oils, followed by multilevel extraction by maceration and UAE to obtain different extracts	Ethanol	Disc diffusion method	<i>Staphylococcus aureus</i>	Imelda, Kusumaningrum, and Faridah (2013)
Leaf	Ultrasound-assisted extraction	Ethanol	Macro dilution method	<i>Staphylococcus aureus</i>	Imelda et al. (2014)
Leaf	Solvent extraction	Water	Agar well diffusion	<i>Staphylococcus aureus</i> , <i>Staphylococcus epidermidis</i> , <i>Staphylococcus aureus</i> and <i>Staphylococcus epidermidis</i> ,	Hassan et al. (2015)
		Methanol			
		Ethanol			
Leaf	Solvent extraction	30% Water -ethanol and 100% Water	Disc diffusion and microplate dilution	<i>Staphylococcus aureus</i>	Abubakar et al. (2015)
Leaf	Solvent extraction	Methanol	Disc diffusion method	<i>Staphylococcus aureus</i>	Zamri (2015)
Leaf and the whole plant	Soxhlet extraction	Methanol and distilled water extracts	Disc diffusion method	<i>Staphylococcus aureus</i>	Hassim et al. (2015)

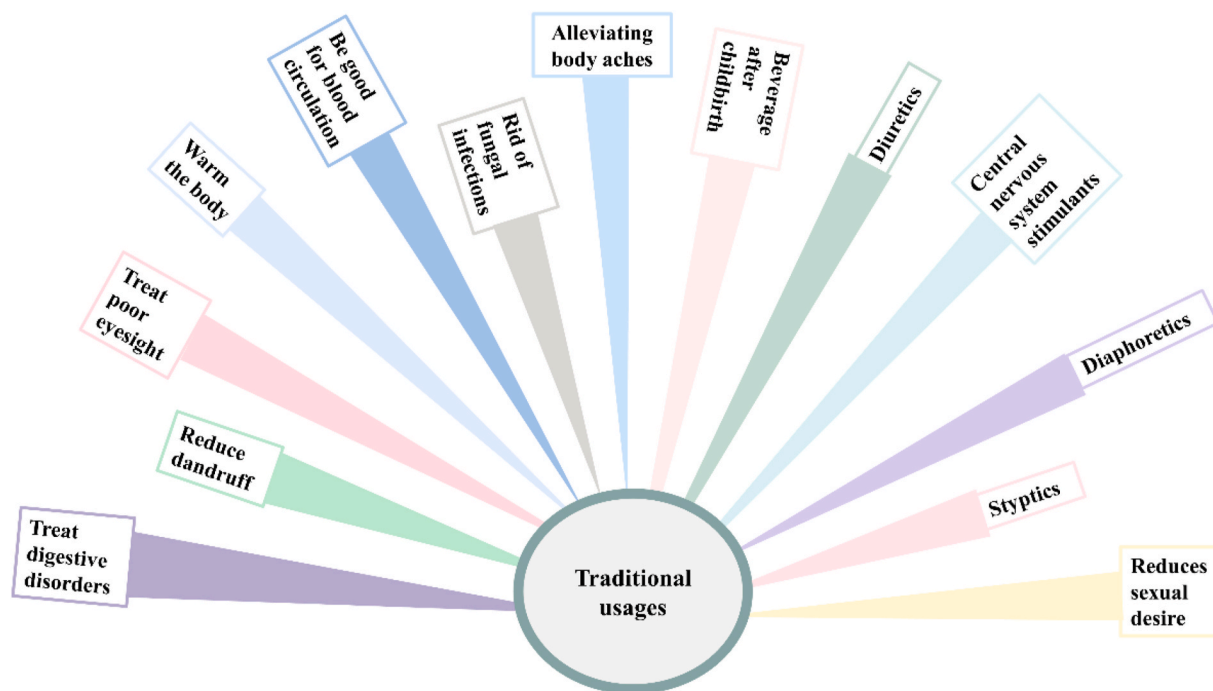


Fig. 6. Traditional medicine uses of PM.

refrigerated storage ($\pm 4\text{ }^{\circ}\text{C}$) over 21 days (Nurul, Ruzita, & Aronal, 2010). In addition, after 21 days of storage, quail meatballs formulated with PM extract exhibited lower TBA, free fatty acid (FFA), PV values, and aerobic plate count compared to the control (Ikhlas, Huda, & Ismail, 2012). Overall, the above study suggests PM extracts have the potential to serve as a natural antioxidant resource for extending the shelf life of meatballs, and resembling the synthetic antioxidant hydroxy anisole

(BHT) (Ikhlas et al., 2012; Nurul et al., 2010). Moreover, since Ikhlas et al. (2012) discovered that PM extract decreased FFA levels in food products, where FFA is known to affect the color and taste of a food leading to reduced consumer acceptance (Sete da Cruz et al., 2022), thus Gervacia and Ratih (2021) investigated the addition of PM leaves to waste cooking oil to reduce FFA content. The results were gratifying, with an average reduction in FFA levels of 7.12% in used cooking oil

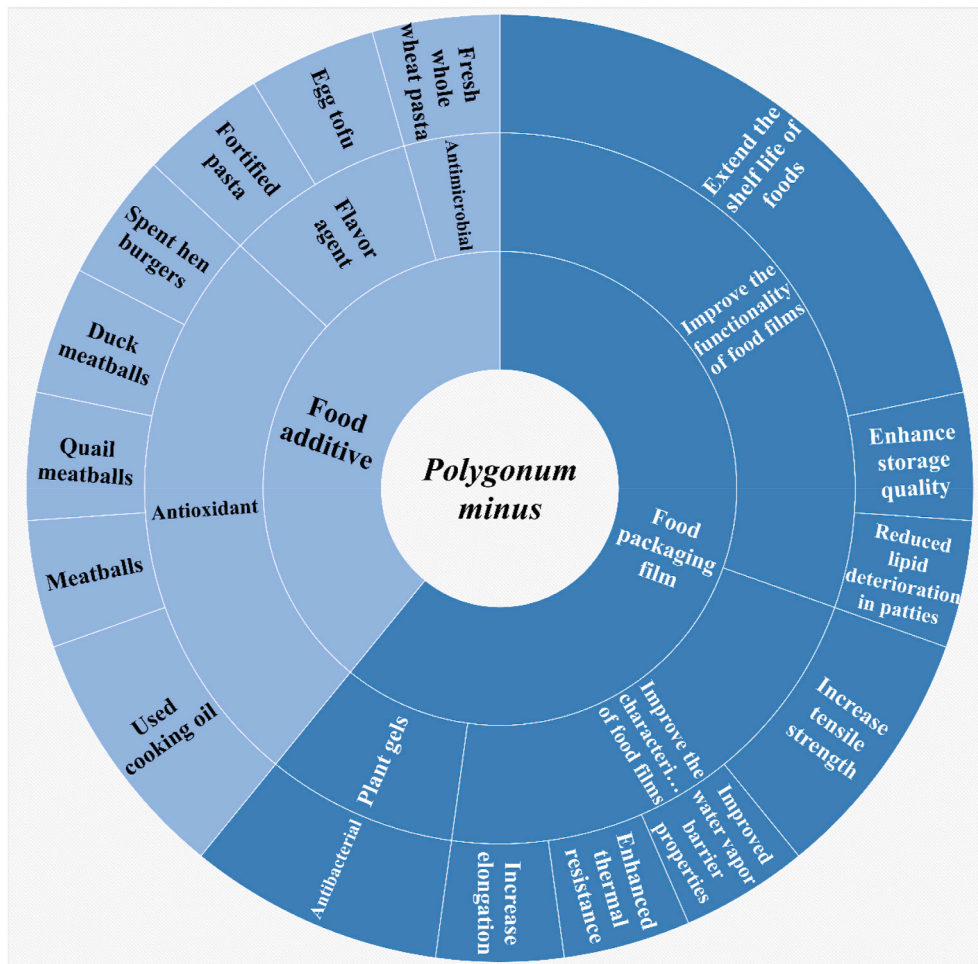


Fig. 7. Application of PM in food products and food packaging film.

before adding PM leaves. A positive correlation was established between the addition of PM leaves and the reduction in fatty acid levels (Gervacia & Ratih, 2021). In essence, achieving lower FFA content in oils by using the antioxidant properties of PM leaves is desirable for enhancing their quality (Mataruco et al., 2023).

PM demonstrates potent antimicrobial properties, as highlighted in a study by Abdul Azziz et al. (2015), wherein the hexane extract of PM exhibited remarkable efficacy against various food bacteria in the disk diffusion method. Afterward, researchers considered whether incorporating PM as an antibacterial agent would impact consumer acceptance of food products. To assess this, they added dry PM powder to fresh whole wheat pasta and stored it under refrigeration (4 °C) and at ambient temperature (24 °C). Notably, the study found that whole wheat pasta supplemented with PM powder showed comparable overall acceptance to control pasta, with no significant differences (Az-Zahra Tashim, Harkani, Abdullah Lim, & Maryam Basri, 2021). This suggests the potential for plant powder additions to serve as potent bio-preservatives and antimicrobial agents without compromising consumer acceptance (Az-Zahra Tashim et al., 2021).

To assess the impact of incorporating PM into food on consumer sensory perception, researchers experimented with adding PM to egg tofu. The results revealed that an increase in PM content significantly enhanced the springiness of the egg tofu, with the optimal formula combination of 0.7% PM, 0.5% *Curcuma longa*, and 0.8% *Zingiber officinale* (Maizura, Aminah, & Wan Aida, 2016). In another investigation, Tashim, Lim, and Basri (2022) observed that adding PM powder to pasta resulted in favorable sensory acceptability. The fortified pasta, enriched with the PM powder mixture significantly increased DPPH scavenging

activity while retaining characteristics indistinguishable from the control pasta, including minimal cooking loss and favorable cohesiveness, springiness, and chewiness (Tashim et al., 2022).

In short, PM holds promising prospects as a food additive. On one hand, its antioxidative properties can effectively delay or prevent the oxidation reaction of food. On the other hand, its antibacterial attributes can prevent food from being contaminated by microbes. Moreover, incorporating PM into food can enhance sensory acceptance among consumers.

5.2. Food packaging film

Food packaging primarily aims to shield food from contaminants and extend its shelf life (Kola & Carvalho, 2023). However, the global food industry has a significant environmental impact, primarily due to the extensive use of petroleum-based plastic packaging films (Babayev et al., 2023; R. Westlake et al., 2023; W. Zhang, Roy, Ezati, Yang, & Rhim, 2023). Considering environmental concerns, there is a growing focus on replacing synthetic polymers with renewable, biodegradable films. These biodegradable films offer advantages including biocompatibility, edibility, non-toxicity, and a wide range of potential applications. This shift towards biodegradable films is regarded as an innovative and effective alternative for extending the shelf life of various food products (Manzoor, Yousuf, Ahmad Pandith, & Ahmad, 2023; Díaz-Montes & Castro-Muñoz, 2021; Kaya et al., 2018; Suvarna, Nair, Mallya, Khan, & Omri, 2022).

PM extracts have been found to improve the characteristics of food films. Yahaya, Husaini, Zaharudin, and Azman (2020) reported that

adding 2% PM extract to semi-refined carrageenan plasticized with glycerol can increase tensile strength and elongation at break values to improve film properties. In other words, adding PM extract to a semi-refined carrageenan-based film can make the film durable and strong (Yahaya et al., 2020). Other researchers had similar reports, and they found that PM at a 10% weight percentage was incorporated into polylactic acid (PLA) to create PLA-10PM packaging films using the solvent-casting method. PLA-10PM demonstrated enhanced thermal resistance and improved water vapor barrier properties (Mohamad et al., 2020). Another study found that adding PM leaf extract increased the tensile strength of edible films made from durian seed starch-chitosan, but also resulted in a decrease in film uniformity and changes in the surface appearance of the film (Lestari, Hartanti, & Permadi, 2020). In a later study, Kong, Heng, and Pui (2022) also found that PM leaf extract and curry leaf extract (CLE) were added to chitosan-based films. With the increase of PM leaf extract and CLE, the mechanical strength of the chitosan film decreased and the water solubility increased. Moreover, the addition of PM and CLE resulted in a darker, greener, less yellow color compared to regular films. Hence, adding PM leaf extract into chitosan-based films remains a challenge, and we still need to optimize the preparation method of this film. Furthermore, polybutylene succinate (PBS) film containing PM displayed a cohesive structure according to morphological analysis (Mohamad et al., 2022).

PM extracts have also been found to improve the functionality of food films. Husna Abd Hamid and the team successfully developed a semi-refined carrageenan film containing α -tocopherol and PM extract that can extend the shelf life of foods with high-fat content (Husna Abd Hamid et al., 2019). Subsequently, Yahaya and the team found that a semi-refined carrageenan film containing 2% PM extract significantly reduced lipid deterioration in patties and reduced metmyoglobin value changes (Yahaya, Almajano, Yazid, & Azman, 2019). As reported, PLA-10PM could preserve chicken meat for up to 15 days while maintaining odorless and firm textural properties (Mohamad et al., 2020). Kong et al. (2022) also had similar research results. Chicken breast meat wrapped with chitosan films containing 2.0% (w/v) PM and 2.0% (w/v) CLE remained viable at 4 °C for 14 days. The film extended the shelf life of chicken breasts by at least 2 days (Kong et al., 2022). In a study by Lestari, Hartanti, and Permadi (2020), the findings indicated that using an edible coating made of durian-chitosan seed starch with PM leaf extract could prolong the shelf life of beef sausage by four months when stored at freezing temperatures (Lestari, Permadi, & Mulyadi, 2022). The quality of chicken fillets coated with active PBS films improved in terms of color attributes when loaded with 15% PM (Mohamad et al., 2022). In addition to meat, edible films containing PM extracts can enhance the lipid oxidation stability and storage quality of apples (Husaini & Azman, 2020). Therefore, active packaging of edible films incorporating PM extracts shows potential as food packaging applications to extend the shelf life of meat, fruits, and vegetables (Husaini & Azman, 2020; Mohamad et al., 2020).

Moreover, in food packaging, in addition to food films, there are also coatings applied around the surface of the food (Mohamed, El-Sakhawy, & El-Sakhawy, 2020). Plant gels have an important role in food preservation as edible coatings (Kahramanoglu, Chen, Chen, & Wan, 2019) and are generally applied by dipping the foods, spraying, or brushing. To date, numerous studies have been conducted on the use of PM gel as an edible coating. Pramita investigated the antibacterial properties of methanol extract from PM leaves and its effectiveness when formulated into a gel product. Gel C (15% methanol extract) exhibited the most potent antimicrobial effect, suggesting its potential as an antiseptic agent. Furthermore, Gel C demonstrated better stability in all variables of the gel stability test compared to Gel A (5% methanol extract) and Gel B (10% methanol extract) (Pramita, 2013). Another study had a similar view. Gel C experienced the least viscosity decrease, while Gel B showed the least reduction in thickening ability, an increase in pH, and improved spreadability (Ansiah, 2014).

In summary, PM extract shows great use and bright prospects in the food industry. On the one hand, it can be added to food as a food additive, and on the other hand, it can also be used as an edible film and gel to protect food from contaminants. An in-depth investigation of the vital components within PM extracts is necessary before processing them further for use in food formulation and packaging.

6. Agricultural field

Given the frequent occurrence of disease in the livestock breeding and aquaculture industry, the use of chemicals and antibiotics is increasingly restricted by governments, as well as the low feed conversion ratio. Thus, herbal feed additives have become a focus of research and development. This section reviewed some studies on the application of PM in livestock poultry breeding and aquaculture industry, as shown in Fig. 8.

6.1. Livestock and poultry

Poultry and livestock products are valuable sources of proteins and minerals (Abd El-Hack et al., 2022). However, the industry faces various environmental challenges like slow growth of animals, and hazards of parasites causing significant economic losses globally (El-Shall et al., 2022; Salem et al., 2022). Currently, there is a global initiative to explore natural drug alternatives to enhance poultry and livestock production while avoiding the residual impact associated with drug use in these products (Rehman et al., 2020).

PM usage as an anti-parasite has also raised substantial concern in animal husbandry. The results of the study by Alawiyah, Kahtan, and Widiyantoro (2016) showed that there was no significant difference in the death time of *Ascaris gallinae* between the PM extract concentration of 2 mg/mL and the albendazole group, indicating that the methanol extract of PM leaves has comparable anthelmintic activity to albendazole. The study found no significant difference in the time of death for the worms between the PM leaves ethanol extract concentration of 2 mg/mL and treated with Albendazole at 0.2 mg/mL. This indicates that the 2 mg/mL concentration of the ethanol extract from PM leaves exhibits comparable anthelmintic activity to Albendazole at 0.2 mg/mL (Maulidya, Kahtan, Natalia, Handini, & Vidiyantoro, 2018). In addition to chicken parasites, PM extract can also inhibit the growth of sheep parasites. *Haemonchus contortus* is a nematode commonly found in sheep, causing issues like hemorrhagic anemia or even death (Flay, Hill, & Muguero, 2022). A previous study evaluated the effectiveness of ethanolic PM extracts in inhibiting egg hatch and larval development assays of *H. contortus*. Results showed that PM extract exhibited larvicidal properties against *H. contortus*, indicating its potential to hinder parasite development (N. N. H. Yahya, 2017).

Additionally, PM can be used as a feed additive to improve chicken production. Basit et al. (2020) found that adding PM leaf meal to the diet improved growth performance, blood parameters, and serum biochemistry in broiler chickens. Remarkably, there were no harmful effects observed in the liver histomorphology. This suggests that PM leaf meal of feed can be safely used as an alternative to traditional growth promoters for broiler chickens. Another study investigated PM as a feed additive in the diet of *Kampung Unggul Balitbangtan* chickens to enhance their overall performance. It has been demonstrated that adding 2% PM powder during the starter period significantly improved the feed gain ratio in these chickens (Febriyanto, Lestari, & Tribudi, 2021).

In short, PM extract can inhibit the growth of parasites and has a significant anthelmintic effect on some parasites (its anthelmintic activity is equivalent to albendazole). PM leaf powder feed can promote the growth of broiler chickens without toxic side effects. Given that PM feed has anti-parasitic qualities, it can be utilized safely as an alternative to conventional growth promoters in broiler farms.

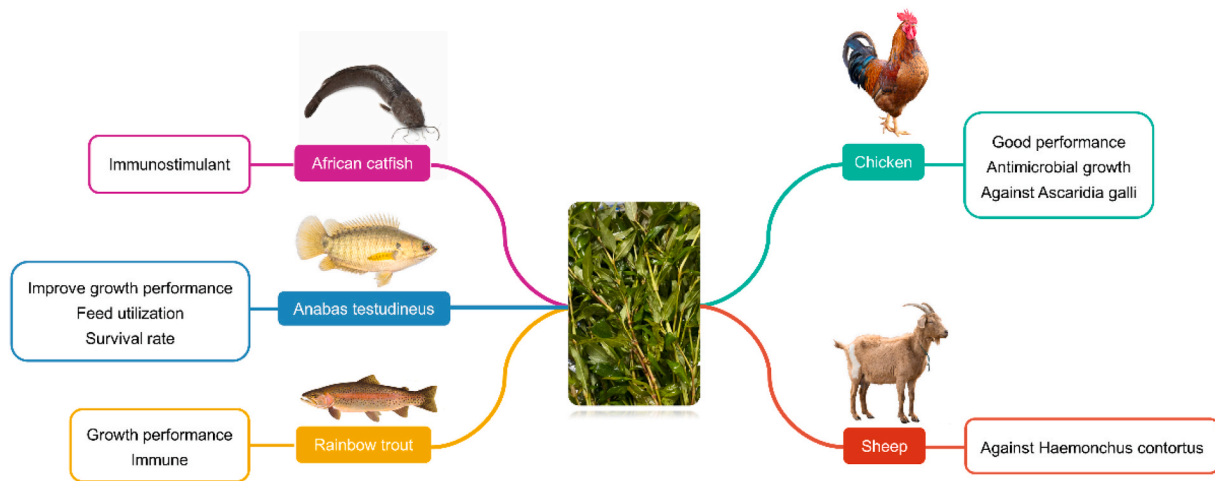


Fig. 8. Application of PM in the breeding industry.

6.2. Aquatic products

The aquaculture industry is a vital source of nutritious and affordable protein, particularly through fish and shrimp farming (Abdel-Latif et al., 2022). Across the globe, farmers are increasingly turning to intensive culture systems to meet the rising demand for these seafood products. However, the high fish biomass in confined spaces of intensive systems often leads to disease outbreaks (Abdel-Latif, Dawood, Menanteau-Ledouble, & El-Matbouli, 2020, 2022; Abdel-Latif & Khafaga, 2020; El-Son, Nofal, & Abdel-Latif, 2021). To address this, researchers have explored various feed supplements for farmed fish, including herbal extracts and essential oils, to promote immune function, support microbiota, improve growth, and prevent diseases (Awad & Awaad, 2017; Bulfon, Volpatti, & Galeotti, 2015; Citarasu, 2010; Galina, Yin, Ardó, & Jeney, 2009; Harikrishnan, Balasundaram, & Heo, 2011; Newaj-Fyzul & Austin, 2015; Reverter, Bontemps, Lecchini, Banaigs, & Sasal, 2014; Van Hai, 2015).

Veerasamy et al. (2014) investigated the impact of the aqueous extract of PM leaves on the hematological parameters and lysozyme activity in *Clarias gariepinus*. The findings suggest that the extract could serve as an immunostimulant for African catfish, but careful monitoring may be necessary for prolonged use. Similarly, Panase and Tipdacho (2018) conducted a study on *Anabas testudineus* fingerlings, applying PM leaf extract at different levels for 60 days. Their results indicated improved growth performance, especially in feed utilization, and increased survival rates during the fingerling nursing phase. Moreover, the use of herbal extracts positively affected white blood cell counts. Adel, Dawood, Shafiei, Sakhaie, and Shekarabi (2020) explored the effects of PM extracts on rainbow trout, focusing on growth, skin mucus, serum immune parameters, and immune-related gene expressions. Rainbow trout-fed diets with varying PM levels (0, 5, 10, and 15 mg/kg) for 8 weeks exhibited significant improvements in final bw, weight gain, and specific growth rate. Therefore, the above studies confirm the positive effects of dietary PM on fish growth and immunity.

7. Medicinal field

Medicinal plants, recognized for their chemical compositions (Hayat et al., 2022), hold crucial pharmaceutical and therapeutic values (Begum, Hamayun, Tabassum Yaseen, Sumbal Akhter, & Muhammad, 2016). They have historically been utilized across cultures (Khumalo, Van Wyk, Feng, & Cock, 2022) and continue to serve as fundamental resources for enhancing human health (Bernardini, Tiezzi, Laghezza Masci, & Ovidi, 2017). Additionally, they contribute significantly to the development of modern medicines, nutraceuticals, food supplements,

and pharmaceutical products (Dias, Urban, & Roessner, 2012). In developing nations, about 80% of the population is dependent on traditional medicines which are reported to be a combination of phytochemical and herbal plants for curing various diseases (Chilverly et al., 2023). PM is a plant that contains several bioactive compounds with potential medicinal properties which are useful in fighting and treating common ailments, as well as inhibiting effects on microbes (Ahmad et al., 2018, 2023; P. V. Christopher, Parasuraman, et al., 2015; Qader et al., 2011; Vikram et al., 2014). The protective effects of PM in body systems are summarized in Fig. 9, and each study is discussed in the following sections.

7.1. Integumentary system

The skin acts as a critical interface between the body and the external environment, serving as a primary barrier against environmental factors and pathogens (Grice, 2014). Its main function is to protect the body from microorganisms, toxins, and various external factors, thus preventing potential harm and maintaining overall health (Goceri, 2019; Grice & Segre, 2011; Larouche, Sheoran, Maruyama, & Martino, 2018; Rousselle, Braye, & Dayan, 2019). Nevertheless, skin infections are a common reason for consultations in primary health care centers, and the most common bacteria causing skin infections (CBSIs) are *Staphylococcus aureus*, *Staphylococcus epidermidis*, and *Streptococcus pyogenes* (Mussin et al., 2021; Severn & Horswill, 2023). Fortunately, according to studies in recent years, a large number of PM extracts have shown anti-CBSIs activity. Table 4 summarizes the antibacterial activity of different solvent extracts of PM from previous studies. Besides, as summarized in part 2 (Phytochemical), PM functional attributes are influenced by postharvest interventions, encompassing processing (utilized extraction techniques and analytical methods), preservation practices, and storage methodologies. Therefore, we suggest that it is necessary to conduct in-depth studies on the anti-CBSIs activity of PM hexane extracts.

It is well known that fungi form a commensal relationship with human skin (Hurabielle et al., 2020). However, when the immune system is compromised, fungi can become opportunistic pathogens, evading the host immune system and leading to frequent and severe infections (Köhler, Casadevall, & Perfect, 2015). The findings of Dewi and her team revealed that the ethanol extract exhibited antifungal activity against *Trichophyton rubrum* (Dewi, Asseggaf, Natalia, & Mahyarudin, 2019). Melinda and her team measured the inhibition zone diameters and found that the ethanol extract exhibited antifungal activity against *Trichophyton mentagrophytes* at concentrations of 40% and 80%, with average inhibition zone diameters of 10.125 mm and 20.625

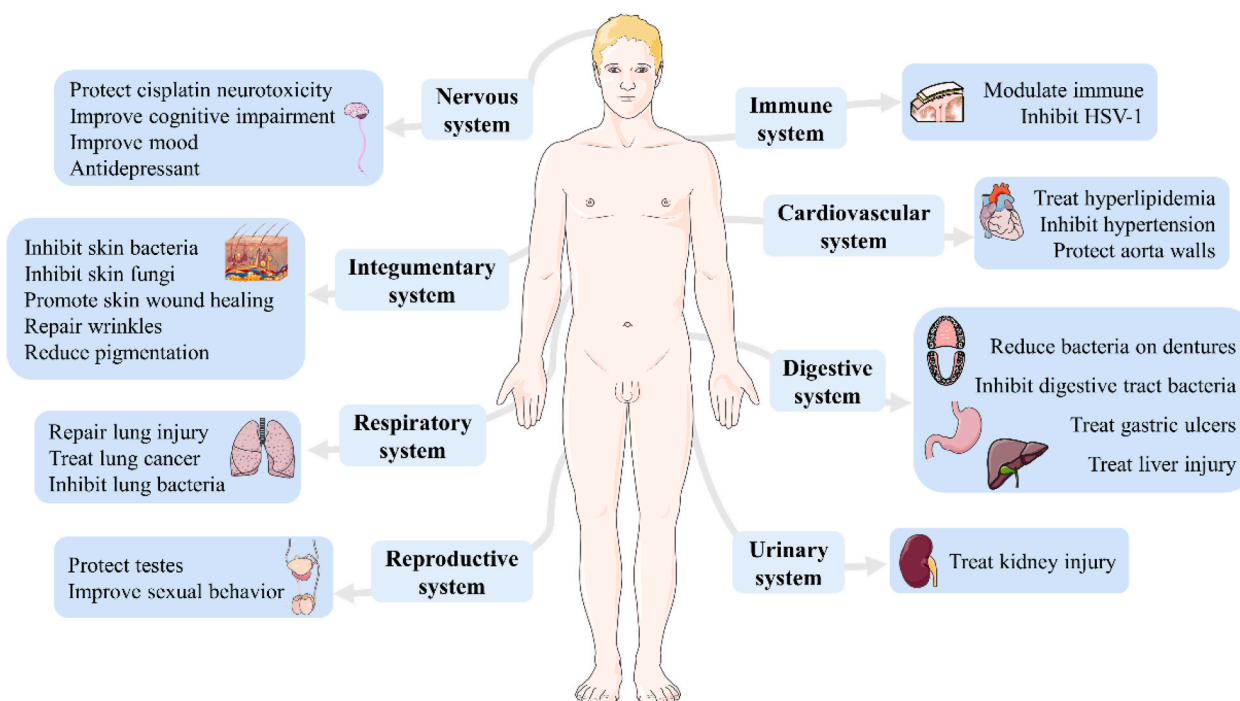


Fig. 9. Protective effects of PM in body systems.

mm, respectively (Melinda, Assegaf, Mahyarudin, & Natalia, 2019). Moreover, the ethanol extract did not create an inhibitory zone on the growth of *Microsporum gypseum*, indicating a lack of antifungal activity against this fungus (Liauw, Assegaf, & Natalia, 2021). Dandruff causes itchy, flaking skin with mild inflammatory reactions and is also one of the most common and widely seen dermatological diseases that affect the majority of the world population (Sheth & Dande, 2021). PM leaves were divided into four shampoo formulas by varying concentrations of PM leaves ethanol extract: 0%, 5 %, 10 %, and 15 %. The shampoo formula of PM leaves ethanol extract showed that it had antifungal activity against the fungi that caused dandruff. The best inhibitor activity was obtained from the third formula, with the diameter of a clear zone at 2.61 cm (Hadiarti, 2017).

