



**HOT-WATER DIPPING EFFECT ON POSTHARVEST QUALITY AND  
METABOLITE PROFILING OF BENTONG GINGER (*Zingiber officinale*  
Roscoe) STORED AT LOW TEMPERATURE**

By

**NUR INDAH BINTI ABDUL SHUKOR**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of  
Doctor of Philosophy**

**July 2021**

**FP 2021 79**

## COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**HOT-WATER DIPPING EFFECT ON POSTHARVEST QUALITY AND METABOLITE PROFILING OF BENTONG GINGER (*Zingiber officinale* Roscoe) STORED AT LOW TEMPERATURE**

By

**NUR INDAH ABDUL SHUKOR**

July 2021

**Chairman : Azizah Misran, PhD**  
**Faculty : Agriculture**

Ginger is a tropical produce that is susceptible to chilling injury (CI). Storing ginger below 15 °C will defect its postharvest quality as it is susceptible to chilling injury (CI). Therefore, the objective of this experiment was to evaluate the effects of storage temperatures and storage durations on the physico-chemical qualities of rhizomes. The collected rhizomes from Bentong were stored at 5, 15, and 25 °C and were kept for 0, 8, 16, and 24 days. The experiment was laid out as a randomized complete block design (RCBD) and analysed as a two-way factorial with four replications. For physical attribute, storage under 15 °C showed that weight loss was concomitant with the firmness of rhizomes and was supported by a positive correlation between weight loss and firmness. For colours, changes of the values of  $h^{\circ}$  were gradual when the rhizomes were stored at 15 °C contrary to chilling of rhizomes at 5 °C suggesting that the colour of the rhizomes was managed to be maintained at such condition. The phytochemical analysis resulted in a similar increment in total phenolic and flavonoid when stored at 5 and 15 °C up to 16 d of storage durations. It also showed that storage at 25 °C could retain 6-gingerol and 6-shogaol up to 16 d of storage. For DPPH activities, the antioxidant capacity became weaker, especially during storage at 25 °C as compared to 5 C of storage.

Hot water treatment is developed for disease control and provides an alternative to the application of synthetic chemicals. In conjunction to mitigate CI symptoms, a study on hot water dip at 45 °C for 0, 5, 10, and 15 min and immediately stored at 5 °C for 0, 8 and 16 d were assessed. Storage at 5 °C was chosen as it represents chilling temperature. The postharvest qualities were expected to be prolonged under chilling temperature after being treated in hot water treatment. The experiments were conducted using RCBD, analysed as a two-way factorial with four replications and evaluated for physico-chemical qualities. From the

results, the rhizome weight loss, increased proportionally with dipping durations and storage durations. For firmness, dipped rhizome for 5 min at 45 °C was found to maintain the rhizome firmness along storage as compared to other dipping durations. The hue ( $h^{\circ}$ ) colour value of undipped rhizome resulted in a significant decline after 16 d of storage. Exposed rhizome for 5, 10 and 15 min seemed to maintain the  $h^{\circ}$  of the rhizome after storage at 5 °C. The lightness of colour ( $L^*$ ) also responded similarly. The decrease in  $h^{\circ}$  from yellow to slightly brown also indicated rhizome browning. This was supported by a sharp increase of browning index in undipped rhizome within 16 d of storage at 5 °C. Rhizome held at 45 °C / 5 min managed to reduce browning as compared to other durations. Dipping for 15 min resulted in a significant increase in TPC, TFC and 6-gingerols, especially at 16 days of storage durations whilst DPPH activity was achieved when dipped for 5 min.

Metabolomic profiling on hot water dipped rhizomes has identified two secondary metabolites groups: gingerols-related compounds and diarylheptanoid group. The distinct separation between hot water dip treatment at 5, 10, and 15 min and control were observed via OPLS-DA. The heat map showed the distribution of the up and down regulation of the identified metabolites. Based on the distribution, heat map assists in suggesting the optimum durations of hot water treatment at 45 °C for 5 min to be the optimum duration in reducing browning index that could be an indicator for chilling injury. In conclusion, preconditioning of ginger rhizomes by hot water treatment at 45 °C increased chilling tolerance upon storage at chilling temperature.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN CELUPAN AIR PANAS TERHADAP KUALITI LEPASTUAI DAN  
PROFIL METABOLOMIK RIZOM HALIA (*Zingiber officinale* Roscoe) PADA  
SUHU PENYIMPANAN RENDAH**

Oleh

**NUR INDAH ABDUL SHUKOR**

Julai 2021

**Pengerusi : Azizah Misran, PhD**  
**Fakulti : Pertanian**

Halia merupakan hasil tanaman tropika yang retan kecederaan kedinginan (CI). Penyimpanan halia pada suhu dingin akan merosakkan kualiti lepas tuai akibat mengalami kecederaan kedinginan (CI). Oleh itu, objektif eksperimen ini adalah untuk menilai kesan suhu penyimpanan dan jangka masa penyimpanan terhadap kualiti fizik-kimia rizom. Rizom yang dikutip dari Bentong disimpan pada suhu 5, 15, dan 25 °C selama 0, 8, 16, dan 24 hari. Eksperimen ini disusun menggunakan reka bentuk rawak blok sepenuhnya (RCBD) dan dianalisis sebagai faktorial dua hala dengan empat replikasi. Untuk atribut fizikal, penyimpanan di bawah suhu 15 °C menunjukkan bahawa berlaku penurunan berat selari dengan ketegasan rizom dan disokong oleh korelasi positif antara penurunan berat dan ketegasan. Untuk warna, perubahan nilai  $h^{\circ}$  berlaku secara beransur-ansur apabila rizom disimpan pada suhu 15 °C berbanding dengan penyejukan pada 5 °C. Hal ini menunjukkan bahawa warna rizom berjaya dikekalkan dalam keadaan suhu sedemikian. Analisis fitokimia menunjukkan peningkatan yang serupa bagi jumlah kandungan fenolik dan flavonoid apabila disimpan pada 5 dan 15 °C selama 16 hari tempoh penyimpanan. Ia menunjukkan bahawa penyimpanan pada suhu 25 °C dapat mengekalkan kandungan 6-gingerol dan 6-shogaol sehingga 16 hari tempoh penyimpanan. Untuk aktiviti DPPH, kapasiti antioksidan semakin berkurangan terutama semasa penyimpanan pada suhu 25 °C berbanding dengan penyimpanan pada suhu 5 °C.

Rawatan air panas ialah rawatan, yang telah dikembangkan bagi tujuan kawalan penyakit, dan merupakan alternatif kepada penggunaan bahan kimia sintetik. Sehubungan itu, bagi mengurangkan simptom CI, kajian mengenai celupan air panas pada suhu 45 °C selama 0, 5, 10 dan 15 min dan disimpan pada 5 °C selama 0, 8 dan 16 hari dinilai. Penyimpanan pada suhu 5 °C telah ditentukan

sebagai suhu dingin. Kualiti lepas tuai dijangka dapat dikekalkan pada suhu dingin setelah dirawat dengan rawatan air panas. Eksperimen dilakukan menggunakan RCBD, dalam susunan factorial dua hala dengan empat replikasi dan dinilai untuk kualiti fizik-kimia. Hasil kajian menunjukkan penurunan berat rizom meningkat secara berkadar dengan jangka masa pencelupan. Untuk ketegasan, rizom dicelupkan selama 5 minit pada 45 °C didapati mengekalkan ketegasan rizom di sepanjang tempoh penyimpanan berbanding dengan jangka masa pencelupan yang lain. Nilai warna rizom ( $h^{\circ}$ ) yang tidak dicelup menunjukkan penurunan yang ketara setelah 16 hari tempoh penyimpanan. Rizom yang terdedah selama 5, 10 dan 15 minit dilihat mengekalkan nilai  $h^{\circ}$  bagi rizom selepas penyimpanan pada suhu 5 °C. Kecerahan warna ( $L^*$ ) juga menunjukkan tindak balas yang sama. Penurunan  $h^{\circ}$  daripada kuning hingga sedikit perang juga menunjukkan pemerangan rizom. Ini disokong oleh peningkatan ketara indeks pemerangan dalam rizom yang tidak dicelup dalam tempoh 16 hari pada suhu 5 °C. Rizom yang dicelup pada 45 °C / 5 min berjaya mengurangkan pemerangan berbanding dengan jangka masa yang lain. Pencelupan selama 15 min menyebabkan peningkatan ketara bagi jumlah kandungan TPC, TFC dan 6-gingerol khususnya selepas 16 hari tempoh penyimpanan. Sementara itu, aktiviti antioksidan (DPPH) pula dapat dicapai pada jangka masa pencelupan 5 min.

Pencirian metabolomik terhadap rizom yang dirawat dengan celupan air panas telah mengenal pasti dua kumpulan metabolit sekunder iaitu gingerol dan diarilheptanoid. Pemerhatian terhadap pemisahan distingtif antara rawatan celupan air panas dalam tempoh 5, 10 dan 15 min dengan kawalan dilakukan menggunakan model OPLS-DA. Peta haba menunjukkan taburan peningkatan dan penurunan oleh metabolit yang dikenal pasti. Berdasarkan taburan tersebut, peta haba dapat membantu memberi cadangan jangka masa optimum 5 in bagi rawatan air panas pada suhu 45 °C sebagai tempoh optimum bagi mengurangkan indeks pemerangan yang menjadi tanda aras kepada kecederaan dingin. Berdasarkan penemuan tersebut, dapat dilihat bahawa metabolomik LCMS adalah pendekatan holistik dalam pengesahan rawatan pasca tuai.

Sebagai kesimpulan, rawatan air panas pada suhu 45 °C sebagai prakondisi ke atas rizom halia dapat dilihat mampu mencetuskan ketahanan sewaktu penyimpanan dalam suhu yang dingin.

## ACKNOWLEDGEMENTS

Alhamdulillah that I have finally managed to complete this journey. I would like to express my gratitude to my beloved mother and my late father for supporting me throughout this journey, sending me back and forth to the faculty until I managed to drive on my own. Thank you for not pressuring me as I took many years to complete the entire story. May Allah bless them. To my first supervisor who are my mother's best friend, the ultimate Dr. Siti Hajar Ahmad for bringing me in to the Faculty of Agriculture and made myself to enrol as a phd student under her supervision. I was her last phd student. She made me believe in my capabilities even though I came from a different background. She wanted me badly to become a phd holder as a gift to my mom for not being able to complete her phd studies. Thank you for sharing your valuable knowledge and experiences in study and life. Thank you for those wonderful moments that we have spent together with the rest of the gang. Thank you for your endless support, courage, and guidance. To my last and current supervisor, Dr. Azizah Misran, thank you for adopting me joining your team. Thank you for the financial support and sharing of knowledge. To my first eating gang Kak Elly, Muni, Bunga, and Azimah, thank you guys for always being there for me. To all my little sisters Xue Yi and Elmira, the ginger gang, Aimi the parthenin gang, Tirah the Rawang fig, Xi Yun the peanut planter, Mai the corn breeder, Musalmah the snail collector, Sha my JPJ instructor, Anis the tissue culturist, Yus the nano emulsifier and Miss K the lovely Korean mint, thank you for the lovely friendship and for giving me a ride to anywhere we go. I pray for success and wish you all the best in everything that you do.

Al-fatihah to late Dr Bokhary Zainal.  
Thank you.

