



GENETIC DIVERSITY OF *Bacillus pumilus* GROUP CAUSING TRUNK BULGES ON RRIM 3001 SUPERCLONE RUBBER TREE (*Hevea brasiliensis* Müll.Arg.) IN PENINSULAR MALAYSIA

By

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Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

GENETIC DIVERSITY OF *Bacillus pumilus* GROUP CAUSING TRUNK BULGES ON RRIM 3001 SUPERCLONE RUBBER TREE (*Hevea brasiliensis* Müll. Arg.) IN PENINSULAR MALAYSIA

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Bacillus pumilus was identified as the pathogen in the initial outbreak of trunk bulges in RRIM 3001 superclone rubber plants in Serdang, Selangor. The occurrences of trunk bulges of RRIM 3001 superclone rubber trees eventually affect the quality and yield of natural rubber. Multilocus sequence analysis (MLSA) of five housekeeping genes (*gyrB*, *pyrE*, *aroE*, *rpoB*, and *trpB*) and repetitive elements-based polymerase chain reaction (rep-PCR) using REP, ERIC and BOX primers were conducted to analyze the diversity and systematic relationship of 20 isolates of *B. pumilus* group from four rubber tree plantations in Peninsular Malaysia (Serdang, Tanah Merah, Baling and Rawang). The analysis of all individual phylogenetic trees revealed the nearly congruent topology structure, with 75-100% bootstrap value across all 20 isolates of *B. altitudinis* and *B. safensis*. Both *B. altitudinis* and *B. safensis* are closely related to each other. Interestingly, none of the 20 bacterial isolates collected from the four collection areas were clustered together, indicating that variations between isolates from various geographical locations were significantly smaller than those between isolates from the same locations. Specifically, these 20 isolates were classified into two primary clusters: Cluster A, which contained 17 isolates of *B. altitudinis*, and Cluster B, which contained three isolates of *B. safensis*. The results were validated by a concatenated phylogenetic tree, which revealed that the isolates were divided into two clusters of *B. altitudinis* and *B. safensis*, with 98% and 100% bootstrap values, respectively. The dendrograms generated by REP-, ERIC-, and BOX-PCRs in rep-PCR analysis revealed that both species separated well. Compared to independent rep-PCR experiments, multi rep-PCR generates a high level of discrimination, while isolates of the two species remain separate in the corresponding dendrogram. The similarity coefficient observed

among isolates in Cluster A (*B. altitudinis* isolates from Kedah, Kelantan and Rawang and five isolates from Serdang, ~35%) and Cluster B (three *B. safensis* isolates from Serdang, ~95%) from geographically separated states indicated that the isolates possibly were lineages of a single virulent isolate. The spread of these infections over Peninsular Malaysia was most likely due to transmission via planting stock. REP-PCR is the best approach for species grouping and separation since REP primers produced the most distinct fingerprinting pattern, followed by ERIC primer and BOX primer. The pathogens responsible for trunk bulges on RRIM 3001 superclone rubber trees in Peninsular Malaysia have been reclassified as *B. altitudinis* and *B. safensis* of the *B. pumilus* group. This finding would be a powerful platform for generating detailed documentation on the genetic diversity of *B. pumilus* group associated with RRIM 3001 trunk bulges in Peninsular Malaysia as well as developing disease control strategies to limit the spread of *B. pumilus* group species to new area.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**KEPELBAGAIAN GENETIK KEATAS KUMPULAN *Bacillus pumilus*
PENYEBAB BONJOLAN BATANG KEPADA KLON RRIM 3001 POKOK
GETAH (*Hevea brasiliensis* Müll. Arg.) DI SEMENANJUNG MALAYSIA**

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Bacillus pumilus dikenal pasti sebagai patogen dalam wabak awal bonjolan batang di pokok getah klon RRIM 3001 di Serdang, Selangor. Kejadian bonjolan batang pokok getah klon RRIM 3001 telah mempengaruhi kualiti dan hasil getah asli. Analisis urutan multilokus (MLSA) lima gen pengemasan (*gyrB*, *pyrE*, *aroE*, *rpoB*, dan *trpB*) dan reaksi berantai polimerase berasaskan elemen berulang (rep-PCR) menggunakan primer REP, ERIC dan BOX dilakukan untuk menganalisis kepelbagaian dan sistematik hubungan 20 pencilan kumpulan *B. pumilus* dari empat kebun pokok getah di Semenanjung Malaysia (Serdang, Tanah Merah, Baling dan Rawang). Analisis pada semua pokok filogenetik menunjukkan struktur topologi hampir sama, dan hasilnya menunjukkan 75-100% nilai butstrap antara semua 20 pencilan dengan *B. altitudinis* dan *B. safensis*. Kedua-dua *B. altitudinis* dan *B. safensis* berkait rapat antara satu sama lain. Menariknya, tiada satu pun daripada 20 pencilan bakteria yang dikumpul daripada empat kawasan pengumpulan dikelompokkan bersama, menunjukkan bahawa variasi antara pencilan dari pelbagai lokasi geografi adalah jauh lebih kecil daripada pencilan antara pencilan dari lokasi yang sama. Secara khusus, 20 pencilan ini diklasifikasikan menjadi dua kelompok utama: Cluster A, yang berisi 17 pencilan *B. altitudinis*, dan Cluster B, yang mengandungi tiga pencilan *B. safensis*. Keputusan telah disahkan oleh pokok filogenetik gabungan, yang mendedahkan bahawa pencilan dibahagikan kepada dua kelompok, *B. altitudinis* dan *B. safensis* dengan nilai butstrap 98% dan 100% masing-masing. Dalam analisis rep-PCR, dendrogram yang dihasilkan oleh REP-, ERIC-, BOX-PCR menunjukkan bahawa kedua-dua spesies berpisah dengan baik. Berbanding dengan eksperimen rep-PCR bebas, multi-PCR menghasilkan tahap diskriminasi yang lebih tinggi sedangkan pencilan kedua spesies tetap

terpisah dalam dendrogram yang sesuai. Pekali kesamaan yang diperhatikan di antara pencilan di Kluster A (pencilan *B. altitudinis* dari Kedah, Kelantan dan Rawang dan lima pencilan dari Serdang, ~ 35%) dan Kluster B (tiga pencilan *B. safensis* dari Serdang, ~ 95%) dari negeri yang terpisah secara geografi ditunjukkan bahawa pencilan mungkin adalah keturunan dari satu pencilan virulen. Penyebaran jangkitan ini ke seluruh Semenanjung Malaysia berkemungkinan besar disebabkan oleh penularan melalui stok tanaman. REP-PCR adalah kaedah yang paling sesuai untuk pengelompokan dan pemisahan spesies kerana primer REP menunjukkan corak cap jari yang paling jelas, diikuti oleh primer ERIC dan primer BOX. Patogen yang bertanggungjawab untuk membonjol batang pada pokok getah klon RRIM 3001 di Semenanjung Malaysia telah diklasifikasikan semula sebagai *B. altitudinis* dan *B. safensis* dari kumpulan *B. pumilus*. Kajian ini akan menjadi platform utama untuk menghasilkan dokumentasi terperinci mengenai kepelbagaian genetik kumpulan *B. pumilus* yang berkaitan dengan penyakit bonjolan batang pada RRIM 3001 di Semenanjung Malaysia dan dapat digunakan untuk merancang strategi pengendalian penyakit untuk membatasi pengenalan spesies kumpulan *B. pumilus* ke wilayah baru.

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LIST OF ABBREVIATIONS

%	percent
°C	degree Celsius
ANOVA	Analysis of Variance
bp	base pair
DNA	deoxyribonucleic acid
FAO	Food and Agriculture Organization
g	gram
h	hour
ha	hectare
kb	kilobase pair
kg	kilogram
L	litre
LTC	latex timber clones
mL	milliliter
M	molar
min	minutes
MRB	Malaysian Rubber Board
NR	natural rubber
rRNA	ribosomal DNA
RRIM	Rubber Research Institute of Malaysia
sec	second
TBE	<i>tris-Borate-EDTA</i>
<i>Taq</i>	<i>Thermus aquaticus</i>
TM	melting temperature
NB	nutrient broth
U	unit
µL	microlitre
UV	ultra-violet
V	voltan/volt

CHAPTER 1

INTRODUCTION

1.1 Background of Study

The rubber tree (*Hevea brasiliensis*) is a native tree species of Amazon forests in Brazil and commercially exploited for the production of high-quality natural rubber derived from latex or sap (Priyadarshan and de Gonçalves, 2003; Webster and Paardekooper, 1989). Rubber tree has become an essential commercial crop in many developing countries, especially in South East Asia (Bakri *et al.*, 2017; Nath *et al.*, 2010; Roy *et al.*, 2014). To fulfill the high demand for planting materials, the Malaysian Rubber Board (MRB) has introduced extraordinary rubber tree clones known as latex timber clones (LTC) that can contribute to the remarkable return of latex and timber (Mayati and Izilawati, 2017). RRIM 3001 superclone rubber tree is one of the successful latex timber clones with several vital features, including vigorous growth, high yield latex and timber production (Mokhatar and Daud, 2011). It can generate high production of natural rubber, about 3 tons/hectare/year (Mazlan *et al.*, 2019b). Additionally, RRIM 3001 superclone rubber tree has been appointed a new name, "Clone 1 Malaysia" (Borneo Post Online, 2011).

