



**ANTIOXIDANT POTENTIAL AND CANCER-SPECIFIC CYTOTOXIC  
EFFECT OF SELECTED MARINE MICROALGAL EXTRACTS**

By

**FERDOUS UMME TAMANNA**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of  
Doctor of Philosophy**

**February 2023**

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## DEDICATION

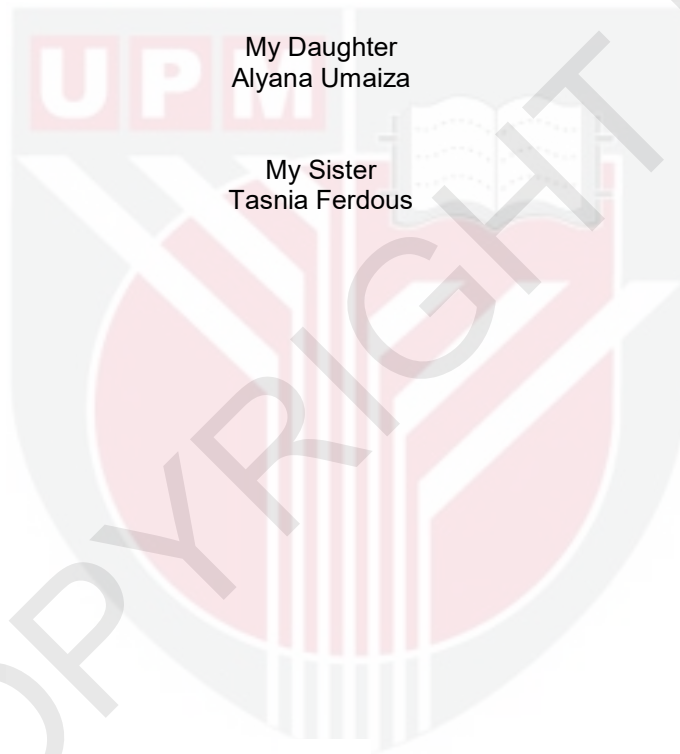
To

My Parents  
Mohammad Abdul Mabud  
Mahinur Nesa

My Husband  
Dr. Md Shahrul Islam

My Daughter  
Alyana Umaiza

My Sister  
Tasnia Ferdous



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

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**February 2023**

**Chairman : Associate Professor Zetty Norhana Balia Yusof, PhD**  
**Institute : Bioscience**

Microalgae derived metabolites have shown potential biological activities, especially antioxidant and cytotoxicity. Marine microalgae are often considered as a mother lode of antioxidant and antitumor compounds due to their inhabitants in the harsh marine environment. Despite having valuable and novel metabolites, the marine microalgae species are still not thoroughly investigated for their pharmaceutical and nutraceutical importance. Since cancer treatment with synthetic drugs shows adverse effects, it is urgent to search for alternative therapy from this promising marine source. Therefore, this study was focused to investigate the crude extracts of six marine microalgae species, *Chlorella* sp., *Tetraselmis* sp., *Nannochloropsis* sp., *Isochrysis* sp., *Chaetoceros* sp., and *Thalassiosira* sp., isolated from Malaysian coastal region in terms of their antioxidant activity and cytotoxicity against human breast cancer cells, MCF-7. These six marine microalgae are considered safe for human and animal usage as well as frequently used as fish feed and dietary supplements. Moreover, microalgae need shorter time, less nutrient and no arable land to grow. These microalgal species were collected and identified based on morphological and molecular characteristics. A total of forty-eight crude extracts from six marine microalgae species were prepared using eight different polarity solvents. From the antioxidant assays, methanol and ethyl acetate extract of *Tetraselmis* sp. exhibited significantly higher ( $p < 0.05$ ) antioxidant activities, revealed through DPPH ( $54.41 \pm 1.18$  mg Trolox Equivalent Antioxidant Capacity or TEAC/g extract) and ABTS ( $41.57 \pm 0.83$  mg TEAC/g extract) radical scavenging activities, respectively than the rest. Ethyl acetate extract of *Tetraselmis* sp. also showed high ferric reducing power ( $113.46 \pm 4.83$  mg TEAC/g extract). On the other hand, ethanol extract of *Isochrysis* sp. reduced the viability of human breast cancer, MCF-7 cells to  $7.24 \pm 0.47\%$  after 72 hours of incubation, at a concentration of  $100 \mu\text{g/ml}$ . The IC<sub>50</sub> (half maximal inhibitory concentration) value was  $13.37 \pm 0.59 \mu\text{g/ml}$  after 24 hours in MCF-7 cells and  $>100 \mu\text{g/ml}$  in non-cancerous human lung fibroblast cells, MRC-5. Ethanol extract of *Isochrysis*

sp. was further investigated for apoptosis induction in MCF-7 cells. With the increasing concentration of the extract, a reduction in MCF-7 cell population was observed. The Annexin V-FITC and propidium iodide staining analysis confirmed that the mode of cell death is mainly apoptosis. Cell cycle analysis revealed the accumulation of cells in the sub-G<sub>0</sub> phase which suggests induction of apoptosis and G<sub>2</sub>/M cycle arrest. RT-qPCR analysis revealed an up-regulation of the proapoptotic *Bax* gene and tumor suppressor *p53* gene. Metabolite profiling by Liquid Chromatography/Mass Spectrometry showed the presence of possible metabolites from fatty acid, sphingolipid, carotenoid, and phenolic classes. In conclusion, marine microalgae from indigenous sources have shown antioxidant and cancer-selective cytotoxicity. The data suggest that crude ethanolic extract from marine *Isochrysis* sp. has induced apoptosis and cell cycle arrest in human breast cancer cells, MCF-7, while methanol and ethyl acetate extracts from marine *Tetraselmis* sp. have good radical scavenging and ferric reduction capability. Therefore, indigenous marine *Isochrysis* sp. and *Tetraselmis* sp. may have potential therapeutic value for treating human breast cancer and nutraceutical use, respectively, which needs further investigation along with extensive *in vivo* study.

Keywords: Antioxidant, Apoptosis, Cytotoxicity, Microalgae, MCF-7

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## **POTENSI ANTIOKSIDAN DAN KESAN CYTOTOXIC KHUSUS KANSER DARI EKSTRAK MIKROAGAL LAUT TERPILIH**

Oleh

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Metabolit terhasil mikroalga telah menunjukkan aktiviti biologi yang berpotensi, terutamanya antioksidan dan sitotoksiti. Mikroalga marin sering dianggap sebagai induk sebatian antioksidan dan antitumor kerana mendiami persekitaran marin yang keras. Walaupun mempunyai metabolit yang berharga dan baru, spesies mikroalga marin masih belum disiasat secara menyeluruh untuk kepentingan farmaseutikal dan nutrasetikalnya. Oleh kerana rawatan kanser dengan ubat sintetik menunjukkan kesan buruk, adalah mendesak untuk mencari terapi alternatif daripada sumber marin yang menjanjikan ini. Oleh itu, kajian ini difokuskan untuk menyiasat ekstrak mentah enam spesies mikroalga marin, *Chlorella* sp., *Tetraselmis* sp., *Nannochloropsis* sp., *Isochrysis* sp., *Chaetoceros* sp., dan *Thalassiosira* sp., yang diasingkan daripada kawasan pantai Malaysia dari segi aktiviti antioksidan dan sitotoksiti mereka terhadap sel kanser payudara manusia, MCF-7. Enam mikroalga marin ini dianggap selamat untuk kegunaan manusia dan haiwan serta kerap digunakan sebagai makanan ikan dan makanan tambahan. Selain itu, mikroalga memerlukan masa yang lebih singkat, kurang nutrien dan tiada tanah yang boleh ditanam untuk tumbuh. Spesies mikroalga dikumpul dan dikenal pasti berdasarkan ciri-ciri morfologi dan molekul. Sebanyak empat puluh lapan ekstrak mentah daripada enam spesies mikroalga marin telah disediakan menggunakan lapan pelarut dengan kekutuban yang berbeza. Ekstrak-ekstrak mentah ini didedahkan kepada ujian antioksidan dan ujian kesitotoksikan kepekatan tunggal. Daripada ujian antioksidan, ekstrak metanol dan ekstrak etil asetat *Tetraselmis* sp. menunjukkan aktiviti antioksidan yang jauh lebih tinggi ( $p < 0.05$ ), yang didedahkan melalui aktiviti penghapusan radikal DPPH ( $54.41 \pm 1.18$  mg TEAC/g) dan ABTS ( $41.57 \pm 0.83$  mg ekstrak TEAC/g) daripada yang lain. Ekstrak etil asetat *Tetraselmis* sp. juga menunjukkan kuasa penurunan ferik yang tinggi ( $113.46 \pm 4.83$  mg TEAC/g ekstrak) yang didedahkan melalui ujian FRAP. Ujian MTT kepekatan tunggal mendedahkan bahawa ekstrak etanol *Isochrysis* sp. mengurangkan kebolehhidupan kanser payudara manusia, sel

MCF-7 kepada  $7.24 \pm 0.47\%$  selepas 72 jam pengeraman, pada kepekatan  $100 \mu\text{g/ml}$ . Nilai IC<sub>50</sub> (separuh kepekatan perencatan maksimum) ialah  $13.37 \pm 0.59 \mu\text{g/ml}$  selepas 24 jam dalam sel MCF-7 dan  $>100 \mu\text{g/ml}$  dalam sel bukan kanser fibroblas paru-paru manusia, MRC-5. Ekstrak etanol *Isochrysis* sp. telah dikaji lebih lanjut untuk induksi apoptosis dalam sel MCF-7. Pemerhatian morfologi di bawah mikroskop cahaya mendedahkan pengecutan sel, pembundaran, pemeluwapan kandungan selular, dan pembleban membran dalam sel MCF-7 yang dirawat berbanding dengan sel yang tidak dirawat. Dengan peningkatan kepekatan ekstrak, pengurangan populasi sel juga diperhatikan. Analisis pewarnaan Annexin V-FITC dan PI mengesahkan bahawa mod kematian sel yang utama adalah apoptosis. Analisis kitaran sel mendedahkan pengumpulan sel dalam fasa sub-G<sub>0</sub> yang mencadangkan induksi apoptosis dan henti kitaran G<sub>2</sub>/M. Analisis RT-qPCR mendedahkan peningkatan gen Bax proapoptotik dan gen p53 penindas tumor. Pemprofilan metabolit oleh LC/MS menunjukkan kemungkinan kehadiran metabolit daripada asid lemak, sfingolipid, karotenoid, dan kelas fenolik. Kesimpulannya, mikroalga laut dari sumber asli telah menunjukkan antioksidan dan sitotoksitas selektif barah. Data menunjukkan bahawa ekstrak etanol kasar dari *Isochrysis* sp laut. telah menyebabkan apoptosis dan penangkapan kitaran sel pada sel barah payudara manusia, MCF-7, sementara ekstrak metanol dan etil asetat dari *Tetraselmis* sp laut. mempunyai keupayaan penyingkiran radikal dan pengurangan ferrik yang baik. Oleh itu, *Isochrysis* sp. laut asli dan *Tetraselmis* sp. mungkin mempunyai nilai terapi yang berpotensi untuk merawat barah payudara manusia dan penggunaan nutraceutical, yang memerlukan penyelidikan lebih lanjut bersama.

Kata kunci Antioksidan, Apoptosis, Sitotoksiti, Mikroalga, MCF-7

## ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and the Most Merciful

All praise to Allah SWT and peace be upon our prophet Muhammad (SAW). Praise Allah for granting me the strength and blessing to pursue and successfully complete this study.

I would like to express my sincere gratitude and appreciation to my supervisor Assoc. Prof. Dr. Zetty Norhana Balia Yusof for her continuous support throughout the study with her guidance, patience, motivation and immense knowledge. I would like to thank my co-supervisors, Prof. Dr. Khozirah Shaari and Dr. Saila Ismail for their insightful comments, suggestions, and unwavering support. My sincerest appreciation to my co-supervisor Dr. Armania Nurdin for her regular guidance, technical support, encouragement, hand-on training and intelligent counseling.

My heartfelt thanks go to all the staff of the Institute of Bioscience, all the research officers of AquaHealth lab, especially Muhammad Farhan Nazarudin and Dr Farah, and also to all the staff of the Faculty of Biotechnology and Biomolecular Science for their support and kindness. I wish to extend my appreciation to my all colleagues and friends who had supported me technically and morally, especially Aisamuddin Ardi Zainal Abidin, Reem A. Dawood, Nor Izzati Husna Noor Hisham, Nur Farhah Nabihan Ismail, Nur Asna Faiqah Johari, Nur Aininie Yusoh, Gan Yian Chee, Tan Hui Teng, Yam Sim Khaw, Nur Amirah Izyan Noor Mazli, Muhammad Raznisyafiq Razak, Chin Yong Kit, Yan Shan, Munirah Adibah, Leen, Ashikin Afzan and all friends from the virology lab, Biotech2.

I would like to acknowledge and thank the Islamic Development Bank (IsDB) for the financial support to carry out my PhD smoothly.

I would like to express my sincerest gratitude to my parents, my father-in-law and my younger sister for their prayers, love and emotional support. I am truly indebted to my husband, Dr. Md. Shaharul Islam for his untiring support, continuous encouragement and understanding. This accomplishment would not have been possible without his support and sacrifice.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

ABTS	2,2'-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid)
Abs	Absorbance
ANOVA	Analysis of variance
ATCC	American type culture collection
<i>Bax</i>	BCL2 Associated X protein
BLAST	Basic Local Alignment Tool
CAT	Catalase
CO <sub>2</sub>	Carbon dioxide
DCM	Dichloromethane
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DPPH	2,2-diphenyl-1-picrylhydrazyl
EDTA	Ethylene diamine tetra-acetic acid
FBS	Fetal bovine serum
F-C	Folin-Ciocalteu
FC	Flow cytometry
FITC	Fluorescein isothiocyanate
FRAP	Ferric reducing antioxidant power
g	Gram
GAE	Gallic acid equivalents
GPx	Glutathione peroxidase
HCl	Hydrochloric acid
I-AQUAS	International Institute of Aquaculture and Aquatic Science
kg	Kilogram
LC-MS	Liquid chromatography-mass spectrometry

mg	Milligram
ml	Milliliter
mM	Milimolar
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
nm	Nanometer
OD	Optical density
MCF-7	Michigan Cancer Foundation-7
MRC-5	Medical Research Council cell strain 5
<i>p</i>	Significance difference
<i>p53</i>	Tumor protein
PCR	Polymerase chain reaction
PI	<i>Propidium iodide</i>
PMS	Premenstrual syndrome
PS	Phosphatidylserine
QE	Quercetin equivalents
<i>r</i>	Pearson correlation coefficient
RNA	Ribonucleic acid
RPMI	Roswell Park Memorial Institute Medium
SDS	Sodium dodecyl sulfate
SEM	Standard error mean
SOD	Superoxide dismutase
SPSS	Statistical package for the social sciences
TAP	Tris-acetate-phosphate
TEAC	Trolox equivalents antioxidant capacity
TFC	Total Flavonoid Content
TPC	Total Phenolic Content

$T_r$	Retention time
UV	Ultraviolet
v/v	Volume per volume
w/v	Weight per volume
w/w	Weight per weight
$\mu\text{g}$	Microgram
$\mu\text{l}$	Microliter
$\mu\text{M}$	Micromolar



## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of the Study

Marine organisms reside in a salty aqueous environment that covers 71% of the earth's surface and accounts for 90% of the earth's biosphere (Wang et al., 2020). This is a gigantic reservoir for diversified marine species, with approximately 2500,000 species so far (Khalifa et al., 2019). The marine environment is defined as a harsh and unfavorable domain where scarcity of light, nutrients, high pH, pressure, and continuous encountering of predators make the marine flora tackle this situation with adaptive and symbiotic mechanisms. Their survival strategy, which includes defense against predators, finding a mate, or beating competitors, helps them to produce a broad range of secondary metabolites (Wang et al., 2020). These secondary metabolites from marine organisms are now exploited to design life-saving drugs and drug leads.