Skin damage has become a clinical problem, that not only affects the body and mind of patients but also causes great loss to society (Liang et al., 2021). It acts as the initial defense barrier against microbes, preventing their unrestricted access to internal organs and potentially fatal consequences (Goceri, 2019; Larouche et al., 2018; Rousselle et al., 2019). Elnehrawy (2015) conducted a study to assess the effectiveness of various PM leaf extracts in promoting wound healing. Aqueous (31 µg/mL) and ethanol extract (63 µg/mL) of PM leaves significantly enhanced cellular migration and wound closure by 71% and 65% on day 1, respectively. The induced scratch was fully healed on day 2 with aqueous and ethanol extracts. Consequently, the aqueous extract of PM leaves may contribute to enhancing the healing capacity of skin cells, offering a safer, more natural, and cost-effective remedy. Hypersensitivity to mosquito bites is characterized by intense local skin responses to mosquito bites and various systemic symptoms (Vander Does, Labib, & Yosipovitch, 2022). In local reactions, mosquito bites can cause clear or hemorrhagic blisters of the skin, accompanied by intense red swelling (Hirai et al., 2023). These reactions can progress to necrosis or ulcers, ultimately healing with lasting scarring on the skin (Tatsuno et al., 2016). Some studies demonstrated that the methanol extract from PM had repellent and knockdown effects on *Musca domestica*. The 10% concentration of the extract showed the highest repellent effect at 61.67%, falling into the moderate category. Additionally, the 25% concentration was more effective in causing knockdown, reaching 75%, with the first knockdown observed in the 40 min (Kumalasari, Rima

Setyawati, & Hepi Yanti, 2015). Nugraha explored whether the n-hexane extract of PM leaf has an effect as a mosquito repellent and how it is effective when formulated in gel products with different concentrations. The results showed that the gel had a few changes in all variables of evaluation, but it was good enough esthetically for a month and could be used on the skin (Nugraha, 2021).

Beyond mere protection, skin health and appearance play crucial roles in self-esteem and social interactions (Khmaladze, Leonardi, Fabre, Messaraa, & Mavon, 2020). Beautiful, even-toned, blemish-free skin is everybody's dream (de Souza, 2008). However, no one can avoid pigmentation and wrinkles in the skin. Therefore, it is important to find bioactive substances that combat pigmentation and wrinkles. Skin hyperpigmentation is a disorder that causes blotchy, discolored, or darker skin tone in comparison with normal skin (M. C. Kang et al., 2020). Cosmeceuticals with biologically active ingredients that demonstrate both therapeutic and pharmaceutical effects are commonly used for reducing melanin content by inhibiting major regulatory steps in melanin synthesis and selectively targeting the hyperplastic melanocytes (Poulose et al., 2020; Sarkar, Arora, & Garg, 2013). In addition, cosmeceuticals with high antioxidant contents have been shown to be effective in reducing hyperpigmentation in the skin induced by UV radiation (Nahhas et al., 2019). In a particular investigation, the compounds from both PM leaves and rhizomes were extracted using hot water (100 °C), fresh water, and 80% ethanol individually. Subsequently, Yusuf discovered the potential of medicinal plant as an antioxidant, suggesting its applicability in producing herbal-based products in cosmetics and pharmaceuticals (Yusuf, 2014). It has been found that a wide variety of new cosmeceuticals and formulas can facilitate the skin to repair wrinkles, leading to a younger healthy-looking face (Mukherjee, Maity, Nema, & Sarkar, 2011). Haris, Ming, Perin, Blanche, and Jinapong (2014) conducted a study to assess the safety and effectiveness of Lineminus + PM cream compared to its placebo in healthy Asian females (split-face). The findings indicated that Lineminus + PM cream is safe exhibits anti-wrinkle effects, and possesses beneficial cosmetic properties for reducing wrinkles. Another study suggested that, along with PM-recognized antioxidant efficacy, the plant extract also demonstrated a noticeable clinical anti-wrinkle effect by reducing crow's feet wrinkles. Muthukumarasamy and colleagues concentrated

on creating and assessing an herbal antioxidant cream using the methanol leaf extract of PM. The resulting antioxidant cream, derived from the methanol leaf extract of PM, demonstrated promising free radical scavenging activity. This discovery highlights the plant's significance in future research and raises its profile in the herbal and cosmetic industry (Muthukumarasamy et al., 2018).

7.2. Respiratory system

Lung diseases involve abnormalities in the lungs, stroma, and alveoli, potentially leading to fatal outcomes (Aboushanab, El-Far, Narala, Ragab, & Kovaleva, 2021). A study by Wibowo, Anwari, Aulanni'am, and Rahman (2010) revealed that the n-hexane extract reduced lung cell proliferation in animal models. Another study focused on the n-hexane fraction impact on inducing apoptosis in lung tissue epithelial cells exposed to benzo(a)pyrene. The results indicated that the n-hexane fraction not only decreased lung reactive oxygen species levels in the animal models but also induced apoptosis in the lung epithelial cells (Wibowo, Purnomo, Widodo, & Aulanni'am, 2012). Subsequently, their team compared the effects of n-hexane, ethyl acetate, and methanol fractions of PM leaf extracts in repairing lung injuries caused by benzo(a)pyrene exposure. The research, using a single dose (100 mg/kg), showed that the n-hexane fraction was the most effective in repairing lung injuries induced by benzo(a)pyrene exposure (Wibowo, Widodo, Purnomo, & A, 2013). Therefore, PM application in the adjuvant treatment of lung cancer and lung injuries merits to be considered. On the other hand, *Streptococcus pneumoniae* is the most frequent bacterial cause of community-acquired pneumonia (Suaya et al., 2021), while *Serratia marcescens* assumes an opportunistic role in respiratory infections (Luttmann, Starnes, Haddad, & Duggan, 2022). Fortunately, extracts from the PM plant exhibit inhibitory effects against these bacteria. A study by Ridzuan et al. showed that both PM leaf n-hexane and methanol extracts possess antibacterial activity against *Streptococcus pneumoniae* (P.M. Ridzuan et al., 2013). Furthermore, PM leaf water, methanol, and ethanol extracts have been observed to exhibit activity against *Serratia marcescens* (Hassan et al., 2015).

7.3. Digestive system

The prevalence of oral fungal infections has significantly changed in recent years, primarily due to an increase in predisposing factors such as immune-compromising situations (e.g., HIV infection, endocrine disorders, immunosuppressive drug use, prolonged antibiotic therapies, malnourishment, and denture use) (Gleiznys, Zdanavicienė, Žilinskas, Eglė, & Juozas Žilinskas, 2015). Consequently, Rezki, Halimah, Maryani, and Setyowati (2020) explored the antibacterial effects of PM infusion extracts on bacterial colonization on acrylic dentures in the mouth. The findings indicated that PM could reduce bacterial colonies on acrylic dentures, and the antibacterial effect increased with higher infusion concentrations.

In addition, there are many bacteria that cause digestive tract diseases including *Salmonella typhi* and *Escherichia coli*, as well as leading agents of induced foodborne diseases, such as *Salmonella Typhimurium* and *Bacillus cereus* (Bernal-Bayard & Ramos-Morales, 2018; Devlin et al., 2022; Jovanovic, Ornelis, Madder, & Rajkovic, 2021; Knecht, Cholewińska, Jankowska-Mąkosza, & Czyż, 2020). However, surprisingly, PM extract was also able to inhibit these bacteria. Researchers found that PM leaf dichloromethane extract had an inhibitory effect on *Salmonella typhi* (P.M. Ridzuan et al., 2013). At the same time, PM leaf methanol extract had an inhibitory effect on *Bacillus cereus* (Zamri, 2015). Then, Imelda and his team found that PM leaf ethanol extract had inhibitory effects on *Escherichia coli* (Imelda et al., 2013). In subsequent experiments, they verified this finding (Imelda et al., 2014). This conclusion was also supported by Hassan et al. (Hassan et al., 2015). Not only that, PM leaf methanol, distilled water, and PM leaf 30% water-ethanol extracts also showed inhibitory effects on *Escherichia coli* (Abubakar et al.,

2015; Hassan et al., 2015; Hassim et al., 2015). PM leaf methanol and water extracts showed inhibitory effects on *Salmonella typhimurium* (Hassan et al., 2015).

Gastric ulcer is the most common digestive tract disease (Beiranvand & Bahramikia, 2020). If not treated in time, gastric ulcers will develop into gastritis, gastric bleeding, and even gastric cancer, endangering the life of patients (Scherübl, 2020). Therefore, finding and applying natural medicines with low toxicity and good biological activity to prevent gastric ulcers has gradually become the focus of researchers (X. Wang, Wang, Wen, & Li, 2023). The findings by Wasman et al. (2010) indicated that PM exhibited anti-ulcer activity, as evidenced by reduced ulcer areas, decreased edema, and decreased leucocyte infiltration in the submucosal layer. Their separate study had similar findings, suggesting that PM could be a promising plant for the treatment of gastric ulcers (S. Wasman, Ameen, Chua, & Hamdan, 2012). Researchers have also used elution solvents to separate PM leaves into different fractions to understand the pharmacological mechanisms behind the anti-ulcer activity of PM. The findings indicated that the ethyl acetate: methanol 1:1 fraction was the most effective in providing mucous protection in the ethanol induction model. The protective mechanisms were linked to the production of antioxidants and prostaglandin E2 (Qader, Abdulla, Chua, Sirat, & Hamdan, 2012).

The worldwide occurrence of liver disease continues to be a significant concern, with liver disease being the foremost cause of illness and mortality globally, resulting in a substantial public health challenge (Asrani, Devarbhavi, Eaton, & Kamath, 2019; Byass, 2014; Xiao et al., 2019). Drug-induced liver injuries from medications, in particular, have become a growing health problem (Fu et al., 2023). A study conducted by Thanapakiam et al. (2016) revealed that both aqueous and methanolic extracts of PM have substantial hepatoprotective properties. This finding is corroborated by other scientists who suggested that incorporating PM leaves into the diet could serve as a recommended approach to counteracting various types of chemical-induced liver damage (P. V. Christopher, Joe, et al., 2016). In addition, Manoontri, Joe, Tanusha, Parasuraman, and Christopher (2016) evaluated that the higher PM dose displayed substantial hepatoprotective activity. The histopathology study further confirmed the protective effect and the efficacy increased with the dosage.

Cisplatin is a well-known platinum-based anticancer agent, which is highly effective in treating various cancers (Abd Rashid et al., 2021). Hepatotoxicity has also been demonstrated in the patients who received low doses of cisplatin, which, probably due to cumulative effect in the liver, causes massive hepatic toxicity, including dissolution of hepatic cords, focal inflammatory lesions, and necrosis (Pratibha, Sameer, Rataboli, Bhiwgade, & Dhume, 2006; Singh & Khajuria, 2015). Fajar, Kusharyanti, and Wahdaningsih (2016) revealed that cisplatin-induced hepatotoxicity, leading to elevated levels of aspartate aminotransferase (AST) and alanine aminotransferase (ALT), along with a significant degree of liver damage compared to the control group. However, the ethyl acetate fraction from PM leaves exhibited hepatoprotective activity, as evidenced by reduced levels of AST and ALT. Moreover, Rashid et al. found PM given at 100 mg/kg had preventive effects against cisplatin-induced hepatotoxicity in rats (Rashid et al., 2019). Furthermore, they discovered PM essential oil at a dose of 100 mg/kg may protect against cisplatin-induced hepatotoxicity, possibly via inhibition of oxidative stress, inflammation, and apoptosis (Rashid et al., 2020).

7.4. Nervous system

Increasing evidence demonstrated that cisplatin could cross the blood-brain barrier and cause central nervous system injury (Andres, Gong, Di, & Bota, 2014). Fortunately, some studies showed that PM ethanolic extract at 100 and 200 mg/kg attenuated cisplatin-induced oxidative stress in the cerebral cortex via its antioxidant activity (Ridzuan et al., 2019). In other words, PM ethanol extract can alleviate drug

damage to the central nervous system. Besides, when there is injury or damage to the central nervous system, it can disrupt the normal functioning of these cognitive processes and lead to the development of cognitive impairment (Greenhalgh, David, & Bennett, 2020; Walker & Tesco, 2013). Cognitive impairment is sometimes neglected but common, which has a profound effect on the daily life of patients (Romo-Araiza & Ibarra, 2020). Evidence shows that patients with mild cognitive impairment have a far higher rate of developing chronic neurodegenerative diseases than cognitively normal persons (Goldman et al., 2018). Meanwhile, available medications merely slow the decline process but cannot reverse it (Sun, Ho, Zhang, Hong, & Zhang, 2023). Thus, alternative treatment approaches for mitigating cognitive impairment are greatly needed.

George, Ng et al. found that PM exhibited antioxidant and anticholinesterase activity, contributing to enhanced cognition *in vivo*. The results suggested that the extract possesses neuroprotective properties (George, Ng, et al., 2014). More than that, several studies have indicated that PM has positive effects on cognitive status and mood. The bioactive compounds present in PM may have neuroprotective effects, although there is still a lack of human studies examining the connection between PM consumption and cognitive status (You, Shahar, Haron, & Mastura Yahya, 2018). Additionally, according to reports, the supplementation of PM extract for six months can notably significantly improve visuospatial memory, reduce tension, anger, confusion, total negative subscales, triglyceride levels, and increase brain-derived neurotrophic factor levels in older adults with Mild Cognitive Impairment (Lau et al., 2020). Christopher et al. investigated the use of PM alleviated both paclitaxel- and scopolamine-induced neuropathic pain and cognitive impairments, showing a dose-dependent effect. A potential mechanism is PM exhibits potential neuroprotective effects attributed to its antioxidant properties, inhibition of lipid peroxidation, and regulation of cholinergic neurotransmitter functions (P. V. Christopher, Muthuraman, Zhang, Jordon, & Jonathan, 2021).

The findings of Yahya et al. indicated that PM supplementation could improve mood and quality of life, and participants did not report any adverse effects after six weeks of supplementation (H. M. Yahya et al., 2017). In addition, Bashir and colleagues evaluated the effects of PM extract on chronic ultra-mild stress-induced anorexia and anhedonia. Chronic administration of PM extract was found to reduce anorexia and improve anhedonia among stressed mice (Bashir, Aziz, & Noor, 2022a). Meanwhile, they suggested that the PM aqueous extract has antidepressant effects, as evidenced by reduced immobility time, enhanced spatial memory, decreased corticosterone levels, increased brain-derived neurotrophic factor levels, and lowered monoamine oxidase-A enzyme levels, leading to increased levels of monoamines (serotonin and norepinephrine) in the hippocampus (Bashir, Aziz, & Noor, 2022b). Besides, they explored the effects of PM aqueous effects on Kruppel-like factor11 and Sirtuin1 levels in the hippocampus of stressed mice. The results showed that PM aqueous extract treatment showed a significant reverse in an elevated level of Kruppel-like factor11 and Sirtuin1 in the hippocampus of stressed mice after treatment of 8 weeks (Bashir, Aziz, & Noor, 2022c). In another study, leaves of PM standardized extract, 1 and 100 mg/L, were used to evaluate the anti-stress effect in the chronic unpredictable stress zebrafish model. Both concentrations showed ameliorating effects only in the exploratory test. No significant changes were detected in the treatment groups. Cortisol analysis showed that after four days, the chronic unpredictable stress effect still affected the zebrafish Hypothalamus–Pituitary–Adrenal axis, but PM extract and fluoxetine treatment might need a more prolonged duration treatment to give the desired effect (Rahim et al., 2022). Moreover, the leaf component of PM has demonstrated significant potential as a future antidepressant option. This is attributed to the presence of flavonoids, particularly those involved in memory enhancement and the elevation of neurotransmitter levels in the brain, as reported. The current research observations suggest that conducting experimental screening for the antidepressant activity of the leaf part of PM is highly

recommended (Bashir et al., 2020).

Chronic depressive disorder or clinical depression has become a highly prevalent mental disorder affecting over 300 million populations worldwide (Haenisch & Bönisch, 2011; James et al., 2018). Natural medicinal plants and healthy food are considered alternative therapies to control the progression of depression and depressive-like symptoms with few side effects (Yeung et al., 2014). Many natural compounds, such as caffeine, green tea catechins, anthocyanins, ginsenosides, and resveratrol, are potential therapeutic agents with antidepressant-like bioactivities (A. Kang et al., 2017; Mao et al., 2020; Nabavi, Daglia, Braid, & Nabavi, 2017).

7.5. Cardiovascular system

Cardiovascular diseases (CVDs), a major cause of global mortality (Liao et al., 2023), are primarily fueled by hyperlipidemia, a substantial risk factor for CVDs (Karr, 2017; Prabhakaran et al., 2018). Christopher, Vikneswaran, et al. (2015) investigated results that revealed the anti-hyperlipidemic effect of leaves of PM in an acute hyperlipidemic rat model. The study also suggested that the methanol extract possesses a higher antihyperlipidemic effect than the aqueous extract (Christopher, Vikneswaran, et al., 2015). Besides, another significant risk factor for CVDs, hypertension, is also severely threatening human health (Castilla-Guerra, 2022; Gumprecht, Domek, Lip, & Shantsila, 2019; Suvila & Niiranen, 2022). Khalid & Salam found that Malaysian tropical plants, especially PM, are potential sources of natural antioxidant and antihypertensive agents (Khalid & Salam, 2018).

Upon chronic exposure, cadmium can accumulate in other body parts, including the heart and aorta, leading to various diseases (Tai et al., 2022). Kusumaningrum et al. (2019) demonstrated that PM leaves ethanol extract could protect the aortic wall against cadmium chloride exposure in mice. Afterward, they conducted another study to investigate the protective effect of PM leaf extract on the histopathological changes in the aorta wall in mice induced by cadmium chloride. The result showed PM leaf extract could protect mice's aorta walls from the damaging effect of cadmium chloride, with the optimum dose being 400 mg/kg bw (Kusumaningrum et al., 2020).

7.6. Urinary system

The kidney plays a pivotal role in eliminating toxic metabolites and is susceptible to toxicity induced by xenobiotics (Hassanen, Fahmi, Shams-Eldin, & Abdur-Rahman, 2020). Acute kidney injury, often caused by drugs, not only increases mortality and morbidity but also prolongs hospital stays, diminishing quality of life (Yu et al., 2021). For instance, cisplatin can cause severe deterioration in kidney function (Khan et al., 2013). The findings of Michael and his team revealed that PM leaf methanol extract had the ability to decrease urea and creatinine concentrations, suggesting its potential to mitigate the side effects of the chemotherapy agent cisplatin (Michael & Isnindar, 2013). Other researchers further studied and found the effective dose of PM leaf methanol fraction, which demonstrated nephroprotective activity in cisplatin-induced rats was 4.547 mg/200 g bw (Rita, Kusharyanti, & Wahdaningsih, 2013). Not only that, it has been proved that 1308 mg/200 g bw of PM leaves n-hexane extract decreased kidney damage and urea and creatinine levels, which indicates that it has the potency to reduce the side effects of chemotherapy agent cisplatin (Tommy, 2013). Kidneys are the first target site for heavy metal (mercury) deposits and toxicity (Vieira et al., 2021). The protective effect of PM leaf extract on the histopathological changes of kidneys induced by mercuric chloride in mice was investigated by Aprianti, Widiyatno, and Sudjarwo (2017). Their results showed that PM leaf extract could protect mice kidneys from the damaging effect of mercuric chloride, with the best dose of PM 400 mg/kg bw (Aprianti et al., 2017).

7.7. Reproductive system

Mercury chloride causes harmful effects on the liver, blood, kidneys, and the male reproductive system (Apaydin, Baş, Kalender, & Kalender, 2016; Uzunhisarcikli, Aslanturk, Kalender, Apaydin, & Bas, 2016). It induces various pathological processes and membrane damage, ultimately resulting in the loss of sperm motility (Agarwal, Saleh, & Bedaiwy, 2003; Vachhrajani K.D. et al., 1988). Zulfar (2017) demonstrated that the ethanol extract of PM leaves can act as a preventive measure against the toxic effects of mercury chloride on the thickness of seminiferous tubule epithelium and the count of spermatocytes. The group treated with 200 mg/kg bw of PM ethanol extract exhibited the most favorable results (Zulfar, 2017).

Cadmium has been shown to lower testosterone and steroidogenic enzyme levels, and decrease sperm concentration and viability, ultimately disrupting the fertilization process (de Angelis et al., 2017; Hernández-Rodríguez et al., 2016). The findings of Siswanto suggested that PM leaves ethanol extract could shield the spermatogenic cells in the testes of mice from the harmful effects of cadmium chloride, with the optimal dose being 400 mg/kg bw (Siswanto, 2018).

The potential of certain substances to enhance sexual behavior has prompted studies on aphrodisiacs derived from plants (Melnyk & Marccone, 2011). Udani et al. revealed that supplementing with PM and the proprietary *Eurycoma longifolia* extract, Physta, for twelve weeks was well-tolerated and more effective than a placebo in improving sexual performance in healthy volunteers (Udani et al., 2014).

7.8. Immune system

A disturbance in the components of the immune system leads to the development of various health problems (Brindha & Pavelic, 2016). Hence, it is of great significance to search for an immunomodulatory agent that affects the immune system (Chen et al., 2022). Recently, natural substances have been attracting increased attention owing to their multiple biological activities, including immunomodulation (Qin et al., 2022; Xie, Huang, Meng, Shi, & Xie, 2022). George, Chinnappan, Choudhary, Bommu, and Sridhar (2014) investigated the immune system-modulating characteristics of PM aqueous extract on Swiss albino mice through the carbon clearance assay. The extract demonstrated a notable enhancement in phagocytosis at doses of 200 and 400 mg/kg bw (George, Chinnappan, et al., 2014).

Furthermore, Herpes simplex virus type 1 (HSV-1) employed several strategies to evade the host's immune system, allowing it to establish latent infections and periodically reactivate (Verzosa et al., 2021). Therefore, there is an increasing need to find new antiviral compounds where pure compounds of plant origin have been shown to possess antiviral activity against HSV types (El-Toumy et al., 2018). Shahar et al. found that aqueous extract of PM could be a potential candidate for anti-HSV-1 activity (Shahar, Hamdan, Baba, & Paul Joko, 2015). Indeed, further studies are required before a conclusive experimental finding is suggested.

Based on these above observations, it is evident that PM is a plant with various bioactive compounds that have the potential for medicinal properties and are effective in combating and treating common illnesses. Numerous *in vitro* and *in vivo* studies have demonstrated the other therapeutic activities of PM against body systems diseases, sexual performance and well-being, cognitive function and mood, and microbial infections.

8. Application of nanotechnology in PM

The tremendous progress of nanotechnology and nanomedicine has brought a variety of opportunities to the development of nanomedicines, which are extensively exploited to ameliorate the solubility of insoluble drugs (Hu, Johnston, & Williams, 2004) and to achieve sustained drug release (Kamaly, Yameen, Wu, & Farokhzad, 2016; Peng et al., 2013),

long-circulating and targeted delivery (Moghimi, Hunter, & Murray, 2001), and spatial-temporally controllable drug release (D. Huang et al., 2020; Y. Wu, Wang, Huang, Yang, & Wang, 2017). Application of nanotechnology in the synthesis of plant material, especially in functional properties, is expected to improve solubility and bioavailability, protect from toxicity, enhance pharmacological activity, enhance stability, improve tissue macrophage distribution, and protect from physical and chemical degradation (Nadzirah et al., 2014). Likewise, PM has been studied and prepared into various nanoparticles due to its wide range of uses and activities (Fig. 10).

8.1. PM extract-loaded gold nanoparticles

Gold nanoparticles (AuNPs) have been applied as an antimicrobial agent, cancer therapy, and diagnostic tools in fluorescence tomography, among others (Firdhouse & Lalitha, 2022; Murphy et al., 2008; Sardar, Funston, Mulvaney, & Murray, 2009; Wei, Famouri, Carroll, Lee, & Famouri, 2013; P. Zhao, Li, & Astruc, 2013). Initially, Borhamdin, Shamsuddin, and Alizadeh (2014) presented a bioinspired method for synthesizing magnetically recoverable AuNPs catalysts on a Fe₃O₄@-SiO₂ support. Subsequently, they investigated an environmentally friendly method for synthesizing AuNPs using an aqueous leaf extract of PM. This biosynthesis is simple, time-saving, and eco-friendly, requiring only 20 min to produce AuNPs with an average particle size of 23 nm (Borhamdin et al., 2014). The newly prepared biostabilized icosahedral AuNPs exhibited good catalytic activity in reducing 4-nitrophenol to 4-aminophenol (Borhamdin, Shamsuddin, & Alizadeh, 2015).

8.2. PM extract-loaded silver nanoparticles

Silver nanoparticles (AgNPs) have been widely studied in various scientific applications due to their unique physicochemical and biological features. These include ease of functionalization or connection with different ligands for customized properties, antimicrobial toxicity, effective cytotoxicity against cancer cells, catalytic applications, and more (Akter et al., 2018; Jain, Huang, El-Sayed, & El-Sayed, 2008).



