



**CHARACTERISTICS, GENETIC DIVERSITY AND PATHOGENIC
VARIABILITY OF *Fusarium oxysporum* f. sp. *niveum* IN WATERMELON
(*Citrullus lanatus* var. *lanatus* (THUNB.) MATSUM. & NAKAI)**

By

MUHAMMAD ZIAUR RAHMAN

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

August 2022

FP 2022 79

COPYRIGHT

All materials contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes by the copyright holder. Commercial use of the material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

The most precious people in my life; my lovely better-half Mst. Fahomida Amin, adorable son Farraj Farihzaad Ziyaam, and daughter Feona Nusaiba Zuha, my beloved parents Late Muhammad Meher Ali, Late Mst. Aeshya Begum, for their everlasting love and prayer that encouraged me to continue it.

To my family members

And

To all my friends who supported me all these year



Abstract of thesis presented to Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Doctor of Philosophy

CHARACTERISTICS, GENETIC DIVERSITY AND PATHOGENIC VARIABILITY OF *Fusarium oxysporum* f. sp. *niveum* IN WATERMELON (*Citrullus lanatus* var. *lanatus* (THUNB.) MATSUM. & NAKAI)

By

MUHAMMAD ZIAUR RAHMAN

August 2022

Chairman : Associate Professor Khairulmazmi bin Ahmad, PhD
Faculty : Agriculture

Watermelon (*Citrullus lanatus* L.) is one of the most economically important horticultural fruit crops. It belongs to the *Cucurbitaceae* family. Presently a major hindrance to this crop production is the soil-borne disease of Fusarium wilt caused by *Fusarium oxysporum* f. sp. *niveum* (Fon). This disease causes yield losses to growers approximately 30-80% or even more. There is limited information on the detection, genetic diversity, and pathogenic variability of the Fon pathogen in Malaysia and Bangladesh. Hence, this study was conducted with the following objectives: to survey and characterize *Fusarium oxysporum* isolates collected from different watermelon growing areas in Peninsular Malaysia and Bangladesh based on morphological and molecular methods; to determine the genetic diversity of *F. oxysporum* f. sp. *niveum* through simple sequence repeat (SSR) markers; and to determine the pathogenic variability and hosts reaction of watermelon commercial cultivars against *F. oxysporum* f. sp. *niveum*. A survey of Fusarium wilt disease of watermelon was done in major watermelon-producing areas in Peninsular Malaysia and in Bangladesh from August, 2019 to December, 2020, using a diagonal sampling method. Based on the cultural and morphological characteristics were used to identify the fungus. The Fon forma speciales-specific primers (Fn-1/Fn-2 and Fon-1/Fon-2) were used to identify the fungus based on the molecular method. The molecular characterization of *Fusarium* species was determined by PCR amplification of different gene regions (*tef1-a*, *IGS*, and *mtSSU*). The analysis of molecular variance (AMOVA) for genetic diversity was done using SSR marker. A pathogenicity test was carried out for 65 Fon isolates to determine their aggressiveness and 12 watermelon commercial cultivars were tested to identify adequate resistance to Fon. The findings of field disease surveys revealed that field disease incidence was 5-45% based on random sampling. The highest disease incidence (45%) was in Pahang, whereas, Terengganu had the lowest disease incidence (5%). The mycelia of the collected isolates were delicate, sparse to numerous, white to pinkish-white, and violet to dark violet pigmentation. Microconidia were small, oval-ellipsoid, straight to curve, non-septate and macroconidia were fusoid-subulate, scarce to abundant, mostly three-septate,

hooked apex and pedicellate base. Whereas, chlamydospores were abundant, formed terminally or intercalary, and found singly or in pairs, chains, or clusters. These typical cultural and morphological characteristics of 65 isolates were identified as *F. oxysporum*. The Fon forma speciales-specific primers (Fn-1/Fn-2 and Fon-1/Fon-2) were amplified amplicons of 320 bp and 174 bp in 100% and 91% of the isolates, respectively. The result of phylogenetic analyses of individual and combined gene sequences of *tef1- α* , *IGS*, and *mtSSU* revealed that 65 isolates belonging to *F. oxysporum* f. sp. *niveum*. Multigene phylogenetic analysis using *tef1- α* , *IGS*, *mtSSU* data set of all isolates were clustered into four main clades, indicating that Fon is polyphyletic. The analysis of molecular variance (AMOVA) showed a total variation of 67% within the population and a 33% total variation among the populations. The results of this investigation indicated that there is a maximum genetic diversity existing within the Fon populations. Based on the pathogenicity test, the 65 Fon isolates were characterized into 4 groups, viz., highly aggressive 15 isolates, moderately aggressive 7 isolates, weakly or low aggressive 29 isolates, and non-pathogenic 14 isolates. Among the 12 watermelon commercial cultivars, Black Giant and Big Family were identified as highly resistant cultivars with disease severity index (DSI) of 5.55-18.15% and 18.11-30.00%, respectively. These results indicated that these two cultivars have the potential to cultivate as resistant varieties against Fon.

Abstraktesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**CIRI-CIRI, KEPELBAGAIAN GENETIK DAN KEBOLEHUBAHAN CIRI
PATOGENIK *Fusarium oxysporum* f. sp. *niveum* PADA TEMBIKAI (*Citrullus
lanatus* var. *lanatus* (THUNB.) MATSUM. & NAKAI)**

Oleh

MUHAMMAD ZIAUR RAHMAN

Ogos 2022

Pengerusi : Profesor Madya Khairulmazmi bin Ahmad, PhD
Fakulti : Pertanian

Tembikai (*Citrullus lanatus* L.) merupakan salah satu tanaman buah hortikultur yang sangat penting dari segi ekonomi. Tembikai tergolong dalam keluarga *Cucurbitaceae*. Pada masa ini, cabaran utama pengeluaran tembikai adalah penyakit bawaan tanah iaitu layu *Fusarium* yang disebabkan oleh *Fusarium oxysporum* f. sp. *niveum* (Fon). Penyakit ini menyebabkan kehilangan hasil kepada penanam sekitar 30-80% atau lebih. Setakat ini, maklumat mengenai pengesanan dan kepelbagaian genetik dan kebolehubahan ciri patogenik patogen Fon di Malaysia dan Bangladesh adalah terhad. Oleh itu, kajian ini dijalankan berdasarkan objektif berikut: untuk meninjau dan mencirikan pencilan *Fusarium oxysporum* yang dikumpul dari kawasan penanaman tembikai yang berbeza di Semenanjung Malaysia dan Bangladesh berdasarkan kaedah morfologi dan molekul; untuk menentukan kepelbagaian genetik *F. oxysporum* f. sp. *niveum* melalui penanda jujukan ringkas (SSR); dan untuk menentukan kebolehubahan kepatogenan dan tindak balas perumah kultivar komersial tembikai terhadap *F. oxysporum* f. sp. *niveum*. Tinjauan penyakit layu *Fusarium* pada tembikai telah dilakukan di kawasan penghasil tembikai utama di Semenanjung Malaysia dan di Bangladesh dari Ogos, 2019 hingga Disember, 2020, menggunakan kaedah pensampelan pepenjuru. Berdasarkan ciri kultura dan morfologi digunakan untuk mengenalpasti kulat. Primer khusus Fon forma (Fn-1/Fn-2 dan Fon-1/Fon-2) digunakan untuk mengenalpasti kulat berdasarkan kaedah molekul. Pencirian molekular spesies *Fusarium* ditentukan oleh amplifikasi PCR kawasan gen yang berbeza (*tef1-a*, *IGS*, dan *mtSSU*). Analisis varians molekul (AMOVA) untuk kepelbagaian genetik dilakukan menggunakan penanda SSR. Ujian patogenik telah dijalankan untuk 65 pencilan Fon untuk menentukan keagresifannya dan 12 kultivar komersial tembikai telah diuji untuk mengenal pasti rintangan yang mencukupi terhadap Fon. Dapatan tinjauan penyakit di lapangan mendedahkan bahawa kejadian penyakit di lapangan adalah 5-45% berdasarkan persampelan rawak. Kejadian penyakit tertinggi (45%) adalah di Pahang, manakala, Terengganu mempunyai kejadian penyakit terendah (5%). Miselia bagi pencilan yang dikumpul adalah halus, jarang kepada banyak, putih hingga putih merah jambu, dan pigmentasi ungu hingga ungu gelap. Mikrokonidia adalah kecil, bujur-

elipsoid, lurus ke lengkung, tanpa septat dan makrokonidia adalah fusoid-subulate, jarang kepada banyak, kebanyakannya tiga septa, puncak bercangkuk dan tapak pediselata. Manakala, klamidospora banyak, terbentuk secara terminal atau interkalari, dan ditemui secara tunggal atau berpasangan, rantai, atau kelompok. Ciri-ciri tipikal kultura dan morfologi bagi 65 isolat ini dikenalpasti sebagai *F. oxysporum*. Primer khusus Fon forma (Fn-1/Fn-2 dan Fon-1/Fon-2) masing-masing menghasilkan ampikon yang bersaiz 320 bp dan 174 bp dengan 100% dan 91% kesamaan bagi isolat masing-masing. Hasil analisis filogenetik bagi jujukan gen individu dan gabungan *tefl- α* , *IGS*, dan *mtSSU* mendedahkan bahawa 65 isolat kepunyaan *F. oxysporum* f. sp. *niveum*. Analisis filogenetik pelbagai gen menggunakan set data *tefl- α* , *IGS*, *mtSSU* bagi semua pencilan dikelompokkan kepada empat klad utama, menunjukkan bahawa Fon adalah polifiletik. Analisis varians molekul (AMOVA) menunjukkan jumlah variasi 67% dalam populasi dan 33% jumlah variasi di kalangan populasi. Keputusan penyiasatan ini menunjukkan bahawa terdapat kepelbagaian genetik maksimum yang wujud dalam populasi Fon. Berdasarkan ujian patogenik, 65 Fon pencilan telah dicirikan kepada 4 kumpulan, iaitu, sangat agresif 15 pencilan, sederhana agresif 7 pencilan, lemah atau rendah agresif 29 pencilan, dan bukan patogenik 14 pencilan. Antara 12 kultivar komersil tembikai, “Black Giant” dan “Big Family” dikenal pasti sebagai kultivar sangat tahan dengan indeks keterukan penyakit (DSI) masing-masing 5.55-18.15% dan 18.11-30.00%. Keputusan ini menunjukkan bahawa kedua-dua kultivar tersebut berpotensi untuk ditanam sebagai varieti tembikai tahan terhadap jangkitan Fon.

ACKNOWLEDGEMENTS

Firstly, I would like to express my greatest gratefulness and gratitude to the Almighty Allah for giving me the strength, health, patience, and wisdom to complete this study successfully. I would like to take this opportunity to thank my supervisor, Assoc. Prof. Dr. Khairulmazmi Ahmad for his keen supervision, advice, guidance, support, and fruitful discussion. I would like to acknowledge his understanding and working group for the working atmosphere. I would like to further extend my supervisory committee members, Dr. Norsazilawati Saad, Assoc. Prof. Dr. Tan Geok Hun, and Dr. Erneeza Mohd Hata for giving astute suggestions, and valuable advice for rendering all possible guidance in carrying out the research work.

I wish to express my deepest thanks to the Deputy Director, Department of Agriculture, Bandar Indera Mahkota, Kuantan, Pahang; Agriculture officer, Mersing District Agriculture Office, Jalan Ibrahim, Mersing Kechil, Mersing, Johor, Mr. Aiman Takrim and Upazila agriculture officer, Cox'sbazar Sadar, Cox'sbazar, Bangladesh for their assistance during sample collection. I express my gratitude to the Late Dr. Abdul Jalil Bhuyan (Ex-Director General of BRR), Dr. Hafizulla (Ex-Director of BARI), and Dr. Abdul Wohab (Ex-Director General of BARI) for their immense help and cooperation during the pursuit of my Ph.D scholarship.

I would like to extend appreciation to all faculty members and staff for their guidance, for permitting using the equipment, and for providing research facilities to conduct my research at the Department of Plant Protection, Faculty of Agriculture, Universiti Putra Malaysia. Moreover, my sincere appreciation also goes to Dr. Shafiqur Rahman, Krishibid Abdur Razzaque, and Mr. Jewel for being helpful in my lab work at Marine Fisheries Technology Station, Cox'sbazar, Bangladesh.

I would like to special dedication to my beloved wife, Mst. Fahomida Amin, son, Farraj Farihzaad Ziyaam, daughter, Feona Nusaiba Zuha, and father, Late Muhammad Meher Ali for their love, moral support, encouragement, and patience throughout the research. I am deeply grateful to my late mother, Mst. Aeshya Begum for her love and affection. I am also thankful to all my friends, especially Md Mahmudul Hasan Khan as well as all of my colleagues for keeping in contact, and also for their motivation, support, and cooperation to strive to reach the goal.

I am grateful to my workplace Regional Agricultural Research Station, Rahmatpur, Barishal-8211, my organization Bangladesh Agricultural Research Institute (BARI), Gazipur-1701, and the Ministry of Agriculture, Government of the People's Republic of Bangladesh for providing deputation to commence my study period September 2018 to August 2022. Lastly, I would like to express my deepest gratitude and thankfulness to the National Agricultural Technology Program-Phase II Project, Bangladesh Agricultural Research Council (BARC), Dhaka, and the Ministry of Agriculture, Government of the People's Republic of Bangladesh for awarding me the fellowship and funding to conduct this research.

I certify that a thesis examination committee has met on 12 August 2022 to conduct the final examination of name Muhammad Ziaur Rahman on his Doctor of Philosophy thesis entitled “Characterization, Genetic Diversity and Pathogenic Variability of *Fusarium oxysporum* f. sp. *niveum* in Watermelon (*Citrullus lanatus* var. *Lanatus* (Thunb.) Matsum. & Naki)” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P. U. (A) 106] 15 March 1998. The committee recommends that the candidate be awarded the Doctor of Philosophy.