An array of natural products, that have pharmaceutical importance, are being offered by marine organisms which can be exploited to treat human diseases, especially cancer. The marine environment represents huge biodiversity with 36 phyla, and it is anticipated that huge taxonomic diversity is related to the broad chemical diversity of these natural products. These diversified marine natural products are assumed to have high bioactivity, especially cytotoxicity, and could be a potential source of anticancer drug candidates. But these colossal reservoirs of natural products are still largely unexplored. Only 18% of marine natural products are discovered so far compared to terrestrial products (Ruiz-Torres et al., 2017). Different medicinal plants, like garlic, ginger, turmeric, black cohosh, burdock, sumac, sundew, and citrus-based plants, showed anticancer activities against breast cancer cell lines, MCF-7, MDA-MB-231 and T47B (Shrihastini et al., 2021). Metabolites isolated from marine invertebrates and seaweed showed cytotoxicity against MCF-7 and MDA-MB-231 cell lines (Veríssimo et al., 2021). Currently, a total of 12 marine-derived cancer drugs, are available on the market but only two of them are for breast cancer treatment and another 23 compounds from the marine organisms are in different phases of cancer clinical trials (<https://www.marinepharmacology.org/>, accessed on 27 September 2022). Most of these drugs, such as Cytarabine, Vidarabine, Eribulin Mesylate, are extracted from marine invertebrates. A major challenge is associated with cultivating these invertebrates for a sustainable supply of the drug leads. Moreover, some of the marine-derived drugs were suspended from clinical trials or withdrawn from the market due to toxicity (Saeed et al., 2021). Therefore, a search for a safe, sustainable, and less toxic source of marine-derived anticancer agents is warranted.

Marine microalgae are accounted for a major portion of oceanic biomass. Microalgae, both eukaryotic and cyanobacteria, comprise more than 30,000 species and contribute up to 40% of global productivity (Sithranga Boopathy &

Kathiresan, 2010). They contain a wide range of phytochemicals like carotenoids, phenolics, flavonoids, fatty acids, alkaloids, polysaccharides, and vitamins. These phytochemicals make them attractive sources of bioactive compounds that are frequently used in the pharmaceutical, cosmetic, aquaculture, and energy-related industry (Sansone & Brunet, 2019; Abd El-Hack et al., 2019). Microalgae are known to produce different anticancer compounds like, polyunsaturated aldehydes, fucoxanthin, violaxanthin, stigmaterol, eicosapentaenoic Acid (EPA), nonyl 8-acetoxy-6-methyloctanoate, monogalactosyl glycerols and polysaccharides (Martínez Andrade et al., 2018).

Microalgae can withstand all environmental extremities, from cold to hydrothermal vents. On a lab-scale or industrial scale, they can be grown all year-round irrespective of any seasonal variation, which also excludes the need for long-term storage and helps to avoid valuable phytochemical degradation. They can be grown with a limited nutritional supply and the advantageous point is that microalgae can be grown in wastewater as a nutrient source, which in turn, reduces carbon footprint and water usage (Gong & Bassi, 2016). Not only that, microalgae can be grown in large photo bioreactors without competing with arable land and disturbing the human food chain (Rajkumar et al., 2014). Moreover, microalgae can grow faster than terrestrial plants. Their fast replicability, easy cultivation, ecological sustainability and wide adaption capability make them an alternative for the production of high-value products. They are important in terms of drug discovery due to their metabolic plasticity. They can produce pharmaceutically important phytochemicals, especially anticancer compounds (Martínez Andrade et al., 2018). Due to the presence of antioxidants, microalgal biomass is used popularly as dietary supplements and also as food additives (Sansone & Brunet, 2019). Hamidi et al., (2020) mentioned that marine microalgae may produce more carotenoids and EPA than marine bacteria.

Prokaryotic microalgae, cyanobacteria, has long been explored for its antitumor activity against different cancer cells. Compounds like Hantupeptin A, Malyngamide, Pitipeptolides from cyanobacteria, *Lyngbya* sp. showed anticancer activity against breast cancer cells, MDA-MB-231 and MCF-7. Ankaraholide A from *Geitlerinema* sp., coibamide A from *Leptolyngbya* sp. exhibited anticancer activity against MDA-MB-231 cells. One of the most studied cyanobacteria species *Symploca* sp. VP642 produce peptide called dolastatins which has shown anticancer effect on MCF-7 cells (Qamar et al., 2021). Some of the cyanobacterial phytochemicals are now in the clinical trial, as a conjugate with other drugs. Cyanobacterial species, *Symploca* sp. and *Caldora penicillata* have successfully ended up with a few drugs, mainly anti-cancer (Zuo & Kwok, 2021). But a significant challenge exists in cultivating the pure culture of these microalgae since they are in a symbiotic relationship with invertebrate host. Moreover, some of the drugs from *C. penicillata*, which were in clinical trials, have been discontinued due to toxicity (Saeed et al., 2021). Therefore, edible, less toxic and cultivable microalgal species should be prioritized for a sustainable and safe source of pharmaceutically important compounds.

Eukaryotic microalgae have been known for their low toxicity. For instance, *Chlorella* sp. is considered as generally recognized as safe (GRAS) which is approved by the U.S. Food and Drug Administration (FDA). No toxin is found from the microalgae species like, *Isochrysis* sp., *Nannochloropsis* sp., *Tetraselmis* sp., and *Thalassiosira* sp. and these microalgae including *Chaetoceros* sp. are now frequently used in aquaculture industries as fish feed (Lucakova et al., 2022). Moreover, green eukaryotic microalgae, *Chlorella* sp., *Nannochloropsis* sp., *Tetraselmis* sp. are now used as commercial food supplement, while brown microalgae *Isochrysis* sp. is used as food additive (Yasir et al., 2022). These eukaryotic microalgae are considered prolific sources of various bioactive compounds like carotenoids, polyphenols, sulfated polysaccharides, fatty acids, minerals, and peptides but extensive bio-prospection from eukaryotic marine microalgae is warranted. Therefore, all these six microalgae from marine sources need thorough investigation for their bioactive properties (Lauritano et al., 2016; Ferdous & Yusof, 2021c).

## 1.2 Problem Statement

Cancer is one of the leading causes of mortality worldwide. Breast cancer is the most common cancer type in the world and also in Malaysia. This is the second most common cause of cancer-related death in Malaysia (Sung et al., 2021; WHO, 2021). Though early diagnosis can reduce the mortality rate, severe side effects from the treatment and multidrug resistance make this disease a major health complication and engender the need for searching for novel anti-cancer biomolecules from natural sources, as more than 60% of clinically useful anticancer drugs were developed thus (Li et al., 2017; Cragg et al., 2009).

Marine eukaryotic microalgae are an excellent source of valuable bioactive compounds which may have antioxidant and anticancer properties (Martínez Andrade et al., 2018). Indigenous eukaryotic marine microalgae species, *Isochrysis galbana* and *Chaetoceros calcitrans*, isolated from Malaysian coastal areas have shown good fatty acids profile (Natrah et al., 2007; Bustamam et al., 2021; Azizan et al., 2020). Total phenolic content and high antioxidant activities were reported for indigenous marine *Tetraselmis tetrathele*, *Nannochloropsis* sp. and *Chaetoceros calcitrans* (Farahin et al., 2016; Foo et al., 2015; Goh et al., 2010). Phang et al., (2015) reported that the Port Dickson region in Malaysia showed the presence of diversified marine microalgal species which may be attributed to the suitable salinity (32-34 ppt) and pH range (6.88-7.49) and solar irradiance of this region. Hossain et al., (2020) also highlighted the suitability of Malaysian weather and location for microalgal growth in terms of nutrient availability, solar irradiance, salinity and temperature. However, these microalgae remain unexplored vastly in terms of their bioactivities. Therefore, this study aims to investigate the antioxidant and cytotoxic activities of the crude extracts from marine indigenous eukaryotic microalgae, *Chlorella* sp., *Tetraselmis* sp., *Isochrysis* sp., *Nannochloropsis* sp., *Chaetoceros* sp., and *Thalassiosira* sp. and to demonstrate their cell killing mechanism in human breast cancer cell line, MCF-7.

### 1.3 Hypothesis

- a. Marine microalgal extracts from different solvents with different polarities show antioxidant activity, especially radical scavenging activity and ferric reduction capability. There is a correlation between antioxidant activity and total phenolic or flavonoid content of the microalgal extracts.
- b. Microalgal extracts are cytotoxic to MCF-7 cancer cell line but show less toxicity towards non-cancerous MRC-5 cell line. The most potential cytotoxic extract induces apoptosis while upregulating apoptosis related genes, *Bax* and *p53* and arrests cell cycle in MCF-7 cells.
- c. Microalgal extract contains several phytochemicals like, fatty acids, carotenoids, lipoids, and polyphenols which are attributed to their antioxidant and cytotoxic properties.

### 1.4 Objectives

#### 1.4.1 Main objective

To study the antioxidant activity of the extracts from six indigenous marine microalgae (*Chlorella* sp., *Tetraselmis* sp., *Isochrysis* sp., *Nannochloropsis* sp., *Chaetoceros* sp., and *Thalassiosira* sp.) and their cytotoxic activity on human breast cancer cell line, MCF-7.

#### 1.4.2 Specific Objectives

1. To identify the morphological and molecular characteristics of six marine microalgae including *Chlorella* sp., *Isochrysis* sp., *Nannochloropsis* sp., *Tetraselmis* sp., *Chaetoceros* sp. and *Thalassiosira* sp.
2. To compare the antioxidant activity, total phenolic and flavonoid contents of different solvent extracts of the marine microalgae.
3. To evaluate the cytotoxic effects, the mechanism of cell death, cell cycle analysis, the expression level of apoptosis-related genes, *Bax* and *p53*, of the selected microalgal extract on human breast cancer cell line (MCF-7)
4. To identify the potential compounds responsible for the cytotoxic activity of the selected microalgae extract using liquid chromatography-mass spectrometry

## REFERENCES

- Abd El-Hack, M. E., Abdelnour, S., Alagawany, M., Abdo, M., Sakr, M. A., Khafaga, A. F., Mahgoub, S. A., Elnesr, S. S., & Gebriel, M. G. (2019a). Microalgae in modern cancer therapy: Current knowledge. *Biomedicine and Pharmacotherapy*, *111*(December 2018), 42–50. <https://doi.org/10.1016/j.biopha.2018.12.069>
- Abd El-Hack, M. E., Abdelnour, S., Alagawany, M., Abdo, M., Sakr, M. A., Khafaga, A. F., Mahgoub, S. A., Elnesr, S. S., & Gebriel, M. G. (2019b). Microalgae in modern cancer therapy: Current knowledge. *Biomedicine and Pharmacotherapy*, *111*(December 2018), 42–50. <https://doi.org/10.1016/j.biopha.2018.12.069>
- Abdi Goushbolagh, N., Farhood, B., Astani, A., Nikfarjam, A., Kalantari, M., & Zare, M. H. (2018). Quantitative Cytotoxicity, Cellular Uptake and Radioprotection Effect of Cerium Oxide Nanoparticles in MRC-5 Normal Cells and MCF-7 Cancerous Cells. *BioNanoScience*, *8*(3), 769–777. <https://doi.org/10.1007/s12668-018-0538-z>
- Abdolmohammadi, M. H., Fallahian, F., Fakhroueian, Z., Kamalian, M., Keyhanvar, P., M. Harsini, F., & Shafiekhani, A. (2017). Application of new ZnO nanoformulation and Ag/Fe/ZnO nanocomposites as water-based nanofluids to consider in vitro cytotoxic effects against MCF-7 breast cancer cells. *Artificial Cells, Nanomedicine and Biotechnology*, *45*(8), 1769–1777. <https://doi.org/10.1080/21691401.2017.1290643>
- Abolhasani, M. H., Safavi, M., Goodarzi, M. T., Kassaei, S. M., & Azin, M. (2018). Identification and anti-cancer activity in 2D and 3D cell culture evaluation of an Iranian isolated marine microalgae Picochlorum sp. RCC486. *DARU, Journal of Pharmaceutical Sciences*, *26*(2), 105–116. <https://doi.org/10.1007/s40199-018-0213-5>
- Abotaleb, M., Kubatka, P., Caprnda, M., Varghese, E., Zolakova, B., Zubor, P., Opatrilova, R., Kruzliak, P., Stefanicka, P., & Büsselberg, D. (2018). Chemotherapeutic agents for the treatment of metastatic breast cancer: An update. *Biomedicine and Pharmacotherapy*, *101*(August 2017), 458–477. <https://doi.org/10.1016/j.biopha.2018.02.108>
- Ahmed, F., Fanning, K., Netzel, M., Turner, W., Li, Y., & Schenk, P. M. (2014). Profiling of carotenoids and antioxidant capacity of microalgae from subtropical coastal and brackish waters. *Food Chemistry*, *165*, 300–306. <https://doi.org/10.1016/j.foodchem.2014.05.107>
- Akhtari-Zavare, M., Mohd-Sidik, S., Periasamy, U., Rampal, L., Fadhilah, S. I., & Mahmud, R. (2018). Determinants of quality of life among Malaysian cancer patients: A cross-sectional study. *Health and Quality of Life Outcomes*, *16*(1), 1–17. <https://doi.org/10.1186/s12955-018-0989-5>
- Amna Kashif, S., Hwang, Y. J., & Park, J. K. (2018). Potent biomedical applications of isolated polysaccharides from marine microalgae



Tetraselmis species. *Bioprocess and Biosystems Engineering*, 41(11), 1611–1620. <https://doi.org/10.1007/s00449-018-1987-z>

- Andriopoulos, V., Gkioni, M. D., Koutra, E., Mastropetros, S. G., Lamari, F. N., Hatziantoniou, S., & Kornaros, M. (2022). Total Phenolic Content , Biomass Composition , and Antioxidant Activity of Selected Marine Microalgal Species with Potential as Aquaculture Feed. *Antioxidants*, 11, 1320.
- Ardi, A., Abidin, Z., Othman, N. A., & Yusoff, F. (2021). Determination of transgene stability in *Nannochloropsis* sp . transformed with immunogenic peptide for oral vaccination against vibriosis Content courtesy of Springer Nature , terms of use apply . Rights reserved . Content courtesy of Springer Nature , ter. *Aquaculture International*, 29, 477–486.
- Arifin, K. T., Sulaiman, S., Saad, S., Damanhuri, H. A., Zurinah, W., Ngah, W., Anum, Y., & Yusof, M. (2017). Elevation of tumour markers TGF-  $\beta$  , M 2 -PK , OV-6 and AFP in hepatocellular carcinoma ( HCC )-induced rats and their suppression by microalgae *Chlorella vulgaris*. *BMC Cancer*, 17(879), 1–9. <https://doi.org/10.1186/s12885-017-3883-3>
- Armania, N., Yazan, L. S., Ismail, I. S., Foo, J. B., Tor, Y. S., Ishak, N., Ismail, N., & Ismail, M. (2013). *Dillenia suffruticosa* extract inhibits proliferation of human breast cancer cell lines (MCF-7 and MDA-MB-231) via Induction of G2/M arrest and apoptosis. *Molecules*, 18(11), 13320–13339. <https://doi.org/10.3390/molecules181113320>
- Arora, M. (2016). *Tetraselmis* : an Introduction. *The Botanica*, 66, 155–175.
- Ashashalini, A., Ali, S., Anuradha, V., Yogananth, N., Suganya, V., & Bhuvana, P. (2018). Cultivation , Quantification and Pharmacognostic Study of methanolic extract of *Thalassiosira weissflogii*. *IOSR Journal of Pharmacy*, 8(10), 36–43.
- Ashfaq, W., Rehman, K., Siddique, M. I., & Khan, Q. A. A. (2019). Eicosapentaenoic Acid and Docosahexaenoic Acid from Fish Oil and Their Role in Cancer Research. *Food Reviews International*, 00(00), 1–20. <https://doi.org/10.1080/87559129.2019.1686761>
- Assefa, A. D., Ko, E. Y., Moon, S. H., & Keum, Y. S. (2016). Antioxidant and antiplatelet activities of flavonoid-rich fractions of three citrus fruits from Korea. *3 Biotech*, 6(1), 1–10. <https://doi.org/10.1007/s13205-016-0424-8>
- Aung, T. N., Qu, Z., Kortschak, R. D., & Adelson, D. L. (2017). Understanding the effectiveness of natural compound mixtures in cancer through their molecular mode of action. *International Journal of Molecular Sciences*, 18(3). <https://doi.org/10.3390/ijms18030656>
- Azizan, A., Maulidiani, M., Rudyanto, R., Shaari, K., Ismail, I. S., Nagao, N., & Abas, F. (2020). Mass spectrometry-based metabolomics combined

with quantitative analysis of the microalgal diatom (*Chaetoceros calcitrans*). *Marine Drugs*, 18(8), 3–5. <https://doi.org/10.3390/MD18080403>

