



**SEROPREVALENCE AND IDENTIFICATION OF PATHOGENIC
LEPTOSPIRA IN RODENTS FROM KUALA LUMPUR WET MARKETS,
MALAYSIA**

By

MOHAMED ASYRAF BIN NOH

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

March 2021

FPSK (m) 2021 50

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**SEROPREVALENCE AND IDENTIFICATION OF PATHOGENIC
LEPTOSPIRA IN RODENTS FROM KUALA LUMPUR WET MARKETS,
MALAYSIA**

By

MOHAMED ASYRAF BIN NOH

March 2021

Chair : Siti Norbaya binti Masri, MD, MPath
Faculty : Medicine and Health Sciences

Pathogenic *Leptospira* are spiral-shaped bacteria that causes leptospirosis in infected victim. It was reported to be isolated from many mammal species all over the world, including human, establishing leptospirosis as the most widespread zoonosis disease. This bacterium species is widely known for having rodents as their main reservoir host, residing in the host's renal tubule and spread to the environment through their host's urine. Malaysia has declared leptospirosis as notifiable disease in 2010 and there has been increasing reported cases annually since then. Due to this, there is a need for a complete list or database of pathogenic *Leptospira* that circulate in Malaysia. Therefore, this study is performed in hope to contribute and provide information towards that goal. The objective of this study is to determine the seroprevalence and species distribution of pathogenic *Leptospira* in rodents captured from Kuala Lumpur wet markets. Four wet markets in Kuala Lumpur were chosen as the sampling sites of this study, namely Pasar Pudu, Pasar Chow Kit, Pasar Datuk Keramat and Pasar Petaling Street, as per recommendation by Kuala Lumpur City Hall (Dewan Bandaraya Kuala Lumpur, DBKL). Sampling was performed by live trapping for the duration of 12 months in 2017. Each rodent caught was euthanised, morphologically identified and processed for the extraction of blood serum and kidney samples. Microscopic Agglutination Test (MAT) were performed on serum samples (against a panel of 24 live *Leptospira* serovar culture) for serology test, while kidney samples were processed and subjected to molecular screening test for pathogenic *Leptospira* detection and cultured in Ellinghausen-McCullough-Johnson-Harris (EMJH) medium for *Leptospira* isolation. For each culture medium that succeeded in growing *Leptospira*, they were morphologically confirmed as *Leptospira* cultures under dark field microscope and underwent molecular characterisation using six loci Multilocus Sequence Typing (MLST) test for *Leptospira* species identification. Sampling yields 144 live captured rodents, all identified as house rat or *Rattus rattus*. MAT gave 50/144 (34.72%) positive samples for leptospiral antibody in rodents, with

the most prominent serovar detected being serovar Malaya, followed by IMR LEP 175. A total of 50/144 (34.7%) samples were detected as pathogenic *Leptospira* from molecular screening test. As for bacteria isolation, 23 out of 144 culture samples (15.97%) were successfully isolated, and from this, MLST analysis identified two *Leptospira* species, with 20/23 (87%) samples identified as *Leptospira borpetersenii* and the other remaining three (13%) samples were identified as ST149 *Leptospira interrogans* serogroup Bataviae serovar Bataviae strain Swart. The findings in this study present the evidence of pathogenic *Leptospira* presence which strive and actively circulating in the rodent population of Kuala Lumpur wet markets. Rodents are indeed the reservoir or maintenance host of pathogenic *Leptospira* in the urban, highly populated setting of Kuala Lumpur. Both serology and molecular screening test results indicate a high endemicity for leptospirosis if the rodent population is left unchecked, as control of *Leptospira* reservoir host is important for preventing future leptospirosis outbreaks in Kuala Lumpur.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**KELAZIMAN SERO DAN PENGENALPASTIAN *LEPTOSPIRA* PATOGENIK
DI DALAM TIKUS DARI PASAR BASAH KUALA LUMPUR, MALAYSIA**

Oleh

MOHAMED ASYRAF BIN NOH

Mac 2021

Pengerusi : Siti Norbaya binti Masri, MD, MPath
Fakulti : Perubatan dan Sains Kesihatan

Leptospira patogenik ialah bakteria berbentuk lingkaran yang menyebabkan leptospirosis pada mangsa yang dijangkiti. *Leptospira* telah dilaporkan berjaya diasinkan daripada banyak spesies mamalia di seluruh dunia, termasuk manusia, menjadikan leptospirosis sebagai penyakit zoonosis yang paling meluas. Spesies bakteria ini terkenal dengan mempunyai tikus sebagai hos rizab utama mereka, tinggal di dalam tubul ginjal hos dan disebar ke persekitaran melalui air kencing hos mereka. Malaysia telah mengisytiharkan leptospirosis sebagai penyakit yang perlu dilaporkan pada tahun 2010 dan terdapat peningkatan kes yang dilaporkan setiap tahun sejak itu. Disebabkan ini, terdapat keperluan untuk senarai lengkap atau pangkalan data *Leptospira* patogenik yang berkitar di Malaysia. Jesteru itu, kajian ini dilakukan dengan harapan untuk menyumbang dan memberikan maklumat ke arah matlamat tersebut. Objektif kajian ini adalah untuk menentukan kelaziman sero dan taburan spesies *Leptospira* patogenik dalam tikus yang ditangkap dari pasar-pasar basah Kuala Lumpur. Empat pasar basah di Kuala Lumpur telah dipilih sebagai tapak persampelan kajian ini, iaitu Pasar Pudu, Pasar Chow Kit, Pasar Datuk Keramat dan Pasar Petaling Street, seperti yang disyorkan oleh Dewan Bandaraya Kuala Lumpur (DBKL). Persampelan dilakukan dengan cara tangkapan hidup selama 12 bulan pada tahun 2017. Setiap tikus yang ditangkap dieutanasi, dikenal pasti secara morfologi dan diproses untuk pengambilan serum darah dan sampel buah pinggang. Ujian Aglutinasi Mikroskopik (MAT) dilakukan pada sampel serum (terhadap panel 24 kultur hidup *Leptospira* serovar) untuk ujian serologi, manakala sampel ginjal diproses dan dikenakan ujian saringan molekul untuk mengesan *Leptospira* patogenik dan dikulturkan dalam Ellinghausen-McCullough-Johnson-Harris (EMJH) media untuk pengasingan *Leptospira*. Untuk setiap medium kultur yang berjaya membiakkkan *Leptospira*, mereka disahkan secara morfologi sebagai kultur *Leptospira* di bawah mikroskop medan gelap dan menjalani pencirian molekul menggunakan ujian enam lokus Penjenisan Jujukan Multi-lokus (MLST) untuk mengenal pasti spesies *Leptospira*. Persampelan menghasilkan 144 tikus hidup yang berjaya ditangkap,

