



**BIOTRANSFORMATION OF GLUTEN-FREE COMPOSITE FLOUR  
MEDIATED BY LACTIC ACID BACTERIA VIA SOLID-STATE  
FERMENTATION PROCESS CONDUCTED UNDER DIFFERENT MOISTURE  
CONTENTS**

By  
**KAREEM KOYUM ADEBAYO**

Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of of Master of  
Science

June 2022

FBSB 2022 21

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement for the degree of Master of Science

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**June 2022**

**Chair : Professor Foo Hooi Ling, PhD**  
**Faculty : Biotechnology and Biomolecular Sciences**

Over the years, the development of staple food products from composite flour is considered a low-cost approach to address the burden of protein-energy malnutrition in developing countries. However, despite the numerous advantages of composite flour, the high amount of anti-nutritional factors typical to food derived from plant origin usually impede nutrient bioavailability. Five strains of lactic acid bacteria (LAB) isolated from Malaysian foods that include *Lactiplantibacillus plantarum* RG-14, *L. plantarum* RI-11, *L. plantarum* RS5, *L. plantarum* IUL-4, and *Pediococcus pentosaceus* UP-2 have been reported for their capabilities to produce various extracellular hydrolytic enzymes via solid-state fermentation (SSF) which can breakdown complex food matrix into smaller absorbable forms and reduced antinutrients. Therefore, the LAB strains were employed in this study to biotransform the gluten-free composite flour derived from rice, sorghum, and soybean. The SSF process was performed under 30-60% moisture content for 7 days, where samples were withdrawn at 24 h intervals for various analyses such as LAB cell viability, pH, total titratable acidity, extracellular protease activity, soluble protein concentration, crude protein content, and *in vitro* protein digestibility. The pH of the biotransformed composite flour showed a significant reduction from the initial range of pH 5.98 - 6.67 to the final pH of 4.36 - 3.65, corresponding to the increase in the percentage of total titratable acidity in the range of 0.28 - 0.47% to 1.07 - 1.65% from Day 0-4 and remained stable till Day 7 of the SSF process. The LAB strains exhibited high extracellular proteolytic activity (0.63 - 1.35 U/mg to 4.21 - 5.13 U/mg) from Day 0-7. In addition, the treated composite flour soluble protein increased significantly ( $p \leq 0.05$ ) (0.58 - 0.60 mg/mL to 0.72 - 0.79 mg/mL) from Day 0-7, crude protein content (12.00 - 12.18% to 13.04 - 14.39%) and protein digestibility (70.05 - 70.72% to 78.46 - 79.95%) from Day 0-4 of SSF. In addition, the antinutritional factors of the biotransformed composite flour showed a significant reduction ( $p \leq 0.05$ ) in the phytic acid (127.11 – 137.73 mg/100 g to 124.84 -

120.24 mg/100 g) and tannin content (89.48 – 93.92 mg/100 g to 63.51 – 39.84 mg/100g). Since lower moisture content promotes flour quality, 50% moisture was selected as the most suitable moisture content to have effectively biotransformed the composite flour, even though a comparable result was observed at 60% moisture content. Overall, *Lactiplantibacillus plantarum* RG-14 was ranked the best strain attributed to the general improvement in composite flour's pH, TTA, protein quality, and antinutritional properties.

Keywords: composite flour; lactic acid bacteria; solid-state fermentation; moisture content; nutritive quality

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**BIOTRANSFORMASI TEPUNG KOMPOSIT BEBAS GLUTEN DIMEDIASI  
BAKTERIA ASID LAKTIK PENAPAIAN PEPEJAL YANG DIJALANKAN DI  
BAWAH KANDUNGAN KELEMBAPAN YANG BERBEZA**

Oleh

**KAREEM KOYUM ADEBAYO**

**Jun 2022**

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**Fakulti : Bioteknologi dan Sains Biomolekul**

Pembangunan produk makanan ruji daripada tepung komposit dari tahun ke tahun dianggap sebagai pendekatan kos rendah untuk menangani beban malnutrisi tenaga protein di negara membangun. Walaupun terdapat banyak kelebihan tepung komposit, namun jumlah faktor anti-pemakanan yang tinggi dan tipikal kepada makanan yang diperoleh daripada asal tumbuhan biasanya menghalang bioavailabiliti nutrien. Lima strain bakteria asid laktik (LAB) iaitu *Lactiplantibacillus plantarum* RG-14, *L. plantarum* RI-11, *L. plantarum* RS5, *L. plantarum* IUL-4, dan *Pediococcus pentosaceus* UP-2 yang telah diasingkan daripada makanan negara Malaysia dilaporkan mempunyai keupayaan untuk menghasilkan pelbagai enzim hidrolitik ekstraselular melalui penapaian keadaan pepejal (SSF) yang boleh memecahkan matrik makanan kompleks kepada bentuk yang lebih kecil dan mudah diserap serta mengurangkan antinutrien. Oleh itu, strain LAB telah digunakan dalam kajian ini untuk biotransformasi tepung komposit bebas gluten yang diperoleh daripada beras, sekoi dan kacang soya. Kaedah SSF dilakukan di bawah 30-60% kandungan lembapan dan dijalankan selama 7 hari. Sampel diambil pada selang 24 jam untuk analisis pH, jumlah keasidan boleh titrasi, aktiviti protease ekstraselular, kepekatan protein larut, kandungan protein kasar, dan kebolehcernaan protein *in vitro*. Nilai pH tepung komposit biotransformasi menunjukkan pengurangan yang ketara, dari nilai pH permulaan 5.98 – 6.67 ke nilai pH akhiran 4.36 – 3.65, sepadan dengan peningkatan peratusan jumlah keasidan boleh titrasi (0.28 – 0.47% ke 1.07 – 1.65%) yang dicatatkan pada hari pertama hingga hari keempat, dan nilai peratusan kekal stabil sehingga hari ketujuh. Strain LAB menunjukkan aktiviti proteolitik ekstraselular yang tinggi (0.63 - 1.35 U/mg ke 4.21 - 5.13 U/mg) dari hari pertama hingga hari ketujuh. Selain itu, nilai peningkatan yang ketara ditunjukkan pada hari pertama hingga hari ketujuh pada protein larut tepung komposit yang dirawat ( $p \leq 0.05$ ) (0.58 - 0.60 mg/mL kepada 0.72 - 0.79 mg/mL), dan kandungan protein kasar (12.00 - 12.18% kepada 13.04 - 14.39%) serta kebolehcernaan protein (70.05 - 70.72% kepada 78.46 - 79.95%) pada hari

pertama ke hari keempat ujian SSF dijalankan. Faktor antinutrisi tepung komposit biotransformasi menunjukkan pengurangan ketara ( $p \leq 0.05$ ) dalam asid fitak (127.11 – 137.73 mg/100 g kepada 124.84 - 120.24 mg/100 g) dan kandungan tanin (89.48 – 93. 100 g hingga 63.51 – 39.84 mg/100g). Kandungan lembapan yang lebih rendah menggalakkan kualiti tepung,dan sebanyak 50% lembapan telah dipilih sebagai kandungan lembapan yang paling sesuai untuk mentransformasikan tepung komposit secara berkesan, walaupun hasil yang setanding diperhatikan pada kandungan lembapan 60%. Secara keseluruhan, *Lactiplantibacillus plantarum* RG-14 disenaraikan sebagai strain terbaik yang dikaitkan dengan peningkatan umum dalam pH tepung komposit, TTA, kualiti protein dan sifat antinutrisi.

Kata Kunci: tepung komposit; bakteria asid laktik; penapaian keadaan pepejal; kandungan lembapan; kualiti pemakanan

## **ACKNOWLEDGEMENTS**

My sincere appreciation goes to my supervisor, Prof. Dr. Foo Hooi Ling that allowed me to embark on this research project and guide me throughout the excitingly challenging task. I would also like to appreciate my Co-supervisor Assoc. Prof. Norhayati Ramli for her support during my study period and appreciate the collective teamwork of the supervisory committee for their collaborations, and recommendations to make this project a dream come true. Also, I much appreciate the staff of the Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia, for their contribution in facilitating the smooth, successful completion of my research work.

Furthermore, I deeply appreciate the profound support of the Queen Elizabeth Commonwealth Scholarships and the School of Graduate Studies (SGS), Universiti Putra Malaysia for finding me worthy of the award during the selection process and providing the finance that facilitated the journey of my research career. Also, my appreciation goes to my family, and friends who have been there for me all these years and finally, I am indebted to the memory of my late father (Mr. Kareem. M. Aderemi) for his special love and moral support. May Almighty Allah reward his efforts and admit him to Al-janat fridaous (Ameen).

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

ANOVA	analysis of variance
BSA	bovine serum albumin
CFU	colony forming unit
EMB	eosin methylene blue agar
HCl	hydrochloric acid
IUL-4	<i>Lactiplantibacillus plantarum</i> IUL-4
IVPD	<i>in vitro</i> protein digestibility
LAB	lactic acid bacteria
MC	moisture content
MRS	de man, rogosa sharpe medium agar
NTC	raw composite flour with no water and culture
PEM	protein-energy malnutrition
PKC	palm kernel cake
RG-14	<i>Lactiplantibacillus plantarum</i> RG-14
RI-11	<i>Lactiplantibacillus plantarum</i> RI-11
RS-5	<i>Lactiplantibacillus plantarum</i> RS-5
RTU	ready to use culture
SDB	sabouraud dextrose agar
SMF	submerged fermentation
SSF	solid-state fermentation
TCA	trichloroacetic acid
TTA	total titratable acidity
UP-2	<i>Pediococcus pentosaceus</i> UP-2
WTC	water-treated composite with no culture

## CHAPTER 1

### INTRODUCTION

Malnutrition resulting from protein-energy deficiencies remains a major health burden in developing countries with approximately 821 million people including 667 million children currently undernourished (FAO/WHO, 2019). Protein-energy malnutrition (PEM) occurs due to insufficient intake of quality protein and the symptoms include marasmus, kwashiorkor, stunted growth, and underweight. According to the global prevalence report on PEM, Africa and Asia had the highest occurrence of the health burden (FAO, 2018). Similarly, several studies have established the consumption of low-quality diets as one of the major factors responsible for the high PEM in developing countries (Adesogan et al., 2019; Nazri et al., 2020). Food grains obtained from cereals and legumes provide the cheapest source of dietary energy such as protein, fat, vitamins, and minerals for people in developing countries (Gilani et al., 2012; Udomkun et al., 2019). According to the global statistics report for cereals and legumes consumption, nearly 3.5 billion people consume rice (FAO, 2016), about 300 million people in Africa depend on sorghum-based staple foods while millions of people globally relied on soybean (Dicko et al., 2006; FAO, 2019). Although rice, sorghum, and soybean are cheap and accessible to vulnerable populations, over-reliance on a single type of food (monotonous diet) can lead to nutritional deficiencies (Govindaraj, 2015), thus, suggesting composting cereals and legumes to produce nutritious foods (Temba et al., 2016).

