



**COMBINATION TREATMENT OF DOXORUBICIN AND SAR405-LOADED
CHITOSAN NANOPARTICLES FOR DUAL TARGETED ANTICANCER
THERAPY ASSESSMENT IN A549 LUNG CANCER CELL LINE**

By

ALAMASSI MOHAMMEDARFAT N K

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

February 2023

FBSB 2023 3

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**COMBINATION TREATMENT OF DOXORUBICIN AND SAR405-LOADED
CHITOSAN NANOPARTICLES FOR DUAL TARGETED ANTICANCER
THERAPY ASSESSMENT IN A549 LUNG CANCER CELL LINE**

By

ALAMASSI MOHAMMEDARFAT N K

February 2023

Chair : Associate Professor Mas Jaffri Masarudin, PhD
Faculty : Biotechnology and Biomolecular Sciences

Conventional anti-cancer drugs including doxorubicin are associated with high toxicity and non-specific distribution in the body resulting in acute side effects. Targeting cancer cell survival pathways like autophagy can be enhanced the effective of therapeutic drugs at low concentration to decrease its side effects. Cancer cells can use autophagy to promote tolerance to the stress caused by anti-cancer agents, leading to resistance induction in advanced tumors. SAR405 is an inhibitor of autophagy activity due to its molecular interactions within the ATP binding site but is prematurely degraded extracellularly, therefore is poorly taken up by cells limiting its use. Chitosan nanoparticles (CNPs) are considered to be biologically degradable, non-toxic, and biocompatible as a drug delivery system to minimize and determine their side effects. In this study, we developed a therapeutic strategy to enhance doxorubicin efficiency while simultaneously inhibiting autophagy, by increasing cellular uptake of SAR405 loaded into CNPs. The synthesized nanoparticles were characterized for hydrodynamic diameters and polydispersity measurements. Encapsulation efficiency and drug loading was then elucidated and subsequent morphological of the nanoparticles were determined by electron microscopy. Cytotoxicity assessment and apoptosis were performed through MTT and Annexin-v apoptosis assays. Following encapsulation, the SAR405-loaded chitosan nanoparticles expanded from 54 nm to 161 nm at 10 μ M SAR405, while the polydispersity index increased from 0.11 to 0.31. When cells were treated with IC₅₀ values of doxorubicin and 10 μ M of SAR405 encapsulated CNP, approximately 47% reduction in viability of cells were observed in Annexin V-FITC/PI assay, compared to doxorubicin alone. Inhibition of autophagy was shown to decrease the resistance of cancer cells to therapeutic drugs and increase their efficacy at low concentration. This study promises targeting of cancer cell survival pathways can be an effective way to Increasing the efficiency of chemotherapeutic drugs.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**GABUNGAN RAWATAN DOKSORUBISIN DAN NANOPARTIKEL KITOSAN
BERISI SAR405 UNTUK PENILAIAN TERAPI ANTIKANSER DWITARGET
DALAM TITISAN SEL KANSER PARU-PARU A549**

Oleh

ALAMASSI MOHAMMEDARFAT N K

February 2023

Pengerusi : Profesor Madya Mas Jaffri Masarudin, PhD
Fakulti : Bioteknologi dan Sains Biomolekul

Ubat anti-kanser konvensional termasuk doxorubicin dikaitkan dengan ketoksikan tinggi dan pengedaran tidak spesifik dalam badan, mengakibatkan kesan sampingan akut. Dengan menyasar tapak jalan kemandirian sel kanser seperti autofagi, kesannya boleh dipertingkatkan dengan menggunakan ubat terapeutik pada kepekatan rendah untuk mengurangkan kesan sampingannya. Sel kanser boleh menggunakan autofagi untuk menggalakkan toleransi terhadap tekanan yang disebabkan oleh agen anti-kanser, yang menyebabkan induksi rintangan dalam tumor yang kritikal. SAR405 ialah perencat aktiviti autofagi kerana interaksi molekulnya dalam tapak pengikatan ATP, namun ia ternyahasli secara pramatang secara ekstraselular; oleh itu, ia kurang diambil oleh sel, lalu mengehadkan penggunaannya. Nanopartikel kitosan dianggap boleh terurai secara biologi, tidak toksik dan biokompatibel sebagai kargo penghantaran ubat. Dalam kajian ini, kami membangunkan strategi terapeutik untuk meningkatkan kecekapan doksorubisin sambil menghalang autofagi secara serentak, dengan meningkatkan pengambilan selular SAR405 yang dimuatkan ke dalam nanopartikel kitosan. Nanozarah yang disintesis telah dicirikan untuk diameter hidrodinamik dan ukuran polidispersi. Kecekapan enkapsulasi dan pemuatan dadah kemudiannya diperincikan dan morfologi nanozarah seterusnya ditentukan oleh mikroskop elektron. Penilaian sitotoksiti dan apoptosis dilakukan melalui ujian apoptosis MTT dan Annexin-v. Berikutkan pengkapsulan, nanozarah kitosan yang dimuatkan SAR405 berkembang daripada 54 nm kepada 161 nm, manakala indeks polidispersi atau penyebaran meningkat daripada 0.11 kepada 0.31 Apabila sel dirawat dengan nilai IC₅₀ doxorubicin dan 10 μ M SAR405, kira-kira 47% pengurangan daya tahan sel dalam ujian Annexin V-FITC/PI, berbanding dengan doksorubisin sahaja. Perencatan autofagi mengurangkan rintangan sel kanser terhadap ubat terapeutik dan meningkatkan keberkesanannya pada kepekatan rendah. Kajian ini menjanjikan bahawa menyasarkan laluan survival sel kanser sebagai cara yang mujarab untuk meningkatkan kecekapan ubat kemoterapi.

ACKNOWLEDGEMENTS

In the name of Allah, praise be to him, who led me to be supervised by the honorable Assoc. Prof. Dr. Mas Jaffri Masarudin, who helped to complete my master's thesis and supported me on a daily basis with everything. He never hesitated to give opportunities, he taught me well in the lab and gave me many tips to enhance my writing skills. Moreover, he used to calm me down when I felt stressed out.

I would like to thank the other members of my supervisory committee, Assoc. Prof. Suet Lin Chia and Assoc. Prof. Che Azurahanim Che Abdullah for guidance and help throughout the research. I sincerely appreciate having each of them as my co-supervisors.

I would also like to extend my gratitude to Assoc. Prof. Dr. Mas Jaffri Masarudin's students, Ruqayah and Galen, who guide me and advise me all the time.

I also appreciate the friendship and assistance from my best friends in Malaysia, Dr Mohamed, Ahmed and, Moneer who shared with me a lot of unforgettable memories.

I also want to give special thanks to my colleague, Dr. Malak. She wasn't just a colleague in the lab but was a sister who shared with me so many memories.

My gratitude also goes to my beloved friend Dr. Aya (Najma). We spent so many hours together, with long talks and discussions, and she supported me in writing my thesis and encouraged me to submit it on time. My memories with her will be the warmest memories I had in this place.

I sincerely thank my number one supporter, my family, my parents, Numan and Abeer, for helping me financially and always praying for me. I also would like to give a special thank to my sister Aseel, for being there for me in many situations, and my little brother Yousef, for doing special designs for my studies and helping with tech issues. And I wouldn't forget my older sister Maha and her husband Mahmoud for checking out on me and supporting me during my master's journey.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mas Jaffri Masarudin, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Suet Lin Chia, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

COPYRIGHT © UPM

Che Azurahanim Che Abdullah, PhD

Associate Professor

Faculty of Sciences

Universiti Putra Malaysia

(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 08 June 2023

Declaration by Members of the Supervisory Committee

This is to confirm that:

- the research and the writing of this thesis were done under our supervision;
- supervisory responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2015-2016) are adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee: _____

Signature: _____

Name of Member of
Supervisory
Committee: _____

Signature: _____

Name of Member of
Supervisory
Committee: _____

Signature: _____

Name of Member of
Supervisory
Committee: _____

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	ii
ACKNOWLEDGEMENTS	iii
APPROVAL	iv
DECLARATION	vi
LIST OF FIGURES	x
LIST OF ABBREVIATIONS	xii
 CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Objectives	5
1.2.1 General objective	5
1.2.2 Specific objectives	5
2 LITERATURE REVIEW	6
2.1 Lung Cancer	6
2.2 Current modalities and treatments for lung cancer	7
2.3 Doxorubicin as a potent Chemotherapeutic Agent	8
2.4 Autophagy and its role as tumor suppressor and promotion	9
2.4.1 Autophagy as Drug-Resistant Factor of Tumors	10
2.4.2 Inhibition of Autophagy as an Alternative Target	
Mechanism towards Cancer Treatment	10
2.4.3 SAR405 as an Autophagy Inhibitor combining	
chemotherapy drugs for cancer treatment	14
2.5 Nanoparticles as a delivery vector for cancer therapy	15
2.6 Chitosan nanoparticles as a promising delivery vector for	
therapy	17
3 MATERIAL AND METHODS	20
3.1 Material	20
3.2 Nanoparticle synthesis and characterization	20
3.2.1 Chitosan Nanoparticles synthesis, CNP	20
3.2.2 Determination of the fraction of free NH ₂ groups by	
the TNBS assay	21
3.2.3 Analysis of nanoparticle size distribution using	
dynamic light scattering (DLS)	22
3.2.4 Determination of encapsulation efficiency of the drug	
by Nano spectrophotometer (nanodrop)	22
3.2.5 Analysis of nanoparticle morphology by transmission	
electron microscopy (TEM)	22
3.3 Cellular Studies Efficacy	23
3.3.1 Establishment of A549 Cell line	23
3.3.2 Cell cytotoxicity assay (MTT)	23
3.3.3 Apoptosis assay	24
3.3.4 Quantitative Analysis of Autophagosome Formation	
24	
3.4 Statistical Analysis	24

