



**USING SILICATE SOLUBILIZING BACTERIA AS BIOCONTROL AGENT TO  
SUPPRESS *Rigidoporus microporus* CAUSING WHITE ROOT ROT  
DISEASE OF RUBBER**

**By**

**NURUL SHAKIRAH BINTI AYOB**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of Master of  
Science**

**December 2021**

**FP 2021 74**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**USING SILICATE SOLUBILIZING BACTERIA AS BIOCONTROL AGENT TO SUPPRESS *Rigidoporus microporus* CAUSING WHITE ROOT ROT DISEASE OF RUBBER**

By

**NURUL SHAKIRAH BINTI AYOB**

**December 2021**

**Chair : Mohd Yusof Bin Abd Samad, PhD**  
**Faculty : Agriculture**

White root rot disease (WRD) caused by *Rigidoporus microporus* is the most prevalent disease in rubber plantation. Current treatment using chemicals has caused environmental pollution and health hazard. An alternative curative for WRD using microbes to degrade naturally occurring silica (Si) in soil has potential to reduce disease incidence of WRD. Si is a beneficial element for plant growth and its amount in soil is abundant. However, Si availability is relatively low due to strong bonding with other elements in soil. The most important aspect in this research is to make full use of the role of microbes to mineralize silicon in soil for plant uptake against stresses despite promoting plant growth. Thus, the goals of this research were: (i) to isolate and screen potential microbes for its ability in mineralizing silicon, suppressing fungal growth and secreting plant growth promoting (PGP) traits, (ii) to study mechanisms of silicon solubilization by bacteria and (iii) to evaluate efficacy of silicate solubilizing bacteria (SSB) applied at different rates on determination of silicon uptake, inhibition of *R. microporus*, stimulation of induced systemic resistance (ISR) and enhancement of plant growth. To reveal the ability of the potential microbes on reducing disease occurrence of WRD, soil samples under rubber rhizosphere of various ages were collected to select bacteria with silicate solubilizing property, antagonism and plant growth promotion traits. Mechanisms underlying silicate solubilization by selected isolates were conducted. Finally, rubber seedling infected with *R. microporus* was challenged with SSB applied at different rates for 16 weeks to evaluate its effectiveness on reducing disease incidence, solubilizing natural silica in soil and promoting plant growth. Results showed that 3 isolates identified by 16S rRNA as *Bacillus* sp. strain NSAMYKJ16 (SSB16), *Proteus* sp. strain NSAMYKJ18 (SSB18) and *Bacillus* sp. strain NSAMYKJ21 (SSB21) were able to mineralize insoluble silicate, inhibit growth of *R. microporus* and produce PGP traits. It was found that all SSB were able to dissociate silicate minerals up to 14 days with

reduction in pH of the cultured medium. Tartaric and succinic acids were major acids involved in solubilization process of silicate minerals by all SSB. In the study of the effect of pH, it was found that all SSB released maximum silicic acid at pH 9 tested on quartz. Meanwhile, magnesium trisilicate was best dissociates only by SSB18 observed at pH 3, pH 6 and pH 9. Based on the ability of SSB to suppress *R. microporus*, results showed that treatment SSB18 applied at 25 mL/seedling (D25) had the lowest disease incidence compared to 50 mL/seedling (D50) and 75 mL/seedling (D75). Meanwhile, SSB16 and SSB21 showed the lowest disease incidence at 50 mL/seedling (D50) and 75 mL/seedling (D75) respectively. Total silica accumulation in plant treated with SSB16, SSB18 and SSB21 were the lowest at 50 mL/seedling (D50), 25 mL/seedling (D25) and 75 mL/seedling (D75) respectively. A correlation analysis between disease suppression and silicon content in root indicated positive association suggesting that disease occurrence increased linearly with silicon accumulation. Lignin content in rubber seedling treated with SSB16, SSB18 and SSB21 were higher in treatment D50 (2.88 mg LTGA g<sup>-1</sup>), D25 (2.82 mg LTGA g<sup>-1</sup>) and D75 (3.74 mg LTGA g<sup>-1</sup>) respectively. Similar trend on the increase of peroxidase enzyme was also shown by SSB16, SSB18 and SSB21 at D50 (4.25 unit mL<sup>-1</sup>), D25 (10.75 unit mL<sup>-1</sup>) and D75 (6.88 unit mL<sup>-1</sup>). Association between both ISR compounds with silicon content in root resulted in negative correlation. Similarly, reduction of disease was also strongly associated with the negative correlation with ISR compounds. Strong positive relationship between lignin and peroxidase reflected that stimulation of peroxidase due to the effect of silicic acid in plant affect lignin formation. Application of SSB significantly increased ( $P \leq 0.05$ ) plant growth attributes and fresh plant biomass were significantly affected by the rate with the highest shown by D50 treatment. Application of SSB increased plant nutrient availability particularly potassium with D50 and D75 rates. Hence, results of this study confirmed that organic acids and pH plays important role in silica dissociation and application of SSB had successfully reduced WRD incidence in rubber seedling.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**PENGUNAAN BAKTERIA PELARUT SILIKA SEBAGAI AGEN  
KAWALAN BIO UNTUK MERENCAT *Rigidoporus microporus*  
PENYEBAB PENYAKIT AKAR PUTIH TANAMAN GETAH**

Oleh

**NURUL SHAKIRAH BINTI AYOB**

Disember 2021

**Pengerusi : Mohd Yusoff Bin Abd Samad, PhD**  
**Fakulti : Pertanian**

Penyakit akar putih (WRD) disebabkan oleh kulat *Rigidoporus microporus* merupakan penyakit yang berleluasa dalam perladangan getah. Rawatan terkini menggunakan kimia menyebabkan pencemaran alam sekitar dan merisikokan kesihatan. Rawatan alternative bagi WRD menggunakan mikro untuk melarutkan silika (Si) semulajadi di dalam tanah berupaya mengurangkan insiden penyakit WRD. Si merupakan unsur bermanfaat bagi tumbesaran tanaman dan jumlahnya di dalam tanah adalah tinggi. Namun, kepadatan Si di dalam tanah adalah rendah kerana terikat oleh unsur lain di dalam tanah. Aspek yang paling utama di dalam kajian ini adalah untuk mengeksplorasi fungsi mikro bagi melarutkan silika di dalam tanah untuk pengambilan tanaman melawan tekanan selain meningkatkan tumbesaran tanaman. Justeru, objektif kajian ini adalah: (i) untuk memencil dan menyaring mikro yang berupaya melarutkan silika, merencat pertumbuhan kulat dan menghasilkan ciri-ciri penggalak pertumbuhan tanaman, (ii) untuk mengkaji mekanisma pelarutan silika oleh bakteria dan (iii) untuk menilai keberkesanan bakteria pelarut silika (SSB) yang diuji pada kadar berlainan bagi penentuan pengambilan silika, perencatan *R. microporus*, perangsangan aruhan rintangan sistemik (ISR) dan peningkatan pertumbuhan tanaman. Bagi membuktikan keupayaan mikro mengurangkan kejadian penyakit WRD, sampel tanah di kawasan akar pokok getah daripada pelbagai usia dikumpulkan bagi memilih bakteria yang mempunyai ciri pelarut silika, anatgonis, dan ciri-ciri perangsang pertumbuhan tanaman. Mekanisma disebalik pelarutan silika oleh bakteria kemudiannya dijalankan. Akhir sekali, anak pokok getah dijangkitkan dengan kulat *R. microporus*, dan dirawat dengan SSB pada kadar berbeza selama 16 minggu bagi menguji keberkesanannya mengurangkan penyakit, melarutkan silika semulajadi di dalam tanah dan merangsang tumbesaran pokok. Keputusan menunjukkan bahawa 3 pencilan yang dikenalpasti menggunakan 16S rRNA sebagai *Bacillus* sp. strain NSAMYKJ16 (SSB16),

*Proteus* sp. strain NSAMYKJ18 (SSB18) dan *Bacillus* sp. strain NSAMYKJ21 (SSB21) berupaya melarutkan silika, merencat pertumbuhan *R. microporus* dan menghasilkan ciri-ciri perangsang pertumbuhan tanaman. Didapati kesemua strain berupaya melarutkan silika sehingga 14 hari dengan penurunan pH di dalam media kultur. Asid tartaric dan suksinik merupakan asid utama terlibat dalam proses pelarutan silika oleh kesemua SSB. Dalam kajian kesan pH ke atas pelarutan silika, didapati kesemua SSB menghasilkan asid silisik maksima pada pH 9 yang diuji ke atas kuartza. Sementara itu, pada magnesium trisilika, pelarutan yang paling baik adalah pada pH 3, pH 6 dan pH 9 yang kesemuanya ditunjukkan oleh SSB18. Berdasarkan keupayaan SSB untuk merencat *R. microporus*, keputusan menunjukkan rawatan SSB pada kadar 25 mL/anak pokok (D25) mempunyai insiden penyakit yang terendah berbanding 50 mL/anak pokok (D50) dan 75 mL/anak pokok (D75). Sementara itu, SSB16 dan SSB21 menunjukkan insiden penyakit yang terendah pada kadar rawatan masing-masing pada 50 mL/anak pokok (D50) dan 75 mL/anak pokok (D75). Jumlah silika terkumpul di dalam tanaman yang dirawat dengan SSB16, SSB18 dan SSB21 adalah terendah pada kadar rawatan masing-masing 50 mL/ anak pokok (D50), 25 mL/ anak pokok (D25) dan 75 mL/ anak pokok. Analisa korelasi antara rencatan penyakit dan jumlah silika di dalam akar menunjukkan hubungan yang positif yang mencadangkan bahawa kejadian penyakit meningkat secara selari dengan jumlah silika terkumpul. Kandungan lignin di dalam anak pokok getah dirawat dengan SSB16, SSB18 dan SSB21 masing-masing adalah tinggi pada kadar D50 (2.82 mg LTGA g<sup>-1</sup>), (2.88 mg LTGA g<sup>-1</sup>) dan D75 (3.74 mg LTGA g<sup>-1</sup>). Trend yang sama pada peningkatan enzim peroksida juga ditunjukkan oleh SSB16, SSB18 dan SSB21 pada kadar D50 (4.25 unit mL<sup>-1</sup>), D25 (10.75 unit mL<sup>-1</sup>) dan D75 (6.88 unit mL<sup>-1</sup>). Hubungan antara kedua-dua kompaun ISR dengan kandungan silika di dalam akar menunjukkan korelasi negatif. Trend serupa juga ditunjukkan oleh pengurangan penyakit dengan korelasi negatif antara pengurangan penyakit dan kompaun ISR. Hubungan positif yang kuat antara lignin dan enzim peroksida menunjukkan rangsangan enzim peroksida hasil daripada asid silisik memberi kesan kepada pembentukan lignin. Penggunaan SSB yang meningkatkan sifat-sifat pertumbuhan tanaman dan jisim segar dengan ketara ( $p \leq 0.05$ ) dipengaruhi oleh kadar aplikasi dengan bacaan tertinggi ditunjukkan oleh rawatan D50. Penggunaan SSB meningkatkan kepadatan nutrien tanaman khususnya potassium yang ditunjukkan pada kadar D50 dan D75. Justeru, keputusan kajian menyimpulkan dan mengesahkan asid organik dan pH berperanan penting dalam proses pelarutan silika dan penggunaan SSB telah berjaya mengurangkan kejadian penyakit WRD pada anak pokok getah.



## ACKNOWLEDGEMENTS

*In the name of Allah, the most Gracious and the most Merciful.*

All praises be to Allah and His blessing for the completion of this thesis. Without His blessing, this journey could not be completed.

I wish to express my deepest appreciation to my supervisor, Associate Professor Dr. Mohd Yusoff Abd Samad, the chairman of the Supervisory Committee, for his invaluable expertise, guidance, diligent advice, helpfulness and profound belief in my abilities throughout my study and preparation of this thesis. I honour his patience, positive encouragement and warm spirit throughout the process.

I am extremely grateful to Associate Professor Dr. Zulkefly Sulaiman and Dr. Siti Izera Ismail, members of the supervisory committee, for their invaluable insight into this research project, helpful contributions and guidance in my research studies, papers and preparation of this thesis. Without their persistent help, this dissertation would not have been possible.

I gratefully acknowledged the Public Service Department Malaysia for awarding me the scholarship towards my master degree. I would also like to recognize the Institute of Plantation Studies for the grant funding that I received for my research.

I greatly appreciate the assistance that I received from all laboratory staffs and officers in the Department of Land Management, especially to Mr. Dzulkifli Duaji, Madam Norizah Mohd Yusof and Madam Nooraishah Abdul Aziz.

A heartfelt thank you to my parents Ayob Abdullah and Tumini Kijo, my husband (Muhamad Fahmie Hairudin), my son (Khalid Affan) and my sisters (Nurul Sakinah and Nurul A'isyah) whom have always believed in me and proud of me. I really appreciate and indebted for their endless prayers, supports, patience, loves and motivations in pursuing my master degree that made my dream to come true.

Last but not least, my special thanks to all my colleagues, for being so helpful in numerous ways, understanding, emotionally and physically supportive throughout the process of completing my research and has made my postgraduate life alive.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Mohd Yusoff Bin Abd Samad, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Zulkefly Bin Sulaiman, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

**Siti Izera Binti Ismail, PhD**

Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 13 April 2023



## Declaration by the Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and the copyright of the thesis are fully-owned by Universiti Putra Malaysia, as stipulated in the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from the supervisor and the office of the Deputy Vice-Chancellor (Research and innovation) before the thesis is published in any written, printed or electronic form (including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials) as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld in accordance with the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Nurul Shakirah Binti Ayob

## Declaration by Members of the Supervisory Committee

This is to confirm that:

- the research and the writing of this thesis were done under our supervision;
- supervisory responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) are adhered to.

