



**EDIBLE BIRD'S NEST TREATMENT EFFECTS ON SUBCHONDRAL BONE  
AND ARTICULAR CARTILAGE CHANGES AND SYNOVIAL FLUID  
PROTEOME PROFILES IN AN OSTEOARTHRITIS  
RABBIT MODEL**

**By**

**SHARIFAH ZAKIAH BINTI SYED SULAIMAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirement for the Degree of Doctor of Philosophy**

**June 2022**

**FPV 2022 31**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
Fulfilment of the requirement for the Degree of Doctor of Philosophy

**EDIBLE BIRD'S NEST TREATMENT EFFECTS ON SUBCHONDRAL BONE  
AND ARTICULAR CARTILAGE CHANGES AND SYNOVIAL FLUID  
PROTEOME PROFILES IN AN OSTEOARTHRITIS  
RABBIT MODEL**

By

**SHARIFAH ZAKIAH BINTI SYED SULAIMAN**

**June 2022**

**Chairman : Assoc. Prof. Lau Seng Fong, PhD**  
**Faculty : Veterinary Medicine**

Osteoarthritis (OA) is characterised by progressive degeneration of articular cartilage, subchondral bone changes and synovium inflammation. Animal models are important and can demonstrate different features depending on method of induction which can resemble primary and secondary OA in humans. The choice of animal models used is based on the type and duration of research and outcome measurements. Currently, the goals for OA treatment mainly focus to alleviate disease signs and symptoms. Numerous natural products including Edible Bird's Nest (EBN) have been studied extensively in order to create potential therapies for inflammatory disorders such as arthritis. Previous studies have reported the antioxidative, anti-inflammatory and bone-strengthening effects of EBN. This study is divided into two parts. The first study is aimed to compare subchondral bone and articular cartilage changes and proteome profiles for surgically induced and chemically induced rabbit model of osteoarthritis at different time points. Based on the first study, more suitable model is chosen for the second study which is aimed to observe the effects of EBN in ameliorating OA development at different time points. For the first part of the study, New Zealand white rabbits underwent either anterior cruciate ligament transection (ACLT) procedure or injected intra-articularly with monosodium iodoacetate (MIA, 8 mg) into the right knee and were further divided into week 4 (n=5), week 8 (n=5) and week 12 (n=5) groups. The joints were subjected to micro-computed tomographic (micro-CT) analysis and histological evaluation. The synovial fluid were subjected to MALDI TOF/TOF analysis. Bone volume over tissue volume for surgically induced group increased in femur (35.35%) and tibia (32.82%) during week 12 which suggested bone remodelling whereas chemically induced group showed persistent bone resorption with decreased value of BV/TV in femur (27.12%) and tibia (26.82%). For histopathological grading of articular cartilage in femur and tibia, surgically induced group showed minimal changes during week 12 with median values of 1 and 2 in femur and tibia, respectively. As for chemically induced group, more severe

changes were recorded with median values of 4.5 and 5 with significant difference with control group ( $p=0.0184, 0.0208$ ) in femur and tibia, respectively. Micro-CT and histopathological analysis revealed that subchondral bone remodelling precede articular cartilage damage in surgically induced group, and vice versa in chemically induced group. Proteome profiles showed peak OA progression during week 12 for surgically induced group with upregulation of gelsolin and serotransferrin protein which involved in advanced OA. For chemically induced group, OA progression is the highest during early stage indicated by high upregulation of apolipoprotein I-IV precursor and serpin peptidase inhibitor protein during week 4 but were later downregulated. The results showed different pathogenic mechanisms for both induction method. For the second part of the study, New Zealand white rabbits were chemically induced (MIA, 8 mg) and divided into four groups; (1) negative control (n=9): non-treated osteoarthritis, (2) positive control (n=15): OA + diclofenac sodium 2 mg/kg daily orally, (3) low dosage (n=15): OA + 75 mg/kg hydrolysed EBN, (4) high dosage (n=15): OA + 150 mg/kg hydrolysed EBN. The joints were subjected to micro-CT analysis and histological evaluation and the synovial fluid subjected to LCMS/MS analysis. Micro-CT analysis showed a 22% increase in BV/TV value and 10% decrease in total porosity (PO) in treatment group that showed bone integrity improvement. Histopathological results revealed comparable changes between positive control group and EBN treatment group. There was upregulation of annexin-1, a protein involved in resolution of inflammation and downregulation carbonic anhydrase II, a protein associated with bone resorption process. Overall, morphology evaluation showed that EBN supplementation can inhibit osteoclastic activity but have no effect on cartilage damage. Protein expression showed action of several proteins involved in bone resorption inhibition and resolution of inflammation via several signalling pathways which includes toll-like receptor (TLR) and NF- $\kappa$ B signalling pathways.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**KESAN RAWATAN SARANG BURUNG WALIT DALAM PERUBAHAN  
TULANG SUBKONDRAL DAN RAWAN ARTIKULAR DAN PENGKELASAN  
PROFIL PROTEIN DALAM MODEL ARNAB OSTEOARTRITIS**

Oleh

**SHARIFAH ZAKIAH BINTI SYED SULAIMAN**

**June 2022**

**Pengerusi : Prof. Madya Lau Seng Fong, PhD**  
**Fakulti : Perubatan Veterinar**

Osteoarthritis dikenalpasti melalui kemerosotan progresif rawan artikular, perubahan tulang subkondral dan keradangan sinovium. Model haiwan adalah penting dan boleh menunjukkan ciri-ciri tertentu bergantung kepada kaedah induksi yang boleh menyerupai osteoarthritis jenis primer atau sekunder dalam manusia. Pemilihan model haiwan adalah bergantung kepada jenis dan tempoh masa kajian dan juga hasil yang ingin dikenalpasti. Pada masa ini, tujuan rawatan OA lebih terfokus kepada pengurangan simptom dan kesan penyakit. Beberapa produk semulajadi termasuk sarang burung walit (EBN) telah dikaji secara mendalam dan berpotensi untuk rawatan gangguan keradangan seperti penyakit artritis. Kajian lepas telah melaporkan kesan EBN seperti anti-oksidatif, anti-radang dan penguatan tulang. Kajian ini dibahagikan kepada dua bahagian. Kajian pertama bertujuan untuk membuat perbandingan diantara kaedah induksi secara pembedahan dan induksi secara bahan kimia untuk menghasilkan model arnab osteoarthritis dalam beberapa tempoh masa yang berbeza. Seterusnya, berdasarkan hasil kajian bagi kajian pertama, model induksi yang lebih sesuai dipilih untuk kajian bahagian kedua yang bertujuan untuk melihat kesan EBN dalam memperbaiki perkembangan OA juga dalam beberapa tempoh masa yang berbeza. Untuk kajian bahagian pertama, arnab putih New Zealand menjalani sama ada pembedahan transeksi ligament krusiat anterior (ACLT) atau disuntik secara intrasendi dengan monosodium iodoacetate (MIA, 8mg) di lutut kanan dan dibahagikan lagi kepada kumpulan minggu 4 (n=5), minggu 8 (n=5) dan minggu 12 (n=5). Tulang sendi yang diperolehi akan melalui proses mikro-CT dan penilaian histologi. Cecair sinovial akan dianalisa menggunakan MALDI TOF/TOF. Nisbah isipadu tulang dan isipadu tisu (BV/TV) dalam kumpulan induksi secara pembedahan menunjukkan peningkatan di femur (35.35%) dan tibia (32.82%) pada minggu ke-12 yang mencadangkan permodelan semula tulang manakala kumpulan induksi secara bahan kimia menunjukkan penyerapan semula tulang yang konsisten diwakili oleh pengurangan nilai BV/TV dalam femur (27.12%) dan tibia (26.82%). Bagi penskoran histopatologi rawan artikular dalam femur dan tibia, kumpulan induksi secara

pembedahan menunjukkan perubahan yang tidak ketara dengan nilai median 1 dan 2 dalam femur dan tibia pada minggu ke-12. Bagi kumpulan induksi secara bahan kimia, perubahan yang lebih ketara telah direkodkan dengan nilai median 4.5 dan 5 yang mempunyai perubahan signifikan berbanding kumpulan kawalan ( $p=0.0184, 0.0208$ ) dalam femur dan tibia. Analisis mikro-CT dan histopatologi menunjukkan bagi kumpulan induksi secara pembedahan, permodelan semula tulang berlaku sebelum kerosakan rawan artikular dan perkara sebaliknya berlaku dalam kumpulan induksi secara bahan kimia. Profil protein menunjukkan progres OA yang tertinggi berlaku ketika minggu ke-12 bagi kumpulan induksi secara pembedahan dengan peningkatan ekspresi protein gelsolin dan serotransferrin yang terlibat dalam OA peringkat akhir. Bagi kumpulan induksi secara bahan kimia, progress OA adalah yang tertinggi ketika peringkat awal yang ditunjukkan oleh peningkatan ekspresi protein apolipoprotein I-IV precursor dan serpin peptidase inhibitor pada minggu ke-4 namun kemudiannya menurun. Hasil kajian menunjukkan mekanisme patogenik yang berbeza bagi kedua-dua kaedah induksi. Untuk kajian bahagian kedua, arnab putih New Zealand disuntik secara intrasendi dengan monosodium iodoacetate (MIA, 8mg) dan dibahagikan kepada empat kumpulan; (1) kawalan negatif ( $n=9$ ): osteoarthritis yang tidak dirawat, (2) kawalan positif ( $n=15$ ): OA + diclofenac sodium 2mg/kg setiap hari secara oral, (3) dos rendah ( $n=15$ ): OA + 75 mg/kg hydrolysed EBN, (4) dos tinggi ( $n=15$ ): OA + 150 mg/kg hydrolysed EBN. Tulang sendi yang diperolehi akan melalui proses mikro-CT dan penilaian histologi manakala cecair sinovial akan dianalisa menggunakan LCMS/MS. Analisis mikro-CT menunjukkan peningkatan isipadu tulang sebanyak 22% dan penurunan jumlah keliangan tulang sebanyak 10% dalam kumpulan rawatan yang menunjukkan kesan EBN dalam peningkatan kekuatan tulang. Hasil analisis histopatologi menunjukkan hasil yang agak sama bagi kumpulan kawalan positif dan juga kumpulan rawatan EBN. Terdapat peningkatan ekspresi protein annexin-1 yang terlibat dalam resolusi keradangan dan pengurangan ekspresi protein carbonic anhydrase II yang terlibat dalam proses penyerapan tulang. Penilaian secara morfologi menunjukkan penambahan EBN merencatkan aktiviti osteoklas namun tidak mempunyai apa-apa kesan terhadap kerosakan rawan. Ekspresi protein menunjukkan kesan beberapa protein yang terlibat dalam perencatan proses penyerapan tulang dan resolusi keradangan melalui beberapa laluan pengisyaratan yang termasuk laluan pengisyaratan toll-like receptor (TLR) dan NF- $\kappa$ B.

## ACKNOWLEDGEMENTS

Alhamdulillah, thanks to Allah Almighty, I finally successfully managed to complete this thesis. First and foremost, I would like to express my sincere gratitude to my supervisor Associate Professor Dr. Lau Seng Fong for the continuous support of my PhD study and research, for her patience, motivation, enthusiasm, and immense knowledge. Her guidance helped me in all the time of research and writing of this thesis. I could not have imagined having a better advisor and supervisor for my PhD study. My utmost appreciation also to my co-supervisors, Dr. Mokrish, Prof. Suhaila, Dr. Rozaihan and Dr. Angela for their expertise and guidance throughout this research.

My sincere thanks also goes to to En. Azmil and En. Hilman from ARF for help during animal study, Puan Norasfaliza (ABI), Dr. Benjamin Lau (MPOB) and Puan Asikin (MPOB) for assisting in MALDI TOF/TOF and LCMS/MS during my proteomics study, Dr. Mohd Fauzi Md Busra for assisting in the use of micro-CT analysis, Puan Latifah Mohd Hanan and Puan Zainatulaishah Manap for preparing the histological slides, En. Hasmadi from Anatomy Laboratory, En. Ghazali from Post-mortem Laboratory and En. Jamil from Makmal Peralatan for help during my labwork.

I am also indebted to my beloved labmates and fellow postgraduates, Dr. Murshidah Mohd Asri, Dr. Amirul Nazhan, Dr. Rachel Tan, Nor Hashikin, Fadzly Salleh, Danish Adli, Dr. Mohd Akmal, Nor Shahirah Solehah, members of Biochemistry Lab, members of Nutrition Lab and my other postgraduate friends for all the fun that we had which make my PhD journey memorable. I thank all for all of our discussion and brainstorming session and I will also cherish our funny as well as awkward moments, our '*makan-makan*' session, and other activities that we spend together and hope that we can cross path again in the future.