Members of the examination committee are as follows:

Lau Wei Hong, PhD
Associate Professor
Department of Plant Protection
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Mohd Rafii Yusop, PhD
Professor
Institute of Tropical Agriculture and Food Security
Universiti Putra Malaysia
(Internal Examiner)

Ganesen Vadamalai, PhD
Associate Professor
Department of Plant Protection
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Mujeebur Rahman Khan, PhD
Department of Plant Protection
Aligarh Muslim University
Aligarh, India
(External Examiner)

SITI SALWA ABD GANI, PhD

Associate Professor ChM. and Deputy Dean
School of Graduate Studies Universiti Putra
Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirements for the degree of Doctor of Philosophy. The members of Supervisory Committee were as follows:

Khairulmazmi bin Ahmad, PhD

Associate Professor
Department of Plant Protection
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Tan Geok Hun, PhD

Associate Professor
Department of Land Management
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Norsazilawati binti Saad, PhD

Senior Lecturer
Department of Plant Protection
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Erneeza binti Mohd Hata, PhD

Research Officer
Sustainable Agronomy and Crop Protection
Institute of Plantation Studies (IKP)
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 8 December 2022

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No: Muhammad Ziaur Rahman

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012- 2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Associate Professor Dr. Khairulmazmi bin Ahmad

Signature: _____
Name of Member
of Supervisory
Committee: Associate Professor Dr. Tan Geok Hun

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Norsazilawati binti Saad

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Erneeza binti Mohd Hata

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xviii
CHAPTER	
1	
INTRODUCTION	1
1.1 Background of study	1
1.2 Problem statements	2
1.3 Objectives of the studies	2
2	
LITERATURE REVIEW	3
2.1 Watermelon	3
2.1.1 Taxonomy, Origin & Distribution	3
2.1.2 Watermelon type	4
2.1.3 Present status of Watermelon production	4
2.1.4 Nutritional Importance and Health benefits	6
2.1.5 Watermelon industry in the world	8
2.2 Diseases of Watermelon	9
2.2.1 Fusarium wilt disease of watermelon	10
2.2.2 Symptoms of Fusarium wilt of watermelon	10
2.2.3 Dissemination, Mode of infection, and favorable condition	11
2.2.4 Pathogenic variability of <i>F. oxysporum</i> f. sp. <i>niveum</i> (Fon)	11
2.2.5 Economic importance of <i>Fusarium oxysporum</i> f. sp. <i>niveum</i> (Fon)	12
2.2.6 Disease cycle and epidemiology	12
2.3 Characterization of <i>F. oxysporum</i> f. sp. <i>niveum</i>	14
2.3.1 Morphological characterization	14
2.3.2 Molecular identification	15
2.3.3 Pathogenic characterization	17
2.4 Genetic diversity of <i>F. oxysporum</i> f. sp. <i>niveum</i> (Fon)	18
2.4.1 Molecular method: simple sequence repeats (SSR)	17
3	
ISOLATION, MORPHOLOGICAL AND MOLECULAR CHARACTERIZATION OF <i>Fusarium oxysporum</i> f. sp. <i>niveum</i> CAUSAL AGENT OF FUSARIUM WILT DISEASE OF WATERMELON IN PENINSULAR MALAYSIA AND BANGLADESH	20
3.1 Introduction	20

3.2	Materials and methods	21
3.2.1	Disease survey and sampling	21
3.2.2	Isolation and purification of <i>F. oxysporum</i> isolates	23
3.2.3	Morphological Characterization	23
3.2.4	Molecular characterization	24
3.2.4.1	Culture preparation and extraction of Genomic DNA	24
3.2.4.2	Quantification and qualitative analysis of DNA	24
3.2.4.3	PCR amplification of Fon-specific primers	25
3.2.4.4	PCR amplification of translation elongation factor (<i>tef1-α</i>) region	26
3.2.4.5	PCR amplification of mitochondrial small subunit (<i>mtSSU</i>) region	26
3.2.4.6	PCR amplification of intergenic spacer (<i>IGS</i>) region	26
3.2.4.7	Agarose gel electrophoresis	27
3.2.4.8	Gene sequencing and alignment	27
3.2.4.9	Phylogenetic analyses	28
3.2.5	Statistical analysis	28
3.3	Results	28
3.3.1	Disease survey and sampling	28
3.3.2	Isolation and morphological characterization	29
3.3.3	Molecular characterization	39
3.3.3.1	Fon-specific PCR assay	39
3.3.3.2	PCR amplification of translation elongation factor (<i>tef1-α</i>) region	41
3.3.3.3	PCR amplification of mitochondrial small subunit (<i>mtSSU</i>) region	44
3.3.3.4	PCR amplification of intergenic spacer (<i>IGS</i>) region	47
3.3.4	Phylogenetic analysis of PCR-generated DNA sequences	49
3.3.4.1	Phylogenetic analysis of the translation elongation factor (<i>tef1-α</i>) gene region	49
3.3.4.2	Phylogenetic analysis of the mitochondrial small subunit (<i>mtSSU</i>) gene region	51
3.3.4.3	Phylogenetic analysis of the intergenic spacer (<i>IGS</i>) gene region	53
3.3.4.4	Phylogenetic analysis for a combined dataset of the <i>tef1-α</i> , <i>mtSSU</i> , and <i>IGS</i>	55
3.4	Discussion	60
3.5	Conclusion	63
4	GENETIC DIVERSITY OF <i>Fusarium oxysporum</i> f. sp. <i>niveum</i> THROUGH SIMPLE SEQUENCE REPEAT (SSR) MOLECULAR MARKERS	64
4.1	Introduction	64

4.2	Materials and Methods	65
4.2.1	Collection of <i>F. oxysporum</i> f. sp. <i>niveum</i> (Fon) isolates	65
4.2.2	DNA extraction	65
4.2.3	PCR amplification for SSR marker	65
4.2.4	Agarose gel electrophoresis and staining	66
4.2.5	SSR Analysis and dendrogram Construction	66
4.3	Results	68
4.3.1	SSR analysis	68
4.3.2	Clustering using SSR markers	69
4.3.3	Genetic diversity within the populations	73
4.3.4	Analysis of molecular variance (AMOVA)	73
4.3.5	Principal component analysis (PCA)	74
4.4	Discussion	76
4.5	Conclusion	79
5	PATHOGENIC VARIABILITY AND REACTION OF WATERMELON COMMERCIAL CULTIVARS AGAINST <i>Fusarium oxysporum</i> f. sp. <i>niveum</i>	80
5.1	Introduction	80
5.2	Materials and Methods	81
5.2.1	Pathogenicity tests	81
5.2.2	Pathogenicity gene identification	84
5.2.2.1	DNA extraction	84
5.2.2.2	PCR amplification of FonSIX homologos	84
5.2.2.3	Agarose gel electrophoresis	84
5.2.2.4	Gene sequencing and alignment	84
5.2.2.5	Phylogenetic analyses of <i>FonSIX</i> homologos	85
5.2.4	Cultivars evaluation	85
5.2.4.1	Host materials	85
5.2.4.2	Fungal isolates	86
5.2.4.3	Location	86
5.2.4.4	Inoculum preparation	87
5.2.4.5	Root dipping inoculation technique	87
5.2.4.6	Disease assessment	87
5.2.5	Statistical analysis	90
5.3	Results	90
5.3.1	Pathogenicity tests	90
5.3.2	Pathogenicity gene identification	96
5.3.2.1	PCR amplification of FonSIX homologos	96
5.3.2.2	Phylogenetic analysis of FonSIX homologos	97
5.3.3	Cultivars evaluation	98
5.3.3.1	The pathogenicity reaction of watermelon cultivars against selected Fon isolates	98
5.3.3.2	The relationship between host disease severity and plant fresh weight	100

5.3.3.3	Host reaction groups	105
5.4	Discussion	107
5.5	Conclusion	110
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	111
6.1	Summary and conclusion	111
6.2	Recommendations for future research	113
	REFERENCES	114
	APPENDICES	139
	BIODATA OF STUDENT	218
	LIST OF PUBLICATIONS	219



LIST OF TABLES

Table	Page
2.1 Nutritional composition of the edible portion of watermelon	7
3.1 Information of Fusarium wilt symptomatic plant samples in Peninsular Malaysia and Bangladesh	22
3.2 List of primer pairs amplified in this study	25
3.3 Isolates, sampling locations, and their year of collection used in this study	30
3.4 Cultural and morphological characteristics of different isolates of <i>Fusarium oxysporum</i>	35
3.5 BLASTn results of 65 isolates for the <i>tefl-a</i> region related to <i>F. oxysporum</i> f. sp. <i>niveum</i>	42
3.6 BLASTn results of 65 isolates for the <i>mtSSU</i> region related to <i>F. oxysporum</i> f. sp. <i>niveum</i>	45
3.7 BLASTn results of 65 isolates for the <i>IGS</i> region related to <i>F. oxysporum</i> f. sp. <i>niveum</i>	48
3.8 Isolates of <i>F. oxysporum</i> f. sp. <i>niveum</i> from different countries were used in this investigation, as well as reference gene sequences from the GenBank database	57
4.1 List of SSR markers, sequences, SSR motifs, and allele sizes	66
4.2 Various features of genetic diversity among the 65 Fon isolates depend on 9 SSR markers	69
4.3 Assessment of genetic diversity among the isolates of Fon	73
4.4 The 65 isolates of Fon were subjected to molecular variance analysis (AMOVA).	74
4.5 Based on SSR markers pairwise analyses of genetic identity and genetic distance among the five Fon populations	74
5.1 Watermelon cultivars bought to use in the experiments	86
5.2 Rating scale used for disease severity assessment	87
5.3 Watermelon seedlings were assessed based on the disease severity category	88
5.4 Assessment of virulence based on pathogenicity of <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates under glasshouse trial	92
5.5 Assessment of variation for disease severity on different watermelon	99

cultivars under glasshouse conditions

- 5.6 Assessment of variation for the area under disease progress curve (AUDPC) on different watermelon cultivars under glasshouse conditions 100
- 5.7 Analysis of variance results for disease index values on the preliminary and final virulence tests 105
- 5.8 Resistance reaction of 12 watermelon cultivars against the 10 *F. oxysporum* f. sp. *niveum* pathogenic isolates 106
- 5.9 Results for screening of 12 watermelon cultivars against 10 pathogenic *F. oxysporum* f. sp. *niveum* isolates 106



LIST OF FIGURES

Figure	Page	
2.1	Percentage production of watermelon in some countries	5
2.2	Watermelon production in Malaysia 2008-2019	5
2.3	Watermelon production in Bangladesh 2011-2021	6
2.4	Distribution Maps of Fusarium wilt disease of watermelon (presented in orange dot) globally	10
2.5	Disease life cycle of Fusarium wilt (<i>F. oxysporum</i> f. sp. <i>niveum</i>) of watermelon	13
3.1	Distribution of sampling location in Peninsular Malaysia and Bangladesh	22
3.2	Typical Fusarium wilting symptoms on watermelon plants in the growing fields	29
3.3	Colony features of <i>F. oxysporum</i> on PDA	32
3.4	Conidia features of <i>F. oxysporum</i> on PDA	33
3.5	Chlamyospore features of <i>F. oxysporum</i> on PDA	34
3.6	PCR amplification by using Fon-specific primers Fn-1/Fn-2 and Fon-1/Fon-2	40
3.7	PCR amplification of 65 <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates using EF1 and EF2 primers, each band had approximately 700 bp amplicon; M is 1kb DNA Ladder (GeneDireX) and Control (C)	40
3.8	PCR analysis of 65 <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates using NMS1 and NMS2 primers, each band had approximately 660 bp amplicon; M is 1kb DNA Ladder (GeneDireX) and Control (C)	44
3.9	PCR analysis of 65 <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates using FIGS11 and FIGS12 primers, each band had approximately 650 bp amplicon; M is 1kb DNA Ladder (GeneDireX)	47
3.10	Phylogenetic relationships of the <i>tef1</i> -a sequence of <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates by Maximum Likelihood method with 1000 bootstrap replications	50
3.11	Phylogenetic relationships of the <i>mtSSU</i> sequence of <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates by Maximum Likelihood method based on 1000 bootstrap replications	52
3.12	Phylogenetic relationships of <i>IGS</i> sequence of <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates by Maximum Likelihood method based on 1000 bootstrap replications	54

3.13	Phylogenetic relationships of combined sequences of <i>tef1-a</i> , <i>mtSSU</i> , and <i>IGS</i> region <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates by Maximum Likelihood method based on 1000 bootstrap replications	56
4.1	Bands pattern of <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates using Fo314 SSR molecular markers, each band had approximately 300 bp amplicon; M is 100 bp DNA Ladder (GeneDireX)	68
4.2	The genetic relationships among the 65 <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates shown in the dendrogram derived from UPGMA Cluster Analysis	71
4.3	Two-dimensional principal coordinate analysis (PCoA) of 65 isolates of <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates from four states of Malaysia and one region of Bangladesh exhibited the pattern of sub-clustering	72
4.4	Principal Component Analysis (PCA) was used to analyse the association among all the isolates in a two-dimensional graph	75
5.1	Various Fusarium wilt disease symptoms	83
5.2	<i>F. oxysporum</i> f. sp. <i>niveum</i> infected plant splitted longitudinally to look for signs of vascular discolouration	89
5.3	Distribution of the <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates virulence across a) the four major watermelon growing regions in Peninsular Malaysia and b) one region in Bangladesh	94
5.4	Relationship of disease severity of Fusarium wilt and fresh weight (g) of inoculated watermelon plant at glasshouse condition	95
5.5	Bands pattern of <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates using FonSIX6F and FonSIX6R molecular markers, each band had approximately 500 bp amplicon; M is 100 bp DNA Ladder (GeneDireX)	96
5.6	Phylogenetic relationships of <i>SIX</i> gene sequences of <i>F. oxysporum</i> f. sp. <i>niveum</i> isolates by Maximum Likelihood method based on 1000 bootstrap replications	97
5.7	Intra-specific variation of pathogenicity-related gene sequences FonSIX6 for the highly virulent, moderately virulent, and low virulent isolates of <i>F. oxysporum</i> f. sp. <i>niveum</i>	98
5.8	Expression of the linear relationship for disease severity and plant fresh weight on different cultivars	102

LIST OF ABBREVIATIONS

AFLP	Amplified Fragment Length Polymorphism
ANOVA	Analysis of Variance
AMOVA	Analysis of Molecular Variance
a.i.	Active Ingredient
AUDPC	The area under the disease progression curve
BLASTn	Nucleotide Basic Local Alignment Search Tool
Bp	Base pair
β -tubulin	Beta tubulin
CABI	Centre for Agriculture and Bioscience International
CRD	Completely Randomized Design
CTAB	Cetyltrimethylammonium Bromide
Cm	Centimetre
DAI	Days after inoculation
Df	Degree of freedom
DI	Disease Incidence
DNA	Deoxyribonucleic acid
dH ₂ O	Distilled water
ddH ₂ O	Double distilled water
DSI	Disease Severity Index
EDTA	Ethylenediaminetetraacetic acid
Fon	<i>Fusarium oxysporum</i> f. sp. <i>niveum</i>
GDP	Gross Domestic Product
G	Gram
GS	Genetic similarity
H	Nei's Gene Diversity

