

Radiofrequency waves increase the brain levels of inflammatory biomarkers, neurotrophin and serotonin

Mansour Azimzadeh^{1,2}, Fatemeh Radmard¹ and Gholamali Jelodar^{1*}

¹ Department of Physiology, School of Veterinary Medicine, Shiraz University, Shiraz, Iran.

² Department of Biomedical Science, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, Selangor, Malaysia.

* Correspondence: jelodar@shirazu.ac.ir; Tel.: +987136138757

Received: 18 January 2024; **Accepted:** 28 April 2024; **Published:** 21 June 2024

Edited by: Sharmili Vidyadaran (Universiti Putra Malaysia, Malaysia)

Reviewed by: Nur Fariesha Md Hashim (Universiti Putra Malaysia, Malaysia);

Gaik Theng Toh (Taylor's University Lakeside Campus, Malaysia)

<https://doi.org/10.31117/neuroscirn.v7i2.326>

Abstract: The increase in mobile technology has raised concerns about the potential health effects of mobile phone radiation. The biological impact of exposure to radiofrequency (RF) waves emitted by electronic devices has been extensively studied and is a concern for the public, policymakers, and health researchers. The study aimed to examine the impact of 900 MHz radiofrequency waves on biomarkers such as interleukin (IL)-1 α , IL-1 β , tumour necrosis factor (TNF)- α , homocysteine, nerve growth factor, and serotonin in rats' serum and brain tissue. Thirty adult male Sprague Dawley rats (200 \pm 20g) were randomly assigned to three groups (n=10): control (not exposed to RF), exposed I (2 hours per day), and exposed II (4 hours per day). The exposed groups were exposed to 900 MHz RFW for 30 consecutive days. The results showed that only the exposed group II significantly increased serum serotonin levels compared to the control group (P=0.0496). IL-1 α , TNF- α , and nerve growth factor levels in brain tissue increased significantly in both exposed groups compared to the control group (P<0.0001). The control group had significantly lower levels of IL-1 β compared to exposed groups I (P=0.0289) and II (P=0.0004). Additionally, serotonin and homocysteine levels in the brains of exposed II were significantly higher compared to the other groups (P<0.0001). The results showed disruptions in all biomarkers, indicating the potential impacts of daily exposure to 900 MHz radiofrequency waves from mobile phones on brain function. This suggests that mobile phone radiation may affect brain function.

Keywords: Mobile phone; Radiofrequency; Cytokines; Homocysteine; Nerve growth factor; Serotonin

©2024 by Azimzadeh *et al.* for use and distribution according to the Creative Commons Attribution (CC BY-NC 4.0) license (<https://creativecommons.org/licenses/by-nc/4.0/>), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original author and source are credited.

1.0 INTRODUCTION

Radiofrequency waves (RFW) are non-ionizing radiation used in mobile phones, wireless networks, and other electronic devices. The long-term effects of exposure to these electromagnetic fields (EMFs) on brain function and health are still not fully understood and are the

subject of ongoing research and debate. As mobile phone usage becomes more common, it is important to understand the effects of long-term exposure to radiofrequency EMF, which falls within the frequency range of 3 kHz to 300 GHz. The Global System for Mobile Communications (GSM) uses 900 MHz radiofrequency

(RF) ([The International Commission on Non-Ionizing Radiation Protection, 2020](#)).

Research has focused on understanding potential health risks and biological mechanisms associated with EMF exposure. Studies have shown that EMF radiation can have detrimental effects on the central nervous system (CNS), including impacts on the blood-brain barrier (BBB), brain trace element balance, memory function, synaptic plasticity, neurotransmitter release, and neuronal viability ([Azimzadeh & Jelodar, 2020a, 2020b](#); [Bertagna et al., 2021](#); [Sirav & Seyhan, 2016](#)).

Cytokines are multifunctional proteins that immune system cells produce in response to injury or pathogens. In the brain, activated neuronal and glial cells continuously produce cytokines ([Bourgognon & Cavanagh, 2020](#)), which play a role in various functions, such as neuronal development ([Monet & Quan, 2023](#)), sleep regulation ([Krueger, 2008](#)), synaptic plasticity, neurotransmitter metabolism ([Zipp et al., 2023](#)), neuroendocrine functions, and BBB modifications ([Yang et al., 2022](#)). IL-1 is a crucial regulator of inflammation and stress in the central nervous system by controlling various innate immune responses ([Park et al., 2018](#)). TNF- α plays a vital role in different central nervous system functions, including synaptic homeostasis, transmission, and scaling. It also influences excitotoxicity, neuroinflammation, and the permeability of the BBB ([Freseigna et al., 2020](#)). Exposure to EMFs has been shown to increase levels of certain cytokines at both the protein and mRNA levels ([Wu et al., 2012](#)).

Homocysteine (Hcy) is a sulfur-containing amino acid that is derived from methionine and is known to be a potent pro-inflammatory factor, stimulating the production of pro-inflammatory cytokines ([Borowska et al., 2021](#); [Li et al., 2015](#)). Elevated levels of Hcy are a well-established risk factor for vascular disorders, brain atrophy, and Alzheimer's disease ([Smith et al., 2018](#)).

Nerve Growth Factor (NGF) is a crucial neurotrophin that supports the development and survival of specific neurons in the CNS and peripheral nervous systems (PNS). It is mainly expressed in the hippocampus, olfactory, and cortex regions, and the sympathetic ganglia ([Berry et al., 2012](#)). Inflammatory conditions, such as cerebral ischemia and reperfusion, can trigger NGF expression in neurons ([Li et al., 2022](#)). NGF also regulates communication between the nervous and immune systems. It is produced by various brain cells, including astrocytes, glial cells, neurons, lymphocytes, and mast cells. It is found in different brain regions, such

as the hippocampus, cortex, basal forebrain, cerebellum, and brainstem ([Minnone et al., 2017](#)).