Signature: \_\_\_\_\_  
Chairman of  
Supervisory  
Committee: Mohd Yusoff Abd Samad, PhD  
\_\_\_\_\_

Signature: \_\_\_\_\_  
Member of  
Supervisory  
Committee: Zulkefly Bin Sulaiman, PhD  
\_\_\_\_\_

Signature: \_\_\_\_\_  
Member of  
Supervisory  
Committee: Siti Izera Binti Ismail, PhD  
\_\_\_\_\_

## TABLE OF CONTENTS

		<b>Page</b>
<b>ABSTRACT</b>		i
<b>ABSTRAK</b>		iii
<b>ACKNOWLEDGEMENTS</b>		v
<b>APPROVAL</b>		vi
<b>DECLARATION</b>		viii
<b>LIST OF TABLES</b>		xv
<b>LIST OF FIGURES</b>		xvii
<b>LIST OF ABBREVIATIONS</b>		xx
<b>CHAPTER</b>		
<b>1</b>	<b>INTRODUCTION</b>	1
<b>2</b>	<b>LITERATURE REVIEW</b>	4
	2.1 Rubber	4
	2.1.1 Rubber Cultivation in Malaysia	4
	2.1.2 Economic Importance of Rubber in Rubber-Based Industry	5
	2.2 White Root Disease of Rubber	7
	2.2.1 Characteristics of <i>Rigidoporus microporus</i> as Causal Agent of WRD	9
	2.2.2 Disease Symptoms	9
	2.2.3 Mechanisms of Disease Infection	10
	2.2.4 Disease Management	10
	2.3 Silicon	13
	2.3.1 Roles of Silica on Plant Growth and Disease Resistance	13
	2.3.2 Silicon Uptake, Deposition and Accumulation in Higher Plant	15
	2.3.3 Availability of Silica in Soil and Factors Affecting Its Solubilisation	16
	2.4 Silicate Solubilizing Bacteria and Its Roles	19
	2.4.1 Mechanisms of Silica Solubilization by Silicate Solubilizing Bacteria	20
	2.4.2 Ability of SSB as Plant Growth Promoting Rhizobacteria (PGPR)	21
<b>3</b>	<b>ISOLATION, SCREENING AND IDENTIFICATION OF BENEFICIAL BACTERIA RESPONSIBLE FOR SILICATE SOLUBILIZATION, BIOCONTROL AND PLANT GROWTH PROMOTION</b>	23
	3.1 Introduction	23
	3.2 Materials and Methods	24
	3.2.1 Isolation of Silicate Solubilizing Bacteria	24

3.2.2	Qualitative Screening of Silicate Solubilizing Bacteria Using Plate Assay	24
3.2.3	Nitrogen Fixation, Phosphate and Potassium Solubilization	25
3.2.4	Antagonism by Dual Culture Assay	25
3.2.5	Volatile Compound	26
3.2.6	Siderophore	26
3.2.7	Production of Hydrolyzing Enzymes Chitinase and Glucanase	27
3.2.8	Indole-3-Acetic Acid (IAA) Production	28
3.2.9	Characterization of Selected Silicate Solubilizing Bacteria	28
3.2.9.1	Gram Staining	28
3.2.9.2	Spore Staining	29
3.2.9.3	Motility Test by Hanging Drop Technique	29
3.2.9.4	Identification of Selected SSB by 16S rRna Sequencing	29
3.2.10	Determination of Bacterial Growth Curve	29
3.2.11	Experimental Design	30
3.2.12	Statistical Analysis	30
3.3	Results and Discussion	30
3.3.1	Isolation of Bacteria	30
3.3.2	First Screening for Solubilization of Silica, Phosphate and Potassium, Antagonism and Nitrogen Fixation	32
3.3.3	Mechanisms of Biological Control Through Volatile Compound, Siderophore, Chitinase and Glucanase Production in Qualitative Assay	37
3.3.4	Quantification of Siderophore in Liquid Assay of Selected SSB	41
3.3.5	Quantification of IAA Produced by Selected SSB	43
3.3.6	Determination of Bacterial Growth Curve	44
3.3.7	Identification of Selected SSB by 16S rRna	45
3.4	Conclusions	47
<b>4</b>	<b>EVALUATION OF SILICATE SOLUBILIZATION BY SILICATE SOLUBILIZING BACTERIA</b>	<b>48</b>
4.1	Introduction	48
4.2	Materials and Methods	50
4.2.1	Quantification of Silicic Acid Solubilized by SSB in Liquid Broth of Different Silica Sources	50
4.2.2	Effect of Incubation Period on Solubility of Silicate Minerals	50

4.2.3	Determination of Organic Acids Produced During Solubilization of Silicate	51
4.2.4	Effect of pH on Silicate Solubilizing Activity	51
4.2.5	Attachment of Microbial Inoculum on Silicate Surface by SEM	51
4.2.6	Experimental Design	52
4.2.7	Statistical Analysis	52
4.3	Results and Discussion	52
4.3.1	Quantification of Silicate Solubilizing Bacteria In Liquid Assay	52
4.3.2	Effect of Incubation Period on Solubility of Different Silicate Mineral by Selected SSB	53
4.3.3	Determination of Organic Acids Produced During Solubilization of Silicate Mineral	57
4.3.4	Effect of pH on Silicate Solubilizing Activity	59
4.3.5	Attachment of Microbial Inoculum on Silica Surface by SEM	63
4.4	Conclusions	64
<b>5</b>	<b>ROLE OF SILICATE SOLUBILIZING BACTERIA ON SUPPRESSION OF WHITE ROOT DISEASE AND PLANT GROWTH PROMOTION OF <i>Hevea brasiliensis</i> UNDER RAIN SHELTER HOUSE</b>	<b>66</b>
5.1	Introduction	66
5.2	Materials and Methods	68
5.2.1	Study Site	68
5.2.2	Soil Preparation	68
5.2.2.1	Determination of Soil Texture by Mechanical Analysis	68
5.2.2.2	Determination of Total Silica in Soil by Gravimetric Method	69
5.2.2.3	Determination of Soil Chemical Properties	70
5.2.3	Planting Materials	70
5.2.4	Agnomical Practices	70
5.2.5	Preparation of Fungal Inoculum on Rubber Wood Block	70
5.2.6	Infestation of Fungal Inoculum on Rubber Seedlings	71
5.2.7	Preparation of Microbial Inoculum Treatment	71
5.2.8	Experimental Design and Treatments	71

5.2.9	Measurement of Vegetative Growth Parameters	73
5.2.9.1	Plant Height of Rubber Seedling	73
5.2.9.2	Girth of Rubber Seedling	74
5.2.9.3	Total Number of Leaves and Chlorophyll Content	74
5.2.10	Determination of Fresh Weight of Rubber Seedling	74
5.2.11	Disease Assessment	74
5.2.11.1	Disease Incidence	74
5.2.11.2	Disease Severity Index (DSI) on Foliar and on Root	75
5.2.11.3	Area Under Diseases Progression Curve and Rate Area Under Disease Progression Curve	76
5.2.11.4	Disease Reduction	76
5.2.12	Effect of SSB on The Production of Plant Secondary Metabolites	77
5.2.12.1	Peroxidase	77
5.2.12.2	Lignin	77
5.2.13	Plant Nutrient Analysis	78
5.2.13.1	Plant Sample Preparation	78
5.2.13.2	Determination of Plant Total Nitrogen (N) and Total Carbon (C)	79
5.2.13.3	Determination of Phosphorus (P) and Potassium (K) in Plant	79
5.2.13.4	Determination of Silica Content in Plant	79
5.2.14	Soil Chemical Analysis	80
5.2.14.1	Soil Preparation	80
5.2.14.2	Determination of Total Nitrogen and Organic Carbon	80
5.2.14.3	Determination of Total Phosphorus	81
5.2.14.4	Determination of Available Phosphorus	81
5.2.14.5	Determination of Cation Exchange Capacity (CEC), Exchangeable Potassium, Calcium, Magnesium, Sodium and Base Saturation of Soil	81
5.2.14.6	Determination of Soil pH	83
5.2.15	Visualization of Silica Body in Plant Cell Wall Under SEM	83
5.2.16	Statistical Analysis	83

5.3	Results and Discussion	84
5.3.1	Evaluation of Silicate Solubilizing Bacteria as Biocontrol Agents for White Rot Root Disease Under Rain Shelter	84
5.3.1.1	Suppression of <i>Rigidoporus Microporus</i> by Selected SSB and Its Association With Silicon Content	84
5.3.1.2	Effect of Silicate Solubilizing Bacteria on Silicon Accumulation in Root And Its Association on Disease Reduction	90
5.3.1.3	Stimulation of Plant Defence and Compound and Its Association with Silicon Accumulation in Root and Disease Suppression	93
5.3.1.4	Effect of Silicate Solubilizing Bacteria on Crop Growth Under Biotic Stress	96
5.3.1.5	Effect of Silicate Solubilizing Bacteria on Plant Nutrient Uptake	101
5.3.2	Evaluation of Silicate Solubilizing Bacteria on Chemical Properties of Soil and Nutrient Content	103
5.3.2.1	Soil Texture, Chemical Properties and Nutritional Content for Potting Media	103
5.3.2.2	Effect of Treatment on Soil Chemical Properties	104
5.3.2.3	Effect of Treatment on Soil Nutrients Properties	107
5.4	Conclusion	110
<b>6</b>	<b>SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>112</b>
	<b>REFERENCES</b>	<b>114</b>
	<b>APPENDICES</b>	<b>142</b>
	<b>BIODATA OF STUDENT</b>	<b>173</b>
	<b>PUBLICATION</b>	<b>174</b>



## LIST OF TABLES

Table		Page
2.1	Distribution and severity of white root disease in Asian's rubber producing countries	8
2.2	Microbial strains inducing systemic resistance for controlling white root rot disease of rubber	22
3.1	Bacterial isolation from different zone of rubber age	31
3.2	Growth of bacterial isolates on nitrogen fixation, silica, phosphate solubilization and antagonistic activity screened on plate assay	33
3.3	Percentage of effective isolates to produce volatile compound against <i>Rigidoporus micropus</i> based on respective PIRG values	38
3.4	Characterization of selected SSB based on gram staining, spore, motility, colour and shape of colony	47
4.1	Organic acids produced by SSB16, SSB18 and SSB21 after 21 days of incubation in liquid culture	59
5.1	Description of treatments	72
5.2	Disease scoring for disease severity index of white root disease of rubber based on foliar symptom	75
5.3	Disease scoring for disease severity index of white root disease of rubber based on root symptom	76
5.4	Percentage of disease severity on root symptom assessed at week 16	87
5.5	Area under disease progression curve (AUDPC) and disease reduction (DR) in rubber seedling inoculated with <i>Rigidoporus micropus</i> and treated with silicate solubilizing bacteria at different rates	88
5.6	Effect of different SSB strains applied at different rates on the accumulation of silicon deposited in the root	91
5.7	Relationship between silicon accumulation in root of rubber seedling in all silicate solubilizing bacteria and disease progression (AUDPC)	91

5.8	Relationship between silicon accumulation in root of rubber seedling and plant induced systemic resistance	94
5.9	Relationship between induced peroxidase and lignin content of rubber root seedling	96
5.10	Summary of ANOVA results on the effect of SSB strain and inoculum with its interaction on plant height and girth increment, number of leaf and chlorophyll content in rubber seedling from week 2 to week 16 after infection	97
5.11	Relationship between growth parameters and silicon accumulation in root of rubber seedling	98
5.12	Effect of silicate solubilizing bacteria applied at different rates on plant nutrient content in foliar	102
5.13	Soil physical and chemical properties of potting soil	103
5.14	Effect of treatment on soil pH	105
5.15	Effect of treatment on soil CEC and base saturation	106
5.16	Effect of treatment on soil organic carbon	107
5.17	Effect of treatment on total P, available P	108
5.18	Effect of treatment on soil total N, exchangeable K, Ca and Mg and C/N ratio	109

## LIST OF FIGURES

Figure		Page
2.1	Planted area of natural rubber in Malaysia under smallholder, estate and total area for year 1990 to 2018.	5
2.2	Malaysia's natural rubber production from 2001 to 2017.	7
2.3	Malaysia's natural rubber production from 2000 to 2017.	7
2.4	Symptom of <i>Rigidoporus micropus</i> infection on leaf, formation of white rhizomorph on above ground and formation of basidiocarp on rubber trunk.	10
2.5	Fraction of silicon in soils.	19
3.1	Scatter diagram of 46 isolated bacteria responsible for silica solubilization, antagonism, nitrogen fixation, phosphate and potassium solubilization grouped under component 1 and component 2 in multi-dimensional preference analysis plot of the PCA procedure based on the ability of isolates with all possible criterion.	37
3.2	Scatter diagram of 46 isolated bacteria responsible for siderophore, volatile metabolite and glucanase grouped under component 1 and component 2 in multi-dimensional preference analysis plot of the PCA procedure based on the ability of isolates with all possible criterion.	39
3.3	Production of siderophore by SSB16 (A), SSB18 (B) and SSB21 (C) at 24, 48 and 72 hours after incubation.	42
3.4	IAA production by selected SSB at 24, 48 and 72 hours after incubation.	44
3.5	Bacterial growth curve of SSB16, SSB18 and SSB21 in liquid nutrient broth during 120 hours incubation period.	45

3.6	Phylogenetic tree of selected silicate solubilizing bacteria constructed based on 16S rRNA gene sequence analysis of <i>Bacillus</i> sp. NSAMYKJ16 (A), <i>Proteus</i> sp. NSAMYKJ18 (B) and <i>Bacillus</i> sp. NSAMYKJ21 (C).	46
4.1	Mineralization of silica in liquid assay by selected SSB at 7 days after incubation.	53
4.2	Effect of incubation period on solubility of quartz and magnesium trisilicate respectively with the changes in pH of the medium inoculated with SSB16, SSB18 and SSB21 after 21 days of incubation.	55
4.3	Effect of pH on solubility of quartz and changes in pH of culture medium inoculated with SSB16 (A and B), SSB18 (E and F) and SSB21 (I and J) and on solubility of magnesium trisilicate and changes in pH of culture medium inoculated with SSB16 (C and D), SSB18 (G and H) and SSB21 (K and L) at weekly interval for 14 days of incubation.	61
4.4	Colonization of silicate solubilizing bacteria on the surface of silicate mineral under SEM in uninoculated (A), SSB16 (B), SSB18 (C) and SSB21 (D).	64
5.1	USDA soil textural triangle chart for soil texture classification.	69
5.2	Trend of disease incidence recorded on the rubber seedling inoculated with SSB strains of SSB16 (A), SSB18 (B) and SSB21 (C) applied at different inoculum rate.	85
5.3	Trend of disease severity recorded on the rubber seedling inoculated with SSB strains of SSB16 (A), SSB18 (B) and SSB21 (C) applied at different inoculum rate.	87
5.4	Accumulation of silicon in the form of silica body in cell wall of rubber seedling in (A) control (+pathogen –SSB), (B) SSB16 (+pathogen +SSB), (C) SSB18 (+pathogen +SSB), and (D) SSB21 (+pathogen +SSB) at 16 weeks after infection.	93

5.5	Concentration of peroxidase in parasitized root tissue of rubber seedling inoculated with beneficial SSB strains applied at different rates.	94
5.6	Concentration of lignin as lignothioglycolic acid (LTGA) in parasitized root tissue of rubber seedling inoculated with beneficial SSB strains applied at different rates.	95
5.7	Effect of treatments on fresh weight of rubber root seedling.	99
5.8	Effect of treatments on fresh weight of rubber shoot seedling.	100



## LIST OF ABBREVIATIONS

%	Percentage
µg	Microgram
µg/mL	Microgram per millilitre
µL	Microliter
µM	Micromolar
µm	Micrometer
°	Degree
°C	Degree celcius
A.R.	Analytical reagent
Al	Aluminium
Al-O	Aluminium oxide bond
AMP	Antimicrobial peptide
ANOVA	Analysis of variance
AUDPC	Area under disease progression curve
C	Carbon
C/N	Carbon to nitrogen ratio
C <sub>10</sub> H <sub>11</sub> NO <sub>5</sub> S	l-amino-2-naphthol-4-sulfonic acid
C <sub>8</sub> H <sub>18</sub> O	Octyl alcohol
Ca	Calcium
CEC	Cation exchange capacity
cfu	Colony forming unit
cm	Centimetre
cmol <sub>c</sub>	Centimole charge
DAI	Day after incubation

DI	Disease incidence
DR	Disease reduction
DS	Disease severity
DSI	Disease severity index
ED	Effective dose
Fe	Iron
FeO	Iron (II) oxide
g	Gram
g	G-force
H <sup>+</sup>	Hydrogen ion
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
H <sub>2</sub> SO <sub>4</sub>	Sulphuric acid
H <sub>4</sub> SiO <sub>4</sub>	Silicic acid
ha.	Hectare
HCl	Hydrochloric acid
HMWOAs	High molecular weight acids
HNO <sub>3</sub>	Nitric acid
IAA	Indole-3-acetic acid
ISR	Induced systemic resistance
K	Potassium
kbp	Kilo-base pair
KH <sub>2</sub> PO <sub>4</sub>	Potassium dihydrogen phosphate
L	Litre



LMWOAs	Low molecular weight acids
ln	Natural log
LTGA	Lignothioglycolic acid
m	Metre
M	Molar
Mg	Magnesium
Mg	Milligram
mL	Millilitre
Mm	Millimetre
mM	Millimolar
Mmol	Millimol
N	Nitrogen
Na	Sodium
NaOH	Sodium hydroxide
NH <sub>4</sub> F	Ammonium fluoride
NH <sub>4</sub> OAc	Ammonium acetate
nm	Nanometre
O.D.	Optical density
OH	Hydroxide
P	Phosphorus
PCA	Principal component analysis
PCR	Polymerase chain reaction
PDA	Potato dextrose agar
PIRG	Percentage inhibition rate growth

pmol	Picomole
POX	Peroxidase
PROC GLM	Procedure generalized linear model
psu	Percent siderophore unit
rpm	Revolution per minute
rRNA	ribosomal RNA
SEM	Scanning electron microscope
Si	Silicon
SI	Solubilization index
Si-O bond	Silicon oxide bond
SiO <sub>2</sub>	Silicon dioxide
SOC	Soil organic carbon
SSB	Silicate solubilizing bacteria
UV	Ultra violet
VOC	Volatile organic compound
w/v	Weight per volume
WRD	White root disease
Zn	Zinc

## CHAPTER 1

### INTRODUCTION

Rubber (*Hevea brasiliensis*) is one of the major plantation crop and main commodity export in Malaysia after oil palm. The cultivation of rubber had seen a progressed from year to year in the area planted with rubber after it was commercially established in 1904 (Setiawan et al., 2007) which was 2,400 hectares in the early 1900s to 218,900 hectares in 1910s. Later, rubber cultivation grown rapidly and became one of the key success in Malaysian economy which contributes to the gross domestic product (GDP) and national gross export up to 50% as well as becoming source of income for small holders. However, the area grown with rubber was reduced due to several constraints such as emergence of synthetic rubber and disturbance of its growth by plethora of diseases. Biotic stress affecting growth of rubber due to diseases had been a major concerned to researchers at global.