4 RESULTS AND DISCUSSION	25
4.1 Chitosan nanoparticles characterization	25
4.1.1 Determination of free amine group using the trinitrobenzene sulfonic (TNBS) Assay	25
4.1.2 The effect of TPP volume on the particle size and polydispersity index (PDI) values of CNP	27
4.1.3 Influence of SAR405 encapsulation on the size and polydispersity index of CNP to form CNP-SAR405	29
4.1.4 The encapsulation efficiency of SAR405 loaded in CNP	31
4.1.5 Morphological analysis of CNP and CNP-SAR405	
	32
4.2 Cellular studies and analysis	34
4.2.1 Cytotoxicity effects of CNP on A549 lung cancer cell line.	34
4.2.2 Cytotoxicity effects of SAR405 and CNP-SAR405	36
4.2.3 Cytotoxic effects of doxorubicin	39
4.2.4 Cytotoxicity effects of SAR405 and doxorubicin combination treatments	41
4.2.5 Evaluation of apoptosis in A549 cells treated with doxorubicin, SAR405, and CNP-SAR405.	43
4.2.6 Evaluation of autophagy formation in A549 cells treated with doxorubicin, SAR405, and CNP-SAR405.	45
5 CONCLUSION AND FUTURE PERSPECTIVES	47
5.1 Conclusion	47
5.2 Future recommendation	49
REFERENCES	50
APPENDICES	66
BIODATA OF STUDENT	68
PUBLICATION	69

LIST OF FIGURES

Figure		Page
1.1	A Schematic representation of the preparation of chitosan-TPP nanoparticles and SAR405-loaded chitosan	3
1.2	A schematic representation of the binary treatment approach using CNPs as a delivery vector for SAR405. Doxorubicin, a potent cellular apoptotic agent may induce autophagic responses in cancer cells	4
2.1	Number of Lung Cancer New Cases and Deaths Worldwide for Both Sexes (2020)	6
2.2	Signaling pathways that regulate autophagy	14
2.3	Schematic representation of different nano vectors	16
4.1	The effect of TPP volume on the free amine group of CS. There was a decrease in free amine group with increasing TPP volume	26
4.2	Shows the effect of TPP volume on the particle size (d.nm) of CNP	28
4.3	Influence of SAR405 concentration on (A) the particle size distribution and (B) polydispersity index (PDI) of CNP-SAR405 after being encapsulated	30
4.4	Morphological analysis of nanoparticle samples using TEM	33
4.5	Shows the cytotoxic effect of CNP towards the A549 cell line after 72 hrs	35
4.6	Shows the cytotoxic effect of different concentrations of SAR405 (2, 4, 6, 8 and 10 μ M) on A549 lung cancer cells after 72 hrs	37
4.7	Shows the effect of SAR405 concentration on cytotoxicity of CNP-SAR405 at different concentration from 2 to 10 μ M after 72hrs	38
4.8	Shows the cytotoxic effect of different concentration of doxorubicin on A549 lung cancer cells after 72hrs	40

4.9	The cytotoxicity effect of doxorubicin 2 μ M alone and in a combination with SAR405 2 μ M, SAR405 10 μ M, CNP, CNP-SAR405 2 μ M and CNP-SAR405 10 μ M after incubation for 72 hrs	42
4.10	The apoptotic effect of doxorubicin 2 μ M alone and in a combination with SAR405 2 μ M, SAR405 10 μ M, CNP, CNP-SAR405 2 μ M and CNP-SAR405 10 μ M after incubation for 72 hrs	44
4.11	Flowcytometry based profiling of autophagy in A549 cells	46

LIST OF ABBREVIATIONS

A549	Adenocarcinomic human alveolar basal epithelial cell line
ULK	Unc-51-like Autophagy-Activating Kinase
A430nm	Absorbance at 430nm
A570nm	Absorbance at 570nm
A335	Absorbance at 570nm
BIF-1	Bax Interacting Factor-1
CNP	Chitosan nanoparticle
CNP-SAR405	Chitosan nanoparticle encapsulated SAR405
CNT	Carbon Nanotube
Conc.	Concentration
CS	Chitosan
Mwt	Molecular weight
dH ₂ O	Deionized distilled water
DLS	Dynamic Light Scattering
NNMT	Nicotinamide N-methyl transferase
DMSO	Dimethyl Sulfoxide
DNA	Deoxyribonucleic acid
DIW	Deionized Water
EE	Encapsulation Efficiency
EDTA	Ethylenediaminetetraacetic acid
ERK	Extracellular Signal-regulated Kinase
FBS	Feta bovine serum
FDA	Food and Drug Administration
TEM	Transmission Electron Microscopy

FITC	Fluorescein isothiocyanate
GFP	Green Fluorescence Protein
HCl	Hydrochloric acid
HUVEC	Human Umbilical Vein Endothelial Cells
IC ₅₀	Half maximal inhibitory concentration
ERK	Extracellular single-regulated kinase
MTT	3-(4 5-dimethylthiazol-2-yl)-2 5-diphenyltetrazolium bromide
MW	Molecular weight
NaOH	Sodium hydroxide
NH ₂	Amine group
NACO ₃	Sodium Carbonate
TNBS	Trinitobenzenesulfonic acid
NSCLC	Non-small cell lung carcinoma
PBS	Phosphate buffer saline
PCA	Protocatechuic acid
pCNP	Palmitoyl chitosan nanoparticle
TPP	Tripolyphosphate
pCS	Palmitoyl chitosan
PDI	Polydispersity Index
PEG	Polyethylene Glycol
PI	Propidium Iodide
PSD	Particle Size Distribution
RNA	Ribonucleic acid
ROS	Reactive Oxygen Species
rpm	Revolutions Per Minute
RPMI	Roswell Park Memorial Institute

°C Degree Celsius

SDS Sodium Dodecyl Sulphate

CHAPTER 1

INTRODUCTION

1.1 Background

Despite recent advancements in medicine, cancer is still considered one of the most researched illnesses owing to its complexity and steadily rising incidence rate over the years. Globally, 19.3 million new cancer cases were diagnosed, with an estimated 295.3 cases of cancer per 100,000 people in areas with very high human development, compared with 115.7 in areas with low human development in 2020. The most often diagnosed malignancies are lung cancer (11.4%), colorectal cancer (10%), prostate cancer (7.3%), and stomach cancer (5.6%). According to estimates, the global cancer burden will reach 28.4 million cases in 2040, up 47% from 2020, with a greater increase in developing (64%) than developed (32%) countries. Despite being the most widespread cancer in the world, lung cancer is also the greatest cause of cancer-related fatality, with just a 15% five-year survival rate (Sung et al., 2021a). Due to this worrying incidence, an efficient therapy must be developed.

Chemotherapy is the primary therapy for cancer due to its high efficacy in killing cells. Doxorubicin (DOX), a non-specific cell cycle inhibitor, was one of the two first anthracycline antibiotics identified from *Streptomyces paucitoxins* and is one of the most commonly used chemotherapeutic agents to date. DOX stops cell proliferation by intercalating into DNA; inhibiting DNA synthesis and function via inhibition of topoisomerase II and formation of free radicals (Shoshan & Linder, 2008). As the pathway exists in all cell types, DOX-induced cytotoxicity also leads to induction of cell death pathways in not only cancer cells but also in normal cells soliciting unwanted side effects such as severe toxicity in normal organs and rapidly dividing cells. Chemotherapeutic persistence has also been reported in cancers treated with regular dosages of doxorubicin, where neoplastic cells develop drug resistance due to their pro-apoptotic ability to activate a number of mechanisms as a resistance to the chemotherapy treatment (Dandotra, 2021). These have included including drug inactivation, inhibition of cell death (apoptosis suppression), alterations in drug metabolism, enhanced DNA repair, gene amplification, and autophagy stimulation. Therefore, these recent findings have led to the notion that therapeutic regimes against cancer should be streamlined towards addressing a more holistic view towards this dynamic disease. Increasing the effectiveness of anticancer therapies is no longer a subject of producing more potent chemotherapy drugs, but instead designing combinatory therapies containing multiple moieties that target more than one aspect of cancer cell biology.

Autophagy is a well conserved response to nutrient restriction, in which starving cells use it as a recycling mechanism to temporarily compensate for the absence of external nutrients (Limpert et al., 2018). Autophagy also plays a role in the

elimination of long-lived or superfluous proteins and organelles. Autophagy, for example, is responsible for the removal of paternal mitochondria following conception. Another key role of this process is the destruction of defective mitochondria, which limits the release of reactive oxygen species and other harmful proteins into the cytosol. Protein aggregation clearance is also important for protecting cells from hazardous cellular waste. Autophagy is required for maintaining cellular homeostasis in addition to its function as a response to cellular stress. As a result, autophagy is a critical catabolic process in cell formation, differentiation, and survival.

However, autophagy has been shown to be triggered in certain cancer cells as a reaction towards the cellular stress caused by exposure to an anticancer medication. In this example, inhibiting autophagy can potentially render cancer cells more sensitive to treatment of cisplatin and increases the lethal effects of chemotherapeutic drugs to 40% (Schlütermann et al., 2018). Autophagy has been characterized as a pro-survival process seen in most advanced malignancies, allowing tumors to adapt to various stressors such as hypoxia and food restriction, and therefore influencing tumor development (Limpert et al., 2018). As a result, many advanced malignancies have more basal autophagic activity than normal tissues, and some have been labelled "autophagy-dependent," such as pancreatic cancer or activated Ras tumors (Pérez-Hernández et al., 2019). This basal autophagy also aids cancer cell adaptation to treatment-induced stressors, resulting in therapy resistance in malignant malignancies, which is a significant clinical problem. Autophagy has also been linked to the survival of latent cancer cells and the recurrence of metastatic tumors. As a result, inhibiting autophagy may not allow cancer cells to adapt to stress when they treat with chemotherapy leading to increasing the cell death pathway. This can open a window for improving chemotherapeutic efficacy and clinical outcomes in the treatment of malignant cancers.