Ho	Expected Homozygosity
He	Expected Heterozygosity
Ha	Hectares
I	Shannon's Information Index
ITS	Internal Transcribed Spacer
IGS	Intergenic spacer
kb	Kilobase pair
LSD	Least significant difference
Mg	Milligram
Min	Minute
ml	Millilitre
mm	Millimetre
MS	Means of Sum Squares
<i>mtSSU</i>	Mitochondrial small subunit
Na	Observed Number of Alleles
NaOCl	Sodium hypochlorite
NCBI	National Centre for Biotechnology Institute
Ne	The effective number of alleles
NTSYS	Numerical Taxonomy and Multivariate Analysis System
PCA	Principal component analysis
PCoA	Principal Coordinate Analysis
PCR	Polymerase Chain Reaction
PDA	Potato dextrose agar
PIC	Polymorphic Information Content
POPGENE	Population Genetic Analysis
RAPD	Random Amplified Polymorphic DNA
RFLP	Restriction Fragment Length Polymorphism

R	Correlation Coefficient
rDNA	Ribosomal DNA
Rpm	Revolutions per minute
SAS	Statistical Analysis Software
SDS	Sodium Dodecyl Sulfate
SDW	Sterile distilled water
SIX	Secreted in Xylem Gene
SS	Sum of Squares
SSR	Simple Sequence Repeats
Secs	Second
TBE	Tris-borate EDTA
Tukey's HSD	Tukey's Honestly Significant Difference
tef-1 α	Translation Elongation Factor -1 Alpha
uHe	Unbiased Expected Heterozygosity
Mg	Microgram
μ l	Microliter
Mm	Micrometre
UV	Ultraviolet
uHe	Unbiased Expected Heterozygosity
V	Voltage
VCG	Vegetative Compatibility Group
UPM	Universiti Putra Malaysia
UPMGA	Unweighted Pair Group Methods with Arithmetic Mean
%	Percentage
% P	Percentage of polymorphic loci
$^{\circ}$ C	Degree Celsius

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Watermelon (*Citrullus lanatus* L.) is one of the most economically important horticultural fruit crops belonging to the *Cucurbitaceae* family. It is widely cultivated for its sugary, fleshy edible fruit (Lv et al., 2018; Wehner, 2008; Xie et al., 2019). The advantages of watermelon cultivation over other fruit crops are its short duration, ease of handling, quick financial returns, and low production costs (Costa et al., 2018). Various factors limit watermelon production, but diseases are critical. However, the wilt disease caused by *F. oxysporum* f. sp. *niveum* (Fon) is the most serious disease that affects watermelon productivity (Egel and Martyn, 2013; Zitter et al., 1996). Fon causes infection only in watermelon. The pathogen (Fon) can rapidly disperse in watermelon cultivating areas and is liable for about 30-80% yield losses (Egel and Martyn, 2013; Lü et al., 2011).

Rapid and accurate identification of pathogenic micro-organisms is essential for successful disease management. Traditionally, *Fusarium* species are identified based on their morphological characteristics, which are usually influenced by environmental and geographical factors (Das et al., 2012). However, these methods are mostly documented to be unreliable and unstable (Bosland & Williams, 1987; Watanabe et al., 2011). On the other hand, molecular methods are progressively used to detect and identify pathogens due to the assurance of better results (accuracy), and phylogenetic analyses (McCartney et al., 2003; Saikia & Kadoo, 2010).

PCR amplification based on the nucleotide sequences which is important in identifying species and distinguishing among the formae speciales of *F. oxysporum*. The important genes are translation elongation factor 1- α (*tef1- α*), calmodulin (*cmdA*), actin, intergenic spacer (*IGS*), the mitochondrial small subunit (*mtSSU*), RNA polymerase II subunits 1 and 2 (RPB1 and RPB2) and β -*tubulin* (TUB) region (Lombard et al., 2019; O'Donnell et al., 2010). These genes provide phylogenetic data that are somewhat similar, and only one of them is usually recommended for giving adequate genetic resolution (O'Donnell et al., 2015; Ramdial et al., 2017).

Knowledge of the genetic diversity within the *F. oxysporum* formae speciales is crucial and is investigated as a primary vestige for the sketch of disease management strategies (McDonald et al., 2002). To evaluate genetic variation within a population of *F. oxysporum* forma speciales, various techniques have already been developed such as vegetative compatibility grouping (VCGs), random amplified polymorphism DNA (RAPD) analysis, amplified fragment length polymorphisms (AFLPs), restriction fragment length polymorphism (RFLP), single-nucleotide polymorphism (SNP), inter-

simple sequence repeats (ISSR), and simple sequence repeat (SSR) (Aguilar-Hawod et al., 2019; Merzoug & Belabid, 2018; Petkar et al., 2019; Zang et al., 2018).

Of these methods, SSRs marker can help understand significant function and evolution based on their abundance and density. As a result, SSR techniques have been extensively used for estimating genetic mapping and diversity analysis (Mahfooz et al., 2015).

1.2 Problem Statements

Fon is a predominant, highly diversified fungus comprising pathogenic and non-pathogenic individuals that affect watermelon yield and quality (Das et al., 2012; LeBlanc et al., 2017; Xiong & Zhan, 2018). In Malaysia, almost 13.0% of the total area of fruit production which is equivalent to 8308 ha and produced 144213 Mt watermelon. The disease incidence of *Fusarium* wilt in major production areas namely Johor, Kedah, Kelantan, Pahang and Terengganu were 10-45% and export decrease around 2.1%. In Bangladesh watermelon is grown on 16,542 ha and about 345955 Mt are produced annually. *Fusarium* wilt disease incidence in commercial fields of main watermelon growing regions in Barishal, Chattagram, and Rajshahi were 30-70% and yield loss as a result of disease is around 23%. Thus, concerning effective pathogen management, current status of the disease and identification up to the species level is essential. In addition, there are many reports of characterization, genetic diversity and pathogenic variability in other species of *Fusarium*. Still, less information is available on the relative importance of Fon, their distribution, diversity and pathogenic variability in Peninsular Malaysia and Bangladesh. For this reason, the present study was undertaken; to evaluate the characterization, genetic diversity, and pathogenic variability of Fon isolates in watermelon growing areas in Peninsular Malaysia and Bangladesh for effective management strategies.

1.3 The objectives of the studies were as follows:

- 1) To survey and characterize *Fusarium oxysporum* isolates collected from different watermelon growing areas in Peninsular Malaysia and Bangladesh based on morphological and molecular methods.
- 2) To determine the genetic diversity of *Fusarium oxysporum* f. sp. *niveum* through Simple Sequence Repeat (SSR) markers.
- 3) To determine the pathogenic variability and host reaction of watermelon commercial cultivars against *Fusarium oxysporum* f. sp. *niveum*.

REFERENCES

- Anonymous. 2021. Yearbook of Agricultural Statistics 2021, Bangladesh Bureau of Statistics.
- Achigan-Dako, E. G., Avohou, E. S., Linsoussi, C., Ahanchede, A., Vodouhe, R. S., & Blattner, F. R. (2015). Phenetic characterization of *Citrullus* spp.(*Cucurbitaceae*) and differentiation of egusi-type (*C. mucosospermus*). *Genetic Resources and Crop Evolution*, 62(8), 1159–1179.
- Adejumo, B. A., Olorunsogo, S. T., & Njoku, C. B. (2015). Qualities of water melon seed oil extracted at different moisture content. *International Journal of Advanced Engineering Research and Technology (IJAERT)*, 3(3), 2348–8190.
- Aguilar-Hawod, K. G. I., de la Cueva, F. M., & Cumagun, C. J. R. (2019). Genetic Diversity of *Fusarium oxysporum* f. sp. *ubense* Causing Panama Wilt of Banana in the Philippines. *Pathogens*, 9(1), 32.
- Ahou, A. G., Kouam eacute, K. K., Nandy, D. F. B., Serge, T. D. B., Hippolyte, H. T., Jean Pierre, B., & Iri eacute, A. Z. B. (2016). Morphological diversity in oleaginous watermelon (*Citrullus mucosospermus*) from the Nangui Abrogoua University germplasm collection. *African Journal of Biotechnology*, 15(21), 917–929.
- ouvette, C., Steinberg, C., & Olivain, C. (2013). The Endophytic Strain *Fusarium oxysporum* Fo47: A Good Candidate for Priming the Defense Responses in Tomato Roots. *Molecular P*
- Aimé, S., *Alablant-Microbe Interactions*®, 26(8), 918–926.
- Alabouvette, C., Olivain, C., Migheli, Q., & Steinberg, C. (2009). Microbiological control of soil-borne phytopathogenic fungi with special emphasis on wilt-inducing *Fusarium oxysporum*. *New Phytologist*, 184(3), 529–544.
- Alam, K. M., Alam, M. M., Islam, M. M., Momotaz, R., Arifunnahar, M., Sultana, N. A., Raihan, H.Z., Mujahidi, T.A., Khatun, F. & Banu, S. P. (2020). First Report on *Fusarium oxysporum* f. sp. *niveum* Causing Watermelon Fusarium Wilt in Bangladesh. *Plant disease*, 104(6), 1859.
- Alexopoulos, C., Mims, C. W., & Blackwell, M. (1996). *th Introductory Mycology*. New York: John Wiley & Sons.
- Álvarez-Hernández, J. C., Castellanos-Ramos, J. Z., Aguirre-Mancilla, C. L., Huitrón-Ramírez, M. V., & Camacho-Ferre, F. (2015). Influence of rootstocks on *Fusarium* wilt, nematode infestation, yield and fruit quality in watermelon production. *Ciência e Agrotecnologia*, 39(4), 323–330.
- Amaradasa, B. S., Beckham, K., Dufault, N., Sanchez, T., Ertek, T. S., Iriarte, F., Paret, M., & Ji, P. (2018). First Report of *Fusarium oxysporum* f. sp. *niveum* Race 3 Causing Wilt of Watermelon in Florida, U.S.A. *Plant Disease*, 102(5), 1029–1029.

- Andeden, E. E., Baloch, F. S., Çakır, E., Toklu, F., & Özkan, H. (2015). Development, characterization and mapping of microsatellite markers for lentil (*Lens culinaris* Medik.). *Plant Breeding*, 134(5), 589–598.
- Anderson, J. A., Churchill, G. A., Autrique, J. E., Tanksley, S. D., & Sorrells, M. E. (1993). Optimizing parental selection for genetic linkage maps. *Genome*, 36(1), 181–186.
- Anderson, J. B., & Kohn, L. M. (1995). Clonality in Soilborne, Plant-Pathogenic Fungi. *Annual Review of Phytopathology*, 33(1), 369–391.
- Anne, C. (2006). Choosing the right molecular genetic markers for studying biodiversity: from molecular evolution to practical aspects. *Genetica*, 127(1), 101–120.
- Aoki, T., & O'Donnell, K. (1999). Morphological and molecular characterization of *Fusarium pseudograminearum* sp. nov., formerly recognized as the Group 1 population of *F. graminearum*. *Mycologia*, 91(4), 597–609.
- Appel, D. J., & Gordon, T. R. (1996). Relationships among pathogenic and nonpathogenic isolates of *Fusarium oxysporum* based on the partial sequence of the intergenic spacer region of the ribosomal DNA. *MPMI-Molecular Plant Microbe Interactions*, 9(2), 125–138.
- Armengol, J., José, C. M., Moya, M. J., Sales, R., Vicent, A., & García-Jiménez, J. (2000). *Fusarium solani* f. sp. *cucurbitae* race 1, a potential pathogen of grafted watermelon production in Spain. *EPPO Bulletin*, 30(2), 179–183.
- Baayen, R. P., O'Donnell, K., Bonants, P. J. M., Cigelnik, E., Kroon, L. P. N. M., Roebroeck, E. J. A., & Waalwijk, C. (2000). Gene genealogies and AFLP analyses in the *Fusarium oxysporum* complex identify monophyletic and nonmonophyletic formae speciales causing wilt and rot disease. *Phytopathology*, 90(8), 891–900.
- Bahar, M., & Shahab, H. (2012). Analysis of Iranian isolates of *Fusarium solani* using morphological, pathogenicity and microsatellite DNA marker characterization. *African Journal of Biotechnology*, 11(2), 474–482.
- Bahari, M., Rafii, M. Y., Saleh, G. B., & Latif, M. A. (2012). Combining Ability Analysis in Complete Diallel Cross of Watermelon (*Citrullus lanatus* (Thunb.) Matsum. & Nakai). *The Scientific World Journal*, 2012.
- Baloch, G. N., Tariq, S., Ehteshamul-Haque, S., Athar, M., Sultana, V., & Ara, J. (2013). Management of root diseases of eggplant and watermelon with the application of asafetida and seaweeds. *Journal of Applied Botany and Food Quality*, 86(1).
- Balogun, O. S., Hirano, Y., Teraoka, T., & Arie, T. (2008). PCR-based analysis of disease in tomato singly or mixed inoculated with *Fusarium oxysporum* f. sp. *lycopersici* races 1 and 2. *Phytopathologia Mediterranea*, 47(1), 50–60.