Fig. 10. Various PM nanoparticles. AuNPs, PM extract-loaded gold nanoparticles; AgNPs, PM extract-loaded silver nanoparticles; CuNPs, PM extract-loaded copper nanoparticles; NiNPs, PM extract-loaded nickel nanoparticles; PMNPs, nano-size PM powder. (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

Ullah et al. introduced a method using aqueous IL and microwave extraction to extract lignocellulosic biomass from PM. They achieved a high yield of nanoparticles using recycled ILs (Ullah, Wilfred, & Shaharun, 2017). Additionally, the PM plant extract was employed in the synthesis of AgNPs from a silver nitrate solution. The initially colorless mixture gradually transformed from yellowish-green to reddish-brown, indicating the reduction of silver ions (Ag⁺) over several minutes. Three types of bacteria—*Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas aeruginosa*—were selected for testing. Morphological changes in bacterial cells treated with AgNPs were observed using a field emission scanning electron microscope (FESEM), revealing the excellent antimicrobial properties of AgNPs against these microorganisms (Nadzirah, Mahamudin, Muhamad, & Latif Ibrahim, 2019). In a different investigation, Kamarudin and colleagues established an environmentally friendly method for producing ultrafine and well-defined spherical AgNPs (5–20 nm) using an IL. The extract from PM leaves served as both a green reducing agent and a capping agent in the synthesis process. The findings indicated that the IL and phenolic compounds worked synergistically to reduce Ag⁺ to AgNPs and stabilize the nanoparticles (Kamarudin et al., 2022).

8.3. PM extract-loaded copper, nickel nanoparticles

Ullah and the team (2018a) showed that bioactive compounds from PM can be extracted using an IL-based microwave-assisted method. The obtained extract was then employed to produce copper nanoparticles (CuNPs). UV/Vis results showed that this synthesis was fast, well dispersed, and nanosized CuNPs in comparison to conventional synthesis. Moreover, the antibacterial activity of the synthesized CuNPs exhibited an effective inhibitory zone against three different bacteria, namely *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas aeruginosa* (Ullah, Wilfred, & Shaharun, 2018a). Ullah and the team (2018b) performed an additional experiment to extract bioactive compounds from PM using 1-ethyl-3-methyl imidazolium chloride along with ultrasonication. The resulting extract was utilized to create nickel nanoparticles (NiNPs). The synthesized NiNPs demonstrated high effectiveness against three different bacteria—*Aeromonas hydrophila*, *Escherichia coli*, and *Staphylococcus aureus*—in disc diffusion tests at various concentrations (Ullah, Wilfred, & Shaharun, 2018b).

8.4. Nano-size PM powder

The recently developed superfine grinding technology as a useful tool for preparing super fine powder has opened a vast area of research and development for the medicinal benefits of natural food products (Archana, Aman, Singh, Kr, & Jabeen, 2021). This approach has the potential to enhance surface area, thereby improving bioavailability, distribution, and absorption (Xiaoyan, Yang, Gai, & Yang, 2009; Ya Ling, Sheu, Lee, & Chau, 2008). Additionally, it contributes to improved dispersibility, solubility, and water-holding capacity (Ramachandraiah & Chin, 2016). Nadzirah et al. utilized a planetary ball mill to reduce the particle sizes of PM powder efficiently, achieving a successful preparation at the nanoscale (Nadzirah, Rusop, & Latif, 2015a). Afterward, the same team explored the changes in the physical properties of PM resulting from the grinding effect. The planetary ball mill successfully reduced the particle size of PM to the range of 227 nm–241 nm. FESEM images demonstrated significant alterations in particle size morphology, featuring a higher degree of cell breakage attributed to the planetary ball mill process. Atomic force microscopy results indicated changes in particle roughness due to the grinding process. Furthermore, energy-dispersive X-ray analysis revealed that the sample exhibited the highest elements of carbon and oxygen, averaging 86%, suggesting the presence of carbohydrates (Nadzirah, Rusop, & Latif, 2015b).

9. Conclusion and future perspectives

PM is an aromatic plant that has gained popularity owing to its excellent nutritional value. It is extensively used for nutritional and medicinal purposes. Despite the usage of PM in food and herbal medicine in countries like Malaysia and India, limited information is available about its therapeutics and pharmacology. Notably, extremely few articles were found on the application potential of this species, with particular reference to its applications in the food, agricultural, medicinal, and nanotechnology fields. This review discusses the above-said issues in detail and in a scientific manner that could be beneficial for the scientific community for the development of PM applications for the benefit of society. However, there are still some problems to be solved in the later application of PM.

First of all, the variability of the composition profile, amounts of individual components, and yield of the extract from the PM materials is due to the influence of different factors. Therefore, selecting the best parts and most effective methods of extracting from PM is essential to preserve the quality and achieve maximum effect.

Second, most of the studies observed positive effects on the characteristics of products like meat, vegetables, and fruits after the addition of PM extracts by controlling the quality deterioration (microbial growth inhibition, antioxidant activity, and preservation of sensory properties). Besides the promising results obtained from the literature, there is much to achieve concerning product quality, shelf life, and consumer acceptance of PM extract-based additives and films on food products. For example, PM extract-based biodegradable films are lacking in the market because there are still obstacles to using sustainable polymers instead of synthetic ones. For this reason, more detailed studies must be done to improve the properties of the additives, films, and coatings for the industrial application of food since the improvement of some material characteristics is generally associated with a decrease in others.

Third, although PM powder/extract has been applied to test all types of animals, some challenges still need to be solved for large-scale feed production. Life cycle assessment of animal feed production using PM should be further investigated for better environmental impact, particularly in the livestock, poultry, and aquaculture industries. In order to maximize the quantity and quality of extracted nutritive values and secondary metabolites in PM, each process, such as drying methods, extraction techniques, and analytical methods, should be optimized. This information can help feed formulators fully exploit the presence of nutritional value and secondary metabolites in PM when devising diets for animals.

Fourth, although insights have been revealed about the pharmacology properties of PM, some gaps between basic studies and clinical application still exist, which limits the use of PM for body disease clinical treatment to a certain extent. Therapeutic and safe doses of active ingredients from PM need to be established based on recent pharmacological and pharmacokinetic trials in nonclinical or clinical settings. At the same time, studies in which the ingredients of PM are extracted and then applied to simultaneously explore the suitable cellular, animal, and clinical levels are lacking.

Finally, nanoparticle biosynthesis by PM is eco-friendly, easy to access, non-toxic, and cost-effective, and it enhances the therapeutic efficacy of herbal drugs. These delivery systems ensure the delivery of plant bio-actives to the targeted site, such as the liver, brain, heart, kidney, etc., with low doses in contrast with conventional plant extracts or phytomolecules. However, despite PM applications in nanotechnology providing enough information, they are not enough. On the one hand, studies are required to explore the mechanism and pathways PM uses through the biosynthesis of nanoparticles. On the other hand, there are still possibilities for the development of new carriers in order to reduce the toxicity and enhance the therapeutic efficacy of herbal drugs. Meanwhile, the factors and conditions of the PM-based NP synthesis need to be developed on a large scale to be suitable for commercial production. Besides, various fields and applications need to study the

impact of PM-NPs.

In addition to the discussed fields, strategic exploration of the understanding and application of PM using biotechnology may be a good approach. These methods aim to optimize the production and utilization of PM bioactive compounds for various fields. Such as metabolic pathways in PM can be improved through genetic engineering to increase the yield and consistency of bioactive compounds. Targeted genetic modification can produce PM varieties with higher therapeutic efficacy and improved nutritional status. In addition, using genomics, proteomics, and metabolomics can provide a comprehensive understanding of the molecular mechanisms behind various PM applications. This information can guide the development of improved extraction methods, and application strategies.

In general, PM and its main bioactive compounds have enormous application potential and represent potential candidates for food, feed, and medicine. Future research is urgently required to exploit of this promising resource with efficient green protocols, and additional alternative uses should also be investigated to facilitate the overall value-added aspect of this vital herb.

CRediT authorship contribution statement

Zhongming Yang: Writing – original draft. **Xi Deng:** Writing – original draft. **Zhongguo Yang:** Writing – original draft. **Mingzhao Han:** Writing – original draft. **Norsharina Ismail:** Writing – review & editing, Funding acquisition, Conceptualization. **Kim Wei Chan:** Writing – review & editing. **Ahmad Faizal Abdull Razis:** Writing – review & editing. **Norhaizan Mohd Esa:** Writing – review & editing. **Ket Li Ho:** Writing – review & editing. **Md Zuki Abu Bakar:** Funding acquisition, Conceptualization.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

Data will be made available on request.

Acknowledgments

This work was supported by the internal Universiti Putra Malaysia, Geran Putra IPS (GP-IPS/2023/9748400) and Universiti Putra Malaysia, Geran Putra-IPM (GP-IPM/2020/9684800).

Glossary

PM	<i>Polygonum minus</i>
bw	body weight
GC-MS	gas chromatography–mass spectrometry
SPME	Solid Phase Microextraction
TPC	total phenolic content
TFC	total flavonoid content
TP	total phenolic
TF	total flavonoid
FRAP	ferric reducing antioxidant power
DPPH	2,2-diphenyl-1-picrylhydrazyl
IL	ionic liquid
ILs	ionic liquids
PV	peroxide value
TBA	thiobarbituric acid
FFA	free fatty acid
PLA	polylactic acid
PBS	polybutylene succinate

CLE	curry leaf extract
AST	aspartate aminotransferase
ALT	alanine aminotransferase
CVDs	cardiovascular diseases
MIC	minimal inhibitory concentrations
MBC	minimal bactericidal concentration
HSV-1	herpes simplex virus 1
AuNPs	gold nanoparticles
AgNPs	silver nanoparticles
Ag+	silver ions
FESEM	field emission scanning electron microscope
CuNPs	copper nanoparticles
NiNPs	nickel nanoparticles
NPs	nanoparticles

References

- Abas, F., Lajis, N. H., Israif, D. A., Khozirah, S., & Kalsom, Y. U. (2006). Antioxidant and nitric oxide inhibition activities of selected Malay traditional vegetables. *Food Chemistry*, 95(4), 566–573. <https://doi.org/10.1016/J.FOODCHEM.2005.01.034>
- Abd El-Hack, M. E., El-Saadony, M. T., Elbestawy, A. R., Gado, A. R., Nader, M. M., Saad, A. M., et al. (2022). Hot red pepper powder as a safe alternative to antibiotics in organic poultry feed: An updated review. *Poultry Science*, 101(4), Article 101684. <https://doi.org/10.1016/j.psj.2021.101684>
- Abd Rashid, N., Abd Halim, S. A. S., Teoh, S. L., Budin, S. B., Hussain, F., Adib Ridzuan, N. R., et al. (2021). The role of natural antioxidants in cisplatin-induced hepatotoxicity. *Biomedicine & Pharmacotherapy*, 144, Article 112328. <https://doi.org/10.1016/J.BIOPHA.2021.112328>
- Abd-ElGawad, A. M., Assaeed, A. M., Al-Rowaily, S. L., Alshahri, M. S., Bonanomi, G., & Elshamy, A. I. (2023). Influence of season and habitat on the essential oils composition, allelopathy, and antioxidant activities of *Artemisia monosperma Delile*. *Separations*, 10(4), 263. <https://doi.org/10.3390/separations10040263>
- Abd-Elgawad, A. M., Elgamal, A. M., El-Amier, Y. A., Mohamed, T. A., El Gendy, A. E. N. G., & Elshamy, A. I. (2021). Chemical composition, allelopathic, antioxidant, and anti-inflammatory activities of sesquiterpenes rich essential oil of cleome amblyocarpa barrante & murb. *Plants*, 10(7), 1294. <https://doi.org/10.3390/plants10071294>
- Abd-ElGawad, A. M., Elshamy, A. I., El-Nasser El Gendy, A., Al-Rowaily, S. L., & Assaeed, A. M. (2019). Preponderance of oxygenated sesquiterpenes and diterpenes in the volatile oil constituents of *Lactuca serriola L.* revealed antioxidant and allelopathic activity. *Chemistry and Biodiversity*, 16(8), Article e1900278. <https://doi.org/10.1002/cbdv.201900278>
- Abdel-Latif, H. M. R., Dawood, M. A. O., Alagawany, M., Faggio, C., Nowosad, J., & Kucharczyk, D. (2022). Health benefits and potential applications of fucoidan (PCD) extracted from brown seaweeds in aquaculture: An updated review. *Fish & Shellfish Immunology*, 122, 115–130. <https://doi.org/10.1016/J.FSI.2022.01.039>
- Abdel-Latif, H. M. R., Dawood, M. A. O., Menanteau-Ledouble, S., & El-Matbouli, M. (2020). The nature and consequences of co-infections in tilapia: A review. *Journal of Fish Diseases*, 43(6), 651–664. <https://doi.org/10.1111/JFD.13164>
- Abdel-Latif, H. M. R., & Khafaga, A. F. (2020). Natural co-infection of cultured Nile tilapia *Oreochromis niloticus* with *Aeromonas hydrophila* and *Gyrodactylus cichlidarum* experiencing high mortality during summer. *Aquaculture Research*, 51(5), 1880–1892. <https://doi.org/10.1111/ARE.14538>
- Abdul Azziz, S. S. S., Talip, M. A., Wong, C. F., Alimon, H., Bakar, N. A., Wan Mahamad, W. R., et al. (2015). Determination of Malaysian herbs and spices as biopreservative agents in food products. *American Journal of Plant Sciences*, 6(5), 718–724. <https://doi.org/10.4236/AJPS.2015.65077>
- Abdullah, M. Z., Mohd Ali, J., Abolmaesoomi, M., Abdul-Rahman, P. S., & Hashim, O. H. (2017). Anti-proliferative, *in vitro* antioxidant, and cellular antioxidant activities of the leaf extracts from *Polygonum minus* Huds: Effects of solvent polarity. *International Journal of Food Properties*, 20(1), S846–S862. https://doi.org/10.1080/10942912.2017.1315591/SUPPL_FILE/LJFP_A_1315591_SM4163.DOCX
- Aboushanab, S. A., El-Far, A. H., Narala, V. R., Ragab, R. F., & Kovaleva, E. G. (2021). Potential therapeutic interventions of plant-derived isoflavones against acute lung injury. *International Immunopharmacology*, 101, Article 108204. <https://doi.org/10.1016/J.INTIMP.2021.108204>
- Abubakar, M. A., Zulkifli, R. M., Atiqah, W. N., Hassan, W., Husni, A., Shariff, M., et al. (2015). Antibacterial properties of *Pescicaria minor* (Huds.) ethanolic and aqueous-ethanolic leaf extracts. *Journal of Applied Pharmaceutical Science*, 5(2), 50–56. <https://doi.org/10.7324/JAPS.2015.58.58>
- Adel, M., Dawood, M. A. O., Shafiei, S., Sakhaie, F., & Shekarabi, S. P. H. (2020). Dietary *Polygonum minus* extract ameliorated the growth performance, humoral immune parameters, immune-related gene expression and resistance against *Yersinia ruckeri* in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*, 519, Article 734738. <https://doi.org/10.1016/J.AQUACULTURE.2019.734738>
- Agarwal, A., Saleh, R. A., & Bedaiwy, M. A. (2003). Role of reactive oxygen species in the pathophysiology of human reproduction. *Fertility and Sterility*, 79(4), 829–843. [https://doi.org/10.1016/S0015-0282\(02\)04948-8](https://doi.org/10.1016/S0015-0282(02)04948-8)
- Ahmad, R., Baharum, S. N., Bunawan, H., Lee, M., Noor, N. M., Rohani, E. R., et al. (2014). Volatile profiling of aromatic traditional medicinal plant, *Polygonum minus* in

- different tissues and its biological activities. *Molecules*, 19(11), 19220–19242. <https://doi.org/10.3390/molecules191119220>
- Ahmad, R., Bunawan, H., Normah, M. N., & Baharum, S. N. (2016). Chemical composition in different tissues of *Polygonum minus* by using GC X GC-TOF MS and direct discrimination by multivariate analysis of fourier transform infrared spectroscopy data. *International Journal of Pharmacognosy and Phytochemical Research*, 8(12), 1986–1992.
- Ahmad, R., Rosandy, A. R., Sahidin, I., Syatila, N., Ghani, A., Noor, N. M., et al. (2023). Bioassay analysis and molecular docking study revealed the potential medicinal activities of active compounds Polygonumins B, C and D from *Polygonum minus* (*Persicaria minor*). *Plants*, 12(1), 59.
- Ahmad, R., Sahidin, I., Taher, M., Low, C., Noor, N. M., Sillapachaiyaporn, C., et al. (2018). Polygonumins A, a newly isolated compound from the stem of *Polygonum minus* Huds with potential medicinal activities. *Scientific Reports*, 8(1), 4202. <https://doi.org/10.1038/s41598-018-22485-5>
- Ahmed, A. F., Attia, F. A. K., Liu, Z., Li, C., Wei, J., & Kang, W. (2019). Antioxidant activity and total phenolic content of essential oils and extracts of sweet basil (*Ocimum basilicum* L.) plants. *Food Science and Human Wellness*, 8(3), 299–305. <https://doi.org/10.1016/j.fshw.2019.07.004>
- Ahsan, T., Chen, J., Zhao, X., Irfan, M., & Wu, Y. (2017). Extraction and identification of bioactive compounds (eicosane and dibutyl phthalate) produced by *Streptomyces* strain KX852460 for the biological control of *Rhizoctonia solani* AG-3 strain KX852461 to control target spot disease in tobacco leaf. *AMB Express*, 7(1), 1–9. <https://doi.org/10.1186/s13568-017-0351-z>
- Ajlouli, R. El, Raji, H., Ouairi, M. El, Chakroun, A., Zeroual, A., Syed, A., et al. (2024). MEDT Study, hemisynthesis via regioselective ring opening of α -Himachalene-Epoxides, ADME survey and docking studies designed to target coronavirus and HIV-1. *Journal of Molecular Liquids*, 394, Article 123707. <https://doi.org/10.1016/j.molliq.2023.123707>
- Akter, M., Sikder, M. T., Rahman, M. M., Ullah, A. K. M. A., Hossain, K. F. B., Banik, S., et al. (2018). A systematic review on silver nanoparticles-induced cytotoxicity: Physicochemical properties and perspectives. *Journal of Advanced Research*, 9, 1–16. <https://doi.org/10.1016/J.JARE.2017.10.008>
- Al-Amin, M., Rahiman, S. S. F., Khairuddean, M., & Salhimi, S. M. (2023). Chemical constituents of curcuma zanthorrhiza and the activity of (R)-(-)- α -curcumene on the migration and invasion of MDA-MB-231 cell line. *Revista Brasileira de Farmacognosia*, 33(6), 1243–1250. <https://doi.org/10.1007/s43450-023-00449-9>
- Alawiyah, F., Kahtan, M. I., & Widiyantoro, A. (2016). Daya antelmintik ekstrak metano daun kesum (*Polygonum minus*) terhadap ascaridia galli secara in vitro abstrak pendahuluan infeksi cacing merupakan kolik di daerah epigastrium atau umbilikus. emesis, konstipasi, perut terkadang juga dapat dijumpai. I. *Jurnal Mahasiswa PSPD FK Universitas Tanjungpura*, 2(4), 657–666.
- Albratty, M., Alhazmi, H. A., Meraya, A. M., Najmi, A., Alam, M. S., Rehman, Z., et al. (2021). Spectral analysis and antibacterial activity of the bioactive principles of *Sargassum tenerrimum* J. Agardh collected from the Red sea, Jazan, Kingdom of Saudi Arabia. *Brazilian Journal of Biology*, 83, Article e249536. <https://doi.org/10.1590/1519-6984.249536>
- Alencar, M. V. O. B., Islam, M. T., Ali, E. S., Santos, J. V. O., Paz, M. F. C. J., Sousa, J. M. C., et al. (2018). Association of phytol with toxic and cytotoxic activities in an antitumor perspective: A meta-analysis and systemic review. *Anti-Cancer Agents in Medicinal Chemistry*, 18(13), 1828–1837. <https://doi.org/10.2174/1871520618666180821113830>
- Ali, A. M., Muhammad, I., Maceken, M., Ei-Sharkawy, S. H., Hamidi, J. A., Ahmadi, B. H., et al. (1996). Antiviral and cytotoxic activities of some plants used in Malaysian indigenous medicine. *Journal of Tropical Agricultural Science*, 19(3), 129–136.
- Alonso-Torre, S., Carrillo, C., & Cavia, M. M. (2012). Role of oleic acid in immune system; mechanism of action; a review. *Nutricion Hospitalaria*, 27(4), 978–990. <https://doi.org/10.3305/nh.2012.27.4.5783>
- Alves, T. M. D. A., Ribeiro, F. L., Kloos, H., & Zani, C. L. (2001). Polygodial, the fungitoxic component from the Brazilian medicinal plant *Polygonum punctatum*. *Memorias Do Instituto Oswaldo Cruz*, 96(6), 831–833. <https://doi.org/10.1590/S0074-02762001000600016>
- Alwazeer, D., Elnasanelkasim, M. A., Çicek, S., Engin, T., Çiğdem, A., & Karaoğlu, E. (2023). Comparative study of phytochemical extraction using hydrogen-rich water and supercritical fluid extraction methods. *Process Biochemistry*, 128, 218–226. <https://doi.org/10.1016/J.PROCBIO.2023.01.022>
- Ambroz, M., Boušová, I., Skarka, A., Hanušová, V., Králová, V., Matoušková, P., et al. (2015). The influence of sesquiterpenes from *Myrica rubra* on the antiproliferative and pro-oxidative effects of doxorubicin and its accumulation in cancer cells. *Molecules*, 20(8), 15343–15358. <https://doi.org/10.3390/molecules200815343>
- Ambroz, M., Matoušková, P., Skarka, A., Zajdlová, M., Žáková, K., & Skálová, L. (2017). The effects of selected sesquiterpenes from myrica rubra essential oil on the efficacy of doxorubicin in sensitive and resistant cancer cell lines. *Molecules*, 22(6), 1021. <https://doi.org/10.3390/molecules22061021>
- Amiruddin, F., Wahab, I. A., & Mohsin, H. F. (2020). Review on *Polygonum minus*: Pharmacological properties and phytochemical compounds. *Journal of Engineering and Health Sciences*, 4, 123–134.
- Ananthi, R., Chandra, N., Santhiya, S. T., & Ramesh, A. (2010). Genotoxic and antigenotoxic effects of *Hemidesmus indicus* R. Br. root extract in cultured lymphocytes. *Journal of Ethnopharmacology*, 127(2), 558–560. <https://doi.org/10.1016/J.JEP.2009.10.034>
- Andres, A. L., Gong, X., Di, K., & Bota, D. A. (2014). Low-doses of cisplatin injure hippocampal synapses: A mechanism for “chemo” brain? *Experimental Neurology*, 255, 137–144. <https://doi.org/10.1016/J.EXPNEURO.2014.02.020>
- Annaz, H., El Fakhouri, K., Ben Bakrim, W., Mahdi, I., El Bouhssini, M., & Sobeh, M. (2023). Bergamotenes: A comprehensive compile of their natural occurrence, biosynthesis, toxicity, therapeutic merits and agricultural applications. *Critical Reviews in Food Science and Nutrition*, 1–20. <https://doi.org/10.1080/10408398.2023.2184766>
- Ansiah, S. W. (2014). Formulation of kesum leaves (*Polygonum minus* Huds) polar fraction as antiseptic gel. *Jurnal Mahasiswa Farmasi Fakultas Kedokteran UNTAN*, 1–8.
- Aparna, V., Dileep, K. V., Mandal, P. K., Karthe, P., Sadasivan, C., & Haridas, M. (2012). Anti-inflammatory property of n-hexadecanoic acid: Structural evidence and kinetic assessment. *Chemical Biology & Drug Design*, 80(3), 434–439. <https://doi.org/10.1111/j.1747-0285.2012.01418.x>
- Apaydin, F. G., Baş, H., Kalender, S., & Kalender, Y. (2016). Subacute effects of low dose lead nitrate and mercury chloride exposure on kidney of rats. *Environmental Toxicology and Pharmacology*, 41, 219–224. <https://doi.org/10.1016/J.ETAP.2015.12.003>
- Aprianti, W., Widiyatno, T. V., & Sudjarwo, S. A. (2017). Effect of *Polygonum minus* (knotweed) leaves extract on the histopathological changes of kidney in mice (*Mus musculus*) induced by mercuric chloride. *KnE Life Sciences*, 3(6), 753. <https://doi.org/10.18502/kl.v3i6.1206>
- Archana, Aman, A. K., Singh, R. K., Kr, N., & Jabeen, A. (2021). Effect of superfine grinding on structural, morphological and antioxidant properties of ginger (*Zingiber Officinale*) nano crystalline food powder. *Materials Today: Proceedings*, 43, 3397–3403. <https://doi.org/10.1016/J.MATPR.2020.09.028>
- Asakawa, Y., Yoyota, M., Takemoto, T., Kubo, I., & Nakanishi, K. (1980). Insect antifeedant secoaromadendrane-type sesquiterpenes from *Plagiochila* species. *Phytochemistry*, 19(10), 2147–2154. [https://doi.org/10.1016/S0031-9422\(00\)82212-8](https://doi.org/10.1016/S0031-9422(00)82212-8)
- Asanai, M. F., Haron, N., Camalxaman, S. N., & Mohamed, E. (2020). A review on medicinal benefits of *Persicaria minor*. *Health Scope*, 3(2), 44–50.
- Asrani, S. K., Devarbhavi, H., Eaton, J., & Kamath, P. S. (2019). Burden of liver diseases in the world. *Journal of Hepatology*, 70(1), 151–171. <https://doi.org/10.1016/J.JHEP.2018.09.014>
- Awad, E., & Awaad, A. (2017). Role of medicinal plants on growth performance and immune status in fish. *Fish & Shellfish Immunology*, 67, 40–54. <https://doi.org/10.1016/J.FSI.2017.05.034>
- Az-Zahra Tashim, N., Harkani, S., Abdullah Lim, S., & Maryam Basri, A. (2021). Shelf-life assessment of fortified wholemeal pasta with *Amaranthus tricolor* and *Polygonum minus*. *Malaysian Journal of Biochemistry and Molecular Biology*, 3, 96–104. <http://mjmbm.org>
- Azad, A. K., Sulaiman, W. M. A. W., & Kundu, S. K. (2022). Toxicity profile of *Phaleria macrocarpa* (Scheff.) Boerl. fruits extract in adult male Sprague Dawley rats. *Advances in Traditional Medicine*, 22(3), 557–567. <https://doi.org/10.1007/S13596-021-00592-5/FIGURES/7>
- Azhari, N. A. M., Markom, M., Ismail, I., & Anuar, N. (2020). Effects of different drying methods on essential oil yield and component profile of *Polygonum minus* root extract. *International Food Research Journal*, 27(1), 96–102.
- Azhari, N. A. M., Markom, M., Ismail, I., & Anuar, N. (2022). Effects of CO₂ flow rate in supercritical fluids extraction of *Polygonum minus* roots is state. *Jurnal Kejuruteraan*, 34(6), 1281–1286.
- Aziman, N., Abdullah, N., Mohd Noor, Z., Zulkifli, K. S., & Wan Kamarudin, W. S. S. (2012). Phytochemical constituents and in vitro bioactivity of ethanolic aromatic herb extracts. *Sains Malaysiana*, 41(11), 1437–1444.
- Babayev, A., Spasojević, L., Jelena, J., JelenaŠkrbić, J., Bučko, S., Kocić-Tanackov, S., et al. (2023). Antimicrobial pseudolatex zein films with encapsulated carvacrol for sustainable food packaging. *Food Packaging and Shelf Life*, 37, Article 101076. <https://doi.org/10.1016/j.fpsl.2023.101076>
- Baharum, S. N., Bunawan, H., Ghani, M. A., Wan, A. W. M., & Noor, N. M. (2010). Analysis of the chemical composition of the essential oil of *Polygonum minus* Huds. using two-dimensional gas chromatography-time-of-flight mass spectrometry (GC-TOF MS). *Molecules*, 15(10), 7006–7015. <https://doi.org/10.3390/molecules15107006>
- Bahi, A., Al Mansouri, S., Al Memari, E., Al Ameri, M., Nurulain, S. M., & Ojha, S. (2014). β -Caryophyllene, a CB2 receptor agonist produces multiple behavioral changes relevant to anxiety and depression in mice. *Physiology and Behavior*, 135, 119–124. <https://doi.org/10.1016/j.physbeh.2014.06.003>
- Bakir, S., Hall, R. D., de Vos, R. C. H., Mumm, R., Kadakal, Ç., & Capanoglu, E. (2023). Effect of drying treatments on the global metabolome and health-related compounds in tomatoes. *Food Chemistry*, 403, Article 134123. <https://doi.org/10.1016/J.FOODCHEM.2022.134123>
- Bakir, B., Him, A., Özbek, H., Düz, E., & Tütüncü, M. (2008). Investigation of the anti-inflammatory and analgesic activities of β -caryophyllene. *International Journal of Essential Oil Therapeutics*, 2(1), 41–44.
- Baradaran, A., HooiLing, F., ChinChin, S., & Rahim, R. A. (2012). Isolation, identification and characterization of lactic acid bacteria from *Polygonum minus*. *Romanian Biotechnological Letters*, 17(3), 7245–7252.
- Basha, R. H., & Sankaranarayanan, C. (2014). β -Caryophyllene, a natural sesquiterpene, modulates carbohydrate metabolism in streptozotocin-induced diabetic rats. *Acta Histochemica*, 116(8), 1469–1479. <https://doi.org/10.1016/j.acthis.2014.10.001>
- Bashir, M. I., Aziz, N. H. K. A., & Noor, D. A. M. (2020). Possible antidepressant potential of a cognitive enhancer *Polygonum minus* based on its major chemical constituents in leaf part. *Drug Invention Today*, 13(4), 549–557.
- Bashir, M. I., Aziz, N. H. K. A., & Noor, D. A. M. (2022a). *Polygonum minus* aqueous extract supplement reduces stress-induced anorexia and anhedonia in mice. *Pakistan Journal of Medical and Health Sciences*, 16(1), 696–701. <https://doi.org/10.53350/pjmhs22161696>
- Bashir, M. I., Aziz, N. H. K. A., & Noor, D. A. M. (2022b). Antidepressant-like effects of *Polygonum minus* aqueous extract in chronic ultra-mild stress-induced depressive mice model. *Behavioral Sciences*, 12(6), 196. <https://doi.org/10.3390/BS12060196>