In 2019, the first outbreak of trunk bulges has been reported in RRIM 3001 superclone rubber tree by Mazlan *et al.* (2019b). The occurrences of trunk bulges of RRIM 3001 superclone rubber trees eventually affect the quality and yield of natural rubber. The symptoms can be seen on the whole trunk of the rubber trees, including numerous tumor-like bacteriosis, canker wounds of different sizes at the tapping zone and bleeding lesions (Mazlan *et al.*, 2019b). Swelling and cracking of the trunk bulges may happen at the most severe stage (Kovaleva *et al.*, 2015; Mazlan *et al.*, 2019b). The tapping method on the uneven bark surface could lead to cambium cells' injury and results in bulges on the RRIM 3001 superclone rubber tree (Zarawi, 2018). *Bacillus pumilus* (*B. pumilus*) has been identified as the causative agent of trunk bulges (Mazlan *et al.*, 2019b).

1.2 Problem Statement

The occurrences of trunk bulges have further diminished smallholder enthusiasm in the rubber industry since it has become one of the severe trunk diseases in Malaysia. It spreads quickly, produces severe yield losses, and has a serious impact on the rubber industry. Rubber has become the second important commodity crop in Malaysia, with a 4.7% contribution to the national gross domestic products (Sharib and Halog, 2017). The occurrence of trunk bulges causes a significant reduction in latex and rubberwood production in Malaysia (Mokhter and Aris, 2018). Thus, trunk bulges in rubber trees caused by *B.*

pumilus become the primary concern towards latex and rubberwood production. To date, there is no in-depth investigation and research related to this disease and the pathogen, as well as the genetic diversity and genetic relationship between *B. pumilus* group isolates. *Bacillus pumilus* group composed of the species *B. pumilus*, *B. safensis*, *B. altitudinis*, *B. stratosphericus*, *B. aerophilus*, *B. xiamenensis* and *B. invictae* that share sequence similarity value over 99.5% in their 16S rRNA genes (Liu *et al.*, 2013). All these species cannot be distinguished by biochemical characteristics, phenotypic tests and 16S rRNA gene sequencing due to high genetic homogeneity. The 16S rRNA gene sequence was introduced by Woese and Fox (1977) and has been widely used as a molecular marker for bacteria identification and to estimate the relationships among bacteria (Amann *et al.*, 1995; Stahl, 1991). Unfortunately, 16S rRNA gene sequence is not a perfect measure of overall sequence divergence between some genera such as *Bacillus pumilus* group (Liu *et al.*, 2013), *Bacillus cereus* group (Ash *et al.*, 1991) and *Burkholderia cepacia* complex (Bcc) (Mahenthiralingam *et al.*, 2000; Payne *et al.*, 2005).

Hence, this research aims to determine and elucidate the taxonomy, genetic diversity, and genetic profiling between *B. pumilus* group isolated from different geographical locations in Peninsular Malaysia. Indeed, the data obtained from this research will be valuable and useful for quarantine purposes and suppression of trunk bulges in our country and enhance the rubber industry in Malaysia.

1.3 Significance of Study

This study will be a powerful platform for generating detailed documentation of the genetic relationship and diversity of *B. pumilus* group isolates associated with trunk bulges on RRIM 3001 superclone rubber tree in Malaysia. Our study would also provide more information about the evolutionary and population biology studies of *B. pumilus* group species. Moreover, our study is the first attempt to compare multilocus sequence analysis (MLSA) and rep-PCR method on analyzing the genetic diversity of *B. pumilus* group isolates causing trunk bulges of RRIM 3001 superclone rubber tree.

1.4 Objectives of Study

Our study was carried out with the following objectives:

1. To determine the genetic relationship of *B. pumilus* group isolates associated with trunk bulges of RRIM 3001 superclone rubber tree from different geographical areas in Peninsular Malaysia by using multilocus sequence analysis (MLSA).
2. To elucidate the genetic diversity of *B. pumilus* group isolates associated with trunk bulges of RRIM 3001 superclone rubber tree from different geographical areas in Peninsular Malaysia using molecular profiling via rep-PCR.

REFERENCES

- Adékambi, T., Shinnick, T. M., Raoult, D., & Drancourt, M. (2008). Complete *rpoB* gene sequencing as a suitable supplement to DNA–DNA hybridization for bacterial species and genus delineation. *International Journal of Systematic and Evolutionary Microbiology*, 58(8), 1807–1814. PMID: 18676461 DOI: 10.1099/ijms.0.65440-0
- Ali, H. M., Rasib, A. W., Hamid, N. R. A., Ghani, Z. A., Mahsuri, I., Yusoff, A. R. M., Idris, K. M., & Dollah, R. (2018, June). Proceedings from In IOP Conference Series: *Earth and Environmental Science Determination of rubber tree clones leaf diseases spectral using Unmanned Aerial Vehicle compact sensor*. IOP Publishing.
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, 215(3), 403–410. DOI: 10.1016/S0022-2836(05)80360-2
- Amann, R. I., Ludwig, W., & Schleifer, K. H. (1995). Phylogenetic identification and in situ detection of individual microbial cells without cultivation. *Microbiological Reviews*, 59(1), 143-169.
- Ansari, M., Taghavi, S. M., Zarei, S., Miri, K., Portier, P., & Osdaghi, E. (2018). Pathogenicity and molecular-phylogenetic analysis revealed a distinct position of the banana finger-tip rot pathogen among the *Burkholderia cenocepacia* genomovars. *Plant Pathology*, ppa.12976. DOI: 10.1111/ppa.12976
- Archer, B. L., Audley, B. G., Cockbain, E. G., & McSweeney, G. P. (1963). The biosynthesis of rubber. Incorporation of mevalonate and isopentenyl pyrophosphate into rubber by *Hevea brasiliensis*-latex fractions. *Biochemical Journal*, 89(3), 565-574.
- Ash, C., Farrow, J. A., Dorsch, M., Stackebrandt, E., & Collins, M. D. (1991). Comparative analysis of *Bacillus anthracis*, *Bacillus cereus*, and related species on the basis of reverse transcriptase sequencing of 16S rRNA. *International Journal of Systematic and Evolutionary Microbiology*, 41(3), 343-346.
- Aunpad, R., & Na-Bangchang, K. (2007). Pumilicin 4, a novel bacteriocin with anti-MRSA and anti-VRE activity produced by newly isolated bacteria *Bacillus pumilus* strain WAPB4. *Current Microbiology*, 55(4), 308-313.
- Azevedo, H., Lopes, F., Silla, P., & Hungria, M. (2015). A database for the taxonomic and phylogenetic identification of the genus *Bradyrhizobium* using multilocus sequence analysis. *BMC Genomics*, 16 Suppl 5(Suppl 5), S10. DOI: 10.1186/1471-2164-16-S5-S10.

- Bakri, A., Mokhtar, S. J., & Daud, N. W. (2017). Physiological and morphological responses of rubber (*Hevea brasiliensis*) RRIM 3001 to different rates of basalt application. *Journal of Tropical Plant Physiology*, 9, 24-35.
- Balsiger, J., Bahdon, J., & Whiteman, A. (2000). *The utilization, processing and demand for rubberwood as a source of wood supply*. Rome: Forestry Policy and Planning Division.
- Banerjee, S., Devaraja, T. N., Shariff, M., & Yusoff, F. M. (2007). Comparison of four antibiotics with indigenous marine *Bacillus* spp. in controlling pathogenic bacteria from shrimp and Artemia. *Journal of Fish Diseases*, 30(7), 383-389.
- Bastiah, A., Wan, C. K., & Akib, M. Y. (1996). Effect of interstock on dry matter production and growth analysis of *Hevea brasiliensis*. *Muell. Arg. Journal of National Rubber Research*, 11(4), 265-296.
- Bhattacharya, D., Sarma, P. M., Krishnan, S., Mishra, S., & Lal, B. (2003). Evaluation of genetic diversity among *Pseudomonas citronellolis* strains isolated from oily sludge-contaminated sites. *Application Environment Microbiology*, 69(3), 1435-1441.
- Borneo Post Online, (2011). RRIM Hasil Getah 1Malaysia. Retrieved from <http://www.theborneopost.com/2011/01/30/rrim-hasil-getah-1malaysia/>. Accessed on 15 December 2014.
- Brady, C., Cleenwerck, I., Venter, S., Vancanneyt, M., Swings, J., & Coutinho, T. (2008). Phylogeny and identification of *Pantoea* species associated with plants, humans and the natural environment based on multilocus sequence analysis (MLSA). *Systematic and Applied Microbiology*, 31, 447-460. DOI: 10.1016/j.syapm.2008.09.004
- Branquinho, R., Meirinhos-Soares, L., Carriço, J. A., Pintado, M., & Peixe, L. V. (2014a). Phylogenetic and clonality analysis of *Bacillus pumilus* isolates uncovered a highly heterogeneous population of different closely related species and clones. *FEMS Microbiology Ecology*, 90(3), 689-698. PMID: 25230950 DOI: 10.1111/1574-6941.12426
- Branquinho, R., Sousa, C., Lopes, J., Pintado, M. E., Peixe, L. V., & Osório, H. (2014b). Differentiation of *Bacillus pumilus* and *Bacillus safensis* using MALDI-TOF-MS. *PLoS One*, 9(10), e110127.
- Brown, A. E., & Soepena, H. (1994). Pathogenicity of *Colletotrichum acutatum* and *C. gloeosporioides* on leaves of *Hevea* spp. *Mycological Research*, 98(3), 264-266.
- Chanroj, V., Rattanawong, R., Phumichai, T., Tangphatsornruang, S., & Ukoskit, K. (2017). Genome-wide association mapping of latex yield and girth in Amazonian accessions of *Hevea brasiliensis* grown in a suboptimal climate zone. *Genomics*, 109(5-6), 475-484.

