



**PHYTOCHEMICALS AND ANTIOXIDANT ACTIVITIES OF LEAF, NODE
AND *IN VITRO*-INDUCED CALLUS OF *Bougainvillea glabra* Choisy
USING DIFFERENT SOLVENTS**

By

NASRAT MOHAMMAD NASIM

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

June 2022

FP 2022 57

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

**This thesis is dedicated to my family
Malika Rahimi, Najla Nasrat, Mohammad Emran Nasrat and Viana Nasrat
and
My respected mentors
Dr. Mohd Hakiman Mansor
Prof. Madya Dr. Siti Zaharah Sakimin**

**Thank you for your endless patience, encouragements, advices,
sacrifices and motivations throughout my master degree journey**



Abstract of the thesis presented to the senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

PHYTOCHEMICALS AND ANTIOXIDANT ACTIVITIES OF LEAF, NODE AND *IN VITRO*-INDUCED CALLUS OF *Bougainvillea glabra* Choisy USING DIFFERENT SOLVENTS

By

NASRAT MOHAMMAD NASIM

June 2022

Chairman : Mohd Hakim Mansor, PhD
Faculty : Agriculture

Bougainvillea, popularly known as 'Bunga kertas' in Malaysia, is thoroughly explored for nutritional and medicinal purposes. *Bougainvillea* has been shown to possess alkaloids, flavonoids, cardiac glycosides, saponins, and beta-cyanins, which are widely used in folk medicine to treat different illnesses. Despite its major conventional therapeutic importance, only limited attempts have been made to investigate this species' chemical and pharmacological properties in relation to its medicinal uses. Therefore, this study was conducted to determine the effect of *in vitro* induced callus under different light conditions and plant growth regulators on phytochemical and antioxidant activities using different extraction solvents. In this study, the leaves and nodes were collected and dried in an oven at a temperature of 40 °C for 48 hours until the weight remained constant. The dried materials made into fine powder using mortar and pestle, then different solvents (aqueous, ethanol, acetone, and hexane) were used for extraction purposes. Subsequently, the phytochemicals (total phenolic and total flavonoid contents), and antioxidant activities such as 2,2-diphenyl-1-picrylhydrazyl (DPPH), 2,2'-azinol-bis (3-ethylbenzothiazoline-6-sulfonic acid (ABTS) and iron (II) chelating activity were assayed using a spectrophotometer. For the second experiment, leaves and nodes of the plants were used for *in vitro* surface sterilization. Different concentrations of Clorox® (10, 20, 30 and 40%) and exposure time (10, 15, and 20 minutes) were applied. For the third experiment, the sterilized nodal segments were used for *in vitro* callus induction. Different concentrations and combinations of 2,4-D (2.5, 5, and 7.5 µM) and BAP (0.5, 1, and 1.5 µM) were used for callus induction purposes under dark and light conditions. Finally, the phytochemicals screening and antioxidant activities of *in vitro*-derived calluses and conventional propagated donor plants were measured as per the first experiment to find out and compare the influence of callus induction on secondary metabolite production potential. The results from the first experiment showed that leaf segment extracted with aqueous extract had a significantly superior effect than other plant parts and solvents where it achieved the highest total phenolic content (58.04 mg GAE/g DW), total flavonoid content (127.93 mg RE/g DW), DPPH free radical scavenging (10.08 mg TE/g DW), ABTS scavenging

activity (1.59 mg TE/g DW), respectively. Based on the results of the callus induction experiment, the maximum days (18.25) to callus initiation recorded when nodal segment cultured on woody plant medium (WPM) supplemented with 7.5 μ M 2,4-D without addition cytokinin under light condition. On the contrary, the minimum days (7) to callus initiation were recorded when nodal treated with 2.5, 5 and 7.5 μ M 2,4-D + 1 and 1.5 μ M BAP under dark conditions, which showed significant differences between various concentrations and combinations of auxin and cytokinin. This study showed that the dark condition is better for callus induction than light condition.



Abstrak thesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan Ijazah Master Sains

**AKTIVITI FITOKIMIA DAN ANTIOKSIDAN DAUN, NOD DAN KALUS DARIPADA
IN VITRO *Bougainvillea glabra* Choisy DALAM
PELBAGAI PELARUT BERBEZA**

Oleh

NASRAT MOHAMMAD NASIM

Jun 2022

Pengerusi : Mohd Hakim Mansor, PhD
Fakulti : Pertanian

Bougainvillea, dikenali sebagai Bunga kertas di Malaysia, diterokai secara menyeluruh bagi tujuan pemakanan dan perubatan. *Bougainvillea* telah terbukti mempunyai alkaloid, flavonoid, glikosida, saponin dan beta-sianin, yang digunakan secara meluas dalam perubatan tradisional untuk merawat pelbagai penyakit. Walaupun terdapat kepentingan terapeutik yang banyak, hanya sedikit kajian telah dijalankan untuk mengetahui sifat kimia dan farmakologi spesies ini berhubung dengan kegunaan perubatannya. Oleh itu, kajian ini dijalankan untuk menentukan kesan induksi kalus secara *in vitro* di bawah keadaan cahaya yang berbeza dan pengawalatur tumbesaran tumbuhan terhadap aktiviti fitokimia dan antioksidan menggunakan pelarut yang berbeza. Dalam kajian ini, daun dan nod diambil dan dikeringkan di dalam ketuhar pada suhu 40 °C selama 48 jam atau sehingga tiada perubahan berat. Sampel kering dijadikan serbuk halus menggunakan mortar dan alu, kemudian pelarut yang berbeza (akuas, etanol, aseton, dan heksana) digunakan untuk pengekstrakan. Selepas itu, fitokimia (jumlah kandungan fenolik dan flavonoid), dan aktiviti antioksidan seperti 2,2-difenil-1-picrilhidrazil (DPPH), asid 2,2'-azinol-bis (3-etilbenzothiazolin-6-sulfonik (ABTS) dan aktiviti *chelation* besi (II) telah diukur menggunakan spektrofotometer. Pada eksperimen kedua, daun dan nod tumbuhan disterilkan permukaan secara *in vitro*. Kepekatan Clorox® yang berbeza (10, 20, 30 dan 40%) dan tempoh masa (10, 15 dan 20 minit) digunakan. Pada eksperimen ketiga, segmen nod yang steril digunakan untuk induksi kalus *in vitro*. Kepekatan dan kobinasi berbeza 2,4-D (2.5, 5 dan 7.5 µM) dan BAP (0.5, 1 dan 1.5 µM) telah digunakan untuk tujuan induksi kalus dalam keadaan gelap dan terang. Akhir sekali, penyaringan fitokimia dan aktiviti antioksidan bagi kalus daripada *in vitro* dan tumbuhan penderma yang dibiakkan secara konvensional diukur sebagai eksperimen pertama untuk mengetahui dan membandingkan pengaruh induksi kalus terhadap potensi pengeluaran metabolit sekunder. Hasil eksperimen pertama menunjukkan bahawa segmen daun yang diekstrak dengan pelarut akueus mempunyai kesan unggul yang ketara berbanding bahagian tumbuhan dan pelarut lain di mana masing-masing mencapai jumlah kandungan fenolik tertinggi (58.04 mg GAE/g DW), jumlah kandungan flavonoid (127.93 mg RE/g DW), DPPH penghapusan radikal bebas

(10.08 mg TE/g DW), ABTS aktiviti penipisan (1.59 mg TE/g DW). Hasil dari eksperimen induksi kalus, hari maksimum (18.25) bagi penghasilan kalus pertama diperhatikan apabila segmen nod dikultur dalam medium tumbuhan berkayu (WPM) ditambah dengan 7.5 μM 2,4-D tanpa penambahan sitokinin dalam keadaan terang. Sebaliknya, hari minimum (7) bagi penghasilan kalus pertama diperhatikan apabila nod dirawat dengan 2.5, 5 dan 7.5 μM 2,4-D + 1 dan 1.5 μM BAP dalam keadaan gelap, menunjukkan perbezaan ketara antara pelbagai kepekatan dan kombinasi auksin dan sitokinin dalam keadaan cahaya yang berbeza.



ACKNOWLEDGEMENTS

First, and most importantly, I am obligated to express gratitude to Allah, most gracious and most merciful, for the blessing granted to me to execute and complete the study. I am very indebted to those who have helped me prepare and conduct the study. I wish to express my deepest gratitude.

In my journey towards this degree, I found a supervisor, a working partner, and a mentor who gave his best to supervise my master's degree, Dr. Mohd Hakiman Mansor. Thank you for being the most understanding person and always with me through my thick and thin in completing this study. Thank you for your kindness, invaluable guidance, and encouragement. Despite my shortcomings in many ways, you have been patient, trusting, and given me many opportunities to improve.

I also want to give my warm appreciation to my committee members, Associate Professor Dr. Siti Zaharah Sakimin for her helpful suggestions, insights, and comments.

I would like to thank the various members of our lab mates with whom I had the opportunity to work, to mention a few Nur Haida Zainol, Nisar Ahmad Zahid, and Narmada. They provided a friendly and cooperative atmosphere at work and also useful guidance and insightful comments on my work. And also the staff of Faculty of Agriculture for their kind support and assistance towards the successful completion of this thesis.

