



**SEED QUALITY OF HYACINTH BEANS (*Lablab purpureus* L.)
INFLUENCED BY DRYING METHODS AND STORAGE TEMPERATURES**

By

ALIYAH BT MOHD YAHAYA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of
Master of Science**

November 2020

FP 2020 51

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**SEED QUALITY OF HYACINTH BEANS (*Lablab purpureus* L.)
INFLUENCED BY DRYING METHODS AND STORAGE TEMPERATURES**

By

ALIYAH BT MOHD YAHAYA

November 2020

Chairman : Professor Uma Rani a/p Sinniah, PhD
Faculty : Agriculture

Lablab purpureus or known as Hyacinth bean is a high-value legume that has potential as an important vegetable crop due to its dietary value which is high in protein content. It also has anti-cancer properties but the crop has remained unexploited. To promote the cultivation of this legume good quality seeds would be a prerequisite. Apart from the field related production factors, problems that often occur in seed production are the post-harvest handling which includes drying and storage. Therefore, this study was conducted to determine suitable drying method and storage temperature for Hyacinth bean seed. Seeds were harvested at 30 days after anthesis (DAA) and subjected to different drying methods namely sun, shade, oven ($35\pm 2^{\circ}\text{C}$) and drying beads to obtain target moisture content (14, 12, 10%). The experiment adopted the Completely Randomized Design (CRD) with three replicates. Dried samples were taken randomly for seed physical characteristics, moisture content and germination. Results showed that, Hyacinth bean seeds dried using oven and drying beads attained 10% moisture content within 24 hours while those dried under the sun and shade required up to 72 hours to reach 10% MC. The most rapid rate of drying was by using drying beads and seeds dried with drying beads recorded the highest germination percentage (71%) and had high seed vigour index (20.46). Both shade and drying beads gave good results with drying beads being more economical for adoption by farmers. In contrast, sun and oven drying had large number of dead seeds. Bead-dried seeds were then stored at two storage temperature which are ambient ($27.7\text{-}34.2^{\circ}\text{C}$; $74\pm 5\%$ RH) and fridge ($2.0\text{-}5.0^{\circ}\text{C}$; $25\pm 5\%$ RH) for six months. Experiment was arranged in CRD with three replicates. The results revealed that after six months of storage, seeds stored under refrigerated temperature at $\leq 5.0^{\circ}\text{C}$ recorded higher germination ($\geq 70\%$) compared with ($\leq 50\%$) in ambient temperature. Seed vigour index was also higher (17.49) when stored under refrigerated temperature compared to ambient (8.67). In addition, under ambient declining in antioxidant enzymes activities, germination performance and seedling growth were recorded after six months of storage. Therefore, it is best stored refrigerated ($\leq 5^{\circ}\text{C}$) in order to retain

storability. It is concluded that Hyacinth seeds were best to bead-dried to 10% MC and further stored under refrigerated temperature in which the germination and defence mechanism of the Hyacinth were maintained.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**KUALITI BIJI BENIH KACANG KARA (*Lablab purpureus* L.) DIPENGARUHI
KAEDAH PENGERINGAN DAN SUHU PENYIMPANAN**

Oleh

ALIYAH BT MOHD YAHAYA

November 2020

Pengerusi : Professor Uma Rani a/p Sinniah, PhD
Fakulti : Pertanian

Lablab purpureus atau Kacang Kara merupakan kekacang bernilai tinggi yang mempunyai potensi untuk menjadi sayuran penting berikutan kandungan protein yang tinggi. Kacang ini juga mempunyai sifat anti-kanser. Walaupun mempunyai pelbagai ciri-ciri yang baik, kacang ini masih tidak dieksploitasi. Masalah yang sering berlaku dalam penghasilan biji benih adalah pengendalian pasca-tuai termasuk pengeringan dan penyimpanan biji benih. Oleh itu, kajian ini telah dijalankan untuk mengenalpasti kaedah pengeringan dan suhu simpanan terbaik bagi Kacang Kara. Biji benih dituai 30 hari selepas anthesis dan dikeringkan menggunakan kaedah pengeringan yang berbeza iaitu matahari, teduh, ketuhar ($35\pm 2^{\circ}\text{C}$) dan manik zeolit. Kajian ini menerima pakai reka bentuk rawak sepenuhnya dengan tiga replikasi untuk penilaian fizikal, kelembapan dan percambahan biji benih. Kaedah yang berlainan telah mempengaruhi masa yang diambil untuk mencapai nilai sasaran kelembapan (14%, 12%, 10%) dan juga peratus percambahan. Keputusan menunjukkan bahawa biji benih Kacang Kara yang dikeringkan menggunakan ketuhar dan manik zeolit mencapai nilai sasaran kelembapan (10%) dalam tempoh 24 jam manakala, kaedah pengeringan di bawah matahari dan teduh, memerlukan sehingga 72 jam untuk mencapai 10%. Kaedah pengeringan terpantas adalah menggunakan manik zeolit dan juga tertinggi bagi peratus percambahan (71%) dan kecergasan biji benih (20.46). Manik zeolit dan pengeringan di bawah teduh berjaya mengekalkan kualiti percambahan biji benih Kacang Kara. Namun, manik zeolit lebih menjimatkan bagi petani dan pengeluar biji benih berikutan penggunaan berulang kali. Sebaliknya, pengeringan di bawah matahari dan ketuhar mencatatkan bilangan biji benih mati yang tinggi. Biji benih dikeringkan oleh zeolit telah disimpan pada dua suhu berlainan iaitu ambien ($27.7\text{-}34.2^{\circ}\text{C}$; $74\pm 5\%$ RH) dan peti sejuk ($2.0\text{-}5.0^{\circ}\text{C}$; $25\pm 5\%$ RH) untuk tempoh 6 bulan. Kajian disusun dalam reka bentuk rawak dengan tiga replikasi. Sepanjang tempoh penyimpanan, peratus percambahan adalah ketara dipengaruhi oleh suhu penyimpanan. Hasil kajian mendedahkan bahawa setelah disimpan 6 bulan, biji benih yang disimpan pada suhu peti sejuk mencatatkan percambahan yang lebih

tinggi ($\geq 70\%$) berbanding dengan ($\leq 50\%$) suhu ambien. Kecergasan biji benih juga lebih tinggi (17.49) apabila disimpan pada suhu peti sejuk berbanding suhu ambien (8.67). Suhu ambien mencatatkan penurunan bagi aktiviti enzim antioksidan, prestasi percambahan dan pertumbuhan anak benih. Dengan itu, biji benih Kacang Kara adalah sesuai untuk disimpan pada suhu peti sejuk untuk memastikan keboleh simpanan benih. Secara kesimpulan, biji benih Kacang Kara disyorkan untuk dikeringkan menggunakan manik zeolit ke 10% dan seterusnya disimpan pada suhu peti sejuk bagi mengekalkan kualiti percambahan dan mekanisme pertahanan biji benih.



