



**SITUATIONAL ANALYSIS ON THE ANTIMICROBIAL RESISTANCE  
PATTERNS AMONG BACTERIAL ISOLATES FROM DISEASED SMALL  
ANIMALS, LIVESTOCK AND WILDLIFE**

By

NURUL ASYIQIN BINTI HAULISAH

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of  
Master of Science**

**February 2021**

**FPV 2021 26**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement for the degree of Master of Science

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**February 2021**

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**Faculty : Veterinary Medicine**

Antimicrobial resistance (AMR) is a global threat of immense significance to the health of animals and humans. While there is an abundance of reports on AMR among clinical isolates from humans, information regarding the patterns of resistance among clinical isolates from animals is scarce. Hence a situational analysis of AMR among the clinical cases received at a veterinary diagnostic laboratory was performed to describe the pattern of resistance demonstrated by clinical isolates. Specifically, this study describes the AMR pattern of clinically important pathogens from pets and livestock using retrospective laboratory data between 2015 and 2017, and determine the resistance patterns of selected pathogens of public health significance from domestic and wildlife between 2018 and 2019. In diseased pets, *Escherichia coli* was the most commonly isolated ( $n=101$ , 13%) bacteria. Additionally, *E. coli* from the pets was highly resistant to amoxicillin (73.2%) and cephalexin (66.3%). Moreover, more than 75% of isolates from diseased pets were multi-drug resistant (MDR). Isolates from cats appeared to have higher level of resistance to multiple antibiotics compared to those from dogs. While in diseased livestock, *Escherichia coli* was the most commonly isolated ( $n=185$ , 26%) bacteria, and (>77%) were resistant to neomycin and streptomycin. Moreover, more than 76% of isolates from livestock were MDR. In general, isolates from non-ruminants have a higher level of resistance to multiple antibiotics compared to ruminants. In conclusion, this study has highlighted the trends and level of AMR among clinical isolates from pets and livestock based on routine antibiotic therapies. This study also discovered the trends of resistance among clinical isolates to those antibiotics of public health significance using the scope of testing as described by the national integrated surveillance for AMR.

Keywords: pet, livestock, wildlife, clinical isolates, AMR, MDR

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**ANALISIS SITUASI CORAK KERINTANGAN ANTIBIOTIK DI KALANGAN  
ISOLAT BAKTERIA DARI HAIWAN PELIHARAAN, TERNAKAN DAN  
HAIWAN LIAR SAKIT**

Oleh

**NURUL ASYIQIN BINTI HAULISAH**

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Kerintangan antibiotik (AMR) adalah ancaman global yang sangat penting kepada kesihatan haiwan dan manusia. Terdapat banyak laporan mengenai kerintangan antibiotik (AMR) dalam kalangan isolat klinikal dari manusia, tetapi maklumat dan laporan kerintangan antibiotik dalam kalangan isolat klinikal haiwan sangat jarang diperolehi. Oleh itu, analisis situasi mengenai kerintangan antibiotik (AMR) dalam kalangan kes-kes klinikal yang diterima dari makmal diagnostik veterinar dilakukan untuk menggambarkan corak kerintangan antibiotik yang ditunjukkan oleh isolat klinikal. Secara khususnya kajian ini menerangkan corak kerintangan antibiotik (AMR) terhadap patogen bakteria penting dari haiwan peliharaan dan dari ternakan menggunakan data retrospektif makmal antara tahun 2015 dan 2017, dan menentukan corak kerintangan antibiotik dari patogen bakteria terpilih yang mempunyai kepentingan kepada kesihatan awam dari haiwan domestik dan liar antara 2018 dan 2019. Dalam haiwan peliharaan sakit, *Escherichia coli* adalah bakteria yang paling sering dijumpai ( $n= 101$ , 13%). Selain itu, isolat *E. coli* dari haiwan peliharaan mempunyai tahap rintangan yang tinggi terhadap *amoxicillin* (73.2%) dan *cephalexin* (66.3%). Tambahan pula, lebih daripada 75% isolat dari kucing dan anjing yang sakit rintang terhadap pelbagai antibiotik (MDR). Isolat dari kucing kelihatan mempunyai tahap kerintangan yang lebih tinggi terhadap pelbagai antibiotik berbanding dengan anjing. Manakala, dalam haiwan ternakan sakit, *Escherichia coli* adalah bakteria yang paling kerap dijumpai ( $n= 185$ , 26%), dan (>77%) rintang terhadap *neomycin* dan *streptomycin*. Malahan, lebih daripada 76% isolat dari ternakan adalah MDR. Secara amnya, isolat dari bukan ruminan mempunyai tahap kerintangan yang lebih tinggi terhadap pelbagai antibiotik berbanding dengan ruminan. Kajian keratan rentas dilakukan di mana *E. coli*, *Klebsiella pneumoniae* dan *Salmonella* spp. telah diperiksa menggunakan protokol bersepada AMR. Analisis menunjukkan (>75%) bakteria terpilih ini rintang kepada *ampicillin*. Secara keseluruhan, 58.6% isolat ini adalah MDR. Kesimpulannya, penemuan

dari kajian ini telah memberi penekanan tentang tren dan tahap AMR di kalangan isolat klinikal dari haiwan peliharaan dan ternakan berdasarkan terapi antibiotik lazim. Kajian ini juga menemui tren ketahanan antara isolat klinikal terhadap antibiotik yang mempunyai kepentingan kesihatan awam dengan menggunakan skop pengujian seperti yang dijelaskan oleh pengawasan bersepadu nasional untuk pengawasan AMR.

Kata kunci: haiwan peliharaan, ternakan, haiwan liar, isolat klinikal, AMR, MDR

## **ACKNOWLEDGEMENTS**

*In The Name of ALLAH, the Most Gracious the Most Merciful*

All the praise to Allah, whom ultimately, we depend for sustenance and guidance, thanks to Him, His blessing makes this thesis successfully complete.

Firstly, I am grateful to my loving parents Haulisah Othman and Nor Aishah Ismail who brought me to life and sacrificed so much to raise me in a nurturing environment. Without their unconditional love, care, encouragement, and support I would not be here today.

I would like to extend my deepest gratitude to my supervisor Prof. Dr. Latiffah Hassan for her patience, guidance, encouragement and advice. She spent so much time helping me improve my thesis, giving valuable comments and suggestions, and responded to my questions and queries so promptly. I thank my co-supervisors Associate Prof. Dr. Siti Khairani Bejo and Dr. Nur Indah Ahmad for their advice, encouragement and support throughout my work. Without my supervisory committee invaluable support, this work would not have been completed.

Special thanks to Bacteriology Laboratory staff Cik Krishnammah Kuppusami, Cik Nur Rabiatuladawiyah Rosli, and Encik Mohammad Azri Roslan for their time, continuous support and sharing knowledge while I was collecting data and sample at the lab. Also, my thanks to Veterinary Public Health Laboratory Staff Puan Fauziah Nordin for her time and technical assistance throughout my study.

Last but not least, to my siblings, namely, Sofiah Haulisah and Muhamad Firdaus Haulisah and friends, Rita Rosmala Dewi, Khaleeda Azalea Dzulkifli, Sabrina Danial Leong, Aina Nazurah Khlubi, Siti Tasnim Makhtar, and Sherryl Nur Farahanim Ghaffar for unconditional love and support. To all who have involved directly and indirectly contributed to this project, my sincere thanks.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

|                      |   |
|----------------------|---|
| AGP                  | Antibiotic growth promotant   |
| AMR                  | Antimicrobial resistance  |
| ARG                  | Antibiotic resistance gene  |
| AST                  | Antibiotic susceptibility testing   |
| ATCC                 | American type culture collection  |
| BLFVM                | Bacteriology Laboratory, Faculty Veterinary Medicine                      |
| Cfu                  | Colony forming unit   |
| CLSI                 | Clinical and laboratory standards institute                               |
| DANMAP               | Danish integrated antibiotic resistance monitoring and research programme |
| DNA                  | Deoxyribonucleic acid   |
| DOF                  | Department of Fisheries   |
| DVS                  | Department of Veterinary Services   |
| <i>E. coli</i>       | <i>Escherichia coli</i>   |
| <i>E. faecalis</i>   | <i>Enterococcus faecalis</i>  |
| ESBL                 | Extended-β-lactamase  |
| FAO                  | Food and agriculture organization of the United Nations                   |
| FCR                  | Feed conversion ratio   |
| I                    | Intermediate  |
| <i>K. pneumoniae</i> | <i>Klebsiella pneumoniae</i>  |
| Kg                   | Kilogram  |
| MDR                  | Multidrug resistant   |
| Mg                   | Milligram   |

|                            |  |
|----------------------------|--|
| MHA                        | Mueller hinton agar  |
| MINDEF                     | Ministry of Defence  |
| MOH                        | Ministry of Health   |
| MOHE                       | Ministry of Higher Education                                 |
| MRSA                       | Methicillin-resistant <i>Staphylococcus aureus</i>           |
| MRSP                       | Methicillin-resistant <i>Staphylococcus pseudintermedius</i> |
| MVDL                       | Minnesota Veterinary Diagnostic Laboratory                   |
| My-AP-AMR                  | Malaysia action plan for antimicrobial resistance            |
| NA                         | Not included   |
| NPCB                       | National pharmaceutical control bureau                       |
| OIE                        | World Organization for Animal Health                         |
| PFGE                       | Pulsed field gel electrophoresis                             |
| <i>P. mirabilis</i>        | <i>Proteus mirabilis</i>                                     |
| R                          | Resistant  |
| S                          | Susceptible  |
| <i>S. aureus</i>           | <i>Staphylococcus aureus</i>                                 |
| <i>S. canis</i>            | <i>Streptococcus canis</i>                                   |
| <i>S. intermedius</i>      | <i>Staphylococcus intermedius</i>                            |
| <i>S. pseudintermedius</i> | <i>Staphylococcus pseudintermedius</i>                       |
| SPSS                       | Statistical package for social sciences                      |

|                 |                                  |
|-----------------|----------------------------------|
| UNISZA          | Universiti Sultan Zainal Abidin  |
| UPM             | Universiti Putra Malaysia        |
| VRE             | Vancomycin-resistant Enterococci |
| WHO             | World Health Organization        |
| $\beta$ -lactam | Beta-lactam                      |



# CHAPTER 1

## INTRODUCTION

### 1.1 Background

Antimicrobial resistance (AMR) is a global health issue of immense importance for both animals and human health (Robinson *et al.*, 2016). Activities related to sustaining human health, animal production and health, and the environment are major drivers for the emergence of AMR (Berendonk *et al.*, 2015; Hernando-Amado *et al.*, 2019; Wegener, 2012). The impact of AMR on human health has been widely reported (Fey *et al.*, 2000; Friedman *et al.*, 2016; Marshall & Levy, 2011; Mund *et al.*, 2017). In the animal production and health component, vast discussions have been presented on activities related to livestock production and husbandry that drives AMR (Landers *et al.*, 2012). However, there has been less emphasis on the impact of AMR on animal health in general especially among companion animals and wildlife.

In (2003), Wegener stated that the use of antimicrobial agents in domestic animals for treating, preventing diseases and growth-promoting contributes to the selection of antimicrobial resistance. It has also been widely reported that antimicrobial resistance in food-producing animals poses risks to humans because of the transmission of resistant zoonotic bacteria via the food chain (Economou & Gousia, 2015; Rozman *et al.*, 2019; Vidovic & Vidovic, 2020). However, until very recently, little attention has been directed to the use of antibiotics in companion animal clinics and the impact of such usage on the resistance level of veterinary pathogens. In a few recent studies, companion animals are reported to be able to disseminate antimicrobial resistance due to their close contact to humans (Amadi *et al.*, 2019; Chung *et al.*, 2017; Wedley *et al.*, 2011).

Little information is available about the level of resistance among clinical isolates from diseased animals in Malaysia. In contrast, there is a growing evidence of the widespread occurrence public health significant pathogens such as vancomycin-resistant enterococci (VRE), methicillin-resistant *Staphylococcus aureus* (MRSA) and multi-drug resistant *Salmonella typhimurium* in pet animals (Guardabassi *et al.*, 2017; Iseppi *et al.*, 2015; Van Den Bunt *et al.*, 2018). There are also studies that have linked bacterial pathogen isolated in livestock and pets to those found in their owners (Subbiah *et al.*, 2020; Yamasaki *et al.*, 2012). Therefore, the gaps of information from veterinary clinical isolates will be explored through the objectives of the current study.

The AMR patterns and trends among clinical isolates from domestic and wildlife have not been well reported. This information is necessary to enhance the understanding about the AMR situation among important disease-causing agents in local animals. In Malaysia, a report from a regional government laboratory has revealed the occurrence of multi-drug resistance *E. coli* from veterinary clinical isolates (Shahaza, *et al.*, 2017). Moreover, global reports on laboratory surveillance of AMR patterns and trends from diagnostic cases have shown high levels of resistance among common disease-causing bacteria in animals (Bourély *et al.*, 2019; Rzewuska *et al.*, 2015; Saputra *et al.*, 2017; Yassin *et al.*, 2017). Therefore, laboratory surveillance data have been highly effective in providing comprehensive information on the local AMR trends and patterns (IACG, 2018; Opintan *et al.*, 2015).

## 1.2 Problem statement

Dramatic increase of AMR among bacterial pathogens of zoonotic potential has been reported in many countries (Garcia-Migura *et al.*, 2014; Kaesbohrer *et al.*, 2012). Some antimicrobial agents used in animals have almost completely lost their efficacy to treat common bacterial infections. The trend of AMR is dependent on the way that antibiotic is used in the population. In Malaysia, antibiotic use in animals (pets or livestock) has not been documented and AMR among isolates from diseased animals is unknown. There is a large gap in knowledge about the AMR among local pets and livestock. Previous study has reported high level of resistance among clinical isolates from domestic and wild animals (Shahaza, *et al.*, 2017). However, more analysis from available data is needed to obtain more information to describe the AMR situation on the country.

### **1.3 Justification of the study**

Previous studies have focused on the trends of antimicrobial-resistant pathogens in clinical isolates from humans. There is limited information on the AMR trends of common bacterial species isolated from clinical veterinary isolates especially from diseased pets and livestock. Analysis of clinical data is necessary to determine the situation of AMR in veterinary clinical application. This could serve to guide future treatments of sick animals. The present study will describe the pattern and trend among clinical isolates from animals' samples submitted at the Bacteriology Laboratory of UPM Serdang. The findings from this study will emphasize the significance of antimicrobial resistance emergence among pathogens in veterinary cases and improves the understanding of the AMR among pathogens of significance in local animals.

### **1.4 Research questions**

This study aims to address these questions:

- i. What is the trend and pattern of AMR among clinically important bacterial pathogens from diseased pets (2015-2017)?
- ii. What are the differences between resistance profile of bacterial isolates from diseased cats and dogs?
- iii. What is the trend and pattern of clinically important bacterial pathogens from diseased livestock (2015-2017)?
- iv. What are the differences between resistance profile of bacterial isolates from diseased ruminants and non-ruminants?
- v. What is the pattern of antimicrobial resistance of selected sentinel pathogens (*E. coli*, *Salmonella* spp. and *K. pneumoniae*) isolated from diseased domestic and wildlife between 2018 and 2019?

## **1.5 Research hypothesis**

- i. The resistance profile of AMR is significantly different between diseased cats and dogs.
- ii. The resistance profile of AMR is significantly different between diseased ruminants and non-ruminants.
- iii. There is an increasing and decreasing trend of antibiotic resistance in diseased pets and livestock from 2015 to 2017.
- iv. The resistance profile of AMR in selected pathogens is significantly different between diseased companion, livestock and wildlife between 2018 and 2019.

## **1.6 General objective**

The present study was carried out with the aim of describing the patterns of resistance among bacteria isolated from clinical cases presented at a diagnostic laboratory in Universiti Putra Malaysia.

## **1.7 Specific objectives**

- i. To determine the antimicrobial resistance patterns of clinically important bacterial pathogens in clinical isolates from diseased pets (2015-2017)
- ii. To identify the antimicrobial resistance patterns of clinically important bacterial pathogens in clinical isolates from diseased livestock (2015-2017)
- iii. To study the antimicrobial resistance patterns of selected pathogens in veterinary clinical isolates from diseased domestic and wildlife (2018-2019).