Serotonin is a neurotransmitter that regulates mood, anxiety, and happiness ([Hensler, 2010](#)). It plays a crucial role in neurodevelopment, learning and memory, neuropsychiatric diseases, and autonomic regulation ([Shah et al., 2018](#); [Witteveen et al., 2013](#)). While the majority of serotonin (90%) is produced in the gastrointestinal tract, a small percentage (1–2%) is produced by neurons in the brain ([Terry & Margolis, 2017](#)). Serotonin in platelets affects cytokine production and is involved in inflammation ([Jenne & Kubes, 2015](#); [Li et al., 2012](#)). Exposure to 1800 MHz for 1 and 2 months significantly increased hippocampal serotonin levels ([Aboul Ezz et al., 2013](#)). Additionally, exposure to 900 MHz RFW for 45 minutes significantly increased plasma serotonin levels ([Eris et al., 2015](#)).

Previous studies have shown that exposure to 900 MHz RFW can have harmful effects on the pancreas and testis tissue, leading to decreased testosterone levels and increased levels of specific biomarkers such as cytokines (IL-1 β and TNF- α) and homocysteine ([Azimzadeh & Jelodar, 2019](#); [Jelodar et al., 2021](#)). Additionally, exposure to RFW from mobile phones can have a greater impact on the brain due to its weak protective enzymes and high lipid content, making it susceptible to lipid peroxidation and oxidative stress ([Fang et al., 2013](#)). In this study, we investigated the effects of 900 MHz RFW on regulatory and functional biomarkers IL-1 α and β , TNF- α , Hcy, NGF, and serotonin in the brain tissue and serum of rats. Our findings demonstrated significant changes in cytokines, homocysteine, NGF, and serotonin levels in brain tissue following exposure to radiofrequency waves, while serum parameters remained essentially unchanged except for serotonin levels.

2.0 MATERIALS AND METHODS

2.1 Animals

Thirty adult male Sprague Dawley rats weighing 200 ± 20 g were obtained from the Shiraz animal house centre. The rats were housed in polycarbonate cages ($42 * 26.5 * 15$ cm³) with a constant temperature of $20 \pm 2^\circ\text{C}$ and a 12-hour light-dark cycle. They had free access to food and water. The exposure time was set between 9 a.m. and 1 p.m. All experiments were conducted in compliance with Shiraz University's ethical guidelines (Code No.IR.AC.REC. 1398.S9530650) and the National Institutes of Health's Guide for the Care and Use of Laboratory Animals ([National Research Council, 2011](#)).

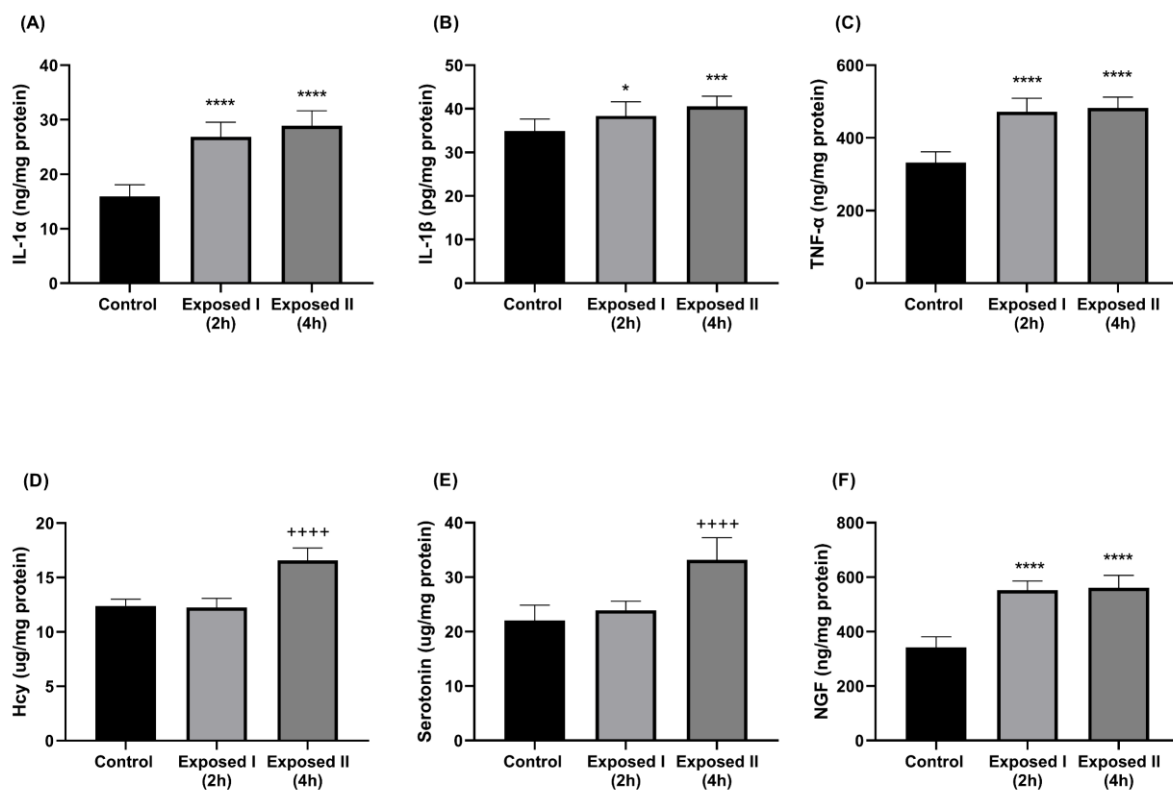


Figure 1. Effects of 900 MHz RFW exposure for 2 and 4 hours per day over 30 consecutive days on the concentrations of (A) IL-1 α , (B) IL-1 β , (C) TNF- α , (D) Hcy, (E) serotonin, and (F) NGF in brain tissue. The columns represent the mean \pm SD, One-way ANOVA, and Tukey's multiple comparison tests. * $p < 0.05$; *** $p < 0.001$; and **** $p < 0.0001$ compared to the control group. +++++ $p < 0.0001$ compared to the control and exposed group I.