I certify that a Thesis Examination Committee has met on (date of viva voce) to conduct the final examination of (student's name) on his (her) thesis entitled "Hot-Water Dipping Effect on Postharvest Quality and Metabolite Profiling of Bentong Ginger (*Zingiber officinale* Roscoe) Stored at Low Temperature" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctoral of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Dato' Abdul Shukor Juraimi, PhD**

Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Hawa ZE Jaafar, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Phebe Ding, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Zora Singh, PhD**

Professor  
Department of Environment and Agriculture  
University of Curtin  
Australia  
(External Examiner)

---

**SITI SALWA ABD GANI, PhD**

Associate Professor ChM. and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of degree of philosophy. The members of the Supervisory Committee were as follows:

**Azizah binti Misran, PhD**

Senior Lecturer  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Juju Nakasha binti Jaafar, PhD**

Senior Lecturer  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

**Nazamid bin Saari, PhD**

Professor  
Faculty of Science and Food Technology  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 11 May 2023

## Declaration by the Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_

Date: \_\_\_\_\_

Name and Matric No: Nur Indah Abdul Shukor

## Declaration by Members of the Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of Chairman  
of Supervisory  
Committee: Dr. Azizah Misran

Signature: \_\_\_\_\_  
Name of Member  
of Supervisory  
Committee: Dr. Juju Nakasha

Signature: \_\_\_\_\_  
Name of Member  
of Supervisory  
Committee: Professor Dr. Nazamid Saari

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xx
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
<b>2 LITERATURE REVIEW</b>	<b>4</b>
2.1 <i>Zingiber officinale</i> Roscoe	4
2.2 Cultivation Characteristics and Harvesting	4
2.3 Chemical Composition, Nutritional Values and Medicinal Values	5
2.4 Storage of Rhizomes at Low Temperature	12
2.5 Chilling Injury (CI)	12
2.5.1 Symptoms of Chilling Injury	13
2.5.2 Effects of Chilling Injury	14
2.5.3 Mechanism of Chilling Injury	14
2.6 Postharvest Treatment in Alleviation of Chilling Injury	16
2.6.1 Heat Treatment	16
2.6.2 Mechanism of Heat Treatment	17
2.6.3 Quality of Evaluation	17
2.7 Metabolomic Profiling	19
2.7.1 Analytical Platform in Metabolomic Profiling using LC-MS	20
2.7.2 Data Processing	21
2.7.3 Chemometric in Metabolomics: PCA, PLS, Correlation and Heatmap	21
2.7.4 Heatmaps	22
<b>3 THE CHANGES IN PHYSICO-CHEMICAL QUALITY OF BENTONG GINGER RHIZOMES STORED AT CHILLING TEMPERATURE</b>	<b>24</b>
3.1 Introduction	24
3.2 Material and Methods	25
3.2.1 Rhizomes Materials and Treatment Conditions	25
3.2.2 Physical Analysis	25
3.2.3 Chemical Analysis	26
3.2.4 Antioxidant Activity	28

	3.2.5	Experimental Design and Statistical Analysis	29
3.3		Result and Discussion	29
	3.3.1	Weight Loss (WL)	29
	3.3.2	Firmness	32
	3.3.3	Colour	34
	3.3.4	Soluble Solid Concentrations (SSC)	39
	3.3.5	Titrateable Acidity (TA)	42
	3.3.6	Browning Index (BI)	46
	3.3.7	Total Phenolic Contents (TPC)	48
	3.3.8	Total Flavonoid Contents (TFC)	50
	3.3.9	Identification and Quantification of 6-gingerol and 6-shogaol	52
	3.3.10	1,1-diphenyl-1-picrylhydrazyl (DPPH) Radical Scavenging Assays	56
3.4		Correlation Analysis	59
3.5		Conclusion	60
<b>4</b>		<b>EFFECTS OF HOT WATER TREATMENT ON POSTHARVEST QUALITIES OF BENTONG GINGER (<i>Zingiber officinale</i>) RHIZOMES</b>	<b>61</b>
	4.1	Introduction	61
	4.2	Material and Methods	62
	4.2.1	Rhizomes Materials and Treatment Conditions	62
	4.2.2	Physical Analysis	62
	4.2.3	Chemical Analysis	63
	4.2.4	Antioxidant Activity	64
	4.2.5	Experimental Design and Statistical Analysis	64
	4.3	Result and Discussion	64
	4.3.1	Weight Loss (WL)	64
	4.3.2	Firmness	67
	4.3.3	Colour	68
	4.3.4	Soluble Solid Concentrations (SSC)	74
	4.3.5	Browning Index (BI)	76
	4.3.6	Total Phenolic Contents (TPC)	79
	4.3.7	Total Flavonoid Contents (TFC)	81
	4.3.8	Identification and Quantification of 6-gingerol and 6-shogaol	83
	4.3.9	1,1-diphenyl-1-picrylhydrazyl (DPPH) Radical Scavenging Capacity Assays	85
	4.4	Correlation Analysis	88
	4.5	Conclusion	89
<b>5</b>		<b>METABOLOMIC PROFILING ON THE EFFECT OF HOT WATER DIP TREATMENT IN GINGER RHIZOMES</b>	<b>90</b>
	5.1	Introduction	90
	5.2	Material and Methods	90
	5.2.1	Sample Preparation	90

5.2.2	Extraction of Metabolites	91
5.2.3	High Performance Liquid Chromatography using Mass Spectrometry	91
5.2.4	Data Processing and Statistical Analysis	91
5.3	Results and Discussions	92
5.3.1	Metabolite Profiling of Hot Water Dip Rhizomes using LC-HRESI-MS	92
5.3.2	Discrimination of Secondary Metabolites as Affected by Dipping Durations of Hot Water Dip Treatment and Storage Durations of Ginger Rhizomes	96
5.4	Conclusion	101
<b>6</b>	<b>CONCLUSION AND RECOMMENDATION</b>	<b>102</b>
	<b>REFERENCES</b>	<b>104</b>
	<b>APPENDICES</b>	<b>125</b>
	<b>BIODATA OF STUDENT</b>	<b>137</b>
	<b>LIST OF PUBLICATIONS</b>	<b>138</b>

## LIST OF TABLES

Table		Page
2.1	Chemical constituents isolated from rhizomes of <i>Zingiber officinale</i> by different solvent extraction	7
3.1	The main and interaction effects of storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16 and 24 d) on weight loss and firmness of ginger	30
3.2	The main and interaction effects of storage temperatures (5, 15, and 25 °C) and storage durations (0, 8, 16, and 24 d) on hue, lightness, and chroma of ginger rhizomes	34
3.3	The main and interaction effects of storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16 and 24 d) on soluble solids concentrations of ginger rhizomes	40
3.4	The main and interaction effects of storage temperatures (5, 15, and 25 °C) and storage durations (0, 8, 16, and 24 d) on titratable acidity of ginger rhizomes	43
3.5	The main and interaction effects of storage temperatures (5, 15, and 25 °C) and storage durations (0, 8, 16, and 24 d) on the browning index of Bentong ginger rhizomes	46
3.6	The main and interaction effects of storage temperatures (5, 15, and 25 °C) and storage durations (0, 8, 16, and 24 d) on total phenolic contents and total flavonoid contents of Bentong ginger rhizomes	48
3.7	The main and interaction effects of storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16 and 24 d) on 6-gingerol and 6-shogaol contents of Bentong ginger rhizomes	53
3.8	The main and interaction effects of storage temperatures (5, 15, and 25 °C) and storage durations (0, 8, 16, and 24 d) on 1,1-diphenyl-1-picrylhydrazyl activities of Bentong ginger rhizomes	57
3.9	Pearson correlation coefficient (r) matrices among the responses of physical quality attributes of hot water dipped rhizomes for 0, 5, 10 and 15 min for 0, 8 and 16 days under storage of 5 °C	59
4.1	The main and interaction effects of dipping durations (0, 5, 10 and 15 min) and storage durations (0, 8, 16 and 24 d) on weight loss and firmness of ginger rhizomes	65

4.2	The main and interaction effects of dipping durations (0, 5, 10 and 15 min) and storage durations (0, 8, 16 and 24 d) on hue ( $h^\circ$ ), lightness ( $L^*$ ) and chroma ( $C^*$ ) of ginger rhizomes	69
4.3	Total colour changes $\Delta E$ of hot water dipped rhizomes (5, 10 and 15 min) at initial and day 16 after storage under 5 °C	73
4.4	The main and interaction effects of dipping durations (0, 5, 10 and 15 min) and storage durations (0, 8, and 16 d) on soluble solid concentrations (%) of ginger rhizomes	74
4.5	The main and interaction effects of dipping durations (0, 5, 10 and 15 min) and storage durations (0, 8, and 16 d) on browning index (abs) of ginger rhizomes	77
4.6	The main and interaction effects of dipping durations (0, 5, 10 and 15 min) and storage durations (0, 8, and 16 d) on total phenolic contents and total flavonoid contents of ginger rhizomes	79
4.7	The main and interaction effects of dipping durations (0, 5, 10 and 15 min) and storage durations (0, 8, and 16 d) on 6-gingerol and 6-shogaol contents of ginger rhizomes	83
4.8	The main and interaction effects of dipping durations (0, 5, 10 and 15 min) and storage durations (0, 8, and 16 d) on 1,1-diphenyl-1-picrylhydrazyl radical scavenging activities of ginger rhizomes	86
4.9	Pearson correlation coefficient ( $r$ ) matrices among the responses of physical quality attributes of hot water dipped rhizomes for 0, 5, 10 and 15 min for 0, 8 and 16 days under storage of 5 °C	88
5.1	Chromatographic and mass spectral characteristics of metabolite contents in hot water dipped rhizomes by LC-ESI-MS from the Antimarin and DNP database	95



## LIST OF FIGURES

Figure	Page
2.1 Chemical structures of common gingerols, shogaols, and paradols. (1) [6]-gingerol, (2) [8]-gingerol, (3) [10]-gingerol, (4) [6]-shogaol, (5) [8]-shogaol, (6) [10]-shogaol, (7) [6]-paradol, (8) [8]-paradol, and (9) [10]-paradol	10
2.2 Symptoms of chilling injury in ginger rhizome. (a) Water-soak appearance (b) Skin peeling (c) Shrivelling (d) Browning (e) Skin purple	13
3.1 The relationship between weight loss of ginger rhizome and storage duration (0, 8, 16 and 24 d) at different storage temperatures (5, 15 and 25 °C) of Bentong ginger rhizomes. $Y(5) = -1.87 \times 10^{-14} + 0.14x + 0.00031x^2 - 0.00019x^3$ ; $R^2 = 0.99$ ; $Y(15) = -2.16 \times 10^{-16} + 0.10x - 0.0037x^2 + 2.60 \times 10^{-16}x^3$ ; $R^2 = 0.99$ and $Y(25) = -1.12 \times 10^{-14} - 0.038x + 0.061x^2 - 0.0022x^3$ ; $R^2 = 0.99$ . Means with the different letters are significantly different at $P=0.05$ . Vertical bars indicate the standard error ( $n = 4$ )	31
3.2 The relationship between firmness and storage duration (0, 8, 16 and 24 d) at different storage temperatures (5, 15 and 25 °C) of Bentong ginger rhizomes. $Y(5) = 56.43 - 1.86x$ ; $R^2 = 0.76$ , $Y(15) = 65.79 - 0.61x$ ; $R^2 = 0.89$ , $Y(25) = 52.31 - 1.19x$ ; $R^2 = 0.43$ . Means with the different letters are significantly different at $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ )	33
3.3 The relationship between (a) hue ( $h^\circ$ ), (b) chroma ( $C^*$ ) and (c) lightness ( $L^*$ ) and storage duration (0, 8, 16 and 24 d) at different storage temperatures (5, 15 and 25 °C) of Bentong ginger rhizomes. $h^\circ$ : $Y(5) = 98.26 - 0.58x + 0.02x^2$ ; $R^2 = 0.77$ ; $Y(15) = 33.0 - 0.86x + 0.021x^2$ ; $R^2 = 0.73$ and $Y(25) = 31.99 - 1.94x - 0.097x^2$ ; $R^2 = 0.99$ . $C^*$ : $Y(5) = 31.5 - 1.73x + 0.064x^2$ ; $R^2 = 0.88$ ; $Y(15) = 33.4 - 0.82x + 0.018x^2$ ; $R^2 = 0.70$ , and $Y(25) = 32.9 - 2.07x + 0.10x^2$ ; $R^2 = 0.98$ . $L^*$ : $Y(5) = 76.98 - 1.92x + 0.071x^2$ ; $R^2=0.98$ , $Y(15) = 76.08 - 1.87x + 0.07x^2$ ; $R^2 = 0.75$ , and $Y(25) = 76.49 - 2.16x + 0.096x^2$ ; $R^2 = 0.92$ . Means with the different letters are significantly different at $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ )	36
3.4 The relationship between soluble solids concentration and storage duration (0, 8, 16 and 24 d) at different storage temperatures (5, 15 and 25 °C) of Bentong ginger rhizomes. $Y(5) = 3.4 - 0.23x + 0.019x^2 - 0.0007x^3$ ; $R^2 = 0.99$ , $Y(15) = 3.4 - 0.21x + 0.024x^2 - 0.001x^3$ ; $R^2 = 0.99$ and $Y(25) = 3.4 - 0.45x + 0.075x^2 + 0.0019x^3$ ; $R^2 = 0.99$ . Means with the different letters	