## REFERENCES

- Aarestrup, F. M., Bager, F., Jensen, N. E., Madsen, M., Meyling, A., & Wegener, H. C. (1998). Resistance to antimicrobial agents used for animal therapy in pathogenic-, zoonotic- and indicator bacteria isolated from different food animals in Denmark: A baseline study for the Danish Integrated Antimicrobial Resistance Monitoring Programme (DANMAP). *APMIS*, 106(7-12), 745-770. doi: 10.1111/j.1699-0463.1998.tb00222.x
- Aarestrup, Frank M. (2015). The livestock reservoir for antimicrobial resistance: A personal view on changing patterns of risks, effects of interventions and the way forward. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 370(1670), 20140085. doi: 10.1098/rstb.2014.0085
- Aarestrup, Frank M., Wegener, H. C., & Collignon, P. (2008). Resistance in bacteria of the food chain: Epidemiology and control strategies. *Expert Review of Anti-Infective Therapy*, 6(5), 733-750. doi: 10.1586/14787210.6.5.733
- Aasmäe, B., Häkkinen, L., Kaart, T., & Kalmus, P. (2019). Antimicrobial resistance of *Escherichia coli* and *Enterococcus* spp. isolated from Estonian cattle and swine from 2010 to 2015. *Acta Veterinaria Scandinavica*, 61(1), 1-8. doi: 10.1186/s13028-019-0441-9
- Abatcha, M. G., Zakaria, Z., Gurmeet, K. D., & Thong, K. T. (2015). Antibiograms, resistance genes, class I integrons and PFGE profiles of zoonotic *Salmonella* in Malaysia. *Tropical Biomedicine*, 32(4), 573-586. [http://eprints.um.edu.my/18469/1/Antibiograms%2C\\_Resistance\\_Genes%2C\\_Class\\_I\\_Integrons\\_and.pdf](http://eprints.um.edu.my/18469/1/Antibiograms%2C_Resistance_Genes%2C_Class_I_Integrons_and.pdf)
- Abbassi, M. (2017). Antimicrobial Resistance in *Escherichia coli* Isolates from healthy poultry, bovine and ovine in Tunisia: A real animal and human health threat. *Journal of Clinical Microbiology and Biochemical Technology*, 3(2), 019-123, doi: 10.17352/jcmbt.000021
- Abo-Shama. (2014). Prevalence and antimicrobial susceptibility of *Staphylococcus aureus* isolated from cattle, buffalo, sheep and goat's raws milk in Sohag Governorate, Egypt. *Journal of Veterinary Medicine*, 60(141), 63-72. [http://www.aun.edu.eg/journal\\_files/160\\_J\\_5513.pdf](http://www.aun.edu.eg/journal_files/160_J_5513.pdf)
- Abraham, S., Jordan, D., Wong, H. S., Johnson, J. R., Toleman, M. A., Wakeham, D. L., Gordon, D. M., Turnidge, J. D., Mollinger, J. L., Gibson, J. S., & Trott, D. J. (2015). First detection of extended-spectrum cephalosporin- and fluoroquinolone-resistant *Escherichia coli* in Australian food-producing animals. *Journal of Global Antimicrobial Resistance*, 3(4), 273-277. doi: 10.1016/j.jgar.2015.08.002

- Adelowo, O. O., Fagade, O. E., & Agersø, Y. (2014). Antibiotic resistance and resistance genes in *Escherichia coli* from poultry farms, southwest Nigeria. *Journal of Infection in Developing Countries*, 8(09), 1103-1112. doi: 10.3855/jidc.4222
- Adzitey, F., Ali, G. R. R., Huda, N., & Ting, S. L. (2013). Antibiotic resistance and plasmid profile of *Escherichia coli* isolated from ducks in Penang, Malaysia. *International Food Research Journal*, 20(3), 1473-1478. doi: [http://www.ifrj.upm.edu.my/20%20\(03\)%202013/65%20IFRJ%2020%20\(03\)%202013%20Adzitey%20\(427\).pdf](http://www.ifrj.upm.edu.my/20%20(03)%202013/65%20IFRJ%2020%20(03)%202013%20Adzitey%20(427).pdf)
- Agarwal, A., Kapila, K., & Kumar, S. (2009). WHONET software for the surveillance of antimicrobial susceptibility. *Medical Journal Armed Forces India*, 65(3), 264-266. doi: 10.1016/S0377-1237(09)80020-8
- Ahlstrom, C. A., Bonnedahl, J., Woksepp, H., Hernandez, J., Olsen, B., & Ramey, A. M. (2018). Acquisition and dissemination of cephalosporin-resistant *E. coli* in migratory birds sampled at an Alaska landfill as inferred through genomic analysis. *Scientific Reports*, 8(1), 1-11. doi: 10.1038/s41598-018-25474-w
- Ahmed, A. M., Motoi, Y., Sato, M., Maruyama, A., Watanabe, H., Fukumoto, Y., & Shimamoto, T. (2007). Zoo animals as reservoirs of gram-negative bacteria harboring integrons and antimicrobial resistance genes. *Applied and Environmental Microbiology*, 73(20), 6686-6690. doi: 10.1128/AEM.01054-07
- Allen, H. K., Donato, J., Wang, H. H., Cloud-Hansen, K. A., Davies, J., & Handelsman, J. (2010). Call of the wild: Antibiotic resistance genes in natural environments. *Nature Reviews Microbiology*, 8(4), 251-259. doi: 10.1038/nrmicro2312
- Allen, H. K., & Stanton, T. B. (2014). Altered egos: Antibiotic effects on food animal microbiomes. *Annual Review of Microbiology*. 68, 297-315. doi: 10.1146/annurev-micro-091213-113052
- Allen, S. E., Boerlin, P., Janecko, N., Lumsden, J. S., Barker, I. K., Pear, D. L., Reid-Smith, R. J., & Jardine, C. (2011). Antimicrobial resistance in generic *Escherichia coli* isolates from wild small mammals living in swine farm, residential, landfill, and natural environments in Southern Ontario, Canada. *Applied and Environmental Microbiology*, 77(3), 882-888. doi:10.1128/AEM.01111-10
- Allos, B. M., Moore, M. R., Griffin, P. M., & Tauxe, R. V. (2004). Surveillance for sporadic foodborne disease in the 21st century: The FoodNet perspective. *Clinical Infectious Diseases*, 38(3), 115-120. doi: 10.1086/381577

- Alvarez-Uria, G., Gandra, S., Mandal, S., & Laxminarayan, R. (2018). Global forecast of antimicrobial resistance in invasive isolates of *Escherichia coli* and *Klebsiella pneumoniae*. *International Journal of Infectious Diseases*, 68, 50-53. doi: 10.1016/j.ijid.2018.01.011
- Amadi, V. A., Hariharan, H., Amadi, O. A., Matthew-Belmar, V., Nicholas-Thomas, R., Perea, M. L., Carter, K., Rennie, E., Kalasi, K., Alhassan, A., Kabuusu, R. M., Alozie, G. U., Fields, P. J., Pinckney, R., & Sharma, R. (2019). Antimicrobial resistance patterns of commensal *Escherichia coli* isolated from feces of non-diarrheic dogs in Grenada, West Indies. *Veterinary World*, 12 (12): 2070-2075. doi: 10.14202/vetworld.2019.2070-2075
- Apun, K., Chong, Y. L., Abdullah, M. T., & Micky, V. (2008). Antimicrobial susceptibilities of *Escherichia coli* isolates from food animals and wildlife animals in Sarawak, East Malaysia. *Asian Journal of Animal and Veterinary Advances*, 3(6), 409-416.  
doi: 10.3923/ajava.2008.409.416
- Auwaerter, P. (2019). *Proteus species*. Johns Hopkins ABX Guide. The Johns Hopkins University:  
[https://www.hopkinsguides.com/hopkins/view/Johns\\_Hopkins\\_ABX\\_Guide/540454/all/Proteus\\_species](https://www.hopkinsguides.com/hopkins/view/Johns_Hopkins_ABX_Guide/540454/all/Proteus_species)
- Awosile, B. B., Heider, L. C., Saab, M. E., & McClure, J. T. (2018). Antimicrobial resistance in mastitis, respiratory and enteric bacteria isolated from ruminant animals from the Atlantic Provinces of Canada from 1994-2013. *Canadian Veterinary Journal*, 59(10), 1099-1104.  
<https://pubmed.ncbi.nlm.nih.gov/30510316/>
- Awosile, B. B., McClure, J. T., Saab, M. E., & Heider, L. C. (2018). Antimicrobial resistance in bacteria isolated from cats and dogs from the Atlantic Provinces, Canada from 1994-2013. *Canadian Veterinary Journal*, 59(8), 885.  
[https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6049328/pdf/cvj\\_08\\_885.pdf](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6049328/pdf/cvj_08_885.pdf)
- Ayukekpong, J. A., Ntemgwa, M., & Atabe, A. N. (2017). The threat of antimicrobial resistance in developing countries: Causes and control strategies. *Antimicrobial Resistance and Infection Control*, 6(1), 1-8. doi: 10.1186/s13756-017-0208-x
- Babic, M., Hujer, A. M., & Bonomo, R. A. (2006). What's new in antibiotic resistance? Focus on beta-lactamases. *Drug Resistance Updates*, 9(3), 142-156. doi: 10.1016/j.drup.2006.05.005
- Balcazar, J. L. (2014). Bacteriophages as vehicles for antibiotic resistance genes in the environment. *PLoS Pathogens*, 10(7), e1004219. doi: 10.1371/journal.ppat.1004219

- Ball, K. R., Rubin, J. E., Chirino-Trejo, M., & Dowling, P. M. (2008). Antimicrobial resistance and prevalence of canine uropathogens at the Western College of Veterinary Medicine Veterinary Teaching Hospital, 2002-2007. *Canadian Veterinary Journal*, 49(10), 985. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2553511/pdf/cvj-10-985.pdf>
- Bantawa, K., Sah, S. N., Limbu, D. S., Subba, P., & Ghimire, A. (2019). Antibiotic resistance patterns of *Staphylococcus aureus*, *Escherichia coli*, *Salmonella*, *Shigella* and *Vibrio* isolated from chicken, pork, buffalo and goat meat in eastern Nepal. *BMC Research Notes*, 12(1), 1-6. doi: 10.1186/s13104-019-4798-7
- Barlow, R. S., McMillan, K. E., Duffy, L. L., Fegan, N., Jordan, D., & Mellor, G. E. (2017). Antimicrobial resistance status of *Enterococcus* from Australian cattle populations at slaughter. *PLoS ONE*, 12(5), e0177728. doi: 10.1371/journal.pone.0177728
- Beco, L., Guaguère, E., Méndez, C. L., Noli, C., Nuttall, T., & Vroom, M. (2013). Suggested guidelines for using systemic antimicrobials in bacterial skin infections (2): Antimicrobial choice, treatment regimens and compliance. *Veterinary Record*, 172(6), 156-160. doi: 10.1136/vr.101070
- Beever, L., Bond, R., Graham, P. A., Jackson, B., Lloyd, D. H., & Loeffler, A. (2015). Increasing antimicrobial resistance in clinical isolates of *Staphylococcus intermedius* group bacteria and emergence of MRSP in the UK. *Veterinary Record*, 176(7), 172. doi: 10.1136/vr.102651
- Bengtsson, B., Franklin, A., & Greko, C. (2006). Swedish Veterinary Antimicrobial Resistance Monitoring – surveillance of resistance in bacteria from animals. *National Veterinary Institute, Uppsala, Sweden*. [https://www.sva.se/media/jpyahig3/rapport\\_svarm\\_2006.pdf](https://www.sva.se/media/jpyahig3/rapport_svarm_2006.pdf)
- Bennani, H., Mateus, A., Mays, N., Eastmure, E., Stärk, K. D. C., & Häslér, B. (2020). Overview of evidence of antimicrobial use and antimicrobial resistance in the food chain. *Antibiotics*, 9(2), 49. doi: 10.3390/antibiotics9020049
- Berendonk, T. U., Manaia, C. M., Merlin, C., Fatta-Kassinos, D., Cytryn, E., Walsh, F., Bürgmann, H., Sørum, H., Norström, M., Pons, M. N., Kreuzinger, N., Huovinen, P., Stefani, S., Schwartz, T., Kisand, V., Baquero, F., & Martinez, J. L. (2015). Tackling antibiotic resistance: The environmental framework. *Nature Reviews Microbiology*, 13(5), 310-317. doi: 10.1038/nrmicro3439
- Bidewell, C. A., Williamson, S. M., Rogers, J., Tang, Y., Ellis, R. J., Petrovska, L., & AbuOun, M. (2018). Emergence of *Klebsiella pneumoniae* subspecies *pneumoniae* as a cause of septicaemia in pigs in England. *PLoS ONE*, 13(2), e0191958. doi: 10.1371/journal.pone.0191958

- Boguta, L., Gradzki, Z., Borges, E., Maurin, F., Kodjo, A., & Winiarczyk, S. (2002). Bacterial flora in foals with upper respiratory tract infections in Poland. *Journal of Veterinary Medicine, Series B*, 49(6), 294-297. doi: 10.1046/j.1439-0450.2002.00570.x
- Boireau, C., Cazeau, G., Jarrige, N., Calavas, D., Madec, J. Y., Leblond, A., Haenni, M., & Gay, É. (2018). Antimicrobial resistance in bacteria isolated from mastitis in dairy cattle in France, 2006–2016. *Journal of Dairy Science*, 101(10), 9451-9462. doi: 10.3168/jds.2018-14835
- Boonmasawai, S., Bangphoomi, N., Sungpradit, S., Pati, N., Tangkoskul, T., Thamthaweechok, N., & Thamlikitkul, V. (2018). Screening of antimicrobial resistant bacteria in dog shelters in Thailand. *Journal of Applied Animal Science*, 11(3), 25-36.  
<https://vs.mahidol.ac.th/jaas/Files/Vol11No3/RS%20K.Sookruetai%20Final.pdf>
- Bourély, C., Cazeau, G., Jarrige, N., Leblond, A., Madec, J. Y., Haenni, M., & Gay, E. (2019). Antimicrobial resistance patterns of bacteria isolated from dogs with otitis. *Epidemiology and Infection*, 147(e121), 1-10. doi: 10.1017/S0950268818003278
- Bourne, J. A., Chong, W. L., & Gordon, D. M. (2019). Genetic structure, antimicrobial resistance and frequency of human associated *Escherichia coli* sequence types among faecal isolates from healthy dogs and cats living in Canberra, Australia. *PLoS ONE*, 14(3), e0212867. doi: 10.1371/journal.pone.0212867
- Boxall, A., & Wilkinson, J. (2019). Identifying hotspots of resistance selection from antibiotic exposure in urban environments around the world. *SETAC Europe 29th Annual Meeting, Helsinki, Finland*.
- Brisse, S., & Van Duijkeren, E. (2005). Identification and antimicrobial susceptibility of 100 *Klebsiella* animal clinical isolates. *Veterinary Microbiology*, 105(3-4), 307-312. doi: 10.1016/j.vetmic.2004.11.010
- Busani, L., Cigliano, A., Taioli, E., Caligiuri, V., Chiavacci, L., Di Bella, C., Battisti, A., Duranti, A., Gianfranceschi, M., Nardella, M. C., Ricci, A., Rolesu, S., Tamba, M., Marabelli, R., & Caprioli, A. (2005). Prevalence of *Salmonella enterica* and *Listeria monocytogenes* contamination in foods of animal origin in Italy. *Journal of Food Protection*. 68(8), 1729-1733. doi: 10.4315/0362-028X-68.8.1729
- Carrique-Mas, J. J., Choisy, M., Van Cuong, N., Thwaites, G., & Baker, S. (2020). An estimation of total antimicrobial usage in humans and animals in Vietnam. *Antimicrobial Resistance & Infection Control*, 9(1), 1-6. doi: 10.1186/s13756-019-0671-7
- Carroll, D., Wang, J., Fanning, S., & McMahon, B. J. (2015). Antimicrobial resistance in wildlife: Implications for public health. *Zoonoses and Public Health*, 62(7), 534–542. doi: 10.1111/zph.12182