A special appreciation to my family, Syed Sulaiman, Sharifah Zaleha, Sharifah Khadijah and Syed Alwi, and my late grandmother Sharifah Zainap for their unconditional love and moral support. I would also like to thank my beloved husband, Qayyum Latip for the never ending support and motivation during this final stage. Once again, thank you so much to everyone for all of your help and support.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Lau Seng Fong, PhD**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Mohd Mokrish bin Md. Ajat, PhD**

Senior Lecturer  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

**Rozaihan binti Mansor, PhD**

Senior Lecturer  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

**Suhaila binti Mohamed, PhD**

Professor  
Institute of Bioscience  
Universiti Putra Malaysia  
(Member)

**Angela Ng Min Hwei, PhD**

Associate Professor  
Tissue Engineering Centre  
Universiti Kebangsaan Malaysia Medical Centre  
(Member)

---

**ZURIATI AHMAD ZUKARNAIN, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 9 February 2023



## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Sharifah Zakiah Binti Syed Sulaiman

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiv
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF APPENDICES</b>	xviii
<b>LIST OF ABBREVIATIONS</b>	xix
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Background and hypothesis	1
1.2 Problem statement and justification	2
1.3 Research objectives	2
1.4 Summary of the chapters	3
<b>2 LITERATURE REVIEW</b>	<b>4</b>
2.1 General overview of stifle joint	4
2.1.1 Anatomy, structure, and function	4
2.1.2 Synovial fluid	5
2.1.3 Articular cartilage	6
2.1.4 Subchondral bone	7
2.2 Osteoarthritis	8
2.2.1 Definition of OA	8
2.2.2 Prevalence of OA	8
2.2.3 Risk Factors of OA	8
2.2.4 OA pathogenesis	10
2.3 Diagnostic modalities of osteoarthritis	14
2.3.1 Diagnostic imaging	14
2.3.2 Diagnostic pathology	15
2.4 Current treatment	17
2.5 Animal mode	21
2.5.1 Classification of OA animal models	21
2.5.2 Common animals models used in OA	22

2.6	Method of OA induction	24
2.6.1	Surgically induced model	24
2.6.2	Chemically induced model	24
2.7	Proteomics studies	25
2.7.1	Technological basis of proteomics	25
2.8	Applications of proteomics	28
2.8.1	Diagnostic applications	28
2.8.1	Therapeutics strategies	29
2.9	Proteomics in OA	29
2.9.1	OA biomarkers	30
2.9.2	Past, current, and future studies of proteomics in OA	30
2.10	Edible bird's nest (EBN)	31
2.10.1	Composition of EBN	31
2.10.2	EBN as treatment	32
2.10.3	Hydrolysate EBN	33
<b>3</b>	<b>EVALUATION OF SUBCHONDRAL BONE AND ARTICULAR CARTILAGE TEMPORAL CHANGES IN SURGICALLY AND CHEMICALLY INDUCED OSTEOARTHRITIS RABBIT MODEL</b>	<b>34</b>
3.1	Introduction and objective	34
3.2	Materials and methods	35
3.2.1	Animals	35
3.2.2	Preparation of animal model of osteoarthritis	35
3.2.3	Sample collection	36
3.2.4	Micro-computed tomography evaluation	36
3.2.5	Histology	37
3.2.6	Statistical analysis	37
3.3	Results	38
3.3.1	General condition of rabbit models	38
3.3.2	Visual inspection of the joint and gross anatomy of the femoral condyle and tibial plateau	38
3.3.3	Two-dimensional micro-CT images assessment	41
3.3.4	Quantitative bone micro-architecture assessment	42
3.3.5	Histopathological grading and statistical analysis	46
3.4	Discussion	52

<b>4</b>	<b>CHARACTERISATION OF SYNOVIAL FLUID PROTEINS IN SURGICALLY INDUCED AND CHEMICALLY INDUCED OSTEOARTHRITIS RABBIT MODEL</b>	<b>56</b>
4.1	Introduction and objective	56
4.2	Materials and methods	57
4.2.1	Animals	57
4.2.2	Samples collection	57
4.2.3	Protein precipitation from synovial fluid	57
4.2.4	Protein concentration estimation	57
4.2.5	Isoelectric focusing	58
4.2.6	Two-dimensional gel electrophoresis (2DGE)	58
4.2.7	Protein digestion and matrix-assisted laser desorption/ionization time of flight/time of flight mass spectrometry (MALDI-TOF/TOF)	58
4.3	Results	59
4.3.1	General condition of rabbit models	59
4.3.2	Evaluation of protein yield and two-dimensional gel electrophoresis analysis	59
4.3.3	Identification of proteins spots	60
4.4	Discussion	62
<b>5</b>	<b>COMPARISON OF SUBCHONDRAL BONE AND ARTICULAR CARTILAGE CHANGES IN BETWEEN EDIBLE BIRD'S NEST TREATED GROUP AND NON-TREATED GROUP OF OSTEOARTHRITIC RABBITS</b>	<b>70</b>
5.1	Introduction and objective	70
5.2	Materials and methods	71
5.2.1	Animals and grouping	71
5.2.2	Preparation of EBN hydrolysate	71
5.2.3	Sample collection	72
5.2.4	Micro-computed tomography evaluation	72
5.2.5	Histology	72
5.2.6	Statistical analysis	73
5.3	Results	73
5.3.1	General condition of rabbit models	73
5.3.2	Gross anatomy of the femoral condyle	73
5.3.3	Quantitative bone micro-architecture assessment	75
5.3.4	Histopathological grading and statistical analysis	78
5.4	Discussion	81

<b>6</b>	<b>DIFFERENTIAL PROTEIN EXPRESSION OF SYNOVIAL FLUID IN BETWEEN EDIBLE BIRD'S NEST TREATED GROUP AND NON-TREATED GROUP OF OSTEOARTHRITIC RABBITS</b>	<b>83</b>
6.1	Introduction and objective	83
6.2	Materials and methods	85
6.2.1	Animals and grouping	85
6.2.2	Sample collection	85
6.2.3	Synovial fluid preparation	85
6.2.4	Protein precipitation from synovial fluid	86
6.2.5	Protein digestion	86
6.2.6	Protein clean up and elution	86
6.2.7	Liquid chromatography-tandem mass spectrometry	86
6.2.8	Protein quantification and data analysis	87
6.3	Results	87
6.3.1	General condition of rabbit model	87
6.3.2	Comparative proteome profiling between EBN treated and non-treated group	87
6.4	Discussion	97
<b>7</b>	<b>SUMMARY, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>102</b>
7.1	Summary and conclusions	102
7.2	Limitations and recommendations for future research	103
	<b>REFERENCES</b>	<b>105</b>
	<b>APPENDICES</b>	<b>132</b>
	<b>BIODATA OF STUDENT</b>	<b>147</b>
	<b>LIST OF PUBLICATIONS</b>	<b>148</b>

## LIST OF TABLES

Tables		Page
1	Kellgren and Lawrence OA grading guidelines.	14
2	Modified Outerbridge scoring.	15
3	The OARSI Cartilage Histopathology Grading System.	16
4	Micro-architecture parameters of the subchondral bone of right stifle joints of femur obtained from surgically and chemically induced OA in rabbit model.	43
5	Micro-architecture parameters of the subchondral bone of right stifle joints of tibia obtained from surgically and chemically induced OA in rabbit model.	43
6	OARSI histopathological grade of femur and tibia in the control, and at week 4, 8, and 12 for the surgically induced and chemically induced group	48
7	List of proteins and their log normalized volume for control, week 4, week 8 and week 12 for both induction groups.	61
8	Micro architectural parameters of the subchondral bone of right stifle joints of femur for negative control, positive control, high dosage EBN treatment and low dosage EBN treatment groups for week 4, week 8 and week 12.	76
9	Result of OARSI histopathological grade for negative control, positive control, low dosage EBN treatment and high dosage EBN treatment group for week 4, week 8 and week 12.	79
10	Proteins with statistically significant upregulation and downregulation in control versus treatment samples	92

## LIST OF FIGURES

Figure		Page
1	Anatomy of stifle joint.	4
2	Composition and structure of articular cartilage.	6
3	Signalling pathways and molecular component involved in OA development.	13
4	Flow of tandem mass spectrometry.	27
5	Gross morphology observation of the femoral condyle of surgically and chemically induced groups for control, week 4, week 8 and week 12 groups.	39
6	Gross morphology observation of the tibial plateau of surgically and chemically induced groups for control, week 4, week 8 and week 12 groups.	40
7	Two-dimensional micro-CT scans of dorsal and sagittal planes of the surgically and chemically induced group.	41
8	Comparison of the bone micro-architecture of the subchondral bone of right stifle joints of the femur between different type for OA induction for control, week 4, week 8, and week 12 groups.	44
9	Comparison of the bone micro-architecture of the subchondral bone of right stifle joints of the tibia between different type for OA induction for control, week 4, week 8, and week 12 groups.	45
10	OARSI histological scores for (A) surgically induced femoral cartilage (B) chemically induced femoral cartilage (C) surgically induced tibial cartilage (D) chemically induced tibial cartilage.	49
11	Representative histology images of femoral articular cartilage. A-D: surgically induced group. E-H: chemically induced group.	50
12	Representative histology images of tibial articular cartilage. A-D: surgically induced group. E-H: chemically induced group.	51

13	Representative 2DGE (pI 3-10) image for rabbit synovial fluid at week 8 in (A) surgically induced group and (B) chemically induced group after staining with colloidal Coomassie stain.	60
14	Characterization of proteins according to biological process functions for surgically induced group.	62
15	Characterization of proteins according to biological process functions for chemically induced group.	62
16	Proposed pathway of surgically induced OA (ACLT) induction mechanism based on proteins identified from MALDI TOF/TOF.	66
17	Proposed pathway of chemically induced OA (MIA) mechanism based on proteins identified from MALDI TOF/TOF.	68
18	Gross morphology observation of the femoral condyle of negative control, positive control, low dosage EBN treatment and high dosage EBN treatment groups for week 4, week 8 and week 12.	74
19	Progression of the bone micro architecture of subchondral bone of negative control, positive control, low dosage EBN treatment and high dosage EBN treatment group for week 4, week 8 and week 12.	77
20	OARSI histological scores for negative control, positive control, low dosage EBN treatment and high dosage EBN treatment group for week 4, week 8 and week 12.	79
21	Representative histology images of femoral articular cartilage for negative control, positive control, low dosage EBN treatment and high dosage EBN treatment group for week 4, week 8 and week 12.	80
22	Principal component analysis plot representing proteomics data from the differential expression analysis of control and treatment group.	88
23	Protein from control and treatment group in week 4, week 8 and week 12 represented in the form of heat map with hierarchical clustering showing the expression levels of differentially expressed proteins as colours.	89



24	Gene Ontology (GO) domain overview of all identified proteins.	91
25	Protein-protein interaction network of treatment effects.	96
26	Proposed pathway of Edible Bird's Nest (EBN) supplementation in ameliorating osteoarthritis (OA) in monosodium iodoacetate (MIA) induced rabbits.	101



## LIST OF APPENDICES

Appendix		Page
A	Rabbits' body weight (kg)	132
B	Visual observation of rabbits' joint from surgically and chemically induced group.	135
C	Protein concentration of chemically and surgically induced groups.	136
D	Surgically induced group two-dimensional gel from four different time groups.	137
E	Chemically induced group two-dimensional gel from four different time groups.	138
F	Histology Protocol (Safranin O).	139
G	Reagents for MALDI TOF/TOF analysis.	141
H	Reagents for LCMS/MS analysis.	146

## LIST OF ABBREVIATIONS

2DGE	Two-dimensional gel electrophoresis
ACLT	Anterior cruciate ligament transection
ADAMTS	A disintegrin and metalloproteinase with thrombospondin motifs
AGE	Advanced glycation end-products
ANOVA	Analysis of variance
BS/BV	Bone surface to volume ratio
BS/TV	Bone surface density
BSA	Bovine serum albumin
BV/TV	Bone volume over tissue ratio
COMP	Serum cartilage oligomeric protein
COX2	Cyclooxygenase-2
CT	Computed tomography
DTT	Dithiothreitol
FCF	Fast green
GAG	Glycosaminoglycan
GDF5	Growth differentiation factor 5
HA	Hyaluronan
HCl	Hydrochloric acid
HIF	Hypoxia inducible factor
HMGB-1	High mobility group box-1
IAA	Iodoacetamide
IACUC	Institutional Animal Care and Use Committee

IGF-1	Insulin growth factor-1
IL	Interleukin
IL-1 $\beta$	Interleukin-1-beta
JSN	Joint space narrowing
JSW	Joint space width
kDA	Kilo Dalton
LCMS/MS	Liquid chromatography mass spectrometry
m/z	Mass over charge
MALDI	Matrix-assisted laser desorption/ionization
MAPK	Mitogen activated protein kinase
MIA	Monosodium iodoacetate
MMP	Matrix metalloproteinases
MRI	Magnetic resonance imaging
MS	Mass spectrometry
MW	Molecular weight
NF- $\kappa$ B	Nuclear factor kappa B
NO	Nitric oxide
NOS	Nitric oxide synthase
NSAIDs	Nonsteroidal anti-inflammatory drugs
OA	Osteoarthritis
OARSI	Osteoarthritis Research Society International
OCT	Optical coherence tomography
OPG	Osteoprotegerin
PG	Proteoglycan

PGE2	Prostaglandin E2
pH	Potential of hydrogen
pI	Isoelectric point
PO	Total porosity
RAGE	Receptor advanced glycation end-products
RANK	Receptor activator of nuclear factor kappa-B
RANKL	Receptor activator of nuclear factor kappa-B ligand
ROS	Reactive oxygen species
SD	Standard deviation
Tb.Sp	Trabecular space
Tb.Th	Trabecular thickness
TCA	Trichloroacetic acid
TGF	Transforming growth factor
TLR	Toll-like receptor
TNF	Tumour necrosis factor
TNF- $\alpha$	Tumor necrosis factor alpha
TOF	Time-of-flight
US	Ultrasound
VEGF	Vascular endothelial growth factor
VOI	Volume of interest
w/v	Weight over volume
WHO	World Health Organisation

# CHAPTER 1

## INTRODUCTION

### 1.1 Background and hypothesis

Osteoarthritis (OA) is characterised by degeneration of the articular cartilage, changes in the subchondral bone, and inflammation of the synovium (Li et al., 2013). OA affects up to 15% of the global human population (Johnson & Hunter, 2014) and about 20% of the canine population over the age of a year (Johnson, 2015). Since one of the traditional OA characteristics is articular cartilage degradation, OA was previously thought to be a disease affecting only articular cartilage (Buckwalter & Mankin, 1998; Pitsillides & Beier, 2011). However, recent findings suggested that subchondral bones are also importantly involved in OA pathogenesis (Mahjoub et al., 2012; Peters et al., 2018). Subchondral bones and articular cartilage are complementarily involved in biomechanical load-bearing joints (Li et al., 2013).

OA is characterised into primary (idiopathic) OA which is due to degenerative changes at the joint, mainly because of aging and secondary OA which related to risk factors that will cause OA such as obesity, joint injury, trauma and congenital disease (Chen et al., 2017). Many factors contribute to the onset of OA, including biomechanical, biochemical, genetics and also biomolecular and signalling feedback mechanisms (Shirai et al., 2011). Osteoarthritic changes are caused by cartilage matrix degradation, which culminates from disruption in the bone remodelling phase, causing further catabolic processes to outnumber anabolic processes, resulting in articular cartilage degradation, osteophytosis and functional limitations (Permuy et al., 2015).