- are significantly different at  $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ ) 41
- 3.5 The relationship between titratable acidity and storage duration (0, 8, 16 and 24 d) at different storage temperatures (5, 15 and 25 °C) of Bentong ginger rhizomes.  $Y(5) = 0.36 + 0.072x - 0.058x^2 + 0.00014x^3$ ,  $R^2 = 0.92$ ;  $y(15) = 0.36 + 0.017x - 0.0023x^2 + 0.00009928x^3$ ,  $R^2 = 0.84$  and  $Y(25) = 0.36 + 0.138x - 0.013x^2 + 0.00035x^3$ ,  $R^2 = 0.96$ . Means with the different letters are significantly different at  $P = 0.05$  44
- 3.6 The relationship between browning index and storage duration (0, 8, 16 and 24 d) at different storage temperatures of ginger (5, 15 and 25 °C).  $Y(5) = 0.27 + 0.0026x + 0.0036x^2 - 0.00010x^3$ ;  $R^2 = 0.99$ ,  $Y(15) = 0.27 + 0.068x - 0.0050x^2 + 0.00013x^3$ ;  $R^2 = 0.99$  and  $Y(25) = 0.27 - 0.039x + 0.010x^2 - 0.00032x^3$ ;  $R^2 = 0.99$ . Means with the different letters are significantly different at  $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ ) 47
- 3.7 The relationship between total phenolic contents and storage duration (0, 8, 16 and 24 d) at different storage temperatures of ginger (5, 15 and 25 °C).  $Y(5) = 30.04 - 6.09x + 0.88x^2 - 0.026x^3$ ;  $R^2 = 0.99$ ,  $Y(15) = 30.04 - 7.26x + 1.02x^2 - 0.03x^3$ ;  $R^2 = 0.99$  and  $Y(25) = 30.04 - 4.20x + 0.58x^2 - 0.017x^3$ ;  $R^2 = 0.99$ . Means with the different letters are significantly different at  $P = 0.05$  49
- 3.8 The relationship between total flavonoid contents and storage duration (0, 8, 16 and 24 d) at different storage temperatures (5, 15 and 25 °C) of Bentong ginger rhizomes.  $Y(5) = 30.78 - 2.44x + 0.71x^2 - 0.023x^3$ ;  $R^2 = 0.99$ ,  $Y(15) = 30.78 - 6.08x + 1.26x^2 - 0.04x^3$ ;  $R^2 = 0.99$  and  $Y(25) = 30.78 + 2.81x - 0.15x^2 + 0.00034x^3$ ;  $R^2 = 0.99$ . Means with the different letters are significantly different at  $P = 0.05$  51
- 3.9 The relationship between 6-gingerol and 6-shogaol contents and storage duration (0, 8, 16 and 24 d) at different storage temperatures (5, 15 and 25 °C) of Bentong ginger rhizomes. (a)  $Y(5) = 34.35 - 3.78x + 0.82x^2 - 0.027x^3$ ;  $R^2 = 0.99$ ,  $Y(15) = 34.35 - 3.54x + 0.68x^2 - 0.021x^3$ ;  $R^2 = 0.99$  and  $Y(25) = 34.35 - 0.68x + 0.23x^2 + 0.0083x^3$ ;  $R^2 = 0.99$ . (b)  $Y(5) = 1.52 - 0.22x + 0.03x^2 - 0.012x^3$ ;  $R^2 = 0.99$ ,  $Y(15) = 1.52 - 0.12x + 0.023x^2 - 0.0007x^3$ ;  $R^2 = 0.99$  and  $Y(25) = 1.52 + 0.13x - 0.0092x^2 + 0.0002x^3$ ;  $R^2 = 0.99$ . Means with the different letters are significantly different at  $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ ) 54
- 3.10 Dehydration of 6-gingerol to 6-shogaol 55

- 3.11 HPLC profile of (1) 6-gingerol and (2) 6-shogaol in the juice of ginger rhizome 56
- 3.12 The relationship between percentage of DPPH radical scavenging activities and storage duration (0, 8, 16, and 24 d) at different storage temperature of Bentong ginger rhizomes (5, 15 and 25 ° C).  $Y(5) = 87.87 + 0.74x - 0.07x^2$ ;  $R^2 = 0.98$ ,  $Y(15) = 85.96 + 7.94x - 0.96x^2 + 0.023x^3$ ;  $R^2 = 0.82$ , and  $Y(25) = 90.95 + 1.77x - 0.65x^2 + 0.020x^3$ ;  $R^2 = 0.59$ . Means with the different letters are significantly different at  $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ ) 57
- 4.1 The relationship between weight loss of ginger rhizome and storage duration (0, 8, 16 and 24 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger.  $Y(0) = 0.039 + 0.089x - 0.0027x^2$ ;  $R^2 = 0.89$ ,  $Y(5) = 0.17 + 0.33x - 0.0049x^2$ ;  $R^2 = 0.96$ ,  $Y(10) = 0.425 + 0.63x - 0.0068x^2$ ;  $R^2 = 0.94$  and  $Y(15) = 0.91 + 0.99x - 0.025x^2 + 0.0059x^3$ ;  $R^2 = 0.56$ . Means with the different letters are significantly different at  $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ ) 66
- 4.2 The relationship between firmness of ginger rhizome and storage duration (0, 8, and 16 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger.  $Y(15) = 49.36 - 1.41x + 0.096x^2$ ;  $R = 0.79$ . Means with the different letters are significantly different at  $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ ) 67
- 4.3 The relationship between hue ( $h^\circ$ ) of ginger rhizome and storage duration (0, 8, and 16 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger.  $h^\circ$ :  $Y(0) = 95.85 - 1.31x$ ;  $R^2 = 0.91$ ;  $Y(5) = 97.66 - 0.18x$ ;  $R^2 = 0.73$ .  $C^* = Y(0) = 28.70 + 0.25x - 0.055x^2$ ;  $R^2 = 0.93$ ;  $Y(5) = 26.84 + 0.56x - 0.04x^2$ ;  $R^2 = 0.74$ , and  $Y(15) = 8.01 - 4.21x + 0.26x^2$ ;  $R^2 = 0.97$ .  $L^* = Y(0) = 65.14 + 0.38x - 0.078x^2$ ;  $R^2 = 0.97$ ;  $Y(5) = 67.78 - 0.28x + 0.025x^2$ ;  $R^2 = 0.53$ ;  $Y(10) = 69.35 + 0.41x - 0.026x^2$ ;  $R^2 = 0.54$ ;  $Y(15) = 75.87 - 0.34x + 0.024x^2$ ;  $R^2 = 0.62$ . Means with the different letters are significantly different at  $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ ) 70
- 4.4 The relationship between soluble solid concentrations (%) of ginger rhizomes and storage duration (0, 8, 16 and 24 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger.  $Y(0) = 5.01 - 0.07x$ ;  $R^2 = 0.71$ ;  $Y(10) = 2.95 - 0.13x$ ;  $R^2 = 0.99$  and  $Y(15) = 4.75 + 0.06x$ ;  $R^2 = 0.92$ . Means with the different letters are significantly different at  $P = 0.05$ . Vertical bars indicate the standard error ( $n = 4$ ) 75
- 4.5 The response for browning index ( $abs_{480\text{ nm}}$ ) of ginger rhizomes and storage duration (0, 8, and 16 d) at different dipping

	duration (0, 5, 10 and 15 min) of Bentong ginger. Vertical bars indicate the standard error (n = 4)	78
4.6	The relationship between total phenolic contents (g GAE kg <sup>-1</sup> FW) of ginger rhizomes and storage duration (0, 8, and 16 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger rhizomes. $Y(0) = 0.00126 - 8.4 \times 10^{-5}x + 4.4 \times 10^{-6}x^2$ ; $R^2 = 0.86$ , $Y(5) = 0.001 - 4.2 \times 10^{-5}x - 1.9 \times 10^{-6}x^2$ ; $R^2 = 0.97$ , $Y(10) = 0.0013 - 5.5 \times 10^{-5}x + 4.7 \times 10^{-6}x^2$ ; $R^2 = 0.96$ , $Y(15) = 0.0014 + 0.00013x - 4.2 \times 10^{-6}x^2$ ; $R^2 = 0.97$ . Vertical bars indicate the standard error (n = 4)	80
4.7	The relationship between total flavonoids contents (g RUE kg <sup>-1</sup> FW) and storage duration (0, 8, and 16 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger rhizomes. $Y(0) = 0.032 - 0.00013 \times 10^{-5}x - 1.2 \times 10^{-5}x^2$ ; $R^2 = 0.92$ , $Y(5) = 0.022 - 0.0013x - 8.9 \times 10^{-5}x^2$ ; $R^2 = 0.54$ , $Y(10) = 0.017 - 0.0025x - 0.00017x^2$ ; $R^2 = 0.89$ , $Y(15) = 0.043 - 0.0052x + 0.00049x^2$ ; $R^2 = 0.99$ . Vertical bars indicate the standard error (n = 4)	82
4.8(a)	The relationship between 6-gingerol contents (g kg <sup>-1</sup> FW) and storage duration (0, 8, and 16 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger rhizomes. $Y(0) = 3.09 \times 10^{-5} + 1.1 \times 10^{-6}x - 6.4 \times 10^{-8}x^2$ ; $R^2 = 0.95$ , $Y(5) = 0.00062 - 0.0001x + 6.4 \times 10^{-6}x^2$ ; $R^2 = 0.99$ , $Y(10) = 0.0014 - 0.00022x + 9.5 \times 10^{-6}x^2$ ; $R^2 = 0.99$ and $Y(15) = 0.0014 - 0.0002x + 8.5 \times 10^{-6}x^2$ ; $R^2 = 0.99$ . Means with the different letters are significantly different at $P = 0.05$ . Vertical bars indicate the standard error (n = 4)	84
4.8(b)	The relationship between 6-shogaol contents (g kg <sup>-1</sup> FW) of ginger rhizomes and storage duration (0, 8, and 16 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger. Vertical bars indicate the standard error (n = 4)	84
4.9	The relationship between percentage of DPPH radical scavenging activities (%) and storage duration (0, 8, and 16 d) at different dipping duration (0, 5, 10 and 15 min) of Bentong ginger rhizomes. $Y(0) = 14.01 + 4.04x - 0.21x^2$ ; $R^2 = 0.99$ , $Y(5) = 77.3 + 0.07x + 0.0038x^2$ ; $R^2 = 0.80$ , $Y(10) = 77.9 - 6.7x + 0.35x^2$ ; $R^2 = 0.98$ , and $Y(15) = 9.2 + 0.27x + 0.049x^2$ ; $R^2 = 0.98$ . Means with the different letters are significantly different at $P = 0.05$ . Vertical bars indicate the standard error (n = 4)	87
5.1	Total ion chromatogram of hot water (45 °C) dipped rhizomes for 5, 10, and 15 min stored under for 0, 8 16	94
5.2	Secondary metabolites that were putatively identified using LCMS. 2,3,5-trihydroxy-1,7-bis(4-hydroxy-3-methoxyphenyl)heptane (1), 1-(4-hydroxy-3,5-dimethoxyphenyl)-7-(4-hydroxy-	

- 3- methoxyphenyl)-3,5-heptanediol (2), Hexahydrocurcumin (3), 1,7-bis(4-hydroxyphenyl)-5-oxo-3-heptanyl B-D-glucopyranoside (4), 6-gingerdiol (5), 6-gingerol (6), 6-shogaol (7), 8-shogaol (8), and 6-gingerdione (9) 96
- 5.3 (a) Unsupervised PCA scores plot ( $R^2 = 0.75$ ,  $Q^2 = 0.42$ ). (b) Supervised OPLS-DA score plot analysis 97
- 5.4 Heat map of secondary metabolites in treated and non-treated ginger rhizomes. The colours indicate the proportional content of each identified metabolites as determined by the average peak response area with  $R$  scale normalization. Three independent replicates were performed for each stage. Con = control; HT (5, 10, and 15) = hot treatment duration in min; D (0, 8, and 16) = storage durations 98

## LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
°C	Degree celsius
AlCl <sub>3</sub>	Aluminium chloride
Aq	Aqueous
BI	Browning index
C*	Chroma
CHCl <sub>3</sub>	Chloroform
CHS	Chalcone synthase
CI	Chilling injury
cm min <sup>-1</sup>	Centimetre per minute
D	Days
DAD	Diode array detector
DCM	Dichloromethane
DD	Dipping duration
DNA	Deoxyribonucleic acid
DNP	Dictionary of Natural Products
DPPH	1, 1-diphenyl-2-picrylhydrazyl
ESI	Electron spray ionization
ESI	Electron spray ionization
EtOH	Ethanol
FW	Fresh weight
g	Gram
g kg <sup>-1</sup>	Gram per kilogram
g L <sup>-1</sup>	Gram per litre
GAE	Gallic acid equivalents
GC	Gas chromatography

h°	Hue
HPLC/EI-MS	High-performance liquid chromatography/electrospray ionization-mass spectrometry
HR-MS	High-resolution mass spectrometry
HSD	Tukey's honestly significant difference
HSP	Heat shock protein
IT	Ion trap
kg	Kilogram
kV	Kilovolt
L	Litre
L*	Lightness
m	Meter
MeOH	Methanol
mg	Milligram
min	Minute
mL	Millilitre
µm	Micro metre
MS	Mass spectrometry
MWA	Multivariate analysis
N	Force
Na <sub>2</sub> CO <sub>3</sub>	Sodium carbonate
NAD-MDH	NAD-dependent malate dehydrogenase
NADP-ME	NADP-malic enzyme
NADPH	
NaNO <sub>3</sub>	Sodium nitrate
NaOH	Sodium hydroxide
nd	Nonsignificant
nm	Nanometre

OPLS-DA	Orthogonal projection to latent structure
<i>P</i>	Probability
PAL	Phenylalanine ammonia lyase
PCA	Principal component analysis
PE	Petroleum ether
PEPC	phosphoenolpyruvate carboxylase
PLS-DA	Partial least squares-discriminant analysis
Ppm	Part per million
PPO	Polyphenol oxidase
PROC GLM	Procedure of the general linear model
PROC REG	Procedure of regression
RCBD	Randomized completely block design
RE	Rutin equivalent
ROS	Reactive oxidative species
SD	Storage duration
SSC	Soluble solid concentrations
ST	Storage temperature
TA	Titrateable acidit
TFC	Total flavonoid content
TIC	Total ion chromatogram
TOF	Time of flight
TPC	Total phenolic content
UPLC	Ultra-performance liquid chromatography
UV	Ultraviolet detector
WL	Weight loss
ANOVA	Analysis of variance



## CHAPTER 1

### INTRODUCTION

Botanically, ginger, known as *Zingiber officinale* (Zingiberaceae), is a perennial herb grown as an annual crop. The thick tuberous rhizome is pale yellow in colour and aromatic in scent is filled with pungent taste (Gupta and Sharma, 2014). Ginger consumption as a medicinal herb has been practiced since ancient times in treating various illnesses such as nausea, menstruation disorder, inflammation, cough and cold, food poisoning, epilepsy, and even cancer (Shukla and Singh, 2007). The ability of ginger to reduce the symptoms of those illnesses is related to the therapeutic properties, including antimicrobial, antioxidant due to the presence of gingerol and shogaol and its derivatives (Kumar et al., 2011).