- Badrulhisham, N. S. R., Ab Hamid, S. N. P., Ismail, M. A. H., Yong, Y. K., Muhamad Zakuan, N., Harith, H. H., Saidi, H. I., & Nurdin, A. (2020). Harvested locations influence the total phenolic content, antioxidant levels, cytotoxic, and anti-inflammatory activities of stingless bee honey. *Journal of Asia-Pacific Entomology*, 23(4), 950–956. <https://doi.org/10.1016/j.aspen.2020.07.015>
- Bajpai, V. K., Shukla, S., Kang, S. M., Hwang, S. K., Song, X., Huh, Y. S., & Han, Y. K. (2018). Developments of cyanobacteria for nano-marine drugs: Relevance of nanoformulations in cancer therapies. *Marine Drugs*, 16(6). <https://doi.org/10.3390/md16060179>
- Balaji, M., Thamilvanan, D., Vinayagam, S. C., & Balakumar, B. S. (2017). Anticancer, Antioxidant activity and GC-MS analysis of selected microalgal members of Chlorophyceae. *International Journal of Pharmaceutical Sciences and Research*, 8(8), 3302–3314. [https://doi.org/10.13040/IJPSR.0975-8232.8\(8\).3302-14](https://doi.org/10.13040/IJPSR.0975-8232.8(8).3302-14)
- Banskota, A. H., Sperker, S., Stefanova, R., McGinn, P. J., & O'Leary, S. J. B. (2019). Antioxidant properties and lipid composition of selected microalgae. *Journal of Applied Phycology*, 31(1), 309–318. <https://doi.org/10.1007/s10811-018-1523-1>
- Baudelet, P. H., Gagez, A. L., Bérard, J. B., Juin, C., Bridiau, N., Kaas, R., Thiéry, V., Cadoret, J. P., & Picot, L. (2013). Antiproliferative activity of *Cyanophora paradoxa* pigments in melanoma, breast and lung cancer cells. *Marine Drugs*, 11(11), 4390–4406. <https://doi.org/10.3390/md11114390>
- Beyerinck, M. W. (1890). Culturversuche mit Zoochlorellen, Lichenengonidien und anderen niederen Algen. *Botanische Zeitung*, 47, 725–739, 741–754, 757–768, 781–785.
- Bhat, A. A., Prabhu, K. S., Kuttikrishnan, S., Krishnankutty, R., Babu, J., Mohammad, R. M., & Uddin, S. (2017). Potential therapeutic targets of Guggulsterone in cancer. *Nutrition and Metabolism*, 14(1), 1–11. <https://doi.org/10.1186/s12986-017-0180-8>
- Bhattacharjya, R., Kiran Marella, T., Tiwari, A., Saxena, A., Kumar Singh, P., & Mishra, B. (2020). Bioprospecting of marine diatoms *Thalassiosira*, *Skeletonema* and *Chaetoceros* for lipids and other value-added products. *Bioresource Technology*, 318(September), 124073. <https://doi.org/10.1016/j.biortech.2020.124073>
- Bhingarde, S., & Nimse, A. (2022). *Spirulina Farming : A Superfood*. 2(6).
- Bin, Q., Rao, H., Hu, J. N., Liu, R., Fan, Y. W., Li, J., Deng, Z. Y., Zhong, X., & Du, F. L. (2013). The caspase pathway of linoelaidic acid (9t, 12t-

C18:2)-induced apoptosis in human umbilical vein endothelial cells. *Lipids*, 48(2), 115–126. <https://doi.org/10.1007/s11745-012-3728-4>

Bongiovani, N., Virginia Sanchez-Puerta, M., Popovich, C., & Leonardi, P. (2014). Identificación molecular y filogenética de una cepa oleaginosa de *nannochloropsis oceanica* (eustigmatophyceae) aislada de la costa atlántica suroeste (argentina). *Revista de Biología Marina y Oceanografía*, 49(3), 615–623. <https://doi.org/10.4067/S0718-19572014000300019>

Borowitzka, M. A. (2018). Biology of Microalgae. In *Microalgae in Health and Disease Prevention* (Issue 1998, pp. 23–72). Elsevier Inc. <https://doi.org/10.1016/b978-0-12-811405-6.00003-7>

Bozarth, A., Maier, U. G., & Zauner, S. (2009). Diatoms in biotechnology: Modern tools and applications. *Applied Microbiology and Biotechnology*, 82(2), 195–201. <https://doi.org/10.1007/s00253-008-1804-8>

Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., & Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians*, 68(6), 394–424. <https://doi.org/10.3322/caac.21492>

Bulut, O., Akın, D., Sönmez, Ç., Öktem, A., Yücel, M., & Öktem, H. A. (2019). Phenolic compounds, carotenoids, and antioxidant capacities of a thermo-tolerant *Scenedesmus* sp. (Chlorophyta) extracted with different solvents. *Journal of Applied Phycology*, 31(3), 1675–1683. <https://doi.org/10.1007/s10811-018-1726-5>

Bustamam, M. S. A., Pantami, H. A., Azizan, A., Shaari, K., Min, C. C., Abas, F., Nagao, N., Maulidiani, M., Banerjee, S., Sulaiman, F., & Ismail, I. S. (2021). Complementary Analytical Platforms of NMR Spectroscopy and LCMS Analysis in the Metabolite Profiling of *Isochrysis galbana*. *Marine Drugs*, 19(3). <https://doi.org/10.3390/md19030139>

Camacho, F., Macedo, A., & Malcata, F. (2019). Potential industrial applications and commercialization of microalgae in the functional food and feed industries: A short review. *Marine Drugs*, 17(6). <https://doi.org/10.3390/md17060312>

Camp, E. R., Patterson, L. D., Kester, M., & Voelkel-Johnson, C. (2017). Therapeutic implications of bioactive sphingolipids: A focus on colorectal cancer. *Cancer Biology and Therapy*, 18(9), 640–650. <https://doi.org/10.1080/15384047.2017.1345396>

Carneiro, B. A., & El-Deiry, W. S. (2020). Targeting apoptosis in cancer therapy. *Nature Reviews Clinical Oncology*, 17(7), 395–417. <https://doi.org/10.1038/s41571-020-0341-y>

Casas-Arrojo, V., Decara, J., Arrojo-Agudo, M. de los Á., Pérez-Manríquez, C., & Abdala-Díaz, R. T. (2021). Immunomodulatory, antioxidant activity

and cytotoxic effect of sulfated polysaccharides from porphyridium cruentum. (s.f.gray) nägeli. *Biomolecules*, 11(4). <https://doi.org/10.3390/biom11040488>

- Chahal, A., Saini, A. K., Chhillar, A. K., & Saini, R. V. (2018). Natural antioxidants as defense system against cancer. *Asian Journal of Pharmaceutical and Clinical Research*, 11(5), 38–44. <https://doi.org/10.22159/ajpcr.2018.v11i5.24119>
- Chaudhuri, D., Ghate, N. B., Deb, S., Panja, S., Sarkar, R., Rout, J., & Mandal, N. (2014). Assessment of the phytochemical constituents and antioxidant activity of a bloom forming microalgae *Euglena tuba*. *Biological Research*, 47(1), 1–11. <https://doi.org/10.1186/0717-6287-47-24>
- Chen, F., Leng, Y., Lu, Q., & Zhou, W. (2021). The application of microalgae biomass and bio-products as aquafeed for aquaculture. *Algal Research*, 60(August), 102541. <https://doi.org/10.1016/j.algal.2021.102541>
- Chen, G., Zhao, L., & Qi, Y. (2015). Enhancing the productivity of microalgae cultivated in wastewater toward biofuel production: A critical review. *Applied Energy*, 137, 282–291. <https://doi.org/10.1016/j.apenergy.2014.10.032>
- Chen, X., Song, L., Wang, H., Liu, S., Yu, H., Wang, X., Li, R., Liu, T., & Li, P. (2019). Partial characterization, the immune modulation and anticancer activities of sulfated polysaccharides from filamentous microalgae *Tribonema* sp. *Molecules*, 24(2), 1–11. <https://doi.org/10.3390/molecules24020322>
- Chern, S., Yusoff, F., Umar, M., & Biau, J. (2018). Increased fucoxanthin in *Chaetoceros calcitrans* extract exacerbates apoptosis in liver cancer cells via multiple targeted cellular pathways. *Biotechnology Reports*, 20, e00296. <https://doi.org/10.1016/j.btre.2018.e00296>
- Chipuk, J. E., Kuwana, T., Bouchier-Hayes, L., Droin, N. M., Newmeyer, D. D., Schuler, M., & Green, D. R. (2004). Direct Activation of Bax by p53 Mediates Mitochondrial Membrane Permeabilization and Apoptosis. *Science*, 303(5660), 1010–1014. <https://doi.org/10.1126/science.1092734>
- Chisti, Y. (2018). Society and microalgae: Understanding the past and present. In *Microalgae in Health and Disease Prevention* (pp. 11–21). Elsevier Inc. <https://doi.org/10.1016/B978-0-12-811405-6.00002-5>
- Chittora, D., Meena, M., Barupal, T., & Swapnil, P. (2020). Cyanobacteria as a source of biofertilizers for sustainable agriculture. *Biochemistry and Biophysics Reports*, 22(February), 100737. <https://doi.org/10.1016/j.bbrep.2020.100737>
- Choudhari, A. S., Mandave, P. C., Deshpande, M., Ranjekar, P., & Prakash, O. (2020). Phytochemicals in cancer treatment: From preclinical studies to

clinical practice. *Frontiers in Pharmacology*, 10(January), 1–17. <https://doi.org/10.3389/fphar.2019.01614>

- Chu, Y., Zhou, X., & Wang, X. (2021). Antibody-drug conjugates for the treatment of lymphoma: clinical advances and latest progress. *Journal of Hematology and Oncology*, 14(1), 1–20. <https://doi.org/10.1186/s13045-021-01097-z>
- Chung, J., Peng, H., Chu, Y., Hsieh, Y., Wang, S., & Chou, S. (2011). Anti-invasion and apoptosis induction of chlorella (*Chlorella sorokiniana*) in Hep G2 human hepatocellular carcinoma cells. *Journal of Functional Foods*, 4, 302–310. <https://doi.org/10.1016/j.jff.2011.12.008>
- Cichoński, J., & Chrzanowski, G. (2022). Microalgae as a Source of Valuable Phenolic Compounds and Carotenoids. *Molecules*, 27(24), 1–21. <https://doi.org/10.3390/molecules27248852>
- Cordoba-Matson, M. V, Arredondo-Vega, B. O., & Carreón-Palau, L. (2013). Evaluation of growth, cell size and biomass of *Isochrysis aff. galbana* (T-ISO) with two LED regimes. *All Res. J. Biol*, 4(April), 7–15.
- Coronado-Reyes, J. A., Acosta-Ramírez, E., Martínez-Olguín, M. V., & González-Hernández, J. C. (2022). Antioxidant Activity and Kinetic Characterization of *Chlorella vulgaris* Growth under Flask-Level Photoheterotrophic Growth Conditions. *Molecules*, 27(19). <https://doi.org/10.3390/molecules27196346>
- Corrêa, P. S., Morais Júnior, W. G., Martins, A. A., Caetano, N. S., & Mata, T. M. (2021). Microalgae biomolecules: Extraction, separation and purification methods. *Processes*, 9(1), 1–40. <https://doi.org/10.3390/pr9010010>
- Costa, J. A. V., Freitas, B. C. B. de, Lisboa, C. R., Santos, T. D., Bruschi, L. R. de F., & de Morais, M. G. (2019). Microalgal biorefinery from CO<sub>2</sub> and the effects under the Blue Economy. *Renewable and Sustainable Energy Reviews*, 99(October 2017), 58–65. <https://doi.org/10.1016/j.rser.2018.08.009>
- Cragg, G. M., Grothaus, P. G., & Newman, D. J. (2009). Impact of Natural Products on Developing New Anti-Cancer Agents †. *Chem Rev*, 109, 3012–3043.
- Cukier, P., Santini, F. C., Scaranti, M., & Hoff, A. O. (2017). Endocrine side effects of cancer immunotherapy. *Endocrine-Related Cancer*, 24(12), T331–T347. <https://doi.org/10.1530/ERC-17-0358>
- Custódio, L., Soares, F., Pereira, H., Barreira, L., Vizetto-Duarte, C., Rodrigues, M. J., Rauter, A. P., Alberício, F., & Varela, J. (2014). Fatty acid composition and biological activities of *Isochrysis galbana* T-ISO, *Tetraselmis* sp. and *Scenedesmus* sp.: Possible application in the pharmaceutical and functional food industries. *Journal of Applied Phycology*, 26(1), 151–161. <https://doi.org/10.1007/s10811-013-0098-0>

- de Morais, M. G., de Morais, E. G., Duarte, J. H., Deamici, K. M., Mitchell, B. G., & Costa, J. A. V. (2019). Biological CO<sub>2</sub> mitigation by microalgae: technological trends, future prospects and challenges. *World Journal of Microbiology and Biotechnology*, 35(5), 1–7. <https://doi.org/10.1007/s11274-019-2650-9>
- Dębowski, M., Zieliński, M., Kazimierowicz, J., Kujawska, N., & Talbierz, S. (2020). Microalgae cultivation technologies as an opportunity for bioenergetic system development—advantages and limitations. *Sustainability*, 12(23), 1–37. <https://doi.org/10.3390/su12239980>
- Del Mondo, A., Smerilli, A., Ambrosino, L., Albini, A., Noonan, D. M., Sansone, C., & Brunet, C. (2021). Insights into phenolic compounds from microalgae: structural variety and complex beneficial activities from health to nutraceuticals. *Critical Reviews in Biotechnology*, 41(2), 155–171. <https://doi.org/10.1080/07388551.2021.1874284>
- Denton, D., & Kumar, S. (2019). Autophagy-dependent cell death. *Cell Death and Differentiation*, 26(4), 605–616. <https://doi.org/10.1038/s41418-018-0252-y>
- Di Meo, S., Reed, T. T., Venditti, P., & Victor, V. M. (2016). Role of ROS and RNS Sources in Physiological and Pathological Conditions. *Oxidative Medicine and Cellular Longevity*, 2016, 1–44. <https://doi.org/10.1155/2016/1245049>
- Dilalla, V., Chaput, G., Williams, T., & Sultanem, K. (2020). Radiotherapy side effects: Integrating a survivorship clinical lens to better serve patients. *Current Oncology*, 27(2), 107–112. <https://doi.org/10.3747/co.27.6233>
- DiPaola, R. S. (2002). To Arrest or Not To G2-M Cell-Cycle Arrest. *Clinical Cancer Research*, 8(November), 3311–3314.
- Diprat, A. B., Silveira Thys, R. C., Rodrigues, E., & Rech, R. (2020). Chlorella sorokiniana: A new alternative source of carotenoids and proteins for gluten-free bread. *Lwt*, 134(April), 109974. <https://doi.org/10.1016/j.lwt.2020.109974>
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food and Drug Analysis*, 22(3), 296–302. <https://doi.org/10.1016/j.jfda.2013.11.001>
- Ebrahimi Nigjeh, S., Yusoff, F. M., Mohamed Alitheen, N. B., Rasoli, M., Keong, Y. S., & Omar, A. R. Bin. (2013). Cytotoxic effect of ethanol extract of microalga, *Chaetoceros calcitrans*, and its mechanisms in inducing apoptosis in human breast cancer cell line. *BioMed Research International*, 2013, 1–9. <https://doi.org/10.1155/2013/783690>
- Ebrahimpzadeh, M. A., Khalili, M., & Dehpour, A. A. (2018). Antioxidant activity of ethyl acetate and methanolic extracts of two marine algae,

nannochloropsis oculata and gracilaria gracilis - An in vitro assay. *Brazilian Journal of Pharmaceutical Sciences*, 54(1). <https://doi.org/10.1590/s2175-97902018000117280>

Eikenberry, J., Harris, A., Torabi, R., Lang, M., Denney, D., Verticchio Vercellin, A., & Siesky, B. (2020). Ocular side effects of target therapy and immunotherapy in patients with cutaneous malignant melanoma. *European Journal of Ophthalmology*. <https://doi.org/10.1177/1120672120930688>

Elkhateeb, W., El-Sayed, H., Fayad, W., Kolaibe, A. G. Al, Emam, M., & Daba, G. (2020). In vitro Anti-breast cancer and antifungal Bio-efficiency of some microalgal extracts Waill. *Journal of Membrane Science and Research*, 6(1), 81–89. <https://doi.org/10.22079/JMSR.2019.112686.1282>

Farahin, A. W., Yusoff, F. M., Nagao, N., Basri, M., & Shariff, M. (2016). Phenolic content and antioxidant activity of Tetraselmis tetrahale (West) Butcher 1959 cultured in annular photobioreactor. *Journal of Environmental Biology*, 37, 91–100.