- Bao, J. R., Fravel, D. R., O'Neill, N. R., Lazarovits, G., & Berkum, P. van. (2002). Genetic analysis of pathogenic and nonpathogenic *Fusarium oxysporum* from tomato plants. *Canadian Journal of Botany*, 80(3), 271–279.
- Barbará, T., Martinelli, G., Fay, M. F., Mayo, S. J., & Lexer, C. (2007). Population differentiation and species cohesion in two closely related plants adapted to neotropical high-altitude 'inselbergs', *Alcantarea imperialis* and *Alcantarea geniculata* (Bromeliaceae). *Molecular Ecology*, 16(10), 1981–1992.
- Barres, B., Dutech, C., Andrieux, A., Caron, H., Pinon, J., & Frey, P. (2006). Isolation and characterization of 15 microsatellite loci in the poplar rust fungus, *Melampsora larici-populina*, and cross-amplification in related species. *Molecular Ecology Notes*, 6(1), 60–64.
- Barve, M. P., Haware, M. P., Sainani, M. N., Ranjekar, P. K., & Gupta, V. S. (2001). Potential of microsatellites to distinguish four races of *Fusarium oxysporum* f. sp. *ciceri* prevalent in India. *Theoretical and Applied Genetics*, 102(1), 138–147.
- Basım, E., Basım, H., Abdulai, M., Baki, D., & Öztürk, N. (2016). Identification and characterization of *Didymella bryoniae* causing gummy stem blight disease of watermelon (*Citrullus lanatus*) in Turkey. *Crop Protection*, 90, 150–156.
- Bayraktar, H., Dolar, F. S., & Maden, S. (2008). Use of RAPD and ISSR Markers in Detection of Genetic Variation and Population Structure among *Fusarium oxysporum* f. sp. *ciceris* Isolates on Chickpea in Turkey. *Journal of Phytopathology*, 156(3), 146–154.
- Bayraktar, Harun, & Dolar, F. S. (2011). Molecular Identification and Genetic Diversity of Fusarium species Associated with Onion Fields in Turkey. *Journal of Phytopathology*, 159(1), 28–34.
- Bayraktar, Harun, & Dolar, F. S. (2012). Pathogenic variability of *Fusarium oxysporum* f. sp. *ciceris* isolates from chickpea in Turkey. *Pakistan Journal of Botany*, 44(2), 821–823.
- Bayraktar, Harun, Türkkán, M., & Dolar, F. S. (2010). Characterization of *Fusarium oxysporum* f.sp. *cepae* from Onion in Turkey Based on Vegetative Compatibility and rDNA RFLP Analysis. *Journal of Phytopathology*, 158(10), 691–697.
- Beckman, C. H., & Roberts, E. M. (1995). On the nature and genetic basis for resistance and tolerance to fungal wilt diseases of plants. In *Advances in Botanical Research* (Vol. 21, pp. 35-77). Academic Press.
- Bertoldo, C., Gilardi, G., Spadaro, D., Gullino, M. L., & Garibaldi, A. (2015). Genetic diversity and virulence of Italian strains of *Fusarium oxysporum* isolated from *Eustoma grandiflorum*. *European Journal of Plant Pathology*, 141(1), 83–97.
- Bharath, B. G., Lokesh, S., & Shetty, H. S. (2005). Effects of fungicides and bioagents on seed Mycoflora, growth and yield of watermelon. *Integrative Biosciences*, 9(2), 75–78.

- Bogale, M., Wingfield, B. D., Wingfield, M. J., & Steenkamp, E. T. (2005). Simple sequence repeat markers for species in the *Fusarium oxysporum* complex. *Molecular Ecology Notes*, 5(3), 622–624.
- Booth, C. (1971). *The genus fusarium*. Commonwealth Mycological Institute, Key, Surrey (England).
- Bosland, P. W., & Williams, P. H. (1987). An evaluation of *Fusarium oxysporum* from crucifers based on pathogenicity, isozyme polymorphism, vegetative compatibility, and geographic origin. *Canadian Journal of Botany*, 65(10), 2067–2073.
- Boughalleb, N., & Mahjoub, M. El. (2007). Frequency of *Fusarium oxysporum* f. sp. *niveum* and *F. solani* f. sp. *Cucurbitae* from Watermelon Seeds and Their Effect on Disease Incidence. *Research Journal of Parasitology*, 2(1), 32–38.
- Branham, S. E., Levi, A., & Wechter, W. P. (2019). QTL Mapping Identifies Novel Source of Resistance to Fusarium Wilt Race 1 in *Citrullus amarus*. *Plant Disease*, 103(5), 984–989.
- Britz, H., Wingfield, B. D., Coutinho, T. A., & Wingfield, M. J. (2002). Sequence characterized amplified polymorphic markers for the pitch canker pathogen, *Fusarium circinatum*. *Molecular Ecology Notes*, 2(4), 577–580.
- Bruns, T. D., White, T. J., & Taylor, J. W. (1991). Fungal Molecular Systematics. *Annual Review of Ecology and Systematics*, 22(1), 525–564.
- Bruns, T. M. S. (1992). Rate and mode differences between nuclear and mitochondrial small-subunit rRNA genes in mushrooms. *Molecular Biology and Evolution*, 9(5), 836–855.
- Bruton, B. D., Fish, W. W., Zhou, X. G., Everts, K. L., & Roberts, P. D. (2007, January). Fusarium wilt in seedless watermelons. In *Proceeding 2007 Southeast Regional Vegetable Conference* (pp. 93-8).
- Chakrabarti, A., Rep, M., Wang, B., Ashton, A., Dodds, P., & Ellis, J. (2011). Variation in potential effector genes distinguishing Australian and non-Australian isolates of the cotton wilt pathogen *Fusarium oxysporum* f.sp. *vasinfectum*. *Plant Pathology*, 60(2), 232–243.
- Chehri, K., Salleh, B., Yli-Mattila, T., Reddy, K. R. N., & Abbasi, S. (2011). Molecular characterization of pathogenic *Fusarium* species in cucurbit plants from Kermanshah province, Iran. *Saudi Journal of Biological Sciences*, 18(4), 341–351.
- Chen, H., Liu, L., Wang, L., Wang, S., Somta, P., & Cheng, X. (2015). Development and Validation of EST-SSR Markers from the Transcriptome of Adzuki Bean (*Vigna angularis*). *PloS one*, 10(7), 0131939.
- Chomicki, G., & Renner, S. S. (2015). Watermelon origin solved with molecular phylogenetics including L innaean material: another example of museomics. *New Phytologist*, 205(2), 526–532.

- Cohen, R., Tyutyunik, J., Fallik, E., Oka, Y., Tadmor, Y., & Edelstein, M. (2014). Phytopathological evaluation of exotic watermelon germplasm as a basis for rootstock breeding. *Scientia Horticulturae*, 165, 203–210.
- Cohen, Ron, Burger, Y., Horev, C., & Koren, A. (2007). Introducing grafted cucurbits to modern agriculture: the Israeli experience. *Plant Disease*, 91(8), 916–923.
- Costa, A. E. S., da Cunha, F. S., Honorato, A. da C., Capucho, A. S., Dias, R. de C. S., Borel, J. C., & Ishikawa, F. H. (2018). Resistance to Fusarium Wilt in watermelon accessions inoculated by chlamydospores. *Scientia Horticulturae*, 228, 181–186.
- Costa, R., Pereira, G., Garrido, I., Tavares-de-Sousa, M. M., & Espinosa, F. (2016). Comparison of RAPD, ISSR, and AFLP molecular markers to reveal and classify orchardgrass (*Dactylis glomerata* L.) germplasm variations. *PloS one*, 11(4), 0152972.
- Costa, S. N., Bragança, C. A. D., Ribeiro, L. R., Amorim, E. P., Oliveira, S. A. S., Dita, M. A., Laranjeira, F. F., & Haddad, F. (2015). Genetic structure of *Fusarium oxysporum* f. sp. *cubense* in different regions from Brazil. *Plant Pathology*, 64(1), 137–146.
- Covey, P. A., Kuwitzky, B., Hanson, M., & Webb, K. M. (2014). Multilocus Analysis Using Putative Fungal Effectors to Describe a Population of *Fusarium oxysporum* from Sugar Beet. *Phytopathology*, 104(8), 886–896.
- Cruz, A. F., Silva, L. F., Sousa, T. V., Nicoli, A., de Paula Junior, T. J., Caixeta, E. T., & Zambolim, L. (2018). Molecular diversity in *Fusarium oxysporum* isolates from common bean fields in Brazil. *European Journal of Plant Pathology*, 152(2), 343–354.
- Czislowski, E., Fraser-Smith, S., Zander, M., O'Neill, W. T., Meldrum, R. A., Tran-Nguyen, L. T. T., Batley, J., & Aitken, E. A. B. (2018). Investigation of the diversity of effector genes in the banana pathogen, *Fusarium oxysporum* f. sp. *cubense*, reveals evidence of horizontal gene transfer. *Molecular Plant Pathology*, 19(5), 1155–1171.
- D'Amore, R., Morra, L., & Parisi, B. (1996). Grafted watermelon: production results. *Colture Protette*, 25(9), 29–31.
- Dane, F., & Liu, J. (2007). Diversity and origin of cultivated and citron type watermelon (*Citrullus lanatus*). *Genetic Resources and Crop Evolution*, 54(6), 1255–1265.
- Daniel, H. C. F., Wilfredo, F. F., Francisco, C. R., Gabriel, G. M., & Epifanio, C. D. Á. (2014). Antibiosis in vitro of Trichoderma strains metabolic extract on mycelial growth and reproductive capacity of *Fusarium oxysporum* isolated from pepper plants (*Capsicum annuum* L.). *British Biotechnology Journal*, 4(4), 387.
- Das, A., Venkataramana, M., Chandranayaka, S., Murali, H. S., & Batra, H. V. (2012). Molecular characterization of *Fusarium oxysporum* f. sp. *cubense* isolates from

- banana. *Pest Management in Horticultural Ecosystems*, 18(2), 171–178.
- Datta, S., Choudhary, R. G., Shamim, M. D., Singh, R. K., & Dhar, V. (2011). Molecular diversity in Indian isolates of *Fusarium oxysporum* f. sp. *lentis* inciting wilt disease in lentil (*Lens culinaris* Medik). *African Journal of Biotechnology*, 10(38), 7314–7323.
- De Cal, A., Szejnberg, A., Sabuquillo, P., & Melgarejo, P. (2009). Management Fusarium wilt on melon and watermelon by *Penicillium oxalicum*. *Biological Control*, 51(3), 480–486.
- de Melo Oliveira, P. R. P., Dianese, A. C., Fragoso, R. R., Cruz, A. F., & Blum, L. E. B. (2014). Variability of *Fusarium* spp. isolates, causal agents of the soybean sudden death syndrome. *Acta Agriculturae Scandinavica, Section B - Soil & Plant Science*, 64(8), 675–682.
- de Sain, M., & Rep, M. (2015). The Role of Pathogen-Secreted Proteins in Fungal Vascular Wilt Diseases. *International Journal of Molecular Sciences*, 16(10), 23970–23993.
- Dhaouadi, S., Rouissi, W., Mougou-Hamdane, A., Hannachi, I., and Nasraoui, B. (2018). Antifungal Activity of Essential Oils of *Origanum majorana* and *Lavender angustifolia* against Fusarium Wilt and Root Rot Disease of Melon Plants. *Tunisian Journal of Plant Protection*, 13(1), 39–55.
- Di, X., Takken, F. L. W., & Tintor, N. (2016). How Phytohormones Shape Interactions between Plants and the Soil-Borne Fungus *Fusarium oxysporum*. *Frontiers in Plant Science*, 7, 170.
- Dissanayake, M. L. M. C., Kashima, R., Tanaka, S., & Ito, S. (2009). Pathogenic variation and molecular characterization of *Fusarium* species isolated from wilted Welsh onion in Japan. *Journal of General Plant Pathology*, 75(1), 37–45.
- Dube, J., Ddamulira, G., & Maphosa, M. (2020). Watermelon production in Africa: challenges and opportunities. *International Journal of Vegetable Science*, 27(3), 211–219.
- Dutech, C., Enjalbert, J., Fournier, E., Delmotte, F., Barrès, B., Carlier, J., Tharreau, D., & Giraud, T. (2007). Challenges of microsatellite isolation in fungi. *Fungal Genetics and Biology*, 44(10), 933–949.
- Edel-Hermann, V., Gautheron, N., & Steinberg, C. (2012). Genetic diversity of *Fusarium oxysporum* and related species pathogenic on tomato in Algeria and other Mediterranean countries. *Plant Pathology*, 61(4), 787–800.
- Egel, D. S., & Martyn, R. D. (2013). Fusarium wilt of watermelon and other cucurbits. *Plant Health Instructor*.
- Egel, D. S., & Hoke, S. (2011). Evaluation of fungicides for the control of anthracnose of watermelon, 2010. *Plant Disease Management Reports*, 5, V022.

- Enya, J., Togawa, M., Takeuchi, T., Yoshida, S., Tsushima, S., Arie, T., & Sakai, T. (2008). Biological and phylogenetic characterization of *Fusarium oxysporum* complex, which causes yellows on *Brassica* spp., and proposal of *F. oxysporum* f. sp. *rapae*, a novel forma specialis pathogenic on *B. rapa* in Japan. *Phytopathology*, 98(4), 475–483.
- Everts, K L, & Hochmuth, M. (2011). Field evaluation of triploid cultivars for resistance to Fusarium wilt of watermelon in Delaware, 2010. *Plant Disease Management Reports*, 5, V175.
- Everts, Kathryn L., Egel, D. S., Langston, D., & Zhou, X.-G. (2014). Chemical management of Fusarium wilt of watermelon. *Crop Protection*, 66, 114–119.
- Everts, Kathryn L., & Himmelstein, J. C. (2015). Fusarium wilt of watermelon: Towards sustainable management of a re-emerging plant disease. *Crop Protection*, 73, 93–99.
- Fall, L. A., Clevenger, J., & McGregor, C. (2018). Assay development and marker validation for marker assisted selection of *Fusarium oxysporum* f. sp. *niveum* race 1 in watermelon. *Molecular Breeding*, 38(11), 130.
- FAOSTAT. (2020). *Food and agriculture organization of the united nations statistics database*. <https://www.fao.org/faostat/en/#home>. Accessed 19 March 2022.
- Fourie, G., Steenkamp, E. T., Gordon, T. R., & Viljoen, A. (2009). Evolutionary Relationships among the *Fusarium oxysporum* f. sp. *cubense* Vegetative Compatibility Groups. *Applied and Environmental Microbiology*, 75(14), 4770–4781.
- Fraser-Smith, S., Czislowski, E., Meldrum, R. A., Zander, M., O'Neill, W., Balali, G. R., & Aitken, E. A. B. (2014). Sequence variation in the putative effector gene SIX8 facilitates molecular differentiation of *Fusarium oxysporum* f. sp. *cubense*. *Plant Pathology*, 63(5), 1044–1052.
- Fravel, D. R. (2005). Commercialization and implementation of biocontrol. *Annual Review of Phytopathology*, 43, 337–359.
- Fulton, J. C., Amaradasa, B. S., Ertek, T. S., Iriarte, F. B., Sanchez, T., Ji, P., Paret, M. L., Hudson, O., Ali, M. E., & Dufault, N. S. (2021). Phylogenetic and phenotypic characterization of *Fusarium oxysporum* f. sp. *niveum* isolates from Florida-grown watermelon. *PLoS one*, 16(3), 0248364.
- García-Mendivil, H. A., Munera, M., Giné, A., Escudero, N., Picó, M. B., Gisbert, C., & Sorribas, F. J. (2019). Response of two *Citrullus amarus* accessions to isolates of three species of Meloidogyne and their graft compatibility with watermelon. *Crop Protection*, 119, 208–213.
- Garmendia, G., Umpierrez-Failache, M., Ward, T. J., & Vero, S. (2018). Development of a PCR-RFLP method based on the transcription elongation factor 1- α gene to differentiate *Fusarium graminearum* from other species within the *Fusarium graminearum* species complex. *Food Microbiology*, 70, 28–32.