In 1964, the Food and Agricultural Organization (FAO) introduce the concept of composite flour technology as an attempt to reduce the financial burden of wheat importation in developing countries (Hasmadi et al., 2020). Composite flour involves the mixture of wheat with varying proportions of legumes, tubers, cereals, and pseudocereals to develop food products (Melini et al., 2020). However, the major concern for wheat usage in food production is the presence of gluten protein which has been linked to the incidence of celiac disease with a global prevalence rate of 2.4% to 44% (Bolarinwa & Oyesiji, 2021; Mahadev & Green, 2011). Hence, recent research focuses on the development of non-wheat composite flour for use in food production (Melini et al., 2017). Rice, sorghum, and soybean are gluten-free and widely consumed in developing countries. Thus, the development of protein-rich composite flour from rice, sorghum, and soybean grains offers a vast potential to alleviate PEM. Food products made from composite flour derived from rice, sorghum, and soybean have been reported to possess features capable of replacing the wheat-based composite (Adeyeye, 2018; Adeyeye et al., 2017; Seth & Rajamanickam, 2012). However, despite the low-cost and high accessibility advantage of composite flour derived from rice, sorghum, and soybean, staple foods derived from plant sources contains a high amount of antinutritional factors that impede nutrient bioavailability (Schönfeldt & Hall, 2012; Udomkun et al., 2019).

Protein quality interrelates with protein digestibility (Boye et al., 2012) and research efforts over the years on composite flour have focused on improving the protein content/lysine limitations in cereal-based food products. A study on food products developed from rice, sorghum, millet, and soybean composite flour (Seth & Rajamanickam, 2012), rice, sorghum, and soybean composite flour (Omwamba & Mahungu, 2014) via extrusion processing method recorded a significant loss in lysine content and does not account for protein digestibility and anti-nutritional factors in composite flour. Food processing is meant to improve nutritive quality in food products (Joye, 2019) and the biotransformation process mediated by wholly microorganisms via solid-state fermentation technique (SSF) holds a promising potential to improve food quality.

SSF technique is a bioprocess carried out in the absence or near-absence of free water, although, the moisture content of the substrate must be sufficient to support microbial growth and metabolic activity (Thomas et al., 2013). SSF is eco-friendly, energy-efficient, low risk of contamination risk, and high bioproducts productivity (Thomas et al., 2013; Webb & Manan, 2017). Apart from moisture content being an important criterion to develop the SSF process, the choice of microorganisms also plays a significant role in determining the success of the bioprocess (Thomas et al., 2013). Fungi are considered the ideal microorganisms for SSF since they mimic their natural habitat, however, the possibilities of mycotoxin contamination have limited fungal application in food production (Adebiyi et al., 2019). Contrarily, bacteria such as the lactic acid bacteria (LAB) that are known to exhibit limited growth under SSF conditions are currently being applied as biotransformation agents for many bioprocesses to develop bioproducts from cheap agricultural biomass (Lee et al., 2019).

LAB are Gram-positive rod or cocci non-respiring, aerotolerant, fastidious, catalase-negative, acid-tolerant, and nonsporulating bacteria. They are generally referred as safe (GRAS) and are widely known for their functional role in food fermentation (Zabidi et al., 2020). In this study, a few strains of LAB such as *Lactiplantibacillus plantarum* RI-11, *L. plantarum* RG-14, *L. plantarum* RS-5, *L. plantarum* IUL-4, and *Pediococcus pentocaseous* UP-2 isolated from Malaysian foods were recently reported for their capabilities to induce extracellular hydrolytic enzymes under SSF conditions (Lee et al., 2019). The extracellular hydrolytic enzymes induced by the LAB are capable of breaking down complex food matrix into digestible forms (Kårlund et al., 2020). However, previous application of these LAB and their postbiotic metabolites have focused on improving feed quality (Lee et al., 2019) and livestock health (Izuddin et al., 2020; Merzza et al., 2019). Therefore, the current study aimed at evaluating the effects of moisture contents on the SSF of composite flour derived from rice sorghum and soybean mediated by the selected LAB via the following specific objectives:

- i. To determine the cell viability of selected LAB, pH, and total titratable acidity of biotransformed gluten-free composite flour mediated by selected LAB under different moisture contents.

- ii. To determine the extracellular protease activity, protein concentration and protein digestibility of biotransformed gluten-free composite flour mediated by selected LAB under different moisture contents.
- iii. To determine the antinutritional factors of biotransformed gluten-free composite flour mediated by selected LAB under different moisture contents.

### **1.1 Research Hypothesis**

Rice and sorghum basically contain low protein. Soybean is cheaper protein source compared to animal origin food. The protein content of gluten-free composite flour derived from rice and sorghum can be improved by adding soybean flour. However, the presence of antinutritional factors in the composite flour usually affect the protein digestibility. Thus, to further improve the protein content/digestibility of the gluten-free composite flour the following hypothesis were proposed:

Ho - Biotransformation process mediated via SSF by various lactic acid bacteria isolated from Malaysian foods will further enhance the protein content/digestibility of gluten-free composite flour

Ho - Moisture content will have significant effect on the biotransformation of gluten-free composite flour mediated via SSF by various lactic acid bacteria isolated from Malaysian foods

## REFERENCES

- Abedi, E., & Hashemi, S. M. B. (2020). Lactic acid production – producing microorganisms and substrates sources-state of art. *Helijon*, 6(2020), 1–32. <https://doi.org/10.1016/j.heliyon.2020.e04974>
- Acevedo-Rocha, C. G., Gronenberg, L. S., Mack, M., Commichau, F. M., & Genee, H. J. (2019). Microbial cell factories for the sustainable manufacturing of B vitamins. *Current Opinion in Biotechnology*, 56, 18–29. <https://doi.org/10.1016/j.copbio.2018.07.006>
- Achi, O. K., & Asamudo, N. U. (2019). Cereal-based fermented foods of africa as functional foods (pp. 1527–1558). Springer Nature Switzerland AG 2019 J.-M. Mérillon, K. G. Ramawat (eds.), Bioactive. <https://doi.org/10.1007/978-3-319-78030-6>
- Adebiyi, J. A., Kayitesi, E., Adebo, O. A., Changwa, R., & Njobeh, P. B. (2019). Food fermentation and mycotoxin detoxification: An african perspective. *Food Control*, 106(May), 106731. <https://doi.org/10.1016/j.foodcont.2019.106731>
- Adebo, O. A. (2020). African sorghum-based fermented foods: past, current and future prospects. *Nutrients Review*, 12(1111), 2–25.
- Adejuwon, O. H., Jideani, A. I. O., & Falade, K. O. (2020). Quality and public health concerns of instant noodles as influenced by raw materials and processing technology. *Food Reviews International*, 36(3), 276–317. <https://doi.org/10.1080/87559129.2019.1642348>
- Adeoye, B. K., Akinbode, B. A., Adesida, A. A., & Akpa, C. T. (2020). Production and evaluation of a protein-enriched meal from composite flour of cassava, rice and soybean. *SDRP Journal of Food Science & Technology*, 5(1), 18–26. <https://doi.org/10.25177/jfst.5.1.ra.10594>
- Adesogan, A. T., Havelaar, A. H., McKune, S. L., Eilitä, M., & Dahl, G. E. (2019). Animal source foods: sustainability problem or malnutrition and sustainability solution? Perspective matters. *Global Food Security*, 1–7. <https://doi.org/10.1016/j.gfs.2019.100325>
- Adeyeye, S. A. O. (2018). Quality evaluation and acceptability of cookies produced from rice (*Oryza glaberrima*) and soybeans (*Glycine max*) flour blends. *Journal of Culinary Science and Technology*, 1–14. <https://doi.org/10.1080/15428052.2018.1502113>
- Adeyeye, S. A. O., Adebayo-Oyetoro, A. O., Fayemi, O. E., Tiamiyu, H. K., Oke, E. K., & Soretire, A. A. (2017). Effect of co-fermentation on nutritional composition, anti-Nutritional factors and acceptability of cookies from fermented sorghum (*Sorghum bicolor*) and soybeans (*Glycine max*) flour blends. *Journal of Culinary Science and Technology*, 17(1), 59–74.

