

# CONSTRUCTION OF TRIPEPTIDE HETEROLIGAND LIBRARY AS CAPTURING AGENT FOR MERCURY PLASMONIC DETECTION

By

KU SYARIDATUL IRMA BT KU ISMAIL

Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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December 2022

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Mercury is one of the priority metals classified as a human carcinogen by the US Environmental Protection Agency and the International Agency for Research on Cancer. This metallic element has a high degree of toxicity and is known to induce multiple organ damage and have severe adverse effects on human health and the environment, even at low levels of exposure. It has many forms in the soil, including inorganic and organic mercury. In this work, two novel tripeptides were designed and synthesized based on the amino-terminal  $Cu^{2+}$  and  $Ni^{2+}$  binding (ATCUN) motif. Two systems, namely monoligand and heteroligand systems, were compared in this work. Tripeptides were individually immobilized onto gold nanoparticles (AuNPs) surfaces via covalent coupling. In a monoligand system, only a particular tripeptide-AuNPs will be used as capturing agents for Hg<sup>2+</sup>, while in a heteroligand system, two different tripeptide-AuNPs will be used simultaneously in a mixture. The heteroligand system was found to be more effective compared to the monoligand system. The interaction of heteroligand enhances the selectivity and sensitivity of the plasmonic sensor for  $Hg^{2+}$ . Upon the addition of metal ions, the red-to-blue color change and the degree of AuNPs aggregation formed by the heteroligand system were doubled when compared to the monoligand system. These two novel tripeptides: 0.10 mM of pH 9 DCH (aspartic acid- cysteinehistidine) and 0.20 mM of pH 11 HCD (histidine-cysteine-aspartic acid) were selected among eleven novel tripeptides and one commercial tripeptide as the best capturing agents for Hg<sup>2+</sup> with an absorbance ratio (A<sub>683</sub>/A<sub>524</sub>) of 1.098. The finding was supported by UV-Vis spectra, Dynamic Light Scattering (DLS) spectroscopy, and Transmission Electron Microscopy (TEM) analysis. The limit of detection (LOD) for Hg<sup>2+</sup> detection was 0.025 parts per millions (ppm) with absorbance reading of 0.094. This new approach can constitute a more effective detection system targeting small molecules such as amino acids, metal ions and fatty acids.

Abstrak tesis yang dikemukakan kepada Senat of Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

## PEMBINAAN PERPUSTAKAAN HETEROLIGAN TRIPEPTIDA SEBAGAI EJEN PENANGKAP UNTUK PENGESANAN PLASMONIK MERKURI

Oleh

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Merkuri adalah salah satu logam utama yang diklasifikasikan sebagai karsinogen manusia menurut Agensi Perlindungan Alam Sekitar A.S. dan Agensi Antarabangsa untuk Penyelidikan Kanser. Unsur logam ini mempunyai tahap ketoksikan yang tinggi, diketahui boleh menyebabkan pelbagai kerosakan organ dan mempunyai kesan buruk terhadap kesihatan manusia dan alam sekitar walaupun pada tahap pendedahan yang rendah. Ia wujud dalam pelbagai bentuk dalam tanah, seperti merkuri tak organik dan merkuri organik. Dalam kajian ini, dua tripeptida novel telah direka dan disintesis berdasarkan motif pengikat terminal amino Cu<sup>2+</sup> and Ni<sup>2+</sup> ATCUN. Dua sistem, iaitu sistem monoligan dan heteroligan telah dibandingkan dalam kajian ini. Tripeptida secara individu dialihkan ke permukaan nanopartikel emas (AuNPs) melalui gandingan kovalen. Dalam sistem monoligan hanya satu tripeptida-AuNP tertentu akan digunakan, manakala dalam sistem heteroligan, dua jenis tripeptida-AuNPs akan digunakan dalam campuran. Sistem heteroligan didapati lebih berkesan berbanding sistem monoligan. Interaksi heteroligand meningkatkan selektiviti dan sensitiviti sensor plasmonik untuk Hg<sup>2+.</sup> Selepas penambahan ion logam, perubahan warna merah-ke-biru, dan tahap pengagregatan AuNPs yang dibentuk oleh sistem heteroligand adalah dua kali ganda jika dibandingkan dengan sistem monoligan. Kedua-dua tripeptida novel: 0.10 mM of pH 9 DCH (aspartat-sisteina-histidina) dan 0.20 mM of pH 11 HCD (histidina-sisteinaaspartat) ini dipilih antara sebelas tripeptida novel dan satu tripeptida komersial sebagai agen penangkap terbaik untuk Hg<sup>2+</sup> dengan nisbah penyerapan (A<sub>683</sub>/A<sub>524</sub>) sebanyak 1.098. Penemuan ini disokong oleh spektrum UV-Vis, spektroskopi Penyebaran Cahaya Dinamik (DLS), dan analisis Transmission Electron Microscopy (TEM). Had pengesanan (LOD) untuk pengesanan merkuri ialah 0.025 bahagian per million (ppm) dengan bacaan penyerapan 0.094. Pendekatan baharu ini berpotensi untuk membentuk sistem pengesanan yang lebih berkesan, terutamanya dalam menyasarkan molekul kecil seperti acid amino, ion metil dan acid lemak.