SAR405 is an autophagy inhibitor compound that was synthesized as a low-molecular-mass with a high specificity for Vps34, a critical protein in the autophagy mechanism. A study was shown the selectivity of SAR405 binding with ATP binding site of Vps34 affects vesicle trafficking between early and late endosomes and prevents autophagy (Pasquier, 2015). However, this compound is administered naked, it has been shown to have poor cellular absorption and stability *in vivo*. Increasing its cellular uptake and accumulation in cancer cells is one potential technique for increasing its efficiency to increase the autophagy inhibition level at which cancer cells use it as a pro-survival pathway. The emphasis of research has been on helping or improving existing medication delivery methods. Over the years, several researchers have reported on the use of nanoparticles to encapsulate cargos such as different medicinal medicines or chemicals, as well as genetic elements.

In this study a nanoparticle-mediated delivery system was adopted to increase the intracellular uptake of SAR405 into A549 lung cancer cell models. Encapsulation within a delivery vector protects the inhibitor from extracellular enzymatic degradation, and repacking it within nano-sized dimensions may

facilitate greater uptake into cells. Chitosan nanoparticles (CNPs) were selected as a delivery system for SAR405, being an inert polymeric nanoparticle system widely applied in drug administration applications (Fu et al., 2016). In contrast to other nanomaterials requiring surface modifications prior to *in vivo* and *in vitro* use, CNPs are a polymer-based organic nanomaterial conferring easier preparation approaches, biologically-compatible, and possess improved *in vivo* distribution (Fu et al., 2016). The SAR405-encapsulated nanoparticle system was formed through complexation of the chitosan polymer (CS) with SAR405, and subsequent cross-linking with sodium tripolyphosphate (TPP) initiating particle formation through ionic gelation processes (Figure 1.1).

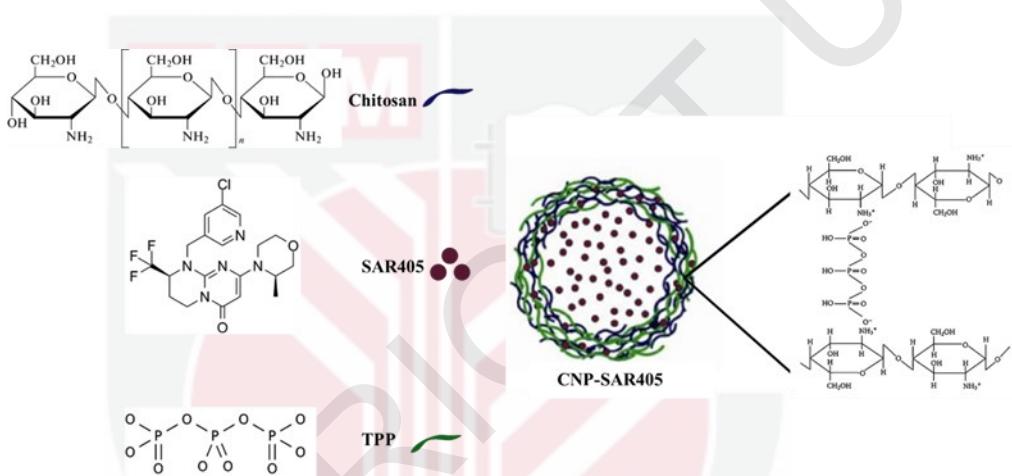


Figure 1.1 : A Schematic representation of the preparation of chitosan-TPP nanoparticles and SAR405-loaded chitosan

The system was then used in a binary treatment regime containing doxorubicin and the SAR405-encapsulated CNPs against A549 lung cancer cells, where the combination incurs apoptotic responses by targeting both cellular-death by apoptotic pathways and sequential inhibition of autophagy. By preventing cancer cells undergoing apoptosis to activate autophagy as a pro-apoptotic response, this system was able to increase the effectiveness of doxorubicin as an anticancer agent (Figure 1.2). Consequent of this, similar cell death rates are achievable at lower doxorubicin concentrations – a proof of concept elucidating the importance of employing a multi-facet approach towards anticancer therapies in the future.

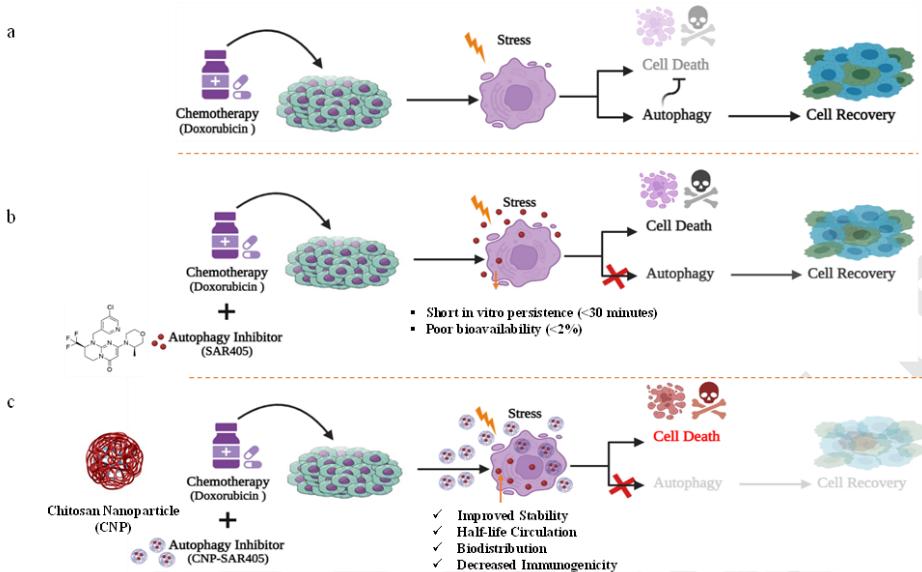


Figure 1.2 : A schematic representation of the binary treatment approach using CNPs as a delivery vector for SAR405. Doxorubicin, a potent cellular apoptotic agent may induce autophagic responses in cancer cells. As a result, this pro-apoptotic event leads to (a) cancer cell recovery and causes doxorubicin resistance in advanced cancers. Alternatively, autophagy inhibitors such as SAR405 may be able to prevent activation of autophagy but instead (b) suffers from limitations including short in vivo persistence as well as poor bioavailability. Therefore, (c) adoption of a nanoparticle delivery system such as CNPs increase the stability, intracellular uptake, and biodistribution of SAR405 in cells. As a consequence, autophagy is arrested in the cancer cells, enhancing the efficacy of doxorubicin by inhibiting autophagy activation leading to increased cell death.

The purpose of this project is to develop a dual treatment system for anticancer therapy which targets both cell death and autophagy pathways of A549 lung cancer cells for enhancing the efficacy of the chemotherapy drugs by inhibiting of autophagy pathway. This involves use of doxorubicin + SAR405, in this project SAR405 is delivered via CNP.

1.2 Objectives

1.2.1 General objective

To develop a dual-treatment system for anticancer therapy which targets both the apoptotic and autophagic pathways of A549 lung cancer cell using Doxorubicin + CNP-SAR405.

1.2.2 Specific objectives

1. To characterize the formation of SAR405-encapsulated chitosan nanoparticles, CNP-SAR405.
2. To determine the efficacy of Doxorubicin in co- treatments with CNP-SAR405 toward A549 lung cancer cells.
3. To assess autophagy progression in A549 cells treated with Doxorubicin in co-treatments with CNP-SAR405.

REFERENCES

- Ai, J., Liao, W., & Ren, Z.-L. (2017). Enhanced anticancer effect of copper-loaded chitosan nanoparticles against osteosarcoma. *RSC Advances*, 7(26), 15971–15977. <https://doi.org/10.1039/C6RA21648J>
- Ali, S. W., Rajendran, S., & Joshi, M. (2011). Synthesis and characterization of chitosan and silver loaded chitosan nanoparticles for bioactive polyester. *Carbohydrate Polymers*, 83(2), 438–446. <https://doi.org/10.1016/j.carbpol.2010.08.004>
- Allmann, S., Mayer, L., Olma, J., Kaina, B., Hofmann, T. G., Tomicic, M. T., & Christmann, M. (2021). Benzo[a]pyrene represses DNA repair through altered E2F1/E2F4 function marking an early event in DNA damage-induced cellular senescence. *Nucleic Acids Research*, 48(21), 12085–12101. <https://doi.org/10.1093/nar/gkaa965>
- Amaravadi, R., Kimmelman, A. C., & White, E. (2016). Recent insights into the function of autophagy in cancer. *Genes & Development*, 30(17), 1913–1930. <https://doi.org/10.1101/gad.287524.116>
- Aredo, J. V., Luo, S. J., Gardner, R. M., Sanyal, N., Choi, E., Hickey, T. P., Riley, T. L., Huang, W.-Y., Kurian, A. W., Leung, A. N., Wilkens, L. R., Robbins, H. A., Riboli, E., Kaaks, R., Tjønneland, A., Vermeulen, R. C. H., Panico, S., Le Marchand, L., Amos, C. I., ... Han, S. S. (2021). Tobacco Smoking and Risk of Second Primary Lung Cancer. *Journal of Thoracic Oncology*, 16(6), 968–979. <https://doi.org/10.1016/j.jtho.2021.02.024>
- Ariff, S. A. Y., Yusoff, K., & Masarudin, M. J. (2017). Encapsulation of miRNA in chitosan nanoparticles as a candidate for an anti-metastatic agent in cancer therapy. *Malaysian Applied Biology*, 46(1), 165–170.
- Artursson, P., Lindmark, T., Davis, S. S., & Illum, L. (1994). Effect of chitosan on the permeability of monolayers of intestinal epithelial cells (Caco-2). *Pharmaceutical Research*, 11(9), 1358–1361. <https://doi.org/10.1023/a:1018967116988>
- Ashley, N., & Poulton, J. (2009). Mitochondrial DNA is a direct target of anti-cancer anthracycline drugs. *Biochemical and Biophysical Research Communications*, 378(3), 450–455. <https://doi.org/10.1016/j.bbrc.2008.11.059>
- Baka, S., Califano, R., Ferraldeschi, R., Aschroft, L., Thatcher, N., Taylor, P., Faivre-Finn, C., Blackhall, F., & Lorigan, P. (2008). Phase III randomised trial of doxorubicin-based chemotherapy compared with platinum-based chemotherapy in small-cell lung cancer. *British Journal of Cancer*, 99(3), Article 3. <https://doi.org/10.1038/sj.bjc.6604480>