Like other crops, various pathogenic diseases influence rubber production (Faÿ et al., 2010). According to Wastie (1975), rubber is a plantation crop that subjected to a plethora of economically important pathological diseases. White root disease (WRD) caused by basidiomycetes fungus *Rigidoporus microporus* is one of the challenging disease posing an epidemic treat in rubber plantation. It is well known destructive disease that infect rubber growing countries in Cameroon, Nigeria, India, Thailand, Indonesia, Ivory Coast, Ghana, Gabon, Sri Lanka, Malaysia and West and Central Africa (Wastie, 1975; Wattanasilakorn et al., 2015). Guyot & Flori (2002) reported that the disease has caused extensive death of rubber and severe loss that lead to the reduction of the tree stand and yield of latex being produced.

In order to maintain the steady supply of natural rubber, preventing and controlling the occurrence of WRD is necessary to prevent the widespread of the disease to the neighbouring trees. Currently, numerous strategies such as breeding of resistant clone, management of cultural practices and application of chemical fungicides had been employed to control the disease. According to (Narayanan & Mydin K., 2012), the commercially cultivated rubber clones has a very narrow genetic base due to its originality produce from hybridization thus there are no known species of rubber clone reported to be resistant towards *R. microporus*. Despite breeding, proper clearing of the land by uprooting the soil and poisoning the old rubber tree before replanting had been known to reduce the disease occurrence but this method requires high cost for small holders. The emergence of chemical fungicide was known to be effective against white root disease but the drawback of chemical usage on environment and health hazard has gained awareness to find for an alternative treatment.

Application of microbes as in biological control in agricultural practice has been considered as a viable alternative to chemical fungicide that can conserve and sustain the environment (Ouda, 2014) which suppress the activities of one organism by one or more other organism. In case of biological control for white root disease of rubber, previous studies showed that *Trichoderma harzianum*, *Trichoderma hamatum* (Jayasuriya & Thennakoon, 2007), *Aspergillus niger*, *Chaetomium bostrychodes*, *Chaetomium cupreus* (Kaewchai & Soyong, 2010), *Hypocrea virens*, *Hypocrea jecorina*, *Hypocrea lixii*, *Trichoderma spirale* (Nicholas et al., 2015), and *Streptomyces* sp. (Nakaew et al., 2015) had significant effect on the reduction of the severity index of disease. Although biological control agents have the ability to suppress the disease, but the efficacy of the introduced biocontrol agent in the field must be fully understood since there are many interrelationships between the antagonist, pathogen, the host plant and other indigenous organisms in the soil. Therefore, selection of the antagonist should be stressed initially starting from isolation to screening of potential antagonist so that the selected biocontrol agent does not only reduce the severity of plant disease but also promote plant growth.

Soil plays a vital role in plant health and fertility as it provides nutrients and protect plant from pathogen. This undoubtedly is related to the complex nature of soil microbial communities in soil ecosystem. In natural soil ecosystem, the diversity and number of microbial communities is higher because of high biomass content compared to the agro-ecosystem soil which had been polluted and had low amount of organic matter. As a result, plant becomes weakening when it is infected with pathogen. Therefore, management practices are essential to reclaim soil microorganisms. Different microorganisms carried out different function and show different performance according to its specified host plant. Variety of microorganisms are available for biological control but not all microorganisms have the ability to produce antagonism, plant growth promoting properties and solubilize naturally occurring mineral in soil.

Currently, research on microorganism capable of solubilizing silicate mineral in soil and suppressing phytopathogenic fungi had gained attention by researchers worldwide. Silicate solubilizing bacteria like *Bacillus flexus*, *Bacillus mucilaginosus*, *Bacillus megaterium* and *Pseudomonas fluorescens* were known to have antagonistic, plant growth promoting properties and solubilized insoluble silica (Vasanthi et al., 2018). Silicon which is the second most abundant mineral in soil was found to be effective in controlling various pests and diseases caused by both fungi and bacteria in different plant species (Ma, 2004). However, most of the research conducted on dissolution and uptake of silica were tested on monocotyledonous plant like rice, wheat, maize and sugarcane. On the other hand, it is important to study the uptake of silica on dicotyledonous plant especially the perennial crop of woody stem. Nonetheless, silicon research had gained attention only in the last two decades (Nawaz et al., 2019). Though silicon was abundant in soil but it is not an indication that it is supplied sufficiently because soluble silicon in the form of monosilicic acid ( $H_4SiO_4$ ) in soil system is regulated by soil pH, and amount of clay, organic matter, minerals and Fe / Al oxides/hydroxides which are associated with the geologic age of the soil (Tubana & Heckman, 2015). In other words, the degree of soil weathering determined the

amount of silica in the soil. In Malaysia, most of the soil used for agricultural activity is highly weathered soil which are from the order Oxisol and Ultisol (Jusop & Ishak, Weathered tropical soils: the ultisols & oxisols, 2010). About 70% of Malaysian soil are from these two types of soil order which are mainly used for rubber and oil palm cultivation. Oxisol and Ultisol contained high amount of complex Al/Fe hydroxide due to the disintegration of silicate mineral as a result of intense weathering. Quartz is the most common mineral found in Malaysian weathered soil and the most stable mineral in the soil. Because of its physico-chemical properties and stable form, availability of soluble silicon in soil solution is less between 1.6 – 1.9 mg L<sup>-1</sup> of dissolved silicon (Dress et al., 1985). With respect to low solubility, dissolved silicon are easily removed from soil solution due to the adsorption by Fe and Al hydroxide (Bruun et al., 1994). Previous studies reported that Fe and Al hydroxide can remove large amount of dissolved silicon because they have strong adsorption capacity. Therefore, introduction of microorganism like bacteria and fungi are essentially important in the dissolution process of insoluble silica in soil. Studies showed that some bacteria and fungi had the ability to mineralize silica and silicates in nature by several modes of action such as ligands of cation, secretion of organic and inorganic acids, alkali process by production of ammonia and amine and production of extracellular polysaccharides (EPS) by the bacteria at acidic pH (Konhauser, 2015). Microbes with the ability to mineralize insoluble silica for plant uptake in strengthening plant cell wall from pathogen attack is potentially to have double protection for plant against pathogen. Thus, this study was conducted with the following objectives:

1. To isolate and screen potential microbes for its ability in mineralizing silicon, suppressing fungal growth and secreting plant growth promoting characteristics.
2. To study mechanisms of silicon solubilization by potential isolates.
3. To evaluate the efficacy of silicate solubilizing bacteria applied at different rates on the silicon uptake, inhibition of *R. microporus*, stimulation of induced systemic resistance and enhancement of plant growth under rain shelter house.

## REFERENCES

- Abdel-Aziz, S. M., Moharam, M. E., Hamed, H. A., & Mouafi, F. E. (2012). Extracellular Metabolites Produced by a Novel Strain, *Bacillus alvei* NRC-14: 1. Some Properties of the Chitinolytic System. *New York Science Journal*.
- Adeleke, R., Cloete, T. E., & Khasa, D. P. (2011). Culturable microorganisms associated with Sishen iron ore and their potential roles in biobeneficiation. *World Journal of Microbiology and Biotechnology*, 28(3), 1057–1070. <https://doi.org/10.1007/s11274-011-0904-2>
- Ahemad, M., & Kibret, M. (2014). Mechanisms and applications of plant growth promoting rhizobacteria: Current perspective. *Journal of King Saud University - Science*, 26(1), 1–20. <https://doi.org/10.1016/j.jksus.2013.05.001>
- Ahmed, E., & Holmström, S. J. (2015). Microbe–mineral interactions: The impact of surface attachment on mineral weathering and element selectivity by microorganisms. *Chemical Geology*, 403, 13–23. <https://doi.org/10.1016/j.chemgeo.2015.03.009>
- Ahmed, E., & Holmström, S. J. M. (2014). Siderophores in environmental research: roles and applications. *Microbial Biotechnology*, 7(3), 196–208. <https://doi.org/10.1111/1751-7915.12117>
- Ali, S., Techato, K., Taweenkun, J., & Gyawali, S. (2018). Assessment of land use suitability for natural rubber using GIS in the U-tapao River basin, Thailand. *Kasetsart Journal of Social Sciences*. <https://doi.org/10.1016/j.kjss.2018.07.002>
- Allen, B., & Hajek, B. (2018). Mineral Occurrence in Soil Environments. *SSSA Book Series*, 199–278. <https://doi.org/10.2136/sssabookser1.2ed.c5>
- Amores, D. R., & Warren, L. A. (2007). Identifying when microbes biosilicify: The interconnected requirements of acidic pH, colloidal SiO<sub>2</sub> and exposed microbial surface. *Chemical Geology*, 240(3–4), 298–312. <https://doi.org/10.1016/j.chemgeo.2007.02.016>
- Amruta, N., Prasanna Kumar, M. K., Puneeth, M. E., Sarika, G., Kandikattu, H. K., Vishwanath, K., & Narayanaswamy, S. (2018). Exploring the Potentiality of Novel Rhizospheric Bacterial Strains against the Rice Blast Fungus *Magnaporthe oryzae*. *The Plant Pathology Journal*, 34(2), 126–138. <https://doi.org/10.5423/ppj.oa.11.2017.0242>
- Apine, O., & Jadhav, J. (2011). Optimization of medium for indole-3-acetic acid production using *Pantoea agglomerans* strain PVM. *Journal of Applied*



*Microbiology*, 110(5), 1235–1244. <https://doi.org/10.1111/j.1365-2672.2011.04976.x>

- Araújo, F. D. D. S., Araújo, W. L., & Eberlin, M. N. (2017). Potential of *Burkholderia seminalis* TC3.4.2R3 as Biocontrol Agent Against *Fusarium oxysporum* Evaluated by Mass Spectrometry Imaging. *Journal of the American Society for Mass Spectrometry*, 28(5), 901–907. <https://doi.org/10.1007/s13361-017-1610-6>
- Arora, N. K., & Verma, M. (2017). Modified microplate method for rapid and efficient estimation of siderophore produced by bacteria. *3 Biotech*, 7(6). <https://doi.org/10.1007/s13205-017-1008-y>
- Arora, P. K., Sharma, A., & Bae, H. (2015). Microbial Degradation of Indole and Its Derivatives. *Journal of Chemistry*, 2015, 1–13. <https://doi.org/10.1155/2015/129159>
- Artyszak, A. (2018). Effect of Silicon Fertilization on Crop Yield Quantity and Quality—A Literature Review in Europe. *Plants*, 7(3), 54. <https://doi.org/10.3390/plants7030054>
- Avakyan, Z. A., Belkanova, N. P., Karavaiko, G. I., & Piskunov, V. P. (1985). Silicon-Compounds In Solution During Bacterial Quartz Degradation. *Microbiology*, 54(2), 250–256.
- Bakri, A., Mokhtar, S. J., & Daud, N. W. (2017). Physiological and morphological responses of rubber (*Hevea brasiliensis*) RRIM 3001 to different rates of basalt application. *Journal of Tropical Plant Physiology*.
- Balogh-Brunstad, Z., Kent Keller, C., Thomas Dickinson, J., Stevens, F., Li, C., & Bormann, B. T. (2008). Biotite weathering and nutrient uptake by ectomycorrhizal fungus, *Suillus tomentosus*, in liquid-culture experiments. *Geochimica et Cosmochimica Acta*, 72(11), 2601–2618. <https://doi.org/10.1016/j.gca.2008.04.003>
- Bazylinski, D. A. (2003). Biologically Controlled Mineralization in Prokaryotes. *Reviews in Mineralogy and Geochemistry*, 54(1), 217–247. <https://doi.org/10.2113/0540217>
- Beckwith, R., & Reeve, R. (1964). Studies on soluble silica in soils. II. The release of monosilicic acid from soils. *Soil Research*, 2(1), 33. <https://doi.org/10.1071/sr9640033>
- Begho, E. R., & Ekpo, E. J. A. (1987). Incidence and pathogenicity of *Fomes lignosus* (Klotzsch) Bres. on *Triplochiton scleroxylon* K. Schum. *Plant and Soil*, 98(3), 439–442. <https://doi.org/10.1007/bf02378366>
- Bekker, T. F., Kaiser, C., & Labuschagne, N. (2006). Efficacy of water soluble silicon against *Phytophthora cinnamomi* root rot of avocado: A progress report. *South African Avocado Growers' Association Yearbook*.