Lastly, I would like to express my profound and heartfelt gratitude to my parents, wife, son and daughter, relatives and all other well-wishers for their interest and immense contributions in prayers, guidance, and moral support towards my success.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd Hakim Mansor, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Siti Zaharah Sakimin, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 10 November 2022

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Dr. Mohd Hakiman Mansor

Signature: _____
Name of Member
of Supervisory
Committee: Associate Prof. Dr. Siti Zaharah Sakimin

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
TABLE OF CONTENTS	x
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xvii
CHAPTER	
1 INTRODUCTION	1
1.1 Background of the study	1
1.2 Statement of the problem	2
1.4 Objectives of the study	2
2 LITERATURE REVIEW	3
2.1 Medicinal plants	3
2.2 <i>Bougainvillea glabra</i> Choisy	3
2.3 The medicinal uses of <i>B. glabra</i> Choisy	6
2.4 Concerns of <i>B. glabra</i> raw materials production	6
2.5 Plant tissue culture	7
2.5.1 Selection of explants	7
2.5.2 Surface sterilization of explants	8
2.5.3 Basal medium	9
2.5.4 Plant growth regulators	10
2.5.4.1 Auxin	11
2.5.4.2 Cytokinin	12
2.5.5 Carbon source	12
2.6 Callus induction	12
2.7 Plant secondary metabolites	15
2.7.1 Selection of plant part	17
2.7.2 Influence of extraction solvents on phytochemicals and antioxidant activities of plant	17
3 GENERAL MATERIALS AND METHODS	19
3.1 Introduction	19
3.2 Experimental location	19
3.3 Plant materials	19
3.4 Chemicals and reagents	19
3.5 Preparation of stock solutions for basal media	20
3.6 Preparation of plant growth regulators	20
3.7 Preparation of basal medium	20

3.9	Glassware	21
3.10	Preparation of aseptic conditions	21
3.11	Culture maintenance	21
3.12	Preparation of extract	22
	Quantification of phenolic content of <i>B. glabra</i> leaves, nodes and <i>in vitro</i> -induced calli	22
3.13	3.12.1 Total phenolic acids content	22
	3.12.2 Total flavonoids content	22
	Quantification of antioxidant properties of <i>B. glabra</i> leaves, nodes and <i>in vitro</i> -induced calli	22
	3.13.1 DPPH free radical scavenging activity	23
3.14	313.2 ABTS scavenging activity	23
	313.3 Iron (II) chelating activity	23
	Statistical analysis	23
4	PHYTOCHEMICAL SCREENING AND ANTIOXIDANT ACTIVITIES OF LEAVES AND NODES OF <i>B. glabra</i> Choisy USING DIFFERENT SOLVENTS	24
4.1	Introduction	24
4.2	Plant materials	25
4.3	Preparation of extract and quantification of phenolic content and antioxidant activities of leaf and nodes of <i>B. glabra</i>	25
4.4	Statistical analysis	25
4.5	Results and discussion	25
	4.5.1 Comparison of phenolic content and antioxidant properties of <i>B. glabra</i> leaves and nodes	25
	4.5.2 The effect of solvents on phenolic content and antioxidant activities of <i>B. glabra</i> leaves and nodes	26
	4.5.3 The synergistic effect of different solvents on total phenolic acids, and flavonoids contents in leaf and node of <i>B. glabra</i>	29
	4.5.4 The synergistic effect of different solvents on antioxidant properties of leaf and node of <i>B. glabra</i>	31
	4.5.5. Correlation analysis between variables	34
4.6	Conclusion	36

5	IN VITRO SURFACE STERILIZATION OF <i>Bougainvillea glabra</i> Choisy LEAF AND NODAL SEGMENT	37
5.1	Introduction	37
5.2	Materials and methods	37
	5.2.1 Effects of different concentration of Clorox® and exposure time on explant surface sterilization and culture	37
5.3	establishment	38
	Results and discussion	38
	5.3.1 The effects of Clorox® and exposure time on explant surface sterilization	41
	5.3.2 Synergistic effects of Clorox®, exposure time and	
5.4	explant types on surface sterilization	43
	Conclusion	
6	IN VITRO CALLUS INDUCTION OF <i>Bougainvillea glabra</i> Choisy NODAL SEGMENT UNDER DIFFERENT PGRs AND LIGHT CONDITIONS	44
6.1	Introduction	44
6.2	Materials and methods	44
	6.2.1 Plant materials	44
	6.2.2 Callus induction of <i>B. glabra</i> as affected by cytokinin and auxin	45
	6.2.3 Synergistic effect of cytokinin, auxin and light regime	45
	6.2.4 Culture maintenance	45
	6.2.5 Fresh and dry weight of callus	45
	6.2.6 Statistical analysis	46
6.3	Results and discussion	46
	6.3.1 Callus induction of <i>B. glabra</i>	46
	6.3.1.1 The effect of 2,4-D, BAP and light regimes on callus induction of <i>B. glabra</i>	46
	6.3.1.2 Synergistic effect of cytokinin, auxin and light regimes on callus induction	51
6.4	Conclusion	56
7	QUANTIFICATION OF PHENOLICS CONTENT AND ANTIOXIDANT PROPERTIES OF CONVENTIONALLY PROPAGATED NODAL PART AND IN VITRO-INDUCED CALLI OF <i>B. glabra</i> USING DIFFERENT EXTRACTION SOLVENTS	57
7.1	Introduction	57
7.2	Plant materials	58
7.3	Preparation of extract and quantification of phenolic content and antioxidant activities	58

	of node and <i>in vitro</i> -induced callus of <i>B. glabra</i>	
7.4	Results and discussion	58
7.4.1	The effect of different solvents on phenolic content and antioxidant activities of <i>B. glabra</i> nodes and <i>in vitro</i> -induced callus	58
7.4.2	The combination effect of different solvents on TPC and TFC in nodes and <i>in vitro</i> -induced callus of <i>B. glabra</i>	63
7.4.3	The combination effect of different solvents on DPPH, ABTS and iron (II) chelating activity in nodes and <i>in vitro</i> -induced callus of <i>B. glabra</i>	66
7.4.4	Correlation analysis between variables	70
7.5	Conclusion	72
8	CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	73
	REFERENCES	76
	APPENDICES	94
	BIODATA OF STUDENT	102
	LIST OF PUBLICATION	103

LIST OF TABLES

Table		Page
2.1	Taxonomy of <i>Bougainvillea glabra</i> Choisy	4
2.2	Effect of various plant growth regulators on callus induction of Nyctaginaceae plant species	14
4.1	The effect of different solvents on total phenolic acid, flavonoid contents, DPPH free radical scavenging activity, ABTS scavenging activity, and iron (II) chelating activity in leaf and node of <i>B. glabra</i>	27
4.2	The synergistic effect of different solvents on total phenolic acids, and flavonoids contents in leaf and node of <i>B. glabra</i>	30
4.3	The synergistic effect of different solvents on DPPH free radical scavenging activity and ABTS scavenging activity in leaf and node of <i>B. glabra</i>	32
4.4	The synergistic effect of different solvents on iron (II) chelating activity in leaf and node of <i>B. glabra</i>	34
4.5	Pearson's correlation analysis between variables	35
5.1	The main effects of different concentration of Clorox®, exposure time and explant types on aseptic cultures and explant survival of <i>B. glabra</i>	39
5.2	Synergistic effects of different concentrations of Clorox®, exposure time and explant types on aseptic culture and explants survival of <i>B. glabra</i>	42
6.1	The effects of 2,4-D and BAP on callus induction in the nodal segment of <i>B. glabra</i> under different light conditions	47
6.2	Interaction effects of 2,4-D and BAP on callus induction in the nodal segment of <i>B. glabra</i> under different light conditions	52
7.1	The effect of different solvents on TPC, TFC, DPPH, ABTS, and iron (II) chelating activity in node and <i>in vitro</i> -induced callus of <i>B. glabra</i>	60
7.2	The combination effect of different solvents on TPC and TFC in node and <i>in vitro</i> -induced callus of <i>B. glabra</i>	65

7.3	The combination effect of different solvents on DPPH free radical scavenging activity and ABTS scavenging activity in node and <i>in vitro</i> -induced callus of <i>B. glabra</i>	67
7.4	The combination effect of different solvents on iron (II) chelating activity in node and <i>in vitro</i> -induced callus of <i>B. glabra</i>	70
7.5	Pearson's correlation analysis between variables	71



LIST OF FIGURES

Figure		Page
2.1	a) <i>B. glabra</i> Choisy plant, b) <i>B. glabra</i> white flowers surrounded with pink bracts	5
5.1	a) leaf segment contaminated with fungal after 7 days on inoculation and b) nodal segments contaminated with fungal and bacteria after 7 days of inoculation.	40
6.1	<i>In vitro</i> callus induction under light incubation condition.	54
6.2	<i>In vitro</i> callus induction under dark incubation condition.	55

LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
UPM	Universiti Putra Malaysia
BAP	6-benzylaminopurine
2,4-D	2,4-dichlorophenoxyacetic acid
TPC	Total phenolic content
TFC	Total flavonoid Content
DPPH	2,2-diphenyl-1-picrylhydrazyl
ABTS	2,2'-azinol-bis (3-ethylbenzothiazoline-6-sulfonic acid)

CHAPTER 1

INTRODUCTION

1.1 Background of the study

The genus of *Bougainvillea* belongs to the family Nyctaginaceae, which is one of the utmost valuable ornamental and medicinal plants native from South America (Brazil, Peru, and northern Argentina). It is commonly used for many industries such as horticulture, medicinal, cosmetic, agriculture, and environmental in tropical and subtropical regions due to broad stability in the various agro-climate areas. The name derives from the French navigator Louis Antoine de Bougainville, who was the first to state this plant in Brazil in 1786 (Naito et al., 2020; Kumar et al., 2020).