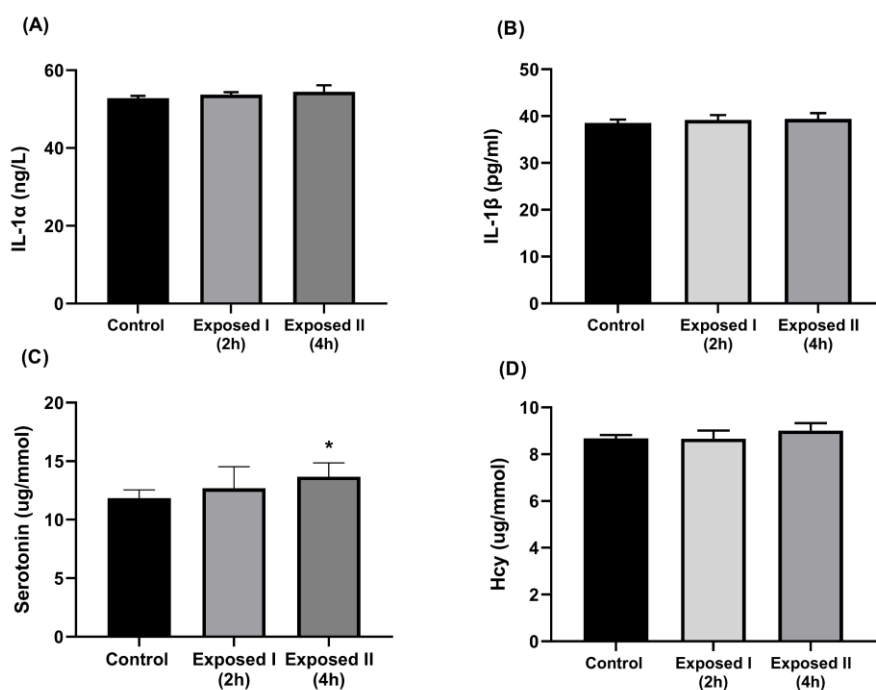


Figure 2. Effects of 900 MHz RFW exposure for 2 and 4 hours per day over 30 consecutive days on the concentrations of (A) IL-1 α , (B) IL-1 β , (C) serotonin, and (D) Hcy levels in the serum. The columns represent the mean \pm SD, One-way ANOVA, and Tukey's multiple comparison tests. * $p < 0.05$ compared to the control group.

2.2 The radiofrequency wave exposure device and power dosimetry

We used a simulator developed by Shiraz University's Faculty of Telecommunication and Electronics Engineering to generate an electromagnetic field at a frequency of 900 MHz. The simulator has a 12 cm antenna that emits 900 MHz RFW circularly. The output power of the simulator was monitored using a spectrum analyzer (FSH6, Rohde and Schwarz, Germany), with the antenna of the spectrum analyzer placed 1 meter away from the simulator during the measurements. The MCS Real-Time Spectrum Analyzer Software recorded the real-time readings during the test. The peak power density recorded from the proximity of the simulator in the downlink band was 0.6789 mW/cm² at 876 MHz. We also detected a specific absorption rate (SAR) value of 0.035 W/kg using a field-probe device (300 kHz -18 GHz, Wave Control, Spain) ([Azimzadeh & Jelodar, 2020c](#)).

2.3 Experimental protocol

All rats were divided into three groups of 10 each: control group (with no exposure); exposed I group (2 hours per day); and exposed II group (4 hours per day). The signal generator was placed one meter away from the cages of the exposed groups. Both exposed groups were irradiated to 900 MHz RFW for 30 consecutive days ([Azimzadeh et al., 2018](#)).

2.4 Sampling and tissue preparation

On the last day of the exposure period, all animals were anaesthetized with a 2% diethyl ether-saturated cotton ball in a chamber for 3–5 minutes and then euthanized by collecting whole blood through heart puncture. The blood was collected in glass tubes and left to clot at room temperature for thirty minutes. It was then centrifuged at 1300 g for 10 minutes. The serum was collected and stored at -70°C for later analysis. Brain tissue was rapidly removed, isolated, and stored at -70°C. The brain tissue was washed once with distilled water, homogenized using a tissue grinder on ice and centrifuged at 2000 rpm for 20 minutes at 4°C to collect the resulting supernatant. The concentration of biomarkers (serotonin, Hcy, NGF, TNF- α , IL-1 α , and β) was measured in the serum and brain supernatant using the ELISA kits according to the manufacturer's instructions (serotonin: IBL, Hamburg, Germany; Hcy: Diazyme, Shanghai, China; NGF, TNF- α , IL-1 α , and β : Crystal Day Biotech, Shanghai, China). The total protein concentration of the brain tissue was evaluated using the Bradford method ([Bradford, 1976](#)).

2.5 Statistical analysis

The data were presented as mean \pm standard deviation (SD). Statistical analysis was performed using one-way ANOVA followed by Tukey's multiple comparison tests with Graph Pad Prism® 8.0.1. A p-value of <0.05 was considered statistically significant in all experiments.

3.0 RESULTS

3.1 The effects of RFW exposure on the brain tissue

Figure 1 shows the concentration of all studied biomarkers in the brain. The mean levels of IL-1 α , IL-1 β , and TNF- α in the exposed groups (I and II) were significantly higher compared to the control group. The one-way ANOVA revealed a significant difference in the mean IL1- α levels between the studied groups (F (2, 24) = 68.12, P<0.0001). The post hoc test indicated that both exposed groups I and II had significantly higher mean IL1- α levels compared to the control group (P<0.0001) (**Figure 1A**). Similarly, there was a significant difference in the mean levels of IL-1 β among the groups (F (2, 27) = 10.20, P=0.0005, one-way ANOVA). The control group had significantly lower mean levels of IL-1 β compared to exposed groups I (P=0.0289) and II (P=0.0004) (**Figure 1B**). The one-way ANOVA also indicated a significant difference in the mean TNF- α concentration (F (2, 27) = 67.67, P<0.0001). The mean TNF- α levels in the control group were significantly lower than in both exposed I and II groups (P<0.0001) (**Figure 1C**).