Ferdous, U. T., Nurdin, A., Ismail, S., & Yusof, Z. N. B. (2022). Evaluation of the antioxidant and cytotoxic activities of crude extracts from marine *Chlorella* sp. *Biocatalysis and Agricultural Biotechnology*, 47, 102551. <https://doi.org/https://doi.org/10.1016/j.bcab.2022.102551>

Ferdous, Umme Tamanna, & Yusof, Z. N. B. (2021a). Algal Terpenoids: A Potential Source of Antioxidants for Cancer Therapy. In *Terpenes and Terpenoids*. IntechOpen. <https://doi.org/10.5772/intechopen.94122>

Ferdous, Umme Tamanna, & Yusof, Z. N. B. (2021b). Insight into Potential Anticancer Activity of Algal Flavonoids : Current Status and Challenges. *Molecules*, 26, 6844. <https://doi.org/https://doi.org/10.3390/molecules26226844>

Ferdous, Umme Tamanna, & Yusof, Z. N. B. (2021c). Medicinal Prospects of Antioxidants From Algal Sources in Cancer Therapy. *Frontiers in Pharmacology*, 12(March), 1–22. <https://doi.org/10.3389/fphar.2021.593116>

Ferreira, R. M., Ribeiro, A. R., Patinha, C., Silva, A. M. S., Cardoso, S. M., & Costa, R. (2019). Water Extraction Kinetics of Bioactive Compounds of *Fucus vesiculosus*. *Molecules*, 24(18), 1–15. <https://doi.org/10.3390/molecules24183408>

FMI. (2022). *Microalgae Market Outlook (2022-2028)*. <https://doi.org/https://www.futuremarketinsights.com/reports/microalgae-market>

Foo, S. C., Yusoff, F. M., Imam, M. U., Foo, J. B., Ismail, N., Azmi, N. H., Tor, Y. S., Khong, N. M. H., & Ismail, M. (2018). Increased fucoxanthin in *Chaetoceros calcitrans* extract exacerbates apoptosis in liver cancer

- cells via multiple targeted cellular pathways. *Biotechnology Reports*, 20(e00296). <https://doi.org/10.1016/j.btre.2018.e00296>
- Foo, S. C., Yusoff, F. M., Ismail, M., Basri, M., Khong, N. M. H., Chan, K. W., & Yau, S. K. (2015). Efficient solvent extraction of antioxidant-rich extract from a tropical diatom, *Chaetoceros calcitrans* (Paulsen) Takano 1968. *Asian Pacific Journal of Tropical Biomedicine*, 5(10), 834–840. <https://doi.org/10.1016/j.apjtb.2015.06.003>
- Gargouch, N., Elleuch, F., Karkouch, I., Tabbene, O., Pichon, C., Gardarin, C., Rihouey, C., Picton, L., Abdelkafi, S., Fendri, I., & Laroche, C. (2021). Potential of exopolysaccharide from *porphyridium marinum* to contend with bacterial proliferation, biofilm formation, and breast cancer. *Marine Drugs*, 19(2). <https://doi.org/10.3390/md19020066>
- Gauthier, M. R., Senhorinho, G. N. A., Basiliko, N., Desjardins, S., & Scott, J. A. (2022). Green Photosynthetic Microalgae from Low pH Environments Associated with Mining as a Potential Source of Antioxidants. *Industrial Biotechnology*, 18(3), 168–175. <https://doi.org/10.1089/ind.2022.0013>
- George, B. P., Chandran, R., & Abrahamse, H. (2021). Role of phytochemicals in cancer chemoprevention: Insights. *Antioxidants*, 10(9). <https://doi.org/10.3390/antiox10091455>
- Glasauer, A., & Chandel, N. S. (2014). Targeting antioxidants for cancer therapy. *Biochemical Pharmacology*, 92(1), 90–101. <https://doi.org/10.1016/j.bcp.2014.07.017>
- GLOBOCAN. (2020). The Global Cancer Observatory - CANCER FACT SHEETS. In *International Agency for Research on Cancer - WHO* (Vol. 419). <https://gco.iarc.fr/today/home>
- Gnanakani, P. E., Santhanam, P., Kumar, K. E., & Dhanaraju, M. D. (2019). Chemical Composition, Antioxidant, and Cytotoxic Potential of *Nannochloropsis* Species Extracts. *Journal of Natural Science, Biology and Medicine*, 10, 167–177. <https://doi.org/10.4103/jnsbm.JNSBM>
- Go, Y., & Jones, D. P. (2017). Redox theory of aging : implications for health and disease. *Clinical Sciences*, 131, 1669–1688. <https://doi.org/10.1042/CS20160897>
- Goh, S.-H., Yusoff, F. M., & Loh, S. P. (2010). A Comparison of the Antioxidant Properties and Total Phenolic Content in a Diatom, *Chaetoceros* sp. and a Green Microalga, *Nannochloropsis* sp. *Journal of Agricultural Science*, 2(3). <https://doi.org/10.5539/jas.v2n3p123>
- Goh, S., Alitheen, N. B., Yusoff, F., Yap, S., & Loh, S. (2014). Crude ethyl acetate extract of marine microalga, *Chaetoceros calcitrans*, induces Apoptosis in MDA-MB-231 breast cancer cells. *Pharmacognosy Magazine*, 10(37), 1–8. <https://doi.org/10.4103/0973-1296.126650>
- Goiris, K., Muylaert, K., Fraeye, I., Foubert, I., De Brabanter, J., & De Cooman,



- L. (2012). Antioxidant potential of microalgae in relation to their phenolic and carotenoid content. *Journal of Applied Phycology*, 24(6), 1477–1486. <https://doi.org/10.1007/s10811-012-9804-6>
- Gong, M., & Bassi, A. (2016). Carotenoids from microalgae: A review of recent developments. *Biotechnology Advances*, 34(8), 1396–1412. <https://doi.org/10.1016/j.biotechadv.2016.10.005>
- Guo, M., Lu, B., Gan, J., Wang, S., Jiang, X., & Li, H. (2021). Apoptosis detection: a purpose-dependent approach selection. *Cell Cycle*, 20(11), 1033–1040. <https://doi.org/10.1080/15384101.2021.1919830>
- Gupta, P., Sinha, D., & Bandopadhyay, R. (2014). Isolation and screening of marine microalgae *Chlorella* sp. for anticancer activity. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(10), 517–519.
- Guzmán, F., Wong, G., Román, T., Cárdenas, C., Álvarez, C., Schmitt, P., Albericio, F., & Rojas, V. (2019). Identification of Antimicrobial Peptides from the Microalgae *Tetraselmis suecica* (Kylin) Butcher and Bactericidal Activity Improvement. *Marine Drugs*, 17(8), 1–19. <https://doi.org/10.3390/md17080453>
- Habibi Roudkenar, M., Mohammadi Roushandeh, A., Delazar, A., Halabian, R., Soleimani Rad, J., Mehdipour, A., Bagheri, M., & Jahanian-Najafabadi, A. (2011). Effects of polygonum aviculare herbal extract on proliferation and apoptotic gene expression of MCF-7. *DARU, Journal of Pharmaceutical Sciences*, 19(5), 326–331.
- Hachicha, R., Elleuch, F., Hlima, H. Ben, Dubessay, P., de Baynast, H., Delattre, C., Pierre, G., Hachicha, R., Abdelkafi, S., Michaud, P., & Fendri, I. (2022). Biomolecules from Microalgae and Cyanobacteria: Applications and Market Survey. *Applied Sciences*, 12(4). <https://doi.org/10.3390/app12041924>
- Hafsa, M. B. E. N., Ismail, M. B. E. N., & Garrab, M. (2017). Antimicrobial, antioxidant, cytotoxic and anticholinesterase activities of water-soluble polysaccharides extracted from microalgae *Isochrysis galbana* and *Nannochloropsis oculata*. *Journal of the Serbian Chemical Society*, 82(5), 509–522.
- Halliwell, B., Gutteridge, J. M. C., & Cross, C. E. (1992). Free radicals, antioxidants, and human disease: where are we now. *Journal of Laboratory and Clinical Medicine*, 119(6), 598–620.
- Hamdy, O., Karim, A., Farouk Gheda, S., Ismail, G. A., Abo-Shady, A. M., & Assistant Of Phycology, T. (2020). Phytochemical Screening and antioxidant activity of *Chlorella vulgaris*. *Delta Journal of Science*, 41, 81–91. <https://djs.journals.ekb.eg/>
- Hamidi, M., Safarzadeh Kozani, P., Safarzadeh Kozani, P., Pierre, G., Michaud, P., & Delattre, C. (2020). Marine bacteria versus microalgae: Who is the

best for biotechnological production of bioactive compounds with antioxidant properties and other biological applications? *Marine Drugs*, 18(1), 1–38. <https://doi.org/10.3390/md18010028>

- Hamouda Ali, I., & Doumandji, A. (2017). Comparative phytochemical analysis and in vitro antimicrobial activities of the cyanobacterium *Spirulina platensis* and the green alga *Chlorella pyrenoidosa*: Potential application of bioactive components as an alternative to infectious diseases. *Bulletin de l'Institut Scientifique, Section Sciences de La Terre*, 39(1), 41–49.
- Haoujar, I., Cacciola, F., Abrini, J., Mangraviti, D., Giu, D., Oulad, Y., Majdoub, E., Kounoun, A., Miceli, N., Taviano, M. F., Mondello, L., Rigano, F., & Senhaji, N. S. (2019). The Contribution of Carotenoids, Phenolic Compounds, and Flavonoids to the Antioxidative Properties of Marine Microalgae Isolated from Mediterranean Morocco. *Molecules*, 24, 4037.
- Hibberd, D. J. (1981). Notes on the taxonomy and nomenclature of the algal classes Eustigmatophyceae and Tribophyceae (synonym Xanthophyceae). *Botanical Journal of the Linnean Society*, 82(2), 93–119. <https://doi.org/10.1111/j.1095-8339.1981.tb00954.x>
- Hossain, F. M., Danos, D. M., Fu, Q., Wang, X., Scribner, R. A., Chu, S. T., Horswell, R. L., Price-Haywood, E. G., Collins-Burow, B. M., Wu, X. C., Ochoa, A. C., & Miele, L. (2022). Association of Obesity and Diabetes With the Incidence of Breast Cancer in Louisiana. *American Journal of Preventive Medicine*, 63(1), S83–S92. <https://doi.org/10.1016/j.amepre.2022.02.017>
- Hossain, N., Hasan, M. H., Mahlia, T. M. I., Shamsuddin, A. H., & Silitonga, A. S. (2020). Feasibility of microalgae as feedstock for alternative fuel in Malaysia: A review. *Energy Strategy Reviews*, 32, 100536. <https://doi.org/10.1016/j.esr.2020.100536>
- Hussein, H. A., & Abdullah, M. A. (2020). Anticancer Compounds Derived from Marine Diatoms. *Marine Drugs*, 18(7). <https://doi.org/10.3390/md18070356>
- Hussein, H. A., Mohamad, H., Ghazaly, M. M., Laith, A. A., & Abdullah, M. A. (2019). Cytotoxic effects of *Tetraselmis suecica* chloroform extracts with silver nanoparticle co-application on MCF-7, 4 T1, and Vero cell lines. *Journal of Applied Phycology*. <https://doi.org/10.1007/s10811-019-01905-7>
- Hussein, H. A., Mohamad, H., Mohd Ghazaly, M., Laith, A. A., & Abdullah, M. A. (2020). Anticancer and antioxidant activities of *Nannochloropsis oculata* and *Chlorella* sp. extracts in co-application with silver nanoparticle. *Journal of King Saud University - Science*, 32(8), 3486–3494. <https://doi.org/10.1016/j.jksus.2020.10.011>

- Hyung, J., Kim, E., Moon, S., Kang, N. S., & Park, J. (2021). *Tetraselmis jejuensis* sp. nov. (Chlorodendrophyceae), a Euryhaline Microalga Found in Supralittoral Tide Pools at Jeju Island, Korea. *Plants*, *10*, 1289.
- Id, C. M. W., Veale, R. B., & Mayne, E. S. (2022). *Inducing apoptosis using chemical treatment and acidic pH , and detecting it using the Annexin V flow cytometric assay*. 1–7. <https://doi.org/10.1371/journal.pone.0270599>
- Iglesias, M. J., Soengas, R., Probert, I., Guilloud, E., Gourvil, P., Mehiri, M., López, Y., Cepas, V., Gutiérrez-del-Río, I., Redondo-Blanco, S., Villar, C. J., Lombó, F., Soto, S., & Ortiz, F. L. (2019). NMR characterization and evaluation of antibacterial and antiobiofilm activity of organic extracts from stationary phase batch cultures of five marine microalgae (*Dunaliella* sp., *D. salina*, *Chaetoceros calcitrans*, *C. gracilis* and *Tisochrysis lutea*). *Phytochemistry*, *164*(April), 192–205. <https://doi.org/10.1016/j.phytochem.2019.05.001>
- Indrayanto, G., Putra, G. S., & Suhud, F. (2021). Validation of in-vitro bioassay methods: Application in herbal drug research. In *Prof. of Drug Substances, Excipients & Related Methodology* (1st ed., Vol. 46). Elsevier Inc. <https://doi.org/10.1016/bs.podrm.2020.07.005>
- Ishiguro, S., Robben, N., Burghart, R., Cote, P., Greenway, S., Thakkar, R., Upreti, D., Nakashima, A., & Suzuki, K. (2020). Cell Wall Membrane Fraction of *Chlorella sorokiniana* Enhances Host Antitumor Immunity and Inhibits Colon Carcinoma Growth in Mice. *Integrative Cancer Therapies*, *19*, 1–10. <https://doi.org/10.1177/1534735419900555>
- Ishika, T., Laird, D. W., Bahri, P. A., & Moheimani, N. R. (2019). Co-cultivation and stepwise cultivation of *Chaetoceros muelleri* and *Amphora* sp. for fucoxanthin production under gradual salinity increase. *Journal of Applied Phycology*, *31*(3), 1535–1544. <https://doi.org/10.1007/s10811-018-1718-5>
- Ismail, A., Prasad, K. N., Chew, L. Y., Khoo, H. E., Kong, K. W., & Azlan, A. (2010). Antioxidant capacities of peel, pulp, and seed fractions of canarium odontophyllum Miq. fruit. *Journal of Biomedicine and Biotechnology*, *2010*. <https://doi.org/10.1155/2010/871379>
- Jan, R., & Chaudhry, G.-S. (2019). Understanding Apoptosis and Apoptotic Pathways Targeted Cancer Therapeutics. *Advanced Pharmaceutical Bulletin*, *9*(2), 205–218. <https://doi.org/10.15171/apb.2019.024>
- Jane, E., Blair, K., Darmanin, S., & Muraglia, M. (2013). Totarol Content and Cytotoxicity Varies Significantly in Different Types of. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, *4*(3), 1047–1058.
- Jaspars, M., De Pascale, D., Andersen, J. H., Reyes, F., Crawford, A. D., & Ianora, A. (2016). The marine biodiscovery pipeline and ocean medicines of tomorrow. *Journal of the Marine Biological Association of*