- Bashir, M. I., Aziz, N. H. K. A., & Noor, D. A. M. (2022c). Influence of *Polygonum minus* aqueous extract on monoamine oxidase-a transcriptional activators KLF-11 and SIRT1 levels in the hippocampus of stress-induced depressed mice. *International Journal of Pharmaceutical Investigation*, 12(3), 375–379. <https://doi.org/10.5530/IJPI.2022.3.63>
- Basile, S., Badalamenti, N., Riccobono, O., Guarino, S., Ilardi, V., Bruno, M., et al. (2022). Chemical composition and evaluation of insecticidal activity of *Calendula incana* subsp. *maritima* and *Laserpitium siler* subsp. *siculum* essential oils against stored products pests. *Molecules*, 27(3), 588. <https://doi.org/10.3390/molecules27030588>
- Basit, M. A., Kadir, A. A., Loh, T. C., Aziz, S. A., Salleh, A., Kaka, U., et al. (2020). Effects of Inclusion of different doses of persicaria odorata leaf meal (POLM) in broiler chicken feed on biochemical and haematological blood indicators and liver histomorphological changes. *Animals*, 10(7), 1209. <https://doi.org/10.3390/ani10071209>
- Basu, S., & Banik, B. K. (2020). Natural spices in medicinal chemistry: Properties and benefits. In *Green approaches in medicinal chemistry for sustainable drug design* (pp. 739–758). Elsevier. <https://doi.org/10.1016/B978-0-12-817592-7.00022-8>.
- Bayala, B., Bassole, I. H. N., Gnoula, C., Nebie, R., Yonli, A., Morel, L., et al. (2014). Chemical composition, antioxidant, anti-inflammatory and anti-proliferative activities of essential oils of plants from Burkina Faso. *PLoS One*, 9(3), Article e92122. <https://doi.org/10.1371/journal.pone.0092122>
- Begum, H. A., Hamayun, M., Tabassum East, Sumbal Akhter, & Muhammad, Shakeel. (2016). Phytochemical analysis, antifungal bioassay and folklore uses of selected medicinal plants of family Rosaceae. *Pure and Applied Biology*, 5(2), 183–192. <https://doi.org/10.19045/BSPAB.2016.50024>
- Beiranvand, M., & Bahramikia, S. (2020). Ameliorating and protective effects mesalazine on ethanol-induced gastric ulcers in experimental rats. *European Journal of Pharmacology*, 888, Article 173573. <https://doi.org/10.1016/j.ejphar.2020.173573>
- Bernal-Bayard, J., & Ramos-Morales, F. (2018). Molecular mechanisms used by salmonella to evade the immune system. *Current Issues in Molecular Biology*, 25(1), 133–168. <https://doi.org/10.21775/CIMB.025.133>
- Bernardini, S., Tiezzi, A., Laghezza Masci, V., & Ovidi, E. (2017). Natural products for human health: an historical overview of the drug discovery approaches. *Natural Product Research*, 32(16), 1926–1950. <https://doi.org/10.1080/14786419.2017.1356838>
- Bettarini, F., Borgonovi, G. E., Fiorani, T., Gagliardi, I., Caprioli, V., Massardo, P., et al. (1993). Antiparasitic compounds from East African plants: Isolation and biological activity of anonaine, matricarianol, canthin-6-one and caryophyllene oxide. *International Journal of Tropical Insect Science*, 14(1), 93–99. <https://doi.org/10.1017/S174275840001345X>
- Bharathy, V., Sumathy, B. M., & Uthayakumari, F. (2012). Determination of phytochemicals by GC-MS in leaves of *Jatropha gossypifolia* L. *Science Research Reporter*, 2(3), 286–290.
- Biazi, B. I., Zanetti, T. A., Baranoski, A., Corveloni, A. C., & Mantovani, M. S. (2017). Cisnerololol induces endoplasmic reticulum stress and cell death in human hepatocellular carcinoma cells through extensive CYP2C19 and CYP1A2 oxidation. *Basic and Clinical Pharmacology and Toxicology*, 121(4), 334–341. <https://doi.org/10.1111/bcpt.12772>
- Bonikowski, R., Świtakowska, P., & Kula, J. (2015). Synthesis, odour evaluation and antimicrobial activity of some geranyl acetone and nerolidol analogues. *Flavour and Fragrance Journal*, 30(3), 238–244. <https://doi.org/10.1002/ffj.3238>
- Borhamdin, S., Shamsuddin, M., & Alizadeh, A. (2014). Gold clusters on thiol-functionalized Fe₃O₄@SiO₂ nanoparticles: A novel bioreduced catalyst for oxidation of benzyl alcohol. In *Conference of academic libraries & national library of Malaysia*, Article 61227. <http://malrep.uum.edu.my/rep/Record/my.utm.61227/Description#tabnav>.
- Borhamdin, S., Shamsuddin, M., & Alizadeh, A. (2015). Biostabilised icosahedral gold nanoparticles: Synthesis, cyclic voltammetric studies and catalytic activity towards 4-nitrophenol reduction. *Journal of Experimental Nanoscience*, 11(7), 518–530. <https://doi.org/10.1080/17458080.2015.1090021>
- Brindha, K., & Pavelic, P. (2016). Identifying priority watersheds to mitigate flood and drought impacts by novel conjunctive water use management. *Environmental Earth Sciences*, 75(5), 1–17. <https://doi.org/10.1007/S12665-015-4989-Z>
- Britto, A. C. S., De Oliveira, A. C. A., Henriques, R. M., Cardoso, G. M. B., Bomfim, D. S., Carvalho, A. A., et al. (2012). *In vitro* and *in vivo* antitumor effects of the essential oil from the leaves of *Guatteria friesiana*. *Planta Medica*, 78(5), 409–414. <https://doi.org/10.1055/s-0031-1298173>
- Bruno, F., Castelli, G., Migliazzo, A., Piazza, M., Galante, A., Verde, V. L., et al. (2015). Cytotoxic screening and *in vitro* evaluation of pentadecane against *Leishmania infantum* promastigotes and amastigotes. *The Journal of Parasitology*, 101(6), 701–705. <https://doi.org/10.1645/15-736>
- Bueno, J., Escobar, P., Martínez, J. R., Leal, S. M., & Stashenko, E. E. (2011). Composition of three essential oils, and their mammalian cell toxicity and antimycobacterial activity against drug resistant-tuberculosis and nontuberculous mycobacteria strains. *Natural Product Communications*, 6(11), 1743–1748. <https://doi.org/10.1177/1934578x1100601143>
- Bulfin, C., Volpatti, D., & Galeotti, M. (2015). Current research on the use of plant-derived products in farmed fish. *Aquaculture Research*, 46(3), 513–551. <https://doi.org/10.1111/ARE.12238>
- Bunawan, H., Choong, C. Y., Md-Zain, B. M., Baharum, S. N., & Noor, N. M. (2011). Molecular systematics of *Polygonum minus* Huds. Based on ITS sequences. *International Journal of Molecular Sciences*, 12(11), 7626–7634. <https://doi.org/10.3390/ijms12117626>
- Bunawan, H., Talip, N., & Noor, N. M. (2011). Foliar anatomy and micromorphology of *Polygonum minus* Huds. and their taxonomic implications. *Australian Journal of Crop Science*, 5(2), 123–127.
- Burkill, I. H. (1966). *A dictionary of the economic products of the Malay peninsula*. Kuala Lumpur, Malaysia: Ministry of Agriculture and Co-operatives.
- Byass, P. (2014). The global burden of liver disease: A challenge for methods and for public health. *BMC Medicine*, 12(1), 1–3. https://doi.org/10.1186/S12916-014-0159-5/MEDIAOBJECTS/12916_2014_159_MOESM1_ESM.GIF
- Cano-Lamadrid, M., Martínez-Zamora, L., Castillejo, N., & Artés-Hernández, F. (2022). From pomegranate byproducts waste to worth: A review of extraction techniques and potential applications for their revalorization. *Foods*, 11(17), 2596. <https://doi.org/10.3390/foods11172596>
- Capon, R. J., & MacLeod, J. K. (1988). New isothiocyanate sesquiterpenes from the Australian marine sponge *acanthella pulcherrima*. *Australian Journal of Chemistry*, 41(6), 979–983. <https://doi.org/10.1071/CH9880979>
- Casillo, A., Papa, R., Ricciardelli, A., Sannino, F., Ziaco, M., Tilotta, M., et al. (2017). Anti-biofilm activity of a long-chain fatty aldehyde from antarctic *Pseudoalteromonas haloplanktis* TAC125 against *Staphylococcus epidermidis* biofilm. *Frontiers in Cellular and Infection Microbiology*, 7(46), Article 241357. <https://doi.org/10.3389/fcimb.2017.00046>
- Castilla-Guerra, L. (2022). Late-life hypertension as a risk factor for cognitive decline and dementia. *Hypertension Research*, 45(10), 1670–1671. <https://doi.org/10.1038/S41440-022-00988-Z>
- Cautela, D., Vella, F. M., & Laratta, B. (2019). The effect of processing methods on phytochemical composition in Bergamot juice. *Foods*, 8(10), 474. <https://doi.org/10.3390/foods8100474>
- Cavaleiro, C., Salgueiro, L., Gonçalves, M. J., Hrimpeng, K., Pinto, J., & Pinto, E. (2015). Antifungal activity of the essential oil of *Angelica* major against *Candida*, *Cryptococcus*, *Aspergillus* and *Dermatophytes* species. *Journal of Natural Medicines*, 69(2), 241–248. <https://doi.org/10.1007/s11418-014-0884-2>
- Chan, W. K., Tan, L. T. H., Chan, K. G., Lee, L. H., & Goh, B. H. (2016). Nerolidol: A sesquiterpene alcohol with multi-faceted pharmacological and biological activities. *Molecules*, 21(5), 529. <https://doi.org/10.3390/molecules21050529>
- Chang, S. T., Chen, P. F., Wang, S. Y., & Wu, H. H. (2001). Antimite activity of essential oils and their constituents from Taiwan cryptomerioides. *Journal of Medical Entomology*, 38(3), 455–457. <https://doi.org/10.1603/0022-2585-38.3.455>
- Chanprapai, P., Kubo, I., & Chavasiri, W. (2018). Anti-rice pathogenic microbial activity of *Persicaria* sp. extracts. *Science & Technologie Alimentaire*, 23(4), 32–41. <https://doi.org/10.14456/scitechasia.2018.30>
- Chaudhary, A., Sood, S., Das, P., Kaur, P., Mahajan, I., Gulati, A., et al. (2014). Synthesis of novel antimicrobial aryl himachalene derivatives from naturally occurring himachalenes. *EXCLI Journal*, 13, 1216–1225.
- Chen, T., Lu, H., Shen, M., Yu, Q., Chen, Y., Wen, H., et al. (2022). Phytochemical composition, antioxidant activities and immunomodulatory effects of pigment extracts from Wugong Mountain purple red rice bran. *Food Research International*, 157, Article 111493. <https://doi.org/10.1016/J.FOODRES.2022.111493>
- Cheng, M., He, J., Wang, H., Li, C., Wu, G., Zhu, K., et al. (2023). Comparison of microwave, ultrasound and ultrasound-microwave assisted solvent extraction methods on phenolic profile and antioxidant activity of extracts from jackfruit (*Artocarpus heterophyllus* Lam.) pulp. *LWT - Food Science and Technology*, 173, Article 114395. <https://doi.org/10.1016/j.lwt.2022.114395>
- Chia, S. R., Foo, S. P., Hew, Y. S., Loh, Y. J., Devadas, V. V., Chew, K. W., et al. (2020). Extraction of phenolic compounds from fresh and wilt kesum plant using liquid biphasic flotation. *Separation and Purification Technology*, 242, Article 116831. <https://doi.org/10.1016/J.SEPUR.2020.116831>
- Chilverly, S., Yelne, A., Khurana, A., Saifi, M. A., Bansod, S., Anchi, P., et al. (2023). Acetaminophen induced hepatotoxicity: An overview of the promising protective effects of natural products and herbal formulations. *Phytomedicine*, 108, Article 154510. <https://doi.org/10.1016/j.phymed.2022.154510>
- Choi, W. H., & Jiang, M. (2014). Evaluation of antibacterial activity of hexanedioic acid isolated from *Hermetia illucens* larvae. *Journal of Applied Biomedicine*, 12(3), 179–189. <https://doi.org/10.1016/j.jab.2014.01.003>
- Choi, D., Kang, W., & Park, T. (2020). Anti-allergic and anti-inflammatory effects of undecane on mast cells and keratinocytes. *Molecules*, 25(7), 1554. <https://doi.org/10.3390/molecules25071554>
- Christopher, P. V., Christina, J. M. A., Asmawi, M. Z., & Murugaiyah, V. (2015). Evaluation of antihyperlipidemic effect of *Polygonum minus*, a South East Asian salad plant in poloxamer 407-induced acute hyperlipidemic rats. *Chinese Journal of Pharmacology and Toxicology*, 29, 118.
- Christopher, P. V., Joe, L. S., Tian, M., Brij Mohan, T. S., Parasuraman, S., Resq Al-Suede, F. S., et al. (2016). Evaluation of methanol extract of *Polygonum minus* Huds. leaves for its hepatoprotective activity. *Malaysian Journal of Microbiology*, 12(5), 345–352.
- Christopher, P. V., Muthuraman, A., Zhang, L., Jordon, K. S., & Jonathan, K. H. (2021). Effect of methanol extract of *Polygonum minus* on neuropathic pain and cognitive dysfunction in rats. *International Journal of Nutrition, Pharmacology, Neurological Diseases*, 11(2), 154–162. <https://doi.org/10.4103/IJNPND.IJNPND.109.20>
- Christopher, P. V., Parasuraman, S., Asmawi, M. Z., & Murugaiyah, V. (2017). Acute and subchronic toxicity studies of methanol extract of *Polygonum minus* leaves in Sprague Dawley rats. *Regulatory Toxicology and Pharmacology*, 86, 33–41. <https://doi.org/10.1016/J.YRTPH.2017.02.005>
- Christopher, P. V., Parasuraman, S., Christina, J. M. A., Asmawi, M. Z., & Vikneswaran, M. (2015). Review on *Polygonum minus* Huds, a commonly used food additive in Southeast Asia. *Pharmacognosy Research*, 7(1), 1–6. <https://doi.org/10.4103/0974-8490.147125>
- Christopher, P. V., Parasuraman, S., Vasanth Raj, P., Saghir, S. A. M., Asmawi, M. Z., & Vikneswaran, M. (2016). Influence of extracting solvent on pharmacological activity and cytotoxicity of *Polygonum minus*, a commonly consumed herb in Southeast Asia.

- Pharmacognosy Magazine, 12(4), 424–430. <https://doi.org/10.4103/0973-1296.191451>
- Christopher, P., Vikneswaran, M., Xin, T., Yuan, G., Parasuraman, S., Leng, L., et al. (2015). Evaluation of analgesic, anti-inflammatory, antipyretic and antiulcer effect of aqueous and methanol extracts of leaves of *Polygonum minus* Huds. (Polygonaceae) in rodents. *Archives of Medicine and Health Sciences*, 3(1), 12. <https://doi.org/10.4103/2321-4848.154919>
- Ciriminna, R., Lomeli Rodriguez, M., Demma Carà, P., Lopez Sanchez, J. A., & Pagliaro, M. (2014). Limonene: A versatile chemical of the bioeconomy. *Chemical Communications*, 50(97), 15288–15296. <https://doi.org/10.1039/c4cc06147k>
- Citarasu, T. (2010). Herbal biomedicines: A new opportunity for aquaculture industry. *Aquaculture International*, 18(3), 403–414. <https://doi.org/10.1007/S10499-009-9253-7/METRICS>
- Dias, D. A., Urban, S., & Roessner, U. (2012). A historical overview of natural products in drug discovery. *Metabolites*, 2(2), 303–336. <https://doi.org/10.3390/METABO2020303>
- Díaz-Montes, E., & Castro-Muñoz, R. (2021). Edible films and coatings as food-quality preservers: An overview. *Foods*, 10(2), 249. <https://doi.org/10.3390/FOODS10020249>
- Da Silva, A. C. R., Lopes, P. M., De Azevedo, M. M. B., Costa, D. C. M., Alviano, C. S., & Alviano, D. S. (2012). Biological activities of α -pinene and β -pinene enantiomers. *Molecules*, 17(6), 6305–6316. <https://doi.org/10.3390/molecules17066317>
- Dahham, S. S., Tabana, Y. M., Iqbal, M. A., Ahamed, M. B. K., Ezzat, M. O., Majid, A. S. A., et al. (2015). The anticancer, antioxidant and antimicrobial properties of the sesquiterpene β -caryophyllene from the essential oil of *Aquilaria crassna*. *Molecules*, 20(7), 11808–11829. <https://doi.org/10.3390/molecules200711808>
- de Angelis, C., Galdiero, M., Pivonello, C., Salzano, C., Gianfrilli, D., Piscitelli, P., et al. (2017). The environment and male reproduction: The effect of cadmium exposure on reproductive functions and its implication in fertility. *Reproductive Toxicology*, 73, 105–127. <https://doi.org/10.1016/j.reprotox.2017.07.021>
- de Oliveira, D. R., da Silva, D. M., Florentino, I. F., de Brito, A. F., Fajemiroye, J. O., da Silva, D. P. B., et al. (2018). Monoamine involvement in the antidepressant-like effect of β -caryophyllene. *CNS & Neurological Disorders - Drug Targets*, 17(4), 309–320. <https://doi.org/10.2174/1871527317666180420150249>
- De Sousa, D. P., Raphael, E., Brocksom, U., & Brocksom, T. J. (2007). Sedative effect of monoterpene alcohols in mice: A preliminary screening. *Zeitschrift Fur Naturforschung - Section C Journal of Biosciences*, 62(7–8), 563–566. <https://doi.org/10.1515/znc-2007-7-816>
- de Souza, M. M. (2008). The concept of skin bleaching in Africa and its devastating health implications. *Clinics in Dermatology*, 26(1), 27–29. <https://doi.org/10.1016/j.clindermatol.2007.10.005>
- Deng, X., Shi, B., Ye, Z., Huang, M., Chen, R., Cai, Y., et al. (2022). Systematic identification of *Ocimum sanctum* sesquiterpenoid synthases and (–)-eremophilene overproduction in engineered yeast. *Metabolic Engineering*, 69, 122–133. <https://doi.org/10.1016/j.ymben.2021.11.005>
- Denver, C. V., Jackson, P., Loakes, D. M., Ellis, M. R., & Young, D. A. B. (1994). Isolation of antirhinoviral sesquiterpenes from ginger (*Zingiber Officinale*). *Journal of Natural Products*, 57(5), 658–662. <https://doi.org/10.1021/np50107a017>
- Devlin, J. R., Santus, W., Mendez, J., Peng, W., Yu, A., Wang, J., et al. (2022). *Salmonella enterica* serovar Typhimurium chitinases modulate the intestinal glycome and promote small intestinal invasion. *PLoS Pathogens*, 18(4), Article e1010167. <https://doi.org/10.1371/journal.ppat.1010167>
- Dewi, S., Assegaf, S. N., Natalia, D., & Mahyudin, M. (2019). Efek ekstrak ethanol daun kesum (*Polygonum minus* Huds.) sebagai antifungi terhadap *Trichophyton rubrum*. *Jurnal Kesehatan Andalas*, 8(2), 198. <https://doi.org/10.25077/jka.v8i2.992>
- Dhmi, N., & Mishra, A. D. (2015). Phytochemical variation: How to resolve the quality controversies of herbal medicinal products? *Journal of Herbal Medicine*, 5(2), 118–127. <https://doi.org/10.1016/J.JHERMED.2015.04.002>
- Djeddi, N., Djebile, K., Hadjbourrega, G., Achour, Z., Argyropoulou, C., & Skaltsa, H. (2012). *In vitro* antimicrobial properties and chemical composition of santolina chamaecyparissus essential oil from Algeria. *Natural Product Communications*, 7(7), 937–940. <https://doi.org/10.1177/1934578x1200700735>
- Dragomanova, S., Tancheva, L., & Georgieva, M. (2018). A review: Biological activity of myrtenal and some myrtenal-containing medicinal plant essential oils. *Scripta Scientifica Pharmaceutica*, 5(2), 22–33. <https://doi.org/10.14748/ssp.v5i2.5614>
- Dragomanova, S., Tancheva, L., Georgieva, M., & Klisurov, R. (2019). Analgesic and anti-inflammatory activity of monoterpene myrtenal in rodents. *Journal of IMAB - Annual Proceeding (Scientific Papers)*, 25(1), 2406–2413. <https://doi.org/10.5272/jimab.2019251.2406>
- Dzoyem, J. P., Kuete, V., McGaw, L. J., & Eloff, J. N. (2014). The 15-lipoxygenase inhibitory, antioxidant, antimycobacterial activity and cytotoxicity of fourteen ethnomedicinally used African spices and culinary herbs. *Journal of Ethnopharmacology*, 156, 1–8. <https://doi.org/10.1016/J.JEP.2014.08.007>
- El-Shall, N. A., Abd El-Hack, M. E., Albaqami, N. M., Khafaga, A. F., Taha, A. E., Swelum, A. A., et al. (2022). Phytochemical control of poultry coccidiosis: A review. *Poultry Science*, 101(1), Article 101542. <https://doi.org/10.1016/J.PSJ.2021.101542>
- El-Son, M. A. M., Nofal, M. I., & Abdel-Latif, H. M. R. (2021). Co-infection of *Aeromonas hydrophila* and *Vibrio parahaemolyticus* isolated from diseased farmed striped mullet (*Mugil cephalus*) in Manzala, Egypt – a case report. *Aquaculture*, 530, Article 735738. <https://doi.org/10.1016/J.AQUACULTURE.2020.735738>
- El-Toumy, S. A., Salib, J. Y., El-Kashak, W. A., Marty, C., Bedoux, G., & Bourgougnon, N. (2018). Antiviral effect of polyphenol rich plant extracts on herpes simplex virus type 1. *Food Science and Human Wellness*, 7(1), 91–101. <https://doi.org/10.1016/J.FSHW.2018.01.001>
- Elias, A., Shebawy, W. N., Nehme, B., Faour, W., Bassil, B. S., & Hakim, J. El (2019). *In vitro* and *in vivo* evaluation of the anticancer and anti-inflammatory activities of 2-himachelen-7-ol isolated from *Cedrus Libani*. *Scientific Reports*, 9(1), Article 12855. <https://doi.org/10.1038/s41598-019-49374-9>
- Elnehrawy, N. Y. F. (2015). *In vitro* study of *Polygonum minus* extract effect on skin cells healing capacity (pp. 1–44). Universiti Teknologi Malaysia, 2015.
- Embong, W. H. W. (2007). Healing herbs of Malaysia. In *Healing herbs of Malaysia* (p. 201). Kuala Lumpur: Federal Land Development Authority (FELDA), 2007.
- Entigu, R., Linton, A., Lihan, S., & Ahmad, I. (2013). The effect of combination of octadecanoic acid, methyl ester and ribavirin against measles virus. *International Journal of Scientific & Technology Research*, 2(10), 181–184.
- Erhirhie, E. O., Ihekwereme, C. P., & Iodigwe, E. E. (2018). Advances in acute toxicity testing: Strengths, weaknesses and regulatory acceptance. *Interdisciplinary Toxicology*, 11(1), 5–12. <https://doi.org/10.2478/intox-2018-0001>
- Fajar, J.P., Kusharyanti, I., & Wahdaningsih, S. (2016). Hepatoprotective activity test of ethyl acetate fraction from leaves of kesum (*Polygonum minus* Huds) on cisplatin induced Wistar strain white rats (p. 1-11). Universitas Tanjungpura Pontianak, 2016.
- Faris, A., Edder, Y., Louchachha, I., Lahcen, I. A., Azzaoui, K., Hammouti, B., et al. (2023). From himachalenes to trans-himachalol: Unveiling bioactivity through hemisynthesis and molecular docking analysis. *Scientific Reports*, 13(1), Article 17653. <https://doi.org/10.1038/s41598-023-44652-z>
- Faudale, M., Viladomat, F., Bastida, J., Poli, F., & Codina, C. (2008). Antioxidant activity and phenolic composition of wild, edible, and medicinal fennel from different Mediterranean countries. *Journal of Agricultural and Food Chemistry*, 56(6), 1912–1920. <https://doi.org/10.1021/jf073083c>
- Febriyanto, W. Y., Lestari, R. B., & Tribudi, Y. A. (2021). The effect of kesum leaf (*Polygonum minus* Huds) on growth performance of KUB chicken at starter phase. *Nutrisi Jurnal Ternak*, 4(2), 124–129. <https://doi.org/10.21776/ub.jnt.2021.004.02.8>
- Felipe Alzate-Arbeláez, A., Dorta, E., López-Alarcón, C., Cortés, F. B., & Rojano, B. A. (2019). Immobilization of andean berry (*Vaccinium meridionale*) polyphenols on nanocellulose isolated from banana residues: A natural food additive with antioxidant properties. *Food Chemistry*, 294, 503–517. <https://doi.org/10.1016/j.foodchem.2019.05.085>
- Ferreira, B. A., Silva, R. F., de Moura, F. B. R., Narduchi, C. T., Deconte, S. R., Sartorelli, P., et al. (2022). α -Zingiberene, a sesquiterpene from essential oil from leaves of *Casearia sylvestris*, suppresses inflammatory angiogenesis and stimulates collagen deposition in subcutaneous implants in mice. *Natural Product Research*, 36(22), 5858–5862. <https://doi.org/10.1080/14786419.2021.2019729>
- Fidyk, K., Fiedorowicz, A., Strzadala, L., & Szumny, A. (2016). β -caryophyllene and β -caryophyllene oxide—natural compounds of anticancer and analgesic properties. *Cancer Medicine*, 5(10), 3007–3017. <https://doi.org/10.1002/cam4.816>
- Firdhouse, M. J., & Lalitha, P. (2022). Biogenic green synthesis of gold nanoparticles and their applications – a review of promising properties. *Inorganic Chemistry Communications*, 143, Article 109800. <https://doi.org/10.1016/J.INOCHE.2022.109800>
- Flay, K. J., Hill, F. I., & Muguero, D. H. (2022). A review: *Haemonchus contortus* infection in pasture-based sheep production systems, with a focus on the pathogenesis of anaemia and changes in haematological parameters. *Animals*, 12(10), 1238. <https://doi.org/10.3390/ani12101238>
- Francomano, F., Caruso, A., Barbarossa, A., Fazio, A., Torre, C. La, Ceramella, J., et al. (2019). β -Caryophyllene: A sesquiterpene with countless biological properties. *Applied Sciences*, 9(24), 5420. <https://doi.org/10.3390/app9245420>
- Fratianni, F., Cefola, M., Pace, B., Cozzolino, R., De Giulio, B., Cozzolino, A., et al. (2017). Changes in visual quality, physiological and biochemical parameters assessed during the postharvest storage at chilling or non-chilling temperatures of three sweet basil (*Ocimum basilicum* L.) cultivars. *Food Chemistry*, 229, 752–760. <https://doi.org/10.1016/J.FOODCHEM.2017.02.137>
- Friedlein, U., Dorn-In, S., & Schwaiger, K. (2021). Antimicrobial effects of plant extracts against *Clostridium perfringens* with respect to food-relevant influencing factors. *Journal of Food Protection*, 84(10), 1809–1818. <https://doi.org/10.4315/JFP-21-139>
- Fu, X., Chen, S., Xian, S., Wu, Q., Shi, J., & Zhou, S. (2023). Dendrobium and its active ingredients: Emerging role in liver protection. *Biomedicine & Pharmacotherapy*, 157, Article 114043. <https://doi.org/10.1016/J.BIOPHA.2022.114043>
- Galina, J., Yin, G., Ardó, L., & Jeney, Z. (2009). The use of immunostimulating herbs in fish: an overview of research. *Fish Physiology and Biochemistry*, 35(4), 669–676. <https://doi.org/10.1007/S10695-009-9304-Z/METRICS>
- Gallily, R., Yekhtin, Z., & Hanus, L. O. (2018). The anti-inflammatory properties of terpenoids from *Cannabis*. *Cannabis and Cannabinoid Research*, 3(1), 282–290. <https://doi.org/10.1089/can.2018.0014>
- Gaspar-Marques, C., Simões, M. F., & Rodríguez, B. (2004). Further labdane and kaurane diterpenoids and other constituents from *Plectranthus fruticosus*. *Journal of Natural Products*, 67(4), 614–621. <https://doi.org/10.1021/np030490j>
- Gattuso, S. J. (2001). Structure and ultrastructure of the secretory glands in the genus, *Polygonum* (L.), section *Persicaria* (Polygonaceae). *Biocell: Official Journal of the Sociedades Latinoamericanas de Microscopia Electronica ... et. Al*, 25(3), 229–233.
- George, A., Chinnappan, S., Choudhary, Y., Bommur, P., & Sridhar, M. (2014). Immunomodulatory activity of an aqueous extract of *Polygonum minus* Huds on Swiss albino mice using carbon clearance assay. *Asian Pacific Journal of Tropical Disease*, 4(5), 398–400. [https://doi.org/10.1016/J.SJT.2012.1808\(14\)60594-6](https://doi.org/10.1016/J.SJT.2012.1808(14)60594-6)
- George, A., Ng, C. P., O'Callaghan, M., Jensen, G. S., & Wong, H. J. (2014). *In vitro* and *ex-vivo* cellular antioxidant protection and cognitive enhancing effects of an extract of *Polygonum minus* Huds (LinemenusTM) demonstrated in a Barnes Maze animal model for memory and learning. *BMC Complementary and Alternative Medicine*, 14, 1–10. <https://doi.org/10.1186/1472-6882-14-161>