The mean levels of Hcy, serotonin, and NGF in the brains of the studied groups showed significant differences (Hcy: F (2, 27) = 73.84, P<0.0001; serotonin: F (2, 21) = 31.27, P<0.0001; NGF: F (2, 27) = 99.02, P<0.0001, one-way ANOVA). Post hoc analysis revealed that the exposed II group had significantly higher mean Hcy and serotonin levels than the exposed I and control groups (P<0.0001, Tukey's multiple comparison tests). Additionally, both exposed groups had significantly higher mean NGF levels compared to the control group (P<0.0001, Tukey's multiple comparison tests) (**Figure 1D-F**).

3.2 The effects of RFW exposure on the serum parameters

Figure 2 shows the levels of IL-1 α , IL1- β , Hcy, and serotonin in the serum of the experimental groups (**Figure 2A-D**). The only significant change observed in the serum was an increase in serotonin concentration (F (2, 26) = 4.414, P=0.0224, one-way ANOVA). Post hoc analysis revealed a significant increase in serotonin levels in group II compared to the control group (P=0.0172, Tukey's multiple comparison tests).

(Figure 2C). It should be noted that due to technical issues, we could not evaluate the TNF- α and NGF levels in the serum.

4.0 DISCUSSION

The study found that exposure to 900 MHz RFW had a significant impact on biomarkers in the brain, while only a few changes were observed in the serum. Inflammatory cytokines (IL-1 α , IL-1 β , and TNF- α) showed a significant increase in the brain tissue in both exposed groups compared to the control group. These results demonstrated an increase in the inflammatory markers assessed, reinforcing concerns about the biological effects of such radiation. The findings suggest that radiofrequency exposure may trigger inflammation in the brain, consistent with previous experiments showing elevated inflammatory cytokine levels in other tissues following exposure to 900 MHz RFW or microwave radiation ([Azimzadeh & Jelodar, 2019](#); [Jelodar et al., 2021](#); [Wu et al., 2012](#)). Increased levels of these mediators have been associated with diseases such as depression, Alzheimer's, and epilepsy ([Bourgognon & Cavanagh, 2020](#)). Elevated levels of IL-1 α and IL-1 β in brain tissue did not correlate with changes in serum levels, suggesting localized production of these cytokines.

Research on the effects of EMF on cytokine levels is limited, and the findings are conflicting. Some studies suggest that short-term exposure to EMF can increase innate immunity cytokines, while long-term exposure may decrease the adaptive immune response ([Mahaki et al., 2019](#)). Exposure to EMF has also been associated with increased production of inflammatory cytokines and reactive oxygen species ([Kim et al., 2017](#); [Patrundo et al., 2018](#)). However, other studies have found different effects ([Mahaki et al., 2020](#)), and some have reported no effects of EMF exposure on cytokine production ([Fan et al., 2015](#); [Ikeda et al., 2002](#)). The discrepancies in findings may be due to differences in study design and methodologies.

The Hcy levels in group II brains were significantly higher than in the exposed I and control groups ($P < 0.05$). Previous research has shown that exposure to 900 MHz RFW increased Hcy levels in pancreatic tissue, with no significant change in testicular tissue ([Jelodar et al., 2021](#)). To our knowledge, there are no other published reports on the effects of EMF on Hcy levels in tissues.

The brain lacks metabolic pathways to eliminate Hcy, making neurons and glial cells more susceptible to its toxic effects, impacting neuronal survival and signalling

([Boldyrev et al., 2013](#); [Škovierová et al., 2015](#)). Our study did not find significant changes in serum Hcy levels, supporting this hypothesis. However, high Hcy levels can adversely affect the CNS, including increased excitatory neurotransmission, neuronal damage, and compromised BBB integrity ([Lehotský et al., 2016](#)). High homocysteine (HHcy) and nitrosative stress can also compromise the integrity of the BBB, leading to cerebrovascular permeability and neuronal degeneration ([Kamat et al., 2016](#)). HHcy inhibits nitric oxide production and bioavailability, induces reactive oxygen species generation, and increases the production of inflammatory cytokines ([Li et al., 2015](#)). Our study found that long-term exposure to 900 MHz RFW led to a significant increase in Hcy levels in brain tissue, accompanied by an increase in the production of inflammatory cytokines (IL-1 α , IL-1 β , and TNF- α), which may have detrimental effects on the CNS.

The levels of NGF in the brain tissues of both exposed groups (I and II) were significantly higher compared to the control group. An increase in NGF may be a compensatory response to counteract the stress and potential damage caused by exposure to RFW. However, persistently elevated levels of neurotrophins may lead to dysregulation of neuronal growth and function, potentially contributing to neuropathology. Previous studies have also reported changes in NGF levels in various tissues following exposure to 900 MHz RFW or pulsed electromagnetic fields ([Azimzadeh & Jelodar, 2019](#); [Jelodar et al., 2021](#); [Longo et al., 1999](#)). Differences in tissue type, size, and capacity for NGF generation may explain variations in results. NGF can activate innate immune responses and regulate inflammation to prevent tissue damage ([Minnone et al., 2017](#)). It can increase inflammatory cytokines ([Bayas et al., 2003](#); [Hepburn et al., 2014](#)) and stimulate the release of anti-inflammatory cytokines ([Liew et al., 2005](#)). The significant increase in NGF concentration in both exposed groups following exposure to RFW may modulate inflammatory cytokines.