Up to 2017, the total number of ginger production has reached 3,038,120 tons worldwide with India is the top country contributing to 35.2 % of production. Malaysia ranked the 13<sup>th</sup> position with total ginger production of 14, 279 tons in 2017, equivalent to only 0.5 % of the world population (FAOSTAT, 2019). In Malaysia, Bentong, Bara, and Tanjung Sepat varieties have been cultivated locally. Bentong is the most popular ginger as the pungency is stronger, low in fibre contents and bigger in size as compared to other varieties (Mohd et al., 2015). The production of Bentong ginger has reached 7,382.399 kg in 2016, which is equivalent to RM 64 million of profits (Harian Metro, 2017).

Bentong rhizomes offered the best quality as compared to imported ginger from Thailand, China and Indonesia. Therefore, Bentong rhizomes have been exported mainly to Singapore followed by South Africa and Indonesia. However, improper temperature management of the rhizomes will reduce the postharvest qualities and marketability (IndexBox, 2021). It is known that ginger is a tropical produce which is prone to chilling injury when stored at a temperature lower than 15 °C (Kasamo et al., 2000). Storing the rhizomes under this temperature may develop physiological breakdown as it alters the physical properties of cell membranes, thus reducing the membrane elasticity (Kasamo et al., 2000).

Exported Bentong ginger follows a series of postharvest standards of procedure. Washed and cleaned rhizomes are treated with anti-fungal solutions at 500 ppm to prevent fungal growth. After the curing process, rhizomes are packed in corrugated fireboard and stored at 10-15 °C at relative humidity of 85-90% to extend its shelf life up to 4-5 months before being transported by ship (Mohd Ismail and Abd Shukor, 1990).

In common practice during transportation, the imported rhizomes were stored under refrigeration at low temperature around  $\pm 2$  °C. They deposited to a local wholesale market and stored in a cold room under the same condition, causing the rhizomes to manifest its CI symptoms such as translucency and shrivelling

when returned to ambient temperature. Eventually, reducing its physical and chemical qualities such as the colour, sweetness, acidity, phytochemical contents, antioxidant, and enzyme activities. These deteriorating effects may imply that ginger rhizomes are intolerant to chilling temperatures and the means to avoid this deterioration needs to be addressed.

There are various ways to control chilling injury depending on the effectiveness of the techniques in achieving the target. Alleviation of chilling injury symptoms can be controlled by increasing the tolerance of commodities or retarding the development of chilling injury symptoms in the tropical and subtropical that are susceptible to chilling injury. Reducing chilling injury symptoms involves modifying the storage environment such as low and high temperature conditioning and intermittent warming (Wang, 2010). Previous studies showed that hot water treatment heat-treated by hot water dip at 45 °C for 4 min, and then stored at 2 °C for 90 days, the CI symptoms were slightly, but significantly reduced after being heat-treated (Mirdehghan et al., 2007). All tubers were treated with hot water (45, 50 and 55 °C), subjected to 10 min and 20 min immersion time was able to maintain the other postharvest qualities such as weight loss, firmness, titratable acidity and total soluble solid (TSS) of the treated sweet potatoes. Other findings on tuber such as potato that dipped into 45 °C hot water for 10 min effectively reduced the weight loss and the disease index of wounded potato tubers (Wang et al., 2020). In this study, hot water treatment was accessed to examine its mitigating effect to alleviate chilling injury symptoms.

Although hot water treatment could control the effect of chilling injury in many fruits (Wang et al., 2014), its effect on ginger rhizomes has yet to be tested, especially on ginger Bentong rhizome. Recently, the evaluation of the commodity reactions to the hot treatment has been widely studied at transcriptome, proteome and metabolome level (Lurie and Pedreschi, 2014). In this study, hot water dip treatment was implemented to control chilling injury symptoms on ginger rhizomes as it is more efficient than hot air (Lurie, 1998). Although hot water treatment is commonly used as pest control and prevention of fungal rot, research on hot water treatment on CI of postharvest qualities has been successful (Gonzales-Aguilar et al., 2000; Wang et al., 2014). The work on ginger rhizome, however, has not been carried out yet.

Evaluation of physico-chemicals postharvest qualities include weight loss, softening (Ketsa et al., 1997), ripening, colour (Wang et al., 2014), phytochemical contents, and antioxidant properties has been carried out. Under such treatment, ginger rhizome needs to be established. To facilitate an understanding between the relationship and correlation between the heat treatment and changes or alterations in metabolites, a metabolic profiling that covers primary and secondary metabolites was executed by high-performance liquid chromatography/electrospray ionization-mass spectrometry (HPLC/EI-MS). With the advance in data processing and analysis by multivariate analysis (MVA) using principal component analysis (PCA), a clear comprehension on the mechanism of hot water treatment in alleviating chilling injury in ginger rhizomes

could be achieved. These metabolomic studies have been successfully adopted in some studies (Tanaka et al., 2015). Even though hot water treatments could pro-long the shelf-life of fresh-cut products or fruits, the literature on the effect of hot water treatments in reducing chilling injury symptoms on rhizomes is still not conclusive and limited.

It is hypothesized that optimum storage temperature would have significant interaction with storage duration in reducing the chilling injury symptoms of rhizomes. Secondly, the hot water treatment could reduce the chilling injury symptoms when the rhizomes are stored under chilling temperature. Therefore, to test the hypotheses, this study is initiated with the objectives:

1. To characterize chilling injury of Bentong ginger rhizome during storage at low temperature by physical and chemical mechanisms.
2. To evaluate the effect of hot water treatment at different dipping durations and storage durations on physical characteristics and chemical mechanisms on Bentong ginger rhizome.
3. To characterize the metabolic profiling of hot water-treated rhizome based on high-performance liquid chromatography coupled with electrospray ionization-mass spectrometry (HPLC-EI-MS).

## REFERENCES

- Abbasi, K.S., Masud, T., Qayyum, A., Khan, S. U., Abbas, S. and Matthew A. J. (2016). Storage stability of potato variety Lady Rosetta under comparative temperature regimes. *Sains Malaysiana*, 45(5): 677– 688.
- Abd. Shukor, R., Aziz, I, A.B. and Ahmad Shokri, O. 1986. Physico-chemical changes of fresh ginger rhizomes as influenced by storage temperature and duration. *MARDI Res. Bull.* 14(3), 243-248
- Aeschbach, R., Loliger, J., Scott B.C. Murcia, A., Butler, J., Halliwell, B. and Oi, A. (1994). Antioxidant actions of thymol, carvacrol, 6- gingerol, zingerone and hydroxytyrosol. *Food Chemistry Toxicology*, 32(1): 31-36.
- Ahmed, A. M. A. R. E. (2018). Preclinical studies of ginger (*Zingiber officinale*) rhizome and its clinical trial as an add-on antiepileptic therapy in children with generalized epilepsies. Doctor of Philosophy. University of Gezira.
- Akram, M., Ibrahim Shah, M., Khan Usmanghan, Mohiuddin, E., Abdul Sami, A.M., Ali Shah, S.M., Khalil, A. and Ghazala, S. (2011). *Zingiber officinale* Roscoe (A Medicinal Plant). *Pakistan Journal of Nutrition*, 10(4): 399-400.
- Aksoy, L., Kolay, E., Agilonu, Y., Aslan, Z. and Kargioglu, M. (2013). Free radical scavenging activity, total phenolic content, total antioxidant status, and total oxidant status of endemic *Thermopsis turcica*. *Saudi Journal of Biological Sciences*, 20(3): 235-239.
- Alberti Á., Riethmüller E. and Béni S. (2018). Characterization of diarylheptanoids: an emerging class of bioactive natural products. *J. Pharm. Biomed. Anal.*, 147:13–34.
- Altunkaya, A. and Gokmen, V. (2009). Effect of various anti-browning agents on phenolic compounds profile of fresh lettuce (*L. sativa*). *Food Chemistry*, 117: 122-126.
- Amalraj A., Pius A., Gopi S. and Gopi S. (2017). Biological activities of curcuminoids, other biomolecules from turmeric and their derivatives—A review. *Journal of Traditional Complement Med.*, 7(2): 205–233.
- Angasu, O. N., Dessalgne, O. G. and Tadese, T. N. (2014). Effect of hot water treatment on quality and incidence of postharvest disease of mango (*Mangifera indica* L.) fruits. *Asian Journal of Plant Sciences*, 13(2): 87-92.
- Aquino-Bolaños, E.N., Cantwell, M.I., Peiser., G. and Mercado-Silva., E. (2000). Changes in the quality of fresh-cut jicama in relation to storage temperatures and controlled atmospheres. *Journal of Food Science*, 65(7): 1238–1243.

- Araya, J. J., Zhang, H., Prisinzano, T. E., Mitscher, L. A. and Timmermann, B. N. (2011). Identification of unprecedented purine-containing compounds, the zingerines, from ginger rhizomes (*Zingiber officinale* Roscoe) using a phase-trafficking approach. *Phytochemistry*, 72: 935-941.
- Arendse, E., Fawole, O. A., and Opara, U. L. (2014). Effects of postharvest storage conditions on phytochemical and radical-scavenging activity of pomegranate fruit (cv. Wonderful). *Scientia Horticulture*, 169: 125-129.
- Armstrong, F. B. and Bennet, T. P. (1979). *Biochemistry*. New York: Oxford University Press, Inc.
- Azu, N. and Onyeagba, R. (2007). Antimicrobial properties of extracts of *Allium cepa* (Onions) and *Zingiber officinale* (Ginger) on *Escherichia coli*, *Salmonella typhi* and *Bacillus subtilis*. *The Internet Journal of Tropical Medicine*, 3:1–10.
- Bambirra, M.L.A., Junqueira, R.G. and Glória, M. B. A. (2002). Influence of post harvest processing conditions on yield and quality of ground turmeric (*Curcuma longa* L.). *Brazilian Archives of Biology and Technology*, 45(4):pp. 423-429.
- Bao, L., Deng, A., Li, Z., Du, G. and Qin, H. (2010). Chemical constituents of rhizomes of *Zingiber officinale*. *Zhongguo Zhong Yao Za Zhi*, 35(5):598-601.
- Bartosz, G. (1997). Oxidative stress in plants. *Acta Physiologiae Plantarum*, 19(1): 47-64.
- Bassal, M. and El-Hamahmy, M. (2011). Hot water dip and preconditioning treatments to reduce chilling injury and maintain postharvest quality of Navel and Valencia oranges during cold quarantine. *Postharvest Biology and Technology*, 60: 186–191.
- Beaulieu, J. C. and Gorny, J. R. (2001). Fresh-cut fruits. In: Gross, K. C., Saltveit, M. E., Wang, C. Y. (Eds). *The commercial storage of fruits, vegetables, and florist and nursery stocks*, p. 1-49. *USDA Handbook 66*: 1-49.
- Becker, C., Klaering, H. P., Kroh, L. W. and Krumbein, A. (2014). Cool-cultivated red leaf lettuce accumulates cyanidin-3-O-(6"-O-malonyl)-glucoside and caffeoylmalic acid. *Food Chemistry*, 146, 404-11.
- Bedair, M. and Sumner, L. W. (2008). Current and emerging mass-spectrometry technologies for metabolomics. *Trends in Analytical Chemistry*, 27(3): 238 – 250.
- Bhattarai, S., Tran, V.H. and Duke, C.C. (2007). Stability of [6]-gingerol and [6]-shogaol in simulated gastric and intestinal fluids. *J Pharm Biomed Anal*, 45(4):648-53.