- Ahirwar, S., Soni, H., Rawat, H. K., Prajapati, B. P., & Kango, N. (2016). Experimental design of response surface methodology used for utilisation of palm kernel cake as solid substrate for optimised production of fungal mannanase. *Mycology*, 7(3). <https://doi.org/10.1080/21501203.2016.1229697>
- Ahure, D., & Ejoha, P. O. (2020). Quality evaluation of cookies from malted sorghum (*Sorghum bicolor*), sprouted soybean (*Glycine max*) and carrot (*Daucus carota*) flour blends. *Asian Journal of Biotechnology and Bioresource Technology*, 6(1), 14–27. <https://doi.org/10.9734/ajb2t/2020/v6i130072>
- Ajani, R., Oboh, G., Adefegha, S. A., Nwokocha, K. E., & Akindahunsi, A. A. (2020). Sensory attributes, nutritional qualities, and glycemic indices of bread blends produced from cocoa powder flavored yellow-fleshed cassava-wheat composite flours. *Journal of Food Processing and Preservation*, 44(9), 1–15. <https://doi.org/10.1111/jfpp.14673>
- Akeson, W. R., & Stahmann, M. A. (1964). a pepsin pancreatin digest index of protein quality evaluation. *The Journal of Nutrition*, 83(64), 257–261. <https://doi.org/10.1093/jn/83.3.257>
- Alshelmani, M. I., Loh, T. C., Foo, H. L., Lau, W. H., & Sazili, A. Q. (2014). Biodegradation of palm kernel cake by cellulolytic and hemicellulolytic bacterial cultures through solid state fermentation. *The Scientific World Journal*, 2014, 1–8. <https://doi.org/10.1155/2014/729852>
- Anal, A. K. (2019). Quality ingredients and safety concerns for traditional fermented foods and beverages from Asia: A review. *Fermentation*, 5(1). <https://doi.org/10.3390/fermentation5010008>
- AOAC. (1990). Official Methods of Analysis. Washington, D.C. *Association of Official Analytical Chemist*.
- AOAC. (1995). Official Methods of Analysis. Washington, D.C. *Association of Official Analytical Chemist*.
- Ayele, H. H., Bultosa, G., Abera, T., Astatkie, T., & Yildiz, F. (2017). Nutritional and sensory quality of wheat bread supplemented with cassava and soybean flours. *Cogent Food & Agriculture*, 3(1331892), 1–13.
- Ayivi, R. D., Gyawali, R., Krastanov, A., Aljaloud, S. O., Worku, M., Tahergorabi, R., Silva, R. C. da, & Ibrahim, S. A. (2020). Lactic acid bacteria: food safety and human health applications. *Dairy*, 1(3), 202–232.
- Azam, M., Zhang, S., Qi, J., Abdelghany, A. M., Shaibu, A. S., Ghosh, S., Feng, Y., Huai, Y., Gebregziabher, B. S., Li, J., Li, B., & Sun, J. (2021). Profiling and associations of seed nutritional characteristics in Chinese and USA soybean cultivars. *Journal of Food Composition and Analysis*, 98(September 2020), 1–10. <https://doi.org/10.1016/j.jfca.2021.103803>

- Bakare, A. H., Adeola, A. A., Otesile, I., Obadina, A. O., Afolabi, W. A., Adegunwa, M. O., Akerele, R. A., Bamgbose, O. O., & Alamu, E. O. (2020). Nutritional, texture, and sensory properties of composite biscuits produced from breadfruit and wheat flours enriched with edible fish meal. *Food Science and Nutrition*, 8(11), 6226–6246. <https://doi.org/10.1002/fsn3.1919>
- Bakare, H. A., Osundahunsi, O. F., Adegunwa, M. O., & Olusanya, J. O. (2014). Pasting characteristics, baking and sensory qualities of cakes from blends of cassava and wheat flours. *Journal of Culinary Science and Technology*, 12(2), 109–127. <https://doi.org/10.1080/15428052.2013.846880>
- Barrios-González, J. (2012). Solid-state fermentation: Physiology of solid medium, its molecular basis and applications. *Process Biochemistry*, 47(2), 175–185. <https://doi.org/10.1016/j.procbio.2011.11.016>
- Bartkiene, E., Krungleviciute, V., Juodeikiene, G., Vidmantiene, D., & Maknickiene, Z. (2014). Solid state fermentation with lactic acid bacteria to improve the nutritional quality of lupin and soya bean. *Journal of the Science of Food and Agriculture*, 95(6), 1336–1342. <https://doi.org/10.1002/jsfa.6827>
- Bartkiene, E., Mozuriene, E., Lele, V., Zokaityte, E., Gruzauskas, R., Jakobsone, I., Juodeikiene, G., Ruibys, R., & Bartkevics, V. (2020). Changes of bioactive compounds in barley industry by-products during submerged and solid state fermentation with antimicrobial *Pediococcus acidilactici* strain LUHS29. In *Food Science and Nutrition* (Vol. 8, Issue 1, pp. 340–350). <https://doi.org/10.1002/fsn3.1311>
- Bartkiene, E., Schleining, G., Rekstyte, T., Krungleviciute, V., Juodeikiene, G., Vaiciulyte-Funk, L., & Maknickiene, Z. (2013). Influence of the addition of lupin sourdough with different lactobacilli on dough properties and bread quality. *International Journal of Food Science and Technology*, 48(12), 2613–2620. <https://doi.org/10.1111/ijfs.12257>
- Bartkiene, E., Skabeikyte, E., Juodeikiene, G., Vidmantiene, D., Bašinskiene, L., Maruška, A., Ragažinskiene, O., & Krunglevičiute, V. (2014). The use of solid state fermentation for food and feed plant material processing. *Veterinarija Ir Zootechnika*, 66(88), 3–11.
- Behera, S. S., Ray, R. C., & Zdolec, N. (2018). *Lactobacillus plantarum* with Functional Properties : an approach to increase safety and shelf-Life of Fermented Foods. 2018, 1–18.
- Benayad, A., Taghouti, M., Benali, A., Aboussaleh, Y., & Benbrahim, N. (2021). Nutritional and technological assessment of durum wheat-faba bean enriched flours, and sensory quality of developed composite bread. *Saudi Journal of Biological Sciences*, 28(1), 635–642. <https://doi.org/10.1016/j.sjbs.2020.10.053>

- Bibek, B., Milagros, P. H.-E., & Buddhi, P. L. (2021). Fermentation performance and nutritional assessment of physically processed lentil and green pea flour. *Journal of the Science of Food and Agriculture*, 50(6), 776–780. <https://doi.org/10.1111/pce.14045>
- Bolarinwa, I. F., Abioye, A. O., Adeyanju, J. A., & Kareem, Z. O. (2016). Production and quality evaluation of biscuits produced from malted sorghum-soy flour blends. *Journal of Advances in Food Science & Technology*, 3(3), 107–113.
- Bolarinwa, I. F., & Oyesiji, O. O. (2021). Gluten free rice-soy pasta: proximate composition, textural properties and sensory attributes. *Heliyon*, 7(1), e06052. <https://doi.org/10.1016/j.heliyon.2021.e06052>
- Boye, J., Wijesinha-Bettoni, R., & Burlingame, B. (2012). Protein quality evaluation twenty years after the introduction of the protein digestibility corrected amino acid score method. *British Journal of Nutrition*, 108(SUPPL. 2). <https://doi.org/10.1017/S0007114512002309>
- Bozoglu, F. T., & Erkmen, O. (2016). Food preservation by reducing water activity. *Food microbiology: principles into practice*. *Food Microbiology*: 44–58.
- Bradford, M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72, 248–254.
- Byanju, B., Hojilla-Evangelista, M. P., & Lamsal, B. P. (2021). Fermentation performance and nutritional assessment of physically processed lentil and green pea flour. *Journal of the Science of Food and Agriculture*. <https://doi.org/10.1002/jsfa.11229>
- Cappelli, A., Oliva, N., & Cini, E. (2020). A systematic review of gluten-free dough and bread: Dough rheology, bread characteristics, and improvement strategies. *Applied Sciences (Switzerland)*, 10(18). <https://doi.org/10.3390/APP10186559>
- Carrizo, S. L., de Moreno de LeBlanc, A., LeBlanc, J. G., & Rollán, G. C. (2020). Quinoa pasta fermented with lactic acid bacteria prevents nutritional deficiencies in mice. *Food Research International*, 127, 108735. <https://doi.org/10.1016/j.foodres.2019.108735>
- Castro-Alba, V., Lazarte, C. E., Perez-Rea, D., Carlsson, N. G., Almgren, A., Bergenstähl, B., & Granfeldt, Y. (2019). Fermentation of pseudocereals quinoa, canihua, and amaranth to improve mineral accessibility through degradation of phytate. *Journal of the Science of Food and Agriculture*, 99(11), 5239–5248. <https://doi.org/10.1002/jsfa.9793>