- Cherif, A., Brusetti, L., Borin, S., Rizzi, A., Boudabous, A., Khyami-Horani, H., & Daffonchio, D. (2003). Genetic relationship in the '*Bacillus cereus* group' by rep-PCR fingerprinting and sequencing of a *Bacillus anthracis*-specific rep-PCR fragment. *Journal of Applied Microbiology*, 94(6), 1108-1119.
- Cherif, A., Ettoumi, B., Raddadi, N., Daffonchio, D., & Boudabous, A. (2007). Genomic diversity and relationship of *Bacillus thuringiensis* and *Bacillus cereus* by multi-REP-PCR fingerprinting. *Canadian Journal of Microbiology*, 53(3), 343-350.
- Chun, J., & Bae, K. S. (2000). Phylogenetic analysis of *Bacillus subtilis* and related taxa based on partial *gyrA* gene sequences. *Antonie van Leeuwenhoek*, 78(2), 123-127.
- Clément-Demange, A., Priyadarshan, P. M., Hoa, T. T. T., & Venkatachalam, P. (2007). *Hevea* rubber breeding and genetics. *Plant Breeding Reviews*, 29, 177.
- Coenye, T., LiPuma, J. J., Henry, D., Hoste, B., Vandemeulebroecke, K., Gillis, M., & Vandamme, P. (2001b). *Burkholderia cepacia* genomovar VI, a new member of the *Burkholderia cepacia* complex isolated from cystic fibrosis patients. *International Journal of Systematic and Evolutionary Microbiology*, 51(2), 271-279.
- Coenye, T., Mahenthiralingam, E., Henry, D., LiPuma, J. J., Laevens, S., Gillis, M., & Vandamme, P. (2001a). *Burkholderia ambifaria* sp. nov., a novel member of the *Burkholderia cepacia* complex including biocontrol and cystic fibrosis-related isolates. *International Journal of Systematic and Evolutionary Microbiology*, 51(4), 1481-1490.
- Coorevits, A., De Jonghe, V., Vandroemme, J., Reekmans, R., Heyrman, J., Messens, W., & Heyndrickx, M. (2008). Comparative analysis of the diversity of aerobic spore-forming bacteria in raw milk from organic and conventional dairy farms. *Systematic and Applied Microbiology*, 31(2), 126-140.
- Cornish, K. (1993). The separate roles of plant cis and trans prenyl transferases in cis-1, 4-polyisoprene biosynthesis. *European Journal of Biochemistry*, 218(1), 267-271.
- Cornish, K. (2001). Similarities and differences in rubber biochemistry among plant species. *Phytochemistry*, 57, 1123-1134.
- da Hora Júnior, B. T., de Macedo, D. M., Barreto, R. W., Evans, H. C., Mattos, C. R. R., Maffia, L. A., & Mizubuti, E. S. (2014). Erasing the past: a new identity for the *Damoclean* pathogen causing South American leaf blight of rubber. *PLoS One*, 9(8): e104750.

- Da Silva, R. B., & Valicente, F. H. (2013). Molecular characterization of *Bacillus thuringiensis* using REP-PCR. *SpringerPlus*, 2(1), 641.
- d'Auzac, J. (1989). Tapping systems and area of drained bark. *Physiology of rubber tree latex: the laticiferous cell and latex: a model of cytoplasm/editors*, Jean d'Auzac, Jean-Louis Jacob, Herv'e Chrestin.
- De Jonghe, V., Coorevits, A., De Block, J., Van Coillie, E., Grijspeerdt, K., Herman, L., & Heyndrickx, M. (2010). Toxinogenic and spoilage potential of aerobic spore-formers isolated from raw milk. *International Journal of Food Microbiology*, 136(3), 318-325.
- De Vos, P. (2011). Multilocus Sequence Determination and Analysis. *Methods in Microbiology*, 38, 385–407. DOI: 10.1016/B978-0-12-387730-7.00017-6
- Dijkman, M. J. (1951). *Hevea, Thirty years of research in the Far East*. University of Miami Press, Coral Gables, Florida. pp. 5-7.
- Elbanna, K., Elnaggar, S., & Bakeer, A. (2014). Characterization of *Bacillus altitudinis* as a new causative agent of bacterial soft rot. *Journal of Phytopathology*, 162(11-12), 712-722. DOI: 10.1111/jph.12250
- Espariz, M., Zuljan, F. A., Esteban, L., & Magni, C. (2016). Taxonomic identity resolution of highly phylogenetically related strains and selection of phylogenetic markers by using genome-scale methods: the *Bacillus pumilus* group case. *PLoS One*, 11(9), e0163098. DOI: 10.1371/journal.pone.0163098.
- FAO (1989). *Commodity Review and Outlook 1988-89*. Rome: Food and Agriculture Organization of the United Nations.
- FAO Regional Office for Asia and the Pacific. (2007). *Report of the twenty-fifth session of the Asia and Pacific Plant Protection Commission*. Bangkok: RAP Publication. Retrieved October 2018
- Farid, A. M., Lee, S. S., Maziah, Z., & Patahayah, M. (2009). Pathogenicity of *Rigidoporus microporus* and *Phellinus noxius* against four major plantation tree species in peninsular Malaysia. *Journal of Tropical Forest Science*, 289-298.
- Fay, E. D., Moraes, L. A. C., & Moraes, V. H. D. F. (2010). Cyanogenesis and the onset of tapping panel dryness in rubber tree. *Pesquisa Agropecuária Brasileira*, 45(12), 1372-1380.
- Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. *Evolution*, 39(4), 783-791.

- Ficarra, F. A., Santecchia, I., Lagorio, S. H., Alarcón, S., Magni, C., & Espariz, M. (2016). Genome mining of lipolytic exoenzymes from *Bacillus safensis* S9 and *Pseudomonas alcaliphila* ED1 isolated from a dairy wastewater lagoon. *Archives of Microbiology*, 198(9), 893-904.
- Fox, G. E., Stackebrandt, E., Hespell, R. B., Gibson, J., Maniloff, J., Dyer, T. A., & Woese, C. R. (1980). The phylogeny of prokaryotes. *Science*, 209(4455), 457-463. doi: 10.1126/science.6771870
- Fox, G. E., Wisotzkey, J. D., & Jurtshuk Jr, P. (1992). How close is close: 16S rRNA sequence identity may not be sufficient to guarantee species identity. *International Journal of Systematic and Evolutionary Microbiology*, 42(1), 166-170. doi: 10.1099/00207713-42-1-166
- Fox, J., & Castella, J. C. (2013). Expansion of rubber (*Hevea brasiliensis*) in Mainland Southeast Asia: what are the prospects for smallholders?. *The Journal of Peasant Studies*, 40(1), 155-170.
- Freitas, D. B., Reis, M. P., Lima-Bittencourt, C. I., Costa, P. S., Assis, P. S., Chartone-Souza, E., & Nascimento, A. M. (2008). Genotypic and phenotypic diversity of *Bacillus* spp. isolated from steel plant waste. *BMC Research Notes*, 1(1), 92.
- Fritze, D. (2002). *Bacillus* identification-traditional approaches. *Applications and Systematics of Bacillus and Relatives*, 100-122.
- Galal, A. A., El-Bana, A. A., & Janse, J. (2006). *Bacillus pumilus*, a new pathogen on mango plants. *Egypt Journal of Phytopathology*, 34(1), 17-29.
- Gao, J. J., Kang, Y. J., Ceng, J., & Yin, S. X. (2012). PGPRs Effects on rice seedlings by method of raising rice seedling with plastic plate. *Soils*, 44, 126-132.
- Gevers, D., Cohan, F. M., Lawrence, J. G., Spratt, B. G., Coenye, T., Feil, E. J., & Swings, J. (2005). Re-evaluating prokaryotic species. *Nature Reviews Microbiology*, 3(9), 733.
- Gillis, M., Van Van, T., Bardin, R., Goor, M., Hebbar, P., Willems, A., ... & Fernandez, M. P. (1995). Polyphasic taxonomy in the genus *Burkholderia* leading to an emended description of the genus and proposition of *Burkholderia vietnamiensis* sp. nov. for N₂-fixing isolates from rice in Vietnam. *International Journal of Systematic and Evolutionary Microbiology*, 45(2), 274-289.
- Glaeser, S. P., & Kämpfer, P. (2015). Multilocus sequence analysis (MLSA) in prokaryotic taxonomy. *Systematic and Applied Microbiology*, 38(4), 237-245.