- Blaesing, D. (2016). Action Planner: A guide to post-harvest management of Australian ginger. Good Practice Action Planner V1.
- Bugaud, C., Deverge, E., Daribo, M. O., Ribeyre, F. (2011). Sensory characterisation enabled the first classification of dessert bananas. *Journal of the Science of Food and Agriculture*, 91(6):992-100.
- Burdurlu, H. S. and Karadeniz, F. (2003). Effect of storage on nonenzymatic browning of apple juice concentrates. *Food Chemistry*, 20: 91-97.
- Çandır, E. E., Temizyürek, F. and Özdemir, A. E. (2009). The effects of hot water dip treatments on the cold storage of Big Top nectarines. *Journal of Applied Botany and Food Quality*, 82: 136 – 140.
- Candrawinata, V.I., Golding, J.B., Roach, P.D. and Stathopoulos, C.E. (2014). Total phenolic content and antioxidant activity of apple pomace aqueous extract: Effect of time, temperature and water to pomace ratio. *International Food Research Journal*, 21(6): 2337-2344.
- Capanoglu, E., Beekwilder, J., Boyacioglu, D., Hall, R. and de vos, C.H. (2008). Changes in antioxidant and metabolite profiles during production tomato paste. *Journal of Agricultural and Food Chemistry*, 56(3): 964-973.
- Cedeño-Maldonado, A., & Bosques-Vega, A. (1990). A new method for rapid multiplication of cocoyam (*Xanthosoma* spp) vegetative material. *The Journal of Agriculture of the University of Puerto Rico*, 74(4), 465-465.
- Chen, I. N., Chang, C. C., Ng, C. C., Wang, C. Y., Shyu, Y. T. and Chang, T. L. (2008). Antioxidant and antimicrobial activity of zingiberaceous plants in Taiwan. *Plants Foods for Human Nutrition*, 63:15–20.
- Chiabrande, V., Giacalone, G., Rolle, L., 2009. Mechanical behaviour and quality traits of highbush blueberry during postharvest storage. *J. Sci. Food Agric.*, 89: 989–992.
- Chinwong, U., Siriphanich, J., Chairat, R. (2006). Quality of fresh-cut galangal harvested at different maturity stages.
- Choi, E.J., Lee, K.A., Kim, B.S. and Ku, K.H. (2012). Effect of pre-treatment and storage conditions on the quality characteristics of ginger paste. *Prev Nutr Food Sci.*,17(1):46-52.
- Chopsri, A., Sekozawa, Y. and Sugaya, S. (2018). Effects of hot air treatment on cell wall-degrading enzymes, pulp softening and ripening in bananas. *International Food Research Journal*, 25(5): 2195-2203.
- Chung, I. M., Ali, M. and Ahmad, A. (2005). Isolation and characterization of water-soluble new gingerlanosterol from *Zingiber officinale* rhizomes. *Asian Journal of Chemistry*, 17(3): 1915-1920.

- Cocci, E., Rocculi, S., Romani and Rosa, M. D., (2006). Changes in nutritional properties of minimally processed apples during storage. *Postharvest Biology and Technology*, 39: 265-271.
- Coyne, D. L., Claudius-Cole, AO., Kenyon, L. and Baimey, H. (2010). Differential effect of hot water treatment on whole tubers versus cut sets of yam (*Dioscorea spp.*). *Pest Management Science*, 66(4): 385-389.
- Couey, H. M. (1989). Heat treatment for control of postharvest diseases and insect pests of fruits. *HortScience*, 24: 198-202.
- Crifò, T., Puglisi, I., Petrone, G., Recupero, G.R. and Lo Piero, A.R. (2011). Expression analysis in response to low temperature stress in blood oranges: implication of the flavonoid biosynthetic pathway. *Gene*, 476(1-2):1-9.
- Crisoto, C. H., Scott, J. R., Luza-Juvenal, G. Crisoto, G.M. (1994). Irrigation regimes affect fruit soluble solid concentration and rate of water loss of O'henry peaches. *HortScience*, 29: 1169-1171.
- Cheyrier, V. and Fulcrand. H. (2003). Analysis of polymeric proanthocyanidins and complex polyphenols. In Santos Buelga, C., Williamson, G. (Eds.), *Methods in Polyphenol Analysis*, p. 284-313. Royal Society of Chemistry, Cambridge.
- Denniff, P., Macleod, I. and Whiting, D. A. (1981) Syntheses of the ( $\pm$ )-[n]-gingerols (pungent principles of ginger) and related compounds through regioselective aldol condensations: relative pungency assays. *Journal of the Chemical Society Perkin Transactions*: 1, 82–87.
- Department of Agriculture (DOA). (2016). Herbs and spices statistics. Putrajaya, Malaysia: Department of Agriculture.
- Diao, S., Jin, M., Sun, J., Zhou, Y., Ye C., Jin, Y., Zhou, W. and Li, Gao. (2019). A new diarylheptanpis and a new diarylheptanoid glycoside isolated from the roots of *Juglans mandshurica* and their anti-inflammatory activities. *Formerly Natural Product Letters*, 33(5): 701-707.
- Duan, W., Huang, Z., Song, X., Liu, T., Liu, H., Hou, X. and Li, Y. (2016). Comprehensive analysis of the polygalacturonase and pectin methylesterase genes in *Brassica rapa* shed light on their different evolutionary patterns. *Scientific Report*, 6, 1-14.
- Dunn, W.B., (2008). Current trends and future requirements or the mass investigation of microbial, mammalian and plant metabolomes. *Physical Biology*, 5: 24.
- Erkan, M., Pekmezci, M. and Wang, C. Y. (2005). Hot water and curing treatments reduce chilling injury and maintain post-harvest quality of 'Valencia' oranges. *International Journal of Food Science and Technology*, 40: 91–96.

- Eun-Jeong, C., Kyung-A, L. and Kyung-Hyung, K. (2012). Effect of pre-treatment and storage conditions on the quality characteristics of ginger paste. *Preventive Nutrition and Food Science*, 17(1): 46-52
- Fallik, W. (2004). Prestorage hot water treatments (immersion, rinsing, and brushing). *Postharvest Biology and Technology*, 32: 125-134.
- FAMA. (2017). Statistik utama pengeluaran FAMA. Retrieved on 1 March, 2019 from <http://www.fama.gov.my>.
- FAO. (2009). Declaration of the World Summit on Food Security, World Summit on Food Security, Rome, 16 – 18 November 2009, <http://www.fao.org/>.
- FAOSTAT. (2019). Ginger, production quantity (tons) – for all countries. Retrieved October 2019, from <http://www.factfish.com>.
- Ferrante, A. and T. Maggiore (2007). Chlorophyll fluorescence measurements to evaluate storage time and temperature of Valeriana leafy vegetables. *Postharvest Biology and Technology*, 45(1): 73-80.
- Ferreira, M. L. F., Rius, S. P. and Casati, P. (2012). Flavonoids: biosynthesis, biological functions, and biotechnological applications. *Frontiers in Plant Science*, 3: 222.
- Fiehn, O., Kind, T. and Barupal, D. K. (2018). 12 Data processing, metabolite databases and pathway analysis. *Biology of Plant Metabolomics*, 43:
- Fraser, P. D., Enfissi, E. M., Goodfellow, M., Eguchi, T. and Bramley, P.M. (2007). Metabolite profiling of plant carotenoids using the matrix-assisted laser desorption ionization time-of-flight mass spectrometry. *The Plant Journal*, 49(3): 552-564.
- Fukumoto, L. R., Toivonen, P. M. A. and Delaquis, P. J. (2002). Effect of wash water temperature and chlorination on phenolic metabolism and browning of stored Iceberg lettuce in photosynthetic and vascular tissues. *Journal Agricultural and Food Chemistry*, 50: 4503-4511.
- Gan, H., Charters, E., Driscoll, R. and Szrednicki, G. (2017). Effects of drying and blanching on the retention of bioactive compounds in ginger and turmeric. *Horticulturae*, 3(13): 1-9.
- Geysen, S., Verlinden, B. E. and Nicolai, B. M. (2005). Thermal treatments of fresh fruit and vegetables. Woodhead Publishing Series in Food Science, Technology and Nutrition, 429-453.
- Ghasemzadeh, A. and Ghasemzadeh, N. (2011). Flavonoids and phenolic acids: Role and biochemical activity in plants and human. *Journal of Medicinal Plants Research*, 5(31): 6697-6703.



- Ghasemzadeh, A., Ashkani, S., Baghdadi, A., Pazoki, A., Jaafar, H. Z. E. and Rahmat, A. (2016). Improvement in flavonoids and phenolic acids production and pharmaceutical quality of sweet basil (*Ocimum basilium* L.) by ultraviolet-B irradiation. *Molecules*, 21(9): 1-15.
- Gonzales-Aguilar, G. A., Gayosso, L., Cruz, R., Fortiz, J., Baez, R. and Wang, C. Y. (2000). Polyamines induced by hot water treatments reduce chilling injury and decay in pepper fruit. *Postharvest Biology and Technology*, 18: 19-26.
- González-Hernández, J., Rosales, L., Jaramillo, M., Alvarez-Navarrete, M., Vera Villa, J. Corona, R. and Chávez-Parga, Peña, A. (2012). Chemical Hydrolysis of the Polysaccharides of the Tamarind Seed. *Journal of the Mexican Chemical Society*, 56(4):395-401.
- Gould, W.P. and Sharp, J.L. (1992). Hot-water immersion quarantine treatment for guavas infested with Caribbean fruit fly (Diptera, Tephritidae). *Journal of Economy and Entomology*, 85: 1235-123.
- Gould, W. P. and McGuire, R. G. (2000). Hot water treatment and insecticidal coating for disinfesting lies of mealybugs (Homoptera: Pseudococcidae). *Journal of Economy and Entomology*, 93: 1017-1020.
- Guo, Y.H. and Zhang, Z.X. (2005). Establishment and plant regeneration of somatic embryogenic cell suspension cultures of the *Zingiber officinale* Rosc. *Scientia Horticulturae*, 107: 90-96.
- Grace, S. C. and Hudson, D. A. (2017). Processing and visualization of metabolomics data using R. pp. 67-91.
- Gupta, S. K. and Sharma, A. (2014). Medicinal properties of *Zingiber officinale* Roscoe - A Review. *Journal of Pharmacy and Biological Sciences*, 9(5): 124-129.
- Harian Metro. (2017). Potensi tinggi pasaran dalam, luar negara. Retrieved on 1 March, 2019 from <https://www.hmetro.com.my/agro/2017/09/265015/potensi-tinggi-pasaran-dalam-luar-negara>.
- Hassimotto, N.M.A., Genovese, M.I. and Lajolo, F.M. (2005). Antioxidant activity of dietary fruits, vegetables, and commercial frozen fruit pulps. *Journal of Agricultural and Food Chemistry*, 53: 2928–2935.
- Hazbavi, I., Khoshtaghaza, M.H., Mostaan, A., Banakar, A. (2015). Effect of postharvest hot-water and heat treatment on quality of date palm (cv. Stamaran). *Journal of the Saudi Society of Agricultural Sciences*, 14(2): 153-159.
- He, X. G., Bernart, M. W., Lian, L. Z., and Lin, L. Z. (1998). High-performance liquid chromatography-electrospray mass spectrometric analysis of pungent constituents of ginger. *Journal of Chromatography A*, 796:327-334.

- Hernanz, D., Gallo, V., Recamales, A. F., Meléndez-Martínez, A. J., González-Miret, M. L. and Heredia, F. J. (2009). Effect of storage on the phenolic content, volatile composition and colour of white wines from the varieties Zalema and Colombard. *Food Chemistry*, 113(2): 530-537.
- Herppich, W. B., Harold, B., Geyer, M. and Gomez, F.(2004). Effects of temperature and water relations on carrots and radish tuber texture. *Journal of Applied Botany*, 78: 11-17.
- Hii, C.L., Law, C.L., Suzannah, S. and Cloke, M. (2009). Polyphenols in cocoa (*Theobroma cacao* L.). *Asian Journal of Food and Agro-Industry*, 2: 702-722.
- Hong, S. S. and Oh, J. S. (2012). Phenylpropanoid ester from *Zingiber officinale* and their inhibitory effects on the production of nitric oxide. *Archives of Pharmacal Research*, 35(2):315-320.
- Iijima, Y. and Joh, A. (2014). Pigment composition responsible for the pale-yellow color of ginger (*Zingiber officinale*) rhizomes. *Food Science and Technology Research*, 20 (5): 971-978.
- Imm, J., Zhang, G., Chan, L-K., Nitteranon, V. and Parkin, K. L. (2010). [6]-Dehydroshogaol, a minor component in ginger rhizome, exhibits quinone reductase inducing and anti-inflammatory activities that rival those of curcumin. *Food Research International*. 43:2208-2213.
- IndexBox. (2021). Malaysia: Ginger Market Overview 2021 – IndexBox. Retrieved on 21 August, 2021 from <https://www.indexbox.io/>.
- Intorasoot, A., Chornchoem, P., Sookkhee, S. and Intorasoot, S. (2017). Bactericidal activity of herbal volatile oil extracts against multidrug-resistant *Acinetobacter baumannii*. *Journal of Intercult Ethnopharmacology*, 6(2):218–222.
- Ioannou, I. and Ghoul, M. (2013). Prevention of enzymatic browning in fruit and vegetables. *European Scientific Journal*, 9, 57-81.
- Ioku, K., Aoyama, Y., Tokuno, A., Terao, J., Nakatani, N. and Takei, Y. (2001). Various cooking methods and the flavonoid content in onion. *J. Nutr. Sci. Vitaminol.*, 47, 78-83.
- Jackman, R., Gibson, H. J. and Stanley, D. W. (2006). Effects of chilling on tomato fruit texture. *Physiologia Plantarum*, 86(4), 600-608.
- Janick, J. (2013). *Horticultural Reviews*, 41.
- Ji Soo, H., Sunmin, L., Hyang Yeon, K. and Choong Hwan, L. (2015). MS-based metabolite profiling of aboveground and root components of *Zingiber mioga* and *officinale*. *Molecules*, 20: 16170-16185.