- Gawehns, F., Houterman, P. M., Ichou, F. A., Michielse, C. B., Hijdra, M., Cornelissen, B. J. C., Rep, M., & Takken, F. L. W. (2014). The *Fusarium oxysporum* Effector Six6 Contributes to Virulence and Suppresses I-2-Mediated Cell Death. *Molecular Plant-Microbe Interactions*, 27(4), 336–348.
- Geiser, D. M., del Mar Jiménez-Gasco, M., Kang, S., Makalowska, I., Veeraraghavan, N., Ward, T. J., Zhang, N., Kuldau, G. A., & O'donnell, K. (2004). FUSARIUM-ID v. 1.0: a DNA sequence database for identifying Fusarium. *European Journal of Plant Pathology*, 110(5), 473–479.
- George Agrios. (2005). *Plant Pathology* (5th Editio). Academic Press.
- Grin, A., Matsum, L., & Nakai, O. (2010). Taxon *Citrullus lanatus* (Thunb) Taxonomy for plants. Germplasm Resources Information Network. *Agricultural Research Service, Beltsville Area*.
- Groenewald, S. (2007). Biology, pathogenicity and diversity of *Fusarium oxysporum*. University of Pretoria.
- Guner*, N., Pesic-VanEsbroeck, Z., & Wehner, T. (2004). Inheritance of Resistance to the Watermelon Strain of Papaya Ringspot Virus in Watermelon. *HortScience*, 39(4), 811C-811.
- Haegi, A., Catalano, V., Luongo, L., Vitale, S., Scotton, M., Ficcadenti, N., & Belisario, A. (2013). A Newly Developed Real-Time PCR Assay for Detection and Quantification of *Fusarium oxysporum* and Its Use in Compatible and Incompatible Interactions with Grafted Melon Genotypes. *Phytopathology*, 103(8), 802–810.
- Hafez, M., Abdelmagid, A., Adam, L. R., & Daayf, F. (2020). Specific Detection and Identification of *Fusarium graminearum* Sensu Stricto Using a PCR-RFLP Tool and Specific Primers Targeting the Translational Elongation Factor 1 α Gene. *Plant Disease*, 104(4), 1076–1086.
- Hall, T. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, 41, 95–98.
- Hall, T., Biosciences, I., & Carlsbad, C. (2011). BioEdit: an important software for molecular biology. *GERF Bulletin of Biosciences*, 2(1), 60–61.
- Hamed, E. R., Abdel-Sayed, M. H. F., & Shehata, H. S. (2009). Suppression of Fusarium wilt of watermelon by biological and chemical control. *Journal of Applied Sciences Research*, 5(10), 1816–1825.
- Hao, W., Ren, L., Ran, W., & Shen, Q. (2010). Allelopathic effects of root exudates from watermelon and rice plants on *Fusarium oxysporum* f. sp. *niveum*. *Plant and Soil*, 336(1), 485–497.
- Hasegawa, M., Kishino, H., & Yano, T. (1985). Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *Journal of Molecular Evolution*, 22(2),

160–174.

- Ho, L. H., Khandaker, M. M., Fah, J. B. C., & Tan, T. C. (2017). Cultivation, common diseases and potential nutraceutical values of watermelon. *Research Updates*, 71.
- Holliday, P. (1970). *Fusarium oxysporum* f.sp. *niveum*. [Descriptions of Fungi and Bacteria]. *IMI Descriptions of Fungi and Bacteria*, 22.
- Hoque, M. S., Uddin M. F., & Islam M. A. (2015). A market model for watermelon with supply under rational expectations: An empirical study on Bangladesh. *European Scientific Journal*, 11(9), 236.
- Houterman, P. M., Speijer, D., Dekker, H. L., De Koster, C. G., Cornelissen, B. J. C., & Rep, M. (2007). The mixed xylem sap proteome of *Fusarium oxysporum* infected tomato plants. *Molecular Plant Pathology*, 8(2), 215–221.
- Hua, G. K. H., Timper, P., & Ji, P. (2019). Meloidogyne incognita intensifies the severity of Fusarium wilt on watermelon caused by *Fusarium oxysporum* f. sp. *niveum*. *Canadian Journal of Plant Pathology*, 41(2), 261–269.
- Huang, C.-H., Roberts, P. D., Gale, L. R., Elmer, W. H., & Datnoff, L. E. (2013). Population structure of *Fusarium oxysporum* f. sp. *radicis-lycopersici* in Florida inferred from vegetative compatibility groups and microsatellites. *European Journal of Plant Pathology*, 136(3), 509–521.
- Hudson, O., Waliullah, S., Fulton, J. C., Ji, P., Dufault, N. S., Keinath, A., & Ali, M. E. (2021). Marker Development for Differentiation of *Fusarium oxysporum* f. sp. *niveum* Race 3 from Races 1 and 2. *International Journal of Molecular Sciences*, 22(2), 822.
- Hussein, S., & Sari, N. (2020). Effects of different rootstocks on seed yield and quality of triploid watermelon grown in greenhouse. *Acta Horticulturae*, 1282, 67–74.
- Karaca, F., Yetişir, H., Solmaz, İ., Candir, E., Kurt, Ş., Sari, N., & Güler, Z. (2012). Rootstock potential of Turkish *Lagenaria siceraria* germplasm for watermelon: plant growth, yield and quality. *Turkish Journal of Agriculture and Forestry*, 36(2), 167–177.
- Karaoglu, H., Lee, C. M. Y., & Meyer, W. (2005). Survey of simple sequence repeats in completed fungal genomes. *Molecular Biology and Evolution*, 22(3), 639–649.
- Karlsson, I., Edel-Hermann, V., Gautheron, N., Durling, M. B., Kolseth, A.-K., Steinberg, C., Persson, P., & Friberg, H. (2016). Genus-specific primers for study of *Fusarium* communities in field samples. *Applied and Environmental Microbiology*, 82(2), 491–501.
- Kashiwa, T., Inami, K., Teraoka, T., Komatsu, K., & Arie, T. (2016). Detection of cabbage yellows fungus *Fusarium oxysporum* f. sp. *conglutinans* in soil by PCR and real-time PCR. *Journal of General Plant Pathology*, 82(5), 240–247.
- Kawabe, M., Kobayashi, Y., Okada, G., Yamaguchi, I., Teraoka, T., & Arie, T. (2005). Three evolutionary lineages of tomato wilt pathogen, *Fusarium oxysporum* f. sp.

lycopersici, based on sequences of IGS, MAT1, and pg1, are each composed of isolates of a single mating type and a single or closely related vegetative compatibility group. *Journal of General Plant Pathology*, 71(4), 263–272.

Kee, Y. J., Zakaria, L., & Mohd, M. H. (2020). Morphology, phylogeny and pathogenicity of *Fusarium* species from *Sansevieria trifasciata* in Malaysia. *Plant Pathology*, 69(3), 442–454.

Keinath, A. P., & Hassell, R. L. (2014). Control of *Fusarium* Wilt of Watermelon by Grafting onto Bottlegourd or Interspecific Hybrid Squash Despite Colonization of Rootstocks by *Fusarium*. *Plant Disease*, 98(2), 255–266.

Keinath, A. P., Wintermantel, W. M., & Zitter, T. A. (Eds.). (2017). *Compendium of Cucurbit Diseases and Pests, Second Edition*. The American Phytopathological Society.

Keinath, Anthony P., DuBose, V. B., Katawczik, M. M., & Wechter, W. P. (2020). Identifying Races of *Fusarium oxysporum* f. sp. *niveum* in South Carolina Recovered From Watermelon Seedlings, Plants, and Field Soil. *Plant Disease*, 104(9), 2481–2488.

Keinath, Anthony P., Hassell, R. L., Everts, K. L., & Zhou, X.-G. (2010). Cover Crops of Hybrid Common Vetch Reduce *Fusarium* Wilt of Seedless Watermelon in the Eastern United States. *Plant Health Progress*, 11(1), 8.

Keinath, A. P., Coolong, T. W., Lanier, J. D., & Ji, P. (2019a). Managing *Fusarium* wilt of watermelon with delayed transplanting and cultivar resistance. *Plant Disease*, 103(1), 44–50.

Keinath, A. P., Wechter, W. P., Rutter, W. B., & Agudelo, P. A. (2019b). Cucurbit Rootstocks Resistant to *Fusarium oxysporum* f. sp. *niveum* Remain Resistant When Coinfected by *Meloidogyne incognita* in the Field. *Plant Disease*, 103(6), 1383–1390.

Kemble, J. M. (2018). *Vegetable Crop Handbook for Southeastern US—2010*.

Khanam, M., & Hafsa, U. (2013). Market model analysis and forecasting behavior of watermelon production in Bangladesh. *Bangladesh Journal of Scientific Research*, 26(1&2), 47–56.

Kimura, M., & Crow, J. F. (1964). The number of alleles that can be maintained in a finite population. *Genetics*, 49(4), 725.

Kleczewski, N. M., & Egel, D. S. (2011). A Diagnostic Guide for *Fusarium* Wilt of Watermelon. *Plant Health Progress*, 12(1), 27.

Klein, E., Katan, J., & Gamliel, A. (2012). Soil suppressiveness to *Meloidogyne javanica* as induced by organic amendments and solarization in greenhouse crops. *Crop Protection*, 39, 26–32.

Komada, H. (1975). Development of a selective medium for quantitative isolation of

- Fusarium oxysporum* from natural soil. *Review of Plant Protection Research*, 8, 114–124.
- Kumar, Sudheer, Maurya, D., Shalini, R. A. I., Kashyap, P. L., & Srivastava, A. K. (2012). Computational mining and genome wide distribution of microsatellite in *Fusarium oxysporum* f. sp. *lycopersici*. *Notulae Scientia Biologicae*, 4(4), 127–131.
- Kumar, Sudhir, Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution*, 35(6), 1547–1549.
- Kuppusamy, A., Amar, G., & Roshni, N. P. (2018). Molecular characterization of *Fusarium oxysporum* by PCR amplification of 18S rRNA gene region.
- Lambel, S., Lanini, B., Vivoda, E., Fauve, J., Patrick Wechter, W., Harris-Shultz, K. R., Massey, L., & Levi, A. (2014). A major QTL associated with *Fusarium oxysporum* race 1 resistance identified in genetic populations derived from closely related watermelon lines using selective genotyping and genotyping-by-sequencing for SNP discovery. *Theoretical and Applied Genetics*, 127(10), 2105–2115.
- Lanubile, A., Ellis, M. L., Marocco, A., & Munkvold, G. P. (2016). Association of Effector Six 6 with Vascular Wilt Symptoms Caused by *Fusarium oxysporum* on Soybean. *Phytopathology*, 106(11), 1404–1412.
- Larkin, R. P., Hopkins, D. L., & Martin, F. N. (1990). Vegetative compatibility within *Fusarium oxysporum* f.sp. *niveum* and its relationship to virulence, aggressiveness, and race. *Canadian Journal of Microbiology*, 36(5), 352–358.
- Laurence, M. H., Summerell, B. A., & Liew, E. C. Y. (2015). *Fusarium oxysporum* f. sp. *canariensis*: evidence for horizontal gene transfer of putative pathogenicity genes. *Plant Pathology*, 64(5), 1068–1075.
- LeBlanc, N., Essarioui, A., Kinkel, L., & Kistler, H. C. (2017). Phylogeny, Plant Species, and Plant Diversity Influence Carbon Use Phenotypes Among *Fusarium* Populations in the Rhizosphere Microbiome. *Phytobiomes Journal*, 1(3), 150–157.
- Lehner, M. S., Paula Júnior, T. J., Hora Júnior, B. T., Teixeira, H., Vieira, R. F., Carneiro, J. E. S., & Mizubuti, E. S. G. (2015). Low genetic variability in *Sclerotinia sclerotiorum* populations from common bean fields in Minas Gerais State, Brazil, at regional, local and micro-scales. *Plant Pathology*, 64(4), 921–931.
- Leskovar, D., Bang, H., Crosby, K., Maness, N., Franco, A., & Perkins-Veazie, P. (2004). Lycopene, carbohydrates, ascorbic acid and yield components of diploid and triploid watermelon cultivars are affected by de@cit irrigation. *The Journal of Horticultural Science and Biotechnology*, 79(1), 75–81.