*the United Kingdom*, 96(1), 151–158. <https://doi.org/10.1017/S0025315415002106>

- Jayappriyan, K. R., Rajkumar, R., Venkatakrisnan, V., Nagaraj, S., & Rengasamy, R. (2013). In vitro anticancer activity of natural  $\beta$ -carotene from *Dunaliella salina* EU5891199 in PC-3 cells. *Biomedicine and Preventive Nutrition*, 3(2), 99–105. <https://doi.org/10.1016/j.bionut.2012.08.003>
- Jayshree, A., Jayashree, S., & Thangaraju, N. (2016a). *Chlorella vulgaris* and *Chlamydomonas reinhardtii*: Effective antioxidant, antibacterial and anticancer mediators. *Indian Journal of Pharmaceutical Sciences*, 78(5), 575–581. <https://doi.org/10.4172/pharmaceutical-sciences.1000155>
- Jayshree, A., Jayashree, S., & Thangaraju, N. (2016b). *Chlorella vulgaris* and *Chlamydomonas reinhardtii*: Effective antioxidant, antibacterial and anticancer mediators. *Indian Journal of Pharmaceutical Sciences*, 78(5), 575–581. <https://doi.org/10.4172/pharmaceutical-sciences.1000155>
- Jerney, J., & Spilling, K. (2020). Large Scale Cultivation of Microalgae: Open and Closed Systems. *Methods in Molecular Biology*, 1980, 1–8. [https://doi.org/10.1007/7651\\_2018\\_130](https://doi.org/10.1007/7651_2018_130)
- Jiang, L., Wang, Y., Liu, G., Liu, H., Zhu, F., Ji, H., & Li, B. (2018). C-Phycocyanin exerts anti-cancer effects via the MAPK signaling pathway in MDA-MB-231 cells. *Cancer Cell International*, 18(1), 1–14. <https://doi.org/10.1186/s12935-018-0511-5>
- Jin, Y., Qiu, S., Shao, N., & Zheng, J. (2018). Fucoxanthin and tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) synergistically promotes apoptosis of human cervical cancer cells by targeting PI3K/Akt/NF- $\kappa$ B signaling pathway. *Medical Science Monitor*, 24, 11–18. <https://doi.org/10.12659/MSM.905360>
- Johnson, K. S., Conant, E. F., & Soo, M. S. (2021). Molecular Subtypes of Breast Cancer: A Review for Breast Radiologists. *Journal of Breast Imaging*, 3(1), 12–24. <https://doi.org/10.1093/jbi/wbaa110>
- Juin, C., Oliveira Junior, R. G. de, Fleury, A., Oudinet, C., Pytowski, L., Bérard, J. B., Nicolau, E., Thiéry, V., Lanneluc, I., Beugeard, L., Prunier, G., Almeida, J. R. G. D. S., & Picot, L. (2018). Zeaxanthin from *Porphyridium purpureum* induces apoptosis in human melanoma cells expressing the oncogenic BRAF V600E mutation and sensitizes them to the BRAF inhibitor vemurafenib. *Brazilian Journal of Pharmacognosy*, 28(4), 457–467. <https://doi.org/10.1016/j.bjp.2018.05.009>
- Jung, J. Y., Lee, H., Shin, W. S., Sung, M. G., Kwon, J. H., & Yang, J. W. (2015). Utilization of seawater for cost-effective cultivation and harvesting of *Scenedesmus obliquus*. *Bioprocess and Biosystems Engineering*, 38(3), 449–455. <https://doi.org/10.1007/s00449-014-1284-4>

- Kadar, N. A., Shaleh, S. R. M., & Ali, E. S. (2018). Molecular and phylogenetic identification of marine microalgae inferred by 18s rDNA gene. *Malaysian Applied Biology*, 47(6), 41–45.
- Kapinova, A., Stefanicka, P., Kubatka, P., Zubor, P., Uramova, S., Kello, M., Mojzis, J., Blahutova, D., Qaradakhi, T., Zulli, A., Caprnda, M., Danko, J., Lasabova, Z., Busselberg, D., & Kruzliak, P. (2017). Are plant-based functional foods better choice against cancer than single phytochemicals? A critical review of current breast cancer research. *Biomedicine and Pharmacotherapy*, 96(August), 1465–1477. <https://doi.org/10.1016/j.biopha.2017.11.134>
- Kapoor, B., Kapoor, D., Gautam, S., Singh, R., & Bhardwaj, S. (2021). Dietary Polyunsaturated Fatty Acids (PUFAs): Uses and Potential Health Benefits. *Current Nutrition Reports*, 10(3), 232–242. <https://doi.org/10.1007/s13668-021-00363-3>
- Katiyar, R., & Arora, A. (2020). Health promoting functional lipids from microalgae pool: A review. *Algal Research*, 46, 101800. <https://doi.org/10.1016/j.algal.2020.101800>
- Khalifa, S. A. M., Elias, N., Farag, M. A., Chen, L., Saeed, A., Hegazy, M. E. F., Moustafa, M. S., El-Wahed, A. A., Al-Mousawi, S. M., Musharraf, S. G., Chang, F. R., Iwasaki, A., Suenaga, K., Alajlani, M., Göransson, U., & El-Seedi, H. R. (2019). Marine natural products: A source of novel anticancer drugs. *Marine Drugs*, 17(9). <https://doi.org/10.3390/md17090491>
- Khan, H. Y., Hadi, S. M., Mohammad, R. M., & Azmi, A. S. (2020). Prooxidant anticancer activity of plant-derived polyphenolic compounds: An underappreciated phenomenon. In *Functional Foods in Cancer Prevention and Therapy*. Elsevier Inc. <https://doi.org/10.1016/b978-0-12-816151-7.00012-0>
- Khan, M. I., Shin, J. H., & Kim, J. D. (2018). The promising future of microalgae: Current status, challenges, and optimization of a sustainable and renewable industry for biofuels, feed, and other products. *Microbial Cell Factories*, 17(1), 1–21. <https://doi.org/10.1186/s12934-018-0879-x>
- Khurana, R. K., Jain, A., Jain, A., Sharma, T., Singh, B., & Kesharwani, P. (2018). Administration of antioxidants in cancer: debate of the decade. *Drug Discovery Today*, 23(4), 763–770. <https://doi.org/10.1016/j.drudis.2018.01.021>
- Kntayya, S. B., Ibrahim, M. D., Ain, N. M., Iori, R., Ioannides, C., & Razis, A. F. A. (2018). Induction of Apoptosis and Cytotoxicity by Isothiocyanate Sulforaphene in Human Hepatocarcinoma HepG2 Cells. *Nutrients*, 10, 718. <https://doi.org/10.3390/nu10060718>
- Ko, E. C., Formenti, S. C., & Formenti, S. C. (2019). Radiation therapy to enhance tumor immunotherapy: a novel application for an established

modality established modality. *International Journal of Radiation Biology*, 95(7), 936–939. <https://doi.org/10.1080/09553002.2019.1623429>

- Kroschinsky, F., Stölzel, F., von Bonin, S., Beutel, G., Kochanek, M., Kiehl, M., & Schellongowski, P. (2017). New drugs, new toxicities: Severe side effects of modern targeted and immunotherapy of cancer and their management. *Critical Care*, 21(1), 1–11. <https://doi.org/10.1186/s13054-017-1678-1>
- Krysko, O., Aaes, T. L., Kagan, V. E., D'Herde, K., Bachert, C., Leybaert, L., Vandenabeele, P., & Krysko, D. V. (2017). Necroptotic cell death in anti-cancer therapy. *Immunological Reviews*, 280(1), 207–219. <https://doi.org/10.1111/imr.12583>
- Kubatka, P., Kapinová, A., Kružliak, P., Kello, M., Výbohová, D., Kajo, K., Novák, M., Chripková, M., Adamkov, M., Pěč, M., Mojžiš, J., Bojková, B., Kassayová, M., Stolarová, N., & Dobrota, D. (2015). Antineoplastic effects of *Chlorella pyrenoidosa* in the breast cancer model. *Nutrition*, 31(4), 560–569. <https://doi.org/10.1016/j.nut.2014.08.010>
- Kultys, E., & Kurek, M. A. (2022). Green Extraction of Carotenoids from Fruit and Vegetable Byproducts: A Review. *Molecules*, 27, 518. <https://doi.org/10.3390/molecules27020518>
- Kumar, S. R., Hosokawa, M., & Miyashita, K. (2013). Fucoxanthin: A marine carotenoid exerting anti-cancer effects by affecting multiple mechanisms. *Marine Drugs*, 11(12), 5130–5147. <https://doi.org/10.3390/md11125130>
- Kunte, M., & Desai, K. (2018). The protein extract of *Chlorella minutissima* inhibits the expression of MMP-1, MMP-2 and MMP-9 in cancer cells through upregulation of TIMP-3 and down regulation of c-Jun. *Cell Journal*, 20(2), 211–219. <https://doi.org/10.22074/cellj.2018.5277>
- Kwan, Y. P., Saito, T., Ibrahim, D., Al-Hassan, F. M. S., Ein Oon, C., Chen, Y., Jothy, S. L., Kanwar, J. R., & Sasidharan, S. (2016). Evaluation of the cytotoxicity, cell-cycle arrest, and apoptotic induction by *Euphorbia hirta* in MCF-7 breast cancer cells. *Pharmaceutical Biology*, 54(7), 1223–1236. <https://doi.org/10.3109/13880209.2015.1064451>
- Lacouture, M., & Sibaud, V. (2018). Toxic Side Effects of Targeted Therapies and Immunotherapies Affecting the Skin, Oral Mucosa, Hair, and Nails. *American Journal of Clinical Dermatology*, 19(s1), 31–39. <https://doi.org/10.1007/s40257-018-0384-3>
- Lafarga, T. (2019). Effect of microalgal biomass incorporation into foods: Nutritional and sensorial attributes of the end products. *Algal Research*, 41(June), 101566. <https://doi.org/10.1016/j.algal.2019.101566>
- Lauritano, C., Andersen, J. H., Hansen, E., Albrigtsen, M., Escalera, L., Esposito, F., Helland, K., Hanssen, K., Romano, G., & Ianora, A. (2016).

- Bioactivity screening of microalgae for antioxidant, anti-inflammatory, anticancer, anti-diabetes, and antibacterial activities. *Frontiers in Marine Science*, 3(MAY), 1–2. <https://doi.org/10.3389/fmars.2016.00068>
- Lee, K. J., Oh, Y. C., Cho, W. K., & Ma, J. Y. (2015). Antioxidant and Anti-Inflammatory Activity Determination of One Hundred Kinds of Pure Chemical Compounds Using Offline and Online Screening HPLC Assay. *Evidence-Based Complementary and Alternative Medicine*, 2015. <https://doi.org/10.1155/2015/165457>
- Lee, S. H., Chang, D. U., Lee, B. J., & Jeon, Y. J. (2009). Antioxidant activity of solubilized tetraselmis suecica and chlorella ellipsoidea by enzymatic digests. *Journal of Food Science and Nutrition*, 14(1), 21–28. <https://doi.org/10.3746/jfn.2009.14.1.021>
- Lee, W. N., Ong, C. P., Khamis, A. S. M., Singaram, N., & Lee, S. H. (2021). Breast cancer awareness and knowledge assessment among men and women in Malaysia. *Journal of Public Health (Germany)*. <https://doi.org/10.1007/s10389-021-01509-x>
- Leuner, O., Havlik, J., Budesinsky, M., Vrkoslav, V., Chu, J., Bradshaw, T. D., Hummelova, J., Miksatkova, P., Lapcik, O., Valterova, I., & Kokoska, L. (2013). Cytotoxic Constituents of Pachyrhizus Tuberosus from Peruvian Amazon. *Natural Product Communications*, 8(10), 1–4. <https://doi.org/10.1177/1934578X1300801022>
- Li, Hua Bin, Cheng, K. W., Wong, C. C., Fan, K. W., Chen, F., & Jiang, Y. (2007). Evaluation of antioxidant capacity and total phenolic content of different fractions of selected microalgae. *Food Chemistry*, 102(3), 771–776. <https://doi.org/10.1016/j.foodchem.2006.06.022>
- Li, Haifeng, Su, L., Chen, S., Zhao, L., Wang, H., Ding, F., Chen, H., Shi, R., Wang, Y., & Huang, Z. (2018). Physicochemical characterization and functional analysis of the polysaccharide from the edible microalga nostoc sphaeroides. *Molecules*, 23(2). <https://doi.org/10.3390/molecules23020508>
- Li, Y., Li, S., Meng, X., Gan, R., Zhang, J., & Li, H. (2017). Dietary Natural Products for Prevention and Treatments of Breast Cancer. *Nutrients*, 9(728), 1–38. <https://doi.org/10.3390/nu9070728>
- Lin, P. Y., Tsai, C. T., Chuang, W. L., Chao, Y. H., Pan, I. H., Chen, Y. K., Lin, C. C., & Wang, B. Y. (2017). Chlorella sorokiniana induces mitochondrial-mediated apoptosis in human non-small cell lung cancer cells and inhibits xenograft tumor growth in vivo. *BMC Complementary and Alternative Medicine*, 17(1), 1–8. <https://doi.org/10.1186/s12906-017-1611-9>
- Lin, S., Li, M., Chuang, K., Lin, N., Chang, C., Wu, H., Chao, Y., Lin, C., Pan, I., Perng, M., & Wen, S. (2020). Chlorella sorokiniana Extract Prevents Cisplatin-Induced Myelotoxicity In Vitro and In Vivo. *Oxidative Medicine*

and *Cellular Longevity*, 2020. <https://doi.org/https://doi.org/10.1155/2020/7353618>

- Lindner, A. V., & Pleissner, D. (2019). Utilization of phenolic compounds by microalgae. *Algal Research*, 42(January), 101602. <https://doi.org/10.1016/j.algal.2019.101602>
- Loh, S. (2017). A Review of Cancer Awareness in Malaysia – What’s Next? *Open Access Journal of Cancer & Oncology*, 1(1), 1–6. <https://doi.org/10.23880/oajco-16000105>
- Lucakova, S., Branyikova, I., & Hayes, M. (2022). Microalgal Proteins and Bioactives for Food, Feed, and Other Applications. *Applied Sciences (Switzerland)*, 12(9). <https://doi.org/10.3390/app12094402>
- Łukasiewicz, S., Czeczelewski, M., Forma, A., Baj, J., Sitarz, R., & Stanisławek, A. (2021). Breast cancer—epidemiology, risk factors, classification, prognostic markers, and current treatment strategies—An updated review. *Cancers*, 13(17), 1–30. <https://doi.org/10.3390/cancers13174287>
- Ma, X. N., Chen, T. P., Yang, B., Liu, J., & Chen, F. (2016). Lipid production from *Nannochloropsis*. *Marine Drugs*, 14(4). <https://doi.org/10.3390/md14040061>
- Maadane, A., Merghoub, N., Ainane, T., El Arroussi, H., Benhima, R., Amzazi, S., Bakri, Y., & Wahby, I. (2015). Antioxidant activity of some Moroccan marine microalgae: Pufa profiles, carotenoids and phenolic content. *Journal of Biotechnology*, 215(January 2019), 13–19. <https://doi.org/10.1016/j.jbiotec.2015.06.400>
- Maligan, J. M., Saksony, A. K., & Widayanti, V. T. (2011). Identification Of Antioxidant and Antibacteria Activity of Marine Microalgae *Tetraselmis chuii* Extract. *5th Young Scientist Seminar, October(May)*, 25–26.
- Mantecón, L., Moyano, R., Cameán, A. M., & Jos, A. (2019). Safety assessment of a lyophilized biomass of *Tetraselmis chuii* (TetraSOD®) in a 90 day feeding study. *Food and Chemical Toxicology*, 133(July), 110810. <https://doi.org/10.1016/j.fct.2019.110810>
- Marino, T., Iovine, A., Casella, P., Martino, M., Chianese, S., Larocca, V., Musmarra, D., & Molino, A. (2020). From haematococcus pluvialis microalgae a powerful antioxidant for cosmetic applications. *Chemical Engineering Transactions*, 79(March 2020), 271–276. <https://doi.org/10.3303/CET2079046>
- Martin-Creuzburg, D., & Merkel, P. (2016). Sterols of freshwater microalgae: Potential implications for zooplankton nutrition. *Journal of Plankton Research*, 38(4), 865–877. <https://doi.org/10.1093/plankt/fbw034>
- Martin, L. J. (2015). Fucoxanthin and its metabolite fucoxanthinol in cancer prevention and treatment. *Marine Drugs*, 13(8), 4784–4798.