Medicinal plants or herbs are enriched in various phytochemicals, which have been shown to have various biological effects and should be researched further (Saleem et al., 2020). One of the medicinal plants that has currently expanded much attention because of its therapeutic constituents is *Bougainvillea glabra* Choisy, locally known as 'Bunga kertas' in Malaysia. Since 1970, the chemical components of the *Bougainvillea* genus have been extensively studied. Phytochemical studies were carried out to classify various bioactive compounds from the extracts of different polarity solvents from buds, leaves, or bracts with or without flowers, bark stems, and roots of the species (Abarca-Vargas and Petricevich, 2018). The type of solvents used in the extraction technique has a big impact on the success of determining biologically active compounds from plant material. Extraction is essential in phytochemical isolation for detecting pharmacologically active constituents in plant parts (Thomas et al., 2020). Selecting a suitable solvent system for extraction is crucial for standardization of pharmaceutical products. It isolates the essential compounds in medicinal plants while excluding the unwanted matrix (Kumarasamy and Selvi, 2020). *Bougainvillea* is mainly grown in tropical and subtropical climates, and there is a lack of raw material in temperate weather to treat the diseases. In such places, this plant is propagated in the greenhouse to protect from cool weather and can produce raw material, but by this method, raw material production is low. A massive amount of raw materials is needed to extract the high amount of secondary metabolites. This problem can be overcome by producing the secondary metabolites in the laboratory by plant tissue culture technique (Haida et al., 2020). Plant tissue culture has arisen as an approaching instrument and forms the foundation of plant biotechnology. Plant tissue culture is conducted by excising any part of the plant and culturing it onto a nutrition medium that has been created artificially in culture vessels under aseptic conditions and maintained under controlled conditions (Colombo et al., 2018).

As far as we know, there are not enough studies on the influence of white light and dark on callus induction and phytochemical/antioxidant activity in *B. glabra*.

Hence, we studied the effect of light quality on morphological and biochemical components of *in vitro* grown node-derived callus cultures of *B. glabra*. This research will help understand the effect of light on the production of commercially essential secondary metabolites and their optimization in the *in vitro* cultures of *B. glabra* Choisy.

1.2 Statement of the problem

B. glabra is vegetatively propagated by stem cuttings. However, in the traditional propagation method, the production of secondary metabolites is known to be unsuitable due to external factors in the environment such as climate, plant pests and diseases, and fertilizer application. Therefore, the plant tissue culture technique is the most suitable technique for the consistent production of secondary metabolites under a controlled environment. Protocol for micro propagation of many woody plants is different, because each species could be influenced by different factors from which plant growth regulators, physical conditions, growth parameters and growing media are the most important ones. Although, micro propagation protocol for *B. glabra* has been carried out before, but no protocol can be found on callogenesis of *B. glabra* Choisy under different light regime. So, there is a need to develop a micro propagation protocol for callogenesis of *B. glabra* not only as a mass of biomass but also as somatic embryogenesis for future work.

1.3.1 Objectives of the study

The objectives of this research were;

1. To determine the suitable extraction solvent on phytochemical and antioxidant activities of leaf and node in *Bougainvillea glabra* Choisy.
2. To compare the effective concentration of Clorox® and exposure time for explant surface sterilization of *Bougainvillea*.
3. To determine the optimum concentration of 2,4-dichlorophenoxyacetic acid (2,4-D) and 6-benzyl amino purine (BAP) on callus induction of *Bougainvillea* under light conditions.
4. To compare the phytochemical and antioxidant activities of *in vitro*-induced callus and conventionally propagated nodal segment.

REFERENCES

- Abarca-Vargas, R., Peña Malacara, C. F., & Petricevich, V. L. (2016). Characterization of chemical compounds with antioxidant and cytotoxic activities in *Bougainvillea x buttiana* holttum and standl, (Var. rose) extracts. *Antioxidants*, 5(4), 1–12. <https://doi.org/10.3390/antiox5040045>
- Abarca-Vargas, R., & Petricevich, V. L. (2018). *Bougainvillea* genus: A review on phytochemistry, pharmacology, and toxicology. *Evidence-Based Complementary and Alternative Medicine*, 2018, 907-927. <https://doi.org/10.1155/2018/9070927>
- Abarca-vargas, R., Zamilpa, A., & Petricevich, V. L. (2019). Development and validation of conditions for extracting flavonoids content and evaluation of antioxidant and cytoprotective activities from *Bougainvillea x buttiana* bracteas (Var. rose). *Antioxidants*, 8(8), 264. <https://doi.org/10.3390/antiox8080264>
- Adibah, M., Zaman, K., Azzeme, A. M., Ramle, I. K., Normanshah, N., Ramli, S. N., Shaharuddin, N. A., Ahmad, S., Nor, S., & Abdullah, A. (2020). Induction, multiplication, and evaluation of antioxidant activity of *Polyalthia bullata* callus, a woody medicinal Plant. *Plants*, 9(8), 1–21. <https://doi.org/doi:10.3390/plants9121772>
- Adil, M., Ren, X., & Jeong, B. R. (2019). Light elicited growth, antioxidant enzymes activities and production of medicinal compounds in callus culture of *Cnidium officinale* Makino. *Journal of Photochemistry and Photobiology*, 196(March), 111509. <https://doi.org/10.1016/j.jphotobiol.2019.05.006>
- Ahmad, A., ul Qamar, M. T., Shoukat, A., Aslam, M. M., Tariq, M., Hakiman, M., & Joyia, F. A. (2021). The effects of genotypes and media composition on callogenesis, regeneration and cell suspension culture of chamomile (*Matricaria chamomilla* L.). *PeerJ*, 9, 1–19. <https://doi.org/10.7717/peerj.11464>
- Ahmad, I., Zamir, R., Shah, S. T., & Wali, S. (2016). *In vitro* surface sterilization of the shoot tips of *Bougainvillea spectabilis* Willd . *Pure and Applied Biology*, 5(4), 1171–1175. <https://doi.org/http://dx.doi.org/10.19045/bspab.2016.50140>
- Ahmed, K. M. I., Opu, S. A., Muttaki, A. A., Islam, T., Das, P. R., & Rahmatullah, M. (2015). Plant remedies of a unani medicinal practitioner in Bhola District, Bangladesh. *World Journal of Pharmacy and Pharmaceutical Sciences*, 4(11), 186–198.
- Ali, M. B., Khatun, S., Hahn, E. J., & Paek, K. Y. (2006). Enhancement of phenylpropanoid enzymes and lignin in *Phalaenopsis orchid* and their influence on plant acclimatisation at different levels of photosynthetic photon flux. *Plant Growth Regulation*, 49(2–3), 137–146. <https://doi.org/10.1007/s10725-006-9003-z>

- Ameya, G., Manilal, A., & Merdekios, B. (2017). *In vitro* antibacterial activity and phytochemical analysis of *Nicotiana tabacum* L. extracted in different organic solvents. *The Open Microbiology Journal*, 11(1), 352–359. <https://doi.org/10.2174/1874285801711010352>
- Anjusha, S., & Gangaprasad, A. (2017). Callus culture and *in vitro* production of anthraquinone in *Gynochthodes umbellata* (L.) Razafim. & B. Bremer (Rubiaceae). *Industrial Crops and Products*, 95, 608–614. <https://doi.org/10.1016/j.indcrop.2016.11.021>
- Anwar, F., & Przybylski, R. (2012). Effect of solvents extraction on total phenolics and antioxidant activity of extracts from flaxseed (*Linum usitatissimum* L.). *Acta Sci. Pol., Technol. Aliment*, 11(3), 293–302.
- Arezki, O., Boxus, P., Kevers, C., & Gaspar, T. (2001). Changes in peroxidase activity, and level of phenolic compounds during light-induced plantlet regeneration from *Eucalyptus camaldulensis* Dehn. nodes *in vitro*. *Plant Growth Regulation*, 33(3), 215–219. <https://doi.org/10.1023/A:1017579623170>
- Asuk, A. A., Agiang, M. A., Dasofunjo, K., & Willie, A. J. (2015). The biomedical significance of the phytochemical, proximate and mineral compositions of the leaf, stem bark and root of *Jatropha curcas*. *Asian Pacific Journal of Tropical Biomedicine*, 5(8), 650–657. <https://doi.org/10.1016/j.apjtb.2015.05.015>
- Azad, M. A. K., Yokota, S., Ohkubo, T., Andoh, Y., Yahara, S., & Yoshizawa, N. (2005). *In vitro* regeneration of the medicinal woody plant *Phellodendron amurense* Rupr. through excised leaves. *Plant Cell, Tissue and Organ Culture*, 80(1), 43–50. <https://doi.org/10.1007/s11240-004-8809-5>
- Barneix, A. J., & Causin, H. F. (1996). The central role of amino acids on nitrogen utilization and plant growth. *Journal of Plant Physiology*, 149(3–4), 358–362. [https://doi.org/10.1016/s0176-1617\(96\)80134-9](https://doi.org/10.1016/s0176-1617(96)80134-9)
- Behbahani, M., Shanehsazzadeh, M., & Hessami, M. J. (2011). Optimization of callus and cell suspension cultures of *Barringtonia racemosa* (Lecythidaceae family) for lycopene production. *Scientia Agricola*, 68(1), 69–76. <https://doi.org/10.1590/s0103-90162011000100011>
- Bhojwani, S. S., & Dantu, P. K. (2013). *Plant Tissue Culture : An Introductory Text*. Springer India. <https://doi.org/10.1007/978-81-322-1026-9>
- Bouasla, I., Hamel, T., Barour, C., Bouasla, A., Hachouf, M., Bouguerra, O. M., & Messarah, M. (2020). Evaluation of solvent influence on phytochemical content and antioxidant activities of two Algerian endemic taxa, *Stachys marrubiifolia* Viv. and *Lamium flexuosum* Ten. (Lamiaceae). In *European Journal of Integrative Medicine*. Elsevier GmbH. <https://doi.org/10.1016/j.eujim.2020.101267>