ACKNOWLEDGEMENTS

Alhamdulillah all praise to Allah for the health, the wisdom and the perseverance that He had bestowed upon me throughout this study. It would not be possible to complete this study without His blessing.

First and foremost, I would like to express my sincere gratitude especially to my supervisor Professor Uma Rani a/p Sinniah, who has whole heartedly guided me in achieving the goal and for the continuous support during this research, for her patience, motivation, enthusiasm and immense knowledge. Without her guidance and persistent help, this dissertation would not have been possible.

Not forgetting my co-supervisor, Dr. Azizah binti Misran and all supportive lecturers for priceless guidance in this journey. I really appreciate the guidance given by other supervisor as well as the panels especially during my project presentation that has improved my presentation skills, thanks to their comments and advice.

A special thanks to my parents, Mohd Yahaya bin Hj. Sheikh Ali and Arfah bt Abd Manan and all my family members for supporting me mentally and physically. I will not have overcome the pressure and obstacles encountered during this journey, if not for them. Words cannot express how grateful I am for all the sacrifices made on my behalf. Your prayer for me was what sustained me this far.

I would like to thank my friends who have been my helping hand throughout this journey. Thanks to my fellow friends who supported me in writing and encouraged me to strive towards my goal, for the stimulating discussions, for the sleepless nights we were working together before the deadlines and for all the fun we had. Thanks for everything and success is meant for us all.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Uma Rani a/p Sinniah, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Azizah binti Misran, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ZALILAH BINTI MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 09 March 2022

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Aliyah bt Mohd Yahaya

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman of
Supervisory
Committee:

Uma Rani a/p Sinniah

Signature: _____

Name of Chairman of
Supervisory
Committee:

Azizah binti Misran

TABLE OF CONTENTS

| | Page |
|--|-------------|
| ABSTRACT | i |
| ABSTRAK | iii |
| ACKNOWLEDGEMENTS | v |
| APPROVAL | vi |
| DECLARATION | vii |
| LIST OF TABLES | xii |
| LIST OF FIGURES | xiv |
| LIST OF ABBREVIATIONS | xvii |
| CHAPTER | |
| 1 INTRODUCTION | 1 |
| 2 LITERATURE REVIEW | 4 |
| 2.1 <i>Lablab purpureus</i> L. and Its Importance | 4 |
| 2.2 Botany of <i>Lablab purpureus</i> L. | 5 |
| 2.3 Seed Quality | 7 |
| 2.3.1 Pre-Harvest Factors Affecting Seed Quality | 7 |
| 2.3.2 Post-Harvest Factors Affecting Seed Quality | 9 |
| 2.4 Seed Drying | 10 |
| 2.4.1 Desiccation Tolerance and Seed Viability | 11 |
| 2.4.2 Open and Closed Drying Technique | 12 |
| 2.4.3 Advantages and Disadvantages of Different Drying Methods | 14 |
| 2.5 Seed Storage | 14 |
| 2.5.1 Responses of Legumes during Storage | 15 |
| 2.5.2 Mechanisms of Seed Deterioration in Association with Enzyme | 16 |
| 2.5.3 Antioxidant Enzyme Systems during Storage | 17 |
| 3 INFLUENCE OF DRYING USING DIFFERENT METHODS ON SEED QUALITY OF HYACINTH BEANS | 19 |
| 3.1 Introduction | 19 |
| 3.2 Materials and Method | 20 |
| 3.2.1 Seed Materials and Drying Treatments | 21 |
| 3.2.2 Experimental Design and Layout | 22 |
| 3.2.3 Seed Quality and Performance Evaluation | 22 |
| 3.2.3.1 Seed Moisture Content | 22 |
| 3.2.3.2 Standard Germination Test | 22 |

| | | | |
|----------|---------|--|-----------|
| | 3.2.3.3 | Electrical Conductivity Test | 23 |
| | 3.2.3.4 | Seedling Vigour Index | 23 |
| | 3.2.3.5 | Germination Rate Index | 23 |
| | 3.2.3.6 | Seedling Length and Dry Weight | 23 |
| | 3.2.4 | Data Analysis | 24 |
| 3.3 | | Results and Discussion | 24 |
| | 3.3.1 | Removal of Moisture as Influenced by the Drying Methods | 24 |
| | 3.3.2 | Germination Performance of Hyacinth Bean Seed | 26 |
| | 3.3.3 | Vigour Assessments of Hyacinth Bean Seed | 30 |
| | 3.3.4 | Seedling Performance of Hyacinth Bean Seed | 34 |
| 3.4 | | Conclusion | 38 |
| 4 | | INFLUENCE OF TWO TEMPERATURES ON STORAGE OF HYACINTH BEAN SEEDS | 39 |
| | 4.1 | Introduction | 39 |
| | 4.2 | Materials and Method | 40 |
| | 4.2.1 | Seed Materials and Storage Temperature Treatments | 40 |
| | 4.2.2 | Seed Quality and Performance Evaluation | 40 |
| | 4.2.3 | Antioxidant Enzyme Analysis | 40 |
| | | 4.2.3.1 Catalase (EC1.11.1.6) Assay | 40 |
| | | 4.2.3.2 Guaiacol Peroxidase (EC1.11.1.7) Assay | 41 |
| | 4.2.4 | Data Analysis | 41 |
| | 4.3 | Results and Discussion | 42 |
| | 4.3.1 | Moisture Content | 42 |
| | 4.3.2 | Germination Performance of Hyacinth Bean Seed | 43 |
| | 4.3.3 | Vigour Assessments of Hyacinth Bean Seed | 44 |
| | 4.3.4 | Seedling Performance of Hyacinth Bean Seed | 48 |
| | 4.3.5 | Antioxidant Enzyme Activities | 52 |
| | 4.4 | Conclusion | 55 |
| 5 | | SUMMARY, CONCLUSION AND RECOMMENDATION | 56 |
| | | REFERENCES | 59 |
| | | APPENDICES | 70 |
| | | BIODATA OF STUDENT | 74 |
| | | PUBLICATION | 75 |