semuanya dikenal pasti sebagai tikus rumah atau *Rattus rattus*. MAT memberikan 50/144 (34.72%) sampel positif untuk antibodi *leptospiral* dalam tikus, dengan serovar paling menonjol dikesan ialah serovar Malaya, diikuti oleh IMR LEP 175. Sebanyak 50/144 (34.7%) sampel dikesan sebagai *Leptospira* patogenik daripada ujian saringan molekul. Bagi pengasingan bakteria, 23 daripada 144 sampel kultur (15.97%) telah berjaya diasingkan, dan daripada ini, analisis MLST berjaya mengenal pasti dua spesies *Leptospira*, dengan 20/23 (87%) sampel dikenal pasti sebagai *Leptospira borpetersenii* dan baki tiga lagi (13%) sampel dikenal pasti sebagai ST149 *Leptospira interrogans* serogroup Bataviae serovar Bataviae strain Swart. Penemuan dalam kajian ini menunjukkan bukti kehadiran *Leptospira* patogenik yang berusaha dan berkitar aktif dalam populasi tikus di pasar-pasar basah Kuala Lumpur. Tikus sememangnya merupakan hos rizab atau perumah penyenggaraan *Leptospira* patogenik di kawasan bandar Kuala Lumpur yang padat dengan penduduk. Kedua-dua keputusan ujian saringan serologi dan molekul menunjukkan endemisiti tinggi untuk leptospirosis jika populasi tikus dibiarkan tanpa kawalan, kerana kawalan terhadap hos rizab *Leptospira* adalah penting untuk mencegah wabak leptospirosis pada masa hadapan di Kuala Lumpur.

ACKNOWLEDGEMENTS

This research is a part of major project titled Isolation and Characterisation of Pathogenic *Leptospira* from Human, Animals and Environment in Various Localities in Selangor and Wilayah Persekutuan NMRR-16-1000-30876 (IIR) by Malaysian Institute for Medical Research (IMR) with principal researcher, Dr. Fairuz Amran, funded by Malaysian Ministry of Health (MOH) MOH-NIH Research Grant (MRG).

Praise to Allah S.W.T the Almighty for giving me the chance and by guiding and granting me strength to finish this research.

I am taking this opportunity to express my greatest gratitude to my supervisor, Assoc. Prof. Dr. Siti Norbaya Masri who gave constant supports, advices, encouragements and toleration to me from the beginning until the completion of this study.

Huge acknowledgement to my internal co-supervisor Assoc. Prof. Dr. Mohd Nasir Mohd Desa for his suggestions and guidance throughout this study.

Special thanks to my external co-supervisor, Dr. Fairuz Amran for giving me the opportunity to take up a portion of her extensive research as the topic for my Master degree. Without the opportunity, her permission and her guidance from the start of this research, I would have never been able to take up further study let alone complete it.

My sincere thanks also go to all staffs and members of Bacteriology Unit of IMR and UPM Department of Medical Microbiology and Parasitology for their cooperation and support for the entirety of this study duration.

Finally, I would also like to thank all my family members and friends for their help, faith and support on me throughout the completion of this study.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Siti Norbaya binti Masri, MPath

Associate Professor (Medical)

Faculty of Medicine and Health Science

Universiti Putra Malaysia

(Chairman)

Mohd Nasir bin Mohd Desa, PhD

Associate Professor, Ts.

Faculty of Medicine and Health Science

Universiti Putra Malaysia

(Member)

Fairuz binti Amran, MPath

Head of Bacteriology (Retired)

Infectious Disease Research Centre

Institute for Medical Research

(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 12 August 2021

Declaration by the Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and the copyright of the thesis are fully-owned by Universiti Putra Malaysia, as stipulated in the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from the supervisor and the office of the Deputy Vice-Chancellor (Research and innovation) before the thesis is published in any written, printed or electronic form (including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials) as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld in accordance with the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: 12 August 2021

Name and Matric No.: Mohamed Asyraf bin Noh

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xiv
 CHAPTER	
1 INTRODUCTION	1
1.1 Introduction	1
1.2 Problem Statement	2
1.3 Objectives	2
1.3.1 General Objective	2
1.3.2 Specific Objectives	2
2 LITERATURE REVIEW	3
2.1 History of Leptospirosis	3
2.2 Pathogenic <i>Leptospira</i>	4
2.2.1 Classification and Nomenclature	4
2.2.2 <i>Leptospira</i> Characteristics	6
2.3 Rodents as <i>Leptospira</i> Carrier and Reservoir	8
2.4 Human and Animal Leptospirosis	10
2.4.1 Source of Infection and Transmission	10
2.4.2 Pathogenesis, Symptoms and Clinical Features	10
2.4.3 Diagnosis	11
2.4.3.1 Serological test	12
2.4.3.2 <i>Leptospira</i> Isolation	13
2.4.3.3 Molecular Detection and Identification	13
2.4.4 Disease Treatment and Control	14
2.4.5 Epidemiology and Disease Burden	15
3 MATERIALS AND METHODS	18
3.1 Study Design	18
3.1.1 Study Sites	18
3.1.2 Sampling Method	20
3.1.3 Ethical Approval	20
3.2 Sampling Protocols	20
3.2.1 Rodent Trapping	20
3.2.2 Rodent Species Identification	21
3.2.3 Sample Collection	22
3.3 Microscopic Agglutination Test	23
3.4 <i>Leptospira</i> Isolation	25
3.4.1 Culture Media	25