- González, A., Hierro, N., Poblet, M., Rozes, N., Mas, A., & Guillamón, J. M. (2004). Application of molecular methods for the differentiation of acetic acid bacteria in a red wine fermentation. *Journal of Applied Microbiology*, 96(4), 853-860.
- Guo, Y., Zheng, W., Rong, X., & Huang, Y. (2008). A multilocus phylogeny of the *Streptomyces griseus* 16S rRNA gene clade: use of multilocus sequence analysis for *streptomycete* systematics. *International Journal of Systematic and Evolutionary Microbiology*, 58(1), 149-159.
- Hakim, S., Liaquat, F., Gul, S., Chaudhary, H. J., & Munis, M. F. H. (2015). Presence of *Bacillus pumilus* causing fruit rot of *Ficus lacor* in Pakistan. *Journal of Plant Pathology*, 97(3), 543. DOI: 10.4454/JPP.V97I3.007.
- Hall, B. G. (2013). Building Phylogenetic Trees from Molecular Data with MEGA. *Molecular Journal of Biological Evolution*, 30, 1229-1235, DOI:10.1093/molbev/mst012
- Hanage, W. P., Fraser, C., & Spratt, B. G. (2005). Fuzzy species among recombinogenic bacteria. *BMC Biology*, 3(1), 1-7. PMID: 15752428 DOI: 10.1186/1741-7007-3-6
- Handtke, S., Volland, S., Methling, K., Albrecht, D., Becher, D., Nehls, J., & Voigt, B. (2014). Cell physiology of the biotechnological relevant bacterium *Bacillus pumilus*—An omics-based approach. *Journal of Biotechnology*, 192, 204-214.
- Hedges, S. B. (1992). The number of replications needed for accurate estimation of the bootstrap P value in phylogenetic studies. *Molecular Biology and Evolution*, 9(2), 366-369.
- Hevea Africa Limited (HAL) (2014), factsheet. <http://www.heveafricaltd.com/rubber-tree/factsheet/>
- Holm, L. G., Plucknett, D. L., Pancho, J. V., & Herberger, J. P. (1977). The world's worst weeds. Distribution and biology: Honolulu, Hawaii, USA, University Press of Hawaii.
- Hong, H. A., Duc, L. H., & Cutting, S. M. (2005). The use of bacterial spore formers as probiotics. *FEMS Microbiology Reviews*, 29(4), 813-835. <http://vital.lgm.gov.my/vital/access/manager/Collection/vital1:78300;jse-sessionid=ED57218D35C06BA5D87A673B74480277>
- Hua, D., Ma, C., Lin, S., Song, L., Deng, Z., Maomy, Z., Zhang, Z., Yu, B., & Xu, P. (2007). Biotransformation of isoeugenol to vanillin by a newly isolated *Bacillus pumilus* strains: identification of major metabolites. *Journal of Biotechnology*, 130(4), 463-470.

- Huang, Q., Peng, Y., Li, X., Wang, H., & Zhang, Y., (2003). Purification and characterization of an extracellular alkaline serine protease with dehairing function from *Bacillus pumilus*. *Current Microbiology*, 46(3), 0169-0173.
- Hulton, C. S. J., Higgins, C. F., & Sharp, P. M. (1991). ERIC sequences: a novel family of repetitive elements in the genomes of *Escherichia coli*, *Salmonella typhimurium* and other enterobacteria. *Molecular Microbiology*, 5(4), 825-834.
- Ibrahim, N. A. G. A. A., Omar, M. N. A. M., El Heba, G. A. A., Moënne-Loccoz, Y., Prigent-Combaret, C., & Muller, D. (2018). Draft genome sequence of plant growth-promoting *Bacillus altitudinis* strain PAE4. *Microbiology Resource Announcements*, 7(13).
- IRSG. 2014. Rubber Statistical Bulletin Oct-Dec 2014. The International Rubber Study Group. 69:4-6.
- Ishii, S., & Sadowsky, M. J. (2009). Applications of the rep-PCR DNA fingerprinting technique to study microbial diversity, ecology and evolution. *Environmental Microbiology*, 11(4), 733-740.
- Ismail, H., & Jeyanayagi, I. (1999). Occurrence and identification of physiological races of *Corynespora cassiicola* of *Hevea*. In *Proceedings of IRRDB Symposium* (pp. 263-272).
- Jacob, C. K. (2006). *Corynespora* leaf disease of *Hevea brasiliensis*: A threat to natural rubber production in *Corynespora* Leaf Disease of *H. brasiliensis*: strategies for management. *Rubber Research Institute of India*, 9-16.
- Jayasinghe, C. K. (2011). White root disease, the most devastating root disease of the rubber tree. *International Rubber Research and Development Board: Malaysia*.
- Jayasinghe, C. K., Fernando, T. H. P. S., & Priyanka, U. M. S. (1997). *Colletotrichum acutatum* is the main cause of *Colletotrichum* leaf disease of rubber in Sri Lanka. *Mycopathologia*, 137(1), 53-56.
- Joung, K. B., & Côté, J. C. (2002). Evaluation of ribosomal RNA gene restriction patterns for the classification of *Bacillus* species and related genera. *Journal of Applied Microbiology*, 92(1), 97-108.
- Junior, H. D. J. E., Ohto, J. M., da Silva, L. L., Palma, H. A. L., & Ballarin, A. W. (2015). Potential of rubberwood (*Hevea brasiliensis*) for structural use after the period of latex extraction: a case study in Brazil. *Journal of Wood Science*, 61(4), 384-390.
- Kathleen, M. M., Samuel, L., Felecia, C., Ng, K. H., Lesley, M. B., & Kasing, A. (2014). (GTG) 5-PCR analysis and 16S rRNA sequencing of bacteria from Sarawak aquaculture environment. *International Food Research Journal*, 21(3).

- Khan, M. H. U., Khattak, J. Z. K., Jamil, M., Malook, I., Khan, S. U., Jan, M., & Fahad, S. (2017). *Bacillus safensis* with plant-derived smoke stimulates rice growth under saline conditions. *Environmental Science and Pollution Research*, 24(30), 23850-23863. PMID: 28868579 DOI: 10.1007/s11356-017-0026-y
- Kim, W., Hong, Y. P., Yoo, J. H., Lee, W. B., Choi, C. S., & Chung, S. I. (2002). Genetic relationships of *Bacillus anthracis* and closely related species based on variable-number tandem repeat analysis and BOX-PCR genomic fingerprinting. *FEMS Microbiology Letters*, 207(1), 21-27.
- Kimouli, M., Vrioni, G., Papadopoulou, M., Koumaki, V., Petropoulou, D., Gounaris, A., & Tsakris, A. (2012). Two cases of severe sepsis caused by *Bacillus pumilus* in neonatal infants. *Journal of Medical Microbiology*, 61(4), 596-599.
- Kimura, M. (1980). A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16(2), 111-120. DOI: 10.1007/BF01731581
- Kitahara, K., and Miyazaki, K. (2013). Revisiting bacterial phylogeny: natural and experimental evidence for horizontal gene transfer of 16S rRNA. *Mobile Genetic Elements*, 3(1), e24210. DOI: 10.4161/mge.24210
- Konstantinidis, K. T., Ramette, A., & Tiedje, J. M. (2006). Toward a more robust assessment of intraspecies diversity, using fewer genetic markers. *Applied and Environmental Microbiology*, 72(11), 7286-7293. PMID: 16980418 DOI: 10.1128/AEM.01398-06.
- Kovaleva, V. A., Shalovylo, Y. I., Gorovik, Y. N., Lagonenko, A. L., Evtushenkov, A. N., & Gout, R. T. (2015). *Bacillus pumilus*-a new phytopathogen of Scots pine. *Journal of Forest Science*, 61(3), 131-137. DOI: 10.17221/16/2014-JFS
- Kumar, A., Kumar, A., & Pratush, A. (2014). Molecular diversity and functional variability of environmental isolates of *Bacillus* species. *SpringerPlus*, 3(1), 312. PMID: 25279279 DOI: 10.1186/2193-1801-3-312
- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution*, 33(7), 1870–1874. DOI: 10.1093/molbev/msw054.
- La Duc, M. T., Kern, R., & Venkateswaran, K. (2004). Microbial monitoring of spacecraft and associated environments. *Microbial Ecology*, 47(2), 150-158.
- Lai, Q., Liu, Y., & Shao, Z. (2014). *Bacillus xiamenensis* sp. nov., isolated from intestinal tract contents of a flathead mullet (*Mugil cephalus*). *Antonie van Leeuwenhoek*, 105(1), 99-107.