- Belton, D. J., Deschaume, O., & Perry, C. C. (2012). An overview of the fundamentals of the chemistry of silica with relevance to biosilicification and technological advances. *FEBS Journal*, 279(10), 1710–1720. <https://doi.org/10.1111/j.1742-4658.2012.08531.x>
- Bennett, P. C., Rogers, J. R., Choi, W. J., & Hiebert, F. K. (2001). Silicates, silicate weathering, and microbial ecology. 18(1) 3-19. *Geomicrobiology Journal*. doi: 10.1080/01490450151079734
- Bennett, P., Melcer, M., Siegel, D., & Hassett, J. (1988). The dissolution of quartz in dilute aqueous solutions of organic acids at 25°C. *Geochimica et Cosmochimica Acta*, 52(6), 1521–1530. [https://doi.org/10.1016/0016-7037\(88\)90222-0](https://doi.org/10.1016/0016-7037(88)90222-0)
- Bernal, P., Allsopp, L. P., Filloux, A., & Llamas, M. A. (2017). The *Pseudomonas putida* T6SS is a plant warden against phytopathogens. *The ISME Journal*, 11(4), 972–987. <https://doi.org/10.1038/ismej.2016.169>
- Beveridge, T. J., & Murray, R. G. (1980). Sites of metal deposition in the cell wall of *Bacillus subtilis*. *Journal of Bacteriology*, 141(2), 876–887. <https://doi.org/10.1128/jb.141.2.876-887.1980>
- Bist, V., Niranjana, A., Ranjan, M., Lehri, A., Seem, K., & Srivastava, S. (2020). Silicon-Solubilizing Media and Its Implication for Characterization of Bacteria to Mitigate Biotic Stress. *Frontiers in Plant Science*, 11. <https://doi.org/10.3389/fpls.2020.00028>
- Blanchette, R. A. (1991). Delignification by Wood-Decay Fungi. *Annual Review of Phytopathology*, 29(1), 381–403. <https://doi.org/10.1146/annurev.py.29.090191.002121>
- Bonneville, S., Smits, M. M., Brown, A., Harrington, J., Leake, J. R., Brydson, R., & Benning, L. G. (2009). Plant-driven fungal weathering: Early stages of mineral alteration at the nanometer scale. *Geology*, 37(7), 615–618. <https://doi.org/10.1130/g25699a.1>
- Borer, P., & Hug, S. J. (2014). Photo-redox reactions of dicarboxylates and  $\alpha$ -hydroxydicarboxylates at the surface of Fe(III)(hydr)oxides followed with in situ ATR-FTIR spectroscopy. *Journal of Colloid and Interface Science*, 416, 44–53. <https://doi.org/10.1016/j.jcis.2013.10.030>
- Bowen, N. L. (1915). The Later Stages of the Evolution of the Igneous Rocks. *The Journal of Geology*, 23(S8), 1–91. <https://doi.org/10.1086/622298>
- Brady, M. S., & Katz, S. E. (1990). Factors Influencing Optimization of Diffusion Assays for Antibiotics. *Journal of AOAC INTERNATIONAL*, 73(2), 202–205. <https://doi.org/10.1093/jaoac/73.2.202>

- Brady, P. V., & Walther, J. V. (1989). Controls on silicate dissolution rates in neutral and basic pH solutions at 25°C. *Geochimica et Cosmochimica Acta*, 53(11), 2823–2830. [https://doi.org/10.1016/0016-7037\(89\)90160-9](https://doi.org/10.1016/0016-7037(89)90160-9)
- Bray, R. H., & Kurtz, L. T. (1945). Determination Of Total, Organic, And Available Forms Of Phosphorus In Soils. *Soil Science*, 59(1), 39–46. <https://doi.org/10.1097/00010694-194501000-00006>
- Broadley, M., Brown, P., Cakmak, I., Ma, J. F., Rengel, Z., & Zhao, F. (2011). Beneficial Elements. In *Marschner's Mineral Nutrition of Higher Plants: Third Edition*. doi: 10.1016/B978-0-12-384905-2.00008-X
- Brogden, K. A. (2005). Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria? *Nature Reviews Microbiology*, 3(3), 238–250. doi: 10.1038/nrmicro1098
- Bruun Hansen, H. C., Raben-Lange, B., Raulund-Rasmussen, K., & Borggaard, O. K. (1994). Monosilicate Adsorption by Ferrihydrite and Goethite at pH 3–6. *Soil Science*, 158(1), 40–46. <https://doi.org/10.1097/00010694-199407000-00005>
- Cai, K., Gao, D., Chen, J., & Luo, S. (2009). Probing the mechanisms of silicon-mediated pathogen resistance. *Plant Signaling & Behavior*, 4(1), 1–3. <https://doi.org/10.4161/psb.4.1.7280>
- Camiletti, B. X., Moral, J., Asensio, C. M., Torrico, A. K., Lucini, E. I., Giménez-Pecci, M. D. L. P., & Michailides, T. J. (2018). Characterization of Argentinian Endemic *Aspergillus flavus* Isolates and Their Potential Use as Biocontrol Agents for Mycotoxins in Maize. *Phytopathology*®, 108(7), 818–828. <https://doi.org/10.1094/phyto-07-17-0255-r>
- Campbell, C. L., & Madden, L. V. (1990). Introduction to plant disease epidemiology. In *New York: Wiley-Interscience*.
- Cao, J., Cheng, C., Yang, J., & Wang, Q. (2015). Pathogen infection drives patterns of nutrient resorption in citrus plants. *Scientific Reports*, 5(1). <https://doi.org/10.1038/srep14675>
- Carneiro, R. G., Mazzafera, P., Ferraz, L. C. C., Muraoka, T., & Trivelin, P. C. O. (2002). Uptake and translocation of nitrogen, phosphorus and calcium in soybean infected with *Meloidogyne incognita* and *M. javanica*. *Fitopatologia Brasileira*, 27(2), 141–150. <https://doi.org/10.1590/s0100-41582002000200004>
- Casey, W. H., & Swaddle, T. W. (2003). Why small? The use of small inorganic clusters to understand mineral surface and dissolution reactions in geochemistry. *Reviews of Geophysics*, 41(2). <https://doi.org/10.1029/2002rg000118>

- Chaiharn, M., Sujada, N., Pathom-aree, W., & Lumyong, S. (2019). Biological Control of *Rigidoporus microporus* the Cause of White Root Disease in Rubber Using PGPRs In vivo. *Chiang Mai Journal Science*, *46*(5), 850–866. [https://www.researchgate.net/publication/337243735\\_Biological\\_Control\\_of\\_Rigidoporus\\_microporus\\_the\\_Cause\\_of\\_White\\_Root\\_Disease\\_in\\_Rubber\\_Using\\_PGPRs\\_In\\_vivo](https://www.researchgate.net/publication/337243735_Biological_Control_of_Rigidoporus_microporus_the_Cause_of_White_Root_Disease_in_Rubber_Using_PGPRs_In_vivo)
- Chandrakala, C., Voleti, S. R., Bandeppa, S., Sunil Kumar, N., & Latha, P. C. (2019). Silicate Solubilization and Plant Growth Promoting Potential of *Rhizobium* Sp. Isolated from Rice Rhizosphere. *Silicon*, *11*(6), 2895–2906. <https://doi.org/10.1007/s12633-019-0079-2>
- Chandrasekhar, T. R. (2012). Evaluation of unconstrained and constrained mathematical functions to model girth growth of rubber trees (*Hevea brasiliensis*) using young age measurements. *Journal of Forestry Research*, *23*(3), 365–375. <https://doi.org/10.1007/s11676-012-0272-2>
- Chapman, H. . (1965). Cation-exchange capacity. In: C. A. Black (ed.) Methods of soil analysis Chemical and microbiological properties. In *Agronomy Journal*.
- Cherif, M. (1994). Defense Responses Induced by Soluble Silicon in Cucumber Roots Infected by *Pythium* spp. *Phytopathology*, *84*(3), 236. <https://doi.org/10.1094/phyto-84-236>
- Chérif, M., Benhamou, N., Menzies, J., & Bélanger, R. (1992). Silicon induced resistance in cucumber plants against *Pythium ultimum*. *Physiological and Molecular Plant Pathology*, *41*(6), 411–425. [https://doi.org/10.1016/0885-5765\(92\)90053-x](https://doi.org/10.1016/0885-5765(92)90053-x)
- Chiang, K., Liu, H., & Bock, C. (2017). A discussion on disease severity index values. Part I: warning on inherent errors and suggestions to maximise accuracy. *Annals of Applied Biology*, *171*(2), 139–154. <https://doi.org/10.1111/aab.12362>
- Contreras-Pérez, M., Hernández-Salmerón, J., Rojas-Solís, D., Rocha-Granados, C., Orozco-Mosqueda, M. D. C., Parra-Cota, F. I., de Los Santos-Villalobos, S., & Santoyo, G. (2019). Draft genome analysis of the endophyte, *Bacillus toyonensis* COPE52, a blueberry (*Vaccinium* spp. var. Biloxi) growth-promoting bacterium. *3 Biotech*, *9*(10). <https://doi.org/10.1007/s13205-019-1911-5>
- Cunha, A. C. M. C. M. D., Oliveira, M. L. D., Caballero, E. C., Martinez, H. E. P., Fontes, P. C. R., & Pereira, P. R. G. (2012b). Growth and nutrient uptake of coffee seedlings cultivated in nutrient solution with and without silicon addition. *Revista Ceres*, *59*(3), 392–398. <https://doi.org/10.1590/s0034-737x2012000300015>

- Datnoff, L. E. (1992). Influence of Silicon Fertilizer Grades on Blast and Brown Spot Development and on Rice Yields. *Plant Disease*, 76(10), 1011. <https://doi.org/10.1094/pd-76-1011>
- Davy, H. (1819). *The Elements of Agricultural Chemistry* (1st ed.). Hartford: Hudson and Co.
- Deahl, K. L. (1993). First Report of Resistance of *Phytophthora infestans* to Metalaxyl in Eastern Washington and Southwestern British Columbia. *Plant Disease*, 77(4), 429C. <https://doi.org/10.1094/pd-77-0429c>
- Debona, D., Rodrigues, F. A., & Datnoff, L. E. (2017). Silicon's Role in Abiotic and Biotic Plant Stresses. *Annual Review of Phytopathology*, 55(1), 85–107. <https://doi.org/10.1146/annurev-phyto-080516-035312>
- di Benedetto, N. A., Campaniello, D., Bevilacqua, A., Cataldi, M. P., Sinigaglia, M., Flagella, Z., & Corbo, M. R. (2019). Isolation, Screening, and Characterization of Plant-Growth-Promoting Bacteria from Durum Wheat Rhizosphere to Improve N and P Nutrient Use Efficiency. *Microorganisms*, 7(11), 541. <https://doi.org/10.3390/microorganisms7110541>
- Dixon, J. B., Weed, S. B., Allen, B. L., & Hajek, B. F. (2002). *Mineral Occurrence in Soil Environments*. doi: 10.2136/sssabookser1.2ed.c5
- Dordas, C. (2008). Role of nutrients in controlling plant diseases in sustainable agriculture. A review. *Agronomy for Sustainable Development*, 28(1), 33–46. <https://doi.org/10.1051/agro:2007051>
- Douglas, L. A. (1971). Chemical Weathering of the Silicate Minerals. *Soil Science*, 111(4), 270. <https://doi.org/10.1097/00010694-197104000-00019>
- Drever, J., & Stillings, L. (1997). The role of organic acids in mineral weathering. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 120(1–3), 167–181. [https://doi.org/10.1016/s0927-7757\(96\)03720-x](https://doi.org/10.1016/s0927-7757(96)03720-x)
- Drzewiecka, D. (2016). Significance and Roles of *Proteus* spp. Bacteria in Natural Environments. *Microbial Ecology*, 72(4), 741–758. <https://doi.org/10.1007/s00248-015-0720-6>
- Duca, D., Lorv, J., Patten, C. L., Rose, D., & Glick, B. R. (2014). Indole-3-acetic acid in plant–microbe interactions. *Antonie van Leeuwenhoek*, 106(1), 85–125. <https://doi.org/10.1007/s10482-013-0095-y>
- Ehrlich, H. L., Newman, D. K., & Kappler, A. (2015). *Ehrlich's Geomicrobiology* (6th ed.). CRC Press. <https://doi.org/10.1201/b19121>
- Eikenberg, J. (1990). On the problem of silica solubility at high pH. *Tech. Ber. Nagra*, (July 1990), 1015–2636.

- Elliott, C. L., & Snyder, G. H. (1991). Autoclave-induced digestion for the colorimetric determination of silicon in rice straw. *Journal of Agricultural and Food Chemistry*, 39(6), 1118–1119. <https://doi.org/10.1021/jf00006a024>
- Epstein, E. (1994). The anomaly of silicon in plant biology. *Proceedings of the National Academy of Sciences*, 91(1), 11–17. <https://doi.org/10.1073/pnas.91.1.11>
- Epstein, E. (2001). Chapter 1 Silicon in plants: Facts vs. concepts. *Silicon in Agriculture*, 1–15. [https://doi.org/10.1016/s0928-3420\(01\)80005-7](https://doi.org/10.1016/s0928-3420(01)80005-7)
- Epstein, E. (2009). Silicon: its manifold roles in plants. *Annals of Applied Biology*, 155(2), 155–160. <https://doi.org/10.1111/j.1744-7348.2009.00343.x>
- Fan, M., Liu, Z., Nan, L., Wang, E., Chen, W., Lin, Y., & Wei, G. (2018). Isolation, characterization, and selection of heavy metal-resistant and plant growth-promoting endophytic bacteria from root nodules of Robinia pseudoacacia in a Pb/Zn mining area. *Microbiological Research*, 217, 51–59. <https://doi.org/10.1016/j.micres.2018.09.002>
- Fan, Q. (2002). Production of 1,3-glucanase and chitinase of two biocontrol agents and their possible modes of action. *Chinese Science Bulletin*, 47(4), 292. <https://doi.org/10.1360/02tb9070>
- Farid, A. M., Lee, S. S., Maziah, Z., & Patahayah, M. (2009). Pathogenicity of rigidoporus microporus and phellinus noxius against four major plantation tree species in Peninsular Malaysia. *Journal of Tropical Forest Science*, 21(4), 289–298. doi: 10.2307/23616753
- Fauteux, F., Rémus-Borel, W., Menzies, J. G., & Bélanger, R. R. (2005). Silicon and plant disease resistance against pathogenic fungi. *FEMS Microbiology Letters*, 249(1), 1–6. <https://doi.org/10.1016/j.femsle.2005.06.034>
- Fawe, A., Abou-Zaid, M., Menzies, J. G., & Bélanger, R. R. (1998). Silicon-Mediated Accumulation of Flavonoid Phytoalexins in Cucumber. *Phytopathology*, 88(5), 396–401. <https://doi.org/10.1094/phyto.1998.88.5.396>
- Ferguson, A. D., & Deisenhofer, J. (2002). TonB-dependent receptors—structural perspectives. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, 1565(2), 318–332. [https://doi.org/10.1016/s0005-2736\(02\)00578-3](https://doi.org/10.1016/s0005-2736(02)00578-3)
- Ferreira, C. M. H., Vilas-Boas, N., Sousa, C. A., Soares, H. M. V. M., & Soares, E. V. (2019). Comparison of five bacterial strains producing siderophores with ability to chelate iron under alkaline conditions. *AMB Express*, 9(1). <https://doi.org/10.1186/s13568-019-0796-3>



