

A Tropical Harpacticoid Copepod, *Nitocra affinis californica* Lang As an Effective Live Feed for Black Tiger Shrimp Larvae *Penaeus monodon* Fabricius

Hazel Monica Matias-Peralta^{1,#4}, Fatimah Md. Yusoff^{1,2*}, Mohamed Shariff¹ and Suhaila Mohamed³

¹Laboratory of Marine Biotechnology, Institute of Bioscience, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

²Department of Aquaculture, Faculty of Agriculture, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

³Laboratory of Molecular Biomedicine, Institute of Bioscience, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

^{#4}Department of Technology and Heritage, Faculty of Science, Technology and Human Development, Universiti Tun Hussein Onn Malaysia, 86400 Parit Raja, Batu Pahat, Johor, Malaysia

ABSTRACT

Survival and specific growth rates of *Penaeus monodon* larvae (post-larval stages 1-15), fed with different live feeds and artificial diets, were evaluated using three different treatments, namely: i) *Nitocra affinis*, ii) combination of *Artemia* nauplii + *N. affinis* and iii) artificial diet. The experiment was carried out in 10-L aquaria with 30% daily water exchange for a period of 16 days. The survival rate (61%) and specific growth rate (16.7 %day⁻¹) were highest (p<0.05) in the treatment with shrimp larvae fed with *N. affinis*. Likewise, the protein contents of *N. affinis* was found to be the highest (p<0.05) among all the diets used. The fatty acids of *N. affinis* was dominated by polyunsaturated fatty acids (PUFA) (22:6n3) forming 19.5% of the total PUFA identified. In fact, *N. affinis* contained the highest (p<0.05) amount of PUFA and the highest (p<0.05) n-3/n-6 ratio amongst the three diets. Analysis of the copepod fed shrimp showed significantly higher (p<0.05) amount of long chain PUFA, both of the n-3 and n-6 series fatty acids, when compared to the artificial diet fed larvae. The results of this study showed that *N. affinis* has the potential to be used as

an effective live feed for *P. monodon* due to their high PUFA contents and broad size range (nauplii to adults) to cater for different shrimp post-larval stages.

Keywords: Harpacticoid copepod, Live feed, *Nitocra affinis californica*, *Penaeus monodon*, Shrimp larvae

ARTICLE INFO

Article history:

Received: 20 March 2009

Accepted: 31 May 2012

E-mail addresses:

monica@uthm.edu.my (Hazel Monica Matias-Peralta),

fatimah@ibs.upm.edu.my (Fatimah Md. Yusoff),

shariff@vet.upm.edu.my (Mohamed Shariff),

mohamed.suhaila@gmail.com (Suhaila Mohamed)

* Corresponding author

Current affiliation

INTRODUCTION

Today, the larviculture of marine decapod crustaceans is still regarded as one of the major bottlenecks impairing the production of several commercially important species (Calado *et al.*, 2008). Meanwhile, live food has been considered to be a limiting factor in the commercial larval production of many fish and crustacean species (Kovalenko *et al.*, 2002), and it forms an important factor in overall production cost. For penaeid larvae, animal protein is the most important component of the diet (Anderson & De Silva, 2003; Perera *et al.*, 2005), and this is generally supplied by live feeds such as rotifers and brine shrimp, *Artemia*. Although live feeds have been proven successful as animal protein source in raising the larvae of many species, inherent problems still remain. The problems include variable nutrient composition and availability, potential introduction of pathogens into the culture system, and in the case of rotifers, the cultures are prone to crashes (Jones *et al.*, 1993; Kovalenko *et al.*, 2002). Adequate production of quality live food microorganisms, at a lowest possible cost, is essential for the successful development of commercial fish or shrimp hatchery since artificial diets developed have so far not been fully adequate, and generally, the use of live food has yielded better results (Tseng & Hsu, 1984; Watanabe & Kiron, 1994; Calado *et al.*, 2008).

The increased global demand for seed-stock experienced by shrimp farming industry, specifically *P. monodon*, during the last two decades has prompted extensive

improvement and refinement of the basic larval rearing techniques (Jory, 1997; Martin *et al.*, 2006). The objective of this study was to obtain predictable production of high quality hatchery reared penaeid post-larvae by improving feeding regimes evaluated using three different treatments (Wilkenfed *et al.*, 1981, 1984; Kuban *et al.*, 1987; Barros & Valenti, 2003).

The harpacticoid, *Nitocra affinis*, is a widely distributed species found commonly in coastal areas, living on fine sand or mud, in shell sand among stones and algae, in mangrove sea, in salty wells, tidal pools and shore reefs (Lang, 1948; 1965; Kunz, 1975). They are epibenthic and euryhaline with a salinity tolerance well beyond the range of most natural salinity fluctuations. Their food requirement is flexible, as they can live and thrive on a variety of food such as algae, bacteria, yeast and artificial food sources (Hicks & Coull, 1983; Gee, 1989; Weiss *et al.*, 1996). Thus, *N. affinis* is a desirable organism for experimental work and a good candidate for mass cultivation. The present study was undertaken to evaluate the survival and relative growth rate of *P. monodon* post-larvae fed with *N. affinis* solely, and in combination with the traditionally used *Artemia* nauplii and artificial shrimp diet for a period of 16 days.

MATERIALS AND METHODS

Treatments

Three treatments were employed in this experiment. *Penaeus monodon* (post-larvae stages 1-15) were fed with i) *N. affinis* (size range: 50-400 μm), ii) a combination of

Artemia nauplii (size range: 410-430 μ m) and *N. affinis*, and iii) artificial diet (ground freeze-dried form, size range: 17-22 μ m). For each treatment, three replicates were randomly assigned. The experiment was run for 16 days. The use of *Artemia*, as sole live food source, was excluded in this study due to the fact that *Artemia* is in no doubt still the prime live feed for the larviculture industry (Jones *et al.*, 1989; Abelin *et al.*, 1991; Samocha *et al.*, 1999; Wouters & Van Horenbeeck, 2003; Robinson *et al.*, 2005). In addition, it is well established that the use of *Artemia* can yield from 50% up to 75% survival of shrimp post larvae in commercial hatcheries throughout the world (Jones *et al.*, 1989; Abelin *et al.*, 1991; Samocha *et al.*, 1999; Wouters & Van Horenbeeck, 2003; Robinson *et al.*, 2005).