In understanding this disease, animal models serve an important complement and simulation for human OA studies. Several small and large animal models such as horses, dogs and rabbits were developed and being used to study OA, specifically for articular cartilage and subchondral bone changes. The choice of animal models used is depending on the type and duration of research, husbandry costs and measurements of the outcome (Kuyinu et al., 2016b). While different induction method proved to successfully develop OA, the pathogeneses and progression are different in each of the model. The difference in time-dependent progression between induction methods is suggested to be caused by different pathogenesis between two types of OA. In depth knowledge of the different pathophysiology of the disease are important in planning targeted therapeutic strategies and specifically monitor disease and treatment outcome.

Current OA medications are palliative in nature, with the aim of reducing pain, which is the disease's primary symptom. Intra-articular injections of hyaluronic acids and steroids, as well as administration of nonsteroidal anti-inflammatory drugs (NSAIDs) are a few treatment strategies used in addressing this disease (Guo et al., 2021). However, these treatments come with several adverse effects such as renal toxicity, diarrhoea, vomiting, gastrointestinal disturbances, nausea, or increased cardiovascular risks (Crofford, 2013).

For the past decades, natural products are gaining a lot of attention as an alternative remedy for a variety of health problems because of their high level of effectiveness and low adverse effects (Goudarzi et al., 2018). Numerous traditional natural products including Edible Bird's Nest (EBN) have been studied extensively in order to create potential therapies for inflammatory disorders such as arthritis (Kang et al., 2019). EBN is associated with wide range of high nutritional values and health benefits such as anti-inflammatory, anti-oxidative, chondroprotective and bone strengthening effects. Therefore, EBN are hypothesized to exhibit anti-inflammatory, chondroprotective and bone strengthening effect in OA, and to reduce pain, cartilage degradation and subchondral bone alteration.

## **1.2 Problem statement and justification**

OA is associated with high economy burden, largely attributable to the effects of disability, comorbid disease, and the expense of treatment. Additionally, OA is usually diagnosed at advanced stages where joint damage already immense and irreversible. Currently, there are no cost-effective diagnostic, treatment and prognostic method for OA in both human and veterinary medicine.

Typically, most OA studies focused only on articular cartilage changes and there is a data paucity regarding the evaluation of changes in both articular cartilage and subchondral bones for different induction methods at different time. This information is useful in understanding the pathophysiology of the disease and subsequently planning targeted treatment strategies and to specifically monitor disease and treatment outcome.

The available treatment of osteoarthritis available is based on reducing symptoms, recover function and delay the progression of the disease (Michael et al. 2010). Up to the present, it can be deduced that the treatments for OA are limited, and that the outcomes are poor due to the high occurrence of adverse side effects. The potential of new, novel non-surgical treatments approach for OA is reassuring and exciting to be explored and natural products such as EBN possess many health benefits that have the potential in ameliorating OA.

## **1.3 Research objectives**

- i. To evaluate the temporal changes of subchondral bone and articular cartilage in surgically and chemically induced osteoarthritis rabbit model using micro-computed tomography (micro-CT) and histology.
- ii. To compare proteome profiles in surgically and chemically induced osteoarthritis rabbit model.
- iii. To evaluate the effects of Edible Bird's Nest (EBN) treatment on subchondral bone and articular cartilage of osteoarthritic rabbit model using micro-computed tomography (micro-CT) and histology.
- iv. To compare differential expression of proteins in synovial fluid of Edible Bird's Nest (EBN) treated group and non-treated group in osteoarthritic rabbit model.

#### 1.4 Summary of the chapters

- Chapter 2 described a review on osteoarthritis, diagnostic methods, current treatment, proteomics research, Edible Bird's Nest and other related topics to increase understanding of this study.
- Chapter 3 evaluated the changes in subchondral bone and articular cartilage in surgically induced and chemically induced OA rabbit model.
- Chapter 4 compared proteome profiles between surgically induced and chemically induced OA rabbit model.

After determining that chemically induced rabbit model is more suitable in studying the effects of EBN in osteoarthritic rabbit model, we then proceeded to observe the efficacy of EBN in OA.

- Chapter 5 evaluated the effects of EBN treatment in subchondral bone and articular cartilage.
- Chapter 6 compared differential expression of proteins between EBN treated group and non-treated group.
- Chapter 7 summarized the important findings and recommendations for future research.



## REFERENCES

- Aigner, T., Rose, J., Martin, J., & Buckwalter, J. (2004). Aging theories of primary osteoarthritis: From epidemiology to molecular biology. In *Rejuvenation Research*. <https://doi.org/10.1089/1549168041552964>
- Altman, R. D., & Gold, G. E. (2007). Atlas of individual radiographic features in osteoarthritis, revised. *Osteoarthritis and Cartilage*, 15(SUPPL. 1), 1–56. <https://doi.org/10.1016/j.joca.2006.06.017>
- Amin, K. (2012). The role of mast cells in allergic inflammation. In *Respiratory Medicine*. <https://doi.org/10.1016/j.rmed.2011.09.007>
- Andia, I., & Maffulli, N. (2013). Platelet-rich plasma for managing pain and inflammation in osteoarthritis. In *Nature Reviews Rheumatology*. <https://doi.org/10.1038/nrrheum.2013.141>
- Arden, N., & Nevitt, M. C. (2006). Osteoarthritis: Epidemiology. In *Best Practice and Research: Clinical Rheumatology*. <https://doi.org/10.1016/j.berh.2005.09.007>
- Babji, A. S., Ety Syarmila, I. K., Nur 'Aliah, D., Nurul Nadia, M., Hadi Akbar, D., Norrakiah, A. S., Ghassem, M., Najafian, L., & Salma, M. Y. (2018). Assessment on bioactive components of hydrolysed edible bird nest. *International Food Research Journal*
- Banios, K., Raoulis, V., Fylos, A., Chytas, D., Mitrousias, V., & Zibis, A. (2022). Anterior and Posterior Cruciate Ligaments Mechanoreceptors: A Review of Basic Science. In *Diagnostics*. <https://doi.org/10.3390/diagnostics12020331>
- Bank, R. A., Bayliss, M. T., Lafeber, F. P. J. G., Maroudas, A., & Tekoppele, J. M. (1998). Ageing and zonal variation in post-translational modification of collagen in normal human articular cartilage: The age-related increase in Non-Enzymatic Glycation affects biomechanical properties of cartilage. *Biochemical Journal*. <https://doi.org/10.1042/bj3300345>
- Banks, R. E., Dunn, M. J., Hochstrasser, D. F., Sanchez, J. C., Blackstock, W., Pappin, D. J., & Selby, P. J. (2000). Proteomics: New perspectives, new biomedical opportunities. *Lancet*. [https://doi.org/10.1016/S0140-6736\(00\)03214-1](https://doi.org/10.1016/S0140-6736(00)03214-1)
- Bateman, T. A., & Countryman, S. (2002). Osteoprotegerin and bone loss associated with spaceflight. In *Drug Discovery Today*. [https://doi.org/10.1016/S1359-6446\(02\)02260-2](https://doi.org/10.1016/S1359-6446(02)02260-2)

- Batiste, D. L., Kirkley, A., Laverty, S., Thain, L. M. F., Spouge, A. R., Gati, J. S., Foster, P. J., & Holdsworth, D. W. (2004). High-resolution MRI and micro-CT in an ex vivo rabbit anterior cruciate ligament transection model of osteoarthritis. *Osteoarthritis and Cartilage*, 12(8), 614–626. <https://doi.org/10.1016/j.joca.2004.03.002>
- Bellido, M., Lugo, L., Roman-Blas, J. A., Castañeda, S., Caeiro, J. R., Dapia, S., Calvo, E., Largo, R., & Herrero-Beaumont, G. (2010). Subchondral bone microstructural damage by increased remodelling aggravates experimental osteoarthritis preceded by osteoporosis. *Arthritis Research and Therapy*. <https://doi.org/10.1186/ar3103>
- Blagojevic, M., Jinks, C., Jeffery, A., & Jordan, K. P. (2010). Risk factors for onset of osteoarthritis of the knee in older adults: a systematic review and meta-analysis. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2009.08.010>
- Bobinac, D., Spanjol, J., Zoricic, S., & Maric, I. (2003). Changes in articular cartilage and subchondral bone histomorphometry in osteoarthritic knee joints in humans. *Bone*. [https://doi.org/10.1016/S8756-3282\(02\)00982-1](https://doi.org/10.1016/S8756-3282(02)00982-1)
- Bodmer, J. L., Schneider, P., & Tschopp, J. (2002). The molecular architecture of the TNF superfamily. In *Trends in Biochemical Sciences*. [https://doi.org/10.1016/S0968-0004\(01\)01995-8](https://doi.org/10.1016/S0968-0004(01)01995-8)
- Boe, C., & Vangness, C. T. (2015). Fish Oil and Osteoarthritis: Current Evidence. *American Journal of Orthopedics* (Belle Mead, N.J.), 44(7), 302–305. <http://www.ncbi.nlm.nih.gov/pubmed/26161757>
- Boris Chan, P. M., Zhu, L., Wen, C. Y., & Chiu, K. Y. (2015). Subchondral bone proteomics in osteoarthritis: Current status and perspectives. In *Journal of Orthopaedic Translation*. <https://doi.org/10.1016/j.jot.2015.02.002>
- Bouchgua, M., Alexander, K., André d'Anjou, M., Girard, C. A., Carmel, E. N., Beauchamp, G., Richard, H., & Laverty, S. (2009). Use of routine clinical multimodality imaging in a rabbit model of osteoarthritis - part I. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2008.06.017>
- Braun, H. J., & Gold, G. E. (2012). Diagnosis of osteoarthritis : Imaging. *Bone*, 51(2), 278–288. <https://doi.org/10.1016/j.bone.2011.11.019>
- Bruins, A. P. (1998). Mechanistic aspects of electrospray ionization. In *Journal of Chromatography A*. [https://doi.org/10.1016/S0021-9673\(97\)01110-2](https://doi.org/10.1016/S0021-9673(97)01110-2)
- Bruno, M. A., Mosher, T. J., & Gold, G. E. (Eds.). (2010). *Arthritis in Color: Advanced Imaging of Arthritis*. *American Journal of Neuroradiology*, 31(1), R1–R1. <https://doi.org/10.3174/ajnr.A1813>

- Buckwalter, J. A., & Mankin, H. J. (1998). Articular cartilage: degeneration and osteoarthritis, repair, regeneration, and transplantation. In Instructional course lectures
- Buckwalter, J. A., Roughley, P. J., & Rosenberg, L. C. (1994). Age-Related changes in cartilage proteoglycans: Quantitative electron microscopic studies. *Microscopy Research and Technique*. <https://doi.org/10.1002/jemt.1070280506>
- Burr, D. B., & Gallant, M. A. (2012). Bone remodelling in osteoarthritis. In *Nature Reviews Rheumatology*. <https://doi.org/10.1038/nrrheum.2012.130>
- Burr, D. B., & Schaffler, M. B. (1997). The involvement of subchondral mineralized tissues in osteoarthritis: Quantitative microscopic evidence. *Microscopy Research and Technique*. [https://doi.org/10.1002/\(SICI\)1097-0029\(19970515\)37:4<343::AID-JEMT9>3.0.CO;2-L](https://doi.org/10.1002/(SICI)1097-0029(19970515)37:4<343::AID-JEMT9>3.0.CO;2-L)
- Butt, A. Q., Ahmed, S., Maratha, A., & Miggin, S. M. (2012). 14-3-3 $\epsilon$  and 14-3-3 $\sigma$  inhibit toll-like receptor (TLR)-mediated proinflammatory cytokine induction. *Journal of Biological Chemistry*. <https://doi.org/10.1074/jbc.M112.367490>
- Cabahug-Zuckerman, P., Frikha-Benayed, D., Majeska, R. J., Tuthill, A., Yakar, S., Judex, S., & Schaffler, M. B. (2016). Osteocyte Apoptosis Caused by Hindlimb Unloading is Required to Trigger Osteocyte RANKL Production and Subsequent Resorption of Cortical and Trabecular Bone in Mice Femurs. *Journal of Bone and Mineral Research*. <https://doi.org/10.1002/jbmr.2807>
- Campos, W. N. S., Souza, M. A., Ruiz, T., Peres, T. P., Néspoli, P. B., Marques, A. T. C., Colodel, E. M., Souza, R. L. De, S., A. C. W. N., Souza, M. A., Ruiz, T., Peres, T. P., Néspoli, P. B., & Marques, A. T. C. (2013). Experimental osteoarthritis in rabbits : lesion progression 1. 32(3), 279–285
- Canapp, S. O. (2007). The Canine Stifle. *Clinical Techniques in Small Animal Practice*. <https://doi.org/10.1053/j.ctsap.2007.09.008>
- Caplan, A. I., & Correa, D. (2011). The MSC: An Injury Drugstore. *Cell Stem Cell*, 9(1), 11–15. <https://doi.org/10.1016/j.stem.2011.06.008>
- Carlson, C. S., Loeser, R. F., Jayo, M. J., Weaver, D. S., Adams, M. R., & Jerome, C. P. (1994). Osteoarthritis in cynomolgus macaques: A primate model of naturally occurring disease. *Journal of Orthopaedic Research*. <https://doi.org/10.1002/jor.1100120305>
- Carlson, C. S., Loeser, R. F., Purser, C. B., Gardin, J. F., & Jerome, C. P. (1996). Osteoarthritis in cynomolgus macaques III: Effects of age, gender, and subchondral bone thickness on the severity of disease. *Journal of Bone and Mineral Research*. <https://doi.org/10.1002/jbmr.5650110904>