- Bourgaud, F., Saurav, M., Pandey, B., & Srivastava, P. (2001). Synthesis of nano-dimensional ZnO and Ga doped ZnO thin films by vapor phase transport and study as transparent conducting oxide. *Plant Science*, 161, 839–851. <https://doi.org/10.1166/jnn.2008.18298>
- Britton, R. S., Leicester, K. L., & Bacon, B. R. (2002). Iron toxicity and chelation therapy. *International Journal of Hematology*, 76(3), 219–228. <https://doi.org/10.1007/BF02982791>
- Bucić-Kojić, A., Planinić, M., Tomas, S., Bilić, M., & Velić, D. (2007). Study of solid-liquid extraction kinetics of total polyphenols from grape seeds. *Journal of Food Engineering*, 81(1), 236–242. <https://doi.org/10.1016/j.jfoodeng.2006.10.027>
- Cai, Y., Luo, Q., Sun, M., & Corke, H. (2004). Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life Sciences*, 74(17), 2157–2184. <https://doi.org/10.1016/j.lfs.2003.09.047>
- Chinnappan, R. S. (2018). Review article on problems and its remedy in plant tissue culture. *Asian Journal of Biological Sciences*, 11(4), 165–172. <https://doi.org/DOI: 10.3923/ajbs.2018.165.172>
- Chye, F. Y., & Kheng, Y. S. (2009). Antioxidative and antibacterial activities of *Pangium edule* seed extracts. *International Journal of Pharmacology*, 5(5), 285–297. <https://doi.org/10.3923/ijp.2009.285.297>
- Colombo, R. C., Da Cruz, M. A., De Carvalho, D. U., Hoshino, R. T., Alves, G. A. C., & De Faria, R. T. (2018). *Adenium obesum* as a new potted flower, Growth management. *Ornamental Horticulture*, 24(3), 197–205. <https://doi.org/10.14295/oh.v24i3.1226>
- Cos, P., Vlietinck, A. J., Berghe, D. Vanden, & Maes, L. (2006). Anti-infective potential of natural products. How to develop a stronger *in vitro* “proof-of-concept.” *Journal of Ethnopharmacology*, 106(3), 290–302. <https://doi.org/10.1016/j.jep.2006.04.003>
- Costa, P., Gonçalves, S., Valentão, P., Andrade, P. B., Coelho, N., & Romano, A. (2012). *Thymus lotocephalus* wild plants and *in vitro* cultures produce different profiles of phenolic compounds with antioxidant activity. *Food Chemistry*, 135(3), 1253–1260. <https://doi.org/10.1016/j.foodchem.2012.05.072>
- Daud, N. H., Jayaraman, S., & Mohamed, R. (2012). Methods paper: An improved surface sterilization technique for introducing leaf, nodal and seed explants of *Aquilaria malaccensis* from field sources into tissue culture. *Asia-Pacific Journal of Molecular Biology and Biotechnology*, 20(2), 55–58.
- Diengngan, S., Murthy, B. N. S., & Mahadevamma, M. (2014). Effective decontamination and regeneration protocol for *in vitro* culture of strawberry

- cultivar. Chandler. *Journal of Horticultural Sciences*, 9(2), 126–130.
- Dinis, T. C. P., Madeira., V. M. C., & Almeida, L. M. (1994). Action of phenolic derivatives (Acetaminophen) as inhibitors of membrane Lipid peroxidation and as peroxy radical scavengers. *Archives of Biochemistry and Biophysics*, 315 (1), 161–169.
- Do, L. T. M., Aree, T., Siripong, P., Vo, N. T., Nguyen, T. T. A., Nguyen, P. K. P., & Tip-Pyang, S. (2018). Cytotoxic flavones from the stem bark of *Bougainvillea spectabilis* Willd. *Planta Medica*, 84(2), 129–134. <https://doi.org/10.1055/s-0043-118102>
- Elkazzaz, A. (2015). Micropropagation of four potato cultivars *in vitro*. *Academia Journal of Agricultural Research*, 3(9), 184-188, January. <https://doi.org/10.15413/ajar.2015.0145>
- Esmaeili, A. K., Taha, R. M., Mohajer, S., & Banisalam, B. (2015). Antioxidant activity and total phenolic and flavonoid content of various solvent extracts from *in vivo* and *in vitro* grown *Trifolium pratense* L. (Red clover). *BioMed Research International*, 2015. <https://doi.org/10.1155/2015/643285>
- Espinosa-Leal, C. A., Puente-Garza, C. A., & García-Lara, S. (2018). *In vitro* plant tissue culture: means for production of biological active compounds. *Planta*, 248(1). <https://doi.org/10.1007/s00425-018-2910-1>
- Eziashi, E. I., Asemota, O., Okwuagwu, C. O., Eke, C. R., Chidi, N. I., & Oruade-Dimaro, E. A. (2014). Screening sterilizing agents and antibiotics for the elimination of bacterial contaminants from oil palm explants for plant tissue culture. *European Journal of Experimental Biology*, 4(4), 111–115.
- Fahim, J. R., Hegazi, G. A. E. M., Abo El-Fadl, R. E. S., Abd Alhady, M. R. A. A., Desoukey, S. Y., Ramadan, M. A., & Kamel, M. S. (2015). Production of rhoifolin and tiliroside from callus cultures of *Chorisia chodatii* and *Chorisia speciosa*. *Phytochemistry Letters*, 13, 218–227. <https://doi.org/10.1016/j.phytol.2015.06.004>
- Farvardin, A., Ebrahimi, A., Hosseinpour, B., & Khosrowshahli, M. (2017). Effects of growth regulators on callus induction and secondary metabolite production in *Cuminum cyminum*. *Natural Product Research*, 31(17), 1963–1970. <https://doi.org/10.1080/14786419.2016.1272105>
- Fatima, H., Khan, K., Zia, M., Ur-Rehman, T., Mirza, B., & Haq, I. ul. (2015). Extraction optimization of medicinally important metabolites from *Datura innoxia* Mill. An *in vitro* biological and phytochemical investigation. *BMC Complementary and Alternative Medicine*, 15(1), 358-376. <https://doi.org/10.1186/s12906-015-0891-1>
- Ferreres, F., Pereira, D. M., Valentão, P., Andrade, P. B., Seabra, R. M., & Sottomayor, M. (2008). New phenolic compounds and antioxidant potential of *Catharanthus roseus*. *Journal of Agricultural and Food Chemistry*, 56(21), 9967–9974. <https://doi.org/10.1021/jf8022723>

- Gaber, M. K. (2015). In vitro propagation and somatic embryogenesis of Marvel of Peru (*Mirabilis jalapa* L.) from nodal and leaf explants. *Alexandria Science Exchange Journal, An International Quarterly Journal of Science Agricultural Environments*, 36(1), 314–324. <https://doi.org/10.21608/asejaiqsae.2015.2937>
- Gao, J., Li, J., Luo, C., Yin, L., Li, S., Yang, G., & He, G. (2011). Callus induction and plant regeneration in *Alternanthera philoxeroides*. *Molecular Biology Reports*, 38(2), 1413–1417. <https://doi.org/10.1007/s11033-010-0245-5>
- García-González, R., Quiroz, K., Carrasco, B., & Caligari, P. (2010). Plant tissue culture: Current status, opportunities and challenges. *Ciencia e Investigación Agraria*, 37(3), 5–30. <https://doi.org/10.4067/s0718-16202010000300001>
- George, E. F., Hall, M. A., & Klerk, G. J. De. (2008). Plant growth regulators, Introduction; Auxins, their analogues and inhibitors. *Plant Propagation by Tissue Culture 3rd Edition*, 1, 175–204. https://doi.org/10.1007/978-1-4020-5005-3_5
- Ghogar, A., Jiraungkoorskul, K., & Jiraungkoorskul, W. (2016). Paper flower, *Bougainvillea spectabilis*: Update antiepileptic and antipsychotic effects of properties of traditional medicinal plant. *Journal of Natural Remedies*, 16(3), 82–87. <https://doi.org/10.18311/jnr/2016/5703>
- Gupta, V., George, M., Joseph, L., Singhal, M., & Singh, H. P. (2009). Evaluation of antibacterial activity of *Bougainvillea glabra* 'snow white' and *Bougainvillea glabra* 'choicy'. *Journal of Chemical and Pharmaceutical Research*, 1(1), 233–237.
- Habibah, N. A., Moeljepawiro, S., Dewi, K., & Indrianto, A. (2018). Callus induction and flavonoid production on the immature seed of *Stelechocarpus burahol*. *Journal of Physics, Conference Series*, 983(1), 78-86. <https://doi.org/10.1088/1742-6596/983/1/012186>
- Haida, Z., Nakasha, J. J., & Hakiman, M. (2020). In vitro responses of plant growth factors on growth, yield, phenolics content and antioxidant activities of *Clinacanthus nutans* (Sabah snake grass). *Plants*, 9(8), 1–17. <https://doi.org/10.3390/plants9081030>
- Hakiman, M., & Maziah, M. (2009). Non enzymatic and enzymatic antioxidant activities in aqueous extract of different *Ficus deltoidea* accessions. *Journal of Medicinal Plants Research*, 3(3), 120–131.
- Hakkim, F., Shankar, C., & Girija, S. (2007). Chemical composition and antioxidant property of holy basil (*Ocimum sanctum* L.) leaves, stems, and inflorescence and their in vitro callus cultures. *Journal of Agricultural and Food Chemistry*, 55, 9109–9117.
- Halliwell, B., & Gutteridge, J. M. C. (2015). Free radicals in biology and medicine. OXFORD UNIVERSITY PRESS, 5th edition, 132.