- Leslie and Summerell, B. A. (2006). *The Fusarium Laboratory Manual*. Blackwell Publishing, Ames, Iowa.
- Leslie, J. F., & Summerell, B. A. (2008). *The Fusarium laboratory manual*. John Wiley & Sons.
- Leslie, J. F., Zeller, K. A., & Summerell, B. A. (2001). Icebergs and species in populations of *Fusarium*. *Physiological and Molecular Plant Pathology*, 59(3), 107–117.
- Levene, H. (1949). On a matching problem arising in genetics. *The Annals of Mathematical Statistics*, 91–94.
- Li, C.X., Fu, X.P., Zhou, X.G., Liu, S.W., Xia, Y., Li, N.H., Zhang, X.X., & Wu, F.Z. (2019). Treatment With Wheat Root Exudates and Soil Microorganisms From Wheat/Watermelon Companion Cropping Can Induce Watermelon Disease Resistance Against *Fusarium oxysporum* f. sp. *niveum*. *Plant Disease*, 103(7), 1693-1702.
- Li, K. N., Rouse, D. I., & German, T. L. (1994). PCR primers that allow intergeneric differentiation of ascomycetes and their application to *Verticillium* spp. *Applied and Environmental Microbiology*, 60(12), 4324-4331.
- Li, S., Tam, Y. K., & Hartman, G. L. (2000). Molecular differentiation of *Fusarium solani* f. sp. *glycines* from other *F. solani* based on mitochondrial small subunit rDNA sequences. *Phytopathology*, 90(5), 491-497.
- Lievens, B., Houterman, P. M., & Rep, M. (2009). Effector gene screening allows unambiguous identification of *Fusarium oxysporum* f. sp. *lycopersici* races and discrimination from other formae speciales. *FEMS Microbiology Letters*, 300(2), 201–215.
- Lin, Y. H., Chen, K. S., Liou, T. D., Huang, J. W., & Chang, P. F. L. (2009a). Development of a molecular method for rapid differentiation of watermelon lines resistant to *Fusarium oxysporum* f. sp. *niveum*. *Botanical Studies*, 50(3), 273–280.
- Lin, Y. H., Chang, J. Y., Liu, E. T., Chao, C. P., Huang, J. W., & Chang, P. F. L. (2009b). Development of a molecular marker for specific detection of *Fusarium oxysporum* f. sp. *cubense* race 4. *European Journal of Plant Pathology*, 123(3), 353-365.
- Lin, Y.-H., Chang, J.-Y., Liu, E.-T., Chao, C.-P., Huang, J.-W., & Chang, P.-F. L. (2009c). Development of a molecular marker for specific detection of *Fusarium oxysporum* f. sp. *cubense* race 4. *European Journal of Plant Pathology*, 123(3), 353–365.
- Lin, Y.H., Chen, K.S., Chang, J.Y., Wan, Y.L., Hsu, C.C., Huang, J.W., & Chang, P.F. L. (2010). Development of molecular methods for rapid detection and differentiation of *Fusarium oxysporum* and *F. oxysporum* f. sp. *niveum* in Taiwan. *New Biotechnology*, 27(4), 409-418.

- Ling, N., Xue, C., Huang, Q., Yang, X., Xu, Y., & Shen, Q. (2010). Development of a mode of application of bioorganic fertilizer for improving the biocontrol efficacy to *Fusarium* wilt. *BioControl*, 55(5), 673–683.
- Ling, N., Zhang, W., Wang, D., Mao, J., Huang, Q., Guo, S., & Shen, Q. (2013). Root Exudates from Grafted-Root Watermelon Showed a Certain Contribution in Inhibiting *Fusarium oxysporum* f. sp. *niveum*. *PLoS one*, 8(5), 63383.
- Lombard, L., Van Doorn, R., & Crous, P. W. (2019). Neotypification of *Fusarium chlamydosporum*-a reappraisal of a clinically important species complex. *Fungal Systematics and Evolution*, 4, 183.
- Louie, M., Louie, L., & Simor, A. E. (2000). The role of DNA amplification technology in the diagnosis of infectious diseases. *Cmaj*, 163(3), 301–309.
- Lü, G., Guo, S., Zhang, H., Geng, L., Song, F., Fei, Z., & Xu, Y. (2011). Transcriptional profiling of watermelon during its incompatible interaction with *Fusarium oxysporum* f. sp. *niveum*. *European Journal of Plant Pathology*, 131(4), 585–601.
- Lum, T., Connolly, M., Marx, A., Beidler, J., Hooshmand, S., Kern, M., Liu, C., & Hong, M. (2019). Effects of Fresh Watermelon Consumption on the Acute Satiety Response and Cardiometabolic Risk Factors in Overweight and Obese Adults. *Nutrients*, 11(3), 595.
- Lv, H., Cao, H., Nawaz, M. A., Sohail, H., Huang, Y., Cheng, F., Kong, Q., & Bie, Z. (2018). Wheat Intercropping Enhances the Resistance of Watermelon to *Fusarium* Wilt. *Frontiers in Plant Science*, 9, 696.
- Lv, P., & Gu, H. (2013). Studies on carotenoids in watermelon flesh.
- Ma, L.J., van der Does, H. C., Borkovich, K. A., Coleman, J. J., Daboussi, M.J., Di Pietro, A., Dufresne, M., Freitag, M., Grabherr, M., Henrissat, B., Houterman, P. M., Kang, S., Shim, W.B., Woloshuk, C., Xie, X., Xu, J.R., Antoniw, J., Baker, S. E., Bluhm, B. H., ... Rep, M. (2010). Comparative genomics reveals mobile pathogenicity chromosomes in *Fusarium*. *Nature*, 464(7287), 367–373.
- Madden, L. V., Hughes, G., & van den Bosch, F. (2017). *The Study of Plant Disease Epidemics* (Volume 19). The American Phytopathological Society.
- Mahfooz, S., Maurya, D. K., Srivastava, A. K., Kumar, S., & Arora, D. K. (2012). A comparative in silico analysis on frequency and distribution of microsatellites in coding regions of three formae speciales of *Fusarium oxysporum* and development of EST-SSR markers for polymorphism studies. *FEMS Microbiology Letters*, 328(1), 54–60.
- Mahfooz, S., Srivastava, A., Srivastava, A. K., & Arora, D. K. (2015). A comparative analysis of distribution and conservation of microsatellites in the transcripts of sequenced *Fusarium* species and development of genic-SSR markers for polymorphism analysis. *FEMS Microbiology Letters*, fnv131.

- Mahmood, K., Saleem, M., & Ahsan, M. (2011). Inheritance of resistance to Fusarium wilts in chickpea. *Pakistan Journal of Agricultural Science*, 48(1), 55–58.
- Maina, P., Wachira, P., Okoth, S., & Kimenju, J. (2017). Cultural, Morphological and Pathogenic Variability among *Fusarium oxysporum* f. sp. *phaseoli* Causing Wilt in French Bean (*Phaseolus vulgaris* L.). *Journal of Advances in Microbiology*, 2(4), 1–9.
- Manikandan, R., Harish, S., Karthikeyan, G., & Raguchander, T. (2018). Comparative Proteomic Analysis of Different Isolates of *Fusarium oxysporum* f.sp. *lycopersici* to Exploit the Differentially Expressed Proteins Responsible for Virulence on Tomato Plants. *Frontiers in Microbiology*, 9.
- Maoto, M. M., Beswa, D., & Jideani, A. I. O. (2019). Watermelon as a potential fruit snack. *International Journal of Food Properties*, 22(1), 355–370.
- Mariod, A. A., Ahmed, Y. M., Matthäus, B., Khaleel, G., Siddig, A., Gabra, A. M., & Abdelwahab, S. I. (2009). A Comparative Study of the Properties of Six Sudanese Cucurbit Seeds and Seed Oils. *Journal of the American Oil Chemists' Society*, 86(12), 1181–1188.
- Martyn, R. D. (2012). Fusarium wilt of watermelon: a historical review. In *Cucurbitaceae 2012. Proceedings of the Xth EUCARPIA Meeting on Genetics and Breeding of Cucurbitaceae, Antalya, Turkey, 15-18 October, 2012* (pp. 136–156). University of Cukurova, Ziraat Fakultesi.
- Martyn, R. D. (1987). *Fusarium oxysporum* f. sp. *niveum* Race 2: A Highly Aggressive Race New to the United States. *Plant Disease*, 71(3), 233.
- Martyn, R.D., & Netzer, D. (1991). Resistance to Races 0, 1, and 2 of Fusarium Wilt of Watermelon in *Citrullus* sp. PI-296341 -FR. *HortScience*, 26(4), 429–432.
- Martyn, Ray D. (2014). Fusarium Wilt of Watermelon: 120 Years of Research. In *Horticultural Reviews: Volume 42* (pp. 349–442). John Wiley & Sons, Inc.
- Martyn, Raymond D., & Vakalounakis, D. J. (2017). Chapter 16: Fusarium wilts of Greenhouse Cucurbits: Melon, Watermelon, and Cucumber. In *Fusarium Wilts of Greenhouse Vegetable and Ornamental Crops* (pp. 159–174). *The American Phytopathological Society*.
- Mbofung, G. Y., Hong, S. G., & Pryor, B. M. (2007). Phylogeny of *Fusarium oxysporum* f. sp. *lactucae* inferred from mitochondrial small subunit, elongation factor 1- α , and nuclear ribosomal intergenic spacer sequence data. *Phytopathology*, 97(1), 87–98.
- McCartney, H. A., Foster, S. J., Fraaije, B. A., & Ward, E. (2003). Molecular diagnostics for fungal plant pathogens. *Pest Management Science: Formerly Pesticide Science*, 59(2), 129–142.
- McDonald, B. A., Linde, C., & McDonald, B. A., & Linde, C. (2002). Pathogen population genetics, evolutionary potential, and durable resistance. *Annual Review of Phytopathology*, 40(1), 349–379.

- Meru, G., & McGregor, C. (2016). Genotyping by sequencing for SNP discovery and genetic mapping of resistance to race 1 of *Fusarium oxysporum* in watermelon. *Scientia Horticulturae*, 209, 31–40.
- Merzoug, A., & Belabid, L. (2018). Relationship between pathogenicity, race and vegetative compatibility grouping among Algerian populations of *Fusarium oxysporum* f. sp. *pisi* causing pea wilt. *Journal of Plant Protection Research*.
- Meyer, A., Todt, C., Mikkelsen, N. T., & Lieb, B. (2010). Fast evolving 18S rRNA sequences from Solenogastres (Mollusca) resist standard PCR amplification and give new insights into mollusk substitution rate heterogeneity. *BMC Evolutionary Biology*, 10(1), 1–12.
- Miguel, A., Maroto, J. V., San Bautista, A., Baixauli, C., Cebolla, V., Pascual, B., López, S., & Guardiola, J. L. (2004). The grafting of triploid watermelon is an advantageous alternative to soil fumigation by methyl bromide for control of Fusarium wilt. *Scientia Horticulturae*, 103(1), 9–17.
- Miller, Nathan F., Standish, J. R., & Quesada-Ocampo, L. M. (2020). Sensitivity of *Fusarium oxysporum* f. sp. *niveum* to Prothioconazole and Pydiflumetofen In Vitro and Efficacy for Fusarium Wilt Management in Watermelon. *Plant Health Progress*, 21(1), 13–18.
- Miller, Nathan Forrest. (2017). Characterization of Fungicide Sensitivity and Analysis of Microsatellites for Population Studies of *Fusarium oxysporum* f. sp. *niveum* Causing Fusarium Wilt of Watermelon.
- Mirhendi, H., Makimura, K., de Hoog, G. S., Rezaei-Matehkolaei, A., Najafzadeh, M. J., Umeda, Y., & Ahmadi, B. (2015). Translation elongation factor 1- α gene as a potential taxonomic and identification marker in dermatophytes. *Medical Mycology*, 53(3), 215–224.
- Mishra, P. K., Fox, R. T. ., & Culham, A. (2003). Development of a PCR-based assay for rapid and reliable identification of pathogenic Fusaria. *FEMS Microbiology Letters*, 218(2), 329–332.
- Mohammadi, M., Aminipour, M., & Banhashemi, Z. (2004). Isozyme Analysis and Soluble Mycelial Protein Pattern in Iranian Isolates of Several formae speciales of *Fusarium oxysporum*. *Journal of Phytopathology*, 152(5), 267–276.
- Mokrini, F., Waeyenberge, L., Viaene, N., & Moens, M. (2012). First Report of the Cereal Cyst Nematode *Heterodera latipons* on Wheat in Morocco. *Plant Disease*, 96(5), 774-774.
- Mondal, S. N., Hossain, A., Hossain, A. E., Islam, M. A., & Bashar, M. A. (1994). Effect of various rootstocks in the graft culture of watermelon in Bangladesh. *Punjab Vegetable Grower*, 29, 15–19.
- Monfared, M. A., Samsampour, D., Sharifi-Sirchi, G. R., & Sadeghi, F. (2018). Assessment of genetic diversity in *Salvadora persica* L. based on inter simple sequence repeat (ISSR) genetic marker. *Journal of Genetic Engineering and*

Biotechnology, 16(2), 661–667.

- Moreno, B., Jacob, C., Rosales, M., Krarup, C., & Contreras, S. (2016). Yield and Quality of Grafted Watermelon Grown in a Field Naturally Infested with Fusarium Wilt. *HortTechnology*, 26(4), 453–459.
- Mujaju, C., Zborowska, A., Werlemark, G., Garkava-Gustavsson, L., Andersen, S. B., & Nybom, H. (2011). Genetic diversity among and within watermelon (*Citrullus lanatus*) landraces in Southern Africa. *The Journal of Horticultural Science and Biotechnology*, 86(4), 353–358.
- Munisse, P., Andersen, S. B., Jensen, B. D., & Christiansen, J. L. (2011). Diversity of landraces, agricultural practises and traditional uses of watermelon (*Citrullus lanatus*) in Mozambique. *African Journal of Plant Science*, 5(2), 75–86.
- Munisse, P., Jensen, B. D., & Andersen, S. B. (2013). Genetic differentiation of watermelon landraces in Mozambique using microsatellite markers. *African Journal of Biotechnology*, 12(36).
- Mwang`Ombe, A. W., Thiong`O, G., Olubayo, F. M., & Kiprop, E. K. (2007). DNA Microsatellite Analysis of Kenyan Isolates of *Rhizoctonia solani* from Common Bean (*Phaseolus vulgaris* L.). *Plant Pathology Journal*, 6(1), 66–71.
- Nasr Esfahani, M. (2018). Genetic and virulence variation in *Fusarium oxysporum* f. sp. *cepae* causing root and basal rot of common onion in Iran. *Journal of Phytopathology*, 166(7–8), 572–580.
- Naz, A., Butt, M. S., Sultan, M. T., Qayyum, M. M. N., & Niaz, R. S. (2014). Watermelon lycopene and allied health claims. *EXCLI Journal*, 13, 650.
- Nelson, P E, Tousson, T. A., & Marasas, W. F. O. (1983). *Fusarium species: An illustrated Manual for identification, USA: The Penn. State University Press, University Park.*
- Nelson, Paul E, Juba, J. H., Ross, P. F., & Rice, L. G. (1994). Fumonisin Production by Fusarium Species on Solid Substrates. *Journal of AOAC International*, 77(2), 522–525.
- Nitschke, E., Nihlgard, M., & Varrelmann, M. (2009). Differentiation of eleven *Fusarium* spp. isolated from sugar beet, using restriction fragment analysis of a polymerase chain reaction–amplified translation elongation factor 1 α gene fragment. *Phytopathology*, 99(8), 921–929.
- Niu, J., Arentshorst, M., Seelinger, F., Ram, A. F. J., & Ouedraogo, J. P. (2016). A set of isogenic auxotrophic strains for constructing multiple gene deletion mutants and parasexual crossings in *Aspergillus niger*. *Archives of Microbiology*, 198(9), 861–868.
- Nourollahi, Kh, & Aliaran, A. (2017). Genetic structure of *Fusarium oxysporum* f. sp. *ciceri* populations from chickpea in Ilam province, Iran. *Mycologia Iranica*, 4(2), 93–102.