- Carvalho, A. C., Barbosa, A. V., Arais, L. R., Ribeiro, P. F., Carneiro, V. C., & Cerqueira, A. M. F. (2016). Resistance patterns, ESBL genes, and genetic relatedness of *Escherichia coli* from dogs and owners. *Brazilian Journal of Microbiology*, 47(1), 150-158. doi: 10.1016/j.bjm.2015.11.005
- Chandrasekaran, D., Venkatesan, P., Tirumurugaan, K. G., Nambi, A. P., Thirunavukkarasu, P. S., Kumaran, K., Vairamuthu, S., & Ramesh, S. (2014). Pattern of antibiotic resistant mastitis in dairy cows. *Veterinary World*, 7(6), 389-394. doi: 10.14202/vetworld.2014.389-394
- Chantziaras, I., Boyen, F., Callens, B., & Dewulf, J. (2014). Correlation between veterinary antimicrobial use and antimicrobial resistance in food-producing animals: A report on seven countries. *Journal of Antimicrobial Chemotherapy*, 69(3), 827-834. doi: 1093/jac/dkt443
- Chattopadhyay, M. K. (2014). Use of antibiotics as feed additives: A burning question. *Frontiers in Microbiology*, 5,(334), 1-3. doi: 10.3389/fmicb.2014.00334
- Chen, X., Zhang, W., Yin, J., Zhang, N., Geng, S., Zhou, X., Wang, Y., Gao, S., & Jiao, X. (2014). *Escherichia coli* isolates from sick chickens in China: Changes in antimicrobial resistance between 1993 and 2013. *Veterinary Journal*, 202(1), 112-115. doi: 10.1016/j.tvjl.2014.06.016
- Cheng, F., Li, Z., Lan, S., Liu, W., Li, X., Zhou, Z., Song, Z., Wu, J., Zhang, M., & Shan, W. (2018). Characterization of *Klebsiella pneumoniae* associated with cattle infections in southwest China using multi-locus sequence typing (MLST), antibiotic resistance and virulence-associated gene profile analysis. *Brazilian Journal of Microbiology*, 49(1), 93-100. doi: 10.1016/j.bjm.2018.06.004
- Cheong, Y. M., Lim, V. K. E., Jegathesan, M., & Suleiman, A. B. (1994). Antimicrobial resistance in 6 Malaysian general hospitals. *Medical Journal of Malaysia*, 49(4), 317–326.  
<https://europepmc.org/article/med/7674966>
- Chomel, B. B. (2014). Emerging and re-emerging zoonoses of dogs and cats. In *Animals*, 4(3), 434-445. doi: 10.3390/ani4030434
- Chung, Y. S., Park, Y. K., Park, Y. H., & Park, K. T. (2017). Probable secondary transmission of antimicrobial-resistant *Escherichia coli* between people living with and without pets. *Journal of Veterinary Medical Science*, 79(3), 486-491. doi: 10.1292/jvms.16-0585
- Clinical Laboratory Standards Institute (2015). Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals: approved standard-fourth edition. CLSI document VET01-A4. clinical laboratory standards institute, Wayne, PA (and performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals: second informational supplement. CLSI document VET01-S2. Clinical laboratory standards institute, Wayne, PS)

- CLSI. (2018). 28th Edition M100 Performance Standards for Antimicrobial Susceptibility Testing. *Journal of Clinical Microbiology*, 28, 1-296. doi: 10.1128/JCM.00655-17
- Cobbold, R. N., Rice, D. H., Davis, M. A., Besser, T. E., & Hancock, D. D. (2006). Long-term persistence of multi-drug-resistant *Salmonella enterica* serovar *Newport* in two dairy herds. *Journal of the American Veterinary Medical Association*, 228(4), 585-591. doi: 10.2460/javma.228.4.585
- Collignon, P. C., Conly, J. M., Andremont, A., McEwen, S. A., Aidara-Kane, A., Griffin, P. M., Agerso, Y., Dang Ninh, T., Donado-Godoy, P., Fedorka-Cray, P., Fernandez, H., Galas, M., Irwin, R., Karp, B., Matar, G., McDermott, P., Mitema, E., Reid-Smith, R., Scott, H. M., Woo, G. J. (2016). World Health Organization ranking of antimicrobials according to their importance in human medicine: A critical step for developing risk management strategies to control antimicrobial resistance from food animal production. *Clinical Infectious Diseases*, 63(8), 1087-1093. doi: 10.1093/cid/ciw475
- Costa, D., Poeta, P., Sáenz, Y., Vinué, L., Coelho, A. C., Matos, M., Rojo-Bezares, B., Rodrigues, J., & Torres, C. (2008). Mechanisms of antibiotic resistance in *Escherichia coli* isolates recovered from wild animals. *Microbial Drug Resistance*, 14(1), 71-77. doi: 10.1089/mdr.2008.0795
- Courtice, R., Sniatynski, M., & Rubin, J. E. (2016). Antimicrobial resistance and beta-lactamase production of *Escherichia coli* causing canine urinary tract infections: Passive surveillance of laboratory isolates in Saskatoon, Canada, 2014. *Canadian Veterinary Journal*, 57(11), 1166. doi: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5081147/>
- Cromwell, G. L. (2002). Why and how antibiotics are used in swine production. *Animal Biotechnology*, 13(1), 7-27. doi: 10.1081/ABIO-120005767
- Cummings, K. J., Aprea, V. A., & Altier, C. (2015). Antimicrobial resistance trends among canine *Escherichia coli* isolates obtained from clinical samples in the northeastern USA, 2004-2011. *Canadian Veterinary Journal*, 56(4), 393. doi: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4357913/>
- Dalhoff, A. (2012). Global fluoroquinolone resistance epidemiology and implications for clinical use. *Interdisciplinary Perspectives on Infectious Diseases*, 2012, 1-37. doi: 10.1155/2012/976273
- DANMAP. (2018). DANMAP 2018. *Statens Serum Institut DTU National Food Institute*, 2018. Retrieved from <file:///C:/Users/pc/Downloads/Rapport-DANMAP-2018.pdf>

- Davies, M., & Stewart, P. R. (2008). Transferable drug resistance in man and animals: genetic relationships between R-plasmids and enteric bacteria from man and domestic pets. *Australian Veterinary Journal*, 54(11), 507–512. doi: 10.1111/j.1751-0813.1978.tb00316.x
- De Briyne, N., Atkinson, J., Borriello, S. P., & Pokludová, L. (2014). Antibiotics used most commonly to treat animals in Europe. *Veterinary Record*, 175(13), 325-325. doi: 10.1136/vr.102462
- De Graef, E. M., Decostere, A., Devriese, L. A., & Haesebrouck, F. (2004). Antibiotic resistance among fecal indicator bacteria from healthy individually owned and kennel dogs. *Microbial Drug Resistance*, 10(1), 65-69. doi: 10.1089/107662904323047826
- Decline, V., Effendi, M. H., Rahmani, R. P., Yanestria, S. M., & Harijani, N. (2020). Profile of antibiotic-resistant and presence of methicillin-resistant *Staphylococcus aureus* from nasal swab of dogs from several animal clinics in Surabaya, Indonesia. *International Journal of One Health*, 6(1), 90-94. doi: 10.14202/IJOH.2020.90-94
- Department of Veterinary Services Malaysia. (2020). National Veterinary Antimicrobials Guidelines. *First Edition*, 74.
- Dore, S., Liciardi, M., Amatiste, S., Bergagna, S., Bolzoni, G., Caligiuri, V., Cerrone, A., Farina, G., Montagna, C. O., Saletti, M. A., Scatassa, M. L., Sotgiu, G., & Cannas, E. A. (2016). Survey on small ruminant bacterial mastitis in Italy, 2013–2014. *Small Ruminant Research*, 141, 91-93. doi: 10.1016/j.smallrumres.2016.07.010
- Dorsch, R., Von Vopelius-Feldt, C., Hartmann, K., Wolf, G., & Straubinger, R. K. (2015). Feline urinary tract pathogens: Prevalence of bacterial species and antimicrobial resistance over a 10-year period. *Veterinary Record*, 176(8), 201-201. doi: 10.1136/vr.102630
- Dosen, R., Prodanov-Radulovic, J., Pusic, I., Stojanov, I., Stojanovic, D., & Ratajac, R. (2011). Resistance *Escherichia coli* isolates to antibiotics from the organ samples originating from swine farms. *Biotechnology in Animal Husbandry*, 27(3), 861-866. doi: 10.2298/bah1103861d
- Doyle, M. E. (2015). Multidrug-resistant pathogens in the food supply. *Foodborne Pathogens and Disease*, 12(4), 261-279. doi: 10.1089/fpd.2014.1865
- Dupouy, V., Abdelli, M., Moyano, G., Arpaillange, N., Bibbal, D., Cadiergues, M. C., Lopez-Pulin, D., Sayah-Jeanne, S., De Gunzburg, J., Saint-Lu, N., Gonzalez-Zorn, B., Andremont, A., & Bousquet-Mélou, A. (2019). Prevalence of beta-lactam and quinolone/fluoroquinolone resistance in Enterobacteriaceae from dogs in France and spain—characterization of ESBL/pAmpC isolates, genes, and conjugative plasmids. *Frontiers in Veterinary Science*, 6(279), 1-10. doi: 10.3389/fvets.2019.00279

- Economou, V., & Gousia, P. (2015). Agriculture and food animals as a source of antimicrobial-resistant bacteria. *Infection and Drug Resistance*, 8, 49-61. doi: 10.2147/IDR.S55778
- Equale, T. (2018). Non-typhoidal *Salmonella* serovars in poultry farms in central Ethiopia: Prevalence and antimicrobial resistance. *BMC Veterinary Research*, 14(1), 1-8. doi: 10.1186/s12917-018-1539-4
- Eltaib, N. O., Abdfarag, E. A., Al-Romaihi, H., Wehedy, E., Mahmoud, M. H., Alawad, O. K., Al-Hajri, M. M., Thani, A. A. A. L., & Yassine, H. M. (2018). Antibiotic resistance profile of commensal *Escherichia coli* isolated from broiler chickens in Qatar. *Journal of Food Protection*, 81(2), 302-307. doi: 10.4315/0362-028X.JFP-17-191
- EMEA. (1998). Committee for veterinary medicinal products. Enrofloxacin (Summary Report 1). *The European Agency for the Evaluation of Medicinal Products*. Retrieved from [https://www.ema.europa.eu/en/documents/mrl-report/enrofloxacin-summary-report-1-committee-veterinary-medicinal-products\\_en.pdf](https://www.ema.europa.eu/en/documents/mrl-report/enrofloxacin-summary-report-1-committee-veterinary-medicinal-products_en.pdf)
- Eng, S. K., Pusparajah, P., Ab Mutalib, N. S., Ser, H. L., Chan, K. G., & Lee, L. H. (2015). *Salmonella*: A review on pathogenesis, epidemiology and antibiotic resistance. *Frontiers in Life Science*, 8(3), 284-293. doi: 10.1080/21553769.2015.1051243
- Enne, V. I., Cassar, C., Sprigings, K., Woodward, M. J., & Bennett, P. M. (2008). A high prevalence of antimicrobial resistant *Escherichia coli* isolated from pigs and a low prevalence of antimicrobial resistant *E. coli* from cattle and sheep in Great Britain at slaughter. *FEMS Microbiology Letters*, 278(2), 193-199. doi: 10.1111/j.1574-6968.2007.00991.x
- Erskine, R. J., Walker, R. D., Bolin, C. A., Bartlett, P. C., & White, D. G. (2002). Trends in antibacterial susceptibility of mastitis pathogens during a seven-year period. *Journal of Dairy Science*, 85(5), 1111-1118. doi: 10.3168/jds.S0022-0302(02)74172-6
- European Centre for Disease Prevention and Control. (2018). Surveillance of antimicrobial resistance in Europe Annual report of the European Antimicrobial Resistance Surveillance Network (EARS-Net) 2017. *ECDC: Surveillance Report*. 1-108. doi: 10.2900/230516
- European Medicines Agency. (2017). ECDC/EFSA/EMA second joint report on the integrated analysis of the consumption of antimicrobial agents and occurrence of antimicrobial resistance in bacteria from humans and food-producing animals. *EFSA Journal*, 15(7), e04872. doi: 10.2903/j.efsa.2017.4872
- European Medicines Agency. (2018). Sales of veterinary antimicrobial agents in 30 European countries in 2016- Trends from 2010 to 2016- Eight ESVAC report. *European Medicines Agency*, 1-178. doi: 10.1016/j.phpro.2011.05.019

FAAIR Scientific Advisory Panel. (2002). The need to improve antimicrobial use in agriculture ecological and human. *Clinical Infectious Diseases*, 34, 1-76. Retrieved from <https://pascal-francis.inist.fr/vibad/index.php?action=getRecordDetail&idt=13346395>

Faires, M. C., Tater, K. C., & Weese, J. S. (2009). An investigation of methicillin-resistant *Staphylococcus aureus* colonization in people and pets in the same household with an infected person or infected pet. *Journal of the American Veterinary Medical Association*, 235(5), 540-543. doi: 10.2460/javma.235.5.540

FDA. (2012). Guidance for Industry- The judicious use of medically important antimicrobial drugs in food-producing animals. *Federal Register*, 1-26. Retrieved from <https://www.fda.gov/media/79140/download>

Ferreira, R. L., Da Silva, B. C. M., Rezende, G. S., Nakamura-Silva, R., Pitondo-Silva, A., Campanini, E. B., Brito, M. C. A., Da Silva, E. M. L., De Melo Freire, C. C., Da Cunha, A. F., & Da Silva Pranchevicius, M. C. (2019). High prevalence of multidrug-resistant *Klebsiella pneumoniae* harboring several virulence and β-lactamase encoding genes in a Brazilian intensive care unit. *Frontiers in Microbiology*, 9(3198), 1-15. doi: 10.3389/fmicb.2018.03198

Ferreira, T. S. P., Moreno, L. Z., Felizardo, M. R., de Gobbi, D. D. S., Filsner, P. H. D. L. N., de Moura Gomes, V. T., Moreno, M., & Moreno, A. M. (2016). Pheno- and genotypic characterization of *Pasteurella multocida* isolated from cats, dogs and rabbits from Brazil. *Comparative Immunology, Microbiology and Infectious Diseases*, 45, 48-52. doi: 10.1016/j.cimid.2016.02.004

Fey, P. D., Safranek, T. J., Rupp, M. E., Dunne, E. F., Ribot, E., Iwen, P. C., Bradford, P. A., Angulo, F. J., & Hinrichs, S. H. (2000). Ceftriaxone-resistant *Salmonella* infection acquired by a child from cattle. *New England Journal of Medicine*, 342(17), 1242-1249. doi: 10.1056/NEJM200004273421703

FAO/WHO/OIE. (2008). Report of the FAO/WHO/OIE Expert meeting FAO. *FAO/WHO/OIE Stakeholders Meeting on Critically Important Antimicrobials*.

Fleming, S. (2009). Ovine and caprine respiratory disease: Infectious agents, management factors, and preventive strategies. *Food Animal Practice*, 194-198. doi: 10.1016/b978-1-4160-3591-6.10044-2

- Flemming Bager, Bortolaia, V., Ellis-Iversen, J., Hendriksen, R. S., Høg, B. B., Jensen, L. B., Jensen, A. N., Knegt, L. de, Korsgaard, H., Dalby, T., Hammerum, A. M., Hasman, H., Kuhn, K. G., Hoffmann, S., Larsen, A. R., Laursen, M., Nielsen, E. M., Olsen, S. S., Petersen, A., Torpdahl, M. (2016). DANMAP 2015 - Use of antimicrobial agents and occurrence of antimicrobial resistance in bacteria from food animals, food and humans in Denmark. *DANMAP*, 1-142. Retrieved from <http://www.danmap.org/~media/Projekt%20sites/Danmap/DANMAP%20reports/DANMAP%202015/DANMAP%202015.ashx>
- Fleming, A. (1944). The discovery of penicillin. *British Medical Bulletin*, 2(1), 4-5. doi: 10.1093/oxfordjournals.bmb.a071032
- French, G. L. (2010). The continuing crisis in antibiotic resistance. *International Journal of Antimicrobial Agents*, 36, S3-S7. doi: 10.1016/S0924-8579(10)70003-0
- Friedman, N. D., Temkin, E., & Carmeli, Y. (2016). The negative impact of antibiotic resistance. *Clinical Microbiology and Infection*, 22(5), 416-422. doi: 10.1016/j.cmi.2015.12.002
- Friese, A., Schulz, J., Laube, H., von Salviati, C., Hartung, J., & Roesler, U. (2013). Faecal occurrence and emissions of livestock-associated methicillin-resistant *Staphylococcus aureus* (laMRSA) and ESbl/AmpC-producing *E. coli* from animal farms in Germany. *Berliner Und Münchener Tierärztliche Wochenschrift*, 126(3-4), 175-180. Retrieved from <https://europepmc.org/article/med/23540202>
- Gahamanyi, N., Mboera, L. E. G., Matee, M. I., Mutangana, D., & Komba, E. V. G. (2020). Prevalence, risk factors, and antimicrobial resistance profiles of thermophilic *Campylobacter* species in humans and animals in Sub-Saharan Africa: A systematic review. *International Journal of Microbiology*, 2020, 1-20. doi: 10.1155/2020/2092478
- Gally, D. L., & Stevens, M. P. (2017). Microbe profile: *Escherichia coli* O157:H7 – notorious relative of the microbiologist's workhorse. *Microbiology*, 163(1), 1-3. doi: 10.1099/mic.0.000387
- Gandra, S., Barter, D. M., & Laxminarayan, R. (2014). Economic burden of antibiotic resistance: How much do we really know? *Clinical Microbiology and Infection*, 20(10), 973-980. doi: 10.1111/1469-0691.12798
- Garcia-Migura, L., Hendriksen, R. S., Fraile, L., & Aarestrup, F. M. (2014). Antimicrobial resistance of zoonotic and commensal bacteria in Europe: The missing link between consumption and resistance in veterinary medicine. *Veterinary Microbiology*, 170(1-2), 1-9.. doi: 10.1016/j.vetmic.2014.01.013