- Gervacia, J. R., & Ratih, I. (2021). The effectiveness of kesum leaves (*Polygonum minus*) in reducing of free fatty acids in used cooking oil. *INTEK: Jurnal Penelitian*, 7(2), 167–170. <https://doi.org/10.31963/intek.v7i2.2715>
- Gleizyns, A., Zdanavičienė, E., Žilinskis, J., Eglė, D. D. S., & Juozas Žilinskis, Z. *D. D. S. (2015). *Candida albicans* importance to denture wearers. a literature review. *Baltic Dental and Maxillofacial Journal*, 17(2), 54–66.
- Goceri, E. (2019). Analysis of deep networks with residual blocks and different activation functions: Classification of skin diseases. *2019 ninth international conference on image processing theory, tools and applications (IPTA)* (pp. 1–6). <https://doi.org/10.1109/IPTA.2019.8936083>
- Goel, R. K., Kaur, D., & Pahwa, P. (2016). Assessment of anxiolytic effect of nerolidol in mice. *Indian Journal of Pharmacology*, 48(4), 450–452. <https://doi.org/10.4103/0253-7613.186188>
- Goldman, J. G., Holden, S. K., Litvan, I., McKeith, I., Stebbins, G. T., & Taylor, J. P. (2018). Evolution of diagnostic criteria and assessments for Parkinson's disease mild cognitive impairment. *Movement Disorders*, 33(4), 503–510. <https://doi.org/10.1002/MDS.27323>
- González, A. M., Tracanna, M. I., Amani, S. M., Schuff, C., Poch, M. J., Bach, H., et al. (2012). Chemical composition, antimicrobial and antioxidant properties of the volatile oil and methanol extract of *Xenophyllum poposum*. *Natural Product Communications*, 7(12), 1663–1666. <https://doi.org/10.1177/1934578x1200701230>
- Graf, M., & Stappen, I. (2022). Beyond the bark: An overview of the chemistry and biological activities of selected bark essential oils. *Molecules*, 27(21), 7295. <https://doi.org/10.3390/MOLECULES27217295>
- Greenhalgh, A. D., David, S., & Bennett, F. C. (2020). Immune cell regulation of glia during CNS injury and disease. *Nature Reviews Neuroscience*, 21(3), 139–152. <https://doi.org/10.1038/s41583-020-0263-9>
- Grice, E. A. (2014). The skin microbiome: Potential for novel diagnostic and therapeutic approaches to cutaneous disease. *Seminars in Cutaneous Medicine and Surgery*, 33(2), 98–103. <https://doi.org/10.12788/j.sder.0087>
- Grice, E. A., & Segre, J. A. (2011). The skin microbiome. *Nature Reviews Microbiology*, 9(4), 244–253. <https://doi.org/10.1038/nrmicro2537>
- Gumprecht, J., Domek, M., Lip, G. Y. H., & Shantsila, A. (2019). Invited review: Hypertension and atrial fibrillation: Epidemiology, pathophysiology, and implications for management. *Journal of Human Hypertension*, 33(12), 824–836. <https://doi.org/10.1038/S41371-019-0279-7>
- Gyrdymova, Y. V., & Rubtsova, S. A. (2022). Caryophyllene and caryophyllene oxide: A variety of chemical transformations and biological activities. *Chemical Papers*, 76(1), 1–39. <https://doi.org/10.1007/s11696-021-01865-8>
- Hadiarti, D. (2017). Shampoo of kesum (*Polygonum minus*) leaves ethanol extract as an anti-dandruff. *AIP Conference Proceedings*, 1823(1), 1–4. <https://doi.org/10.1063/1.4978075>
- Hadidi, M., Orellana-Palacios, J. C., Aghababaei, F., Gonzalez-Serrano, D. J., Moreno, A., & Lorenzo, J. M. (2022). Plant by-product antioxidants: Control of protein-lipid oxidation in meat and meat products. *LWT - Food Science and Technology*, 169, Article 114003. <https://doi.org/10.1016/j.lwt.2022.114003>
- Haenisch, B., & Bönisch, B. (2011). Depression and antidepressants: Insights from knockout of dopamine, serotonin or noradrenaline re-uptake transporters. *Pharmacology & Therapeutics*, 129(3), 352–368. <https://doi.org/10.1016/J.PHARMTHERA.2010.12.002>
- Hamid, A. A., Aminuddin, A., Yunus, M. H. M., Murthy, J. K., Hui, C. K., & Ugusman, A. (2020). Antioxidative and anti-inflammatory activities of *Polygonum minus*: A review of literature. *Reviews in Cardiovascular Medicine*, 21(2), 275–288. https://doi.org/10.31083/J.RCM.2020.02.50/2153-8174-21-2-275/IMG_4.JPG
- Han, M., Kasim, S., Yang, Z., Deng, X., Uddin, M. K., Saidi, N. B., et al. (2024). Application of *Polygonum minus* extract in enhancing drought tolerance in maize by regulating osmotic and antioxidant system. *Phyton-International Journal of Experimental Botany*, 93(2), 213–226. <https://doi.org/10.32604/PHTON.2024.047150>
- Harada, A., Sakata, K., & Ina, K. (1984). A new screening method for antifouling substances using the blue mussel, *Mytilus edulis* L. *Agricultural and Biological Chemistry*, 48(3), 641–644. <https://doi.org/10.1080/00021369.1984.10866197>
- Harikrishnan, R., Balasundaram, C., & Heo, M. S. (2011). Impact of plant products on innate and adaptive immune system of cultured finfish and shellfish. *Aquaculture*, 317(1–4), 1–15. <https://doi.org/10.1016/J.AQUACULTURE.2011.03.039>
- Haris, H. H. B., Ming, Y. K., Perin, F., Blanche, C., & Jinapong, N. (2014). Split-face placebo controlled evaluation of the *in vivo* anti-ageing efficacy of lineminus™ cream (*Polygonum minus* extract) in healthy asian skin type female subjects. *Asian Journal of Pharmaceutical and Clinical Research*, 7(3), 7–13.
- Hassan, K. Z., Noor, H. M., & Kader, J. (2015). Antibacterial efficacy of three different extracts of *Polygonum minus* (Huds.). *International conference on waste management, ecology and biological sciences* (p. 41). <https://doi.org/10.17758/ER1515231>
- Hassan Sakar, E., Zeroual, A., Mahjoubi, F., Chaouq, M., Chaqroune, A., & Taleb, M. (2021). Effects of extraction technique and solvent on phytochemicals, antioxidant, and antimicrobial activities of cultivated and wild rosemary (*Rosmarinus officinalis* L.) from taounate region (Northern Morocco). *Biointerface Research in Applied Chemistry*, 12(6), 8441–8452. <https://doi.org/10.33263/BRIAC126.84418452>
- Hassanen, N. H. M., Fahmi, A., Shams-Eldin, E., & Abdur-Rahman, M. (2020). Protective effect of rosemary (*Rosmarinus officinalis*) against diethylnitrosamine-induced renal injury in rats. *Biomarkers*, 25(3), 281–289. <https://doi.org/10.1080/1354750X.2020.1737734>
- Hassim, N., Markom, M., Anuar, N., & Baharum, S. N. (2014). Solvent selection in extraction of essential oil and bioactive compounds from *Polygonum minus*. *Journal of Applied Sciences*, 14(13), 1440–1444. <https://doi.org/10.3923/jas.2014.1440.1444>
- Hassim, N., Markom, M., Anuar, N., Dewi, K. H., Baharum, S. N., & Mohd Noor, N. (2015). Antioxidant and antibacterial assays on *Polygonum minus* extracts: Different extraction methods. *International Journal of Chemical Engineering*, 2015, 1–10. <https://doi.org/10.1155/2015/826709>
- Hayat, S., Ahmad, A., Ahmad, H., Hayat, K., Khan, M. A., & Runan, T. (2022). Garlic, from medicinal herb to possible plant bioprotectant: A review. *Scientia Horticulturae*, 304, Article 111296. <https://doi.org/10.1016/J.SCIEN.2022.111296>
- Henriques, H., Hassan, A. H. E., Sarkar, R., Dragomanova, S., Andonova, V., Volcho, K., et al. (2023). Therapeutic potential of myrtenal and its derivatives—a review. *Life*, 13(10), 2086. <https://doi.org/10.3390/LIFE13102086>
- Hernández-Rodríguez, J., Togno-Peirce, C., López De Jesús, P., Pérez-Aguirre, S. G., Arenas-Ríos, E., Viguera-Villaseñor, R. M., et al. (2016). Efecto del cadmio en la maduración espermática epididimaria. *Revista Iberoamericana de Ciencias*, 3(3), 11–21. www.reibci.org
- Hirai, Y., Asada, H., Hamada, T., Kawada, J. ichi, Kimura, H., Arai, A., et al. (2023). Diagnostic and disease severity determination criteria for hydroa vacciniforme lymphoproliferative disorders and severe mosquito bite allergy. *Journal of Dermatology*, 50(7), e198–e205. <https://doi.org/10.1111/1346-8138.16842>
- Ho, C. L., Wang, E. I. C., Hsu, K. P., Lee, P. Y., & Su, Y. C. (2009). Composition and antimicrobial activity of the leaf essential oil of *Litsea kostermansii* from Taiwan. *Natural Product Communications*, 4(8), 1123–1126. <https://doi.org/10.1177/1934578x0900400822>
- Hollingsworth, R. G. (2005). Limonene, a citrus extract, for control of mealybugs and scale insects. *Journal of Economic Entomology*, 98(3), 772–779. <https://doi.org/10.1603/0022-0493-98.3.772>
- Hu, J., Johnston, K. P., & Williams, R. O. (2004). Nanoparticle engineering processes for enhancing the dissolution rates of poorly water soluble drugs. *Drug Development and Industrial Pharmacy*, 30(3), 233–245. <https://doi.org/10.1081/DDC-120030422>
- Huang, D., Ren, J., Li, R., Guan, C., Feng, Z., Bao, B., et al. (2020). Tooth regeneration: Insights from tooth development and spatial-temporal control of bioactive drug release. *Stem Cell Reviews and Reports*, 16(1), 41–55. <https://doi.org/10.1007/S12015-019-09940-0/FIGURES/4>
- Huang, M. H., Wu, S. N., Wang, J. P., Lin, C. H., Lu, S. I., Liao, L. F., et al. (2003). Biological study of naphthalene derivatives with antiinflammatory activities. *Drug Development Research*, 60(4), 261–269. <https://doi.org/10.1002/ddr.10327>
- Hubert, T. D., Okunade, A. L., & Wiemer, D. F. (1987). Quadrangolide, a heliangolide from *Eupatorium quadrangulare*. *Phytochemistry*, 26(6), 1751–1753. [https://doi.org/10.1016/S0031-9422\(00\)82282-7](https://doi.org/10.1016/S0031-9422(00)82282-7)
- Huda-Faujan, N., Norriham, A., Norrakiah, A., & Babji, A. (2009). Antioxidant activity of plants methanolic extracts containing phenolic compounds. *African Journal of Biotechnology*, 8(3), 484–489.
- Huda-Fujan, N., Norriham, A., Norrakiah, A. S., & Babji, A. S. (2006). Antioxidant effects of *Cosmos caudatus*, *Polygonum minus* and *Murraya koenigii* in spent hen burger. *Malaysian Journal of Animal Science*, 1(1), 72–77.
- Hurabelle, C., Link, V. M., Bouladoux, N., Han, S. J., Merrill, E. D., Lightfoot, Y. L., et al. (2020). Immunity to commensal skin fungi promotes psoriasisiform skin inflammation. *Proceedings of the National Academy of Sciences*, 117(28), 16465–16474. <https://doi.org/10.1073/PNAS.2003022117>
- Husaini, N. binti, & Azman, N. A. binti M. (2020). Active packaging film for food protection. *Research Communication in Engineering Science & Technology*, 3(2019), 2019.
- Husna Abd Hamid, K., Amnin Wan Yahaya, W., Bebe Mohd Nor, N., Syahiera Ghazali, A., Kholijah Abdul Mudalip, S., Mat Zain, N., et al. (2019). Semirefined carrageenan (Src) film incorporated with α -tocopherol and persicaria minor for meat patties application. *Indonesian Journal of Chemistry*, 19(4), 1008–1018. <https://doi.org/10.22146/ijc.40884>
- Ibrahim, S. R. M., & Mohamed, G. A. (2016). Naturally occurring naphthalenes: Chemistry, biosynthesis, structural elucidation, and biological activities. *Phytochemistry Reviews*, 15, 279–295. <https://doi.org/10.1007/s11101-015-9413-5>
- Iguchi, K., Okumura, N., Usui, S., Sajiki, H., Hirota, K., & Hirano, K. (2001). Myristoleic acid, a cytotoxic component in the extract from *Serenoa repens*, induces apoptosis and necrosis in human prostatic LNCaP cells. *The Prostate*, 47(1), 59–65. <https://doi.org/10.1002/pros.1047>
- Ikhlas, B., Huda, N., & Ismail, N. (2012). Effect of cosmos caudatus, *Polygonum minus* and bht on physical properties, oxidative process, and microbiology growth of quail meatball during refrigeration storages. *Journal of Food Processing and Preservation*, 36(1), 55–66. <https://doi.org/10.1111/J.1745-4549.2011.00552.X>
- Imelda, F., Faridah, D. N., & Kusumaningrum, H. D. (2014). Bacterial inhibition and cell leakage by extract of *Polygonum minus* Huds. leaves. *International Food Research Journal*, 21(2), 553–560.
- Imelda, F., Kusumaningrum, H. D., & Faridah, D. N. (2013). Detection of antibacterial compound from kesum leaves (*Polygonum minus* Huds.) by TLC-bioautography method and the effect to *escherichia coli* and *staphylococcus aureus* membrane. Bogor Agricultural University]. <http://repository.ipb.ac.id/handle/123456789/67409>
- Iornumbe, E. N., Yiase, S. G., & Ato, R. S. (2016). Studies on the synthetic and biological activity of some organotin (iv) derivatives of hexanedioic acid. *IOSR Journal of Applied Chemistry*, 9(7), 23–32. <https://doi.org/10.9790/5736-0907012332>
- Islam, M. T., Ali, E. S., Uddin, S. J., Shaw, S., Islam, M. A., Ahmed, M. I., et al. (2018). Phytol: A review of biomedical activities. *Food and Chemical Toxicology*, 121, 82–94. <https://doi.org/10.1016/j.fct.2018.08.032>
- Jain, P. K., Huang, X., El-Sayed, I. H., & El-Sayed, M. A. (2008). Noble metals on the nanoscale: Optical and photothermal properties and some applications in imaging, sensing, biology, and medicine. *Accounts of Chemical Research*, 41(12), 1578–1586. https://doi.org/10.1021/AR700280A/ASSET/IMAGES/MEDIUM/AR-2007-002084_0001.GIF