<https://doi.org/10.1093/acprof:oso/9780198717478.003.0001>

- Harsha, K., Jaishree, R., Mousumi, B., & Avinash, U. (2011). *In-vitro* callus induction and shoot regeneration in *Boerhaavia diffusa* L. *Scholars. Annals of Biological Research*, 2(1), 142–148.
- Harshal, B. A., & Gauam, S. P. (2014). Plant tissue culture: A review. *World Journal of Pharmaceutical Sciences*, 2(6), 565–572. <https://doi.org/10.1007/BF00057619>
- Hassan, K., Hosni, A., Hewidy, M., & Abd El razik, A. (2019). Micropropagation and evaluation of genetic stability of foxglove tree (*Paulownia tomentosa*). *Arab Universities Journal of Agricultural Sciences*, 26(6), 2287–2296. <https://doi.org/10.21608/ajs.2018.35343>
- Hayouni, E. A., Abedrabba, M., Bouix, M., & Hamdi, M. (2007). The effects of solvents and extraction method on the phenolic contents and biological activities *in vitro* of Tunisian *Quercus coccifera* L. and *Juniperus phoenicea* L. fruit extracts. *Food Chemistry*, 105(3), 1126–1134. <https://doi.org/10.1016/j.foodchem.2007.02.010>
- Hesami, M., & Daneshvar, M. H. (2018). *In vitro* adventitious shoot regeneration through direct and indirect organogenesis from seedling-derived hypocotyl segments of *Ficus religiosa* L., An important medicinal plant. *HortScience*, 53(1), 55–61. <https://doi.org/10.21273/HORTSCI12637-17>
- Hill, K., & Schaller, G. E. (2013). Enhancing plant regeneration in tissue culture: A molecular approach through manipulation of cytokinin sensitivity. *Plant Signaling and Behavior*, 8(10), 162–168. <https://doi.org/10.4161/psb.25709>
- Hoque, A., Razvy, M., Biswas, M. ., & Kbir, A. . (2006). Micropropagation of water chestnut (*Trapa sp.*) through local varieties of Rajshahi Division. *Asian Journal of Plant Sciences*, 5(3), 409–413.
- Hu, Shang lian, Zhou, J. ying, Cao, Y., Lu, X. qin, Duan, N., Ren, P., & Chen, K. (2011). *In vitro* callus induction and plant regeneration from mature seed embryo and young shoots in a giant sympodial bamboo, *Dendrocalamus farinosus* (Keng et Keng f.) Chia et H.L. Fung. *African Journal of Biotechnology*, 10(16), 3210–3215. <https://doi.org/10.5897/ajb10.2014>
- Hu, Sheng, Ma, Y., Jiang, H., Feng, D., Yu, W., Dai, D., & Mei, L. (2015). Production of paeoniflorin and albiflorin by callus tissue culture of *Paeonia lactiflora* Pall. *Chinese Journal of Chemical Engineering*, 23(2), 451–455. <https://doi.org/10.1016/j.cjche.2014.06.036>
- Ikeuchi, M., Ogawa, Y., Iwase, A., & Sugimoto, K. (2016). Plant regeneration: Cellular origins and molecular mechanisms. *Development (Cambridge)*, 143(9), 1442–1451. <https://doi.org/10.1242/dev.134668>
- Iloki-Assanga, S. B., Lewis-Luján, L. M., Lara-Espinoza, C. L., Gil-Salido, A. A., Fernandez-Angulo, D., Rubio-Pino, J. L., & Haines, D. D. (2015). Solvent

- effects on phytochemical constituent profiles and antioxidant activities, using four different extraction formulations for analysis of *Bucida buceras* L. and *Phoradendron californicum* Complementary and Alternative Medicine. *BMC Research Notes*, 8(1), 1–14. <https://doi.org/10.1186/s13104-015-1388-1>
- Ishida, T., & Rossky, P. J. (2001). Solvent effects on solute electronic structure and properties: Theoretical study of a betaine dye molecule in polar solvents. *Journal of Physical Chemistry A*, 105(3), 558–565. <https://doi.org/10.1021/jp0041104>
- Islam, M. Z., Hossain, M. T., Hossen, F., Akter, M. S., & Mokammel, M. A. (2016). *In-vitro* antioxidant and antimicrobial activity of *Bougainvillea glabra* flower. *Research Journal of Medicinal Plant*, 10(3), 228–236. <https://doi.org/10.3923/rjmp.2016.228.236>
- Jacob, A., & Malpathak, N. (2005). Plantlet regeneration enhances solasodine productivity in hairy root cultures of *Solanum khasianum* Clarke. *In vitro Cellular and Developmental Biology, Plant*, 41(3), 291–295. <https://doi.org/10.1079/IVP2005637>
- Jadid, N., Hidayati, D., Hartanti, S. R., Arraniry, B. A., Rachman, R. Y., & Wikanta, W. (2017). Antioxidant activities of different solvent extracts of *Piper retrofractum* Vahl. using DPPH assay. *AIP Conference Proceedings*, 1854(June 2017). <https://doi.org/10.1063/1.4985410>
- Jan, A., Bhat, K. M., Mir, M. A., Bhat, M. A., & Wani, I. A. (2013). Surface sterilization method for reducing microbial contamination of field grown strawberry explants intended for *in vitro* culture. *African Journal of Biotechnology*, 12(39), 5749–5753. <https://doi.org/10.5897/AJB2013.12918>
- Jaramillo, J., & Summers, W. L. (2019). Dark–light treatments influence induction of tomato anther callus. *HortScience*, 26(7), 915–916. <https://doi.org/10.21273/hortsci.26.7.915>
- Juntachote, T., & Berghofer, E. (2005). Antioxidative properties and stability of ethanolic extracts of Holy basil and Galangal. *Food Chemistry*, 92(2), 193–202. <https://doi.org/10.1016/j.foodchem.2004.04.044>
- Kainama, H., Fatmawati, S., Santoso, M., Papilaya, P. M., & Ersam, T. (2020). The relationship of free radical scavenging and total phenolic and flavonoid contents of *Garcinia lasoar* PAM. *Pharmaceutical Chemistry Journal*, 53(12), 1151–1157. <https://doi.org/10.1007/s11094-020-02139-5>
- Kaisoon, O., Konczak, I., & Siriamornpun, S. (2012). Potential health enhancing properties of edible flowers from Thailand. *Food Research International*, 46(2), 563–571. <https://doi.org/10.1016/j.foodres.2011.06.016>
- Kannamba, B., Winnie, T., M, S., & B, L. (2017). Steady characteristics of the water-lubricated conical bearings. *Derpharmachemica*, 9(9), 152–156.

- Kaufmann, B., & Christen, P. (2002). Recent extraction techniques for natural products: Microwave-assisted extraction and pressurised solvent extraction. *Phytochemical Analysis*, 13(2), 105–113. <https://doi.org/10.1002/pca.631>
- Khafagi, I. K. (2007). Variation of callus induction and active metabolite accumulation in callus cultures of two varieties of *Ricinus communis* L. *Biotechnology*, 6 (2), 193–201. <https://doi.org/10.3923/biotech.2007.193.201>
- Khan, J. A., Jaskani, M. J., Abbas, H., & Khan, M. M. (2006). Effect of light and dark culture conditions on callus induction and growth in citrus (*Citrus reticulata* blanco.). *Int. J. Biol. Biotech*, 3(4), 669–672.
- Khan Rohela, G., Damera, S., Bylla, P., Korra, R., Pendli, S., & Thammidala, C. (2016). Somatic embryogenesis and indirect regeneration in *Mirabilis jalapa* Linn. *Materials Today, Proceedings*, 3(10), 3882–3891. <https://doi.org/10.1016/j.matpr.2016.11.045>
- Kudo, T., Kiba, T., & Sakakibara, H. (2010). Metabolism and long-distance translocation of cytokinins. *Journal of Integrative Plant Biology*, 52(1), 53–60. <https://doi.org/10.1111/j.1744-7909.2010.00898.x>
- Kumarasamy, S., & Selvi, S. (2020). Extraction of phytochemicals of artocarpus altilis (Parkinson) fosberg (seedless) fruit pulp using non-polar and polar solvents. *International Journal Of Scientific Resarch in Engineering and Management(IJSREM)*, 04(03), 1–13.
- Lai, P., Li, K. Y., Lu, S., & Chen, H. H. (2009). Phytochemicals and antioxidant properties of solvent extracts from Japonica rice bran. *Food Chemistry*, 117(3), 538–544. <https://doi.org/10.1016/j.foodchem.2009.04.031>
- Larson, R. A. (1988). The antioxidants of higher plants. *Phytochemistry*, 27(4), 969–978. [https://doi.org/10.1016/0031-9422\(88\)80254-1](https://doi.org/10.1016/0031-9422(88)80254-1)
- Lee, J. H., Renita, M., Fioritto, R. J., St. Martin, S. K., Schwartz, S. J., & Vodovotz, Y. (2004). Isoflavone characterization and antioxidant activity of Ohio Soybeans. *Journal of Agricultural and Food Chemistry*, 52(9), 2647–2651. <https://doi.org/10.1021/jf035426m>
- Lee, O. N., Ak, G., Zengin, G., Cziáky, Z., Jekó, J., Rengasamy, K. R. R., Park, H. Y., Kim, D. H., & Sivanesan, I. (2020). Phytochemical composition, antioxidant capacity, and enzyme inhibitory activity in callus, somaclonal variant, and normal green shoot tissues of *Catharanthus roseus* (L) G. Don. *Molecules (Basel, Switzerland)*, 25(21), 1–23. <https://doi.org/10.3390/molecules25214945>
- Leifert, C., Morris, C. E., & Waites, W. M. (1994). Ecology of microbial saprophytes and pathogens in tissue culture and field-grown plants: Reasons for contamination problems *in vitro*. *Critical Reviews in Plant*