- Jiang, H., Sólyom, A.M., Timmermann, B.N. and Gang, D.R. (2005). Characterization of gingerol-related compounds in ginger rhizome (*Zingiber officinale* Roscoe) by high-performance liquid chromatography / electrospray ionization mass spectrometry. *Rapid communications in mass spectrometry: RCM*, 19(20): 2957-2964.
- Jimenez-Monreal, A.M., Garcí'a-Diz, L., Martí'nez-Tomé, M., Mariscal, M., Murcia, M. A. (2009). Influence of cooking methods on antioxidant activity of vegetables. *Journal of Food Science*, 74: 97-103.
- Jolad, S. D., Lantz, R. C., Chen, G. J., Bates, R. B. and Timmermann, B. N. (2005). Commercially processed dry ginger (*Zingiber officinale*): composition and effects on LPS-stimulated PGE2 production. *Phytochemistry*, 65:1937-1954.
- Jouyban Z., Hasanzade R. and Sharafi S. (2013): Chilling stress in plants. *International Journal of Agriculture and Crop Sciences*, 5: 2961–2968.
- Jung, S. K. and Watkins, C. B. (2011). Involvement of ethylene in browning development of controlled atmosphere-stored 'Empire' apple fruit. *Postharvest Biology and Technology*, 59: 219–226.
- Jun, Z., Aa Jiye, Guang-ji, W., Feng-yi, Z., Rong-rong, G., Xin-wen, W., Chun-yan, Z., Meng-jie, L., Jian, S., Bei, C., Tian, Z., Lin-sheng, L., Sheng G. and Jin-ao, D. (2011). Visualization of multivariate metabolomic data. *Chinese Herbal Medicines*, 3(4): 289-293.
- Junot, C., Fenaille, F., Colsch, B. and Bécher, F. (2014). High resolution mass spectrometry-based techniques at the crossroads of metabolic pathways. *Mass Spectrometry Review*, 33: 471-500.
- Kader, A. (2008). Flavor quality of fruits and vegetables. *Journal of the Science of Food and Agriculture*, 88 (2008): 1863-1868.
- Kang, H. M. and Saltveit, M. (2001). Enzymatic antioxidant defense systems in chilled and heat shocked cucumber seedlings radicles. *Physiologia Plantarum*, 113(4): 548-556.
- Kang, H. M. and Saltveit, M. E. (2002a). Reduced chilling tolerance in elongating cucumber seedling radicles is related to their reduced antioxidant enzyme and DPPH-radical scavenging activity. *Physiologia Plantarum*, 115: 244-250.
- Kang, H. M. and Saltveit, M. E. (2002b). Effect of chilling on antioxidant enzymes and DPPH-radical scavenging activity of high- and low-vigour cucumber seedling radicles. *Plant, Cell and Environment*, 25: 1233–1238.
- Karna, P., Chagani, S., Gundala, S. R., Rida, P. C., Asif, G., Sharma, V., Gupta, M. V. and Aneja, R. (2012). Benefits of whole ginger extract in prostate cancer. *British Journal of Nutrition*, 107:473–484.

- Kasamo K., Yamaguchi M. and Nakamura Y. (2000). Mechanism of the chilling-induced decrease in proton pumping across the tonoplast of rice cells. *Plant and Cell Physiology*, 41(7): 840–849.
- Katajama, M. and Data, O. M. (2007). Data processing for mass spectrometry-based metabolomics. *Journal of Chromatography*, 1158: 318-328.
- Kays, S. J. and Paull, R.E. (2004). *Postharvest Biology*. Exon Press, Athens, GA.
- Ketsa, S., Chidtragool, S., Klein, J. D. and Lurie, S. (1997). Effect of heat treatment on changes in softening, pectic substances and activities of polygalacturonase, pectinesterase and  $\beta$ -galactosidase of ripening mango. *Journal of Plant Physiology*, 153: 457-461.
- Key, M. (2012). A tutorial in displaying mass spectrometry based proteomic data using heatmaps. *BMC Bioinformatics*, 13 (Suppl 16): S10.
- Khademi, O., Zamani, Z., Kalantari, S., Ebadi, M. and Sepahvand, E. (2013). Hot water treatment maintains postharvest quality of 'Karaj' persimmon fruit during cold storage. *Acta Horticulturae*, 1012: 485-489.
- Khalil, S.A., Ayub, M., Zamir, R., Sajid, M., Muhammad, A., Wahid, F. and Faiq, M. (2012). Influence of postharvest hot water dip treatment on quality of peach fruit (*Prunus persica* L.). *Journal of Medicinal Plants Research*, 6(1): 108-113.
- Kikuzaki, H., and Nakatan, N. (1996). Cyclic diarylheptanoids from rhizomes of *Zingiber officinale*. *Phytochemistry*, 43(1):273-277.
- Kirana, C, McIntosh, G. H., Record, I. R., Jones, G. P. (2003). Antitumor activity of extract of *Zingiber aromaticum* and its bioactive sesquiterpenoid zerumbone. *Nutrition Cancer*, 45:218–225.
- Kontunen-Soppela, S., Lankila, J., Lahdesmaki, P. and Laine, Kari. (2002). Response of protein and carbohydrate metabolism of Scot pine seedling to low temperature. *Journal of Plant Physiology*, 159(2): 175-180.
- Kou, X., Ke, Y., Wang, X., Rahman, M. R. T., Xie, Y., Chen, S. and Wang, H. (2018). Simultaneous extraction of hydrophobic and hydrophilic bioactive compounds from ginger (*Zingiber officinale* Roscoe). *Food Chemistry*, 257: 223–229.
- Kratsch, H. and Wise, R. R. (2000). The ultrastructure of chilling stress. *Plant Cell and Environment*, 23(4): 337-350.
- Kühlbrandt, W. (2015). Structure and function of mitochondrial membrane protein complexes. *BMC Biol.*, 13: 89.
- Kumar, G., Kathie, L. and Rao, K. V. B. (2011). A review on pharmacological and phytochemical properties of *Zingiber officinale* Roscoe (Zingiberaceae), *Journal of Pharmacy Research*, 4(9): 2963-2966.

- Kusano, M., Fukushima, A., Redestig, H. and Saito, K. (2011). Metabolomic approaches toward understanding nitrogen metabolism in plants. *Journal of Experimental Botany*, 62: 1439-1453.
- Lan, W. Zhang, H., Chen, X. and Yang, F. (2016). Gelatinization and decrystallization of cellulose by newly isolated *Arthrobotrys* sp. CX1 to facilitate cellulose degradability. *Cellulose*, 23(6): 3543-3554.
- Larsen, K., Ibrahim, H., Khaw, S.H. and Saw, L.G. (1999). Gingers of Peninsular Malaysia and Singapore. Kota Kinabalu. *Nat. Hist. Publ. (Borneo)* p.135.
- Lee, S. W., Lim, J. H., Kim, M. S., Jeong, J. H., Song, G. Y., Lee, W. S. and Rho, M. C. (2011). Phenolic compounds isolated from *Zingiber officinale* roots inhibit cell adhesion. *Food Chemistry*, 128: 778-782.
- Lei, Z., Li, H., Chang, J., Zhao, P. X. and Sumner, L. W. (2012). MET-IDEA version 2.06; improved efficiency and additional functions for mass spectrometry-based metabolomics data processing. *Metabolomics*, 8: 105-110.
- Leonel, M., Suman, P. A. and Garcia, E. L. (2015). Production of ginger vinegar. *Ciência e Agrotecnologia*, 39(2): 183-190.
- Li, F., Wang, Y., Parkin, K. L., Nitteranon, V., Liang, J., Yang, W., Li, Y., Zhang, G. and Hu, Q. (2011). Isolation of quinone reductase (QR) inducing agents from ginger rhizome and their in vitro anti-inflammatory activity. *Food Research International*, 44:1597–1603.
- Lin, R. J., Chen, C. Y., Chung, L. Y. and Yen, C. M. (2010). Larvicidal activities of ginger (*Zingiber officinale*) against *Angiostrongylus cantonensis*. *Acta Tropica*, 115: 69-76.
- Loaiza-Velarde, J.G. and Saltveit, M.E. (2001). Heat shocks applied either before or after wounding reduce browning of lettuce leaf tissue. *Journal of American Social and Horticultural Science*, 126: 227 - 234.
- Luo, Z., Li, D., Xie, J., Feng, S. and Wang, Y. (2014). Effects of heat treatment on quality and browning of fresh-cut sugarcane. *Journal of Food Processing and Preservation*, 39: 688-696.
- Lurie, S. (1998). Review: Postharvest heat treatments. *Postharvest Biology and Technology*, 14: 257-269.
- Lurie, S. and Pedreschi, R. (2014). Review: Fundamental aspects of postharvest heat treatments. *Horticulture Research*, 30: 1- 7.
- Lyons, J. M. (1973). Chilling injury in plants. *Annual Review of Plant Biology*, 24: 445–466

- Ma, X.H., Ying Ma, Tang, J.F., He, Y.L., Liu, Y.C, Ma, X.J., Shen, Y. Cui, G. H., H. X. Lin, Rong, Q.X., Guo, J. and Huang, L.Q. (2015). The biosynthetic pathways of tanshinones and phenolic acids in *Salvia miltiorrhiza*. *Molecules*, 20: 16235-16254.
- Maizura, M., Aminah, A. and Wan Aida, W. M. (2011). Total phenolic content and antioxidant activity of kesum (*Polygonum minus*), ginger (*Zingiber officinale*) and turmeric (*Curcuma longa*) extract. *International Food Research Journal*, 18: 529-534.
- Makeredza, B., Marais, H., Schmeisser, M. and Lotze, E. (2015). Ripening associated red color development masks sunburn browning in apple peel. *HortScience: a publication of the American Society for Horticultural Science*, 50(6):814-81.
- Malakou, A. and Nanos, G.D., (2005). A combination of hot water treatment and modified atmosphere packaging maintains quality of advanced maturity 'Caldesi 2000' nectarines and 'Royal Glory' peaches. *Postharvest Biology Technology*, 38: 106-114.
- Martin-Diana, A. B., Rico, D., Barry-Ryan, C., Mulcahy, J., Frias, J. and Henehan, G. T. M. (2005). Effect of heat shock on browning-related enzymes in minimal processed iceberg lettuce and crude extracts. *Bioscience Biotechnology and Biochemistry*, 69(9): 1677-1685.
- Maskan, M. (2001). Kinetics of colour change of kiwifruits during hot air and microwave drying. *Journal of Food Engineering*, 48: 169-175.
- Mama, S., Yemer, J. and Woelore, W. (2016). Effect of hot water treatments on shelf life of tomato (*Lycopersicon esculantum* Mill). *Journal of Natural Sciences Research*, 6(7): 69-77.
- Manach, C., Scalbert, A., Morand, C., Rémésy, C. and Jiménez, L. (2004). Polyphenols: food sources and bioavailability. *American Journal of Clinical Nutrition*, 79:727-747.
- Mangprayool, T., Kupittayanant, S. and Chudapongse, N. (2013). Participation of citral in the bronchodilatory effect of ginger oil and possible mechanism of action. *Fitoterapia*, 89:68–73
- Mansor, P., Muhamad Ghawas, M. and Sentoor, K. (2005). Halia (*Zingiber officinale* Ross). In: Musa, Y., Muhammad Ghawas, M., Mansor, P. Penanaman tubuhan ubatan dan beraroma. Serdang: MARDI. p 29-35.
- Manzocco, L., Calligaris, S., Mastrocola, D., Nicoli, M.C. and Lericci, C.R. (2001). Review of non-enzymatic browning and antioxidant capacity in processed foods. *Trends Food Science Technology*, 11:340-346.
- Masuda Y, Kikuzaki H, Hisamoto M, Nakatani N. (2004). Antioxidant properties of gingerol related compounds from ginger. *Biofactors*, 21:293–296.