- Bao, L., Jaramillo, M. C., Zhang, Z., Zheng, Y., Yao, M., Zhang, D. D., & Yi, X. (2015). Induction of autophagy contributes to cisplatin resistance in human ovarian cancer cells. *Molecular Medicine Reports*, 11(1), 91–98. <https://doi.org/10.3892/mmr.2014.2671>
- Barkoula, N. M., Alcock, B., Cabrera, N. O., & Peijs, T. (2008). Flame-Retardancy Properties of Intumescent Ammonium Poly(Phosphate) and Mineral Filler Magnesium Hydroxide in Combination with Graphene. *Polymers and Polymer Composites*, 16(2), 101–113. <https://doi.org/10.1002/pc>
- Behzadi, S., Serpooshan, V., Tao, W., Hamaly, M. A., Alkawareek, M. Y., Dreaden, E. C., Brown, D., Alkilany, A. M., Farokhzad, O. C., & Mahmoudi, M. (2017). Cellular Uptake of Nanoparticles: Journey Inside the Cell. *Chemical Society Reviews*, 46(14), 4218. <https://doi.org/10.1039/c6cs00636a>
- Berger, J., Reist, M., Mayer, J. M., Felt, O., Peppas, N. A., & Gurny, R. (2004). Structure and interactions in covalently and ionically crosslinked chitosan hydrogels for biomedical applications. *European Journal of Pharmaceutics and Biopharmaceutics*, 57(1), 19–34. [https://doi.org/10.1016/S0939-6411\(03\)00161-9](https://doi.org/10.1016/S0939-6411(03)00161-9)
- Boffetta, P., Autier, P., Boniol, M., Boyle, P., Hill, C., Aurengo, A., Masse, R., Thé, G. de, Valleron, A.-J., Monier, R., & Tubiana, M. (2010). An estimate of cancers attributable to occupational exposures in France. *Journal of Occupational and Environmental Medicine*, 52(4), 399–406. <https://doi.org/10.1097/JOM.0b013e3181d5e355>
- Brigger, I., Dubernet, C., & Couvreur, P. (2002). Nanoparticles in cancer therapy and diagnosis. *Advanced Drug Delivery Reviews*, 54(5), 631–651. [https://doi.org/10.1016/S0169-409X\(02\)00044-3](https://doi.org/10.1016/S0169-409X(02)00044-3)
- Bruno, B. J., Miller, G. D., & Lim, C. S. (2013). Basics and recent advances in peptide and protein drug delivery. *Therapeutic Delivery*, 4(11), 1443–1467. <https://doi.org/10.4155/tde.13.104>
- Cai, M., Hu, Z., Liu, J., Gao, J., Liu, C., Liu, D., Tan, M., Zhang, D., & Lin, B. (2014). Beclin 1 expression in ovarian tissues and its effects on ovarian cancer prognosis. *International Journal of Molecular Sciences*, 15(4), 5292–5303. <https://doi.org/10.3390/ijms15045292>
- Carvalho, C., Santos, R. X., Cardoso, S., Correia, S., Oliveira, P. J., Santos, M. S., & Moreira, P. I. (2009). Doxorubicin: The good, the bad and the ugly effect. *Current Medicinal Chemistry*, 16(25), 3267–3285. <https://doi.org/10.2174/092986709788803312>
- Chen, C., Lu, L., Yan, S., Yi, H., Yao, H., Wu, D., He, G., Tao, X., & Deng, X. (2018). Autophagy and doxorubicin resistance in cancer. *Anti-Cancer Drugs*, 29(1), 1–9. <https://doi.org/10.1097/CAD.0000000000000572>

- Chen, Z., Jiang, Q., Zhu, P., Chen, Y., Xie, X., Du, Z., Jiang, L., & Tang, W. (2019). NPRL2 enhances autophagy and the resistance to Everolimus in castration-resistant prostate cancer. *The Prostate*, 79(1), 44–53. <https://doi.org/10.1002/pros.23709>
- Choudhary, S., Gupta, L., Rani, S., Dave, K., & Gupta, U. (2017). Impact of Dendrimers on Solubility of Hydrophobic Drug Molecules. *Frontiers in Pharmacology*, 8, 261. <https://doi.org/10.3389/fphar.2017.00261>
- Clark, K., MacKenzie, K. F., Petkevicius, K., Kristariyanto, Y., Zhang, J., Choi, H. G., Peggie, M., Plater, L., Pedrioli, P. G. A., McIver, E., Gray, N. S., Arthur, J. S. C., & Cohen, P. (2012). Phosphorylation of CRTC3 by the salt-inducible kinases controls the interconversion of classically activated and regulatory macrophages. *Proceedings of the National Academy of Sciences of the United States of America*, 109(42), 16986–16991. <https://doi.org/10.1073/pnas.1215450109>
- Crew, E., Tessel, M. A., Rahman, S., Razzak-Jaffar, A., Mott, D., Kamundi, M., Yu, G., Tchah, N., Lee, J., Bellavia, M., & Zhong, C.-J. (2012). MicroRNA conjugated gold nanoparticles and cell transfection. *Analytical Chemistry*, 84(1), 26–29. <https://doi.org/10.1021/ac202749p>
- Cunha, R. A., Soares, T. A., Rusu, V. H., Pontes, F. J. S., Franca, E. F., Lins, R. D., Cunha, R. A., Soares, T. A., Rusu, V. H., Pontes, F. J. S., Franca, E. F., & Lins, R. D. (2012). The Molecular Structure and Conformational Dynamics of Chitosan Polymers: An Integrated Perspective from Experiments and Computational Simulations. In *The Complex World of Polysaccharides*. IntechOpen. <https://doi.org/10.5772/51803>
- Dandot, S. (2021). Mechanisms adopted by cancer cells to escape apoptosis—A review. *BIOCELL*, 45(4), Article 4. <https://doi.org/10.32604/biocell.2021.013993>
- Desai, M. P., Labhasetwar, V., Walter, E., Levy, R. J., & Amidon, G. L. (1997). The mechanism of uptake of biodegradable microparticles in Caco-2 cells is size dependent. *Pharmaceutical Research*, 14(11), 1568–1573. <https://doi.org/10.1023/a:1012126301290>
- Dingemans, A.-M. C., Früh, M., Ardizzone, A., Besse, B., Faivre-Finn, C., Hendriks, L. E., Lantuejoul, S., Peters, S., Reguart, N., Rudin, C. M., Ruysscher, D. D., Schil, P. E. V., Vansteenkiste, J., & Reck, M. (2021). Small-cell lung cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up☆. *Annals of Oncology*, 32(7), 839–853. <https://doi.org/10.1016/j.annonc.2021.03.207>
- Dombb, A., Miillerd, R. H., Langerf, R., Gref, R., Domb, A., Quellec, P., Blunk, T., Müller, R. H., Verbavatz, J. M., & Langer, R. (1995). The controlled intravenous delivery of drugs using PEG-coated sterically stabilized nanospheres. *Advanced Drug Delivery Reviews*, 16(2–3), 215–233.