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As for myself, congratulations!

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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## LIST OF ABREVIATIONS

$As^{3+}$	Arsenic (III) ions
ATCUN motif	Amino terminal Cu <sup>2+</sup> and Ni <sup>2+</sup> -binding motif
Au	Gold
AuNPs	Gold nanoparticles
BME	2-mercaptoethanol/ß-mercaptoethanol
Cd <sup>2+</sup>	Cadmium (II) ions
Co <sup>2+</sup>	Cobalt (II) ions
-СООН	Carboxyl group
Cr <sup>3+</sup>	Chromium (III) ions
Cu <sup>2+</sup>	Copper (II) ions
DNA	Deoxyribonucleic acid
Fe <sup>3+</sup>	Iron (III) ions
fM	Femtomolar
FOM	Figure of merit
h	Hour
Hg <sup>2+</sup>	Mercury (II) ions
М	Molar
MDL	Minimum detection limit
mg	Milligram
min	Minute
mL	Millilitre
mM	Millimolar
Ν	Nitrogen
Ni <sup>2+</sup>	Nickel (II) ions

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nm	Nanometre
-NH <sub>2</sub>	Amino group
Pb <sup>2+</sup>	Lead (II) ions
ppb	Part per billion
ppm	Part per million
rpm	Revolution per minute
S	Sulfur
-SH	Thiol group
ТЕМ	Transmission electron microscope
Tripeptide-AuNPs	Tripeptide functionalized AuNPs
UV-vis	Ultraviolet-visible
Zn <sup>2+</sup>	Zinc (II) ions
%	Percent
μL	Microlitre
°C	Degree Celsius