- Nourollahi, K., & Madahjalali, M. (2017). Analysis of population genetic structure of Iranian *Fusarium oxysporum* f. sp. *lentis* isolates using microsatellite markers. *Australasian Plant Pathology*, 46(1), 35–42.
- O'Donnell, K., Kistler, H. C., Cigelnik, E., & Ploetz, R. C. (1998). Multiple evolutionary origins of the fungus causing Panama disease of banana: concordant evidence from nuclear and mitochondrial gene genealogies. *Proceedings of the National Academy of Sciences*, 95(5), 2044–2049.
- O'Donnell, K., Sutton, D. A., Fothergill, A., McCarthy, D., Rinaldi, M. G., Brandt, M. E., Zhang, N., & Geiser, D. M. (2008). Molecular Phylogenetic Diversity, Multilocus Haplotype Nomenclature, and In Vitro Antifungal Resistance within the *Fusarium solani* Species Complex. *Journal of Clinical Microbiology*, 46(8), 2477–2490.
- O'Donnell, K., Sutton, D. A., Rinaldi, M. G., Sarver, B. A. J., Balajee, S. A., Schroers, H.-J., Summerbell, R. C., Robert, V. A. R. G., Crous, P. W., & Zhang, N. (2010). Internet-accessible DNA sequence database for identifying fusaria from human and animal infections. *Journal of Clinical Microbiology*, 48(10), 3708–3718.
- O'Donnell, K., Ward, T. J., Robert, V. A. R. G., Crous, P. W., Geiser, D. M., & Kang, S. (2015). DNA sequence-based identification of *Fusarium*: Current status and future directions. *Phytoparasitica*, 43(5), 583–595.
- Oda, J. L. M., & Lee, M. (2003). Grafting of herbaceous vegetable and ornamental crops. *Horticultural Reviews*, 28, 61–124.
- Ohara, T., & Tsuge, T. (2004). FoSTUA, encoding a basic helix-loop-helix protein, differentially regulates development of three kinds of asexual spores, macroconidia, microconidia, and chlamydospores, in the fungal plant pathogen *Fusarium oxysporum*. *Eukaryotic Cell*, 3(6), 1412–1422.
- Olivain, C., & Alabouvette, C. (1999). Process of tomato root colonization by a pathogenic strain of *Fusarium oxysporum* f. sp. *lycopersici* in comparison with a non-pathogenic strain. *New Phytologist*, 141(3), 497–510.
- Pakshir, K., Farazmand, F., Ghasemi, F., Mirhend, H., Zomorodian, K., Kharazi, M., Alborzi Pour, R., Golestani, H., & Motamedi, M. (2020). Translation elongation factor 1-alpha gene as a marker for diagnosing of *Candida onychomycosis*. *Current Medical Mycology*.
- Paris, H. S. (2015). Origin and emergence of the sweet dessert watermelon, *Citrullus lanatus*. *Annals of Botany*, 116(2), 133–148.
- Paris, H. S., Daunay, M.C., & Janick, J. (2013). Medieval iconography of watermelons in Mediterranean Europe. *Annals of Botany*, 112(5), 867–879.
- Park, Y., & Cho, S. (2012). Watermelon production and breeding in South Korea. *Israel Journal of Plant Sciences*, 60(4), 415–423.
- Pasquali, M., Marena, L., Fiora, E., Piatti, P., Gullino, M. L., & Garibaldi, A. (2004).

- Real-time PCR for the identification of a highly pathogenic group of *Fusarium oxysporum* f. sp. *chrysanthemi* on *Argyranthemum frutescens* L. *Journal of Plant Pathology*, 86(1), 51–57.
- Peakall, R. O. D. and, & Smouse, P. E. (2006). GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Molecular Ecology Notes*, 6(1), 288–295.
- Petkar, A., Harris-Shultz, K., Wang, H., Brewer, M. T., Sumabat, L., & Ji, P. (2019). Genetic and phenotypic diversity of *Fusarium oxysporum* f. sp. *niveum* populations from watermelon in the southeastern United States. *PLoS one*, 14(7), e0219821.
- Petkar, A., Langston, D. B., Buck, J. W., Stevenson, K. L., & Ji, P. (2017). Sensitivity of *Fusarium oxysporum* f. sp. *niveum* to Prothioconazole and Thiophanate-Methyl and Gene Mutation Conferring Resistance to Thiophanate-Methyl. *Plant Disease*, 101(2), 366–371.
- Petrov, N. B., Vladychenskaya, I. P., Drozdov, A. L., & Kedrova, O. S. (2016). Molecular genetic markers of intra-and interspecific divergence within starfish and sea urchins (Echinodermata). *Biochemistry (Moscow)*, 81(9), 972–980.
- Pietro, A. Di, Madrid, M. P., Caracuel, Z., Delgado-Jarana, J., & Roncero, M. I. G. (2003). *Fusarium oxysporum*: exploring the molecular arsenal of a vascular wilt fungus. *Molecular Plant Pathology*, 4(5), 315–325.
- Pinaria, A. G., Laurence, M. H., Burgess, L. W., & Liew, E. C. Y. (2015). Phylogeny and origin of *Fusarium oxysporum* f. sp. *vanillae* in Indonesia. *Plant Pathology*, 64(6), 1358–1365.
- Powell, W., Morgante, M., Andre, C., Hanafey, M., Vogel, J., Tingey, S., & Rafalski, A. (1996). The comparison of RFLP, RAPD, AFLP and SSR (microsatellite) markers for germplasm analysis. *Molecular Breeding*, 2(3), 225–238.
- Prados-Ligero, A. M., Basallote-Ureba, M. J., López-Herrera, C. J., & Melero-Vara, J. M. (2007). Evaluation of Susceptibility of Carnation Cultivars to Fusarium Wilt and Determination of *Fusarium oxysporum* fsp. *dianthi* Races in Southwest Spain. *HortScience*, 42(3), 596–599.
- Quesada-Ocampo, L. (2018). Fusarium Wilt of Watermelon. *NC State Extension: Raleigh, NC, USA*,.
- Rabbany, G. M., Airin Rahman, Sharmin Afrin and Fazlul Haque. 2013. An analysis of cost of production of watermelon and profitability at Gopalganj district in Bangladesh. *European Journal of Banking and Finance*, 10, 2013.
- Rahman, M. Z., Kibria, M. G., Talukder, M. M. R., Akhter, M. S. and Amin, M. F. 2019. Evaluation of fungicides for control of gummy stem blight of watermelon caused by *Didymella bryoniae*. *Bangladesh Journal of Plant Pathology*, 35(1&2), 47-52.

- Rahman, M. Z., Ahmad, K., Siddiqui, Y., Saad, N., Hun, T. G., Mohd Hata, E., Rashed, O., Hossain, M. I., & Kutawa, A. B. (2021a). First Report of *Fusarium* wilt disease on Watermelon Caused by *Fusarium oxysporum* f. sp. *niveum* (FON) in Malaysia. *Plant Disease*, 105(12), 4169.
- Rahman, M. Z., Ahmad, K., Siddiqui, Y., Saad, N., Hun, T. G., Mohd Hata, E., Rashed, O., & Hossain, M. I. (2021). First Report of *Fusarium equiseti*, Causing Fruit Rot Disease of Watermelon in Malaysia. *Plant Disease*, 106(1), 326.
- Rahman, M. Z., Ahmad, K., Bashir Kutawa, A., Siddiqui, Y., Saad, N., Geok Hun, T., Hata, E. M., & Hossain, M. I. (2021c). Biology, Diversity, Detection and Management of *Fusarium oxysporum* f. sp. *niveum* Causing Vascular Wilt Disease of Watermelon (*Citrullus lanatus*): A Review. *Agronomy*, 11(7), 1310.
- Ramdial, H., Latchoo, R. K., Hosein, F. N., & Rampersad, S. N. (2017). Phylogeny and Haplotype Analysis of Fungi Within the *Fusarium incarnatum-equiseti* Species Complex. *Phytopathology*, 107(1), 109–120.
- Raveau, R., Fontaine, J., & Lounès-Hadj Sahraoui, A. (2020). Essential Oils as Potential Alternative Biocontrol Products against Plant Pathogens and Weeds: A Review. *Foods*, 9(3), 365.
- Rebib, H., Bouraoui, H., Rouaissi, M., Brygoo, Y., Boudabbous, A., Hajlaoui, M. R., & Sadfi-Zouaoui, N. (2014). Genetic diversity assessed by SSR markers and chemotyping of *Fusarium culmorum* causal agent of foot and root rot of wheat collected from two different fields in Tunisia. *European Journal of Plant Pathology*, 139(3), 481–495.
- Reddy, A. C., Lavanya, B., Tejaswi, T., Sreenivasa Rao, E., & Lakshmana Reddy, D. C. (2019). Isolation and Characterization of NBS-Encoding Disease Resistance Gene Analogs in Watermelon against *Fusarium* Wilt. *Current Science*, 117(4), 617.
- Ren, L., Su, S., Yang, X., Xu, Y., Huang, Q., & Shen, Q. (2008). Intercropping with aerobic rice suppressed *Fusarium* wilt in watermelon. *Soil Biology and Biochemistry*, 40(3), 834–844.
- Ren, Y., Di Jiao, Gong, G., Zhang, H., Guo, S., Zhang, J., & Xu, Y. (2015). Genetic analysis and chromosome mapping of resistance to *Fusarium oxysporum* f. sp. *niveum* (FON) race 1 and race 2 in watermelon (*Citrullus lanatus* L.). *Molecular Breeding*, 35(9), 183.
- Rep, M., Van Der Does, H. C., Meijer, M., Van Wijk, R., Houterman, P. M., Dekker, H. L., De Koster, C. G., & Cornelissen, B. J. C. (2004). A small, cysteine-rich protein secreted by *Fusarium oxysporum* during colonization of xylem vessels is required for I-3-mediated resistance in tomato. *Molecular Microbiology*, 53(5), 1373–1383.
- Robinson, J. (2005). Desertification and disarray: the threats to plant genetic resources of southern Darfur, western Sudan. *Plant Genetic Resources*, 3(1), 3–11.

- Robinson, R. W., & Decker-Walters, D. S. (1999). Cucurbits. CAB International, Wallingford. *Oxon (GB)*. Pp, 226.
- Rohlf, F. J. (1998). *NTSYSpc numerical taxonomy and multivariate analysis system version 2.0 user guide*.
- Romdhane, M. Ben, Haddar, A., Ghazala, I., Jeddou, K. Ben, Helbert, C. B., & Ellouz-Chaabouni, S. (2017). Optimization of polysaccharides extraction from watermelon rinds: Structure, functional and biological activities. *Food Chemistry*, 216, 355–364.
- Rouxel, M., Papura, D., Nogueira, M., Machefer, V., Dezette, D., Richard-Cervera, S., Carrere, S., Mestre, P., & Delmotte, F. (2012). Microsatellite markers for characterization of native and introduced populations of *Plasmopara viticola*, the causal agent of grapevine downy mildew. *Applied and Environmental Microbiology*, 78(17), 6337–6340.
- Rozhan Abu Dardak. (2019). Trends in Production, Trade, and Consumption of Tropical Fruit in Malaysia. *FFTC Agricultural Policy Platform (FFTC-AP)*. <https://ap.fftc.org.tw/article/1381>. Accessed 19 December 2021.
- Rusli, M. H. (2017). Disease epidemiology and genetic diversity of *Fusarium oxysporum* f. sp. *elaeidis*, Cause of Fusarium wilt of Oil Palm (*Elaeis guineensis* Jacq.). *Journal of Oil Palm Research*.
- Saikia, R., & Kadoo, N. (2010). Molecular detection and identification of *Fusarium oxysporum*. In *Molecular identification of fungi* (pp. 131–157). Springer.
- Saleh, A. A., Sharafaddin, A. H., El_Komy, M. H., Ibrahim, Y. E., & Hamad, Y. K. (2021). Molecular and physiological characterization of *Fusarium* strains associated with different diseases in date palm. *Plos one*, 16(7), 0254170.
- Sakata, Y., Ohara, T., & Sugiyama, M. (2007). The history and present state of the grafting of cucurbitaceous vegetables in japan. *Acta Horticulturae*, 731, 159–170.
- Sanders, I. R. (2002). Ecology and evolution of multigenomic arbuscular mycorrhizal fungi. *The American Naturalist*, 160(S4), S128–S141.
- Santos, G. R. dos, Leão, E. U., Gonçalves, C. G., & Cardon, C. H. (2013). Manejo da adubação potássica e da irrigação no progresso de doenças fúngicas e produtividade da melancia. *Horticultura Brasileira*, 31(1), 36–44.
- Sasseron, G. R., Benchimol-Reis, L. L., Perseguini, J. M. K. C., Paulino, J. F. C., Bajay, M. M., Carbonell, S. A. M., & Chiorato, A. F. (2020). *Fusarium oxysporum* f. sp. *phaseoli* genetic variability assessed by new developed microsatellites. *Genetics and Molecular Biology*, 43(2).
- Schmidt, S. M., Houterman, P. M., Schreiver, I., Ma, L., Amyotte, S., Chellappan, B., Boeren, S., Takken, F. L. W., & Rep, M. (2013). MITEs in the promoters of effector genes allow prediction of novel virulence genes in *Fusarium oxysporum*. *BMC Genomics*, 14(1), 119.