- Castañeda, S., Roman-Blas, J. A., Largo, R., & Herrero-Beaumont, G. (2012). Subchondral bone as a key target for osteoarthritis treatment. In *Biochemical Pharmacology*. <https://doi.org/10.1016/j.bcp.2011.09.018>
- Castrogiovanni, P., Trovato, F. M., Loreto, C., Nsir, H., Szychlinska, M. A., & Musumeci, G. (2016). Nutraceutical supplements in the management and prevention of osteoarthritis. In *International Journal of Molecular Sciences*. <https://doi.org/10.3390/ijms17122042>
- Chang, S. H., Mori, D., Kobayashi, H., Mori, Y., Nakamoto, H., Okada, K., Taniguchi, Y., Sugita, S., Yano, F., Chung, U. il, Kim-Kaneyama, J. ri, Yanagita, M., Economides, A., Canalis, E., Chen, D., Tanaka, S., & Saito, T. (2019). Excessive mechanical loading promotes osteoarthritis through the gremlin-1–NF- $\kappa$ B pathway. *Nature Communications*. <https://doi.org/10.1038/s41467-019-09491-5>
- Chappard, C., Peyrin, F., Bonnassie, A., Lemineur, G., Brunet-Imbault, B., Lespessailles, E., & Benhamou, C. L. (2006). Subchondral bone micro-architectural alterations in osteoarthritis: A synchrotron micro-computed tomography study. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2005.09.008>
- Chen, D., Shen, J., Zhao, W., Wang, T., Han, L., Hamilton, J. L., & Im, H. J. (2017). Osteoarthritis: Toward a comprehensive understanding of pathological mechanism. In *Bone Research*. <https://doi.org/10.1038/boneres.2016.44>
- Chen, Y. Y., Lin, S. Y., Yeh, Y. Y., Hsiao, H. H., Wu, C. Y., Chen, S. T., & Wang, A. H. J. (2005). A modified protein precipitation procedure for efficient removal of albumin from serum. *Electrophoresis*. <https://doi.org/10.1002/elps.200410381>
- Cheng, F., Shen, Y., Mohanasundaram, P., Lindström, M., Ivaska, J., Ny, T., & Erikss, J. E. (2016). Vimentin coordinates fibroblast proliferation and keratinocyte differentiation in wound healing via TGF- $\beta$ -Slug signaling. *Proceedings of the National Academy of Sciences of the United States of America*. <https://doi.org/10.1073/pnas.1519197113>
- Choi, Jo, Park, Kang, & Park. (2019). NF-B Signaling Pathways in Osteoarthritic Cartilage Destruction. *Cells*. <https://doi.org/10.3390/cells8070734>
- Chu, C. R., Lin, D., Geisler, J. L., Chu, C. T., Fu, F. H., & Pan, Y. (2004). Arthroscopic Microscopy of Articular Cartilage Using Optical Coherence Tomography. *American Journal of Sports Medicine*. <https://doi.org/10.1177/0363546503261736>

- Chua, K.-H., Lee, T.-H., Nagandran, K., Md Yahaya, N. H., Lee, C.-T., Tjih, E. T. T., & Abdul Aziz, R. (2013). Edible Bird's nest extract as a chondro-protective agent for human chondrocytes isolated from osteoarthritic knee: *in vitro* study. *BMC Complementary and Alternative Medicine*, 13(1), 19. <https://doi.org/10.1186/1472-6882-13-19>
- Clark, D. P., & Badea, C. T. (2014). Micro-CT of rodents: State-of-the-art and future perspectives. In *Physica Medica*. <https://doi.org/10.1016/j.ejmp.2014.05.011>
- Cray, C., Zaias, J., & Altman, N. H. (2009). Acute phase response in animals: A review. In *Comparative Medicine*
- Crofford, L. J. (2013). Use of NSAIDs in treating patients with arthritis. In *Arthritis Research and Therapy*. <https://doi.org/10.1186/ar4174>
- Dai, S. M., Shan, Z. Z., Nakamura, H., Masuko-Hongo, K., Kato, T., Nishioka, K., & Yudoh, K. (2006). Catabolic stress induces features of chondrocyte senescence through overexpression of caveolin 1: Possible involvement of caveolin 1-induced down-regulation of articular chondrocytes in the pathogenesis of osteoarthritis. *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.21639>
- Dakappagari, N., Neely, L., Tangri, S., Lundgren, K., Hipolito, L., Estrellado, A., Burrows, F., & Zhang, H. (2010). An investigation into the potential use of serum Hsp70 as a novel tumour biomarker for Hsp90 inhibitors. *Biomarkers*. <https://doi.org/10.3109/13547500903261347>
- Davies, N. M., Røseth, A. G., Appleyard, C. B., Mcknight, W., Del Soldato, P., Calignano, A., Cirino, G., & Wallace, J. L. (1997). NO-naproxen vs. naproxen: Ulcerogenic, analgesic and anti-inflammatory effects. *Alimentary Pharmacology and Therapeutics*. <https://doi.org/10.1046/j.1365-2036.1997.115286000.x>
- Day-Williams, A. G., Southam, L., Panoutsopoulou, K., Rayner, N. W., Esko, T., Estrada, K., Helgadottir, H. T., Hofman, A., Ingvarsson, T., Jonsson, H., Keis, A., Kerkhof, H. J. M., Thorleifsson, G., Arden, N. K., Carr, A., Chapman, K., Deloukas, P., Loughlin, J., McCaskie, A., ... Zeggini, E. (2011). A variant in MCF2L is associated with osteoarthritis. *American Journal of Human Genetics*. <https://doi.org/10.1016/j.ajhg.2011.08.001>
- De Seny, D., Cobraiville, G., Charlier, E., Neuville, S., Lutteri, L., Goff, C. Le, Malaise, D., Malaise, O., Chapelle, J. P., Relic, B., & Malaise, M. G. (2015). Apolipoprotein-A1 as a damage-associated molecular patterns protein in osteoarthritis: Ex vivo and *in vitro* pro-inflammatory properties. *PLoS ONE*. <https://doi.org/10.1371/journal.pone.0122904>

- DeGroot, J., Verzijl, N., Budde, M., Bijlsma, J. W. J., Lafeber, F. P. J. G., & TeKoppele, J. M. (2001). Accumulation of advanced glycation end products decreases collagen turnover by bovine chondrocytes. *Experimental Cell Research*. <https://doi.org/10.1006/excr.2001.5224>
- Dempster, D. W. (2000). The contribution of trabecular architecture to cancellous bone quality. In *Journal of Bone and Mineral Research*. <https://doi.org/10.1359/jbmr.2000.15.1.20>
- Di Martino, A., Di Matteo, B., Papio, T., Tentoni, F., Selleri, F., Cenacchi, A., Kon, E., & Filardo, G. (2019). Platelet-Rich Plasma Versus Hyaluronic Acid Injections for the Treatment of Knee Osteoarthritis: Results at 5 Years of a Double-Blind, Randomized Controlled Trial. *American Journal of Sports Medicine*. <https://doi.org/10.1177/0363546518814532>
- Dick, M. K., & Limaïem, F. (2019). Histology, Fibroblast. In *StatPearls*.
- Elron-Gross, I., Glucksam, Y., Melikhov, D., & Margalit, R. (2008). Cyclooxygenase inhibition by diclofenac formulated in bioadhesive carriers. *Biochimica et Biophysica Acta - Biomembranes*, 1778(4), 931–936. <https://doi.org/10.1016/j.bbamem.2008.01.002>
- English, K., & Mahon, B. P. (2011). Allogeneic mesenchymal stem cells: Agents of immune modulation. In *Journal of Cellular Biochemistry*. <https://doi.org/10.1002/jcb.23119>
- Evangelou, E., Valdes, A. M., Kerkhof, H. J. M., Styrkarsdottir, U., Zhu, Y. Y., Meulenbelt, I., Lories, R. J., Karassa, F. B., Tylzanowski, P., Bos, S. D., Rayner, N. W., Southam, L., Zhai, G., Elliott, K. S., Hunt, S. E., Blackburn, H., Potter, S. C., Day-Williams, A. G., Beazley, C., ... Spector, T. D. (2011). Meta-analysis of genome-wide association studies confirms a susceptibility locus for knee osteoarthritis on chromosome 7q22. *Annals of the Rheumatic Diseases*. <https://doi.org/10.1136/ard.2010.132787>
- Felson, D. T., & Neogi, T. (2004). Osteoarthritis: Is It a Disease of Cartilage or of Bone? In *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.20051>
- Felson, D. T., Anderson, J. J., Naimark, A., Walker, A. M., & Meenan, R. F. (1988). Obesity and knee osteoarthritis. The Framingham Study. *Annals of Internal Medicine*. <https://doi.org/10.7326/0003-4819-109-1-18>
- Felson, David T., Couropmitree, N. N., Chaisson, C. E., Hannan, M. T., Zhang, Y., Mcalindon, T. E., Lavalley, M., Levy, D., & Myers, R. H. (1998). Evidence for a Mendelian gene in a segregation analysis of generalized radiographic osteoarthritis: The Framingham Study. In *Arthritis and Rheumatism*. [https://doi.org/10.1002/1529-0131\(199806\)41:6<1064::AID-ART13>3.0.CO;2-K](https://doi.org/10.1002/1529-0131(199806)41:6<1064::AID-ART13>3.0.CO;2-K)

- Florea, C., Malo, M. K. H., Rautiainen, J., Mäkelä, J. T. A., Fick, J. M., Nieminen, M. T., Jurvelin, J. S., Davidescu, A., & Korhonen, R. K. (2015). Alterations in subchondral bone plate, trabecular bone and articular cartilage properties of rabbit femoral condyles at 4 weeks after anterior cruciate ligament transection. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2014.11.023>
- Foo, C. N., Manohar, A., Rampal, L., Lye, M.-S., Mohd-Sidik, S., & Osman, Z. J. (2017). Knee Pain and Functional Disability of Knee Osteoarthritis Patients Seen at Malaysian Government Hospitals. *Malaysian Journal of Medicine and Health Sciences*
- Frank, C. B. (2004). Ligament structure, physiology and function. In *Journal of Musculoskeletal Neuronal Interactions*
- Freitag, J., Bates, D., Wickham, J., Shah, K., Huguenin, L., Tenen, A., Paterson, K., & Boyd, R. (2019). Adipose-derived mesenchymal stem cell therapy in the treatment of knee osteoarthritis: A randomized controlled trial. *Regenerative Medicine*. <https://doi.org/10.2217/rme-2018-0161>
- García-Coronado, J. M., Martínez-Olvera, L., Elizondo-Omaña, R. E., Acosta-Olivo, C. A., Vilchez-Cavazos, F., Simental-Mendía, L. E., & Simental-Mendía, M. (2019). Effect of collagen supplementation on osteoarthritis symptoms: a meta-analysis of randomized placebo-controlled trials. In *International Orthopaedics*. <https://doi.org/10.1007/s00264-018-4211-5>
- Gharbi, M., Deberg, M., & Henrotin, Y. (2011). Application for proteomic techniques in studying osteoarthritis: A review. *Frontiers in Physiology*. <https://doi.org/10.3389/fphys.2011.00090>
- Gobezie, R., Kho, A., Krastins, B., Sarracino, D. A., Thornhill, T. S., Chase, M., Millett, P. J., & Lee, D. M. (2007). High abundance synovial fluid proteome: Distinct profiles in health and osteoarthritis. *Arthritis Research and Therapy*, 9(2), 1–15. <https://doi.org/10.1186/ar2172>
- Goldring SR, Goldring MB (2016) Changes in the osteochondral unit during osteoarthritis: Structure, function and cartilage bone crosstalk. *Nature Reviews Rheumatology*
- Golightly, Y. M., Allen, K. D., Helmick, C. G., Renner, J. B., & Jordan, J. M. (2009). Symptoms of the knee and hip in individuals with and without limb length inequality. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2008.11.005>

- Golightly, Y. M., Allen, K. D., Renner, J. B., Helmick, C. G., Salazar, A., & Jordan, J. M. (2007). Relationship of limb length inequality with radiographic knee and hip osteoarthritis. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2007.01.009>
- Goudarzi, R., Reid, A., & McDougall, J. J. (2018). Evaluation of the novel avocado/soybean unsaponifiable Arthrocen to alter joint pain and inflammation in a rat model of osteoarthritis. *PLoS ONE*. <https://doi.org/10.1371/journal.pone.0191906>
- Grebe, S. K. G., & Singh, R. J. (2011). LC-MS / MS in the Clinical Laboratory – Where to From Here ? 32(February), 5–31.
- Grossin, L., Cournil-Henrionnet, C., Pinzano, A., Gaborit, N., Dumas, D., Etienne, S., Stoltz, J. F., Terlain, B., Netter, P., Mir, L. M., & Gillet, P. (2006). Gene transfer with HSP 70 in rat chondrocytes confers cytoprotection *in vitro* and during experimental osteoarthritis. *The FASEB Journal*. <https://doi.org/10.1096/fj.04-2889com>
- Guilak, F. (2011). Biomechanical factors in osteoarthritis. In *Best Practice and Research: Clinical Rheumatology*. <https://doi.org/10.1016/j.berh.2011.11.013>
- Guingamp, C., Gegout-Pottie, P., Philippe, L., Terlain, B., Netter, P., & Gillet, P. (1997). Mono-iodoacetate-induced experimental osteoarthritis: A dose-response study of loss of mobility, morphology, and biochemistry. *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.1780400917>
- Guo, C. T., Takahashi, T., Bukawa, W., Takahashi, N., Yagi, H., Kato, K., Hidari, K. I. P. J., Miyamoto, D., Suzuki, T., & Suzuki, Y. (2006). Edible bird's nest extract inhibits influenza virus infection. *Antiviral Research*. <https://doi.org/10.1016/j.antiviral.2006.02.005>
- Guo, H., Yin, W., Zou, Z., Zhang, C., Sun, M., Min, L., Yang, L., & Kong, L. (2021). Quercitrin alleviates cartilage extracellular matrix degradation and delays ACLT rat osteoarthritis development: An *in vivo* and *in vitro* study. *Journal of Advanced Research*. <https://doi.org/10.1016/j.jare.2020.06.020>
- Guzman, R. E., Evans, M. G., Bove, S., Morenko, B., & Kilgore, K. (2003). Mono-Iodoacetate-Induced Histologic Changes in Subchondral Bone and Articular Cartilage of Rat Femorotibial Joints: AN Animal Model of Osteoarthritis. *Toxicologic Pathology*. <https://doi.org/10.1080/01926230390241800>
- Hall, M. P., Band, P. A., Meislin, R. T., Jazrawi, L. M., & Cardone, D. A. (2009). Platelet-rich plasma: Current concepts and application in sports medicine. *Journal of the American Academy of Orthopaedic Surgeons*. <https://doi.org/10.5435/00124635-200910000-00002>