Exposure to 900 MHz RF increased serotonin levels in both the serum and brain. This finding is particularly interesting because it suggests that exposure to RFW may affect emotional states and cognitive functions regulated by serotonin. Altered serotonin levels have complex implications. Increased serotonin may improve mood and reduce anxiety, but imbalances in serotonin levels have been linked to psychiatric disorders such as depression and schizophrenia ([Celada et al., 2013a](#); [Jenkins et al., 2016](#)).

Previous research has also found decreased pancreatic serotonin levels following similar exposure ([Jelodar et al., 2021](#)). Studies on the effects of EMF on neurotransmitters such as serotonin, acetylcholine, and catecholamines have shown conflicting results. For example, exposure to 1800 MHz EMF increased serotonin and decreased dopamine levels in the hippocampus and hypothalamus ([Aboul Ezz et al., 2013](#)), while exposure to 900 MHz RFW increased blood serotonin levels ([Eris et al., 2015](#)). Furthermore, exposure to 800 MHz EMF decreased the release of acetylcholine, while exposure to 1800 MHz EMF had no effect ([Li et al., 2017](#); [Testylier et al., 2002](#)). Additionally, exposure to different frequencies (900, 1800, 2100 MHz) reduced brain serotonin levels in newborn rats ([Ismail et al., 2015](#)).

Several potential mechanisms have been proposed for increased serotonin levels following RFW exposure. These include increased catabolism due to the heightened activity of monoamine oxidase ([Said et al., 2012](#)), EMF-induced damage to the ileal mucosa ([Herrera et al., 1995](#)), and decreased synthesis and absorption of the serotonin precursor tryptophan. Since 90% of the body's serotonin is produced in the gastrointestinal tract, the significant increase in serum serotonin concentrations in the current study is due to this source. Additionally, exposure to different EMF frequencies can modify blood-brain barrier permeability ([Salford et al., 2003](#); [Zhou et al., 2013](#)), suggesting that the primary source of increased brain serotonin levels is the serum, originating from the gastrointestinal tract.

Serotonin-producing neurons are widely distributed in the brain, and evidence shows that several serotonin receptor subtypes are densely expressed throughout the brain ([Carhart-Harris & Nutt, 2017](#); [Celada et al., 2013b](#)). Pro-inflammatory cytokines such as TNF- α and IL-1 β have been reported to positively correlate with elevated brain serotonin levels ([Masson & Hamon, 2009](#)), indicating that serotonergic neurons also

contribute to the increase in serotonin concentration in the brain.

Further research, including replication studies and investigations into underlying mechanisms, may be needed to establish the significance and broader implications of the findings fully. Nonetheless, the findings contribute valuable insights into the potential effects of radiofrequency wave exposure on brain health, laying the groundwork for future research and discussions in the scientific community.

5.0 CONCLUSION

Our study revealed that daily exposure to 900 MHz radiofrequency waves for 30 days resulted in significant alterations in cytokines (IL-1 α , IL-1 β , and TNF- α), Hcy, NGF, and serotonin levels in the brain tissue with long-term exposure (4 hours). Short-term exposure also caused significant changes in cytokines (IL-1 α , IL-1 β , and TNF- α), and NGF levels in the brain tissue. There were no significant changes in serum parameters, except for serotonin levels, indicating that the effects of electromagnetic fields may be limited to specific tissues. These findings suggest that radiofrequency waves (900 MHz) could disrupt brain function by affecting the neuroendocrine, neurotransmitter, and immune systems.

Acknowledgements:

We thank the Department of Telecommunication and Electronics Engineering at Shiraz University for providing the signal generator. Additionally, we are grateful for the technical support from Professor Naziefies's laboratory.

Author Contributions:

All authors participated in the design, interpretation of the studies, analysis of the data, and review of the manuscript; MA and FR conducted the experiments. GJ supervised, designed the study, and edited the article.

Conflicts of Interest:

The authors declared no conflict of interest.

REFERENCES

- Aboul Ezz, H. S., Khadrawy, Y. A., Ahmed, N. A., Radwan, N. M., & El Bakry, M. M. (2013). The effect of pulsed electromagnetic radiation from mobile phones on the levels of monoamine neurotransmitters in four different areas of rat brain. *European Review for Medical and Pharmacological Sciences*, 17(13), 1782–1788. <https://pubmed.ncbi.nlm.nih.gov/23852905>
- Azimzadeh, M., & Jelodar, G. (2019). Alteration of testicular regulatory and functional molecules following long-time exposure to 900 MHz RFW emitted from BTS. *Andrologia*, 51(9), e13372. <https://doi.org/10.1111/and.13372>
- Azimzadeh, M., & Jelodar, G. (2020a). The protective effect of vitamin supplementation (E and E + C) on passive avoidance learning and memory during exposure to 900 MHz RFW emitted from BTS. *Toxicology and Industrial Health*, 36(2), 93–98. <https://doi.org/10.1177/0748233720912058>