#### **CHAPTER 1**

## INTRODUCTION

## 1.1 Introduction

Heavy metal pollution harms the environment because of its non-biodegradability and ability to accumulate in living organisms (E. Pehlivan et al., 2009). This is an inevitable cost of industrialization pressures that have increased irresponsible human activities, discharging byproducts directly into the rivers or other water reservoirs (Banares & Alvarez, 2015). Mercury is arguably the most hazardous metal pollutant in the environment. It causes severe diseases due to its physiological toxicity and neurotoxicity effects. The Occupational Health and Safety Authorities (OSHA) across the globe only permit 0.1 ppb level of its exposure, while contacting it at ten ppb will have an immediate danger (Buratti et al., 2019: Priyadarshini & Pradhan., 2017). Hg<sup>0</sup> and Hg<sup>2+</sup> pollution are common contaminants released from both natural sources such as volcano eruptions and anthropogenic emissions from industrial processes such as mining and fossil fuel combustion (Tangahu et al., 2011). Mercury can be emitted into the air and subsequently settle into the water. More noteworthy, the water-soluble mercury tends to be converted to methylmercury, the most toxic form of mercury, through biomethylation by microorganisms. This will eventually result in contamination of the food chain, which leads to poisoning or even malignancy (Priyadarshini & Pradhan., 2017: Fu et al., 2019). The World Health Organization (WHO) reports that among some subsistence fishing populations, between 1.5/1000 and 17/1000 kids displayed cognitive impairment (severe mental retardation) caused on by fish consumption containing mercury. These included the populations of Brazil, Canada, China, Columbia, and Greenland. In 2010, consumption of methylmercury was linked to 7,360 fatal heart attacks and a 0.14-point decline in the IQ of each foetus, according to a map of Hg-related health concerns in China. Chinese anthropogenic factors are responsible for about 61.8% (4532 fatal heart attacks) and 60.8% (0.08 points) of IQ declines. The remaining statistics relate to emissions from both domestic and foreign anthropogenic sources as well as natural processes such as volcanic eruptions, crustal weathering, and oceanic evasions (Chen, Liang & Liu, 2019). Therefore, close monitoring of mercury-befouled water in real-time becomes a crucial task.

### 1.2 **Problem statement**

Moreover, determining the pollutant in the environment is vital to discover a targeted region for the remediation process, which is at the frontline of research (Wang et al., 2020). Several advanced and sophisticated instruments were used to identify heavy metal ions in river or seawater samples, such as microwave plasma atomic emission spectroscopy (MP-AES (Ríos, Peňuela & Botero, 2017), atomic absorption spectrometry (AAS) (Bannon & Chilson, 2001), and inductively coupled plasma mass spectrometry (ICP-MS) (Yamakawa, Moriya & Yoshinaga, 2017). These instruments are operative and reliable. However, they come at a very high cost. Because these facilities are

laboratory bound and often operated by highly trained personnel, conducting a robust insitu analysis is less practical. Therefore, developing a new system that is easy to use and selective toward specific metals of concern is significant.

To detect such small targets, a colorimetric and plasmonic approach using gold nanoparticles (AuNPs) is an optimal method thanks to its unique properties of localized plasmon resonance and plasmon coupling effect (Priyadarshini & Pradhan, 2017). A standalone localized plasma of well dispersed AuNPs gives its solution a red-to-orange range of colors depending on the size and shape of the AuNPs. Meanwhile, when two or more particles come into proximity with a distance of less than 4 nm, plasmon coupling will occur, thus causing the red-shift of the absorbance peak spectra, which is associated with a red-to-purple or red-to-blue color change of the AuNPs solution. Governing such aggregation are molecular interaction forces between the AuNPs-surface bound capturing agent and the target (ions, molecules, etc.). Peptides, aptamers, and DNA are the usual capturing agent candidates to capture a specific target selectively. These capturing agents often serve as monoligands, using only one specific surface-bound bioreceptor. However, the aggregation has an entropic obstacle for the aggregates to be fixed in an optimal condition.

In this study, we propose using a heteroligand system to control the AuNP aggregation more efficiently in detecting target molecules. This study constructs a heteroligand tripeptide library, consisting of seventy-eight pairs of capturing agents combined from eleven novel and one commercial tripeptide. The tripeptides were designed based on the ATCUN (amino-terminal  $Cu^{2+}$  and  $Ni^{2+}$  binding) motif structure, with a thiol group at the center back of the tripeptide to anchor onto the AuNP surface via thiol-gold interaction. Tripeptides were individually conjugated onto the AuNPs surface and the interaction between seventy-eight pairs of capturing agents upon the addition of ten metal ions was studied. With a combination of two different tripeptides, the heteroligand-functionalized AuNPs detection system formed a more stable complex with the targeted  $Hg^{2+}$  than the monoligand systems.