- Heaney, R. P. (2004). Phosphorus Nutrition and the Treatment of Osteoporosis. In Mayo Clinic Proceedings. <https://doi.org/10.4065/79.1.91>
- Henrotin, Y., Mobasher, A., & Marty, M. (2012). Is there any scientific evidence for the use of glucosamine in the management of human osteoarthritis? In Arthritis Research and Therapy. <https://doi.org/10.1186/ar3657>
- Hermeto, L. C., Rafael De Rossi, Henrique, P., Jardim, D. A., Evangelista, A., Iv, S., & De, J. (2016). MODELS , BIOLOGICAL Comparison between two different experimental models of osteoarthritis in rabbits . Intra- articular collagenase injection and anterior cruciate ligament transection 1. Acta Cirurgica Brasileira, 31(9), 602–607
- Herzog, W., Diet, S., Suter, E., Mayzus, P., Leonard, T. R., Müller, C., Wu, J. Z., & Epstein, M. (1998). Material and functional properties of articular cartilage and patellofemoral contact mechanics in an experimental model of osteoarthritis. Journal of Biomechanics. [https://doi.org/10.1016/S0021-9290\(98\)00136-5](https://doi.org/10.1016/S0021-9290(98)00136-5)
- Ho, C. S., Lam, C. W. K., Chan, M. H. M., Cheung, R. C. K., Law, L. K., Lit, L. C. W., Ng, K. F., Suen, M. W. M., & Tai, H. L. (2003). Electrospray Ionisation Mass Spectrometry : Principles and Clinical Applications. 24(February), 3–12
- Holmdahl, D. E., & Ingelmark, B. E. (1951). The contact between the articular cartilage and the medullary cavities of the bones. Acta Anatomica. <https://doi.org/10.1159/000140554>
- Hosseini, S., Weis, M. A., Rai, J., Kim, L., Funk, S., Dahlberg, L. E., & Eyre, D. R. (2016). Evidence for enhanced collagen type III deposition focally in the territorial matrix of osteoarthritic hip articular cartilage. Osteoarthritis and Cartilage. <https://doi.org/10.1016/j.joca.2016.01.001>
- Hou, C. H., Fong, Y. C., & Tang, C. H. (2011). HMGB-1 induces IL-6 production in human synovial fibroblasts through c-Src, Akt and NF- $\kappa$ B pathways. Journal of Cellular Physiology. <https://doi.org/10.1002/jcp.22541>
- Hou, T. Y., Chiang-Ni, C., & Teng, S. H. (2019). Current status of MALDI-TOF mass spectrometry in clinical microbiology. In Journal of Food and Drug Analysis. <https://doi.org/10.1016/j.jfda.2019.01.001>
- Hou, Z. ping, Tang, S. ying, Ji, H. ru, He, P. yuan, Li, Y. hong, Dong, X. ling, Du, M. nan, Maznah, I., & He, W. jing. (2021). Edible Bird's Nest Attenuates Menopause-Related Bone Degeneration in Rats via Increasing Bone Estrogen-Receptor Expression. Chinese Journal of Integrative Medicine. <https://doi.org/10.1007/s11655-019-3209-1>

- Hou, Z., Imam, M. U., Ismail, M., Ooi, D. J., Ideris, A., & Mahmud, R. (2015). Nutrigenomic effects of edible bird's nest on insulin signaling in ovariectomized rats. *Drug Design, Development and Therapy*. <https://doi.org/10.2147/DDDT.S80743>
- Hunter, D. J., Zhang, Y. Q., Tu, X., LaValley, M., Niu, J. B., Amin, S., Guermazi, A., Genant, H., Gale, D., & Felson, D. T. (2006). Change in joint space width: Hyaline articular cartilage loss or alteration in meniscus? *Arthritis and Rheumatism*, 54(8), 2488–2495. <https://doi.org/10.1002/art.22016>
- Huskisson, E. C., Berry, H., Gishen, P., Jubb, R. W., & Whitehead, J. (1995). Effects of antiinflammatory drugs on the progression of osteoarthritis of the knee. *Journal of Rheumatology*
- Iannitti, T., Elhensheri, M., Bingöl, A. Ö., & Palmieri, B. (2013). Preliminary histopathological study of intra-articular injection of a novel highly cross-linked hyaluronic acid in a rabbit model of knee osteoarthritis. *Journal of Molecular Histology*, 44(2), 191–201. <https://doi.org/10.1007/s10735-012-9457-4>
- Intema, F., Hazewinkel, H. A. W., Gouwens, D., Bijlsma, J. W. J., Weinans, H., Lafeber, F. P. J. G., & Mastbergen, S. C. (2010). In early OA, thinning of the subchondral plate is directly related to cartilage damage: Results from a canine ACLT-menisectomy model. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2010.01.004>
- Janusz, M. J., Hookfin, E. B., Heitmeyer, S. A., Woessner, J. F., Freemont, A. J., Hoyland, J. A., Brown, K. K., Hsieh, L. C., Almstead, N. G., De, B., Natchus, M. G., Pikul, S., & Taiwo, Y. O. (2001). Moderation of iodoacetate-induced experimental osteoarthritis in rats by matrix metalloproteinase inhibitors. *Osteoarthritis and Cartilage*. <https://doi.org/10.1053/joca.2001.0472>
- Ji, X., & Zhang, H. (2019). Current Strategies for the Treatment of Early Stage Osteoarthritis. *Frontiers in Mechanical Engineering*, 5, 57. <https://doi.org/10.3389/fmech.2019.00057>
- Ji, X., Yan, Y., Sun, T., Zhang, Q., Wang, Y., Zhang, M., Zhang, H., & Zhao, X. (2019). Glucosamine sulphate-loaded distearoyl phosphocholine liposomes for osteoarthritis treatment: Combination of sustained drug release and improved lubrication. *Biomaterials Science*. <https://doi.org/10.1039/c9bm00201d>
- Jiang, L., Li, L., Geng, C., Gong, D., Jiang, L., Ishikawa, N., Kajima, K., & Zhong, L. (2013). Monosodium iodoacetate induces apoptosis via the mitochondrial pathway involving ROS production and caspase activation in rat chondrocytes *in vitro*. *Journal of Orthopaedic Research*. <https://doi.org/10.1002/jor.22250>

- Jiřík, M., Bartoš, M., Tomášek, P., Malečková, A., Kural, T., Horáková, J., Lukáš, D., Suchý, T., Kochová, P., Hubálek Kalbáčová, M., Králíčková, M., & Tonar, Z. (2018). Generating standardized image data for testing and calibrating quantification of volumes, surfaces, lengths, and object counts in fibrous and porous materials using X-ray microtomography. *Microscopy Research and Technique*. <https://doi.org/10.1002/jemt.23011>
- Johnson, V. L., & Hunter, D. J. (2014). The epidemiology of osteoarthritis. In *Best Practice and Research: Clinical Rheumatology*. <https://doi.org/10.1016/j.berh.2014.01.004>
- Jones, H. W., Bailey, R., Zhang, Z., Dunne, K. A., Blake, D. R., Cox, N. L., Morris, C. J., & Winyard, P. G. (1998). Inactivation of antithrombin III in synovial fluid from patients with rheumatoid arthritis. *Annals of the Rheumatic Diseases*. <https://doi.org/10.1136/ard.57.3.162>
- Jones, I. A., Togashi, R., Wilson, M. L., Heckmann, N., & Vangsness, C. T. (2019). Intra-articular treatment options for knee osteoarthritis. In *Nature Reviews Rheumatology*. <https://doi.org/10.1038/s41584-018-0123-4>
- Jose, A. (2011). Trends In Proteomics: A Tool In Disease Diagnosis And Drug Discovery. *Asian Journal of Pharmaceutical and Health Sciences*, 1(1)
- Judex, S., Gross, T. S., Bray, R. C., & Zernicke, R. F. (1997). Adaptation of bone to physiological stimuli. *Journal of Biomechanics*. [https://doi.org/10.1016/S0021-9290\(96\)00060-7](https://doi.org/10.1016/S0021-9290(96)00060-7)
- Kahan, A., Uebelhart, D., De Vathaire, F., Delmas, P. D., & Reginster, J. Y. (2009). Long-term effects of chondroitins 4 and 6 sulfate on knee osteoarthritis: The study on osteoarthritis progression prevention, a two-year, randomized, double-blind, placebo-controlled trial. *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.24255>
- Kamada, T., Kurokawa, M. S., Kato, T., Takenouchi, K., Takahashi, K., Yoshioka, T., Uchida, T., Mitsui, H., Suematsu, N., Okamoto, K., Yudo, K., Katayama, Y., & Nakamura, H. (2012). Proteomic analysis of bone marrow-adherent cells in rheumatoid arthritis and osteoarthritis. *International Journal of Rheumatic Diseases*. <https://doi.org/10.1111/j.1756-185X.2012.01702.x>
- Kang, Y. H., Lee, H. J., Lee, C. J., & Park, J. S. (2019). Natural products as sources of novel drug candidates for the pharmacological management of osteoarthritis: A narrative review. In *Biomolecules and Therapeutics*. <https://doi.org/10.4062/biomolther.2019.139>
- Karas, M., & Kru, R. (2003). Ion Formation in MALDI: The Cluster Ionization Mechanism. <https://doi.org/10.1021/cr010376a>

- Karsdal, M. A., Leeming, D. J., Dam, E. B., Henriksen, K., Alexandersen, P., Pastoureau, P., Altman, R. D., & Christiansen, C. (2008). Should subchondral bone turnover be targeted when treating osteoarthritis? In *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2008.01.014>
- Kellgren, J. H., & Lawrence, J. S. (1957). Radiological assessment of osteo-arthritis. *Annals of the Rheumatic Diseases*. <https://doi.org/10.1136/ard.16.4.494>
- Kennedy, O. D., Herman, B. C., Laudier, D. M., Majeska, R. J., Sun, H. B., & Schaffler, M. B. (2012). Activation of resorption in fatigue-loaded bone involves both apoptosis and active pro-osteoclastogenic signaling by distinct osteocyte populations. *Bone*. <https://doi.org/10.1016/j.bone.2012.01.025>
- Khushairay, E. S. I., Ayub, M. K., & Babji, A. S. (2014). Effect of enzymatic hydrolysis of pancreatin and alcalase enzyme on some properties of edible bird's nest hydrolysate. *427(2014)*, 427–432. <https://doi.org/10.1063/1.4895235>
- Kitts, D., & Weiler, K. (2003). Bioactive Proteins and Peptides from Food Sources. Applications of Bioprocesses used in Isolation and Recovery. *Current Pharmaceutical Design*. <https://doi.org/10.2174/1381612033454883>
- Kon, E., Filardo, G., Drobnic, M., Madry, H., Jelic, M., van Dijk, N., & della Villa, S. (2012). Non-surgical management of early knee osteoarthritis. In *Knee Surgery, Sports Traumatology, Arthroscopy*. <https://doi.org/10.1007/s00167-011-1713-8>
- Korfmacher, W. A. (2004). Using mass spectrometry for drug metabolism studies. In *Using Mass Spectrometry for Drug Metabolism Studies*. <https://doi.org/10.1201/9781420092219>
- Korhonen, H., & Pihlanto, A. (2006). Bioactive peptides: Production and functionality. In *International Dairy Journal*. <https://doi.org/10.1016/j.idairyj.2005.10.012>
- Kothari, P., Sinha, S., Sardar, A., Tripathi, A. K., Girme, A., Adhikary, S., Singh, R., Maurya, R., Mishra, P. R., Hingorani, L., & Trivedi, R. (2020). Inhibition of cartilage degeneration and subchondral bone deterioration by: *Spinacia oleracea* in human mimic of ACLT-induced osteoarthritis. *Food and Function*. <https://doi.org/10.1039/d0fo01125h>
- Kujala, U. M., Kettunen, J., Paananen, H., Aalto, T., Battié, M. C., Impivaara, O., Videman, T., & Sarna, S. (1995). Knee osteoarthritis in former runners, soccer players, weight lifters, and shooters. *Arthritis & Rheumatism*. <https://doi.org/10.1002/art.1780380413>

- Kuyinu, E. L., Narayanan, G., Nair, L. S., & Laurencin, C. T. (2016). Animal models of osteoarthritis: classification, update, and measurement of outcomes. *Journal of Orthopaedic Surgery and Research*, 11(1), 19. <https://doi.org/10.1186/s13018-016-0346-5>
- Langelier, E., Suetterlin, R., Hoemann, C. D., Aebi, U., & Buschmann, M. D. (2000). The chondrocyte cytoskeleton in mature articular cartilage: Structure and distribution of actin, tubulin, and vimentin filaments. *Journal of Histochemistry and Cytochemistry*. <https://doi.org/10.1177/002215540004801002>
- Lavigne, P., Benderdour, M., Lajeunesse, D., Reboul, P., Shi, Q., Pelletier, J. P., Martel-Pelletier, J., & Fernandes, J. C. (2005). Subchondral and trabecular bone metabolism regulation in canine experimental knee osteoarthritis. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2004.12.015>
- Lee, D. G., Park, S. Y., Chung, W. S., Park, J. H., Hwang, E., Mavlonov, G. T., Kim, I. H., Kim, K. Y., & Yi, T. H. (2015). Fucoidan Prevents the Progression of Osteoarthritis in Rats. *Journal of Medicinal Food*. <https://doi.org/10.1089/jmf.2014.3334>
- Lee, J. H., Chun, K. J., Kim, H. S., Kim, S. H., Han, P., Jun, Y., & Lim, D. (2012). Alteration patterns of trabecular bone microarchitectural characteristics induced by osteoarthritis over time. *Clinical Interventions in Aging*. <https://doi.org/10.2147/CIA.S32513>
- Lee, T. H., Wani, W. A., Lee, C. H., Cheng, K. K., Shreaz, S., Wong, S., Hamdan, N., & Azmi, N. A. (2021). Edible Bird's Nest: The Functional Values of the Prized Animal-Based Bioproduct From Southeast Asia—A Review. In *Frontiers in Pharmacology*. <https://doi.org/10.3389/fphar.2021.626233>
- Lehenkari, P., Hentunen, T. A., Laitala-Leinonen, T., Tuukkanen, J., & Väänänen, H. K. (1998). Carbonic anhydrase II plays a major role in osteoclast differentiation and bone resorption by effecting the steady state intracellular pH and Ca<sup>2+</sup>. *Experimental Cell Research*. <https://doi.org/10.1006/excr.1998.4071>
- Lelovas, K. L. P. (2014). Useful animal models for the research of osteoarthritis. 263–271. <https://doi.org/10.1007/s00590-013-1205-2>
- Levick, J. R., & McDonald, J. N. (1995). Fluid movement across synovium in healthy joints: Role of synovial fluid macromolecules. *Annals of the Rheumatic Diseases*. <https://doi.org/10.1136/ard.54.5.417>
- Li, G., Yin, J., Gao, J., Cheng, T. S., Pavlos, N. J., Zhang, C., & Zheng, M. H. (2013). Subchondral bone in osteoarthritis: Insight into risk factors and microstructural changes. In *Arthritis Research and Therapy*. <https://doi.org/10.1186/ar4405>