- Mazlan, N. W., Tate, R., Yusoff, Y. M., Clements, C. and Edrada-Ebel, R. (2019). Metabolomics-guided isolation of anti-trypanosomal compounds from endophytic fungi of the mangrove plant *Avicennia lanata*. *Current Medicinal Chemistry*, 26: 1-20.
- McGuire, R.G. (1992) Reporting of Objective Color Measurements. *HortScience*, 27, 1254-1255.
- Mesomo, M. C., Corazza, M. L., Ndiaye, P. M., Dalla Santa, O. R., Cardozo, L., and Scheer, A. P. (2013). Supercritical CO<sub>2</sub> extracts and essential oil of ginger (*Zingiber officinale* R.): Chemical composition and antibacterial activity. *Journal of Supercritical Fluids*, 80(Supplement C):44–49.
- Minolta, K. (2007). Precise color communication. Konica Minolta Sensing. Inc. Japan.
- Mirdehghan, S. H., Rahemi, M., Mart´inez-Romero, D., Guillen, F., Valverde, J. M., Zapata, P. J., Serrano, M. and Valero, D.(2007). Reduction of pomegranate chilling injury during storage after heat treatment: Role of polyamines. *Postharvest Biology and Technology*, 44: 19–25.
- Mittler, R., Finka, A. and Goloubinoff, P. (2012). How do plants feel the heat? *Trends in Biochemical Science*, 37, 118–125.
- Mohammadian, M. A., Mobrami, Z. and Sajedi, R. H. (2011). Bioactive compounds and antioxidant capacities in the flavedo tissue of two citrus cultivars under low temperature. *Brazilian Journal of Plant Physiology*, 23(3): 1677-9452.
- Mohd, Y. S., Manas, M. A., Sidik, N. J., Ahmad, R. and Yaacob, A. (2015). Effects of organic substrates on growth and yield of ginger cultivated using soilless culture. *Malaysian Applied Biology*, 44(3): 63–68.
- Mohd Ismail, A. and Abd. Shukor, A R. (1990). Pengendalian halia untuk eksport ke Eropah. Kuala Lumpur: Institut Penyelidikan dan Kemajuan Pertanian Malaysia (MARDI).
- Mu, X., Wang, P., Du, J., Gao, Y. G. and Zhang, J. (2018). Comparison of fruit organic acids and metabolism-related gene expression between *Cerasus humilis* (Bge.) Sok and *Cerasus glandulosa* (Thunb.) Lois. *PLoS One*, 13(4):1-14.
- Murata, T., Shinohara, M. and Miyamoto, M. (1972). Isolation of hexahydrocurcumin, dihydrogingerol and two additional pungent principles from ginger. *Chemical and Pharmaceutical Bulletin*, 20(10): 2291-2292.
- Mukherjee, P. K. (2019). High-performance liquid chromatography for analysis of herbal drugs. *Quality Control and Evaluation of Herbal Drugs*, 421-458.

- Mustafa, K. and Ghalem, S. A. (2007). Effect of heat treatment on polyphenol oxidase and peroxidase activities in Algerian stored dates. *African Journal of Biotechnology*, 6(6): 790-794.
- Nägele, T. and Heyer, A. (2013). Approximating subcellular organisation of carbohydrate metabolism during cold acclimation in different natural accessions of *Arabidopsis thaliana*. *New Phytologist*, 198(3):
- Nakabayashi, R., Mori, T. and Saito, K. (2014). Alternation of flavonoid accumulation under drought stress in *Arabidopsis thaliana*. *Plant Signal and Behavior*, 9(8): 1-3.
- National Agricultural Research Institute. 2004. Ginger: Postharvest care and market preparation. Postharvest Handling Technical Bulletin. Retrieved 18 August 2014 from pdf.usaid.gov. pp.1-10.
- Noh, S. A., Park, S. H., Huh, G. H., Paek, K. H., Shin, J. S. and Bae, J. M. (2009). Growth retardation and differential regulation of expansin genes in chilling-stressed sweet potato. *Plant Biotechnology Report*, 3:75–85.
- Novacky, A. (1991). The Plant Membrane and Its Response to Disease. In: Cole G.T., Hoch H.C. (eds) *The Fungal Spore and Disease Initiation in Plants and Animals*. Springer, Boston, MA.
- Ok, S. and Woo-Sik, J. (2012). Optimization of extraction conditions for the 6-shogaol-rich extract from ginger (*Zingiber officinale* Roscoe). *Preventive Nutrition and Food Science*, 17(2): 166-171.
- Pan, M.H., Hsieh, M. C., Kuo, J. M., Lai, C. S., Wu, H., Sang, S. and Ho, C.T. (2008). [6]-Shogaol induces apoptosis in human colorectal carcinoma cells via ROS production, caspase activation, and GADD 153 expression. *Molecular Nutrition Food Research*, 52:527–37.
- Panche, A. N., Diwan, A. D. and Chandra, S.R. (2016). Flavonoids: an overview. *J. Nutr. Sci.*, 5: 1-15.
- Paniagua, A. C., East, A. R., Hindmarsh, J. P. and Heyes, J. A. (2013). Moisture loss is the major cause of firmness change during postharvest storage of blueberry. *Postharvest Biology and Technology*, 79: 13-19.
- Park, E.J. and Pizzuto, J.M. (2002). Botanicals in cancer chemoprevention. *Cancer Metastasis Review*, 21: 231-255.
- Park, J. S. and Jung, M. Y. (2012). Development of high-performance liquid chromatography-time-of-flight mass spectrometry for the simultaneous characterization and quantitative analysis of gingerol-related compounds in ginger products. *Journal of Agricultural and Food Chemistry*, 60(40):10015-26.
- Paull, R. E. (1990). Chilling injury of crops of tropical and subtropical origin. In: C. Y. Wang (ed), *Chiling injury of Horticultural Crops*. CRC Press, Boca Raton, FL.



- Pereira, D. M., Valentão, P., Pereira, J. A. and Andrade, P. B. (2009). Phenolics: From chemistry to biology. *Molecules*, 14: 2202-2211.
- Pluskal, T., Korf, A., Smirnov, A. and Schmid, R. (2020). Chapter 7: Metabolomic data analysis using MZmine. Processing Metabolomics and Proteomics Data with Open Software. Royal Society of Chemistry. Pp 232-254.
- Policegoudra, R. S., Divakar, S. and Aradhya, S. M. (2007). Identification of difurocumenonol, a new antimicrobial compound from mango ginger (*Curcuma amada* Roxb.) rhizome. *Journal of Applied Microbiology*, 102(6): 1594-1602.
- Prochazkova, D., Bousova, I., and Wilhelmova, N. (2011). Antioxidant and prooxidant properties of flavonoids. *Fitoterapia*, 82(4):513-23.
- Provart, N. J., Gil, P., Chen, W., Han, B., Chang, H. S., Wang, X., and Zhu, T. (2003). Gene expression phenotypes of *Arabidopsis* associated with sensitivity to low temperatures. *Plant Physiology*, 132: 893-906.
- Proulx, E., Yagiz, Y., Cecilia, M., Nunes, N., and Emond, J. P. (2010). Quality attributes limiting snap bean (*Phaseolus vulgaris* L.) postharvest life at chilling and non-chilling temperatures. *HortScience*, 45(8): 1238-1249.
- Queiroz, C., Lopes, M. L. M., Fialho, E. and Valente-Mesquita, V. L. (2008). Polyphenol oxidase: characteristics and mechanisms of browning control. *Food Reviews International*, 24(4):361-375.
- Rahmani, A. H., Al-Shabrmi, F. M. and Aly, S. M. (2014). Active ingredients of ginger as potential candidates in the prevention and treatment of diseases via modulation of biological activities. *Int. J. Physiol Pathophysiol Pharmacol.* 2014; 6(2): 125–136.
- Ramirez-Ahumada M, del C., Timmermann, B.N. and Gang, D.R. (2006). Biosynthesis of curcuminoids and gingerols in turmeric (*Curcuma longa*) and ginger (*Zingiber officinale*): identification of curcuminoid synthase and hydroxycinnamoyl-CoA thioesterases. *Phytochemistry*, 67(18): 2017-2029
- Rasouli, M. and Khademi, O. (2014). Extending postharvest life of Karaj persimmon by hot water and 1-MCP treatments. *International Journal of Biosciences*, 4 (3): 31-38.
- Rahuman, A. A., Gopalakrishnan, G., Venkatesan, P., Geetha, K. and Bagavan, A. (2008). Mosquito larvicidal activity of isolated compounds from the rhizome of *Zingiber officinale*. *Phytotherapy Research*, 22(8): 1035-1039.
- Reddy, A.A. and Lokesh, B.R. (1992). Studies on spice principles as antioxidants in the inhibition of lipid peroxidation of rat liver microsomes. *Molecular and Cellular Biochemistry*, 111: 117–124.

- Robinson, P. K. (2015). Enzymes: principles and biotechnological applications. *Essays Biochem*, 59: 1-41
- Rocha, A. and Morais, A.M.M.B. (2002). Polyphenoloxidase activity and total phenolic content as related to browning of minimally process 'Jonagored'apple. *Journal of the Science of Food and Agriculture*,82(1): 120 – 126.
- Rodríguez, M., Canales, E. and Borrás-Hidalgo, O. (2005). Molecular aspects of abiotic stress in plants. *Biotechnology Appl.*, 22: 1–10.
- Saccetti, E., Hoefsloot, H., Smilde, A. K., Westerhuis, J. A. and Hendriks, M. M. (2014). Reflections on univariate and multivariate analysis of metabolomics data. *Metabolomics*, 10: 361 – 374.
- Sacks, E. J. and Francis, D. M. (2001). Genetic and environmental variation for tomato flesh color in a population of modern breeding lines. *Journal of the American Society for Horticultural Science*, 126(2): 221-226.
- 126(2), 221–226
- Sala and Lafuente, (2000). Catalase enzyme activity is related to tolerance of mandarin fruits to chilling. *Postharvest Biology and Technology*, 20: 81–89.
- Sanwal, S. K., Rai, N., Singh, J. and Buragohain, J. (2010). Antioxidant phytochemicals and gingerol content in diploid and tetraploid clones of ginger (*Zingiber officinale* Roscoe). *Scientia Horticulturae*, 124: 280-285.
- Scheibe R. (2004). Malate valves to balance cellular energy supply. *Physiologia Plantarum*, 120: 21–26.
- Schreiner, M., Krumbein, A., Schonhof, I. and Huyskens-Keil, S. (2003). Quality determination of red radish by nondestructive root color measurement. *Journal of the American Society for Horticultural Science*, 128 (3): 397-403.
- Scholz, M. and Selbig, J. (2007). Visualization and analysis of molecular data. In: Weckwerth W. *Metabolomics: methods and protocols. Methods in Molecular Biology*, 358: 87-104.
- Seid, A., Tasew, D. and Tsedaley, B. (2017). Effect of hot water treatment on development of anthracnose (*Colletotrichum gloeosporioides*) and quality of mango fruit at Jimma southwest Ethiopia. *Journal of Archives of Phytopathology and Plant Protection*, 50 (7-8): 303-316.
- Semwal, R. B., Semwal, D.K., Combrinck, S. and Viljoen, A. M. (2015). Gingerols and shogaols: important nutraceutical principles from ginger. *Phytochemistry*, 117:554–568.

- Sharma, G. (2014). The CIEDE2000 Color-Difference Formula: Implementation Notes, Supplementary Test Data, and Mathematical Observations. Retrieved 20 February 2020, from <http://www.ece.rochester.edu/~gsharma/ciede2000/ciede2000not eCRNA.pdf>
- Sharma, K., Young Kou, E., Assefa, A. D., Haa, S., Nile, S. H., Tai Lee, E. and Won Park, S. (2015). Temperature-dependent studies on the total phenolics, flavonoids, antioxidant activities, and sugar content in six onion varieties. *Journal of Food and Drug Analysis*, 23 (2):243-252.
- Sharma, K. Sarma, S., Bohra, A., Mitra, A., Sharma, N. K. and Kumar, A. (2018). Plant metabolomics: An emerging technology for crop improvement. In book: Plant Science (Ed. Özge Çelik). IntechOpen, UK. pp: 65-7
- Sharma, K. and Sahai, M. (2018). Chemical constituents of *Zingiber officinale* rhizome. *Journal of Medicinal Plants Studies*; 6(1): 146-149.
- Sharp, J. L. (1990). Immersion in heated water as a quarantine treatment for California stone fruits infested with the Caribbean fruit fly (Diptera, Tephritidae). *Journal of Economy Entomology*, 83: 1468-1470.
- Shellie, K.C. and Mangan, R.L. (2000). Postharvest disinfestation heat treatments: response of fruit and fruit fly larvae to different heating media. *Postharvest Biology Technology*, 21: 51-60.
- Shen, Y, Zhong, L., Sun, Y., Chen, J., Liu, D. and Ye, X. (2013). Influence of hot water dip on fruit quality, phenolic compounds, and antioxidant capacity of Satsuma mandarin during storage. *Food Science Technology Int.*, 19(6): 511-21.
- Shewfelt, R. L., and del Rosario, B. A. (2000). The role of lipid peroxidation in the mechanism of membrane-associated disorders of fresh fruits and vegetable. *HortScience*, 35: 575-579.
- Shukla, Y. and Singh, M. (2007). Cancer preventive properties of ginger: a brief review. *Food Chemical and Toxicology*, 45(5): 683-690.
- Shukor, N. I. A., Misran, A., Ahmad, S. H., Xue, Y. T., Mahmud, T.M.M. and Saari, N. (2020). The changes in chemical quality of ginger during postharvest storage at chilling temperature. *Food Research*, 4(5): 1653-1662.
- Shi, Y., Jiang, L., Zhang, L., Kang, R. and Yu, Z. (2014). Dynamic changes in proteins during apple (*Malus x domestica*) fruit ripening and storage. *Horticulture Research*, 1(6): 1-21.
- Singh, G., Kapoor, I. P., Singh, P., de Heluani, C. S., de Lampasona, M. P. and Catalan, C. A. (2008). Chemistry, antioxidant and antimicrobial investigations on essential oil and oleoresins of *Zingiber officinale*. *Food. Chem. Toxicol.* 46: 3295-302.