3.4.2	<i>Leptospira</i> Culture	25
3.5	Molecular Test	26
3.5.1	DNA Extraction and Polymerase Chain Reaction	26
3.5.2	Pathogenic <i>Leptospira</i> Screening	27
3.5.3	Multilocus Sequence Typing	28
4	RESULTS	31
4.1	Rodent Distribution and Species Identification	31
4.2	Seropositivity of Anti-Leptospiral Antibody in Rats	31
4.3	<i>Leptospira</i> Serovar Distribution	32
4.4	Pathogenic <i>Leptospira</i> Detection	34
4.4.1	Molecular Screening with Amplification of <i>flaB</i> Gene	34
4.4.2	Pathogenic <i>Leptospira</i> Isolation	34
4.5	Result Comparison of Positive Samples on MAT, PCR <i>flaB</i> Amplification and <i>Leptospira</i> Isolation	35
4.6	Pathogenic <i>Leptospira</i> Identification	36
4.6.1	Multilocus Sequence Typing	36
4.6.2	Genetic and Phylogenetic Analysis	37
5	DISCUSSION	40
5.1	Rodent Distribution	40
5.2	Leptospirosis in Rats	40
5.2.1	Seropositivity of Anti-leptospiral Antibody in Rats	40
5.2.2	<i>Leptospira</i> Serovar Distribution	41
5.3	Pathogenic <i>Leptospira</i> Detection and Isolation	42
5.4	Comparison of Positive Results on MAT, PCR, and Isolation Test	43
5.5	Pathogenic <i>Leptospira</i> Identification	44
5.6	Suggestions for Leptospirosis Control	46
6	CONCLUSION	47
6.1	Summary and Conclusion	47
6.2	Limitations and Recommendations for Future Research	48
REFERENCES/BIBLIOGRAPHY		50
APPENDICES		59
BIODATA OF STUDENT		68
LIST OF PUBLICATIONS		69

LIST OF TABLES

Table		
		Page
2.1	List of identified <i>Leptospira</i> species.	5
2.2	List of <i>Leptospira</i> Species and Serovar found in Malaysian Animals.	17
3.1	List of Sampling Sites and Their Approximate Size.	18
3.2	List of <i>Leptospira</i> Cultures Used in MAT	24
3.3	PCR Procedure and Parameter of <i>flaB</i> Amplification.	28
3.4	List of MLST3 Loci and Primers.	29
3.5	MLST3 PCR Procedure and Parameters.	29
4.1	Distribution of Captured Rodents.	31
4.2	Seropositivity of Anti-Leptospiral Antibody in Captured Rats.	32
4.3	<i>Leptospira</i> Serovar Distribution and Titer Reading of Positive MAT Samples.	32
4.4	Positive MAT Sample with Multiple Serovar Reaction.	33
4.5	Positive Results of MAT, PCR and Culture.	35
4.6	Sequence Analysis Based on <i>Leptospira</i> PubMLST Database (MLST3)	38
4.7	BLAST Analysis of SG1-SG5.	38

LIST OF FIGURES

Figure		Page
2.1	First Observation of <i>Leptospira</i> by Arthur Stimson.	3
2.2	Phylogenetic Tree of <i>Leptospiraceae</i> 16S rRNA Gene Sequences by Maximum Likelihood Method Based on Tamura-Nei Model Using MEGA5.	6
2.3	Scanning Electron Micrograph of <i>Leptospira interrogans</i> Strain RGA.	7
2.4	Visual Representation of <i>Leptospira</i> Morphology Observed Under Dark Field Microscope.	8
2.5	Illustration of <i>Rattus rattus</i> and <i>Rattus norvegicus</i> .	9
2.6	The Appropriate Diagnosis Methods of Leptospirosis Based on Time of Disease Onset.	11
3.1	Study Site Locations in Kuala Lumpur.	19
3.2	Grouped Captured Rodents After Trapping Session.	21
3.3	Morphological Identification of Captured Rodents.	22
3.4	Sample Collection from Captured Rodents.	23
3.5	<i>Leptospira</i> Culture Procedure.	25
3.6	DNA Extraction Procedure (Boiling Method).	26
4.1	Representative Gel Image of <i>flaB</i> Amplification in 1.5% Agarose Gel Electrophoresis.	34
4.2	Representative Gel Image of <i>LipL32</i> Amplification in 1.5% Agarose Gel Electrophoresis.	35
4.3	Representative Gel Image of All MLST3 6 Loci Amplification in 1.5% Agarose Gel Electrophoresis.	36
4.4	Trimmed Maximum Likelihood Tree of MLST3 <i>Leptospira</i> Genetic Analysis Using SeaView (version 5) Utilising GTR Model and 1000 Bootstrap Replicates.	39

LIST OF ABBREVIATIONS

°C	Celsius
BLAST	Basic Local Alignment Search Tool
bp	Base Pair
BSA	Bovine Serum Albumin
DBKL	Dewan Bandaraya Kuala Lumpur
DNA	Deoxyribonucleic Acid
ELISA	Enzyme-Linked Immunosorbent Assay
EMJH	Ellinghausen, Mccullough, Johnson, Harris Media
<i>flaB</i>	Flagellin B
ft ²	Square Feet
Ictero	Icterohaemorrhagiae
IMR	Institute for Medical Research
L.	<i>Leptospira</i>
M	Molar
m ²	Square Meter
MAT	Microscopic Agglutination Test
MgCl ²	Magnesium Chloride
min	Minute
ml	Millilitre
MLST	Multi Locus Sequence Typing
MLST1	MLST Scheme 1
MLST3	MLST Scheme 3
NCBI	National Centre of Biotechnology
NMRR	National Medical Research Register

PBS	Phosphate Buffered Saline
PCR	Polymerase Chain Reaction
pH	Potential Of Hydrogen
R.	<i>Rattus</i>
sec	Second
ST	Sequence Type
TE	Tris-EDTA
WHO	World Health Organisation
μl	Micrometer
μm	Micrometer
μM	Micromolar

CHAPTER 1

INTRODUCTION

1.1 Introduction

Leptospirosis is a disease caused by bacterial infection of pathogenic *Leptospira* species that occur in both human and animal. *Leptospira* has been isolated from almost all mammalian species around the world which make leptospirosis recognised as the most widespread zoonosis disease. This disease is endemic in tropical regions, having high incidence during rainy season and natural disasters such as flood or tsunami.

Leptospira are grouped into two; saprophytic (non-pathogenic) and pathogenic. Saprophytic *Leptospira* are natural, free-living bacteria that live in the environment and do not cause leptospirosis. Pathogenic *Leptospira*, that causes leptospirosis are mainly found in the renal tubules of its reservoir or carrier host which they are later secreted through the host's urine into the environment. Natural reservoirs and carriers of pathogenic *Leptospira* includes domestic and wild animals. Rodents are the most common and well-known reservoir host for *Leptospira* that are responsible for disease transmission in both urban and rural settings where they are abundant in number.