- Li, Hao, Zhang, H., Tang, Z., & Hu, G. (2008). Micro-computed tomography for small animal imaging: Technological details. In *Progress in Natural Science*. <https://doi.org/10.1016/j.pnsc.2008.01.002>
- Li, Heng, Wang, D., Yuan, Y., & Min, J. (2017). New insights on the MMP-13 regulatory network in the pathogenesis of early osteoarthritis. In *Arthritis Research and Therapy*. <https://doi.org/10.1186/s13075-017-1454-2>
- Liao, W., Li, Z., Zhang, H., Li, J., Wang, K., & Yang, Y. (2015). Proteomic analysis of synovial fluid as an analytical tool to detect candidate biomarkers for knee osteoarthritis. *International Journal of Clinical and Experimental Pathology*
- Liu, C.-C., Lee, H.-C., Peng, Y.-S., Tseng, A. H., Wu, J.-L., Tsai, W.-Y., Wong, C.-S., & Su, L.-J. (2019). Transcriptome Analysis Reveals Novel Genes Associated with Cartilage Degeneration in Posttraumatic Osteoarthritis Progression. *CARTILAGE*. <https://doi.org/10.1177/1947603519847744>
- Liu, G., Cai, M., Zhou, F., & Liu, W. (2014). Charged polymer brushes-grafted hollow silica nanoparticles as a novel promising material for simultaneous joint lubrication and treatment. *Journal of Physical Chemistry B*. <https://doi.org/10.1021/jp500074g>
- Liu, J., Zhao, Z., Sun, Z., Liu, C., Cheng, X., Ruge, F., Yang, Y., Jiang, W. G., & Ye, L. (2020). Increased expression of Psoriasin is correlated with poor prognosis of bladder transitional cell carcinoma by promoting invasion and proliferation. *Oncology Reports*. <https://doi.org/10.3892/or.2019.7445>
- Liu-Bryan, R. (2013). Synovium and the innate inflammatory network in osteoarthritis progression topical collection on osteoarthritis. *Current Rheumatology Reports*. <https://doi.org/10.1007/s11926-013-0323-5>
- Loeser, R. F., Goldring, S. R., Scanzello, C. R., & Goldring, M. B. (2012). Osteoarthritis: A disease of the joint as an organ. In *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.34453>
- Lopez, H. L. (2012). Nutritional Interventions to Prevent and Treat Osteoarthritis. Part II: Focus on Micronutrients and Supportive Nutraceuticals. In *PM and R*. <https://doi.org/10.1016/j.pmrj.2012.02.023>
- Lu, Q. Y., Han, Q. H., Li, X., Li, Z. C., Pan, Y. T., Liu, L., & Fu, Q. G. (2014). Analysis of differentially expressed genes between rheumatoid arthritis and osteoarthritis based on the gene co-expression network. *Molecular Medicine Reports*. <https://doi.org/10.3892/mmr.2014.2166>

- Lyons, T. J., McClure, S. F., Stoddart, R. W., & McClure, J. (2006). The normal human chondro-osseous junctional region: Evidence for contact of uncalcified cartilage with subchondral bone and marrow spaces. *BMC Musculoskeletal Disorders*. <https://doi.org/10.1186/1471-2474-7-52>
- Ma, F., & Liu, D. (2012). Sketch of the edible bird's nest and its important bioactivities. In *Food Research International*. <https://doi.org/10.1016/j.foodres.2012.06.001>
- Madry, H., van Dijk, C. N., & Mueller-Gerbl, M. (2010). The basic science of the subchondral bone. *Knee Surgery, Sports Traumatology, Arthroscopy*. <https://doi.org/10.1007/s00167-010-1054-z>
- Madzuki, I. N., Lau, S. F., Che Ahmad Tantowi, N. A., Mohd Ishak, N. I., & Mohamed, S. (2018). *Labisia pumila* prevented osteoarthritis cartilage degeneration by attenuating joint inflammation and collagen breakdown in postmenopausal rat model. *Inflammopharmacology*. <https://doi.org/10.1007/s10787-018-0452-6>
- Mahjoub, M., Berenbaum, F., & Houard, X. (2012). Why subchondral bone in osteoarthritis? the importance of the cartilage bone interface in osteoarthritis. *Osteoporosis International*. <https://doi.org/10.1007/s00198-012-2161-0>
- Mahla, R. S. (2016). Stem cells applications in regenerative medicine and disease therapeutics. In *International Journal of Cell Biology*. <https://doi.org/10.1155/2016/6940283>
- Makarov, A. (2000). Electrostatic axially harmonic orbital trapping: A high-performance technique of mass analysis. *Analytical Chemistry*. <https://doi.org/10.1021/ac991131p>
- Man, G. S., & Mologhianu, G. (2014). Osteoarthritis pathogenesis - a complex process that involves the entire joint. In *Journal of medicine and life*
- Manderson, G. A., Martin, M., Önerfjord, P., Saxne, T., Schmidtchen, A., Mollnes, T. E., Heinegård, D., & Blom, A. M. (2009). Interactions of histidine-rich glycoprotein with immunoglobulins and proteins of the complement system. *Molecular Immunology*. <https://doi.org/10.1016/j.molimm.2009.07.011>
- Mankin, H. J. (1971). Biochemical and metabolic aspects of osteoarthritis. *The Orthopedic Clinics of North America*, 2(1), 19–31
- Marcone, M. F. (2005). Characterization of the edible bird's nest the "Caviar of the East." *Food Research International*. <https://doi.org/10.1016/j.foodres.2005.02.008>
- Marker, C. L., & Pomonis, J. D. (2012). The monosodium iodoacetate model of osteoarthritis pain in the rat. *Methods in Molecular Biology*. [https://doi.org/10.1007/978-1-61779-561-9\\_18](https://doi.org/10.1007/978-1-61779-561-9_18)

- Martel-pelletier, J. (2004). Pathophysiology of osteoarthritis. 31–33. <https://doi.org/10.1016/j.joca.2003.10.002>
- Martel-Pelletier, J., Barr, A. J., Cicuttini, F. M., Conaghan, P. G., Cooper, C., Goldring, M. B., Goldring, S. R., Jones, G., Teichtahl, A. J., & Pelletier, J. P. (2016). Osteoarthritis. In *Nature Reviews Disease Primers*. <https://doi.org/10.1038/nrdp.2016.72>
- Martin, J. A., Ellerbroek, S. M., & Buckwalter, J. A. (1997). Age-related decline in chondrocyte response to insulin-like growth factor-I: The role of growth factor binding proteins. *Journal of Orthopaedic Research*. <https://doi.org/10.1002/jor.1100150403>
- Matas, J., Orrego, M., Amenabar, D., Infante, C., Tapia-Limonchi, R., Cadiz, M. I., Alcayaga-Miranda, F., González, P. L., Muse, E., Khoury, M., Figueroa, F. E., & Espinoza, F. (2019). Umbilical Cord-Derived Mesenchymal Stromal Cells (MSCs) for Knee Osteoarthritis: Repeated MSC Dosing Is Superior to a Single MSC Dose and to Hyaluronic Acid in a Controlled Randomized Phase I/II Trial. *Stem Cells Translational Medicine*. <https://doi.org/10.1002/sctm.18-0053>
- Mateos, J., Lourido, L., Fernández-puente, P., Calamia, V., Fernández-lópez, C., Oreiro, N., Ruiz-romero, C., & Blanco, F. J. (2012). Differential protein profiling of synovial fluid from rheumatoid arthritis and osteoarthritis patients using LC – MALDI TOF / TOF ☆. *Journal of Proteomics*, 75(10), 2869–2878. <https://doi.org/10.1016/j.jprot.2011.12.042>
- Matharoo-Ball, B., Ball, G., & Rees, R. (2007). Clinical proteomics: Discovery of cancer biomarkers using mass spectrometry and bioinformatics approaches-A prostate cancer perspective. In *Vaccine*. <https://doi.org/10.1016/j.vaccine.2007.06.040>
- Matsukawa, N., Matsumoto, M., Bukawa, W., Chiji, H., Nakayama, K., Hara, H., & Tsukahara, T. (2011). Improvement of Bone Strength and Dermal Thickness Due to Dietary Edible Bird's Nest Extract in Ovariectomized Rats. *Bioscience, Biotechnology, and Biochemistry*, 75(3), 590–592. <https://doi.org/10.1271/bbb.100705>
- McWilliams, D. F., Leeb, B. F., Muthuri, S. G., Doherty, M., & Zhang, W. (2011). Occupational risk factors for osteoarthritis of the knee: A meta-analysis. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2011.02.016>
- Mellon, F. A. (2003). MASS SPECTROMETRY | Principles and Instrumentation. In *Encyclopedia of Food Sciences and Nutrition*. <https://doi.org/10.1016/b0-12-227055-x/00746-x>
- Michael, J. W., Schluter-Brust, K. U., & Eysel, P. (2010). The epidemiology, etiology, diagnosis, and treatment of osteoarthritis of the knee. *Dtsch Arztebl Int*, 107(9), 152–162. <https://doi.org/10.3238/arztebl.2010.0152>



- Michel, B. A., Stucki, G., Frey, D., De Vathaire, F., Vignon, E., Bruehlmann, P., & Uebelhart, D. (2005). Chondroitins 4 and 6 sulfate in osteoarthritis of the knee: A randomized, controlled trial. *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.20867>
- Milz, S., & Putz, R. (1994). Quantitative morphology of the subchondral plate of the tibial plateau. *Journal of Anatomy*
- Min, T., Sheng, L. Y., Chao, C., Jian, T., Guang, G. S., & Hua, L. G. (2015). Correlation between osteopontin and caveolin-1 in the pathogenesis and progression of osteoarthritis. *Experimental and Therapeutic Medicine*. <https://doi.org/10.3892/etm.2015.2433>
- Miosge, N., Hartmann, M., Maelicke, C., & Herken, R. (2004). Expression of collagen type I and type II in consecutive stages of human osteoarthritis. *Histochemistry and Cell Biology*. <https://doi.org/10.1007/s00418-004-0697-6>
- Mobasheri, A. (2011). Applications of proteomics to osteoarthritis, a musculoskeletal disease characterized by aging. *Frontiers in Physiology*. <https://doi.org/10.3389/fphys.2011.00108>
- Mobasheri, A., & Henrotin, Y. (2010). Identification, validation and qualification of biomarkers for osteoarthritis in humans and companion animals: Mission for the next decade. In *Veterinary Journal*. <https://doi.org/10.1016/j.tvjl.2010.05.026>
- Munier, C. C., Ottmann, C., & Perry, M. W. D. (2021). 14-3-3 modulation of the inflammatory response. In *Pharmacological Research*. <https://doi.org/10.1016/j.phrs.2020.105236>
- Musumeci, G., Trovato, F. M., Pichler, K., Weinberg, A. M., Loreto, C., & Castrogiovanni, P. (2013). Extra-virgin olive oil diet and mild physical activity prevent cartilage degeneration in an osteoarthritis model: An *in vivo* and *in vitro* study on lubricin expression. *Journal of Nutritional Biochemistry*. <https://doi.org/10.1016/j.jnutbio.2013.07.007>
- Myers, S. L. (1999). Synovial fluid markers in osteoarthritis. In *Rheumatic Disease Clinics of North America* (Vol. 25, Issue 2, pp. 433–449). [https://doi.org/10.1016/S0889-857X\(05\)70077-6](https://doi.org/10.1016/S0889-857X(05)70077-6)
- Nakagawa, H., Hama, Y., Sumi, T., Li, S. C., Maskos, K., Kalayanamitra, K., Mizumoto, S., Sugahara, K., & Li, Y. T. (2007). Occurrence of a nonsulfated chondroitin proteoglycan in the dried saliva of *Collocalia swiftlets* (edible bird's-nest). *Glycobiology*, 17(2), 157–164. <https://doi.org/10.1093/glycob/cwl058>