Species Isolation

Nitocra affinis californica was isolated from the coastal waters of Peninsular Malaysia. The copepods were collected using Schindler Patalas trap, along the sandy beach with water level reaching 0.35 m. The copepods were transferred into a 1-L beaker with filtered (0.20 μ m filter) seawater, and provided with 24 hour-aeration for conditioning. Then, each copepod carrying egg sacs was separately transferred into a new container filled with filtered (0.20 μ m filter) seawater, while mixed algae (*Chaetoceros calcitrans*, *Nannochloropsis oculata* and *Tetraselmis tetrahele*) were provided as food sources. To ensure a monospecies stock, the culture was started

with one gravid female grown continuously for several generations in the laboratory.

Feed Preparation

Small scale mass production of *N. affinis* was started by transferring 150 gravid female copepods (from the original stock) in 2-L of filtered aged seawater to each culture vessel. The 3-L culture vessels were cylindrical basins (semi transparent and lightly coloured plastics with smooth surface on the inside and out) of polypropylene material, with a base area equals to 177 cm², a mouth area of 416 cm², and a height of 10 cm. The aged seawater (seawater stocked for a period of time; chlorine free and with stable water chemistry) was filtered in a 0.45 μ m membrane filter paper before use. The culture was maintained at salinity 30 g·L⁻¹ and at temperature levels between 25°C to 35°C (seawater at room temperature changed from a minimum of 25°C, occurring in the early morning while a maximum of 35°C usually in the mid afternoon). When salinity increased, fresh water was added to the culture vessel until the desired salinity was achieved. The cultures were exposed to natural lighting condition (ranging from 25-40 μ mol m²s⁻¹) and photoperiod (12h light:12h dark cycles). All the above parameters provided for in this study were determined to provide the maximum population growth of *N. affinis* (Matias-Peralta *et al.*, 2005). Maintenance of copepod culture involves siphoning of waste water, replenishing the vessels with fresh aged water and feeding (Matias-Peralta *et*

al., 2011). In other words, copepods were harvested daily, in such a way that gravid females were separated from nauplii and copepodids (Matias-Peralta *et al.*, 2011). The procedure described above was able to produce 14.9×10^3 copepod L^{-1} for a period of 14 days, with a total of 75 gravid females L^{-1} as inoculum (Matias-Peralta *et al.*, 2011).

The *Artemia* nauplii were produced by incubating cysts for 18 hours in seawater under optimal conditions of temperature (28-30°C), salinity (30-32 $g L^{-1}$), pH (7.5-8.0) and continuous illumination using the Philips white light tubes. The cysts were incubated in a 1-L capacity conical shaped container at a density of 2 g cysts L^{-1} of seawater. The production of *Artemia* nauplii without enrichment followed the standard procedure adapted by shrimp hatcheries in Malaysia (Shrimp hatchery operators in Negeri Sembilan, Malacca and Johor, Malaysia, pers. com.).

Artificial feed was prepared by weighing the recommended amount of feed according to the number of shrimp larvae used in the experiment. This procedure was done according to the manufacturer's recommendation.

Source of P. monodon Post larvae (PL)

The shrimp larvae were purchased from a private hatchery located in Port Dickson, Negeri Sembilan (approximately 100 km from the laboratory). Shrimps at their first mysis stage were transported from the hatchery to the Aquatic Animal Health Unit for acclimation and further rearing until molting to PL1 (11 days old). The mysis

were fed ad libitum four times daily, with a diatom, *Skeletonema* sp., until they were stocked into the experimental tanks.

Tank Preparation

Twelve 10 L capacity plastic aquaria (made of transparent acrylic plastic) with the dimension of 31.5 cm x 16.5 cm x 24 cm were used in the experiment. The tanks were thoroughly washed with bactericidal soap, soaked in 100 $mg L^{-1}$ chlorine, rinsed and dried to prevent any disease transfer during the culture. Then, the tanks were filled with 4 L of chlorinated (aerated under high pressure air bubbles for at least three days before use), filtered (1 μm) seawater a day before stocking. A few pieces of stones and 20 mm diameter plastic pipes cut into a length of 40 mm (which were soaked in 100 $mg L^{-1}$ chlorine for 24 hours, washed, rinsed and dried) were placed inside the tanks to serve as refuge for the weak shrimp during the molting period. Each tank was provided with individual aeration of low pressure air bubbles.

Stocking

The shrimp post larvae (PL1), with an average initial weight of 34.24 ± 1.02 mg, were hauled from the acclimation tank using soft net into a basin with fresh seawater. The larvae were collected from the basin using a wide mouth plastic bottle with attached soft net, counted and transferred into a tared beaker with seawater and weighed in a Vibra (Shinko Denshi) digital tuning fork scale. These procedures were done quickly and carefully so as to avoid stress that might

lead to shrimp mortality. Individual weight was calculated from the bulk weight over the total number of individuals. Each tank was stocked at a density of 45 PLs L⁻¹ of seawater.

Feeding

Larval live feeds (*Artemia* nauplii, Superior 90 Brine Shrimp Eggs, USA; and *N. affinis*) and artificial shrimp diet (Mixed Feed for *P. monodon*, Higashimaru Co. Ltd.) were given ad libitum four times a day at six hours interval (0600, 1200, 1800 and 2400 hours, respectively). Live feeds were counted, while artificial diet was weighed prior to feeding. Live feeds were also washed with filtered seawater upon harvest from their respective culture containers before counting. Live feed counts were done by determining the number of individuals/ml in three 1 ml samples using graduated pipette. The counts were done using 1 ml gridded counting chamber under Leitz Diavert inverted microscope. The counts were also done daily before feeding. The dry weights of the live food were also determined to be compared with that of the artificial diet (Table 3).