- Fox, J., & Castella, J. C. (2013). Expansion of rubber (*Hevea brasiliensis*) in Mainland Southeast Asia: what are the prospects for smallholders? *Journal of Peasant Studies*, 40(1), 155–170. <https://doi.org/10.1080/03066150.2012.750605>
- Fox, R. A. (1965). The role of biological eradication in root-disease control in replantings of *Hevea brasiliensis*. *Ecology of Soil-Borne Plant Pathogens: Prelude to Biological Control*.
- Fox, R. A. (1970). A comparison of methods of dispersal, survival, and parasitism in some fungi causing root diseases of tropical plantation crops. *Root Diseases and Soil-borne Pathogens*, 2, 179.
- Fox, R. A. (1970). A comparison of methods of dispersal, survival, and parasitism in some fungi causing root diseases of tropical plantation crops. *Root Diseases and Soil-borne Pathogens*, 2, 179.
- Fox, R. L., Silva, J. A., Plucknett, D. L., & Teranishi, D. Y. (1969). Soluble and total silicon in sugar cane. *Plant and Soil*, 30(1), 81–92. <https://doi.org/10.1007/bf01885263>
- Frew, A., Weston, L. A., Reynolds, O. L., & Gurr, G. M. (2018). The role of silicon in plant biology: a paradigm shift in research approach. *Annals of Botany*, 121(7), 1265–1273. <https://doi.org/10.1093/aob/mcy009>
- Frey, B., Rieder, S. R., Brunner, I., Plötze, M., Koetzsch, S., Lapanje, A., Brandl, H., & Furrer, G. (2010). Weathering-Associated Bacteria from the Damma Glacier Forefield: Physiological Capabilities and Impact on Granite Dissolution. *Applied and Environmental Microbiology*, 76(14), 4788–4796. <https://doi.org/10.1128/aem.00657-10>
- Gascho, G. J. (2001). Chapter 12 Silicon sources for agriculture. *Silicon in Agriculture*, 197–207. [https://doi.org/10.1016/s0928-3420\(01\)80016-1](https://doi.org/10.1016/s0928-3420(01)80016-1)
- Gaur, S., Kumar, J., Kumar, D., Chauhan, D. K., Prasad, S. M., & Srivastava, P. K. (2020). Fascinating impact of silicon and silicon transporters in plants: A review. *Ecotoxicology and Environmental Safety*, 202, 110885. <https://doi.org/10.1016/j.ecoenv.2020.110885>
- Geiger, J. P., Rio, B., Nicole, M., & Nandris, D. (1986). Biodegradation of *Hevea brasiliensis* wood by *Rigidoporus lignosus* and *Phellinus noxius*. *Forest Pathology*, 16(3), 147–159. <https://doi.org/10.1111/j.1439-0329.1986.tb01055.x>
- Geiger, J., Rio, B., Nandris, D., & Nicole, M. (1989). Peroxidase production in tissues of the rubber tree following infection by root rot fungi. *Physiological and Molecular Plant Pathology*, 34(3), 241–256. [https://doi.org/10.1016/0885-5765\(89\)90047-7](https://doi.org/10.1016/0885-5765(89)90047-7)

- Gerami, M., & Rameeh, V. (2012). Study of silicon and nitrogen effects on yield components and shoot ions nutrient composition in rice. *Agriculture (Pol'nohospodárstvo)*, 58(3), 93–98. <https://doi.org/10.2478/v10207-012-0011-x>
- Ghosh, S. K., Pal, S., & Chakraborty, N. (2015). The qualitative and quantitative assay of siderophore production by some microorganisms and effect of different media on its production. *International Journal of Chemical Sciences*.
- Głazowska, S., Baldwin, L., Mravec, J., Bukh, C., Hansen, T. H., Jensen, M. M., Fangel, J. U., Willats, W. G. T., Glasius, M., Felby, C., & Schjoerring, J. K. (2018). The impact of silicon on cell wall composition and enzymatic saccharification of *Brachypodium distachyon*. *Biotechnology for Biofuels*, 11(1). <https://doi.org/10.1186/s13068-018-1166-0>
- Gong, H., Chen, K., Chen, G., Wang, S., & Zhang, C. (2003). Effects of Silicon on Growth of Wheat Under Drought. *Journal of Plant Nutrition*, 26(5), 1055–1063. <https://doi.org/10.1081/pln-120020075>
- Gooch, J. W. (2011). Schaeffer-Fulton Spore Staining. *Encyclopedic Dictionary of Polymers*, 922. [https://doi.org/10.1007/978-1-4419-6247-8\\_14746](https://doi.org/10.1007/978-1-4419-6247-8_14746)
- Gotor-Vila, A., Teixidó, N., di Francesco, A., Usall, J., Ugolini, L., Torres, R., & Mari, M. (2017). Antifungal effect of volatile organic compounds produced by *Bacillus amyloliquefaciens* CPA-8 against fruit pathogen decays of cherry. *Food Microbiology*, 64, 219–225. <https://doi.org/10.1016/j.fm.2017.01.006>
- Greger, M., Landberg, T., & Vaculík, M. (2018). Silicon Influences Soil Availability and Accumulation of Mineral Nutrients in Various Plant Species. *Plants*, 7(2), 41. <https://doi.org/10.3390/plants7020041>
- Grob, A. (1896). *Beiträge zur Anatomie der Epidermis der Gramineenblätter*. Stuttgart: Verlag von Erwin Nägele (Druck von Fiedr. Scheel, Cassel). Retrieved from <https://www.biodiversitylibrary.org/item/145907>
- Guan, N., & Liu, L. (2020). Microbial response to acid stress: mechanisms and applications. *Applied Microbiology and Biotechnology*, 104(1), 51–65. <https://doi.org/10.1007/s00253-019-10226-1>
- Guntzer, F., Keller, C., & Meunier, J. D. (2011). Benefits of Plant Silicon for Crops: A Review. *Agronomy for Sustainable Development*, 32(1), 201–213. <https://doi.org/10.1007/s13593-011-0039-8>
- Guo, H. C., Wang, W. B., Luo, X. H., & Wu, X. P. (2015). Characteristics of rhizosphere and bulk soil microbial communities in rubber plantations in Hainan Island, China. *Journal of Tropical Forest Science*. doi: 10.2307/43582385



- Hai-Chao, G. U. O., Wang, W. Bin, Xue-Hua, L. U. O., & Xiao-Ping, W. U. (2013). Variations in rhizosphere microbial communities of rubber plantations in hainan island, China. *Journal of Rubber Research*.
- Hayashi, Y. (2009). Production of natural rubber from Para rubber tree. *Plant Biotechnology*, 26(1), 67–70. <https://doi.org/10.5511/plantbiotechnology.26.67>
- Hernández-Rivera, M., Ojeda-Morales, M., Martínez-Vázquez, J., Villegas-Cornelio, V., & Córdova-Bautista, Y. (2011). Optimal Parameters For In Vitro Development of the Hydrocarbonoclastic Microorganism *Proteus* sp. *Journal of Soil Science and Plant Nutrition*, 11(1), 29–43. <https://doi.org/10.4067/s0718-95162011000100003>
- Heine, G., Tikum, G., & Horst, W. J. (2006). The effect of silicon on the infection by and spread of *Pythium aphanidermatum* in single roots of tomato and bitter gourd. *Journal of Experimental Botany*, 58(3), 569–577. <https://doi.org/10.1093/jxb/erl232>
- Heinen, W. (1960). *Arch Mikrob*.
- Heydari, A., & Pessarakli, M. (2010). A Review on Biological Control of Fungal Plant Pathogens Using Microbial Antagonists. *Journal of Biological Sciences*, 10(4), 273–290. <https://doi.org/10.3923/jbs.2010.273.290>
- Hiemstra, T., Barnett, M. O., & van Riemsdijk, W. H. (2007). Interaction of silicic acid with goethite. *Journal of Colloid and Interface Science*, 310(1), 8–17. <https://doi.org/10.1016/j.jcis.2007.01.065>
- Hodson, M. J., White, P. J., Mead, A., & Broadley, M. R. (2005). Phylogenetic Variation in the Silicon Composition of Plants. *Annals of Botany*, 96(6), 1027–1046. <https://doi.org/10.1093/aob/mci255>
- Holiday, P. (1980). Fungus diseases of tropical crops. *Australasian Plant Pathology*, 9(4), 120. <https://doi.org/10.1007/bf03213662>
- Horneck, D. A., & Miller, R. O. (1998). Determination of total nitrogen in plant tissue. In *Handbook of reference methods for plant analysis*.
- Hu, Cai, & Jeong. (2019). Silicon Affects Root Development, Tissue Mineral Content, and Expression of Silicon Transporter Genes in Poinsettia (*Euphorbia pulcherrima* Willd.) Cultivars. *Plants*, 8(6), 180. <https://doi.org/10.3390/plants8060180>
- Hu, L. (2019). Application Of Bryophyte Rhizoid-Associated Bacteria Increases Silicon Accumulation And Growth In Maize (*Zea Mays* L.) Seedlings. *Applied Ecology and Environmental Research*, 17(6). [https://doi.org/10.15666/aeer/1706\\_1342313433](https://doi.org/10.15666/aeer/1706_1342313433)

- Huang, F., Wen, X. H., Cai, Y. X., & Cai, K. Z. (2018). Silicon-Mediated Enhancement of Heavy Metal Tolerance in Rice at Different Growth Stages. *International Journal of Environmental Research and Public Health*, *15*(10), 2193. <https://doi.org/10.3390/ijerph15102193>
- Huang, Y. T., Lowe, D. J., Churchman, G. J., Schipper, L. A., Cursons, R., Zhang, H., Chen, T. Y., & Cooper, A. (2016). DNA adsorption by nanocrystalline allophane spherules and nanoaggregates, and implications for carbon sequestration in Andisols. *Applied Clay Science*, *120*, 40–50. <https://doi.org/10.1016/j.clay.2015.11.009>
- Hunt, R. (1990). Absolute growth rates. In *Basic growth analysis* (pp. 17–24). Springer.
- Iler. (1979). the chemistry of silica chapter 6: The Surface Chemistry of Silica. *The Chemistry of Silica*.
- Inanaga, S., Okasaka, A., & Tanaka, S. (1995). Does silicon exist in association with organic compounds in rice plant? *Soil Science and Plant Nutrition*, *41*(1), 111–117. <https://doi.org/10.1080/00380768.1995.10419564>
- Iroque, V. (2012). Effects of White Root Rot Disease on Hevea brasiliensis (Muell. Arg.) – Challenges and Control Approach. *Plant Science*. <https://doi.org/10.5772/54024>
- Janda, J. M., & Abbott, S. L. (2007). 16S rRNA Gene Sequencing for Bacterial Identification in the Diagnostic Laboratory: Pluses, Perils, and Pitfalls. *Journal of Clinical Microbiology*, *45*(9), 2761–2764. <https://doi.org/10.1128/jcm.01228-07>
- Jang, S. W., Kim, Y., Khan, A. L., Na, C. I., & Lee, I. J. (2018). Exogenous short-term silicon application regulates macro-nutrients, endogenous phytohormones, and protein expression in *Oryza sativa* L. *BMC Plant Biology*, *18*(1). <https://doi.org/10.1186/s12870-017-1216-y>
- Jayasuriya, K. E., & Thennakoon, B. I. (2007). Biological control of Rigidoporus microporus, the cause of white root disease in rubber. *Ceylon Journal of Science (Biological Sciences)*.
- Jones, D. L. (1998). Organic acids in the rhizosphere - A critical review. *Plant and Soil*, *205*(1), 25–44. <https://doi.org/10.1023/a:1004356007312>
- Jones, L. H. P., & Handreck, K. A. (1963). Effects of Iron and Aluminium Oxides on Silica in Solution in Soils. *Nature*, *198*(4883), 852–853. <https://doi.org/10.1038/198852a0>
- Kabata-Pendias, A., & Mukherjee, A. B. (2007). *Trace Elements From Soil to Human* (2007th ed.). Springer.

- Monger, H. C., & Kelly, E. F. (2002). Silica Minerals. *Soil Mineralogy with Environmental Applications*, 611–636. <https://doi.org/10.2136/sssabookser7.c20>
- Napper, R. P. N. (1933). Root disease investigations. *Rep. Rubb. Res. Inst. Malaya*, 105.
- Petch, T. (1921). The Diseases and Pests of the Rubber Tree. *The Diseases and Pests of the Rubber Tree*.
- Suryanto, D., Munthe, R. A., Nurwahyuni, I., & Munir, E. (2017). An Assay On Potential of Local Trichoderma Spp. to Control White Root Rot Disease Caused by Rigidoporus Microporus in Rubber Plant Stump. *Journal of Pure and Applied Microbiology*, 11(2), 717–723. <https://doi.org/10.22207/jpam.11.2.10>
- Wu, L., Huang, Z., Li, X., Ma, L., Gu, Q., Wu, H., Liu, J., Borriss, R., Wu, Z., & Gao, X. (2018). Stomatal Closure and SA-, JA/ET-Signaling Pathways Are Essential for *Bacillus amyloliquefaciens* FZB42 to Restrict Leaf Disease Caused by *Phytophthora nicotianae* in *Nicotiana benthamiana*. *Frontiers in Microbiology*, 9. <https://doi.org/10.3389/fmicb.2018.00847>
- Jusop, S., & Ishak, F. (2010). *Weathered Tropical Soils: The Ultisols & Oxisols*. Universiti Putra Malaysia Press. Retrieved from <https://books.google.com.my/books?id=wq2hYgEACAAJ>
- Jusop, S. (2011). *Methods in Soil Mineralogy*. Amsterdam University Press.
- Kaewchai, S., & Soyong, K. (2010). Application of biofungicides against *Rigidoporus microporus* causing white root disease of rubber trees. *International Journal of Agricultural Technology*, 6, 349-363.
- Kang, S. M., Waqas, M., Shahzad, R., You, Y. H., Asaf, S., Khan, M. A., Lee, K. E., Joo, G. J., Kim, S. J., & Lee, I. J. (2017). Isolation and characterization of a novel silicate-solubilizing bacterial strain *Burkholderia eburnea* CS4-2 that promotes growth of japonica rice (*Oryza sativa* L. cv. Dongjin). *Soil Science and Plant Nutrition*, 1–9. <https://doi.org/10.1080/00380768.2017.1314829>
- Karavaiko, G. I., Krutsko, V. S., Melnikova, E. O., Avakyan, Z. A., & Ostroushko, Y. I. (1980). Role of microorganisms in spodumene degradation. *Microbiology*, 49(3), 402–406.
- Kaur, H., & Greger, M. (2019). A Review on Si Uptake and Transport System. *Plants*, 8(4), 81. <https://doi.org/10.3390/plants8040081>
- Kaur, S., Kaur, N., Siddique, K. H. M., & Nayyar, H. (2015). Beneficial elements for agricultural crops and their functional relevance in defence against stresses. *Archives of Agronomy and Soil Science*, 62(7), 905–920. <https://doi.org/10.1080/03650340.2015.1101070>