- Gebru, E., Damte, D., Choi, M. J., Lee, S. J., Kim, Y. H., & Park, S. C. (2012). Mutant prevention concentration and phenotypic and molecular basis of fluoroquinolone resistance in clinical isolates and in vitro-selected mutants of *Escherichia coli* from dogs. *Veterinary Microbiology*, 154(3-4), 384-394. doi: 10.1016/j.vetmic.2011.07.033
- Ghasemzadeh, I., & Namazi, S. H. (2015). Review of bacterial and viral zoonotic infections transmitted by dogs. *Journal of Medicine and Life*, 8(4), 1-5. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5319273/>
- Ghosh, A. N., Bhatta, D. R., Ansari, M. T., Tiwari, H. K., Mathuria, J. P., Gaur, A., Supram, H. S., & Gokhale, S. (2013). Application of WHONET in the antimicrobial resistance surveillance of uropathogens: A first user experience from Nepal. *Journal of Clinical and Diagnostic Research*, 7(5), 845-848. doi: 10.7860/JCDR/2013/5193.2955
- Gocke, T. M., & Finland, M. (1951). Cross-resistance to antibiotics. Effect of repeated exposure of bacteria to aureomycin, terramycin, chloramphenicol or neomycin on the resistance to all of these antibiotics and to streptomycin and penicillin. *Journal of Laboratory and Clinical Medicine*, 38(5), 719-35. doi: 10.5555/uri:pii:002221435190008X
- Gómez-Poveda, B., & Moreno, M. A. (2018). Antimicrobial prescriptions for dogs in the Capital of Spain. *Frontiers in Veterinary Science*, 5(309), 1-9. doi: 10.3389/fvets.2018.00309
- Goni, M., Gurmeet, D., Thong, K. L., & Zakaria, Z. (2014). Occurrence of antibiotic resistant *Salmonella* isolated from dogs in Klang Valley, Malaysia. *Malaysia Journal of Microbiology*, 10, 219-224. doi: 10.21161/mjm.58213
- Gootz, T. D. (1990). Discovery and development of new antimicrobial agents. *Clinical Microbiology Reviews*, 3(1), 13–31. doi: 10.1128/CMR.3.1.13
- Gouvêa, R., Dos Santos, F. F., De Aquino, M., & Pereira, VL de A (2015). Fluoroquinolones in industrial poultry production, bacterial resistance and food residues: A review. *Revista Brasileira de Ciencia Avicola*, 17, 1-10. doi: 10.1590/1516-635x17011-10
- Graham, J. P., Boland, J. J., & Silbergeld, E. (2007). Growth promoting antibiotics in food animal production: An economic analysis. *Public Health Reports*, 122(1), 79-87. doi: 10.1177/003335490712200111
- Grant, A., Hashem, F., & Parveen, S. (2016). *Salmonella* and *Campylobacter*: Antimicrobial resistance and bacteriophage control in poultry. *Food Microbiology*, 53, 104-109. doi: 10.1016/j.fm.2015.09.008

- Grönthal, T., Eklund, M., Thomson, K., Piiparinen, H., Sironen, T., & Rantala, M. (2017). Antimicrobial resistance in *Staphylococcus pseudintermedius* and the molecular epidemiology of methicillin-resistant *S. pseudintermedius* in small animals in Finland. *Journal of Antimicrobial Chemotherapy*, 72(4), 1021-1030. doi: 10.1093/jac/dkw559
- Guardabassi, L. (2004). Pet animals as reservoirs of antimicrobial-resistant bacteria: Review. *Journal of Antimicrobial Chemotherapy*, 54(2), 321–332. doi: 10.1093/jac/dkh332
- Guardabassi, L., Jensen, L. B., & Kruse, H. (2009). Guide to antimicrobial use in animals. *Guide to antimicrobial use in animals*. doi: 10.1002 /9781444302639
- Guardabassi, L., Schwarz, S., & Lloyd, D. H. (2017). Pet animals as reservoirs of antimicrobial-resistant bacteria. 54(2), 321–332. doi: 10.1093/jac /dkh332
- Guenther, S., Grobbel, M., Heidemanns, K., Schlegel, M., Ulrich, R. G., Ewers, C., & Wieler, L. H. (2010). First insights into antimicrobial resistance among faecal *Escherichia coli* isolates from small wild mammals in rural areas. *Science of the Total Environment*, 408(17), 3519-3522. doi: 10.1016 /j.scitotenv.2010.05.005
- Guerin, M. T., Martin, S. W., Darlington, G. A., & Rajic, A. (2005). A temporal study of *Salmonella* serovars in animals in Alberta between 1990 and 2001. *Canadian Journal of Veterinary Research*, 69(2), 88. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1142175/>
- Gustafson, R.H., & Bowen, R. E. (1997). Antibiotic use in animal agriculture. *Journal of Applied Microbiology*, 83(5), 531–541. doi: 10.1046/j.1365-2672.1997.00280.x
- Gustafson, R. H. (1991). Use of antibiotics in livestock and human health concerns. *Journal of Dairy Science*, 74(4), 1428-1432. doi: 10.3168/jds.S0022-0302(91)78299-4
- Hæggman, S., Löfdahl, S., Paauw, A., Verhoef, J., & Brisse, S. (2004). Diversity and evolution of the class A chromosomal beta-lactamase gene in *Klebsiella pneumoniae*. *Antimicrobial Agents and Chemotherapy*, 48(7), 2400-2408. doi: 10.1128/AAC.48.7.2400-2408.2004
- Haenni, M., Saras, E., Châtre, P., Médaille, C., Bes, M., Madec, J. Y., & Laurent, F. (2012). A USA300 variant and other human-related methicillin-resistant *Staphylococcus aureus* strains infecting cats and dogs in France. *Journal of Antimicrobial Chemotherapy*, 67(2), 326-329. doi: 10.1093/jac/dkr499

- HAIAP. (2013). Antibiotic use and antibiotic resistance in food animals in Malaysia: A threat to human and animal health. *Health Action International Asia Pacific (HAIAP)*. Retrieved from <http://www.haiasiapacific.org/wp-content/uploads/2014/06/Memo-on-Antibiotics-in-animal-feeds-the-case-for-Malaysia-21-Nov-2013-V1.pdf>
- Han, P., Luo, L., Shahid, M., Li, M., Wang, S., Nobrega, D. B., Sabir, N., Barkema, H. W., Han, B., & Gao, J. (2020). Prevalence, genetic diversity and antimicrobial resistance of *Proteus mirabilis* isolated from dogs hospitalized in Beijing. *Pakistan Veterinary Journal*, 40(1), 61-66. doi: 10.29261/pakvetj/2019.101
- Harada, K., Shimizu, T., Mukai, Y., Kuwajima, K., Sato, T., Usui, M., Tamura, Y., Kimura, Y., Miyamoto, T., Tsuyuki, Y., Ohki, A., & Kataoka, Y. (2016). Phenotypic and molecular characterization of antimicrobial resistance in *Klebsiella* spp. isolates from companion animals in Japan: Clonal dissemination of multidrug-resistant extended-spectrum β-lactamase-producing *Klebsiella pneumoniae*. *Frontiers in Microbiology*, 7, 1-12. doi: 10.3389/fmicb.2016.01021
- Hariharan, H., Matthew, V., Fountain, J., Snell, A., Doherty, D., King, B., Shemer, E., Oliveira, S., & Sharma, R. N. (2011). Aerobic bacteria from mucous membranes, ear canals, and skin wounds of feral cats in Grenada, and the antimicrobial drug susceptibility of major isolates. *Comparative Immunology, Microbiology and Infectious Diseases*, 34(2), 129-134. doi: 10.1016/j.cimid.2010.05.001
- Hartantyo, S. H. P., Chau, M. L., Fillon, L., Ariff, A. Z. B. M., Kang, J. S. L., Aung, K. T., & Gutiérrez, R. A. (2018). Sick pets as potential reservoirs of antibiotic-resistant bacteria in Singapore. *Antimicrobial Resistance and Infection Control*, 7(1), 1-3. doi: 10.1186/s13756-018-0399-9
- Hassali, M. A., Yann, H. R., Verma, A. K., Hussain, R., & Sivaraman, S. (2018). Antibiotic use in food animals : Malaysia overview. *Discipline of Social & Administrative Pharmacy. School of Pharmaceutical Sciences. Universiti Sains Malaysia*. Retrieved from [https://www.reactgroup.org/wp-content/uploads/2018/11/Antibiotic\\_Use\\_in\\_Food\\_Animals\\_MalaysiaOverview\\_2018web.pdf](https://www.reactgroup.org/wp-content/uploads/2018/11/Antibiotic_Use_in_Food_Animals_MalaysiaOverview_2018web.pdf)
- Hassan, A. N., & Frank, J. F. (2011). Microorganisms associated with milk. *Encyclopedia of Dairy Sciences: Second Edition*, 447-457. doi: 10.1016/B978-0-12-374407-4.00309-5
- Hegde, N. V., Cook, M. L., Wolfgang, D. R., Love, B. C., Maddox, C. C., & Jayarao, B. M. (2005). Dissemination of *Salmonella enterica* subsp. *enterica* serovar *Typhimurium* var. *Copenhagen* clonal types through a contract heifer-raising operation. *Journal of Clinical Microbiology*, 43(8), 4208-4211. doi: 10.1128/JCM.43.8.4208-4211.2005

- Hernández-Cortez, C., Palma-Martínez, I., Gonzalez-Avila, L. U., Guerrero-Mandujano, A., Solís, R. C., & Castro-Escarpulli, G. (2017). Food poisoning caused by bacteria (food toxins). *Poisoning - From Specific Toxic Agents to Novel Rapid and Simplified Techniques for Analysis*, 33. doi: 10.5772/intechopen.69953
- Hernando-Amado, S., Coque, T. M., Baquero, F., & Martínez, J. L. (2019). Defining and combating antibiotic resistance from one health and global health perspectives. *Nature Microbiology*, 4(9), 1432-1442. doi: 10.1038/s41564-019-0503-9
- Hesp, A., Veldman, K., van der Goot, J., Mevius, D., & van Schaik, G. (2019). Monitoring antimicrobial resistance trends in commensal *Escherichia coli* from livestock, the Netherlands, 1998 to 2016. *Eurosurveillance*, 24(25), 1800438. doi: 10.2807/1560-7917.ES.2019.24.25.1800438
- Heuer, O. E., Jensen, V. F., & Hammerum, A. M. (2005). Antimicrobial drug consumption in companion animals. *Emerging Infectious Diseases*, 11(2), 344. doi: 10.3201/eid1102.040827
- Hoelzer, K., Wong, N., Thomas, J., Talkington, K., Jungman, E., & Coukell, A. (2017). Antimicrobial drug use in food-producing animals and associated human health risks: What, and how strong, is the evidence? *BMC Veterinary Research*, 13(1), 1-38. doi: 10.1186/s12917-017-1131-3
- Hong, S., Rovira, A., Davies, P., Ahlstrom, C., Muellner, P., Rendahl, A., Olsen, K., Bender, J. B., Wells, S., Perez, A., & Alvarez, J. (2016). Serotypes and antimicrobial resistance in *Salmonella enterica* recovered from clinical samples from cattle and swine in minnesota, 2006 to 2015. *PLoS ONE*, 11(12), e0168016. doi: 10.1371/journal.pone.0168016
- Hu, Y., & Cheng, H. (2014). Research opportunities for antimicrobial resistance control in China's factory farming. *Environmental Science & Technology*, 48(10), 5364–5365. doi: 10.1021/es502032c
- Huang, X., Yu, L., Chen, X., Zhi, C., Yao, X., Liu, Y., Wu, S., Guo, Z., Yi, L., Zeng, Z., & Liu, J. H. (2017). High prevalence of colistin resistance and mcr-1 gene in *Escherichia coli* isolated from food animals in China. *Frontiers in Microbiology*, 8, 1-5. doi: 10.3389/fmicb.2017.00562
- Huber, H., Zweifel, C., Wittenbrink, M. M., & Stephan, R. (2013). ESBL-producing uropathogenic *Escherichia coli* isolated from dogs and cats in Switzerland. *Veterinary Microbiology*, 162(2-4), 992-996. doi: 10.1016/j.vetmic.2012.10.029
- Hur, J., Jawale, C., & Lee, J. H. (2012). Antimicrobial resistance of *Salmonella* isolated from food animals: A review. *Food Research International*, 45(2), 819-830. doi: 10.1016/j.foodres.2011.05.014

- Hutchings, M. I., Truman, A. W., & Wilkinson, B. (2019). Antibiotics: past, present and future. *Current Opinion in Microbiology*, 51, 72–80. doi: 10.1016/j.mib.2019.10.008
- IACG. (2018). Interagency coordination group on antimicrobial resistance. Antimicrobial resistance: national action plans. *National Action Plans*, 1-16. doi: 10.1080/17441692.2018.1449230
- Iannini, P. B., Paladino, J. A., Lavin, B., Singer, M. E., & Schentag, J. J. (2007). A case series of macrolide treatment failures in community acquired pneumonia. *Journal of Chemotherapy*, 19(5), 536-545. doi: 10.1179/joc.2007.19.5.536
- Igbinosa & Beshiru. (2018). Detection and antibiotic-resistance of *Salmonella* species and *Escherichia coli* from selected captive animals in Ogbia zoological garden. *Nigerian Journal of Pure and Applied Sciences*, 31 (1), 3171-3079. doi: 10.7910/DVN/YXZH6Z
- Irizarry, R., Amadi, V., Brathwaite-Sylvester, E., Nicholas-Thomas, R., Sharma, R., & Hariharan, H. (2017). Update on urinary tract infections in dogs in a tropical island and antimicrobial susceptibility of *Escherichia coli* isolates for the period 2010-2016. *Veterinary Medicine - Open Journal*, 1(3), 56-61. doi: 10.17140/vmoj-2-110
- Iseppi, R., Messi, P., Anacarso, I., Bondi, M., Sabia, C., Condò, C., & De Niederhausern, S. (2015). Antimicrobial resistance and virulence traits in *Enterococcus* strains isolated from dogs and cats. *New Microbiologica*, 38(3), 369-378. Retrieved from <https://iris.unimore.it/retrieve/handle/11380/1105314/79666/369.pdf>
- Jacoby, G. A., & Munoz-Price, L. S. (2005). The new β-lactamases. *New England Journal of Medicine*, 352(4), 380-391. doi: 10.1056/nejmra041359
- Jang, S. S., Biberstein, E. L., & Hirsh, D. C. (2008). A Diagnostic: Manual of Veterinary Clinical Bacteriology and Mycology. University of California Davis.
- Jajere, S. M., Latiffah, H., Zunita, Z., Saleha, A. A., & Jalila, A. (2019). Epidemiology, antibiograms and multilocus sequence typing of *Salmonella enterica* among ayam kampung (*Gallus domesticus*) from the South Central, Peninsular Malaysia [Doctoral dissertation, Universiti Putra Malaysia].
- Jessen, L. R., Sørensen, T. M., Bjornvad, C. R., Nielsen, S. S., & Guardabassi, L. (2015). Effect of antibiotic treatment in canine and feline urinary tract infections: A systematic review. *Veterinary Journal*, 203(3), 270-277. doi: 10.1016/j.tvjl.2014.12.004

Jiang, H. X., Lü, D. H., Chen, Z. L., Wang, X. M., Chen, J. R., Liu, Y. H., Liao, X. P., Liu, J. H., & Zeng, Z. L. (2011). High prevalence and widespread distribution of multi-resistant *Escherichia coli* isolates in pigs and poultry in China. *Veterinary Journal*, 187(1), 99-103. doi: 10.1016/j.tvjl.2009.10.017

Joosten, P., Ceccarelli, D., Odent, E., Sarrazin, S., Graveland, H., Van Gompel, L., Battisti, A., Caprioli, A., Franco, A., Wagenaar, J. A., Mevius, D., & Dewulf, J. (2020). Antimicrobial usage and resistance in companion animals: A cross-sectional study in three european countries. *Antibiotics*, 9(2), 87-104. doi: 10.3390/antibiotics9020087

Kaesbohrer, A., Schroeter, A., Tenhagen, B. A., Alt, K., Guerra, B., & Appel, B. (2012). Emerging antimicrobial resistance in commensal *Escherichia coli* with public health relevance. *Zoonoses and Public Health*, 59, 158-165. doi: 10.1111/j.1863-2378.2011.01451.x