- Azimzadeh, M., & Jelodar, G. (2020b). Trace elements homeostasis in brain exposed to 900 MHz RFW emitted from a BTS–antenna model and the protective role of vitamin E. *Journal of Animal Physiology and Animal Nutrition*, 104(5), 1568–1574. <https://doi.org/10.1111/jpn.13360>
- Azimzadeh, M., & Jelodar, G. (2020c). Prenatal and early postnatal exposure to radiofrequency waves (900 MHz) adversely affects passive avoidance learning and memory. *Toxicology and Industrial Health*, 36(12), 1024–1030. <https://doi.org/10.1177/0748233720973143>
- Azimzadeh, M., Jelodar, G., Namazi, F., & Soleimani, F. (2018). Exposure to radiofrequency wave (RFW) generated by a base transceiver stations (BTS) antenna model affects learning and memory in female more than male rats. *International Journal of Radiation Research*, 16(4), 487–491. <http://ijrr.com/article-1-2402-en.html>
- Bayas, A., Kruse, N., Moriabadi, N., Weber, F., Hummel, V., Wohleben, G., Gold, R., Toyka, K., & Rieckmann, P. (2003). Modulation of cytokine mRNA expression by brain-derived neurotrophic factor and nerve growth factor in human immune cells. *Neuroscience Letters*, 335(3), 155–158. [https://doi.org/10.1016/s0304-3940\(02\)01152-7](https://doi.org/10.1016/s0304-3940(02)01152-7)
- Berry, A., Bindocci, E., & Alleva, E. (2012). NGF, brain and behavioral plasticity. *Neural Plasticity*, 2012, 1–9. <https://doi.org/10.1155/2012/784040>
- Bertagna, F., Lewis, R., Silva, S. R. P., McFadden, J., & Jeevaratnam, K. (2021). Effects of electromagnetic fields on neuronal ion channels: a systematic review. *Annals of the New York Academy of Sciences*, 1499(1), 82–103. <https://doi.org/10.1111/nyas.14597>
- Boldyrev, A., Bryushkova, E., Mashkina, A., & Vladychenskaya, E. (2013). Why is homocysteine toxic for the nervous and immune systems? *Current Aging Science*, 6(1), 29–36. <https://doi.org/10.2174/18746098112059990007>
- Borowska, M., Winiarska, H., Dworacka, M., Wesołowska, A., Dworacki, G., & Mikołajczak, P. Ł. (2021). The effect of homocysteine on the secretion of IL-1B, IL-6, IL-10, IL-12 and RANTES by peripheral blood mononuclear cells - an In vitro study. *Molecules*, 26(21), 6671. <https://doi.org/10.3390/molecules26216671>
- Bourgognon, J., & Cavanagh, J. (2020). The role of cytokines in modulating learning and memory and brain plasticity. *Brain and Neuroscience Advances*, 4, 239821282097980. <https://doi.org/10.1177/2398212820979802>
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1–2), 248–254. [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- Carhart-Harris, R., & Nutt, D. (2017). Serotonin and brain function: a tale of two receptors. *Journal of Psychopharmacology*, 31(9), 1091–1120. <https://doi.org/10.1177/0269881117725915>
- Celada, P., Bortolozzi, A., & Artigas, F. (2013a). Serotonin 5-HT1A receptors as targets for agents to treat psychiatric disorders: rationale and current status of research. *CNS Drugs*, 27(9), 703–716. <https://doi.org/10.1007/s40263-013-0071-0>
- Celada, P., Puig, M. V., & Artigas, F. (2013b). Serotonin modulation of cortical neurons and networks. *Frontiers in Integrative Neuroscience*, 7, 25. <https://doi.org/10.3389/fnint.2013.00025>
- Eris, A. H., Kiziltan, H. S., Meral, I., Genc, H., Trabzon, M., Seyithanoglu, H., Yagci, B., & Uysal, O. (2015). Effect of short-term 900 MHz low level electromagnetic radiation exposure on blood serotonin and glutamate levels. *Bratislava Medical Journal*, 116(02), 101–103. https://doi.org/10.4149/bll_2015_019
- Fan, W., Qian, F., Ma, Q., Zhang, P., Chen, T., Chen, C., Zhang, Y., Deng, P., Zhou, Z., & Yu, Z. (2015). 50 Hz electromagnetic field exposure promotes proliferation and cytokine production of bone marrow mesenchymal stem cells. *International Journal of Clinical and Experimental Medicine*, 8(5), 7394–7404. <https://pubmed.ncbi.nlm.nih.gov/26221281>
- Fang, K., Cheng, F., Huang, Y., Chung, S., Jian, Z., & Lin, M. (2013). Trace element, antioxidant activity, and lipid peroxidation levels in brain cortex of gerbils after cerebral ischemic injury. *Biological Trace Element Research*, 152(1), 66–74. <https://doi.org/10.1007/s12011-012-9596-1>
- Freseigna, D., Bullitta, S., Musella, A., Rizzo, F. R., De Vito, F., Guadalupi, L., Caioli, S., Balletta, S., Sanna, K., Dolcetti, E., Vanni, V., Bruno, A., Buttari, F., Bassi, M. S., Mandolesi, G., Centonze, D., & Gentile, A. (2020). Re-examining the role of TNF in MS pathogenesis and therapy. *Cells*, 9(10), 2290. <https://doi.org/10.3390/cells9102290>
- Hensler, J. G. (2010). Serotonin in mood and emotion. *Handbook of Behavioral Neuroscience*, 21, 367–378. [https://doi.org/10.1016/s1569-7339\(10\)70090-4](https://doi.org/10.1016/s1569-7339(10)70090-4)
- Hepburn, L., Prajsnar, T. K., Klapholz, C., Moreno, P., Loynes, C. A., Ogryzko, N. V., Brown, K., Schiebler, M., Hegyi, K., Antrobus, R., Hammond, K. L., Connolly, J., Ochoa, B., Bryant, C., Otto, M., Surewaard, B., Seneviratne, S. L., Grogono, D. M., Cachat, J., . . . Floto, R. A. (2014). A Spaetzle-like role for nerve growth factor in vertebrate immunity to *Staphylococcus aureus*. *Science*, 346(6209), 641–646. <https://doi.org/10.1126/science.1258705>
- Herrera, J. L., Vigneulle, R. M., Gage, T., MacVittie, T. J., Nold, J. B., & Dubois, A. (1995). Effect of radiation and radioprotection on small intestinal function in canines. *Digestive Diseases and Sciences*, 40(1), 211–218. <https://doi.org/10.1007/bf02063968>