- James, S. L., Abate, D., Abate, K. H., Abay, S. M., Abbafati, C., Abbasi, N., et al. (2018). Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: A systematic analysis for the global burden of disease study 2017. *The Lancet*, 392(10159), 1789–1858. [https://doi.org/10.1016/S0140-6736\(18\)32279-7](https://doi.org/10.1016/S0140-6736(18)32279-7)
- Jansen, B. J. M., & De Groot, A. (2004). Occurrence, biological activity and synthesis of drimane sesquiterpenoids. *Natural Product Reports*, 21(4), 449–477. <https://doi.org/10.1039/b311170a>
- Jarukamjorn, K., & Nemoto, N. (2008). Pharmacological aspects of *Andrographis paniculata* on health and its major diterpenoid constituent andrographolide. *Journal of Health Science*, 54(4), 370–381. <https://doi.org/10.1248/jhs.54.370>
- Javed, H., Azimullah, S., Abul Khair, S. B., Ojha, S., & Haque, M. E. (2016). Neuroprotective effect of nerolidol against neuroinflammation and oxidative stress induced by rotenone. *BMC Neuroscience*, 17(1), 1–12. <https://doi.org/10.1186/s12868-016-0293-4>
- Javed, H., Azimullah, S., Haque, M. E., & Ojha, S. K. (2016). Cannabinoid type 2 (CB2) receptors activation protects against oxidative stress and neuroinflammation associated dopaminergic neurodegeneration in rotenone model of Parkinson's disease. *Frontiers in Neuroscience*, 10, 1–14. <https://doi.org/10.3389/fnins.2016.00321>
- Jin, J. H., Lee, D. U., Kim, Y. S., & Kim, H. P. (2011). Anti-allergic activity of sesquiterpenes from the rhizomes of *Cyperus rotundus*. *Archives of Pharmacological Research*, 34(2), 223–228. <https://doi.org/10.1007/s12272-011-0207-z>
- Johnny, L., Yusuf, U. K., & Nulit, R. (2011). Antifungal activity of selected plant leaves crude extracts against a pepper anthracnose fungus, *Colletotrichum capsici* (Sydow) butler and bisby (Ascomycota: Phyllachorales). *African Journal of Biotechnology*, 10(20), 4157–4165. <https://doi.org/10.5897/AJB10.2085>
- Joshi, A., Sunil Krishnan, G., & Kaushik, V. (2020). Molecular docking and simulation investigation: Effect of beta-sesquiphellandrene with ionic integration on SARS-CoV2 and SFTS viruses. *Journal of Genetic Engineering and Biotechnology*, 18(1), 1–8. <https://doi.org/10.1186/s43141-020-00095-x>
- Jou, Y. J., Hua, C. H., Lin, C. S., Wang, C. Y., Wan, L., Lin, Y. J., et al. (2016). Anticancer activity of γ -bisabolene in human neuroblastoma cells via induction of p53-mediated mitochondrial apoptosis. *Molecules*, 21(5), 601. <https://doi.org/10.3390/molecules21050601>
- Jovanovic, J., Ornelis, V. F. M., Madder, A., & Rajkovic, A. (2021). Bacillus cereus food intoxication and toxicoinfection. *Comprehensive Reviews in Food Science and Food Safety*, 20(4), 3719–3761. <https://doi.org/10.1111/1541-4337.12785>
- Köhler, J. R., Casadevall, A., & Perfect, J. (2015). The spectrum of fungi that infects humans. *Cold Spring Harbor Perspectives in Medicine*, 5(1), a019273. <https://doi.org/10.1101/CSHPERSPECT.A019273>
- Kahramanoğlu, I., Chen, C., Chen, J., & Wan, C. (2019). Chemical constituents, antimicrobial activity and food preservative characteristics of aloe vera gel. *Agronomy*, 9(12), 831. <https://doi.org/10.3390/agronomy9120831>
- Kamaly, N., Yameen, B., Wu, J., & Farokhzad, O. C. (2016). Degradable controlled-release polymers and polymeric nanoparticles: Mechanisms of controlling drug release. *Chemical Reviews*, 116(4), 2602–2663. https://doi.org/10.1021/ACS.CHEMREV.5B00346/ASSET/IMAGES/ACS.CHEMREV.5B00346.SOCIAL.JPEG_V03
- Kamarudin, N. S., Jusoh, R., Sukor, N. F., Jalil, A. A., Setiabudi, H. D., & Salleh, N. F. M. (2022). Facile electro-assisted green synthesis of size-tunable silver nanoparticles and its photodegradation activity. *Journal of Cluster Science*, 33(3), 985–997. <https://doi.org/10.1007/s10876-021-02028-1/METRICS>
- Kang, M. C., Lee, J. W., Lee, T. H., Subedi, L., Wahedi, H. M., Do, S. G., et al. (2020). UP256 inhibits hyperpigmentation by tyrosinase expression/dendrite formation via rho-dependent signaling and by primary cilium formation in melanocytes. *International Journal of Molecular Sciences*, 21(15), 5341. <https://doi.org/10.3390/IJMS21155341>
- Kang, A., Xie, T., Zhu, D., Shan, J., Di, L., & Zheng, X. (2017). Suppressive effect of ginsenoside rg3 against lipopolysaccharide-induced depression-like behavior and neuroinflammation in mice. *Journal of Agricultural and Food Chemistry*, 65(32), 6861–6869. <https://doi.org/10.1021/ACS.JAFC.7B02386>
- Karr, S. (2017). Epidemiology and management of hyperlipidemia. *American Journal of Managed Care*, 23(9), S139–S148.
- Kartikasari, D., Rahman, I. R., Puspasari, H., & Ridha, A. (2022). The spectrometric quantification of total content of flavonoid, phenol, and alkaloid in kesum leaf (*Polygonum minus* Huds) from West Borneo with methanol and ethanol solvent. *Majalah Obat Tradisional*, 27(1), 1–6. <https://doi.org/10.22146/mot.68497>
- Kaya, M., Ravikumar, P., Ilk, S., Mujtaba, M., Akyuz, L., Labidi, J., et al. (2018). Production and characterization of chitosan based edible films from Berberis crataegina's fruit extract and seed oil. *Innovative Food Science & Emerging Technologies*, 45, 287–297. <https://doi.org/10.1016/j.ifset.2017.11.013>
- Kazemi, M. (2014). Phytochemical composition, antioxidant, anti-inflammatory and antimicrobial activity of *Nigella sativa* L. essential oil. *Journal of Essential Oil-Bearing Plants*, 17(5), 1002–1011. <https://doi.org/10.1080/0972060X.2014.914857>
- Khalid, N. M., & Salam, A. B. (2018). Antioxidative and antihypertensive activities of selected Malaysian ulam (salad), vegetables and herbs. *Journal of Food Research*, 7(3), 27–37. <https://doi.org/10.5539/jfr.v7n3p27>
- Khan, M. A. H., Liu, J., Kumar, G., Skapek, S. X., Falck, J. R., & Imig, J. D. (2013). Novel orally active epoxyeicosatrienoic acid (EET) analogs attenuate cisplatin nephrotoxicity. *Federation of American Societies for Experimental Biology Journal*, 27(8), 2946–2956. <https://doi.org/10.1096/FJ.12-218040>
- Khmaladze, I., Leonardi, M., Fabre, S., Messara, C., & Mavon, A. (2020). The skin interactome: A holistic “genome-microbiome-exposome” approach to understand and modulate skin health and aging. *Clinical, Cosmetic and Investigational Dermatology*, 13, 1021–1040. <https://doi.org/10.2147/CCID.S239367>
- Khumalo, G. P., Van Wyk, Feng, Y., & Cock, I. E. (2022). A review of the traditional use of southern African medicinal plants for the treatment of inflammation and inflammatory pain. *Journal of Ethnopharmacology*, 283, 114436. <https://doi.org/10.1016/J.JEP.2021.114436>
- Kim, D. H., Park, M. H., Choi, Y. J., Chung, K. W., Park, C. H., Jang, E. J., et al. (2013). Molecular study of dietary heptadecane for the anti-inflammatory modulation of NF- κ B in the aged kidney. *PLoS One*, 8(3), Article e59316. <https://doi.org/10.1371/journal.pone.0059316>
- Kim, T. H., Yoo, S. D., Lee, H. S., Lee, K. M., Seok, S. H., Kim, M. G., et al. (2015). *In vivo* absorption and disposition of α -cedrene, a sesquiterpene constituent of cedarwood oil, in female and male rats. *Drug Metabolism and Pharmacokinetics*, 30(2), 168–173. <https://doi.org/10.1016/j.dmpk.2014.12.003>
- Knecht, D., Cholewińska, P., Jankowska-Mąkosa, A., & Czyż, K. (2020). Development of swine's digestive tract microbiota and its relation to production indices—a review. *Animals*, 10(3), 527. <https://doi.org/10.3390/ani10030527>
- Kola, V., & Carvalho, I. S. (2023). Plant extracts as additives in biodegradable films and coatings in active food packaging. *Food Bioscience*, 54, Article 102860. <https://doi.org/10.1016/j.fbio.2023.102860>
- Konfo, T. R. C., Djouhou, F. M. C., Koudoro, Y. A., Dahouenon-Ahoussi, E., Avlessi, F., Sohounhloue, C. K. D., et al. (2023). Essential oils as natural antioxidants for the control of food preservation. *Food Chemistry Advances*, 2, Article 100312. <https://doi.org/10.1016/j.focha.2023.100312>
- Kong, I., Heng, Z. W., & Pui, L. P. (2022). Development of chitosan edible film incorporated with curry leaf and kesum for the packaging of chicken breast meat. *Asia-Pacific Journal of Molecular Biology and Biotechnology*, 30(3), 91–104. <https://doi.org/10.35118/apjmbb.2022.030.3.08>
- Krishnan, K., Mani, A., & Jasmine, S. (2014). Cytotoxic activity of bioactive compound 1, 2-benzene dicarboxylic acid, mono 2- ethylhexyl ester extracted from a marine derived *Streptomyces* sp. VITSJK8. *International Journal of Molecular and Cellular Medicine*, 3(4), 246.
- Kumalasari, E., Rima Setyawati, T., & Hepi Yanti, A. (2015). Daya tolak ekstrak metanol daun kesum (*Polygonum minus* Huds.) terhadap lalat rumah (*Musca domestica* L.). *Jurnal Protobiont*, 4(2), 40–47. <https://doi.org/10.26418/PROTOBIONT.V4I2.10867>
- Kundu, A., Saha, S., Walia, S., Ahluwalia, V., & Kaur, C. (2013). Antioxidant potential of essential oil and cadinene sesquiterpenes of *Eupatorium adenophorum*. *Toxicological and Environmental Chemistry*, 95(1), 127–137. <https://doi.org/10.1080/02772248.2012.759577>
- Kusumaningrum, P. A., Yustinasari, L. R., Hamid, I. S., Sudjarwo, S. A., Santoso, K. P., & Anwar, C. (2020). Effect of *Polygonum minus* (kesum) leaves ethanolic extract on histopathological changes on the wall aorta of mice (*Mus musculus*) induced by cadmium chloride antioxidant. *Journal of Basic Medical Veterinary*, 8(2), 66. <https://doi.org/10.20473/v8i2.20408>
- Kusumaningrum, P. A., Yustinasari, L. R., Hamid, I. S., Sudjarwo, S. A., Santoso, K. P., Anwar, C., et al. (2019). Protective effect of *Polygonum minus* leaves ethanol extract on cadmium chloride-induced alteration of aortic histopathology in mice (*Mus musculus*). *Indian Veterinary Journal*, 96(7), 39–42.
- Langenheim, J. H. (1994). Higher plant terpenoids: A phyto-centric overview of their ecological roles. *Journal of Chemical Ecology*, 20(6), 1223–1280. <https://doi.org/10.1007/BF02059809>
- Larouche, J., Sheoran, S., Maruyama, K., & Martino, M. M. (2018). Immune regulation of skin wound healing: Mechanisms and novel therapeutic targets. *Advances in Wound Care*, 7(7), 209–231. <https://doi.org/10.1089/WOUND.2017.0761/ASSET/IMAGES/LARGE/FIGURE6.JPEG>
- Lau, H., Shahar, S., Mohamad, M., Rajab, N. F., Yahya, H. M., Din, N. C., et al. (2020). The effects of six months persicaria minor extract supplement among older adults with mild cognitive impairment: A double-blinded, randomized, and placebo-controlled trial. *BMC Complementary Medicine and Therapies*, 20(1), 1–15. <https://doi.org/10.1186/s12906-020-03092-2>
- Lee, S. J., Han, J. I., Lee, G. S., Park, M. J., Choi, I. G., Na, K. J., et al. (2007). Antifungal effect of eugenol and nerolidol against *Microrhizum gypseum* in a Guinea pig model. *Biological and Pharmaceutical Bulletin*, 30(1), 184–188. <https://doi.org/10.1248/bpb.30.184>
- Legault, J., & Pichette, A. (2010). Potentiating effect of β -caryophyllene on anticancer activity of α -humulene, isocaryophyllene and paclitaxel. *Journal of Pharmacy and Pharmacology*, 59(12), 1643–1647. <https://doi.org/10.1211/jpp.59.12.0005>
- Lestari, R. B., Hartanti, L., & Permadi, E. (2020). Effects of kesum leaf extract supplementation on characteristics of durian seeds starch (*Durio Zibethinus*)-chitosan edible film. *Scientific Study & Research. Chemistry & Chemical Engineering, Biotechnology, Food Industry*, 21(4), 473–482.
- Lestari, R. B., Permadi, E., & Harahap, R. P. (2020). Decrease quality during storage packaged beef sausage edible coating by durian seeds starch-chitosan with the addition of kesum leaf extract. *IOP Conference Series: Earth and Environmental Science*, 478(1), Article 012036. <https://doi.org/10.1088/1755-1315/478/1/012036>
- Lestari, R. B., Permadi, E., & Mulyadi, A. (2022). Application of edible coating of durian seed starch composites with kesum leaves extracts on microbiological quality and TVB-N of beef sausage. *Jurnal Ilmu Dan Teknologi Hasil Ternak*, 17(1), 56–63. <https://doi.org/10.21776/ub.jitek.2022.017.01.7>
- Li, Y. H., Sun, X. P., Zhang, Y. Q., & Wang, N. S. (2008). The antithrombotic effect of borneol related to its anticoagulant property. *American Journal of Chinese Medicine*, 36(4), 719–727. <https://doi.org/10.1142/S0192415X08006181>
- Liang, J., Cui, L., Li, J., Guan, S., Zhang, K., & Li, J. (2021). Aloe vera: A medicinal plant used in skin wound healing. *Tissue Engineering Part B Reviews*, 27(5), 455–474. <https://doi.org/10.1089/ten.teb.2020.0236>
- Liao, L., Tang, Y., Li, B., Tang, J., Xu, H., Zhao, K., & Zhang, X. (2023). Stachydrine, a potential drug for the treatment of cardiovascular system and central nervous system

- diseases. *Biomedicine & Pharmacotherapy*, 161, 114489. <https://doi.org/10.1016/J.BIOPHA.2023.114489>
- Liauw, J. J., Assegaf, S. N., & Natalia, D. (2021). The antifungal effect of kesum leaf (*Polygonum minus* Huds) in ethanol extract on *Microsporum gypseum*. *Sains Medika*, 11(1), 26–33.
- Lin, W. Y., Kuo, Y. H., Chang, Y. L., Teng, C. M., Wang, E. C., Ishikawa, T., et al. (2003). Anti-platelet aggregation and chemical constituents from the rhizome of *Gynura japonica*. *Planta Medica*, 69(8), 757–764. <https://doi.org/10.1055/s-2003-42796>
- Liu, K., Chen, Q., Liu, Y., Zhou, X., & Wang, X. (2012). Isolation and biological activities of decanal, linalool, valencene, and octanal from sweet orange oil. *Journal of Food Science*, 77(11), C1156–C1161. <https://doi.org/10.1111/j.1750-3841.2012.02924.x>
- Liu, J., Chen, C., Wan, X., Yao, G., Bao, S., Wang, F., et al. (2022). Identification of the sesquiterpene synthase AcTPS1 and high production of (–)-germacrene D in metabolically engineered *Saccharomyces cerevisiae*. *Microbial Cell Factories*, 21(1), 89. <https://doi.org/10.1186/s12934-022-01814-4>
- Luo, W., Du, Z., Zheng, Y., Liang, X., Huang, G., Zhang, Q., et al. (2019). Phytochemical composition and bioactivities of essential oils from six *Lamiaceae* species. *Industrial Crops and Products*, 133, 357–364. <https://doi.org/10.1016/j.indcrop.2019.03.025>
- Luttmann, K., Starnes, V. R., Haddad, M., & Duggan, J. (2022). *Serratia marcescens*, a rare and devastating cause of endocarditis: A case report and review of the literature. *Cureus*, 14(6), Article e25572. <https://doi.org/10.7759/cureus.25572>
- Ma, G., Chai, X., Hou, G., Zhao, F., & Meng, Q. (2022). Phytochemistry, bioactivities and future prospects of mulberry leaves: A review. *Food Chemistry*, 372, Article 131335. <https://doi.org/10.1016/J.FOODCHEM.2021.131335>
- Machado, K. da C., Paz, M. F. C. J., Oliveira Santos, J. V. de, da Silva, F. C. C., Tchekalarova, J. D., Salehi, B., et al. (2020). Anxiety therapeutic interventions of β -caryophyllene: A laboratory-based study. *Natural Product Communications*, 15(10), 1–15. <https://doi.org/10.1177/1934578X20962229>
- Magalhães, L. A. M. I., Da Paz Lima, M., Marques, M. O. M., Facanali, R., Da Silva Pinto, A. C., & Tadei, W. P. (2010). Chemical composition and larvicidal activity against *Aedes aegypti* larvae of essential oils from four *Guarea* species. *Molecules*, 15(8), 5734–5741. <https://doi.org/10.3390/molecules15085734>
- Mahanom, H., Azizah, A. H., & Dzulkifly, M. H. (1999). Effect of different drying methods on concentrations of several phytochemicals in herbal preparation of 8 medicinal plants leaves. *Malaysian Journal of Nutrition*, 5(1–2), 47–54.
- Maizura, M., Aminah, A., & Wan Aida, W. M. (2016). Antioxidant capacity and consumer acceptability of herbal egg tofu. *LWT - Food Science and Technology*, 65, 549–556. <https://doi.org/10.1016/J.LWT.2015.07.062>
- Malaysia Biodiversity Information System (MyBIS) 2024. Ministry of Natural Resources and Environment Sustainability, Malaysia Biodiversity Centre & Forest Research Institute Malaysia. Internet: <https://www.mybis.gov.my/sp/28319>. (accessed June 8, 2024).
- Mancini, E., De Martino, L., Malova, H., & De Feo, V. (2013). Chemical composition and biological activities of the essential oil from *Calamintha nepeta* plants from the wild in southern Italy. *Natural Product Communications*, 8(1), 139–142. <https://doi.org/10.1177/1934578x1300800134>
- Manontri, T., Joe, L. S., Tanusha, S. B. M., Parasuraman, S., & Christopher, P. V. (2016). Hepatoprotective effect of methanol extract of *Polygonum minus* leaves in carbon tetrachloride-induced liver damage in rats. In *The 3rd regional conference on biosensors, biodiagnostics, biochips and biotechnology* (Vol. 45).
- Manzoor, A., Yousuf, B., Ahmad Pandith, J., & Ahmad, S. (2023). Plant-derived active substances incorporated as antioxidant, antibacterial or antifungal components in coatings/films for food packaging applications. *Food Bioscience*, 53, Article 102717.
- Mao, Z. F., Ouyang, S. H., Zhang, Q. Y., Wu, Y. P., Wang, G. E., Tu, L. F., et al. (2020). New insights into the effects of caffeine on adult hippocampal neurogenesis in stressed mice: Inhibition of CORT-induced microglia activation. *Federation of American Societies for Experimental Biology Journal*, 34(8), 10998–11014. <https://doi.org/10.1096/FJ.202000146RR>
- Markom, M., Hassim, N., Anuar, N., & Baharum, S. N. (2012). Co-solvent selection for supercritical fluid extraction of essential oil and bioactive compounds from *Polygonum minus*. *ASEAN Journal of Chemical Engineering*, 12(2), 19–26. <https://doi.org/10.22146/ajche.49739>
- Marques, R. C. P., De Medeiros, S. R. B., Da Silva Dias, C., Barbosa-Filho, J. M., & Agnez-Lima, L. F. (2003). Evaluation of the mutagenic potential of yangambin and of the hydroalcoholic extract of *Ocotea duckei* by the Ames test. *Mutation Research: Genetic Toxicology and Environmental Mutagenesis*, 536(2), 117–120. [https://doi.org/10.1016/S1383-5718\(03\)00040-8](https://doi.org/10.1016/S1383-5718(03)00040-8)
- Mataruco, L. de S., Henrique Maldonado da Silva, L., Stevanato, N., da Silva, C., Rodrigo Fink, J., Cardozo Filho, L., et al. (2023). Pressurized n-propane extraction improves bioactive compounds content, fatty acid profile, and biological activity of Mandacaru (*Cereus jamaicaru* DC.) seed oil. *Industrial Crops and Products*, 195, Article 116367. <https://doi.org/10.1016/j.indcrop.2023.116367>
- Matsuo, A., Atsumi, K., Nakayama, M., & Hayashi, S. (1981). Structures of ent-2,3-secoalloaromadendrane sesquiterpenoids, which have plant-growth-inhibitory activity, from *Plagiochila semidecurrens* (liverwort). *Journal of the Chemical Society, Perkin Transactions*, 1, 2816–2824. <https://doi.org/10.1039/P19810002816>
- Mauliyda, D. A., Kahtan, M. I., Natalia, D., Handini, M., & Vidiyantoro, A. (2018). Anthelmintic activity of ethanol extract of *Polygonum minus* leaves against *Ascaridia galli*. *Journal of Microbiology and Infectious Diseases*, 8, 23–26. <https://doi.org/10.5799/jmid.403142>
- McGinty, D., Letizia, C. S., & Api, A. M. (2010). Fragrance material review on 2-hexadecan-1-ol, 3,7,11,15-tetramethyl. *Food and Chemical Toxicology*, 48(3), S101–S102. <https://doi.org/10.1016/j.fct.2009.11.023>
- Melinda, T., Assegaf, S. N., Mahyarudin, & Natalia, D. (2019). Aktivitas anti jamur ekstrak ethanol daun kesum (*Polygonum minus* Huds) terhadap jamur *Trichophyton mentagrophytes*. *Majalah Kedokteran Andalas*, 42(3S), 48–56. <http://jurnalmlka.fk.una.ac.id/index.php/art/article/view/616>
- Melnyk, J. P., & Marcone, M. F. (2011). Aphrodisiacs from plant and animal sources—A review of current scientific literature. *Food Research International*, 44(4), 840–850. <https://doi.org/10.1016/J.FOODRES.2011.02.043>
- Melo-Reis, P. R., Bezerra, L. S. A., Vale, M. A. B., Canhête, R. F. R., & Chen-Chen, L. (2011). Assessment of the mutagenic and antimutagenic activity of *Synadenium umbellatum* Pax latex by micronucleus test in mice. *Brazilian Journal of Biology*, 71(1), 169–174. <https://doi.org/10.1590/S1519-69842011000100024>
- Messer, A., McCormick, K., Sunjaya, Hagedorn, H. H., Tumbel, F., & Meinwald, J. (1990). Defensive role of tropical tree resins: Antitermitic sesquiterpenes from Southeast Asian *dipterocarpaceae*. *Journal of Chemical Ecology*, 16(12), 3333–3352. <https://doi.org/10.1007/BF00982102>
- Michael, Indri Kusharyanti, & Isnindar (2013). Effect of kesum leaves (*Polygonum minus* huds.) methanol extract on increase of creatinine and urea concentration on cisplatin induced Wistar strain white rats (p. 1–8). Universitas Tanjungpura. 2013.
- Ming, Y., Zulkawi, N. B. T., Cy, V. K., & Choudhary, Y. K. (2013). Acute and sub-acute oral toxicity of *Polygonum minus* aqueous extract (biotropics®PM101) in Wistar rats. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(2), 120–124.
- Moghimi, S. M., Hunter, A. C., & Murray, J. C. (2001). Long-circulating and target-specific nanoparticles: Theory to practice. *Pharmacological Reviews*, 53(2), 283–318. [https://doi.org/10.1016/0165-6147\(94\)90314-x](https://doi.org/10.1016/0165-6147(94)90314-x)
- Mohamad, N., Mazlan, M. M., Tawakkal, I. S. M. A., Talib, R. A., Kian, L. K., Fouad, H., et al. (2020). Development of active agents filled polylactic acid films for food packaging application. *International Journal of Biological Macromolecules*, 163, 1451–1457. <https://doi.org/10.1016/j.ijbiomac.2020.07.209>
- Mohamad, N., Mazlan, M. M., Tawakkal, I. S. M. A., Talib, R. A., Kian, L. K., & Jawaid, M. (2022). Characterization of active polybutylene succinate films filled essential oils for food packaging application. *Journal of Polymers and the Environment*, 30(2), 585–596. <https://doi.org/10.1007/s10924-021-02198-z>
- Mohamed, S. A. A., El-Sakhawy, M., & El-Sakhawy, M. A. M. (2020). Polysaccharides, protein and lipid based natural edible films in food packaging: A review. *Carbohydrate Polymers*, 238, Article 116178. <https://doi.org/10.1016/j.carbpol.2020.116178>
- Mohd Ghazali, M. A., Al-Naqeb, G., Krishnan Selvarajan, K., Hazizul Hasan, M., & Adam, A. (2014). Apoptosis induction by *Polygonum minus* is related to antioxidant capacity, alterations in expression of apoptotic-related genes, and S-phase cell cycle arrest in HepG2 cell line. *BioMed Research International*, 2014, Article 539607. <https://doi.org/10.1155/2014/539607>
- Moiteiro, C., Esteves, T., Ramalho, L., Rojas, R., Alvarez, S., Zacchino, S., et al. (2013). Essential oil characterization of two Azorean *Cryptomeria japonica* populations and their biological evaluations. *Natural Product Communications*, 8(12), 1785–1790. <https://doi.org/10.1177/1934578x1300801233>
- Moreira, I. C., Lago, J. H. G., Young, M. C. M., & Roque, N. F. (2003). Antifungal aromadendrane sesquiterpenoids from the leaves of *Xylopia brasiliensis*. *Journal of the Brazilian Chemical Society*, 14(5), 828–831. <https://doi.org/10.1590/S0103-50532003000500020>
- Muhammad, N., Mansor, E., Sang, Y. C., Shariffudin, N. S., Dahdi, A., Alias, A. F., et al. (2013). Acute and subacute toxicity of *Persicaria minor* in Wistar rats. *The Asian Journal of Animal Science*, 7(2), 47–55. <https://doi.org/10.3923/ajas.2013.47.55>
- Muhammad, Z., & Mustafa, A. M. (2015). Traditional Malay medicinal plants. In *Itbm* (pp. 39–41). Institut Terjemahan Negara Malaysia.
- Mujeeb, F., Bajpai, P., & Pathak, N. (2014). Phytochemical evaluation, antimicrobial activity, and determination of bioactive components from leaves of *Aegle marmelos*. *BioMed Research International*, 2014, 1–11. <https://doi.org/10.1155/2014/497606>
- Mukherjee, P. K., Maity, N., Nema, N. K., & Sarkar, B. K. (2011). Bioactive compounds from natural resources against skin aging. *Phytomedicine*, 19(1), 64–73. <https://doi.org/10.1016/j.phymed.2011.10.003>
- Murakami, A., Ali, A. M., Mat-Salleh, K., Koshimizu, K., & Ohigashi, H. (2000). Screening for the *in vitro* anti-tumor-promoting activities of edible plants from Malaysia. *Bioscience Biotechnology and Biochemistry*, 64(1), 9–16. <https://doi.org/10.1271/bbb.64.9>
- Murata, M., Yamakoshi, Y., Homma, S., Hori, K., Ohashi, Y., & Aida, K. (1990). Macrocarpal A, a novel antibacterial compound from *Eucalyptus macrocarpa*. *Agricultural and Biological Chemistry*, 54(12), 3221–3226. <https://doi.org/10.1271/bbb1961.54.3221>
- Murphy, C. J., Gole, A. M., Stone, J. W., Sisco, P. N., Alkhalil, A. M., Goldsmith, E. C., et al. (2008). Gold nanoparticles in biology: Beyond toxicity to cellular imaging. *Accounts of Chemical Research*, 41(12), 1721–1730. https://doi.org/10.1021/AR800035U/ASSET/IMAGES/MEDIUM/AR-2008-00035U_0001.GIF
- Mussin, J., Robles-Botero, V., Casañas-Pimentel, R., Rojas, F., Angiolella, L., San Martín-Martínez, E., et al. (2021). Antimicrobial and cytotoxic activity of green synthesis silver nanoparticles targeting skin and soft tissue infectious agents. *Scientific Reports*, 11(1), Article 14566. <https://doi.org/10.1038/s41598-021-94012-y>
- Muthukumarasamy, R., Asmaq, N., Mahasan, B., Nabilah, F., Fuad, B. M., Rosli, N. B., et al. (2018). Formulation and evaluation of antioxidant cream from methanol leaves extract of *Polygonum minus*. *Indo American Journal of Pharmaceutical Sciences*, 5(4), 2244–2251. <https://doi.org/10.5281/zenodo.1216048>
- Nabavi, S. M., Daglia, M., Braid, N., & Nabavi, S. F. (2017). Natural products, micronutrients, and nutraceuticals for the treatment of depression: A short review. *Nutritional Neuroscience*, 20(3), 180–194. <https://doi.org/10.1080/1028415X.2015.1103461>
- Nadzirah, A. S., Mahamudin, M., Muhammad, S., & Latif Ibrahim, A. (2019). Green synthesis of silver nanoparticles from *Polygonum minus* extract and its antimicrobial properties. *Journal of Academia*, 7(2), 46–55.