LIST OF TABLES

| Table | | Page |
|-------|---|------|
| 3.1 | Drying treatments of Hyacinth bean seeds used in this study | 21 |
| 3.2 | Hyacinth bean seed germination percentage after drying using different methods | 26 |
| 3.3 | Hyacinth bean seedling vigour as influenced after drying using different methods | 31 |
| 3.4 | Hyacinth bean seed testa permeability after drying using different methods | 33 |
| 3.5 | Hyacinth bean seedling height after drying using different methods | 35 |
| 3.6 | Hyacinth bean seedling root length after drying using different methods | 36 |
| 3.7 | Hyacinth bean seedling dry weight after drying using different methods | 37 |
| 4.1 | Hyacinth bean seed moisture content after being stored for 6 months under different storage temperature | 42 |
| 4.2 | Hyacinth bean seed testa permeability after being stored for 6 months under different storage temperature | 45 |
| 4.3 | Hyacinth bean seedling vigour after being stored for 6 months under different storage temperature | 47 |
| 4.4 | Hyacinth bean seed germination rapidity after being stored for 6 months under different storage temperature | 48 |
| 4.5 | Hyacinth bean seed seedlings height after being stored for 6 months under different storage temperature | 49 |
| 4.6 | Hyacinth bean seedling root length after being stored for 6 months under different storage temperature | 50 |
| 4.7 | Hyacinth seedling dry weight after being stored for 6 months under different storage temperature | 51 |
| 4.8 | CAT activity of Hyacinth bean seeds after being stored for 6 months under different storage temperature | 53 |

4.9

POD activity of Hyacinth bean seeds after being stored
for 6 months under different storage temperature

54



LIST OF FIGURES

| Figure | | Page |
|--------|--|------|
| 2.1 | Varietal diversity of <i>Lablab purpureus</i> L. sweet in terms of pod size, shape and colour harvested during the mature stage. These varieties were collected from diverse habitat and geographical areas of Eastern Uttar Pradesh, India | 5 |
| 2.2 | Purple Hyacinth bean with purple flower, suitable for food and widely growing in gardens due to attractive colour | 6 |
| 2.3 | Development of seed dry weigh, viability, vigour (relative values) and moisture content (%) in time from seed filling to storage and also indicating the period of maturation drying (MD) between the moment of physiological maturity (PM) and harvest maturity (HM). | 8 |
| 2.4 | Drying beads are used in drying for improving seed quality for smallholders | 13 |
| 3.1 | Visual aspects of <i>Lablab purpureus</i> L. purple variety at each growing stage (A) vegetative growth (B) flowering (C) pod formation | 20 |
| 3.2 | Effects of different drying methods (A) drying beads (B) oven (C) shade drying and (D) sun drying on drying duration of Hyacinth bean seeds to achieve target moisture content of 14%, 12% and 10% MC | 25 |
| 3.3 | Effects of sun, oven, drying beads and shade drying of Hyacinth bean seeds on the number of germinated seedlings after 14 th DAS | 27 |
| 3.4 | (A) Effects of different position of seeds within the pod on germination of Hyacinth seeds B) Variation of seed size within Hyacinth pod at middle part of the pod and at tip of the pod | 29 |
| 3.5 | Pod formation and maturation of <i>Lablab purpureus</i> L., and changes in pod colour occurs from purple to light brown starting at 30-40DAA | 30 |

| | | |
|-----|---|----|
| 3.6 | Effects of different drying methods and moisture content on seedling vigour index of Hyacinth bean seeds | 32 |
| 3.7 | Comparison on the seed membrane integrity of freshly harvested seeds and when subjected to two different drying methods (drying beads-membrane integrity was less reduced) and (oven drying-membrane integrity was greatly reduced). | 34 |
| 3.8 | Effects of different drying methods to 10% target moisture content on seedlings height of Hyacinth bean seedlings | 35 |
| 4.1 | (A) Effects of different storage temperature (ambient and refrigerator) on germination of Hyacinth bean seeds after 6 months of storage (B) Effects of different moisture content (14%, 12%, 10%) on germination of Hyacinth bean seeds after 6 months of storage | 43 |
| 4.2 | Effects of different storage temperature and seed moisture content on seed physical conditions of Hyacinth bean seeds. | 44 |
| 4.3 | Comparison on the seed membrane integrity of seeds stored under ambient (membrane integrity was greatly reduced) and when seeds stored under cold temperature (drying beads-membrane integrity was less reduced) | 46 |

LIST OF APPENDICES

| Appendix | | Page |
|----------|---|------|
| 1 | Origin and characteristics of Hyacinth bean cultivar used in this thesis | 70 |
| 2 | Identification of Hyacinth bean plant (purple variety) used in this study by Malaysian Agricultural Research and Development Institute (MARDI) | 70 |
| 3 | Data on temperature and relative humidity of storage environment under A) ambient and B) fridge condition for 6 months | 71 |
| 4 | Mean square values for analysis of variance on the effect of different drying methods and seed moisture content on Hyacinth bean seed quality | 72 |
| 5 | Mean square values for analysis of variance on the effect of different storage temperature and seed moisture content on Hyacinth germination performance, seedling vigour assessments, seedling performance and changes in antioxidant enzyme activities at 6 months of storage | 73 |

LIST OF ABBREVIATIONS

| | |
|-----------------|---|
| μm | Micromolar |
| μmol | Micromole |
| AA | Accelerated Aging |
| AOSA | Association of Official Seed Analyst |
| ANOVA | Analysis of Variance |
| CAT | Catalase |
| cm | Centimetre |
| CRD | Completely Randomized Design |
| df | Degree of Freedom |
| EC | Electrical Conductivity of Seed Leachates |
| GRI | Germination Rate Index |
| HM | Harvest Maturity |
| hr | hour |
| ISTA | International Seed Testing Association |
| LSD | Least Significant Difference |
| M | Molarity |
| MD | Maturity Desiccation |
| min | minutes |
| mm | Millimolar |
| mmol | Millimole |
| mol | Mole |
| nmol | Nanomole |
| PM | Physiological Maturity |

| | |
|-----|-------------------------|
| POD | Peroxidase |
| RH | Relative Humidity |
| ROS | Reactive Oxygen Species |
| rpm | Revolution per Minute |
| SVI | Seedling Vigour Index |
| WRR | Water Rate Removal |



CHAPTER 1

INTRODUCTION

Legumes belonging to the family Fabaceae are major components in human diet and it is often consumed as vegetables. Generally, legumes are rich in protein, fibre, iron, B-vitamins and low in fat (Graham and Vance, 2003). Legume seeds are the major source of food second only to cereals (Poaceae) and highly nutritious. There are a number of popular legumes in Malaysia which is used as vegetable such as long beans, winged beans, French beans and green peas. Some legumes have been used commonly while others have remained underutilized. One such example of an underutilized bean in Malaysia is *Lablab purpureus* or commonly known as Hyacinth bean. Hyacinth bean originated from Africa, but today, Bangladesh is the leading producer in the world with a total production area of around 48,000 ha. This bean is widely distributed in many tropical countries such as Malaysia, Indonesia and Philippines (Gowda, 2013).

In Malaysia, Hyacinth bean is known as 'Kacang Kara' and according to Cook et al. (2005) it is a high-value legume that has the potential of being an important vegetable crop because of its high protein content (20-28%) nearly three times compared with most beans. In addition it is also known to have numerous medicinal advantages such as anti-cancer, anti-diabetic, anti-inflammatory, helps in eczema and also used for the treatment of anaemia (Al-snafi, 2017). In Asia, Hyacinth bean is mainly grown for its pods and consumed as vegetable, while dry seeds are used in making tofu and tempeh (Subagio and Morita, 2008).