- Ikedo, K., Shinmura, Y., Mizoe, H., Yoshizawa, H., Yoshida, A., Kanao, S., Sumitani, H., Hasebe, S., Motomura, T., Yamakawa, T., Mizuno, F., Otaka, Y., & Hirose, H. (2002). No effects of extremely low frequency magnetic fields found on cytotoxic activities and cytokine production of human peripheral blood mononuclear cells in vitro. *Bioelectromagnetics*, 24(1), 21–31. <https://doi.org/10.1002/bem.10062>
- Ismail, S., Ali, R., Hassan, H., & Abd El-Rahman, D. (2015). Effect of exposure to electromagnetic fields (EMFs) on monoamine neurotransmitters of newborn rats. *Biochemistry & Physiology*, 4(2), 156. <https://doi.org/doi:10.4172/2168-9652.1000156>
- Jelodar, G., Azimzadeh, M., Radmard, F., & Darvishhoo, N. (2021). Alteration of intrapancreatic serotonin, homocysteine, TNF- α , and NGF levels as predisposing factors for diabetes following exposure to 900-MHz waves. *Toxicology and Industrial Health*, 37(8), 496–503. <https://doi.org/10.1177/07482337211022634>
- Jenkins, T. A., Nguyen, J. C. D., Polglaze, K. E., & Bertrand, P. P. (2016). Influence of tryptophan and serotonin on mood and cognition with a possible role of the gut-brain axis. *Nutrients*, 8(1), 56. <https://doi.org/10.3390/nu8010056>
- Jenne, C. N., & Kubes, P. (2015). Platelets in inflammation and infection. *Platelets*, 26(4), 286–292. <https://doi.org/10.3109/09537104.2015.1010441>
- Kamat, P. K., Kyles, P., Kalani, A., & Tyagi, N. (2016). Hydrogen sulfide ameliorates homocysteine-Induced Alzheimer's Disease-Like pathology, Blood-Brain barrier disruption, and synaptic disorder. *Molecular Neurobiology*, 53(4), 2451–2467. <https://doi.org/10.1007/s12035-015-9212-4>
- Kim, J. H., Yu, D., Huh, Y. H., Lee, E. H., Kim, H., & Kim, H. R. (2017). Long-term exposure to 835 MHz RF-EMF induces hyperactivity, autophagy and demyelination in the cortical neurons of mice. *Scientific Reports*, 7, 41129. <https://doi.org/10.1038/srep41129>
- Krueger, J. M. (2008). The role of cytokines in sleep regulation. *Current Pharmaceutical Design*, 14(32), 3408–3416. <https://doi.org/10.2174/138161208786549281>
- Lehotský, J., Tothová, B., Kovalská, M., Dobrota, D., Beňová, A., Kalenská, D., & Kaplán, P. (2016). Role of homocysteine in the ischemic stroke and development of ischemic tolerance. *Frontiers in Neuroscience*, 10, 538. <https://doi.org/10.3389/fnins.2016.00538>
- Li, C., Li, J., Li, Y., Lang, S., Yougbaré, I., Zhu, G., Chen, P., & Ni, H. (2012). Crosstalk between platelets and the immune system: old systems with new discoveries. *Advances in Hematology*, 2012, 1–14. <https://doi.org/10.1155/2012/384685>
- Li, F., Chang, J., Lv, Y., Xu, D., Chen, J., & Sun, X. (2017). Impact of electromagnetic irradiation produced by 3G mobile phone on brain neurotransmitters in mice during growth and development period. *Biomedical Research*, 28(14), 6220–6224. <https://www.alliedacademies.org/articles/impact-of-electromagnetic-irradiation-produced-by-3g-mobile-phone-on-brain-neurotransmitters-in-mice-during-growth-and-development-7959.html>
- Li, J., Li, Q., Du, H., Wang, Y., You, S., Wang, F., Xu, X., Cheng, J., Cao, Y., Liu, C., & Hu, L. (2015). Homocysteine triggers inflammatory responses in macrophages through inhibiting CSE-H2S signaling via DNA hypermethylation of CSE promoter. *International Journal of Molecular Sciences*, 16(12), 12560–12577. <https://doi.org/10.3390/ijms160612560>
- Li, Y., Wu, F., Zhou, M., Zhou, J., Cui, S., Guo, J., Wu, J., & He, L. (2022). ProNGF/NGF modulates autophagy and apoptosis through PI3K/Akt/mTOR and ERK signaling pathways following cerebral ischemia-reperfusion in rats. *Oxidative Medicine and Cellular Longevity*, 2022, 1–16. <https://doi.org/10.1155/2022/6098191>
- Liew, F. Y., Xu, D., Brint, E. K., & O'Neill, L. a. J. (2005). Negative regulation of toll-like receptor-mediated immune responses. *Nature Reviews. Immunology*, 5(6), 446–458. <https://doi.org/10.1038/nri1630>
- Longo, F., Yang, T., Hamilton, S., Hyde, J., Walker, J., Jennes, L., Stach, R., & Siskin, B. (1999). Electromagnetic fields influence NGF activity and levels following sciatic nerve transection. *Journal of Neuroscience Research*, 55(2), 230–237. [https://doi.org/10.1002/\(sici\)1097-4547\(19990115\)55:2](https://doi.org/10.1002/(sici)1097-4547(19990115)55:2)
- Mahaki, H., Jabarivasal, N., Sardarian, K., & Zamani, A. (2020). Effects of various densities of 50 Hz electromagnetic field on serum IL-9, IL-10, and TNF-A levels. *International Journal of Occupational and Environmental Medicine*, 11(1), 24–32. <https://doi.org/10.15171/ijocem.2020.1572>
- Mahaki, H., Tanzadehpanah, H., Jabarivasal, N., Sardanian, K., & Zamani, A. (2019). A review on the effects of extremely low frequency electromagnetic field (ELF-EMF) on cytokines of innate and adaptive immunity. *Electromagnetic Biology and Medicine*, 38(1), 84–95. <https://doi.org/10.1080/15368378.2018.1545668>
- Masson, J., & Hamon, M. (2009). Monoamine transporters: Focus on the regulation of serotonin transporter by cytokines. *Encyclopedia of Neuroscience*, 2009, 921–929. <https://doi.org/10.1016/b978-008045046-9.01150-5>
- Minnone, G., De Benedetti, F., & Bracci-Laudiero, L. (2017). NGF and its receptors in the regulation of inflammatory response. *International Journal of Molecular Sciences*, 18(5), 1028. <https://doi.org/10.3390/ijms18051028>
- Monet, M. C., & Quan, N. (2023). Complex neuroimmune involvement in neurodevelopment: a mini-review. *Journal of Inflammation Research*, 16, 2979–2991. <https://doi.org/10.2147/jir.s410562>
- National Research Council. (2011). *Guide for the care and use of Laboratory animals* (8th ed.). National Academies Press, Washington. <https://doi.org/10.17226/12910>