Water Quality

Water parameters, such as salinity, pH, dissolved oxygen, and temperature, were measured *in situ* twice (morning and noon) daily. Salinity and temperature readings were done using YSI 30 Salinity/Conductivity/Temperature meter, while YSI 52 Dissolved Oxygen meter and ORION

portable pH meter were used for dissolved oxygen and pH reading, respectively. On the other hand, water samples for total ammonia nitrogen and nitrite nitrogen were collected (before cleaning the tanks) and analyzed daily in the laboratory following the procedures by Parsons *et al.* (1984).

Meanwhile, daily water exchange was carried out at 30%. Uneaten food and faeces were siphoned out using a 1 mm flexible hose. Using this hose, the water level was gradually reduced to provide minimal disturbance to the shrimp larvae and also to prevent the larvae from being siphoned out. Feeding was commenced as soon as the desired amount of seawater had been removed and the refilling was begun.

Feeding was stopped eight hours before harvesting. Upon harvest, all the shrimp larvae were collected, weighed and measured. All the collected samples were washed thoroughly with double distilled water to remove salt before freeze drying (prior to analysis). Protein was determined following the method described by Meyer and Walther (1988). Fatty acid methyl esters (FAME) were prepared according to the direct methylation techniques by Divakaran and Ostrowski (1989). The freeze dried samples (100 mg) were refluxed at 100°C for 10 minutes with 10 mL of 2% methanolic NaOH. The samples were further refluxed with 6.25 mL 14% Boron Trifluoride and Heptane. Then, 2 mL of saturated NaCl was added to make the FAME float, and 1 g of anhydrous Na₂SO₄ was added to absorb the remaining water in the FAME for best recovery.

The FAME of the samples were analyzed using gas liquid chromatograph (Shimadzu GC-8A) equipped with a FID and BPX-70 (SGE) or Supelco 2330 capillary column. Individual peaks of FAME were identified by comparing them with the retention times of known standards obtained from Sigma Chemicals Company and using cod liver oil as a secondary standard. A Chromatopac (SHIMADZU C-R3A) quantified the magnitude of the peaks of each chromatographic reading.

Data Analysis

The specific growth rate (SGR) was calculated from the body weight using the following formula:

$$\text{SGR} = 100 [(\exp G) - 1]$$

Where:

$$G = \frac{\ln(wf) - \ln(wi)}{\Delta t}$$

where:

$\ln(wf)$ is the natural logarithm of the weight at time (t),

and $\ln(wi)$ is the natural logarithm of the initial wt of the shrimp larvae.

The collected data were analyzed using one-way analysis of variance (ANOVA). Significant differences among the individual treatment effects were determined using Tukey's honestly significant different test (T-HSD) at 0.05 level of probability. The data (whenever appropriate) were arcsine-transformed to satisfy the condition of homogeneity of variance (Gomez & Gomez, 1983; Zar, 1984). Statistical analyses were done using the Statistical Analysis System (SAS Inc. 1992) computer package (version 6.07).

RESULTS

Shrimp larvae fed with *N. affinis* alone achieved the highest ($p < 0.05$) survival rate (60.6%), whereas those fed with the combination of *Artemia* + *N. affinis* had the lowest survival rate (24.1%) (see Table 1). Similarly, the highest ($p < 0.05$) specific growth rate (SGR) was achieved by the shrimps fed with *N. affinis* alone, while those fed with the combination of *Artemia* and harpacticoid copepod have the lowest ($p < 0.05$) growth rate (14.3%) (see Table 1).

TABLE 1

Mean (\pm S.E.) survival, specific growth rate of shrimp larvae and total weight of different diets fed to *Penaeus monodon* larvae for a period of 15 days. Means in the rows with the same superscript are not significant ($p < 0.05$).

Parameters	Treatment		
	I	II	Control
Survival (%)	60.6 ^a ±6.8	24.1 ^c ±1.0	43.0 ^b ±3.2
Specific growth rate (% d ⁻¹)	16.7 ^a ±0.2	14.3 ^c ±0.2	15.8 ^b ±0.2
Total weight of feed used (mg dry weight)	94.9 ^b ±2.4	53.0 ^c ±0.6	109.8 ^a ±4.5

Note: Treatment I = fed *N. affinis* (size range 50-400 μm)

Treatment II = combination of *Artemia* nauplii (size range 410-430 μm) and *N. affinis*

Treatment III = artificial diet (grounded freeze-dried form, size range 17-22 μm).

Meanwhile, significant differences in the total amount of feed consumed were found among the treatments (Table 1). The shrimp larvae consumed significantly ($p < 0.05$) higher amount of artificial diet compared to the live feed used.

The results also revealed that the highest ($p < 0.05$) level of protein in the copepod tissue ($52\% = 52.32 \pm 2.13$) compared to the two other diets used (namely, *Artemia* nauplii = 34% or 33.82 ± 0.35 and artificial diet = 40% or 39.82 ± 0.39). The fatty acids of *N. affinis* were dominated by polyunsaturated fatty acids (PUFA), 22:6n-3 accounting 19.5% of the total PUFA identified (see Table 2). On the other hand, *Artemia* nauplii and artificial diet were dominated by the monounsaturated fatty acids (MUFA), 18:1n-9 contributing 31.3% and 21.3% of the total MUFA identified, respectively. Among the three diets, the artificial diet was found to contain the highest ($p < 0.05$) amount of saturated fatty acids (SFA), whereas *Artemia* was found to contain the highest ($p < 0.05$) MUFA, and *N. affinis* contained the highest ($p < 0.05$) amount of PUFA (Table 2). The results also showed that *N. affinis* contained the highest ($p < 0.05$) n-3/n-6 ratio compared to the other feed used.