- Douglas, K. L., & Tabrizian, M. (2005). Effect of experimental parameters on the formation of alginate-chitosan nanoparticles and evaluation of their potential application as DNA carrier. *Journal of Biomaterials Science, Polymer Edition*, 16(1), 43–56. <https://doi.org/10.1163/1568562052843339>
- Dowdle, W. E., Nyfeler, B., Nagel, J., Elling, R. A., Liu, S., Triantafellow, E., Menon, S., Wang, Z., Honda, A., Pardee, G., Cantwell, J., Luu, C., Cornell-Taracido, I., Harrington, E., Fekkes, P., Lei, H., Fang, Q., Digan, M. E., Burdick, D., ... Murphy, L. O. (2014). Selective VPS34 inhibitor blocks autophagy and uncovers a role for NCOA4 in ferritin degradation and iron homeostasis in vivo. *Nature Cell Biology*, 16(11), 1069–1079. <https://doi.org/10.1038/ncb3053>
- Dupont, M., Huart, M., Lauvinerie, C., Bidet, A., Guitart, A. V., Villacreces, A., Vigon, I., Desplat, V., El Habhab, A., Pigneux, A., Ivanovic, Z., Brunet De la Grange, P., Dumas, P.-Y., & Pasquet, J.-M. (2022). Autophagy Targeting and Hematological Mobilization in FLT3-ITD Acute Myeloid Leukemia Decrease Repopulating Capacity and Relapse by Inducing Apoptosis of Committed Leukemic Cells. *Cancers*, 14(2), 453. <https://doi.org/10.3390/cancers14020453>
- Dyczynski, M., Yu, Y., Otracka, M., Parpal, S., Braga, T., Henley, A. B., Zazzi, H., Lerner, M., Wennerberg, K., Viklund, J., Martinsson, J., Grandér, D., De Milito, A., & Pokrovskaja Tamm, K. (2018). Targeting autophagy by small molecule inhibitors of vacuolar protein sorting 34 (Vps34) improves the sensitivity of breast cancer cells to Sunitinib. *Cancer Letters*, 435, 32–43. <https://doi.org/10.1016/j.canlet.2018.07.028>
- Egan, D. F., Chun, M. G. H., Vamos, M., Zou, H., Rong, J., Miller, C. J., Lou, H. J., Raveendra-Panicker, D., Yang, C.-C., Sheffler, D. J., Teriete, P., Asara, J. M., Turk, B. E., Cosford, N. D. P., & Shaw, R. J. (2015). Small Molecule Inhibition of the Autophagy Kinase ULK1 and Identification of ULK1 Substrates. *Molecular Cell*, 59(2), 285–297. <https://doi.org/10.1016/j.molcel.2015.05.031>
- Fan, W., Yan, W., Xu, Z., & Ni, H. (2012). Formation mechanism of monodisperse, low molecular weight chitosan nanoparticles by ionic gelation technique. *Colloids and Surfaces B: Biointerfaces*, 90(1), 21–27. <https://doi.org/10.1016/j.colsurfb.2011.09.042>
- Farago, A. F., Drapkin, B. J., Lopez-Vilarino de Ramos, J. A., Galmarini, C. M., Núñez, R., Kahatt, C., & Paz-Ares, L. (2019). ATLANTIS: A Phase III study of irubinectedin/doxorubicin versus topotecan or cyclophosphamide/doxorubicin/vincristine in patients with small-cell lung cancer who have failed one prior platinum-containing line. *Future Oncology*, 15(3), 231–239. <https://doi.org/10.2217/fon-2018-0597>
- Fatima, M., Munir, N., Mahmood, Z., & Jahangir, M. (n.d.). Association of Lung Cancer with Tobacco smoking; a review. 2020.

- Fu, S., Xia, J., & Wu, J. (2016). Functional Chitosan Nanoparticles in Cancer Treatment. *Journal of Biomedical Nanotechnology*, 12(8), 1585–1603. <https://doi.org/10.1166/jbn.2016.2228>
- Fung, C., Lock, R., Gao, S., Salas, E., & Debnath, J. (2008). Induction of Autophagy during Extracellular Matrix Detachment Promotes Cell Survival. *Molecular Biology of the Cell*, 19(3), 797–806. <https://doi.org/10.1091/mbc.e07-10-1092>
- Gan, Q., & Wang, T. (2007). Chitosan nanoparticle as protein delivery carrier- Systematic examination of fabrication conditions for efficient loading and release. *Colloids and Surfaces B: Biointerfaces*, 59(1), 24–34. <https://doi.org/10.1016/j.colsurfb.2007.04.009>
- Garg, U., Chauhan, S., Nagaich, U., & Jain, N. (2019). Current Advances in Chitosan Nanoparticles Based Drug Delivery and Targeting. *Advanced Pharmaceutical Bulletin*, 9(2), 195–204. <https://doi.org/10.15171/apb.2019.023>
- Gogishvili, M., Melkadze, T., Makharadze, T., Giorgadze, D., Dvorkin, M., Penkov, K., Laktionov, K., Nemsadze, G., Nechaeva, M., Rozhkova, I., Kalinka, E., Gessner, C., Moreno-Jaime, B., Passalacqua, R., Li, S., McGuire, K., Kaul, M., Paccaly, A., Quek, R. G. W., ... Rietschel, P. (2022). Cemiplimab plus chemotherapy versus chemotherapy alone in non-small cell lung cancer: A randomized, controlled, double-blind phase 3 trial. *Nature Medicine*, 28(11), Article 11. <https://doi.org/10.1038/s41591-022-01977-y>
- Guo, H., Li, H., Zhu, L., Feng, J., Huang, X., & Baak, J. P. A. (2021). "How Long Have I Got?" in Stage IV NSCLC Patients With at Least 3 Months Up to 10 Years Survival, Accuracy of Long-, Intermediate-, and Short-Term Survival Prediction Is Not Good Enough to Answer This Question. *Frontiers in Oncology*, 11. <https://www.frontiersin.org/articles/10.3389/fonc.2021.761042>
- Guo, X., Zhang, Y., Zheng, L., Zheng, C., Song, J., Zhang, Q., Kang, B., Liu, Z., Jin, L., Xing, R., Gao, R., Zhang, L., Dong, M., Hu, X., Ren, X., Kirchhoff, D., Roider, H. G., Yan, T., & Zhang, Z. (2018). Global characterization of T cells in non-small-cell lung cancer by single-cell sequencing. *Nature Medicine*, 24(7), 978–985. <https://doi.org/10.1038/s41591-018-0045-3>
- Guzman-Villanueva, D., El-Sherbiny, I. M., Vlassov, A. V., Herrera-Ruiz, D., & Smyth, H. D. C. (2014). Enhanced cellular uptake and gene silencing activity of siRNA molecules mediated by chitosan-derivative nanocomplexes. *International Journal of Pharmaceutics*, 473(1–2), 579–590. <https://doi.org/10.1016/j.ijpharm.2014.07.026>
- Hanna, A. D., Lam, A., Tham, S., Dulhunty, A. F., & Beard, N. A. (2014). Adverse Effects of Doxorubicin and Its Metabolic Product on Cardiac RyR2 and

- SERCA2A. Molecular Pharmacology, 86(4), 438–449.
<https://doi.org/10.1124/mol.114.093849>
- Hare, J. I., Lammers, T., Ashford, M. B., Puri, S., Storm, G., & Barry, S. T. (2017). Challenges and strategies in anti-cancer nanomedicine development: An industry perspective. Advanced Drug Delivery Reviews, 108, 25–38.
<https://doi.org/10.1016/j.addr.2016.04.025>
- Harhaji-Trajkovic, L., Vilimanovich, U., Kravic-Stevovic, T., Bumbasirevic, V., & Trajkovic, V. (2009). AMPK-mediated autophagy inhibits apoptosis in cisplatin-treated tumour cells. Journal of Cellular and Molecular Medicine, 13(9 B), 3644–3654. <https://doi.org/10.1111/j.1582-4934.2009.00663.x>
- Hassan, U. A., Hussein, M. Z., Alitheen, N. B., Ariff, S. A. Y., & Masarudin, M. J. (2018). In vitro cellular localization and efficient accumulation of fluorescently tagged biomaterials from monodispersed chitosan nanoparticles for elucidation of controlled release pathways for drug delivery systems. International Journal of Nanomedicine, 13, 5075–5095.
<https://doi.org/10.2147/IJN.S164843>
- He, Q., He, X., Deng, B., Shi, C., Lin, L., Liu, P., Yang, Z., Yang, S., & Xu, Z. (2018). Sorafenib and indocyanine green co-loaded in photothermally sensitive liposomes for diagnosis and treatment of advanced hepatocellular carcinoma. Journal of Materials Chemistry B, 6(36), 5823–5834. <https://doi.org/10.1039/C8TB01641K>
- Hinde, E., Thammasiraphop, K., Duong, H. T. T., Yeow, J., Karagoz, B., Boyer, C., Gooding, J. J., & Gaus, K. (2017). Pair correlation microscopy reveals the role of nanoparticle shape in intracellular transport and site of drug release. Nature Nanotechnology, 12(1), Article 1.
<https://doi.org/10.1038/nnano.2016.160>
- Huang, F., Wang, B.-R., & Wang, Y.-G. (2018). Role of autophagy in tumorigenesis, metastasis, targeted therapy and drug resistance of hepatocellular carcinoma. World Journal of Gastroenterology, 24(41), 4643–4651. <https://doi.org/10.3748/wjg.v24.i41.4643>
- Huang, J., Deng, Y., Tin, M. S., Lok, V., Ngai, C. H., Zhang, L., Lucero-Prisno, D. E., Xu, W., Zheng, Z.-J., Elcarte, E., Withers, M., & Wong, M. C. S. (2022). Distribution, Risk Factors, and Temporal Trends for Lung Cancer Incidence and Mortality: A Global Analysis. Chest, 161(4), 1101–1111.
<https://doi.org/10.1016/j.chest.2021.12.655>
- Jaggi, P. (2017). A Review Article on Lung Cancer Diagnosis & Treatment. Research and Reviews: Journal of Medical and Health Sciences.
<https://www.semanticscholar.org/paper/A-Review-Article-on-Lung-Cancer-Diagnosis-%26-Jaggi/b04a317314bc47d3e1f68e4f6600494d3aed03bf>