Human infection of *Leptospira* occurred through direct contact with the urine of infected carriers or through contact with contaminated environmental elements such as wet soil or stagnant water. *Leptospira* exposure in human usually happened through occupational or recreational activities in places where infected or reservoir host are plenty. In human, symptoms of leptospirosis are broad which include headache, myalgia, arthralgia, brain conjunctival suffusion and meningeal irritation. These symptoms are too broad and similar with other diseases such as dengue resulting in difficulties for quick and accurate diagnose. Owing to this reason, mortality rate of leptospirosis cases is 5–15% involving multisystem disease and organ failure (Costa et al., 2015). Combined with high global annual estimated severe leptospirosis cases (around 350 000-500 000 cases) and difficulties in quick and accurate diagnose, leptospirosis is a serious and worrisome disease that may threat human lives.

Since 2010, there are gradually increasing reported cases of human leptospirosis in Malaysia as leptospirosis was declared as a notifiable disease in Malaysia. A total of 1976 cases were reported in 2010 and the cases have quadrupled to 7806 in 2014 (Wahab, 2015). Due to these high annual cases, it is important to have a data on what kind or species of *Leptospira* that is the most commonly harboured by rodents in Malaysia especially in urban area where

rodents are abundant and have frequent interaction with human population in the area.

1.2 Problem Statement

This study was performed considering the lack of multiple information on pathogenic *Leptospira* in Malaysia. Until this day, study on detection of leptospiral antibody in Malaysian small mammals especially rodents are still very limited because most previous *Leptospira* studies mainly focused of on human leptospirosis. Data on circulating *Leptospira* serovars in rodents is important as it can be used as reference when dealing with both human and animal leptospirosis. In addition, details on detection of pathogenic *Leptospira* presence in highly populated urban setting such as Kuala Lumpur is lacking. Highly populated urban areas are at great risk for *Leptospira* infection. Hence, there is a need for source tracking of the bacteria for outbreak intervention. This study was also performed in order to obtain information or list on pathogenic *Leptospira* species that are thriving and prevailing in Malaysia. Current knowledge on this subject is insufficient since there are still only few reports on isolation and identification of local *Leptospira* strains. Information on local pathogenic *Leptospira* species is crucial since it can provide genetic variation data on local strains and the information can also be used to increase microscopic agglutination test (MAT) sensitivity by addition of local strains in MAT panel for diagnosis or serosurvey purpose.

1.3 Objectives

1.3.1 General Objective

To study the seroprevalence and distribution of pathogenic *Leptospira* in rodents captured from Kuala Lumpur wet markets.

1.3.2 Specific Objectives

1. To determine the seropositivity of anti-leptospiral antibody in rodents from Kuala Lumpur wet markets.
2. To isolate, characterise and identify pathogenic *Leptospira* in captured rodents.
3. To determine the most common pathogenic *Leptospira* species in rodents Kuala Lumpur wet markets.

REFERENCES

- Abdullah, F. (Producer). (2021, 9.6.2021). MDKS tawar RM3 seekor tikus hidup atau mati. *SelangorTV*.
- Abdullah, N. M., Mohammad, W., Shafei, M. N., Sukeri, S., Idris, Z., et al. (2019). Leptospirosis and its prevention: knowledge, attitude and practice of urban community in Selangor, Malaysia. *BMC Public Health*, 19(1), 628. doi:10.1186/s12889-019-6981-0.
- Academic Medical Center (AMC), L. R. C. (2020a). Serovar Bataviae, Bataviae Swart 1926. Retrieved from <https://leptospira.amc.nl/filters/serovar/bataviae/>
- Academic Medical Center (AMC), L. R. C. (2020b). Serovar Malaya, Malaya H 1955. Retrieved from <https://leptospira.amc.nl/filters/serovar/malaya/>
- Adler, B. (2015). History of Leptospirosis and Leptospira. In Adler, B. (Ed.), *Leptospira and Leptospirosis* (pp. 1-8). Heidelberg: Springer.
- Agudelo-Florez, P., Arango, J. C., Merizalde, E., Londono, A. F., Quiroz, V. H., et al. (2010). [Serological evidence of Leptospira spp circulation in naturally-exposed rats (*Rattusnorvegicus*) in a Colombian urban area]. *Rev Salud Publica (Bogota)*, 12(6), 990-999.
- Ahmed, A., Engelberts, M. F., Boer, K. R., Ahmed, N., & Hartskeerl, R. A. (2009). Development and validation of a real-time PCR for detection of pathogenic leptospira species in clinical materials. *PLoS One*, 4(9), e7093. doi:10.1371/journal.pone.0007093.
- Ahmed, A., Thaipadungpanit, J., Boonsilp, S., Wuthiekanun, V., Nalam, K., et al. (2011). Comparison of two multilocus sequence based genotyping schemes for Leptospira species. *PLoS Negl Trop Dis*, 5(11), e1374. doi:10.1371/journal.pntd.0001374.
- Ahmed, N., Devi, S. M., Valverde, M. d. I. A., Vijayachari, P., Machang'u, R. S., et al. (2006). Multilocus sequence typing method for identification and genotypic classification of pathogenic Leptospira species. *Ann Clin Microbiol Antimicrob*, 5, 28-28. doi:10.1186/1476-0711-5-28.
- Alashraf, A. R., Lau, S. F., Khor, K. H., Khairani-Bejo, S., Bahaman, A. R., et al. (2019). Serological Detection of Anti-Leptospira Antibodies in Shelter Cats in Malaysia. *Top Companion Anim Med*, 34, 10-13. doi:10.1053/j.tcam.2018.12.002.
- Alia, S. N., Joseph, N., Philip, N., Azhari, N. N., Garba, B., et al. (2019). Diagnostic accuracy of rapid diagnostic tests for the early detection of leptospirosis. *J Infect Public Health*, 12(2), 263-269. doi:10.1016/j.jiph.2018.10.137.