The copepod fed shrimp showed significantly higher ($p < 0.05$) amount of long chain PUFA, both of the n-3 and n-6 series fatty acids (Table 3), as compared to the artificial diet fed larvae. The fatty acid of the artificial diet fed shrimp was dominated by MUFA, specifically 18:1 and 16:1. However, the MUFA content of

shrimp larvae fed with either copepod or artificial diet did not show any significant difference (Table 3).

No significant difference ($p > 0.05$) was detected in the water with regards to salinity, pH, dissolved oxygen and temperature from all the treatments (Table 4). Nonetheless, significantly high ($p < 0.05$) total ammonia nitrogen (TAN) and nitrite nitrogen ($\text{NO}_2\text{-N}$) were detected from the tanks containing the artificial feed (control) (Table 4). On the other hand, the tanks with *N. affinis* (treatment II) was found to contain the lowest ($p < 0.05$) concentrations of TAN and $\text{NO}_2\text{-N}$ compared to the other treatments (Table 4).

DISCUSSION

This study illustrated that *N. affinis* was the preferred food by the *P. monodon* larvae. Meanwhile, the *Nitocra affinis* fed shrimp larvae achieved the highest survival rate and SGR compared to those fed with other food items. According to Mock *et al.* (1980), food organism in an intermediate size between algae ($\leq 8\text{-}14\mu\text{m}$) and freshly hatched *Artemia* nauplii ($\leq 430\mu\text{m}$) could increase shrimp survival and growth rates. In addition, Motoh (1981) suggested that a gradual change from algal diet during acclimatization to a diet of *Artemia* nauplii, which was fairly large ($\leq 430\mu\text{m}$) for small shrimp larvae with carapace length ranging from 0.5-2.2 mm with intermediate sized food items could reduce stress and mortality. In this study, harpacticoid copepod from copepodid to adult (ranging from 50 to $400\mu\text{m}$) served as an appropriate sized prey

TABLE 2

Means (\pm S.E.) of different fatty acids (% of total fatty acids) of food sources for the post larvae *Penaeus monodon* Means in the rows with the same superscript are not significant ($p < 0.05$).

Fatty acids	<i>Nitocra affinis</i>	<i>Artemia nauplii</i>	Artificial diet
C12:0	nd	nd	11.76 ^a \pm 1.13
C14:0	0.65 ^b \pm 0.02	0.51 ^c \pm 0.00	1.47 ^a \pm 0.02
C14:1	0.60 ^b \pm 0.01	0.65 ^b \pm 0.00	1.37 ^a \pm 0.29
C14:2	6.65 ^a \pm 1.14	nd	nd
C15:0	0.65 ^b \pm 0.11	0.46 ^b \pm 0.00	14.39 ^a \pm 3.83
C16:0	11.83 ^b \pm 0.16	12.55 ^a \pm 0.00	5.06 ^c \pm 1.31
C16:1	3.26 ^a \pm 0.27	nd	1.14 ^b \pm 0.21
C16:1n - 7	1.31 ^b \pm 0.03	13.24 ^a \pm 0.00	nd
C18:0	nd	5.57 ^a \pm 0.00	1.54 ^b \pm 0.27
C18:1	nd	nd	4.61 ^a \pm 1.58
C18:1n - 9	14.10 ^c \pm 0.10	31.30 ^a \pm 0.00	21.26 ^b \pm 2.99
C18:2n - 6	9.73 ^a \pm 0.08	6.23 ^a \pm 0.00	
C18:3n - 3	3.55 ^b \pm 0.15	8.08 ^a \pm 0.00	0.48 ^c \pm 0.00
C18:3n - 6	0.37 ^a \pm 0.15	nd	0.13 ^b \pm 0.00
C18:4	0.76 ^a \pm 0.05	nd	0.37 ^b \pm 0.00
C18:4n - 3	0.86 ^b \pm 0.00	5.09 ^a \pm 0.00	nd
C20:1n-9	0.15 ^b \pm 0.02	nd	3.73 ^a \pm 1.79
C20:2	2.32 ^a \pm 0.26	nd	0.30 ^b \pm 0.11
C20:4n - 6	2.32 ^b \pm 0.26	1.41 ^c \pm 0.00	5.13 ^a \pm 0.06
C20:5n - 3	11.26 ^a \pm 0.73	nd	0.64 ^b \pm 0.19
C22:1n - 11	0.88 ^b \pm 0.18	0.62 ^a \pm 0.00	nd
22:5n - 3	2.57 ^a \pm 0.23	nd	nd
C22:6n - 3	19.50 ^a \pm 0.29	nd	8.07 ^b \pm 1.87
Total SFA	13.13 ^c \pm 0.14	19.09 ^b \pm 0.00	28.73 ^a \pm 6.11
Total MUFA	20.30 ^c \pm 0.26	45.81 ^a \pm 0.00	32.10 ^b \pm 6.87
Total PUFA	59.89 ^a \pm 1.27	20.81 ^b \pm 0.00	15.11 ^c \pm 1.63
Total n - 3	37.74 ^a \pm 0.62	13.17 ^b \pm 0.00	9.19 ^c \pm 1.68
Total n - 6	12.42 ^a \pm 0.80	7.64 ^b \pm 0.00	5.26 ^c \pm 0.06
n - 3/n - 6	3.04 ^a \pm 0.22	1.72 ^b \pm 0.00	1.75 ^b \pm 0.30

nd = not detectable; SFA = saturated fatty acids;

MUFA = monounsaturated fatty acids;

PUFA = polyunsaturated fatty acids.

TABLE 3

Mean (\pm S.E.) fatty acid composition (% of total fatty acids) of post larvae (PL 15) *Penaeus monodon* fed with different diets. Means in the rows with the same letters are not significant ($p < 0.05$).