- Khan, H. I. (2018). Appraisal of Biofertilizers in Rice: To Supplement Inorganic Chemical Fertilizer. *Rice Science*, 25(6), 357–362. <https://doi.org/10.1016/j.rsci.2018.10.006>
- Khosro, M., & Yousef, S. (2012). Bacterial Biofertilizers for Sustainable Crop Production : a Aeviu. *Journal of Agricultural and Biological Science*.
- Kim, Y. H., Kim, C. Y., Song, W. K., Park, D. S., Kwon, S. Y., Lee, H. S., Bang, J. W., & Kwak, S. S. (2007). Overexpression of sweetpotato swpa4 peroxidase results in increased hydrogen peroxide production and enhances stress tolerance in tobacco. *Planta*, 227(4), 867–881. <https://doi.org/10.1007/s00425-007-0663-3>
- Knight, C. T., & Kinrade, S. D. (2001). Chapter 4 A primer on the aqueous chemistry of silicon. *Silicon in Agriculture*, 57–84. [https://doi.org/10.1016/s0928-3420\(01\)80008-2](https://doi.org/10.1016/s0928-3420(01)80008-2)
- Köhl, J., Kolnaar, R., & Ravensberg, W. J. (2019). Mode of Action of Microbial Biological Control Agents Against Plant Diseases: Relevance Beyond Efficacy. *Frontiers in Plant Science*, 10. <https://doi.org/10.3389/fpls.2019.00845>
- Konhauser, K. (2015). Geomicrobial interactions with silicon. In *Ehrlich's Geomicrobiology, Sixth Edition*. doi: 10.1201/b19121
- Kumar, V., & Jha, G. K. (2014). Use of Fly Ash in Agriculture : Indian Scenario. *WACAU-2014, Israel International Workshop on Agricultural Coal Ash Uses*.
- Lam Melaka (Malaysia). ICI Agriculture Research Centre), C. H. (ICI A. (M) S. B., & Chiu, S. Bin. (n.d.). Hexaconazole (Anvil 5SC), a cost-effective fungicide for controlling white root disease in immature rubber [*Hevea brasiliensis*]. *Planter (Malaysia)*.
- Lauwers, A. M., & Heinen, W. (1974). Bio-degradation and utilization of silica and quartz. *Archives of Microbiology*, 95(1), 67–78. <https://doi.org/10.1007/bf02451749>
- Łażniewska, J., Macioszek, V. K., & Kononowicz, A. K. (2012). Plant-fungus interface: The role of surface structures in plant resistance and susceptibility to pathogenic fungi. *Physiological and Molecular Plant Pathology*, 78, 24–30. <https://doi.org/10.1016/j.pmpp.2012.01.004>
- Lee, K. J., Oh, B. T., & Seralathan, K. K. (2012). Advances in Plant Growth Promoting Rhizobacteria for Biological Control of Plant Diseases. *Bacteria in Agrobiology: Disease Management*, 1–13. [https://doi.org/10.1007/978-3-642-33639-3\\_1](https://doi.org/10.1007/978-3-642-33639-3_1)

- Lee, K. E., Adhikari, A., Kang, S. M., You, Y. H., Joo, G. J., Kim, J. H., Kim, S. J., & Lee, I. J. (2019). Isolation and Characterization of the High Silicate and Phosphate Solubilizing Novel Strain Enterobacter ludwigii GAK2 that Promotes Growth in Rice Plants. *Agronomy*, 9(3), 144. <https://doi.org/10.3390/agronomy9030144>
- Lee, K. H., Park, S. J., Choi, S. J., & Park, J. Y. (2017). Proteus vulgaris and Proteus mirabilis Decrease Candida albicans biofilm formation by suppressing morphological transition to its hyphal form. *Yonsei Medical Journal*, 58(6), 1135. <https://doi.org/10.3349/ymj.2017.58.6.1135>
- Liyanage, G. W., Liyanage, A. D. S., Peries, O. S., & Halangoda, L. (1977). Studies on the variability and pathogenicity of Rigidoporus lignosus. *Journal of the Rubber Research Institute of Sri Lanka*, 54(1), 363-372.
- Liang, Y. C., Sun, W. C., Si, J., & Romheld, V. (2005). Effects of foliar- and root-applied silicon on the enhancement of induced resistance to powdery mildew in Cucumis sativus. *Plant Pathology*, 54(5), 678-685. <https://doi.org/10.1111/j.1365-3059.2005.01246.x>
- Liang, Y. L., Zhang, Z., Wu, M., Wu, Y., & Feng, J. X. (2014). Isolation, Screening, and Identification of Cellulolytic Bacteria from Natural Reserves in the Subtropical Region of China and Optimization of Cellulase Production by Paenibacillus terrae ME27-1. *BioMed Research International*, 2014, 1-13. <https://doi.org/10.1155/2014/512497>
- Liang, Y., Si, J., & Römheld, V. (2005). Silicon uptake and transport is an active process in Cucumis sativus. *New Phytologist*, 167(3), 797-804. <https://doi.org/10.1111/j.1469-8137.2005.01463.x>
- Lieberei, R. (2007). South American Leaf Blight of the Rubber Tree (Hevea spp.): New Steps in Plant Domestication using Physiological Features and Molecular Markers. *Annals of Botany*, 100(6), 1125-1142. <https://doi.org/10.1093/aob/mcm133>
- Liu, W., Xu, X., Wu, X., Yang, Q., Luo, Y., & Christie, P. (2006). Decomposition of silicate minerals by Bacillus mucilaginosus in liquid culture. *Environmental Geochemistry and Health*, 28(1-2), 133-140. <https://doi.org/10.1007/s10653-005-9022-0>
- Lu, G., Jian, W., Zhang, J., Zhou, Y., & Cao, J. (2008). Suppressive Effect of Silicon Nutrient on Phomopsis Stem Blight Development in Asparagus. *HortScience*, 43(3), 811-817. <https://doi.org/10.21273/hortsci.43.3.811>
- Lugtenberg, B., & Kamilova, F. (2009). Plant-Growth-Promoting Rhizobacteria. *Annual Review of Microbiology*, 63(1), 541-556. <https://doi.org/10.1146/annurev.micro.62.081307.162918>



- Lux, A., Luxová, M., Abe, J., Tanimoto, E., Hattori, T., & Inanaga, S. (2003). The dynamics of silicon deposition in the sorghum root endodermis. *New Phytologist*, 158(3), 437–441. <https://doi.org/10.1046/j.1469-8137.2003.00764.x>
- Luyckx, M., Hausman, J. F., Lutts, S., & Guerriero, G. (2017). Impact of Silicon in Plant Biomass Production: Focus on Bast Fibres, Hypotheses, and Perspectives. *Plants*, 6(4), 37. <https://doi.org/10.3390/plants6030037>
- Ma, J., Miyake, Y., & Takahashi, E. (2001). Chapter 2 Silicon as a beneficial element for crop plants. *Silicon in Agriculture*, 17–39. [https://doi.org/10.1016/s0928-3420\(01\)80006-9](https://doi.org/10.1016/s0928-3420(01)80006-9)
- Ma, J. F., & Yamaji, N. (2008). Functions and transport of silicon in plants. *Cellular and Molecular Life Sciences*, 65(19), 3049–3057. <https://doi.org/10.1007/s00018-008-7580-x>
- Ma, J. F. (2004). Role of silicon in enhancing the resistance of plants to biotic and abiotic stresses. *Soil Science and Plant Nutrition*, 50(1), 11–18. <https://doi.org/10.1080/00380768.2004.10408447>
- Ma, J. F. (2010). Silicon Transporters in Higher Plants. *MIPs and Their Role in the Exchange of Metalloids*, 99–109. [https://doi.org/10.1007/978-1-4419-6315-4\\_8](https://doi.org/10.1007/978-1-4419-6315-4_8)
- Ma, J. F., & Takahashi, E. (2002). *Soil, Fertilizer, and Plant Silicon Research in Japan* (1st ed.). Elsevier Science.
- Ma, J. F., & Yamaji, N. (2006). Silicon uptake and accumulation in higher plants. *Trends in Plant Science*, 11(8), 392–397. <https://doi.org/10.1016/j.tplants.2006.06.007>
- Madushani, H., Fernando, T., Wijesundara, R., & Siriwardane, D. (2014). First Report of white root disease of *Artocarpus nobilis* in Sri Lanka caused by *Rigidoporus microporus*. *Journal of the National Science Foundation of Sri Lanka*, 42(2), 197. <https://doi.org/10.4038/jnsfsr.v42i2.6998>
- Mahdavi, S., Kafi, M., Fallahi, E., Shokrpour, M., & Tabrizi, L. (2016). Water stress, nano silica, and digoxin effects on minerals, chlorophyll index, and growth in ryegrass. *International Journal of Plant Production*, 10(2), 251–264.
- Maleva, M., Borisova, G., Koshcheeva, O., & Sinenko, O. (2018). Biofertilizer Based On Silicate Solubilizing Bacteria Improves Photosynthetic Function Of Brassica Juncea. *Agrofor*, 2(3). <https://doi.org/10.7251/agreng1703013m>
- Malinovskaya, I. M., Kosenko, L. V., Votselko, S. K., & Podgorsky, V. S. (1990). The role of *Bacillus mucilaginosus* polysaccharide in the destruction of silicate minerals. *Mikrobiologiya*.

- Mandlik, R., Thakral, V., Raturi, G., Shinde, S., Nikolić, M., Tripathi, D. K., Sonah, H., & Deshmukh, R. (2020). Significance of silicon uptake, transport, and deposition in plants. *Journal of Experimental Botany*, 71(21), 6703–6718. <https://doi.org/10.1093/jxb/eraa301>
- Martínez-Viveros, O., Jorquera, M., Crowley, D., Gajardo, G., & Mora, M. (2010). Mechanisms And Practical Considerations Involved In Plant Growth Promotion By Rhizobacteria. *Journal of Soil Science and Plant Nutrition*, 10(3). <https://doi.org/10.4067/s0718-95162010000100006>
- Matejovic, I. (1997). Determination of carbon and nitrogen in samples of various soils by the dry combustion. *Communications in Soil Science and Plant Analysis*, 28(17–18), 1499–1511. <https://doi.org/10.1080/00103629709369892>
- Maurice, P. A., Vierkorn, M. A., Hersman, L. E., Fulghum, J. E., & Ferryman, A. (2001). Enhancement Of Kaolinite Dissolution by An Aerobic Pseudomonas mendocina Bacterium. *Geomicrobiology Journal*. 18(1), 21–35. <https://doi.org/10.1080/01490450151079752>
- McKeague, J. A., & Cline, M. G. (1963). Silica In Soil Solutions: I. The Form and Concentration of Dissolved Silica In Aqueous Extracts of Some Soils. *Canadian Journal of Soil Science*, 43(1), 70–82. <https://doi.org/10.4141/cjss63-010>
- McKeague, J. A., & Cline, M. G. (1963). Silica In Soil Solutions: Ii. The Adsorption Of Monosilicic Acid by Soil and by Other Substances. *Canadian Journal of Soil Science*, 43(1), 83–96. <https://doi.org/10.4141/cjss63-011>
- Mclean, E. O. (2015). *Soil pH and Lime Requirement*. doi: 10.2134/agronmonogr9.2.2ed.c12
- Meena, V. D., Dotaniya, M. L., Coumar, V., Rajendiran, S., Ajay, Kundu, S., & Subba Rao, A. (2014). A Case for Silicon Fertilization to Improve Crop Yields in Tropical Soils. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 84(3), 505–518. <https://doi.org/10.1007/s40011-013-0270-y>
- Mengel, K., E. A. Kirkby, H. K. and T. A. (2001). Principles of Plant Nutrition 5th Edition. In *Journal of experimental biology*. doi: 10.1038/278101a0
- Menzies, J. G. (1991). Effects of Soluble Silicon on the Parasitic Fitness of *Sphaerotheca fuliginea* on *Cucumis sativus*. *Phytopathology*, 81(1), 84. <https://doi.org/10.1094/phyto-81-84>
- Metcalf, C. R. (1967). Distribution of latex in the plant kingdom. *Economic Botany*, 21(2), 115–127. <https://doi.org/10.1007/bf02897859>



- Miranda, J. R. P., Carvalho, J. G., Santos, D. R., Freire, A. L. O., Bertoni, J. C., Melo, J. R. M., & Caldas, A. L. (2002). Silício e cloreto de sódio na nutrição mineral e produção de matéria seca de plantas de moringa (*Moringa oleifera* Lam.). *Revista Brasileira de Ciência Do Solo*, 26(4), 957–965. <https://doi.org/10.1590/s0100-06832002000400013>
- Mitani, N. (2005). Uptake system of silicon in different plant species. *Journal of Experimental Botany*, 56(414), 1255–1261. <https://doi.org/10.1093/jxb/eri121>
- Mitchell, M. R., Link, R. E., Zhang, L., Zhao, Z. M., Jin, X. D., Liu, L., & Chen, G. (2010). A Novel Measurement System for Dry Rubber Content in Concentrated Latex Based on Y-Type Optical Fiber. *Journal of Testing and Evaluation*, 38(5), 102778. <https://doi.org/10.1520/jte102778>
- Mohaghegh, P., Khoshgoftarmanesh, A. H., Shirvani, M., Sharifnabi, B., & Nili, N. (2011). Effect of Silicon Nutrition on Oxidative Stress Induced by *Phytophthora melonis* Infection in Cucumber. *Plant Disease*, 95(4), 455–460. <https://doi.org/10.1094/pdis-05-10-0379>
- Monreal, J., & Reese, E. T. (1969). The chitinase of *Serratia marcescens*. *Canadian Journal of Microbiology*, 15(7), 689–696. <https://doi.org/10.1139/m69-122>
- Montesinos, E. (2007). Antimicrobial peptides and plant disease control. *FEMS Microbiology Letters*, 270(1), 1–11. <https://doi.org/10.1111/j.1574-6968.2007.00683.x>
- Mooibroek, H., & Cornish, K. (2000b). Alternative sources of natural rubber. *Applied Microbiology and Biotechnology*, 53(4), 355–365. <https://doi.org/10.1007/s002530051627>
- Morikawa, C., & Saigusa, M. (2002). Si amelioration of Al toxicity in barley (*Hordeum vulgare* L.) growing in two Andosols. *Plant and Soil*, 240(1), 161–168. <https://doi.org/10.1023/a:1015804401190>
- Muir, J. W. (1951). The Determination of Total Phosphorus in Soil. *The Macaulay Institute for Soil Research*, 313–317.
- Naher, U. (2018). Biofertilizer as a Supplement of Chemical Fertilizer for Yield Maximization of Rice. *Journal of Agriculture Food and Development*, 2(1), 16–22. <https://doi.org/10.30635/2415-0142.2016.02.3>
- Najihah, N. I., Hanafi, M. M., Idris, A. S., & Hakim, M. A. (2015). Silicon treatment in oil palms confers resistance to basal stem rot disease caused by *Ganoderma boninense*. *Crop Protection*, 67, 151–159. <https://doi.org/10.1016/j.cropro.2014.10.004>

- Nakaew, N., Rangjaroen, C., & Sungthong, R. (2015). Utilization of rhizospheric *Streptomyces* for biological control of *Rigidoporus* sp. causing white root disease in rubber tree. *European Journal of Plant Pathology*, 142(1), 93–105. <https://doi.org/10.1007/s10658-015-0592-0>
- Nandris, D. (1987). Root Rot Diseases of Rubber Trees. *Plant Disease*, 71(4), 298. <https://doi.org/10.1094/pd-71-0298>
- Nandris, D., Chadoeuf, J., Pierrat, J. C., Joannes, H., Geiger, J. P., & Nicole, M. (1996). Modelling rubber-tree root diseases, simulations of various inoculum rates and methods of control. *Forest Pathology*, 26(1), 25–44. <https://doi.org/10.1111/j.1439-0329.1996.tb00707.x>
- Narayanan, C., & Mydin K., K. (Eds.). (2012). *Breeding for disease resistance in Hevea spp. - Status, potential threats, and possible strategies*. Pacific Southwest Research Station.
- Naureen, Z., Aqeel, M., Hassan, M. N., Gilani, S. A., Bouqellah, N., Mabood, F., Hussain, J., & Y. Hafeez, F. (2015). Isolation and Screening of Silicate Bacteria from Various Habitats for Biological Control of Phytopathogenic Fungi. *American Journal of Plant Sciences*, 06(18), 2850–2859. <https://doi.org/10.4236/ajps.2015.618282>
- Neumann, D., & zur Nieden, U. (2001). Silicon and heavy metal tolerance of higher plants. *Phytochemistry*, 56(7), 685–692. [https://doi.org/10.1016/s0031-9422\(00\)00472-6](https://doi.org/10.1016/s0031-9422(00)00472-6)
- Neumann, D. (2003). Silicon in Plants. *Silicon Biomineralization*, 149–160. [https://doi.org/10.1007/978-3-642-55486-5\\_6](https://doi.org/10.1007/978-3-642-55486-5_6)
- Ng, L., Anuar, S., Jong, J., & Elham, M. (2016). Phytobeneficial and Plant Growth-promotion Properties of Silicon-solubilising Rhizobacteria on the Growth and Control of Rice Sheath Blight Disease. *Asian Journal of Plant Sciences*, 15(3–4), 92–100. <https://doi.org/10.3923/ajps.2016.92.100>
- Nicole, M., Chamberland, H., Rioux, D., Lecours, N., Rio, B., Geiger, J. P., & Ouellette, G. B. (1993). A Cytochemical Study of Extracellular Sheaths Associated with *Rigidoporus lignosus* during Wood Decay. *Applied and Environmental Microbiology*, 59(8), 2578–2588. <https://doi.org/10.1128/aem.59.8.2578-2588.1993>
- Nicole, M., Geiger, J. P., & Nandris, D. (1986). Root rot diseases of *Hevea brasiliensis*. *Forest Pathology*, 16(1), 37–55. <https://doi.org/10.1111/j.1439-0329.1986.tb01050.x>
- Noran, A. S., Maiden, N. A., & Ahmad Fauzi, M. A. F. (2015). Rhizospheric Microorganisms From *Hevea Brasiliensis* And Their Antagonism On *Rigidoporus Microporus*. *International Journal Of Agriculture, Forestry and Plantation*, 1, 59–65.