## 1.3 Objectives

The objectives of this study are:

- To construct a tripeptide heteroligand library as capturing agents for mercury plasmonic detection
  - 1. To design eleven novel tripeptides that mimics ATCUN motif features for metal ions detection
  - 2. To optimize the tripeptide-AuNPs in order to enhance the sensing signal
  - 3. To study the effectiveness of tripeptide-AuNPs in targeting Hg<sup>2+</sup> and compare between the monoligand and heteroligand system

#### REFERENCES

- Aaryasomayajula, V.S.R., Severs, T., Ghosh, K., DeLong, R., Zhang, X., Talapatra, S., and Wanekaya, A.K. (2014) Assembly of a Dual Aptamer Gold Nanoparticle Conjugate Ensemble in the Specific Detection of Thrombin when Coupled with Dynamic Light Scattering Spectroscopy. *Journal of Nanomedicine & Nanotechnology*. 5:4
- Ali, H., Khan, E., & Sajad, M. A. (2013). Phytoremediation of heavy metals– concepts and applications. *Chemosphere*, 91(7), 869-881.
- Arai, Y., Lanzirotti, A., Sutton, S., Davis, J. A., & Sparks, D. L. (2003). Arsenic speciation and reactivity in poultry litter. *Environmental Science & Technology*, 37(18), 4083-4090
- Barbara, G., Wojciech, D., & Aneta H. B. (2020). Mercury in the terrestrial environment: A review. *Environmental Sciences Europe* 32:128
- Bannon, D. I., & Chisolm, J. J. (2001). Anodic stripping voltammetry compared with graphite furnace atomic absorption spectrophotometry for blood lead analysis. *Clinical Chemistry*, 47, 1703-1704
- Besra, L., & Liu, M. (2007). A review on fundamentals and applications of electrophoretic deposition (EPD). *Progress in Materials Science*, 52,1-61
- Byeong, J. Y., Byoung, G. K., Man, J. J., Se, Y. K., Hawn, C. K., Tae, W.K., Hong, J.C., Won, J.C., Mi, N.H., & Young, S.H. (2016). Evaluation of mercury exposure level, clinical diagnosis and treatment for mercury intoxication. *Annals of Occupational and Environmental Medicine*, 28:5
- Bañares, C. B., & Alvarez, M. L. C. (2015). Detection of the presence and concentration of heavy metals in selected rivers in the Province of Samar. *International Journal of Research-Granthaalayah*, 9,70-86
- Biplab, K. M., Nidhi, G., Taraknath, K., José, J.G. M. (2020). Designed Metal-ATCUN Derivatives: Redox- and Non-Redox-Based Applications Relevant for Chemistry, Biology, and Medicine. *iScience*, 23(12) 101792
- Burratti, L., Ciotta, E., Bolli, E., Kaciulis, S., Casalboni, M., Matteis F. D., Garzón-Manjón, A., Scheu, C., Pizzoferratoa, R., & Prosposito, P. (2019) Fluorescence enhancement induced by the interaction of silver nanoclusters with lead ions in water. *Colloids and Surfaces A*, 579,123634
- Boruah, B. S., & Biswas, R. (2018). An optical fiber-based surface plasmon resonance technique for sensing of lead ions: A toxic water pollutant. *Optical Fiber Technology*, 46, 152–156