Fatty acids	<i>Nitocra affinis</i> fed larvae	Artificial diet fed larvae
C14:0	3.25 ^a \pm 0.00	2.24 ^b \pm 0.12
C14:1	0.05 ^b \pm 0.00	1.80 ^a \pm 1.08
C15:0	0.29 ^a \pm 0.00	Nd
C16:0	11.25 ^a \pm 0.35	10.00 ^a \pm 0.99
C16:1	1.23 ^b \pm 0.00	4.10 ^a \pm 0.46
C16:1 $n-7$	3.57 ^a \pm 0.47	Nd
C18:0	4.90 ^b \pm 0.00	8.05 ^a \pm 0.46
C18:1	Nd	13.62 ^a \pm 2.49
C18:1 $n-9$	17.43 ^b \pm 1.54	21.17 ^a \pm 0.21
C18:2 $n-6$	2.60 ^a \pm 0.42	Nd
C18:3 $n-3$	1.80 ^a \pm 0.00	0.95 ^b \pm 0.03
C18:3 $n-6$	Nd	0.38 ^a \pm 0.21
C18:4	Nd	0.47 ^a \pm 0.01
C18:4 $n-3$	2.15 ^a \pm 0.14	Nd
C20:1 $n-9$	4.30 ^a \pm 0.00	3.04 ^b \pm 0.35
C20:2	Nd	0.98 ^a \pm 0.25
C20:4 $n-6$	4.71 ^a \pm 0.01	2.86 ^b \pm 0.76
C20:5 $n-3$	15.62 ^a \pm 0.47	1.08 ^b \pm 0.11
C22:1 $n-11$	1.13 ^b \pm 0.00	3.57 ^a \pm 0.11
C22:5 $n-3$	1.80 ^a \pm 0.00	Nd
C22:6 $n-3$	16.59 ^a \pm 0.02	16.33 ^a \pm 0.59
Total SFA	19.69 ^a \pm 0.35	19.78 ^a \pm 2.28
Total MUFA	27.71 ^a \pm 2.02	28.28 ^a \pm 1.19
Total PUFA	45.26 ^a \pm 0.16	23.04 ^b \pm 0.28
Total $n-3$	37.95 ^a \pm 0.59	18.36 ^b \pm 0.45
Total $n-6$	7.31 ^a \pm 0.43	3.24 ^b \pm 0.97

nd = not detectable; SFA = saturated fatty acids;

MUFA = monounsaturated fatty acids;

PUFA = polyunsaturated fatty acids.

TABLE 4

Mean (\pm S.E.) of different water quality parameters measured. Means in the rows with the same superscript are not significant ($p < 0.05$).

Parameters	Treatment		
	I	II	Control
Salinity (gL-1)	30.14 ^a \pm 0.09	30.14 ^a \pm 0.11	30.04 ^a \pm 0.10
Temperature (°C)	27.51 ^a \pm 0.10	27.52 ^a \pm 0.04	27.51 ^a \pm 0.10
pH (range)	7.90-8.31	7.90-8.31	7.92-8.30
Dissolved Oxygen (mg L-1)	5.85 ^a \pm 0.05	5.88 ^a \pm 0.04	6.20 ^a \pm 0.08
Total Ammonia-N (μ gL-1)	18.4 ^c \pm 0.06	19.5 ^a \pm 0.08	19.2 ^b \pm 0.08
Nitrite-N (μ gL-1)	24.3 ^c \pm 0.15	29.1 ^a \pm 0.15	28.3 ^b \pm 0.07

Note: Treatment I = fed *N. affinis* (size range 50-400 μ m)

Treatment II = combination of *Artemia* nauplii (size range 410-430 μ m) and *N. affinis*

Treatment III = artificial diet (ground freeze-dried form, size range 17-22 μ m).

for the shrimp larvae as they did not have to spend excess energy manipulating big sized prey.

Harpacticoid copepods have been widely used as a live prey for hatchery reared fish and crustaceans (Kahan *et al.*, 1982; Watanabe *et al.*, 1983; Szyper, 1989; Kraul *et al.*, 1992; 1993; Nanton & Castell, 1998; Shields *et al.*, 1999) because they have relatively high caloric content per unit weight and superior nutritional value (Kahan *et al.*, 1982; Volk *et al.*, 1984; Chandler, 1986; Gee, 1989). Although the absolute nutritional requirements of penaeid larvae are not fully known (Bienenbach *et al.*, 1989), Colvin and Brand (1977) reported that a relatively high level of protein is important for the growth and survival of the early post-larval stages of many important penaeid species. In fact, they further suggested that larval protein requirements must be equal to or greater than 44%. In this case, the copepod *N. affinis* has provided the much needed protein

requirement (52%) for the shrimp larvae. In addition, Leger *et al.* (1986), Norsker and Støttrup (1994), and McEvoy *et al.* (1995) have reported that harpacticoids are richer in essential fatty acids, most notably 22:6n-3 and 20:5n-3 and higher DHA:EPA ratio compared to *Artemia* (even after they have been fed with diet rich in DHA) and rotifer *Brachionus*. Meanwhile, several studies (e.g. Millamena *et al.*, 1988; Ozkizilcik & Chu, 1994; Sorgeloos & Leger, 1992) have shown that the content of long-chain PUFA, in particular the n-3 series, is mainly responsible for the nutritional value of live feed for marine animal larvae. In this study, compared to *Artemia* (13.17 % total fatty acid) and the artificial feed (9.19 % total fatty acid) used, *N. affinis* (37.74 % total fatty acid) was found to contain significantly higher amount of long-chain PUFA, particularly of the n-3 series. According to Jones *et al.* (1979), highly unsaturated fatty acids (HUFA), specifically EPA and docosahexaenoic (DHA, 22:6n-

3), play an important role in penaeid larval nutrition and are required in larval diets. In addition, Kreeger *et al.* (1991) and Gonzalez-Felix *et al.* (2002) suggested the importance of EPA and DHA for the survival and growth of many important marine crustacean species. In the present study, it was evident that *N. affinis* provided adequate nutritional requirements for fatty acids, as reflected by the high survival and growth of the shrimp. Diets containing higher level of n-3 HUFA would improve growth and survival of penaeid larvae, since DHA is essential for penaeid larval growth (Gonzalez-Felix *et al.*, 2002).