Due to the goodness of this bean, Malaysia is importing it from Thailand and Bangladesh (UC Davis, 2017). Despite the suitable environmental condition to grow this bean, the crop has remained unexploited locally. In view of its high protein content, other medicinal uses as well as the growing popularity among Malaysian to consume this bean, initiatives should be taken to recommend this crop as future legume in fulfilling food security demand. This is in line with the agenda mentioned in the National Agricultural Policy 4 (NAP4) whereby vegetables are prioritized for development in Malaysia for food security. It is important to know that Hyacinth bean is grown using seeds as the planting material and low supply of good quality seeds will be a major constraint for Hyacinth bean crop production.

Good quality seeds are crucial in any crop production system, and determined both by pre-harvest management and post-harvest handling. In many instances the post-harvest component is underestimated and less attention is given. Shelar et al, (2008) stated the drop in seed quality under tropical conditions is much more extreme because environmental conditions in these areas are challenging to retain seed viability. As a leading producer of Hyacinth bean

seeds, Bangladesh recorded large postharvest losses due to traditional method of drying seeds (UC Davis, 2017). Bangladeshi seed companies estimate a loss of 5-10% of their seeds worth millions of dollars when dried on tarpaulin under the sun due to poor drying techniques.

Method of drying has been reported to effect seed quality. A number of drying techniques including natural and artificial drying used in legumes have been reported by many researches (Scariot et al., 2017; Ellis and Roberts 1991). Drying is mostly done naturally by exposing seeds to direct sunlight or under shade in open fields. Although it is a cost-saving method, sun-drying has been reported to be slow and weather dependent (Muckle and Stirling, 1971). For example, a study by Ali et al., (2015) suggested soybean seeds can be sun dried to $\leq 8\%$ MC without affecting seed quality. However, groundnut deteriorated rapidly when sun-dried and resulted in lower germination (Hon, 1977). Groundnut was best dried using oven method. Larger companies and seed producers preferred oven drying where constant temperature can be maintained (Babiker et al., 2010).

It has been shown that a wide gradient of desiccation sensitivity or tolerance exists among legume species (Berjak and Pammenter, 2008). Ellis and Roberts (1991) have proposed other methods of drying such as sun, shade, freeze and refrigeration drying with low relative humidity but concluded that response to drying can be species specific which depends on the ability to tolerate desiccation. Hence, there is a need to determine proper drying method for Hyacinth which is still unknown. A simple, low-cost but efficient method for farmers and seed producers to adopt would be useful.

Recently, drying using zeolite drying beads have been utilized particularly in small-scale drying such as for gene bank. Drying beads, a zeolite-based desiccant was developed by Rhino Tech Thailand which potentially enables seedsman to dry seeds to safe moisture content. It is said to be effective, economical and can be used repeatedly over time. Drying using the beads ensured better quality seeds and was able to prolong storage. A study by Nassari et al., (2014) on tomato seeds confirmed that rapid drying took place when drying beads were used compared with sun-drying with maintained viability and vigour. Another study by Atilia and Zaitialia (2016) in chilly, seeds dried using bead drying had better quality compared with sun dried seeds. Furthermore, the application of drying beads in drying 200 tons of vegetable seed in Bangladesh proved to provide an estimated 100,000 farmers with superior seed (Anon, 2018). Thus, this study attempts to use zeolite beads on Hyacinth bean seed to see its drying efficacy.

According to Gupta et al., (2005) when seeds are in dry state, it is necessary to store them appropriately. Seed quality can remain at its initial level or drop to an unacceptable level for planting during storage. Surki et al., (2012) stated that unfavourable storage temperature contributes to acceleration of seed deterioration. Deterioration is related to reactive oxygen species (ROS) and

yearly losses due to deterioration can amount to 25% of harvested crop. ROS are usually produced from stress conditions such as high temperature during storage and could be demolished by scavenger enzymes such as catalase and peroxides (Jyoti and Malik, 2013). Peroxidase (POD), catalase (CAT) and superoxide dismutase (SOD), and are main enzymatic antioxidants that function as a detoxifying mechanism that stops the degrading radicals produced during stress and thus prevents seed from deteriorating. Few studies have revealed connection concerning seed deterioration and deprivation of specific antioxidant enzyme activities in legumes seed such as pea (Yao et al., 2012) and soybean (Tian et al., 2008). However, currently no specific research has been conducted to understand the connection between storage temperature and deterioration related with antioxidant enzyme activities in Hyacinth bean seeds.

Hyacinth bean has a potential to be commercialized widely in Malaysia however, a good protocol on how to dry and store the seeds is yet to be established and is very much needed. Therefore, this study was conducted with the following objectives:

1. To determine the influence of drying using different methods on seed quality of *Lablab purpureus*
2. To evaluate the effects of temperature on seed viability of *Lablab purpureus* during storage

REFERENCES

- Abdul-Baki, A.A., and Anderson, J.D. (1973). Vigor determination in soybean seed by multiple criteria 1. *Crop science*, 13(6): 630-633.
- Asbrouck, V.J., and Taridno, P. (2009). Using the single seed oxygen consumption measurement as a method of determination of different seed quality parameters for commercial tomato seed samples. *Asian J. Food & Agro-Industry*, 2: 88-95.
- Adebisi, A.A., and Bosch, C.H. (2004). *Lablab purpureus* L. sweet. *Plant resources of tropical Africa (PROTA)*, (2): 343-348.
- Aebi, H. (1984). Catalase in vitro. *Method of enzymology*, 105:121-126
- Afonso Júnior, P.C., and Corrêa, P.C. (2000). Immediate and latent drying effects of bean seeds harvested with different moisture levels. *Ciência e Agrotecnologia*, 24: 33-40.
- Ajouri, A., Asgedom, H., and Becker, M. (2004). Seed priming enhances germination and seedling growth of barley under conditions of P and Zn deficiency. *Journal of Plant Nutrition and Soil Science*, 167(5): 630-636.
- Alam, M.M., Alam, A.K.M., Khandker, S., Alim, M.A., and Haque, S. (2009). Effect of sulfur on late jute seed production in different AEZ of Bangladesh. *Interl. J. Sustainable Crop Produ*, 4(3): 33-37.
- Ali, M., Rahman, M., Rahman, M., Hossain, M., and Asaduzzaman, M. (2015). Effect of Drying Method on Quality of Soybean Seed. *Bangladesh Agronomy Journal*, 18(1): 53-57.
- Almeida, D.P., Mendes, U.C., Resende, O., Donadon, J.R., Da Rocha, A.C., and Oliveira, D.E.C. (2016). Effect of drying on the physical properties of adzuki bean. *Semina: Ciências Agrárias*, 37(6): 3871-3880.
- Al-Snafi, A.E. (2017). The pharmacology and medical importance of *Dolichos lablab* (*Lablab purpureus*) -A review. *IOSR Journal of Pharmacy*, 7(2):22-30.
- Altuntas, E., and Demirtola, H. (2007). Effect of moisture content on physical properties of some grain legume seeds. *New Zealand Journal of Crop and Horticultural Science*, 35(4): 423-433.
- Andrade, A.C.S. (2001). The effect of moisture content and temperature on the longevity of heart of palm seeds (*Euterpe edulis*). *Seed Science and Technology*, 29(1): 171-182.
- Angelovici, R., Galili, G., Fernie, A.R., and Fait, A. (2010). Seed desiccation: a bridge between maturation and germination. *Trends in plant science*, 15(4): 211-218.