Sciences, 13(2), 139-183. <https://doi.org/10.1080/07352689409701912>

- Leong, L. P., & Shui, G. (2002). An investigation of antioxidant capacity of fruits in Singapore markets. *Food Chemistry*, 76(1), 69–75. [https://doi.org/10.1016/S0308-8146\(01\)00251-5](https://doi.org/10.1016/S0308-8146(01)00251-5)
- Lian, T. T., Cha, S. Y., Moe, M. M., Kim, Y. J., & Bang, K. S. (2019). Effects of different colored LEDs on the enhancement of biologically active ingredients in callus cultures of *Gynura procumbens* (Lour.) Merr. *Molecules*, 24(23), 4336–4346. <https://doi.org/10.3390/molecules24234336>
- López-Laredo, A. R., Ramírez-Flores, F. D., Sepúlveda-Jiménez, G., & Trejo-Tapia, G. (2009). Comparison of metabolite levels in callus of *Tecoma stans* (L.) Juss. ex Kunth. cultured in photoperiod and darkness. *In vitro Cellular and Developmental Biology - Plant*, 45(5), 550–558. <https://doi.org/10.1007/s11627-009-9250-6>
- Lulai, E. C., Suttle, J. C., Olson, L. L., Neubauer, J. D., Campbell, L. G., & Campbell, M. A. (2016). Wounding induces changes in cytokinin and auxin content in potato tuber, but does not induce formation of gibberellins. *Journal of Plant Physiology*, 191, 22–28. <https://doi.org/10.1016/j.jplph.2015.11.006>
- Maharik, N., Souad, E., & Hussein, T. (2009). Anthocyanin production in callus cultures of *Cleome rosea*: Modulation by culture conditions and characterization of pigments by means of HPLC-DAD/ESIMS. *Plant Physiology and Biochemistry*, 47(10), 895–903. <https://doi.org/10.1016/j.plaphy.2009.06.005>
- Mahendra, M., M., G., M., & M.S., S. (2020). Biopotentiality of leaf and leaf derived callus extracts of *Salacia macrosperma* Wight. An endangered medicinal plant of Western Ghats. *Industrial Crops and Products*, 143(2019), 111921. <https://doi.org/10.1016/j.indcrop.2019.111921>
- Mahmad, N., Taha, R. M., Rawi, N., & Mohajer, S. (2015). The effects of picloram and 2,4-dichlorophenoxyacetic acid on the induction of red colored callus from *Celosia plumose* as an attractive ornamental plant. *Journal of Applied and Physical Sciences*, 1(1), 9–12. <https://doi.org/10.20474/japs-1.1.2>
- Mahmoud, S. N., & Al-ani, N. K. (2016). Effect of different sterilization methods on contamination and viability of nodal segments of *Cestrum nocturnum* L. *International Journal of Research Studies in Biosciences*, 4(1), 4–9. <https://doi.org/10.20431/2349-0365.0401002>
- Majda, M., & Robert, S. (2018). The role of auxin in cell wall expansion. *International Journal of Molecular Sciences*, 19(4), 951. <https://doi.org/10.3390/ijms19040951>
- Manian, R., Anusuya, N., Siddhuraju, P., & Manian, S. (2008). The antioxidant activity and free radical scavenging potential of two different solvent extracts of *Camellia sinensis* (L.) O. Kuntz, *Ficus bengalensis* L. and *Ficus racemosa* L. *Food Chemistry*, 107(3), 1000–1007.

<https://doi.org/10.1016/j.foodchem.2007.09.008>

Marasini, P., & Khanal, A. . (2018). Assessing rooting media and hormone on rooting potential of stem cuttings of *Bougainvillea*. *Journal of the Institute of Agriculture and Animal Science*, 35(1), 197–201. <https://doi.org/10.3126/jjaas.v35i1.22541>

Marinova, D., Ribarova, & Atanassova. (2005). Total phenolics and total flavonoids in Bulgarian fruits and vegetables. *Journal of the university of chemical technology and metallurgy*, 40(3), 255–260.

Matkowski, A. (2008). Plant *in vitro* culture for the production of antioxidants - A review. *Biotechnology Advances*, 26(6), 548–560. <https://doi.org/10.1016/j.biotechadv.2008.07.001>

Meera & Sivakumar. (2019). Screening of medicinal plants for iron chelating and antioxidant activity. *Biotechnology International*, 12(2)(January), 30–36.

Mofokeng, M. M., du Plooy, C. P., Araya, H. T., Amoo, S. O., Mokgehle, S. N., Pofu, K. M., & Mashela, P. W. (2022). Medicinal plant cultivation for sustainable use and commercialisation of high-value crops. *South African Journal of Science*, 118(7/8), 1–7. <https://doi.org/10.17159/sajs.2022/12190>

Mohammad, S., Khan, M. A., Ali, A., Khan, L., Khan, M. S., & Mashwani, Z. ur R. (2019). Feasible production of biomass and natural antioxidants through callus cultures in response to varying light intensities in olive (*Olea europaea*. L) cult. Arbosana. *Journal of Photochemistry and Photobiology B: Biology*, 193(December 2018), 140–147. <https://doi.org/10.1016/j.jphotobiol.2019.03.001>

Mohan, M., Khanam, S., & Shivananda, B. G. (2013). Optimization of microwave assisted extraction of andrographolide from andrographis paniculata and its comparison with refluxation extraction method. *Journal of Pharmacognosy and Phytochemistry*, 2(1), 342–348.

Mok, D. W. S., & Mok, M. C. (2001). Cytokinin metabolism and action. *Annu. Rev. Plant Physiol. Plant Mol. Biol*, 52(39), 89–118. <http://www.ncbi.nlm.nih.gov/pubmed/11337393>

Moniruzzaman, M., Yaakob, Z., & Taha, R. A. (2017). In vitro production of fig (*Ficus carica* L.) plantlets. *Acta Horticulturae*, 1173(October), 231–235. <https://doi.org/10.17660/ActaHortic.2017.1173.40>

Mostafiz, S., & Wagiran, A. (2018). Efficient callus induction and regeneration in selected Indica rice. *Agronomy*, 8(5), 77-95. <https://doi.org/10.3390/agronomy8050077>

Murali, M., & Prabakaran, G. (2018). Effect of different solvents system on antioxidant activity and phytochemical screening in various habitats of *Ocimum basilicum* L . (sweet basil) leaves. *International Journal of Zoology and Applied Biosciences*, 3(5), 375–381.

<https://doi.org/https://doi.org/10.5281/zenodo.1439290>

- Murashige, T. (1974). Plant propagation through tissue cultures. *Annual Review of Plant Physiology*, 25(1), 135–166. <https://doi.org/10.1146/annurev.pp.25.060174.001031>
- Muttaleb, Q. A., Abdullah, T. L., Rashid, A. A., & Hassan, S. A. (2017). Rooting of stem cuttings with different indole 3 butyric acid (IBA) treatments and development of Mmcropropagation protocol for *Piper betle* L. node culture. *American Journal of Plant Sciences*, 08(12), 3084–3100. <https://doi.org/10.4236/ajps.2017.812208>
- Naito, F. Y. B., de Nazaré Almeida dos Reis, L., Batista, J. G., Nery, F. M. B., Rossato, M., Melo, F. L., & de Cássia Pereira-Carvalho, R. (2020). Complete genome sequence of *Bougainvillea* chlorotic vein banding virus in *Bougainvillea spectabilis* from Brazil. *Tropical Plant Pathology*, 45(2), 159–162. <https://doi.org/10.1007/s40858-020-00349-6>
- Nawaz, H., Aslam, M., & Muntaha, S. T. (2019). Effect of solvent polarity and extraction method on phytochemical composition and antioxidant potential of corn silk. *Free Radicals and Antioxidants*, 9(1), 05–11. <https://doi.org/10.5530/fra.2019.1.2>
- Neha, K., Haider, M. R., Pathak, A., & Yar, M. S. (2019). Medicinal prospects of antioxidants: A review. *European Journal of Medicinal Chemistry*, 178, 687–704. <https://doi.org/10.1016/j.ejmech.2019.06.010>
- Nguyen, Pham, H. N. T., Bowyer, M. C., Van Altena, I. A., & Scarlett, C. J. (2016). Influence of solvents and novel extraction methods on bioactive compounds and antioxidant capacity of *Phyllanthus amarus*. *Chemical Papers*, 70(5), 556–566. <https://doi.org/10.1515/chempap-2015-0240>
- Nguyen, T. T. A., Nguyen, H. L., Pham, T. N. K., Nguyen, P. K. P., Huynh, T. T. N., Sichaem, J., & Do, L. T. M. (2021). Bougainvinones N–P, three new flavonoids from *Bougainvillea spectabilis*. *Fitoterapia*, 149(January), 104832. <https://doi.org/10.1016/j.fitote.2021.104832>
- Nimse, S. B., & Pal, D. (2015). Free radicals, natural antioxidants, and their reaction mechanisms. *RSC Advances*, 5(35), 27986–28006. <https://doi.org/10.1039/c4ra13315c>
- Nobossé, P., Fombang, E. N., & Mbofung, C. M. F. (2018). Effects of age and extraction solvent on phytochemical content and antioxidant activity of fresh *Moringa oleifera* L. leaves. *Food Science and Nutrition*, 6(8), 2188–2198. <https://doi.org/10.1002/fsn3.783>
- Ogunwande, I. A., Avoseh, O. N., Olasunkanmi, K. N., Lawal, O. A., Ascrizzi, R., & Flamini, G. (2019). Chemical composition, anti-nociceptive and anti-inflammatory activities of essential oil of *Bougainvillea glabra*. *Journal of Ethnopharmacology*, 232, 188–192. <https://doi.org/10.1016/j.jep.2018.12.017>