According to D'Abramo and Sheen (1993), the combined percentage of at least 15% of C-20 PUFAs of the n-3 and n-6 families (C 20:4 n-6; 20:5 n-3; 22:5 n-3; 22:6 n-3) in the whole body tissues of an animal serves as an indicator that the optimal growth requirement for essential fatty acids has been met. In this study, the combined percentage of these fatty acids for the *N. affinis* fed shrimp was >38%, while those fed with artificial diet had approximately 20%.

Merican and Shim (1996) suggested that linolenic (LNA, 18:3n-3) and DHA have been identified as essential fatty acid for *P. monodon*. Similarly, Glencross and Smith (1999) and Glencross *et al.* (2002) noted that linoleic acid (LOA, 18:2n-6) and EPA also have considerable growth-promoting value for shrimp. In fact, they suggested that additional growth would occur when two or more of the essential fatty acids were provided in an optimal combination.

Growth was significantly reduced when either LNA or DHA was not provided in the diet. Moreover, Merican and Shim (1996) also showed that without any HUFA in the feed, the survival of shrimp was reduced to 52%. In this study, shrimp larvae obtained adequate supply of essential fatty acids (Table 2) from *N. affinis*, as illustrated by the significantly higher survival and growth rates compared to those fed with other diets, even with the combined *Artemia* + *N. affinis*.

In the present study, the tanks containing *Artemia* + *N. affinis* (treatment II) showed the highest ($P < 0.05$) concentrations of TAN and $\text{NO}_2\text{-N}$ as compared to other treatments. However, the levels found were within the range suitable for shrimp/prawn hatcheries. Olivar *et al.* (2000) reported that the unconsumed *Artemia* could have contributed to the higher ammonia levels in hatchery tanks. It is interesting to note that the treatment tanks using the artificial feed had significantly ($p < 0.05$) lower levels of TAN and $\text{NO}_2\text{-N}$ compared to those with *Artemia* + *N. affinis*, although one would have expected artificial feed to produce higher levels of the same. Moreover, artificial diet is easily consumed because of the small particles size appropriate for the requirements of the shrimp larvae with little or no excess. However, the levels of ammonia were primarily contributed by the shrimp fed with artificial diet, as reported by Shishehchian *et al.* (1999).

CONCLUSION

The results of this study suggest that *N. affinis* could be used as a live food source

for culturing penaeid shrimp larvae. This is because the shrimp larvae fed with *N. affinis* showed the highest survival percentage as compared to those fed with artificial diet and the combination of *N. affinis* and *Artemia*. In addition, the relative growth rate of shrimp larvae fed with copepod was significantly higher than that of *Artemia* nauplii combined with *N. affinis*. Furthermore, *N. affinis* was found to contain the highest amounts of protein and PUFA as compared to both *Artemia* and artificial diet. Another benefit of using copepods as shrimp larval feed is the maintenance of good water quality of the culture tanks. It is evident from this study that the amount of nitrogenous substances in the water was kept at significantly lower level compared to other treatments although similar methods of maintenance were applied in all the treatment tanks.

ACKNOWLEDGEMENTS

The authors would like to acknowledge the Government of Malaysia through the Intensification of Research in Priority Areas (IRPA) Project No.01-02-04-0392 for supporting this study. Thanks to Mr. Eduardo D. Peralta for the technical assistance.

REFERENCES

- Abelin, P., Tackaert, W., & Sorgeloos, S. (1991). Ensiled *Artemia* biomass: a promising and practical feed for Penaeid shrimp postlarvae. In P. Lavens, P. Sorgeloos, E. Jaspers, & F. Z. Ollevier (Eds). Larvi '91: Fish and Crustacean Larviculture Symposium. *European Aquaculture Society Special Publication No. 15* (p 125–127) Gent, Belgium.
- Anderson, T., & De Silva, S. (2003). Nutrition. In Lucas, J. S., & P. C. Southgate (Eds.), *Aquaculture: Farming aquatic animals and plants* (pp. 146–171). Oxford: Blackwell Publishing Ltd.
- Barros, H. P. & Valenti, W. C. (2003). Ingestion rates of *Artemia* nauplii for different larval stages of *Macrobrachium rosenbergii*. *Aquaculture*, 217, 223–233.
- Biendenbach, J. M., Smith, L. L., Thomsen, T. K., & Lawrence, A. L. (1989). Use of nematode *Penagrellus redivivus* as an *Artemia* replacement in a larval penaeid diet. *Journal of the World Aquaculture Society*, 2, 61-71.
- Calado, R., Pimentel, T., Vitorino, A., Dionísio, G., & Dinis, M. T. (2008). Technical improvements of a rearing system for the culture of decapod crustacean larvae, with emphasis on marine ornamental species. *Aquaculture*, 285, 264–269.
- Chandler, T. G. (1986). High-density culture of meiobenthic harpacticoid copepods within a muddy sediment substrate. *Canadian Journal of Fisheries and Aquatic Sciences*, 43, 53-59.
- Colvin, L. B., & Brand, C. W. (1977). The protein requirement of penaeid shrimp at various life cycle stages in controlled environment systems. *Proceedings of the World Mariculture Society*, 8, 821-840.
- D'Abramo, L. R., & Sheen, S. S. (1993). Polyunsaturated fatty acid nutrition in juvenile freshwater prawn *Macrobrachium rosenbergii*. *Aquaculture*, 115, 63-86.
- Divakaran, S., & Ostrowski, A. C. (1989). Fatty acid analysis of fish eggs without solvent extraction. *Aquaculture*, 80, 371-375.
- Gee, J. M. (1989). An ecological and economic review of meiofauna as food for fish. *Zoological Journal of the Linnean Society*, 96, 243-261.