- Cayres, C. A., Ascheri, J. L. R., Couto, M. A. P. G., & Almeida, E. L. (2020). Impact of pregelatinized composite flour on nutritional and functional properties of gluten-free cereal-based cake premixes. *Journal of Food Measurement and Characterization*, 15(1), 769–781.
- Cerda, A., Artola, A., Barrena, R., Font, X., Gea, T., & Sánchez, A. (2019). Innovative production of bioproducts from organic waste through solid-state fermentation. *Frontiers in Sustainable Food Systems*, 3(August), 1–6. <https://doi.org/10.3389/fsufs.2019.00063>
- Chang, H. M., Foo, H. L., Loh, T. C., Lim, E. T. C., & Abdul Mutualib, N. E. (2021). Comparative studies of inhibitory and antioxidant activities, and organic acids compositions of postbiotics produced by probiotic *Lactiplantibacillus plantarum* strains isolated from Malaysian foods. *Frontiers in Veterinary Science*, 7(602280), 1–14. <https://doi.org/10.3389/fvets.2020.602280>
- Chen, H. (2013). Modern Solid state fermentation. *Modern Solid State Fermentation*. <https://doi.org/10.1007/978-94-007-6043-1>
- Chen, L., Vadlani, P. V., Madl, R. L., & Gibbons, W. (2016). Degradation of phytic acid and soy protein in soy meal via co-fermentation of *Aspergillus oryzae* and *Aspergillus ficuum*. *JAOCS, Journal of the American Oil Chemists' Society*, 93(1), 45–50. <https://doi.org/10.1007/s11746-015-2754-9>
- Chhikara, N., Abdulahi, B., Munezero, C., Kaur, R., Singh, G., & Panghal, A. (2019). Exploring the nutritional and phytochemical potential of sorghum in food processing for food security. *Nutrition and Food Science*, 49(2), 318–332. <https://doi.org/10.1108/NFS-05-2018-0149>
- Chis, M. S., Paucean, A., Man, S. M., Bonta, V., Pop, A., Stan, L., Beldean (Tatar), B. V., Pop, C. R., Muresan, V., & Muste, S. (2020). Effect of rice flour fermentation with *Lactobacillus spicheri* DSM 15429 on the nutritional features of gluten-free muffins. *Foods*, 9(822), 1–21.
- Coda, R., Melama, L., Rizzello, C. G., Curiel, J. A., Sibakov, J., Holopainen, U., Pulkkinen, M., & Sozer, N. (2015). Effect of air classification and fermentation by *Lactobacillus plantarum* VTT E-133328 on faba bean (*Vicia faba* L.) flour nutritional properties. *International Journal of Food Microbiology*, 193, 34–42.
- Correa, D., Alejandra, M., Rodríguez de Olmos, A., & Garro, M. S. (2018). Solid state fermentation to obtain vegetable products bio-enriched with isoflavone aglycones using lactic cultures. *Revista Argentina de Microbiología*, 51(3), 201–207. <https://doi.org/10.1016/j.ram.2018.04.006>
- Correa Deza, M. A., Rodríguez de Olmos, A., & Garro, M. S. (2019). Solid state fermentation to obtain vegetable products bio-enriched with isoflavone aglycones using lactic cultures. *Revista Argentina de Microbiología*, 51(3), 201–207. <https://doi.org/10.1016/j.ram.2018.04.006>

- Corsetti, A., Perpetuini, G., Schirone, M., Tofalo, R., & Suzzi, G. (2012). Application of starter cultures to table olive fermentation: An overview on the experimental studies. *Frontiers in Microbiology*, 3, 248.
- Cotârlet, M., Stănciu, N., & Bahrim, G. E. (2020). *Yarrowia lipolytica* and *Lactobacillus paracasei* solid state fermentation as a valuable biotechnological tool for the pork lard and okara's biotransformation. *Microorganisms*, 8(8), 1–12. <https://doi.org/10.3390/microorganisms8081098>
- Croplife International. (2021, July). Beyond the big four – staple crops around the world. <https://croplife.org/news/beyond-the-big-four-staple-crops-around-the-world/>
- Cubas-can, E., & González-fernández, C. (2018). Biotechnological advances in lactic acid production by lactic acid bacteria : lignocellulose as. *Biofuels Bioproducts and Biorefining*, 1–14. <https://doi.org/10.1002/bbb.1852>
- Danbaba, N., Nkama, I., & Badau, M. H. (2019). Use of response surface methodology (RSM) for composite blends of low grade broken rice fractions and full-fat Soybean flour by a twin-screw extrusion cooking process. *International Journal of Food Studies*, 8(1), 14–29. <https://doi.org/10.7455/ijfs/8.1.2019.a2>
- De Pasquale, I., Pontonio, E., Gobbetti, M., & Rizzello, C. G. (2020). Nutritional and functional effects of the lactic acid bacteria fermentation on gelatinized legume flours. *International Journal of Food Microbiology*, 316, 108426. <https://doi.org/10.1016/j.ijfoodmicro.2019.108426>
- Devraj, L., Panoth, A., Kashampur, K., Kumar, A., & Natarajan, V. (2020). Study on physicochemical, phytochemical, and antioxidant properties of selected traditional and white rice varieties. *Journal of Food Process Engineering*, 43(3), 1–13. <https://doi.org/10.1111/jfpe.13330>
- Díaz-Gómez, J., Twyman, R. M., Zhu, C., Farré, G., Serrano, J. C., Portero-Otin, M., Muñoz, P., Sandmann, G., Capell, T., & Christou, P. (2017). Biofortification of crops with nutrients: factors affecting utilization and storage. *Current Opinion in Biotechnology*, 44, 115–123. <https://doi.org/10.1016/j.copbio.2016.12.002>
- Dicko, M. H., Gruppen, H., Traoré, A. S., Voragen, A. G. J., & Van Berkel, W. J. H. (2006). Sorghum grain as human food in Africa: Relevance of content of starch and amylase activities. *African Journal of Biotechnology*, 5(5), 384–395. <https://doi.org/10.5897/AJB>
- Divate, R. D., Wang, C. C., Chou, S. T., Chang, C. T., Wang, P. M., & Chung, Y. C. (2017). Using wheat bran and soybean meal as solid state fermentation substances for the production of *Xylaria nigripes* with bioactivities. *Journal of the Taiwan Institute of Chemical Engineers*, 70, 127–133. <https://doi.org/10.1016/j.jtice.2016.11.003>

- Duguma, H. T., Forsido, S. F., Belachew, T., & Hensel, O. (2021). Changes in anti-nutritional factors and functional properties of extruded composite flour. *Frontiers in Sustainable Food Systems*, 5(November), 1–11. <https://doi.org/10.3389/fsufs.2021.713701>
- Duliński, R., Starzyńska-Janiszewska, A., Byczyński, Ł., & Błaszczyk, U. (2017). Myo-inositol phosphates profile of buckwheat and quinoa seeds: Effects of hydrothermal processing and solid-state fermentation with *Rhizopus oligosporus*. *International Journal of Food Properties*, 20(9), 2088–2095. <https://doi.org/10.1080/10942912.2016.1230871>
- Edet, E. E., Onwuka, G. I., & Orike, C. O. M. (2017). Nutritonal properties of composite flour (blends of rice (*Oryza sativa*), acha (*Digitaria exilis*) and soybeans (*Glycine max*) and sensory properties of noodles produced from the flour. *Asian Journal of Advances in Agricultural Research*, 1(2). <https://doi.org/10.9734/ajaar/2017/34429>
- El-Bakry, M., Abraham, J., Cerda, A., Barrena, R., Ponsá, S., Gea, T., & Sanchez, A. (2015). From wastes to high value added products: Novel aspects of SSF in the production of enzymes. *Critical Reviews in Environmental Science and Technology*, 45(18), 1999–2042. <https://doi.org/10.1080/10643389.2015.1010423>
- EU. (2016). Commission Implementing Regulation (EU) 2017/66 of 14 December 2016 concerning the authorisation of tannic acid as a feed additive for all animal species. *Official Journal of the European Union*, 2017(66), 10–13.
- Evanson Inyang, U. (2018). Physical Properties, Nutritional composition and sensory evaluation of cookies prepared from rice, unripe banana and sprouted soybean flour blends. *International Journal of Food Science and Biotechnology*, 3(2), 70. <https://doi.org/10.11648/j.ijfsb.20180302.15>
- FAO/IFAD/UNICEF/WFP/WHO. (2019). Estimation of forest canopy height over mountainous areas using satellite lidar. In the state of food security and nutrition in the world 2019. Safeguarding against economic slowdowns and downturns. *Rome*, FAO. (Vol. 7, Issue 7). <https://doi.org/10.1109/JSTARS.2014.2300145>
- FAO. (2013). Dietary protein quality evaluation in human nutrition. Report of an FAQ Expert Consultation. In *FAO food and nutrition paper* (Vol. 92).
- FAO. (2018). Protein quality assessment in follow-up formula for young children and ready to use therapeutic foods (Issue November).
- FAO. (2019). Faostat crop production database. Faostat Crop Production Database.
- Fatimah, M. A. (2017). Food policy in malaysian rice products. *Food Science*.

- Fekri, A., Torbati, M., Yari Khosrowshahi, A., Bagherpour Shamloo, H., & Azadmard-Damirchi, S. (2020). Functional effects of phytate-degrading, probiotic lactic acid bacteria and yeast strains isolated from Iranian traditional sourdough on the technological and nutritional properties of whole wheat bread. *Food Chemistry*, 306(2020), 125620. <https://doi.org/10.1016/j.foodchem.2019.125620>
- Fellers, D. A., & Bean, M. M. (1988). Composite flours. *Food Reviews International*, 4(2), 213–235. <https://doi.org/10.1080/87559128809540831>
- Fernandes, P., & Carvalho, F. (2017). Microbial enzymes for the food industry. in biotechnology of microbial enzymes: *Production, Biocatalysis and Industrial Applications*. Elsevier Inc. <https://doi.org/10.1016/B978-0-12-803725-6.00019-4>
- Flourpedia. (2018). Gluten-free flour and starch glossary. <https://www.flourpedia.com/2021/04/gluten-free-flour-and-starch-glossary.html>
- Foo, H. . . , Loh, T. C., Lai, P. W., Lim, Y. Z., Kuflı, C. N., & Gulam, R. (2003). Effects of adding *Lactobacillus plantarum* I-UL4 metabolites in drinking water of rats. *Pakistan Journal of Nutrition*, 2(5), 283–288. <https://doi.org/10.3923/pjn.2003.283.288>
- Gallo, M., Federica, N., Francesca, P., Dana, S., Paola, S., Andrea, B., & Roberto, N. (2019). Lactic fermentation of cereal flour: feasibility tests on rice, oat and wheat. *Applied Food Biotechnology*, 6(3), 165–172. <https://doi.org/10.22037/afb.v%vi%.24299>
- Gallo, M., Passannanti, F., Cante, R. C., Nigro, F., Salameh, D., Schiattarella, P., Schioppa, C., Budelli, A., & Nigro, R. (2020). Effects of the glucose addition during lactic fermentation of rice, oat and wheat flours. *Applied Food Biotechnology*, 7(1), 21–30. <https://doi.org/10.22037/afb.v7i1.26975>
- Gao, Y., Shang, C., Saghai Maroof, M. A., Biyashev, R. M., Grabau, E. A., Kwanyuen, P., Burton, J. W., & Buss, G. R. (2007). A modified colorimetric method for phytic acid analysis in soybean. *Crop Science*, 47(5), 1797–1803. <https://doi.org/10.2135/cropsci2007.03.0122>
- García-cano, I., Rocha-mendoza, D., Ortega-anaya, J., Wang, K., Kosmerl, E., & Jiménez-flores, R. (2019). Lactic acid bacteria isolated from dairy products as potential producers of lipolytic , proteolytic and antibacterial proteins. *Applied Microbiology and Biotechnology*, 103, 5243–5257.
- Ghosh, J. S. (2016). Solid state fermentation and food processing: A short review. *Journal of Nutrition & Food Sciences*, 06(01), 1–7. <https://doi.org/10.4172/2155-9600.1000453>