- AOSA. (2002). *Seed vigor testing handbook*. Association of Official Seed Analysts. Las Cruces, NM.
- Arah, I.K., Amaglo, H., Kumah, E.K., and Ofori, H. (2015). Preharvest and postharvest factors affecting the quality and shelf life of harvested tomatoes: a mini review. *International Journal of Agronomy*, 1-7.
- Atici, O.K.K.E.S., Agar, G.U.L.E.R.A.Y., and Battal, P.E.Y.A.M.I. (2007). Influence of long term storage on plant growth substance levels, germination and seedling growth in legume seeds stored for 37 years. *Indian Journal of Plant Physiology*, 12(1): 1.
- Babiker, A.Z., Dulloo, M.E., Balla, M.A.M. El, and Ibrahim, E.T. (2010). Effects of low cost drying methods on seed quality of *Sorghum bicolor* L. monech. *African Journal of Plant Science*, 4(9): 339–345.
- Bailly, C., Bogatek-Leszczynska, R., Côme, D., and Corbineau, F. (2002). Changes in activities of antioxidant enzymes and lipoxygenase during growth of sunflower seedlings from seeds of different vigour. *Seed Science Research*, 12(1): 47.
- Balesevic-Tubic, S., Tatic, M., Dordevic, V., Nikolic, Z. and Dukic, V. (2010). Seed viability of oil crops depending on storage conditions. *Helia*, 33(52):153-160.
- Bashir, A.A., Sinha, J.P., Jha, G. K., and Chopra, S. (2016). Drying kinetics for vegetable seeds with zeolite beads. *Indian Journal of Agricultural Sciences*, 86:1630-1634.
- Berjak, P., and Pammenter, N.W. (2008). From Avicennia to Zizania: seed recalcitrance in perspective. *Annals of botany*, 101(2):213-228.
- Bewley, J.D., Bradford, K.J., Hilhorst, H.W., and Nonogaki, H. (2013). Longevity, storage, and deterioration. *Seeds*, 341-376.
- Bhardwaj, R., Sood, M., and Thakur, U. (2014). Effect of storage temperature and period on seed germination of *Rheum australe* : an endangered medicinal herb of high altitude Himalaya. *International Journal of Farm Sciences*, 4(2): 139-147.
- Braga Jr, R.A., Borem, F.M., Dal Fabbro, I.M., Arizaga, R.A., Rabal, H.J., and Trivi, M. (2000). Seed characterization by dynamic speckle patterns: a proposal. *Biological Quality and Precision Agriculture II*, 4203:55-62.
- Champa, W.A.H., Weerakkody, W.A.P., and Palipane, K.B. (2008). Maturity indices for harvesting of Beans (*Phaseolous vulgaris* L.) variety Kentucky wonder green in dry zone cultivations in Sri Lanka. *Tropical Agricultural Research*, 20:123-133.
- Chance, B., and Maehly, A.C. (1955). Assay of catalases and peroxidases, methods in Enzymology, 136(2):764-775.

- Chen, H., Osuna, D., Colville, L., Lorenzo, O., Graeber, K., Kuester, H., and Kranner, I. (2013). Transcriptome-wide mapping of pea seed ageing reveals a pivotal role for genes related to oxidative stress and programmed cell death. *Plos one*, 8(10): 78471.
- Cheng, H.Y., Zheng, G.H., Wang, X.F., Liu, Y., Yan, Y.T., and Lin, J. (2005). Possible involvement of K⁺/Na⁺ in assessing the seed vigor index. *Journal of Integrative Plant Biology*, 47(8): 935-941.
- Copeland, L.O., and McDonald M.B.(1995). Principles of seed science and technology, pp. 113-119. New York and London: Experimental Agriculture.
- Cook, B.G., Pengelly, B.C., Brown, S.D., Donnelly, J.L., Eagles, D.A., Franco, M.A., Hanson, J., Mullen, B.F., Partridge, I.J., Peters, M., and Schultze-Kraft, R. (2005). *Tropical forages*. CSIRO, DPI&F(Qld), CIAT and ILRI, Brisbane, Australia
- Das, S.S., and Fakir, M.S.A. (2014). Pod growth and seed composition in two genotypes of *Lablab purpureus*. *Legume Research*, 37(3): 306–310.
- Deaker, R., Roughley, R. J., and Kennedy, I.R. (2004). Legume seed inoculation technology—a review. *Soil Biology and Biochemistry*, 36(8): 1275-1288.
- Demirkaya, M., Dietz, K.J., and Sivritepe, H.O. (2010). Changes in antioxidant enzymes during ageing of onion seeds. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 38(1): 49-52.
- Devaraj, V.R. (2016). Evaluation of appropriate reference gene for normalization of microRNA expression by real-time PCR in *Lablab purpureus* under abiotic stress conditions. *Biologia*, 71(6): 660-668.
- Dona, M., Balestrazzi, A., Mondoni, A., Rossi, G., Ventura, L., Buttafava, A., and Carbonera, D. (2013). DNA profiling, telomere analysis and antioxidant properties as tools for monitoring ex situ seed longevity. *Annals of Botany*, 111(5): 987-998.
- Donahaye, E.J. (2000). Current status of non-residual control methods against stored product pests. *Crop Protection*, 19(8-10): 571-576.
- Ellis, R.H., and Roberts, E.H. (1980). The influence of temperature and moisture on seed viability period in barley (*Hordeum distichum* L.). *Annals of Botany*, 45(1): 31-37.
- Ellis, R.H., Hong, T.D., and Roberts, E.H. (1991). Seed moisture content, storage, viability and vigour. *Seed Science Research*, 1(4): 275-279.
- Fatin , H.H., Sinniah, U.R., and Martini M.Y. (2019). Importance of Maturity Stage on Quality of *Lablab purpureus* Seeds. ISSAAS International Congress. 18-20th October, Universiti Putra Malaysia, Serdang Selangor Malaysia