- Amran, F., Mohd Khalid, M. K., Mohamad, S., Mat Ripen, A., Ahmad, N., et al. (2016). Draft Genome Sequence of *Leptospira interrogans* Serovar Bataviae Strain LepIMR 22 Isolated from a Rodent in Johor, Malaysia. *Genome Announc*, 4(5). doi:10.1128/genomeA.00956-16.
- Azaman, A. (2018). Kempen kebersihan DBKL berjaya hapus 78,000 tikus. *Astro Awani*.
- Azhari, N. N., Ramli, S. N. A., Joseph, N., Philip, N., Mustapha, N. F., et al. (2018). Molecular characterization of pathogenic *Leptospira* sp. in small mammals captured from the human leptospirosis suspected areas of Selangor state, Malaysia. *Acta Trop*, 188, 68-77. doi:10.1016/j.actatropica.2018.08.020.
- Bahaman, A. R., Ibrahim, A. L., & Adam, H. (1987). Serological prevalence of leptospiral infection in domestic animals in West Malaysia. *Epidemiol Infect*, 99(2), 379-392. doi:10.1017/s0950268800067868.
- Bahaman, A. R., Ibrahim, A. L., Stallman, N. D., & Tinniswood, R. D. (1988). The bacteriological prevalence of leptospiral infection in cattle and buffaloes in West Malaysia. *Epidemiol Infect*, 100(2), 239-246. doi:10.1017/s0950268800067376.
- Benacer, D., Mohd Zain, S. N., Ahmed, A. A., Mohd Khalid, M. K. N., Hartskeerl, R. A., et al. (2016a). Predominance of the ST143 and ST50 *Leptospira* clones in the urban rat populations of Peninsular Malaysia. *J Med Microbiol*, 65(6), 574-577. doi:10.1099/jmm.0.000262.
- Benacer, D., Mohd Zain, S. N., Amran, F., Galloway, R. L., & Thong, K. L. (2013). Isolation and molecular characterization of *Leptospira interrogans* and *Leptospira borgpetersenii* isolates from the urban rat populations of Kuala Lumpur, Malaysia. *Am J Trop Med Hyg*, 88(4), 704-709. doi:10.4269/ajtmh.12-0662.
- Benacer, D., Mohd Zain, S. N., Sim, S. Z., Mohd Khalid, M. K., Galloway, R. L., et al. (2016b). Determination of *Leptospira borgpetersenii* serovar Javanica and *Leptospira interrogans* serovar Bataviae as the persistent *Leptospira* serovars circulating in the urban rat populations in Peninsular Malaysia. *Parasit Vectors*, 9, 117. doi:10.1186/s13071-016-1400-1.
- Blasdell, K. R., Morand, S., Perera, D., & Firth, C. (2019). Association of rodent-borne *Leptospira* spp. with urban environments in Malaysian Borneo. *PLoS Negl Trop Dis*, 13(2), e0007141. doi:10.1371/journal.pntd.0007141.
- Boey, K., Shiokawa, K., & Rajeev, S. (2019). Leptospira infection in rats: A literature review of global prevalence and distribution. *PLoS Negl Trop Dis*, 13(8), e0007499. doi:10.1371/journal.pntd.0007499.

- Bojiraj, M., Porteen, K., Gunaseelan, L., & Sureshkannan, S. (2017). Seroprevalence of Leptospirosis in Animals and Its Public Health Significance. *Int. Journal of Livestock Research*, 7(11). doi:10.5455/ijlr.20170812041853.
- Boonsilp, S., Thaipadungpanit, J., Amornchai, P., Wuthiekanun, V., Chierakul, W., et al. (2011). Molecular detection and speciation of pathogenic *Leptospira* spp. in blood from patients with culture-negative leptospirosis. *BMC Infect Dis*, 11, 338. doi:10.1186/1471-2334-11-338.
- Bourhy, P., Collet, L., Brisse, S., & Picardeau, M. (2014). *Leptospira mayottensis* sp. nov., a pathogenic species of the genus *Leptospira* isolated from humans. *Int J Syst Evol Microbiol*, 64(Pt 12), 4061-4067. doi:10.1099/ijs.0.066597-0.
- Brenner, D. J., Kaufmann, A. F., Sulzer, K. R., Steigerwalt, A. G., Rogers, F. C., et al. (1999). Further determination of DNA relatedness between serogroups and serovars in the family *Leptospiraceae* with a proposal for *Leptospira alexanderi* sp. nov. and four new *Leptospira* genomospecies. *Int J Syst Bacteriol*, 49 Pt 2, 839-858. doi:10.1099/00207713-49-2-839.
- Cameron, C. E. (2015). Leptospiral Structure, Physiology, and Metabolism. In Adler, B. (Ed.), *Leptospira and Leptospirosis* (pp. 21-41). Heidelberg: Springer.
- Cosson, J. F., Picardeau, M., Mielcarek, M., Tatard, C., Chaval, Y., et al. (2014). Epidemiology of leptospira transmitted by rodents in southeast Asia. *PLoS Negl Trop Dis*, 8(6), e2902. doi:10.1371/journal.pntd.0002902.
- Costa, F., Hagan, J. E., Calcagno, J., Kane, M., Torgerson, P., et al. (2015). Global Morbidity and Mortality of Leptospirosis: A Systematic Review. *PLoS Negl Trop Dis*, 9(9), e0003898. doi:10.1371/journal.pntd.0003898.
- Daud, A., Mohd Fuzy, N. M. H., Arshad, M. M., Kamarudin, S., Wan Mohammad, W. M. Z., et al. (2018a). Leptospirosis seropositivity and its serovars among cattle in Northeastern Malaysia. *Vet World*, 11(6), 840-844. doi:10.14202/vetworld.2018.840-844.
- Daud, A., Mohd Fuzy, N. M. H., Wan Mohammad, W. M. Z., Amran, F., Ismail, N., et al. (2018b). Leptospirosis and Workplace Environmental Risk Factors among Cattle Farmers in Northeastern Malaysia. *Int J Occup Environ Med*, 9(2), 88-96. doi:10.15171/ijoem.2018.1164.
- El Jalii, I. M., & Bahaman, A. R. (2004). A review of human leptospirosis in Malaysia. *Trop Biomed*, 21(2), 113-119.
- Ellis, W. A. (2015). *Animal Leptospirosis* (Vol. 387). Heidelberg: Springer.
- Everard, J. D., & Everard, C. O. R. (1993). Leptospirosis in the Caribbean. *Rev Med Microbiol* ;, 4, 114–122.