- Bhavtosh, S., & Shweta, T. (2013). Simplification of metal ion analysis in freshwater samples by atomic absorption spectroscopy for laboratory students. *Journal of Laboratory Chemical Education*, 1(3), 54–58
- Bunz, U.H.F., & Rotello, V.M. (2010). Gold nanoparticle-flurophore complexes: sensitive and discerning noses for biosystems sensing, *Angewandte Chemie International Edition*, 49 (19), 3268–3279
- Camerman, N., Camerman, A., & Sarkar, B. (1975). Molecular design to mimic the copper (II) transport site of human albumin. The crystal and molecular structure of copper (II) – glycylglycyl-L-histidine-N-methyl amide monoaquo complex. *Canadian Journal of Chemistry*, 54, 1309-1316
- Cheng, H., Wu, C., Shen, L., Liu, J., & Xu, Z. (2014). Online anion exchange column preconcentration and high-performance liquid chromatographic separation with inductively coupled plasma mass spectrometry detection for mercury speciation analysis. *Analytica Chimica Acta*, 828, 9–16
- Chen, X., Han, C., Cheng, H., Wang, Y., Liu, J., Xu, Z., & Hu, L. (2013). Rapid speciation analysis of mercury in seawater and marine fish by cation exchange chromatography hyphenated with inductively coupled plasma mass spectrometry. *Journal of Chromatography A*, 1314, 86–93
- Chen, C., & Wang, J. (2020). Optical Biosensors: an exhaustive and comprehensive review. *The Analyst*, 145(5)
- Chen, L., Liang, S., & Liu, M. (2019) Trans-provincial health impacts of atmospheric mercury emissions in China. *Nature Communication* 10, 1484 (2019)
- Charles, T. D., Robert, P. M., Hing, M. C., Daniel, J. J., & Nicola, P. (2013). Mercury as a Global Pollutant: Sources, Pathways, and Effects. *Environment Science & Technology*, 47, 4967–4983
- Clifton, J.C. (2007) Mercury exposure and public health. *Pediatric Clinic North America*, 54(2), 237-269
- Colin, D. B., Barry, T., Yu, T.T., Joseph, E., George, M. W., Ralph, G. N. (1989).
  Formation of monolayer films by the spontaneous assembly of organic thiols from solution onto gold. *Journal of the American Chemical Society*, 111, 321–335
- Daryoush, M., Abbas, Z. K., Akif, K., & Wei, D. (2011). "Nano-plasmonic biosensors: A review," *The 2011 IEEE/ICME International Conference on Complex Medical Engineering*, pp. 31-36
- Elena, A. E., Mark, M. J. R., Nico, S., Gert, S. G., Joke, A. B., Aimee, L. B., & Alexander K. (2020). One Peptide for Them All: Gold Nanoparticles of Different Sizes Are Stabilized by a Common Peptide Amphiphile, ACS Nano, 14 (5), 5874-5886

- Eustis, S., & El-Sayed, M.A. (2006). Why gold nanoparticles are more precious than pretty gold: noble metalsurface plasmon resonance and its enhancement of the radiative and nonradiative properties of nanocrystals of different shapes, *Chemical Society Reviews*, 35,209–217
- Estevez, M.C., Alvarez, M., & Lechuga, L. (2012). Integrated optical devices for labon-a-chip biosensing applications. *Laser & Photonics Review*, 6, 463
- Fu, L., Xie, K., Wang, A., Lyu, F., Ge, J., Zhang, L., Zhang, H., Su, W., Hou, Y., Zhou, C., Wang, C., & Ruan, S. (2019). High selective detection of mercury (II) ions by thioether side groups on metal-organic frameworks. *Analytica Chimica Acta*, 1081, 51-58
- Guangyang, L., Meng, L., Xiaodong, H., Tengfei, Li., & Donghui, Xu. (2018). Application of gold-nanoparticle Colorimetric Sensing to Rapid Food Safety Screening. Sensors, 18, 4166
- Guo, Y., Wang, Z., Qu, W., Shao, H., & Jiang, X. (2011). Colorimetric detection of mercury, lead and copper ions simultaneously using protein-functionalized gold nanoparticles. *Biosensors and Bioelectronics*, 26(10), 4064-4069.
- Hai, C., Kai, Z., Guanghua, Z. (2018). Gold nanoparticles: From synthesis, properties to their potential application as colorimetric sensors in food safety screening, *Trends in Food Science & Technology*, 78, 83-94
- Han, F. X., Patterson, W. D., Xia, Y., Sridhar, B. B. M., & Su, F. (2006). Rapid determination of mercury in plant and soil samples using inductively coupled plasma atomic emission spectroscopy, a comparative study. *Water, Air, and Soil Pollution*, 170(1–4), 161–171
- Harford, C., & Sarkar, B. (1997). Amino terminal Cu (II)- and Ni (II)-binding (ATCUN) motif of proteins and peptides: metal binding, DNA cleavage, and other properties. *Accounts of Chemical Research*, 30, 123-130
- Hu, X.F., Lowe, M., Chan, H.M. (2021). Mercury exposure, cardiovascular disease, and mortality: A systematic review and dose-response meta-analysis. *Environmental Research*, 193:110538
- Imre, S., & Katalin, O. (2006). Metal ion selectivity of oligopeptides. *The Royal* Society of Chemistry Dalton Transaction, 32:3841–3854
- Ismail, K.S.I.K., Tajudin, A.A., Ikeno, S., Amir, S.A.H. (2022). Heteroligand nanoarchitectonics of functionalized gold nanoparticle for Hg2+ detection. *Journal of Nanoparticle Research*, 24, 253
- Jain, P.K., Lee, K.S., El-Sayed, I.H., & El-Sayed, M.A. (2006). Calculated absorption and scattering properties of gold nanoparticles of different size, shape, andcomposition: applications in biological imaging and biomedicine, *Journal* of Physical Chemistry B, 110,7238–7248