Kalhoro, D. H., Kalhoro, M. S., Mangi, M. H., Jahejo, A. R., Kumbhar, S., Lochi, G. M., Mari, G. M., Kaka, A., Lund, A. K., & Liu, Y. J. (2019). Antimicrobial resistance of *Staphylococci* and *Streptococci* isolated from dogs. *Tropical Biomedicine*, 36(2), 468-474. Retrieved from <https://msptm.org/files/Vol36No2/468-474-Kalhoro-DH.pdf>

Kanwar, N., Scott, H. M., Norby, B., Loneragan, G. H., Vinasco, J., Cottell, J. L., Chalmers, G., Chengappa, M. M., Bai, J., & Boerlin, P. (2014). Impact of treatment strategies on cephalosporin and tetracycline resistance gene quantities in the bovine fecal metagenome. *Scientific Reports*, 4(1), 5100. doi: 10.1038/srep05100

Karczmarczyk, M., Abbott, Y., Walsh, C., Leonard, N., & Fanning, S. (2011). Characterization of multidrug-resistant *Escherichia coli* isolates from animals presenting at a University Veterinary Hospital. *Applied and Environmental Microbiology*, 77(20), 7104-7112. doi: 10.1128/AEM.00599-11

Karesh, W. B., Dobson, A., Lloyd-Smith, J. O., Lubroth, J., Dixon, M. A., Bennett, M., Aldrich, S., Harrington, T., Formenty, P., Loh, E. H., MacHalaba, C. C., Thomas, M. J., & Heymann, D. L. (2012). Ecology of zoonoses: Natural and unnatural histories. *The Lancet*, 380(9857), 1936-1945. doi: 10.1016/S0140-6736(12)61678-X

Katakweba, A. A. S. (2014). Prevalence of antimicrobial resistance and characterization of fecal indicator bacteria and *Staphylococcus aureus* from farm animals, wildlife, pets and humans in Tanzania Abdul. [Doctoral dissertation, Sokoine University of Agriculture]. doi:10.1590/S1517-83822011000200008

Kazemnia, A., Ahmadi, M., & Dilmaghani, M. (2014). Antibiotic resistance pattern of different *Escherichia coli* phylogenetic groups isolated from human urinary tract infection and avian colibacillosis. *Iranian Biomedical Journal*, 18(4), 219. doi: 10.6091/ibj.1394.2014

- Kirchhelle, C. (2018). Swann song: Antibiotic regulation in British livestock production (1953-2006). *Bulletin of the History of Medicine*, 92(2), 317-350. doi: 10.1353/bhm.2018.0029
- Ko, W. C., Paterson, D. L., Sagnimeni, A. J., Hansen, D. S., Von Gottberg, A., Mohapatra, S., Casellas, J. M., Goossens, H., Mulazimoglu, L., Trenholme, G., Klugman, K. P., McCormack, J. G., & Yu, V. L. (2002). Community-acquired *Klebsiella pneumoniae* bacteremia: Global differences in clinical patterns. *Emerging Infectious Diseases*, 8(2), 160. doi: 10.3201/eid0802.010025
- Köck, R., Daniels-Haardt, I., Becker, K., Mellmann, A., Friedrich, A. W., Mevius, D., Schwarz, S., & Jurke, A. (2018). Carbapenem-resistant Enterobacteriaceae in wildlife, food-producing, and companion animals: a systematic review. *Clinical Microbiology and Infection*, 24(12), 1241-1250. doi: 10.1016/j.cmi.2018.04.004
- Kolumna, A., & Dikici, A. (2013). Antimicrobial resistance of emerging foodborne pathogens: Status quo and global trends. *Critical Reviews in Microbiology*, 39(1), 57-69. doi: 10.3109/1040841X.2012.691458
- Kroemer, S., El Garch, F., Galland, D., Petit, J. L., Woehrle, F., & Boulouis, H. J. (2014). Antibiotic susceptibility of bacteria isolated from infections in cats and dogs throughout Europe (2002-2009). *Comparative Immunology, Microbiology and Infectious Diseases*, 37(2), 97-108. doi: 10.1016/j.cimid.2013.10.001
- Kuan, E. C., Yoon, A. J., Vijayan, T., Humphries, R. M., & Suh, J. D. (2016). Canine *Staphylococcus pseudintermedius* sinonasal infection in human hosts. *International Forum of Allergy and Rhinology*, 6(7), 710-715. doi: 10.1002/alr.21732
- KuKanich, K., Lubbers, B., & Salgado, B. (2020). Amoxicillin and amoxicillin-clavulanate resistance in urinary *Escherichia coli* antibiograms of cats and dogs from the Midwestern United States. *Journal of Veterinary Internal Medicine*, 34(1), 227-231. doi: 10.1111/jvim.15674
- Kunz, F., Corti, S., Giezendanner, N., Stephan, R., Wittenbrink, M. M., & Zweifel, C. (2011). Antimicrobial resistance of *Staphylococcus aureus* and coagulase negative *Staphylococci* isolated from mastitis milk samples from sheep and goats. *Schweizer Archiv Fur Tierheilkunde*, 153(2), 63-69. doi: 10.1024/0036-7281/a000152
- Landers, T. F., Cohen, B., Wittum, T. E., & Larson, E. L. (2012). A review of antibiotic use in food animals: Perspective, policy, and potential. *Public Health Reports*, 127(1), 4-22. doi: 10.1177/003335491212700103

- Lanz, R., Kuhnert, P., & Boerlin, P. (2003). Antimicrobial resistance and resistance gene determinants in clinical *Escherichia coli* from different animal species in Switzerland. *Veterinary Microbiology*, 91(1), 73-84. doi: 10.1016/S0378-1135(02)00263-8
- Lappin, M. R., Blondeau, J., Boothe, D., Breitschwerdt, E. B., Guardabassi, L., Lloyd, D. H., Papich, M. G., Rankin, S. C., Sykes, J. E., Turnidge, J., & Weese, J. S. (2017). Antimicrobial use guidelines for treatment of respiratory tract disease in dogs and cats: Antimicrobial guidelines working group of the international society for companion animal infectious diseases. *Journal of Veterinary Internal Medicine*, 31(2), 279-294. doi: 10.1111/jvim.14627
- Lee, J. H., Park, K. S., Jeon, J. H., & Lee, S. H. (2018). Antibiotic resistance in soil. *The Lancet Infectious Diseases*, 18(12), 1306–1307. doi: 10.1016/S1473-3099(18)30675-3
- Leenstra, T., Monen, J., S, W., Altorf-van der Kuil, W., Sande-Bruinsma, van de, Lo Fo Wong, D., Nahrgang, S., Giske, C. G., & Skov, R. (2014). Central Asian and Eastern European surveillance of antimicrobial resistance annual report 2014. *World Health Organization Press*, 1-72. Retrieved from [https://www.euro.who.int/\\_\\_data/assets/pdf\\_file/0006/285405/CAESAR-Surveillance-Antimicrobial-Resistance2014.pdf](https://www.euro.who.int/__data/assets/pdf_file/0006/285405/CAESAR-Surveillance-Antimicrobial-Resistance2014.pdf)
- Lei, T., Tian, W., He, L., Huang, X. H., Sun, Y. X., Deng, Y. T., Sun, Y., Lv, D. H., Wu, C. M., Huang, L. Z., Shen, J. Z., & Liu, J. H. (2010). Antimicrobial resistance in *Escherichia coli* isolates from food animals, animal food products and companion animals in China. *Veterinary Microbiology*, 146(1-2), 85-89. doi: 10.1016/j.vetmic.2010.04.025
- Leonard, E. K., Pearl, D. L., Finley, R. L., Janecko, N., Reid-Smith, R. J., Peregrine, A. S., & Weese, J. S. (2012). Comparison of antimicrobial resistance patterns of *Salmonella* spp. and *Escherichia coli* recovered from pet dogs from volunteer households in Ontario (2005-06). *Journal of Antimicrobial Chemotherapy*, 67(1), 174-181. doi: 10.1093/jac/dkr430
- Lessing, A. (2010). Killing us softly: How sub-therapeutic dosing of livestock causes drug-resistant bacteria in humans causes drug-resistant bacteria. *Boston College Environmental Affairs Law Review*, 37, 463. Retrieved from <https://heinonline.org/HOL/LandingPage?handle=hein.journals/bcenv37&div=18&id=&page=>
- Liljeblad, K. A., Hofacre, C. L., White, D. G., Ayers, S., Lee, M. D., & Maurer, J. J. (2017). Diversity of antimicrobial resistance phenotypes in *Salmonella* isolated from commercial poultry farms. *Frontiers in Veterinary Science*, 4, 96. doi: 10.3389/fvets.2017.00096

- Liu, X. Q., Wang, J., Li, W., Zhao, L. Q., Lu, Y., Liu, J. H., & Zeng, Z. L. (2017). Distribution of cfr in *Staphylococcus* spp. and *Escherichia coli* strains from pig farms in China and characterization of a novel cfr-carrying F43: A- B-plasmid. *Frontiers in Microbiology*, 8, 329. doi: 10.3389/fmicb.2017.00329
- Liu, X., Thungrat, K., & Boothe, D. M. (2016). Occurrence of oxa-48 carbapenemase and other β-lactamase genes in Esbl-producing multidrug resistant: *Escherichia coli* from dogs and cats in the united states, 2009-2013. *Frontiers in Microbiology*, 7, 1057. doi: 10.3389/fmicb.2016.01057
- Ljubojevic, D., Pelic, M., Puvača, N., & Milanov, D. (2017). Resistance to tetracycline in *Escherichia coli* isolates from poultry meat: Epidemiology, policy and perspective. *World's Poultry Science Journal*, 73(2), 409-417. doi: 10.1017/S0043933917000216
- Looft, T., Johnson, T. A., Allen, H. K., Bayles, D. O., Alt, D. P., Stedtfeld, R. D., Sul, W. J., Stedtfeld, T. M., Chai, B., Cole, J. R., Hashsham, S. A., Tiedje, J. M., & Stanton, T. B. (2012). In-feed antibiotic effects on the swine intestinal microbiome. *Proceedings of the National Academy of Sciences of the United States of America*, 109(5), 1691-1696. doi: 10.1073/pnas.1120238109
- Love, D. N. (1996). *Escherichia coli* in domestic animals and humans. *Veterinary Microbiology*, 1(52), 181-182. doi: 10.1016/0378-1135(96)00021-1
- Ludwig, C., de Jong, A., Moyaert, H., El Garch, F., Janes, R., Klein, U., Morrissey, I., Thiry, J., & Youala, M. (2016). Antimicrobial susceptibility monitoring of dermatological bacterial pathogens isolated from diseased dogs and cats across Europe (ComPath results). *Journal of Applied Microbiology*, 121(5), 1254-1267. doi: 10.1111/jam.13287
- Magiorakos, A. P., Srinivasan, A., Carey, R. B., Carmeli, Y., Falagas, M. E., Giske, C. G., Harbarth, S., Hindler, J. F., Kahlmeter, G., Olsson-Liljequist, B., Paterson, D. L., Rice, L. B., Stelling, J., Struelens, M. J., Vatopoulos, A., Weber, J. T., & Monnet, D. L. (2012). Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: An international expert proposal for interim standard definitions for acquired resistance. *Clinical Microbiology and Infection*, 18(3), 268-281. doi: 10.1111/j.1469-0691.2011.03570.x
- Maniam, R., Abdullah, F. F. J., Zakaria, Z., Saad, M. Z., & Salleh, A. (2020). The level of endotoxin in organs, antibiotic sensitivity, and serotyping of bacteria isolated from cats and dogs with septicaemia. *Advances in Animal and Veterinary Sciences*, 8(6), 614-623. doi: 10.17582/journal.aavs /2020/8.6.614.623

- Manyi-Loh, C., Mamphweli, S., Meyer, E., & Okoh, A. (2018). Antibiotic use in agriculture and its consequential resistance in environmental sources: Potential public health implications. *Molecules*, 23(4), 795. doi: 10.3390/molecules23040795
- Marquardt, R. R., & Li, S. (2018). Antimicrobial resistance in livestock: Advances and alternatives to antibiotics. *Animal Frontiers*, 8(2), 30-37. doi: 10.1093/af/vfy001
- Marques, C., Belas, A., Aboim, C., Cavaco-Silva, P., Trigueiro, G., Gama, L. T., & Pomba, C. (2019). Evidence of sharing of *Klebsiella pneumoniae* strains between healthy companion animals and cohabiting humans. *Journal of Clinical Microbiology*, 57(6), 1-12. doi: 10.1128/JCM.01537-18
- Marques, C., Belas, A., Franco, A., Aboim, C., Gama, L. T., & Pomba, C. (2018). Increase in antimicrobial resistance and emergence of major international high-risk clonal lineages in dogs and cats with urinary tract infection: 16 year retrospective study. *Journal of Antimicrobial Chemotherapy*, 73(2), 377-384. doi: 10.1093/jac/dkx401
- Marques, C., Gama, L. T., Belas, A., Bergström, K., Beurlet, S., Briand-Marchal, A., Broens, E. M., Costa, M., Criel, D., Damborg, P., van Dijk, M. A. M., van Dongen, A. M., Dorsch, R., Espada, C. M., Gerber, B., Kritsepi-Konstantinou, M., Loncaric, I., Mion, D., Misic, D., & Pomba, C. (2016). European multicenter study on antimicrobial resistance in bacteria isolated from companion animal urinary tract infections. *BMC Veterinary Research*, 12(1), 1-17. doi: 10.1186/s12917-016-0840-3
- Marques, C., Menezes, J., Belas, A., Aboim, C., Cavaco-Silva, P., Trigueiro, G., Gama, L. T., & Pomba, C. (2019). *Klebsiella pneumoniae* causing urinary tract infections in companion animals and humans: Population structure, antimicrobial resistance and virulence genes. *Journal of Antimicrobial Chemotherapy*, 74(3), 594-602. doi: 10.1093/jac/dky499
- Marshall, B. M., & Levy, S. B. (2011). Food animals and antimicrobials: Impacts on human health. *Clinical Microbiology Reviews*, 24(4), 718-733. doi: 10.1128/CMR.00002-11
- Martinez, J. L. (2008). Antibiotics and antibiotic resistance genes in natural environments. *Science*, 321(5887), 365–367. doi: 10.1126/science.1159483
- Martins, T., Rosa, A. F., Castelani, L., Miranda, M. S. D., Arcaro, J. R. P., & Pozzi, C. R. (2016). Intramammary treatment with gentamicin in lactating cows with clinical and subclinical mastitis. *Pesquisa Veterinaria Brasileira*, 36(4), 283-289. doi: 10.1590/S0100-736X2016000400006
- Matin, M. A., Islam, M. A., & Khatun, M. M. (2017). Prevalence of colibacillosis in chickens in greater Mymensingh district of Bangladesh. *Veterinary World*, 10(1), 29. doi: 10.14202/vetworld.2017.29-33

- McEachran, A. D., Blackwell, B. R., Hanson, J. D., Wooten, K. J., Mayer, G. D., Cox, S. B., & Smith, P. N. (2015). Antibiotics, bacteria, and antibiotic resistance genes: aerial transport from cattle feed yards via particulate matter. *Environmental Health Perspectives*, 123(4), 337–343. doi: 10.1289/ehp.1408555
- McEwen, S. A., & Fedorka-Cray, P. J. (2002). Antimicrobial use and resistance in animals. *Clinical Infectious Diseases*, 34(3), 93-106. doi: 10.1086/340246
- McMeekin, C. H., Hill, K. E., Gibson, I. R., Bridges, J. P., & Benschop, J. (2017). Antimicrobial resistance patterns of bacteria isolated from canine urinary samples submitted to a New Zealand veterinary diagnostic laboratory between 2005–2012. *New Zealand Veterinary Journal*, 65(2), 99-104. doi: 10.1080/00480169.2016.1259594
- Meade E, Slattery MA, G. M. (2017). Antimicrobial resistance: an agent in zoonotic disease and increased morbidity. *Journal of Clinical Experimental Toxicology*, 1(1), 30-37. Retrieved from <https://www.alliedacademies.org/articles/antimicrobial-resistance-an-agent-in-zoonotic-disease-and-increased-morbidity-9144.html>
- Mendonça, N., Figueiredo, R., Mendes, C., Card, R. M., Anjum, M. F., & da Silva, G. J. (2016). Microarray evaluation of antimicrobial resistance and virulence of *Escherichia coli* isolates from Portuguese poultry. *Antibiotics*, 5(1), 4. doi: 10.3390/antibiotics5010004
- Merkeviciene, L., Klimiene, I., Siugzdiniene, R., Virgailis, M., Mockeliunas, R., & Ruzauskas, M. (2018). Prevalence and molecular characteristics of multi-resistant *Escherichia coli* in wild birds. *Acta Veterinaria Brno*, 87(1), 9–17. doi: 10.2754/avb201887010009
- Miles, T. D., McLaughlin, W., & Brown, P. D. (2006). Antimicrobial resistance of *Escherichia coli* isolates from broiler chickens and humans. *BMC Veterinary Research*, 2(1), 1-9. doi: 10.1186/1746-6148-2-7
- Ministry of Health Malaysia. (2017a). Malaysian action plan on antimicrobial resistance (MyAP-AMR) 2017-2021. *Ministry of Health Malaysia*. Retrieved from [https://www.moh.gov.my/moh/resources/Penerbitan/Garis%20Panduan/Garis%20panduan%20Umum%20\(Awam\)/National\\_Action\\_Plan\\_-\\_FINAL\\_29\\_june.pdf](https://www.moh.gov.my/moh/resources/Penerbitan/Garis%20Panduan/Garis%20panduan%20Umum%20(Awam)/National_Action_Plan_-_FINAL_29_june.pdf)
- Ministry of Health Malaysia. (2017c). National Surveillance of Antibiotic Resistance (NSAR) Report 2017. *Institute for medical research*. doi: org/10.1002/art.23552