- Faine, S. (1982). Guidelines for the control of leptospirosis / edited by S. Faine. In Geneva: World Health Organization.
- Faine, S., & Stallman, N. D. (1982). Amended Descriptions of the Genus *Leptospira* Noguchi 1917 and the Species *L. interrogans* (Stimson 1907) Wenyon 1926 and *L. biflexa* (Wolbach and Binger 1914) Noguchi 1918. 32(4), 461-463. doi:<https://doi.org/10.1099/00207713-32-4-461>.
- Francis, C. M., & Barrett, P. (2008). *A field guide to the mammals of South-East Asia*. London: New Holland.
- Goh, S. H., Ismail, R., Lau, S. F., Megat Abdul Rani, P. A., Mohd Mohidin, T. B., et al. (2019). Risk Factors and Prediction of Leptospiral Seropositivity Among Dogs and Dog Handlers in Malaysia. *Int J Environ Res Public Health*, 16(9). doi:10.3390/ijerph16091499.
- Haake, D. A., & Levett, P. N. (2015). Leptospirosis in Humans. In Adler, B. (Ed.), *Leptospira and Leptospirosis* (pp. 65-97). Heidelberg: Springer.
- Hartskeerl, R. A., & Smythe, L. D. (2015). Leptospirosis in Humans. In Adler, B. (Ed.), *The Role of Leptospirosis Reference Laboratories* (pp. 273-288). Heidelberg: Springer.
- Himsworth, C. G., Jardine, C. M., Parsons, K. L., Feng, A. Y., & Patrick, D. M. (2014). The characteristics of wild rat (*Rattus spp.*) populations from an inner-city neighborhood with a focus on factors critical to the understanding of rat-associated zoonoses. *PLoS One*, 9(3), e91654. doi:10.1371/journal.pone.0091654.
- Ido, Y., Hoki, R., Ito, H., & Wani, H. (1917). The Rat As A Carrier Of Spirochaeta Icterohaemorrhagiae, The Causative Agent Of Weil's Disease (Spirochaetosis Icterohaemorrhagica). *J Exp Med*, 26(3), 341-353. doi:10.1084/jem.26.3.341.
- Inada, R., Ido, Y., Hoki, R., Kaneko, R., & Ito, H. (1916). The Etiology, Mode Of Infection, And Specific Therapy Of Weil's Disease (Spirochaetosis Icterohaemorrhagica). *J Exp Med*, 23(3), 377-402. doi:10.1084/jem.23.3.377.
- Kanagavel, M., Princy Margreat, A. A., Arunkumar, M., Prabhakaran, S. G., Shanmughapriya, S., et al. (2016). Multilocus sequence typing (MLST) of leptospiral strains isolated from two geographic locations of Tamil Nadu, India. *Infect Genet Evol*, 37, 123-128. doi:10.1016/j.meegid.2015.11.008.
- Kattan, G. A., Bahaman, A. R., Bejo, S. K., Zakaria, Z., & Garba, B. (2017). Serological and molecular prevalence of *Leptospira* infection in Rat populations in Kuala Lumpur. *Aust J Basic & Appl Sci*, 11(1), 62-72.

- Koay, T. K., Nirmal, S., Noitie, L., & Tan, E. (2004). An epidemiological investigation of an outbreak of leptospirosis associated with swimming, Beaufort, Sabah. *Med J Malaysia*, 59(4), 455-459.
- Koizumi, N., Miura, K., Sanai, Y., Takemura, T., Ung, T. T. H., et al. (2019). Molecular epidemiology of *Leptospira interrogans* in *Rattus norvegicus* in Hanoi, Vietnam. *Acta Trop*, 194, 204-208. doi:10.1016/j.actatropica.2019.02.008.
- Kudo, Y., Vansith, K., Rin, E., Uchida, K., Kodama, S., et al. (2018). Molecular Epidemiological Survey of Leptospira Infection of Wild Rodents in the Urban Settlement of Cambodia. *Vector Borne Zoonotic Dis*, 18(3), 144-150. doi:10.1089/vbz.2017.2198.
- Latifah, I., Abdul Halim, A., Rahmat, M. S., Nadia, M. F., Ubil, Z. E., et al. (2017). Isolation by culture and PCR identification of LipL32 gene of pathogenic *Leptospira* spp. in wild rats of Kuala Lumpur. *Malays J Pathol*, 39(2), 161-166.
- Levett, P. N. (2003). Usefulness of serologic analysis as a predictor of the infecting serovar in patients with severe leptospirosis. *Clin Infect Dis*, 36(4), 447-452. doi:10.1086/346208.
- Levett, P. N. (2015). Systematics of Leptospiraceae. In Adler, B. (Ed.), *Leptospira and Leptospirosis* (pp. 11-20). Heidelberg: Springer.
- Levett, P. N., Morey, R. E., Galloway, R. L., & Steigerwalt, A. G. (2006). *Leptospira broomii* sp. nov., isolated from humans with leptospirosis. *Int J Syst Evol Microbiol*, 56(Pt 3), 671-673. doi:10.1099/ijsm.0.63783-0.
- Livingeshan, K. (2013). Pasar Raja Bot Di Chow Kit Bakal Lalui Pembangunan Semula RM200 Juta. *mStar*.
- Loan, H. K., Van Cuong, N., Takhampunya, R., Kiet, B. T., Campbell, J., et al. (2015). How important are rats as vectors of leptospirosis in the Mekong Delta of Vietnam? *Vector Borne Zoonotic Dis*, 15(1), 56-64. doi:10.1089/vbz.2014.1613.
- Marshall, R. (1992). International committee on systematic bacteriology subcommittee on the taxonomy of *Leptospira*: minutes of the meetings, 13 and 15 September 1990, Osaka, Japan. *Int J Syst Bacteriol*(42), 330–334.
- Matthias, M. A., Ricaldi, J. N., Cespedes, M., Diaz, M. M., Galloway, R. L., et al. (2008). Human leptospirosis caused by a new, antigenically unique *Leptospira* associated with a *Rattus* species reservoir in the Peruvian Amazon. *PLoS Negl Trop Dis*, 2(4), e213. doi:10.1371/journal.pntd.0000213.