- Servin, A., Elmer, W., Mukherjee, A., De la Torre-Roche, R., Hamdi, H., White, J. C., Bindraban, P., & Dimkpa, C. (2015). A review of the use of engineered nanomaterials to suppress plant disease and enhance crop yield. *Journal of Nanoparticle Research*, 17(2), 92.
- Shahzad, R., Khan, A. L., Bilal, S., Asaf, S., & Lee, I.J. (2017). Plant growth-promoting endophytic bacteria versus pathogenic infections: an example of *Bacillus amyloliquefaciens* RWL-1 and *Fusarium oxysporum* f. sp. *lycopersici* in tomato. *PeerJ*, 5, 3107.
- Sharma, M., Gupta, S. K., & Sharma, T. R. (2005). Characterization of Variability in *Rhizoctonia solani* by Using Morphological and Molecular Markers. *Journal of Phytopathology*, 153(7–8), 449–456.
- Shi, L., Du, N., Shu, S., Sun, J., Li, S., & Guo, S. (2017). *Paenibacillus polymyxa* NSY50 suppresses *Fusarium* wilt in cucumbers by regulating the rhizospheric microbial community. *Scientific Reports*, 7(1), 41234.
- Shirzadi, T., & Nourollahi, K. (2020). Genetic structure analysis of *Fusarium oxysporum* f. sp. *phaseoli* populations on common bean by using SSR markers. *Mycologia Iranica*, 7(1), 105–114.
- Shlevin, E., Gamliel, A., Katan, J., & Shtienberg, D. (2018). Multi-study analysis of the added benefits of combining soil solarization with fumigants or non-chemical measures. *Crop Protection*, 111, 58–65.
- Silva, D. N., Talhinhos, P., Cai, L., Manuel, L., Gichuru, E. K., Lourejro, A., Varzea, V., Paulo, O. S., & Batista, D. (2012). Host-jump drives rapid and recent ecological speciation of the emergent fungal pathogen *Colletotrichum kahawae*. *Molecular Ecology*, 21(11), 2655–2670.
- Silva, F. P. da, Vechiato, M. H., & Harakava, R. (2014). EF-1 α gene and *IGS* rDNA sequencing of *Fusarium oxysporum* f. sp. *vasinfectum* and *F. oxysporum* f. sp. *phaseoli* reveals polyphyletic origin of strains. *Tropical Plant Pathology*, 39(1), 64–73.
- Singh, N., Pal, A. K., Roy, R. K., Tamta, S., & Rana, T. S. (2017). Development of cpSSR markers for analysis of genetic diversity in *Gladiolus* cultivars. *Plant Gene*, 10, 31–36.
- Skovgaard, K., Bødker, L., & Rosendahl, S. (2002). Population structure and pathogenicity of members of the *Fusarium oxysporum* complex isolated from soil and root necrosis of pea (*Pisum sativum* L.). *FEMS Microbiology Ecology*, 42(3), 367–374.
- Skovgaard, K., Nirenberg, H. I., O'Donnell, K., & Rosendahl, S. (2001). Evolution of *Fusarium oxysporum* f. sp. *vasinfectum* Races Inferred from Multigene Genealogies. *Phytopathology*, 91(12), 1231–1237.
- Smith, E. F. (1894). The watermelon disease of the South. *Proceedings of the American Association for the Advancement Science*, 43, 289–290.

- Smolinska, U. (2000). Survival of *Sclerotium cepivorum* Sclerotia and *Fusarium oxysporum* Chlamydo spores in Soil Amended with Cruciferous Residues. *Journal of Phytopathology*, 148(6), 343–349.
- Srinivasan, K., Spadaro, D., Poli, A., Gilardi, G., Gullino, M. L., & Garibaldi, A. (2012). Genetic diversity and pathogenicity of *Fusarium oxysporum* isolated from wilted rocket plants in Italy. *Phytoparasitica*, 40(2), 157–170.
- Subburaj, S., Lee, K., Jeon, Y., Tu, L., Son, G., Choi, S., Lim, Y.-P., McGregor, C., & Lee, G.-J. (2019). Whole genome resequencing of watermelons to identify single nucleotide polymorphisms related to flesh color and lycopene content. *PloS one*, 14(10), 0223441.
- Summerell, B. A., Salleh, B., & Leslie, J. F. (2003). A utilitarian approach to *Fusarium* identification. *Plant Disease*, 87(2), 117–128.
- Sun, S.-K., & Huang, J. W. (1985). Mechanisms of control of *Fusarium* wilt diseases by amendment of soil with SH mixture. *Plant Protection Bulletin, Taiwan*, 27(3), 159–169.
- Szecsí, A., & Dobrovolszky, A. (1985). Genetic distance in fungus genus *Fusarium* measured by comparative computer analysis of DNA thermal denaturation profiles. *Mycopathologia*, 89(2), 95–100.
- Tautz, D. (1989). Hypervariability of simple sequences as a general source for polymorphic DNA markers. *Nucleic Acids Research*, 17(16), 6463–6471.
- Thanassouloupoulos, C. C., Giannopolitis, C. N., & Kitsos, G. T. (1970). Vol. 54, No. 7. Plant Disease Reporter-" July 1970 561 Control of *Fusarium* wilt of tomato and watermelon with benomyl. *The Plant Disease Reporter*, 54, 561.
- Tran-Nguyen, L. T. T., Condé, B. D., Smith, S. H., & Ulyatt, L. I. (2013). Outbreak of *Fusarium* wilt in seedless watermelon seedlings in the Northern Territory, Australia. *Australasian Plant Disease Notes*, 8(1), 5–8.
- Umesha, S., Jyothi, N., & Roohie, R. K. (2015). Detection of bacterial and *Fusarium* wilt pathogens in cabbage by multiplex PCR. *Journal of Plant Science*, 3, 185–190.
- van Dam, P., Fokkens, L., Schmidt, S. M., Linmans, J. H. J., Kistler, H. C., Ma, L.-J., & Rep, M. (2016). Effector profiles distinguish formae speciales of *Fusarium oxysporum*. *Environmental Microbiology*, 18(11), 4087–4102.
- van der Does, H. C., Duyvesteyn, R. G. E., Goltstein, P. M., van Schie, C. C. N., Manders, E. M. M., Cornelissen, B. J. C., & Rep, M. (2008). Expression of effector gene SIX1 of *Fusarium oxysporum* requires living plant cells. *Fungal Genetics and Biology*, 45(9), 1257–1264.
- Watanabe, M., Yonezawa, T., Lee, K., Kumagai, S., Sugita-Konishi, Y., Goto, K., & Hara-Kudo, Y. (2011). Evaluation of genetic markers for identifying isolates of the species of the genus *Fusarium*. *Journal of the Science of Food and*

Agriculture, 91(13), 2500–2504.

- Wechter, W. P., Kousik, C., McMillan, M., & Levi, A. (2012). Identification of Resistance to *Fusarium oxysporum* f. sp. *niveum* Race 2 in *Citrullus lanatus* var. *citroides* Plant Introductions. *HortScience*, 47(3), 334–338.
- Wehner et al. (2019). *Cucurbits*. CAB International.
- Wehner, T. C. (2008). Watermelon. In *Vegetables I* (pp. 381–418). Springer.
- Weiberg, A., Wang, M., Lin, F.-M., Zhao, H., Zhang, Z., Kaloshian, I., Huang, H.-D., & Jin, H. (2013). Fungal Small RNAs Suppress Plant Immunity by Hijacking Host RNA Interference Pathways. *Science*, 342(6154), 118–123.
- Williams PH, & Palmer MJ. (1996). Fusarium wilt. NC state watermelon disease handbook. 1996. Available from: <https://content.ces.ncsu.edu/fusarium-wilt-of-watermelon>. Accessed 20 February 2022.
- Wingfield, M. J., de Beer, Z. W., Slippers, B., Wingfield, B. D., Groenewald, J. Z., Lombard, L., & Crous, P. W. (2012). One fungus, one name promotes progressive plant pathology. *Molecular Plant Pathology*, 13(6), 604–613.
- Wu, H-S, Gao, Z., Zhou, X., Shi, X., Wang, M., Shang, X., Liu, Y., Gu, D., & Wang, W. (2013). Microbial dynamics and natural remediation patterns of *F. usarium* infested watermelon soil under 3-yr of continuous fallow condition. *Soil Use and Management*, 29(2), 220–229.
- Wu, Hong-sheng, Yang, X., Fan, J., Miao, W., Ling, N., Xu, Y., Huang, Q., & Shen, Q. (2009). Suppression of Fusarium wilt of watermelon by a bio-organic fertilizer containing combinations of antagonistic microorganisms. *BioControl*, 54(2), 287–300.
- Xia, J. W., Sandoval-Denis, M., Crous, P. W., Zhang, X. G., & Lombard, L. (2019). Numbers to names - restyling the *Fusarium incarnatum-equiseti* species complex. *Persoonia-Molecular Phylogeny and Evolution of Fungi*, 43(1), 186–221.
- Xie, X.G., Huang, C.Y., Cai, Z.D., Chen, Y., & Dai, C.C. (2019). Targeted Acquisition of *Fusarium oxysporum* f. sp. *niveum* Toxin-Deficient Mutant and Its Effects on Watermelon Fusarium Wilt. *Journal of Agricultural and Food Chemistry*, 67(31), 8536–8547.
- Xiong, W., & Zhan, A. (2018). Testing clustering strategies for metabarcoding-based investigation of community-environment interactions. *Molecular Ecology Resources*, 18(6), 1326–1338.
- Xu, W., Wang, H., Lv, Z., Shi, Y., & Wang, Z. (2019). Antifungal activity and functional components of cell-free supernatant from *Bacillus amyloliquefaciens* LZN01 inhibit *Fusarium oxysporum* f. sp. *niveum* growth. *Biotechnology & Biotechnological Equipment*, 33(1), 1042–1052.

- Yang, N., Ma, G., Chen, K., & Wu, X. (2018). The Population Genetics of *Alternaria tenuissima* in Four Regions of China as Determined by Microsatellite Markers Obtained by Transcriptome Sequencing. *Frontiers in Microbiology*, 9, 2904.
- Yeh, F. C. (1997). Population genetic analysis of co-dominant and dominant marker and quantitative traits. *Belgium Journal of Botany*, 130, 129–157.
- Yetisir, H., & Sari, N. (2003). Effect of different rootstock on plant growth, yield and quality of watermelon. *Australian Journal of Experimental Agriculture*, 43(10), 1269.
- Yu, H., Chen, S., Zhang, X., Zhou, X., & Wu, F. (2019). Rhizosphere bacterial community in watermelon-wheat intercropping was more stable than in watermelon monoculture system under *Fusarium oxysporum* f. sp. *niveum* invasion. *Plant and Soil*, 445(1–2), 369–381.
- Zang, R., Zhao, Y., Guo, K., Hong, K., Xi, H., & Wen, C. (2018). The Population Genetic Variation Analysis of Bitter Gourd Wilt Caused by *Fusarium oxysporum* f. sp. *momordicae* in China by Inter Simple Sequence Repeats (ISSR) Molecular Marker. *BioRxiv*, 424077.
- Zarrin, M., Ganj, F., & Faramarzi, S. (2016). Development of a polymerase chain reaction-restriction fragment length polymorphism method for identification of the *Fusarium* genus using the transcription elongation factor-1 α gene. *Biomedical Reports*, 5(6), 705–708.
- Zhang, J., Wang, P., Tian, H., Wang, Y., & Jiang, H. (2019). Using a new hybrid rootstock significantly increases the grafted plant rate and watermelon yield. *International Agrophysics*, 33(1).
- Zhang, M., Xu, J. H., Liu, G., Yao, X. F., Li, P. F., & Yang, X. P. (2015). Characterization of the watermelon seedling infection process by *Fusarium oxysporum* f.sp. *niveum*. *Plant Pathology*, 64(5), 1076–1084.
- Zhang, Y., Yu, L., Yung, K.-F., Leung, D. Y. C., Sun, F., & Lim, B. L. (2012). Over-expression of AtPAP2 in *Camelina sativa* leads to faster plant growth and higher seed yield. *Biotechnology for Biofuels*, 5(1), 19.
- Zhang, Z., Zhang, J., Wang, Y., & Zheng, X. (2005). Molecular detection of *Fusarium oxysporum* f. sp. *niveum* and *Mycosphaerella melonis* in infected plant tissues and soil. *FEMS Microbiology Letters*, 249(1), 39–47.
- Zheng, G. F. (1995). A fine watermelon stock, ChaoFeng F1, and its cultivation. *Henan Nongye Kexue*, 3, 24–25.
- Zhou, X. G., & Everts, K. L. (2003). Races and Inoculum Density of *Fusarium oxysporum* f. sp. *niveum* in Commercial Watermelon Fields in Maryland and Delaware. *Plant Disease*, 87(6), 692–698.
- Zhou, X. G., & Everts, K. L. (2004a). Quantification of Root and Stem Colonization of Watermelon by *Fusarium oxysporum* f. sp. *niveum* and Its Use in Evaluating

- Resistance. *Phytopathology*, 94(8), 832–841.
- Zhou, X. G., & Everts, K. L. (2004b). Suppression of Fusarium Wilt of Watermelon by Soil Amendment with Hairy Vetch. *Plant Disease*, 88(12), 1357–1365.
- Zhou, X. G., & Everts, K. L. (2006). Suppression of Fusarium Wilt of Watermelon Enhanced by Hairy Vetch Green Manure and Partial Cultivar Resistance. *Plant Health Progress*, 7(1), 23.
- Zhou, X. G., Everts, K. L., & Bruton, B. D. (2010). Race 3, a New and Highly Virulent Race of *Fusarium oxysporum* f. sp. *niveum* Causing Fusarium Wilt in Watermelon. *Plant Disease*, 94(1), 92–98.
- Zhu, H., Song, P., Koo, D.H., Guo, L., Li, Y., Sun, S., Weng, Y., & Yang, L. (2016). Genome wide characterization of simple sequence repeats in watermelon genome and their application in comparative mapping and genetic diversity analysis. *BMC Genomics*, 17(1), 557.
- Zhu, Z., Zheng, L., Pan, L., Hsiang, T., & Huang, J. (2014). Identification and characterization of Fusarium species associated with wilt of *Eleocharis dulcis* (Chinese water chestnut) in China. *Plant Disease*, 98(7), 977–987.
- Zitter, T. A., Hopkins, D. L., & Thomas, C. E. (1996). *Compendium of cucurbit diseases* (Issue 635.6 632). *American Phytopathological Society*.
- Zohary, D., Hopf, M., & Weiss, E. (2012). *Domestication of Plants in the Old World*. Oxford University Press.
- Zvirin, T., Herman, R., Brotman, Y., Denisov, Y., Belausov, E., Freeman, S., & Perl-Treves, R. (2010). Differential colonization and defence responses of resistant and susceptible melon lines infected by *Fusarium oxysporum* race 1-2. *Plant Pathology*, 59(3), 576–585.