- Moawad, A. A., Hotzel, H., Awad, O., Tomaso, H., Neubauer, H., Hafez, H. M., & El-Adawy, H. (2017). Occurrence of *Salmonella enterica* and *Escherichia coli* in raw chicken and beef meat in northern Egypt and dissemination of their antibiotic resistance markers. *Gut Pathogens*, 9(1), 1-13. doi: 10.1186/s13099-017-0206-9
- Mohamed, M. A., Abdul-Aziz, S., Dhaliwal, G. K., Bejo, S. K., Goni, M. D., Bitrus, A. A., & Muhammad, J. I. (2017). Antibiotic resistance profiles of *Staphylococcus pseudintermedius* isolated from dogs and cats. *Malaysian Journal of Microbiology*, 13(3), 180-186. doi: 10.21161/mjm.84616
- Montso, K. P., Dlamini, S. B., Kumar, A., Ateba, C. N., & Garcia-Perdomo, H. A. (2019). Antimicrobial resistance factors of extended-spectrum beta-lactamases producing *Escherichia coli* and *Klebsiella pneumoniae* isolated from cattle farms and raw beef in North-West Province, South Africa. *BioMed Research International*, 2019, 1-13. doi: 10.1155/2019/4318306
- Moodley, A., Damborg, P., & Nielsen, S. S. (2014). Antimicrobial resistance in methicillin susceptible and methicillin resistant *Staphylococcus pseudintermedius* of canine origin: Literature review from 1980 to 2013. *Veterinary Microbiology*, 171(3-4), 337-341. doi: 10.1016/j.vetmic.2014.02.008
- Morley, P. S., Apley, M. D., Besser, T. E., Burney, D. P., Fedorka-Cray, P. J., Papich, M. G., Traub-Dargatz, J. L., & Weese, J. S. (2005). Antimicrobial drug use in veterinary medicine. *Journal of Veterinary Internal Medicine*, 19(4), 617-629. doi: 10.1892/0891-6640
- Moulin, G., Cavalié, P., Pellanne, I., Chevance, A., Laval, A., Millemann, Y., Colin, P., & Chauvin, C. (2008). A comparison of antimicrobial usage in human and veterinary medicine in France from 1999 to 2005. *Journal of Antimicrobial Chemotherapy*, 62(3), 617-625. doi: 10.1093/jac/dkn213
- Moyaert, H., de Jong, A., Simjee, S., Rose, M., Youala, M., El Garch, F., Vila, T., Klein, U., Rzewuska, M., & Morrissey, I. (2019). Survey of antimicrobial susceptibility of bacterial pathogens isolated from dogs and cats with respiratory tract infections in Europe: ComPath results. *Journal of Applied Microbiology*, 127(1), 29-46. doi: 10.1111/jam.14274
- Moyaert, Hilde, Morrissey, I., De Jong, A., El Garch, F., Klein, U., Ludwig, C., Thiry, J., & Youala, M. (2017). Antimicrobial susceptibility monitoring of bacterial pathogens isolated from urinary tract infections in dogs and cats across Europe: ComPath results. *Microbial Drug Resistance*, 23(3), 391-403. doi: 10.1089/mdr.2016.0110
- Mund, M. D., Khan, U. H., Tahir, U., Mustafa, B. E., & Fayyaz, A. (2017). Antimicrobial drug residues in poultry products and implications on public health: A review. *International Journal of Food Properties*, 20(7), 1433-1446. doi: 10.1080/10942912.2016.1212874

- Munoz, M. A., Welcome, F. L., Schukken, Y. H., & Zadoks, R. N. (2007). Molecular epidemiology of two *Klebsiella pneumoniae* mastitis outbreaks on a dairy farm in New York State. *Journal of Clinical Microbiology*, 45(12), 3964-3971. doi: 10.1128/JCM.00795-07
- Mwanyika, G., Call, D. R., Rugumisa, B., Luanda, C., Murutu, R., Subbiah, M., & Buza, J. (2016). Load and prevalence of antimicrobial-resistant *Escherichia coli* from fresh goat meat in Arusha, Tanzania. *Journal of Food Protection*, 79(9), 1635-1641. doi: 10.4315/0362-028X.JFP-15-573
- National Pharmaceutical Regulatory Agency. (2014). Registration Guidelines of Veterinary Products (REGOVP). Third Version. Retrieved from [https://www.npra.gov.my/images/Guidelines\\_Central/Guidelines\\_on\\_Veterinary/REGOVP\\_JULY2014\\_191015.pdf](https://www.npra.gov.my/images/Guidelines_Central/Guidelines_on_Veterinary/REGOVP_JULY2014_191015.pdf)
- Navon-Venezia, S., Kondratyeva, K., & Carattoli, A. (2017). *Klebsiella pneumoniae*: A major worldwide source and shuttle for antibiotic resistance. *FEMS Microbiology Reviews*, 41(3), 252-275. doi: 10.1093/femsre/fux013
- Nidaullah, H., Abirami, N., Shamila-Syuhada, A. K., Chuah, L. O., Huda, N., Tan, T. P., Zainal Abidin, F. W., & Rusul, G. (2017). Prevalence of *Salmonella* in poultry processing environments in wet markets in Penang and Perlis, Malaysia. *Veterinary World*, 10(3), 286. doi: 10.14202/vetworld.2017.286-292
- Nor, F. M., Shahari, A. S., Palaniasamy, N. K., Rustam, F. R. M., M-Zain, Z., Lee, B. P. K., & Soh, T. S. T. (2019). Multidrug resistant (MDR) *Acinetobacter baumannii*: rate of occurrence from a tertiary hospital, Malaysia. *International Journal of Infectious Diseases*, 79, 46-47. doi: 10.1016/j.ijid.2018.11.126
- Nordmann, P., & Poirel, L. (2014). The difficult-to-control spread of carbapenemase producers among Enterobacteriaceae worldwide. *Clinical Microbiology and Infection*, 20(9), 821-830. doi: 10.1111/1469-0691.12719
- Norlida, O., & Bahaman, A. R. (2015). Prevalence of subclinical mastitis and antibiotic resistant bacteria in three selected cattle farms in Serdang, Selangor and Kluang, Johor. *Malaysia Journal of Veterinary*, 17(1), 27-31. Retrieved from <http://psasir.upm.edu.my/id/eprint/41537/1/0001.pdf>
- Normand, E. H., Gibson, N. R., Reid, S. W. J., Carmichael, S., & Taylor, D. J. (2000). Antimicrobial-resistance trends in bacterial isolates from companion-animal community practice in the UK. *Preventive Veterinary Medicine*, 46(4), 267-278. doi: 10.1016/S0167-5877(00)00149-5
- Nyachuba, D. G. (2010). Foodborne illness: Is it on the rise? *Nutrition Reviews*, 68(5), 257-269. doi: 10.1111/j.1753-4887.2010.00286.x

- O'Neill, J. (2014). Antimicrobial Resistance : Tackling a crisis for the health and wealth of nations. *Review on Antimicrobial Resistance*, 1–16. Retrieved from  
[https://amr-review.org/sites/default/files/160525\\_Final%20paper\\_with%20cover.pdf](https://amr-review.org/sites/default/files/160525_Final%20paper_with%20cover.pdf)
- Oh, J. Y., Kwon, Y. K., Tamang, M. D., Jang, H. K., Jeong, O. M., Lee, H. S., & Kang, M. S. (2016). Plasmid-mediated quinolone resistance in *Escherichia coli* isolates from wild birds and chickens in South Korea. *Microbial Drug Resistance*, 22(1), 69–79. doi: 10.1089/mdr.2015.0090
- OIE. (2007). OIE World organization for animal health. List of Antimicrobials of Veterinary Importance. Retrieved from  
[https://www.oie.int/fileadmin/Home/eng/International\\_Standard\\_Setting/documents/pdf/OIE\\_list\\_antimicrobials.pdf](https://www.oie.int/fileadmin/Home/eng/International_Standard_Setting/documents/pdf/OIE_list_antimicrobials.pdf)
- OÖsterblad, M., Norrdahl, K., Korpimaäki, E., & Huovinen, P. (2001). How wild are wild mammals? *Nature*, 409(6816), 37–38. doi: 10.1038/35051176
- Opintan, J. A., Newman, M. J., Arhin, R. E., Donkor, E. S., Gyansa-Lutterodt, M., & Mills-Pappoe, W. (2015). Laboratory-based nationwide surveillance of antimicrobial resistance in Ghana. *Infection and Drug Resistance*, 8, 379. doi: 10.2147/IDR.S88725
- Organisation for Economic Co-operation and Development. (2015). Global antimicrobial use in the livestock sector. *Working Party on Agricultural Policies and Markets*, 34, 1–43. Retrieved from  
[https://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=TAD/CA/APM/WP\(2014\)34/FINAL&docLanguage=En](https://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=TAD/CA/APM/WP(2014)34/FINAL&docLanguage=En)
- Oz, T., Guvenek, A., Yildiz, S., Karaboga, E., Tamer, Y. T., Mumcuyan, N., Ozan, V. B., Senturk, G. H., Cokol, M., Yeh, P., & Toprak, E. (2014). Strength of selection pressure is an important parameter contributing to the complexity of antibiotic resistance evolution. *Molecular Biology and Evolution*, 31(9), 2387–2401. doi: 10.1093/molbev/msu191
- Page, S. W., & Gautier, P. (2012). Use of antimicrobial agents in livestock. *OIE Revue Scientifique et Technique*, 31(1), 145. doi: 10.20506/rst.31.1.2106
- Park, Y. H., Hwang, S. Y., Hong, M. K., & Kwon, K. H. (2012). Use of antimicrobial agents in aquaculture. *Revue Scientifique et Technique de l'OIE*, 31(1), 189–197. doi: 10.20506/rst.31.1.2105
- Paveenkittiporn, W., Apisarnthanarak, A., Dejsirilert, S., Trakulsomboon, S., Thongmali, O., Sawanpanyalert, P., & Aswapeeke, N. (2009). Five-year surveillance for *Burkholderia pseudomallei* in Thailand from 2000 to 2004: prevalence and antimicrobial susceptibility. *Journal of the Medical Association of Thailand*, 92(8), 46–53. Retrieved from  
<https://pubmed.ncbi.nlm.nih.gov/21298845/>.

- Pedersen, K., Pedersen, K., Jensen, H., Finster, K., Jensen, V. F., & Heuer, O. E. (2007). Occurrence of antimicrobial resistance in bacteria from diagnostic samples from dogs. *Journal of Antimicrobial Chemotherapy*, 60(4), 775-781. doi: 10.1093/jac/dkm269
- Pellegrini, C., Mercuri, P. S., Celenza, G., Galleni, M., Segatore, B., Sacchetti, E., Volpe, R., Amicosante, G., & Perilli, M. (2009). Identification of blaIMP-22 in *Pseudomonas* spp. in urban wastewater and nosocomial environments: biochemical characterization of a new IMP metallo-enzyme variant and its genetic location. *Journal of Antimicrobial Chemotherapy*, 63(5), 901–908. doi: 10.1093/jac/dkp061
- Perrin-Guyomard, A., Jouy, E., Urban, D., Chauvin, C., Granier, S. A., Mourand, G., Chevance, A., Adam, C., Moulin, G., & Kempf, I. (2020). Decrease in fluoroquinolone use in French poultry and pig production and changes in resistance among *E. coli* and *Campylobacter*. *Veterinary Microbiology*, 243, 108637. doi: 10.1016/j.vetmic.2020.108637
- Peters, L., Olson, L., Khu, D. T. K., Linnros, S., Le, N. K., Hanberger, H., Hoang, N. T. B., Tran, D. M., & Larsson, M. (2019). Multiple antibiotic resistance as a risk factor for mortality and prolonged hospital stay: A cohort study among neonatal intensive care patients with hospital-acquired infections caused by gram-negative bacteria in Vietnam. *PLOS ONE*, 14(5), e0215666. doi: 10.1371/journal.pone.0215666
- Phophi, L., Petzer, I. M., & Qekwana, D. N. (2019). Antimicrobial resistance patterns and biofilm formation of coagulase-negative *Staphylococcus* species isolated from subclinical mastitis cow milk samples submitted to the Onderstepoort Milk Laboratory. *BMC veterinary research*, 15(1), 1-9. doi: 10.1186/s12917-019-2175-3
- Podder, M. P., Rogers, L., Daley, P. K., Keefe, G. P., Whitney, H. G., & Tahlan, K. (2014). *Klebsiella* species associated with bovine mastitis in Newfoundland. *PLoS ONE*, 9(9), e106518. doi: 10.1371/journal.pone.0106518
- Podschun, R., & Ullmann, U. (1998). *Klebsiella* spp. as nosocomial pathogens: Epidemiology, taxonomy, typing methods, and pathogenicity factors. *Clinical Microbiology Reviews*, 11(4), 589-603. doi.org/10.1128/cmr.11.4.589
- Pomba, C., Couto, N., & Moodley, A. (2010). Treatment of a lower urinary tract infection in a cat caused by a multi-drug methicillin-resistant *Staphylococcus pseudintermedius* and *Enterococcus faecalis*. *Journal of Feline Medicine and Surgery*, 12(10), 802-806. doi: 10.1016/j.jfms.2010.04.006

Pomba, C., Rantala, M., Greko, C., Baptiste, K. E., Catry, B., Van Duijkeren, E., Mateus, A., Moreno, M. A., Pyörälä, S., Ružauskas, M., Sanders, P., Teale, C., John Threlfall, E., Kunsagi, Z., Torren-Edo, J., Jukes, H., & Törneke, K. (2017). Public health risk of antimicrobial resistance transfer from companion animals. *Journal of Antimicrobial Chemotherapy*, 72(4), 957-968. doi: 10.1093/jac/dkw481

Pompilio, A., De Nicola, S., Crocetta, V., Guarnieri, S., Savini, V., Carretto, E., & Di Bonaventura, G. (2015). New insights in *Staphylococcus pseudintermedius* pathogenicity: Antibiotic-resistant biofilm formation by a human wound-associated strain. *BMC Microbiology*, 15(1), 1-14. doi: 10.1186/s12866-015-0449-x

Powers, J. H. (2004). Antimicrobial drug development – the past, the present, and the future. *Clinical Microbiology and Infection*, 10(4), 23–31. doi: 10.1111/j.1465-0691.2004.1007.x

Prescott, J. F., Brad Hanna, W. J., Reid-Smith, R., & Drost, K. (2002). Antimicrobial drug use and resistance in dogs. *Canadian Veterinary Journal*, 43(2), 107. doi: 10.1111/j.1751-0813.2002.tb11003.x

Priyantha, R., Gaunt, M. C., & Rubin, J. E. (2016). Antimicrobial susceptibility of *Staphylococcus pseudintermedius* colonizing healthy dogs in Saskatoon, Canada. *Canadian Veterinary Journal*, 57(1), 65. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4677612/>

Qekwana, D. N., Ogunju, J. W., Sithole, F., & Odoi, A. (2017). Burden and predictors of *Staphylococcus aureus* and *S. pseudintermedius* infections among dogs presented at an academic veterinary hospital in South Africa (2007-2012). *PeerJ*, 5, e3198. doi: 10.7717/peerj.3198

Quinn, P. J., Markey, B. K., Leonard, F. C., FitzPatrick, E. S., Fanning, S., & Hartigan, P. (2011). Bacterial causes of bovine mastitis. *Veterinary Microbiology and Microbial Disease*. John Wiley & Sons. Retrieved from <https://www.researchgate.net/publication/236026922>