- Gilani, G. S., Xiao, C. W., & Cockell, K. A. (2012). Impact of antinutritional factors in food proteins on the digestibility of protein and the bioavailability of amino acids and on protein quality. *British Journal of Nutrition*, 108(Suppl. 2). <https://doi.org/10.1017/S0007114512002371>
- Giri, S. S., Sen, S. S., Saha, S., Sukumaran, V., & Park, S. C. (2018). Use of a potential probiotic, *Lactobacillus plantarum* L7, for the preparation of a rice-based fermented beverage. *Frontiers in Microbiology*, 9(473), 1–11. <https://doi.org/10.3389/fmicb.2018.00473>
- Gitau, P. W., Kunyanga, C. N., Abong, G. O., Ojiem, J. O., & Muthomi, J. W. (2019). Assessing sensory characteristics and consumer preference of legume-cereal-root based porridges in Nandi county. *Journal of Food Quality*, 2019. <https://doi.org/10.1155/2019/3035418>
- Govindaraj, M. (2015). Is fortification or bio fortification of staple food crops will offer a simple solution to complex nutritional disorder in developing countries? *Journal of Nutrition & Food Sciences*, 5(2), 1–4. <https://doi.org/10.4172/2155-9600.10003>
- Gunasekaran, Y. K., Lele, V., Sakiene, V., Zavistanaviciute, P., Zokaityte, E., Klupsaite, D., Bartkevics, V., Guiné, R. P. F., & Bartkiene, E. (2020). Plant-based proteinaceous snacks: effect of fermentation and ultrasonication on end-product characteristics. *Food Science and Nutrition*, May, 1–11.
- Hashemi, S. M. B., Gholamhosseinpour, A., & Mousavi Khaneghah, A. (2019). Fermentation of acorn dough by *lactobacilli* strains: Phytic acid degradation and antioxidant activity. *Lebensmittel-wissenschaft & Technologie*, 100, 144–149. <https://doi.org/10.1016/j.lwt.2018.10.054>
- Hasmadi, M., Noorfarahzilah, M., Lee, J. S., Sharifudin, M. S., & Mohd Fadzelly, A. B. (2014). Applications of composite flour in development of food products. *International Food Research Journal*, 21(6), 2061–2074.
- Hasmadi, M., Noorfarahzilah, M., Noraidah, H., Zainol, M. K., & Jahurul, M. H. A. (2020). Functional properties of composite flour: A review. *Food Research*, 4(6), 1820–1831. [https://doi.org/10.26656/fr.2017.4\(6\).419](https://doi.org/10.26656/fr.2017.4(6).419)
- Hatti-kaul, R., Chen, L., Dishisha, T., & Enshasy, H. El. (2018). Lactic acid bacteria: from starter cultures to producers of chemicals. *FEMS microbiology letters*. July, 1–20. <https://doi.org/10.1093/femsle/fny213>
- Hotz, C., & McClafferty, B. (2007). From harvest to health: Challenges for developing biofortified staple foods and determining their impact on micronutrient status. *Food and Nutrition Bulletin*, 28(2 SUPPL.), 271–279.
- Hu, S., Zhu, Q., Ren, A., Ge, L., He, J., Zhao, M., & He, Q. (2022). Roles of water in improved production of mycelial biomass and lignocellulose-degrading enzymes by water-supply solid-state fermentation of *Ganoderma lucidum*. *Journal of Bioscience and Bioengineering*, 133(2), 126–132.

- Hussin, A. S. M., Farouk, A. E. A., Greiner, R., Salleh, H. M., & Ismail, A. F. (2007). Phytate-degrading enzyme production by bacteria isolated from Malaysian soil. *World Journal of Microbiology and Biotechnology*, 23(12), 1653–1660. <https://doi.org/10.1007/s11274-007-9412-9>
- Izuddin, W. I., Humam, A. M., Loh, T. C., Foo, H. L., & Samsudin, A. A. (2020). Dietary postbiotic *Lactobacillus plantarum* improves serum and ruminal antioxidant activity and upregulates hepatic antioxidant enzymes and ruminal barrier function in post-weaning lambs. *Antioxidants*, 9(3). <https://doi.org/10.3390/antiox9030250>
- J. De Vries-Ten Have, J., Owolabi, A., Steijns, J., Kudla, U., & Melse-Boonstra, A. (2020). Protein intake adequacy among Nigerian infants, children, adolescents and women and protein quality of commonly consumed foods. *Nutrition Research Reviews*, 33(1), 102–120. <https://doi.org/10.1017/S0954422419000222>
- Jiang, H., Hettiarachchy, N. S., & Horax, R. (2018). Physical properties and estimated glycemic index of protein-enriched sorghum based chips. *Journal of Food Science and Technology*, 55(3), 891–898. <https://doi.org/10.1007/s13197-017-2993-x>
- Jiménez, N., Esteban-Torres, M., Mancheño, J. M., De las Rivas, B., & Muñoz, R. (2014). Tannin degradation by a novel tannase enzyme present in some *Lactobacillus plantarum* strains. *Applied and Environmental Microbiology*, 80(10), 2991–2997. <https://doi.org/10.1128/AEM.00324-14>
- Johansen, P. G., Owusu-Kwarteng, J., Parkouda, C., Padonou, S. W., & Jespersen, L. (2019). Occurrence and importance of yeasts in indigenous fermented food and beverages produced in Sub-saharan Africa. *Frontiers in Microbiology*, 10(August). <https://doi.org/10.3389/fmicb.2019.01789>
- Joye, I. (2019). Protein digestibility of cereal products. *Foods*, 8(199), 1–14. <https://doi.org/10.3390/foods8060199>
- Junior, M. V. J., Teixeira, C. B., & Macedo, G. A. (2013). Biotransformation and bioconversion of phenolic compounds obtainment: an overview. *Critical review in biotechnology*, 8551, 1–7. <https://doi.org/10.3109/07388551.2013.803020>
- Kahlert, H., Meyer, G., & Albrecht, A. (2016). Colour maps of acid–base titrations with colour indicators: how to choose the appropriate indicator and how to estimate the systematic titration errors. *ChemTexts*, 2(2), 1–28. <https://doi.org/10.1007/s40828-016-0026-4>
- Kanagarajah, P. (2018). Quality evaluation of biscuits prepared from the composite flour of sprouted sorghum, soybean and finger millet. *Journal of Food Processing & Technology*, 09. <https://doi.org/10.4172/2157-7110-c10-104>

- Kårlund, A., Carlos, G., Korhonen, J., Palo-oja, O.-M., El-nezami, H., & Kolehmainen, M. (2020). Harnessing microbes for sustainable development: food fermentation as a tool for improving the nutritional quality of alternative protein sources. *Nutrients*, 12, 1–26.
- Kavitake, D., Kandasamy, S., Devi, P. B., & Shetty, P. H. (2018). Recent developments on encapsulation of lactic acid bacteria as potential starter culture in fermented foods – A review. *Food Bioscience*, 21(November 2017), 34–44. <https://doi.org/10.1016/j.fbio.2017.11.003>
- Kechagia, M., Basoulis, D., Konstantopoulou, S., Dimitriadi, D., Gyftopoulou, K., Skarmoutsou, N., & Fakiri, E. M. (2013). Health benefit of probiotic: A review. *Hindawi Publishing Corporation*, 2013, 1–7.
- Kwaw, E., Sackey, A. S., Apaliya, M. T., & Tchabo, W. (2017). Utilization of composite flours as breading agents for deep frying of chicken breast. *Journal of Food Measurement and Characterization*, 11(3), 1523–1530. <https://doi.org/10.1007/s11694-017-9531-4>
- Le, B., Thi, P., Anh, N., Jinhua, J. K., Seung, C., & Yang, H. (2019). Rice bran fermentation by lactic acid bacteria to enhance antioxidant activities and increase the ferulic acid,  $\rho$ -coumaric acid, and  $\gamma$ -oryzanol content. *Food science* 62, 257–264.
- Lee, F. H., Wan, S. Y., Foo, H. L., Loh, T. C., Mohamad, R., Abdul Rahim, R., & Idrus, Z. (2019). Comparative study of extracellular proteolytic, cellulolytic, and hemicellulolytic enzyme activities and biotransformation of palm kernel cake biomass by lactic acid bacteria isolated from Malaysian foods. *International Journal of Molecular Sciences*, 20(4979), 1–26.
- Li, S., Jin, Z., Hu, D., Yang, W., Yan, Y., Nie, X., Lin, J., Zhang, Q., Gai, D., Ji, Y., & Chen, X. (2020). Effect of solid-state fermentation with *Lactobacillus casei* on the nutritional value, isoflavones, phenolic acids and antioxidant activity of whole soybean flour. *Lebensmittel-wissenschaft & Technologie*, 125, 1–8. <https://doi.org/10.1016/j.lwt.2020.109264>
- Li, Ying, Yin, Z., Zhang, Y., Liu, J., Cheng, Y., Wang, J., Pi, F., Zhang, Y., & Sun, X. (2020). Perspective of microbe-based minerals fortification in nutrition security. *Food Reviews International*,. Pp 1–14.
- Li, Yongfu, Cheng, X., Shi, F., Wang, L., Li, Y., & Chen, Z. (2019). Effect of solid-state fermentation by *Lactobacillus plantarum* on the cooking quality, microstructure, and physicochemical properties of brown rice. *Starch/Staerke*, 71, 1–34. <https://doi.org/10.1002/star.201800160>
- Lim, Y. H., Foo, H. L., Loh, T. C., Mohamad, R., & Abdullah, N. (2019). Comparative studies of versatile extracellular proteolytic activities of lactic acid bacteria and their potential for extracellular amino acid productions as feed supplements. *Journal of Animal Science and Biotechnology*, 10(15), 1–13. <https://doi.org/10.1186/s40104-019-0323-z>