- Pan, Z., Zhu, S., Guan, R., & Deng, X. (2010). Identification of 2,4-D-responsive proteins in embryogenic callus of Valencia sweet orange (*Citrus sinensis* Osbeck) following osmotic stress. *Plant Cell, Tissue and Organ Culture*, 103(2), 145–153. <https://doi.org/10.1007/s11240-010-9762-0>
- Pandey, A., Verma, O., & Chand, S. (2019). *In vitro* propagation of *Boerhaavia diffusa* L. An important medicinal plant of family nyctaginaceae. *Indian Journal of Genetics and Plant Breeding*, 79(1), 89–95. <https://doi.org/10.31742/IJGPB.79.1.12>
- Park, J. S., Seong, Z. K., Kim, M. S., Ha, J. H., Moon, K. B., Lee, H. J., Lee, H. K., Jeon, J. H., Park, S. U., & Kim, H. S. (2020). Production of flavonoids in callus cultures of *Sophora flavescens* aiton. *Plants*, 9(6), 1–13. <https://doi.org/10.3390/plants9060688>
- Pavan Kumar, P., Janakiram, T., & Bhat, K. V. (2020). Microsatellite based DNA fingerprinting and assessment of genetic diversity in bougainvillea cultivars. *Gene*, 753, 144794. <https://doi.org/10.1016/j.gene.2020.144794>
- Pérez-Jiménez, M., López-Soto, M. B., & Cos-Terrer, J. (2013). *In vitro* callus induction from adult tissues of peach (*Prunus persica* L. Batsch). *In vitro Cellular and Developmental Biology - Plant*, 49(1), 79–84. <https://doi.org/10.1007/s11627-012-9466-8>
- Priya, J., & Patric, D. D. (2008). Anti-bacterial activity studies of *Jasminum grandiflorum* and *Jasminum sambac*. *Ethnobotanical Leaflets*, 12, 481–483.
- Qasim, M., Aziz, I., Rasheed, M., Gul, B., & Ajmal Khan, M. (2016). Effect of extraction solvents on polyphenols and antioxidant activity of medicinal halophytes. *Pakistan Journal of Botany*, 48(2), 621–627.
- Quezada, N., & Cherian, G. (2012). Lipid characterization and antioxidant status of the seeds and meals of *Camelina sativa* and flax. *European Journal of Lipid Science and Technology*, 114(8), 974–982. <https://doi.org/10.1002/ejlt.201100298>
- Quideau, S., Deffieux, D., Douat-Casassus, C., & Pouységu, L. (2011). Plant polyphenols: Chemical properties, biological activities, and synthesis. *Angewandte Chemie - International Edition*, 50(3), 586–621. <https://doi.org/10.1002/anie.201000044>
- Raju, N., Kanwar, S., & Kumar, S. (2016). Standardization of protocol for in vitro multiplication of bougainvillea. *Indian Journal of Agricultural Sciences*, 86(4), 516–521.
- Rameshkumar, R., Satish, L., Pandian, S., Rathinapriya, P., Rency, A. S., Shanmugaraj, G., Pandian, S. K., Leung, D. W. M., & Ramesh, M. (2018). Production of squalene with promising antioxidant properties in callus cultures of *Nilgiranthus ciliatus*. *Industrial Crops and Products*,

126(October), 357–367. <https://doi.org/10.1016/j.indcrop.2018.10.031>

- Rashid, S. A., Rehmani, F. S., Arman, M., Ibrahim, M., & Shafique, S. (2011). Estimation of moisture content & metal ions in white flowers of *Bougainvillea spectabilis* and purple flowers of *Bougainvillea glabra* in Pakistan. *Pakistan Journal of Chemistry*, 1(4), 190–192. <https://doi.org/10.15228/2011.v01.i04.p08>
- Re, R., Brühwiler, P., Mourad, S., Verdejo, R., & Shaffer, M. (1999). Development and characterisation of carbon nanotube-reinforced polyurethane foams. *Free Radical Biology & Medicine*, 26(9/10), 1231–1237.
- Renu, S., Kharb, P., & Rani, K. (2011). Rapid micropropagation and callus induction of *Catharanthus roseus* in vitro using different explants. *World Journal of Agricultural Sciences*, 7(6), 699–704.
- Rice-Evans, C. A., & Miller, N. J. (1996). Antioxidant activities of flavonoids as bioactive components of food. *Biochemical Society Transactions*, 24(3), 790–795. <https://doi.org/10.1042/bst0240790>
- Ridzuan, N. I., Abdullah, N., Vun, Y. L., & Supramaniam, C. V. (2020). Micropropagation and defence enzymes assessment of *Moringa oleifera* L. plantlets using nodal segments as explant. *South African Journal of Botany*, 129, 56–61. <https://doi.org/10.1016/j.sajb.2018.12.010>
- Rodrigues, F. R., Almeida, W. A. B. De, Ledo, C. A. D. S., Soares, T. L., Rossi, M. L., & Santana, J. R. F. De. (2020). *In vitro* callus induction and development of *Vernonia condensata* baker with embryogenic potential. *Ciencia e Agrotecnologia*, 44. <https://doi.org/10.1590/1413-7054202044026719>
- Rownaq, A., & Hossain, M. . (2010). *In vitro* direct and indirect plantlets regeneration from nodal segment of *Boerhaavia repens* L. *International Journal of Sustainable Crop Production*, 5(4), 5-10.
- Saha, S., Barua, B., & Sikdar, D. (2017). Phytochemical screening, phenolic content and antioxidant activity of wild date palm (*Phoenix sylvestris* Roxb.) fruit extracted with different solvents. *International Food Research Journal*, 24(6), 2534–2542.
- Sajjad, Y., Jaskani, M. J., Asif, M., & Qasim, M. (2017). Application of plant growth regulators in ornamental plants - A review. *Pakistan Journal of Agricultural Sciences*, 54(2), 327–333. <https://doi.org/10.21162/PAKJAS/17.3659>
- Saleem, H., Thet., H., Naidu, R., Sirajudeen, A., & Ahmed, N. (2020a). HPLC–PDA polyphenolic quantification, UHPLC–MS secondary metabolite composition, and in vitro enzyme inhibition potential of *Bougainvillea glabra*. *Plants*, 9(3), 388-399. <https://doi.org/10.3390/plants9030388>
- Saleem, Hammad, Htar, T. T., Naidu, R., Zengin, G., Ahmad, I., & Ahemad, N.

- (2020). Phytochemical profiling, antioxidant, enzyme inhibition and cytotoxic potential of *Bougainvillea glabra* flowers. *Natural Product Research*, 34(18), 2602–2606. <https://doi.org/10.1080/14786419.2018.1543684>
- Saleem, Hammad, Usman, A., Mahomoodally, M. F., & Ahemad, N. (2021). *Bougainvillea glabra* (choisy), A comprehensive review on botany, traditional uses, phytochemistry, pharmacology and toxicity. *Journal of Ethnopharmacology*, 266(August 2020), 113356. <https://doi.org/10.1016/j.jep.2020.113356>
- Saleem, Hammad, Zengin, G., Ahmad, I., Lee, J. T. B., Htar, T. T., Mahomoodally, F. M., Naidu, R., & Ahemad, N. (2019). Multidirectional insights into the biochemical and toxicological properties of *Bougainvillea glabra* (Choisy.) aerial parts. A functional approach for bioactive compounds. *Journal of Pharmaceutical and Biomedical Analysis*, 170, 132–138. <https://doi.org/10.1016/j.jpba.2019.03.027>
- Samuoliene, G., Brazaityte, A., Urbonavičiute, A., Šabajeviene, G., & Duchovskis, P. (2010). The effect of red and blue light component on the growth and development of frigo strawberries. *Zemdirbyste-Agriculture*, 97(2), 99–104.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. ., & Yoga, L. (2011). Extraction, isolation and characterization of bioactive compounds from plants extracts. *African Journal Tradit Complement*, 8(1), 1–10. https://doi.org/10.1007/978-3-642-56936-4_2
- Sati, P., Pandey, A., Rawat, S., & Rani, A. (2013). Phytochemicals and antioxidants in leaf extracts of *Ginkgo biloba* with reference to location, seasonal variation and solvent system. *Journal of Pharmacy Research*, 7(9), 804–809. <https://doi.org/10.1016/j.jopr.2013.09.001>
- Savita, V., Virk, G. S., & Avinash, N. (2010). Effect of explant type and different plant growth regulators on callus induction and plantlet regeneration in *Citrus jambhiri* Lush. *Environ. We Int. J. Sci. Tech.*, 5, 97–106.
- Shah, M., Ullah, M. A., Drouet, S., Younas, M., Tungmunnithum, D., Giglioli-Guivarc'h, N., Hano, C., & Abbasi, B. H. (2019). Interactive effects of light and melatonin on biosynthesis of silymarin and anti-inflammatory potential in callus cultures of *Silybum marianum* (L.) gaertn. *Molecules*, 24(7), 1207. <https://doi.org/10.3390/molecules24071207>
- Sharma, P., Industry, P., & Patil, D. (2017). Effect of culture media and growth hormones on callus induction in *Crataeva tapia* L. *International Journal of Pharmaceutical Research*, 9(2), 70–76.
- Sheikh, B. Y., & Gabr, S. (2016). Influence of extraction solvents and phytochemical analysis in the evaluation of *in-vitro* antioxidant activity of Saudi arabian olive leaves extract. *American Journal of Chemistry and*

Application 2016, 3(2), 6–12.