- Jia, Y., Yan, X., Guo, X., Zhou, G., Liu, P., & Li, Z. (2019) One Step Preparation of Peptide-Coated Gold Nanoparticles with Tunable Size. *Materials* (*Basel*),12(13):2107
- Jun H, Fengjuan C, Junxia S, Fu X, Baodui W (2018) Porous Wood Members-Based Amplified Colorimetric Sensor for Hg<sup>2+</sup> Detection through Hg<sup>2+</sup>-Triggered Methylene Blue Reduction Reactions. *Analytical Chemistry*, 90 (7), 4909-4915
- Kevin, M. R., Ernest, M., Walker, J., Miaozong, W., Chris, G., & Eric, R. B. (2014). Environment Mercury and Its Toxic Effects. *Journal of Preventive Medicine* and Public Health, 47,74-83
- Kim, Y. R., Mahajan, R. K., Kim, J. S., & Kim, H. (2010) Highly sensitive gold nanoparticle-based colorimetric sensing of mercury (II) through simple ligand exchange reaction in aqueous media. ACS Applied Materials and Interfaces, 2(1), 292–295
- K. Lance, Kelly., Eduardo, C., Lin, L. Z., & George, C. S. (2003). The Optical Properties of Metal Nanoparticles: The Influence of Size, Shape, and Dielectric Environment. *The Journal of Physical Chemistry B*, 107 (3), 668-677
- Khan, A.U., Zhao, S., & Liu, G. (2016). Key Parameter Controlling the Sensitivity of Plasmonic Metal Nanoparticles: Aspect Ratio. *Journal Physical Chemistry C*, 120(34),19353
- Lerner, N., Ohaion-Raz, T., Zeiri O. (2020) Improving the properties of a gold nanoparticle barium sensor through mixed-ligand shells. *Talanta*, 208:120370
- Lévy, R.; Thanh, N.T.K.; Doty, R.C.; Hussain, I.; Nichols, R.J.; Schiffrin, D.J.; Brust, M.; Fernig, D.G. (2004) Rational and Combinatorial Design of Peptide Capping Ligands for Gold Nanoparticles. *Journal of the American Chemical Society*. 126, 10076–10084
- Long, F., Zhu, A., Shi, H., Wang, H., & Liu, J. (2013). Rapid on-site/in-situ detection of heavy metal ions in environmental water using a structure-switching DNA optical biosensor. *Scientific reports*, 3:2308
- Lin, H. Y., Huang, C. H., Chen, S. H., Liu, Y. C., Chang, W. Z., & Chau, L. K. (2013). Tubular waveguide evanescent field absorption biosensor based on particle plasmon resonance for multiplex label-free detection. *Biosensors and Bioelectronics*, 41, 268-274
- Liu, J., Jalali, M., Mahshid, S., & Wachsmann-Hogiu, S. (2020). Are plasmonic optical biosensors ready for use in point-of-need applications? *Analyst*, 145(2), 364–384
- Maruccio, G., Primiceri, E., Marzo, P., Arima, V., Torre, A. D., Rinaldi, R., Pellegrino, T., Krahne, R., & Cingolani, R. (2009). A nanobiosensor to detect single hybridization events. *Analyst*, 134, 2458–2461