- Lateef, A., Adelere, I. A., & Gueguim-Kana, E. B. (2015). The biology and potential biotechnological applications of *Bacillus safensis*. *Biologia*, 70(4), 411-419.
- Lau, N. S., Makita, Y., Kawashima, M., Taylor, T. D., Kondo, S., Othman, A. S., & Matsui, M. (2016). The rubber tree genome shows expansion of gene family associated with rubber biosynthesis. *Scientific Reports*, 6, 28594. DOI: 10.1038/srep28594.
- Lemjiber, N., Naamani, K., Merieau, A., Dihazi, A., Zhar, N., Jediyyi, H., & Boukerb, A. M. (2021). Identification and Genomic Characterization of Pathogenic *Bacillus altitudinis* from Common Pear Trees in Morocco. *Agronomy*, 11(7), 1344.
- Leong, S.K. (1979). Propagation and establishment methods in Hevea, In Samsudin, T. (ed.) RRIM training manual on rubber planting and nursery techniques, Rubber Research Institute of Malaysia, Kuala Lumpur, pp. 15-21.
- Li, B., Qiu, W., Tan, Q. M., Su, T., Fang, Y., & Xie, G. L. (2009). Association of a *Bacillus* species with leaf and twig dieback of Asian pear (*Pyrus pyrifolia*) in China. *Journal of Plant Pathology*, 91(3), 705-708.
- Li, C., Lai, Q., Li, G., Liu, Y., Sun, F., & Shao, Z. (2014). Multilocus sequence analysis for the assessment of phylogenetic diversity and biogeography in *Hyphomonas* bacteria from diverse marine environments. *PLoS One*, 9(7).
- Li, X., Bi, Z., Di, R., Liang, P., He, Q., Liu, W., & Zheng, F. (2016). Identification of powdery mildew responsive genes in *Hevea brasiliensis* through mRNA differential display. *International Journal of Molecular Sciences*, 17(2), 181.
- Librado, P., Rozas, J. (2009). DNASP v. 5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics*, 25, 1451-2.
- Lieberei, R. (2007). South American leaf blight of the rubber tree (*Hevea* spp.): new steps in plant domestication using physiological features and molecular markers. *Annals of Botany*, 100(6), 1125-1142.
- Linu, K. (2017). *Micro RNAs of Hevea brasiliensis role in abiotic stress responsive gene regulation* (Doctoral dissertation, Mahadma Gandhi University).
- Lisha, L. P. (2017). *Molecular studies on abiotic stress and identification of stress tolerant genes in Hevea brasiliensis* (Doctoral dissertation, Mahadma Gandhi University).
- Liu, X. J., Yang, Y. T., & Leng, H. Q. (1987). Identification of species and forms of *Colletotricum gloeosporioides* in rubber growing regions in south China. *China Journal of Tropical Crops*, 8, 93-01.

- Liu, Y., Lai, Q., & Shao, Z. (2017). A multilocus sequence analysis scheme for phylogeny of *Thioclava* bacteria and proposal of two novel species. *Frontiers in Microbiology*, 8, 1321.
- Liu, Y., Lai, Q., Dong, C., Sun, F., Wang, L., Li, G., & Shao, Z. (2013). Phylogenetic diversity of the *Bacillus pumilus* group and the marine ecotype revealed by multilocus sequence analysis. *PloS One*, 8(11), e80097. PMID: 24244618 DOI: 10.1371/journal.pone.0080097
- Liu, Y., Lai, Q., Du, J., & Shao, Z. (2015a). Reclassification of *Bacillus invictae* as a later heterotypic synonym of *Bacillus altitudinis*. *International Journal of Systematic and Evolutionary Microbiology*, 65(8), 2769-2773.
- Liu, Y., Lai, Q., Göker, M., Meier-Kolthoff, J. P., Wang, M., Sun, Y., & Shao, Z. (2015b). Genomic insights into the taxonomic status of the *Bacillus cereus* group. *Scientific Reports*, 5(1), 1-11. DOI: 10.1038/srep14082.
- Logan, N. A., Berge, O., Bishop, A. H., Busse, H. J., De Vos, P., Fritze, D., & Seldin, L. (2009). Proposed minimal standards for describing new taxa of aerobic, endospore-forming bacteria. *International Journal of Systematic and Evolutionary Microbiology*, 59(8), 2114-2121.
- López-Hermoso, C., de la Haba, R. R., Sánchez-Porro, C., Papke, R. T., & Ventosa, A. (2017). Assessment of multilocus sequence analysis as a valuable tool for the classification of the Genus *Salinivibrio*. *Frontiers in Microbiology*, 8, 1107. DOI: 10.3389/fmicb.2017.01107.
- Louws, F. J., Fulbright, D. W., Stephens, C. T., & De Bruijn, F. J. (1994). Specific genomic fingerprints of phytopathogenic *Xanthomonas* and *Pseudomonas* pathogens and strains generated with repetitive sequences and PCR. *Application of Environment Microbiology*, 60(7), 2286-2295.
- Louws, F. J., Rademaker, J. L. W., & De Bruijn, F. J. (1999). The three Ds of PCR-based genomic analysis of phytobacteria: diversity, detection, and disease diagnosis. *Annual Review of Phytopathology*, 37(1), 81-125.
- Macheras, E., Roux, A. L., Bastian, S., Leão, S. C., Palaci, M., Sivadon-Tardy, V., & Bodmer, T. (2011). Multilocus sequence analysis and *rpoB* sequencing of *Mycobacterium abscessus* (sensu lato) strains. *Journal of Clinical Microbiology*, 49(2), 491-499.
- Mahenthalingam, E., Bischof, J., Byrne, S. K., Radomski, C., Davies, J. E., Av-Gay, Y., & Vandamme, P. (2000). DNA-based diagnostic approaches for identification of *Burkholderia cepacia* complex, *Burkholderia vietnamiensis*, *Burkholderia multivorans*, *Burkholderia stabilis*, and *Burkholderia cepacia genomovars* I and III. *Journal of Clinical Microbiology*, 38(9), 3165-3173.

- Mahmud, R. S., Ulyanova, V., Malanin, S., Dudkina, E., Vershinina, V., & Ilinskaya, O. (2015). Draft whole-genome sequence of *Bacillus altitudinis* strain B-388, a producer of extracellular RNase. *Genome Announcements*, 3(1).
- Makita, Y., Ng, K. K., Veera Singham, G., Kawashima, M., Hirakawa, H., Sato, S., & Matsui, M. (2017). Large-scale collection of full-length cDNA and transcriptome analysis in *Hevea brasiliensis*. *Dna Research*, 24(2), 159-167.
- Malaysian Rubber Board, 2009. Rubber Plantation and Processing Technologies. Malaysian Rubber Board.
- Manigundan, K., Kumar, P. P., Singh, R., Sakthivel, K., Gautam, R. K., Singh, P. K., & Laha, G. S. (2017). Rep-PCR distinguishes rice bacterial blight pathogen (*Xanthomonas oryzae* pv. *oryzae*) strains of Indian mainland and bay islands. *Journal of Plant Pathology*, 99(3), 773-778. DOI: 10.4454/jpp.v99i3.3988
- Manju, M. J. (2011). *Epidemiology and Management of Corynespora leaf Fall Disease of Rubber caused by Corynespora cassicola* (Berk & Curt.) Wei (Doctoral dissertation, UAS Dharward).
- Mantello, C. C., Cardoso-Silva, C. B., da Silva, C. C., de Souza, L. M., Junior, E. J. S., de Souza Gonçalves, P., & de Souza, A. P. (2014). De novo assembly and transcriptome analysis of the rubber tree (*Hevea brasiliensis*) and SNP markers development for rubber biosynthesis pathways. *PloS One*, 9(7), e102665. DOI: 10.1371/journal.pone.0102665
- Martin, B., Humbert, O., Camara, M., Guenzi, E., Walker, J., Mitchell, T., & Morrison, D. A. (1992). A highly conserved repeated DNA element located in the chromosome of *Streptococcus pneumoniae*. *Nucleic Acids Research*, 20(13), 3479-3483.
- Mayati, N. C. H., & Izilawati, M (2017). Regeneration of Latex Timber Clone RRIM 3001 via Somatic Embryogenesis. *Advances in Plant Science and Technology*, 130-133.
- Mazlan, S., Jaafar, N. M., Wahab, A., Sulaiman, Z., Rajandas, H., & Zulperi, D. (2021). Molecular characterization and phylogenetic analysis of *Bacillus pumilus* causing trunk bulges of RRIM 3001 superclone rubber tree in Malaysia. *European Journal of Plant Pathology*, 1-14.
- Mazlan, S., Zulperi, D., Wahab, A., Jaafar, N. M., Sulaiman, Z., & Rajandas, H. (2019b). First Report of *Bacillus pumilus* Causing Trunk Bulges of Rubber Tree (*Hevea brasiliensis*) in Malaysia. *Plant Disease*, 103(5), 1016. DOI: 10.1094/PDIS-08-18-1409-PDN