- Jain, R. K., & Stylianopoulos, T. (2010). Delivering nanomedicine to solid tumors. *Nature Reviews Clinical Oncology*, 7(11), 653–664. <https://doi.org/10.1038/nrclinonc.2010.139>
- Jarudilokkul, S., Tongthammachat, A., & Boonamnuayvittaya, V. (2011). Preparation of chitosan nanoparticles for encapsulation and release of protein. *Korean Journal of Chemical Engineering*, 28(5), 1247–1251. <https://doi.org/10.1007/s11814-010-0485-z>
- Kang, R., Zeh, H. J., Lotze, M. T., & Tang, D. (2011). The Beclin 1 network regulates autophagy and apoptosis. *Cell Death & Differentiation*, 18(4), Article 4. <https://doi.org/10.1038/cdd.2010.191>
- Kciuk, M., Gielecińska, A., Mujwar, S., Kołat, D., Kałuzińska-Kołat, Ż., Celik, I., & Kontek, R. (2023). Doxorubicin-An Agent with Multiple Mechanisms of Anticancer Activity. *Cells*, 12(4), 659. <https://doi.org/10.3390/cells12040659>
- Kerpel-Fronius, A., Tammämägi, M., Cavic, M., Henschke, C., Jiang, L., Kazerooni, E., Lee, C.-T., Ventura, L., Yang, D., Lam, S., Huber, R. M., Yang, D., Zulueta, J., Viola, L., Mohan, A., Lee, C.-T., Covic, M., Schmidt, H., Kazerooni, E., ... Huber, R. (2022). Screening for Lung Cancer in Individuals Who Never Smoked: An International Association for the Study of Lung Cancer Early Detection and Screening Committee Report. *Journal of Thoracic Oncology*, 17(1), 56–66. <https://doi.org/10.1016/j.jtho.2021.07.031>
- Khani, Y., Pourgholam-Amiji, N., Afshar, M., Otroshi, O., Sharifi-Esfahani, M., Sadeghi-Gandomani, H., Vejdani, M., & Salehiniya, H. (2018). Tobacco Smoking and Cancer Types: A Review. *Biomedical Research and Therapy*, 5(4), Article 4. <https://doi.org/10.15419/bmrat.v5i4.428>
- Kim, A.-S., Ko, H.-J., Kwon, J.-H., & Lee, J.-M. (2018). Exposure to Secondhand Smoke and Risk of Cancer in Never Smokers: A Meta-Analysis of Epidemiologic Studies. *International Journal of Environmental Research and Public Health*, 15(9), Article 9. <https://doi.org/10.3390/ijerph15091981>
- Kim, J.-H., Kim, Y.-S., Kim, S., Park, J. H., Kim, K., Choi, K., Chung, H., Jeong, S. Y., Park, R.-W., Kim, I.-S., & Kwon, I. C. (2006). Hydrophobically modified glycol chitosan nanoparticles as carriers for paclitaxel. *Journal of Controlled Release: Official Journal of the Controlled Release Society*, 111(1–2), 228–234. <https://doi.org/10.1016/j.jconrel.2005.12.013>
- Kim, M. S., Jeong, E. G., Ahn, C. H., Kim, S. S., Lee, S. H., & Yoo, N. J. (2008). Frameshift mutation of UVRAg, an autophagy-related gene, in gastric carcinomas with microsatellite instability. *Human Pathology*, 39(7), 1059–1063. <https://doi.org/10.1016/j.humpath.2007.11.013>
- Klionsky, D. J. (2008). Autophagy revisited: A conversation with Christian de Duve. *Autophagy*, 4(6), 740–743. <https://doi.org/10.4161/auto.6398>

- Koukaras, E. N., Papadimitriou, S. A., Bikaris, D. N., & Froudakis, G. E. (2012). Insight on the formation of chitosan nanoparticles through ionotropic gelation with tripolyphosphate. *Molecular Pharmaceutics*, 9(10), 2856–2862. <https://doi.org/10.1021/mp300162j>
- Kuen. (2020). Polymers Increased Cytotoxic Efficiency of Protocatechuic Acid in. *Polymers*.
- Kumar, M. N. V. R., Muzzarelli, R. a. A., Muzzarelli, C., Sashiwa, H., & Domb, A. J. (2004). Chitosan chemistry and pharmaceutical perspectives. *Chemical Reviews*, 104(12), 6017–6084. <https://doi.org/10.1021/cr030441b>
- Li, J., Hou, N., Faried, A., Tsutsumi, S., Takeuchi, T., & Kuwano, H. (2009). Inhibition of autophagy by 3-MA enhances the effect of 5-FU-induced apoptosis in colon cancer cells. *Annals of Surgical Oncology*, 16(3), 761–771. <https://doi.org/10.1245/s10434-008-0260-0>
- Liang, Y., Huang, W., Zeng, D., Huang, X., Chan, L., Mei, C., Feng, P., Tan, C.-H., & Chen, T. (2018). Cancer-targeted design of bioresponsive prodrug with enhanced cellular uptake to achieve precise cancer therapy. *Drug Delivery*, 25(1), 1350–1361. <https://doi.org/10.1080/10717544.2018.1477862>
- Limpert, A. S., Lambert, L. J., Bakas, N. A., Bata, N., Brun, S. N., Shaw, R. J., & Cosford, N. D. P. (2018). Autophagy in Cancer: Regulation by Small Molecules. *Trends in Pharmacological Sciences*, 39(12), 1021–1032. <https://doi.org/10.1016/j.tips.2018.10.004>
- Liu, D., Yang, Y., Liu, Q., & Wang, J. (2011). Inhibition of autophagy by 3-MA potentiates cisplatin-induced apoptosis in esophageal squamous cell carcinoma cells. *Medical Oncology* (Northwood, London, England), 28(1), 105–111. <https://doi.org/10.1007/s12032-009-9397-3>
- Liu, F., Liu, D., Yang, Y., & Zhao, S. (2013). Effect of autophagy inhibition on chemotherapy-induced apoptosis in A549 lung cancer cells. *Oncology Letters*, 5(4), 1261–1265. <https://doi.org/10.3892/ol.2013.1154>
- Liu, H., Lv, L., & Yang, K. (2015). Chemotherapy targeting cancer stem cells. *American Journal of Cancer Research*, 5(3), 880–893.
- Mai, Y., Dou, L., Yao, Z., Madla, C. M., Gavins, F. K. H., TaherAli, F., Yin, H., Orlu, M., Murdan, S., & Basit, A. W. (2021). Quantification of P-Glycoprotein in the Gastrointestinal Tract of Humans and Rodents: Methodology, Gut Region, Sex, and Species Matter. *Molecular Pharmaceutics*, 18(5), 1895–1904. <https://doi.org/10.1021/acs.molpharmaceut.0c00574>
- Marsh, T., & Debnath, J. (2015). Ironing out VPS34 inhibition. *Nature Cell Biology*, 17(1), 1–3. <https://doi.org/10.1038/ncb3089>

- Martin, K. R., Celano, S. L., Solitro, A. R., Gunaydin, H., Scott, M., O'Hagan, R. C., Shumway, S. D., Fuller, P., & MacKeigan, J. P. (2018). A Potent and Selective ULK1 Inhibitor Suppresses Autophagy and Sensitizes Cancer Cells to Nutrient Stress. *IScience*, 8, 74–84. <https://doi.org/10.1016/j.isci.2018.09.012>
- Maruyama, C. R., Guilger, M., Pascoli, M., Bileshy-José, N., Abhilash, P. C., Fraceto, L. F., & De Lima, R. (2016). Nanoparticles Based on Chitosan as Carriers for the Combined Herbicides Imazapic and Imazapyr. *Scientific Reports*, 6(November 2015). <https://doi.org/10.1038/srep19768>
- Masarudin, M. J., Cutts, S. M., Evison, B. J., Phillips, D. R., & Pigram, P. J. (2015). Factors determining the stability, size distribution, and cellular accumulation of small, monodisperse chitosan nanoparticles as candidate vectors for anticancer drug delivery: Application to the passive encapsulation of [14C]-doxorubicin. *Nanotechnology, Science and Applications*, 8, 67–80. <https://doi.org/10.2147/NSA.S91785>
- Mitchison, T. J. (2012). The proliferation rate paradox in antimitotic chemotherapy. *Molecular Biology of the Cell*, 23(1), 1–6. <https://doi.org/10.1091/mbc.e10-04-0335>
- Mohammed, M. A., Syeda, J. T. M., Wasan, K. M., & Wasan, E. K. (2017). An Overview of Chitosan Nanoparticles and Its Application in Non-Parenteral Drug Delivery. *Pharmaceutics*, 9(4), 53. <https://doi.org/10.3390/pharmaceutics9040053>
- Nallamuthu, I., Devi, A., & Khanum, F. (2015). Chlorogenic acid loaded chitosan nanoparticles with sustained release property, retained antioxidant activity and enhanced bioavailability. *Asian Journal of Pharmaceutical Sciences*, 10(3), 203–211. <https://doi.org/10.1016/j.ajps.2014.09.005>
- Naruphontjirakul, P., & Viravaidya-Pasuwat, K. (2011). Development of Doxorubicin – Core Shell O-succinyl Chitosan Graft Pluronic®127 Copolymer Nanoparticles to Treat Human Cancer. *International Journal of Bioscience, Biochemistry and Bioinformatics*, 131–136. <https://doi.org/10.7763/IJBBB.2011.V1.24>
- Navya, P. N., Kaphle, A., Srinivas, S. P., Bhargava, S. K., Rotello, V. M., & Daima, H. K. (2019). Current trends and challenges in cancer management and therapy using designer nanomaterials. *Nano Convergence*, 6(1), 23. <https://doi.org/10.1186/s40580-019-0193-2>
- Noman, M. Z., Parpal, S., Van Moer, K., Xiao, M., Yu, Y., Viklund, J., De Milito, A., Hasmim, M., Andersson, M., Amaravadi, R. K., Martinsson, J., Berchem, G., & Janji, B. (2020). Inhibition of Vps34 reprograms cold into hot inflamed tumors and improves anti-PD-1/PD-L1 immunotherapy. *Science Advances*, 6(18), eaax7881. <https://doi.org/10.1126/sciadv.aax7881>