- Norton, L. (1993). Micromorphology of silica cementation in soils. *Soil Micromorphology: Studies in Management and Genesis*, 811–824. [https://doi.org/10.1016/s0166-2481\(08\)70465-3](https://doi.org/10.1016/s0166-2481(08)70465-3)
- Nusaibah, S., Saad, G., & Hun, T. G. (2017). Antagonistic Efficacy of *Trichoderma harzianum* and *Bacillus cereus* against *Ganoderma* Disease of Oil Palm via Dip, Place and Drench (DPD) Artificial Inoculation Technique. *International Journal of Agriculture and Biology*, 19(02), 299–306. <https://doi.org/10.17957/ijab/15.0280>
- Ogbebor, N. O., Adekunle, A. T., Eghafona, O. N., & Ogboghodo, A. I. (2015). Biological control of *Rigidoporus lignosus* in *Hevea brasiliensis* in Nigeria. *Fungal Biology*, 119(1), 1–6. <https://doi.org/10.1016/j.funbio.2014.10.002>
- Oghenekaro, A. O., Kovalchuk, A., Raffaello, T., Camarero, S., Gressler, M., Henrissat, B., Lee, J., Liu, M., Martínez, A. T., Miettinen, O., Mihaltcheva, S., Pangilinan, J., Ren, F., Riley, R., Ruiz-Dueñas, F. J., Serrano, A., Thon, M. R., Wen, Z., Zeng, Z., . . . Asiegbu, F. O. (2020). Genome sequencing of *Rigidoporus microporus* provides insights on genes important for wood decay, latex tolerance and interspecific fungal interactions. *Scientific Reports*, 10(1). <https://doi.org/10.1038/s41598-020-62150-4>
- Omorusi, V.I, Eguavo, O.I, Ogbebor, N.O, Bosah, B.O., Orumwense, K, Ijie, K. (2014). Control of White Root Rot Disease in Rubber Plantations in Nigeria. *International Journal of Microbiology and Immunology Research*.
- Irogué, V. (2012). Effects of White Root Rot Disease on *Hevea brasiliensis* (Muell. Arg.) – Challenges and Control Approach. *Plant Science*. <https://doi.org/10.5772/54024>
- Opfergelt, S., de Bournonville, G., Cardinal, D., André, L., Delstanche, S., & Delvaux, B. (2009). Impact of soil weathering degree on silicon isotopic fractionation during adsorption onto iron oxides in basaltic ash soils, Cameroon. *Geochimica et Cosmochimica Acta*, 73(24), 7226–7240. <https://doi.org/10.1016/j.gca.2009.09.003>
- Ouda, S. M. (2014). Biological control by microorganisms and ionizing radiation. *International Journal of Advanced Research*, 2, 314–356. Retrieved from <http://www.journalijar.com>
- Padan, E., Bibi, E., Ito, M., & Krulwich, T. A. (2005). Alkaline pH homeostasis in bacteria: New insights. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, 1717(2), 67–88. <https://doi.org/10.1016/j.bbamem.2005.09.010>
- Palma, S. I. C. J., Traguedo, A. P., Porteira, A. R., Frias, M. J., Gamboa, H., & Roque, A. C. A. (2018). Machine learning for the meta-analyses of microbial pathogens' volatile signatures. *Scientific Reports*, 8(1). <https://doi.org/10.1038/s41598-018-21544-1>

- Pandey, A., Sharma, E., & Palni, L. M. S. (1998). Influence of bacterial inoculation on maize in upland farming systems of the Sikkim Himalaya. *Soil Biology and Biochemistry*, 30(3), 379–384. [https://doi.org/10.1016/s0038-0717\(97\)00121-1](https://doi.org/10.1016/s0038-0717(97)00121-1)
- Pasternak, T., Groot, E. P., Kazantsev, F. V., Teale, W., Omelyanchuk, N., Kovrizhnykh, V., Palme, K., & Mironova, V. V. (2019). Salicylic Acid Affects Root Meristem Patterning via Auxin Distribution in a Concentration-Dependent Manner. *Plant Physiology*, 180(3), 1725–1739. <https://doi.org/10.1104/pp.19.00130>
- Patil, N. B., Gajbhiye, M., Ahiwale, S. S., Gunjal, A. B., & Kapadnis, B. P. (2011). Optimization of Indole 3 - acetic acid ( IAA ) production by *Acetobacter diazotrophicus* L1 isolated from Sugarcane. *International Journal of Environmental Sciences*. doi: 10.6088/ijes.00202010031
- Petch, T. (2018). *The Diseases and Pests of the Rubber Tree*. Van Duuren Media.
- Pieterse, C. M., Zamioudis, C., Berendsen, R. L., Weller, D. M., van Wees, S. C., & Bakker, P. A. (2014). Induced Systemic Resistance by Beneficial Microbes. *Annual Review of Phytopathology*, 52(1), 347–375. <https://doi.org/10.1146/annurev-phyto-082712-102340>
- Piperno, D. R. (1988). *Phytolith Analysis: An Archaeological and Geological Perspective*. Academic Press.
- Pozza, E. A., Pozza, A. A. A., & Dos Santos Botelho, D. M. (2011). Silicon and plant diseases. A review. *Agronomía Colombiana*.
- Qin, G. Z., & Tian, S. P. (2005). Enhancement of Biocontrol Activity of *Cryptococcus laurentii* by Silicon and the Possible Mechanisms Involved. *Phytopathology*, 95(1), 69–75. <https://doi.org/10.1094/phyto-95-0069>
- Raaijmakers, J. M., & Weller, D. M. (1998). Natural Plant Protection by 2,4-Diacetylphloroglucinol-Producing *Pseudomonas* spp. in Take-All Decline Soils. *Molecular Plant-Microbe Interactions*, 11(2), 144–152. <https://doi.org/10.1094/mpmi.1998.11.2.144>
- Ranganathan, S., Suvarchala, V., Rajesh, Y. B. R. D., Prasad, M., Padmakumari, A. P., & Voleti, S. R. (2006). Effects of silicon sources on its deposition, chlorophyll content, and disease and pest resistance in rice. *Biologia Plantarum*, 50(4), 713–716. <https://doi.org/10.1007/s10535-006-0113-2>
- Ratnasingam, J., Ioras, F., & Wenming, L. (2011). Sustainability of the Rubberwood Sector in Malaysia. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 39(2), 305. <https://doi.org/10.15835/nbha3927195>

- Reuss, J. O., & Johnson, D. W. (1986). Soil Sensitivity. *Acid Deposition and the Acidification of Soils and Waters*, 73–80. [https://doi.org/10.1007/978-1-4419-8536-1\\_8](https://doi.org/10.1007/978-1-4419-8536-1_8)
- Reynolds, O. L., Padula, M. P., Zeng, R., & Gurr, G. M. (2016). Silicon: Potential to Promote Direct and Indirect Effects on Plant Defense Against Arthropod Pests in Agriculture. *Frontiers in Plant Science*, 7. <https://doi.org/10.3389/fpls.2016.00744>
- Ribeiro, I. D. A., Volpiano, C. G., Vargas, L. K., Granada, C. E., Lisboa, B. B., & Passaglia, L. M. P. (2020). Use of Mineral Weathering Bacteria to Enhance Nutrient Availability in Crops: A Review. *Frontiers in Plant Science*, 11. <https://doi.org/10.3389/fpls.2020.590774>
- Ribeiro, R. V., Silva, L. D., Ramos, R. A., Andrade, C. A. D., Zambrosi, F. C. B., & Pereira, S. P. (2011). O alto teor de silício no solo inibe o crescimento radicular de cafeeiros sem afetar as trocas gasosas foliares. *Revista Brasileira de Ciência Do Solo*, 35(3), 939–948. <https://doi.org/10.1590/s0100-06832011000300028>
- Rice, E. W., Baird, R. B., & Eaton, A. D. (2018). 4500-SiO<sub>2</sub> SILICA. In E. W. Rice, R. B. Baird, & A. D. Eaton (Eds.), *Standard Methods For the Examination of Water and Wastewater, 23rs Edition* (23rd ed., pp. 174–179). American Public Health Association, American Water Works Association, Water Environment Federation. doi:10.2105/SMWW.2882.095
- Robinson, S. W. O. (1945). The Fusion Analysis Of Soils: Determination Of Si, Ti, Al, Fe, Mn, Ca, Mg, K, Na. *Soil Science*, 59(1). Retrieved From [https://journals.lww.com/soilsci/Fulltext/1945/01000/The\\_Fusion\\_Analysis\\_of\\_Soils\\_\\_Determination\\_of\\_Si.3.aspx](https://journals.lww.com/soilsci/Fulltext/1945/01000/The_Fusion_Analysis_of_Soils__Determination_of_Si.3.aspx)
- Rodrigues, F. A., & Datnoff, L. E. (2015). Silicon and plant diseases. In *Silicon and Plant Diseases*. doi: 10.1007/978-3-319-22930-0
- Rogers, J. R., & Bennett, P. C. (2004). Mineral stimulation of subsurface microorganisms: release of limiting nutrients from silicates. *Chemical Geology*, 203(1–2), 91–108. <https://doi.org/10.1016/j.chemgeo.2003.09.001>
- Sachs, J. (1862). Ergebnisse einiger neuerer untersuchungen uber die in pflanzen enthaltene Kieselsaure. *Flora*, 33, 65-71.
- Saihua, L., Yunhe, X., Ji, X., Juan, H., Bocharnikova, E. A., & Matichenkov, V. V. (2018). Microwave Digestion for Colorimetric Determination of Total Si in Plant and Mineral Samples. *Communications in Soil Science and Plant Analysis*, 49(7), 840–847. <https://doi.org/10.1080/00103624.2018.1435685>



- Saima, Kuddus, M., Roohi, & Ahmad, I. (2013). Isolation of novel chitinolytic bacteria and production optimization of extracellular chitinase. *Journal of Genetic Engineering and Biotechnology*, 11(1), 39–46. <https://doi.org/10.1016/j.jgeb.2013.03.001>
- The neighbor-joining method: a new method for reconstructing phylogenetic trees. (1987). *Molecular Biology and Evolution*. <https://doi.org/10.1093/oxfordjournals.molbev.a040454>
- Santi, L. P., & Goenadi, D. H. (2017). Solubilization of silicate from quartz mineral by potential silicate solubilizing bacteria (Pelarutan silika asal mineral kuarsa oleh bakteri pelarut silika potensial ). *E-Journal Menara Perkebunan*, 85(2). <https://doi.org/10.22302/iribb.jur.mp.v85i2.247>
- Santi, L. P., Nurhaimi-Haris, & Mulyanto, D. (2018). Effect of bio-silica on drought tolerance in plants. *IOP Conference Series: Earth and Environmental Science*, 183, 012014. <https://doi.org/10.1088/1755-1315/183/1/012014>
- Santi, L. P., Nurhaimi-Haris, & Mulyanto, D. (2018). Effect of bio-silica on drought tolerance in plants. *IOP Conference Series: Earth and Environmental Science*, 183, 012014. <https://doi.org/10.1088/1755-1315/183/1/012014>
- Sauer, M., Porro, D., Mattanovich, D., & Branduardi, P. (2008). Microbial production of organic acids: expanding the markets. *Trends in Biotechnology*, 26(2), 100–108. <https://doi.org/10.1016/j.tibtech.2007.11.006>
- Savant, N., Snyder, G., & Datnoff, L. (1996). Silicon Management and Sustainable Rice Production. *Advances in Agronomy*, 151–199. [https://doi.org/10.1016/s0065-2113\(08\)60255-2](https://doi.org/10.1016/s0065-2113(08)60255-2)
- Schulze, D. G. (2018). An Introduction to Soil Mineralogy. *SSSA Book Series*, 1–34. <https://doi.org/10.2136/sssabookser1.2ed.c1>
- Setiawati, T. C., & Mutmainnah, L. (2016). Solubilization of Potassium Containing Mineral by Microorganisms From Sugarcane Rhizosphere. *Agriculture and Agricultural Science Procedia*, 9, 108–117. <https://doi.org/10.1016/j.aaspro.2016.02.134>
- Shamshuddin, J., & Wan, N. (2011). Classification and Management of Highly Weathered Soils in Malaysia for Production of Plantation Crops. *Principles, Application and Assessment in Soil Science*. <https://doi.org/10.5772/29490>
- Shanti Naidu, K. (2011). Characterization and purification of protease enzyme. *Journal of Applied Pharmaceutical Science*.
- Sheng, X. F., Zhao, F., He, L. Y., Qiu, G., & Chen, L. (2008). Isolation and characterization of silicate mineral-solubilizing *Bacillus globisporus* Q12 from the surfaces of weathered feldspar. *Canadian Journal of Microbiology*, 54(12), 1064–1068. <https://doi.org/10.1139/w08-089>