- Filho, C.P., Goneli, A.L.D., Masetto, T.E., Martins, E.A.S., and Oba, G.C. (2016). The effect of drying temperatures and storage of seeds on the growth of soybean seedlings. *Journal of Seed Science*, 38(4): 287-295.
- Finch-Savage, W.E., and Bassel, G.W. (2016). Seed vigour and crop establishment: extending performance beyond adaptation. *Journal of experimental botany*, 67(3): 567-591.
- Galani, S., Naz, F., Soomro, F., Jamil, I., Azhar, A., and Ashraf, A. (2011). Seed storage protein polymorphism in ten elite rice (*Oryza sativa* L.) genotypes of Sindh. *African Journal of biotechnology*, 10(7): 1106-1111.
- George, R.A.T. (2011). Leguminosae-beans and related crops. *Tropical vegetable production*, 132-146.
- Gesch, R.W., and Johnson, B.L. (2012). Seed moisture at physiological maturity in oilseed and confectionary sunflower hybrids in the northern US. *Field Crops Research*, 133: 1-9.
- Golezani, G.K., and Mahootchy, H.A. (2009). Changes in seed vigour of faba bean (*Vicia faba* L.) cultivars during development and maturity. *Seed Science and Technology*, 37(3): 713-720.
- Goel, A., Goel, A.K., and Sheoran, I.S. (2003). Changes in oxidative stress enzymes during artificial ageing in cotton (*Gossypium hirsutum* L.) seeds. *Journal of plant physiology*, 160(9), 1093-1100.
- Govindaraj, M., Masilamani, P., Albert, V.A., and Bhaskaran, M. (2017). Role of antioxidant in seed quality-A review. *Agricultural Reviews*, 38(3): 180-190.
- Gowda, R. (1992). Studies on seed senescence and seed vigour in some genotypes of rice (*Oryza Sativa* L.) (Doctoral dissertation, Tamil Nadu Agricultural University, Coimbatore).
- Gupta, M.L., George, D.L., and Parwata, I.G.M.A. (2005). Effect of harvest time and drying on supersweet sweet corn seed quality. *Seed Science and technology*, 33(1): 167-176.
- Graham, P.H., and Vance, C.P. (2003). Legumes: importance and constraints to greater use. *Plant physiology*, 131(3): 872-877.
- Groot, S.P.C., Surki, A.A., De Vos, R.C.H., and Kodde, J. (2012). Seed storage at elevated partial pressure of oxygen, a fast method for analysing seed ageing under dry conditions. *Annals of Botany*, 110(6): 1149-1159.
- Guan, L.M., and Scandalios, J.G. (2002). Catalase gene expression in response to auxin-mediated developmental signals. *Physiologia Plantarum*, 114(2): 288-295.
- Hampton, J.G. (2002). What is seed quality?. *Seed Science and Technology*, 30(1): 1-10.

- Hor, Y.L. (1977). Storage of field crop seeds under Malaysia conditions. *Seed Technology in the Tropics*, 124-134.
- Hong, T.D., and Ellis, R.H. (1996). A protocol to determine seed storage behaviour (1). Bioversity International.
- Illipronti Jr, R.A., Lommen, W.J.M., Langerak, C.J., and Struik, P.C. (2000). Time of pod set and seed position on the plant contribute to variation in quality of seeds within soybean seed lots. *NJAS-Wageningen Journal of Life Sciences*, 48(2): 165-180.
- ISTA (2016). *International rules for seed testing*, International Seed Testing Association, Bassersdorf, Switzerland.
- Jacoby, R.P., Li, L., Huang, S., Pong Lee, C., Millar, A.H., and Taylor, N.L. (2012). Mitochondrial Composition, Function and Stress Response in Plants F. *Journal of Integrative Plant Biology*, 54(11): 887-906.
- Justice, O.L., and Louis, N. (1978). Bass. Principles and Practices of Seed Storage. *Agriculture Handbook*, (506). Department of Agriculture, Science and Education Administration.
- Jyoti, C.P., and Malik. (2013). Seed Deterioration. *International Journal of Life Sciences Biotechnology and Pharma Research*, 2(3): 374-385.
- Kader, M.A. (2005). A Comparison of Seed Germination Calculation Formulae and the Associated Interpretation of Resulting Data. *Journal & Proceeding of the Royal Society of New South Wales*, 138: 65-75.
- Kaewnaee, P., Vichitphan, S., Klanrit, P., Siri, B., and Vichitphan, K. (2011). Effect of accelerated aging process on seed quality and biochemical changes in sweet pepper (*Capsicum annuum* Linn.) seeds. *Biotechnology*, 10(2): 175-182.
- Kandil, A.A., Sharief, A.E., and Sheteiwy, M.S. (2013). Effect of seed storage periods, conditions and materials on germination of some soybean seed cultivars. *American Journal of Experimental Agriculture*, 3(4): 1020.
- Kante, K., and Reddy, C.S. (2013). Anti-diabetic activity of Dolichos lablab (seeds) in Streptozotocin-Nicotinamide induced diabetic rats. *Journal Of Drugs and Medicine*. 5(1): 32-40.
- Kaushal, N., Gupta, K., Bhandhari, K., Kumar, S., Thakur, P., and Nayyar, H. (2011). Proline induces heat tolerance in chickpea (*Cicer arietinum* L.) plants by protecting vital enzymes of carbon and antioxidative metabolism. *Physiology and Molecular Biology of Plants*, 17(3): 203.
- Kapoor, N., Arya, A., Siddiqui, M.A., Amir, A., and Kumar, H. (2010). Seed deterioration in chickpea (*Cicer arietinum* L.) under accelerated ageing. *Asian Journal of Plant Sciences*, 9(3): 158-162.
- Kibinza, S., Vinel, D., Come, D., Bailly, C., and Corbineau, F. (2006). Sunflower

seed deterioration as related to moisture content during ageing, energy metabolism and active oxygen species scavenging. *Physiologia Plantarum*, 128(3): 496-506.

Kranner, I., Chen, H., Pritchard, H.W., Pearce, S.R., and Birtic, S. (2011). Internucleosomal DNA fragmentation and loss of RNA integrity during seed ageing. *Plant Growth Regulation*, 63(1): 63-72.

Krishna, S.P., and Gejjalagere, H.C. (2018). Advancement of Biodegradable Polysaccharide Nanocarriers for Delivery of Herbal Extracts and Bio-Actives. *CRC Press*, 191-205.

Krzyzanowski, F.C., West, S.H., and França Neto, J.D.B. (2006). Drying peanut seed using air ambient temperature at low relative humidity. *Revista brasileira de sementes*, 28(3): 1-5.

Kumar, C., Karim, M.A., and Joardder, M.U. (2014). Intermittent drying of food products: A critical review. *Journal of Food Engineering*, 121: 48-57.

Lassim, M.B., Chin, H.F., (1987). Some trends in the development and maturation of cowpea (*Vigna unguiculata* L.) seeds. *Acta Hort.* 215: 25-30.