Radhouani, H., Silva, N., Poeta, P., Torres, C., Correia, S., & Igrejas, G. (2014). Potential impact of antimicrobial resistance in wildlife, environment, and human health. *Frontiers in Microbiology*, 2(23), 1-12. doi: 10.3389/fmicb.2014.00023

Rahman, M. A., Rahman, A. K. M. A., Islam, M. A., & Alam, M. M. (2018). Antimicrobial resistance of *Escherichia coli* isolated from milk, beef and chicken meat in Bangladesh. *Bangladesh Journal of Veterinary Medicine*, 15(2), 141-146. doi: 10.3329/bjvm.v15i2.35525

Raji, M. A., Kwaga, J. O., Bale, J. O., & Henton, M. (2007). Biochemical and serological characterization of *Escherichia coli* isolated from colibacillosis and dead-in-shell embryos in poultry in Zaria, Nigeria. *Nigerian Veterinary Journal*, 27(2), 33-40. doi: 10.4314/nvj.v27i2.3513

- Ramey, A. M., Hernandez, J., Tyrlöv, V., Uher-Koch, B. D., Schmutz, J. A., Atterby, C., Järhult, J. D., & Bonnedahl, J. (2018). Antibiotic-resistant *Escherichia coli* in migratory birds inhabiting remote Alaska. *EcoHealth*, 15(1), 72–81. doi: 10.1007/s10393-017-1302-5
- Robb, A. R., Wright, E. D., Foster, A. M. E., Walker, R., & Malone, C. (2017). Skin infection caused by a novel strain of *Staphylococcus pseudintermedius* in a Siberian husky dog owner. *JMM Case Reports*, 4(3), 1-4. doi: 10.1099/jmmcr.0.005087
- Robinson, T. P., Bu, D. P., Carrique-Mas, J., Fèvre, E. M., Gilbert, M., Grace, D., Hay, S. I., Jiwakanon, J., Kakkar, M., Kariuki, S., Laxminarayan, R., Lubroth, J., Magnusson, U., Thi Ngoc, P., Van Boekel, T. P., & Woolhouse, M. E. J. (2016). Antibiotic resistance is the quintessential one health issue. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 110(7), 377-380. doi: 10.1093/trstmh/trw048
- Rozman, V., Bogovič Matijašić, B., & Smole Možina, S. (2019). Antimicrobial resistance of common zoonotic bacteria in the food chain: an emerging threat. *Antimicrobial Resistance - A Global Threat*. doi: 10.5772/intechopen.80782. Retrieved from <https://www.intechopen.com/books/antimicrobial-resistance-a-global-threat/antimicrobial-resistance-of-common-zoonotic-bacteria-in-the-food-chain-an-emerging-threat>
- Rubin, J. E., & Pitout, J. D. D. (2014). Extended-spectrum β-lactamase, carbapenemase and AmpC producing Enterobacteriaceae in companion animals. *Veterinary Microbiology*, 170(1-2), 10-18. doi: 10.1016/j.vetmic.2014.01.017
- Rzewuska, M., Czopowicz, M. B., U, M. K.-, Chrobak, D., Borys, B. B., & Binek, M. (2015). Multidrug resistance in *Escherichia coli* strains isolated from infections in dogs and cats in Poland ( 2007 – 2013 ). *The Scientific World Journal*, 2015, 1-8. doi: 10.1155/2015/408205
- Sáenz, Y., Zarazaga, M., Briás, L., Lantero, M., Ruiz-Larrea, F., & Torres, C. (2001). Antibiotic resistance in *Escherichia coli* isolates obtained from animals, foods and humans in Spain. *International Journal of Antimicrobial Agents*, 18(4), 353-358. doi: 10.1016/S0924-8579(01)00422-8
- Saidi, B., Mafirakureva, P., & Mbanga, J. (2013). Antimicrobial resistance of *Escherichia coli* isolated from chickens with colibacillosis in and around Harare, Zimbabwe. *Avian Diseases*, 57(1), 152-154. doi: 10.1637/10325-081512-case.1
- Saidi, R., Mimoune, N., Baazizi, R., Benaissa, M. H., Khelef, D., & Kaidi, R. (2019). Antibiotic susceptibility of *Staphylococci* isolated from bovine mastitis in Algeria. *Journal of Advanced Veterinary and Animal Research*, 6(2), 231. doi: 10.5455/javar.2019.f337

- Santajit, S., & Indrawattana, N. (2016). Mechanisms of antimicrobial resistance in ESKAPE pathogens. *BioMed Research International*, 2016, 1-16. doi: 10.1155/2016/2475067
- Saputra, S., Jordan, D., Mitchell, T., Wong, H. S., Abraham, R. J., Kidsley, A., Turnidge, J., Trott, D. J., & Abraham, S. (2017). Antimicrobial resistance in clinical *Escherichia coli* isolated from companion animals in Australia. *Veterinary Microbiology*, 211, 43-50. doi: 10.1016/j.vetmic.2017.09.014
- Saputra, S., Jordan, D., Worthing, K. A., Norris, J. M., Wong, H. S., Abraham, R., Trott, D. J., & Abraham, S. (2017). Antimicrobial resistance in coagulase-positive *Staphylococci* isolated from companion animals in Australia: A one year study. *PloS one*, 12(4), e0176379. doi: 10.1371/journal.pone.0176379
- Sárközy, G. (2001). Quinolones: A class of antimicrobial agents. *Veterinarni Medicina*, 46(9/10), 257-274. doi: 10.17221/7883-vetmed
- Sayah, R. S., Kaneene, J. B., Johnson, Y., & Miller, R. A. (2005). Patterns of antimicrobial resistance observed in *Escherichia coli* isolates obtained from domestic- and wild-animal fecal samples, human sepsis, and surface water. *Applied and Environmental Microbiology*, 71(3), 1394-1404. doi: 10.1128/AEM.71.3.1394-1404.2005
- Scallan, E., Griffin, P. M., Angulo, F. J., Tauxe, R. V., & Hoekstra, R. M. (2011). Foodborne illness acquired in the United States—unspecified agents. *Emerging Infectious Diseases*, 17(1), 7. doi: 10.3201/eid1701.P21101
- Schmithausen, R. M., Schulze-Geisthoevel, S. V., Heinemann, C., Bierbaum, G., Exner, M., Petersen, B., & Steinhoff-Wagner, J. (2018). Reservoirs and transmission pathways of resistant indicator bacteria in the biotope pig stable and along the food chain: A review from a one health perspective. *Sustainability (Switzerland)*, 10(11), 3967. doi: 10.3390/su10113967
- Schmithausen, R. M., Schulze-Geisthoevel, S. V., Stemmer, F., El-Jade, M., Reif, M., Hack, S., Meilaender, A., Montabauer, G., Fimmers, R., Parcina, M., Hoerauf, A., Exner, M., Petersen, B., Bierbaum, G., Bekeredjian-Ding, I., & Smith, T. C. (2015). Analysis of transmission of MRSA and ESBL-E among pigs and farm personnel. *PloS one*, 10(9), e0138173. doi: 10.1371/journal.pone.0138173
- Schulz, J., Kemper, N., Hartung, J., Janusch, F., Mohring, S. A. I., & Hamscher, G. (2019). Analysis of fluoroquinolones in dusts from intensive livestock farming and the co-occurrence of fluoroquinolone-resistant *Escherichia coli*. *Scientific Reports*, 9(1), 1-7. doi: 10.1038/s41598-019-41528-z

- Schwarz, S., Loeffler, A., & Kadlec, K. (2017). Bacterial resistance to antimicrobial agents and its impact on veterinary and human medicine. *Advances in Veterinary Dermatology*, 8, 95-110. doi: 10.1111/vde.12362
- Scott, A. M., Beller, E., Glasziou, P., Clark, J., Ranakusuma, R. W., Byambasuren, O., Bakhit, M., Page, S. W., Trott, D., & Mar, C. Del. (2018). Is antimicrobial administration to food animals a direct threat to human health? A rapid systematic review. *International Journal of Antimicrobial Agents*, 52(3), 316-323. doi: 10.1016/j.ijantimicag.2018.04.005
- Seepersadsingh, N., Adesiyun, A. A., & Seebaransingh, R. (2004). Prevalence and antimicrobial resistance of *Salmonella* spp. in non-diarrhoeic dogs in Trinidad. *Journal of Veterinary Medicine Series B: Infectious Diseases and Veterinary Public Health*, 51(7), 337-342. doi: 10.1111/j.1439-0450.2004.00785.x
- Sefton, M. A. (2002). Mechanisms of antimicrobial resistance: Their clinical relevance in the new millennium. *Drugs*, 62, 557–566. doi: 10.2165/00003495-200262040-00001
- Shahaza, O., Mohd. Azizul, O., Zakirah, S., Muhammad Azim, F. A., Syamsyul, A., & Maswati, M. A. (2017). Antimicrobial resistance in veterinary clinical isolates of *Escherichia coli* from northern region of Peninsular Malaysia. *Malaysian Journal of Veterinary Research*, 8(2), 1-7. Retrieved from <https://www.cabdirect.org/cabdirect/abstract/20183247760>
- Shaikh, S., Fatima, J., Shakil, S., Rizvi, S. M. D., & Kamal, M. A. (2015). Antibiotic resistance and extended spectrum beta-lactamases: Types, epidemiology and treatment. *Saudi Journal of Biological Sciences*, 22(1): 90–101. doi: 10.1016/j.sjbs.2014.08.002
- Sharma, A., & Grover, P. S. (2004). Application of whonet for the surveillance of antimicrobial resistance. *Indian Journal of Medical Microbiology*, 22(2), 115-118. <https://pubmed.ncbi.nlm.nih.gov/17642708/>.
- Sharma, C., Rokana, N., Chandra, M., Singh, B. P., Gulhane, R. D., Gill, J. P. S., Ray, P., Puniya, A. K., & Panwar, H. (2018). Antimicrobial resistance: Its surveillance, impact, and alternative management strategies in dairy animals. *Frontiers in Veterinary Science*, 4, 237. doi: 10.3389/fvets.2017.00237
- Sheldon, J. (2001). Concise Oxford textbook of medicine.: JGG Ledingham and David A Warrell (eds). Oxford University Press, 2000. *Family Practice*, 2056. doi: 10.1093/fampra/ 18.3.345

- Singh, F., Sonawane, G. G., Kumar, J., Dixit, S. K., Meena, R. K., & Tripathi, B. N. (2019). Antimicrobial resistance and phenotypic and molecular detection of extended-spectrum  $\beta$ -lactamases among extraintestinal *Escherichia coli* isolated from pneumonic and septicemic sheep and goats in Rajasthan, India. *Turkish Journal of Veterinary and Animal Sciences*, 43(6), 754-760. doi: 10.3906/vet-1905-1
- Smith, M. C., & Sherman, D. M. (2009). Goat Medicine. *Goat Medicine: Second Edition*. John Wiley and Sons. 871. doi: 10.1002/9780813818825
- Smith, M., King, C., Davis, M., Dickson, A., Park, J., Smith, F., Currie, K., & Flowers, P. (2018). Pet owner and vet interactions: exploring the drivers of AMR. *Antimicrobial Resistance & Infection Control*, 7(1), 46. doi: 10.1186/s13756-018-0341-1
- Srisanga, S., Angkititrakul, S., Sringsam, P., Le Ho, P. T., Vo, A. T. T., & Chuanchuen, R. (2017). Phenotypic and genotypic antimicrobial resistance and virulence genes of *Salmonella enterica* isolated from pet dogs and cats. *Journal of Veterinary Science*, 18(3), 273. doi: 10.4142/jvs.2017.18.3.273
- Srivastava, M. K. (2010). Antibiotic growth-promoters in food animals. *Pharma Times*, 42(3):17-21. Retrieved from [https://www.researchgate.net/publication/288584281\\_Antibiotic\\_growth-promoters\\_in\\_food\\_animals](https://www.researchgate.net/publication/288584281_Antibiotic_growth-promoters_in_food_animals)
- Stelling, J. M., & O'Brien, T. F. (1997). Surveillance of antimicrobial resistance: the whonet program. *Clinical Infectious Diseases*, 24(1), 157-168. doi: 10.1093/clinids/24.supplement\_1.s157
- Subbiah, M., Caudell, M. A., Mair, C., Davis, M. A., Matthews, L., Quinlan, R. J., Quinlan, M. B., Lyimo, B., Buza, J., Keyyu, J., & Call, D. R. (2020). Antimicrobial resistant enteric bacteria are widely distributed amongst people, animals and the environment in Tanzania. *Nature Communications*, 11(1), 1-12. doi: 10.1038/s41467-019-13995-5
- Suriyasathaporn, W., Chupia, V., Sing-Lah, T., Wongsawan, K., Mektrirat, R., & Chaisri, W. (2012). Increases of antibiotic resistance in excessive use of antibiotics in smallholder dairy farms in Northern Thailand. *Asian-Australasian Journal of Animal Sciences*, 25(9), 1322. doi: 10.5713/ajas.2012.12023
- Swanson, S. J., Snider, C., Braden, C. R., Boxrud, D., Wünschmann, A., Rudroff, J. A., Lockett, J., & Smith, K. E. (2007). Multidrug-resistant *Salmonella enterica* serotype *typhimurium* associated with pet rodents. *New England Journal of Medicine*, 356(1), 21-28. doi: 10.1056/NEJMoa060465
- Sykes, J. E. (2013). Antimicrobial drug use in dogs and cats. *Antimicrobial Therapy in Veterinary Medicine*, 5, 1-683. doi: 10.1002/9781118675014. ch28

- Szmolka, A., & Nagy, B. (2013). Multidrug resistant commensal *Escherichia coli* in animals and its impact for public health. *Frontiers in Microbiology*, 4, 258. doi: 10.3389/fmicb.2013.00258
- Tacconelli, E., & Pezzani, M. D. (2019). Public health burden of antimicrobial resistance in Europe. *The Lancet Infectious Diseases*, 19(1), 4-6. doi: 10.1016/S1473-3099(18)30648-0
- Tadesse, D. A., Zhao, S., Tong, E., Ayers, S., Singh, A., Bartholomew, M. J., & McDermott, P. F. (2012). Antimicrobial drug resistance in *Escherichia coli* from humans and food animals, United States, 1950-2002. *Emerging Infectious Diseases*, 18(5), 741. doi: 10.3201/eid1805.111153
- Teichmann-Knorrn, S., Reese, S., Wolf, G., Hartmann, K., & Dorsch, R. (2018). Prevalence of feline urinary tract pathogens and antimicrobial resistance over five years. *Veterinary Record*, 183(1), 21-21. doi: 10.1136/vr.104440
- Teillant, A., & Laxminarayan, R. (2015). Economics of antibiotic use in US swine and poultry production. *Choices*, 30(1), 1-11. Retrieved from <http://www.jstor.org/stable/choices.30.1.06>.
- Teuber, M., & Perreten, V. (2000). Role of milk and meat products as vehicles for antibiotic-resistant bacteria. *Acta veterinaria Scandinavica. Supplementum*, (93), 75-87. Retrieved from <https://www.cabdirect.org/cabdirect/abstract/20000404445>
- Thaller, M. C., Migliore, L., Marquez, C., Tapia, W., Cedeño, V., Rossolini, G. M., & Gentile, G. (2010). Tracking acquired antibiotic resistance in commensal bacteria of Galápagos land iguanas: No man, no resistance. *PLoS one*, 5(2), e8989. doi: 10.1371/journal.pone.0008989
- The European Union summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in 2017. (2018). *EFSA Journal*, 16(12), e05500. doi: 10.2903/j.efsa.2018.5500
- Thung, T. Y., Radu, S., Mahyudin, N. A., Rukayadi, Y., Zakaria, Z., Mazlan, N., Tan, B. H., Lee, E., Yeoh, S. L., Chin, Y. Z., Tan, C. W., Kuan, C. H., Basri, D. F., & Wan Mohamed Radzi, C. W. J. (2018). Prevalence, virulence genes and antimicrobial resistance profiles of *Salmonella* serovars from retail beef in Selangor, Malaysia. *Frontiers in Microbiology*, 8, 2697. doi: 10.3389/fmicb.2017.02697
- Tsai, H. J., Huang, H. C., Lin, C. M., Lien, Y. Y., & Chou, C. H. (2007). *Salmonella* and *Campylobacter* in household and stray dogs in Northern Taiwan. *Veterinary Research Communications*, 31(8), 931-939. doi: 10.1007/s11259-007-0009-4