- Meintanis, C., Chalkou, K. I., Kormas, K. A., Lympelopoulou, D. S., Katsifas, E. A., Hatzinikolaou, D. G., & Karagouni, A. D. (2008). Application of *rpoB* sequence similarity analysis, REP-PCR and BOX-PCR for the differentiation of species within the genus *Geobacillus*. *Letters in Applied Microbiology*, 46(3), 395-401.
- Mohapatra, B. R., Broersma, K., & Mazumder, A. (2007). Comparison of five rep-PCR genomic fingerprinting methods for differentiation of fecal *Escherichia coli* from humans, poultry and wild birds. *FEMS Microbiology Letters*, 277(1), 98-106. DOI: 10.1111/j.1574-6968.2007.00948.x
- Mokhatar, S. J., & Daud, N. W. (2011). Performance of hevea brasiliensis on haplic acrisol soil as affected by different source of fertilizer. *International Journal of Applied*, 1(1).
- Mokhter, M. S. & Aris, M. N. M. *Protuberance on RRIM 3001 tree trunks: wood anatomy and mechanical properties*. Paper presented at the meeting of the Seminar RRIM 3001, Akademi Hevea Malaysia. July 2018.
- Mooibroek, H., & Cornish, K. (2000). Alternative sources of natural rubber. *Applied Microbiology and Biotechnology*, 53(4), 355-365.
- Moretti, C., Vinatzer, B. A., Onofri, A., Valentini, F., Buonauro, R. (2017). Genetic and phenotypic diversity of Mediterranean populations of the olive knot pathogen, *Pseudomonas savastanoi* pv. *savastanoi*. *Plant Pathology*, 66, 595–605. DOI: 10.1111/ppa.12614
- MRB, Malaysian Rubber Board (2012). Malaysia eyes 1.2 mln ha of rubber by 2020. Borneo Post Online, 1–7.
- MRB. *Pocket Book 2020*; Malaysian Rubber Board: Kuala Lumpur, Malaysia, 2020.
- Mulet, M., Lalucat, J., & García-Valdés, E. (2010). DNA sequence-based analysis of the *Pseudomonas* species. *Environmental Microbiology*, 12(6), 1513-1530.
- Musigamart, N. (2015). *Study of the role of lipids from matured coagula from Hevea brasiliensis latex on natural rubber behavior in oxidative conditions* (Doctoral dissertation, Biochemistry, Molecular Biology. Montpellier SupAgro).
- Narayanan, C., & Mydin, K. K. (2012). Breeding for disease resistance in Hevea spp.-status, potential threats, and possible strategies. In *In: Sniezko, Richard A.; Yanchuk, Alvin D.; Kliejunas, John T.; Palmieri, Katharine M.; Alexander, Janice M.; Frankel, Susan J., tech. coords. Proceedings of the fourth international workshop on the genetics of host-parasite interactions in forestry: Disease and insect resistance in forest trees. Gen. Tech. Rep. PSW-GTR-240. Albany, CA: Pacific Southwest*

Research Station, Forest Service, US Department of Agriculture. pp. 240-251 (Vol. 240, pp. 240-251).

Nath, T.K., Inoue, M. & De Zoysa, M. (2010). Rubber planting for forest rehabilitation and enhancement of commercial livelihood: a comparative study in three south asian countries. In: *18th Commonwealth Forestry Conference*, Edinburgh.

Ng, A. P. (1983). *Performance of rootstocks*. Planters' bulletin, Rubber Research Institute of Malaysia; Kuala Lumpur. no. 175: 56-63.

Nicholson, W. L., Munakata, N., Horneck, G., Melosh, H. J., & Setlow, P. (2000). Resistance of *Bacillus* endospores to extreme terrestrial and extraterrestrial environments. *Microbiology and Molecular Biology Reviews*, 64(3), 548-572.

Nieminen, T., Rintaluoma, N., Andersson, M., Taimisto, A. M., Ali-Vehmas, T., Seppälä, A., & Salkinoja-Salonen, M. (2007). Toxinogenic *Bacillus pumilus* and *Bacillus licheniformis* from mastitic milk. *Veterinary Microbiology*, 124(3-4), 329-339.

Ntambo, M. S., Meng, J.-Y., Rott, P. C., Royer, M., Lin, L.-H., Zhang, H.-L., & Gao, S.-J. (2019). Identification and characterization of *Xanthomonas albilineans* causing sugarcane leaf scald in China using multilocus sequence analysis. *Plant Pathology*, 68(2), 269–277. DOI: 10.1111/ppa.12951

Nurmi-Rohayu, A.H., Rasyidah, M.R., Zarawi, A.G., Mohd-Nasaruddin, M.A., Noorliana, M.Z., Nor-Afiqah, M. and Aizat-Shamin, N. (2015). MRB clone recommendations 2013. In *Buletin Sains dan Teknologi*, LGM. Bil. 1/2015 Jilid 13. ISSN 1985-0557. Pp. 10-12.

Nwokoro, O., & Dibia, M. E. U. (2014). Degradation of soil cyanide by single and mixed cultures of *Pseudomonas stutzeri* and *Bacillus subtilis*. *Archives of Industrial Hygiene and Toxicology*, 65(1).

Ooi, C.B. (1978). Propagation and planting materials in *Hevea*, In Samsudin, T. (ed.) RRIM short course on rubber planting and nursery techniques, Rubber Research Institute of Malaysia, Kuala Lumpur, pp. 8-16.

Osdaghi, E., Taghavi, S. M., Koebnik, R., & Lamichhane, J. R. (2018). Multilocus sequence analysis reveals a novel phylogroup of *Xanthomonas euvesicatoria* pv. perforans causing bacterial spot of tomato in Iran. *Plant Pathology*, 67(7), 1601–1611. DOI: 10.1111/ppa.12864

Otto, M., Petersen, Y., Roux, J., Wright, J., & Coutinho, T. A. (2018). Bacterial canker of cherry trees, *Prunus avium*, in South Africa. *European Journal of Plant Pathology*, 151(2), 427–438. DOI: 10.1007/s10658-017-1384-5

Oueslati, M., Mulet, M., Gomila, M., Berge, O., Hajlaoui, M. R., Lalucat, J. & García-Valdés, E. (2019). New species of pathogenic *Pseudomonas*

- isolated from citrus in Tunisia: Proposal of *Pseudomonas kairouanensis* sp. nov. and *Pseudomonas nabeulensis* sp. nov. *Systematic and Applied Microbiology*. DOI: 10.1016/J.SYAPM.2019.03.002
- Pakianathan, S.W., Haridas, G. and d'Auzac, J. (1989). Water relations and latex flow. In *Physiology of rubber tree latex*, CRC Press, Inc, Florida.
- Papke, R. T., White, E., Reddy, P., Weigel, G., Kamekura, M., Minegishi, H., & Ventosa, A. (2011). A multilocus sequence analysis approach to the phylogeny and taxonomy of the Halobacteriales. *International Journal of Systematic and Evolutionary Microbiology*, 61(12), 2984-2995. PMID: 21296924 DOI: 10.1099/ijs.0.029298-0
- Parvathi, A., Krishna, K., Jose, J., Joseph, N., & Nair, S. (2009). Biochemical and molecular characterization of *Bacillus pumilus* isolated from coastal environment in Cochin, India. *Brazilian Journal of Microbiology*, 40(2), 269-275.
- Pasanen, T., Koskela, S., Mero, S., Tarkka, E., Tissari, P., Vaara, M., & Kirveskari, J. (2014). Rapid molecular characterization of *Acinetobacter baumannii* clones with rep-PCR and evaluation of carbapenemase genes by new multiplex PCR in Hospital District of Helsinki and Uusimaa. *PloS One*, 9(1), e85854.
- Pascual, J., Macián, M. C., Arahál, D. R., Garay, E., & Pujalte, M. J. (2010). Multilocus sequence analysis of the central clade of the genus *Vibrio* by using the 16S rRNA, *recA*, *pyrH*, *rpoD*, *gyrB*, *rctB* and *toxR* genes. *International Journal of Systematic and Evolutionary Microbiology*, 60(1), 154-165.
- Patil, H. J., Srivastava, A. K., Kumar, S., Chaudhari, B. L., & Arora, D. K. (2010). Selective isolation, evaluation and characterization of antagonistic actinomycetes against *Rhizoctonia solani*. *World Journal of Microbiology and Biotechnology*, 26(12), 2163-2170.
- Payne, G. W., Vandamme, P., Morgan, S. H., LiPuma, J. J., Coenye, T., Weightman, A. J., & Mahenthiralingam, E. (2005). Development of a *recA* gene-based identification approach for the entire *Burkholderia* genus. *Applied and Environmental Microbiology*, 71(7), 3917-3927.
- Peeters, C., Zlosnik, J. E., Spilker, T., Hird, T. J., LiPuma, J. J., & Vandamme, P. (2013). *Burkholderia pseudomultivorans* sp. nov., a novel *Burkholderia cepacia* complex species from human respiratory samples and the rhizosphere. *Systematic and Applied Microbiology*, 36(7), 483-489.
- Peng, Q., Yuan, Y., & Gao, M. (2013). *Bacillus pumilus*, a novel ginger rhizome rot pathogen in China. *Plant Disease*, 97(10), 1308-1315. DOI: 10.1094/PDIS-12-12-1178-RE