<https://doi.org/10.3390/md13084784>

- Martínez Andrade, K. A., Lauritano, C., Romano, G., & Ianora, A. (2018). Marine microalgae with anti-cancer properties. *Marine Drugs*, 16(165), 1–17. <https://doi.org/10.3390/md16050165>
- Matos, Â. P., Teixeira, M. S., Corrêa, F. M. P. S., Machado, M. M., Werner, R. I. S., Aguiar, A. C., Cubas, A. L. V., Sant'Anna, E. S., & Moecke, E. H. S. (2019). Disruption of nannochloropsis gaditana (Eustigmatophyceae) rigid cell wall by non-thermal plasma prior to lipid extraction and its effect on fatty acid composition. *Brazilian Journal of Chemical Engineering*, 36(4), 1419–1428. <https://doi.org/10.1590/0104-6632.20190364s20190097>
- Matos, J., Cardoso, C., Gomes, A., Campos, A. M., Falé, P., Afonso, C., & Bandarra, N. M. (2019). Bioprospection of: Isochrysis galbana and its potential as a nutraceutical. *Food and Function*, 10(11), 7333–7342. <https://doi.org/10.1039/c9fo01364d>
- Maynardo, J. J., Doshi, V., Rajanren, J. R., & Rajasekaran, R. (2015). the Optimization of Light Intensity and Drying Temperature on Lipid Content of Microalgae NANNOCHLOROPSIS OCULATA. *Journal of Engineering Science and Technology*, January, 112–121.
- Medina, E., Cerezal, P., Morales, J., & Ruiz-Domínguez, M. C. (2019). Fucoxanthin from marine microalga Isochrysis galbana: Optimization of extraction methods with organic solvents. *DYNA (Colombia)*, 86(210), 174–178. <https://doi.org/10.15446/dyna.v86n210.72932>
- Mehariya, S., Goswami, R. K., Karthikeyan, O. P., & Verma, P. (2021). Microalgae for high-value products: A way towards green nutraceutical and pharmaceutical compounds. *Chemosphere*, 280, 130553. <https://doi.org/10.1016/j.chemosphere.2021.130553>
- Mei, C. H., Zhou, S. C., Zhu, L., Ming, J. X., Zeng, F. D., & Xu, R. (2017). Antitumor effects of laminaria extract fucoxanthin on lung cancer. *Marine Drugs*, 15(2), 1–12. <https://doi.org/10.3390/md15020039>
- Meybodi, N. M., Mortazavian, A. M., Monfared, A. B., Sohrabvandi, S., & Meybodi, F. A. (2017). Phytochemicals in cancer prevention: A review of the evidence. *International Journal of Cancer Management*, 10(1). <https://doi.org/10.17795/ijcp-7219>
- Micol, V., Mateo, C. R., Shapiro, S., Aranda, F. J., & Villalaín, J. (2001). Effects of (+)-totarol, a diterpenoid antibacterial agent, on phospholipid model membranes. *Biochimica et Biophysica Acta - Biomembranes*, 1511(2), 281–290. [https://doi.org/10.1016/S0005-2736\(01\)00284-X](https://doi.org/10.1016/S0005-2736(01)00284-X)
- Mishra, A. P., Salehi, B., Sharifi-Rad, M., Pezzani, R., Kobarfard, F., Sharifi-Rad, J., & Nigam, M. (2018). Programmed Cell Death, from a Cancer Perspective: An Overview. *Molecular Diagnosis and Therapy*, 22(3), 281–295. <https://doi.org/10.1007/s40291-018-0329-9>

- Mishra, N., & Mishra, N. (2018). EXPLORING THE BIOLOGICALLY ACTIVE METABOLITES OF ISOCHRYSIS GALBANA IN PHARMACEUTICAL INTEREST: AN OVERVIEW. *International Journal for Pharmaceutical Sciences and Research*, 9(6), 2162–2174. [https://doi.org/10.13040/IJPSR.0975-8232.9\(6\).2162-74](https://doi.org/10.13040/IJPSR.0975-8232.9(6).2162-74)
- Mohamadnia, S., Tavakoli, O., Faramarzi, M. A., & Shamsollahi, Z. (2020). Production of fucoxanthin by the microalga *Tisochrysis lutea*: A review of recent developments. In *Aquaculture* (Vol. 516). Elsevier B.V. <https://doi.org/10.1016/j.aquaculture.2019.734637>
- Mohd Azamai, E. S., Sulaiman, S., Mohd Habib, S. H., Looi, M. L., Das, S., Abdul Hamid, N. A., Wan Ngah, W. Z., & Mohd Yusof, Y. A. (2009). *Chlorella vulgaris* triggers apoptosis in hepatocarcinogenesis-induced rats. *Journal of Zhejiang University: Science B*, 10(1), 14–21. <https://doi.org/10.1631/jzus.B0820168>
- Mohy Eldin Abd EL-Fattah Abd EL-Atty, M. M. A.-D. and B. S. E. M. (2019). Chemoprotective Effects of *Chlorella Vulgaris* and *Spirulina Platensis* on Colon Cancer Induced By 1, 2 Dimethylhydrazine. *International Journal of Current Research in Life Sciences*, 8(01), 3043–3049. [http://vipspublisher.com/ijcris.com/sites/default/files/issues-pdf/14877\\_0.pdf](http://vipspublisher.com/ijcris.com/sites/default/files/issues-pdf/14877_0.pdf)
- Momenimovahed, Z., & Salehiniya, H. (2019). Epidemiological characteristics of and risk factors for breast cancer in the world. *Breast Cancer: Targets and Therapy*, 11, 151–164. <https://doi.org/10.2147/BCTT.S176070>
- Monteiro, M., Santos, R. A., Iglesias, P., Couto, A., Serra, C. R., Gouvinhas, I., Barros, A., Oliva-Teles, A., Enes, P., & Díaz-Rosales, P. (2020). Effect of extraction method and solvent system on the phenolic content and antioxidant activity of selected macro- and microalgae extracts. *Journal of Applied Phycology*, 32(1), 349–362. <https://doi.org/10.1007/s10811-019-01927-1>
- Montone, C. M., Capriotti, A. L., Cavaliere, C., La Barbera, G., Piovesana, S., Zenezini Chiozzi, R., & Laganà, A. (2018). Peptidomic strategy for purification and identification of potential ACE-inhibitory and antioxidant peptides in *Tetrademus obliquus* microalgae. *Analytical and Bioanalytical Chemistry*, 410(15), 3573–3586. <https://doi.org/10.1007/s00216-018-0925-x>
- More, A. (2021). *Chlorella Market Size 2021 with CAGR of 7.3%, Top Growth Companies: FEMICO, Taiwan Chlorella, Vedan, and, End-User, SWOT Analysis in Industry 2026. MarketWatch*. <https://www.marketwatch.com/press-release/chlorella-market-size-2021-with-cagr-of-73-top-growth-companies-femico-taiwan-chlorella-vedan-and-end-user-swot-analysis-in-industry-2026-2021-06-14>
- Moreau, D., Tomasoni, C., Jacquot, C., Kaas, R., Le Guedes, R., Cadoret, J. P., Muller-Feuga, A., Kontiza, I., Vagias, C., Roussis, V., & Roussakis, C.

- (2006). Cultivated microalgae and the carotenoid fucoxanthin from *Odontella aurita* as potent anti-proliferative agents in bronchopulmonary and epithelial cell lines. *Environmental Toxicology and Pharmacology*, 22(1), 97–103. <https://doi.org/10.1016/j.etap.2006.01.004>
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65(1–2), 55–63. [https://doi.org/10.1016/0022-1759\(83\)90303-4](https://doi.org/10.1016/0022-1759(83)90303-4)
- Mtaki, K., Kyewalyanga, M. S., & Mtolera, M. S. P. (2020). Assessment of Antioxidant Contents and Free Radical-Scavenging Capacity of *Chlorella vulgaris* Cultivated in Low Cost Media. *Applied Sciences*, 10, 8611.
- Mut-Salud, N., Álvarez, P. J., Aránega, A., Garrido, J. M., Carrasco, E., & Rodríguez-Serrano, F. (2015). Antioxidant Intake and Antitumor therapy: toward nutritional recommendations for optimal results. *Oxidative Medicine and Cellular Longevity*, 2016, 1–19.
- Muthuirulappan, S., & Francis, S. P. (2013). Anti-cancer mechanism and possibility of nano-suspension formulation for a marine algae product fucoxanthin. *Asian Pacific Journal of Cancer Prevention*, 14(4), 2213–2216. <https://doi.org/10.7314/APJCP.2013.14.4.2213>
- Nagappan, S., Das, P., AbdulQuadir, M., Thaher, M., Khan, S., Mahata, C., Al-Jabri, H., Vatland, A. K., & Kumar, G. (2021). Potential of microalgae as a sustainable feed ingredient for aquaculture. *Journal of Biotechnology*, 341(August), 1–20. <https://doi.org/10.1016/j.jbiotec.2021.09.003>
- Natrah, F. M. I., Yusoff, F. M., Shariff, M., Abas, F., & Mariana, N. S. (2007). Screening of Malaysian indigenous microalgae for antioxidant properties and nutritional value. *Journal of Applied Phycology*, 19(6), 711–718. <https://doi.org/10.1007/s10811-007-9192-5>
- Nethravathy, M. U., Mehar, J. G., Mudliar, S. N., & Shekh, A. Y. (2019). Recent Advances in Microalgal Bioactives for Food, Feed, and Healthcare Products: Commercial Potential, Market Space, and Sustainability. *Comprehensive Reviews in Food Science and Food Safety*, 18(6), 1882–1897. <https://doi.org/10.1111/1541-4337.12500>
- Neumann, U., Derwenskus, F., Flister, V. F., Schmid-Staiger, U., Hirth, T., & Bischoff, S. C. (2019). Fucoxanthin, a carotenoid derived from *Phaeodactylum tricornutum* exerts antiproliferative and antioxidant activities in vitro. *Antioxidants*, 8(6), 1–11. <https://doi.org/10.3390/antiox8060183>
- Ngamdee, P., Wichai, U., & Jiamyangyuen, S. (2016). Correlation between phytochemical and mineral contents and antioxidant activity of black glutinous rice bran, and its potential chemopreventive property. *Food Technology and Biotechnology*, 54(3), 282–289. <https://doi.org/10.1016/j.fctb.2016.03.003>

- Nguyen, N. V. T., Duong, N. T., Nguyen, K. N. H., Bui, N. T., Pham, T. L. T., Nguyen, K. T., Le, P. H., & Kim, K. H. (2022). Effect of extraction solvent on total phenol, flavonoid content, and antioxidant activity of *avicennia officinalis*. *Biointerface Research in Applied Chemistry*, 12(2), 2678–2690. <https://doi.org/10.33263/BRIAC122.26782690>
- NIH. (2021). *Types of Cancer Treatment*. <https://www.cancer.gov/about-cancer/treatment/side-effects>
- Niki, E. (2016). Oxidative stress and antioxidants: Distress or eustress? *Archives of Biochemistry and Biophysics*, 595, 19–24. <https://doi.org/10.1016/j.abb.2015.11.017>
- Nunes, M. C., Fernandes, I., Vasco, I., Sousa, I., & Raymundo, A. (2020). Tetraselmis chuii as a sustainable and healthy ingredient to produce gluten-free bread: Impact on structure, colour and bioactivity. *Foods*, 9(5), 1–15. <https://doi.org/10.3390/foods9050579>
- Olasehinde, T. A., Odjadjare, E. C., Mabinya, L. V., Olaniran, A. O., & Okoh, A. I. (2019). Chlorella sorokiniana and Chlorella minutissima exhibit antioxidant potentials, inhibit cholinesterases and modulate disaggregation of  $\beta$ -amyloid fibrils. *Electronic Journal of Biotechnology*, 40, 1–9. <https://doi.org/10.1016/j.ejbt.2019.03.008>
- Oslan, S. N. H., Tan, J. S., Oslan, S. N., Matanjun, P., Mokhtar, R. A. M., Shapawi, R., & Huda, N. (2021). Haematococcus pluvialis as a potential source of astaxanthin with diverse applications in industrial sectors: Current research and future directions. *Molecules*, 26(21). <https://doi.org/10.3390/molecules26216470>
- Ou, A., Zhao, X., & Lu, Z. (2022). The potential roles of p53 signaling reactivation in pancreatic cancer therapy. *BBA - Reviews on Cancer*, 1877(1), 188662. <https://doi.org/10.1016/j.bbcan.2021.188662>
- Oun, R., Moussa, Y., & Wheate, N. (2018). The side effects of platinum-based chemotherapy drugs: a review for chemists. *Dalton Transactions*, 1–29. <https://doi.org/10.1039/C8DT00838H>
- Padanilam, B. J. (2003). Cell death induced by acute renal injury: A perspective on the contributions of apoptosis and necrosis. *American Journal of Physiology - Renal Physiology*, 284(4 53-4), 608–627. <https://doi.org/10.1152/ajprenal.00284.2002>
- Pagels, F., Guedes, A. C., Amaro, H. M., Kijjoa, A., & Vasconcelos, V. (2019). Phycobiliproteins from cyanobacteria: Chemistry and biotechnological applications. *Biotechnology Advances*, 37(3), 422–443. <https://doi.org/10.1016/j.biotechadv.2019.02.010>
- Pal, A., Verma, P., Paul, S., Majumder, I., & Kundu, R. (2021). Two species of *Ulva* inhibits the progression of cervical cancer cells SiHa by means of