- Ly, D., Mayrhofer, S., & Domig, K. J. (2018). Significance of traditional fermented foods in the lower Mekong subregion: A focus on lactic acid bacteria. *Food Bioscience*, 26, 113–125. <https://doi.org/10.1016/j.fbio.2018.10.004>
- Ma, Z. Q., Yi, C. P., Wu, N. N., & Tan, B. (2020). Reduction of phenolic profiles, dietary fiber, and antioxidant activities of rice after treatment with different milling processes. *Cereal Chemistry*, 97(6), 1158–1171. <https://doi.org/10.1002/cche.10336>
- Mahadev, S., & Green, P. H. R. (2011). Celiac disease: A challenge for all physicians. *Gastroenterology and Hepatology*, 7(8), 554–556.
- Maka Taga, C., Jiokap Nono, Y., Icard-Vernière, C., Desmorieux, H., Kapseu, C., & Mouquet-Rivier, C. (2019). Formulation and processing of gruels made from local ingredients, thin enough to flow by gravity in enteral tube feeding. *Journal of Food Science and Technology*, 56(8), 3609–3619. <https://doi.org/10.1007/s13197-019-03787-6>
- Mansor, A., Ramli, M. S., Abdul Rashid, N. Y., Sharifudin, S. A., & Raseetha, S. (2019). Evaluation of selected agri-industrial residues as potential substrates for enhanced tannase production via solid-state fermentation. *Biocatalysis and Agricultural Biotechnology*, 20, 101–216.
- Mao, M., Wang, P., Shi, K., Lu, Z., Bie, X., Zhao, H., Zhang, C., & Lv, F. (2020). Effect of solid state fermentation by *Enterococcus faecalis* M2 on antioxidant and nutritional properties of wheat bran. *Journal of Cereal Science*, 94(5), 1–7. <https://doi.org/10.1016/j.jcs.2020.102997>
- Mayasti, N. K. I., Ushada, M., & Ainuri, M. (2019). Improvement of spaghetti composite quality based on local flours (mocaf, corn, rice and soybean). *AIP Conference Proceedings*, 2175(November).
- Melini, F., Melini, V., Luziatelli, F., & Ruzzi, M. (2017). Current and forward-looking approaches to technological and nutritional improvements of gluten-free bread with legume flours: a critical review. *comprehensive reviews in food science and food safety*, 16(5), 1101–1122.
- Melini, V., Melini, F., & Acquistucci, R. (2020). Phenolic compounds and bioaccessibility thereof in functional pasta. *Antioxidants*, 9(4), 1–30. <https://doi.org/10.3390/antiox9040343>
- Menon, L., Majumdar, S. D., & Ravi, U. (2015). Development and analysis of composite flour bread. *Journal of Food Science and Technology*, 52(7), 4156–4165. <https://doi.org/10.1007/s13197-014-1466-8>
- Merzza, H., Loh, T., Foo, H. L., Samsudin, A. A., Noordin, M. M., Zulkifli, I., & Wan Ibrahim, I. (2019). Effects of feeding different postbiotics produced by *Lactobacillus plantarum* on growth performance, carcass yield, intestinal morphology, gut microbiota composition, immune status, and growth gene expression in broilers under heat stress. *Animals*, 9, 1–20.

- Mette, L., & Peter, S. L. (2019). Application of lactic acid bacteria in green biorefineries. *FEMS Microbiology Letters*, 366, 1–8.
- Mir, S. A., Shah, M. A., Bosco, S. J. D., Sunoj, K. V., & Farooq, S. (2020). A review on nutritional properties, shelf life, health aspects, and consumption of brown rice in comparison with white rice. *Cereal Chemistry*, 97(5), 895–903. <https://doi.org/10.1002/cche.10322>
- Moghadam, M. S., Foo, H. L., Leow, T. C., Rahim, R. A., & Loh, T. C. (2010). Novel bacteriocinogenic *Lactobacillus plantarum* strains and their differentiation by sequence analysis of 16S rDNA, 16S-23S and 23S-5S intergenic spacer regions and randomly amplified polymorphic DNA analysis. *Food Technology and Biotechnology*, 48(4), 476–483.
- Mohammadi-Kouchesfahani, M., Hamidi-Esfahani, Z., & Azizi, M. H. (2019). Isolation and identification of lactic acid bacteria with phytase activity from sourdough. *Food Science and Nutrition*, 7(11), 3700–3708.
- Mohapatra, D., Patel, A. S., Kar, A., Deshpande, S. S., & Tripathi, M. K. (2019). Effect of different processing conditions on proximate composition, anti-oxidants, anti-nutrients and amino acid profile of grain sorghum. *Food Chemistry*, 1–29. <https://doi.org/10.1016/j.foodchem.2018.07.196>
- Mugalavai, V. K., Aduol, K. O., & Onkware, A. O. (2021). Nutritional characteristics of rice (*Oryza sativa*) composite flours obtained by food fortification. *European Journal of Agriculture and Food Sciences*, 3(1), 79–83. <https://doi.org/10.24018/ejfood.2021.3.1.224>
- Muhamad Nor, N., Mohamad, R., Foo, H. L., & Rahim, R. A. (2010). Improvement of folate biosynthesis by lactic acid bacteria using response surface methodology. *Food Technology and Biotechnology*, 48(2), 243–250.
- Muninggar DHR, & Damanik, M. R. M. (2019). Formulation of instant surabi from composite rice-soybean flours and supplemented with torbangun powder as an alternative snack for ADHD children. *Annals of Nutrition & Metabolism*, 75(3), 152-.
- Nazri, N. S., Vanoh, D., & Leng, S. K. (2020). Malnutrition, low diet quality and its risk factors among older adults with low socio- economic status: a scoping review. *The Nutrition Society*.
- NDSU. (2020). Wheat quality & carbohydrate research. <https://www.ndsu.edu/faculty/simsek/wheat/flour.html>
- Nieto-Mazzocco, E., Saldaña-Robles, A., Franco-Robles, E., Rangel-Contreras, A. K., Cerón-García, A., & Ozuna, C. (2020). Optimization of sorghum, rice, and amaranth flour levels in the development of gluten-free bakery products using response surface methodology. *Journal of Food Processing and Preservation*, 44(1), 1–9.

- Nieto, E., Rangel, A., Saldaña, A., Juárez, M., & Ozuna, C. (2018). Caracterización de harinas libres de gluten y su incorporación en productos de panificación. *Investigación y Desarrollo En Ciencia y Tecnología de Alimentos*, 3.
- Nisa, K., Rosyida, V. T., Nurhayati, S., Indrianingsih, A. W., Darsih, C., & Apriyana, W. (2019). Total phenolic contents and antioxidant activity of rice bran fermented with lactic acid bacteria. *IOP Conference Series: Earth and Environmental Science*, 251(1). <https://doi.org/10.1088/1755-1315/251/1/012020>
- Nkhata, S. G., Ayua, E., Kamau, E. H., & Shingiro, J. B. (2018a). Fermentation and germination improve nutritional value of cereals and legumes through activation of endogenous enzymes. *Food Science and Nutrition*, 6, 2446–2458. <https://doi.org/10.1002/fsn3.846>
- Nkhata, S. G., Ayua, E., Kamau, E. H., & Shingiro, J. B. (2018b). Fermentation and germination improve nutritional value of cereals and legumes through activation of endogenous enzymes. *Food Science and Nutrition*, 6(8), 2446–2458. <https://doi.org/10.1002/fsn3.846>
- Noorfarahzilah, M., Hasmadi, M., Lee, J. S., Sharifudin, M. S., & Mohd Fadzelly, A. B. (2014). Applications of composite flour in development of food products. *International Food Research Journal*, 21(6), 2061–2074. <https://doi.org/10.15740/has/ijae/11.sp.issue/65-69>
- Noorfarahzilah, M., Lee, J. S., Sharifudin, M. S., Mohd Fadzelly, A. B., & Hasmadi, M. (2014). Applications of composite flour in development of food products. *International Food Research Journal*, 21(6), 2061–2074. <https://doi.org/10.15740/has/ijae/11.sp.issue/65-69>
- Nusrat, Iqbal Amrish Agrawal, S. D. and J. K. (2020). Role of decomposers in agricultural waste management. *Intech Open*.
- Nwanekezi, E. C. (2013). Composite flours for baked products and possible challenges – a review. *Nigerian Food Journal*, 31(2), 8–17.
- Ogodo, A. C., Chinyere Ugbogu, O., Onyeagba, R. A., & Okereke, H. C. (2018). Proximate composition and in-vitro starch/protein digestibility of bambara groundnut flour fermented with lactic acid bacteria (LAB)-consortium isolated from cereals. *Fermentation Technology*, 07(01), 1–9. <https://doi.org/10.4172/2167-7972.1000148>
- Ogodo, A. C., Ugbogu, O. C., Onyeagba, R. A., & Okereke, H. C. (2018). In-vitro starch and protein digestibility and proximate composition of soybean flour fermented with lactic acid bacteria (LAB) consortia. *Agriculture and Natural Resources*, 52(2018), 503–509. <https://doi.org/10.1016/j.anres.2018.10.001>