- Ong, S. G. M., Ming, L. C., Lee, K. S., & Yuen, K. H. (2016). Influence of the encapsulation efficiency and size of liposome on the oral bioavailability of griseofulvin-loaded liposomes. *Pharmaceutics*, 8(3). <https://doi.org/10.3390/pharmaceutics8030025>
- Othman, N., Siti Nurul, S. N. A., Masarudin, M. J., Abdullah, L. C., Daik, R., & Sarman, N. S. (2020). L-Ascorbic acid and thymoquinone dual-loaded palmitoyl-chitosan nanoparticles: Improved preparation method, encapsulation and release efficiency. *Processes*, 8(9). <https://doi.org/10.3390/pr8091040>
- Panyam, J., & Labhasetwar, V. (2003). Biodegradable nanoparticles for drug and gene delivery to cells and tissue. *Advanced Drug Delivery Reviews*, 55(3), 329–347. [https://doi.org/10.1016/S0169-409X\(02\)00228-4](https://doi.org/10.1016/S0169-409X(02)00228-4)
- Pasquier, B. (2015). SAR405, a PIK3C3/VPS34 inhibitor that prevents autophagy and synergizes with MTOR inhibition in tumor cells. *Autophagy*, 11(4), 725–726. <https://doi.org/10.1080/15548627.2015.1033601>
- Patel, B., & Priefer, R. (2022). Impact of chronic obstructive pulmonary disease, lung infection, and/or inhaled corticosteroids use on potential risk of lung cancer. *Life Sciences*, 294, 120374. <https://doi.org/10.1016/j.lfs.2022.120374>
- Peng, Y.-F., Shi, Y.-H., Ding, Z.-B., Ke, A.-W., Gu, C.-Y., Hui, B., Zhou, J., Qiu, S.-J., Dai, Z., & Fan, J. (2013). Autophagy inhibition suppresses pulmonary metastasis of HCC in mice via impairing anoikis resistance and colonization of HCC cells. *Autophagy*, 9(12), 2056–2068. <https://doi.org/10.4161/auto.26398>
- Pérez-Hernández, M., Arias, A., Martínez-García, D., Pérez-Tomás, R., Quesada, R., & Soto-Cerrato, V. (2019). Targeting Autophagy for Cancer Treatment and Tumor Chemosensitization. *Cancers*, 11(10), 1599. <https://doi.org/10.3390/cancers11101599>
- Peter, S., Alven, S., Maseko, R. B., & Aderibigbe, B. A. (2022). Doxorubicin-Based Hybrid Compounds as Potential Anticancer Agents: A Review. *Molecules*, 27(14), Article 14. <https://doi.org/10.3390/molecules27144478>
- Phillips, R. M. (2016). Targeting the hypoxic fraction of tumours using hypoxia-activated prodrugs. *Cancer Chemotherapy and Pharmacology*, 77(3), 441–457. <https://doi.org/10.1007/s00280-015-2920-7>
- Quagliariello, V., Masarone, M., Armenia, E., Giudice, A., Barbarisi, M., Caraglia, M., Barbarisi, A., & Persico, M. (2019). Chitosan-coated liposomes loaded with butyric acid demonstrate anticancer and anti-inflammatory activity in human hepatoma HepG2 cells. *Oncology Reports*, 41(3), 1476–1486. <https://doi.org/10.3892/or.2018.6932>

- Rajadurai, P., How, S. H., Liam, C. K., Sachithanandan, A., Soon, S. Y., & Tho, L. M. (2020). Lung Cancer in Malaysia. *Journal of Thoracic Oncology*, 15(3), 317–323. <https://doi.org/10.1016/j.jtho.2019.10.021>
- Rajan, M., Raj, V., Al-Arfaj, A. A., & Murugan, A. M. (2013). Hyaluronidase enzyme core-5-fluorouracil-loaded chitosan-PEG-gelatin polymer nanocomposites as targeted and controlled drug delivery vehicles. *International Journal of Pharmaceutics*, 453(2), 514–522. <https://doi.org/10.1016/j.ijpharm.2013.06.030>
- Rajan, R. K., Hussein, M. Z., Fakurazi, S., Yusoff, K., & Masarudin, M. J. (2019). Increased ROS scavenging and antioxidant efficiency of chlorogenic acid compound delivered via a chitosan nanoparticulate system for efficient in vitro visualization and accumulation in human renal adenocarcinoma cells. *International Journal of Molecular Sciences*, 20(19). <https://doi.org/10.3390/ijms20194667>
- Ramana, L. N., Sethuraman, S., Ranga, U., & Krishnan, U. M. (2010). Development of a liposomal nanodelivery system for nevirapine. *Journal of Biomedical Science*, 17(1), 57. <https://doi.org/10.1186/1423-0127-17-57>
- Rathos, M. J., Khanwalkar, H., Joshi, K., Manohar, S. M., & Joshi, K. S. (2013). Potentiation of in vitro and in vivoantitumor efficacy of doxorubicin by cyclin-dependent kinase inhibitor P276-00 in human non-small cell lung cancer cells. *BMC Cancer*, 13(1), 29. <https://doi.org/10.1186/1471-2407-13-29>
- Robert, N. J., Vogel, C. L., Henderson, I. C., Sparano, J. A., Moore, M. R., Silverman, P., Overmoyer, B. A., Shapiro, C. L., Park, J. W., Colbern, G. T., Winer, E. P., & Gabizon, A. A. (2004). The role of the liposomal anthracyclines and other systemic therapies in the management of advanced breast cancer. *Seminars in Oncology*, 31(6 Suppl 13), 106–146. <https://doi.org/10.1053/j.seminoncol.2004.09.018>
- Ronan, B., Flamand, O., Vescovi, L., Dureuil, C., Durand, L., Fassy, F., Bachelot, M.-F., Lamberton, A., Mathieu, M., Bertrand, T., Marquette, J.-P., El-Ahmad, Y., Filoche-Romme, B., Schio, L., Garcia-Echeverria, C., Goulaouic, H., & Pasquier, B. (2014). A highly potent and selective Vps34 inhibitor alters vesicle trafficking and autophagy. *Nature Chemical Biology*, 10(12), 1013–1019. <https://doi.org/10.1038/nchembio.1681>
- Rushton, L., Hutchings, S. J., Fortunato, L., Young, C., Evans, G. S., Brown, T., Bevan, R., Slack, R., Holmes, P., Bagga, S., Cherrie, J. W., & Van Tongeren, M. (2012). Occupational cancer burden in Great Britain. *British Journal of Cancer*, 107 Suppl 1(Suppl 1), S3-7. <https://doi.org/10.1038/bjc.2012.112>
- S, Y., & Hu, S. (2009). Autophagy in cancer and chemotherapy. *Results and Problems in Cell Differentiation*, 49. https://doi.org/10.1007/400_2008_25

- Salata, C., deAlmeida, C. E., Ferreira-Machado, S. C., Barroso, R. C., Nogueira, L. P., Mantuano, A., Pickler, A., Mota, C. L., & de Andrade, C. B. V. (2021). Preliminary pre-clinical studies on the side effects of breast cancer treatment. *International Journal of Radiation Biology*, 97(7), 877–887. <https://doi.org/10.1080/09553002.2021.1919782>
- Salatin, S., Maleki Dizaj, S., & Yari Khosroushahi, A. (2015). Effect of the surface modification, size, and shape on cellular uptake of nanoparticles. *Cell Biology International*, 39(8), 881–890. <https://doi.org/10.1002/cbin.10459>
- Salatin, S., & Yari Khosroushahi, A. (2017). Overviews on the cellular uptake mechanism of polysaccharide colloidal nanoparticles. *Journal of Cellular and Molecular Medicine*, 21(9), 1668–1686. <https://doi.org/10.1111/jcmm.13110>
- Schlütermann, D., Skowron, M. A., Berleth, N., Böhler, P., Deitersen, J., Stuhldreier, F., Wallot-Hieke, N., Wu, W., Peter, C., Hoffmann, M. J., Niegisch, G., & Stork, B. (2018). Targeting urothelial carcinoma cells by combining cisplatin with a specific inhibitor of the autophagy-inducing class III PtdIns3K complex. *Urologic Oncology: Seminars and Original Investigations*, 36(4), 160.e1-160.e13. <https://doi.org/10.1016/j.urolonc.2017.11.021>
- Sercombe, L., Veerati, T., Moheimani, F., Wu, S. Y., Sood, A. K., & Hua, S. (2015). Advances and Challenges of Liposome Assisted Drug Delivery. *Frontiers in Pharmacology*, 6, 286. <https://doi.org/10.3389/fphar.2015.00286>
- Shao, S., Li, S., Qin, Y., Wang, X., Yang, Y., Bai, H., Zhou, L., Zhao, C., & Wang, C. (2014). Spautin-1, a novel autophagy inhibitor, enhances imatinib-induced apoptosis in chronic myeloid leukemia. *International Journal of Oncology*, 44(5), 1661–1668. <https://doi.org/10.3892/ijo.2014.2313>
- Shimizu, S. (2015). Autophagic Cell Death and Cancer Chemotherapeutics. In K. Nakao, N. Minato, & S. Uemoto (Eds.), *Innovative Medicine* (pp. 219–226). Springer Japan. https://doi.org/10.1007/978-4-431-55651-0_18
- Shin, J. H., Park, C. W., Yoon, G., Hong, S. M., & Choi, K. Y. (2018). NNMT depletion contributes to liver cancer cell survival by enhancing autophagy under nutrient starvation. *Oncogenesis*, 7(8), 58. <https://doi.org/10.1038/s41389-018-0064-4>
- Shoshan, M. C., & Linder, S. (2008). Target specificity and off-target effects as determinants of cancer drug efficacy. *Expert Opinion on Drug Metabolism and Toxicology*, 4(3), 273–280. <https://doi.org/10.1517/17425255.4.3.273>
- Sideratou, Z., Kontoyianni, C., Drossopoulou, G. I., & Paleos, C. M. (2010). Synthesis of a folate functionalized PEGylated poly(propylene imine) dendrimer as prospective targeted drug delivery system. *Bioorganic &*

Medicinal Chemistry Letters, 20(22), 6513–6517.
<https://doi.org/10.1016/j.bmcl.2010.09.058>

Sishi, B. J. N., Loos, B., van Rooyen, J., & Engelbrecht, A.-M. (2013). Autophagy upregulation promotes survival and attenuates doxorubicin-induced cardiotoxicity. *Biochemical Pharmacology*, 85(1), 124–134.
<https://doi.org/10.1016/j.bcp.2012.10.005>