- Nadzirah, A. S., Rusop, M., & Latif, I. A. (2015a). Effect of nano size powder of *Polygonum minus* by ball milling. *Advanced Materials Research*, 1109, 333–339. <https://doi.org/10.4028/www.scientific.net/amr.1109.333>.
- Nadzirah, A. S., Rusop, M., & Latif, I. A. (2015b). Surface morphology of herbal prepared using nanotechnology. *Advanced Materials Research*, 1109, 328–332. <https://doi.org/10.4028/www.scientific.net/amr.1109.328>.
- Nadzirah, A. S., Rusop, M., & Noriham, A. (2014). A review of potential of antioxidant properties using *Polygonum minus*. *Advanced Materials Research*, 832, 659–664. <https://doi.org/10.4028/www.scientific.net/AMR.832.659>.
- Nahhas, A. F., Abdel-Malek, Z. A., Kohli, I., Braunberger, T. L., Lim, H. W., & Hamzavi, I. H. (2019). The potential role of antioxidants in mitigating skin hyperpigmentation resulting from ultraviolet and visible light-induced oxidative stress. *Photodermatology, Photoimmunology & Photomedicine*, 35(6), 420–428. <https://doi.org/10.1111/PHPP.12423>
- Nakakaawa, L., Gbala, I. D., Cheseto, X., Bargul, J. L., & Wesonga, J. M. (2023). Oral acute, sub-acute toxicity and phytochemical profile of *Brassica carinata* A. Braun microgreens ethanolic extract in Wistar rats. *Journal of Ethnopharmacology*, 305, Article 116121. <https://doi.org/10.1016/j.jep.2022.116121>
- Namata Abba, B., Ilagouma, A. T., Amadou, I., & Romane, A. (2021). Chemical profiling, antioxidant and antibacterial activities of essential oil from *Englerastrum gracillimum* Th. C. E. Fries growing in Niger. *Natural Product Communications*, 16(3), 1–7. <https://doi.org/10.1177/1934578X211002422>
- Narasimhulu, G., & Mohamed, J. (2014). Medicinal phytochemical and pharmacological properties of kesum (*Polygonum minus* Linn.): A mini review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(4), 682–688.
- Narasimhulu, G., Reddy, K. K., & Mohamed, J. (2014). The genus *Polygonum* (*Polygonaceae*): An ethnopharmacological and phytochemical perspectives - review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(2), 21–45.
- Newaj-Fyzul, A., & Austin, B. (2015). Probiotics, immunostimulants, plant products and oral vaccines, and their role as feed supplements in the control of bacterial fish diseases. *Journal of Fish Diseases*, 38(11), 937–955. <https://doi.org/10.1111/JFD.12313>
- Nibret, E., & Wink, M. (2010). Trypanocidal and antileukaemic effects of the essential oils of *Hagenia abyssinica*, *Leonotis ocymlifolia*, *Moringa stenopetala*, and their main individual constituents. *Phytomedicine*, 17(12), 911–920. <https://doi.org/10.1016/j.phymed.2010.02.009>
- Niu, S., Liu, D., Shao, Z., Liu, J., Fan, A., & Lin, W. (2021). Chemical epigenetic manipulation triggers the production of sesquiterpenes from the deep-sea derived *Eutypella fungus*. *Phytochemistry*, 192, Article 112978. <https://doi.org/10.1016/j.phytochem.2021.112978>
- Nogueira Neto, J. D., De Almeida, A. A. C., Da Silva Oliveira, J., Dos Santos, P. S., De Sousa, D. P., & De Freitas, R. M. (2013). Antioxidant effects of nerolidol in mice hippocampus after open field test. *Neurochemical Research*, 38(9), 1861–1870. <https://doi.org/10.1007/s11064-013-1092-2>
- Noor Hashim, N. H., Abas, F., Shaari, K., & Lajis, N. H. (2012). LC–DAD–ESIMS/MS characterization of antioxidant and anticholinesterase constituents present in the active fraction from *Persicaria hydropiper*. *LWT - Food Science and Technology*, 46(2), 468–476. <https://doi.org/10.1016/j.lwt.2011.11.021>
- Nugraha, F. (2021). Test the effectiveness of kesum leaf n-hexana extract gel (*Polygonum minus* Huds) as a repellent against mosquitoes. *Journal of Science, Technology, and Engineering*, 1(2), 91–97. <https://doi.org/10.33603/JSTE.V1I2.6206>
- Nurul, H., Ruzita, A., & Aronal, A. P. (2010). The antioxidant effects of *Cosmos caudatus* and *Polygonum minus* in refrigerated duck meatballs. *International Food Research Journal*, 9(4), 893–904.
- Omar, A. H., Kue, C. S., Dianita, R., & Yu, K. X. (2020). Teratogenic potential of traditional Malaysian vegetables (ulam) in the zebrafish model. *British Food Journal*, 122(10), 3089–3098. <https://doi.org/10.1108/BJFJ-02-2020-0118/FULL/PDF>
- Ong, H. C., & Nordiana, M. (1999). Malay ethno-medico botany in Machang, Kelantan, Malaysia. *Fitoterapia*, 70(5), 502–513. [https://doi.org/10.1016/S0367-326X\(99\)00077-5](https://doi.org/10.1016/S0367-326X(99)00077-5)
- Osmaniye, D., Sağlık, B. N., Khalilova, N., Levent, S., Bayazit, G., Gül, Ü. D., et al. (2023). Design, synthesis, and biological evaluation studies of novel naphthalene-chalcone hybrids as antimicrobial, anticandidal, anticancer, and VEGFR-2 inhibitors. *ACS Omega*, 8(7), 6669–6678. <https://doi.org/10.1021/acsomega.2c07256>
- Othman, A. R., Abdullah, N., Ahmad, S., Ismail, I. S., & Zakaria, M. P. (2015). Elucidation of *in vitro* anti-inflammatory bioactive compounds isolated from *Jatropha curcas* L. plant root. *BMC Complementary and Alternative Medicine*, 15(1), 1–10. <https://doi.org/10.1186/s12906-015-0528-4>
- Owolabi, K. A. A., L. L., & B. J. (2018). Biological activity of oleic acid and its primary amide : Experimental and computational studies. *Journal Of Chemical Society Of Nigeria*, 43(2), 9–18.
- Pérez-Hernández, N., Ponce-Monter, H., Ortiz, M. I., Carriño-Cortés, R., & Joseph-Nathan, P. (2009). Structure-activity relationships of aromadendranes in uterus-relaxant activity. *Zeitschrift Fur Naturforschung C*, 64(11–12), 840–846. <https://doi.org/10.1515/znc-2009-11-1214>
- Panase, P., & Tıpdaço, P. (2018). Preliminary use of *Polygonum minus* Linn. leaf extract on growth performance, feed utilization, and some hematological indices of *Anabas testudineus* (Bloch, 1792). *Comparative Clinical Pathology*, 27(1), 147–153. <https://doi.org/10.1007/s00580-017-2566-4>
- Paritalac, V., Chiruvella, K. K., Mohammed, A., Vikram, P., Kumar Chiruvella, K., Husna, I., et al. (2014). A recent review on phytochemical constituents and medicinal properties of kesum (*Polygonum minus* Linn.). *Asian Pacific Journal of Tropical Biomedicine Journal Homepage*, 4(1), 930–935. <https://doi.org/10.12980/APJTB.4.2014C1255>
- Paz, C., Viscardi, S., Iturra, A., Marin, V., Miranda, F., Barra, P. J., et al. (2020). Antifungal effects of drimane sesquiterpenoids isolated from *Drimys winteri* against *Gaeumannomyces graminis* var. *tritici*. *Applied and Environmental Microbiology*, 86(24), Article e01834. <https://doi.org/10.1128/AEM.1834-20.20>
- Peng, Q., Sun, X., Gong, T., Wu, C. Y., Zhang, T., Tan, J., et al. (2013). Injectable and biodegradable thermosensitive hydrogels loaded with PHBHHX nanoparticles for the sustained and controlled release of insulin. *Acta Biomaterialia*, 9(2), 5063–5069. <https://doi.org/10.1016/j.actbio.2012.09.034>
- Pham, E. C., Van, L. V., Nguyen, C. V., Duong, N. T. N., Le Thi, T. V., & Truong, T. N. (2023). Acute and sub-acute toxicity evaluation of *Merremia tridentata* (L.) stem extract on mice. *Toxicon*, 227, Article 107093. <https://doi.org/10.1016/j.toxicon.2023.107093>
- Phatik, T., Das, J., & Boruah, P. (2014). Antifungal activity of *Polygonum hydropiper* and *Solanum melongena* against plant pathogenic fungi. *Plant Archives*, 14(1), 15–17. <https://doi.org/10.13140/2.1.1011.1689>
- Poulose, N., Sajayan, A., Ravindran, A., Sreechithra, T. V., Vardhan, V., Selvin, J., et al. (2020). Photoprotective effect of nanomelanin-seaweed concentrate in formulated cosmetic cream: With improved antioxidant and wound healing properties. *Journal of Photochemistry and Photobiology B: Biology*, 205, Article 111816. <https://doi.org/10.1016/J.JPHOTOBIO.2020.111816>
- Prabhakaran, D., Jeemon, P., Sharma, M., Roth, G. A., Johnson, C., Harikrishnan, S., et al. (2018). The changing patterns of cardiovascular diseases and their risk factors in the states of India: The global burden of disease study 1990–2016. *Lancet Global Health*, 6(12), e1339–e1351. [https://doi.org/10.1016/S2214-109X\(18\)30407-8](https://doi.org/10.1016/S2214-109X(18)30407-8)
- Pramita, F. Y. (2013). Formulasi sediaan gel antiseptik ekstrak metanol daun kesum (*Polygonum minus* Huds). *Jurnal Mahasiswa Farmasi Fakultas Kedokteran UNTAN*, 3(1), 1–9.
- Pratibha, R., Sameer, R., Rataboli, P. V., Bhiwade, D. A., & Dhume, C. Y. (2006). Enzymatic studies of cisplatin induced oxidative stress in hepatic tissue of rats. *European Journal of Pharmacology*, 532(3), 290–293. <https://doi.org/10.1016/J.EJPHAR.2006.01.007>
- Premanath, R., James, J. P., Karunasagar, I., Vaňková, E., & Scholtz, V. (2022). Tropical plant products as biopreservatives and their application in food safety. *Food Control*, 141, Article 109185. <https://doi.org/10.1016/J.FOODCONT.2022.109185>
- Punia Bangar, S., Dunno, K., Kumar, M., Mostafa, H., & Maqsood, S. (2022). A comprehensive review on lotus seeds (*Nelumbo nucifera* gaertn.): Nutritional composition, health-related bioactive properties, and industrial applications. *Journal of Functional Foods*, 89, Article 104937. <https://doi.org/10.1016/J.JFF.2022.104937>
- Qader, S. W., Abdulla, M. A., Chua, L. S., Najim, N., Zain, M. M., & Hamdan, S. (2011). Antioxidant, total phenolic content and cytotoxicity evaluation of selected Malaysian plants. *Molecules*, 16(4), 3433–3443. <https://doi.org/10.3390/molecules16043433>
- Qader, S. W., Abdulla, M. A., Chua, L. S., Sirat, H. M., & Hamdan, S. (2012). Pharmacological mechanisms underlying gastroprotective activities of the fractions obtained from *Polygonum minus* in Sprague Dawley rats. *International Journal of Molecular Sciences*, 13(2), 1481–1496. <https://doi.org/10.3390/IJMS13021481>
- Qader, S. W., Ameen Abdulla, M., Chua, L. S., & Hamdan, S. (2012). Potential bioactive property of *Polygonum minus* Huds (kesum) review. *Scientific Research and Essays*, 7(2), 90–93. <https://doi.org/10.5897/SRE11.1789>
- Qin, L., Chen, S., Xie, L., Yu, Q., Chen, Y., Shen, M., et al. (2022). Mechanisms of RAW264.7 macrophages immunomodulation mediated by polysaccharide from mung bean skin based on RNA-seq analysis. *Food Research International*, 154, Article 111017. <https://doi.org/10.1016/J.FOODRES.2022.111017>
- Rahim, N. A., Nordin, N., Ahmad Rasedi, N. I. S., Mohd Kauli, F. S., Wan Ibrahim, W. N., & Zakaria, F. (2022). Behavioral and cortisol analysis of the anti-stress effect of *Polygonum minus* (Huds) extracts in chronic unpredictable stress (CUS) zebrafish model. *Comparative Biochemistry and Physiology - Part C: Toxicology & Pharmacology*, 256, Article 109303. <https://doi.org/10.1016/J.CBPC.2022.109303>
- Rahnamaie-Tajadod, R., Goh, H. H., & Mohd Noor, N. (2019). Methyl jasmonate-induced compositional changes of volatile organic compounds in *Polygonum minus* leaves. *Journal of Plant Physiology*, 240, Article 152994. <https://doi.org/10.1016/J.JPLPH.2019.152994>
- Rahnamaie-Tajadod, R., Loke, K. K., Goh, H. H., & Noor, N. M. (2017). Differential gene expression analysis in *Polygonum minus* leaf upon 24h of methyl jasmonate elicitation. *Frontiers in Plant Science*, 8, 1–14. <https://doi.org/10.3389/fpls.2017.00109>
- Ramachandriah, K., & Chin, K. B. (2016). Evaluation of ball-milling time on the physicochemical and antioxidant properties of persimmon by-products powder. *Innovative Food Science & Emerging Technologies*, 37, 115–124. <https://doi.org/10.1016/J.IJFSET.2016.08.005>
- Rashid, N. A., Hussain, F., Hamid, A., Ridzuan, N. R. A., Halim, S. A. S. A., Jalil, N. A. A., et al. (2020). *Polygonum minus* essential oil modulates cisplatin-induced hepatotoxicity through inflammatory and apoptotic pathways. *EXCLI Journal*, 19, 1265. <https://doi.org/10.17179/EXCLI2020-2355>
- Rashid, N. A., Hussain, F., Hamid, A., Ridzuan, N. R. A., Lin, T. S., & Budin, S. B. (2019). Preventive effects of *Polygonum minus* essential oil on cisplatin-induced hepatotoxicity in Sprague Dawley rats. *Sains Malaysiana*, 48(9), 1975–1988. <https://doi.org/10.17576/jsm-2019-4809-19>
- Regner, G. G., Giancesini, J., Von Borowski, R. G., Silveira, F., Semedo, J. G., Ferraz, A. de B. F., et al. (2011). Toxicological evaluation of *Pterocaulon polystachyum* extract: A medicinal plant with antifungal activity. *Environmental Toxicology and Pharmacology*, 31(1), 242–249. <https://doi.org/10.1016/J.ETAP.2010.11.003>
- Rehman, A., Arif, M., Sajjad, N., Al-Ghadi, M. Q., Alagawany, M., Abd El-Hack, M. E., et al. (2020). Dietary effect of probiotics and prebiotics on broiler performance, carcass, and immunity. *Poultry Science*, 99(12), 6946–6953. <https://doi.org/10.1016/j.psj.2020.09.043>
- Renugadevi, K., Valli Nachiyar, C., & Zaveri, M. (2021). Bioactivity of dodecanoic acid extracted from *Geitlerinema* sp. Trv57. *Indian Journal of Pharmaceutical Education and Research*, 55(1), 224–231. <https://doi.org/10.5530/ijper.55.1.25>

- Reverter, M., Bontemps, N., Lecchini, D., Banaigs, B., & Sasal, P. (2014). Use of plant extracts in fish aquaculture as an alternative to chemotherapy: Current status and future perspectives. *Aquaculture*, 433, 50–61. <https://doi.org/10.1016/j.aquaculture.2014.05.048>
- Reza, A. S. M. A., Haque, M. A., Sarker, J., Nasrin, M. S., Rahman, M. M., Tareq, A. M., et al. (2021). Antiproliferative and antioxidant potentials of bioactive edible vegetable fraction of *Achyranthes ferruginea* Roxb. in cancer cell line. *Food Science and Nutrition*, 9(7), 3777–3805. <https://doi.org/10.1002/fsn3.2343>
- Rezki, S., Halimah, H., Maryani, Y., & Setyowati, N. (2020). The influence of *Polygonum minus* huds on bacteria to acrylic denture. *Jurnal Kesehatan Gigi*, 7(1), 68–72. <https://doi.org/10.31983/jkg.v7i1.5777>
- Ribeiro-Santos, R., Andrade, M., Madella, D., Martinazzo, A. P., de Aquino Garcia Moura, L., de Melo, N. R., et al. (2017). Revisiting an ancient spice with medicinal purposes: Cinnamon. *Trends in Food Science and Technology*, 62, 154–169. <https://doi.org/10.1016/j.tifs.2017.02.011>
- Ridzuan, P. M., Hairul Aini, H., Norazian, M. H., Shah, A., Roesnita, & Aminah, K. S. (2013). Antibacterial and antifungal properties of *Persicaria odorata* leaf against pathogenic bacteria and fungi. *The Open Conference Proceedings Journal*, 4(1), 71–74. <https://doi.org/10.2174/2210289201304020071>
- Ridzuan, N. A., Teoh, S., Abdul Rashid, N., Othman, F., Baharum, S., & Hussain, F. (2019). *Polygonum minus* ethanolic extracts attenuate cisplatin-induced oxidative stress in the cerebral cortex of rats via its antioxidant properties. *Asian Pacific Journal of Tropical Biomedicine*, 9(5), 196–203. <https://doi.org/10.4103/2221-1691.258999>
- Rita, Kusharyanti, I., & Wahdaningsih, S. (2013). Nephroprotective activity of kesum leaves (*Polygonum minus* Huds.) metanolic fraction on Wistar male rats (*Rattus norvegicus*) induced by cisplatin. *Jurnal Mahasiswa Farmasi Fakultas Kedokteran UNTAN*, 1(1), 1–15.
- Rohin, M. A. K., Hadi, N. A., & Ridzuan, N. (2020). Different polarity extracts of *Polygonum minus* towards cytotoxic activities against colon cancer cell lines (HT-29, HCT-116, CT-26). *Journal of Pharmaceutical Research International*, 32(14), 168–180. <https://doi.org/10.9734/jpri/2020/v32i1430617>
- Romo-Araiza, A., & Ibarra, A. (2020). Prebiotics and probiotics as potential therapy for cognitive impairment. *Medical Hypotheses*, 134, Article 109410. <https://doi.org/10.1016/j.mehy.2019.109410>
- Roslan, N. D., Yusop, J. M., Baharum, S. N., Othman, R., Mohamed-Hussein, Z. A., Ismail, I., et al. (2012). Flavonoid biosynthesis genes putatively identified in the aromatic plant *Polygonum minus* via expressed sequences tag (EST) analysis. *International Journal of Molecular Sciences*, 13(3), 2692–2706. <https://doi.org/10.3390/ijms13032692>
- Rousselle, P., Braye, F., & Dayan, G. (2019). Re-Epithelialization of adult skin wounds: Cellular mechanisms and therapeutic strategies. *Advanced Drug Delivery Reviews*, 146, 344–365. <https://doi.org/10.1016/j.addr.2018.06.019>
- Rufino, A. T., Ribeiro, M., Judas, F., Salgueiro, L., Lopes, M. C., Cavaleiro, C., et al. (2014). Anti-inflammatory and chondroprotective activity of (+)- α -pinene: Structural and enantiomeric selectivity. *Journal of Natural Products*, 77(2), 264–269. <https://doi.org/10.1021/np400828x>
- Rusdi, N. A., Goh, H. H., & Baharum, S. N. (2016). GC-MS/Olfactometric characterisation and aroma extraction dilution analysis of aroma active compounds in *Polygonum minus* essential oil. *Plant Omics*, 9(4), 289–294. <https://doi.org/10.21475/poj.16.09.04.p7901>
- Russo, E. B. (2011). Taming THC: Potential cannabis synergy and phytocannabinoid-terpenoid entourage effects. *British Journal of Pharmacology*, 163(7), 1344–1364. <https://doi.org/10.1111/j.1476-5381.2011.01238.x>
- Śęczyk, L., Ahmet Ozdemir, F., & Kołodziej, B. (2022). *In vitro* bioaccessibility and activity of basil (*Ocimum basilicum* L.) phytochemicals as affected by cultivar and postharvest preservation method – convection drying, freezing, and freeze-drying. *Food Chemistry*, 382, Article 132363. <https://doi.org/10.1016/j.foodchem.2022.132363>
- Sabaragamuwa, R., Perera, C. O., & Pedrizzi, B. (2018). *Centella asiatica* (Gotu kola) as a neuroprotectant and its potential role in healthy ageing. *Trends in Food Science and Technology*, 79, 88–97. <https://doi.org/10.1016/j.tifs.2018.07.024>
- Sabulal, B., Dan, M., J. A. J., Kurup, R., Pradeep, N. S., Valsamma, R. K., et al. (2006). Caryophyllene-rich rhizome oil of *Zingiber nimonii* from south India: Chemical characterization and antimicrobial activity. *Phytochemistry*, 67(22), 2469–2473. <https://doi.org/10.1016/j.phytochem.2006.08.003>
- Saganuwan, S. A. (2017). Toxicity studies of drugs and chemicals in animals: An overview. *Bulgarian Journal of Veterinary Medicine*, 20(4), 291–318. <https://doi.org/10.15547/bjvm.983>
- Sahi, N. M. (2016). Evaluation of insecticidal activity of bioactive compounds from *Eucalyptus citriodora* against *Tribolium castaneum*. *International Journal of Pharmacognosy and Phytochemical Research*, 8(8), 1256–1270.
- Sain, S., Naoghare, P., Devi, S., Daiwile, A., Krishnamurthi, K., Arrigo, P., et al. (2014). Beta caryophyllene and caryophyllene oxide, isolated from *Aegle marmelos*, as the potent anti-inflammatory agents against lymphoma and neuroblastoma cells. *Antiinflamm Agents Med Chem*, 13(1), 45–55. <https://doi.org/10.2174/18715230113129990016>
- Saito, A. Y., Marin Rodriguez, A. A., Menchaca Vega, D. S., Sussmann, R. A. C., Kimura, E. A., & Katzin, A. M. (2020). Antimalarial activity of the terpene nerolidol. *International Journal of Antimicrobial Agents*, 48(6), 641–646. <https://doi.org/10.1016/j.ijantimicag.2016.08.017>
- Salem, H. M., Khatib, M. S., Yehia, N., El-Hack, M. E. A., El-Saadony, M. T., Alhimaidi, A. R., et al. (2022). Morphological and molecular characterization of *Ascaridia columbae* in the domestic pigeon (*Columba livia domestica*) and the assessment of its immunological responses. *Poultry Science*, 101(2), Article 101596. <https://doi.org/10.1016/J.PS.2021.101596>
- Santos, P.C.M., dos, da Silva, L. M. R., Magalhaes, F. E. A., Cunha, F. E. T., Ferreira, M. J. G., & de Figueiredo, E. A. T. (2022). Garlic (*Allium sativum* L.) peel extracts: From industrial by-product to food additive. *Applied Food Research*, 2(2), Article 100186. <https://doi.org/10.1016/j.afres.2022.100186>
- Sardar, R., Funston, A. M., Mulvaney, P., & Murray, R. W. (2009). Gold nanoparticles: Past, present, and future. *Langmuir*, 25(24), 13840–13851. https://doi.org/10.1021/LA9019475/ASSET/IMAGES/MEDIUM/LA-2009-019475_0013.GIF
- Sarkar, R., Arora, P., & Garg, K. V. (2013). Cosmeceuticals for hyperpigmentation: What is available? *Journal of Cutaneous and Aesthetic Surgery*, 6(1), 4–11. <https://doi.org/10.4103/0974-2077.110089>
- Scherübl, H. (2020). Alcohol use and gastrointestinal cancer risk. *Visceral Medicine*, 36(3), 175–181. <https://doi.org/10.1159/000507232>
- Seshadri, V. D., Ouyouni, A. A. A., Bawazir, W. M., Alsagaby, S. A., Alsharif, K. F., Albrakati, A., et al. (2022). *Zingiberene* exerts chemopreventive activity against 7,12-dimethylbenz(a)anthracene-induced breast cancer in Sprague Dawley rats. *Journal of Biochemical and Molecular Toxicology*, 36(10), Article e23146. <https://doi.org/10.1002/jbt.23146>
- Sete da Cruz, R. M., da Silva, C., da Silva, E. A., Hegel, P., Barão, C. E., & Cardozo-Filho, L. (2022). Composition and oxidative stability of oils extracted from *Zophobas morio* and *Tenebrio molitor* using pressurized n-propane. *The Journal of Supercritical Fluids*, 181, Article 105504. <https://doi.org/10.1016/J.SUPFLU.2021.105504>
- Severn, M. M., & Horswill, A. R. (2023). *Staphylococcus epidermidis* and its dual lifestyle in skin health and infection. *Nature Reviews Microbiology*, 21(2), 97–111. <https://doi.org/10.1038/s41579-022-00780-3>
- Shaaban, M. T., Ghaly, M. F., & Fahmi, S. M. (2021). Antibacterial activities of hexadecanoic acid methyl ester and green-synthesized silver nanoparticles against multidrug-resistant bacteria. *Journal of Basic Microbiology*, 61(6), 557–568. <https://doi.org/10.1002/jobm.202100061>
- Shahar, S., Hamdan, S., Baba, S., & Paul Joko, A. D. (2015). *In vitro* antiviral activity of aqueous extracts of *Polygonum minus* huds leaves against herpes simplex virus 1. *29th Scientific Meeting of Malaysian Society of Pharmacology and Physiology (MSPP)*, 1.
- Shaheena, S., Chintagunta, A. D., Dirisala, V. R., & Sampath Kumar, N. S. (2019). Extraction of bioactive compounds from *Psidium guajava* and their application in dentistry. *AMB Express*, 9(1), 1–9. <https://doi.org/10.1186/s13568-019-0935-x>
- Sharma, N., & Gulati, A. (2023). Natural vitamins as food antimicrobials in stem and thorn extracts of *Hippophae species* studied by HPLC-ESI-MS. *Food and Humanity*, 1, 415–420. <https://doi.org/10.1016/J.FOOHUM.2023.06.017>
- Sharma, N., Kumar, V., Chopra, M. P., Sourirajan, A., Dev, K., & El-Shazly, M. (2020). *Thalictrum foliolosum*: A lesser unexplored medicinal herb from the himalayan region as a source of valuable benzyl isoquinoline alkaloids. *Journal of Ethnopharmacology*, 255, Article 112736. <https://doi.org/10.1016/J.JEP.2020.112736>
- Sharma, P., & Shah, G. C. (2015). Composition and antioxidant activity of *Senecio nudicaulis* wall. Ex DC. (Asteraceae): A medicinal plant growing wild in Himachal Pradesh, India. *Natural Product Research*, 29(9), 883–886. <https://doi.org/10.1080/14786419.2014.990904>
- Sheth, U., & Dande, P. (2021). *Pityriasis capitis*: Causes, pathophysiology, current modalities, and future approach. *Journal of Cosmetic Dermatology*, 20(1), 35–47. <https://doi.org/10.1111/jocd.13488>
- Shevchenko, A. S., Muzychikina, R. A., Ross, S. A., & Korul'kin, D. Y. (2019). Procedures used for isolation of complex biological active constituents from *Polygonum L.* *Journal of Chemical Technology and Metallurgy*, 54(3), 508–513.
- Shin, Y., & Lee, Y. (2013). Cytotoxic activity from *Curcuma zedoaria* through mitochondrial activation on ovarian cancer cells. *Toxicological Research*, 29(4), 257–261. <https://doi.org/10.5487/TR.2013.29.4.257>
- Shin, I. S., Seo, C. S., Ha, H. K., Lee, M. Y., Huang, D. S., Huh, J. I., et al. (2011). Genotoxicity assessment of Pyungwi-san (PWS), a traditional herbal prescription. *Journal of Ethnopharmacology*, 133(2), 696–703. <https://doi.org/10.1016/J.JEP.2010.10.050>
- Shoge, M., Garba, S., & Labaran, S. (2017). Antimicrobial activity of 1,2-benzenedicarboxylic acid, butyldecyl ester isolated from the seeds and pods of *Acacia nilotica* Linn. *Basic Research Journal of Microbiology*, 3(2), 8–11.
- Shukor, M. F. A., Ismail, I., Zainal, Z., & Noor, N. M. (2013). Development of a *Polygonum minus* cell suspension culture system and analysis of secondary metabolites enhanced by elicitation. *Acta Physiologiae Plantarum*, 35(5), 1675–1689. <https://doi.org/10.1007/s11738-012-1210-9>
- Sianipar, N. F., Purnamaningsih, R., & Rosaria. (2016). Bioactive compounds of fourth generation gamma-irradiated *Typhonium flagelliforme* Lodd. mutants based on gas chromatography-mass spectrometry. *IOP Conference Series: Earth and Environmental Science*, 41(1), Article 012025. <https://doi.org/10.1088/1755-1315/41/1/012025>
- Silva, T. de M., Miranda, R. R. S., Ferraz, V. P., Pereira, M. T., de Siqueira, E. P., & Alcântara, A. F. C. (2013). Changes in the essential oil composition of leaves of *Echinodorus macrophyllus* exposed to γ -radiation. *Revista Brasileira de Farmacognosia*, 23(4), 600–607. <https://doi.org/10.1590/S0102-695X2013005000049>
- Singh, N., & Khajuria, V. (2015). Effect of cisplatin on liver of male albino rats. *Journal of Evolution of Medical and Dental Sciences*, 4(52), 8993–8998. <https://doi.org/10.14260/jemds/2015/1305>
- Siswadi, S., & Saragih, G. S. (2021). Phytochemical analysis of bioactive compounds in ethanolic extract of *Sterculia quadrifida* R.Br. *AIP Conference Proceedings*, 2353, Article 030098. <https://doi.org/10.1063/5.0053057>
- Siswanto, A. N. (2018). *Protective effect of Polygonum minus leaves ethanolic extract on cadmium chloride-induced toxicity on spermatogenic cells of mice (Mus musculus)* [Universitas Airlangga]. <http://lib.unair.ac.id>
- Song, Y., Seo, S., Lamichhane, S., Seo, J., Hong, J. T., Cha, H. J., et al. (2021). Limonene has anti-anxiety activity via adenosine A2A receptor-mediated regulation of