- Ogodo, A. C., Ugbogu, O. C., Onyeagba, R. A., & Okereke, H. C. (2019). Microbiological quality, proximate composition and in vitro starch/protein digestibility of *Sorghum bicolor* flour fermented with lactic acid bacteria consortia. *Chemical and Biological Technologies in Agriculture*, 6(7), 1–9. <https://doi.org/10.1186/s40538-019-0145-4>
- Ohanenye, I. C., Emenike, C. U., Mensi, A., Medina-Godoy, S., Jin, J., Ahmed, T., Sun, X., & Udenigwe, C. C. (2021). Food fortification technologies: Influence on iron, zinc and vitamin A bioavailability and potential implications on micronutrient deficiency in sub-Saharan Africa. *Scientific African*, 11, e00667. <https://doi.org/10.1016/j.sciaf.2020.e00667>
- Ojha, B. K., Singh, P. K., & Shrivastava, N. (2018). Enzymes in the animal feed industry. In *Enzymes in Food Biotechnology: Production, Applications, and Future Prospects* (pp. 93–109). <https://doi.org/10.1016/B978-0-12-813280-7.00007-4>
- Ojha, P., Adhikari, R., Karki, R., Mishra, A., Subedi, U., & Karki, T. B. (2018). Malting and fermentation effects on antinutritional components and functional characteristics of sorghum flour. *Food Science and Nutrition*, 6(1), 47–53. <https://doi.org/10.1002/fsn3.525>
- Olojede, A. O., Sanni, A. I., & Banwo, K. (2020). Rheological, textural and nutritional properties of gluten-free sourdough made with functionally important lactic acid bacteria and yeast from Nigerian sorghum. *Lebensmittel-wissenschaft & Technologie*, 120. <https://doi.org/10.1016/j.lwt.2019.108875>
- Olukomaiya, O. O., Adiamo, O. Q., Fernando, W. C., Mereddy, R., Li, X., & Sultanbawa, Y. (2020). Effect of solid-state fermentation on proximate composition, anti-nutritional factor, microbiological and functional properties of lupin flour. *Food Chemistry*, 315, 126238. <https://doi.org/10.1016/j.foodchem.2020.126238>
- Omwamba, M., & Mahungu, S. M. (2014). Development of a protein-rich ready-to-eat extruded snack from a composite blend of rice, sorghum and soybean flour. *Food and Nutrition Sciences*, 05(14), 1309–1317. <https://doi.org/10.4236/fns.2014.514142>
- Osman, M. A. (2011). Effect of traditional fermentation process on the nutrient and antinutrient contents of pearl millet during preparation of Lohoh. *Journal of the Saudi Society of Agricultural Sciences*, 10(1), 1–6. <https://doi.org/10.1016/j.jssas.2010.06.001>
- Park, J., Sung, J. M., Choi, Y. S., & Park, J. D. (2020). Effect of natural fermentation on milled rice grains: Physicochemical and functional properties of rice flour. *Food Hydrocolloids*, 108(2020), 1–10. <https://doi.org/10.1016/j.foodhyd.2020.106005>

- Pérez-Ramos, A., Mohedano, M. L., López, P., Spano, G., Fiocco, D., Russo, P., & Capozzi, V. (2017). In situ  $\beta$ -glucan fortification of cereal-based matrices by *Pediococcus parvulus* 2.6: technological aspects and prebiotic potential. *International Journal of Molecular Sciences*, 18(7). <https://doi.org/10.3390/ijms18071588>
- Petrova, P., & Petrov, K. (2020). Lactic acid fermentation of cereals and pseudocereals: Ancient nutritional biotechnologies with modern applications. *Nutrients*, 12(4), 1–26. <https://doi.org/10.3390/nu12041118>
- Pranoto, Y., Anggraiani, S., & Efendi, Z. (2013). Effect of natural and *Lactobacillus plantarum* fermentation on *in-vitro* protein and starch digestibilities of sorghum flour. *Food Bioscience*, 2(2013), 46–52. <https://doi.org/10.1016/j.fbio.2013.04.001>
- Ramesh, M. V., & Lonsane, B. K. (1990). Critical importance of moisture content of the medium in alpha-amylase production by *Bacillus licheniformis* M27 in a solid-state fermentation system. *Applied Microbiology and Biotechnology*, 33(5), 501–505. <https://doi.org/10.1007/BF00172541>
- Rani, A., Kumar, V., Shukla, S., Jha, P., Tayalkar, T., & Mittal, P. (2020). Changes in storage protein composition on genetic removal of Kunitz trypsin inhibitor maintain protein content in soybean (*Glycine max*). *Journal of Agriculture and Food Research*, 2(February), 100065. <https://doi.org/10.1016/j.jafr.2020.100065>
- Rawal, R., Kumar, V., Rani, A., & Gokhale, S. M. (2020). Genetic elimination of off-flavour generating lipoxygenase-2 gene of soybean through marker assisted backcrossing and its effect on seed longevity. *Plant Breeding and Biotechnology*, 8(2), 163–173. <https://doi.org/10.9787/PBB.2020.8.2.163>
- Reale, A., Konietzny, U., Coppola, R., Sorrentino, E., & Greiner, R. (2007). The importance of lactic acid bacteria for phytate degradation during cereal dough fermentation. *Journal of Agricultural and Food Chemistry*, 55(8), 2993–2997. <https://doi.org/10.1021/jf063507n>
- Rizzo, G., & Baroni, L. (2018). Soy, soy foods and their role in vegetarian diets. In *Nutrients* (Vol. 10, Issue 1). <https://doi.org/10.3390/nu10010043>
- Rodríguez de Olmos, A., Bru, E., & Garro, M. S. (2014). Optimization of fermentation parameters to study the behavior of selected lactic cultures on soy solid state fermentation. *International Journal of Food Microbiology*, 196, 16–23. <https://doi.org/10.1016/j.ijfoodmicro.2014.11.030>
- Rodríguez de Olmos, Antonieta, & Garro, M. S. (2020). Metabolic profile of *Lactobacillus paracasei* subsp. *paracasei* CRL 207 in solid state fermentation using commercial soybean meal. *Food Bioscience*, 35. <https://doi.org/10.1016/j.fbio.2020.100584>

- Rodríguez de Olmos, Antonieta, Garro, M. S., & Correa Deza, M. A. (2017). Selected *lactobacilli* and *bifidobacteria* development in solid state fermentation using soybean paste. *Revista Argentina de Microbiología*, 49(1), 62–69. <https://doi.org/10.1016/j.ram.2016.08.007>
- Rollán, G. C., Gerez, C. L., & Leblanc, J. G. (2019). Lactic fermentation as a strategy to improve the nutritional and functional values of pseudocereals. *Frontiers in nutrition*, 6(98), 1–16. <https://doi.org/10.3389/fnut.2019.00098>
- Rui, X., Wang, M., Zhang, Y., Chen, X., Li, L., Liu, Y., & Dong, M. (2017). Optimization of soy solid-state fermentation with selected lactic acid bacteria and the effect on the anti-nutritional components. *Journal of Food Processing and Preservation*, 1–7. <https://doi.org/10.1111/jfpp.13290>
- Sadh, P. K., Kumar, S., Chawla, P., & Duhan, J. S. (2018). Fermentation: a boon for production of bioactive compounds by processing of food industries wastes (By-Products). *Molecules* (Vol. 23, Issue 10). <https://doi.org/10.3390/molecules23102560>
- Saleh, A. S. M., Wang, P., Wang, N., Yang, L., & Xiao, Z. (2019). Brown rice versus white rice: nutritional quality, potential health benefits, development of food products, and preservation technologies. *Comprehensive Reviews in Food Science and Food Safety*. <https://doi.org/10.1111/1541-4337.12449>
- Sawangwan, T., Porncharoennop, C., & Nimraksa, H. (2021). Antioxidant compounds from rice bran fermentation by lactic acid bacteria. *AIMS Agriculture and Food*, 6(2), 578–587. <https://doi.org/10.3934/AGRFOOD.2021034>
- Saxena, R. K., Saran, S., Isar, J., & Kaushik, R. (2016). Production and Applications of succinic acid. *current developments in biotechnology and Bioengineering: Production, Isolation and Purification of Industrial Products. Applied Microbiology*. 601–630. <https://doi.org/10.1016/B978-0-444-63662-1.00027-0>
- Schmidt, C. G., & Furlong, E. B. (2012). Effect of particle size and ammonium sulfate concentration on rice bran fermentation with the fungus *Rhizopus oryzae*. *Bioresource Technology*, 123, 36–41.
- Schönfeldt, H. C., & Hall, N. G. (2012). Dietary protein quality and malnutrition in Africa. *British Journal of Nutrition*, 108(Suppl. 2). <https://doi.org/10.1017/S0007114512002553>
- Seth, D., & Rajamanickam, G. (2012). Original article development of extruded snacks using soy , sorghum , millet and rice blend – a response surface methodology approach. *International Journal of Food Science and Technology*, 47, 1526–1531. <https://doi.org/10.1111/j.1365-2621.2012.03001.x>