- Nakashima, T., Hayashi, M., Fukunaga, T., Kurata, K., Oh-Hora, M., Feng, J. Q., Bonewald, L. F., Kodama, T., Wutz, A., Wagner, E. F., Penninger, J. M., & Takayanagi, H. (2011). Evidence for osteocyte regulation of bone homeostasis through RANKL expression. *Nature Medicine*. <https://doi.org/10.1038/nm.2452>
- Naveen, S. V., Ahmad, R. E., Hui, W. J., Suhaeb, A. M., Murali, M. R., Shanmugam, R., & Kamarul, T. (2013). Histology, glycosaminoglycan level and cartilage stiffness in monoiodoacetate-induced osteoarthritis: Comparative analysis with anterior cruciate ligament transection in rat model and human osteoarthritis. *International Journal of Medical Sciences*. <https://doi.org/10.7150/ijms.6964>
- Neogi, T., & Zhang, Y. (2013). Epidemiology of OA. *Rheum Dis Clin North Am*. <https://doi.org/10.1016/j.rdc.2012.10.0>
- Ng, H. Y., Alvin Lee, K. X., & Shen, Y. F. (2017). Articular Cartilage: Structure, Composition, Injuries and Repair. *JSM Bone and Joint Dis*.
- Nguyen, K. C. T., & Cho, K. A. (2017). Versatile Functions of Caveolin-1 in Aging-related Diseases. *Chonnam Medical Journal*. <https://doi.org/10.4068/cmj.2017.53.1.28>
- Nguyen, L. T., Sharma, A. R., Chakraborty, C., Saibaba, B., Ahn, M. E., & Lee, S. S. (2017). Review of prospects of biological fluid biomarkers in osteoarthritis. In *International Journal of Molecular Sciences*. <https://doi.org/10.3390/ijms18030601>
- Niu, J., Zhang, Y. Q., Torner, J., Nevitt, M., Lewis, C. E., Aliabadi, P., Sack, B., Clancy, M., Sharma, L., & Felson, D. T. (2009). Is obesity a risk factor for progressive radiographic knee osteoarthritis? *Arthritis Care and Research*. <https://doi.org/10.1002/art.24337>
- Nurul Nadia, M., Babji, A. S., Ayub, M. K., & Nur 'Aliah, D. (2017). Effect of Enzymatic Hydrolysis on Antioxidant Capacity of Cave Edible Bird's Nests Hydrolysate. *International Journal of ChemTech Research*
- Nylund, R., Lemola, E., Hartwig, S., Lehr, S., Acheva, A., Jahns, J., Hildebrandt, G., & Lindholm, C. (2014). Profiling of low molecular weight proteins in plasma from locally irradiated individuals. *Journal of Radiation Research*. <https://doi.org/10.1093/jrr/rru007>
- Oates, K. M. N., Krause, W. E., & Colby, R. H. (2002). Using rheology to probe the mechanism of joint lubrication: Polyelectrolyte/protein interactions in synovial fluid. *Materials Research Society Symposium - Proceedings*. <https://doi.org/10.1557/proc-711-ff4.7.1>

- Osborn, T. M., Verdrengh, M., Stossel, T. P., Tarkowski, A., & Bokarewa, M. (2008). Research article Decreased levels of the gelsolin plasma isoform in patients with rheumatoid arthritis. *10*(5), 1–9. <https://doi.org/10.1186/ar2520>
- Ostergaard, K., Andersen, C. B., Petersen, J., Bendtzen, K., & Salter, D. M. (1999). Validity of histopathological grading of articular cartilage from osteoarthritic knee joints. *Annals of the Rheumatic Diseases*. <https://doi.org/10.1136/ard.58.4.208>
- Palotie, A., Ott, J., Elima, K., Cheah, K., Väisänen, P., Ryhänen, L., Vikkula, M., Vuorio, E., & Peltonen, L. (1989). Predisposition To Familial Osteoarthritis Linked To Type II Collagen Gene. *The Lancet*. [https://doi.org/10.1016/S0140-6736\(89\)92507-5](https://doi.org/10.1016/S0140-6736(89)92507-5)
- Pan, J., Zhou, X., Li, W., Novotny, J. E., Doty, S. B., & Wang, L. (2009). In situ measurement of transport between subchondral bone and articular cartilage. *Journal of Orthopaedic Research*. <https://doi.org/10.1002/jor.20883>
- Parasuraman, S., R, A., Balamurugan, S., Muralidharan, S., Kumar, K. J., & Vijayan, V. (2014). An Overview of Liquid Chromatography-Mass Spectroscopy Instrumentation. *Pharmaceutical Methods*. <https://doi.org/10.5530/phm.2014.2.2>
- Park, S., Hwang, I. K., Kim, S. Y., & Lee, S. (2006). Characterization of plasma gelsolin as a substrate for matrix metalloproteinases. 1192–1199. <https://doi.org/10.1002/pmic.200500402>
- Pásztói, M., Nagy, G., Géher, P., Lakatos, T., Tóth, K., Wellinger, K., Pócza, P., György, B., Holub, M. C., Kittel, Á., Pálóczy, K., Mazán, M., Nyirkos, P., Falus, A., & Buzas, E. I. (2009). Gene expression and activity of cartilage degrading glycosidases in human rheumatoid arthritis and osteoarthritis synovial fibroblasts. *Arthritis Research and Therapy*. <https://doi.org/10.1186/ar2697>
- Pauly, H. M., Larson, B. E., Coatney, G. A., Button, K. D., DeCamp, C. E., Fajardo, R. S., Haut, R. C., & Haut Donahue, T. L. (2015). Assessment of cortical and trabecular bone changes in two models of post-traumatic osteoarthritis. *Journal of Orthopaedic Research*. <https://doi.org/10.1002/jor.22975>
- Permy, M., Guede, D., López-Peña, M., Muñoz, F., Caeiro, J. R., & González-Cantalapiedra, A. (2015). Effects of diacerein on cartilage and subchondral bone in early stages of osteoarthritis in a rabbit model. *BMC Veterinary Research*. <https://doi.org/10.1186/s12917-015-0458-x>
- Peters, A. E., Akhtar, R., Comerford, E. J., & Bates, K. T. (2018). The effect of ageing and osteoarthritis on the mechanical properties of cartilage and bone in the human knee joint. *Scientific Reports*. <https://doi.org/10.1038/s41598-018-24258-6>

- Piccioli, P., & Rubartelli, A. (2013). The secretion of IL-1 $\beta$  and options for release. In *Seminars in Immunology*. <https://doi.org/10.1016/j.smim.2013.10.007>
- Piskin, A., Gulbabar, M. Y., Tomak, Y., Gulman, B., Hokelek, M., Kerimoglu, S., Koksak, B., Alic, T., & Kabak, Y. B. (2007). Osteoarthritis models after anterior cruciate ligament resection and medial meniscectomy in rats: A histological and immunohistochemical study. *Saudi Medical Journal*
- Pitcher, T., Sousa-Valente, J., & Malcangio, M. (2016). The monoiodoacetate model of osteoarthritis pain in the mouse. *Journal of Visualized Experiments*. <https://doi.org/10.3791/53746>
- Pitsillides, A. A., & Beier, F. (2011). Cartilage biology in osteoarthritis - Lessons from developmental biology. In *Nature Reviews Rheumatology*. <https://doi.org/10.1038/nrrheum.2011.129>
- Plotkin, L. I., Gortazar, A. R., Davis, H. M., Condon, K. W., Gabilondo, H., Maycas, M., Allen, M. R., & Bellido, T. (2015). Inhibition of osteocyte apoptosis prevents the increase in osteocytic receptor activator of nuclear factor  $\kappa$ B Ligand (RANKL) but does not stop bone resorption or the loss of bone induced by unloading. *Journal of Biological Chemistry*. <https://doi.org/10.1074/jbc.M115.642090>
- Pradhan, A. D., Rifai, N., & Ridker, P. M. (2002). Soluble intercellular adhesion molecule-1, soluble vascular adhesion molecule-1, and the development of symptomatic peripheral arterial disease in men. *Circulation*. <https://doi.org/10.1161/01.CIR.0000025636.03561.EE>
- Pritzker, K. P. H., Gay, S., Jimenez, S. A., Ostergaard, K., Pelletier, J. P., Revell, K., Salter, D., & van den Berg, W. B. (2006). Osteoarthritis cartilage histopathology: Grading and staging. *Osteoarthritis and Cartilage*, 14(1), 13–29
- Quintero-Fabián, S., Arreola, R., Becerril-Villanueva, E., Torres-Romero, J. C., Arana-Argáez, V., Lara-Riegos, J., Ramírez-Camacho, M. A., & Alvarez-Sánchez, M. E. (2019). Role of Matrix Metalloproteinases in Angiogenesis and Cancer. In *Frontiers in Oncology*. <https://doi.org/10.3389/fonc.2019.01370>
- Radin, E. L., Burr, D. B., Caterson, B., Fyhrrie, D., Brown, T. D., & Boyd, R. D. (1991). Mechanical determinants of osteoarthrosis. *Seminars in Arthritis and Rheumatism*. [https://doi.org/10.1016/0049-0172\(91\)90036-Y](https://doi.org/10.1016/0049-0172(91)90036-Y)
- Radin, E. L., Martin, R. B., Burr, D. B., Caterson, B., Boyd, R. D., & Goodwin, C. (1984). Effects of mechanical loading on the tissues of the rabbit knee. *Journal of Orthopaedic Research*. <https://doi.org/10.1002/jor.1100020303>

- Ramachandran, R., Babji, A. S., & Sani, N. A. (2018). Antihypertensive potential of bioactive hydrolysate from edible bird's nest. *AIP Conference Proceedings*. <https://doi.org/10.1063/1.5028014>
- Rashed, A. A., & Wan Nazaim, W. M. (2010). Effect of Edible Bird's Nest on Caco-2 Cell Proliferation. *Journal of Food Technology*. <https://doi.org/10.3923/jftech.2010.126.130>
- Rigoglou, S., & Papavassiliou, A. G. (2013). The NF- $\kappa$ B signalling pathway in osteoarthritis. *The International Journal of Biochemistry & Cell Biology*, 45(11), 2580–2584. <https://doi.org/10.1016/j.biocel.2013.08.018>
- Roh, K. B., Lee, J., Kim, Y. S., Park, J., Kim, J. H., Lee, J., & Park, D. (2012). Mechanisms of edible bird's nest extract-induced proliferation of human adipose-derived stem cells. *Evidence-Based Complementary and Alternative Medicine*. <https://doi.org/10.1155/2012/797520>
- Rose, B. J., & Kooyman, D. L. (2016). A Tale of Two Joints: The Role of Matrix Metalloproteases in Cartilage Biology. 2016. <https://doi.org/10.1155/2016/4895050>
- Rosenberg, J. H., Rai, V., Dilisio, M. F., Sekundiak, T. D., & Agrawal, D. K. (2017). Increased expression of damage-associated molecular patterns (DAMPs) in osteoarthritis of human knee joint compared to hip joint. *Molecular and Cellular Biochemistry*. <https://doi.org/10.1007/s11010-017-3078-x>
- Roth, S. H. (2013). Diclofenac in the treatment of osteoarthritis. *International Journal of Clinical Rheumatology*. <https://doi.org/10.2217/ijr.13.5>
- Rutgers, M., van Pelt, M. J. P., Dhert, W. J. A., Creemers, L. B., & Saris, D. B. F. (2010). Evaluation of histological scoring systems for tissue-engineered, repaired and osteoarthritic cartilage. In *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2009.08.009>
- Saad, S., Bendall, L. J., Gottlieb, D. J., Bradstock, K. F., & Overall, C. M. (2002). Cancer cell-associated fibronectin induces release of matrix metalloproteinase-2 from normal fibroblasts. *Cancer Research*
- Salter, D. M., Su, S. L., & Lee, H. S. (2014). Epidemiology and genetics of osteoarthritis. In *Journal of Medical Sciences (Taiwan)*. <https://doi.org/10.4103/1011-4564.147251>
- Saltzman, B. M., Leroux, T., Meyer, M. A., Basques, B. A., Chahal, J., Bach, B. R., Yanke, A. B., & Cole, B. J. (2017). The Therapeutic Effect of Intra-articular Normal Saline Injections for Knee Osteoarthritis: A Meta-analysis of Evidence Level 1 Studies. *American Journal of Sports Medicine*. <https://doi.org/10.1177/0363546516680607>

- Sanchez, C., Deberg, M. A., Bellahcène, A., Castronovo, V., Msika, P., Delcour, J. P., Crielaard, J. M., & Henrotin, Y. E. (2008). Phenotypic characterization of osteoblasts from the sclerotic zones of osteoarthritic subchondral bone. *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.23159>
- Sarkar, A., Carvalho, E., D'Souza, A. A., & Banerjee, R. (2019). Liposome-encapsulated fish oil protein-tagged gold nanoparticles for intra-articular therapy in osteoarthritis. *Nanomedicine*. <https://doi.org/10.2217/nnm-2018-0221>
- Scotece, M., Koskinen-Kolasa, A., Moilanen, T., Moilanen, E., & Vuolteenaho, K. (2018). Novel adipokine associated with osteoarthritis: retinol binding protein 4 is produced by cartilage and correlates with matrix metalloproteinases in osteoarthritis patients. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2018.02.276>
- Sen, R. (2020). High-throughput approaches of diagnosis and therapies for COVID-19: antibody panels, proteomics and metabolomics. In *Future Drug Discovery*. <https://doi.org/10.4155/fdd-2020-0027>
- Shah, K., & Sumer, H. (2019). Outcome of safety and efficacy of allogeneic mesenchymal stromal cell derived from umbilical cord for the treatment of osteoarthritis in a randomized blinded placebo-controlled trial. *Annals of Translational Medicine*. <https://doi.org/10.21037/atm.2019.06.39>
- Shirai, T., Kobayashi, M., Nishitani, K., Satake, T., Kuroki, H., Nakagawa, Y., & Nakamura, T. (2011). Chondroprotective effect of alendronate in a rabbit model of osteoarthritis. *Journal of Orthopaedic Research*. <https://doi.org/10.1002/jor.21394>
- Silacci, P., Mazzolai, L., Gauci, C., Stergiopoulos, N., Yin, H. L., & Hayoz, D. (2004). Gelsolin superfamily proteins: Key regulators of cellular functions. In *Cellular and Molecular Life Sciences*. <https://doi.org/10.1007/s00018-004-4225-6>
- Sipe, J. D. (1995). Acute-phase proteins in osteoarthritis. *Seminars in Arthritis and Rheumatism*. [https://doi.org/10.1016/S0049-0172\(95\)80020-4](https://doi.org/10.1016/S0049-0172(95)80020-4)
- Smyth, T., Harris, H. J., Brown, A., Töttemeyer, S., Farnfield, B. A., Maskell, D. J., Matsumoto, M., Plevin, R., Alldridge, L. C., & Bryant, C. E. (2006). Differential modulatory effects of annexin 1 on nitric oxide synthase induction by lipopolysaccharide in macrophages. *Immunology*. <https://doi.org/10.1111/j.1365-2567.2005.02307.x>