- Mohd Radi, M. F., Hashim, J. H., Jaafar, M. H., Hod, R., Ahmad, N., et al. (2018). Leptospirosis Outbreak After the 2014 Major Flooding Event in Kelantan, Malaysia: A Spatial-Temporal Analysis. *Am J Trop Med Hyg*, 98(5), 1281-1295. doi:10.4269/ajtmh.16-0922.
- Murray, C. K., Gray, M. R., Mende, K., Parker, T. M., Samir, A., et al. (2011). Use of patient-specific Leptospira isolates in the diagnosis of leptospirosis employing microscopic agglutination testing (MAT). *Trans R Soc Trop Med Hyg*, 105(4), 209-213. doi:10.1016/j.trstmh.2010.12.004.
- Musso, D., & La Scola, B. (2013). Laboratory diagnosis of leptospirosis: a challenge. *J Microbiol Immunol Infect*, 46(4), 245-252. doi:10.1016/j.jmii.2013.03.001.
- Natarajaseenivasan, K., Vedhagiri, K., Sivabalan, V., Prabagaran, S. G., Sukumar, S., et al. (2011). Seroprevalence of Leptospira borgpetersenii serovar javanica infection among dairy cattle, rats and humans in the Cauvery river valley of southern India. *Southeast Asian J Trop Med Public Health*, 42(3), 679-686.
- Natarajaseenivasan, K., Vijayachari, P., Sharma, S., Sugunan, A. P., Vedhagiri, K., et al. (2010). FlaB PCR-based identification of pathogenic leptospiral isolates. *J Microbiol Immunol Infect*, 43(1), 62-69. doi:10.1016/s1684-1182(10)60009-6.
- Neela, V. K., Azhari, N. N., Joseph, N., Mimie, N. P., Ramli, S. N. A., et al. (2019). An outbreak of leptospirosis among reserve military recruits, Hulu Perdik, Malaysia. *Eur J Clin Microbiol Infect Dis*, 38(3), 523-528. doi:10.1007/s10096-018-03450-6.
- Perolat, P., Chappel, R. J., Adler, B., Baranton, G., Bulach, D. M., et al. (1998). Leptospira fainei sp. nov., isolated from pigs in Australia. *Int J Syst Bacteriol*, 48 Pt 3, 851-858. doi:10.1099/00207713-48-3-851.
- Philip, N., Bahtiar Affendy, N., Ramli, S. N. A., Arif, M., Raja, P., et al. (2020). Leptospira interrogans and Leptospira kirschneri are the dominant Leptospira species causing human leptospirosis in Central Malaysia. *PLoS Negl Trop Dis*, 14(3), e0008197. doi:10.1371/journal.pntd.0008197.
- Picardeau, M. (2013). Diagnosis and epidemiology of leptospirosis. *Med Mal Infect*, 43(1), 1-9. doi:10.1016/j.medmal.2012.11.005.
- Puche, R., Ferrés, I., Caraballo, L., Rangel, Y., Picardeau, M., et al. (2018). Leptospira venezuelensis sp. nov., a new member of the intermediate group isolated from rodents, cattle and humans. *Int J Syst Evol Microbiol*, 68(2), 513-517. doi:10.1099/ijsem.0.002528.

- Pui, C. F., Bilung, L. M., Apun, K., & Su'ut, L. (2017). Diversity of Leptospira spp. in Rats and Environment from Urban Areas of Sarawak, Malaysia. *J Trop Med*, 2017, 3760674. doi:10.1155/2017/3760674.
- Rahman, M., Hairon, S. M., Hamat, R. A., Jamaluddin, T., Shafei, M. N., et al. (2018). Seroprevalence and distribution of leptospirosis serovars among wet market workers in northeastern, Malaysia: a cross sectional study. *BMC Infect Dis*, 18(1), 569. doi:10.1186/s12879-018-3470-5.
- Ramadass, P., Jarvis, B. D., Corner, R. J., Penny, D., & Marshall, R. B. (1992). Genetic characterization of pathogenic Leptospira species by DNA hybridization. *Int J Syst Bacteriol*, 42(2), 215-219. doi:10.1099/00207713-42-2-215.
- Rao, M., Amran, F., Kamaruzaman, A. A., Hakim Esa, H. A., Abdul Hameed, A., et al. (2021). Case Report: Fatal Human Leptospirosis Caused by Leptospira interrogans Genotype ST149. *Am J Trop Med Hyg*, 104(1), 216-218. doi:10.4269/ajtmh.20-0267.
- Saito, M., Villanueva, S. Y., Chakraborty, A., Miyahara, S., Segawa, T., et al. (2013). Comparative analysis of Leptospira strains isolated from environmental soil and water in the Philippines and Japan. *Appl Environ Microbiol*, 79(2), 601-609. doi:10.1128/aem.02728-12.
- Samsudin, S., Masri, S. N., Tengku Jamaluddin, T. Z. M., Saudi, S. N. S., Md Ariffin, U. K., et al. (2015). Seroprevalence of Leptospiral Antibodies among Healthy Municipal Service Workers in Selangor. *Advances in Public Health*, 2015, 208145. doi:10.1155/2015/208145.
- Samsudin, S., Sakinah, S. N. S., Malina, O., Norliza, B. A., Noh, M. A., et al. (2018). Seroprevalence of leptospiral antibodies among market workers and food handlers in the central state of Malaysia. *Trop Med Int Health*, 23(3), 327-333. doi:10.1111/tmi.13033.
- Sapian, M., Khair, M. T., How, S. H., Rajalingam, R., Sahhir, K., et al. (2012). Outbreak of melioidosis and leptospirosis co-infection following a rescue operation. *Med J Malaysia*, 67(3), 293-297.
- Sejvar, J., Bancroft, E., Winthrop, K., Bettinger, J., Bajani, M., et al. (2003). Leptospirosis in "Eco-Challenge" Athletes, Malaysian Borneo, 2000. *Emerging Infectious Disease journal*, 9(6), 702. doi:10.3201/eid0906.020751.
- Slack, A. T., Kalambaheti, T., Symonds, M. L., Dohnt, M. F., Galloway, R. L., et al. (2008). Leptospira wolffii sp. nov., isolated from a human with suspected leptospirosis in Thailand. *Int J Syst Evol Microbiol*, 58(Pt 10), 2305-2308. doi:10.1099/ijsm.0.64947-0.