- Sharma, S., Kaur, M., Goyal, R., & Gill, B. S. (2014). Physical characteristics and nutritional composition of some new soybean (*Glycine max* (L.) Merrill) genotypes. *Journal of Food Science and Technology*, 51(3), 551–557. <https://doi.org/10.1007/s13197-011-0517-7>
- Singh Nee Nigam, P., & Pandey, A. (2009). Biotechnology for agro-industrial residues utilisation: utilisation of agro-residues. *Biotechnology for Agro-Industrial Residues Utilisation* (Issue January 2009). <https://doi.org/10.1007/978-1-4020-9942-7>
- Singh, U., Praharaj, C. S., Singh, S. S., & Singh, N. P. (2016). Biofortification of food crops. *Food Crops* (pp. 3–18). <https://doi.org/10.1007/978-81-322-2716-8>
- Soccol, C. R., Costa, E. S. F. da, Letti, L. A. J., Karp, S. G., Woiciechowski, A. L., & Vandenberghe, L. P. de S. (2017). Recent developments and innovations in solid state fermentation. *Biotechnology Research and Innovation*, 1(1), 52–71. <https://doi.org/10.1016/j.biori.2017.01.002>
- Songlin, L., Jin, Z., Hu, D., Yang, W., Yan, Y., Nie, X., Lin, J., Zhang, Q., Gai, D., Ji, Y., & Chen, X. (2020). Effect of solid-state fermentation with *Lactobacillus casei* on the nutritional value, isoflavones, phenolic acids and antioxidant activity of whole soybean flour. *Lebensmittel-wissenschaft & Technologie*, 125, 109264. <https://doi.org/10.1016/j.lwt.2020.109264>
- Sugathan, S., Pradeep, N. S., & Abdulhameed, S. (2017). Bioresources and bioprocess in biotechnology. *Bioresources and Bioprocess in Biotechnology*, 2, 127–150. <https://doi.org/10.1007/978-981-10-4284-3>
- Tays, C., Guarnieri, M. T., Sauvageau, D., & Stein, L. Y. (2018). Combined effects of carbon and nitrogen source to optimize growth of proteobacterial methanotrophs. *Frontiers in Microbiology*, 9(SEP), 1–14. <https://doi.org/10.3389/fmicb.2018.02239>
- Temba, M. C., Njobeh, P. B., Adebo, O. A., Olugbile, A. O., & Kayitesi, E. (2016). The role of compositing cereals with legumes to alleviate protein energy malnutrition in Africa. *International Journal of Food Science and Technology*, 51, 543–554. <https://doi.org/10.1111/ijfs.13035>
- Thomas, L., Larroche, C., & Pandey, A. (2013). Current developments in solid-state fermentation. *Biochemical Engineering Journal*, 81, 146–161.
- Thung, T. Y. (2012). Isolation and purification of proteolytic enzyme produced by lactic acid bacteria from Budu and Bambangan. *Master's Thesis, Universiti Putra Malaysia, Selangor, Malaysia*.
- Toe, C. J., Foo, H. L., Loh, T. C., Mohamad, R., Rahim, R. A., & Idrus, Z. (2019). Extracellular proteolytic activity and amino acid production by lactic acid bacteria isolated from Malaysian foods. *International Journal of Molecular Sciences*, 20(1777), 1–22. <https://doi.org/10.3390/ijms20071777>

- Try, S., Voilley, A., Chunhieng, T., De-Coninck, J., & Waché, Y. (2018). Aroma compounds production by solid state fermentation, importance of in situ gas-phase recovery systems. *Applied Microbiology and Biotechnology*, 102(17), 7239–7255. <https://doi.org/10.1007/s00253-018-9157-4>
- Tsao, R. (2019). Technologies for improving the nutritional quality of cereals. *Bioactive Factors and Processing Technology for Cereal Foods*, 19–31. [https://doi.org/10.1007/978-981-13-6167-8\\_2](https://doi.org/10.1007/978-981-13-6167-8_2)
- Twinomuhwezi, H., Awuchi, C. G., & Rachael, M. (2020). Comparative study of the proximate composition and functional properties of composite flours. *American Journal of Food Science and Nutrition*, 6(1), 6–19.
- Udomkun, P., Tirawattanawanich, C., Ilukor, J., Sridonpai, P., Njukwe, E., Nimbona, P., & Vanlauwe, B. (2019). Promoting the use of locally produced crops in making cereal-legume-based composite flours: an assessment of nutrient, antinutrient, mineral molar ratios, and aflatoxin content. *Food Chemistry*, 286(October 2018), 651–658.
- Vafopoulou-Mastrojiannaki, A., & Litopoulou-Tzanetaki, E Tzanetakis, N. (1994). Proteinase, peptidase and esterase activity of crude cell-free extracts of *Pediococcus pentosaceus* isolated from cheese. *Lebensmittelwissenschaft & Technologie*, 27(4), 342–346.
- Venkatasubbaiah, R., & Rajesh, S. K. (2020). Food processing and fermentation studies on reduction of phytic acid in *Triticum aestivum* and *Sorghum bicolor* (L.). *Journal of Microbiology, Biotechnology and Food Sciences*, 10(2), 166–169. <https://doi.org/10.15414/jmbfs.2020.10.2.166-169>
- Verni, M., Rizzello, C. G., Coda, R., & Bran, W. (2019). Fermentation biotechnology applied to cereal industry by-products : *Nutritional and Functional Insights*. 6(April), 1–13.
- Vinicio De Melo Pereira, G., De Carvalho Neto, D. P., Junqueira, A. C. D. O., Karp, S. G., Letti, L. A. J., Magalhães Júnior, A. I., & Soccol, C. R. (2020). A review of selection criteria for starter culture development in the food fermentation industry. *Food Reviews International*, 36(2), 135–167.
- Wanjala, W. N., Mary, O., & Symon, M. (2020). Optimization of protein content and dietary fibre in a composite flour blend containing rice (*Oryza sativa*), sorghum (*Sorghum bicolor* (L.) Moench] and bamboo (*Yushania alpine*) shoots. *Food and Nutrition Sciences*, 11(08), 789–806. <https://doi.org/10.4236/fns.2020.118056>
- Watanabe, M., Techapun, C., Kuntiya, A., Leksawasdi, N., Seesuriyachan, P., Chaiyaso, T., Takenaka, S., Maeda, I., Koyama, M., & Nakamura, K. (2017). Extracellular protease derived from lactic acid bacteria stimulates the fermentative lactic acid production from the by-products of rice as a biomass refinery function. *Journal of Bioscience and Bioengineering*, 123(2), 245–251. <https://doi.org/10.1016/j.jbiosc.2016.08.011>

- Webb, C., & Manan, M. A. (2017). Design aspects of solid state fermentation as applied to microbial bioprocessing. *Journal of Applied Biotechnology & Bioengineering*, 4(1). <https://doi.org/10.15406/jabb.2017.04.00094>
- Wenche, F., Heddle, M., & Inge, T. (2011). Phytate- a natural component in plant food. *National Food Institute*, 1–2. <https://fuldkorn.dk/wp-content/uploads/2019/08/Phytate-a-natural-component-in-plant-food-1.pdf>
- Wulandari, E., Sukarminah, E., Lanti, I., & Sufmawati, I. (2017). Organoleptic characteristics of cookies from sorghum composites flour. *KnE Life Sciences*, 2(6), 506. <https://doi.org/10.18502/kls.v2i6.1071>
- Xing, Q., Dekker, S., Kyriakopoulou, K., Boom, R. M., Smid, E. J., & Schutyser, M. A. I. (2019). Enhanced nutritional value of chickpea protein concentrate by dry separation and solid state fermentation. *Innovative Food Science and Emerging Technologies*, 59, 102269. <https://doi.org/10.1016/j.ifset.2019.102269>
- Xiong, Y., Zhang, P., Warner, R. D., & Fang, Z. (2019). Sorghum grain: from genotype, nutrition, and phenolic profile to its health benefits and food applications. *Comprehensive Reviews in Food Science and Food Safety* (Vol. 18, Issue 6, pp. 2025–2046). <https://doi.org/10.1111/1541-4337.12506>
- Xu, Y., Zhou, X., Zhang, M., Zhang, Z., Song, L., Wang, G., & Zhang, J. (2020). Metabolism analysis for enhanced nutritional profile of chestnuts subjected to anaerobic solid-state fermentation by probiotic lactic acid bacteria. *Journal of Food Processing and Preservation*, 44(3), 1–11.
- Yang, A., Smyth, H., Chaliha, M., & James, A. (2016). Sensory quality of soymilk and tofu from soybeans lacking lipoxygenases. *Food Science and Nutrition*, 4(2), 207–215. <https://doi.org/10.1002/fsn3.274>
- Zabidi, N. A. M., Foo, H. L., Loh, T. C., Mohamad, R., & Rahim, R. A. (2020). Enhancement of versatile extracellular cellulolytic and hemicellulolytic enzyme productions. 25(2607): 1-32 *Molecules*.
- Zayas, J. . (1997). Solubility of proteins. in: functionality of proteins in food (pp. 6–7). Springer Berlin Heidelberg. [https://doi.org/https://doi.org/10.1007/978-3-642-59116-7\\_2](https://doi.org/https://doi.org/10.1007/978-3-642-59116-7_2)
- Zhang, H., & Yu, H. (2019). Enhanced biotransformation of soybean isoflavone from glycosides to aglycones using solid-state fermentation of soybean with effective microorganisms (EM) strains. *Journal of Food Biochemistry*, 43(4), 1–10. <https://doi.org/10.1111/jfbc.12804>
- Zhang, N., Li, D., Zhang, X., Shi, Y., & Wang, H. (2015). Solid-state fermentation of whole oats to yield a synbiotic food rich in lactic acid bacteria and prebiotics. *Food and Function*, 6(8), 2620–2625.

Zhang, S., Shi, Y., Zhang, S. L., Shang, W., Gao, X. Q., & Wang, H. K. (2014). Whole soybean as probiotic lactic acid bacteria carrier food in solid-state fermentation. *Food Control*, 41(1), 1–6. <https://doi.org/10.1016/j.foodcont.2013.12.026>

Zhang, S. T., Shi, Y., Zhang, S. L., Shang, W., Gao, X. Q., & Wang, H. K. (2014). Whole soybean as probiotic lactic acid bacteria carrier food in solid-state fermentation. *Food Control*, 41(2014), 1–6. <https://doi.org/10.1016/j.foodcont.2013.12.026>