Lehner, A., Mamadou, N., Poels, P., Come, D., Bailly, C., and Corbineau, F. (2008). Changes in soluble carbohydrates, lipid peroxidation and antioxidant enzyme activities in the embryo during ageing in wheat grains. *Journal of Cereal Science*, 47(3): 555-565.

Leprince, O., Hendry, G.A.F., and McKersie, B.D. (1993). The mechanisms of desiccation tolerance in developing. *Seed Science Research*, 3: 231-246.

Lu, J.M., Lin, P.H., Yao, Q., and Chen, C. (2010). Chemical and molecular mechanisms of antioxidants: experimental approaches and model systems. *Journal of cellular and molecular medicine*, 14(4): 840-860.

Manish, K.V., Sushil, P., Chitra, D.P., and Yasin, J. (2015). Impact of drying methods on the seed quality of sorghum (*Sorghum bicolor* (L.) Moench). *African Journal of Agricultural Research*, 10(16): 1898–1903.

Marthandan, V., and Jerlin, R. (2017). Effects of seed storage conditions on biochemical changes of freshly harvested high moisture undried rice seeds cv. CO 51. *Int. J. Curr. Microbiol. App. Sci*, 6(12): 2807-2813.

Maass, B.L., and Usongo, M.F. (2007). Changes in seed characteristics during the domestication of the lablab bean (*Lablab purpureus* L. Sweet: Papilionoideae). *Australian Journal of Agricultural Research*, 58(1):9-19.

Mbofung, G.C., Goggi, A.S., Leandro, L.F., and Mullen, R.E. (2013). Effects of storage temperature and relative humidity on viability and vigor of treated soybean seeds. *Crop Science*, 53(3): 1086-1095.

McDonald, M.B. (2004). Orthodox seed deterioration and its repair. *Handbook of Seed Physiology: Applications to Agriculture, Benech-Arnold, RL and*

RA Sanchez (Eds.). *Food Products Press, New York*, 273-304.

- Muasya, R.M. (2001). Crop physiological analysis of seed quality variation in common bean (*Phaseolus vulgaris* L.). (Doctoral dissertation, Wageningen University, Netherlands).
- Muckle, T.B., and Stirling, H.G. (1971). Review of the drying of cereals and legumes in the tropics. *Trop Stored Prod Inform.* 1971 (22): 11-30.
- Murthy, P., and Kumar, B.M. (2004). Studies on seed development and physiological maturity in held bean (*Lablab purpureus* L. sweet). *Legume Research-An International Journal*, 27(2): 134-136.
- Mohammadi, H., Soltani, A., Sadeghipour, H. R., and Zeinali, E. (2012). Effects of seed aging on subsequent seed reserve utilization and seedling growth in soybean. *International Journal of Plant Production*, 5(1): 65-70.
- Moller, I.M., Jensen, P.E., and Hansson, A. (2007). Oxidative modifications to cellular components in plants. *Annu. Rev. Plant Biol.*, 58: 459-481.
- Morris, J.B. (2009). Morphological and reproductive characterization in hyacinth bean, *Lablab purpureus* (L.) Sweet germplasm with clinically proven nutraceutical and pharmaceutical traits for use as a medicinal food. *Journal of Dietary Supplements*, 6(3): 263-279.
- Nagaveni, P.K. (2005). Effect of storage conditions, packing material and seed treatment on viability and vigour of onion seeds (Doctoral dissertation, UAS, Dharwad).
- Nautiyal, P. (2009). Seed and seedling vigour traits in groundnut (*Arachis hypogaea* L.). *Seed Science and Technology*, 37: 721–735.
- Nassari, P.J. Keshavulu K, Manohar R.K. and Reddy, A. R. (2014). Postharvest Drying Of Tomato Seeds To Ultra Low Moisture Safe For Storage Using Desiccant(Zeolite) Beads And Their Effects On Seed Quality. *American Journal Of Research Communication*, 2(4): 74–83.
- Panda, B.B., Mohapatra, S., Mallik, S., and Acharya, P. (2010). Effect of Tamarind seed mucilage on rheological properties: evaluation of suspending properties. *Int. RJ Pharm. Sci*, 1: 8-10.
- Park, S.J., Huang, Y., and Ayoubi, P. (2006). Identification of expression profiles of sorghum genes in response to greenbug phloem-feeding using cDNA subtraction and microarray analysis. *Planta*, 223(5): 932-947.
- Pukacka, S., and Ratajczak, E. (2006). Antioxidative response of ascorbate–glutathione pathway enzymes and metabolites to desiccation of recalcitrant *Acer saccharinum* seeds. *Journal of Plant Physiology*, 163(12): 1259-1266.
- Pradhan, B.K., and Badola, H.K. (2012). Effect of storage conditions and storage periods on seed germination in eleven populations of *Swertia*

- chirayita: a critically endangered medicinal herb in Himalaya. *The Scientific World Journal*, 1-9.
- Pratap, A., and Kumar, J. (2011). History, origin and evolution. *Biology and Breeding of Food Legumes*, 1-18.
- Pratima, K.A.P. (2014). Evaluation of zeolite beads technology for drying vegetable seeds to low moisture content prior to long-term storage in Nepal. *International Journal of Research*, 1(8):140-149.
- Probert, R.J. (2003). Seed viability under ambient conditions, and the importance of drying. *Seed Conservation: Turning Science Into Practice*, 337-365.
- Quiles, M.J. and Lopez, N.I. (2004). Photoinhibition of photosystems I and II induced by exposure to high light intensity during oat plant growth: effects on the chloroplast NADH dehydrogenase complex. *Plant Science*, 166(3): 815-823.
- Raja, K. (2003). Investigations on nursery and main field management techniques for quality seed production of rice hybrid (Doctoral dissertation, Tamil Nadu Agricultural University Coimbatore).
- Rajjou, L., and Debeaujon, I. (2008). Seed longevity: survival and maintenance of high germination ability of dry seeds. *Comptes Rendus Biologies*, 331(10): 796-805.
- Ramanadane, T., Srimathi, P., Malarkodi, K., and Natarajan, K. (2003). Seed extraction in jamun (*Syzygium cuminii* Skeels). *Progressive Horticulture*, 35(2): 221-223.
- Ramos-Solano, B., Algar, E., Gutierrez-Mañero, F.J., Bonilla, A., Lucas, J.A., and García-Seco, D. (2015). Bacterial bioeffectors delay postharvest fungal growth and modify total phenolics, flavonoids and anthocyanins in blackberries. *LWT-Food Science and Technology*, 61(2): 437-443.
- Roberts, E.H. (1972). Storage environment and the control of viability. *Viability of seeds*, 14-58.
- Saha, R.R., and Sultana, W. (2008). Influence of seed ageing on growth and yield of soybean. *Bangladesh Journal of Botany*, 37(1): 21-26.
- Satish, S. and Bhaskaran, M. (2013). Seed dry dressing with botanicals to improve physiological performance of fresh and aged seeds of blackgram (*Vigna mungo* L.) *African Journal of Agricultural Research*, 8(29):4049-4057.
- Scariot, M.A., Tiburski, G., Reichert Júnior, F.W., Radünz, L. L., and Meneguzzo, M.R.R. (2017). Moisture content at harvest and drying temperature on bean seed quality. *Pesquisa Agropecuária Tropical*, 47(1): 93-101.