Soares, A. F., Carvalho, R. de A., & Veiga, F. (2007). Oral administration of peptides and proteins: Nanoparticles and cyclodextrins as biocompatible delivery systems. *Nanomedicine*, 2(2), 183–202.
<https://doi.org/10.2217/17435889.2.2.183>

Sritharan, S., & Sivalingam, N. (2021). A comprehensive review on time-tested anticancer drug doxorubicin. *Life Sciences*, 278, 119527.
<https://doi.org/10.1016/j.lfs.2021.119527>

Steichen, S. D., Caldorera-Moore, M., & Peppas, N. A. (2013). A review of current nanoparticle and targeting moieties for the delivery of cancer therapeutics. *European Journal of Pharmaceutical Sciences: Official Journal of the European Federation for Pharmaceutical Sciences*, 48(3), 416–427. <https://doi.org/10.1016/j.ejps.2012.12.006>

Sui, X., Kong, N., Wang, X., Fang, Y., Hu, X., Xu, Y., Chen, W., Wang, K., Li, D., Jin, W., Lou, F., Zheng, Y., Hu, H., Gong, L., Zhou, X., Pan, H., & Han, W. (2014). RETRACTED ARTICLE: JNK confers 5-fluorouracil resistance in p53-deficient and mutant p53-expressing colon cancer cells by inducing survival autophagy. *Scientific Reports*, 4(1), Article 1.
<https://doi.org/10.1038/srep04694>

Sun, T., Zhang, Y. S., Pang, B., Hyun, D. C., Yang, M., & Xia, Y. (2014). Engineered nanoparticles for drug delivery in cancer therapy. *Angewandte Chemie (International Ed. in English)*, 53(46), 12320–12364.
<https://doi.org/10.1002/anie.201403036>

Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., & Bray, F. (2021a). Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer Journal for Clinicians*, 71(3), 209–249.
<https://doi.org/10.3322/caac.21660>

Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., & Bray, F. (2021b). Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer Journal for Clinicians*, 71(3), 209–249.
<https://doi.org/10.3322/caac.21660>

Takahashi, Y., Coppola, D., Matsushita, N., Cualing, H. D., Sun, M., Sato, Y., Liang, C., Jung, J. U., Cheng, J. Q., Mul, J. J., Pledger, W. J., & Wang, H.-G. (2007). Bif-1 interacts with Beclin 1 through UVRAG and regulates

- autophagy and tumorigenesis. *Nature Cell Biology*, 9(10), Article 10. <https://doi.org/10.1038/ncb1634>
- Takamura, A., Komatsu, M., Hara, T., Sakamoto, A., Kishi, C., Waguri, S., Eishi, Y., Hino, O., Tanaka, K., & Mizushima, N. (2011). Autophagy-deficient mice develop multiple liver tumors. *Genes & Development*, 25(8), 795–800. <https://doi.org/10.1101/gad.2016211>
- Thai, A. A., Solomon, B. J., Sequist, L. V., Gainor, J. F., & Heist, R. S. (2021). Lung cancer. *The Lancet*, 398(10299), 535–554. [https://doi.org/10.1016/S0140-6736\(21\)00312-3](https://doi.org/10.1016/S0140-6736(21)00312-3)
- Tikhomirov, A. S., Shtil, A. A., & Shchekotikhin, A. E. (2018). Advances in the Discovery of Anthraquinone-Based Anticancer Agents. *Recent Patents on Anti-Cancer Drug Discovery*, 13(2), 159–183. <https://doi.org/10.2174/1574892813666171206123114>
- Tiyaboonchai, W. (2013). Chitosan Nanoparticles: A Promising System for Drug Delivery. *Naresuan University Journal: Science and Technology (NUJST)*, 11(3), Article 3.
- Trang, P., Wiggins, J. F., Daige, C. L., Cho, C., Omotola, M., Brown, D., Weidhaas, J. B., Bader, A. G., & Slack, F. J. (2011). Systemic Delivery of Tumor Suppressor microRNA Mimics Using a Neutral Lipid Emulsion Inhibits Lung Tumors in Mice. *Molecular Therapy*, 19(6), 1116–1122. <https://doi.org/10.1038/mt.2011.48>
- Vijay, K., Sowmya, P. R.-R., Arathi, B. P., Shilpa, S., Shwetha, H. J., Raju, M., Baskaran, V., & Lakshminarayana, R. (2018). Low-dose doxorubicin with carotenoids selectively alters redox status and upregulates oxidative stress-mediated apoptosis in breast cancer cells. *Food and Chemical Toxicology*, 118, 675–690. <https://doi.org/10.1016/j.fct.2018.06.027>
- Wang, A. Z., Langer, R., & Farokhzad, O. C. (2012). Nanoparticle delivery of cancer drugs. *Annual Review of Medicine*, 63, 185–198. <https://doi.org/10.1146/annurev-med-040210-162544>
- Wang, F., Wang, Y.-C., Dou, S., Xiong, M.-H., Sun, T.-M., & Wang, J. (2011). Doxorubicin-tethered responsive gold nanoparticles facilitate intracellular drug delivery for overcoming multidrug resistance in cancer cells. *ACS Nano*, 5(5), 3679–3692. <https://doi.org/10.1021/nn200007z>
- Whitaker, J. R., & Granum, P. E. (1980). An absolute method for protein determination based on difference in absorbance at 235 and 280 nm. *Analytical Biochemistry*, 109(1), 156–159. [https://doi.org/10.1016/0003-2697\(80\)90024-X](https://doi.org/10.1016/0003-2697(80)90024-X)
- Woodman, C., Vundu, G., George, A., & Wilson, C. M. (2021). Applications and strategies in nanodiagnosis and nanotherapy in lung cancer. *Seminars in*

- Cancer Biology, 69, 349–364.
<https://doi.org/10.1016/j.semancer.2020.02.009>
- Wu, Y., Yang, W., Wang, C., Hu, J., & Fu, S. (2005). Chitosan nanoparticles as a novel delivery system for ammonium glycyrrhizinate. International Journal of Pharmaceutics, 295(1–2), 235–245.
<https://doi.org/10.1016/j.ijpharm.2005.01.042>
- X, W., XI, W., HI, C., D, W., Jx, C., Xx, W., RI, L., Jh, H., L, M., X, C., Yq, W., & W, J. (2014). Ghrelin inhibits doxorubicin cardiotoxicity by inhibiting excessive autophagy through AMPK and p38-MAPK. Biochemical Pharmacology, 88(3). <https://doi.org/10.1016/j.bcp.2014.01.040>
- Xu, W.-H., Han, M., Dong, Q., Fu, Z.-X., Diao, Y.-Y., Liu, H., Xu, J., Jiang, H.-L., Zhang, S.-Z., Zheng, S., Gao, J.-Q., & Wei, Q.-C. (2012). Doxorubicin-mediated radiosensitivity in multicellular spheroids from a lung cancer cell line is enhanced by composite micelle encapsulation. International Journal of Nanomedicine, 7, 2661–2671. <https://doi.org/10.2147/IJN.S30445>
- Y, K., T, K., R, S., & S, K. (2005). The role of autophagy in cancer development and response to therapy. Nature Reviews. Cancer, 5(9).
<https://doi.org/10.1038/nrc1692>
- Yamamoto, T., Yokoyama, M., Opanasopit, P., Hayama, A., Kawano, K., & Maitani, Y. (2007). What are determining factors for stable drug incorporation into polymeric micelle carriers? Consideration on physical and chemical characters of the micelle inner core. Journal of Controlled Release: Official Journal of the Controlled Release Society, 123(1), 11–18. <https://doi.org/10.1016/j.jconrel.2007.07.008>
- Yan, C., Gu, J., Hou, D., Jing, H., Wang, J., Guo, Y., Katsumi, H., Sakane, T., & Yamamoto, A. (2015). Synthesis of Tat tagged and folate modified N-succinyl-chitosan self-assembly nanoparticles as a novel gene vector. International Journal of Biological Macromolecules, 72, 751–756. <https://doi.org/10.1016/j.ijbiomac.2014.09.031>
- Yang, D., Liu, Y., Bai, C., Wang, X., & Powell, C. A. (2020). Epidemiology of lung cancer and lung cancer screening programs in China and the United States. Cancer Letters, 468, 82–87.
<https://doi.org/10.1016/j.canlet.2019.10.009>
- Yang, F., Teves, S. S., Kemp, C. J., & Henikoff, S. (2014). Doxorubicin, DNA torsion, and chromatin dynamics. Biochimica et Biophysica Acta, 1845(1), 84–89. <https://doi.org/10.1016/j.bbcan.2013.12.002>
- Yu, Y., Yan, Y., Niu, F., Wang, Y., Chen, X., Su, G., Liu, Y., Zhao, X., Qian, L., Liu, P., & Xiong, Y. (2021). Ferroptosis: A cell death connecting oxidative stress, inflammation and cardiovascular diseases. Cell Death Discovery, 7(1), Article 1. <https://doi.org/10.1038/s41420-021-00579-w>

Zauner, W., Farrow, N. A., & Haines, A. M. R. (2001). In vitro uptake of polystyrene microspheres: Effect of particle size, cell line and cell density. *Journal of Controlled Release*, 71(1), 39–51. [https://doi.org/10.1016/S0168-3659\(00\)00358-8](https://doi.org/10.1016/S0168-3659(00)00358-8)

Zhang, Y., Xu, Y., Xi, X., Shrestha, S., Jiang, P., Zhang, W., & Gao, C. (2017). Amino acid-modified chitosan nanoparticles for Cu²⁺ chelation to suppress CuO nanoparticle cytotoxicity. *Journal of Materials Chemistry. B*, 5(19), 3521–3530. <https://doi.org/10.1039/c7tb00344g>