- Pérez-García, A., Romero, D., & De Vicente, A. (2011). Plant protection and growth stimulation by microorganisms: biotechnological applications of Bacilli in agriculture. *Current Opinion in Biotechnology*, 22(2), 187-193.
- Priyadarshan, P. M., and de Gonçalves, P. S. (2003). *Hevea* gene pool for breeding. *Genetic Resources and Crop Evolution*, 50, 101–114. DOI:10.1023/A:1022972320696.
- Purseglove, J. W. (1987). Tropical Crops: Dicotyledons Longman Group Ltd. *Burnt Hill*.
- Rademaker, J. L., de Bruijn F. J. (1997). Characterization and classification of microbes by rep-PCR genomic fingerprinting and computer assisted pattern analysis. *DNA markers: Protocols, Applications, and Overviews*, 151-171.
- Rademaker, J.L.W., Louws, F.J., Versalovic, J.V., and de Bruijn, F.J. (2008) Characterization of the diversity of ecologically important microbes by rep-PCR genomic fingerprinting. In *Molecular Microbial Ecology Manual*. Kowalchuck, G.A., de
- Rahman, A. Y. A., Usharraj, A. O., Misra, B. B., Thottathil, G. P., Jayasekaran, K., Feng, Y., & Tan, H. S. (2013). Draft genome sequence of the rubber tree *Hevea brasiliensis*. *BMC Genomics*, 14(1), 1-15. DOI: 10.1186/1471-2164-14-75.
- Rajput, R., & Gupta, R. (2014). Expression of *Bacillus pumilus* keratinase rK 27 in *Bacillus subtilis*: enzyme application for developing renewable flocculants from bone meal. *Annals of Microbiology*, 64(3), 1257-1266.
- Ratnasingam J, Ioras F, Kaner J, Wenming L (2011). Sustainability of the rubberwood sector in Malaysia. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 39(2), 305–311. DOI: 10.15835/nbha3927195
- Ratnasingam, J., Ramasamy, G., Ioras, F., Kaner, J., & Wenming, L. (2012). Production potential of rubberwood in Malaysia: Its economic challenges. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 40(2), 317-322.
- Reyes-Ramirez, A., & Ibarra, J. E. (2005). Fingerprinting of *Bacillus thuringiensis* type strains and isolates by using *Bacillus cereus* group-specific repetitive extragenic palindromic sequence-based PCR analysis. *Application Environment Microbiology*, 71(3), 1346-1355.
- Ross, T. L., Merz, W. G., Farkosh, M., & Carroll, K. C. (2005). Comparison of an automated repetitive sequence-based PCR microbial typing system to pulsed-field gel electrophoresis for analysis of outbreaks of methicillin-resistant *Staphylococcus aureus*. *Journal of Clinical Microbiology*, 43(11), 5642-5647. doi: 10.1128/JCM.43.11.5642-5647.2005.

- Roy, M., Saha, A., & Roy, M. (2014). Ecological impact of rubber plantations: Tripura perspective. *International Journal of Current Research*, 6(11), 10334-10340.
- Rüger, H. J. (1989). Benthic studies of the northwest African upwelling region: psychrophilic and psychrotrophic bacterial communities from areas with different upwelling intensities. *Marine Ecology Progress Series*, 57, 45-52.
- Sakdapipanich, J.T. (2007). Structural characterization of natural rubber based on recent evidence from selective enzymatic treatments. *Journal of Bioscience and Bioengineering*, 103, 287-292. DOI:10.1263/jbb.103.287.
- Saleh, O. I., Huang, P. Y., & Huang, J. S. (1997). *Bacillus pumilus*, the cause of bacterial blotch of immature balady peach in Egypt. *Journal of Phytopathology*, 145(10), 447-453.
- Sambrook, J., Fritsch, E. F., & Maniatis, T. (1989). Molecular cloning: a laboratory manual (2nd Editio). New York: *Cold Spring Harbor Laboratory Press*.
- Saranya, P., Kumari, H. S., Rao, B. P., & Sekaran, G. (2014). Lipase production from a novel thermo-tolerant and extreme acidophile *Bacillus pumilus* using palm oil as the substrate and treatment of palm oil-containing wastewater. *Environmental Science and Pollution Research*, 21(5), 3907-3919.
- Satomi, M., La Duc, M. T., & Venkateswaran, K. (2006). *Bacillus safensis* sp. nov., isolated from spacecraft and assembly-facility surfaces. *International Journal of Systematic and Evolutionary Microbiology*, 56(8), 1735-1740.
- Sawabe, T., Ogura, Y., Matsumura, Y., Feng, G., Amin, A. R., Mino, S. & Hayashi, T. (2013). Updating the *Vibrio* clades defined by multilocus sequence phylogeny: proposal of eight new clades, and the description of *Vibrio tritonius* sp. nov. *Frontiers in Microbiology*, 4, 414. DOI: 10.3389/fmicb.2013.00414
- Sharib, S., & Halog, A. (2017). Enhancing value chains by applying industrial symbiosis concept to the Rubber City in Kedah, Malaysia. *Journal of Cleaner Production*, 141, 1095-1108.
- Song, J. H., Wu, Z. R., Zhang, L. X., Tan, G. J., Wang, S., & Wang, J. J. (2018). First Report of *Bacillus pumilus* Causing Fruit Rot on Muskmelon (*Cucumis melo*) in China. *Plant Disease*, 102(2), 439-439. DOI: 10.1094/PDIS-08-17-1169-PDN.
- Stahl, D. A. (1991). Development and application of nucleic acid probes. *Nucleic Acid Techniques in Bacterial Systematics*, 205-248.

- Suárez-Moreno, Z. R., Vinchira-Villarraga, D. M., Vergara-Morales, D. I., Castellanos, L., Ramos, F. A., Guarnaccia, C. & Moreno-Sarmiento, N. (2019). Plant-Growth promotion and biocontrol properties of three *Streptomyces* spp. isolates to control bacterial rice pathogens. *Frontiers in Microbiology*, 10, 290. DOI: 10.3389/fmicb.2019.00290.
- Suominen, I., Andersson, M. A., Andersson, M. C., Hallaksela, A. M., Kämpfer, P., Rainey, F. A., & Salkinoja-Salonen, M. (2001). Toxic *Bacillus pumilus* from indoor air, recycled paper pulp, Norway spruce, food poisoning outbreaks and clinical samples. *Systematic and Applied Microbiology*, 24(2), 267-276.
- Teoh, Y. P., Don, M. M., & Ujang, S. (2011). Assessment of the properties, utilization, and preservation of rubberwood (*Hevea brasiliensis*): a case study in Malaysia. *Journal of Wood Science*, 57(4), 255-266.
- Thomas, L., Ushasree, M. V., & Pandey, A. (2014). An alkali-thermostable xylanase from *Bacillus pumilus* functionally expressed in *Kluyveromyces lactis* and evaluation of its deinking efficiency. *Bioresource Technology*, 165, 309-313.
- Thomson. (2017, January). Retrieved from Rubber diseases: <http://rubberdisease.blogspot.com/>
- Thwaites, R., Mansfield, J., Eden-Green, S., & Seal, S. (1999). RAPD and rep PCR-based fingerprinting of vascular bacterial pathogens of *Musa* spp. *Plant Pathology*, 48(1), 121-128.
- Tian, R.M., Cai, L., Zhang, W. P., Cao, H. L., and Qian, P. Y. (2015). Rare events of intragenus and intraspecies horizontal transfer of the 16S rRNA gene. *Genome Biological Evolution*. 7, 2310–2320. DOI: 10.1093/gbe/evv143
Biol. Evol. 7, 2310–2320. DOI: 10.1093/gbe/evv143
- Timilsina, S., Jibrin, M. O., Potnis, N., Minsavage, G. V., Kebede, M., Schwartz, A., & Pruvost, O. (2015). Multilocus sequence analysis of *Xanthomonads* causing bacterial spot of tomato and pepper plants reveals strains generated by recombination among species and recent global spread of *Xanthomonas gardneri*. *Application of Environment Microbiology*, 81(4), 1520-1529.
- Van Beilen, J. B., & Poirier, Y. (2007). Establishment of new crops for the production of natural rubber. *TRENDS in Biotechnology*, 25(11), 522-529. DOI: 10.1016/j.tibtech.2007.08.009.
- Van Belkum, A., Scherer, S., Van Alphen, L., & Verbrugh, H. (1998). Short-sequence DNA repeats in prokaryotic genomes. *Microbiology and Molecular Biology Reviews*, 62(2), 275-293.
- Vandamme, P., Henry, D., Coenye, T., Nzula, S., Vancanneyt, M., LiPuma, J. J., & Mahenthiralingam, E. (2002). *Burkholderia anthina* sp. nov. and *Burkholderia pyrrocinia*, two additional *Burkholderia cepacia* complex