- Mahato, K., Nagpal, S., Shah, M. A., Srivastava, A., Maurya, P. K., Roy, S., Jaiswal, A., Singh, R., & Chandra, P. (2019). Gold nanoparticle surface engineering strategies and their applications in biomedicine and diagnostics. *Biotech*, 9(2), 0.
- Melnick, J. G., & Parkin, G. (2007). Cleaving mercury-alkyl bonds: a functional model for mercury detoxification by *MerB. Science*, 317(5835), 225-227
- Mudarikwa, L. V., Nyoni, M. S., Munyuki, G., & Nyoni, S. (2020) Development of a colorimetric probe for the semi-quantitative detection of mercury levels in water. *Results in Chemistry*, 2, 100076
- Myroshnychenko, V., Rodriguez-Fernandez, J., Pastoriza-Santos, I., Funston, A.M., Novo, C., Mulvaney, P., Liz-Marzan, L.M., De Abajo, F.J.G. (2008). Modelling the optical response of gold nanoparticles. *Chemical Society Reviews*, 37, 1792– 1805
- Narayana, R. I., David, L. S., & Tuan, V. (1998). Surface-Enhanced Raman Gene Probe for HIV Detection. *Analytical Chemistry*. 70 (7), 1352-1356
- Njoki, P.N., Lim, I.I.S., Mott, D., Park, H.Y., Khan, B., Mishra, S., Sujakumar, R., Luo, J., & Zhong, C.J. (2007). Size correlation of optical and spectroscopic properties for gold nanoparticles, *Journal of Physical Chemistry C*, 111 (40)14664–14669
- Omichinski, J. G. (2007). Toward methylmercury bioremediation. *Science*, 317(5835), 205-206
- Prashant, K. J., Xiaohua, H., Ivan, H. E., & Mostafa, A. E. (2008). Noble Metals on the Nanoscale: Optical and Photothermal Properties and Some Applications in Imaging, Sensing, Biology, and Medicine. Accounts of Chemical Research, 41 (12), 1578-1586
- Peters, T. (1960). Interaction of one mole of copper with the alpha amino group of bovine serum albumin. *BBA Biochimica et Biophysica Acta*, 39(3), 546–547
- Pehlivan, E., Ozkan, A. M., Dinc, S., & Parlayici, S. (2009). Adsorption of Cu<sup>2+</sup> and Pb<sup>2+</sup> ion on dolomite powder. *Journal of Hazardous Materials*, 167, 1044–1049
- Priyadarshini, E., Pradhan, N. (2017). Gold nanoparticles as efficient sensors in colorimetric detection of toxic metal ions: A Review. Sensors and Actuators B: Chemical, 238, 888-902
- Ríos, S. E. G., Peñuela, G. A., & Botero, C. M. R. (2017). Method validation for the determination of mercury, cadmium, lead, arsenic, copper, iron, and zinc in fish through Microwave-Induced Plasma Optical Emission Spectrometry (MIP OES). *Food Analytical Methods*, 10, 3407-3414