- dopaminergic and GABAergic neuronal function in the striatum. *Phytomedicine*, 83, Article 153474. <https://doi.org/10.1016/j.phymed.2021.153474>
- Su, Z. S., Yin, S., Zhou, Z. W., Wu, Y., Ding, J., & Yue, J. M. (2008). Sesquiterpenoids from *Hedyosmum orientale*. *Journal of Natural Products*, 71(8), 1410–1413. <https://doi.org/10.1021/np800240v>
- Suaya, J. A., Fletcher, M. A., Georgalis, L., Arguedas, A. G., McLaughlin, J. M., Ferreira, G., et al. (2021). Identification of *Streptococcus pneumoniae* in hospital-acquired pneumonia in adults. *Journal of Hospital Infection*, 108, 146–157. <https://doi.org/10.1016/j.jhin.2020.09.036>
- Sun, Y., Ho, C. T., Zhang, Y., Hong, M., & Zhang, X. (2023). Plant polysaccharides utilized by gut microbiota: New players in ameliorating cognitive impairment. *Journal of Traditional and Complementary Medicine*, 13(2), 128–134. <https://doi.org/10.1016/j.jtcm.2022.01.003>
- Sun, Y., Qiao, H., Ling, Y., Yang, S., Rui, C., Pelosi, P., et al. (2011). New analogues of (E)- β -farnesene with insecticidal activity and binding affinity to aphid odorant-binding proteins. *Journal of Agricultural and Food Chemistry*, 59(6), 2456–2461. <https://doi.org/10.1021/jf104712c>
- Suvarna, V., Nair, A., Mallya, R., Khan, T., & Omri, A. (2022). Antimicrobial nanomaterials for food packaging. *Antibiotics*, 11(6), 1–31. <https://doi.org/10.3390/antibiotics11060729>
- Suvila, K., & Niiranen, T. J. (2022). Interrelations between high blood pressure, organ damage, and cardiovascular disease: No more room for doubt. *Hypertension*, 79(3), 516–517. <https://doi.org/10.1161/HYPERTENSIONAHA.121.18786>
- Syahmina, A., & Usuki, T. (2020). Ionic liquid-assisted extraction of essential oils from *Thujaopsis dolabrata* (Hiba). *ACS Omega*, 5(45), 29618–29622. <https://doi.org/10.1021/acsomega.0c04860>
- Syaiful, Jayuska, A., & Harlia. (2015). Pengaruh waktu distilasi terhadap komponen minyak atsiri pada daun kesum (*Polygonum minus* Huds.). *Jurnal Kimia Khatulistiwa*, 4 (1), 18–23.
- Türker, H., Çelik, K., & Toğar, B. (2014). Effects of copaene, a tricyclic sesquiterpene, on human lymphocytes cells *in vitro*. *Cytotechnology*, 66(4), 597–603. <https://doi.org/10.1007/s10616-013-9611-1>
- Tada, H., & Yasuda, F. (1985). Metabolites from the marine sponge *Epipolasis kushimotoensis*. *Chemical and Pharmaceutical Bulletin*, 33(5), 1941–1945. <https://doi.org/10.1248/cpb.33.1941>
- Tai, Y. T., Chou, S. H., Cheng, C. Y., Ho, C. T., Lin, H. C., Jung, S. M., et al. (2022). The preferential accumulation of cadmium ions among various tissues in mice. *Toxicology Reports*, 9, 111–119. <https://doi.org/10.1016/j.toxrep.2022.01.002>
- Takao, Y., Kuriyama, I., Yamada, T., Mizoguchi, H., Yoshida, H., & Mizushima, Y. (2012). Antifungal properties of Japanese cedar essential oil from waste wood chips made from used sake barrels. *Molecular Medicine Reports*, 5(5), 1163–1168. <https://doi.org/10.3892/mmr.2012.821>
- Tan, T., Li, J., Luo, R., Wang, R., Yin, L., Liu, M., et al. (2021). Recent advances in understanding the mechanisms of elemene in reversing drug resistance in tumor cells: A review. *Molecules*, 26(19), 5792. <https://doi.org/10.3390/molecules26195792>
- Tashim, N. A. Z., Lim, S. A., & Basri, A. M. (2022). Synergistic antioxidant activity of selected medicinal plants in Brunei Darussalam and its application in developing fortified pasta. *Journal of the Science of Food and Agriculture*, 102(15), 7331–7342. <https://doi.org/10.1002/JSSFA.12099>
- Tatman, D., & Mo, H. (2002). Volatile isoprenoid constituents of fruits, vegetables and herbs cumulatively suppress the proliferation of murine B16 melanoma and human HL-60 leukemia cells. *Cancer Letters*, 175(2), 129–139. [https://doi.org/10.1016/S0304-3835\(01\)00723-6](https://doi.org/10.1016/S0304-3835(01)00723-6)
- Tatsuno, K., Fujiyama, T., Matsuoka, H., Shimauchi, T., Ito, T., & Tokura, Y. (2016). Clinical categories of exaggerated skin reactions to mosquito bites and their pathophysiology. *Journal of Dermatological Science*, 82(3), 145–152. <https://doi.org/10.1016/j.jdermsci.2016.04.010>
- Terés, S., Barceló-Coblijn, G., Benet, M., Álvarez, R., Bressani, R., Halver, J. E., et al. (2008). Oleic acid content is responsible for the reduction in blood pressure induced by olive oil. *Proceedings of the National Academy of Sciences of the United States of America*, 105(37), 13811–13816. <https://doi.org/10.1073/pnas.0807500105>
- Thanapakiam, G., Lim, S. Y., Grace, K. Y. X., Loo, S. W., Amutha, V. V., Pavitra, L., et al. (2016). Effect of aqueous and methanol extracts *Polygonum minus* leaves on drug-induced hepatotoxicity in rats toxicity of herbal extract (acute and chronic). *In The 3rd regional conference on biosensors, biodiagnostics, biochips and biotechnology* (Vol. 1).
- To, N. B., Nguyen, Y. T. K., Moon, J. Y., Ediriweera, M. K., & Cho, S. K. (2020). Pentadecanoic acid, an odd-chain fatty acid, suppresses the stemness of MCF-7/SC human breast cancer stem-like cells through JAK2/STAT3 signaling. *Nutrients*, 12(6), 1663. <https://doi.org/10.3390/nu12061663>
- Tommasi, N. De, Pizca, C., Conti, C., Orsi, N., & Stein, M. L. (1990). Structure and *in vitro* antiviral activity of sesquiterpene glycosides from *Calendula arvensis*. *Journal of Natural Products*, 53(4), 830–835. <https://doi.org/10.1021/np50070a009>
- Tommy, T. (2013). *Renoprotective effect of kesum leaves (Polygonum minus Huds.) nhexane fraction as co-chemotherapy on male Wistar rat induced by cis-diammine dichloridoplatinum*. Tanjungpura University.
- Tsoyi, K., Jang, H. J., Lee, Y. S., Kim, Y. M., Kim, H. J., Seo, H. G., et al. (2011). (+)-Nootkatone and (+)-valencene from rhizomes of *Cyperus rotundus* increase survival rates in septic mice due to heme oxygenase-1 induction. *Journal of Ethnopharmacology*, 137(3), 1311–1317. <https://doi.org/10.1016/j.jep.2011.07.062>
- Tutunchi, H., Ostadrahimi, A., & Saghaei-Asl, M. (2020). The effects of diets enriched in monounsaturated oleic acid on the management and prevention of obesity: a systematic review of human intervention studies. *Advances in Nutrition*, 11(4), 864–877. <https://doi.org/10.1093/ADVANCES/NMAA013>
- Udani, J. K., George, A. A., Musthapa, M., Pakdaman, M. N., & Abas, A. (2014). Effects of a proprietary freeze-dried water extract of *Eurycoma longifolia* (Physta) and *Polygonum minus* on sexual performance and well-being in men: A randomized, double-blind, placebo-controlled study. *Evidence-based Complementary and Alternative Medicine*, 2014, 1–10. <https://doi.org/10.1155/2014/179529>
- Ullah, H., Wilfred, C. D., & Shaharun, M. S. (2017). Synthesis of silver nanoparticles using ionic-liquid-based microwave-assisted extraction from *Polygonum minus* and photodegradation of methylene blue. *Journal of the Chinese Chemical Society*, 64(10), 1164–1171. <https://doi.org/10.1002/JCCS.201700144>
- Ullah, H., Wilfred, C. D., & Shaharun, M. S. (2018a). Green synthesis of copper nanoparticle using ionic liquid-based extraction from *Polygonum minus* and their applications. *Environmental Technology*, 40(28), 3705–3712. <https://doi.org/10.1080/09593330.2018.1485751>
- Ullah, H., Wilfred, C. D., & Shaharun, M. S. (2018b). Synthesis of nickel nanoparticle using ionic liquid based extract from *Polygonum minus* and their applications. *Desalination and Water Treatment*, 125, 32–39. <https://doi.org/10.5004/dwt.2018.22938>
- Ullah, H., Wilfred, C. D., & Shaharun, M. S. (2019). Comparative assessment of various extraction approaches for the isolation of essential oil from *Polygonum minus* using ionic liquids. *Journal of King Saud University Science*, 31(2), 230–239. <https://doi.org/10.1016/j.jksus.2017.05.014>
- Urzúa, A., Santander, R., Echeverría, J., Cabezas, N., Palacios, S. M., & Rossi, Y. (2010). Insecticide properties of the essential oils from *Haploppappus foliosus* and *Bahia Ambrosioides* against the house fly, *Musca Domestica* L. *Journal of the Chilean Chemical Society*, 55(3), 392–395. <https://doi.org/10.4067/S0717-97072010000300026>
- Utegenova, G. A., Pallister, K. B., Kushnarenko, S. V., Özek, G., Özek, T., Abidkulova, K. T., et al. (2018). Chemical composition and antibacterial activity of essential oils from *Ferula* L. species against methicillin-resistant *Staphylococcus aureus*. *Molecules*, 23(7), 1679. <https://doi.org/10.3390/molecules23071679>
- Uzunhisarcikli, M., Aslanturk, A., Kalender, S., Apaydin, F. G., & Bas, H. (2016). Mercuric chloride induced hepatotoxic and hematological changes in rats: The protective effects of sodium selenite and vitamin E. *Toxicology and Industrial Health*, 32(9), 1651–1662. <https://doi.org/10.1177/0748233715572561>
- Vachhrājani, K. D., Makhija, S., Chinoy, N. J., & Chowdhury, A. R. (1988). Structural and functional alterations in testis of rats after mercuric chloride treatment. *Journal of Reproductive Biology and Comparative Endocrinology*, 8, 97–104.
- Van Hai, N. (2015). The use of medicinal plants as immunostimulants in aquaculture: A review. *Aquaculture*, 446, 88–96. <https://doi.org/10.1016/J.AQUACULTURE.2015.03.014>
- van Wyk, A. S., & Prinsloo, G. (2020). Health, safety and quality concerns of plant-based traditional medicines and herbal remedies. *South African Journal of Botany*, 133, 54–62. <https://doi.org/10.1016/J.SAJB.2020.06.031>
- Vander Does, A., Labib, A., & Yosipovitch, G. (2022). Update on mosquito bite reaction: Itch and hypersensitivity, pathophysiology, prevention, and treatment. *Frontiers in Immunology*, 13, Article 1024559. <https://doi.org/10.3389/fimmu.2022.1024559>
- Veerasamy, R., Min, L. S., Mohanraj, Pauline, R., Sivadasan, S., Varghese, C., et al. (2014). Effect of aqueous extract of *Polygonum minus* leaf on the immunity and survival of African catfish (*Clarias gariepinus*). *Journal of Coastal Life Medicine*, 2(3), 209–213. <https://doi.org/10.12980/jclm.2.2014ajltb-2014-0007>
- Venn-Watson, S. K., & Butterworth, C. N. (2022). Broader and safer clinically-relevant activities of pentadecanoic acid compared to omega-3: Evaluation of an emerging essential fatty acid across twelve primary human cell-based disease systems. *PLoS One*, 17(5), Article e0268778. <https://doi.org/10.1371/journal.pone.0268778>
- Venn-Watson, S., Lumpkin, R., & Dennis, E. A. (2020). Efficacy of dietary odd-chain saturated fatty acid pentadecanoic acid parallels broad associated health benefits in humans: Could it be essential? *Scientific Reports*, 10(1), 8161. <https://doi.org/10.1038/s41598-020-64960-y>
- Verzosa, A. L., McGeever, L. A., Bhark, S. J., Delgado, T., Salazar, N., & Sanchez, E. L. (2021). Herpes simplex virus 1 infection of neuronal and non-neuronal cells elicits specific innate immune responses and immune evasion mechanisms. *Frontiers in Immunology*, 12, Article 644664. <https://doi.org/10.3389/fimmu.2021.644664>
- Vieira, J. V., dos, A., Marques, V. B., Vieira, L. V., Crajoinas, R. de O., Shimizu, M. H. M., et al. (2021). Changes in the renal function after acute mercuric chloride exposure in the rat are associated with renal vascular endothelial dysfunction and proximal tubule NHE3 inhibition. *Toxicology Letters*, 341, 23–32. <https://doi.org/10.1016/J.TOXLET.2021.01.014>
- Vikram, P., Chiruvella, K. K., Ripain, I. H. A., & Arifullah, M. (2014). A recent review on phytochemical constituents and medicinal properties of kesum (*Polygonum minus* Huds.). *Asian Pacific Journal of Tropical Biomedicine*, 4(6), 430–435. <https://doi.org/10.12980/APJTb.4.2014C1255>
- Vimala, S., Adenan, M. I., Ahmad, A. R., & Shahdan, R. (2003). *Nature's choice to wellness: Antioxidant vegetables/ulam*. Forest Research Institute Malaysia (FRIM).
- Vimala, S., Rohana, S., Rashih, A., & M, J. (2011). Antioxidant evaluation in Malaysian medicinal plant: *Persicaria minor* (Huds.) leaf. *Science Journal of Medicine and Clinical Trials*, 2012, 9–16. <http://www.sjpub.org/sjmet/abstract/sjmet-215.html>
- Vlad, P. F. (2006). Synthetic investigations in the field of drimane sesquiterpenoids. *Studies in Natural Products Chemistry*, 33, 393–432. [https://doi.org/10.1016/S1572-5995\(06\)80031-1](https://doi.org/10.1016/S1572-5995(06)80031-1)
- Wahab, N. Z. A., Bunawan, H., & Ibrahim, N. (2015). Cytotoxicity and antiviral activity of methanol extract from *Polygonum minus*. *AIP Conference Proceedings*, 1678(1), Article 030036. <https://doi.org/10.1063/1.4931257>
- Walker, K. R., & Tesco, G. (2013). Molecular mechanisms of cognitive dysfunction following traumatic brain injury. *Frontiers in Aging Neuroscience*, 5, 29. <https://doi.org/10.3389/fnagi.2013.00029>
- Wan-Ibrahim, W. I., Sidik, K., & Kuppasamy, U. R. (2010). A high antioxidant level in edible plants is associated with genotoxic properties. *Food Chemistry*, 122(4), 1139–1144. <https://doi.org/10.1016/J.FOODCHEM.2010.03.101>

- Wang, X., Wang, X., Wen, M., & Li, X. (2023). Bibliometric analysis of literature on prevention and treatment of gastric ulcer with natural medicines. *Journal of Future Foods*, 3(3), 225–233. <https://doi.org/10.1016/J.JFUTFO.2023.02.004>
- Wang, C., Zada, B., Wei, G., & Kim, S. W. (2017). Metabolic engineering and synthetic biology approaches driving isoprenoid production in *Escherichia coli*. *Bioresource Technology*, 241, 430–438. <https://doi.org/10.1016/j.biortech.2017.05.168>
- Wasman, S., Ameen, M., Chua, L. S., & Hamdan, S. (2012). Gastric protection ability of some medicinal Malaysian plants against ethanol induction model in Sprague Dawley rats. *Jurnal Teknologi (Sciences and Engineering)*, 57(1), 199–209. <https://doi.org/10.11113/JT.V57.1534>
- Wasman, S. Q., Mahmood, A. A., Salehuddin, H., Zahra, A. A., & Salmah, I. (2010). Cytoprotective activities of *Polygonum minus* aqueous leaf extract on ethanol-induced gastric ulcer in rats. *Journal of Medicinal Plants Research*, 4(24), 2658–2665. <https://doi.org/10.5897/jmpr09.412>
- Wei, M. Y., Famouri, L., Carroll, L., Lee, Y., & Famouri, P. (2013). Rapid and efficient sonomechanical formation of gold nanoparticles under ambient conditions using functional alkoxy silane. *Ultrasonics Sonochemistry*, 20(1), 610–617. <https://doi.org/10.1016/J.ULTSONCH.2012.07.028>
- Westlake, R., Tran, J. W., Yunhong Jiang, M., Xinyu Zhang, D., Burrows, A., & Xie, M. (2023). Biodegradable biopolymers for active packaging: Demand, development and directions. *Sustainable Food Technology*, 1(1), 50–72. <https://doi.org/10.1039/D2FB00004K>
- Wiart, C. (2006). *Medicinal plants of Asia and the pacific*. CRC Press. <https://doi.org/10.1201/9781420006803/MEDICINAL-PLANTS-ASIA-PACIFIC-CHRISTOPHE-WIART>
- Wibowo, M. A., Anwari, M. S., Aulanni'am, & Rahman, F. (2010). Antiproliferation effect of the n-hexanal extract of kesum (*Polygonum minus*) at the cells of the rat lung cancer after exploiting of kesum as drug of lung cancer. *Media Kedokteran Hewan*, 26(2), 128–133.
- Wibowo, M. A., Purnomo, B. B., Widodo, M. A., & Aulanni'am, A. (2012). The n-hexane fraction of kesum (*Polygonum minus* L.) induce apoptosis the lung epithelial cells of the ratu novergicus that exposed by benzopyrene. *Berkala Penelitian Hayati*, 18(1), 69–73. <https://doi.org/10.23869/bphjbr.18.1.201211>
- Wibowo, M. A., Widodo, M. A., Purnomo, B. B., & A, A. (2013). Ekstrak daun kesum (*Polygonum minus*) memperbaiki kerusakan paru melalui ditekannya produksi reactive oxygen species (ROS). *Jurnal Kedokteran Hewan - Indonesian Journal of Veterinary Sciences*, 7(2), 160–162. <https://doi.org/10.21157/J.KED.HEWAN.V7I2.936>
- Wong, K. H., Li, G. Q., Li, K. M., Razmovski-Naumovski, V., & Chan, K. (2011). Kudzu root: Traditional uses and potential medicinal benefits in diabetes and cardiovascular diseases. *Journal of Ethnopharmacology*, 134(3), 584–607. <https://doi.org/10.1016/J.JEP.2011.02.001>
- Wu, L. C., Ho, J. A. A., Shieh, M. C., & Lu, I. W. (2005). Antioxidant and antiproliferative activities of *Spirulina* and *Chlorella* water extracts. *Journal of Agricultural and Food Chemistry*, 53(10), 4207–4212. <https://doi.org/10.1021/jf0479517>
- Wu, Y., Wang, K., Huang, S., Yang, C., & Wang, M. (2017). Near-infrared light-responsive semiconductor polymer composite hydrogels: Spatial/temporal-controlled release via a photothermal “sponge” effect. *ACS Applied Materials and Interfaces*, 9(15), 13602–13610. https://doi.org/10.1021/ACSAMI.7B01016/SUPPL_FILE/AM7B01016_SI_002.AVI
- Xiao, J., Wang, F., Wong, N. K., He, J., Zhang, R., Sun, R., et al. (2019). Global liver disease burdens and research trends: Analysis from a Chinese perspective. *Journal of Hepatology*, 71(1), 212–221. <https://doi.org/10.1016/J.JHEP.2019.03.004>
- Xiaoyan, Z., Yang, Z., Gai, G., & Yang, Y. (2009). Effect of superfine grinding on properties of ginger powder. *Journal of Food Engineering*, 91(2), 217–222. <https://doi.org/10.1016/J.JFOODENG.2008.08.024>
- Xie, L., Huang, Z., Meng, H., Shi, X., & Xie, J. (2022). Immunomodulation effect of polysaccharides from liquid fermentation of *Monascus purpureus* 40269 via membrane TLR-4 to activate the MAPK and NF- κ B signaling pathways. *International Journal of Biological Macromolecules*, 201, 480–491. <https://doi.org/10.1016/J.IJBIOMAC.2022.01.045>
- Xiong, Z. Y., Xiao, F. M., Xu, X., Wu, Y. F., & Jiang, X. M. (2013). Studies on pharmacological activity of borneol. *China Journal of Chinese Materia Medica*, 38(6), 786–790. <https://doi.org/10.4268/cjcm20130602>
- Ya Ling, H., Sheu, F., Lee, M. H., & Chau, C. F. (2008). Effects of particle size reduction of insoluble fibres by micron technology on various caecal and faecal indices. *Journal of the Science of Food and Agriculture*, 88(3), 435–441. <https://doi.org/10.1002/J.SFA.3104>
- Yahaya, W. A. W., Almajano, M. P., Yazid, N. A., & Azman, N. A. M. (2019). Antioxidant activities and total phenolic content of Malaysian herbs as components of active packaging film in beef patties. *Antioxidants*, 8(7), 204. <https://doi.org/10.3390/antiox8070204>
- Yahaya, W. A. W., Husaini, N. H., Zaharudin, F. N. H., & Azman, N. A. M. (2020). Characterization of semi-refined carrageenan film plasticized with glycerol incorporated with *Piscaria minor* extract as antioxidant additives. *IOP Conference Series: Materials Science and Engineering*, 736(2), Article 022012. <https://doi.org/10.1088/1757-899X/736/2/022012>
- Yahya, N.N.H. (2017). Preliminary screening on the effect of piscaria minor (kesum) against haemochus contortus (p. 1-10). Universiti Teknologi MARA, Negeri Sembilan. 2017.
- Yahya, H. M., Shahar, S., Arina Ismail, S. N., Aziz, A. F., Din, N. C., & Abdul Hakim, B. N. (2017). Mood, cognitive function and quality of life improvements in middle aged women following supplementation with *Polygonum minus* extract. *Sains Malaysiana*, 46(2), 245–254. <https://doi.org/10.17576/jsm-2017-4602-09>
- Yang, I. J., Lee, D. U., & Shin, H. M. (2016). Inhibitory effect of valencene on the development of atopic dermatitis-like skin lesions in NC/Nga mice. *Evidence-based Complementary and Alternative Medicine*, 2016, 1–11. <https://doi.org/10.1155/2016/9370893>
- Yang, D., Michel, L., Chaumont, J. P., & Millet-Clerc, J. (2000). Use of caryophyllene oxide as an antifungal agent in an *in vitro* experimental model of onychomycosis. *Mycopathologia*, 148(2), 79–82. <https://doi.org/10.1023/A:1007178924408>
- Yeung, W. F., Chung, K. F., Ng, K. Y., Yu, Y. M., Ziea, E. T. C., & Ng, B. F. L. (2014). A systematic review on the efficacy, safety and types of Chinese herbal medicine for depression. *Journal of Psychiatric Research*, 57(1), 165–175. <https://doi.org/10.1016/J.JPSYCHIRES.2014.05.016>
- Yi, L., Cao, J., Cao, H., & Xiao, J. (2019). Report of the 3rd international symposium on phytochemicals in medicine and food (August 25–30th, 2018, Kunming, China). *Food Chemistry*, 289, 671–672. <https://doi.org/10.1016/J.FOODCHEM.2019.03.104>
- Ying, Y. M., Fang, C. A., Yao, F. Q., Yu, Y., Shen, Y., Hou, Z. N., et al. (2017). Bergamotane sesquiterpenes with alpha-glucosidase inhibitory activity from the plant pathogenic fungus *Penicillium expansum*. *Chemistry and Biodiversity*, 14(1), Article e1600184. <https://doi.org/10.1002/CBDV.201600184>
- You, Y. X., Shahar, S., Haron, H., & Mastura Yahya, H. (2018). More ulam for your brain: A review on the potential role of ulam in protecting against cognitive decline. *Sains Malaysiana*, 47(11), 2713–2729. <https://doi.org/10.17576/jsm-2018-4711-15>
- Yu, C., Guo, D., Yao, C., Zhu, Y., Liu, S., & Kong, X. (2021). Development and validation of a nomogram for predicting drug-induced acute kidney injury in hospitalized patients: A case-control study based on propensity-score matching. *Frontiers in Pharmacology*, 12, Article 657853. <https://doi.org/10.3389/FPHAR.2021.657853>
- Yusuf, S.B. (2014). Antioxidative properties of selected Malaysian herbal plants. Universiti Putra Malaysia (p. 1-82). 2014.
- Zahin, M., Ahmad, I., & Aqil, F. (2017). Antioxidant and antimutagenic potential of *Psidium guajava* leaf extracts. *Drug and Chemical Toxicology*, 40(2), 146–153. <https://doi.org/10.1080/01480545.2016.1188397>
- Zakaria, N., Baba, S., Ku Bahaudin, K. N. A., & Hamdana, S. (2015). Total phenolic content and antioxidant activity of pure and formulated extracts of kesum (*Polygonum minus*), bawang putih (*Allivium sativum*), pegaga (*Centella asiatica*), and ulam raja (*Cosmos caudatus*). *Malaysian Journal of Fundamental and Applied Sciences*, 11(4), 179–183. <https://doi.org/10.11113/mjfas.v11n4.399>
- Zamri, N. A. F. M. (2015). *Evaluation of total phenolic content and antibacterial activity of polygonum minus leaves methanolic extract against food borne pathogens*. Universiti Teknologi MARA.
- Zhang, W., Roy, S., Ezati, P., Yang, D. P., & Rhim, J. W. (2023). Tannic acid: A green crosslinker for biopolymer-based food packaging films. *Trends in Food Science & Technology*, 136, 11–23. <https://doi.org/10.1016/J.TIFS.2023.04.004>
- Zhang, L., Ye, M., Shi, Y., Zhu, H., Chi, L., Pan, C., et al. (2021). Phytochemical components and biological activities of essential oils from three selected medicinal plants. *Industrial Crops and Products*, 160, Article 113127. <https://doi.org/10.1016/j.indcrop.2020.113127>
- Zhao, P., Li, N., & Astruc, D. (2013). State of the art in gold nanoparticle synthesis. *Coordination Chemistry Reviews*, 257(3–4), 638–665. <https://doi.org/10.1016/J.CCR.2012.09.002>
- Zhao, Y., Zhu, K., Li, J., Zhao, Y., Li, S., Zhang, C., et al. (2021). High-efficiency production of bisabolene from waste cooking oil by metabolically engineered *Yarrowia lipolytica*. *Microbial Biotechnology*, 14(6), 2497–2513. <https://doi.org/10.1111/1751-7915.13768>
- Zhuang, X., Köllner, T. G., Zhao, N., Li, G., Jiang, Y., Zhu, L., et al. (2012). Dynamic evolution of herbivore-induced sesquiterpene biosynthesis in sorghum and related grass crops. *The Plant Journal*, 69(1), 70–80. <https://doi.org/10.1111/j.1365-3113.2011.04771.x>
- Zulfar, M.R.A.A. (2017). Effect of *Polygonum minus* leaves ethanol extract as a preventive against mercuric chloride toxicity in histological features of mice (*Mus musculus*) (p. 1). Universitas Airlangga. 2017.