- autophagic cell death induction. *3 Biotech*, *11*(2), 1–11. <https://doi.org/10.1007/s13205-020-02576-9>
- Pan, Z., Zhang, X., Yu, P., Chen, X., Lu, P., Li, M., Liu, X., Li, Z., Wei, F., Wang, K., Zheng, Q., & Li, D. (2019). Cinobufagin Induces Cell Cycle Arrest at the G2/M Phase and Promotes Apoptosis in Malignant Melanoma Cells. *Frontiers in Oncology*, *9*(September), 1–11. <https://doi.org/10.3389/fonc.2019.00853>
- Panda, S. K., Ray, S., Nayak, S. R., Behera, S., Bhanja, S. S., & Acharya, V. (2020). A Review on Cell Cycle Checkpoints in Relation to Cancer. *The Journal of Medical Sciences*, *5*(4), 88–95. <https://doi.org/10.5005/jp-journals-10045-00138>
- Pang, J. R., How, S. W., Wong, K. H., Lim, S. H., Phang, S. M., & Yow, Y. Y. (2022). Cholinesterase inhibitory activities of neuroprotective fraction derived from red alga *Gracilaria manilaensis*. *Fisheries and Aquatic Sciences*, *25*(2), 49–63. <https://doi.org/10.47853/FAS.2022.e6>
- Pantami, H. A., Bustamam, M. S. A., Lee, S. Y., Ismail, I. S., Faudzi, S. M. M., Nakakuni, M., & Shaari, K. (2020). Comprehensive GCMS and LC-MS/MS metabolite profiling of *Chlorella vulgaris*. *Marine Drugs*, *18*(7). <https://doi.org/10.3390/MD18070367>
- Park, M., & Kim, M. (2017). Analysis of Antioxidant and Anti-Inflammatory Activities of Solvent Fractions from *Rhynchosia nulubilis* Cultivated with *Ganoderma lucidum* Mycelium. *Preventive Nutrition and Food Science*, *22*(4), 365–371. <https://doi.org/10.3746/pnf.2017.22.4.365>
- Parra-Riofrío, G., García-Márquez, J., Casas-Arrojo, V., Uribe-Tapia, E., & Abdala-Díaz, R. T. (2020). Antioxidant and Cytotoxic Effects on Tumor Cells of Exopolysaccharides From *Tetraselmis suecica* (Kylin) Butcher Grown Under Autotrophic and Heterotrophic Conditions. *Marine Drugs*, *18*(11), 1–23. <https://doi.org/10.3390/md18110534>
- Pasquet, V., Morisset, P., Ihammouine, S., Chepied, A., Aumailley, L., Berard, J. B., Serive, B., Kaas, R., Lanneluc, I., Thiery, V., Lafferriere, M., Piot, J. M., Patrice, T., Cadoret, J. P., & Picot, L. (2011). Antiproliferative activity of violaxanthin isolated from bioguided fractionation of *Dunaliella tertiolecta* extracts. *Marine Drugs*, *9*(5), 819–831. <https://doi.org/10.3390/md9050819>
- Patras, D., Moraru, C. V., & Socaciu, C. (2018). Screening of bioactive compounds synthesized by microalgae: A progress overview on extraction and chemical analysis. *Studia Universitatis Babeş-Bolyai Chemia*, *63*(1), 21–35. <https://doi.org/10.24193/subbchem.2018.1.02>
- Peña-Blanco, A., & García-Sáez, A. J. (2018). Bax, Bak and beyond: mitochondrial performance in apoptosis. *The FEBS Journal*, *285*(3), 416–431. <https://doi.org/10.1111/ijlh.12426>

- Perou, C. M., Sørile, T., Eisen, M. B., Van De Rijn, M., Jeffrey, S. S., Renshaw, A. A., Pollack, J. R., Ross, D. T., Johnsen, H., Akslen, L. A., Fluge, Ø., Pergammenschlkov, A., Williams, C., Zhu, S. X., Lønning, P. E., Børresen-Dale, A. L., Brown, P. O., & Botstein, D. (2000). Molecular portraits of human breast tumours. *Nature*, *406*(6797), 747–752. <https://doi.org/10.1038/35021093>
- Pfeffer, C. M., & Singh, A. T. K. (2018). Apoptosis: A target for anticancer therapy. *International Journal of Molecular Sciences*, *19*(2). <https://doi.org/10.3390/ijms19020448>
- Phang, S. M., Mustafa, E. M., Ambati, R. R., Sulaiman, N. M. N., Lim, P. E., Majid, N. A., Dommange, X., Schwob, C., & Liew, K. E. (2015). Checklist of microalgae collected from different habitats in Peninsular Malaysia for selection of algal biofuel feedstocks. *Malaysian Journal of Science*, *34*(2), 141–167. <https://doi.org/10.22452/mjs.vol34no2.2>
- Pradhan, B., Patra, S., Dash, S. R., Nayak, R., Behera, C., & Jena, M. (2021). Evaluation of the anti-bacterial activity of methanolic extract of *Chlorella vulgaris* Beyerinck [Beijerinck] with special reference to antioxidant modulation. *Future Journal of Pharmaceutical Sciences*, *7*, 17. <https://doi.org/10.1186/s43094-020-00172-5>
- Pumas, P., & Pumas, C. (2014). Proximate composition, total phenolics content and antioxidant activities of microalgal residue from biodiesel production. *Maejo International Journal of Science and Technology*, *8*(2), 122–128. <https://doi.org/10.14456/mijst.2014.11>
- Qamar, H., Hussain, K., Soni, A., Khan, A., Hussain, T., & Chénais, B. (2021). Cyanobacteria as Natural Therapeutics and Pharmaceutical Potential: Role in Antitumor Activity and as Nanovectors. *Molecules*, *26*(1), 1–30. <https://doi.org/10.3390/MOLECULES26010247>
- Raikar, S. M., Kalebar, V. U., & Adhoni, S. A. (2018). Screening of Pharmacological and Cytotoxic Activities of Fresh Water Lake Isolated Microalgae *Chlorella vulgaris* As-13 and *Chlorella pyrenoidosa* AS-6. *International Journal of Bio-Technology and Research*, *8*(4), 1–8.
- Rajkumar, R., Yaakob, Z., & Takriff, M. S. (2014). Potential of the micro and macro algae for biofuel production: A brief review. *BioResources*, *9*(1), 1606–1633. <https://doi.org/10.15376/biores.9.1.1606-1633>
- Ramos, A. L., Torello, C. O., & Queiroz, M. L. S. (2010). *Chlorella vulgaris* modulates immunomyelopoietic activity and enhances the resistance of tumor-bearing mice. *Nutrition and Cancer*, *62*(8), 1170–1180. <https://doi.org/10.1080/01635581.2010.513801>
- Randhir, A., Laird, D. W., Maker, G., Trengove, R., & Moheimani, N. R. (2020). Microalgae: A potential sustainable commercial source of sterols. *Algal Research*, *46*(July 2019), 101772. <https://doi.org/10.1016/j.algal.2019.101772>

- Rani, K., Sandal, N., & Sahoo, P. K. (2018). A comprehensive review on chlorella- its composition , health benefits , market and regulatory scenario. *The Pharma Innovation*, 7(7), 584–589.
- Rasheed, R., Saadaoui, I., Bounnit, T., Cherif, M., Ghazal, G. Al, & Jabri, H. Al. (2020). Sustainable Food Production and Nutraceutical Applications from Qatar Desert. *Animals*, 10(8), 1413.
- Renju, G. L., Kurup, G. M., & Bandugula, V. R. (2014). Effect of lycopene isolated from *Chlorella marina* on proliferation and apoptosis in human prostate cancer cell line PC-3. *Tumor Biol.* <https://doi.org/10.1007/s13277-014-2339-5>
- Reyna-Martinez, R., Gomez-Flores, R., López-Chuken, U., Quintanilla-Licea, R., Caballero-Hernandez, D., Rodríguez-Padilla, C., Beltrán-Rocha, J. C., & Tamez-Guerra, P. (2018). Antitumor activity of *Chlorella sorokiniana* and *Scenedesmus* sp. microalgae native of Nuevo León State, México. *PeerJ*, 2018(2), 1–15. <https://doi.org/10.7717/peerj.4358>
- Rijstenbil, J. W. (2003). Effects of UVB radiation and salt stress on growth, pigments and antioxidative defence of the marine diatom *Cylindrotheca closterium*. *Marine Ecology Progress Series*, 254(June 2003), 37–48. <https://doi.org/10.3354/meps254037>
- Robles-Bañuelos, B., Durán-Riveroll, L. M., Rangel-López, E., Pérez-López, H. I., & González-Maya, L. (2022). Marine Cyanobacteria as Sources of Lead Anticancer Compounds: A Review of Families of Metabolites with Cytotoxic, Antiproliferative, and Antineoplastic Effects. *Molecules*, 27(15). <https://doi.org/10.3390/molecules27154814>
- Rock, K. L., & Kono, H. (2008). The inflammatory response to cell death. *Annual Review of Pathology-Mechanisms of Disease*, 3, 67–97. <https://doi.org/10.1146/annurev.path>
- Rokicka, M., Zieliński, M., Dudek, M., & Dębowski, M. (2021). Effects of Ultrasonic and Microwave Pretreatment on Lipid Extraction of Microalgae and Methane Production from the Residual Extracted Biomass. *Bioenergy Research*, 14(3), 752–760. <https://doi.org/10.1007/s12155-020-10202-y>
- Ruiz-Torres, V., Encinar, J. A., Herranz-López, M., Pérez-Sánchez, A., Galiano, V., Barrajón-Catalán, E., & Micol, V. (2017). An updated review on marine anticancer compounds: The use of virtual screening for the discovery of small-molecule cancer drugs. *Molecules*, 22(7). <https://doi.org/10.3390/molecules22071037>
- Ryckebosch, E., Bruneel, C., Muylaert, K., & Foubert, I. (2012). Microalgae as an alternative source of omega-3 long chain polyunsaturated fatty acids. *Lipid Technology*, 24(6), 128–130. <https://doi.org/10.1002/lite.201200197>

- Sadeer, N. B., Montesano, D., Albrizio, S., Zengin, G., & Mahomoodally, M. F. (2020). The versatility of antioxidant assays in food science and safety—chemistry, applications, strengths, and limitations. *Antioxidants*, 9(8), 1–39. <https://doi.org/10.3390/antiox9080709>
- Saeed, A. F. U. H., Su, J., & Ouyang, S. (2021). Marine-derived drugs: Recent advances in cancer therapy and immune signaling. *Biomedicine and Pharmacotherapy*, 134(November 2020), 111091. <https://doi.org/10.1016/j.biopha.2020.111091>
- Safafar, H., Wagenen, J. Van, Møller, P., & Jacobsen, C. (2015). Carotenoids, phenolic compounds and tocopherols contribute to the antioxidative properties of some microalgae species grown on industrial wastewater. *Marine Drugs*, 13(12), 7339–7356. <https://doi.org/10.3390/md13127069>
- Sanjeeva, K. K. A., Fernando, I. P. S., Samarakoon, K. W., Lakmal, H. H. C., Kim, E. A., Kwon, O. N., Dilshara, M. G., Lee, J. B., & Jeon, Y. J. (2016). Anti-inflammatory and anti-cancer activities of sterol rich fraction of cultured marine microalga nannochloropsis oculata. *Algae*, 31(3), 277–287. <https://doi.org/10.4490/algae.2016.31.6.29>
- Sansone, C., & Brunet, C. (2019). Promises and Challenges of Microalgal Antioxidant Production. *Antioxidants*, 8(7), 199. <https://doi.org/10.3390/antiox8070199>
- Sansone, C., Galasso, C., Orefice, I., Nuzzo, G., Luongo, E., Cutignano, A., Romano, G., Brunet, C., Fontana, A., Esposito, F., & Ianora, A. (2017). The green microalga *Tetraselmis suecica* reduces oxidative stress and induces repairing mechanisms in human cells. *Scientific Reports*, 7(December 2015), 1–12. <https://doi.org/10.1038/srep41215>
- Saranya, C., Hemalatha, A., Parthiban, C., & Anantharaman., P. (2014). Evaluation of Antioxidant Properties, Total Phenolic and Carotenoid Content of *Chaetoceros calcitrans*, *Chlorella salina* and *Isochrysis galbana*. *International Journal of Current Microbiology and Applied Sciences*, 3(8), 365–377. <https://doi.org/10.1108/09574099710805673>
- Sayegh, F., Elazzazy, A., Bellou, S., Moustogianni, A., Elkady, A. I., Baeshen, M. N., & Aggelis, G. (2016a). Production of polyunsaturated single cell oils possessing antimicrobial and anticancer properties. *Annals of Microbiology*, 66(3), 937–948. <https://doi.org/10.1007/s13213-015-1176-0>
- Sayegh, F., Elazzazy, A., Bellou, S., Moustogianni, A., Elkady, A. I., Baeshen, M. N., & Aggelis, G. (2016b). Production of polyunsaturated single cell oils possessing antimicrobial and anticancer properties. *Annals of Microbiology*, 66(3), 937–948. <https://doi.org/10.1007/s13213-015-1176-0>
- Senousy, H. H., Ellatif, S. A., & Ali, S. (2020). Assessment of the antioxidant and anticancer potential of different isolated strains of cyanobacteria and



microalgae from soil and agriculture drain water. *Environmental Science and Pollution Research*. <https://doi.org/https://doi.org/10.1007/s11356-020-08332-z>

- Shanab, S. M. M., Mostafa, S. S. M., Shalaby, E. A., & Mahmoud, G. I. (2012). Aqueous extracts of microalgae exhibit antioxidant and anticancer activities. *Asian Pacific Journal of Tropical Biomedicine*, 2(8), 608–615. [https://doi.org/10.1016/S2221-1691\(12\)60106-3](https://doi.org/10.1016/S2221-1691(12)60106-3)
- Sharifi, S., Barar, J., Hejazi, M. S., & Samadi, N. (2014). Roles of the Bcl-2/Bax ratio, caspase-8 and 9 in resistance of breast cancer cells to paclitaxel. *Asian Pacific Journal of Cancer Prevention*, 15(20), 8617–8622. <https://doi.org/10.7314/APJCP.2014.15.20.8617>
- Shin, J., Song, M. H., Oh, J. W., Keum, Y. S., & Saini, R. K. (2020). Pro-oxidant actions of carotenoids in triggering apoptosis of cancer cells: A review of emerging evidence. *Antioxidants*, 9(6), 1–17. <https://doi.org/10.3390/antiox9060532>
- Shishodia, S., Sethi, G., Ahn, K. S., & Aggarwal, B. B. (2007). Guggulsterone inhibits tumor cell proliferation, induces S-phase arrest, and promotes apoptosis through activation of c-Jun N-terminal kinase, suppression of Akt pathway, and downregulation of antiapoptotic gene products. *Biochemical Pharmacology*, 74(1), 118–130. <https://doi.org/10.1016/j.bcp.2007.03.026>
- Shrihastini, V., Muthuramalingam, P., Adarshan, S., Sujitha, M., Chen, J. T., Shin, H., & Ramesh, M. (2021). Plant derived bioactive compounds, their anti-cancer effects and in silico approaches as an alternative target treatment strategy for breast cancer: An updated overview. *Cancers*, 13(24). <https://doi.org/10.3390/cancers13246222>
- Sies, H. (2020). Oxidative eustress and oxidative distress: Introductory remarks. In *Oxidative Stress*. Elsevier Inc. <https://doi.org/10.1016/b978-0-12-818606-0.00001-8>
- Sigamani, S., Jayaraj, P., Balaji, R., Ramamurthy, D., & Natarajan, H. (2019). Antiproliferative activity of the *Chlorella* sp., SRD3 crude extracts against MCF-7 and Hep2 cell lines. *International Journal of Life Sciences Research*, 7(2), 145–150.
- Singh, J., & Saxena, R. C. (2015). An Introduction to Microalgae: Diversity and Significance. Diversity and Significance. In *Handbook of Marine Microalgae: Biotechnology Advances* (pp. 11–24). <https://doi.org/10.1016/B978-0-12-800776-1.00002-9>
- Singh, S., Sharma, B., Kanwar, S. S., & Kumar, A. (2016). Lead phytochemicals for anticancer drug development. *Frontiers in Plant Science*, 7(November 2016), 1–13. <https://doi.org/10.3389/fpls.2016.01667>

- Sirohi, P., Verma, H., Singh, S. K., Singh, V. K., Pandey, J., Khusharia, S., Kumar, D., Kaushalendra, Teotia, P., & Kumar, A. (2022). Microalgal Carotenoids: Therapeutic Application and Latest Approaches to Enhance the Production. *Current Issues in Molecular Biology*, 44(12), 6257–6279. <https://doi.org/10.3390/cimb44120427>
- Sithranga Boopathy, N., & Kathiresan, K. (2010). Anticancer drugs from marine flora: An overview. *Journal of Oncology*, 2010(214186), 1–18. <https://doi.org/10.1155/2010/214186>
- Soto-Sierra, L., Stoykova, P., & Nikolov, Z. L. (2018). Extraction and fractionation of microalgae-based protein products. *Algal Research*, 36(October), 175–192. <https://doi.org/10.1016/j.algal.2018.10.023>
- Spoerri, L., Oo, Z. Y., Larsen, J. E., Haass, N. K., Gabrielli, B., & Pavey, S. (2015). Stress response pathways in cancer: From molecular targets to novel therapeutics. In *Stress Response Pathways in Cancer* (pp. 29–49). Springer. <https://doi.org/10.1007/978-94-017-9421-3>
- Sun, L., Wang, L., Li, J., & Liu, H. (2014). Characterization and antioxidant activities of degraded polysaccharides from two marine Chrysophyta. *Food Chemistry*, 160, 1–7. <https://doi.org/10.1016/j.foodchem.2014.03.067>
- Sun, Y., Wang, H., Guo, G., Pu, Y., & Yan, B. (2014). The isolation and antioxidant activity of polysaccharides from the marine microalgae *Isochrysis galbana*. *Carbohydrate Polymers*, 113, 22–31. <https://doi.org/10.1016/j.carbpol.2014.06.058>
- Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., & Bray, F. (2021). Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA: A Cancer Journal for Clinicians*, 71(3), 209–249. <https://doi.org/10.3322/caac.21660>
- Surai, P. F., Karadas, F., & Sparks, N. H. (2003). The importance of antioxidants in poultry. *Nineteenth Annual Carolina Poultry Conference*.
- Swamy, M. K., Arumugam, G., Kaur, R., Ghasemzadeh, A., Yusoff, M. M., & Sinniah, U. R. (2017). GC-MS Based Metabolite Profiling, Antioxidant and Antimicrobial Properties of Different Solvent Extracts of Malaysian *Plectranthus amboinicus* Leaves. *Evidence-Based Complementary and Alternative Medicine*, 2017. <https://doi.org/10.1155/2017/1517683>
- Syahril, M. . M., Roshani, O., Hasyimah, R. N., Hafiz, M. . M., Sharida, M. ., & Ahmed, H. . (2011). Screening of Anticancer Activities of Crude Extracts of Unicellular Green Algae (*Chlorella vulgaris*) and Filamentous Blue Green Algae (*Spirulina platensis*) on Selected Cancer Cell Lines. *International Conference on Applied Sciences, Mathematics and Humanities*, 82–87.