- Sniekers, Y. H., Intema, F., Lafeber, F. P. J. G., van Osch, G. J. V. M., van Leeuwen, J. P. T. M., Weinans, H., & Mastbergen, S. C. (2008). A role for subchondral bone changes in the process of osteoarthritis; a micro-CT study of two canine models. *BMC Musculoskeletal Disorders*, 9(1), 20. <https://doi.org/10.1186/1471-2474-9-20>
- Sophia Fox, A. J., Bedi, A., & Rodeo, S. A. (2009). The basic science of articular cartilage: Structure, composition, and function. *Sports Health*. <https://doi.org/10.1177/1941738109350438>
- Spector, T. D., Cicuttini, F., Baker, J., Loughlin, J., & Hart, D. (1996). Genetic influences on osteoarthritis in women: a twin study. *BMJ*. <https://doi.org/10.1136/bmj.312.7036.940>
- Spencer A. Johnson. (2015). Osteoarthritis: Joint Anatomy, Physiology, and Pathobiology. *Veterinary Clinics of North America: Small Animal Practice*
- Speth, C., Rambach, G., Würzner, R., Lass-Flörl, C., Kozarcanin, H., Hamad, O. A., Nilsson, B., & Ekdahl, K. N. (2015). Complement and platelets: Mutual interference in the immune network. In *Molecular Immunology*. <https://doi.org/10.1016/j.molimm.2015.03.244>
- Srikanth, V. K., Fryer, J. L., Zhai, G., Winzenberg, T. M., Hosmer, D., & Jones, G. (2005). A meta-analysis of sex differences prevalence, incidence and severity of osteoarthritis. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2005.04.014>
- Stenberg, J., Rüetschi, U., Skiöldebrand, E., Kärrholm, J., & Lindahl, A. (2013). Quantitative proteomics reveals regulatory differences in the chondrocyte secretome from human medial and lateral femoral condyles in osteoarthritic patients. *Proteome Science*. <https://doi.org/10.1186/1477-5956-11-43>
- Strassle, B. W., Mark, L., Leventhal, L., Piesla, M. J., Jian Li, X., Kennedy, J. D., Glasson, S. S., & Whiteside, G. T. (2010). Inhibition of osteoclasts prevents cartilage loss and pain in a rat model of degenerative joint disease. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2010.06.007>
- Sugimoto, M. A., Vago, J. P., Teixeira, M. M., & Sousa, L. P. (2016). Annexin A1 and the Resolution of Inflammation: Modulation of Neutrophil Recruitment, Apoptosis, and Clearance. In *Journal of Immunology Research*. <https://doi.org/10.1155/2016/8239258>
- Sulzbacher, I. (2013). Osteoarthritis: histology and pathogenesis. *Wiener Medizinische Wochenschrift*, 163(9–10), 212–219. <https://doi.org/10.1007/s10354-012-0168-y>

- Sun, Y., Zhang, H., Wang, Y., & Wang, Y. (2017). Charged polymer brushes-coated mesoporous silica nanoparticles for osteoarthritis therapy: a combination between hydration lubrication and drug delivery. *Journal of Controlled Release*. <https://doi.org/10.1016/j.jconrel.2017.03.114>
- Suri, S., & Walsh, D. A. (2012). Osteochondral alterations in osteoarthritis. *Bone*. <https://doi.org/10.1016/j.bone.2011.10.010>
- Takahashi, I., Matsuzaki, T., & Hosono, M. (2017). Long-term histopathological developments in knee-joint components in a rat model of osteoarthritis induced by monosodium iodoacetate. *Journal of Physical Therapy Science*. <https://doi.org/10.1589/jpts.29.590>
- Takahashi, K., Kubo, T., Goomer, R. S., Amiel, D., Kobayashi, K., Imanishi, J., Teshima, R., & Hirasawa, Y. (1997). Analysis of heat shock proteins and cytokines expressed during early stages of osteoarthritis in a mouse model. *Osteoarthritis and Cartilage*. [https://doi.org/10.1016/S1063-4584\(97\)80036-2](https://doi.org/10.1016/S1063-4584(97)80036-2)
- Tang, D. D. (2018). The Dynamic Actin Cytoskeleton in Smooth Muscle. In *Advances in Pharmacology*. <https://doi.org/10.1016/bs.apha.2017.06.001>
- Tat, S. K., Pelletier, J. P., Lajeunesse, D., Fahmi, H., Duval, N., & Martel-Pelletier, J. (2008). Differential modulation of RANKL isoforms by human osteoarthritic subchondral bone osteoblasts: Influence of osteotropic factors. *Bone*. <https://doi.org/10.1016/j.bone.2008.04.006>
- Teepel, E., Jay, G. D., Elsaid, K. A., & Fleming, B. C. (2013). Animal models of osteoarthritis: Challenges of model selection and analysis. In *AAPS Journal*. <https://doi.org/10.1208/s12248-013-9454-x>
- Teh, S.-S., & Ma, Z.-F. (2018). Bioactive Components and Pharmacological Properties of Edible Bird's Nest. *International Proceedings of Chemical, Biological and Environmental Engineering*
- Thomas, A., Eichenberger, G., Kempton, C., Pape, D., York, S., Decker, A. M., & Kohia, M. (2009). Recommendations for the treatment of knee osteoarthritis, using various therapy techniques, based on categorizations of a literature review. In *Journal of Geriatric Physical Therapy*. <https://doi.org/10.1519/00139143-200932010-00007>
- Ucar, H. I., Tok, M., Atalar, E., Dogan, O. F., Oc, M., Farsak, B., Guvener, M., Yilmaz, M., Dogan, R., Demircin, M., & Pasaoglu, I. (2007). Predictive significance of plasma levels of interleukin-6 and high-sensitivity C-reactive protein in atrial fibrillation after coronary artery bypass surgery. *The Heart Surgery Forum*. <https://doi.org/10.1532/HSF98.20061175>



- Usui, M., Xing, L., Drissi, H., Zuscik, M., O'Keefe, R., Chen, D., & Boyce, B. F. (2008). Murine and chicken chondrocytes regulate osteoclastogenesis by producing RANKL in response to BMP2. *Journal of Bone and Mineral Research*. <https://doi.org/10.1359/jbmr.071025>
- Valdes, A. M., Evangelou, E., Kerkhof, H. J. M., Tamm, A., Doherty, S. A., Kisand, K., Tamm, A., Kerna, I., Uitterlinden, A., Hofman, A., Rivadeneira, F., Cooper, C., Dennison, E. M., Zhang, W., Muir, K. R., Ioannidis, J. P. A., Wheeler, M., Maciewicz, R. A., Van Meurs, J. B., ... Doherty, M. (2011). The GDF5 rs143383 polymorphism is associated with osteoarthritis of the knee with genome-wide statistical significance. In *Annals of the Rheumatic Diseases*. <https://doi.org/10.1136/ard.2010.134155>
- Van Der Poll, T., & Wiersinga, W. J. (2017). Sepsis. *Infectious Diseases*, 415-426.e1. <https://doi.org/10.1016/B978-0-7020-6285-8.00047-2>
- Verzijl, N., Bank, R. A., TeKoppele, J. M., & DeGroot, J. (2003). AGEing and osteoarthritis: A different perspective. In *Current Opinion in Rheumatology*. <https://doi.org/10.1097/00002281-200309000-00016>
- Verzijl, N., DeGroot, J., Zaken, C. Ben, Braun-Benjamin, O., Maroudas, A., Bank, R. A., Mizrahi, J., Schalkwijk, C. G., Thorpe, S. R., Baynes, J. W., Bijlsma, J. W. J., Lafeber, F. P. J. G., & TeKoppele, J. M. (2002). Crosslinking by advanced glycation end products increases the stiffness of the collagen network in human articular cartilage: A possible mechanism through which age is a risk factor for osteoarthritis. *Arthritis and Rheumatism*. [https://doi.org/10.1002/1529-0131\(200201\)46:1<114::AID-ART10025>3.0.CO;2-P](https://doi.org/10.1002/1529-0131(200201)46:1<114::AID-ART10025>3.0.CO;2-P)
- Vignon, E., Bejui, J., Mathieu, P., Hartmann, J. D., Ville, G., Evreux, J. C., & Descotes, J. (1987). Histological cartilage changes in a rabbit model of osteoarthritis. *Journal of Rheumatology*
- Vignon, E., Garnero, P., Delmas, P., Avouac, B., Bettica, P., Boers, M., Ehrich, E., MacKillop, N., Rovati, L., Serni, U., Spector, T., & Reginster, J. Y. (2001). Recommendations for the registration of drugs used in the treatment of osteoarthritis: An update on biochemical markers. *Osteoarthritis and Cartilage*, 9(4), 289–293. <https://doi.org/10.1053/joca.2000.0387>
- Vinod, E., Boopalan, P. R. J. V. C., Arumugam, S., & Sathishkumar, S. (2018). Creation of monosodium iodoacetate-induced model of osteoarthritis in rabbit knee joint. In *Indian Journal of Medical Research*. [https://doi.org/10.4103/ijmr.IJMR\\_2004\\_16](https://doi.org/10.4103/ijmr.IJMR_2004_16)
- Wachsmuth, L., Keiffer, R., Juretschke, H. P., Raiss, R. X., Kimmig, N., & Lindhorst, E. (2003). *In vivo* contrast-enhanced micro MR-imaging of experimental osteoarthritis in the rabbit knee joint at 7.1T1. *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2003.08.008>

- Wakefield, R. J., Gibbon, W. W., & Emery, P. (1999). The current status of ultrasonography in rheumatology. In *Rheumatology*. <https://doi.org/10.1093/rheumatology/38.3.195>
- Wang, L. J., Zeng, N., Yan, Z. P., Li, J. T., & Ni, G. X. (2020). Post-traumatic osteoarthritis following ACL injury. In *Arthritis Research and Therapy*. <https://doi.org/10.1186/s13075-020-02156-5>
- Wanner, J. P., Subbaiah, R., Skomorovska-prokvolit, Y., Shishani, Y., Boilard, E., Mohan, S., Gillespie, R., Miyagi, M., & Gobezie, R. (2013). Proteomic profiling and functional characterization of early and late shoulder osteoarthritis. *Arthritis Research & Therapy*, 15(6), 1. <https://doi.org/10.1186/ar4369>
- Weyrich, A. S. (2014). Platelets: More than a sack of glue. *Hematology*. <https://doi.org/10.1182/asheducation-2014.1.400>
- Wojdasiewicz, P., Poniatowski, A. A., & Szukiewicz, D. (2014). The Role of Inflammatory and Anti-Inflammatory Cytokines in the Pathogenesis of Osteoarthritis. 2014. <https://doi.org/10.1155/2014/561459>
- Woo, S. L.-Y., Buckwalter, J. A., & Fung, Y. C. (1989). Injury and Repair of the Musculoskeletal Soft Tissues. *Journal of Biomechanical Engineering*. <https://doi.org/10.1115/1.3168347>
- Woolf, A. D., & Pfleger, B. (2003). Burden of major musculoskeletal conditions. *Bulletin of the World Health Organization*, 81(9), 646–656. <https://doi.org/S0042-96862003000900007> [pii]
- World Health Organization. (2013). Chronic rheumatic conditions. In *Chronic diseases and health promotion*
- Wright, G. L., & Semmes, O. J. (2003). Proteomics in Health and Disease. In *Journal of Biomedicine and Biotechnology*. <https://doi.org/10.1155/S1110724303002997>
- Xie, L., Lin, A. S. P., Kundu, K., Levenston, M. E., Murthy, N., & Guldberg, R. E. (2012). Quantitative imaging of cartilage and bone morphology, reactive oxygen species, and vascularization in a rodent model of osteoarthritis. *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.34370>
- Xu, J., Yan, L., Yan, B., Zhou, L., Tong, P., & Shan, L. (2020). Osteoarthritis pain model induced by intra-articular injection of mono-iodoacetate in rats. *Journal of Visualized Experiments*. <https://doi.org/10.3791/60649>

- Yamagiwa, H., Sarkar, G., Charlesworth, M. C., McCormick, D. J., & Bolander, M. E. (2003). Two-dimensional gel electrophoresis of synovial fluid: Method for detecting candidate protein markers for osteoarthritis. *Journal of Orthopaedic Science*, 8(4), 482–490. <https://doi.org/10.1007/s00776-003-0657-3>
- Yang, Y., & Morand, E. (2006). The Anti-Inflammatory Role of Annexin-1 in Arthritis. *Current Rheumatology Reviews*. <https://doi.org/10.2174/157339706778699887>
- Yates, J. R. (2011). A century of mass spectrometry: From atoms to proteomes. In *Nature Methods*. <https://doi.org/10.1038/nmeth.1659>
- Yida, Z., Imam, U. U., & Ismail, M. (2014). *In vitro* bioaccessibility and antioxidant properties of edible bird's nest following simulated human gastro-intestinal digestion. *BMC Complementary and Alternative Medicine*. <https://doi.org/10.1186/1472-6882-14-468>
- Yu, L., Wang, L., & Chen, S. (2010). Endogenous toll-like receptor ligands and their biological significance. *Journal of Cellular and Molecular Medicine*. <https://doi.org/10.1111/j.1582-4934.2010.01127.x>
- Yuan, X. L., Meng, H. Y., Wang, Y. C., Peng, J., Guo, Q. Y., Wang, A. Y., & Lu, S. B. (2014). Bone-cartilage interface crosstalk in osteoarthritis: Potential pathways and future therapeutic strategies. In *Osteoarthritis and Cartilage*. <https://doi.org/10.1016/j.joca.2014.05.023>
- Zainal Abidin, F., Hui, C., Luan, N., Mohd Ramli, E. S., Hun, L., & Abd Ghafar, N. (2011). Effects of edible bird's nest (EBN) on cultured rabbit corneal keratocytes. *BMC Complementary and Alternative Medicine*, 11(1), 94. <https://doi.org/10.1186/1472-6882-11-94>
- Zhang, Y., Hunter, D. J., Nevitt, M. C., Xu, L., Niu, J., Lui, L. Y., Yu, W., Aliabadi, P., & Felson, D. T. (2004). Association of Squatting with Increased Prevalence of Radiographic Tibiofemoral Knee Osteoarthritis: The Beijing Osteoarthritis Study. *Arthritis and Rheumatism*. <https://doi.org/10.1002/art.20127>
- Zou, H., Stoppani, E., Volonte, D., & Galbiati, F. (2011). Caveolin-1, cellular senescence and age-related diseases. *Mechanisms of Ageing and Development*. <https://doi.org/10.1016/j.mad.2011.11.0>