- Park, S. Y., Kang, M. J., & Han, J. S. (2018). Interleukin-1 beta promotes neuronal differentiation through the Wnt5a/RhoA/JNK pathway in cortical neural precursor cells. *Molecular Brain*, 11(1), 39. <https://doi.org/10.1186/s13041-018-0383-6>
- Patrino, A., Ferrone, A., Costantini, E., Franceschelli, S., Pesce, M., Speranza, L., Amerio, P., D' Angelo, C., Felaco, M., Grilli, A., & Reale, M. (2018). Extremely low-frequency electromagnetic fields accelerates wound healing modulating MMP-9 and inflammatory cytokines. *Cell Proliferation*, 51(2), e12432. <https://doi.org/10.1111/cpr.12432>
- Said, U. Z., Saada, H. N., Abd-Alla, M. S., Elsayed, M. E., & Amin, A. M. (2012). Hesperidin attenuates brain biochemical changes of irradiated rats. *International Journal of Radiation Biology*, 88(8), 613–618. <https://doi.org/10.3109/09553002.2012.694008>
- Salford, L. G., Brun, A. E., Eberhardt, J. L., Malmgren, L., & Persson, B. R. R. (2003). Nerve cell damage in mammalian brain after exposure to microwaves from GSM mobile phones. *Environmental Health Perspectives*, 111(7), 881–883. <https://doi.org/10.1289/ehp.6039>
- Shah, R., Courtiol, E., Castellanos, F. X., & Teixeira, C. M. (2018). Abnormal serotonin levels during perinatal development lead to behavioral deficits in adulthood. *Frontiers in Behavioral Neuroscience*, 12, 114. <https://doi.org/10.3389/fnbeh.2018.00114>
- Sirav, B., & Seyhan, N. (2016). Effects of GSM modulated radio-frequency electromagnetic radiation on permeability of blood–brain barrier in male & female rats. *Journal of Chemical Neuroanatomy*, 75, 123–127. <https://doi.org/10.1016/j.chemneu.2015.12.010>
- Škovierová, H., Mahmood, S., Blahovcová, E., Hatok, J., Lehotský, J., & Murín, R. (2015). Effect of homocysteine on survival of human glial cells. *Physiological Research*, 64(5), 747–754. <https://doi.org/10.33549/physiolres.932897>
- Smith, A. D., Refsum, H., Bottiglieri, T., Fenech, M., Hooshmand, B., McCaddon, A., Miller, J. W., Rosenberg, I. H., & Obeid, R. (2018). Homocysteine and dementia: an international consensus statement. *Journal of Alzheimer's Disease*, 62(2), 561–570. <https://doi.org/10.3233/jad-171042>
- Terry, N., & Margolis, K. G. (2017). Serotonergic mechanisms regulating the GI tract: experimental evidence and therapeutic relevance. *Handbook of experimental pharmacology*, 239, 319–342. https://doi.org/10.1007/164_2016_103
- Testylier, G., Tonduli, L., Malabiau, R., & Debouzy, J. (2002). Effects of exposure to low level radiofrequency fields on acetylcholine release in hippocampus of freely moving rats. *Bioelectromagnetics*, 23(4), 249–255. <https://doi.org/10.1002/bem.10008>
- The International Commission on Non-Ionizing Radiation Protection (2020). Guidelines for limiting exposure to electromagnetic fields (100 kHz to 300 GHz). *Health Physics*, 118(5), 483-524. <https://doi.org/10.1097/hp.0000000000001210>
- Witteveen, J. S., Middelman, A., Van Hulten, J. A., Martens, G. J. M., Homberg, J. R., & Kolk, S. M. (2013). Lack of serotonin reuptake during brain development alters rostral raphe-prefrontal network formation. *Frontiers in Cellular Neuroscience*, 7, 143. <https://doi.org/10.3389/fncel.2013.00143>
- Wu, H., Wang, D., Shu, Z., Zhou, H., Zuo, H., Wang, S., Li, Y., Xu, X., Li, N., & Peng, R. (2012). Cytokines produced by microwave-radiated Sertoli cells interfere with spermatogenesis in rat testis. *Andrologia*, 44, 590–599. <https://doi.org/10.1111/j.1439-0272.2011.01232.x>
- Yang, J., Ran, M., Li, H., Lin, Y., Ma, K., Yang, Y., Fu, X., & Yang, S. (2022). New insight into neurological degeneration: Inflammatory cytokines and blood–brain barrier. *Frontiers in Molecular Neuroscience*, 15, 1013933. <https://doi.org/10.3389/fnmol.2022.1013933>
- Zhou, J. X., Ding, G. R., Zhang, J., Zhou, Y. C., Zhang, Y. J., & Guo, G. Z. (2013). Detrimental effect of electromagnetic pulse exposure on permeability of in vitro blood-brain-barrier model. *Biomedical and Environmental Sciences*, 26(2), 128–137. <https://doi.org/10.3967/0895-3988.2013.02.007>
- Zipp, F., Bittner, S., & Schafer, D. P. (2023). Cytokines as emerging regulators of central nervous system synapses. *Immunity*, 56(5), 914–925. <https://doi.org/10.1016/j.immuni.2023.04.011>