- Ripp, S., DiClaudio, M. L., & Sayler, G. S. (2010). Biosensors as environmental monitors. *Environmental Microbiology*, 2, 213-233
- Salimi, F., Kiani, M., Karami, C., & Taher, M. A. (2018). Colorimetric sensor of detection of Cr (III) and Fe (II) ions in aqueous solutions using gold nanoparticles modified with methylene blue. *Optik*, 158, 813–825
- Sarah, U., Ian, B., Jie, H., & Laura, S. (2015). Localized Surface Plasmon Resonance Biosensing: Current Challenges and Approaches. Sensors, 15, 15684-15716
- Schlucker, S. (2014). Surface-Enhanced Raman Spectroscopy: Concepts and Chemical Applications. Angewandte Chemie International Edition, 53, 4756– 4795
- Sharpe, J.C., Mitchell, J.S., Lin, L., Sedoglavich, H., & Blaikie, R.J. (2008). Gold nanohole array substrates as immunobiosensors. *Analytical Chemistry*, 80, 2244–2249
- Shouting Z, Dongxu Z, Xuehong Z, Denghui S, Zhonghua X, Duoliang S, Xiaoquan L (2017) Ultratrace Naked-Eye Colorimetric Detection of Hg<sup>2+</sup> in Wastewater and Serum Utilizing Mercury-Stimulated Peroxidase Mimetic Activity of Reduced Graphene Oxide-PEI-Pd Nanohybrids. *Analytical Chemistry*, 89 (6), 3538-3544
- Tangahu, B. V., Sheikh Abdullah, S. R., Basri, H., Idris, M., Anuar, N. & Mukhlisin, M. (2011). A review on heavy metals (As, Pb, and Hg) uptake by plants through phytoremediation. *International Journal of Chemical Engineering*, (2011) 939161
- Turdean, G. L. (2011). Design and development of biosensors for the detection of heavy metal toxicity. *International Journal of Electrochemistry*, 1–15
- Turner, A. P. (2013). Biosensors: sense and sensibility. *Chemical Society Reviews*, 42(8), 3184-3196
- Verma, N., & Singh, M. (2005). Biosensors for heavy metals. *BioMetals*, 18(2), 121–129
- Wang, L., Hou, D., Cao, Y., Sik, O. Y., Tack, F. M. G., Rinklebe, J., & O'Connor, D. (2020). Remediation of mercury contaminated soil, water, and air: A review of emerging materials and innovative technologies. *Environment International*, 134,105281
- Willets, K.A., & Van Duyne, R.P. (2007). Localized surface plasmon resonance spectroscopy and sensing. Annual Review of Physical Chemistry, 58:267-97
- Wu, L., Long, Z., Liu, L., Zhou, Q., Lee, Y. I., & Zheng, C. (2012). Microwaveenhanced cold vapor generation for speciation analysis of mercury by atomic fluorescence spectrometry. *Talanta*, 94, 146–151

- Weiyang, L., Pedro, H.C. C., Xianmao, Lu., & Younan, X. (2009). Dimers of Silver Nanospheres: Facile Synthesis and Their Use as Hot Spots for Surface-Enhanced Raman Scattering. *Nano Letters*, 9(1), 485–490
- Wu, Y., Zhan, S., Wang, F., He, L., Zhi, W., & Zhou, P. (2012). Cationic polymers and aptamers mediated aggregation of gold nanoparticles for the colorimetric detection of arsenic (III) in aqueous solution. *Chemical Communications*, 48(37):4459-61
- Yamakawa, A., Moriya, K., & Yoshinaga, J. (2017). Determination of isotopic composition of atmospheric mercury in urban-industrial and coastal regions of Chiba, Japan, using cold vapor multi-collector inductively coupled plasma mass spectrometry. *Chemical Geology*, 448, 84-92
- Yang, Y., Yildiz, U. H., Peh, J., & Liedberg, B. (2015). Ternary DNA chip based on a novel thymine spacer group chemistry. *Colloids and Surfaces B: Biointerfaces*, 125, 270–276
- Yu, C. J., & Tseng, W. L. (2008). Colorimetric detection of mercury (II) in a highsalinity solution using gold nanoparticles capped with 3-mercaptopropionate acid and adenosine monophosphate. *Langmuir*, 24(21), 12717-12722