- Slack, A. T., Khairani-Bejo, S., Symonds, M. L., Dohnt, M. F., Galloway, R. L., et al. (2009). Leptospira kmetyi sp. nov., isolated from an environmental source in Malaysia. *Int J Syst Evol Microbiol*, 59(Pt 4), 705-708. doi:10.1099/ijss.0.002766-0.
- Smythe, L., Adler, B., Hartskeerl, R. A., Galloway, R. L., Turenne, C. Y., et al. (2013). Classification of Leptospira genomospecies 1, 3, 4 and 5 as Leptospira alstonii sp. nov., Leptospira vanthielii sp. nov., Leptospira terpstrae sp. nov. and Leptospira yanagawae sp. nov., respectively. *Int J Syst Evol Microbiol*, 63(Pt 5), 1859-1862. doi:10.1099/ijss.0.047324-0.
- Smythe, L. D., Wuthiekanun, V., Chierakul, W., Suputtamongkol, Y., Tiengrim, S., et al. (2009). The microscopic agglutination test (MAT) is an unreliable predictor of infecting Leptospira serovar in Thailand. *Am J Trop Med Hyg*, 81(4), 695-697. doi:10.4269/ajtmh.2009.09-0252.
- Stimson, A. M. (1907). Note on an Organism Found in Yellow-Fever Tissue. *Public Health Reports (1896-1970)*, 22(18), 541.
- Suut, L., Mazlan, M. N., Arif, M. T., Yusoff, H., Abdul Rahim, N. A., et al. (2016). Serological Prevalence of Leptospirosis Among Rural Communities in the Rejang Basin, Sarawak, Malaysia. *Asia Pac J Public Health*, 28(5), 450-457. doi:10.1177/1010539516648003.
- Takabe, K., Nakamura, S., Ashihara, M., & Kudo, S. (2013). Effect of osmolarity and viscosity on the motility of pathogenic and saprophytic Leptospira. *Microbiol Immunol*, 57(3), 236-239. doi:10.1111/1348-0421.12018.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., et al. (2011). MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol*, 28(10), 2731-2739. doi:10.1093/molbev/msr121.
- Tan, W. L., Soelar, S. A., Mohd Suan, M. A., Hussin, N., Cheah, W. K., et al. (2016). Leptospirosis Incidence and Mortality in Malaysia. *Southeast Asian J Trop Med Public Health*, 47(3), 434-440.
- Tantibhedhyangkul, W., Wongsawat, E., Chongtrakool, P., Tiengrim, S., Thaipadungpanit, J., et al. (2020). Case Report and Literature Review: Recovery of Pathogenic Leptospira spp. from Routine Aerobic Blood Culture Bottles. *Am J Trop Med Hyg*. doi:10.4269/ajtmh.20-0204.
- Thayaparan, S., Robertson, I., & Abdullah, M. T. (2015a). Serological and molecular detection of Leptospira spp. from small wild mammals captured in Sarawak, Malaysia. *Malaysian Journal of Microbiology*, 11(1), 93-101.
- Thayaparan, S., Robertson, I., Fairuz, A., Suut, L., & Abdullah, M. T. (2013a). Serological Prevalence of Leptospiral Infection in Wildlife in Sarawak, Malaysia. *Borneo J. Resour. Sci. Tech.*, 2(2), 79-82.

- Thayaparan, S., Robertson, I., Fairuz, A., Suut, L., Gunasekera, U. C., et al. (2015b). Seroepidemiological study of leptospirosis among the communities living in periurban areas of Sarawak, Malaysia. *Med J Malaysia*, 70(5), 288-294.
- Thayaparan, S., Robertson, I. D., Fairuz, A., Suut, L., & Abdullah, M. T. (2013b). Leptospirosis, an emerging zoonotic disease in Malaysia. *Malays J Pathol*, 35(2), 123-132.
- Utusan Melayu, M. B. (Producer). (2019, 21.1.2020). Pasar Raja Bot Siap 2020.
- Vallée, E., Heuer, C., Collins-Emerson, J. M., Benschop, J., Ridler, A. L., et al. (2018). Effects of natural infection by *L. borgpetersenii* serovar Hardjo type Hardjo-bovis and *L. interrogans* serovar Pomona, and leptospiral vaccination, on sheep growth. *Prev Vet Med*, 159, 196-202. doi:10.1016/j.prevetmed.2018.09.017.
- Victoriano, A. F., Smythe, L. D., Gloriani-Barzaga, N., Cavinta, L. L., Kasai, T., et al. (2009). Leptospirosis in the Asia Pacific region. *BMC Infect Dis*, 9, 147. doi:10.1186/1471-2334-9-147.
- Villanueva, S. Y., Ezoe, H., Baterna, R. A., Yanagihara, Y., Muto, M., et al. (2010). Serologic and molecular studies of *Leptospira* and leptospirosis among rats in the Philippines. *Am J Trop Med Hyg*, 82(5), 889-898. doi:10.4269/ajtmh.2010.09-0711.
- Wahab, Z. A. (2015). *Epidemiology and Current Situation of Leptospirosis in Malaysia*. Jabatan Kerajaan Tempatan (JKT), Kementerian Perumahan Dan Kerajaan Tempatan Malaysia: Jabatan Kerajaan Tempatan (JKT), Kementerian Perumahan Dan Kerajaan Tempatan Malaysia Retrieved from http://jkt.kpkt.gov.my/jkt/resources/PDF/Persidangan_2015/persidangan%20kesihatan/Leptospirosis_in_Malaysia.pdf.
- Yasuda, P. H., Steigerwalt, A. G., Sulzer, K. R., Kaufmann, A. F., Rogers, F., et al. (1987). Deoxyribonucleic Acid Relatedness between Serogroups and Serovars in the Family Leptospiraceae with Proposals for Seven New *Leptospira* Species. *Int J Syst Bact*, 37(4), 407-415. doi:<https://doi.org/10.1099/00207713-37-4-407>.
- Zhang, C., Xu, J., Zhang, T., Qiu, H., Li, Z., et al. (2019). Genetic characteristics of pathogenic *Leptospira* in wild small animals and livestock in Jiangxi Province, China, 2002-2015. *PLoS Negl Trop Dis*, 13(6), e0007513. doi:10.1371/journal.pntd.0007513.
- Zuerner, R. L. (2005). Laboratory maintenance of pathogenic *Leptospira*. *Curr Protoc Microbiol*, Chapter 12, Unit 12E.11. doi:10.1002/9780471729259.mc12e01s00.