- bacteria, may confound results of new molecular diagnostic tools. *FEMS Immunology & Medical Microbiology*, 33(2), 143-149.
- Vandamme, P., Holmes, B., Coenye, T., Goris, J., Mahenthiralingam, E., LiPuma, J. J., & Govan, J. R. (2003). *Burkholderia cenocepacia* sp. nov.- a new twist to an old story. *Research in Microbiology*, 154(2), 91-96.
- Vandamme, P., Holmes, B., Vancanneyt, M., Coenye, T., Hoste, B., Coopman, R., & Govan, J. R. W. (1997). Occurrence of Multiple Genomovars of *Burkholderia cepacia* in Cystic Fibrosis Patients and Proposal of *Burkholderia multivorans* sp. nov. *International Journal of Systematic and Evolutionary Microbiology*, 47(4), 1188-1200.
- Vandamme, P., Mahenthiralingam, E., Holmes, B., Coenye, T., Hoste, B., De Vos, P., & Speert, D. P. (2000). Identification and population structure of *Burkholderia stabilis* sp. nov. (formerly *Burkholderia cepacia* genomovar IV). *Journal of Clinical Microbiology*, 38(3), 1042-1047.
- Venkatachalam, P., Geetha, N., Sangeetha, P., & Thulaseedharan, A. (2013). Natural rubber producing plants: An overview. *African Journal of Biotechnology*, 12(12).
- Vera Cruz, C. M., Halda-Alija, L., Louws, F. J., Skinner, D. Z., George, M. L., Nelson, R. J., & Leach, J. E. (1995). Repetitive sequence-based polymerase chain reaction of *Xanthomonas oryzae* pv. *oryzae* and *Pseudomonas* species. *International Rice Research Notes*, 20, 23-24.
- Versalovic, J., Koeuth, T., & Lupski, R. (1991). Distribution of repetitive DNA sequences in eubacteria and application to fingerprinting of bacterial genomes. *Nucleic Acids Research*, 19(24), 6823-6831.
- Versalovic, J., Schneider, M., De Bruijn, F. J., & Lupski, J. R. (1994). Genomic fingerprinting of bacteria using repetitive sequence-based polymerase chain reaction. *Methods in Molecular and Cellular Biology*, 5(1), 25-40.
- Vettath, V. K., Junqueira, A. C. M., Uchida, A., Purbojati, R. W., Houghton, J. N., Chénard, C., ... & Lau, K. J. (2017). Complete genome sequence of *Bacillus altitudinis* type strain SGAir0031 isolated from tropical air collected in Singapore. *Genome Announcements*, 5(45).
- Vinuesa, P. (2010). "Multilocus sequence analysis and bacterial species phylogeny" in *Molecular Phylogeny of Microorganisms*, eds O. Aharon and R. Thane Papke (Caister: Academic Press), 45-64.
- Vinuesa, P., Rojas-Jiménez, K., Contreras-Moreira, B., Mahna, S. K., Prasad, B. N., Moe, H., & Werner, D. (2008). Multilocus sequence analysis for assessment of the biogeography and evolutionary genetics of four *Bradyrhizobium* species that nodulate soybeans on the Asiatic continent. *Application of Environment Microbiology*, 74(22), 6987-6996.

- Waleron, M., Misztak, A., Jońca, J., & Waleron, K. (2019). First Report of *Pectobacterium polaris* causing soft rot of potato in Poland. *Plant Disease*, 103(1), 144–144. DOI: 10.1094/PDIS-05-18-0861-PDN
- Watson, G.A. (1989). Climate and soil. In Webster C.C. and W.J. Baulkwill (ed) Rubber. Longman Singapore Publishers (Pte) Ltd, Singapore, pp125-164.
- Webster, C.C., Paardekooper, E. C. (1989). The botany of the rubber tree. In Webster CC, Baulkwill WJ Rubber. Longman Singapore Publishers (Pte) Ltd, Singapore, pp57-84
- Whatmore, A. M., Koylass, M. S., Muchowski, J., Edwards-Smallbone, J., Gopaul, K. K., & Perrett, L. L. (2016). Extended multilocus sequence analysis to describe the global population structure of the genus *Brucella*: phylogeography and relationship to biovars. *Frontiers in Microbiology*, 7, 2049.
- Woese, C. R. (1987). Bacterial evolution. *Microbiological reviews*, 51(2), 221-271.
- Woese, C. R., & Fox, G. E. (1977). Phylogenetic structure of the prokaryotic domain: the primary kingdoms. *Proceedings of the National Academy of Sciences*, 74(11), 5088-5090.
- Wongcharoen, A. (2010). *Biological activities of soils under rubber trees (Hevea brasiliensis) and interactions with trunk phloem necrosis* (Doctoral dissertation, Université Pierre et Marie Curie-Paris VI).
- Wu, T., Xu, J., Liu, J., Guo, W. H., Li, X. B., Xia, J. B., & Wang, R. Q. (2019). Characterization and initial application of endophytic *Bacillus safensis* strain ZY16 for improving phytoremediation of oil-contaminated saline soils. *Frontiers in Microbiology*, 10, 991. DOI: 10.3389/fmicb.2019.00991 PMID: 31134029
- Wycherley, P. R. (1992). The genus *Hevea*: botanical aspects. *Natural Rubber: Biology, Cultivation and Technology*, 50-66.
- Yahiaoui, N., Chéron, J.-J., Ravelomanantsoa, S., Hamza, A. A., Petrouse, B., Jeetah, R. & Poussier, S. (2017). Genetic Diversity of the *Ralstonia solanacearum* Species Complex in the Southwest Indian Ocean Islands. *Frontiers in Plant Science*, 8, 2139. DOI: 10.3389/fpls.2017.02139
- Yarza, P., Yilmaz, P., Pruesse, E., Glöckner, F. O., Ludwig, W., Schleifer, K. H., & Rosselló-Móra, R. (2014). Uniting the classification of cultured and uncultured bacteria and archaea using 16S rRNA gene sequences. *Nature Reviews Microbiology*, 12(9), 635.
- Yi, H. S., Yang, J. W., & Ryu, C. M. (2013). ISR meets SAR outside: additive action of the endophyte *Bacillus pumilus* INR7 and the chemical inducer,

benzothiadiazole, on induced resistance against bacterial spot in field-grown pepper. *Frontiers in Plant Science*, 4, 122.

Yijun, K. A. N. G., Min, S. H. E. N., Huanli, W. A. N. G., Qingxin, Z. H. A. O., & Shixue, Y. I. N. (2012). Biological control of tomato bacterial wilt caused by *Ralstonia solanacearum* with *Erwinia persicinus* RA2 and *Bacillus pumilus* WP8. *Chinese Journal of Biological Control*, 28(2), 255.

Yuan, Y., & Gao, M. (2015). Genomic analysis of a ginger pathogen *Bacillus pumilus* providing the understanding to the pathogenesis and the novel control strategy. *Scientific Reports*, 5, 10259.

Zarawi, A.G. *Performance of RRIM 3001*. Paper presented at the meeting of the Seminar RRIM 3001, Akademi Hevea Malaysia. July 2018.

Zarei, S., Taghavi, S. M., Banihashemi, Z., Hamzehzarghani, H., & Osdaghi, E. (2019). Etiology of leaf spot and fruit canker symptoms on stone fruits and nut trees in Iran. *Journal of Plant Pathology*, 1–10. DOI: 10.1007/s42161-019-00283-w

Zhang, L., Shi, Y., Wu, Z., & Tan, G. (2018). Characterization of response regulator GacA involved in phaseolotoxin production, hypersensitive response and cellular processes in *Pseudomonas syringae* pv. *actinidiae* A18. *Physiological and Molecular Plant Pathology*, 103, 137–142. DOI: 10.1016/J.PMPP.2018.07.001

Zulperi, D., Sijam, K., Ahmad, Z. A. M., Awang, Y., Ismail, S. I., Asib, N., & Hata, E. M. (2016). Genetic diversity of *Ralstonia solanacearum* phylotype II sequevar 4 strains associated with Moko disease of banana (*Musa* spp.) in Peninsular Malaysia. *European Journal of Plant Pathology*, 144(2), 257-270.