- Turutoglu, H., Ercelik, S., & Ozturk, D. (2006). Antibiotic resistance of *Staphylococcus aureus* and coagulase-negative *Staphylococci* isolated from bovine mastitis. *Bulletin of the Veterinary Institute in Pulawy*, 50(1), 41-45. Retrieved from [https://www.researchgate.net/profile/Dilek-Ozturk-3/publication/221670171\\_Antibiotic\\_resistance\\_of\\_Staphylococcus\\_aureus\\_and\\_coagulase-negative\\_staphylococci\\_isolated\\_from\\_bovine\\_mastitis/links/0fcfd50584cabc7d91000000/Antibiotic-resistance-of-Staphylococcus-aureus-and-coagulase-negative-staphylococci-isolated-from-bovine-mastitis.pdf](https://www.researchgate.net/profile/Dilek-Ozturk-3/publication/221670171_Antibiotic_resistance_of_Staphylococcus_aureus_and_coagulase-negative_staphylococci_isolated_from_bovine_mastitis/links/0fcfd50584cabc7d91000000/Antibiotic-resistance-of-Staphylococcus-aureus-and-coagulase-negative-staphylococci-isolated-from-bovine-mastitis.pdf)
- Tyrrell, C., Burgess, C. M., Brennan, F. P., & Walsh, F. (2019). Antibiotic resistance in grass and soil. *Biochemical Society Transactions*, 47(1), 477–486. doi: 10.1042/bst20180552
- Tyson, G. H., Nyirabahizi, E., Crahey, E., Kabera, C., Lam, C., Rice-Trujillo, C., McDermott, P. F., & Tate, H. (2018). Prevalence and antimicrobial resistance of *Enterococci* isolated from retail meats in the United States, 2002 to 2014. *Applied and Environmental Microbiology*, 84(1), 1-9. doi: 10.1128/AEM.01902-17
- Ullah, H., & Ali, S. (2017). Classification of anti-bacterial agents and their functions. *Antibacterial Agents*, 10, 1-108. doi: 10.5772/intechopen.68695
- United States Department of Agriculture. (2010). Dairy 2007 - Heifer calf health and Management practices on U.S. Operations. USDA:APHIS:VS,CEAH. Fort Collins, CO. Retrieved from [https://www.aphis.usda.gov/animal\\_health/nahms/dairy/downloads/dairy07/Dairy07\\_ir\\_CalfHealth\\_1.pdf](https://www.aphis.usda.gov/animal_health/nahms/dairy/downloads/dairy07/Dairy07_ir_CalfHealth_1.pdf)
- Ünüvar, S. (2018). Microbial foodborne diseases. *Foodborne Diseases*, 1-31. doi: 10.1016/B978-0-12-811444-5.00001-4
- US Food and Drug Administration. (2012). The judicious use of medically important antimicrobial drugs in food-producing animals. *Federal Register*, 1-26. Retrieved from <https://www.fda.gov/media/79140/download>
- Van Boeckel, T., & Laxminarayan, R. (2017). Correction to global antibiotic consumption data. *The Lancet Infectious Diseases*, 17(5), 476-477. doi: 10.1016/S1473-3099(17)30187-1
- Van Boeckel, T. P., Brower, C., Gilbert, M., Grenfell, B. T., Levin, S. A., Robinson, T. P., Teillant, A., & Laxminarayan, R. (2015). Global trends in antimicrobial use in food animals. *Proceedings of the National Academy of Sciences of the United States of America*, 112(18), 5649-5654. doi: 10.1073/pnas.1503141112

- Van Den Bogaard, A. E., & Stobberingh, E. E. (2000). Epidemiology of resistance to antibiotics: Links between animals and humans. *International Journal of Antimicrobial Agents*, 14(4), 327-335. doi: 10.1016/S0924-8579(00)00145-X
- Van den Bunt, G., Top, J., Hordijk, J., De Greeff, S. C., Mughini-Gras, L., Corander, J., van Pelt, W., Bonten, M. J. M., Fluit, A. C., & Willems, R. J. L. (2018). Intestinal carriage of ampicillin- and vancomycin-resistant *Enterococcus faecium* in humans, dogs and cats in the Netherlands. *Journal of Antimicrobial Chemotherapy*, 73(3), 607-614. doi: 10.1093/jac/dkx455
- Van Duijkeren, E., Kamphuis, M., van der Mije, I. C., Laarhoven, L. M., Duim, B., Wagenaar, J. A., & Houwers, D. J. (2011). Transmission of methicillin-resistant *Staphylococcus pseudintermedius* between infected dogs and cats and contact pets, humans and the environment in households and veterinary clinics. *Veterinary Microbiology*, 150(3-4), 338-343. doi: 10.1016/j.vetmic.2011.02.012
- Van, T. T. H., Nguyen, H. N. K., Smooker, P. M., & Coloe, P. J. (2012). The antibiotic resistance characteristics of non-typhoidal *Salmonella enterica* isolated from food-producing animals, retail meat and humans in South East Asia. *International Journal of Food Microbiology*, 154(3), 98-106. doi: 10.1016/j.ijfoodmicro.2011.12.032
- Van, T. T. H., Yidana, Z., Smooker, P. M., & Coloe, P. J. (2020). Antibiotic use in food animals worldwide, with a focus on Africa: Pluses and minuses. *Journal of Global Antimicrobial Resistance*, 20, 170-177. doi: 10.1016/j.jgar.2019.07.031
- Ventola, C. L. (2015). The antibiotic resistance crisis: causes and threats. *Pharmacy and Therapeutics*, 40(4), 277-283. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4378521/>
- Vidal, A., Baldomà, L., Molina-López, R. A., Martin, M., & Darwich, L. (2017). Microbiological diagnosis and antimicrobial sensitivity profiles in diseased free-living raptors. *Avian Pathology*, 46(4), 442-450. doi: 10.1080/03079457.2017.1304529
- Videla, R., Solyman, S. M., Brahmbhatt, A., Sadeghi, L., Bemis, D. A., & Kania, S. A. (2018). Clonal complexes and antimicrobial susceptibility profiles of *Staphylococcus pseudintermedius* isolates from dogs in the United States. *Microbial Drug Resistance*, 24(1), 83-88. doi: 10.1089/mdr.2016.0250
- Vidovic, N., & Vidovic, S. (2020). Antimicrobial resistance and food animals: Influence of livestock environment on the emergence and dissemination of antimicrobial resistance. *Antibiotics*, 9(2), 52. doi: 10.3390/antibiotics9020052

- Vignaroli, C., Zandri, G., Aquilanti, L., Pasquaroli, S., & Biavasco, F. (2011). Multidrug-resistant *Enterococci* in animal meat and faeces and co-transfer of resistance from an *Enterococcus durans* to a human *Enterococcus faecium*. *Current Microbiology*, 62(5), 1438-1447. doi: 10.1007/s00284-011-9880-x
- Vigo, G. B., Leotta, G. A., Caffer, M. I., Salve, A., Binsztein, N., & Pichel, M. (2011). Isolation and characterization of *Salmonella enterica* from Antarctic wildlife. *Polar Biology*, 34(5), 675-681. doi: 10.1007/s00300-010-0923-8
- Walther, B., Hermes, J., Cuny, C., Wieler, L. H., Vincze, S., Elnaga, Y. A., Stamm, I., Kopp, P. A., Kohn, B., Witte, W., Jansen, A., Conraths, F. J., Semmler, T., Eckmanns, T., & Lübke-Becker, A. (2012). Sharing more than friendship - nasal colonization with coagulase-positive *Staphylococci* (CPS) and co-habitation aspects of dogs and their owners. *PloS one*, 7(4), e35197. doi: 10.1371/journal.pone.0035197
- Wang, J., Ma, Z. B., Zeng, Z. L., Yang, X. W., Huang, Y., & Liu, J. H. (2017). The role of wildlife (wild birds) in the global transmission of antimicrobial resistance genes. *Zoological research*, 38(2), 55-80. doi: 10.24272/j.issn.2095-8137.2017.003
- Wedley, A. L., Westgarth, C., Pinchbeck, G. L., Williams, N. J., Dawson, S., Coyne, K. P., & Maddox, T. W. (2011). Prevalence of antimicrobial-resistant *Escherichia coli* in dogs in a cross-sectional, community-based study. *Veterinary Record*, 168(13), 354-354. doi: 10.1136/vr.d1540
- Weese, J. S., Giguère, S., Guardabassi, L., Morley, P. S., Papich, M., Ricciuto, D. R., & Sykes, J. E. (2015). ACVIM consensus statement on therapeutic antimicrobial use in animals and antimicrobial resistance. *Journal of Veterinary Internal Medicine*, 29(2), 487-498. doi: 10.1111/jvim.12562
- Wegener, H. (2012). Antibiotic resistance-Linking human and animal health. *Improving Food Safety Through a One Health Approach: Workshop Summary*, 331. Retrieved from <https://www.ncbi.nlm.nih.gov/books/NBK114485/>
- Wegener, H. C. (2003). Antibiotics in animal feed and their role in resistance development. *Current Opinion in Microbiology*, 6(5), 439-445. doi: 10.1016/j.mib.2003.09.009
- Weiss, D., Wallace, R. M., Rwego, I. B., Gillespie, T. R., Chapman, C. A., Singer, R. S., & Goldberg, T. L. (2018). Antibiotic-resistant *Escherichia coli* and class 1 integrons in humans, domestic animals, and wild primates in rural Uganda. *Applied and Environmental Microbiology*, 84(21), 1-10. doi: 10.1128/AEM.01632-18
- Wichmann, F., Udikovic-Kolic, N., Andrew, S., & Handelsman, J. (2014). Diverse antibiotic resistance genes in dairy cow manure. *MBio*, 5(2), e01017-01013. doi: 10.1128/mBio.01017-13

- Wilkinson, J., & Boxall, A. (2019). The first global study of pharmaceutical contamination in riverine environments. *SETAC Europe 29th Annual Meeting, Helsinki, Finland*, 1-432. Retrieved from [https://cdn.ymaws.com/www.setac.org/resource/resmgr/abstract\\_books/setac-helsinki-abstract-book.pdf](https://cdn.ymaws.com/www.setac.org/resource/resmgr/abstract_books/setac-helsinki-abstract-book.pdf)
- Wong, C., Epstein, S. E., & Westropp, J. L. (2015). Antimicrobial susceptibility patterns in urinary tract infections in dogs (2010-2013). *Journal of Veterinary Internal Medicine*, 29(4), 1045-1052. doi: 10.1111/jvim.13571
- World Health Organization. (2014). Antimicrobial resistance. Global report on surveillance. doi: 10.1007/s13312-014-0374-3
- World Health Organization. (2017a). Critically important antimicrobials for human medicine: 5th revision. doi: 10.1017/CBO9781107415324.004
- World Health Organization. (2017b). Integrated surveillance of antimicrobial resistance in foodborne bacteria: application of a one health approach. Retrieved from <https://apps.who.int/iris/bitstream/handle/10665/255747/9789241512411-eng.pdf?sequence=1>
- World Health Organization. (2006a). WHONET tutorial data analysis 1 for the surveillance of antimicrobial resistance. Retrieved from <https://whonet.org/Docs/WHONET%204.Data%20analysis%201.doc>
- World Health Organization. (2006b). WHONET tutorial data analysis 2 for the surveillance of antimicrobial resistance. Retrieved from <https://whonet.org/Docs/WHONET%205.Data%20analysis%202.doc>
- World Health Organization. (2017). WHO list of critically important antimicrobials for human medicine (WHO CIA list). Retrieved from <https://www.who.int/foodsafety/publications/WHO-CIA-list-6flyer-EN.pdf?ua=1>
- World Health Organization. (2015). *World health statistics 2015*. Retrieved from <https://www.who.int/docs/default-source/gho-documents/world-health-statistic-reports/world-health-statistics-2015.pdf>
- World Health Organization. (2018). GLASS | Global antimicrobial resistance surveillance system (GLASS) report. Retrieved from <https://www.who.int/glass/en/>
- World Health Organization. (2014). The evolving threat of antimicrobial resistance: Options for action. Retrieved from [https://apps.who.int/iris/bitstream/handle/10665/44812/9789241503181\\_eng.pdf?sequence=1](https://apps.who.int/iris/bitstream/handle/10665/44812/9789241503181_eng.pdf?sequence=1)
- World Health Organization. (2006). WHONET tutorial data analysis 1 for the surveillance of antimicrobial resistance. Retrieved from <https://whonet.org/Docs/WHONET%204.Data%20analysis%201.doc>

World Health Organization. (2006). WHONET tutorial data analysis 2 for the surveillance of antimicrobial resistance. Retrieved from <https://whonet.org/Docs/WHONET%205.Data%20analysis%202.doc>

Worthing, K. A., Brown, J., Gerber, L., Trott, D. J., Abraham, S., & Norris, J. M. (2018). Methicillin-resistant *Staphylococci* amongst veterinary personnel, personnel-owned pets, patients and the hospital environment of two small animal veterinary hospitals. *Veterinary Microbiology*, 223, 79-85. doi: 10.1016/j.vetmic.2018.07.021

Wu, F., Ying, Y., Yin, M., Jiang, Y., Wu, C., Qian, C., Chen, Q., Shen, K., Cheng, C., Zhu, L., Li, K., Xu, T., Bao, Q., & Lu, J. (2019). Molecular characterization of a multidrug-resistant *Klebsiella pneumoniae* strain R46 isolated from a rabbit. *International Journal of Genomics*, 2019, 1-12. doi: 10.1155 /2019/5459190

Wu, X., Angkititrakul, S., L. Richards, A., Pulsrikarn, C., Khaengair, S., Keosengthong, A., Siriwong, S., & Suksawat, F. (2020). Risk of antimicrobial resistant non-typhoidal *Salmonella* during asymptomatic infection passage between pet dogs and their human caregivers in Khon Kaen, Thailand. *Antibiotics*, 9(8), 477. doi: 10.3390/antibiotics9080477

Xiong, W., Sun, Y., & Zeng, Z. (2018). Antimicrobial use and antimicrobial resistance in food animals. *Environmental Science and Pollution Research*, 25(19), 18377–18384. doi: 10.1007/s11356-018-1852-2

Yamasaki, Y., Nomura, R., Nakano, K., Naka, S., Matsumoto-Nakano, M., Asai, F., & Ooshima, T. (2012). Distribution of periodontopathic bacterial species in dogs and their owners. *Archives of Oral Biology*, 57(9), 1183-1188. doi: 10.1016/j.archoralbio.2012.02.015

Yassin, A. K., Gong, J., Kelly, P., Lu, G., Guardabassi, L., Wei, L., Han, X., Qiu, H., Price, S., Cheng, D., & Wang, C. (2017). Antimicrobial resistance in clinical *Escherichia coli* isolates from poultry and livestock, China. *PLoS ONE*, 12(9), e0185326. doi: 10.1371/journal.pone.0185326

Yohannes, S., Awji, E. G., Lee, S. J., & Park, S. C. (2015). Pharmacokinetics and pharmacokinetic/pharmacodynamic integration of marbofloxacin after intravenous and intramuscular administration in beagle dogs. *Xenobiotica*, 45(3), 264-269. doi: 10.3109/00498254.2014.969794

Yong, M. K., Busing, K. L., Cheng, A. C., & Thrusky, K. A. (2010). Improved susceptibility of Gram-negative bacteria in an intensive care unit following implementation of a computerized antibiotic decision support system. *Journal of Antimicrobial Chemotherapy*, 65(5), 1062-1069. doi: 10.1093/jac/dkq058

Zakeri, A., & Kashefi, P. (2012). Antimicrobial susceptibilities of avian *Escherichia coli* isolates in Tabriz, Iran. *African Journal of Biotechnology*, 11(19), 4467-4470. doi: 10.5897/ajb11.3168

- Zhang, S., Yang, G., Ye, Q., Wu, Q., Zhang, J., & Huang, Y. (2018). Phenotypic and genotypic characterization of *Klebsiella pneumoniae* isolated from retail foods in China. *Frontiers in Microbiology*, 9, 289. doi: 10.3389/fmicb.2018.00289
- Zhao, S., McDermott, P. F., White, D. G., Qaiyumi, S., Friedman, S. L., Abbott, J. W., Glenn, A., Ayers, S. L., Post, K. W., Fales, W. H., Wilson, R. B., Reggiardo, C., & Walker, R. D. (2007). Characterization of multidrug resistant *Salmonella* recovered from diseased animals. *Veterinary Microbiology*, 123(1-3), 122-132. doi: 10.1016/j.vetmic.2007.03.001
- Zhu, Y. G., Johnson, T. A., Su, J. Q., Qiao, M., Guo, G. X., Stedtfeld, R. D., Hashsham, S. A., & Tiedje, J. M. (2013). Diverse and abundant antibiotic resistance genes in Chinese swine farms. *Proceedings of the National Academy of Sciences of the United States of America*, 110(9), 3435–3440. doi: 10.1073/pnas.1222743110