- Singh, C. K., Raj, S. R., Jaiswal, P. S., Patil, V. R., Punwar, B. S., Chavda, J. C., & Subhash, N. (2015). Effect of plant growth regulators on *in vitro* plant regeneration of sandalwood (*Santalum album* L.) via organogenesis. *Agroforestry Systems*, 90(2), 281–288. <https://doi.org/10.1007/s10457-015-9853-3>
- Singh, V., & Vipul, A. (2018). Phytochemical analysis and *in vitro* antioxidant activities of leaves, stems, flowers, and roots extracts of *Bougainvillea spectabilis* Willd. *International Journal of Green Pharmacy*, 12(4), 277–284.
- Singleton, V. L., & Rossi, J. A. J. (1965). Colorimetry to total phenolics with phosphomolybdic acid reagents. *American Journal of Enology and Viniculture*, 16(48), 144–158
<http://garfield.library.upenn.edu/classics1985/A1985AUG6900001.pdf>.
- Siwach, P., Gill, A. R., & Kumari, K. (2011). Effect of season, explants, growth regulators and sugar level on induction and long term maintenance of callus cultures of *Ficus religiosa* L. *African Journal of Biotechnology*, 10(24), 4879–4886. <https://doi.org/10.5897/AJB10.2119>
- Skůpa, P., Opatrný, Z., & Petrášek, J. (2014). Auxin biology, Applications and the mechanisms behind. *Plant Cell Monographs*, 22, 69–102. https://doi.org/10.1007/978-3-642-41787-0_3
- Song, K., Sivanesan, I., Ak, G., Zengin, G., Cziáky, Z., Jekő, J., Rengasamy, K. R. R., Lee, O. N., & Kim, D. H. (2020). Screening of bioactive metabolites and biological activities of calli, shoots, and seedlings of *Mertensia maritima* (L.) gray. *Plants*, 9(11), 1–20. <https://doi.org/10.3390/plants9111551>
- Sotiropoulos, T. E., Molassiotis, A. N., Mouhtaridou, G. I., Papadakis, I., Dimassi, K. N., Therios, I. N., & Diamantidis, G. (2006). Sucrose and sorbitol effects on shoot growth and proliferation *in vitro*, nutritional status and peroxidase and catalase isoenzymes of M 9 and MM 106 apple (*Malus domestica* Borkh.) rootstocks. *European Journal of Horticultural Science*, 71(3), 114–119.
- Souza, J. M. M., Berkov, S., & Santos, A. S. (2014). Improvement of friable callus production of *Boerhaavia paniculata* rich and the investigation of its lipid profile by GC/MS. *Anais Da Academia Brasileira de Ciencias*, 86(3), 1015–1027. <https://doi.org/10.1590/0001-3765201420130098>
- Sturm, A., & Tang, G. Q. (1999). The sucrose-cleaving enzymes of plants are crucial for development, growth and carbon partitioning. *Trends in Plant Science*, 4(10), 401–407. [https://doi.org/10.1016/S1360-1385\(99\)01470-3](https://doi.org/10.1016/S1360-1385(99)01470-3)
- Tariq, U., Ali, M., & Abbasi, B. H. (2014). Morphogenic and biochemical variations under different spectral lights in callus cultures of *Artemisia*

absinthium L. *Journal of Photochemistry and Photobiology*, 130, 264–271.
<https://doi.org/10.1016/j.jphotobiol.2013.11.026>

- Thammina, C., He, M., Lu, L., Cao, K., Yu, H., Chen, Y., Tian, L., Chen, J., Mcavoy, R., Ellis, D., Zhao, D., Wang, Y., Zhang, X., & Li, Y. (2011). *In vitro* regeneration of triploid plants of euonymus alatus “compactus” (burning bush) from endosperm tissues. *HortScience*, 46(8), 1141–1147.
<https://doi.org/10.21273/hortsci.46.8.1141>
- Thi, K. T., Oo, K. S., & Mon, Y. (2018). Establishment of efficient surface Sterilization protocol on different types of field grown strawberry explants (*Fragaria x ananassa* Duch.). *Journal of Scientific and Innovative Research*, 7(3), 70–74.
- Thomas, J., Barley, A., Willis, S., Thomas, J., Verghese, M., & Boateng, J. (2020). Effect of different solvents on the extraction of phytochemicals in colored potatoes. *Food and Nutrition Sciences*, 11(10), 942–954.
<https://doi.org/10.4236/fns.2020.1110066>
- Tiwari, M., Dubey, V., & Srivastava, N. (2020). Pharmacognostic analysis of *Bougainvillea glabra*. *European Journal of Molecular & Clinical Medicine*, 7(11), 7091–7116.
- Tomar, R. ., & Sisodia, S. . (2013). Estimation of phenolic content, total flavonoids and *in vitro* antioxidant activity of *Annona squamosa* Linn. and *Bougainvillea glabra* Choisy. *Journal of Global Pharma Technology*, 3(5), 11–14.
- Truong, D. H., Nguyen, D. H., Ta, N. T. A., Bui, A. V., Do, T. H., & Nguyen, H. C. (2019). Evaluation of the use of different solvents for phytochemical constituents, antioxidants, and *in vitro* anti-inflammatory activities of *Severinia buxifolia*. *Journal of Food Quality*, 2019.
<https://doi.org/10.1155/2019/8178294>
- Usman, A., Mohammed, Y., Muhammed, H. O., Usman, N. L., & Zakari, A. H. (2020). Phytochemical Screening and Antioxidant Activity of *Balanites aegyptiaca* Root Bark Extracts: Influence of solvent. *Communication in Physical Sciences*, 5(2), 156–164. <https://doi.org/https://journalcps.com/>
- Wakeel, A., Jan, S. A., Ullah, I., Shinwari, Z. K., & Xu, M. (2019). Solvent polarity mediates phytochemical yield and antioxidant capacity of *Isatis tinctoria*. *PeerJ*, 7(10), 1–19. <https://doi.org/10.7717/peerj.7857>
- Waszkowiak, K., Gliszczyńska-Źwigło, A., Barthet, V., & Skrety, J. (2015). Effect of extraction method on the phenolic and cyanogenic glucoside profile of flaxseed extracts and their antioxidant capacity. *JAOCs, Journal of the American Oil Chemists' Society*, 92(11–12), 1609–1619.
<https://doi.org/10.1007/s11746-015-2729-x>
- Wesely, E., Johnson, M., MS, R., & Kavitha, P. and. (2010). *In vitro* propagation of *Boerhaavia diffusa* L. through direct and indirect organogenesis. *Journal*

- of *Chemical and Pharmaceutical Research*, 2(5), 339–347.
<https://doi.org/10.1021/ac00220a739>
- Wong, & Kitts, D. (2006). Studies on the dual antioxidant and antibacterial properties of parsley (*Petroselinum crispum*) and cilantro (*Coriandrum sativum*) extracts. *Food Chemistry*, 97(3), 505–515.
<https://doi.org/10.1016/j.foodchem.2005.05.031>
- Wong, S. P., Leong, L. P., & William Koh, J. H. (2006). Antioxidant activities of aqueous extracts of selected plants. *Food Chemistry*, 99(4), 775–783.
<https://doi.org/10.1016/j.foodchem.2005.07.058>
- Yaseen, M., Ahmad, T., Sablok, G., Standardi, A., & Hafiz, I. A. (2013). Review: role of carbon sources for *in vitro* plant growth and development. *Molecular Biology Reports*, 40(4), 2837–2849. <https://doi.org/10.1007/s11033-012-2299-z>
- Yilmaz, Y., & Toledo, R. T. (2004). Health aspects of functional grape seed constituents. *Trends in Food Science and Technology*, 15(9), 422–433.
<https://doi.org/10.1016/j.tifs.2004.04.006>
- Younas, M., Drouet, S., Nadeem, M., Giglioli-Guivarc'h, N., Hano, C., & Abbasi, B. H. (2018). Differential accumulation of silymarin induced by exposure of *Silybum marianum* L. callus cultures to several spectres of monochromatic lights. *Journal of Photochemistry and Photobiology*, 184(April), 61–70.
<https://doi.org/10.1016/j.jphotobiol.2018.05.018>
- Zahid, N. A., Jaafar, H. Z. E., & Hakimian, M. (2021). Micropropagation of ginger (*Zingiber officinale* Roscoe) 'bentong' and evaluation of its secondary metabolites and antioxidant activities compared with the conventionally Propagated Plant. *Plants*, 10, 630.
<https://doi.org/doi.org/10.3390/plants10040630>
- Zahir, A., Ahmad, W., Nadeem, M., Giglioli-Guivarc'h, N., Hano, C., & Abbasi, B. H. (2018). *In vitro* cultures of *Linum usitatissimum* L. Synergistic effects of mineral nutrients and photoperiod regimes on growth and biosynthesis of lignans and neolignans. *Journal of Photochemistry and Photobiology*, 187(April), 141–150. <https://doi.org/10.1016/j.jphotobiol.2018.08.009>
- Zang, Q., Zhou, L., Zhuge, F., Yang, H., Wang, X., & Lin, X. (2016). Callus induction and regeneration via shoot tips of *Dendrocalamus hamiltonii*. *SpringerPlus*, 5, 1799. <https://doi.org/10.1186/s40064-016-3520-7>
- Zarei, M., Salehi, H., & Jowkar, A. (2017). Effects of temperature and season on *in vitro* establishment and shoot multiplication of *Picea abies* (L.) H. Karst. *International Journal of Horticultural Science and Technology*, 4(1), 51–56.
<https://doi.org/10.22059/ijhst.2018.204818.115>
- Zengin, G., & Aktumsek, A. (2014). Investigation of antioxidant potentials of solvent extracts from different anatomical parts of *Asphodeline anatolica* E. *African Journal of Traditional, Complementary and Alternative Medicines*,

11(2), 481–488. <https://doi.org/http://dx.doi.org/10.4314/ajtcam.v11i2.37>

Zhang, Q. (2015). Effects of extraction solvents on phytochemicals and antioxidant activities of walnut (*Juglans regia* L.) green husk extracts. *European Journal of Food Science and Technology*, 3(5), 15–21.

Zhong, J. -j, Seki, T., Kinoshita, S. -i, & Yoshida, T. (1991). Effect of light irradiation on anthocyanin production by suspended culture of *Perilla frutescens*. *Biotechnology and Bioengineering*, 38(6), 653–658. <https://doi.org/10.1002/bit.260380610>

Zhou, X., Yang, B., Stanton, C., Ross, R. P., Zhao, J., Zhang, H., & Chen, W. (2020). Comparative analysis of *Lactobacillus gasserii* from Chinese subjects reveals a new species-level taxa. *BMC Genomics*, 21(1), 1–16. <https://doi.org/10.1186/s12864-020-6527-y>

Zoratti, L., Karppinen, K., Escobar, A. L., Häggman, H., & Jaakola, L. (2014). Light-controlled flavonoid biosynthesis in fruits. *Frontiers in Plant Science*, 5, 1–17. <https://doi.org/10.3389/fpls.2014.00534>