- Glencross, B. D., & Smith, D. M. (1999). The linoleic and linolenic acids requirements of the prawn, *Penaeus monodon*. *Aquaculture Nutrition*, *5*, 53-64.
- Glencross, B. D., Smith, D. M., Thomas, M. R., & Williams, K. C. (2002). Optimizing the essential fatty acids in the diet for weight gain of the prawn, *Penaeus monodon*. *Aquaculture*, *204*, 85-99.
- Gomez, K. A., & Gomez, A. A. (1983). *Statistical Procedure for Agricultural Research* (pp.680). New York: John Wiley and Sons.
- Gonzalez-Felix, M. L., Gatlin III, D. M., Lawrence, A. L., & Perez-Velasquez, M. (2002). Effect of dietary phospholipids on essential fatty acid requirements and tissue lipid composition of *Litopenaeus vannamei* juveniles. *Aquaculture*, *207*, 151-167.
- Hicks, G. R. F., & Coull, B. C. (1983). The ecology of marine meiobenthic harpacticoid copepods. *Oceanography and Marine Biology Annual Review*, *21*, 67-175.
- Jones, D. A., Amjad, S., & Chitravadivelu, K. (1989). Comparison of artificial feeds used in penaeid shrimp hatcheries. in *Proceedings of the 3rd Egyptian-British Conference on Animal, Fish and Poultry Production*, (15-20). Alexandria, Egypt.
- Jones, D. A., Kamarudin, M. S., & Le Vay, L. (1993). The potential replacement of live feeds in larval culture. *Journal of the World Aquaculture Society*, *24*, 199-210.
- Jones, D. A., Kanazawa, A. & Ono, K. (1979). Studies on the nutritional requirements of the larval stages of *Penaeus japonicus* using micro-encapsulated diets. *Marine Biology*, *54*, 261-267.
- Jory, D. E. (1997). Penaeid shrimp hatcheries: Part III, larval rearing. *Aquaculture Magazine*, *23*, 67-75.
- Kahan, D., Uhlig, G., Schwenzer, D., & Horowitz, L. (1982). A simple method for cultivating harpacticoid copepods and offering them to fish larvae. *Aquaculture*, *26*, 303-310.
- Kovalenko, E. E., D'Abramo, L. R., Ohs, C. L., & Buddington, R. K. (2002). A successful microbound diet for the larval culture of freshwater prawn *Macrobrachium rosenbergii*. *Aquaculture*, *210*, 385-395.
- Kraul, S., Ako, H., Nelson, A., Brittain, K., & Ogasawara, A. (1992). Evaluation of live feeds for larval and post-larval mahimahi, *Coryphaena hippurus*. *Journal of the World Aquaculture Society*, *23*, 299-306.
- Kraul, S., Brittain K., Cantrell R., Nagao, T., Ako, H., Ogasawara A., & Kitagawa, H. (1993). Nutritional factors affecting stress resistance in the larval mahimahi, *Coryphaena hippurus*. *Journal of the World Aquaculture Society*, *24*, 186-193.
- Kreeger, K. E., Kreeger, D. A., Langdon, C. J., & Lowry, R. R. (1991). The nutritional value of *Artemia* and *Tigriopus californicus* (Baker) for two Pacific mysid species, *Metamysidopsis elongata* (Holmes) and *Mysidopsis intii* (Holmquist). *Journal of Experimental Marine Biology and Ecology*, *148*, 147-158.
- Kuban, F. D., Lawrence, A. L., & Winkelfeld, J. S. (1987). Survival, metamorphosis and growth of larvae from four penaeid shrimp species fed six food combinations. *Aquaculture*, *47*, 151-162.
- Kunz, H. (1975). Copepoda Harpacticoidea aus dem litoral des sudlichen Afrika. I teil. *Kieler meeresforsch*, *31*, 179-212.
- Lang, K. (1948). *Monographie der Harpacticiden*. Hakan Ohlssons Boktryckeri, Lund (Sweden).
- Lang, K. (1965) Copepoda Harpacticoidea from the Californian Pacific coast. *Kongliga Svenska Vetenskaps-Akademiens Handlingar*, *10*, 1-566.
- Leger, P., Bengston, D. A, Simpson, K. L., & Sorgeloos, P. (1986). The use and nutritional value of *Artemia* as food source. *Oceanography*