- Shetty, R., Jensen, B., Shetty, N. P., Hansen, M., Hansen, C. W., Starkey, K. R., & Jørgensen, H. J. L. (2011). Silicon induced resistance against powdery mildew of roses caused by *Podosphaera pannosa*. *Plant Pathology*, *61*(1), 120–131. <https://doi.org/10.1111/j.1365-3059.2011.02493.x>
- Shokri, D., & Emtiazi, G. (2010). Indole-3-Acetic Acid (IAA) Production in Symbiotic and Non-Symbiotic Nitrogen-Fixing Bacteria and its Optimization by Taguchi Design. *Current Microbiology*, *61*(3), 217–225. <https://doi.org/10.1007/s00284-010-9600-y>
- Siddiqui, M. H., & Al-Whaibi, M. H. (2014). Role of nano-SiO<sub>2</sub> in germination of tomato (*Lycopersicon esculentum* seeds Mill.). *Saudi Journal of Biological Sciences*, *21*(1), 13–17. <https://doi.org/10.1016/j.sjbs.2013.04.005>
- Silva, R. V., Oliveira, R. D. L., Nascimento, K. J. T., & Rodrigues, F. A. (2010). Biochemical responses of coffee resistance against *Meloidogyne exigua* mediated by silicon. *Plant Pathology*, *59*(3), 586–593. <https://doi.org/10.1111/j.1365-3059.2009.02228.x>
- Singh, R., Singh, S., Parihar, P., Mishra, R. K., Tripathi, D. K., Singh, V. P., Chauhan, D. K., & Prasad, S. M. (2016). Reactive Oxygen Species (ROS): Beneficial Companions of Plants' Developmental Processes. *Frontiers in Plant Science*, *7*, 1299. <https://doi.org/10.3389/fpls.2016.01299>
- Siri-udom, S., Suwannarach, N., & Lumyong, S. (2017). Applications of volatile compounds acquired from *Muscodor heveae* against white root rot disease in rubber trees (*Hevea brasiliensis* Müll. Arg.) and relevant allelopathy effects. *Fungal Biology*, *121*(6–7), 573–581. <https://doi.org/10.1016/j.funbio.2017.03.004>
- Soekirman, P., & Budi, S. (2009). Integrated disease management of white root disease on Hevea rubber using *Trichoderma* and antagonistic. In I. R. Institute, *Disease Management Strategies in Plantations* (pp. 4-8). Jogjakarta: AusAID sponsored workshop.
- Sommer, M., Kaczorek, D., Kuzyakov, Y., & Breuer, J. (2006). Silicon pools and fluxes in soils and landscapes—a review. *Journal of Plant Nutrition and Soil Science*, *169*(3), 310–329. <https://doi.org/10.1002/jpln.200521981>
- Song, A., Li, P., Fan, F., Li, Z., & Liang, Y. (2014). The Effect of Silicon on Photosynthesis and Expression of Its Relevant Genes in Rice (*Oryza sativa* L.) under High-Zinc Stress. *PLoS ONE*, *9*(11), e113782. <https://doi.org/10.1371/journal.pone.0113782>
- Song, A., Xue, G., Cui, P., Fan, F., Liu, H., Yin, C., Sun, W., & Liang, Y. (2016). The role of silicon in enhancing resistance to bacterial blight of hydroponic- and soil-cultured rice. *Scientific Reports*, *6*(1). <https://doi.org/10.1038/srep24640>



- Sousa, R. S., Rodrigues, F. V., Schurt, D. A., Souza, N. F. A., & Cruz, M. F. A. (2013). Cytological aspects of the infection process of *Pyricularia oryzae* on leaves of wheat plants supplied with silicon. *Tropical Plant Pathology*, 38(6), 472–477. <https://doi.org/10.1590/s1982-56762013000600002>
- Stewart, A., Brownbridge, M., Hill, R. A., & Jackson, T. A. (2010). Utilizing Soil Microbes for Biocontrol. *Soil Microbiology and Sustainable Crop Production*, 315–371. [https://doi.org/10.1007/978-90-481-9479-7\\_9](https://doi.org/10.1007/978-90-481-9479-7_9)
- Stoops, G., Marcelino, V., & Mees, F. (2010). *Interpretation of Micromorphological Features of Soils and Regoliths* (1st ed.). Elsevier Science.
- Subhan, S., Mujahid, T., Wahab, A., Masnoon, J., Ahmed, N., & Abbas, T. (2015). Effects of Different Physical and Chemical Parameters on Phosphate Solubilization Activity of Plant Growth Promoting Bacteria Isolated from Indigenous Soil. *Journal of Pharmacy and Nutrition Sciences*, 5, 64–70. doi: 10.6000/1927-5951.2015.05.01.10
- Sun, W., Zhang, J., Fan, Q., Xue, G., Li, Z., & Liang, Y. (2010). Silicon-enhanced resistance to rice blast is attributed to silicon-mediated defence resistance and its role as physical barrier. *European Journal of Plant Pathology*, 128(1), 39–49. <https://doi.org/10.1007/s10658-010-9625-x>
- Suzuki, S., Ma, J. F., Yamamoto, N., Hattori, T., Sakamoto, M., & Umezawa, T. (2012). Silicon deficiency promotes lignin accumulation in rice. *Plant Biotechnology*, 29(4), 391–394. <https://doi.org/10.5511/plantbiotechnology.12.0416a>
- Tahir, H. A. S., Gu, Q., Wu, H., Niu, Y., Huo, R., & Gao, X. (2017). *Bacillus volatiles* adversely affect the physiology and ultra-structure of *Ralstonia solanacearum* and induce systemic resistance in tobacco against bacterial wilt. *Scientific Reports*, 7(1). <https://doi.org/10.1038/srep40481>
- Takahashi, E., Ma, J. F., & Miyake, Y. (1990). The possibility of silicon as an essential element for higher plants. *Comments on Agricultural and Food Chemistry*.
- Tateoka, R. (1956). A weathering sequence of minerals and rock-grains in sand-dune soils. *Soil Science and Plant Nutrition*, 2(1), 1–3. <https://doi.org/10.1080/00380768.1956.10431845>
- Tokura, A. M., Furtini Neto, A. E., Curi, N., Carneiro, L. F., & Alovise, A. A. (2007). Silicon and phosphorus in soils cultivated with upland rice. *Acta Scientiarum-Agronomy*, 29(1), 9–16.
- Treguer, P. (2016). Silica. *Encyclopedia of Marine Geosciences*, 1. [https://doi.org/10.1007/978-94-007-6644-0\\_95-3](https://doi.org/10.1007/978-94-007-6644-0_95-3)

- Trudinger, P., & Bubela, B. (1967). Microorganisms and the natural environment. *Mineralium Deposita*, 2(3). <https://doi.org/10.1007/bf00201911>
- Tubana, B. S., Babu, T., & Datnoff, L. E. (2016). A Review of Silicon in Soils and Plants and Its Role in US Agriculture. *Soil Science*, 181(9/10), 393–411. <https://doi.org/10.1097/ss.0000000000000179>
- Tubaña, B. S., & Heckman, J. R. (2015). Silicon in Soils and Plants. *Silicon and Plant Diseases*, 7–51. [https://doi.org/10.1007/978-3-319-22930-0\\_2](https://doi.org/10.1007/978-3-319-22930-0_2)
- Uroz, S., Calvaruso, C., Turpault, M. P., & Frey-Klett, P. (2009). Mineral weathering by bacteria: ecology, actors and mechanisms. *Trends in Microbiology*, 17(8), 378–387. <https://doi.org/10.1016/j.tim.2009.05.004>
- Uroz, S., Kelly, L. C., Turpault, M. P., Lepleux, C., & Frey-Klett, P. (2015). The Mineralosphere Concept: Mineralogical Control of the Distribution and Function of Mineral-associated Bacterial Communities. *Trends in Microbiology*, 23(12), 751–762. <https://doi.org/10.1016/j.tim.2015.10.004>
- Urrutia, M. M., & Beveridge, T. J. (1994). Formation of fine-grained metal and silicate precipitates on a bacterial surface (*Bacillus subtilis*). *Chemical Geology*, 116(3–4), 261–280. [https://doi.org/10.1016/0009-2541\(94\)90018-3](https://doi.org/10.1016/0009-2541(94)90018-3)
- van Beilen, J. B., & Poirier, Y. (2007). Establishment of new crops for the production of natural rubber. *Trends in Biotechnology*, 25(11), 522–529. <https://doi.org/10.1016/j.tibtech.2007.08.009>
- van Bockhaven, J., de Vleeschauwer, D., & Höfte, M. (2012). Towards establishing broad-spectrum disease resistance in plants: silicon leads the way. *Journal of Experimental Botany*, 64(5), 1281–1293. <https://doi.org/10.1093/jxb/ers329>
- Vasanthi, N., Saleena, L. M., Anthoni Raj, S., Rom, S., Biootech, V., Vijay, D. R., & Pvt, B. (2013). Evaluation of media for isolation and screening of silicate solubilising bacteria. *International Journal of Current Research*, 5(2), 3–5.
- Vasanthi, N., Saleena, L. M., & Raj, S. A. (2016). Silica Solubilization Potential of Certain Bacterial Species in the Presence of Different Silicate Minerals. *Silicon*, 10(2), 267–275. <https://doi.org/10.1007/s12633-016-9438-4>
- Vulavala, V. K. R., Elbaum, R., Yermiyahu, U., Fogelman, E., Kumar, A., & Ginzberg, I. (2015). Silicon fertilization of potato: expression of putative transporters and tuber skin quality. *Planta*, 243(1), 217–229. <https://doi.org/10.1007/s00425-015-2401-6>
- Wang, M., Gao, L., Dong, S., Sun, Y., Shen, Q., & Guo, S. (2017). Role of Silicon on Plant–Pathogen Interactions. *Frontiers in Plant Science*, 8. <https://doi.org/10.3389/fpls.2017.00701>

- Wang, Q., Sheng, X. F., He, L. Y., & Shan, Y. (2018). Improving bio-desilication of a high silica bauxite by two highly effective silica-solubilizing bacteria. *Minerals Engineering*, 128, 179–186. <https://doi.org/10.1016/j.mineng.2018.08.041>
- Wang, R. R., Wang, Q., He, L. Y., Qiu, G., & Sheng, X. F. (2015). Isolation and the interaction between a mineral-weathering *Rhizobium tropici* Q34 and silicate minerals. *World Journal of Microbiology and Biotechnology*, 31(5), 747–753. <https://doi.org/10.1007/s11274-015-1827-0>
- Wastie, R. L. (1975). Diseases of Rubber and their Control. *PANS Pest Articles & News Summaries*, 21(3), 268–288. <https://doi.org/10.1080/09670877509411408>
- Wattanasilakorn, S., Sdoodee, S., Nualsri, C., & Bunratchoo, S. (2015). Screening of rubber rootstock by the assessment of root growth and genetic background. *Kasetsart Journal - Natural Science*.
- Wattanasilakorn, S., Sdoodee, S., Nualsri, C., & Chuenchit, S. (2012). Screening of rubber (*Hevea brasiliensis* Muell. Arg.) rootstocks for the white root disease resistance. *Journal of Agricultural Technology*.
- Watteau, F., & Villemin, G. (2001). Ultrastructural study of the biogeochemical cycle of silicon in the soil and litter of a temperate forest. *European Journal of Soil Science*, 52(3), 385–396. <https://doi.org/10.1046/j.1365-2389.2001.00391.x>
- Weiner, S. (2003). An Overview of Biomineralization Processes and the Problem of the Vital Effect. *Reviews in Mineralogy and Geochemistry*, 54(1), 1–29. <https://doi.org/10.2113/0540001>
- Welch, S. A., & Ullman, W. J. (1993). The effect of organic acids on plagioclase dissolution rates and stoichiometry. *Geochimica et Cosmochimica Acta*, 57(12), 2725–2736. [https://doi.org/10.1016/0016-7037\(93\)90386-b](https://doi.org/10.1016/0016-7037(93)90386-b)
- White, A., & Buss, H. (2014). Natural Weathering Rates of Silicate Minerals. *Treatise on Geochemistry*, 115–155. <https://doi.org/10.1016/b978-0-08-095975-7.00504-0>
- White, P. J., & Brown, P. H. (2010). Plant nutrition for sustainable development and global health. *Annals of Botany*, 105(7), 1073–1080. <https://doi.org/10.1093/aob/mcq085>
- Woraathasi, N., Nakkanong, K., & Nualsri, C. (2017). Cloning and Expression Analysis of HbPR-1b and HbPR-3 in *Hevea brasiliensis* During Inoculation with *Rigidoporus microporus*. *Pakistan Journal of Biological Sciences*, 20(5), 233–243. <https://doi.org/10.3923/pjbs.2017.233.243>

- Wu, H., Gu, Q., Xie, Y., Lou, Z., Xue, P., Fang, L., Yu, C., Jia, D., Huang, G., Zhu, B., Schneider, A., Blom, J., Lasch, P., Borriss, R., & Gao, X. (2019). Cold-adapted Bacilli Isolated from the Qinghai–Tibetan Plateau are Able to Promote Plant Growth in Extreme Environments. *Environmental Microbiology*, 21(9), 3505–3526. <https://doi.org/10.1111/1462-2920.14722>
- Wu, Y. W., Zhang, J. C., Wang, L. J., & Wang, Y. X. (2017). A rock-weathering bacterium isolated from rock surface and its role in ecological restoration on exposed carbonate rocks. *Ecological Engineering*, 101, 162–169. <https://doi.org/10.1016/j.ecoleng.2017.01.023>
- Youssef, S. A., Tartoura, K. A., & Abdelraouf, G. A. (2016). Evaluation of *Trichoderma harzianum* and *Serratia proteamaculans* effect on disease suppression, stimulation of ROS-scavenging enzymes and improving tomato growth infected by *Rhizoctonia solani*. *Biological Control*, 100, 79–86. <https://doi.org/10.1016/j.biocontrol.2016.06.001>
- Yu, S., Teng, C., Bai, X., Liang, J., Song, T., Dong, L., Jin, Y., & Qu, J. (2017). Optimization of Siderophore Production by *Bacillus* sp. PZ-1 and Its Potential Enhancement of Phytoextraction of Pb from Soi. *Journal of Microbiology and Biotechnology*, 27(8), 1500–1512. <https://doi.org/10.4014/jmb.1705.05021>
- Zargar, S. M., Mahajan, R., Bhat, J. A., Nazir, M., & Deshmukh, R. (2019). Role of silicon in plant stress tolerance: opportunities to achieve a sustainable cropping system. *3 Biotech*, 9(3). <https://doi.org/10.1007/s13205-019-1613-z>
- Zehnder, G. W., Murphy, J. F., Sikora, E. J., & Kloepper, J. W. (2001). Application of rhizobacteria for induced resistance. *European Journal of Plant Pathology*, 107(1), 39–50. <https://doi.org/10.1023/a:1008732400383>
- Zhan, S., Liu, J., Chen, Y., & Sun, D. (2013). Single and Cooperative Bauxite Bioleaching by Silicate Bacteria. *IERI Procedia*, 5, 172–177. <https://doi.org/10.1016/j.ieri.2013.11.088>
- Zhang, G., Cui, Y., Ding, X., & Dai, Q. (2013). Stimulation of phenolic metabolism by silicon contributes to rice resistance to sheath blight. *Journal of Plant Nutrition and Soil Science*, 176(1), 118–124. <https://doi.org/10.1002/jpln.201200008>
- Zhang, M., Liang, Y., & Chu, G. (2017). Applying silicate fertilizer increases both yield and quality of table grape (*Vitis vinifera* L.) grown on calcareous grey desert soil. *Scientia Horticulturae*, 225, 757–763. <https://doi.org/10.1016/j.scienta.2017.08.019>
- Zhao, X., Zhou, Z. J., Han, Y., Wang, Z. Z., Fan, J., & Xiao, H. Z. (2013). Isolation and identification of antifungal peptides from *Bacillus* BH072, a novel bacterium isolated from honey. *Microbiological Research*, 168(9), 598–606. <https://doi.org/10.1016/j.micres.2013.03.001>

Zhou, Y. J., Li, J. H., Ross Friedman, C., & Wang, H. F. (2017). Variation of Soil Bacterial Communities in a Chronosequence of Rubber Tree (*Hevea brasiliensis*) Plantations. *Frontiers in Plant Science*, 8. <https://doi.org/10.3389/fpls.2017.00849>