- Syed Abdul Rahman, S. N., Abdul Wahab, N., & Abd Malek, S. N. (2013). In vitro morphological assessment of apoptosis induced by antiproliferative constituents from the rhizomes of curcuma zedoaria. *Evidence-Based Complementary and Alternative Medicine*, 2013. <https://doi.org/10.1155/2013/257108>
- Talero, E., García-Mauriño, S., Ávila-Román, J., Rodríguez-Luna, A., Alcaide, A., & Motilva, V. (2015). Bioactive compounds isolated from microalgae in chronic inflammation and cancer. *Marine Drugs*, 13(10), 6152–6209. <https://doi.org/10.3390/md13106152>
- Tan, M. M., Ho, W. K., Yoon, S. Y., Mariapun, S., Hasan, S. N., Shin-Chi Lee, D., Hassan, T., Lee, S. Y., Phuah, S. Y., Sivanandan, K., Pei-Sze Ng, P., Rajaram, N., Jaganathan, M., Jamaris, S., Islam, T., Rahmat, K., Fadzli, F., Vijayanathan, A., Rajadurai, P., ... Teo, S. H. (2018). A case-control study of breast cancer risk factors in 7,663 women in Malaysia. *PLoS ONE*, 13(9), 1–12. <https://doi.org/10.1371/journal.pone.0203469>
- Tan, S. P., Parks, S. E., Stathopoulos, C. E., & Roach, P. D. (2014). Extraction of Flavonoids from Bitter Melon. *Food and Nutrition Sciences*, 05(05), 458–465. <https://doi.org/10.4236/fns.2014.55054>
- Tan, Y., Chen, Q., Li, X., Zeng, Z., Xiong, W., Li, G., Li, X., Yang, J., Xiang, B., & Yi, M. (2021). Pyroptosis: a new paradigm of cell death for fighting against cancer. *Journal of Experimental and Clinical Cancer Research*, 40(1), 1–15. <https://doi.org/10.1186/s13046-021-01959-x>
- Tannin-Spitz, T., Bergman, M., Van-Moppes, D., Grossman, S., & Arad, S. (2005). Antioxidant activity of the polysaccharide of the red microalga *Porphyridium* sp. *Journal of Applied Phycology*, 17(3), 215–222. <https://doi.org/10.1007/s10811-005-0679-7>
- Teh, H. S., & Woon, Y. L. (2021). Burden of cancers attributable to modifiable risk factors in Malaysia. *BMC Public Health*, 21(1), 1–10. <https://doi.org/10.1186/s12889-021-10412-9>
- Thavamoney, N., Sivanadian, L., Tee, L. H., Khoo, H. E., Prasad, K. N., & Kong, K. W. (2018). Extraction and recovery of phytochemical components and antioxidative properties in fruit parts of *Dacryodes rostrata* influenced by different solvents. *Journal of Food Science and Technology*, 55(7), 2523–2532. <https://doi.org/10.1007/s13197-018-3170-6>
- The Global Cancer Observatory. (2020). *Malaysia Globocan 2020*. International Agent for Research on Cancer - WHO. <https://gco.iarc.fr/today/data/factsheets/populations/458-malaysia-factsheets.pdf>
- Tiong, I. K. R., Nagappan, T., Abdul Wahid, M. E., Tengku Muhammad, T. S., Tatsuki, T., Satyantini, W. H., Mahasri, G., Sorgeloos, P., & Sung, Y. Y.

- (2020). Antioxidant capacity of five microalgae species and their effect on heat shock protein 70 expression in the brine shrimp *Artemia*. *Aquaculture Reports*, 18(May). <https://doi.org/10.1016/j.aqrep.2020.100433>
- Trentin, R., Custódio, L., Rodrigues, M. J., Moschin, E., Sciuto, K., da Silva, J. P., & Moro, I. (2022). Total Phenolic Levels, In Vitro Antioxidant Properties, and Fatty Acid Profile of Two Microalgae, *Tetraselmis marina* Strain IMA043 and Naviculoid Diatom Strain IMA053, Isolated from the North Adriatic Sea. *Marine Drugs*, 20, 207. <https://doi.org/10.3390/md20030207>
- Trung, T. S., Huyen, N. T. K., Minh, N. C., Le Trang, T. T., & Han, N. T. (2016). Optimization of harvesting of microalgal *Thalassiosira pseudonana* biomass using chitosan prepared from shrimp shell waste. *Asian Journal of Agricultural Research*, 10(5), 162–174. <https://doi.org/10.3923/ajar.2016.162.174>
- Truong, D. H., Nguyen, D. H., Ta, N. T. A., Bui, A. V., Do, T. H., & Nguyen, H. C. (2019). Evaluation of the use of different solvents for phytochemical constituents, antioxidants, and in vitro anti-inflammatory activities of *Severinia buxifolia*. *Journal of Food Quality*, 2019. <https://doi.org/10.1155/2019/8178294>
- Udayan, A., Pandey, A. K., Sharma, P., Sreekumar, N., & Kumar, S. (2021). Emerging industrial applications of microalgae: challenges and future perspectives. *Systems Microbiology and Biomanufacturing*, 1(4), 411–431. <https://doi.org/10.1007/s43393-021-00038-8>
- Udayan, A., Sirohi, R., Sreekumar, N., Sang, B. I., & Sim, S. J. (2022). Mass cultivation and harvesting of microalgal biomass: Current trends and future perspectives. *Bioresource Technology*, 344(PB), 126406. <https://doi.org/10.1016/j.biortech.2021.126406>
- Ummalyma, S. B., Sukumaran, R. K., & Pandey, A. (2020). Evaluation of Freshwater Microalgal Isolates for Growth and Oil Production in Seawater Medium. *Waste and Biomass Valorization*, 11(1), 223–230. <https://doi.org/10.1007/s12649-018-0393-8>
- Vander Wiel, J. B., Mikulicz, J. D., Boysen, M. R., Hashemi, N., Kalgren, P., Nauman, L. M., Baetzold, S. J., Powell, G. G., He, Q., & Hashemi, N. N. (2017). Characterization of *Chlorella vulgaris* and *Chlorella protothecoides* using multi-pixel photon counters in a 3D focusing optofluidic system. *RSC Advances*, 7(8), 4402–4408. <https://doi.org/10.1039/C6RA25837A>
- Veetil, S. K., Lim, K. G., Chaiyakunapruk, N., Ching, S. M., & Abu Hassan, M. R. (2017). Colorectal cancer in Malaysia: Its burden and implications for a multiethnic country. *Asian Journal of Surgery*, 40(6), 481–489. <https://doi.org/10.1016/j.asjsur.2016.07.005>

- Velasco, L. A., Carrera, S., & Barros, J. (2016). Aislamiento, cultivo y evaluación de *Chaetoceros muelleri* del Caribe como alimento para los pectínidos nativos, *Argopecten nucleus* y *Nodipecten nodosus*. *Latin American Journal of Aquatic Research*, 44(3), 557–568. <https://doi.org/10.3856/vol44-issue3-fulltext-14>
- Verissimo, A. C. S., Pacheco, M., Silva, A. M. S., & Pinto, D. C. G. A. (2021). Secondary metabolites from marine sources with potential use as leads for anticancer applications. *Molecules*, 26(14), 1–13. <https://doi.org/10.3390/molecules26144292>
- Vilakazi, H., Olasehinde, T. A., & Olaniran, A. O. (2021). Chemical characterization, antiproliferative and antioxidant activities of polyunsaturated fatty acid-rich extracts from *Chlorella* sp. s14. *Molecules*, 26(14), 1–13. <https://doi.org/10.3390/molecules26144109>
- Vona, D., Urbano, L., Bonifacio, M. A., De Giglio, E., Cometa, S., Mattioli-Belmonte, M., Palumbo, F., Ragni, R., Cicco, S. R., & Farinola, G. M. (2016). Data from two different culture conditions of *Thalassiosira weissflogii* diatom and from cleaning procedures for obtaining monodisperse nanostructured biosilica. *Data in Brief*, 8, 312–319. <https://doi.org/10.1016/j.dib.2016.05.033>
- Wali, A. F., Dhaheri, Y. Al, Pillai, J. R., Mushtaq, A., Rao, P. G. M., Rabbani, S. A., Firdous, A., Elshikh, M. S., & Al Farraj, D. A. (2020). Lc-ms phytochemical screening, in vitro antioxidant, antimicrobial and anticancer activity of microalgae *nannochloropsis oculata* extract. *Separations*, 7(4), 1–11. <https://doi.org/10.3390/separations7040054>
- Wan Rosli, W. I., Siti Nur Haffizah, R., & Nurrahana, H. (2018). Antioxidative and scavenging properties of polyphenolic rich-fraction of cornett's (Young zea mays). *International Journal of Recent Technology and Engineering*, 7(4), 5–8.
- Wang, E., Sorolla, M. A., Krishnan, P. D. G., & Sorolla, A. (2020). From seabed to bedside: A review on promising marine anticancer compounds. *Biomolecules*, 10(2). <https://doi.org/10.3390/biom10020248>
- Wang, W., Xiong, P., Zhang, H., Zhu, Q., Liao, C., & Jiang, G. (2021). Analysis, occurrence, toxicity and environmental health risks of synthetic phenolic antioxidants: A review. *Environmental Research*, 201(March), 111531. <https://doi.org/10.1016/j.envres.2021.111531>
- Wang, X., & Zhang, X. (2013). Separation, antitumor activities, and encapsulation of polypeptide from *Chlorella pyrenoidosa*. *Biotechnology Progress*, 29(3), 681–687. <https://doi.org/10.1002/btpr.1725>
- WHO. (2021). *Globocan 2020: Malaysia*. International Agency for Research on Cancer. <https://gco.iarc.fr/today/data/factsheets/populations/458-malaysia-fact-sheets.pdf>

- Widowati, I., Zainuri, M., Kusumaningrum, H. P., Susilowati, R., Hardivillier, Y., Leignel, V., Bourgougnon, N., & Mouget, J.-L. (2017). Antioxidant activity of three microalgae *Dunaliella salina*, *Tetraselmis chuii* and *Isochrysis galbana* clone Tahiti. *IOP Conference Series: Earth and Environmental Science*, 55, 012067. <https://doi.org/10.1088/1742-6596/755/1/011001>
- Yamada, T., & Sugimoto, K. (2016). Guggulsterone and its role in Chronic diseases. In *Advances in Experimental Medicine and Biology* (Vol. 929). [https://doi.org/10.1007/978-3-319-41342-6\\_15](https://doi.org/10.1007/978-3-319-41342-6_15)
- Yang, S., Wan, H., Wang, R., & Hao, D. (2019). Sulfated polysaccharides from *Phaeodactylum tricornutum*: Isolation, structural characteristics, and inhibiting HepG2 growth activity in vitro. *PeerJ*, 2019(2). <https://doi.org/10.7717/peerj.6409>
- Yasir, S., Siddiki, A., Mofijur, M., Kumar, P. S., Forruque, S., Chyuan, H., & Mahlia, T. M. I. (2022). Microalgae biomass as a sustainable source for biofuel, biochemical and biobased value-added products: An integrated biorefinery concept. *Fuel*, 307(August 2021), 121782. <https://doi.org/10.1016/j.fuel.2021.121782>
- Yen, H. W., Hu, I. C., Chen, C. Y., Ho, S. H., Lee, D. J., & Chang, J. S. (2013). Microalgae-based biorefinery - From biofuels to natural products. *Bioresource Technology*, 135, 166–174. <https://doi.org/10.1016/j.biortech.2012.10.099>
- Youn, H. J., & Han, W. (2020). A review of the epidemiology of breast cancer in Asia: Focus on risk factors. *Asian Pacific Journal of Cancer Prevention*, 21(4), 867–880. <https://doi.org/10.31557/APJCP.2020.21.4.867>
- Yu, C. C., Chen, H. W., Chen, M. J., Chang, Y. C., Chien, S. C., Kuo, Y. H., Yang, F. L., Wu, S. H., Chen, J., Yu, H. H., & Chao, L. K. P. (2010). Chemical composition and bioactivities of the marine alga *isochrysis galbana* from Taiwan. *Natural Product Communications*, 5(12), 1941–1944. <https://doi.org/10.1177/1934578x1000501222>
- Yusoh, N. A., Leong, S. W., Chia, S. L., Harun, S. N., Rahman, M. B. A., Vallis, K. A., Gill, M. R., & Ahmad, H. (2020). Metallointercalator [Ru(dppz)<sub>2</sub>(PIP)]<sup>2+</sup> Renders BRCA Wild-Type Triple-Negative Breast Cancer Cells Hypersensitive to PARP Inhibition. *ACS Chemical Biology*, 15(2), 378–387. <https://doi.org/10.1021/acscchembio.9b00843>
- Zainoddin, H. A. H., Hamzah, A., Jamari, Z., & Omar, W. A. W. (2018). Chemical profiles of methanolic extracts from two species of microalgae, *nannochloropsis* sp. and *Spirulina* sp. *Pertanika Journal of Tropical Agricultural Science*, 41(3), 1085–1096.
- Zhang, J., Liu, L., & Chen, F. (2019). Production and characterization of exopolysaccharides from *Chlorella zofingiensis* and *Chlorella vulgaris* with anti-colorectal cancer activity. *International Journal of Biological*

*Macromolecules*, 134, 976–983. <https://doi.org/10.1016/j.ijbiomac.2019.05.117>

Zhang, J., Liu, L., Ren, Y., & Chen, F. (2019a). Characterization of exopolysaccharides produced by microalgae with antitumor activity on human colon cancer cells. *International Journal of Biological Macromolecules*, 128, 761–767. <https://doi.org/10.1016/j.ijbiomac.2019.02.009>

Zhang, J., Liu, L., Ren, Y., & Chen, F. (2019b). Characterization of exopolysaccharides produced by microalgae with antitumor activity on human colon cancer cells. *International Journal of Biological Macromolecules*, 128, 761–767. <https://doi.org/10.1016/j.ijbiomac.2019.02.009>

Zhuang, D., He, N., Khoo, K. S., Ng, E. P., Chew, K. W., & Ling, T. C. (2022). Application progress of bioactive compounds in microalgae on pharmaceutical and cosmetics. *Chemosphere*, 291(P2), 132932. <https://doi.org/10.1016/j.chemosphere.2021.132932>

Zuo, W., & Kwok, H. F. (2021). Development of Marine-Derived Compounds for Cancer Therapy. *Marine Drugs*, 19(6), 342. <https://doi.org/10.3390/md19060342>