- Sheahan, C.M. (2012). Plant guide for lablab (*Lablab purpureus*). USDA-Natural Resources Conservation Service, Cape May Plant Materials Center. Cape May, NJ, 8210.
- Shelar, V.R., Shaikh, R.S., and Nikam, A.S. (2008). Soybean seed quality during storage: a review. *Agricultural Reviews*, 29(2): 125-131.
- Siddique, A.B., and Wright, D. (2003). Effects of different drying time and temperature on moisture percentage and seed quality (viability and vigour) of pea seeds (*Pisum sativum* L.). *Asian Journal of Plant Sciences*. 2(13): 978-982.
- Simic, B., Popovic, R., Sudaric, A., Rozman, V., Kalinovic, I., and Cosic, J. (2007). Influence of storage condition on seed oil content of maize, soybean and sunflower. *Agriculturae Conspectus Scientificus*, 72(3): 211-213.
- Sisman, C.B. (2005). Quality Losses in Temporary Sunflower Seed Stores and Influences of Storage Conditions on Quality Losses During Storage. *Journal of Central European Agriculture*, 6(2): 143-150.
- Singh, A., and Abhilash, P.C. (2019). Varietal dataset of nutritionally important *Lablab purpureus* L. sweet from Eastern Uttar Pradesh, India. *Journal Data in Brief*, 24: 103935.
- Singh, U.P., and Prithviraj, B. (1997). Neemazal, a product of neem (*Azadirachta indica*), induces resistance in pea (*Pisum sativum*) against *Erysiphe pisi*. *Physiological and Molecular Plant Pathology*, 51(3): 181-194.
- Souza, F.H., and Filho, J.Ú.L.I.O. (2001). The seed coat as a modulator of seed-environment relationships in Fabaceae. *Brazilian Journal of Botany*, 24(4): 365-375.
- Subagio, A., and Morita, N. (2008). Effects of protein isolate from hyacinth beans (*Lablab purpureus* L. Sweet) seeds on cake characteristics. *Food science and Technology Research*, 14(1): 12-17.
- Surki, A.A., Sharifzadeh, F., and Afshari, R.T. (2012). Effect of drying conditions and harvest time on soybean seed viability and deterioration under different storage temperature. *African Journal of Agricultural Research*, 7(36): 5118-5127.
- Tabatabaei, S.A., (2014). The changes of germination characteristics and enzyme activity of barley seeds under accelerated aging. *Cercetari Agronomice in Moldova*, 48(2): 61-67.
- TeKrony, D.M., Egli, D. B., and Phillips, A.D. (1980). Effect of field weathering on the viability and vigor of soybean seed. *Agronomy journal*, 72(5): 749-753.
- TeKrony, D.M. (2003). Precision is an essential component in seed vigour

testing. *Seed science and technology*, 31(2): 435-447.

- Tian, X., Song, S., and Lei, Y. (2008). Cell death and reactive oxygen species metabolism during accelerated ageing of soybean axes. *Russian Journal of Plant Physiology*, 55(1): 33-40.
- Tilebeni, H.G., and Golpayegani, A. (2011). Effect of seed ageing on physiological and biochemical changes in rice seed (*Oryza sativa* L.). *International Journal of AgriScience*, 1(3): 138-143.
- Tunes, L.M., Corlett, F.M., de A Rufino, C., Vieira, J.F., Tavares, L.C., and Barros, A.C. (2014). The influence of seed coating on the vigor and early seedling growth of barley. *Ciencia e investigación agraria: revista latinoamericana de ciencias de la agricultura*, 41(1): 129-136.
- UC Davis (2017). Drying beads help Bangladesh farmers access better seed. Retrieved on February 20th, 2020, from <https://horticulture.ucdavis.edu/blog/drying-beads-help-bangladesh-farmers-access-better-seed>
- Ushimaru, T., Kanematsu, S., Katayama, M., and Tsuji, H. (2001). Antioxidative enzymes in seedlings of *Nelumbo nucifera* germinated under water. *Physiologia Plantarum*, 112(1): 39-46.
- Walters, C., Mira, S., González-Benito, M.E., and Hill, L.M. (2010). Characterization of volatile production during storage of lettuce (*Lactuca sativa*) seed. *Journal of experimental botany*, 61(14): 3915-3924.
- White, N.D.G., and Jayas, D.S. (1991). Control of insects and mites with carbon dioxide in wheat stored at cool temperatures in nonairtight bins. *Journal of Economic Entomology*, 84(6): 1933-1942.
- Wong, C.C. (2000). The place of silage in ruminant production in the humid tropics. *Fao Plant Production And Protection Papers*: 5-6.
- Xia, F., Chen, L., Sun, Y., and Mao, P. (2015). Relationships between ultrastructure of embryo cells and biochemical variations during ageing of oat (*Avena sativa* L.) seeds with different moisture content. *Acta physiologiae plantarum*, 37(4): 89.
- Xin, X., Tian, Q., Yin, G., Chen, X., Zhang, J., Ng, S., and Lu, X. (2014). Reduced mitochondrial and ascorbate–glutathione activity after artificial ageing in soybean seed. *Journal of Plant Physiology*, 171(2): 140-147.
- Yao, Z., Liu, L., Gao, F., Rampitsch, C., Reinecke, D.M., Ozga, J.A., and Ayele, B.T. (2012). Developmental and seed aging mediated regulation of antioxidative genes and differential expression of proteins during pre-and post-germinative phases in pea. *Journal of plant physiology*, 169(15): 1477-1488.
- Zaitialia, M., and Atilia H. (2016). Evaluation on the effectiveness of rapid drying method and its effects on chilli (*Capsicum annum*) seed quality.

Trans. Malaysian Soc.Plant Physiol, 23: 68–73.

Zamani, A., Nimrouzi, M., Ruyvaran, M., Nasiri, K., and Akbari, A. (2010). Oil and extract of safflower seed improve fructose induced metabolic syndrome through modulating the homeostasis of trace elements, TNF- α , and fatty acids metabolism. *Journal of Ethnopharmacology*, 112721.

Zeng, F., Zhu, S., Chen, F., Gao, Q., and Yu, S. (2006). Effect of different drying methods on the structure and digestibility of short chain amylose crystals. *Food Hydrocolloids*, 52: 721-731.