- and *Marine Biology Annual Review*, 24, 521-623.
- Martin, L., Arenal, A., Fajardo, J., Pimentel, E., Hidalgo, L., Pacheco, M., Garcia, C., & Santiesteban, D. (2006). Complete and partial replacement of *Artemia* nauplii by *Moina micrura* during early postlarval culture of white shrimp (*Litopenaeus schmitti*). *Aquaculture Nutrition*, 12, 89–96.
- Matias-Peralta, H., Yusoff, F. M., Shariff M., & Arshad A. (2005). Effects of some environmental parameters on the reproduction and development of a tropical marine harpacticoid copepod *Nitocra affinis californica* Lang. *Marine Pollution Bulletin*, 51, 722-728.
- Matias-Peralta H., Yusoff, F. M., Shariff, M., & Suhaila M. (2011). Small-scale continuous production of a tropical marine copepod, *Nitocra affinis californica* Lang and its potential as live food for aquaculture. *African Journal of Agricultural Research*, 6(6), 1611-1620.
- McEvoy, L. A., Navarro, J. C., Bell, J. G., & Sargent, J. R. (1995). Oxidation of oil emulsions during *Artemia* enrichment process. *Aquaculture*, 134, 110-112.
- Merican, Z. O., & Shim, K. F. (1996). Qualitative requirements of essential fatty acids for juvenile *Penaeus monodon*. *Aquaculture*, 147, 275-291.
- Meyer, E., & Walther, A. (1988). Methods for estimation of protein, lipid, carbohydrates and chitin levels in freshwater invertebrates. *Archivum fur Hydrobiologie*, 113, 161-177.
- Millamena, O. M., Bombeo, N. A. Jumalon, N. A., & Simpson, K. L. (1988). Effects of various diets on the nutritional value of *Artemia* sp. as food for the prawn, *Penaeus monodon*. *Marine Biology*, 98, 217-221.
- Mock, C. R., Fontaine, C. T., & Revera, D. B. (1980). Improvements in rearing larval penaeid shrimp by the Galveston laboratory method. In G. Persoone, P. Sorgeloos, O. Roels, & E. Jaspers (Eds.), *The Brine Shrimp, Artemia*, 3, *Ecology, Culturing, Use in Aquaculture* (p 331-342). Wetteren, Belgium: Universal Press.
- Motoh, H. (1981). Studies on the fisheries biology of the giant tiger prawn, *Penaeus monodon* in the Philippines. *Technical Report No. 7*. Tigbauan, Iloilo: SEAFDEC Aquaculture Department.
- Nanton, D. A., & Castell, J. D. (1998). The effects of dietary fatty acids on the fatty acid composition of the harpacticoid copepod, *Tisbe* sp., for use as live food for marine fish larvae. *Aquaculture*, 163, 251-261.
- Norsker, N. H., & Støttrup J. G. (1994). The importance of dietary HUFAs for fecundity and HUFA content in the harpacticoid, *Tisbe holothuriae* Humes. *Aquaculture*, 125, 155-166.
- Olivar, M. P., Ambrosio, P. P., & Catalan, I. A. (2000). A closed water recirculation system for ecological studies in marine fish larvae: growth and survival of sea bass larvae fed with live prey. *Aquatic Living Resources*, 13, 29-35.
- Ozkizilcik, S., & Chu, F. E. (1994). Evaluation of omega-3 fatty acid enrichment of *Artemia* nauplii as food for striped bass *Morone saxatilis* Walbaum larvae. *Journal of the World Aquaculture Society*, 25, 147-154.
- Parsons, T. R., Maita, Y., & Lalli, C. M. (1984). *A Manual of Chemical and Biological Methods of Seawater Analysis*. New York: Pergamon Press.
- Perera, E., Fraga, I., Carrillo, O., Diaz-Iglesias, E., Cruz, R., Baez, M., & Galich, G. S. (2005). Evaluation of practical previous term diets next term for the Caribbean spiny lobster *Panulirus argus* (Latreille, 1804): Effects of previous term protein next term sources on substrate metabolism and digestive proteases. *Aquaculture*, 244, 251–262.
- Robinson, C. B., Samocho, T. M., Fox, J. M., Gandy, R. L., & McKee, D. A. (2005). The use of

- inert artificial commercial food sources as replacements of traditional live food items in the culture of larval shrimp, *Farfantepenaeus aztecus*. *Aquaculture*, 245, 135-147.
- Samocha, T. M., Matsumoto, T., Jones, E. R., & Torano, M. (1999). Use of artificial diets to reduce *Artemia* nauplii requirements for production of *Litopenaeus vannamei* postlarvae. *The Israeli Journal of Aquaculture – Bamidgeh*, 51(4), 157-168.
- SAS. (1992). *Statistical Analysis System*. SAS Institute Inc. Cary NC, USA. Science and technology, Marcel Dekker, Inc. New York and Basel USA.
- Shields, R. J., Bell, J. G., Luizi, F. S., & Gara, B. (1999). Natural copepods are superior to enriched *Artemia* nauplii as feed for halibut larvae (*Hippoglossus hippoglossus*) in terms of survival, pigmentation and retinal morphology; relation to dietary essential fatty acids. *The Journal of Nutrition*, 129, 1186-1194.
- Shishehchian, F., Yusoff, F. M., Omar, H. & Kamarudin, M. S. (1999). Nitrogenous excretion of *Penaeus monodon* postlarvae fed with different diets. *Marine Pollution Bulletin*, 39, 224-227.
- Sorgeloos, P., & Leger, P. (1992). Improved larviculture outputs of marine fish, shrimp and prawn. *Journal of the World Aquaculture Society*, 23, 251-264.
- Szyper, J. P. (1989). Nutritional depletion of the aquaculture feed organisms *Euterpina acutifrons*, *Artemia sp.* and *Brachionus plicatilis* during starvation. *Journal of the World Aquaculture Society*, 20, 162-169.
- Tseng, W-Y., & Hsu, C-K. (1984). Studies on the culture of the marine copepod *Tigriopus japonicus* Mori and its value as a food for juveniles of the yellow-fin sea bream *Acanthopagrus latus* (Houttuyn). *Crustaceana. International Journal of Crustacean Research Supplement*, 7, 381-389.
- Volk, E. C., Wissmar, R. C., Simenstad, C. A., & Eggers, D. M. (1984). Relationship between otolith microstructure and the growth of juvenile chum salmon (*Oncorhynchus keta*) under different prey rations. *Canadian Journal of Fisheries and Aquacultural Science*, 41, 126-133.
- Watanabe, T., & Kiron, V. (1994). Prospects in larval fish dietetics. *Aquaculture*, 124, 223-251.
- Watanabe, T., Kitajima, C., & Fujita, S. (1983). Nutritional values of live food organisms used in Japan for mass propagation of fish: a review. *Aquaculture*, 34, 115-143.
- Weiss, G. M., McManus, G. B., & Harvey, R. H. (1996). Development and lipid composition of harpacticoid copepod *Nitocra spinipes* reared on different diets. *Marine Ecology Progress Series*, 132, 57-61.
- Wilkenfed, J. S., Fox, J. M., & Lawrence, A. L. (1981). Consumption of frozen and live *Artemia* by protozoa by *Penaeus satiferus*. *Journal of the World Mariculture Society*, 12, 151.
- Wilkenfed, J. S., Lawrence, A. L., & Kuban, F. D. (1984). Survival, metamorphosis and growth of penaeid shrimp larvae reared on a variety of algal and animal foods. *Journal of the World Mariculture Society*, 15, 31-49.
- Wouters, R., & Van Horenbeeck, T. (2003). Larval Shrimp Feeds: Current status. In D.E. Jory (Ed). *Responsible Aquaculture for a Secure Future: Proceedings of a Special Session on Shrimp Farming* (pp. 90-109). The World Aquaculture Society, Baton Rouge, Louisiana 70803 United States.
- Zar, J. H. (1984). *Biostatistical Analysis* (2nd edition) New York: Prentice Hall Inc. Englewood Cliffs.