- Singh, V., Guizani, N., Essa, M.M., Hakkim, F.L. and Rahman, M.S. (2012). Comparative analysis of total phenolics, flavonoid content and antioxidant profile of different date varieties (*Phoenix dactylifera* L.) from Sultanate of Oman. *International Food Research Journal*, 9: 1063-1070.
- Singla, R., Rattanpal, H. S. and Singh, G. (2018). Storage performance of hot water treated kinnow fruits under ambient conditions. *International Journal of Current Microbiology and Applied Sciences*, 7(6): 3775-3782.
- Singldinger B., Dunkel A., and Hofmann, T. (2017). The cyclic diarylheptanoid asadanin as the main contributor to the bitter off-taste in hazelnuts (*Corylus avellane* L.). *J. Agric. Food Chem.*, 65(8):1677–1683.
- Smith, C. A., Want, E. J., O'Maille, G., Abagyan, R. and Siuzdak, G. (2006). XCMS: processing mass spectrometry data for metabolite profiling using nonlinear peak alignment, matching, and identification. *Anal Chem*, 78(3):779-87.
- Strauss, A. J. Kruger, G., Strasser, R. J. and van Heerden, P. D. R. (2007). The role of low soil temperature in the inhibition of growth and PSII function during dark chilling in soybean genotypes of contrasting tolerance. *Physiologia Plantarum*, 131(1):89-105.
- Soo Han, J., Lee, S., Yeon Kim, H. and Hwan Lee, C. (2015). MS-Based metabolite profiling of aboveground and root components of *Zingiber mioga* and *officinale*. *Molecules*, 20(9), 16170-16185.
- Suresh, K., Manoharan, S., Vijayaanand, M. A. and Sugunadevi, G. (2010). Chemopreventive and antioxidant efficacy of (6)-paradol in 7,12-dimethylbenz(a)anthracene induced hamster buccal pouch carcinogenesis. *Pharmacology Reports*, 62:1178–85.
- Suzuki, N. and Mittler, R. (2006). Reactive oxygen species and temperature stresses: A delicate balance between signaling and destruction. *Physiologia Plantarum*, 126: 45–51.
- Tanaka, K., Arita, M., Sakurai, H, Ono, N. and Tezuka, Y. (2015). Analysis of chemical properties of edible and medicinal ginger by metabolomics approach. *Biomed Res Int*. 2015: 1-7.
- Tang, Y., Cai, W. and Xu, B. (2015). Profiles of phenolics, carotenoids and antioxidative capacities of thermal processed white, yellow, orange and purple sweet potatoes grown in Guilin, China. *Food Science and Human Wellness*, 4(3): 123-132.
- Tadesse, T. N. and Abteu, W. G. (2016). Effect of hot water treatment on reduction of chilling injury and keeping quality in tomato (*Solanum lycopersicum* L.) fruits. *Journal of Stored Products and Postharvest Research*, 7(7): 61-68.

- Toivonen, P. M. A. and Brummell, D. A. (2008). Biochemical bases of appearance and texture changes in fresh-cut fruit and vegetables. *Postharvest Biology and Technology*, 48(1), 1-14.
- Toor, R.K. and G.P. Savage. (2006). Changes in major antioxidant components of tomatoes during post-harvest storage. *Food Chemistry*, 99, 724-727.
- Trygg, J. and Lundstedt, T. (2007). Chemometrics techniques for metabolomics. In J. C. Lindon, J. K. Nicholson, and E. Holmes (Eds.), *The Handbook of Metabonomics and Metabolomics* (pp. 171 – 199). Amsterdam, The Netherlands, Elsevier B. V.
- Tsouvaltzis, P., Deltsidis, A. and Brecht, J. K. (2011). Hot water treatment and pre-processing storage reduce browning development in fresh-cut potato slices. *HortScience*, 46(9): 1282 – 11286.
- Uarotta, V. G. and Maraschin, M. (2015). Metabolomic, enzymatic, and histochemical analysis of cassava roots during postharvest physiological deterioration. *BMC Research Notes*, 648.
- Ullah J. (2009). Storage of fresh tomatoes to determine the level of coating and optimum temperature for extended shelf life. Post-Doctoral Fellowship Report. Department of Food Science and Technology, NWFP Agricultural University Peshawar, Pakistan. 43p.
- Ullah, F., Sajid, M., Gul, S. L., Zainub, B. and Khan, M. (2018). Influence of hot water treatments on the storage life of sweet orange cv. Sherkana-1. *Sarhad Journal of Agriculture*, 34(1): 220-224.
- Vairappan, C. S., Jit Beng, O., Nagappan, T., Gobilik, J., and Ramachandram, T. (2012). Essential oil profiles of major populations *Zingiber officinale* Rosc. utilized in Malaysia for traditional medicine. *Journal of Tropical Biology and Conservation*, 9(2): 206-212.
- Verlinden, S., Silva, S.M., Herner, R. C. and Beaudry, R.M. (2014). Time-dependent changes in the longitudinal sugar and respiratory profiles of asparagus spears during storage at 0 °C. *Journal of American Society for Horticultural Science*, 139(4): 339–348.
- Vicente, A. R., Costa, M. L. Martinez, G.A. and Chaves, A.R. (2005). Effect of heat treatments on cell wall degradation and softening in strawberry fruits. *Postharvest Biology and Technology*, 38(3): 213-222.
- Vinãa, S.Z. and Chaves, A.R. (2006). Antioxidant responses in minimally processed celery during refrigerated storage. *Food Chemistry*, 94, 68–74.
- Wall, M.W., Sivakumar, D., and Korsten, L. (2011). Rambutan (*Nephelium lappaceum* L.). In E. M. Yahia (Eds.) *Postharvest Biology and technology of tropical and subtropical fruits* (pp 312-333). Woodhead Publishing Series in Food Science, Technology and Nutrition, Foreword.

- Wang, C. Y. (1994). Chilling Injury of Tropical Horticultural Commodities. *Hortscience*, 29(9): 986-988.
- Wang, W., Vinocur, B., Shoseyov, O. and Altman, A. (2003). Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. *Planta*, 29: 1-14.
- Wang, N., Fang, W., Han, H., Sui, N., Li B., and Meng, Q. W. (2008) Overexpression of zeaxanthin epoxidase gene enhances the sensitivity of tomato PSII photoinhibition to high light and chilling stress. *Physiologia Plantarum*, 132(3): 384–396.
- Wang, C. Y. (2010). Alleviation of chilling injury in tropical and subtropical fruits. *Acta Horticulturae*, 864: 267-273.
- Wang, C.Y. (2013). Managing chilling injury in vegetables. Proceeding 7th International Postharvest Symposium.
- Wang, Q., Nie, X. and Cantwell, M. (2014). Hot water and ethanol treatments can effectively inhibit the discoloration of fresh-cut sunchoke (*Helianthus tuberosus* L.) tubers. *Postharvest Biology and Technology*, 94: 49-57.
- Wang, X., Kong, D., Ma, Z. and Zhao, R. (2015). Effect of carrot puree edible films on quality preservation of fresh-cut carrots. *Irish Journal of Agricultural and Food Research*, 54(1): 64-71.
- Wang, S., Alseekh, S., Fernie, A. R. and Luo, J. (2019). The structure and function of major plant metabolite modifications. *Molecular Plant*, 12(7): 899-919.
- Wang, Y., Naber, M. R. and Crosby, T. W. (2020). Effects of wound healing management on potato post-harvest storability. *Agronomy*, 10(512): 1-17.
- Wilkinson, L. and Friendly, M. (2009) The history of the cluster heat map. *The American Statistician*, 63(2):179-184.
- Williamson, E. M. (2002). Major herbs of Ayurveda. Churchill Livingstone.
- Wills, R.B.H., McGlasson, W.B., Graham, D and Joyce, D.C. (2007). Postharvest: An introduction to the physiology and handling of fruit, vegetables and ornamentals. 5<sup>th</sup> ed. University of New South Wales Press.
- Winkel-Shirley, B. (2001). Flavonoid biosynthesis. A colorful model for genetics, biochemistry, cell biology, and biotechnology. *Plant Physiology*, 126(2): 485-93.
- Wisal, S., Ullah, J., Zeb, A. and Khan, M. Z. (2013). Effect of refrigeration temperature, sugar concentrations and different chemicals preservatives on the storage stability of strawberry juice. *International Journal of Engineering and Technology*, 13(2): 160-168.

- Wismer, V. W. (2003). Low temperature as a causative agent of oxidative stress in postharvest crops. In: Hodges, D. M. (ed). Postharvest oxidative stress in horticultural crops. The Haworth Press, Inc CRC Press. Binghamton, NY.
- Wohlmuth, H., (2008). Phytochemistry and pharmacology of plants from the ginger family, Zingiberaceae (PhD thesis). Southern Cross University, Lismore, NSW.
- Wojakowska, A., Chekan, M., Widlak, P. and Pietrowska, M. (2015). Application of metabolomics in thyroid cancer research. *International Journal of Endocrinology*, 258-763.
- Workneh, T. S., Osthoff, G. and Steyn, M. S. (2011). Physiological and chemical quality of carrots subjected to pre-and postharvest treatments. *African Journal of Agricultural Research*, 6(12): 2715-2724.
- Worley, B. and Powers, R. (2013). Multivariate Analysis in Metabolomics. *Curr Metabolomics*, 1(1):92-107.
- Xu, B. and Chang, S.K.C. (2008). Effect of soaking, boiling and steaming on total phenolic content and antioxidant activities of cool season food legumes. *Food Chemistry*, 110: 1-13.
- Xue Yi, T., Ahmad, S. H., Shukor, N. I. A, Saari, N., and Ismail, M. F. (2017). Postharvest quality of chilling injured ginger rhizomes (*Zingiber officinale* Roscoe Cv. Bentong) as affected by maturity stages, storage temperatures, and durations. *Indian Journal of Applied Research*, 7(4): 595-601.
- Yamahara, J., Hatakeyama, S., Taniguchi, K., Kawamura, M. and Yoshikawa, M. (1992). Stomachic principles in ginger. II. Pungent and anti-ulcer effects of low polar constituents isolated from ginger, the dried rhizoma of *Zingiber officinale* Roscoe cultivated in Taiwan. The absolute stereostructure of a new diarylheptanoid]. *Yakugaku Zasshi*, 112(9): 645-655.
- Yamamoto-Ribeiro, M. M., Grespan, R., Kohiyama, C. Y., Ferreira, F. D., Mossini, S. A., and Silva, E. L. (2013). Effect of *Zingiber officinale* essential oil on *Fusarium verticillioides* and fumonisin production. *Food Chemistry*, 141(3):3147–3152
- Yang, L., Wen, K. S., Ruan, X., Zhao, Y. X., Wei, F. and Wang, Q. (2018). Response of plant secondary metabolites to environmental factors. *Molecules*, 23(4):1-26.
- Yang, W. Q. and Harpole, J. (2008). Evaluating berry firmness and total soluble solids of newly released highbush blueberry cultivars. *Acta Horticulturae*, 810: 863-867.
- Yasser, S.M. and Mohamad, A.M. (2012). Commercial ginger production using fertigation system. *Buletin Teknologi MARDI*, 1, 97-105.

- Young, H. Y., Luo, Y. L., Cheng, H. Y., Hsieh, W. C., Liao, J. C. and Peng, W. H. (2005). Analgesic and anti-inflammatory activities of [6]-gingerol. *Journal of Ethnopharmacology*, 96: 207–310.
- Yun, Z., Gao, H. and Liu, P. (2013). Comparative proteomic and metabolomic profiling of citrus fruit with enhancement of disease resistance by postharvest heat treatment. *BMC Plant Biology*, 13:44–46.
- Zhang, B., Guo, K., Lin, L. Wei, C. (2018). Comparison of structural and functional properties of starches from rhizome and bulbil of Chinese yam. *Molecules*, 23(2): 1-12.
- Zick, S. M., Djuric, Z., Ruffin, M. T., Litzinger, A. J., Normolle, D. P., Alrawi, S., Feng, M. R., and Brenner, D. E. (2008). Pharmacokinetics of 6-gingerol, 8-gingerol, 10-gingerol, and 6-shogaol and conjugate metabolites in healthy human subjects. *Cancer Epidemiol Biomarkers Prev.*, 17: 1930-1936.